



hsa_circ_0072309 Expression Profiling in Non-small-Cell Lung Carcinoma and Its Implications for Diagnosis and Prognosis

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OPEN ACCESS

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Specialty section:

This article was submitted to Visceral Surgery, a section of the journal Frontiers in Surgery

Received: 23 December 2021 Accepted: 20 January 2022 Published: 25 February 2022

Citation:

Zhou Y, Tong Z, Zhu X, Huang S,
Dong Z and Ye Z (2022)
hsa_circ_0072309 Expression
Profiling in Non-small-Cell Lung
Carcinoma and Its Implications for
Diagnosis and Prognosis.
Front. Surg. 9:842292.
doi: 10.3389/fsurg.2022.842292

Circular RNAs (circRNAs), which fall into the category of endogenous ncRNAs, are linked to disease progression of neoplastic diseases. Whereas, it remains uncharacterized regarding hsa_circ_0072309's function and implications in lung carcinoma (LC). Gene Expression Omnibus (GEO) database was utilized for identifying circRNAs with aberrantly expression in LC. qRT-PCR was responsible for determining hsa_circ_0072309 levels in lung adenocarcinoma (LAC). Also, its involvement in LC cell progression was investigated. Experimentally, hsa_circ_0072309 was identified as one of the most aberrantly down-regulated circRNAs in the GEO database (GSE101684 and GSE112214). qRT-PCR revealed notably down-regulated hsa_circ_0072309 in LAC tissue, which had a close association with adverse 3-year survival, as well as LNM and advanced TNM stage. Based on ROC, the AUC of hsa_circ_0072309 was determined to be 0.887, and its specificity and susceptibility can be improved by combined detection of either CYFRA21-1 or CEA. In a word, hsa_circ_0072309 is lowly expressed in lung cancer patients and the survival rate of lowly expressed patients is significantly lower, a candidate marker with prognostic utility for the disease.

Keywords: hsa_circ_0072309, NSCLC, diagnosis, prognosis, GEO

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INTRODUCTION

Lung carcinoma (LC), as a prevalent respiratory malignancy (1). Epidemiological statistics reveal 4.3 million new cancer cases and an associated death toll of 2.9 million in 2018 in China, including 770,000 new LC cases and 690,000 associated deaths (2). Therefore, in the future, not only will the number of LC patients increase, but also the number of elderly patients (3). The predisposing factors of LC in the elderly include smoking, environmental exposure, ionizing radiation, and vocational diseases, heredity (4). However, due to the relatively concealed pathogenesis, the non-specific initial clinical manifestation, and the lack of good clinical diagnostic indicators, the disease has developed into the mid-late stage in some patients when they are hospitalized, resulting in losing the optimal time for operation (5). In addition, the long-term treatment cycle and dim outcomes of advanced LC aggravate the pressure of sufferers and the family economical load (6). Therefore, searching for a diagnostic index with high specificity and sensitivity is pressing to address this problem.

Recent evidence has indicated that non-coding RNAs (ncRNAs), closely linked to human life process, are vital in modulating a wide spectrum of illnesses, especially neoplastic diseases (7). As vital members of ncRNAs, miRNAs and lncRNAs have always been the focus of research (8). circRNAs, on the other hand, are recently found RNAs capable of forming loops that are covalently closed and continuous, with 3' and 5'RNA ends binded, more favorable stability than linear types, and no polyadenosinic acid counterpart (9). hsa_circ_0072309, or LIFR, is located on chr5:38523520-38530768 (10), with its expression profiling being found in LC (11). But there is no research to prove its connection with patient prognosis and its diagnostic implications in non-small-cell lung carcinoma (NSCLC). Herein, we screened GSE101684 and GSE112214 microarrays and found that hsa_circ_0072309 had high levels in LC, suggesting its role of a candidate target for NSCLC.

Therefore, we have experimentally analyzed the expression of hsa_circ_0072309 in NSCLC patients and its diagnostic significance, providing a new potential target for clinical diagnosis and paving the way for our subsequent studies.

METHODS AND MATERIALS

Microarray (GEO-Chip) Analysis

From GEO database (URL: https://www.ncbi.nlm.nih.gov/gds/), two microarrays (GSE101684 and GSE112214) were obtained. In the GSE101123 microarray, we found 4 tumor samples and 4 normal counterparts. And three tumor samples and three normal counterparts were found in the GSE112214 microarray. Then Series Matrix File(s) was downloaded, and GPL21825 and GPL19978 were sorted out into a matrix file. VLOOKUP function was utilized to correct the circRNA name and limma package to identify the circRNAs with differences (logFC = 1, P < 0.05). Volcano plots and heat maps were drawn.

Clinical Data

Between February 2017 and February 2019, 46 NSCLC patients who received treatment in our hospital were included, with their peripheral blood samples collected before treatment. Meanwhile, peripheral blood specimens of 40 healthy people with concurrent physical examinations were used as control samples. Informed consents were got before patient enrollment. This study, in compliance with the Helsinki Declaration, has been ethically ratified by our hospital. The two arms were similar in age, sex and BMI (P > 0.05).

Eligibility Criteria

Inclusion criteria: Diagnosis of NSCLC by imaging and pathology; In compliance with the TNM staging standard issued by AJCC (8th Edition) (12); Treatmentnaive patients who had not received surgical resection, chemoradiotherapy, immunotherapy or molecular targeted therapy before participation.

Exclusion criteria: Other tumors; Estimated survival < 3 months; No follow-up data; Incomplete case date.

qRT-PCR Detection

Serum was analyzed in this study. TRIzol-extracted (Takara Bio, Shiga, Japan) total RNA from the collected serum was subjected to purity and concentration identification and the subsequent cDNA synthesis with use of the PrimeScript RT kit. Then the synthesized cDNA was subjected to amplification by PCR via the SYBR Premix Ex Taq kit, with the reaction system and reaction conditions configured following the instructions provided with the kit. The primers used are listed in **Table 1**. hsa_circ_0072309 expression normalized with β -actin (endogenous control) was obtained via the formula $2^{-\Delta \Delta CT}[\Delta CT(test) = CT(target, test) - CT(ref, test)\Delta CT(calibrator) = CT(target, calibrator)-CT(ref, calibrator), <math display="inline">\Delta \Delta CT = \Delta CT(test) - \Delta CT(calibrator)]$. In this experiment, the ABI 7500 PCR system was used for analysis.

CEA and CYFRA21-1 Detection

For all experiments, blood samples were collected from an elbow vein in the early morning on an empty stomach prior to the start of treatment. Serum tumor markers: 10 ml of blood was collected. CEA and CYFRA21-1 were measured using a Beckman Coulter AU5800 automated biochemistry analyser.

Endpoints

Primary endpoints: The differential circRNAs in GSE101684 and GSE112214 microarrays were analyzed and heat maps and volcano plots were drawn. Serum and carcinoma tissue hsa_circ_0072309 expression in cases was tested. ROC curves were drawn to visualize hsa_circ_0072309's diagnostic implications in NSCLC.

Secondary endpoints: The connection between hsa_circ_0072309 and patient clinical data was analyzed. In addition, patients were followed up for 3 years to observe the connection between hsa_circ_0072309 and patient prognosis.

Statistical Processing

The above index data were input into SPSS21.0 and GraphPad Prism 8.0 for statistical processing and plotting, respectively. Quantitative variables, described in (Mean \pm SD), were analyzed via the independent samples T test. While represented by n, categorical variables were examined using the χ^2 test. ROC curves were used for assessing the diagnostic implications of serum hsa_circ_0072309. Patient survival was evaluated by the K-M algorithm. The R software limma package was used to analyse the differential genes, the library package to map the differential gene volcanoes and the heatmap package to map the differential gene heat map. Significance was set at the probability (P) level < 0.05.

RESULTS

GEO-Chip Analysis

In this study, the GSE101684 microarray was analyzed, and 249 differential circRNAs were identified, of which 157 were over-expressed and 92 were under-expressed (**Table 1**, **Figures 1A,B**). One hundred differential circRNAs were found through analyzing the GSE112214 microarray, including 11 over-expressed and 89 were under-expressed circRNAs

TABLE 1 | The top five up-down-regulated circRNAs in the GSE101684 microarray.

Symbol	logFC	AveExpr	t-value	P-Value	adj.P.Val	β
Up						
hsa_circ_0072088	2.830	10.003	5.380	4.212E-04	0.011	0.339
hsa_circ_00723098	2.828	11.026	8.856	8.780E-06	0.003	4.135
hsa_circ_0067301	2.689	9.446	7.046	5.560E-05	0.005	2.358
hsa_circ_0001806	2.571	13.307	7.611	3.010E-05	0.004	2.955
hsa_circ_0006349	2.459	12.740	9.572	4.590E-06	0.003	4.739
Down						
hsa_circ_0000317	-2.627	10.286	-5.631	3.033E-04	0.009	0.670
hsa_circ_0001640	-2.057	7.020	-4.820	9.056E-04	0.016	-0.433
hsa_circ_0013480	-2.042	8.534	-6.318	1.290E-04	0.006	1.525
hsa_circ_0000320	-2.005	9.213	-4.839	8.815E-04	0.016	-0.406
hsa_circ_0000319	-1.938	7.523	-5.343	4.430E-04	0.012	0.289

TABLE 2 | The top five up-down-regulated circRNAs in the GSE112214 microarray.

Symbol	logFC	AveExpr	t-value	P-value	adj.P.Val	β	
Up							
hsa_circ_0007580	1.257	6.451	7.041	2.083E-04	1.769E-02	1.232	
hsa_circ_0001998	1.223	8.145	4.843	1.894E-03	3.874E-02	-1.045	
hsa_circ_0017956	1.183	7.915	4.684	2.276E-03	4.295E-02	-1.237	
hsa_circ_0003028	1.132	7.414	6.669	2.911E-04	1.974E-02	0.892	
hsa_circ_0008758	1.087	8.681	7.245	1.743E-04	1.568E-02	1.412	
Down							
hsa_circ_0072309	-2.968	6.912	-9.212	3.770E-05	1.020E-02	2.918	
hsa_circ_0008234	-2.450	9.592	-6.210	4.486E-04	2.251E-02	0.450	
hsa_circ_0006677	-2.372	9.231	-11.186	1.050E-05	6.183E-03	4.104	
hsa_circ_0001947	-2.293	8.022	-7.425	1.495E-04	1.568E-02	1.566	
hsa_circ_0072305	-2.181	6.274	-9.070	4.170E-05	1.020E-02	2.822	

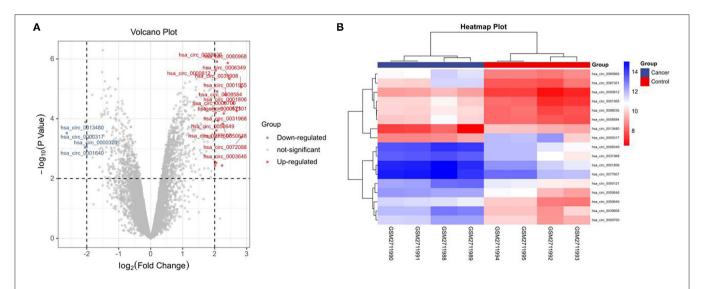


FIGURE 1 | Volcanic plot and heat map of circRNAs with aberrant expression in the GSE101684 microarray. (A). Volcanic plot of circRNAs with aberrant expression in the GSE101684 microarray: Red, blue and gray represent those with high, low, and undifferential expression, respectively. (B). Heat map of circRNAs with aberrant expression in the GSE101684 microarray.

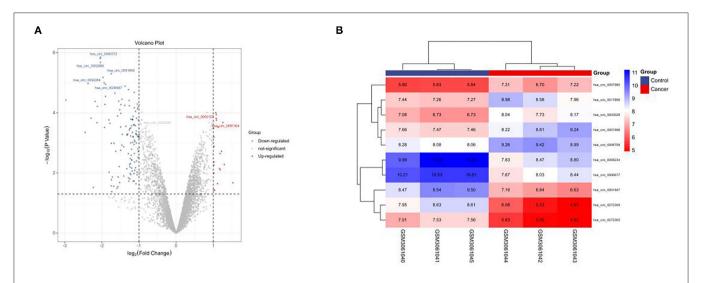
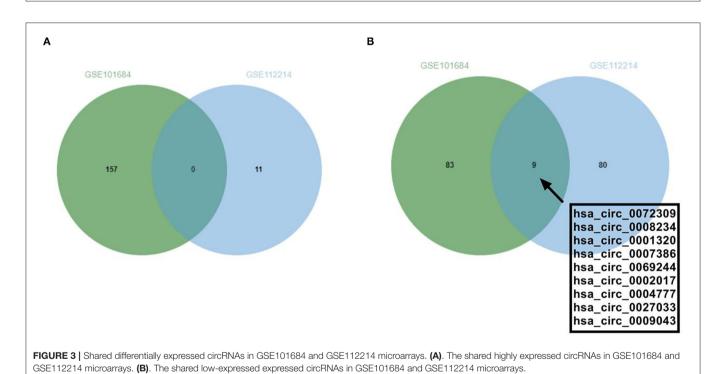


FIGURE 2 | Volcanic plot and heat map of circRNAs with aberrant expression in the GSE112214 microarray. (A). Volcanic plot of differentially expressed circRNAs in the GSE112214 microarray: Red, blue and gray indicate those with high, low, and undifferential expression, respectively. (B). Heat map of circRNAs with aberrant expression in the GSE112214 microarray.



(Table 2, Figures 2A,B). In order to find shared differentially expressed circRNAs, we drew a Venn diagram to find zero shared highly expressed circRNAs in the two microarrays while nine common lowly expressed circRNAs (Figures 3A,B). hsa_circ_0072309 with the most significant difference and the largest LogFC was selected for follow-up study.

hsa_circ_0072309 in Cases

We compared serum hsa_circ_0072309 levels between cases and controls and found statistically

higher hsa_circ_0072309 levels in cases (**Figure 4**, P < 0.001).

Connection Between hsa_circ_0072309 and Patient Clinical Data

Based on median hsa_circ_0072309 expression, cases were subgrouped (high/expression groups, 23 cases each). The comparison of clinical data revealed an evidently elevated chance of developing III-IV LC and lymph node metastasis (LNM) in patients

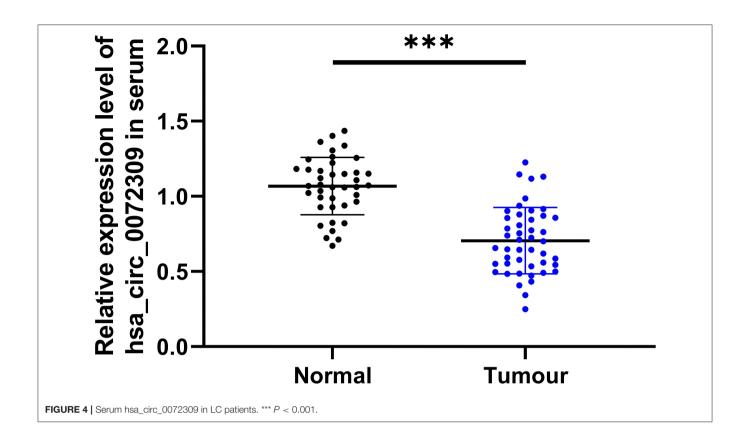


TABLE 3 | Connection between hsa_circ_0072309 and patient clinical data.

Variables		Relative expression	P-value	
		High expression (n = 23)	Low expression (n = 23)	
Age				0.552
	≥60 (<i>n</i> = 20)	9	11	
	<60 (n = 26)	14	12	
Gender				0.555
	Male $(n = 22)$	12	10	
	Female $(n = 24)$	11	13	
Tumor size (cm)				0.546
	≥3 (n = 18)	10	8	
	<3 (n = 28)	13	15	
Clinical staging				0.003
	I-II $(n = 26)$	18	8	
	III-IV ($n = 20$)	5	15	
LNM				0.013
	With $(n = 16)$	4	12	
	Without $(n = 30)$	19	11	
Tumor types				0.767
	Squamous cell carcinoma ($n = 21$)	11	10	
	Adenocarcinoma ($n=25$)	12	13	

TABLE 4 | ROC curve parameters.

Variables AUC 95 CI% Specificity Sensitivity Youden index hsa_circ_0072309 0.887 0.818-0.956 82.50% 86.95% 69.45% CEA 0.815 0.720-0.911 97.50% 63.04% 60.54% CYFRA21-1 0.834 0.743-0.924 77.50% 84.78% 62.28%	
CEA 0.815 0.720-0.911 97.50% 63.04% 60.54% CYFRA21-1 0.834 0.743-0.924 77.50% 84.78% 62.28%	Cut-off
CYFRA21-1 0.834 0.743-0.924 77.50% 84.78% 62.28%	0.921
	6.001
han also 0070000 (OEA 0.040 0.000 0.	3.128
hsa_circ_0072309+CEA 0.940 0.889-0.990 82.50% 95.65% 78.15%	0.337
hsa_circ_0072309+CYFRA21-1 0.944 0.898-0.989 90.00% 84.78% 74.78%	0.614

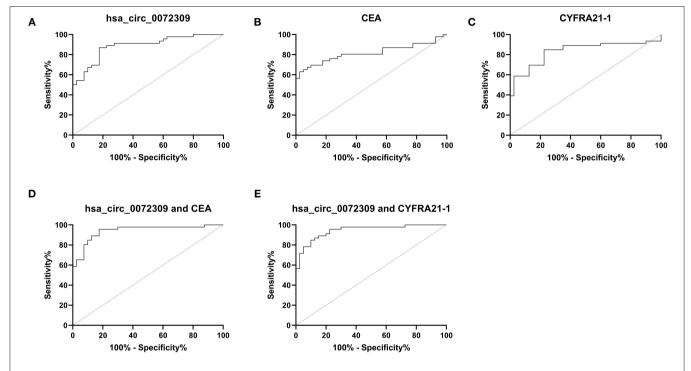


FIGURE 5 | hsa_circ_0072309 for NSCLC diagnosis by ROC analysis. (A). hsa_circ_0072309's implications for NSCLC diagnosis by ROC. (B). CEA's implications for NSCLC diagnosis by ROC. (C). CYFRA21-1's implications for NSCLC diagnosis by ROC. (D). Diagnosistic implications of hsa_circ_0072309+CEA for NSCLC by ROC. (E). Diagnosistic implications of hsa_circ_0072309+CYFRA21-1 for NSCLC by ROC.

with under-expressed hsa_circ_0072309 (**Table 3**, P < 0.05).

Diagnostic Implications of hsa_circ_0072309 in NSCLC Patients

ROC curves were drawn to understand hsa_circ_0072309's diagnostic implications in NSCLC patients. Through analysis, hsa_circ_0072309's AUC, specificity and susceptibility were determined to be 0.887, 82.50, and 86.95%, respectively, which were evidently higher than those of CEA (0.815, 97.50, and 63.04%) and CYFRA21-1 (0.834, 77.50, and 84.78%). Further, hsa_circ_0072309 could improve the sensitivity and specificity of CEA and CYFRA21-1 through joint prediction (**Table 4**, **Figure 5**).

Cox Regression Analysis

Finally, we conducted statistics on the 3-year survival of patients and Cox regression analysis by combining patient's medical

records. Univariate analysis revealed that clinical stage, LNM, hsa_circ_0072309 were prognostic factors for NSCLS patients (**Figure 6**). Furthermore, multivariate Cox regression analysis by further incorporating differentially expressed factors in univariate analysis identified the independence of LNM and hsa_circ_0072309 as prognostic factors for NSCLS (**Table 5**).

DISCUSSION

Due to the current lack of basic biological understanding of NSCLC, the feasible biomarkers for detection and effective medicines for treatment are lacking (13, 14). Relevant literature has revealed a close connection between the survival of LC patients and staging, with the 5-year survival rate decreasing from 82 to 6% as the staging (IA-IV) increases (15). However, early diagnosis is too hard given the non-specific early symptoms (16). Consequently, most cases are confirmed at the late stage when the best treatment opportunity has been missed (17).

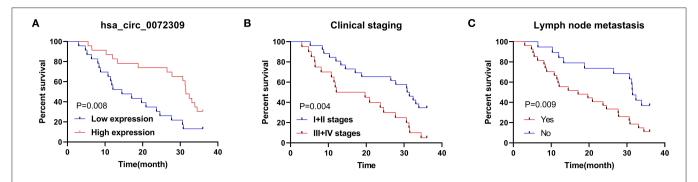


FIGURE 6 | K-M survival curve of factors with differences in univariate analysis. (A). Three-year survival of NSCLC patients with up- and down-regulated hsa_circ_0072309 by K-M curves. (B). Three-year survival of NSCLC patients with I-II or III-IV LC by K-M curves. (C). Three-year survival of NSCLC patients with and without lymph node metastasis by K-M curves.

TABLE 5 | Cox regression.

Factors	Un	Univariate			Multivariate	
	P-value	HR	95%CI	P-value	HR	95%CI
Age	0.708	0.881	0.455-1.707			
Gender	0.749	0.898	0.466-1.731			
Tumor size	0.268	1.482	0.739-2.975			
Clinical staging	0.005	2.582	1.323-5.042	0.180	1.666	0.790-3.515
Lymph node metastasis	0.004	0.364	0.185-0.719	0.010	0.399	0.198-0.804
Tumor types	0.249	1.479	0.760-2.876			
hsa_circ_0072309	0.010	2.426	1.234-4.767	0.025	2.219	1.104-4.460

Therefore, finding effective diagnostic markers is beneficial to improve patient prognosis and survival, which is also a current clinical challenge.

CircRNA are recently found endogenous ncRNAs with closed-loop structure (18). Because of their high tissue specificity and stablility in structure, they are less prone to digestion by exonuclease (RNase) (19), which also results in circRNAs being potentially favorable biomarkers for diagnosing and predicting outcomes of various illnesses (20). Earlier, research has revealed the close association between circRNA-100876 and prognosis of colorectal cancer (21). Another research identified markedly upregulated hsa_circ_0004585 in colorectal cancer and its function as a feasible biomarker for the disease (22). All these studies suggest that circRNAs are feasible biomarkers for cancer diagnosis.

For the purpose of determining the expression of circRNAs in NSCLC, GSE101684 and GSE112214 microarray datasets were retrieved through the GEO database in this study for analysis. Additionally, qRT-PCR was carried out for identifying the circRNAs with aberrant expression in clinical samples. It identified underexpressed hsa_circ_0072309 in cases, which agreed the findings of Mo et al. (11). However, their study did not further analyze hsa_circ_0072309's clinical application in NSCLC. Through analysis, we further confirmed a notably increased probability of LNM and clinical stage III-IV in patients with hsa_circ_0072309 underexpression. Therefore, it may interfere with NSCLC cell growth, progression and

migration. Furthermore, we probed into hsa_circ_0072309's clinical implications in diagnosing NSCLC. As indicated by ROC curve analysis, hsa circ 0072309's AUC was 0.887, notably higher than that of CEA and CYFRA21-1. AUC is considered to be the best index to evaluate the diagnostic value, with a value of <0.5, 0.5-<0.7, 0.7-<0.9 and ≥ 0.9 indicating no diagnostic value, lower accuracy, certain accuracy, and higher accuracy respectively. This suggests that hsa circ 0072309 is a feasible diagnostic index for NSCLC (23). We then performed joint prediction based on Logistic regression, and found improved sensitivity of CEA and CYFRA21-1 by hsa_circ_0072309. Finally, we discussed the connection between hsa circ 0072309 and patient prognosis. As shown by Cox regression analysis, hsa_circ_0072309 was independently linked to patient prognosis, suggesting its potential as an essential marker for adverse prognosis in NSCLC patients.

This research experimentally confirmed down-regulated_circ_0072309 in NSCLC and its potential to be a marker with diagnosis and prognosis utility for NSCLC. However, there are some limitations to be addressed. We only collected samples from LC patients, but not patients with benign lung diseases. The occurrence of LC is a long process, whether hsa_circ_0072309 can distinguish benign lung diseases from LC remains to be defined. Second, this study has a single sample type. Recent studies have found that circRNAs exist in exosomes, peripheral blood mononuclear cells and alveolar fluid. Whether hsa_circ_0072309 is more significantly differentially

expressed in other samples needs further verification. Third, this time, the mechanism of hsa_circ_0072309 has not been deeply explored, and it is hoped that the later expansion through bioinformatics analysis will lay the foundation for our follow-up basic experiments.

In conclusion, hsa_CIRC_0072309 is a candidate marker with prognostic utility in lung cancer patients with low expression and a significantly lower survival rate in patients with low expression.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

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AUTHOR CONTRIBUTIONS

YZ and ZT are the mainly responsible for the writing of the article. XZ is mainly responsible for research design. SH is mainly responsible for data analysis. ZD and ZY are responsible for the guidance of the entire research. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by Natural Science Foundation of Zhejiang Province (No. LQ21H160014), Medical Health Science and Technology Project of Zhejiang Provincial Health Commission (No. 2018KY689), and Ningbo Health Branding Subject Fund (No. PPXK201805).

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