



OPEN ACCESS

EDITED BY

Mangqian Cha,
Chinese Academy of Agricultural Sciences
(CAAS), China

REVIEWED BY

Chengzeng Luo,
Chinese Academy of Agricultural Sciences
(CAAS), China
Zhigao An,
Zhejiang A&F University, China

*CORRESPONDENCE

Jian Ma

✉ crazyma0411@163.com

RECEIVED 06 February 2025

ACCEPTED 10 March 2025

PUBLISHED 26 March 2025

CITATION

Li T, Raja BR, Liao J, Zheng L, Yin F, Gan S,
Sun X, Lyu G and Ma J (2025) The
characteristics, influence factors, and
regulatory strategies of growth retardation in
ruminants: a review.
Front. Vet. Sci. 12:1566427.
doi: 10.3389/fvets.2025.1566427

COPYRIGHT

© 2025 Li, Raja, Liao, Zheng, Yin, Gan, Sun,
Lyu and Ma. This is an open-access article
distributed under the terms of the [Creative
Commons Attribution License \(CC BY\)](#). The
use, distribution or reproduction in other
forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in
this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

The characteristics, influence factors, and regulatory strategies of growth retardation in ruminants: a review

Tao Li¹, Bakhtawar Riaz Raja¹, Jie Liao¹, Longqing Zheng¹,
Fuquan Yin¹, Shangquan Gan¹, Xuemei Sun², Gang Lyu² and
Jian Ma^{1,2*}

¹College of Coastal Agricultural Sciences, Guangdong Ocean University, Zhanjiang, China, ²Xinjiang Taikun Group Co., Ltd., Changji, China

Growth retardation represents a main barrier to affect the productivity and efficiency of ruminants production, which is characterized by low growth rate, a disparity between skeletal and physiological maturation, gastrointestinal dysfunction and reduced reproductive performance. This review provides a concise overview of growth retardation in ruminants, and summarizes the key factors that influence their growth and development, including genetics, nutrition, microbiota and environment. Also, this review emphasizes the central role of nutritional management and gastrointestinal development, as well as the regulatory mechanisms involved in growth processes. In addition, recent advances in these aspects are discussed to form an integrative framework aimed at improving physiological function in ruminants. This review provides a comprehensive perspective for understanding the complex mechanism of growth retardation in ruminants, puts forward a theoretical basis for optimizing the production efficiency of ruminants industry and emphasizes the importance of multidisciplinary collaboration to provide a reference for advancing systematic research on growth and development of ruminants.

KEYWORDS

ruminant, growth retardation, nutritional regulation, oxidative stress, gastrointestinal function

1 Introduction

Growth retardation has been a critical issue in livestock production, which affects animal welfare, reduces productivity and economic benefits and restricts the development of ruminants industry (1). A common strategy to enhance growth performance of animals is to increase the energy content in the feed. Huo et al. (2) reported that cows fed high-energy diets exhibited approximately a 30% higher average daily weight gain compared to those fed low-energy diets. The interaction between genetics and nutrition has become a central focus of research as recent advances in molecular biology. Genetics and nutrition have improved our understanding of mechanisms involved in ruminants growth. A previous study has shown that the growth and development of ruminants were not only affected by genetic, nutritional and environmental factors, but also the genetic modification of nutrients was an effective strategy to regulate the growth and development of ruminants (3). The main causes of growth retardation in animals are nutritional deficiencies, particularly in protein, energy, vitamins, and

minerals. These deficiencies highlight the need for nutritional interventions as they directly affect growth rate and body condition. Specific nutrients supplementation can improve growth performance and immune resistance in ruminants. However, the optimal nutritional programs can not completely mitigate growth retardation, because the healthy development of gastrointestinal tracts is the premise of normal growth and development of animals. The rumen, a unique digestive organ in ruminants, is critical for the digestion of high-fiber diets. Meanwhile, the gut plays a dual role in nutrients absorption and immune support (1). Although recent progress has been made in the regulation of growth in ruminants, most studies mainly focus on the effects of individual nutrients and feed formulations, and often these factors are analyzed in isolation from each other. The comprehensive analysis of the interaction in multiple regulatory measures remains scarce. Therefore, on the basis of published literature and our previous studies, this review primarily summarized recent findings in growth retardation of ruminants, concluded the influencing factors and mechanisms involved in the process of growth retardation and presented a systematic framework for alleviating the ruminants' growth retardation.

2 Comprehensive insights into growth and development of ruminants

From the fertilized egg through embryonic, juvenile, and adult stages to senescence and death, individual growth and development is a lifelong process for animals. Throughout this life cycle, to support the ongoing development and functional optimization of tissues and organs, animals continuously accumulate nutrients and perform metabolic activities to maintain a dynamic balance in cell number and size. This process enables a continuous increase in body weight (BW) until the animal reaches its mature adult weight (4, 5). The growth and development of animals can be broadly divided into two phases including prenatal and postnatal stages. Individual growth status is commonly assessed through periodic measurements, which are conducted by observing age-related changes in overall weight, specific body parts, organs and tissues. The most obvious sign of growth is the continuous rise in BW with age, which is usually in the form of a distinctive "S". Figure 1 shows a theoretical representation of growth rate curves over different time periods of animals. From birth to adolescence, animals grow rapidly and their body weight increases rapidly. From adolescence to adulthood, the growth rate of animals is slow, and their body weight remains stable in adulthood until death, when there is a slight decrease in body weight (6).

The interaction of genotype and environmental factors results in growth and development. Normal growth and development, with characteristic patterns of weight gain as well as unbalanced, non-isokinetic and sequential changes in body, are regulated by a balanced combination of nutrients, hormones, enzymes and a healthy physiological state (7). Genetic mutations, nutritional deficiencies, hormonal or enzymatic imbalances and drastic environmental changes often result in animals with BW or size that deviate from the population average for their breed and age. Unfortunately, the concept of growth retardation in ruminants remains undefined. In pigs research, Jones et al. (8) compared the

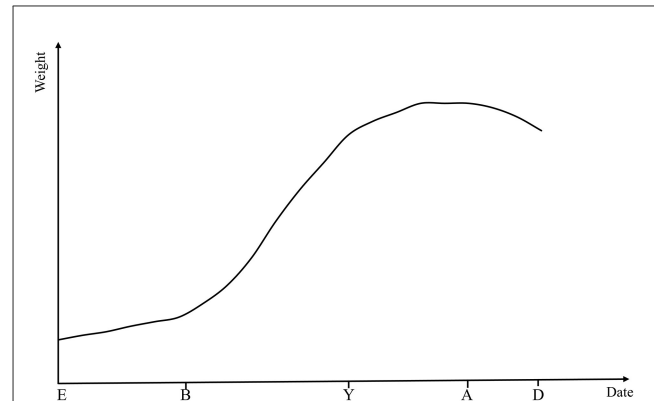


FIGURE 1
Theoretical growth rate curves across key stages of animal development. E, embryo; B, birth; Y, young; A, adult; D, death.

physiological indexes between growth-retarded and normal pigs, and defined that pigs with BW in the lowest 10% were deemed to be growth retardation within the same breed and age. In human medicine, the World Health Organization classifies infants with a weight Z-score reduction >1.0 as growth retardation, defining it as a BW below the 5% normal infants or a growth rate two standard deviations below the norm (9, 10). In our previous studies of growth-retarded yaks, growth retardation referred to individuals whose growth performance is much lower than the average level under the condition of the same breed and age, which were mainly manifested as low BW, high mortality and reduced feed efficiency and immunity (11). Accordingly, in this review paper, in combination of relevant studies, growth retardation is defined as BW that the BW of one ruminant with the same breed and age is in the lowest 10% of population under the same environment and feeding conditions.

3 The characteristics of growth retardation in ruminants

3.1 Reduced weight gain and fat deposition ability

The weight gain rate is a key parameter to evaluate the production performance and health status of animals. Compared with normally developing animals, the growth-retarded ruminants typically have a lower BW, slower weight gain, poor muscle development, and limited fat deposition. Underweight mainly includes low birth weight and postnatal growth retardation due to genetic factor or nutrient deficiency, which results in a weight of $<10\%$ of the similar age (1). These characteristics have a direct impact on the final production efficiency and economic value of animals. As a central driver of glucose and lipid metabolism imbalances, adipose tissue dysfunction is closely linked to metabolic syndrome. Studies in growth-retarded animal models revealed that diminished fat deposition promotes pathogenic lipid redistribution, which exacerbates metabolic risks through mechanisms such as chronic inflammation (12). Nutritional

deficiencies, including inadequate maternal nutrition or placental dysfunction leading to intrauterine growth retardation (IGR), are a major cause of low birth weight in ruminants. The IGR severely affects survival rate of ruminants after birth and can cause irreversible damage to subsequent production performance (13). Research of Zhang et al. (14) showed that placental dysfunction can reduce the efficiency of nutrients transport from the mother to the fetus. Consequently, IGR lambs had a significantly lower initial and final BW as well as reduced net weight gain and average daily gain when compared to lambs with normal intrauterine growth. Sutherland et al. (15) also reported that IGR had irreversible effects on normal growth trajectory, with IGR lambs having significantly lower BW after 127 days of birth. Early rearing ways, especially when the supply of essential nutrients for maintenance and growth is inadequate, are critical in determining later growth performance. Under such conditions, animals are unable to effectively synthesize the nutrients required for the formation of body tissues (16). In most animals, the weight at puberty is typically ~60% of the weight at maturity (7). A previous research by our group has demonstrated that the average daily gain and final BW of growth-retarded yaks were lower than those of normal yaks under same feeding condition (17). Inadequate management of ruminants causes slow weight gain and underdeveloped skeleton and muscles, especially thin subcutaneous fat layers on the back, abdomen and hip, which finally can lead to a long time to reach market weight.

3.2 Discrepancy in body size, bone age, and physiological maturity

The body size is an important index to assess the growth and development of animals, which indirectly reflects the developmental situation of organs and tissues. In ruminants production, the measurement of body size can be used for the selection of breeding stock and adjustment of feeding strategy. Normal ruminants have unbalanced and uneven growth characteristics. They generally grow from head to tail, then tail to head, and finally meet at the waist. In addition, until achievement of a coordinated balance, limb growth usually precedes trunk growth (14, 18). A mismatch between bone maturity and physiological age is often found in growth-retarded ruminants, which typically have shorter body length, smaller chest circumference and lower body height. Wang et al. (19) found that compared with normal yaks, the growth-retarded yaks had significantly lower body height, length, chest circumference and depth, as well as lower mean daily growth rate of body size. These reductions in skeletal dimensions are primarily attributed to growth hormone (GH) and insulin-like growth factor-1 (IGF-1) deficiencies, which impair chondrocyte proliferation in the growth plates of long bones (20, 21). A shorter body length is often correlated with delayed or poor skeletal development. A smaller chest circumference may indicate underdeveloped visceral organs (e.g., limited cardiopulmonary function), and a lower body height may indicate abnormal limb bone development (22). This imbalance has a negative impact on overall health, production performance, and economic benefit of ruminants. Research has shown that abnormal hormone secretion, malnutrition and genetic mutations, which collectively affect

growth and development of animals, often contribute to the mismatch between bone maturity and physiological age (7).

3.3 Digestive system maldevelopment

Digestive tract health is influenced by macroscopic aspects of development and physiological integrity, as well as the microscopic balance between gut microbiota and immune status (23). Severe malnutrition in the early stage of life can damage the structure and function of the gastrointestinal tracts, leading to growth retardation in ruminants, which in turn limits the development of the digestive system (24). Study showed that growth-retarded yaks had a lower weight of rumen and intestine when compared to that of normal yaks (25). The rumen, a unique digestive organ of ruminants, has important function in digesting crude fiber. Ruminal microorganisms and enzymes produced by microorganisms can digest fiber into nutrients that can be utilized by the body, which has significant effects on growth of ruminants (26). In addition, the ruminal epithelium has a vital role in the absorption of volatile fatty acids (VFAs), and its healthy development is crucial for nitrogen transport and urea recycling (27). The height and width of rumen papillae are the important parameters to evaluate the relative development of ruminal epithelium. Our previous research found that compared with normal yaks, the growth-retarded yaks had a significantly lower ruminal papilla height and jejunal villus height, width, surface area and villus/crypt ratio (17). Moreover, another study in sheep has also shown that the small intestinal villus height and villus/crypt ratio of growth-retarded lambs were lower than those of normal lambs, suggesting that the gastrointestinal tracts of normal ruminants had a greater absorptive area and efficiency to absorb the nutrients, and then meet growth and production demands (28). Rumen dysgenesis can impair barrier function by reducing the expression of tight junction proteins (e.g., ZO-1 and Occludin) and increasing the risk of endotoxin and pathogens invasion (1, 23, 29). As a result, animals with growth retardation become more susceptible to microbial disturbances and inflammation, further hindering nutrient digestion and absorption.

Microbiota in the digestive system of animals are essential for growth, health and productivity and interact symbiotically with the host. Ruminal microbiota serve as the primary site for digestion and absorption of VFAs and excess ammonia nitrogen, while gut microbiota absorb nutrients that are not absorbed in the rumen and play a critical role in the immune system (30, 31). Underdeveloped digestive systems of ruminants is usually accompanied by a series of physiological function disorders, including imbalance of gastrointestinal microbiota. Then, some beneficial microorganisms can not be effectively established and maintained, leading to susceptibility to digestive diseases, reduced energy supply, impaired nutrient absorption, and ultimately inhibiting the growth performance of ruminants. Zhuang et al. (32) isolated a strain of *Bifidobacterium longum* (*B. longum* 1109) from calf feces using phenotype clustering analysis and strain isolation techniques, and mouse experiments demonstrated that this strain can promote the growth of young livestock by modulating microbial function and host metabolism, suggesting its potential application in animal

nutrition. Du et al. (33) found that growth-retarded calves had a lower relative abundances of *Proteobacteria*, *Rhodospirillaceae*, *Campylobacter*, and *Butyrivimonas*, which played an important role in energy and VFAs production, in the feces when compared to normal calves, whereas the pathogenic bacteria such as *Anaeroplasma* and *Eubacterium* were more abundant in growth-retarded calves. Our previous findings also showed that the relative abundances of bacteria involved in oligosaccharide, starch and cellulose degradation (e.g., *Ruminococcaceae*, *Spirochaetaceae*, *Clostridiaceae*, *Prevotellaceae*, and *Rikenellaceae*) were higher in normal yaks than those in growth-retarded yaks, while the *Christensenellaceae_R-7*, unclassified *Chitinophagaceae* and *Eubacterium tenue* were significantly higher in growth-retarded yaks (34). These results suggest that growth-retarded ruminants have a disrupted gastrointestinal microbiome, reduced gut immunity and nutrient utilization efficiency and increased susceptibility to digestive diseases such as diarrhea or enteritis, which further affects feed efficiency and growth rate.

3.4 Reduction of reproductive performance

The imbalance between body size and physiological development leads to a delayed sexual maturity and reduced reproductive performance. Many factors, including gene, nutritional status, environmental conditions and disease, can influence the reproductive performance of ruminants. Reproductive performance is regulated by the hypothalamic-pituitary-gonadal (HPG) axis, which controls gonadal function by regulating the secretion of gonadotropins, including the luteinizing hormone and follicle-stimulating hormone levels, thereby affecting follicular development, ovulation and spermatogenesis. The HPG axis is the core of the endocrine mechanisms that control the reproductive performance of ruminants and other mammals (35, 36). Growth-retarded animals often exhibit inadequate secretion of IGF-1 and GH, which affect reproductive performance together with the HPG axis. A previous study has shown that IGF-1 has a critical role in mammary gland development and milk synthesis, particularly in the regulation of fat and protein content (37). Ruminants with growth retardation have abnormal BW and reduced milk quality, which significantly affect offspring survival rates, which indirectly leads to a reduction in reproductive performance. In addition, *in vivo* studies have shown that IGF-1 and GH can directly act on the regulation and growth of bovine follicles at different stages, thus promoting follicular development (38, 39). Hormonal imbalances severely affect overall reproductive efficiency, leading to delayed sexual maturity, irregular estrous cycles, reduced embryo survival after conception and increased risk of abortion. The GH also regulates the development and metabolism of the male gonads through growth-promoting effects. Males with growth retardation tend to have smaller body size and underdeveloped gonads compared to normal females, which result in mating difficulty. IGF-1 protects sperm cells from oxidative stress through the antioxidant properties (40, 41). Kumar et al. (42) found that IGF-1 can increase the stability of the structure and function of buffalo sperm after cryopreservation, which significantly improved the fertilization ability of the sperm. This

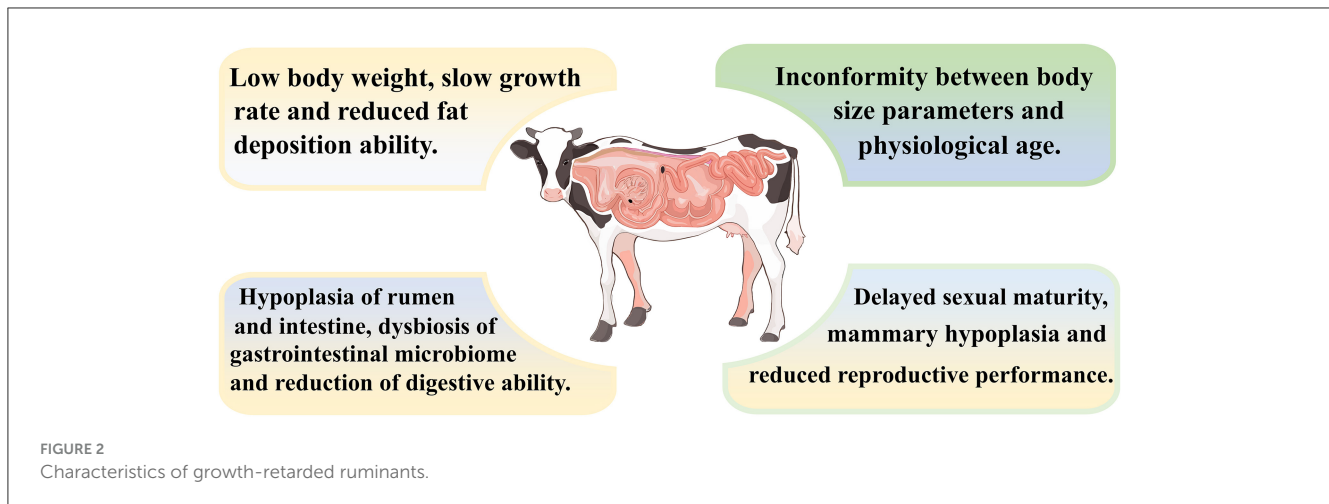
finding suggested that IGF-1 increased the activity of antioxidant enzymes in sperm cells, reduced the accumulation of free radicals, improved the viability of sperm and enhanced the reproductive performance of the male. In conclusion, the decline in reproductive rates in growth-retarded ruminants is primarily due to hormonal imbalances involving IGF-1, GH and the HPG axis, resulting in poor development of oocyte, sperm and gonad.

Overall, as shown in Figure 2, ruminants with growth retardation exhibit characteristics such as low body weight, slow growth rate and reduced fat deposition ability. In addition, the rumen and intestine of growth-retarded ruminants are dysplastic, and the nutrients digestion and absorption are decreased.

4 Key factors contributing to growth retardation in ruminants

4.1 Genetic determinants of growth in ruminants

Ultimately, gene expression and regulatory changes control the growth and development as well as metabolic processes (43). At the molecular level, a large number of genes and their associated signaling pathways play a critical role in cell proliferation, differentiation, nutrient metabolism and endocrine regulation during the growth of animals. The growth and development of animals will be significantly affected by abnormal mutations or expression of certain genes. Hu et al. (25) analyzed the effects of growth hormone-releasing hormone (GHRH) in the pituitary gland and the levels of GH and insulin-like IGF-1 in the blood between growth-retarded and normal yaks, and found a significant correlation between IGF-1 concentration and BW gain. Similarly, study by Saleh et al. (44) showed that the insulin-like hormone IGF-1 helped cells grow and differentiate, and it was also a regulator of cell development and DNA synthesis. GH is secreted by the pituitary gland and stimulates the liver to produce IGF-1. Then, IGF-1 binds to its receptor IGF1R, which activates the downstream PI3K/Akt and MAPK signaling pathways to promote cell proliferation and growth (45, 46). Li et al. (47) has shown that the growth of animals muscle fiber occurred primarily through protein deposition, which promoted muscle fiber hypertrophy, and IGF-1 stimulated protein synthesis via the PI3K/Akt/mTOR signaling pathway to increase muscle fiber cross-sectional area and promote muscle hypertrophy, while IGF-1 level decline inhibited this process. Meanwhile, Hu et al. (25) showed that injection of GH releasing peptide-2 (GHRP-2) into growth-retarded yaks significantly increased serum levels of GH and IGF-1, which not only improved the muscle fiber morphology of the longissimus dorsi and semitendinosus muscles, but also upregulated the expressions of PI3K, Akt, and mTOR mRNA, finally contributing to improved growth of growth-retarded yaks. Another experiment has also verified that long-term utilization of GHRP-2 significantly increased daily weight gain in Holstein cows, with increase of up to 36.4% (48). GHRP-2 is a GH secretinotropin peptide, and GH is the natural ligand of GHRP, which can act on internal receptors and stimulate the secretion of GH and IGF-1 (49). On the other hand, IGF-1 promotes muscle protein synthesis by activating the PI3K/Akt/mTOR pathways and increasing the phosphorylation of



4E binding protein (4EBP-1) and S6 kinase (S6K1) (50). Therefore, the abnormal secretion of GH and IGF-1 hormones is an important factor to induce growth retardation in ruminants, which not only causes slow weight growth and body size, but also indirectly results in the reproductive performance to be reduced.

In addition to the significant effects of GH and IGF-1 on cell proliferation and protein synthesis, other genes play a critical role in the regulation of ruminal development and skeletal growth, thereby affecting the growth of animals. Meng et al. (51) study found that gene knockout of the G-protein coupled receptor 41 (GPR41) in cattle significantly inhibited the ruminal epithelial cell proliferation and reduced the cyclin D2 and cyclin E2 levels. Ruminal cell proliferation has an important function in maintaining rumen homeostasis, enhancing rumen barrier and promoting nutrient digestion and absorption in ruminants. Additionally, myostatin (MSTN), a negative regulator of muscle growth, has been associated with skeletal size variation. Its inactivation significantly alters the gut microbiota composition in sheep, enhancing growth without compromising meat quality (52). Liu et al. (53) analyzed Notch1 gene polymorphism in Qinchuan cattle and found that five novel SNP loci (g.A48250G, g.A49068G, g.A49239C, g.C49307T, g.C49343A) were significantly associated with growth traits including body height, weight and hip height, which suggested that Notch1 can be used as a candidate gene for beef cattle breeding program. Kong et al. (54) investigated the regulatory role of long non-coding RNAs (lncRNAs) in the liver of Leiqiong cattle and identified differential expression of 1,124 mRNAs and 24 lncRNAs between growth-retarded and normal Leiqiong cattle. The differentially expressed mRNAs were not only associated with growth factor binding, K63-linked ubiquitination, and cellular protein metabolism, but also significantly correlated with growth and development pathways such as the PPAR signaling pathway and vitamin B6, glyoxylate and dicarboxylate metabolism. Among the lncRNAs, Lnc_002583 was found to play a synergistic regulatory role in the growth and development of Leiqiong cattle by influencing the expression of IFI44 and IFI44L genes. Yang et al. (55) identified CircDyrk1A in the muscle tissue of beef cattle and demonstrated its role in muscle development. Further analysis revealed that the CircDyrk1A-miR-21-5p-KLF5 axis regulates myoblast differentiation, providing a theoretical basis

for genetic improvement in beef cattle breeding. In a recent study, genome-wide association analysis and quantitative trait loci mapping in sheep were used to identify MAP3K1, ANKRD55, ABCB1, MEF2C, and TRNAW-CCA-8 as key regulators of growth and development. These five candidate genes were significantly associated with BW traits in sheep, offering valuable insights into the genetic mechanisms underlying BW variation and their potential applications in selective breeding (56). In addition, studies have reported that fibroblast growth factors (FGFs) are critical in regulating bone and cartilage development in animals. Mutations in FGFs can cause cartilage developmental disorder of animals, resulting in skeletal maldevelopment which were manifested as dwarfism and growth retardation (57, 58).

To sum up, genetic factors significantly influence the healthy growth of ruminants. The interplay of hormones and genes, together with environmental and nutritional factors, regulate gene expression through epigenetic mechanisms, ultimately affecting individual growth and development.

4.2 Nutritional influences on growth and development of ruminants

4.2.1 Disparity between nutrient supply and growth demand in ruminants

Nutrient imbalance refers to a situation where the nutrients consumed by animals do not meet their actual physiological requirements during the rearing process. This imbalance has a direct impact on health and production performance of animals. During the growth and development of ruminants, the barrier of nutrient transfer between dam and offspring, as well as interaction between different nutrient components in the diet, are major contributors to the mismatch of nutrient supply and demand.

4.2.1.1 Barrier of nutrient transfer from dam to offspring

Pregnancy and lactation in dams are critical for the growth and development of their offspring. During pregnancy, placental dysfunction is a major cause of fetal malnutrition, which not only reduces survival rate of offspring, but also affects the economic benefits of livestock production (59). A previous research has

shown that placental dysfunction can slow blood flow in the uterus and umbilical cord, impair the transport of oxygen and amino acids across the placenta, and reduce the concentration of growth factors involved in metabolic processes in the fetal circulation, ultimately limiting the fetal growth and development. This dysfunction is a primary factor responsible for fetal growth deviating from genetic trajectories and preventing the achievement of genetically determined growth potential (60, 61).

In sheep, trophoblast cells invasion and differentiation during embryo implantation are critical for placentation and fetal development. Study has shown that the chemokine CXCL12 regulated the placental implantation and growth through its receptor CXCR4, and inhibition of the CXCL12/CXCR4 pathway increased the genes expression of fetal FGF2, vascular endothelial growth factor (VEGF-A) and placental growth factor (PLGF), which subsequently tended to increase the placental weight and the number of placental clusters (62). The increase in the weight and number of placentas can increase the flow of blood through the umbilical cord and enhance the expression of placental glucose and amino acid transporters, which is conducive to improving the delivery of maternal oxygen and nutrients to the fetus (63, 64). Moreover, Britt et al. (65) fed pregnant ewes with tall fescue seeds infected by endophytes and analyzed placental gene expression, and results showed that endophyte infection altered the expression of microRNA-21 in the placenta. MicroRNA-21 is a potential biomarker of fetal hypoxia and its up-regulated expression is associated with placental pathology, including placental insufficiency and fetal growth restriction. During fetal development, skeletal muscle development is highly sensitive to nutrients supply. Inadequate nutrients supply will reduce the number of skeletal muscle fibers and alter fiber composition, which then affect fetal fat deposition and production performance (66, 67). This is because that the muscle fibers of animals are derived from myoblasts proliferation and differentiation in early fetal development. A decrease in the effectiveness of nutrients and growth factors impairs the periodic activity of myoblasts, especially inhibition in the function of leucine transporters that uniquely regulate muscle protein synthesis, thereby reducing the number of nucleus of each muscle fiber, resulting in fewer muscle fibers in the offspring, and ultimately affecting normal growth and development (68–72).

The genetic background of animals determines its specific nutrient requirements, metabolic pathways and absorption efficiency. Nutrition, in turn, can regulate gene expression, thereby influencing growth and reproductive performance of animals. The interaction between genetics and nutrition is essential to support growth and development of animals (73). Li et al. (74) highlighted that maternal supplementation with fish oil during late pregnancy in mammals can enhance the expression of the GPR120 signaling pathway, improve placental nutrient transport efficiency and promote the growth and development of offspring. In addition, studies have shown that fat deposition in animals was also regulated by the Wnt signaling pathway. During muscle formation in ruminants, Wnt signaling will be enhanced and the activity of PPAR γ is regulated by the Wnt pathway via GSK-3 β and β -catenin. When maternal nutrient supply is inadequate or placental function is compromised, GSK-3 β activity will be

reduced to cause proteolysis of β -catenin and suppression of PPAR γ target gene expression, which affects muscle growth and fat deposition, resulting in lower birth weight and intrauterine growth retardation (75–77).

The growth and development of the offspring are directly influenced by the quality and quantity of the maternal milk. Milk is not only rich in proteins, fats, minerals, and other nutrients that provide the energy and necessary nutrients for the growth and development of neonate, but also contains several immune factors, such as immunoglobulins, lactoferrin and lysozymes, that boost the newborn immunity and protect it from disease (78). Among the polyunsaturated fatty acids, arachidonic acid (AA), eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA), and docosahexaenoic acid (DHA) are the highest concentrations in colostrum, which is attributed to that AA and DHA are components of neurophospholipid membranes and critical for the development of the central nervous system and vision (79). Therefore, the high content of unsaturated fatty acids in colostrum is essential for the health, growth and development of ruminants in the later stages. Jagusch and Mitchell (80) studied the effect of lambs on the utilization of metabolic energy from maternal milk by using a comparative slaughter method, and results showed that lambs fed the lowest level of nutrition mobilized body fat and had a negative energy balance, besides, the rate of weight gain of these lambs was lower than that of lambs in a positive energy balance. After weaning, the nutrients that satisfy the growth and development of ruminants mainly come from feed. Carbohydrates in the feed are fermented in the rumen to produce VFAs, which provide energy for the body. Proteins are decomposed into amino acids and ammonia by microorganisms to provide raw materials for synthetic proteins and microbial proteins. Trace elements and minerals play an important role in the regulation of GHs and enzymes. In the cattle research of Sakase et al. (81) feeding a high-energy diet to calves after birth increased the concentration of insulin-like peptide 3 (INSL3), which in turn increased IGF-1 levels and promoted growth and muscle development.

Although numerous nutrition studies suggested that high-energy diets can regulate physiological status, promote growth and development, increase meat and milk production and improve meat quality (82, 83). Ford et al. (84) claimed that in growth-retarded lambs, there was still an imbalance between muscle and fat deposition, even under a high energy diet condition, and besides, at 9 months of age, the ratio of muscle to fat of lambs was reduced by 25%, leading to maldevelopment of the body. This is because ruminants with growth retardation, due to early nutritional deficiencies, reorganize and redistribute the nutrients, resulting in restricted expression of relevant genes or pathways, which results in incomplete organ or tissue development, such as an underdeveloped rumen and small intestine, and reduces nutrient absorption efficiency (1).

4.2.1.2 Effects of nutrient interactions on growth of ruminants

Nutrient balance, with different nutrients interacting synergistically or antagonistically, is critical to growth and development of animals. These interactions have a direct impact on the efficiency with which animals utilize the nutrients in

the feed, ultimately affecting growth, health and productivity of animals. In ruminants, the interaction between energy and protein is particularly important. Insufficient fiber intake leads to a decrease in rumination and salivary secretion, which reduce the buffering capacity of the rumen and may induce ruminal acidosis. In addition, the change in ruminal pH can weaken the function of ruminal microorganisms, inhibit the activity of fiber-degrading bacteria and reduce feed digestibility (85). Consequently, insufficient energy intake causes the body's reserves of fat and protein to be metabolized into energy, which weakens their role in tissue construction and repair, suppressing growth and development of ruminants. As a key nutrient for maintaining health and production performance, protein is essential for growth, tissue repair and maintenance of physiological functions of animals (86). The digestion and absorption of protein occurs primarily in the form of amino acids and small peptides. Amino acids are classified as essential (EAA) or non-essential amino acids (NEAA) according to whether animals can synthesize them. The synergistic interactions between EAAs and NEAA are critical for maintaining protein synthesis and metabolic balance. In addition, interactions of other nutrients not only affect the bioavailability and metabolic pathways of the feed, but also play a key role in regulating overall health and productivity of animals. Table 1 shows the interactions between different amino acids and other nutrients.

4.2.2 Impaired nutrients absorption and metabolism in ruminants

The VFAs produced by microbial fermentation in the rumen can be absorbed directly by the ruminal epithelial cells, providing ~80% of the energy requirements for ruminants (87). Additionally, the microbial proteins that are synthesized in the rumen can also reach the small intestine, where they are absorbed and utilized (88). If the rumen is underdeveloped, the colonization and function of the ruminal microbiota will be impaired, leading to difficulty in the digestion of complex carbohydrates such as cellulose or ruminal acidosis (34). Propionate, a fermentation product of the ruminal microbiota, can stimulate free fatty acid receptors in the colonic epithelial L-cells and promote the secretion of GLP-1. GLP-1, in turn, transmits signals to the hypothalamus that affect the appetite (89). Numerous studies have shown that the maturation of ruminal microorganisms, such as fiber (e.g., *Ruminococcus flavefaciens*, *Fibrobacter succinogenes*), starch (e.g., *Ruminobacter amylophilus*, *Streptococcus bovis*) and protein-degrading bacteria (e.g., *Prevotella ruminicola*, *Butyrivibrio fibrisolvens*), directly affect the digestibility of feed, thereby influencing the growth and health of ruminants (90–92). Furthermore, a previous study has shown that ruminal acidosis altered the permeability of ruminal epithelial cells and damaged the ruminal barrier, allowing endotoxins to enter the bloodstream, triggering systemic inflammation and reducing growth efficiency of ruminants (93). The ruminal epithelial cells play a vital role in regulating nutrients absorption and preventing harmful substances from crossing the epithelium into the bloodstream. Previously, a study suggested that the ruminal epithelial cells absorb VFAs via the monocarboxylate transporter (MCT1), which works with protons to facilitate transmembrane transport, thereby supporting cellular energy

supply (94). Simultaneously, ruminal epithelial barrier function is maintained by tight junctions to ensure dynamic homeostasis between ruminal contents and cells and prevent the invasion of harmful pathogens. The changes in ruminal pH and nutritional status can damage the structural integrity of tight junctions, which lead to a decrease in the expression of ZO-1, occludin and claudin-1, thereby compromising epithelial barrier function (95). The dynamic balance between the external environment and the physiological metabolism of body is therefore closely linked and plays an essential role in maintaining health and growth of animals.

In ruminants, the normal microbiota in the intestine helps to metabolize large molecules from external and endogenous sources, such as rumen-protected proteins, microbial proteins, vitamins and minerals. A well-developed gut has tight cell junctions and a robust immune barrier that effectively prevents the invasion of pathogens and harmful substances (34, 96). However, if the gut is underdeveloped, these barriers can be compromised, increasing the possibility of bacterial, viral and parasitic infections and leading to gastrointestinal diseases such as enteritis and diarrhea (23). Previous research from our team compared the gastrointestinal microbiota of growth-retarded and normal yaks by using 16S rRNA gene sequencing, and principal coordinate analysis revealed significant differences in bacterial composition across all gastrointestinal segments. The normal yaks had a higher relative abundances of bacteria associated with oligosaccharide, starch and cellulose degradation when compared to the growth-retarded yaks (34). In addition, we observed that the villus height and crypt area in the intestine of growth-retarded yaks were significantly lower than those of normal yaks, while the expression levels of inflammatory factors such as tumor necrosis factor were significantly higher in growth-retarded yaks, indicating that the gastrointestinal barrier function was impaired in growth-retarded yaks (17). Balancing the gut microbiota and improving gut barrier function may be effective solutions to promote growth in growth-retarded ruminants. Previous studies found that the gut microbiota can promote the differentiation of immune cells through regulatory T cells and Th17 cells, and disruption of the gut microbiota can lead to bacterial translocation and impaired gut barrier function, which in turn affected overall health of animals (97, 98). Therefore, improving the ruminal and intestinal health are essential to increase nutrients utilization efficiency and promote growth in ruminants. In production, strategies such as optimizing feed formulations and using probiotics can help maintain microbial balance and gut health, thereby improving growth, development and health of ruminants.

4.3 Role of immunity and oxidative stress in health and growth of ruminants

Healthy problems, including chronic disease, parasitic infection, immune dysfunction and oxidative stress, induce the disturbance of energy balance and nutrient metabolism, ultimately affecting growth rate, weight gain, and mortality (99, 100). Chronic disease (e.g., rumenitis, enteritis, and pneumonia) trigger a sustained immune response that consumes significant energy reserves to maintain immune function, thereby reducing

TABLE 1 Interactions among amino acids and various nutrients.

Items	Nutrients	Interacting nutrients	Function type physiological	Physiological function	References
Amino acid	Lysine	Arginine	Antagonistic effect ^a	Affects protein synthesis and causes growth retardation and low immunity	(144, 145)
		Methionine	Synergistic effect	Improve growth rate and feed conversion rate	(146)
		Threonine	Synergistic effect	Participate in immunity and protein synthesis to support intestinal health	(147)
	Methionine	Cysteine	Synergistic effect	Jointly participate in the synthesis of glutathione and resist oxidative stress	(148)
	Glycine	Serine	Synergistic effect	Synthesis of precursors for other amino acids and biomolecules, involved in the synthesis of creatine, purine, and other important molecules	(1, 149, 150)
	Leucine	Isoleucine Valine	Synergistic effect	Participate in the synthesis of muscle proteins and energy metabolism, balance three amino acids to promote muscle growth and prevent muscle breakdown	(151, 152)
Vitamin	Vitamin A	Zinc	Synergistic effect	Zinc can enhance the utilization of vitamin A and improve immunity	(153)
	Vitamin E	Selenium	Synergistic effect	Jointly participating in the antioxidant defense system can effectively reduce oxidative damage to cell membranes and prevent muscle diseases	(129)
Mineral	Calcium	Phosphorus	Synergistic effect	Support bone development and prevent osteoporosis	(154)
	Magnesium	Potassium	Synergistic effect	Participate in the regulation of neuromuscular function, muscle contraction, and maintenance of cardiac function	(154)
	Sulfur	Selenium	Synergistic effect	Jointly acting on antioxidant defense mechanisms, such as the synthesis of glutathione peroxidase, to combat oxidative stress	(154)
	Sodium	Chlorine	Synergistic effect	Jointly maintain fluid electrolyte balance, regulate blood pressure and acid-base balance, and ensure normal neuromuscular function	(154)
Fatty acid	n-3	n-6	Antagonistic effect ^b	Causing an increase in inflammatory response and affecting growth performance and health status	(155, 156)

^aLysine and arginine are antagonistic: Both are absorbed by the intestinal alkaline amino acid transport system (such as the CAT family proteins), and excess lysine competitively inhibits arginine absorption.

^bThe antagonistic effect of n-3 and n-6 fatty acids: Both are based on the conversion of $\Delta 6$ and $\Delta 5$ dehydrogenases to their respective active metabolites in the body, e.g. n-3 to EPA and DHA and n-6 to AA. The antagonistic effect of the two is mainly achieved by enzyme competition and metabolite confrontation.

growth and production efficiency. In a study of growth-retarded lambs, the levels of inflammatory cytokines, including IL-1 β , IL-6 and TNF- α , were significantly elevated in the blood when compared to normal lambs (101). In addition to the ability to inhibit the production of IGF-1 in the liver, these cytokines can activate the NF- κ B pathway to suppress the hypothalamic-pituitary-gonadal-liver (HPGH) axis and GH secretion. IGF-1 is a key regulator of growth, and its deficiency directly limits the development of bones, muscles and organs in animals (102, 103). In addition, parasitic infection rapidly triggers an immune response that induces a Th2-type response and promotes the secretion of large amounts of pro-inflammatory cytokines (104). Razavi et al. (105) observed that coccidiosis infection in sheep induced an acute phase response and the liver to produce a large amount of acute phase proteins (APPs). This immune response leads to systemic metabolic reprogramming and causes the liver to increase glucose production through gluconeogenesis to meet the energy needs of immune cells. However, the process results in weight loss and growth restriction. Meanwhile, toxins secreted by parasites such as protease inhibitors also inhibit the nutrients absorption and damage intestinal and liver cells,

which further aggravate the metabolic burden of the host (104, 106).

Oxidative stress produces excessive reactive oxygen species (ROS) through lipid peroxidation, which can disrupt the integrity of lipid bilayer of cell membrane, alter enzyme activity, receptor function and conduction effects of signaling pathways. ROS can also attack mitochondria, cause mitochondrial autophagy via the Parkin and PTEN-induced kinase 1 (PINK1) pathways and impair cellular proteins and DNA (107–109). These effects will weaken mitochondrial dysfunction, affect ATP production and reduce energy supply, all of which limit growth and development of animals, particularly in tissues with high energy metabolism. Studies suggested that elevated IGF-1 levels activated the Nrf2/ARE pathway, which increased the expression of endogenous antioxidant enzymes such as GPx, helping to mitigate the physiological effects of ROS and alleviate oxidative stress (108, 110, 111). Therefore, IGF-1 not only promotes cell growth and differentiation, but also plays a key role in enhancing the antioxidant capacity and immune defense of the body, thereby indirectly supporting growth and development. In summary, effective management and disease prevention strategies aimed

at improving oxidative stress resistance and immune function may significantly improve health and enhance growth and developmental efficiency of ruminants.

4.4 Environmental factors of growth in ruminants

Environmental factors are important external conditions that affect the growth and development of ruminants. These factors include stress (mental stress), temperature and humidity, light exposure, noise, air quality and stocking density, all of which can be categorized as environmental stressors. Sub-optimal environmental conditions can induce stress responses in animals that adversely affect physiological functions and metabolic processes, ultimately leading to growth retardation. In sheep, Du (112) found that maternal stress inhibited the development of fetal muscle and adipose tissue by enhancing DNA methylation within the promoter region of peroxisome proliferator-activated receptor-gamma coactivator 1-alpha (PGC-1alpha), and this epigenetic modification persisted in the muscle and fat tissues of the offspring and had long-lasting effects on their development. Bisphenol A (BPA) is a common environmental pollutant. Zhang et al. (113) found that pregnant ewes exposed by BPA reduced the placental and trophoblast progesterone (P4) levels, and besides, BPA exposure activated mitochondrial apoptotic pathways, which significantly reduced Bcl-2 levels and increased caspase-3 and -9 expression, ultimately reducing the number of viable cells in the placenta, inducing cell apoptosis and leading to IGR in lambs. Temperature also affects the reproductive performance of ruminants. A previous study reported that elevated temperatures can impair ovarian granulosa cell function in cows and cause oxidative damage and cell apoptosis, resulting in infertility and a negative impact on reproductive performance (114). Another factor that can affect fetal development is the ambient temperature during pregnancy. In hot environments, fetal umbilical blood flow will be reduced and the expression of the glucose transporter GLUT-8 in the placenta as well as insulin levels are significantly reduced, which limit fetal growth within the uterus (115, 116).

During the process of growth, animals become unable to effectively dissipate or generate heat when external temperatures exceeds their thermoregulatory capacity, which leads to a series of physiological and metabolic changes. In a high temperature environment, animals exhibit increased respiratory rates, reduced feed intake and elevated body temperatures, as well as increased tissue catabolism and reduced anabolism processes, which not only reduce nutrients intake, but also accelerate nutrients catabolism, ultimately leading to slow weight growth (117). In addition, heat stress disrupts the endocrine balance in animals by activating the hypothalamic-pituitary-adrenal (HPA) axis, which increases the secretion of corticotropin-releasing factor, adrenocorticotropic hormone and cortisol. Higher cortisol will increase blood glucose levels and catabolism and inhibit protein and fat synthesis, thereby reducing growth efficiency of animals (118, 119). Noordhuizen (120) reported that under heat stress conditions, each 1°C increase

in temperature resulted in a 0.85 kg reduction in feed intake and a 600–900 kg reduction in annual milk production in dairy cows. Heat stress also causes a redistribution of internal resources, including fat, protein and energy, and alters absorption and metabolism regardless of changes in feed intake (121). Conversely, in a cold environment, the basal metabolic rate of animals will be increased and more energy is used to maintenance requirement, which reduce the efficiency of feed conversion and slow down growth rates. Our previous investigation found that the harsh plateau environment of the yaks during pregnancy and after birth, characterized by the depletion of forage resources, leads to inadequate early nutrition of the yak calves, which contributes significantly to growth retardation in later life (1). In addition, light exposure plays a key role in regulating growth, development, reproduction and behavior of animals. Photoperiod affects the secretion of melatonin, which in turn affects circadian rhythms and the secretion of GHs (122). Proper humidity levels, good ventilation and optimal stocking density are also crucial factors that contribute to the healthy growth and development of animals. Therefore, it is essential in livestock management practices to optimize environmental conditions, reduce stressors and provide the best conditions for growth and development.

According to the researches mentioned earlier, the growth and development of ruminants are affected by a variety of internal and external factors. These factors include changes in the structure and function of genetic materials, imbalances in nutrients supply and demand, interactions between genetics and nutrition, immune function, oxidative stress levels and environmental conditions, which are intricately interrelated and collectively influence hormone levels, endocrine regulation, nutrients digestion and absorption, energy metabolism, immune responses and oxidative stress, ultimately regulating the growth rate and health status of ruminants.

5 Regulation strategies of growth retardation in ruminants

5.1 Advancing growth performance and efficiency through genetic selection and improvement in ruminants

The important strategies for improving growth performance and disease resistance in ruminants are genetic selection and improvement. Genetic selection identifies and maintains individuals with superior growth traits and disease resistance, while genetic improvement utilizes modern molecular biology techniques to precisely modify the genome to achieve targeted improvements in specific traits. Gene editing technologies such as CRISPR-Cas9 have been widely used in the genetic improvement of livestock to improve growth rates, feed conversion efficiency, and disease resistance (123). The CRISPR-Cas9 system uses the Cas9 nuclease to recognize and cleave the target genome sequence using guide RNA to form DNA double-strand broken fragments, which rely on cells to initiate the connection or homologous recombination of non-homologous ends to achieve

gene knockout or insertion (124). Recent successful animal gene editing applications have demonstrated that the ability to modify expression of genes such as IGF-1 and GH receptors can improve production efficiency and growth performance of animals (125, 126). In practical livestock production, hybrid vigor is also an effective approach to improving growth performance. Hybridization combines the beneficial traits of different breeds to produce offspring that are more adaptable and higher yielding. For example, the milk yield and growth performance of the hybrid cattle are significantly higher than that of the purebred individuals (127). The continued advancement and refinement of gene-editing technologies is therefore a powerful tool for improving growth and productivity of ruminants.

5.2 Optimizing feeding strategies and implementing targeted nutritional regulation for enhanced growth of ruminants

Proper nutritional management is essential to improve growth and production efficiency of ruminants. From a molecular nutrition perspective, nutrients regulate metabolic pathways and gene expression in ruminants, ultimately improving growth performance. Ruminants rely primarily on VFAs, produced by microbial fermentation of fibrous feedstuffs in the rumen, as the primary source of energy (87). A previous research found that the type of carbohydrate in the diet can influence the composition of the ruminal microbiota, thereby affecting the production and ratio of VFAs, which in turn modulates energy metabolism in the host (90). Our previous research also found that supplementing glutamine in the diet of growth retardation yaks can improve the function of the gastrointestinal barrier and promote compensatory growth in growth retardation yaks (11). EAA, NEAA, vitamins and minerals not only have different physiological functions, they also interact to produce synergistic effects that positively influence growth and development of animals. Supplementation of specific nutrients in the diet can alleviate growth retardation and improve growth performance in ruminants. Politis et al. (128) showed that adding 3,000 IU of vitamin E per cow per day to the diet of dairy cows effectively suppressed neutrophil and macrophage function in early postpartum, thus alleviating inflammation. Similarly, June et al. (129) found that supplementation of vitamin E and selenium significantly reduced blood malondialdehyde levels, increased superoxide dismutase activity, attenuated oxidative stress and improved feed intake in pregnant dairy heifers. These findings highlight the potential of appropriate nutritional supplementation to alleviate stress responses and regulate growth performance in ruminants.

Feed additives such as probiotics, prebiotics and synbiotics can also influence growth and development of ruminants. These additives regulate the composition and metabolic function of the gut microbiota to improve nutrients absorption and feed utilization efficiency. Some researches has confirmed that the addition of feed additives in diets can promote feed digestion,

stabilize ruminal pH, strengthen the ruminal barrier and improve the gut microbiota, leading to improved feed intake and utilization (130–132). Zhang et al. (28) reported that the addition of L-arginine and N-carbamoyl-glutamine to the diet significantly increased the levels of ATP, ADP, total amino nitrogen (TAN) and AEC in the duodenum, jejunum and ileum of IGR lambs, and besides, the activity of key enzymes involved in the tricarboxylic acid (TCA) cycle was also enhanced, thereby improving gut function and the efficiency of feed digestion and absorption. Another study also showed that dietary supplementation with cysteine hydrochloride and active dry yeast reduced serum LPS levels, up-regulated the expression of genes related to VFAs absorption (SLC26A3, PAT1, MCT1) and cell junctions (CLDN1, CDH1, OCLN) in the rumen and down-regulated the expression of complement system genes (C2, C7) of growth-retarded yaks, which finally promoted growth (133). Sun et al. (134) revealed that rumen microbial metabolites, such as indole-3-carboxaldehyde and prostaglandin D2, regulate rumen development through specific signaling pathways, underscoring the pivotal role of microbial-metabolite interactions in precision nutrition. These findings lay a theoretical foundation for precision feeding strategies in young livestock by leveraging functional strain screening and targeted metabolite supplementation, providing new insights into the development of feed additives for young ruminants. In livestock production, the expression of growth-related genes can be effectively modulated by the rational regulation of dietary components through nutritional strategies to improve growth performance of ruminants. While the above studies highlight the positive effects of nutritional regulation on ruminants growth, they are often limited by small sample sizes and experimental conditions that may not fully reflect the complexity of different farming environments. Future research should include more comprehensive multi-factorial analysis to explore the synergistic effects of different nutrients or the interaction between nutrition and environment, with the aim of identifying optimal nutritional strategies.

5.3 Ensuring gastrointestinal microbial balance for promoting growth in ruminants

As a key factor in growth performance, the gastrointestinal microbiota plays a critical role in nutrient absorption, metabolism, and health of ruminants. Researches have shown that increasing fermentable fiber in the diet can stimulate the growth of fiber-degrading bacteria such as *Fibrobacter* and *Ruminococcus*, thereby promoting the production of acetic and butyric acids (91, 135). The diversity and functional composition of the gastrointestinal microbiota are closely related to feed digestibility, metabolic product formation and host gene expression (136). Our previous study found that the dietary addition of glutamine increased the relative abundance of unclassified *Peptostreptococcaceae*, *Romboutsia*, *Lachnospiraceae*, and *Clostridium* sensu stricto 1 in the gastrointestinal tract of growth-retarded yaks, and besides, the protease activity in the jejunum and ileum was also increased, which was beneficial for improving nutrients digestibility (137). Du

et al. (33) reported that the weight gain, feed intake and GH/IGF-1 levels were significantly improved by adding probiotics in the diet of growth-retarded calves. Moreover, probiotics intervention increased the abundance of beneficial bacteria related to VFAs production, such as *Bacteroides*, *Prevotella*, *Campylobacter*, and *Butyrivibrio*, while reduced the prevalence of pathogens. Thus, regulating the composition of the gastrointestinal microbiota can improve nutrients digestibility, gastrointestinal fermentation, and digestive enzyme activity, thereby promoting growth and development of ruminants. In addition to contributing to nutrients metabolism, the gut microbiota also regulates host gene expression through metabolites such as VFAs and lactic acid. Studies have shown that VFAs, including acetic, propionic and butyric acids, can up-regulate the expression of tight junction proteins (occludin and claudin) in the intestinal epithelial cells via histone deacetylases (HDACs) and G protein-coupled receptors (GPR41 or GPR43), thereby enhancing intestinal barrier function. Activation of GPR41 also promotes the secretion of gut hormones such as GLP-1, which are involved in the regulation of energy balance and appetite in the host (89, 95, 138). Recent advances in microbiomics technologies have made it possible to precisely analyze how different nutritional strategies affect the composition and function of the microbiota, providing a scientific basis for optimizing feed formulations and improving production performance of animals (139).

5.4 Strategies for disease prevention and stress management in ruminants

In modern intensive farming systems, animals are often subjected to high density and production conditions. Under such circumstances, sub-optimal physiological conditions, including disease and stress, are major contributors to growth retardation and reduced production efficiency. Effective disease prevention and control measures that maintain the health status of ruminants can minimize production losses and improve growth performance and farm profitability. Razavi et al. (105) showed that when sheep were affected by diseases such as coccidiosis, the malondialdehyde and total homocysteine concentrations increased significantly, while the levels of superoxide dismutase and glutathione reductase were decreased, and the utilization of trace elements such as zinc, manganese and selenium was also reduced. Disease signals are transmitted to the hypothalamus, activating the hypothalamic-pituitary-adrenal axis and the sympathetic-adrenal axis, which triggers the release of glucocorticoids and catecholamines to stimulate macrophages and lymphocytes to produce large amounts of pro-inflammatory cytokines, thus causing an inflammatory response.

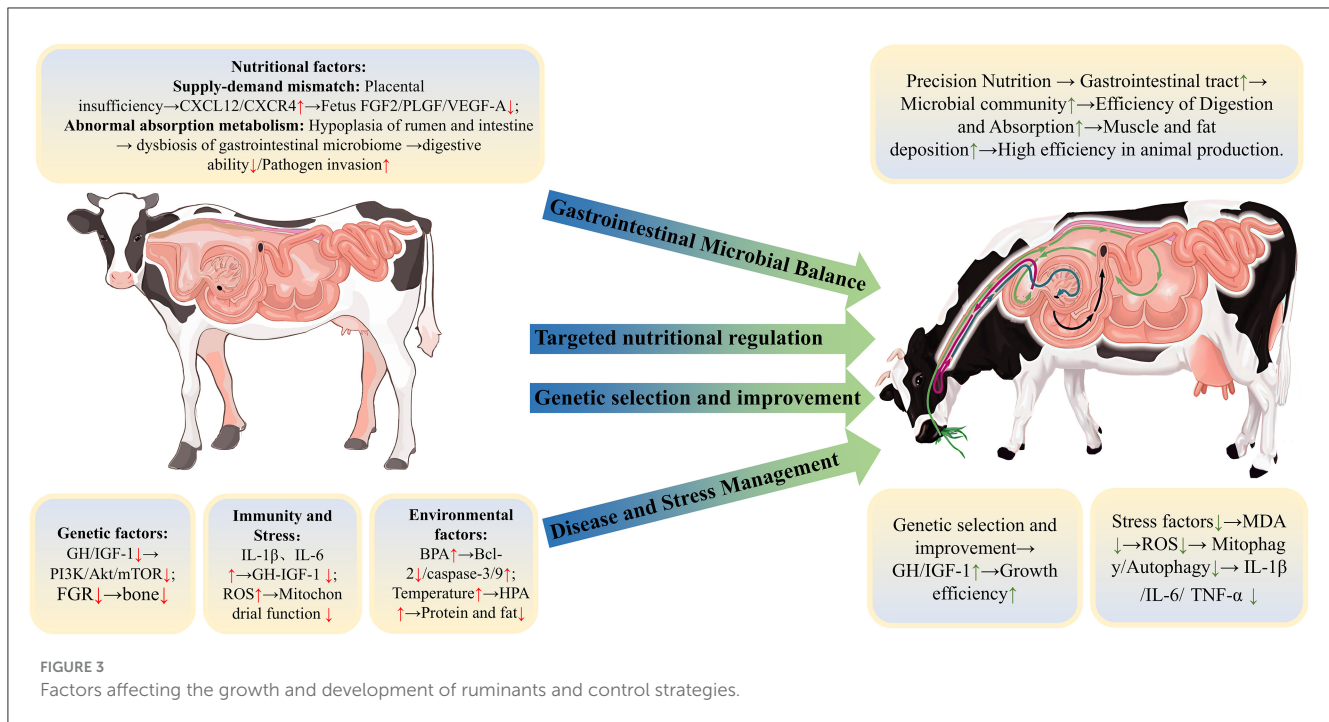
In addition, optimal environmental conditions and welfare standards not only help reduce stress responses and disease incidence, but also promote growth and development of animals. Ruminants have a limited range of tolerance to temperature and humidity, and unfavorable conditions can adversely affect their metabolism, immune system and growth performance. High temperatures can reduce feed intake, increase respiratory rates and induce heat stress, which negatively affects weight gain

and milk production in dairy cows (118). Kaushik et al. (140) investigated the expression of heat shock protein 70 (HSP70) genes in goats of different ages, and found that HSP70 expression was highest at 9 months of age and decreased as the age increasing. Notably, HSP70 expression not only reflects the degree of heat stress, but is also closely associated with growth, energy metabolism efficiency and protein processing in animals (141, 142). Stresses, a common trigger for disease outbreaks and health problems, include transport, weaning, and feed transition. In practical farming, reducing environmental fluctuations, optimizing transport schedules, adjusting weaning procedures, and gradually changing diets can significantly reduce stress responses and the risk of disease outbreaks. Feed additives are widely used to reduce stress in animals. Su et al. (143) showed that renal cells from Mongolian sheep exposed to high-sugar conditions exhibited decreased vitality and increased levels of ROS. The addition of γ -aminobutyric acid (GABA) alleviated metabolic stress by enhancing glycolysis and promoting the TCA cycle, as well as up-regulating the expression of antioxidant genes to reduce oxidative damage in cells. Optimizing the rearing environment and improving animal welfare are essential strategies for promoting growth and improving farm efficiency of ruminants. In highly modernized production systems, the timely integration of new technologies and equipment is essential to improve production efficiency and health of ruminants.

6 Future perspectives and concluding insights

Growth retardation poses a complex and multifaceted challenge in ruminants production, significantly impeding efforts to enhance productivity and efficiency. This review systematically defines and characterizes growth retardation, examines its extensive physiological impacts on ruminants, and analyzes the diverse factors influencing growth and development. As illustrated in Figure 3, the growth and development of ruminants are regulated by a complex interplay of factors, including genetics, nutrition, immunity, antioxidant capacity, and environmental conditions, which collectively shape the growth process. This review underscores the critical roles of nutrition and gastrointestinal health in fostering ruminants growth, integrating insights from genetics, nutrition, microbiology, and environmental sciences to reveal the synergistic effects of these factors on growth regulation. A holistic and multifactorial approach is therefore essential to optimize physiological functions and enhance production performance in ruminants. Additionally, fostering interdisciplinary collaboration that merges perspectives from genetics, microbiology, nutrition, and animal welfare is vital for building more efficient and sustainable livestock production systems. This synthesis highlights the importance of collaborative research that considers the interrelated nature of these factors, ultimately providing a basis for the development of targeted strategies to support sustainable growth.

Future research, guided by advances in genetic analysis, microbiome profiling and real-time health monitoring, should prioritize the development of precision-based approaches that



tailor interventions to the specific needs of individual ruminants or herds. Emerging genomic and transcriptomic tools can further enable selective breeding for growth-enhancing traits, while novel nutritional strategies, such as targeted supplementation or prebiotic and probiotic interventions, can improve gut health and nutrient absorption. Continued investigation of the gut microbiome, particularly in relation to its interaction with immune function and stress resilience, offers promising avenues for improving animal health and growth performance of ruminants. Integrating stress management practices, such as optimized housing and handling systems, with nutritional and microbial interventions may contribute to a more holistic approach to mitigating growth retardation. Ultimately, the management of ruminants with growth retardation will benefit from a systems-level perspective that considers not only individual factors, but also how they collectively influence growth. The future of the field lies in advancing technologies and multidisciplinary collaborations that enable practical, evidence-based solutions and promote more productive, sustainable and humane ruminant production systems.

Author contributions

TL: Conceptualization, Writing – original draft, Writing – review & editing. BR: Visualization, Writing – review & editing. JL: Visualization, Writing – review & editing. LZ: Formal analysis, Writing – review & editing. FY: Supervision, Writing – review & editing. SG: Supervision, Writing – review & editing. XS: Resources, Writing – review & editing. GL: Resources, Writing – review & editing. JM: Conceptualization, Funding acquisition, Supervision, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research and/or publication of this article. This work was supported by the Program for Scientific Research Start-up funds of Guangdong Ocean University (060302052318 and 060302052308) and Innovation Team Project of Guangdong Province (2024KCXTD040).

Conflict of interest

XS, GL, and JM were employed by Xinjiang Taikun Group Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Generative AI statement

The author(s) declare that no Gen AI was used in the creation of this manuscript.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Ma J. *Regulation of glutamine on gastrointestinal barrier function and bacterial community structure in delayed growth yaks* (dissertation). Sichuan Agricultural University, Chengdu, China (2021).
- Huo L, Cao Y, Gao Y, Li Y, Li Q, Li J. The effect of dietary energy level on the growth performance and rumen fermentation of Holstein cattle. *Acta Vet Zootech Sin.* (2019) 50:332–42. doi: 10.11843/j.issn.0366-6964.2019.02.011
- Jiao P, Lu H, Hao L, Degen AA, Cheng J, Yin Z, Mao S, Xue Y. Nutrigenetic and epigenetic mechanisms of maternal nutrition-induced glucolipid metabolism changes in the offspring. *Nutr Rev.* (2024) nuae048. doi: 10.1093/nutrit/nuae048
- Dayton WR, White ME. Cellular and molecular regulation of muscle growth and development in meat animals. *J Anim Sci.* (2008) 86:E217–25. doi: 10.2527/jas.2007-0456
- Chen MM, Zhao YP, Zhao Y, Deng SL, Yu K. Regulation of myostatin on the growth and development of skeletal muscle. *Front Cell Dev Biol.* (2021) 9:785712. doi: 10.3389/fcell.2021.785712
- Yang F. *Animal Nutrition*. Beijing: China Agriculture Press (2003). p. 234.
- Duittoz AH, Kenny DA. Review: early and late determinants of puberty in ruminants and the role of nutrition. *Animal.* (2023) 17:100812. doi: 10.1016/j.animal.2023.100812
- Jones CK, Gabler NK, Main RG, Patience JF. Characterizing growth and carcass composition differences in pigs with varying weaning weights and postweaning performance. *J Anim Sci.* (2012) 90:4072–80. doi: 10.2527/jas.2011-4793
- Abdollahi M, Ajami M, Abdollahi Z, Kalantari N, Houshiarrad A, Fozouni F, et al. Zinc supplementation is an effective and feasible strategy to prevent growth retardation in 6 to 24 month children: a pragmatic double blind, randomized trial. *Heliyon.* (2019) 5:e02581. doi: 10.1016/j.heliyon.2019.e02581
- Fatemi MJ, Dianatinasab M, Sharifnia G, Moravej H, Fararouei M. Growth retardation among children in southern Iran: a 7-year population based cohort study. *BMC Public Health.* (2020) 20:1392. doi: 10.1186/s12889-020-09511-w
- Ma J, Shah AM, Wang ZS, Hu R, Zou HW, Wang XY, et al. Dietary supplementation with glutamine improves gastrointestinal barrier function and promotes compensatory growth of growth-retarded yaks. *Animal.* (2021) 15:100108. doi: 10.1016/j.animal.2020.100108
- Maiorana A, Del Bianco C, Cianfarani S. Adipose tissue: a metabolic regulator. Potential implications for the metabolic outcome of subjects born Small for Gestational Age (SGA). *Rev Diabet Stud.* (2007) 4:134–46. doi: 10.1900/RDS.2007.4.134
- Zhang Y, Ren J, Chen L, Yan H, Zou T, Zhang H, et al. Effects of equol supplementation on growth performance, redox status, intestinal health and skeletal muscle development of weanling piglets with intrauterine growth retardation. *Animals.* (2023) 13:1469. doi: 10.3390/ani13091469
- Zhang H, Jin Y, Wang M, Looor JJ, Wang H. N-Carbamylglutamate and L-arginine supplementation improve hepatic antioxidant status in intrauterine growth-retarded suckling lambs. *RSC Adv.* (2020) 10:11173–81. doi: 10.1039/C9RA09316H
- Sutherland AE, White TA, Rock CR, Piscopo BR, Dudink I, Inocencio IM, et al. Phenotype of early-onset fetal growth restriction in sheep. *Front Endocrinol.* (2024) 15:1374897. doi: 10.3389/fendo.2024.1374897
- Chai J-M, Ma T, Wang H-C, Qi M-L, Tu Y, Diao Q-Y, et al. Effect of early weaning age on growth performance, nutrient digestibility, and serum parameters of lambs. *Anim Prod Sci.* (2017) 57:110. doi: 10.1071/AN15079
- Ma J, Shah AM, Wang Z, Hu R, Zou H, Wang X, et al. Comparing the gastrointestinal barrier function between growth-retarded and normal yaks on the Qinghai-Tibetan Plateau. *Peer J.* (2020) 8:e9851. doi: 10.7717/peerj.9851
- Taylor SC. Genetic aspects of mammalian growth and survival in relation to body size. *Proc Nutr Soc.* (1990) 49:421–2. doi: 10.1079/PNS19900049
- Wang YJ, Wang ZS, Hu R, Peng QH, Xia K, Zhao SN, et al. The effects of different nutritional regulation methods on the growth and development, blood routine, plasma antioxidant and immune indicators of yak stiff cattle. *Chin J Anim Nutr.* (2018) 30:1344–52. doi: 10.3969/j.issn.1006-267x.2018.04.017
- Svensson J, Sjögren K, Lawenius L, Koskela A, Tuukkanen J, Nilsson KH, et al. Bone-derived IGF-I regulates radial bone growth in adult male mice. *Endocrinology.* (2023) 8:bqad104. doi: 10.1210/endo/bqad104
- Dixit M, Poudel SB, Yakar S. Effects of GH/IGF axis on bone and cartilage. *Mol Cell Endocrinol.* (2021) 519:111052. doi: 10.1016/j.mce.2020.111052
- Ma MP. *The effect of neutral detergent fiber sources in the diet on the growth performance and rumen development of weaned calves* (master's thesis). Gansu Agricultural University, Lanzhou, China (2020).
- Liu S, Yang L, Zhang Y, Chen H, Li X, Xu Z, et al. Review of yeast culture concerning the interactions between gut microbiota and young ruminant animals. *Front Vet Sci.* (2024) 11:1335765. doi: 10.3389/fvets.2024.1335765
- Hu L, Peng X, Chen H, Yan C, Liu Y, Xu Q, et al. Effects of intrauterine growth retardation and *Bacillus subtilis* PB6 supplementation on growth performance, intestinal development and immune function of piglets during the suckling period. *Eur J Nutr.* (2017) 56:1753–65. doi: 10.1007/s00394-016-1223-z
- Hu R, Wang Z, Peng Q, Zou H, Wang H, Yu X, et al. Effects of GHRP-2 and cysteamine administration on growth performance, somatotrophic axis hormone and muscle protein deposition in yaks (*Bos grunniens*) with growth retardation. *PLoS ONE.* (2016) 11:e0149461. doi: 10.1371/journal.pone.0149461
- Pokhrel B, Jiang H. Postnatal growth and development of the rumen: integrating physiological and molecular insights. *Biology.* (2024) 13:269. doi: 10.3390/biology13040269
- Zhao F, Ren W, Zhang A, Jiang N, Liu W, Wang F. Effects of different amylose to amylpectin ratios on rumen fermentation and development in fattening lambs. *Asian-Austr J Anim Sci.* (2018) 31:1611–8. doi: 10.5713/ajas.17.0833
- Zhang H, Peng A, Guo S, Wang M, Looor JJ, Wang H. Dietary N-carbamylglutamate and L-arginine supplementation improves intestinal energy status in intrauterine-growth-retarded suckling lambs. *Food Funct.* (2019) 10:1903–14. doi: 10.1039/C8FO01618F
- Huang Y, Wang G, Zhang Q, Chen Z, Li C, Wang W, et al. Effects of milk replacer feeding level on growth performance, rumen development and the ruminal bacterial community in lambs. *Front Microbiol.* (2023) 13:1069964. doi: 10.3389/fmicb.2022.1069964
- Wesemann DR. Microbes and B cell development. *Adv Immunol.* (2015) 125:155–78. doi: 10.1016/bs.ai.2014.09.005
- Henderson G, Cox F, Ganesh S, Jonker A, Young W, Abecia L, et al. Rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. *Sci Rep.* (2015) 5:14567. doi: 10.1038/srep14567
- Zhuang Y, Gao D, Jiang W, Xu Y, Liu G, Hou G, et al. Core microbe Bifidobacterium in the hindgut of calves improves the growth phenotype of young hosts by regulating microbial functions and host metabolism. *Microbiome.* (2025) 1:13. doi: 10.1186/s40168-024-02010-9
- Du R, Jiao S, Dai Y, An J, Lv J, Yan X, et al. Probiotic bacillus amyloliquefaciens C-1 improves growth performance, stimulates GH/IGF-1, and regulates the gut microbiota of growth-retarded beef calves. *Front Microbiol.* (2018) 9:2006. doi: 10.3389/fmicb.2018.02006
- Ma J, Zhu Y, Wang Z, Yu X, Hu R, Wang X, et al. Comparing the bacterial community in the gastrointestinal tracts between growth-retarded and normal yaks on the Qinghai-Tibetan Plateau. *Front Microbiol.* (2020) 11:600516. doi: 10.3389/fmicb.2020.600516
- Mohapatra SS, Mukherjee J, Banerjee D, Das PK, Ghosh PR, Das K. RFamide peptides, the novel regulators of mammalian HPG axis: a review. *Vet World.* (2021) 14:1867–73. doi: 10.14202/vetworld.2021.1867-1873
- Pei S, Wang Z, Liu Y, Xu Y, Bai J, Li W, et al. Transcriptomic analysis of the HPG axis-related tissues reveals potential candidate genes and regulatory pathways associated with testicular size in Hu sheep. *Theriogenology.* (2024) 216:168–76. doi: 10.1016/j.theriogenology.2024.01.002
- Kumar S, Dahiya SP, Magotra A, Ratwan P, Bangar Y. Influence of single nucleotide polymorphism in the IGF-1 gene on performance and conformation traits in Munjal sheep. *Zygote.* (2023) 31:70–7. doi: 10.1017/S0967199422000545
- Gong JG, Baxter G, Bramley TA, Webb R. Enhancement of ovarian follicle development in heifers by treatment with recombinant bovine somatotrophin: a dose-response study. *Reproduction.* (1997) 110:91–7. doi: 10.1530/jrf.0.1100091
- Jimenez CR, De Azevedo JL, Ciro Alexandre Alves T, Penitente-Filho JM, Gonçalves WG. Sequential medium with GH and IGF-1 improved in vitro development of bovine preantral follicles enclosed in ovarian tissue. *Reprod Domest Anim.* (2018) 53:1103–13. doi: 10.1111/rda.13210
- Simopoulou M, Philippou A, Maziotis E, Sfakianoudis K, Nitsos N, Bakas P, et al. Association between male infertility and seminal plasma levels of growth hormone and insulin-like growth factor-1. *Andrologia.* (2018) 50:e13048. doi: 10.1111/and.13048
- Bahadir GB, Gollu G, Ilkay H, Bagriacik U, Hasirci N, Bingol-Kologlu M. Local-IGF-1 and GH application IMPROVES germ cell histology, spermatogenesis and fertility after experimental testicular torsion and detorsion. *J Pediatr Urol.* (2022) 18:410.e1–e8. doi: 10.1016/j.jpuro.2022.07.001
- Kumar A, Singh G, Jerome A, Kumar P, Arjun V, Bala R, et al. IGF-1 supplementation in semen affects mitochondrial functional and calcium status of buffalo sperm following cryopreservation. *Anim Reprod Sci.* (2021) 231:106783. doi: 10.1016/j.anireprosci.2021.106783
- McClelland EK, Chan MTT, Lin X, Sakhrani D, Vincelli F, Kim J-H, et al. Loci associated with variation in gene expression and growth in juvenile salmon are influenced by the presence of a growth hormone transgene. *BMC Genomics.* (2020) 21:185. doi: 10.1186/s12864-020-6586-0

44. Saleh AA, Hassan TGM, EL-Hedainy DKA, El-Barbary ASA, Sharaby MA, Hafez EE, et al. IGF-I and GH genes polymorphism and their association with milk yields, composition and reproductive performance in Holstein–Friesian dairy cattle. *BMC Vet Res.* (2024) 20:341. doi: 10.1186/s12917-024-04188-4
45. Dees WL, Hiney JK, Srivastava VK. IGF-1 influences gonadotropin-releasing hormone regulation of puberty. *Neuroendocrinology.* (2021) 111:1151–63. doi: 10.1159/000514217
46. Yan J, Charles JF. Gut microbiota and IGF-1. *Calcif Tissue Int.* (2018) 102:406–14. doi: 10.1007/s00223-018-0395-3
47. Li B, Feng L, Wu X, Cai M, Yu JJ, Tian Z. Effects of different modes of exercise on skeletal muscle mass and function and IGF-1 signaling during early aging in mice. *J Exp Biol.* (2022) 225:21. doi: 10.1242/jeb.244650
48. Roh S-G, Matsunaga N, Hidaka S, Hidari H. Characteristics of growth hormone secretion responsiveness to growth hormone-releasing peptide-2 (GHRP-2 or KP102) in calves. *Endocr J.* (1996) 43:291–8. doi: 10.1507/endocrj.43.291
49. Bowers CY, Momany FA, Reynolds GA, Hong A. On the *in vitro* and *in vivo* activity of a new synthetic hexapeptide that acts on the pituitary to specifically release growth hormone. *Endocrinology.* (1984) 114:1537–45. doi: 10.1210/endo-114-5-1537
50. Ascenzi F, Barberi L, Dobrowolny G, Villa Nova Bacurau A, Nicoletti C, Rizzuto E, et al. Effects of IGF-1 isoforms on muscle growth and sarcopenia. *Aging Cell.* (2019) 18:e12954. doi: 10.1111/acel.12954
51. Meng Z, Tan D, Cheng Z, Jiang M, Zhan K. GPR41 regulates the proliferation of BRECs via the PIK3-AKT-mTOR pathway. *Int J Mol Sci.* (2023) 24:4203. doi: 10.3390/ijms24044203
52. Du C, Zhou X, Zhang K, Huang S, Wang X, Zhou S, et al. Inactivation of the MSTN gene expression changes the composition and function of the gut microbiome in sheep. *BMC Microbiol.* (2022) 1:273. doi: 10.1186/s12866-022-02687-8
53. Liu M, Zhang C, Lai X, Xue J, Lan X, Lei C, et al. Associations between polymorphisms in the NICD domain of bovine Notch-1 gene and growth traits in Chinese Qinchuan cattle. *J Appl Genetics.* (2017) 58:241–7. doi: 10.1007/s13353-016-0377-y
54. Kong L, Liu G, Deng M, Lian Z, Han Y, Sun B, et al. Growth retardation-responsive analysis of mRNAs and long noncoding RNAs in the liver tissue of Leiqiong cattle. *Sci Rep.* (2020) 10:14254. doi: 10.1038/s41598-020-71206-4
55. Yang P, Li X, Du L, Lyu S, Zhang Z, Lin F, et al. CircDyrk1a regulates bovine myoblasts development by binding miR21-5p to affect KLF5 gene expression. *Anim Res One Health.* (2024) 4:431–45. doi: 10.1002/aro.276
56. Xiang X, Peng C, Cao D, Chen Z, Jin H, Nie S, et al. Whole genome sequencing reveals that five genes are related to BW trait in sheep. *Animal.* (2024) 18:101282. doi: 10.1016/j.animal.2024.101282
57. Rupp T, Amling M, Rolvien T. FGF23 and bone microarchitecture. *Osteoporos Int.* (2019) 30:2359–2359. doi: 10.1007/s00198-019-05124-1
58. Phan P, Saikia BB, Sonnila S, Agrawal S, Alraawi Z, Kumar TKS, et al. The saga of endocrine FGFs. *Cells.* (2021) 10:2418. doi: 10.3390/cells10092418
59. Grismaldo RA, Luévano-Martínez LA, Reyes M, García-Márquez G, García-Rivas G, Sobrevia L. Placental mitochondrial impairment and its association with maternal metabolic dysfunction. *J Phys.* (2024) JP285935. doi: 10.1113/JP285935
60. Brown LD, Palmer C, Teynor L, Boehmer BH, Stremming J, Chang EI, et al. Fetal sex does not impact placental blood flow or placental amino acid transfer in late gestation pregnant sheep with or without placental insufficiency. *Reprod Sci.* (2022) 29:1776–89. doi: 10.1007/s43032-021-00750-9
61. Brown LD, Hay WW. Impact of placental insufficiency on fetal skeletal muscle growth. *Mol Cell Endocrinol.* (2016) 435:69–77. doi: 10.1016/j.mce.2016.03.017
62. Ashley RL, Trigo EM, Ervin JM. Placental insufficiency and heavier placentas in sheep after suppressing CXCL12/CXCR4 signaling during implantation. *Biol Reprod.* (2023) 109:982–93. doi: 10.1093/biolre/iaod122
63. Ticiani E, Lazzarotto CR, Gerger RPC, Wilhelm BR, Rodrigues VHV, Rodrigues JL, et al. Evidence of metabolic compartmentalization or zonation in the bovine placenta: significance for the regulation of placental function and fetal growth. *Reprod Fertil Dev.* (2018) 30:168. doi: 10.1071/RDv30n1Ab58
64. Daida T, Shin BC, Cepeda C, Devaskar SU. Neurodevelopment is dependent on maternal diet: placenta and brain glucose transporters GLUT1 and GLUT3. *Nutrients.* (2024) 14:2363. doi: 10.3390/nu16142363
65. Britt JL, Feltus A, Miller ME, Koch BM, Miller MC, Duckett SK. 324 altered expression of placental microRNA in ewes with uteroplacental insufficiency due to consumption of endophyte-infected tall fescue seed. *J Anim Sci.* (2017) 95:160–160. doi: 10.2527/asasann.2017.324
66. Zhu MJ, Ford SP, Means WJ, Hess BW, Nathanielsz PW, Du M. Maternal nutrient restriction affects properties of skeletal muscle in offspring. *J Physiol.* (2006) 575:241–50. doi: 10.1113/jphysiol.2006.112110
67. Daniel ZC, Brameld JM, Craigan J, Scollan ND, Buttery PJ. Effect of maternal dietary restriction during pregnancy on lamb carcass characteristics and muscle fiber composition. *J Anim Sci.* (2007) 85:1565–76. doi: 10.2527/jas.2006-743
68. Mohammadabadi M, Bordbar F, Jensen J, Du M, Guo W. Key genes regulating skeletal muscle development and growth in farm animals. *Animals.* (2021) 11:835. doi: 10.3390/ani11030835
69. Bayol S, Jones D, Goldspink G, Stickland NC. The influence of undernutrition during gestation on skeletal muscle cellularity and on the expression of genes that control muscle growth. *Br J Nutr.* (2004) 91:331–9. doi: 10.1079/BJN20031070
70. Yan X, Zhu M-J, Dodson MV, Du M. Developmental programming of fetal skeletal muscle and adipose tissue development. *J Genomics.* (2013) 1:29–38. doi: 10.7150/jgen.3930
71. Stickland NC, Widdowson EM, Goldspink G. Effects of severe energy and protein deficiencies on the fibres and nuclei in skeletal muscle of pigs. *Br J Nutr.* (1975) 34:421–8. doi: 10.1017/S0007114575000487
72. Zaugg J, Ziegler F, Nuoffer J-M, Moser-Hässig R, Albrecht C. Efficient leucine transport across the placental barrier depends on physiological, oppositely directed leucine gradients. *Placenta.* (2019) 83:e113. doi: 10.1016/j.placenta.2019.06.358
73. Agrawal P, Kaur J, Singh J, Rasane P, Sharma K, Bhadaria V, et al. Genetics, nutrition, and health: a new frontier in disease prevention. *J Am Nutr Assoc.* (2024) 43:326–38. doi: 10.1080/27697061.2023.2284997
74. Li Q, Zhang Q, Su S, Yang S, Shao J, Guan W, et al. Maternal fish oil supplementation enhances nutrient transport in the placenta and milk biosynthesis in the mammary gland via the GPR120 signaling pathway. *J Adv Res.* (2024) 24:0607–6. doi: 10.1016/j.jare.2024.12.029
75. Okamura M, Kudo H, Wakabayashi K, Tanaka T, Nonaka A, Uchida A, et al. COUP-TFII acts downstream of Wnt/ β -catenin signal to silence PPAR γ gene expression and repress adipogenesis. *Proc Natl Acad Sci USA.* (2009) 106:5819–24. doi: 10.1073/pnas.0901676106
76. De Winter TJJ, Nusse R. Running against the Wnt: How Wnt/ β -Catenin suppresses adipogenesis. *Front Cell Dev Biol.* (2021) 9:627429. doi: 10.3389/fcell.2021.627429
77. Lorzadeh S, Kohan L, Ghavami S, Azarpira N. Autophagy and the Wnt signaling pathway: a focus on Wnt/ β -catenin signaling. *Biochim Biophys Acta BBA - Mol Cell Res.* (2021) 1868:118926. doi: 10.1016/j.bbamcr.2020.118926
78. Dror DK, Allen LH. Overview of nutrients in human milk. *Adv Nutr.* (2018) 9:278S–94S. doi: 10.1093/advances/nmy022
79. Guiso ME, Battacone G, Canu L, Deroma M, Langasco I, Sanna G, et al. Essential and toxic mineral content and fatty acid profile of colostrum in dairy sheep. *Animals.* (2022) 12:2730. doi: 10.3390/ani12202730
80. Jagusch KT, Mitchell RM. Utilisation of the metabolisable energy of ewe's milk by the lamb. *N Z J Agric Res.* (1971) 14:434–41. doi: 10.1080/00288233.1971.10427106
81. Sakase M, Kitagawa K, Kibushi M, Kawate N, Weerakoon WW, Hannan MA, et al. Relationships of plasma insulin-like peptide 3, testosterone, inhibin, and insulin-like growth factor-I concentrations with scrotal circumference and testicular weight in Japanese black beef bull calves. *J Reprod Dev.* (2018) 64:401–7. doi: 10.1262/jrd.2018-034
82. Su Y, Sun X, Zhao S, Hu M, Li D, Qi S, et al. Dietary alfalfa powder supplementation improves growth and development, body health, and meat quality of Tibetan sheep. *Food Chem.* (2022) 396:133709. doi: 10.1016/j.foodchem.2022.133709
83. Ma X, Han M, Li D, Hu S, Gilbreath KR, Bazer FW, et al. L-Arginine promotes protein synthesis and cell growth in brown adipocyte precursor cells via the mTOR signal pathway. *Amino Acids.* (2017) 49:957–64. doi: 10.1007/s00726-017-2399-0
84. Ford SP, Hess BW, Schwoppe MM, Nijland MJ, Gilbert JS, Vonnahme KA, et al. Maternal undernutrition during early to mid-gestation in the ewe results in altered growth, adiposity, and glucose tolerance in male offspring. *J Anim Sci.* (2007) 85:1285–94. doi: 10.2527/jas.2005-624
85. Woodman HE, Stewart J. The mechanism of cellulose digestion in the ruminant organism: II. The transformation of cellulose into glucose by the agency of cellulose-splitting bacteria. *J Agric Sci.* (1928) 18:713–23. doi: 10.1017/S0021859600009291
86. Gurevich VV. Protein multi-functionality: introduction. *Cell Mol Life Sci.* (2019) 76:4405–6. doi: 10.1007/s00018-019-03271-6
87. Bergman EN. Energy contributions of volatile fatty acids from the gastrointestinal tract in various species. *Physiol Rev.* (1990) 70:567–90. doi: 10.1152/physrev.1990.70.2.567
88. Weimer PJ. Redundancy, resilience, and host specificity of the ruminal microbiota: implications for engineering improved ruminal fermentations. *Front Microbiol.* (2015) 6:e00296. doi: 10.3389/fmicb.2015.00296
89. Psichas A, Sleeth ML, Murphy KG, Brooks L, Bewick GA, Hanyaloglu AC, et al. The short chain fatty acid propionate stimulates GLP-1 and PYY secretion via free fatty acid receptor 2 in rodents. *Int J Obes.* (2015) 39:424–9. doi: 10.1038/ijo.2014.153
90. Russell JB, Rychlik JL. Factors that alter rumen microbial ecology. *Science.* (2001) 292:1119–22. doi: 10.1126/science.1058830
91. Steele MA, Penner GB, Chaucheyras-Durand F, Guan LL. Development and physiology of the rumen and the lower gut: targets for improving gut health. *J Dairy Sci.* (2016) 99:4955–66. doi: 10.3168/jds.2015-10351

92. Henderson G, Cox F, Ganesh S, Jonker A, Young W, Janssen PH, et al. Erratum: rumen microbial community composition varies with diet and host, but a core microbiome is found across a wide geographical range. *Sci Rep.* (2016) 6:19175. doi: 10.1038/srep19175
93. Wang K, Lei Q, Ma H, Jiang M, Yang T, Ma Q, et al. Phloretin protects bovine rumen epithelial cells from LPS-induced injury. *Toxins.* (2022) 14:337. doi: 10.3390/toxins14050337
94. Penner GB, Steele MA, Aschenbach JR, McBride BW. Ruminant nutrition symposium: molecular adaptation of ruminal epithelia to highly fermentable diets. *J Anim Sci.* (2011) 89:1108–19. doi: 10.2527/jas.2010-3378
95. Górka P, Kowalski ZM, Zabielski R, Guilloteau P. Invited review: use of butyrate to promote gastrointestinal tract development in calves. *J Dairy Sci.* (2018) 101:4785–800. doi: 10.3168/jds.2017-14086
96. Vincent TJ, Nolasco LR, Suriyapha C, Johnson TA, Schoonmaker JP. Effects of dietary antimicrobials on the ruminant gastrointestinal tract microbiome. *J Anim Sci.* (2024) 102:215–6. doi: 10.1093/jas/skae102.244
97. Chopyk DM, Grakoui A. Contribution of the intestinal microbiome and gut barrier to hepatic disorders. *Gastroenterology.* (2020) 159:849–63. doi: 10.1053/j.gastro.2020.04.077
98. Peng J, Tang Y, Huang Y. Gut health: the results of microbial and mucosal immune interactions in pigs. *Anim Nutr.* (2021) 7:282–94. doi: 10.1016/j.aninu.2021.01.001
99. Lv J, Hou B, Song J, Xu Y, Xie S. The relationship between ferroptosis and diseases. *J Multidiscip Health Volume.* (2022) 15:2261–75. doi: 10.2147/JMDH.S382643
100. Chen Y, Guo X, Zeng Y, Mo X, Hong S, He H, et al. Oxidative stress induces mitochondrial iron overload and ferroptotic cell death. *Sci Rep.* (2023) 13:15515. doi: 10.1038/s41598-023-42760-4
101. Zhang H, Sun H, Peng A, Guo S, Wang M, Looor JJ, et al. N-carbamylglutamate and L-arginine promote intestinal function in suckling lambs with intrauterine growth restriction by regulating antioxidant capacity via a nitric oxide-dependent pathway. *Food Funct.* (2019) 10:6374–84. doi: 10.1039/C9FO01752F
102. Witkowska-Sedek E, Pyrzak B. Chronic inflammation and the growth hormone/insulin-like growth factor-1 axis. *Cent Eur J Immunol.* (2020) 45:469–75. doi: 10.5114/ceji.2020.103422
103. Reed S. 500 roles for inflammation in livestock muscle growth and repair. *J Anim Sci.* (2018) 96:276–7. doi: 10.1093/jas/sky404.607
104. McIntire KM, Juliano SA. Detrimental effects of a failed infection by a Co-Invasive parasite on a native congeneric parasite and its native host. *Biol Invasions.* (2021) 23:1637–48. doi: 10.1007/s10530-021-02464-0
105. Razavi SM, Soltan MS, Abbasian K, Karami A, Nazifi S. Acute phase response and oxidative stress in coccidiosis: a review in domestic animals. *Vet Parasitol.* (2024) 331:110286. doi: 10.1016/j.vetpar.2024.110286
106. Schatz AM, Park AW. Host and parasite traits predict cross-species parasite acquisition by introduced mammals. *Proc R Soc B Biol Sci.* (2021) 288:20210341. doi: 10.1098/rspb.2021.0341
107. Zhang H, Ma Y, Wang M, Elsabagh M, Looor JJ, Wang H. Dietary supplementation of L-Arginine and N-Carbamylglutamate enhances duodenal barrier and mitochondrial functions and suppresses duodenal inflammation and mitophagy in suckling lambs suffering from Intrauterine-Growth-Restriction. *Food Funct.* (2020) 11:4456–70. doi: 10.1039/D0FO00019A
108. Oke OE, Akosile OA, Oni AI, Opopoye IO, Ishola CA, Adebisi JO, et al. Oxidative stress in poultry production. *Poult Sci.* (2024) 103:104003. doi: 10.1016/j.psj.2024.104003
109. Sies H, Berndt C, Jones DP. Oxidative stress. *Annu Rev Biochem.* (2017) 86:715–48. doi: 10.1146/annurev-biochem-061516-045037
110. Lodjak J, Mägi M. Crosstalk between growth and somatic maintenance in young animals. *J Avian Biol.* (2017) 48:1360–3. doi: 10.1111/jav.01519
111. Bartosz G. Book review. *Cell Biol Int.* (2000) 24:764–764. doi: 10.1006/cbir.1999.0607
112. Du M. 195 Maternal impact on fetal muscle and adipose tissue development. *J Anim Sci.* (2021) 99:103–4. doi: 10.1093/jas/skab235.187
113. Zhang H, Zha X, Zheng Y, Liu X, Elsabagh M, Wang H, et al. Mechanisms underlying the role of endoplasmic reticulum stress in the placental injury and fetal growth restriction in an ovine gestation model. *J Anim Sci Biotechnol.* (2023) 14:117. doi: 10.1186/s40104-023-00919-z
114. Wang Y, Yang C, Elsheikh NAH, Li C, Yang F, Wang G, et al. HO-1 reduces heat stress-induced apoptosis in bovine granulosa cells by suppressing oxidative stress. *Aging.* (2019) 11:5535–47. doi: 10.18632/aging.102136
115. De Vrijer B, Davidsen ML, Wilkening RB, Anthony RV, Regnault TR. Altered placental and fetal expression of IGFs and IGF-binding proteins associated with intrauterine growth restriction in fetal sheep during early and mid-pregnancy. *Pediatr Res.* (2006) 60:507–12. doi: 10.1203/01.PDR.0000242364.78002.71
116. Limesand SW, Regnault TR, Hay Jr, WW. Characterization of glucose transporter 8 (GLUT8) in the ovine placenta of normal and growth restricted fetuses. *Placenta.* (2004) 25:70–7. doi: 10.1016/j.placenta.2003.08.012
117. Habeeb AA, Osman SF, Teama FEI, Gad AE. The detrimental impact of high environmental temperature on physiological response, growth, milk production, and reproductive efficiency of ruminants. *Trop Anim Health Prod.* (2023) 55:388. doi: 10.1007/s11250-023-03805-y
118. Tang LP, Liu YL, Zhang JX, Ding KN, Lu MH, He YM. Heat stress in broilers of liver injury effects of heat stress on oxidative stress and autophagy in liver of broilers. *Poult Sci.* (2022) 101:102085. doi: 10.1016/j.psj.2022.102085
119. Shephard R, Maloney S. A review of thermal stress in cattle. *Aust Vet J.* (2023) 101:417–29. doi: 10.1111/avj.13275
120. Noordhuizen J. Heat stress in dairy cattle: major effects and practical management measures for prevention and control. *Symbiosis J Vet Sci.* (2015) 1:1–7. doi: 10.15226/2381-2907/1/1/00103
121. Belhadji Slimen I, Najar T, Ghram A, Abdrabba M. Heat stress effects on livestock: molecular, cellular and metabolic aspects, a review. *J Anim Physiol Anim Nutr.* (2016) 100:401–12. doi: 10.1111/jpn.12379
122. Duan C, Xu J, Zhang Y, Zhang W, Sun Y, Jia Z. Effects of melatonin implantation on cashmere growth, hormone concentrations and cashmere yield in Cashmere-Perennial-Type Liaoning cashmere goats. *Anim Prod Sci.* (2017) 57:60. doi: 10.1071/AN15183
123. Doudna JA, Charpentier E. The new frontier of genome engineering with CRISPR-Cas9. *Science.* (2014) 346:1258096. doi: 10.1126/science.1258096
124. Ding W, Zhang Y, Shi S. Development and application of CRISPR/Cas in microbial biotechnology. *Front Bioeng Biotechnol.* (2020) 8:711. doi: 10.3389/fbioe.2020.00711
125. Perisse IV, Fan Z, Singina GN, White KL, Polejaeva IA. Improvements in gene editing technology boost its applications in livestock. *Front Genet.* (2021) 11:614688. doi: 10.3389/fgene.2020.614688
126. Garcia A. 50 gene editing applications in livestock production. *J Anim Sci.* (2023) 101:43–4. doi: 10.1093/jas/skad068.050
127. Olson KM, Cassell BG, McAllister AJ, Washburn SP. Dystocia, stillbirth, gestation length, and birth weight in holstein, jersey, and reciprocal crosses from a planned experiment. *J Dairy Sci.* (2009) 92:6167–75. doi: 10.3168/jds.2009-2260
128. Politis I, Hidiroglou M, Batra TR, Gilmore JA, Gorewit RC, Scherf H. Effects of vitamin E on immune function of dairy cows. *Am J Vet Res.* (1995) 56:179–84. doi: 10.2460/ajvr.1995.56.02.179
129. Jung DJS, Kim DH, Beak SH, Cho IG, Hong SJ, Lee J, et al. Effects of vitamin E and selenium administration on transportation stress in pregnant dairy heifers. *J Dairy Sci.* (2023) 106:9576–86. doi: 10.3168/jds.2023-23463
130. Zhang X, Liu X, Chang S, Zhang C, Du W, Hou F. Effect of cistanche deserticola on rumen microbiota and rumen function in grazing sheep. *Front Microbiol.* (2022) 13:840725. doi: 10.3389/fmicb.2022.840725
131. Ren Z, Yao R, Liu Q, Deng Y, Shen L, Deng H, et al. Effects of antibacterial peptides on rumen fermentation function and rumen microorganisms in goats. *PLoS ONE.* (2019) 14:e0221815. doi: 10.1371/journal.pone.0221815
132. Ouyang J, Wang M, Hou Q, Feng D, Pi Y, Zhao W. Effects of dietary mulberry leaf powder in concentrate on the rumen fermentation and ruminal epithelium in fattening Hu sheep. *Animals.* (2019) 9:218. doi: 10.3390/ani9050218
133. Hu R, Zou H, Wang Z, Cao B, Peng Q, Jing X, et al. Nutritional interventions improved rumen functions and promoted compensatory growth of growth-retarded yaks as revealed by integrated transcripts and microbiome analyses. *Front Microbiol.* (2019) 10:318. doi: 10.3389/fmicb.2019.00318
134. Sun D, Bian G, Zhang K, Liu N, Yin Y, Hou Y, et al. Early-life ruminal microbiome-derived indole-3-carboxaldehyde and prostaglandin D2 are effective promoters of rumen development. *Genome Biol.* (2024) 1:64. doi: 10.1186/s13059-024-03205-x
135. Flint HJ, Duncan SH, Scott KP, Louis P. Links between diet, gut microbiota composition and gut metabolism. *Proc Nutr Soc.* (2015) 74:13–22. doi: 10.1017/S0029665114001463
136. Jami E, Mizrahi I. Composition and similarity of bovine rumen microbiota across individual animals. *PLoS ONE.* (2012) 7:e33306. doi: 10.1371/journal.pone.0033306
137. Ma J, Zhu Y, Wang Z, Yu X, Hu R, Wang X, et al. Glutamine supplementation affected the gut bacterial community and fermentation leading to improved nutrient digestibility in growth-retarded yaks. *FEMS Microbiol Ecol.* (2021) 97:fiab084. doi: 10.1093/femsec/fiab084
138. Koh A, De Vadder F, Kovatcheva P, Bäckhed F. From dietary fiber to host physiology: short-chain fatty acids as key bacterial metabolites. *Cell.* (2016) 165:1332–45. doi: 10.1016/j.cell.2016.05.041
139. Stewart RD, Auffret MD, Warr A, Walker AW, Roehle R, Watson M. Compendium of 4,941 rumen metagenome-assembled genomes for rumen

- microbiome biology and enzyme discovery. *Nat Biotechnol.* (2019) 37:953–61. doi: 10.1038/s41587-019-0202-3
140. Kaushik R, Goel A, Rout PK. Differential expression and regulation of HSP70 gene during growth phase in ruminants in response to heat stress. *Sci Rep.* (2022) 12:18310. doi: 10.1038/s41598-022-22728-6
141. Cui Y, Liu P, Yu S, He J, Afedo SY, Zou S, et al. Expression analysis of molecular chaperones Hsp70 and Hsp90 on development and metabolism of different organs and testis in cattle (cattle–yak and yak). *Metabolites.* (2022) 12:1114. doi: 10.3390/metabo12111114
142. Shen H, Fan X, Zhang Z, Xi H, Ji R, Liu Y, et al. Effects of elevated ambient temperature and local testicular heating on the expressions of heat shock protein 70 and androgen receptor in boar testes. *Acta Histochem.* (2019) 121:297–302. doi: 10.1016/j.acthis.2019.01.009
143. Su R, Chang L, Zhou T, Meng F, Zhang D. Effects of GABA on oxidative stress and metabolism in high-glucose cultured mongolian sheep kidney cells. *Int J Mol Sci.* (2024) 25:10033. doi: 10.3390/ijms251810033
144. Zhang H, Zhao F, Peng A, Dong L, Wang M, Yu L, et al. Effects of dietary L-arginine and N-Carbamylglutamate supplementation on intestinal integrity, immune function, and oxidative status in Intrauterine-Growth-Retarded suckling lambs. *J Agric Food Chem.* (2018) 66:4145–54. doi: 10.1021/acs.jafc.8b00726
145. Sindhu R, Supreeth M, Prasad SK, Thanmaya M. Shuttle between arginine and lysine: influence on cancer immunonutrition. *Amino Acids.* (2023) 55:1461–73. doi: 10.1007/s00726-023-03327-9
146. Silva JT, Miqueo E, Torrezan TM, Rocha NB, Slanzon GS, Virginio Júnior GF, et al. Lysine and methionine supplementation for dairy calves is more accurate through the liquid than the solid diet. *Animals.* (2021) 11:332. doi: 10.3390/ani11020332
147. Kheiri F, Alibeyghi M. Effect of different levels of lysine and threonine on carcass characteristics, intestinal microflora and growth performance of broiler chicks. *Ital J Anim Sci.* (2017) 16:580–7. doi: 10.1080/1828051X.2017.1302824
148. Bin P, Huang R, Zhou X. Oxidation resistance of the sulfur amino acids: methionine and cysteine. *BioMed Res Int.* (2017) 2017:1–6. doi: 10.1155/2017/9584932
149. Li P, Wu G. Roles of dietary glycine, proline, and hydroxyproline in collagen synthesis and animal growth. *Amino Acids.* (2018) 50:29–38. doi: 10.1007/s00726-017-2490-6
150. Siegert W, Rodehutsord M. The relevance of glycine and serine in poultry nutrition: a review. *Br Poult Sci.* (2019) 60:579–88. doi: 10.1080/00071668.2019.1622081
151. De Lima MB, Sakomura NK, Dorigam JCP, Da Silva EP, Ferreira NT, Fernandes JBK. Maintenance valine, isoleucine, and tryptophan requirements for poultry. *Poult Sci.* (2016) 95:842–50. doi: 10.3382/ps/pev380
152. Veira AM, Santos LS, Gobi JP, Silva WC, Fraga AZ, Hauschild L. Individual responses of growing pigs to diets with valine and isoleucine to lysine ratios. *Anim Feed Sci Technol.* (2021) 279:115037. doi: 10.1016/j.anifeedsci.2021.115037
153. Sheftel J, Loechl C, Mokhtar N, Tanumihardjo SA. Use of stable isotopes to evaluate bioefficacy of provitamin a carotenoids, vitamin a status, and bioavailability of iron and zinc. *Adv Nutr.* (2018) 9:625–36. doi: 10.1093/advances/nmy036
154. O'Dell BL. Mineral interactions relevant to nutrient requirements. *J Nutr.* (1989) 119:1832–8. doi: 10.1093/jn/119.12_Suppl.1832
155. Wolf C, Ulbrich SE, Kreuzer M, Berard J, Giller K. Differential partitioning of rumen-protected n-3 and n-6 fatty acids into muscles with different metabolism. *Meat Sci.* (2018) 137:106–13. doi: 10.1016/j.meatsci.2017.11.007
156. Lands B, Bibus D, Stark KD. Dynamic interactions of n-3 and n-6 fatty acid nutrients. *Prostag leukotr Ess.* (2018) 136:15–21. doi: 10.1016/j.plefa.2017.01.012