



Physiologic Factors Influencing the Arterial-To-End-Tidal CO₂ Difference and the Alveolar Dead Space Fraction in Spontaneously Breathing Anesthetised Horses

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OPEN ACCESS

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Specialty section:

This article was submitted to
Veterinary Surgery and
Anesthesiology,
a section of the journal
Frontiers in Veterinary Science

Received: 01 November 2017

Accepted: 12 March 2018

Published: 28 March 2018

Citation:

Mosing M, Böhm SH, Rasis A, Hoosgood G, Auer U, Tusman G, Bettschart-Wolfensberger R and Schramel JP (2018) Physiologic Factors Influencing the Arterial-To-End-Tidal CO₂ Difference and the Alveolar Dead Space Fraction in Spontaneously Breathing Anesthetised Horses. *Front. Vet. Sci.* 5:58. doi: 10.3389/fvets.2018.00058

The arterial to end-tidal CO₂ difference (P_{(a-ET)CO₂}) and alveolar dead space fraction (VDalv_{frac} = P_{(a-ET)CO₂}/PaCO₂), are used to estimate Enghoff's "pulmonary dead space" (V/Q_{Eng}), a factor which is also influenced by venous admixture and other pulmonary perfusion abnormalities and thus is not just a measure of dead space as the name suggests. The aim of this experimental study was to evaluate which factors influence these CO₂ indices in anesthetized spontaneously breathing horses. Six healthy adult horses were anesthetized in dorsal recumbency breathing spontaneously for 3 h. Data to calculate the CO₂ indices (response variables) and dead space variables were measured every 30 min. Bohr's physiological and alveolar dead space variables, cardiac output (CO), mean pulmonary pressure (MPP), venous admixture (Q_s/Q_t), airway dead space, tidal volume, oxygen consumption, and slope III of the volumetric capnogram were evaluated (explanatory variables). Univariate Pearson correlation was first explored for both CO₂ indices before V/Q_{Eng} and the explanatory variables with rho were reported. Multiple linear regression analysis was performed on P_{(a-ET)CO₂} and VDalv_{frac} assessing which explanatory variables best explained the variance in each response. The simplest, best-fit model was selected based on the maximum adjusted R² and smallest Mallows' p (C_p). The R² of the selected model, representing how much of the variance in the response could be explained by the selected variables, was reported. The highest correlation was found with the alveolar part of V/Q_{Eng} to alveolar tidal volume ratio for both, P_{(a-ET)CO₂} (r = 0.899) and VDalv_{frac} (r = 0.938). Venous admixture and CO best explained P_{(a-ET)CO₂} (R² = 0.752; C_p = 4.372) and VDalv_{frac} (R² = 0.711; C_p = 9.915). Adding MPP (P_{(a-ET)CO₂}) and airway dead space (VDalv_{frac}) to the models improved them only marginally. No "real" dead space variables from Bohr's equation contributed to the explanation of the variance of the two CO₂ indices. P_{(a-ET)CO₂} and VDalv_{frac} were closely associated with the alveolar part of V/Q_{Eng} and as such, were also influenced by variables representing a dysfunctional pulmonary perfusion. Neither P_{(a-ET)CO₂} nor VDalv_{frac} should be considered pulmonary dead space, but used as global indices of V/Q mismatching under the described conditions.

Keywords: equine, spontaneous ventilation, volumetric capnography, airway dead space, pulmonary perfusion

INTRODUCTION

Ventilation/perfusion (V/Q) mismatch develops rapidly in anesthetized horses (1). Venous admixture and pulmonary dead space mark the two “extremes” of the spectrum of this mismatch; the former representing perfused but not ventilated alveoli ($V/Q = 0$) while the latter representing alveoli which are only ventilated but not perfused ($V/Q = \infty$).

Several oxygen indices including arterial-to-alveolar partial pressure of oxygen (PO_2) difference and P_aO_2 /inspiratory fraction of oxygen (FiO_2) ratio or f -shunt have been tested to estimate venous admixture in people and horses (2) and are frequently applied in clinical routine. The index used to estimate physiologic dead space is the difference between the arterial and end-tidal partial pressure of CO₂ [$P_{(a-ET)CO_2}$], which was evaluated in spontaneously breathing and ventilated people several decades ago (3). The $P_{(a-ET)CO_2}$ was divided by P_aCO_2 and denoted as an estimate of alveolar dead space, and called alveolar dead space fraction [$VD_{alv\frac{frac}{frac}} = P_{(a-ET)CO_2}/P_aCO_2$] (4). The concept of using P_aCO_2 and $P_{ET}CO_2$ to estimate “dead space” was adapted in equine anesthesia over 40 years ago (5, 6) and has been used extensively ever since to estimate “dead space” in many animal species (7).

“True” dead space measurements, however, include the measurement of mixed-expired partial pressure of CO₂ (P_ECO_2) and the mean alveolar CO₂ (P_aCO_2) (8). Bohr used these two CO₂ measurements to calculate the famous physiologic dead space ratio [$VD_{Bohr} = (P_aCO_2 - P_ECO_2)/P_aCO_2$]. As P_aCO_2 was difficult to obtain, Enghoff suggested more than 50 years later that the partial pressure of CO₂ in the arterial blood would be representative of the P_aCO_2 . Therefore, he modified Bohr’s formula substituting P_aCO_2 by P_aCO_2 to derive the following formula: $VD_{Eng} = (P_aCO_2 - P_ECO_2)/P_aCO_2$ (9). What Enghoff failed to account for is the fraction of shunting blood volume that cannot unload CO₂ during lung passage and thereby adding CO₂-rich blood to the arterial side (10). This fact is the reason why P_aCO_2 almost invariably exceeds P_aCO_2 . The contribution of this blood flow equals venous admixture and has been shown to cause VD_{Eng} to overestimate the “true” physiologic dead space (11–13). On a positive note, it should be used as a global index of V/Q mismatching since VD_{Eng} includes pulmonary dead space and venous admixture alike (10, 12). It is important to note that all studies concluding that $P_{(a-ET)CO_2}$ and $VD_{alv\frac{frac}{frac}}$ is valid representative of pulmonary dead space used VD_{Eng} as their reference values for dead space (3–5). As VD_{Eng} does not represent the “true” physiologic dead space, it is not surprising that more recent studies show the unreliability of $P_{(a-ET)CO_2}$ and $VD_{alv\frac{frac}{frac}}$ to predict dead space or changes thereof (14–16). Following aforementioned assumptions the main factors causing an inaccuracy in dead space calculation using $P_{(a-ET)CO_2}$ and $VD_{alv\frac{frac}{frac}}$ are venous admixture and atelectasis formation (13, 17, 18). Furthermore, other physiologic factors like tidal volume, oxygen consumption, respiratory rate, and homogeneity of lung emptying have been shown to change the difference between the arterial and the end-tidal CO₂ values (14, 15, 19). To account for the fact that Enghoff’s “dead space” is influenced by $V/Q = \infty$, $V/Q = 0$ and the global V/Q mismatch, for clarity, the term will be referred to as Enghoff’s approach (V/Q_{Eng}) in this manuscript.

Based on the above considerations, there is a clinical need for a reliable and yet easy to obtain alternative index of dead space overcoming the time consuming difficult measurement of CO₂ variables needed for Bohr’s equation. Until two decades ago, mixed-expired CO₂, which is employed in Bohr’s equation and Enghoff’s approach, could only be measured by using a mixing chamber or Douglas bag (20). More recently, volumetric capnography (VCap) provides an easier and clinically more convenient way to measure P_ECO_2 . To obtain the VCap curve the partial pressure of CO₂ is plotted over the expired tidal volume—therefore the term VCap (13). P_ECO_2 is the average of the partial pressure of CO₂ during exhalation. It is calculated from the area under the curve (the integral of the VCap curve during exhalation) divided by tidal volume (10). Beside P_ECO_2 also P_aCO_2 and airway dead space can be derived from the VCap curve (Figure 1) (21). For this evaluation, the exhaled gas flow and partial pressure of CO₂ needs to be measured simultaneously. Due to the time delay of the CO₂ sensor in side-stream capnography, a mainstream CO₂ sensor combined with a pneumotachograph close to the endotracheal tube (ETT) is the most commonly used set-up to measure VCap. The VCap monitors developed for humans cannot cope with the high inspiratory and expiratory flow rates in horses. In order to allow the use of these pneumotachographs during equine anesthesia a custom-made flow-partitioning device can be inserted between ETT and the Y-piece of the anesthetic breathing circle (22). This device splits the flow into four equal parts, making it possible to calculate the correct volume by multiplying the measured values by four.

The aim of this study was to discern factors that best explain the variability in $P_{(a-ET)CO_2}$ and $VD_{alv\frac{frac}{frac}}$ in anesthetized spontaneously breathing horses in dorsal recumbency. We hypothesized that: (1) $P_{(a-ET)CO_2}$ and $VD_{alv\frac{frac}{frac}}$ is strongly associated with V/Q_{Eng} and derived variables, but much less with “true” dead space

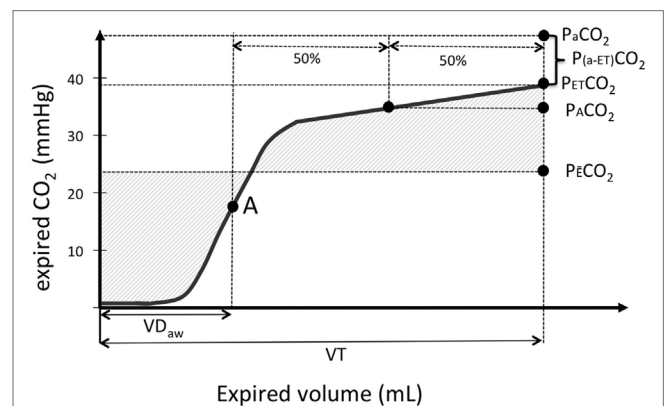


FIGURE 1 | Graphical illustration of a volumetric capnography (VCap) curve where the partial pressure of expired CO₂ (Exp. CO₂) is plotted over the volume of one tidal breath (VT). “A” is the inflection point of VCap that separates per definition the airways from alveolar compartment and is used to calculate airway dead space (VD_{aw}). P_aCO_2 is the CO₂ value at the midpoint between the inflection point “A” and the end-tidal CO₂ ($P_{ET}CO_2$). The mixed-expired CO₂ (P_ECO_2) represents the mean CO₂ in the expired gas. The difference between $P_{ET}CO_2$ and arterial oxygen partial pressure (P_aCO_2) is plotted as well.

variables derived from Bohr's original formula, (2) $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$ is highly influenced by venous admixture and other factors influencing lung perfusion.

MATERIALS AND METHODS

This experimental study had ethical approval by the local committee for animal experimentation of the Swiss government (TV-4985). It was part of an original study performed to evaluate the effects of CPAP in anesthetized horses (23). Only those horses in which no CPAP was applied were included in this current study.

Animals

Six horses were included in this experimental study out of the control group of a former study (23). Horses were acquired from local breeders and housed in boxes at the hospital for at least a week before commencing this study. All horses were considered healthy based on thorough clinical examination, routine blood work, and arterial blood gas analysis before anesthesia. They were fasted for 12 h, but had free access to water until 2 h prior to induction.

Before premedication, a 8F 110-cm angiography balloon catheter (Arrow Swiss, Edwards Lifesciences, Switzerland) was placed *via* the jugular vein in the pulmonary artery using pressure guidance to determine correct positioning.

Anesthesia

Premedication consisted of medetomidine (0.007 mg kg⁻¹) (Dorbene, Graeb AG, Switzerland) IV and phenylbutazone (4 mg kg⁻¹) IV (Butadion, Streuli Pharma AG, Switzerland). Anesthesia was induced with diazepam (0.02 mg kg⁻¹) (Valium, Roche, Switzerland) and ketamine (2 mg kg⁻¹) (Ketanarkon 100, Streuli Pharma AG, Switzerland) IV. After induction, a 26-mm internal diameter cuffed silicone ETT was inserted into the trachea and connected to a circle breathing system (Tafonius, Vetronic Services Ltd., UK) after placing the horse on a mattress in dorsal recumbency. Anesthesia was maintained using isoflurane (Attane Isoflurane, Provet, Switzerland) in an oxygen/air mixture (inspiratory O₂ fraction: $FiO_2 = 0.5$). Medetomidine was infused as a continuous rate infusion (0.0035 mg kg⁻¹ h⁻¹). Ketamine (50–100 mg bolus) was administered IV in case of spontaneous movement.

Two multi-parameter monitors (Datex-Ohmeda S/3 Anesthesia Monitor, Datex-Ohmeda and Solomon monitoring system, Vetronics, UK) were used to measure standard cardiopulmonary parameters throughout anesthesia as well as mean pulmonary artery pressure. The pressure transducer was zeroed to atmospheric pressure and leveled at the height of the shoulder joint. New pressure transducers were used for each horse.

Ringer's lactate (Ringer-Lactat Fresenius, Fresenius Kabi, Switzerland) was infused at a rate of 10 mL kg⁻¹ h⁻¹ and hydroxyethyl starch (HAES-steril 10% ad us. vet., Fresenius Kabi, Switzerland) was started 2 h after induction (1 mL kg⁻¹ h⁻¹) to account for the expected fluid losses due to medetomidine. Dobutamine was infused intravenously to maintain mean arterial pressure between 75 and 85 mmHg.

If arterial PCO₂ (P_aCO_2) exceeded 100 mmHg (13.3 kPa) at any measurement point horses were excluded from further data acquisition and mechanical ventilation was started.

In the original study all horses were anesthetized for 6 h but only data from the first 3 h were used in this study. At the end of the study period, horses were allowed to recover unassisted.

Monitoring and Data Collection

All cardiovascular data including mean pulmonary pressure (MPP) were recorded every 30 min during the first 3 h of anesthesia (T30, T60, T90, T120, T150, T180). Cardiac output (CO) was measured in triplicate at T60, T120, and T180 using the lithium dilution technique (LiDCO; LiDCO Group, UK) (24).

A flow-partitioning device was placed between the ETT and the Y-piece of the circle system dividing the total flow into four parts by two flow splitting adapters connected to each other by means of four silicon tubes equipped with four identical human adult NICO capnograph connectors (Respironics, Wallingford, CT, USA) (22). These connectors include a fixed-orifice resistance for flow measurement and a connection site for the mainstream sensor of the capnography unit of the NICO. Only one of the four connectors was connected to the spirometer and mainstream capnography unit of the NICO.

At time points of cardiovascular evaluation, VCap and spirometry data were recorded for 3 min using the dedicated software Datacoll (Respironics, Wallingford, CT, USA). Furthermore, simultaneous arterial and mixed venous blood samples were taken under anaerobic conditions at each of these time points and PO₂, carbon dioxide tension (PCO₂) and arterial hemoglobin concentration ([Hb]) analyzed immediately (Rapidpoint, Siemens, Switzerland).

All devices were calibrated following manufacturer's guidelines before each anesthesia; the capnograph of the NICO device was calibrated with room air before each experiment (infrared sensor with a response time <60 ms and accuracy of ± 2 mmHg) and the accuracy of the pneumotachograph was verified with a 100-mL calibration syringe before and after all measurements (allowed accuracy of $\pm 3\%$ following manufacturers guidelines for the calibration set-up). The accuracy of the volume measurement of the flow-partitioning device used in our set-up in connection with the NICO device was verified in an *in vitro* experiment published elsewhere (25). The difference between the reading of the flow-partitioning device in combination with the NICO device and the 10 L calibration syringe was $0.73 \pm 4.3\%$ (mean \pm SD).

Data Analysis

After importing CO₂ values into an Excel spread sheet, $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$ were calculated. VCap data were imported into a custom-made macro routine in Excel (Excel; Microsoft Corporation, WA, USA) and the recorded breaths analyzed. The mean value of all recorded breaths per time point and horse was used for statistical analysis. Bohr's dead space ratio (VD_{Bohr}) and V/Q_{Eng} were calculated following curve approximation by the solver function according to Tusman et al. (26). The ratio of alveolar dead space (difference between physiologic VDs and VD_{aw}) to alveolar tidal volume was calculated based on Bohr's ($VD_{alv,Bohr}/VT_{alv}$) and Enghoff's ($V/Q_{alv,Eng}/VT_{alv}$) respective

equations. Airway dead space was defined as the volume at the inflection point of the VCap curve and was normalized by the tidal volume (VD_{aw}/VT). The slope of phase III of the volumetric capnogram (SIII) was determined. Venous admixture ($\dot{Q}s/\dot{Q}t$) and oxygen consumption (VO_2) were calculated using standard equations (27).

Statistics

Continuous data were summarized and reported as mean \pm SD and range. All continuous variables were deemed to follow a normal distribution based on Q-Q plots and failure to reject the null hypothesis of normality was tested using Shapiro-Wilk's statistic at $P \leq 0.05$.

For the purpose of explaining the variance of the responses $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$, the explanatory variables were grouped according to what physiologic state they best represented; VD_{Bohr} , $VD_{alv,Bohr}/VT_{alv}$, and VD_{aw}/VT were considered "true" dead space variables; CO and MPP were considered variables related to pulmonary blood flow; $\dot{Q}s/\dot{Q}t$ was considered a functional variable mirroring collapsed lung areas with low V/Q causing CO₂ admixture to the arterial side; SIII was treated as a factor characterizing homogeneity of lung emptying; and VT and VO_2 were considered possible explanatory variables based on evidence in the literature.

Initial explorations of the association between Enghoff's variables (V/Q_{Eng} , $V/Q_{alv,Eng}/VT_{alv}$) and $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$ were performed using Pearson's correlation with rho reported for each variable. All data points from each horse were used. Multiple linear regression analysis was then performed on $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$, assessing all combinations of explanatory variables to determine which explanatory variables best explained the variance of each response. The simplest, best-fit model was selected based on the maximum adjusted R^2 and the smallest Mallows's

p (C_p). The R^2 of the selected model, representing how much of the variance in the response could be explained by the variance of the selected variables, was reported. Enghoff's approach and the associated variables (representing high and low V/Q ratios) were not included in the multi-variate analysis as the aim of this analysis was to show if the two indices represent $V/Q = \infty$ and areas of high V/Q ratio, or $V/Q = 0$ and areas of low V/Q ratio.

RESULTS

Six healthy adult horses with a mean (SD) age of 10 ± 6 years and body weight of 539 ± 35 kg (one Thoroughbred, five Freiberger) completed the 3 h study period and complete data sets of all horses could be analyzed.

Respiratory variables, VCap, and specific cardiovascular data of all horses are given in **Table 1**.

The $P_{(a-ET)}CO_2$ ranged from -1 to 39 mmHg with the highest values at T120 (one horse), T150 (two horses), and T180 (three horses).

There was a high correlation of $V/Q_{alv,Eng}/VT_{alv}$ and a moderate correlation of V/Q_{Eng} , $\dot{Q}s/\dot{Q}t$, and CO with $P_{(a-ET)}CO_2$ (**Figure 2; Table 2**).

The variance of $P_{(a-ET)}CO_2$ was best explained by $\dot{Q}s/\dot{Q}t$ and CO, accounting for 75% ($R^2 = 0.7516$). Adding MPP to the model improved the explanation by additional 2% ($R^2 = 0.7860$) (**Table 3**).

$VD_{alv,frac}$ ranged from -0.02 to 0.36 with the highest values observed at the same time points as the highest $P_{(a-ET)}CO_2$, except in one horse. This horse had the highest $P_{(a-ET)}CO_2$ earlier than the others showing consistently lower COs than all other horses during the entire study period.

Correlation of $VD_{alv,frac}$ with $V/Q_{alv,Eng}/VT_{alv}$ was high and moderate with V/Q_{Eng} and $\dot{Q}s/\dot{Q}t$ (**Figure 3; Table 2**).

TABLE 1 | Mean \pm SD of the arterial to end-tidal CO₂ difference, alveolar dead space fraction, dead space and cardiorespiratory parameters in six spontaneously breathing horses under general isoflurane anesthesia in dorsal recumbency at different time points (T30–180).

	T30	T60	T90	T120	T150	T180
$P_{(a-ET)}CO_2$	13.14 \pm 8.42	11.15 \pm 6.90	12.07 \pm 4.34	13.39 \pm 4.95	21.93 \pm 8.89	21.40 \pm 8.97
$VD_{alv,frac}$	0.20 \pm 0.12	0.17 \pm 0.09	0.19 \pm 0.05	0.20 \pm 0.07	0.27 \pm 0.08	0.26 \pm 0.07
P_ECO_2	48.54 \pm 3.96	49.42 \pm 3.59	51.80 \pm 4.41	54.02 \pm 4.37	56.57 \pm 4.14	58.91 \pm 8.13
$PaCO_2$	61.68 \pm 8.90	58.66 \pm 6.55	63.87 \pm 7.29	67.33 \pm 5.79	77.43 \pm 10.85	80.32 \pm 14.49
ETCO ₂	48.54 \pm 3.93	49.42 \pm 3.59	51.80 \pm 4.41	54.02 \pm 4.37	56.57 \pm 4.14	58.91 \pm 8.13
V/Q_{Eng}	0.55 \pm 0.06	0.49 \pm 0.01	0.52 \pm 0.04	0.51 \pm 0.03	0.56 \pm 0.06	0.57 \pm 0.06
$V/Q_{alv,Eng}/VT_{alv}$	0.26 \pm 0.11	0.23 \pm 0.07	0.26 \pm 0.04	0.27 \pm 0.06	0.33 \pm 0.09	0.33 \pm 0.06
VD_{Bohr}	0.39 \pm 0.08	0.35 \pm 0.04	0.37 \pm 0.04	0.35 \pm 0.04	0.37 \pm 0.05	0.38 \pm 0.06
$VD_{alv,Bohr}/VT_{alv}$	0.02 \pm 0.02	0.03 \pm 0.01	0.03 \pm 0.01	0.04 \pm 0.01	0.04 \pm 0.01	0.03 \pm 0.01
VD_{aw}/VT	0.38 \pm 0.10	0.33 \pm 0.05	0.35 \pm 0.04	0.33 \pm 0.05	0.34 \pm 0.05	0.36 \pm 0.07
CO		28.16 \pm 4.91		39.85 \pm 6.64		46.10 \pm 7.88
MPP	12.20 \pm 4.87	14.50 \pm 5.01	12.60 \pm 7.70	14.33 \pm 6.50	13.83 \pm 5.34	12.00 \pm 5.93
$\dot{Q}s/\dot{Q}t$	0.23 \pm 0.11	0.28 \pm 0.10	0.31 \pm 0.11	0.32 \pm 0.09	0.34 \pm 0.11	0.34 \pm 0.10
VO_2		104 \pm 16		115 \pm 14		111 \pm 18
VT	5,876 \pm 2,135	6,342 \pm 1,492	5,985 \pm 631	6,470 \pm 1,197	6,427 \pm 1,245	6,085 \pm 1,206
SIII	0.006 \pm 0.006	0.003 \pm 0.001	0.003 \pm 0.001	0.002 \pm 0.001	0.003 \pm 0.001	0.004 \pm 0.002

$P_{(a-ET)}CO_2$ (mmHg), arterial to end-tidal CO₂ partial pressure difference; $VD_{alv,frac}$, alveolar dead space fraction; P_ECO_2 (mmHg), mixed-expired partial pressure of CO₂; $PaCO_2$ (mmHg), arterial partial pressure of CO₂; ETCO₂ (mmHg), end-tidal partial pressure of CO₂; V/Q_{Eng} , Enghoff's dead space ratio; $V/Q_{alv,Eng}/VT_{alv}$, alveolar dead space as a ratio to alveolar tidal volume calculated from VD_{Eng} ; VD_{Bohr} , Bohr's dead space ratio; $VD_{alv,Bohr}/VT_{alv}$, alveolar dead space as a ratio to alveolar tidal volume calculated from VD_{Bohr} ; VD_{aw}/VT , airway dead space per tidal volume; CO (L/min), cardiac output; MPP (mmHg), mean pulmonary pressure; $\dot{Q}s/\dot{Q}t$, venous admixture; VO_2 (mL/min), oxygen consumption; V_T (L), tidal volume; SIII (mmHg/mL), slope of phase III of the volumetric capnogram.

The variance of $VD_{alv,frac}$ was best explained by $\dot{Q}s/\dot{Q}t$ and CO accounting for 71% of it ($R^2 = 0.7115$). Adding VD_{aw}/VT to the model improved the explanation by another 3% ($R^2 = 0.7444$) (Table 3).

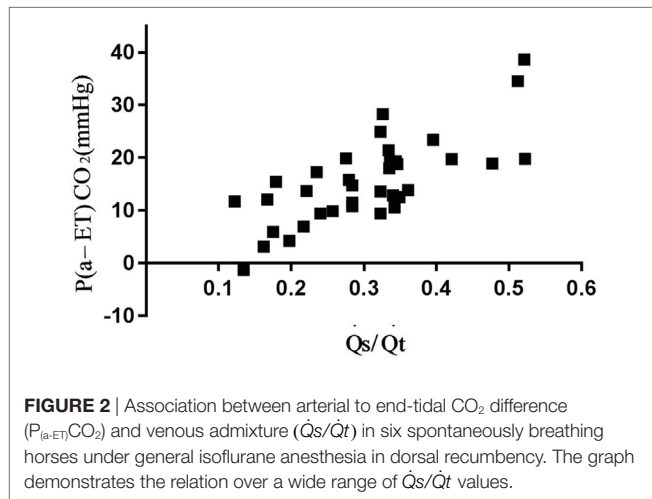


TABLE 2 | Pearson's correlation coefficient (rho) for the arterial to end-tidal CO₂ difference and alveolar dead space fraction with dead space and cardiorespiratory parameters in six spontaneously breathing horses under general isoflurane anesthesia in dorsal recumbency.

	$P_{(a-ET)}CO_2$	$VD_{alv,frac}$
V/Q_{Eng}	0.718	0.659
$V/Q_{alv,Eng}/VT_{alv}$	0.899	0.938
VD_{Bohr}	0.121	0.262
$VD_{alv,Bohr}/VT_{alv}$	0.188	0.207
VD_{aw}/VT	0.130	0.247
CO	0.773	0.288
Mean pulmonary pressure	0.161	0.159
$\dot{Q}s/\dot{Q}t$	0.694	0.684
VO_2	0.045	0.084
VT	0.418	0.317
SIII	0.116	0.130

Descriptions, see Table 1.

TABLE 3 | The results of multi linear regression analysis for factors associated with the arterial to end-tidal CO₂ difference ($P_{(a-ET)}CO_2$) and alveolar dead space fraction ($VD_{alv,frac}$) in six spontaneously breathing horses under general isoflurane anesthesia in dorsal recumbency.

Selected variable	$P_{(a-ET)}CO_2$			$VD_{alv,frac}$		
	R^2	Adjusted R^2	C_p	R^2	Adjusted R^2	C_p
$\dot{Q}s/\dot{Q}t$	0.626	0.603	10.62	0.632	0.609	13.92
CO	0.130	0.075	43.35	0.083	0.083	55.68
VD_{aw}/VT	0.002	-0.060	51.75	0.049	0.049	58.28
MPP	0.192	0.142	39.24	0.138	0.138	51.47
$\dot{Q}s/\dot{Q}t + CO$	0.752	0.718	4.37	0.711	0.673	9.91
$\dot{Q}s/\dot{Q}t + CO + MPP$	0.786	0.740	4.10			
$\dot{Q}s/\dot{Q}t + CO + VD_{aw}/VT$				0.744	0.690	9.42

Venous admixture ($\dot{Q}s/\dot{Q}t$) and CO best explained the variance in the responses, based on the adjusted R^2 and Mallows's p (C_p). The overall R^2 of the selected model is reported, representing how much of the variance in the response is explained by the selected variables.

$P_{(a-ET)}CO_2$ (mmHg), arterial to end-tidal CO₂ partial pressure difference; $VD_{alv,frac}$, alveolar dead space fraction; $\dot{Q}s/\dot{Q}t$, venous admixture; CO (L/min), cardiac output; VD_{aw}/VT , airway dead space per tidal volume; MPP (mmHg), mean pulmonary pressure.

Neither Bohr's dead space variables, nor SIII, VT, or VO_2 showed any significant correlation or provided any further explanation of the variance of $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$.

DISCUSSION

The results of this study support the first hypothesis in that both indices, $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$ can be used to estimate Enghoff's approach, but do not represent "true" dead spaces as derived from Bohr's equation in spontaneously breathing horses. The results of this study also support the second hypothesis in that venous admixture and factors related to pulmonary perfusion, like CO and pulmonary pressure influence $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$. Therefore, $P_{(a-ET)}CO_2$ and $VD_{alv,frac}$ did not represent $V/Q = \infty$ (physiologic or alveolar dead space) in these spontaneously breathing horses in dorsal recumbency, but a global mismatch in the pulmonary ventilation/perfusion ratio, a finding supported by studies in other species.

Looking at the difference between $PaCO_2$ and $ETCO_2$ from the theoretical side every increase of the difference can have two causes: (1) a decrease in $ETCO_2$ relative to $PaCO_2$ or (2) any increase in $PaCO_2$ relative to $ETCO_2$. In the following, we want to discuss the physiologic background for both causes related to the results found in this study.

- (1) A drop in $ETCO_2$ in relation to $PaCO_2$ is indicative of an increase in ventilation of non-perfused ($V/Q = \infty$; alveolar dead space) or poorly perfused (areas of high V/Q ratio) alveoli, resulting in an increase in physiologic dead space. This fact directed Nunn and Hill (3) and Severinghaus et al. (4) toward the conclusion that physiologic or alveolar dead space could be estimated from these two parameters.

No drop in $ETCO_2$ was seen in any of our horses at any time point. In fact $ETCO_2$ increased continuously together with $PaCO_2$ between the measurement points in all horses (Table 1). This indicates that physiologic dead space itself did not increase at all, or if so, only marginally. The VCap results confirmed this by showing only small changes in Bohr's physiologic and alveolar dead space values throughout the study period. The relatively small amount of "real" physiologic and alveolar dead space and minimal changes could be expected in healthy horses

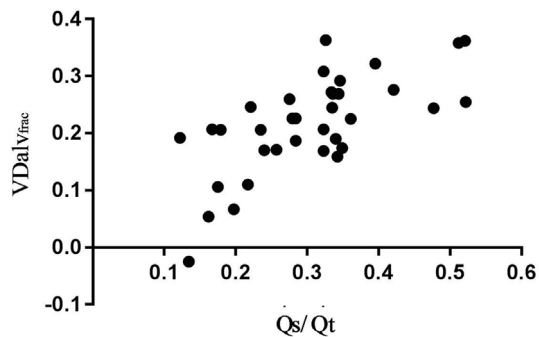


FIGURE 3 | Association between alveolar dead space fraction ($VDalV_{frac}$) and venous admixture (\dot{Q}_s/\dot{Q}_t) in six spontaneously breathing horses under general isoflurane anesthesia in dorsal recumbency. The graph demonstrates the relation over a wide range of \dot{Q}_s/\dot{Q}_t values.

with high CO under spontaneous ventilation. We consider this the most likely reason why we did not find any interaction between the two CO₂ indices and our measured dead space variables. However, in states of low CO, overdistension of alveolar tissue during positive pressure ventilation, excessive PEEP or pulmonary thromboembolism the amount of well-ventilated but poorly or non-perfused alveoli (high V/Q and high Bohr's alveolar dead space) will increase (10, 19, 27–29). These changes also affect V/Q_{Eng} as a factor for the global V/Q mismatch including dead space. This has been shown in a horse suffering from venous air embolism where V/Q_{Eng} nearly doubled during the unintended air infusion (29). As the two CO₂ indices highly correlated with V/Q_{Eng} , we can assume that in situations of high V/Q and high Bohr's alveolar dead space $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ will also increase. Future studies will need to verify this assumption.

The only dead space variable with a relevant influence on $VDalV_{frac}$ in our study was airway dead space. This parameter, which neither influences Enghoff's nor Bohr's equation, is derived from the VCap curve using mathematical modeling of the expired CO₂ over the measured volume (26). Interestingly this was not the first time airway dead space was identified as a factor influencing $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ in a model simulating human physiology airway dead space was detected as one of two main factors, the second one being venous admixture, causing inaccuracies in the calculation of alveolar dead space when using these CO₂ indices (15). As our horses were breathing spontaneously, only small changes in VD_{aw}/VT were expected. However, the impact of VD_{aw}/VT might become more relevant when using positive pressure ventilation, especially in combination with positive end-expiratory pressure, which has been shown by Lumb and others to increase airway dead space (27, 28). Thus, the increase in VD_{aw}/VT can possibly increase $VDalV_{frac}$ under controlled mechanical ventilation.

- (2) The second change, which can influence the difference between $PaCO_2$ and $ETCO_2$, namely an increase in $PaCO_2$ in relation to $ETCO_2$, occurs when CO₂ cannot be eliminated from the blood into the alveoli. This is the case when

the blood flows through atelectatic lung tissue or areas with low V/Q ratios. This fact makes $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ highly variable in situations of high venous admixture (15, 30, 31). Our results confirmed the direct influence of venous admixture on $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ in horses with a broad range of venous admixture volumes. As horses develop extensive venous admixture during anesthesia (1), this species is well suited to show the direct influence of this pathological lung condition on the two CO₂ indices in question. Our horses had between 12 and 52% of their CO perfusing poorly or non-ventilated lung units ($V/Q = 0$; venous admixture). This spectrum represents the expected amount of venous admixture in anesthetized horses in dorsal recumbency (32). This large difference in venous admixture was reflected by a wide range of $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ values. Strang et al. suggested that $P_{(a-ET)CO_2}$ can be used as an index of oxygenation as it correlated well with the amount of atelectasis in pigs seen on computed tomography being even more predictive of the amount of atelectasis than the most accepted oxygen index PaO_2/FiO_2 ratio (18). The strong relationship between $P_{(a-ET)CO_2}$ and an impaired oxygenation has also been shown by means of pulse oximetry in human cardiac patients (33).

To confirm our hypothesis that $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ are influenced by parameters determining lung perfusion even further, we tested the influence of CO and pulmonary arterial pressure, where especially high CO values increased $P_{(a-ET)CO_2}$ and $VDalV_{frac}$. The influence of CO on the two CO₂ indices was very likely due to its direct effect on venous admixture. It is known that increases in CO increase pulmonary pressure whereby pulmonary vessels perfusing collapsed lung areas are recruited. The blood flowing through these vessels increase venous admixture (34) also explaining why we saw an effect of pulmonary pressure on the $VDalV_{frac}$.

Our results also confirm earlier findings in humans that both CO₂ indices have a good correlation with Enghoff's approach and derived variables (3). The fact that Enghoff's approach represents pulmonary dead space as well as venous admixture is well accepted (10, 12, 16, 30, 31, 35). The fact, however, that venous admixture also affects $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ is not even well established in the human anesthesia literature (18, 30, 31) and has not made its way into clinical veterinary anesthesia. This is why the two CO₂ indices are still frequently used in equine anesthesia to estimate pulmonary dead space (36). There are only a few papers specifically studying the two factors (6, 37–39). Most of the variables which have been shown to effect $P_{(a-ET)CO_2}$ and $VDalV_{frac}$ in horses like duration of anesthesia, body weight, dorsal recumbency, and ventilation mode are known to influence venous admixture, CO, and/or pulmonary pressure. Our findings suggest that the idea of $P_{(a-ET)CO_2}$ or $VDalV_{frac}$ being solely representative of dead space is erroneous or at least questionable in cases of high venous admixture typically observed in anesthetized horses.

The high correlation found in this study between the two CO₂ indices with V/Q_{Eng} and $V/Q_{alvEng}/VT_{alv}$ suggests that $P_{(a-ET)CO_2}$ or/and $VDalV_{frac}$ might be good estimates of global V/Q

mismatching in horses. This assumption is based on the fact that variables based on Enghoff's approach have been shown to represent the sum of dead space and venous admixture (10, 16). This might provide equine anesthetist with a new clinical tool to evaluate the lung's status; a high $P_{(a-ET)}CO_2$ or $VD_{alv,frac}$ with an adequate oxygenation (PaO_2) might indicate a high physiological dead space, which points toward low CO and/or overdistension of lung tissue during controlled mechanical ventilation and recruitment maneuvers or even lung embolism (29, 40). On the other hand low PaO_2 values in conjunction with a high $P_{(a-ET)}CO_2$ or/and $VD_{alv,frac}$ might point toward high \dot{Q}_s/\dot{Q}_t probably in combination with high CO and/or pulmonary pressures as seen in our horses. However, further clinical validation in horses is required to support our assumptions.

We also evaluated the influence of other factors like VT, VO_2 , and SIII on the two CO₂ indices as they might have caused changes in the arterial and end-tidal CO₂ difference (14, 15, 19). None of the three parameters contributed any significant variance to either CO₂ index. However, we did not see high variability in our data set for VO_2 and SIII.

One limitation of our study is that VCap has not been directly validated in horses against the multiple inert gas elimination technique for the measurement of Bohr's dead space in the same rigorous way as it has been done for pigs (35). However, as the principle of VCap is based on respiratory physiology, it can be applied to all mammals as long as the trachea and lung follow similar architectures. The special flow-partitioning device for large animals and mathematical correction for the splitting of the tidal volume into four equal parts has already been shown to provide valid VCap parameters in horses (22, 29, 40, 41).

Only six horses were included in this study as the number was set by the primary investigation (23). Hence, no power calculation was performed prior to commencing this study. However, we were able to answer our study question with the collected data sets specifically for healthy spontaneously breathing horses in dorsal recumbency. No inferences can be drawn on how pulmonary disease or mechanical ventilation would affect the two CO₂ indices. Most importantly future studies in horses with higher alveolar Bohr's dead space need to evaluate the influence of this V/Q mismatch "extreme" on the two studied CO₂ indices.

REFERENCES

- Nyman G, Funkquist B, Kvart C, Frostell C, Tokics L, Strandberg A, et al. Atelectasis causes gas exchange impairment in the anaesthetised horse. *Equine Vet J* (1990) 22(5):317–24. doi:10.1111/j.2042-3306.1990.tb04280.x
- Briganti A, Portela DA, Grasso S, Sgorbini M, Tayari H, Bassini JR, et al. Accuracy of different oxygenation indices in estimating intrapulmonary shunting at increasing infusion rates of dobutamine in horses under general anaesthesia. *Vet J* (2015) 204(3):351–6. doi:10.1016/j.tvjl.2015.04.002
- Nunn JF, Hill DW. Respiratory dead space and arterial to end-tidal carbon dioxide tension difference in anesthetized man. *J Appl Physiol* (1960) 15:383–9. doi:10.1152/jappl.1960.15.3.383
- Severinghaus JW, Stupfel MA, Bradley AF. Alveolar dead space and arterial to end-tidal carbon dioxide differences during hypothermia in dog and man. *J Appl Physiol* (1957) 10(3):349–55. doi:10.1152/jappl.1957.10.3.349
- Mitchell B, Littlejohn A. Influence of anaesthesia and posture on arterial oxygen and carbon dioxide tensions, alveolar dead space and pulse rate in the horse. *Vet Anaesth Analg* (1972) 3(1):61–74.

The anesthetic protocol and inspired gas composition might have influenced the measurements and results might vary with different drug protocols and FiO₂ level. Furthermore the hemoglobin analysis was performed with a photometric method not validated for whole horse blood, which might have induced an error in the absolute values of \dot{Q}_s/\dot{Q}_t and VO_2 reported here.

These limitations may have an impact on the predictor validity of the investigated parameters. Future studies should investigate the effect of the listed limiting factors influencing the measurements and their potential clinical influences on the two CO₂ indices.

CONCLUSION

Both CO₂ indices, $P_{(a-ET)}CO_2$, or $VD_{alv,frac}$ represent global indices of the inefficiency of gas exchange and V/Q mismatching, and both are affected by venous admixture in healthy, anesthetized, spontaneously breathing horses. Under such conditions, the indices should not be used to estimate pulmonary dead space.

ETHICS STATEMENT

This experimental study was carried out in accordance with the recommendations of the ARRIVE guidelines and had ethical approval by the local committee for animal experimentation of the Swiss government (TV-4985).

AUTHOR CONTRIBUTIONS

All authors contributed to conception and design of research. MM, UA, and RB performed experiments. AR, SB, GH, GT, and JS analyzed data and did statistical analysis. All authors interpreted results. MM, SB, AR, GT, and JS drafted manuscript. All authors edited, revised, and approved final version of the manuscript.

FUNDING

The want to thank the "Stiftung Forschung für das Pferd" for financing this project (Grant number: PR 2012/02).

- Moens Y. Arterial-alveolar carbon dioxide tension difference and alveolar dead space in halothane anaesthetised horses. *Equine Vet J* (1989) 21(4):282–4. doi:10.1111/j.2042-3306.1989.tb02168.x
- McDonell WN, Kerr C. Physiology, pathophysiology, and anesthetic management of patients with respiratory disease. 5th ed. In: Grimm KA, Lamont LA, Tranquilli WJ, Greene SA, Robertson SA, editors. *Veterinary Anesthesia and Analgesia*. Iowa, US: Wiley Blackwell (2015). p. 513–55.
- Bohr C. Über die Lungenathmung. *Centralblatt für Physiologie* (1887) 1(14): 236–68.
- Enghoff H. Volumen inefficax. Bemerkungen zur Frage des schädlichen Raumes. *Uppsala Lak Forhandl* (1938) 44:191–218.
- Tusman G, Sipmann FS, Bohm SH. Rationale of dead space measurement by volumetric capnography. *Anesth Analg* (2012) 114(4):866–74. doi:10.1213/ANE.0b013e318247f6cc
- Wagner PD. Causes of a high physiological dead space in critically ill patients. *Crit Care* (2008) 12(3):148. doi:10.1186/cc6888
- Suarez-Sipmann F, Santos A, Bohm SH, Borges JB, Hedenstierna G, Tusman G. Corrections of Enghoff's dead space formula for shunt effects still overestimate

- Bohr's deadspace. *Respir Physiol Neurobiol* (2013) 189(1):99–105. doi:10.1016/j.resp.2013.06.020
13. Verscheure S, Massion PB, Verschuren F, Damas P, Magder S. Volumetric capnography: lessons from the past and current clinical applications. *Crit Care* (2016) 20(1):184. doi:10.1186/s13054-016-1377-3
 14. Breen PH, Mazumdar B, Skinner SC. Comparison of end-tidal PCO₂ and average alveolar expired PCO₂ during positive end-expiratory pressure. *Anesth Analg* (1996) 82(2):368–73. doi:10.1097/0000539-199602000-00027
 15. Hardman JG, Aitkenhead AR. Estimation of alveolar deadspace fraction using arterial and end-tidal CO₂: a factor analysis using a physiological simulation. *Anaesth Intensive Care* (1999) 27(5):452–8.
 16. Suarez-Sipmann F, Bohm SH, Tusman G. Volumetric capnography: the time has come. *Curr Opin Crit Care* (2014) 20(3):333–9. doi:10.1097/MCC.0000000000000095
 17. Burrows FA. Physiologic dead space, venous admixture, and the arterial to end-tidal carbon dioxide difference in infants and children undergoing cardiac surgery. *Anesthesiology* (1989) 70(2):219–25. doi:10.1097/0000542-198902000-00007
 18. Strang CM, Hachenberg T, Freden F, Hedenstierna G. Development of atelectasis and arterial to end-tidal PCO₂-difference in a porcine model of pneumoperitoneum. *Br J Anaesth* (2009) 103(2):298–303. doi:10.1093/bja/aep102
 19. Tusman G, Suarez-Sipmann F, Bohm SH, Pech T, Reissmann H, Meschino G, et al. Monitoring dead space during recruitment and PEEP titration in an experimental model. *Intensive Care Med* (2006) 32(11):1863–71. doi:10.1007/s00134-006-0371-7
 20. Douglas CG. A method for determining the total respiratory exchange in man. *J Physiol* (1911) 42:17–8.
 21. Fletcher R. Deadspace during anaesthesia. *Acta Anaesthesiol Scand Suppl* (1990) 94:46–50. doi:10.1111/j.1399-6576.1990.tb03222.x
 22. Schramel JP, Wimmer K, Ambrisko TD, Moens YP. A novel flow partition device for spirometry during large animal anaesthesia. *Vet Anaesth Analg* (2014) 41(2):191–5. doi:10.1111/vaa.12099
 23. Mosing M, MacFarlane P, Bardell D, Luthi L, Cripps PJ, Bettschart-Wolfensberger R. Continuous positive airway pressure (CPAP) decreases pulmonary shunt in anaesthetized horses. *Vet Anaesth Analg* (2016) 43(6):611–22. doi:10.1111/vaa.12357
 24. Kalchofner KS, Picek S, Ringer SK, Jackson M, Hassig M, Bettschart-Wolfensberger R. A study of cardiovascular function under controlled and spontaneous ventilation in isoflurane-medetomidine anesthetized horses. *Vet Anaesth Analg* (2009) 36(5):426–35. doi:10.1111/j.1467-2995.2009.00477.x
 25. Kobler A, Hartnack S, Sacks M, Bettschart-Wolfensberger R. Evaluation of tidal volume measurements of the anaesthesia device Tafonius® in vitro and in vivo. *Pferdeheilkunde* (2016) 32(5):449–56. doi:10.21836/PEM20160505
 26. Tusman G, Scandurra A, Bohm SH, Suarez-Sipmann F, Clara F. Model fitting of volumetric capnograms improves calculations of airway dead space and slope of phase III. *J Clin Monit Comput* (2009) 23(4):197–206. doi:10.1007/s10877-009-9182-z
 27. Lumb AB. *Nunn's Applied Respiratory Physiology*. 7th ed. Oxford, England: Elsevier (2010).
 28. Blankman P, Shono A, Hermans BJ, Wesseliuss T, Hasan D, Gommers D. Detection of optimal PEEP for equal distribution of tidal volume by volumetric capnography and electrical impedance tomography during decreasing levels of PEEP in post cardiac-surgery patients. *Br J Anaesth* (2016) 116(6):862–9. doi:10.1093/bja/aew116
 29. Sacks M, Mosing M. Volumetric capnography to diagnose venous air embolism in an anaesthetised horse. *Vet Anaesth Analg* (2017) 44(1):189–90. doi:10.1111/vaa.12383
 30. Fletcher R, Jonson B, Cumming G, Brew J. The concept of deadspace with special reference to the single breath test for carbon dioxide. *Br J Anaesth* (1981) 53(1):77–88. doi:10.1093/bja/53.1.77
 31. Drummond GB, Fletcher R. Deadspace: invasive or not? *Br J Anaesth* (2006) 96(1):4–7. doi:10.1093/bja/aei289
 32. Kerr CL, McDonnell WN. Oxygen supplementation and ventilatory support. In: Muir WW, Hubbell JAE, editors. *Equine Anesthesia: Monitoring and Emergency Therapy*. St. Louis, Missouri, US: Saunders Elsevier (2009). p. 332–52.
 33. Fletcher R. The relationship between the arterial to end-tidal PCO₂ difference and hemoglobin saturation in patients with congenital heart disease. *Anesthesiology* (1991) 75(2):210–6. doi:10.1097/0000542-199108000-00007
 34. Benumof JL, Wahrenbrock EA. Blunted hypoxic pulmonary vasoconstriction by increased lung vascular pressures. *J Appl Physiol* (1975) 38(5):846–50. doi:10.1152/jappl.1975.38.5.846
 35. Tusman G, Sipmann FS, Borges JB, Hedenstierna G, Bohm SH. Validation of Bohr dead space measured by volumetric capnography. *Intensive Care Med* (2011) 37(5):870–4. doi:10.1007/s00134-011-2164-x
 36. Robinson NE. The respiratory system. In: Muir WW, Hubbell JAE, editors. *Equine Anesthesia: Monitoring and Emergency Therapy*. St. Louis, Missouri: Saunders (2009). p. 11–36.
 37. Rainger JE, Dart CM, Perkins NR. Factors affecting the relationship between arterial and end-tidal carbon dioxide pressures in the anaesthetised horse. *Aust Vet J* (2010) 88(1–2):13–9. doi:10.1111/j.1751-0813.2009.00535.x
 38. Neto FJ, Luna SP, Massone F, Thomassian A, Vargas JL, Junior JR, et al. The effect of changing the mode of ventilation on the arterial-to-end-tidal CO₂ difference and physiological dead space in laterally and dorsally recumbent horses during halothane anesthesia. *Vet Surg* (2000) 29(2):200–5. doi:10.1111/j.1532-950X.2000.00200.x
 39. Meyer RE, Short CE. Arterial to end-tidal CO₂ tension and alveolar dead space in halothane- or isoflurane-anesthetized ponies. *Am J Vet Res* (1985) 46(3):597–9.
 40. Ambrisko TD, Schramel J, Hopster K, Kastner S, Moens Y. Assessment of distribution of ventilation and regional lung compliance by electrical impedance tomography in anaesthetized horses undergoing alveolar recruitment manoeuvres. *Vet Anaesth Analg* (2017) 44(2):264–72. doi:10.1016/j.vaa.2016.03.001
 41. Mosing M, Auer U, MacFarlane P, Bardell D, Schramel J, Bohm SH, et al. Regional ventilation distribution and dead space in anaesthetised horses treated with and without continuous positive airway pressure: novel insights by electrical impedance tomography and volumetric capnography. *Vet Anaesth Analg* (2018) 45(1):31–40. doi:10.1016/j.vaa.2017.06.004
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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