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# Global prevalence of *Eimeria* species in goats: a systematic review and meta-analysis

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**Background:** Coccidiosis is a protozoal disease caused by *Eimeria* species, the main symptom of which is diarrhea. *Eimeria* spp. infection can cause weight loss and ill-thrift in goats, and in severe cases, it can lead to mortality in kids, resulting in economic losses for the goat industry. This study aimed to determine the global prevalence of *Eimeria* spp. in goats and to identify the possible predictors of heterogeneity among selected studies.

**Methods:** Data were retrieved from five databases of major global importance (PubMed, Web of Science, CAB Direct, Scopus, and Google Scholar), with 255 studies published between 1963 and 2022 being included. A random-effects model was used to calculate pooled prevalence estimates with 95% confidence intervals (CI), followed by subgroup meta-analysis and meta-regression analysis to identify factors contributing to high prevalence and explore sources of heterogeneity among studies.

**Results:** The estimated global prevalence of *Eimeria* spp. in goats was 62.9% (95% CI: 58.6–67.2). Our results indicated high inter-study variability (inconsistency index ( $I^2$ ) = 99.7%, p < 0.01). Among the variables analyzed, regions and quality of studies were the most significant predictors of heterogeneity. According to the region-based subgroup meta-analysis, North America had the highest estimated prevalence of Eimeria spp. (92.2, 95% CI: 82.7-98.2), followed by Europe (86.6, 95% CI: 79.8-92.3), while Asia had the lowest prevalence (52.0, 95% CI: 45.9–58.1). Most countries (n = 42/56) had an estimated prevalence above the overall pooled estimate (>62.9%). The subgroup of studies conducted in 2000 or later presented a lower prevalence of 59.6% (95% CI: 54.7-64.3). Studies with a score of 5–7 had a significantly higher prevalence (72.4, 95% CI: 66.2-78.2) than studies with low or medium scores (p < 0.01). The prevalence of Eimeria spp. in goats detected with conventional and molecular methods was 67.3% (95% CI: 47.0-84.7). Only 47% (119/255) of the studies provided details on identifying Eimeria at the species level. Overall, more than 26 Eimeria spp. have been identified in goats globally. Among these, the most frequently reported and pathogenic species were E. arloingi (115/119), E. ninakohlyakimovae (108/119), E. christenseni (94/119), and E. caprina (71/119). Other valid species that were reported less frequently include E. alijevi, E. hirci, E. caprovina, E. aspheronica and E. jolchijevi.

**Conclusion:** These findings suggest that the pathogenic *Eimeria* spp. are widespread in goats globally. Given the high prevalence and the extensive

distribution of pathogenic *Eimeria* spp. in goats, it is recommended that integrated parasite management approaches be implemented for the effective control of coccidiosis in goats.

KEYWORDS

Eimeria, goat, global prevalence, meta-analysis, systematic review

## **1** Introduction

There are estimated to be more than one billion domestic goats worldwide (1), and the global goat population has more than doubled in the last four decades (2). Despite significant growth in the global goat population, the productivity of the goat industry is challenged by health, management and production constraints (3). Among various health concerns, gastrointestinal diseases (e.g., parasitic gastroenteritis) can lead to significant economic losses for the global goat industry (4).

For instance, coccidiosis, a parasitic disease caused by the intracellular protozoan parasites of the genus *Eimeria* (Apicomplexa: Eimeriidae) (5), is a significant concern for goat farmers due to its economic impact. Globally, several species of *Eimeria* (also known as coccidia) infect goats (6–16), leading to significant economic losses due to poor growth and lower productivity. Although the economic impact of coccidiosis is believed to be substantial (17), there is a lack of data to substantiate such a statement regarding small ruminant coccidiosis.

To date, the number of *Eimeria* spp. that are considered parasites of goats remains variable and controversial (14, 18, 19) and depends on the acceptance of the validity of certain *Eimeria* spp. (12). For instance, Levine (20) reported 13 species of *Eimeria* as the true parasites of goats, including *Eimeria arloingi, E. ninakohlyakimovae, E. christenseni, E. caprina, E. caprovina, E. alijevi, E. hirci, E. jolchijevi* and *E. aspheronica*, which are distributed globally (6–16). Among these species, *E. arloingi, E. ninakohlyakimovae, E. christenseni* and *E. caprina* are considered the most pathogenic and prevalent (21), while others are non-pathogenic.

Goats between 1 and 6 months of age are most susceptible to Eimeria spp. which inhabit the small and large intestines (22, 23). The oocysts are passed in feces, infecting other animals after further development (i.e., sporulation - asexual reproduction) in the environment. When a susceptible host ingests a sporulated oocyst, the sporozoites are released in the gastrointestinal tract and invade intestinal epithelial cells. Following a number of predetermined generations of development (schizogony - asexual reproduction) in intestinal cells, the female and male gametes form a zygote, which develops into an oocyst (24). The damage to the host occurs due to cell disruption during schizogony and later during gametogony (sexual reproduction). The more prevalent form of coccidiosis is a subclinical disease resulting in poor growth while the clinical form of the disease is most commonly characterized by diarrhea (25). The economic losses due to coccidiosis (26) are attributed to reduced productivity, reduced weight gain, mortality (22, 23) and treatment costs (27). The damage done to the kid's intestines may be permanent in severe cases, resulting in kids that exhibit ill-thrift for life, i.e., they remain "poor doers" as they fail to recover fully after treatment (25). These kids are more susceptible to other diseases, such as respiratory infections, due to their lower immunity (28).

Given that the global prevalence of Eimeria spp. in goats is variable (29-35), with more than 90% prevalence reported in some regions (11, 14, 35-39), effective control measures are essential to minimize losses associated with subclinical and clinical coccidiosis (40). Currently, control of coccidiosis is based on sound management, using preventive medications and treating clinical cases using anticoccidial drugs (23). Although coccidiosis is well-studied in poultry, sheep and cattle, our understanding of goat coccidiosis is limited despite the remarkable growth in the global goat industry in recent decades (14). As a first step to ascertain the current state of play, an exploration of the prevalence and geographical distribution of Eimeria spp. in goats could pave the way for global efforts to control goat coccidiosis. Therefore, we conducted a systematic review and meta-analysis to estimate the global prevalence of Eimeria spp. infecting goats, with an emphasis on temporal and spatial trends, frequency and spatial distribution of species, diagnostic methods, sample size and quality of selected studies. The findings of this study could be used by veterinarians, researchers, goat farmers and policymakers to make informed decisions about the effective control of coccidiosis in goats worldwide.

# 2 Materials and methods

### 2.1 Search strategy

This study was designed and analyzed according to the Preferred Reporting Items for Systematic Review and Meta-analysis (PRISMA) protocol (41) (Supplementary Table S1). Goats of any age and sex constituted the study population in this study. Two searches across four electronic databases, including PubMed, Scopus, CAB Direct, and Web of Science were performed to retrieve the maximum number of publications. A manual search was also carried out on Google Scholar and reference lists of reviews and included studies. The search queries were designed based on the medical subject headlines (MeSH) and Boolean logic. The MeSH terms and keywords were used to retrieve all relevant articles from the above databases (Supplementary Table S2). The same keywords were used in all electronic databases, and the final database search was completed on March 09, 2023.

## 2.2 Inclusion and exclusion criteria

The search results were imported into an online systematic review platform, Covidence.<sup>1</sup> Duplicate references were then

<sup>1</sup> https://app.covidence.org

removed, followed by assessment and screening in two steps (Figure 1). The first screening step involved removing irrelevant studies based on the information available from titles and abstracts. In the next step, full texts of selected studies were retrieved. If full texts were unavailable online, they were obtained through interlibrary loans from the University of Melbourne library and were also subjected to the set assessment criteria. To ensure the quality of included studies, the following inclusion criteria were used where (i) the study defined the number of goats examined and the number testing positive, (ii) the study was published in the English language, (iii) the studies contained a full text, (iv) samples indicated the prevalence of Eimeria in goats and (v) individual samples were taken from each goat (i.e., samples were not pooled). In addition, we excluded those studies that were not original research articles, involved plagiarism or used the same data for multiple publications, exhibited internal data conflict or insufficient data, only reported other parasites or Eimeria reported in host species other than goats or studies with experimental findings only (Figure 1).

## 2.3 Data extraction

The first author, EA, performed the data extraction. Any discrepancies were discussed with co-authors, who acted as secondary reviewers whenever required. The selected studies were coded, and data were collected using a format prepared in a Microsoft Excel® spreadsheet. The format included the first author's name, research type, title, year of publication, aims of the study, type of diagnostic sample, country of publication and origin of the sample, continent, diagnostic method, study design, sampling period, sampling method, sample size and methods of sample size calculation, the number of goats tested positive, prevalence, age, sex, method of Eimeria spp. identification, the identification keys used, and the number of Eimeria spp. isolated and/or reported. Data related to molecular methods and markers used for Eimeria spp. identification, and sequencing information were also extracted. During the study period, we contacted the authors of some articles to obtain more information and included unpublished data. The extracted data were checked at least twice for accuracy.



## 2.4 Quality assessment

We evaluated the quality of the eligible studies using a scoring method described previously (42–45). The method assessed specific points, including (i) random sampling, (ii) clarity of the detection method described, (iii) detailed description of the sampling method, (iv) inclusion of the sampling period, (v) calculation of sample size, (vi) aim of the study and (vii) species-level identification of *Eimeria*. Each point was scored as one, and articles were assigned to low (0–2 points), medium (3–4), or high (5–7) levels based on their scores (Supplementary Table S3). Data extracted from the studies included were summarized and edited using Microsoft Excel<sup>®</sup> spreadsheet Version 16.

## 2.5 Statistical analyses

All the analyses and visualization were performed in RStudio 4.3.1 (46) using the "meta" package (47). The bar plots were built using GraphPad Prism 10.2.0.<sup>2</sup> The mean species richness value was calculated at the country level considering the common nine *Eimeria* spp. of goats reported globally. A random effects model was used to estimate the global pooled prevalence of *Eimeria* spp. of goats, along with a 95% confidence interval (CI) (48). The estimated pooled prevalence was presented as a percentage [(number of positive samples/total samples tested) \*100] and displayed using forest plots. To stabilize the variance, we used the Freeman-Tukey double arcsine transformation (referred to as "PFT" in the "meta" package) (49).

Cochran's Q values and the inconsistency index (I2) test statistics were used to assess study heterogeneity. The I<sup>2</sup> estimates the percentage of variability in effect estimates due to heterogeneity rather than sampling error or chance differences. Therefore, the I<sup>2</sup> test measures the level of statistical heterogeneity among studies. I2 scores of 25, 50 and 75% indicate low, moderate and high degrees of heterogeneity, respectively (50, 51). To evaluate the possibility of publication bias, we utilized funnel plots, Egger's asymmetry test (52) and Begg's rank correlation test (53). In the funnel plot, we examined the symmetry of the figure, and if the dots (representing included studies) in the funnel plot were symmetrically distributed on both sides of the mid-line, it indicated no publication bias. If they were asymmetric, it suggested publication bias among the included studies. Begg's and Egger's significance tests were also employed to determine the presence of bias. The stability of this study was evaluated by the Duval and Tweedie's trim and fill analysis (54). Sensitivity analysis was conducted to verify the reliability and robustness of the meta-analysis.

The Baujat plot was used to identify sources of heterogeneity. In the Baujat plot, the horizontal (x) axis represents the contribution of each study to the general statistics of the Cochran's Q test for heterogeneity, while the vertical (y) axis represents the influence of each study on the overall estimate. The most heterogeneous studies are represented in the upper right area of the graph (55). Subgroup and meta-regression analyses were conducted to investigate the possible sources of heterogeneity. The variables included in the subgroup and meta-regression analyses were year of publication (before 2000 and 2000 or

later), region (Asia, Africa, Europe, North America, Oceania and South America), diagnostic methods (conventional, and conventional and molecular methods), the score level of studies (1–2, 3–4 and 5–7), sample size ( $n = \langle 400; n = 400-1,000; n = \rangle 1,000$ ) and country (n = 56).

Following the subgroup and univariate meta-regression analyses, a multivariate meta-regression analysis was performed on all response variables to identify the best model that explains the between-study variability in effect size estimates. In the meta-regression, the variable x represents study characteristics (such as region, country, year, sample size, score level and detection method), which are used to predict the study effect size ( $\hat{\theta}_{\hat{k}}$ ) as shown in the following model (Equation 1):

$$\hat{\theta}_{\hat{k}} = \theta + \beta x_k + \epsilon_k + \zeta_k \tag{1}$$

Where:  $\hat{\theta}_{\hat{k}}$  is the observed effect size,  $\theta$  is the intercept,  $\beta x_k$  is a predictor (or covariate)  $x_k$  with a regression coefficient  $\beta$  (fixed effect),  $\in_k$  is the sampling error and  $\zeta_k$  is the between-study error (random effect).

The first step in multivariate meta-regression analysis involved all explanatory variables in a full model. Subsequently, multi-predictor models were manually reduced using backward selection of variables until all predictors were statistically significant (p < 0.05). We constructed mixed-effects regression models (for the meta-regression analysis). We applied Akaike's information and Bayesian information criteria to compare and select the models. We assessed the goodness of fit for the meta-regression by calculating the correlation analog coefficient ( $\mathbb{R}^2$ ) using the following formula (Equation 2):

$$R^2 = \frac{\tau^2_{REM} - \tau^2_{MEM}}{\tau^2_{REM}} \tag{2}$$

Where:  $\tau^2_{REM}$  represents the estimated total heterogeneity based on the random effects model and  $\tau^2_{MEM}$  represents the total heterogeneity of the mixed effects regression model.

In all analyses, *p*-value <0.05 were used to determine a statistically significant association.

## **3** Results

## 3.1 Characteristics of eligible studies

During the literature search, 5,466 studies were retrieved from five databases (CAB Direct = 2,257, PubMed = 1,504, Scopus = 927, Web of Science = 591 and Google Scholar = 186) and one unpublished thesis, and 255 of them met the inclusion criteria (Figure 1). Most of these studies were original research papers (n = 218), followed by short communications (n = 27) and postgraduate theses (n = 4) published from 1963 to 2022 (Figure 2). An evaluation of the quality of 255 studies showed that 79 scored high (5–7), 149 medium (3–4) and 27 low (1–2) points (Supplementary Table S3).

Two hundred and fifty-five eligible studies originated from 56 countries across six continents (Figure 3) and most of them were from Asia (n = 128) followed by Africa (n = 72), Europe (n = 34) and others (n = 21). The eligible studies tested 131,407 goat fecal samples and 75,669 were positive for *Eimeria*. The apparent prevalence in studies

<sup>2</sup> www.graphpad.com





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ranged from 1.6 to 100% (Supplementary Table S4). Various diagnostic methods were used, including flotation (using saturated sodium chloride, sucrose, sodium nitrate, zinc sulfate and magnesium sulfate), the McMaster method (n = 140) in combination with direct smear (n = 28), histopathology and/or post-mortem examination (n = 11), and molecular tests (n = 10) (real-time polymerase chain reaction (qPCR), nested PCR and conventional PCR) (Supplementary Table S4).

Morphological identification of Eimeria spp. was conducted based on the morphology of sporulated oocysts. For this purpose, fecal samples that tested positive for Eimeria were artificially incubated in 2% or 2.5% potassium dichromate for several days (usually 7–10 days) at room temperature. The species of Eimeria were then identified using various parameters, including the size and shape index (SI) of oocysts, the presence/absence of a polar cap and micropyle, and the size and SI of sporocysts along with sporulation time. However, only 119 (47%) studies conducted morphological identification of Eimeria at the species level using sporulated oocysts and histopathological findings. Globally, more than 26 Eimeria spp. (Table 1; Supplementary Table S4) were recorded in goats, with recognized species, including E. arloingi, E. ninakohlyakimovae, E. christenseni, E. alijevi, E. caprina, E. hirci, E. caprovina, E. aspheronica and E. jolchijevi being the most frequently reported (Figure 4), while other species were reported less frequently (Table 1). Mixed species infections were observed in most studies, with an average or mean species richness value of 7.3 and a median of 8 Eimeria spp. reported per study (Figure 5). Only 10 studies from 8 countries corroborated their identification using molecular methods. Molecular-based studies primarily targeted the small subunit of the nuclear ribosomal RNA gene (18S rRNA), followed by mitochondrial cytochrome c oxidase 1 (COX1), and/or the first and second internal transcribed spacers of nuclear ribosomal DNA (ITS-1, ITS-2) to genetically characterize four Eimeria spp. (E. arloingi, E. ninakohlyakimovae, E. christenseni, and E. hirci) (Figure 6). As of June 25th 2024, more than 70 nucleotide sequences belonging to E. arloingi (37 sequences), E. christenseni (30 sequences), E. hirci (8 sequences), E. ninakohlyakimovae (2 sequences), and unidentified Eimeria spp. (4 sequences) were available via GenBank<sup>3</sup>. However, only a few of these sequences have been published in peer-reviewed journals.

## 3.2 Meta-analysis

A random-effects meta-analysis model was computed using the Freeman-Tukey double-arcsine transformed proportion. This model was chosen due to the expected variation among studies. Based on this model, the estimated global prevalence of *Eimeria* spp. in goats was 62.9% (95% CI: 58.6–67.2) (Figure 7; Table 2). The forest plot revealed high heterogeneity among the studies reporting the prevalence of *Eimeria* spp. ( $\tau^2 = 0.1293$ , I<sup>2</sup> = 99.7%, Q = 72,791.38, degrees of freedom = 254, *p* < 0.01). The random effects of studies were weighted similarly to the common (fixed) effects of studies, ranging from 0.0 to 9.6% (see Supplementary Figure S1). The Baujat plot shows that the study conducted by Rahman et al. (56) from India significantly impacted the pooled estimate and contributed the most to the overall heterogeneity (Figure 8A).

TABLE 1	The frequency of	of <i>Eimeria</i> spp	. of goats report	ed in the eligible
studies.				

Eimeria species	Frequency
Eimeria arloingi <sup>1</sup>	115
E. ninakohlyakimovae <sup>2</sup>	108
E. christenseni <sup>3</sup>	94
E. alijevi <sup>4</sup>	72
E. caprina ⁵	71
E. hirci <sup>6</sup>	70
E. jolchijevi <sup>7</sup>	53
E. aspheronica <sup>8</sup>	51
E. caprovina <sup>9</sup>	50
E. parva	29
E. pallida	25
E. faurei	23
E. crandallis	18
E. granulosa	17
E. ahsata	14
E. intricata	13
E. kocharli	7
E. punctuia	4
E. parbhaniensis	3
E. capralis	2
E. hawkin	2
E. charlestoni	2
E. megaembryonica	1
E. tunisiensis	1
E. masseyensi	1
E. zuernii	1
Unidentified Eimeria spp.ª	4
Total studies*	119

\* Total studies refer to studies in which *Eimeria* was identified and/or reported at species level in goats (see Supplementary Table S4 for more detail).

<sup>1-9</sup> These species are epidemiologically well-established and of major veterinary importance in the goat industry.

<sup>a</sup> Studies from India and Slovakia reported unidentified Eimeria spp.

## 3.3 Subgroup meta-analysis

The subgroup analysis revealed significant heterogeneity between studies in all subgroups. The Baujat plots illustrating the studies that contributed the most to the heterogeneity in each continent are also presented (see Figure 8). The highest prevalence was estimated in North America (92.2, 95% CI: 82.7–98.2) followed by Europe (86.6, 95% CI: 79.8–92.3) while it was the lowest in Asia (52.0, 95% CI: 45.9–58.1) (Figure 7; Table 2). Among the 56 countries included in the eligible studies, 75% (n = 42) of them had an estimated prevalence (63–100%) above the overall pooled estimate (>62.9%). On the other hand, Papua New Guinea, Saudi Arabia, and Kuwait reported the lowest estimates with pooled prevalences of 16.4% (95% CI: 7.8–28.8), 19.8% (95% CI: 18.5–21.0) and 19.8% (95% CI: 15.0–25.4), respectively (Figure 9; Supplementary Figure S2).

<sup>3</sup> www.ncbi.nlm.nih.gov



#### FIGURE 4

The spatial distribution and diversity of the valid *Eimeria* species commonly reported in goats globally. The length of each bar indicates the number of studies which reported the respective *Eimeria* spp. from each country.



Mean species richness of nine common *Limeria* species of goats reported globally. The dashed vertical line is the overall mean species richness value. See Supplementary Table S4 for more detail in each country.

The prevalence of *Eimeria* spp. in goats sampled before 2000 was 74.3% (95% CI: 65.2–82.4) and 59.6% (95% CI: 54.7–64.3) in 2000 or later, showing a statistically significant downward trend (p < 0.05). Based on sample size, the prevalence of *Eimeria* spp. was estimated to

be 55.1% (95% CI: 44.6–65.3) in studies with a sample size ranging from 400 to 1,000. In studies with a sample size greater than 1,000, the estimated prevalence was 68.2% (95% CI: 57.7–77.8). However, these differences were not statistically significant (p > 0.05). Based on the



#### FIGURE 6

Geographical distribution of studies reporting molecular characterization of *Eimeria* spp. in goats. Three molecular markers, including 18S (small subunit of the nuclear ribosomal RNA), *coxI* (mitochondrial cytochrome *c* oxidase 1) and ITS (the internal transcribed spacer of the nuclear ribosomal DNA) were used in 10 studies.

	Number of	Interaction				
Subgroup	Studies	P-value	Random Effects Model	Proportion	95%-CI	
Region						
Africa	72	< 0.01		0.64	[0.56; 0.71]	
Oceania	5			0.67	[0.39; 0.89]	
Asia	128			0.52	[0.46; 0.58]	
North America	10			0.92	[0.83; 0.98]	
South America	6			0.66	[0.35; 0.91]	
Europe	34			0.87	[0.80; 0.92]	
Year of publication						
After 2000	199	< 0.01		0.60	[0.55; 0.64]	
Before 2000	56			0.74	[0.65; 0.82]	
Sample size						
Small	167	0.18		0.65	[0.59; 0.70]	
Medium	55			0.55	[0.44; 0.66]	
Large	33			0.68	[0.58; 0.78]	
Detection method						
Conventional	245	0.66		0.63	[0.58; 0.67]	
Conventional and molecul	ar 10			0.67	[0.47; 0.85]	
Study quality						
High quality	79	< 0.01		0.72	[0.66; 0.78]	
Low guality	27		— <mark>—</mark> — !	0.46	[0.33; 0.60]	
Medium quality	149			0.61	[0.55; 0.66]	
Random effects model				0.63	[0.59; 0.67]	

#### FIGURE 7

Forest plot displaying the pooled prevalence estimates of *Eimeria* spp. in goats from subgroup meta-analysis. The "Proportion" column shows the prevalence of *Eimeria* spp. in each subgroup, while the 95% CI represents the corresponding confidence interval (CI). The dashed line represents the global pooled prevalence estimate based on the random effects model. The length of the horizontal lines represents the 95% CIs. The estimated global prevalence is the red diamond at the bottom of the plot.

TABLE 2 Global J	pooled prevalence	estimates of	Eimeria spp.	infection in goats.

Predictors		Pooled prevalence				Heterogeneity			Test for subgroup differences (REM)		
	Categories	No. of studies	Proportion	% Prevalence (95% CI)	Q	l <sup>2</sup> %	τ <sup>2</sup>	p- value	X²	p- value	
Region	Asia	128	35,570/78,625	52.0 (45.9-58.1)	33,027.96	99.6	0.1216	0.00			
	Africa	72	17,895/27,372	63.9 (56.2–71.2)	12,666.25	99.4	0.1111	0.00			
	Europe	34	17,169/18,929	86.6 (79.8-92.3)	2,396.90	98.6	0.0704	0.00	70.7	< 0.01	
	North America	10	2,626/3,032	92.2 (82.7-98.2)	695.10	98.7	0.0526	< 0.01			
	South America	6	1,062/1,590	65.7 (34.8-90.7)	857.70	99.4	0.1514	< 0.0001			
	Oceania	5	1,347/1,859	66.6 (38.9-89.2)	266.85	98.5	0.1000	< 0.01			
Year	Before 2000	56	23,337/32,917	74.3 (65.2-82.4)	15932.30	99.7	0.1411	0.00	7.8	< 0.01	
	2000 or later	199	52,332/98,490	59.6 (54.7-64.3)	52271.89	99.6	0.1211	0.00			
Sample size	Large	33	37,125/64,850	68.2 (57.7–77.8)	32625.82	99.9	0.1004	0.00			
	Medium	57	18,905/34,649	55.1 (44.6-65.3)	22092.78	99.7	0.1628	0.00	3.5	0.18	
	Small	165	19,639/31,908	64.6 (59.3-69.7)	17569.80	99.1	0.1218	0.00			
Detection	Conventional	245	73,337/127,972	62.8 (58.3-67.1)	71497.69	99.7	0.1305	0.00	0.2	0.66	
methods	Conventional & molecular	10	2,332/3,435	67.3 (47.0-84.7)	1152.20	99.2	0.1079	<0.01			
Score levels	1–2	27	4,090/7,455	46.3 (33.2-59.6)	4,123.55	99.4	0.1200	0.00			
	3-4	149	44,077/84,945	60.6 (54.6-66.5)	46,330.97	99.7	0.1414	0.00	15.1	< 0.01	
	5-7	79	27,502/39,007	72.4 (66.2–78.2)	16,942.56	99.5	0.0917	0.00			
Overall	-	255	75,669/131,407	62.9 (58.6-67.2)	72,791.38	99.7	0.1293	0.00	-	-	

CI, Confidence interval, REM: random effects model, sample size was categorized as small when n < 400, medium, n = 400-1,000 and large, n > 1,000, conventional methods refer to diagnostic tests like floatation, MacMaster, stoll egg counting, direct smear, histopathology, post-mortem, whereas molecular tests include real time polymerase chain reaction (qPCR), nested PCR and conventional PCR. Score level was categorized as low (1–2) medium (3–4) and high (5–7) quality. Q is Cochran's measure of heterogeneity,  $\tau^2$  is the variance of the effect size parameters across studies.

score level subgrouping, studies with score levels of 5–7 reported the highest prevalences (72.4, 95% CI: 66.2–78.2) compared to low or medium score levels, with statistically significant differences (p < 0.01). The prevalence of *Eimeria* spp. in goats detected using conventional and molecular methods was 67.3% (95% CI: 47.0–84.7), which was higher than that detected by conventional methods alone, but these differences were statistically non- significant (Table 2).

## 3.4 Meta-regression models

Univariate and multivariate meta-regression analyses were used to further explore the heterogeneity of data. Among the moderators considered in the univariate analysis, the region, year of publication, sample size and quality level of studies had statistically significant effects on the observed variability between the reports (Table 3). The results of univariate meta-regression indicate that the region was the covariate that contributed the most to explaining the heterogeneity of the total prevalence ( $R^2 = 15.71$ ). In the multivariate meta-regression, the first approach was to build a full model including all moderators, where almost all covariates were significant (p < 0.05) except the detection method. The  $R^2$  of the full model was 27.3. The best model included the covariates of region, year, sample size, country and score level with an  $R^2$  of 31.7 (Table 3).

In the univariate regression, a strong positive relationship was observed between the prevalence of Eimeria in goats and the European (+0.27\*\*\*) and North American (+0.58\*\*) regions. Conversely, in the multivariate approach, a negative relationship was found between Eimeria prevalence and the Oceania region  $(-0.86^{***})$ . In relation to the year of publication of the studies analyzed, both meta-regression approaches yielded similar results, indicating that studies published before 2000 were associated with a higher prevalence of Eimeria in goats. Interestingly, the sample size of the studies was identified as a significant factor contributing to the heterogeneity observed in both meta-regression approaches. This was evidenced by a negative relationship between the prevalence level and a medium-sized sample. Finally, the univariate  $(-0.26^{***})$  and multivariate  $(-0.19^{*})$  regression analyses indicated that studies with a lower quality tended to report a lower prevalence of Eimeria. However, the univariate regression also identified that medium-quality studies were associated with lower Eimeria prevalence (Table 3).

## 3.5 Publication bias assessment

In the funnel plot, all studies were symmetrically distributed (Figure 10). Additionally, the results of Egger's regression (b = 3.87; p = 0.06) and rank correlation tests (z = -1.03; p = 0.30) for funnel



plot asymmetry were not statistically significant for the global pooled estimate (Supplementary Tables S5, S6). Egger's test did not show significance, nor did the funnel plot exhibit asymmetry for the African (b = -4.88; p = 0.166) and North American continents (b = 3.7479;p = 0.736). However, publication bias was significant for the pooled prevalence estimate of Europe (b = -5.59; p = 0.02) and Asia (b = 7.42; p = 0.005) (Supplementary Table S7). Funnel plots for each continent are presented in Supplementary Figure S3. Furthermore, there was evidence of missing studies that could be included using Duval and Tweedie's trim and fill analysis, which would address the asymmetry seen in the plot (Supplementary Figure S4). Eighty-one studies were found in the trim and fill analysis, resulting in a final change to the pooled estimate (Supplementary Figure S5). A sensitivity test showed that the reconstructed data were not affected by the removal of any study, suggesting the rationality and reliability of our analysis (Supplementary Figure S6).

# 4 Discussion

This is the first study on the global prevalence of *Eimeria* spp. infection in goats using a substantial number of studies (n = 255) retrieved from five databases. The overall global prevalence of *Eimeria* spp. in goats is 62.9%. The 95% prediction interval (PI) is estimated at 4–100%. The highest prevalence was found in North America (92.2%), followed by Europe (86.6%), while it was the lowest in Asia (52.0%). Surprisingly, in 42 out of 56 countries, the

estimated prevalence (63–100%) was higher than the overall pooled estimate (62.9%).

The subgroup analysis by region and country showed that the highest pooled prevalence of Eimeria spp. was found in North America (92.2%), and the meta-regression showed a positive beta coefficient (b = 0.58). Within this region, the USA had a higher prevalence of Eimeria spp. in goats (92.4%), whereas it was lowest in Mexico (71.2%). In the USA, the higher prevalence of Eimeria spp. in goats was attributed to risk factors such as season, age, farm management, and the use of deep litter straw bedding materials (57). Additionally, the intensive production system and climate conditions contributed to the higher prevalence of *Eimeria* spp. in the USA (9). However, no studies reported the prevalence of Eimeria spp. in goats in other North American countries with significant goat populations such as Haiti and Cuba (1). The study conducted by Cantú-Martínez et al. (58) from Mexico contributed most to the observed heterogeneity on the continent (Figure 8F). Interestingly, no publication bias was detected in this region (Supplementary Table S7).

The estimated prevalence of *Eimeria* spp. in goats in Europe was 86.6%. A meta-regression analysis showed a positive beta coefficient (b = 0.27, p < 0.01) (Table 2). In Europe, 80% (12/15) of countries had a prevalence of *Eimeria* spp. higher than 70%. The lowest estimated prevalence was 57.1% in Serbia, which is still higher than the estimated prevalence of the Asian continent (52.0%). The goat sector in Europe is specialized in milk production and is highly commercially oriented, dominated by an intensive production system, as reviewed by Miller and Lu (59). This may have contributed to the higher prevalence of

	Number of	Interaction	Dandam Effects Media	Duencutie	05% 01
ubgroup	Studies	P-value	Random Effects Model	Proportion	95%-CI
ountry					
Igeria	1	0		0.89	[0.82; 0.94]
ustralia	4			0.78	[0.64: 0.89]
angladesh	2			0.47	[0.41: 0.52]
atewana	1			0.90	[0.97:0.01]
rozil	1			0.09	[0.07, 0.91]
razii	4		_	0.58	[0.16, 0.94]
ameroon	2			0.42	[0.32; 0.54]
hina	3			0.94	[0.88; 0.98]
zech Republic	3			0.94	[0.90; 0.98]
enmark	1			1.00	[0.98; 1.00]
cuador	2			0.80	[0.55; 0.97]
gypt	16			0.65	[0.52; 0.77]
ngland	1		1	0.98	[0.97: 0.99]
thiopia	8			0.67	[0.46: 0.86]
hana	2		-	0.92	[0.61: 1.00]
reece	1		-	0.61	[0.52: 0.69]
renada	1			0.01	[0.02, 0.09]
dia	70		_	0.76	[0.71, 0.01]
loia	13			0.46	[0.38; 0.54]
an	4		_	0.71	[0.31; 0.98]
aq	4			0.44	[0.20; 0.70]
aly	5			0.78	[0.53; 0.95]
ordan	1			0.54	[0.47; 0.61]
eneya	10			0.61	[0.38; 0.81]
orea	2			0.85	[0.48; 1.00]
uwait	2			0.19	[0.12: 0.26]
aos	1			0.72	[0.68: 0.76]
esotho	1		-	0.26	[0 23: 0 29]
thuania	1		-	0.20	[0.07:0.00]
lolovcio	11			0.90	[0.07, 0.99]
laraysia	11			0.05	[0.40, 0.83]
lexico	2			0.79	[0.41; 1.00]
lozambique	1			0.42	[0.29; 0.54]
lyanmar	2			0.77	[0.42; 0.99]
epal	2		·	0.87	[0.74; 0.96]
etherlands	1			0.78	[0.72; 0.84]
igeria	12			0.51	[0.31; 0.71]
akistan	8			0.45	[0.27; 0.63]
apua New Guinea	1			0.16	[0.08: 0.27]
hilinnines	2		-	0.72	[0 53 0 88]
oland	5			0.72	[0.56: 0.06]
ortugal	0			0.00	[0.00, 0.90]
onugai	2			0.99	[0.96, 1.00]
omania	1			0.95	[0.94; 0.96]
wanda	1		_ 1 =	0.83	[0.77; 0.89]
audi Arabia	5			0.46	[0.10; 0.84]
enegal	1			0.85	[0.82; 0.88]
erbia	2			0.65	[0.01; 1.00]
lovakia	1			1.00	[1.00; 1.00]
outh Africa	5			0.86	[0.52: 1.00]
nain	4			0.94	[0.82.1.00]
rilanka	1			0.94	[0.02, 1.00]
udan	2			0.07	[0.02, 0.91]
uudii	2		-	0.25	[0.11, 0.43]
anzania	1		_	0.66	[0.38; 0.88]
nailand	4			0.30	[0.08; 0.60]
urkey	6			0.74	[0.61; 0.85]
kraine	1			1.00	[0.99; 1.00]
SA	7			0.96	[0.89; 1.00]
ietnam	1		-	0.87	[0.82; 0.91]
imbabwe	2			0.69	[0.19: 1.00]
	2		-	0.00	[0.10, 1.00]
andom effects mode	el			0.63	[0.59; 0.67]
		Г			,
		-0.3	2 0 0.2 0.4 0.6 0.8	1	
andom effects mode	2	г -0.1	2 0 0.2 0.4 0.6 0.8	0. 0. 1	69 63

FIGURE 9

Forest plot displaying the pooled prevalence estimates of *Eimeria* spp. in goats stratified by country. The "Proportion" column shows the prevalence of *Eimeria* spp. in respective country, while the 95% CI represents the corresponding confidence interval (CI). The dashed vertical line denotes the global pooled prevalence estimate, derived from the random effects model. The length of the horizontal line represents the 95% CI. The white and black bars, respectively, denotes a very narrow and wide CIs. The estimated global prevalence is the red diamond at the bottom of the plot.

*Eimeria* spp. in goats reported in this region, because the intensification of dairy goat production presents challenges in limiting the spread of infectious and parasitic diseases (e.g., coccidiosis), which are

facilitated by environmental stressors such as high stocking density (60, 61). A major challenge in commercially oriented dairy goat production systems arises when multiple kidding events occur

Modelª	Covariates <sup>b</sup>	R <sup>2</sup>	Significant levels of covariates					
Univariate regression analysis								
Region	Asia	15.71	-0.02*					
	North America		+0.58**					
	South America		+0.29					
	Africa		Reference					
	Europe		+0.27***					
	Oceania		+0.03					
Year	Before 2000		+0.15**					
	2000 or later	2.83	Reference					
Sample size	Small	0.8	-0.03					
	Medium		-0.13*					
	Large		Reference					
Detection methods	Conventional	0.0	Reference					
	Conventional & molecular		+0.04					
Score level	Low quality	4.13	-0.26***					
	Medium quality		-0.12*					
	High quality		Reference					
Multivariate I	regression analysis							
Full model	Region+Year+Sample size+Detection method+Score level + country	27.27						
Best model	Region	31.71	Oceania -0.86***					
	Country		Australia +0.65* Kuwait –0.81* Sudan –0.65* Thailand –0.60*					
	Year		Before 2000 + 0.18***					
	Sample size		Medium sample size -0.21*					
	Score level		Low quality -0.19*					

TABLE 3 Univariate and multivariate approach of meta-regression of estimated global pooled prevalence of *Eimeria* spp. in goats.

<sup>a</sup>Covariates used in the univariate regression model.

<sup>b</sup>Levels of the covariates used in the univariate regression models.

R<sup>2</sup>, Amount of heterogeneity accounted for the observed variation.

p < 0.05, p < 0.01, p < 0.01, p < 0.001

throughout the year to maintain a consistent milk supply. If the same pens are used repeatedly for successive batches or if newly born kids are introduced to a pen already housing older animals, the later-born kids are immediately exposed to a heavy challenge. They can develop severe coccidiosis in the first few weeks of life (62). Moreover, herd size, age and climatic conditions were associated with varying levels of *Eimeria* spp. prevalence in Europe (40). The studies conducted by Ruiz et al. (40) in Spain and Corrias et al. (63) in Italy influenced the estimated prevalence and contributed to the observed heterogeneity (Figure 8E). We also detected the probability of publication bias in Europe (b = -5.59; p = 0.02) (Supplementary Table S7; Supplementary Figure S3), which may have further contributed to the heterogeneity.

In Oceania, the estimated prevalence of *Eimeria* spp. in goats was 66.6%. However, it is worth mentioning that almost all studies were conducted in Australia, with limited prevalence data available from countries such as Fiji, New Zealand, Vanuatu, and French Polynesia, which have a significant goat population (64). The studies conducted by O'Callaghan (13) and Al-Habsi et al. (65), had an impact on the pooled estimate and contributed to the heterogeneity (Figure 8G). The number of studies accessible in Oceania was insufficient (< 10) to perform Egger's test.

In South America, the estimated prevalence of *Eimeria* spp. in goats was 65.7%. The apparent prevalence of *Eimeria* spp. in goats ranged from 4.0 to 91.2%, and almost all studies were conducted in Brazil. This meta-analysis identified the highest prevalence in southern Ecuador and the lowest in Brazil (Supplementary Table S4). The high prevalence of *Eimeria* spp. in goats from Ecuador is often related to the animals' exposure to risk factors such as age, presence of cattle, type of pasture and body condition (66). In addition, this may be due to the typical situation in goat pens in Ecuador with moist and dark environments, ideal conditions for oocyst sporulation to occur (66). However, countries with a goat population greater than 1 million, such as Bolivia, Peru and Venezuela (1), do not have data on the prevalence of *Eimeria* spp. in goats. A study conducted by Cardoso et al. (67) from Brazil influenced the pooled estimate in this region and contributed to heterogeneity (Figure 8D).

The estimated prevalence of *Eimeria* spp. in goats in Africa was 63.9%. Despite Africa being home to over 40% of the world's goat population (1), only 72 studies from 17 countries reported the prevalence of *Eimeria* spp. in goats in Africa. The majority of these studies were conducted in five countries, including Egypt (n = 16), Nigeria (n = 12), Kenya (n = 10), Ethiopia (n = 8) and Tanzania (n = 7) (Figure 5). The highest estimated prevalence of *Eimeria* were found in Algeria (89.0%) and Botswana (89.0%), while the lowest estimate was recorded in Lesotho (26.0%). However, it is important to note that the estimates for these countries were based solely on one study, which may not accurately reflect the true prevalence of *Eimeria* spp. The studies conducted in Africa were more heterogeneous than those of other continents. The heterogeneity was influenced by studies from Egypt (68) and Tanzania (69) (Figure 8C). Egger's test did not identify publication bias in the prevalence estimates in Africa.

Asia has recognized the importance of dairy goat husbandry in the face of climate change, leading to significant investments in dairy goat projects over the past few decades [see the review by (59)]. This explains the large number of studies (n = 128) that report the prevalence of *Eimeria* spp. of goat in Asia. Interestingly, the estimated prevalence of *Eimeria* spp. in Asia, which is home to the world's largest goat population (1), is 52.0%. This result is significantly lower than the global pooled estimate (Table 2). Moreover, the meta-regression analysis showed a negative beta coefficient (b = -0.02, p < 0.05). The highest estimated prevalence in Asia was found in China (95.1%), a global leader in goat populations (1). This result differs from the previous study reported by Diao et al. (42), who estimated the pooled prevalence of *Eimeria* spp. in goats in China to be 78.7% (95% CI: 68.2–87.7%). This discrepancy



evaluates the relationship between study results and their precision.

could be due to the differences in the number of studies (70 studies) and (3 studies in present estimate) included in the meta-analysis and the study periods. Conversely, studies from Saudi Arabia, and Kuwait, Asian countries that reported the lowest prevalence of Eimeria spp., were conducted in extensively managed goat flocks and in regions with higher annual average temperatures and lower relative humidity (arid areas) (70, 71). These environmental conditions are known to negatively affect the sporulation and survival of Eimeria oocysts in the environment which could be the reason for the lower prevalence. The lower prevalence of Eimeria spp. in Saudi Arabia was also attributed to sanitation efforts in management programs introduced by goat producers or ecological differences (8). Most studies used to estimate the prevalence of Eimeria spp. in goats in Asia were conducted in India (n = 73), Malaysia (n = 11)and Pakistan (n = 8). However, countries with large goat populations, such as Bangladesh, Indonesia, Mongolia, Nepal and Myanmar (1), had limited or no available data on the prevalence of Eimeria spp. In this region, Egger's test identified statistically significant publication bias (b = 7.42; p = 0.005) and the greatest contribution to heterogeneity was shared by Rahman et al. (56) (Figure 8B).

It was believed that Eimeria spp. in goats and sheep were the same for a long time because their oocysts have strikingly similar morphologies (12, 72). However, cross-infection studies disproved this assumption in the late 20th century (73, 74). In this meta-analysis, we conducted a subgroup analysis based on the year of publication to investigate any discrepancies between studies conducted before and after the assumption was made. We also aimed to identify any changes in the temporal trend of reported cases over time. The prevalence of Eimeria spp. in goats sampled before 2000 was 74.3%, whereas in the samples collected in 2000 or later, it was 59.6%, which clearly showed a statistically significant decline (p < 0.05). The rationale behind this trend remains uncertain, but it is possible that the previous misconception concerning Eimeria spp. in goats and sheep contributed to these differences. Further comprehensive research and scientific justifications are needed to determine whether the decrease in Eimeria spp. infection in goats is genuine or not.

The antemortem diagnosis of *Eimeria* spp. infection traditionally relies on the concentration and/or quantification of *Eimeria* oocysts per gram (OPG) in the feces through microscopic examination using flotation techniques and/or the morphometry of sporulated oocysts (24, 75). Most studies (n = 202) in this review used fecal flotation techniques to detect and/or enumerate *Eimeria* fecal oocyst count. However, determining a threshold OPG value that indicates clinical coccidiosis is challenging (76). For instance, while some studies suggest that an OPG of 50,000–100,000 could indicate clinical coccidiosis, non-pathogenic *Eimeria* spp. can also be excreted in large numbers without clinical signs (76). Therefore, it is crucial to differentiate between pathogenic and non-pathogenic species to confirm clinical coccidiosis (24).

Our systematic literature review shows that morphological identification is the primary method used to differentiate Eimeria spp. in goats. Interestingly, only 47% (119/255) of the studies included in the review documented the identifications of Eimeria spp. worldwide. More than 26 Eimeria spp. have been reported in goats, including recognized/valid species such as E. arloingi, E. ninakohlyakimovae, E. christenseni, E. caprina, E. caprovina, E. alijevi, E. hirci, E. jolchijevi and E. aspheronica as well as other less characterized species. The spatial distribution of the recognized/valid Eimeria spp. shows no distinct regional pattern, suggesting their widespread presence (Figure 4). However, there is considerable variation and controversy regarding the number of Eimeria spp. parasitising goats (14, 18, 19), and this variation depends on the acceptance of certain Eimeria spp. as valid (12). Mixed species infections were commonly observed in most studies, with an average of 7.3 and a median of 8 Eimeria spp. reported per study. This review also supports the ongoing controversy that some studies have reported Eimeria spp. typically found in sheep (E. faurei, E. crandallis, and E. intricata) and cattle (E. zuernii), raising concerns about misinterpretation despite evidence of the absence of cross-host species transmission of Eimeria (73, 74). Recent studies in South Korea (77), India (78), and Slovakia (34) described unspecified Eimeria spp. Moreover, species such as E. masseyensis and

*E. charlestoni* (19) from New Zealand, *E. hawkin* and *E. charlestoni* (33, we 79) from India, *E. minasensis* (80) from Brazil, *E. sundarbanensis* (81) calc from India, *E. megaembryonica* (82) from Iraq, and *E. tunisiensis* and *E. masseyensis* (83) from Nepal have been reported despite these species never been previously recorded or described globally. *Eimeria capralis* was first reported in New Zealand (19), with subsequent the reports in Iraq (84) and Nepal (83). Furthermore, at least 14 studies allow

reused the same data and/or published their findings twice in different journals, raising significant concerns about the validity of the description of new *Eimeria* spp. or morphological identification of previously described species.

The accurate identification of *Eimeria* spp. is paramount for understanding their epidemiology and assessing the effectiveness of anticoccidial drugs, as only a few *Eimeria* spp. are pathogenic to goats (36, 76, 85). This systematic review showed that the pathogenic species of *Eimeria* (*E. arloingi, E. ninakohlyakimovae, E. christenseni* and *E. caprina*) are widely distributed. For goat farmers, the widespread presence of these pathogenic *Eimeria* spp., along with a 62.9% estimated prevalence of *Eimeria* spp. poses significant economic losses (42).

Despite the morphological characterization of sporulated oocysts being the primary method for identifying Eimeria spp. in goats, this approach has notable limitations, including low sensitivity, the extended time required (1-2 weeks) for oocysts to sporulate under varying conditions (86), labor intensive requirements, requires experienced microscopists (87) and difficulty in differentiating morphologically similar oocysts in certain species of Eimeria (12). Although more than 26 Eimeria spp. have been documented globally, molecular characterization has only been partially achieved for a few Eimeria spp., including E. arloingi, E. christenseni, E. hirci, E. ninakohlyakimovae, and one unidentified Eimeria spp. Only a limited number of studies (n = 10) across eight countries have used molecular techniques, primarily using PCR amplification of 18S, and/ or COX1, and ITS-1 or ITS-2. The lack of combined morphological and molecular methods could lead to the erroneous identification of certain Eimeria spp. in goats. As of June 25th, 2024, at least 78 nucleotide sequences have been deposited in GenBank<sup>4</sup> along with their accession numbers. Most of these sequences were based on partial amplification of 18S gene. Surprisingly, most of these nucleotide sequences (>50%) are not accompanied by peer-reviewed publications, casting doubts on their reliability and validation. Given these challenges, adopting a combined approach using morphological and molecular methods is imperative to accurately identify Eimeria spp. More nucleotide sequencing is needed, particularly for Eimeria spp. that have yet to be characterized. Furthermore, advanced molecular-based studies that include the genetic characterization of new Eimeria spp. and utilizing next-generation sequencing tools could help address the challenges associated with Eimeria.

In this systematic review and meta-analysis, there were only 27 low-quality studies with a score of 1–2, but 149 with a score of 3–4. The reasons for fewer points in some studies were: (1) they needed to clarify whether the sampling was random or not, and the sampling method needed to be detailed. (2) Furthermore, neither was the sample size calculated in most studies nor *Eimeria* reported at the species level. When investigating the prevalence of *Eimeria* spp. in goats,

we recommend researchers report and/or identify *Eimeria* spp., calculate the sample size, apply representative sampling techniques, and collect and present as much information as possible. Detailed data on potential risk factors, such as age, production systems, study period, and climatic conditions is also quite important. Such data would enhance the understanding of the factors driving *Eimeria* spp. prevalence and allow for more robust risk factor analyses in future meta-analyses.

This meta-analysis has shown high I<sup>2</sup> and Cochran's Q statistics, suggesting significant heterogeneity among the studies reporting the prevalence of Eimeria spp. in goats worldwide. The wide range of the 95% PI (4 to 100%) further supports this finding. This variation could be due to several reasons, including geographical factors, differences in production systems, the immune status of the host, differences in the age of goats included in the studies, sample size, diversity of the study populations, breed differences, sampling methods, sex and study periods (88–93). Our meta-regression analyses further showed that region, year of publication, sample size, and the quality level of studies were significant sources of heterogeneity, while the diagnostic methods did not have an impact. Although the improvement in R<sup>2</sup> was minimal (4.4%) compared to the full and best models, the full model allows for improved balance, fit and parsimony, making it the preferred model that achieves a good fit with fewer predictors. Additionally, the study by Rahman et al. (56) notably influenced the pooled estimate and contributed the most to the overall heterogeneity.

The estimated pooled prevalence reported in this systematic review and meta-analysis should be interpreted with caution due to the following limitations. Firstly, we found high heterogeneity and publication biases (for the Asian and European continents). Although we applied relevant statistical methods, these may not completely eliminate the impact of heterogeneity and publication bias on the interpretation of the pooled results. Secondly, our database search was conducted only in five databases, and the search strategy might have overlooked some research, particularly that published in languages other than English. Thirdly, we excluded a substantial number of studies published in languages other than English, which could introduce potential bias. However, considering the narrow range of the 95% CI, the missed studies are unlikely to significantly affect the present estimate. Furthermore, the wider range of the 95% PI (4-100%) is likely to encompass future primary studies reporting the prevalence of Eimeria spp. in goats. Despite these limitations, the authors strongly believe that this systematic review and meta-analysis provide a reliable reflection of the true global prevalence of Eimeria spp. in goats.

# 5 Conclusion and future work

This study presents the global prevalence of *Eimeria* spp. in goats based on data collected from approximately 30% (n = 56) of countries worldwide. The results of the meta-analysis indicate variations in the prevalence of *Eimeria* spp. in goats globally, with significant heterogeneity observed between studies. Nevertheless, the narrow range of the 95% CI (58.6–67.2%) suggests a precise and reliable estimate of the pooled prevalence of *Eimeria* spp. in goats (62.9%). This finding indicates that the included studies reported similar prevalences, instilling high confidence in the accuracy of the current estimate. However, the wide 95% PI (4–100%) reflects substantial heterogeneity and underscores the need for further studies utilizing advanced molecular tools to resolve the ongoing controversy regarding

<sup>4</sup> www.ncbi.com

the number of *Eimeria* spp. parasitising goats. Although a limited number of studies have reported the prevalence of and/or characterized *Eimeria* spp. using molecular data, this technique is known for its sensitivity and accuracy. More sensitive molecular-based approaches, such as next-generation sequencing and genomic analyses based on single oocyst isolation for mixed infections, could offer more precise insights. Our study provides the first meta-analysis of prevalence data on *Eimeria* spp. globally, thereby serving as a valuable reference for the prevention and control of *Eimeria* spp. in goats. Considering the high prevalence and the widespread presence of pathogenic *Eimeria* spp. in goats globally, it is recommended that integrated parasite management approaches be implemented for the effective control of coccidiosis in goats.

## Author contributions

EA: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Software, Validation, Visualization, Writing – original draft, Writing – review & editing. AG: Data curation, Investigation, Writing – review & editing. JA-H: Formal analysis, Writing – review & editing. MY: Formal analysis, Visualization, Writing – review & editing. CG: Resources, Writing – review & editing. IB: Supervision, Writing – review & editing. SB: Writing – review & editing. AJ: Conceptualization, Funding acquisition, Investigation, Methodology, Project administration, Resources, Supervision, Visualization, Writing – review & editing.

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# References

1. FAOSTAT. (2023). In production indices national. . (https://www.fao.org/faostat/en/#data).

2. Utaaker K, Chaudhary S, Kifleyohannes TR. Global goat! Is the expanding goat population an important reservoir of *Cryptosporidium*? *Front Vet Sci.* (2021) 8:648500. doi: 10.3389/fvets.2021.648500

3. Ruvuga PR, Maleko DD. Dairy goats' management and performance under smallholder farming systems in eastern Africa: a systematic review and meta-analysis. *Trop Anim Health Prod.* (2023) 55:255. doi: 10.1007/s11250-023-03661-w

4. Fthenakis GC, Papadopoulos E. Impact of parasitism in goat production. *Small Rumin Res.* (2018) 163:21–3. doi: 10.1016/j.smallrumres.2017.04.001

5. Taylor MA, Coop RL, Wall RL. Veterinary parasitology. Oxford, United Kingdom: Blackwell Publishing (2007).

 Balicka-Ramisz A, Ramisz A, Vovk S, Snitynskyj V. Prevalence of coccidia infection in goats in Western Pomerania (Poland) and West Ukraine region. *Ann Parasitol.* (2012) 58:167–71.

7. Hassanen EAA, Anter RGA, El-Neshwy WM, Elsohaby I. Prevalence and phylogenetic analysis of *Eimeria* species in sheep and goats in Sharkia governorate, Egypt. *Pak Vet J.* (2020) 40:437–42. doi: 10.29261/pakvetj/2020.064

8. Ibrahim MM. Prevalence of *Eimeria* species of the domestic goats *Capra hircus* Linnaeus, 1758 in Al-Baha area, Saudi Arabia. *Egypt Acad J Biol Sci B Zool.* (2012) 4:165–72. doi: 10.21608/eajbsz.2012.14297

9. Kahan TB, Greiner EC. Coccidiosis of goats in Florida, USA. Open J Vet Med. (2013) 3:209-12. doi: 10.4236/ojvm.2013.33033

10. Koudela B, Boková A. Coccidiosis in goats in the Czech Republic. Vet Parasitol. (1998) 76:261–7. doi: 10.1016/S0304-4017(97)00147-7

11. Liang G, Yang X, Liu D, Li Y, Wang J, Chen X, et al. Molecular characterization of 18S rDNA, ITS-1, ITS-2, and COI from *Eimeria christenseni* and *E. Arloingi* in goats

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# **Conflict of interest**

The authors declare that the research was conducted without any commercial or financial relationships that could be perceived as a potential conflict of interest.

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## Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fvets.2024.1537171/ full#supplementary-material

from Shaanxi Province, northwestern China. Animals. (2022) 12:1340. doi: 10.3390/ani12111340

12. Norton CC. Coccidia of the domestic goat *Capra hircus*, with notes on *Eimeria ovinoidalis* and *E. Bakuensis* (syn. *E. ovina*) from the sheep *Ovis aries*. *Parasitology*. (1986) 92:279–89. doi: 10.1017/S0031182000064052

13. O'Callaghan MG. Coccidia of domestic and feral goats in South Australia. Vet Parasitol. (1989) 30:267-72. doi: 10.1016/0304-4017(89)90095-2

14. Silva LMR, Carrau T, Vila-Viçosa MJM, Musella V, Rinaldi L, Failing K, et al. Analysis of potential risk factors of caprine coccidiosis. *Vet Parasitol Reg Stud Reports*. (2020) 22:100458. doi: 10.1016/j.vprsr.2020.100458

15. Verma R, Sharma DK, Gururaj K, Paul S, Banerjee PS, Tiwari J. Molecular epidemiology and point mutations in ITS1 and 18S rDNA genes of *Eimeria ninakohlyakimovae* and *E. christenseni* isolated from Indian goats. *Vet Parasitol Reg Stud Reports*. (2017) 9:51–62. doi: 10.1016/j.vprsr.2017.04.008

16. Waruiru RM, Gichanga EJ, Kimoro CO, Karanu FN. Prevalence of gastrointestinal nematodes, coccidia and lungworms in Ol'Magogo dairy goats. *Bull Anim Health Prod Afr.* (1994) 50:291–5.

17. Bangoura B, Bardsley KD. Ruminant coccidiosis. Vet Clin North Am Food Anim Pract. (2020) 36:187-203. doi: 10.1016/j.cvfa.2019.12.006

18. Hashemnia M, Khodakaram-Tafti A, Razavi SM, Nazifi S. Experimental caprine coccidiosis caused by *Eimeria arloingi*: morphopathologic and electron microscopic studies. *Vet Res Commun.* (2012) 36:47–55. doi: 10.1007/s11259-011-9511-9

19. Soe AK, Pomroy WE. New species of *Eimeria* (Apicomplexa: Eimeriidae) from the domesticated goat *Capra hircus* in New Zealand. *Syst Parasitol.* (1992) 23:195–202. doi: 10.1007/bf00010872

20. Levine ND. Progress in taxonomy of the apicomplexan protozoa. J Protozool. (1988) 35:518–20. doi: 10.1111/j.1550-7408.1988.tb04141.x

22. Lima JD. Coccidiose dos ruminantes domésticos. *Rev Bras Parasitol Vet.* (2004) 13:9–13.

23. Ruiz A, Guedes AC, Muñoz MC, Molina JM, Hermosilla C, Martín S, et al. Control strategies using diclazuril against coccidiosis in goat kids. *Parasitol Res.* (2012) 110:2131–6. doi: 10.1007/s00436-011-2746-0

24. Keeton STN, Navarre CB. Coccidiosis in large and small ruminants. Vet Clin North Am Food Anim Pract. (2018) 34:201–8. doi: 10.1016/j.cvfa.2017.10.009

25. Matthews J. Diseases of the goat. Hoboken, NJ: Wiley Blackwell, John Wiley & Sons Ltd. (2016).

26. de Macedo LO, Bezerra-Santos MA, de Mendonça CL, Alves LC, Ramos RAN, de Carvalho GA. Prevalence and risk factors associated with infection by *Eimeria* spp. in goats and sheep in northeastern Brazil. *J Parasit Dis.* (2020) 44:607–12. doi: 10.1007/s12639-020-01235-3

27. Fitzgerald PR. The economic impact of coccidiosis in domestic animals. *Adv Vet Sci Comp Med.* (1980) 24:121–43.

28. Andrews AH. Some aspects of coccidiosis in sheep and goats. *Small Rumin Res.* (2013) 110:93–5. doi: 10.1016/j.smallrumres.2012.11.011

29. De la Fuente C, Alunda JM. A quantitative study of *Eimeria* infections of goats from Central Spain. *Vet Parasitol.* (1992) 41:7–15. doi: 10.1016/0304-4017(92)90003-R

30. Githigia SM, Munyua WK, Kyvsgaard NC, Thamsborg SM. Helminth infection levels in goats in a semi arid area of Kenya. *Bull Anim Health Prod Afr.* (1998) 46:209–10.

31. Harper CK, Penzhorn BL. Seasonal occurrence of coccidia in a mixed herd of sheep and goats at Nebo, Northern Province, South Africa: research communication. *J South Afr Vet Assoc.* (1998) 69:93–4. doi: 10.4102/jsava.v69i3.824

32. Lima JD. Prevalence of coccidia in domestic goats from Illinois, Indiana, Missouri and Wisconsin. Int Goat Sheep Res. (1980) 1:234-41.

33. Sharma RL, Bhattacharya D, Laha R, Biswas JC, Rangarao GSC. Preliminary observations on intestinal coccidiosis in pashmina (cashmere) goats in India. *J Appl Anim Res.* (1997) 12:107–12. doi: 10.1080/09712119.1997.9706193

34. Vasilková Z, Krupicer I, Legáth J, Kovalkovicova N, Peťko B. Coccidiosis of small ruminants in various regions of Slovakia. *Acta Parasitol.* (2004) 49:272–5.

35. Zhao GH, Lei L-H, Shang C-C, Gao M, Zhao YQ, Chen C-X, et al. High prevalence of *Eimeria* infection in dairy goats in Shaanxi province, northwestern China. *Trop Anim Health Prod.* (2012) 44:943–6. doi: 10.1007/s11250-011-9997-8

36. Cavalcante ACR, Teixeira M, Monteiro JP, Lopes CWG. *Eimeria* species in dairy goats in Brazil. *Vet Parasitol.* (2012) 183:356–8. doi: 10.1016/j.vetpar.2011.07.043

37. Juszczak M, Sadowska N, Udala J. Parasites of the digestive tract of sheep and goats from organic farms in Western Pomerania, Poland. *Ann Parasitol*. (2019) 65:245–50. doi: 10.17420/ap6503.206

38. Mohamaden WI, Sallam NH, Abouelhassan EM. Prevalence of *Eimeria* species among sheep and goats in Suez governorate, Egypt. *Int J Vet Sci Med.* (2018) 6:65–72. doi: 10.1016/j.ijvsm.2018.02.004

39. Penzhorn BL, Rognlie MC, Hall LL, Knapp SE. Enteric coccidia of cashmere goats in southwestern Montana, USA. *Vet Parasitol.* (1994) 55:137–42. doi: 10.1016/0304-4017(94)90064-7

40. Ruiz A, González JF, Rodríguez E, Martín S, Hernández YI, Almeida R, et al. Influence of climatic and management factors on *Eimeria* infections in goats from semi-arid zones. J Veterinary Med Ser B. (2006) 53:399–402. doi: 10.1111/j.1439-0450.2006.00985.x

41. Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The PRISMA 2020 statement: an updated guideline for reporting systematic reviews. *BMJ*. (2021) 372:790–9. doi: 10.1136/bmj.n71

42. Diao N-C, Zhao B, Chen Y, Wang Q, Chen Z-Y, Yang Y, et al. Prevalence of *Eimeria* spp. among goats in China: a systematic review and meta-analysis. *Front Cell Infect Microbiol.* (2022) 12:222. doi: 10.3389/fcimb.2022.806085

43. Gong Q-L, Zhao W-X, Wang Y-C, Zong Y, Wang Q, Yang Y, et al. Prevalence of coccidia in domestic pigs in China between 1980 and 2019: a systematic review and meta-analysis. *Parasit Vectors*. (2021) 14:248. doi: 10.1186/s13071-021-04611-x

44. Guyatt GH, Oxman AD, Vist GE, Kunz R, Falck-Ytter Y, Alonso-Coello P, et al. GRADE: an emerging consensus on rating quality of evidence and strength of recommendations. *BMJ*. (2008) 336:924–6. doi: 10.1136/bmj.39489.470347.AD

45. Li X-M, Geng H-L, Wei Y-J, Yan W-L, Liu J, Wei X-Y, et al. Global prevalence and risk factors of *Cryptosporidium* infection in *Equus*: a systematic review and meta-analysis. *Front Cell Infect Microbiol*. (2022) 12:1072385. doi: 10.3389/fcimb.2022.1072385

46. RCoreTeam. R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing (2023).

47. Balduzzi S, Rücker G, Schwarzer G. How to perform a meta-analysis with R: a practical tutorial. *Evid Based Ment Health.* (2019) 22:153–60. doi: 10.1136/ebmental-2019-300117

48. DerSimonian R, Laird N. Meta-analysis in clinical trials revisited. *Contemp Clin Trials*. (2015) 45:139–45. doi: 10.1016/j.cct.2015.09.002

49. Freeman MF, Tukey JW. Transformations related to the angular and the square root. *Ann Math Stat.* (1950) 21:607–11. doi: 10.1214/aoms/1177729756

50. Haidich AB. Meta-analysis in medical research. Hippokratia. (2010) 14:29-37.

51. Rücker G, Schwarzer G, Carpenter JR, Schumacher M. Undue reliance on I<sup>2</sup> in assessing heterogeneity may mislead. *BMC Med Res Methodol.* (2008) 8:79. doi: 10.1186/1471-2288-8-79

52. Egger M, Smith GD, Schneider M, Minder C. Bias in meta-analysis detected by a simple, graphical test. *BMJ*. (1997) 315:629–34. doi: 10.1136/bmj.315.7109.629

53. Begg CB, Mazumdar M. Operating characteristics of a rank correlation test for publication bias. *Biometrics*. (1994) 50:1088–101. doi: 10.2307/2533446

54. Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. *Biometrics*. (2000) 56:455–63. doi: 10.1111/j.0006-341X.2000.00455.x

55. Baujat B, Mahé C, Pignon JP, Hill C. A graphical method for exploring heterogeneity in meta-analyses: application to a meta-analysis of 65 trials. *Stat Med.* (2002) 21:2641–52. doi: 10.1002/sim.1221

56. Rahman H, Papri P, Bandyopadhyay S, Chatlod LR. Epidemiology of gastrointestinal parasitism in goats in Sikkim. *Indian J Anim Sci.* (2012) 82:355–8.

57. Lloyd S, Soulsby EJ. Survey of parasites in dairy goats. Am J Vet Res. (1978) 39:1057-9.

58. Cantú-Martínez MA, González-Sáenz IS, Pereira-Berto B, Zamora-Ávila DE, Ávalos-Ramírez R, Vázquez-Cisneros KW, et al. Identificación de especies de *Eimeria* presentes en caprinos (*Capra aegagrus hircus*) en Nuevo León, Mexico. *Rev MVZ Cordoba*. (2022) 27:e2560. doi: 10.21897/rmvz.2560

59. Miller BA, Lu CD. Current status of global dairy goat production: an overview. Asian Australas J Anim Sci. (2019) 32:1219–32. doi: 10.5713/ajas.19.0253

60. Perry BD, Grace D, Sones K. Current drivers and future directions of global livestock disease dynamics. *Proc Natl Acad Sci USA*. (2013) 110:20871–7. doi: 10.1073/ pnas.1012953108

61. Ridler A. Disease threats to sheep associated with intensification of pastoral farming, NZ Vet J. (2008) 56:270–3. doi: 10.1080/00480169.2008.36846

62. Taylor MA, Coop RL, Wall RL. Veterinary parasitology. Ames, IA: Wiley Blackwell (2016).

63. Corrias F, Brajon G, Salari F, Dal Prà A, Ragona G, Lombardo A, et al. Health evaluation in the native Garfagnina goat. *Small Rumin Res.* (2012) 104:191–4. doi: 10.1016/j.smallrumres.2011.10.005

64. Asia-Pacific Association of Agricultural Research Institutions. (2021). Sheep and goats in Fiji and Papua New Guinea – success story. Available at: https://apaari.org/web/what-we-do/apcoab/publications/.

65. Al-Habsi K, Yang R, Ryan U, Miller DW, Jacobson C. Morphological and molecular characterization of three *Eimeria* species from captured rangeland goats in Western Australia. *Vet Parasitol Reg Stud Reports.* (2017) 9:75–83. doi: 10.1016/j.vprsr.2017.05.001

66. Celi K, Guzmán L, Rey-Valeirón C. Apicomplexans in goat: prevalence of *Neospora caninum, toxoplasma gondii, Cryptosporidium* spp., *Eimeria* spp., and risk factors in farms from Ecuador. *Animals.* (2022) 12:2224. doi: 10.3390/ani12172224

67. Cardoso CP, Cardozo LL, Silva BF, Amarante AFT. Gastrointestinal parasites in goats from Monte Castelo, Santa Catarina, Brazil. *Rev Bras Parasitol Vet.* (2012) 21:148–50. doi: 10.1590/S1984-29612012000200014

68. Abdel-Salam M, Mahran OM. Some biochemical changes in blood serum of balady goats infested with internal and external parasites at Assiut governorate. *Assiut Vet Med J.* (2004) 50:145–56. doi: 10.21608/avmj.2004.178957

69. Mhoma JRL, Kanyari PWN, Kagira JM. The prevalence of gastrointestinal parasites in goats in urban and peri-urban areas of Mwanza City, Tanzania. *Sci Parasitol.* (2011) 12:191–6.

70. El-Bahy MM, Omer OH, Al-Sadrani AA. Temperature difference and parasite infection at Qassim region, Saudi Arabia. *Res J Parasitol*. (2008) 3:114–22. doi: 10.3923/jp.2008.114.122

71. Mottelib AA, Haroun EM, Magzoub M, El-Basheer E. The effect of gastrointestinal parasites on blood picture in sheep and goats at Al-Gassim. *Assiut Vet Med J*. (1992) 28:215–23.

72. Reeg KJ, Gauly M, Bauer C, Mertens C, Erhardt G, Zahner H. Coccidial infections in housed lambs: oocyst excretion, antibody levels and genetic influences on the infection. *Vet Parasitol.* (2005) 127:209–19. doi: 10.1016/j.vetpar.2004.10.018

73. Lima JD. The Coccidia (Protozoa: Eimeriidae) of the domestic goat, *Capra hircus*. Illinois: University of Illinois at Urbana-Champaign (1980).

74. McDougald LR. Attempted cross-transmission of coccidia between sheep and goats and description of *Eimeria ovinoidalis* sp. n. *J Protozool*. (1979) 26:109–13. doi: 10.1111/j.1550-7408.1979.tb02741.x

75. Bangoura B, Bhuiya MAI, Kilpatrick M. *Eimeria* infections in domestic and wild ruminants with reference to control options in domestic ruminants. *Parasitol Res.* (2022) 121:2207–32. doi: 10.1007/s00436-022-07564-x

76. Chartier C, Paraud C. Coccidiosis due to *Eimeria* in sheep and goats: a review. *Small Rumin Res.* (2012) 103:84–92. doi: 10.1016/j.smallrumres.2011.10.022 77. Roh SG, Kim J, Ku B-K, Lee K. Case study: pathological and phylogenetic analysis of coccidiosis in two goats with heavy infection of unrecorded *Eimeria* sp. *Parasitol Int.* (2023) 92:102662. doi: 10.1016/j.parint.2022.102662

78. Khillare BS, Narladkar BW. Epidemiology of coccidiosis in caprines of Marathwada region of Maharashtra age, sex, breed and season wise prevalence. *J Vet Parasitol.* (2014) 28:7–13.

79. Jha D, Subramanian G. Incidence of *Eimeria* species in goats of Uttar Pradesh. *Indian Vet J.* (1966) 43:588–91.

80. Silva AC, Lima JD. *Eimeria minasensis* n. sp. (Apicomplexa: Eimeriidae) in the domestic goat *Capra hircus*, from Brazil. *Mem Inst Oswaldo Cruz*. (1998) 93:741–4. doi: 10.1590/s0074-02761998000600009

81. Bandyopadhyay PK. A new coccidium *Eimeria* sundarbanensis n. sp. (Protozoa: Apicomplexa: Sporozoea) from *Capra hircus* (Mammalia: Artiodactyla). *Protistology.* (2004) 3:223–5.

82. Al-Bayati SM, Al-Rekani AM, Hamed AA. Diagnosis of *Eimeria* spp. in *Capra ibex* (local meriz goat). *Iraqi J Vet Med.* (2016) 40:47–52. doi: 10.30539/iraqijvm. v40i1.137

83. Ghimire TR, Adhikari RB, Bhattarai N. Diversity and prevalence of *Eimeria* species in goats of Nepal. *J Hellenic Vet Med Soc.* (2021) 72:3299–306. doi: 10.12681/jhvms.29363

84. Shaheed HA, Al-Azizz S. Epidemiological characterization on eimeriosis in small ruminants in Basrah City of southern Iraq. *Plant Arch.* (2020) 20:6010–4.

85. de Macedo LO, Santos MAB, da Silva NMM, do Rêgo Barros GMM, Alves LC, Giannelli A, et al. Morphological and epidemiological data on *Eimeria* species infecting small ruminants in Brazil. *Small Rumin Res.* (2019) 171:37–41. doi: 10.1016/j. smallrumres.2018.12.006

86. Carvalho FS, Wenceslau AA, Teixeira M, Carneiro JAM, Melo ADB, Albuquerque GR. Diagnosis of *Eimeria* species using traditional and molecular methods in field studies. *Vet Parasitol.* (2011) 176:95–100. doi: 10.1016/j. vetpar.2010.11.015

87. Khodakaram-Tafti A, Hashemnia M, Razavi SM, Sharifiyazdi H, Nazifi S. Genetic characterization and phylogenetic analysis of *Eimeria arloingi* in Iranian native kids. *Parasitol Res.* (2013) 112:3187–92. doi: 10.1007/s00436-013-3494-0

88. Balicka-Ramisz A. Studies on coccidiosis in goats in Poland. *Vet Parasitol.* (1999) 81:347–9. doi: 10.1016/S0304-4017(98)00258-1

89. Harper CK, Penzhorn BL. Occurrence and diversity of coccidia in indigenous, Saanen and crossbred goats in South Africa. *Vet Parasitol.* (1999) 82:1–9. doi: 10.1016/S0304-4017(98)00266-0

90. Hashemnia M, Khodakaram-Tafti A, Razavi SM, Nazifi S. Changing patterns of acute phase proteins and inflammatory mediators in experimental caprine coccidiosis. *Korean J Parasitol.* (2011) 49:213–9. doi: 10.3347/kjp.2011.49.3.213

91. Khodakaram-Tafti A, Hashemnia M. An overview of intestinal coccidiosis in sheep and goats. *Rev Med Vet.* (2017) 168:1–3.

92. Ruiz A, Matos L, Muñoz MC, Hermosilla C, Molina JM, Andrada M, et al. Isolation of an *Eimeria ninakohlyakimovae* field strain (Canary Islands) and analysis of its infection characteristics in goat kids. *Res Vet Sci.* (2013) 94:277–84. doi: 10.1016/j.rvsc.2012.08.003

93. Zvinorova PI, Halimani TE, Muchadeyi FC, Matika O, Riggio V, Dzama K. Prevalence and risk factors of gastrointestinal parasitic infections in goats in low-input low-output farming systems in Zimbabwe. *Small Rumin Res.* (2016) 143:75–83. doi: 10.1016/j.smallrumres.2016.09.005