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iValiD-TB: a fully characterized *Mycobacterium tuberculosis* dataset for antimicrobial resistance bioinformatics workflow validations

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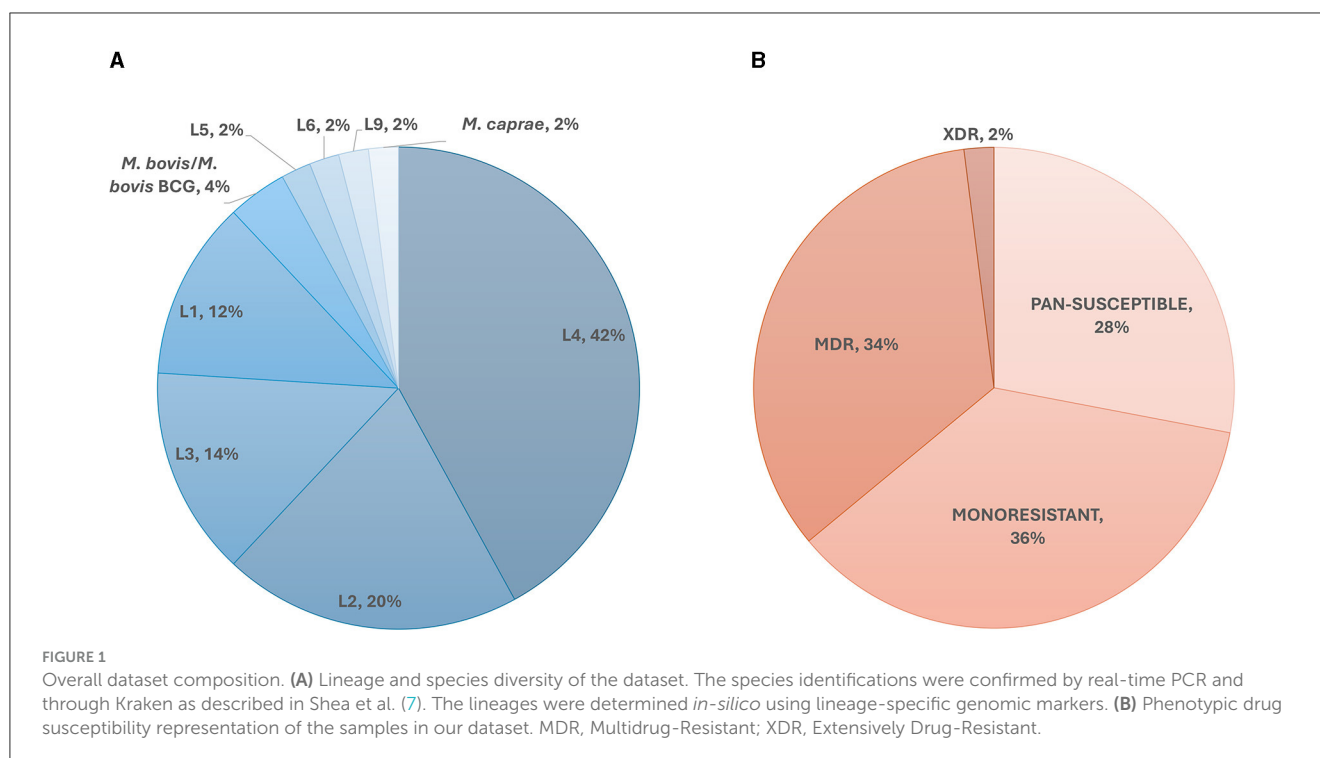
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Introduction

Tuberculosis has been a bane of humanity for centuries and, still to this day, is estimated to affect more than two billion people worldwide (1). The slow growth rate of *Mycobacterium tuberculosis* (MTB) is a major challenge for timely diagnosis and appropriate treatment of cases (2, 3). Diagnostic delays can lead to increased disease burden, cost increases and risk of treatment failures (4). A crucial aspect for clinical assay validations is the availability of well-characterized samples to assess the specificity and sensitivity of the methodology being tested (10). Due to logistical constraints, the nature and availability of specimens, and geographic diversity of the strains, many laboratories struggle with access to adequate clinical MTB specimens for their validation needs (11). Consequently, validation studies may not include representative of the myriad of clinical samples and drug resistant profiles that a clinical laboratory may receive. Therefore, diverse and well-characterized datasets for standardized next generation sequencing (NGS) assay validations for MTB NGS tests are needed. Reference datasets of clinical TB samples and synthetic genomes were released in the past with limited phenotypic drug susceptibility testing (DST) information for research, development and proficiency testing purposes (5, 6). Here, we have assembled a comprehensive dataset of well-characterized whole genome sequences (WGS) from *Mycobacterium tuberculosis* strains to aid in the development of clinical assays for this pathogen. This dataset includes complete whole genome sequences paired-end read sets obtained through Illumina MiSeq sequencing, along with detailed profiles of drug susceptibility patterns and mutations known to be associated with antimicrobial resistance (AR) to nine MTB drugs. This dataset has been curated to be inclusive of a broad range of lineage diversity, drug susceptibility profiles, and mutation types. As such, this dataset only contains two separate pairs of strains that are phylogenetically closely related (iValiD-TB-S22 and iValiD-TB-S23 with 0 SNP differences and iValiD-TB-S6 and iValiD-TB-S46 with 6 SNPs differences) based on our pipeline



results. A complete SNP matrix has been included in [Supplementary Table 3](#). The sequence reads dataset has been made available for bioinformatics pipeline development, and for clinical assay validation of the bioinformatic analysis pipeline, serving as a valuable resource to advance research and enhance the development of clinical MTB NGS assays.

A total of 50 members of the *Mycobacterium tuberculosis* complex (MTBC) were sequenced, which includes 47 strains of *Mycobacterium tuberculosis*, one *Mycobacterium caprae* strain, one *Mycobacterium bovis* strain and one *Mycobacterium bovis*-BCG strain ([Figure 1](#)). These strains were part of our collection of samples obtained from New York State patients since the implementation in 2013 of our clinical diagnostic and reporting TB NGS assay (7). Of the MTBC, 6 strains are from Lineage 1, 10 from Lineage 2, seven from Lineage 3, 21 from Lineage 4, and 1 representative of each of Lineages 5, 6, and 9. Of the 50 samples, 14 were determined to be pan-susceptible to nine drugs (rifampin, isoniazid, ethambutol, pyrazinamide, streptomycin, ethionamide, fluoroquinolones, kanamycin, and amikacin) by phenotypic drug susceptibility testing, 18 were mono-resistant, 17 multi-drug resistant (MDR) and one was extensively drug resistant (XDR; [Figure 1](#), [Supplementary Table 1](#)). A total of 1,073 different mutations present in the 53 screened loci ([Supplementary Table 2](#)) characterized by WGS in this dataset, of which, 107 mutations were identified to be associated with drug resistance, most of which are part of the World Health Organization 2023 Catalog of mutations in *Mycobacterium tuberculosis* (8). The New York State Department of Health implemented this assay for clinical diagnostic before the WHO catalog was released and as such, we are using our own susceptibility interpretation criteria. Consequently, the users of this dataset will have to use their own decision criteria based on

their individual workflow characteristics and interpretations. The characterized mutations included single nucleotide polymorphisms (SNP), stop codons, promoter mutations, small insertion and deletions (indels), and large genomic deletions. The locations of the mutations, types, effect on drug resistance, as well as DST results, lineage information, spoligotype and expected mapping statistics are all listed in an individual report card for each sample ([Figure 2](#)). The release of this fully characterized dataset will facilitate the development and benchmarking of bioinformatics tools for MTB NGS diagnostics and aid in the validation of these clinical assays. The read sequences are accessible from the NCBI SRA Bioproject PRJNA980174. The associated AR reports cards for the 50 samples are available in Dryad at: <https://doi.org/10.5061/dryad.4j0zpc8m8>.

Methods

Genomic DNA extraction, sequencing library preparation and bioinformatics pipeline methods were described in Shea et al. (7). DSTs were determined by either the agar proportion method on solid 7H10 agar or Becton Dickinson 960 system MGIT SIRE-P assay according to the Clinical and Laboratory Standards Institute's recommendations (9). The following concentrations were used for DST determinations: Streptomycin 1.0 µg/ml, Isoniazid 0.1, 0.2, 0.4, and 1.0 µg/ml, Rifampin 1.0 µg/ml, Ethambutol 5.0 and 10.0 µg/ml, Pyrazinamide 100 µg/ml, Kanamycin 5.0 µg/ml, and ofloxacin (1.0, 2.0, and 4.0 µg/ml). Ofloxacin is used in our laboratory as a representative of the fluoroquinolone (FQ) drug class. Genotypic identification, mapping statistics and *in-silico* spoligotyping were done as described in Shea et al. (7).

Sample: iValid_TB_S2

Genotypic-based IG:
Lineage 2 (Beijing)
Mycobacterium tuberculosis

Mapping Statistics:
Genome Coverage = 98.68%
Average Depth = 101.6x

In-silico Spoligotype (binary, octal code, SIT):
0000000000000000000000000000011111111, 000000000003771, 1

All Mutations in screened loci:

Genomic Position	Codon	Nucl. Change	A.A. Change	Locus	Gene	AR	Haplotype/Notes
7362	21	GAG -> CAG	Glu -> Gln	Rv0006	gyrA	Fluoroquinolones	1/1 (Pure) (Neutral)
7585	95	AGC -> ACC	Ser -> Thr	Rv0006	gyrA	Fluoroquinolones	1/1 (Pure) (Neutral)
9304	668	GGC -> GAC	Gly -> Asp	Rv0006	gyrA	Fluoroquinolones	1/1 (Pure) (Neutral)
491742	320	TTT -> TTC	Phe -> Phe	Rv0407	fgd1	Delamanid/Pretomanid	1/1 (Pure) (Silent)
575907	187	GCA -> GTA	Ala -> Val	Rv0486	mshA	Ethionamid	1/1 (Pure) (Unknown)
761155	450	TCG -> TTG	Ser -> Leu	Rv0667	rpoB	Rifampin	1/1 (Pure) (HC mutation)
763031	1075	GCT -> GCC	Ala -> Ala	Rv0667	rpoB	Rifampin	1/1 (Pure) (Silent)
776182	767	GAC -> AAC	Asp -> Asn	Rv0676c	mmpL5	Clofazimine/Bedaquiline	1/1 (Pure) (Neutral)
776100	794	ACC -> ATC	Thr -> Ile	Rv0676c	mmpL5	Clofazimine/Bedaquiline	1/1 (Pure) (Neutral)
775639	948	ATT -> GTT	Ile -> Val	Rv0676c	mmpL5	Clofazimine/Bedaquiline	1/1 (Pure) (Neutral)
781687	43	AAG -> AGG	Lys -> Arg	Rv0682	rpsL	Streptomycin	1/1 (Pure) (HC mutation)
1917972	11	CTA -> CTG	Leu -> Leu	Rv1694	tlyA	Aminoglycosides	1/1 (Pure) (Silent)
2155168	315	AGC -> ACC	Ser -> Thr	Rv1908c	katG	Isoniazid	1/1 (Pure) (HC mutation)
2154724	463	CGG -> CTG	Arg -> Leu	Rv1908c	katG	Isoniazid	1/1 (Pure) (Neutral)
2715342	-10	G -> A		intergenic	eis promoter region	Kanamycin/Amikacin	1/1 (Pure) (HC mutation)
3336825	365	ACA -> GCA	Thr -> Ala	Rv2981c	ddlA	D-Cycloserine	1/1 (Pure) (Neutral)
3878416	31	GGC -> GCC	Gly -> Ala	Rv3457c	rpoA	Rifampin compensatory	1/1 (Pure) (Unknown)
4242643	927	CGC -> CGT	Arg -> Arg	Rc3793	embC	Ethambutol	1/1 (Pure) (Silent)
4243346	38	CAA -> CAG	Gln -> Gln	Rv3794	embA	Ethambutol	1/1 (Pure) (Silent)
4243460	76	TGC -> TGT	Cys -> Cys	Rv3794	embA	Ethambutol	1/1 (Pure) (Silent)
4247469	319	TAT -> TGT	Tyr -> Cys	Rv3795	embB	Ethambutol	1/1 (Pure) (Unknown)
4327054	140	TAC -> TAG	Tyr -> Stop	Rv3854c	ethA	Ethionamide	1/1 (Pure) (Unknown)
4407927	92	GAA -> GAC	Glu -> Asp	RV3919c	gidB	Streptomycin	1/1 (Pure) (Neutral)
4407588	205	GCA -> GCG	Ala -> Ala	RV3919c	gidB	Streptomycin	1/1 (Pure) (Silent)

Final AR Profile:

Antimicrobial	AR Genotype	MGIT	AP	Final Interpretation
Rifampin	rpoB Ser450Leu	RIF 1.0	RIF 1.0	RESISTANT
Isoniazid	katG Ser315Thr	INH 0.1, 0.4	INH 0.2, 1.0	RESISTANT
Pyrazinamide	---	---	---	Susceptible
Ethambutol	embB Tyr319Cys	EMB 5.0	EMB 5.0	RESISTANT
Streptomycin	rpsL Lys43Arg	SM 1.0	SM 2.0	RESISTANT
Kanamycin/Amikacin	eis G-10A	---	AMI 2.0, 4.0	RESISTANT
Fluoroquinolones	---	---	---	Susceptible
Ethionamide	ethA Tyr140STOP	---	ETH 5.0	RESISTANT

This strain was susceptible to Amikacin at 2.0 and 4.0. Kanamycin and Amikacin are the same drug class, however eis mutations only confer kanamycin resistance. We do not have kanamycin DST for this strain.

FIGURE 2

Sample report card. Example of report card for one of the samples in the dataset listing the locations of the mutations, mutation types, drug susceptibility testing results, the final interpretation of the drug resistance, lineage information, spoligotype, and expected mapping metrics. The report cards are available in Dryad at: <https://doi.org/10.5061/dryad.4j0zpc8m8>. RIF, Rifampin; INH, Isoniazid; EMB, Ethambutol; SM, Streptomycin; AMI, Amikacin; ETH, Ethionamide; PZA, Pyrazinamide; FQ, Fluoroquinolones. DSTs concentrations are in µg/ml.

Data availability statement

The datasets presented in this study can be found in online repositories. The data is available in Dryad at: <https://doi.org/10.5061/dryad.4j0zpc8m8>.

Author contributions

PL: Conceptualization, Data curation, Software, Writing – original draft, Writing – review & editing. JS: Data curation, Investigation, Validation, Writing – review & editing. SM: Validation, Writing – review & editing. CS: Validation, Writing – review & editing. DK: Supervision, Validation, Writing – review & editing. MD: Supervision, Validation, Writing – review & editing. KM: Conceptualization, Supervision, Writing – review & editing. VE: Conceptualization, Supervision, Validation, Writing – review & editing.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/ftubr.2024.1441923/full#supplementary-material>

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