Check for updates

OPEN ACCESS

EDITED BY Ozge Yilmaz Manisa Celal Bayar University, Türkiye

REVIEWED BY Moiz Ashraf Ansari, Texas A&M University, United States Avsen Yıldırım, Manisa Celal Bayar University, Türkiye

*CORRESPONDENCE Ute Fischer ute.fischer@med.uni-duesseldorf.de Ersen Kameri ersen.kameri@med.uni-duesseldorf.de

†These authors have contributed equally to this work

RECEIVED 06 June 2024 ACCEPTED 20 December 2024 PUBLISHED 13 January 2025

CITATION

Kameri E, Jepsen VH, Stachura P, Rüchel N, Bhave R, Benitez L, Crispi F, Gratacos E, Dragano N, Janssen S, Borkhardt A, Pandyra A, Kögler G and Fischer U (2025) A gut instinct for childhood leukemia prevention: microbiome-targeting recommendations aimed at parents and caregivers. *Front. Public Health* 12:1445113. doi: [10.3389/fpubh.2024.1445113](https://doi.org/10.3389/fpubh.2024.1445113)

COPYRIGHT

© 2025 Kameri, Jepsen, Stachura, Rüchel, Bhave, Benitez, Crispi, Gratacos, Dragano, Janssen, Borkhardt, Pandyra, Kögler and Fischer. This is an open-access article distributed under the terms of the [Creative](http://creativecommons.org/licenses/by/4.0/) [Commons Attribution License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

A gut instinct for childhood leukemia prevention: microbiome-targeting [recommendations aimed at](https://www.frontiersin.org/articles/10.3389/fpubh.2024.1445113/full) parents and caregivers

Ersen Kameri^{1,2,3*}, Vera Helena Jepsen¹, Pawel Stachura^{1,4}, Nadine Rüchel¹, Rigveda Bhave³, Leticia Benitez⁵, Fatima Crispi⁵, Eduard Gratacos⁵, Nico Dragano^{6†}, Stefan Janssen^{7†}, Arndt Borkhardt^{1,8†}, Aleksandra Pandyra^{1,9,10†}, Gesine Kögler^{2,3†} and Ute Fischer^{1,2,8*†}

¹Department of Pediatric Oncology, Hematology and Clinical Immunology, Medical Faculty, Centre of Child and Adolescent Health, Heinrich-Heine-University, Düsseldorf, Germany, ²Cancer Prevention-Graduate School, German Cancer Research Center (DKFZ), Heidelberg, Germany, ³ Institute of Transplantation Diagnostics and Cell Therapeutics, Medical Faculty, Heinrich-Heine-University, Düsseldorf, Germany, ⁴Department of Molecular Medicine II, Medical Faculty, Heinrich-Heine-University, Düsseldorf, Germany, ⁵BCNatal, Fetal Medicine Research Center (Hospital Clinic and Hospital Sant Joan de Déu), University of Barcelona, Institut d'Investigacions Biomédiques August Pi i Sunyer (IDIBAPS), Barcelona, Spain, ⁶Institute of Medical Sociology, Centre for Health and Society, Medical Faculty, Heinrich-Heine-University, Düsseldorf, Germany, ⁷Algorithmic Bioinformatics, Department of Biology and Chemistry, Justus Liebig University Gießen, Gießen, Germany, ⁸German Cancer Consortium (DKTK), Partner Site Essen/Düsseldorf, Düsseldorf, Germany, 9 Institute of Clinical Chemistry and Clinical Pharmacology, University Hospital Bonn, Bonn, Germany, ¹⁰German Centre for Infection Research (DZIF), Partner Site Bonn-Cologne, Bonn, Germany

Childhood leukemia accounts for 30% of all pediatric cancer cases with acute lymphoblastic leukemia (ALL) being the most common subtype. Involvement of the gut microbiome in ALL development has recently garnered interest due to an increasing recognition of the key contribution the microbiome plays in maintaining the immune system's homeostatic balance. Commensal gut microbiota provide a first line of defense against different pathogens and gut microbiome immaturity has been implicated in ALL pathogenesis. Several environmental factors such as nutrition, mode of delivery, breastfeeding and, early social or livestock contacts are known to alter the composition of the gut microbiota. Variations in these factors influence the risk of childhood leukemia onset. This review aims to elucidate the risk factors influencing microbial composition in the context of childhood ALL. The link between gut microbiome diversity and childhood ALL offers the opportunity to develop risk-reducing strategies that can be communicated to a broad target population of (future) parents and caregivers for childhood leukemia prevention. Here, we summarize evidence on how promoting a diverse gut microbiome in newborns through simple measures such as increasing social contacts early in life may decrease the risk of developing ALL in these children later on.

KEYWORDS

childhood leukemia, risk factors, gut microbiome, prevention, recommendations, public health

1 Introduction

Cancer is the second most frequent cause of death among children in developed countries and acute lymphoblastic leukemia (ALL) is the most common subtype accounting for 30% of childhood cancer cases [\(1\)](#page-10-0). ALL incidence peaks at 2–4 years of age [\(Figure 1A\)](#page-2-0) and is increasing steadily [\(2\)](#page-10-1). Around 80% of childhood ALL cases are characterized by proliferation of abnormal B-cell progenitors characterized as B-cell precursor ALL (BCP-ALL) [\(3\)](#page-10-2). Although the 5-year survival rate of children with BCP-ALL has improved significantly, there are several adverse side effects associated with treatment [\(4\)](#page-10-3). Today, there are about half a million childhood cancer survivors living in Europe and two-thirds of them suffer from acute and late treatment-related toxicities, accounting for a large proportion of deaths [\(4,](#page-10-3) [5\)](#page-10-4). For instance, chemotherapy treatment for childhood ALL can affect all organs and cause acute and persistent organ damage [\(4\)](#page-10-3). Common acute adverse effects include opportunistic infections, mucositis, neuropathy, thromboembolism, bone toxicities, endocrinopathies, hypersensitivity, pancreatitis, nephrotoxicity, thrombosis, and hyperlipidemia [\(3,](#page-10-2) [4\)](#page-10-3). Long-term toxicity stemming from the treatment such as cognitive impairment, osteonecrosis, secondary cancers, infertility and, depression, can be severe and alter the socioeconomic participation of at least half of those affected [\(6\)](#page-10-5). The increasing childhood cancer incidence in Europe highlights the need to shift the current paradigm from therapy to prevention [\(7\)](#page-10-6). Preventive strategies could circumvent traumatic and toxic treatments, associated life-long health sequelae, and the experience of relapse or treatment resistant leukemia subtypes, which occur in about 20% of cases [\(8\)](#page-10-7). In addition, adopting preventive strategies could significantly reduce the cumulative public health burden incurred by an increasing number of adult cancer survivors that suffer from the aftereffects of treatment and a decreased life quality [\(5\)](#page-10-4).

Childhood ALL is frequently triggered by genetic mutations somatically acquired before birth [\(Figure 1B\)](#page-2-0) [\(3\)](#page-10-2). The most common event is the translocation t(12,21)(p13;q22) generating the *ETV6::RUNX1* fusion gene [\(3\)](#page-10-2). Secondary oncogenic gene alterations are necessary for overt leukemia and are likely driven by modifiable environmental and lifestyle factors, therefore making ALL in principle a preventable disease [\(9\)](#page-10-8). Several factors known to increase the risk of developing ALL also have a strong impact on the composition of the gut microbiota [\(Figure 2\)](#page-3-0).

Indeed, a potential causative involvement of the microbiome in the development of ALL is increasingly recognized [\(10\)](#page-10-9). This is particularly relevant in the context of Greaves' delayed infection theory of childhood leukemia development proposing that exposure to common infectious agents may trigger leukemia in genetically predisposed children [\(11\)](#page-10-10). The delayed infection theory highlights the role of typical common childhood infections, including respiratory or gastrointestinal pathogens, which might influence immune system dysregulation in genetically susceptible children [\(11\)](#page-10-10). There are reports of BCP-ALL space-time clusters associated with different specific pathogens: adenovirus for 13 patients in the Fallon cluster, streptococcal fever for 8 same-school patients in the Niles cluster, and influenza A H1N1 swine flu virus for seven patients in the Milan cluster [\(12–](#page-10-11)[15\)](#page-10-12). Furthermore, Christoph et al. identified virus sequences corresponding to common human pathogens, such as *Anelloviridae, Herpesviridae*, and *Parvoviridae* family in 11 B-ALL cases analyzed by whole genome sequencing [\(16\)](#page-10-13). In addition, a study in UK observed peaks of BCP-ALL ∼6 months after seasonal influenza epidemics. According to Greaves (as illustrated in [Figure 1B\)](#page-2-0), if the exposure of a child's immune system to infectious triggers is delayed due to lack of social contacts or an exaggerated hygiene, the eventual immune response may be dysregulated and lead to the progression of pre-leukemic cells [\(3\)](#page-10-2).

The gut microbiome and immune system co-develop during early infancy and childhood. During this period the gut microbiome plays a major role in shaping host's immunity [\(10\)](#page-10-9). Commensal microbiota provide a first line of defense against different pathogens, and gut microbiome immaturity due to genetic or lifestyle factors results in untrained immunity which could promote the switch toward overt leukemia in genetically predisposed children [\(10\)](#page-10-9). *In vivo* mouse models demonstrated that the lack of commensal microbiome alone could be sufficient to promote leukemia in genetically predisposed mice, simply by inducing a microbiome disruption via early-life antibiotic treatment [\(2\)](#page-10-1). Infections combined with an altered microbiome composition may not only provide sufficient proliferative stimuli for pre-leukemic cells to induce the acquisition of secondary oncogenic driver mutations but also drive a pro-inflammatory and immunosuppressive hematopoietic niche supporting a leukemiafavorable microenvironment [\(3,](#page-10-2) [10\)](#page-10-9).

The gut microbiome and the immune system develop simultaneously starting *in-utero*. Different prenatal and postnatal factors influence this process [\(Figure 3\)](#page-4-0) [\(17\)](#page-10-14). During pregnancy, placental transmission of bacterial-derived metabolites originating from maternal diet and microbiota initiate the priming and development of the immune system, emphasizing the importance of a healthy maternal nutrition in preventing ALL [\(17,](#page-10-14) [18\)](#page-10-15). Although essential components of both innate and adaptive immunity already develop at the prenatal stage, they predominantly evolve after birth alongside with the diversification of the microbiome [\(17,](#page-10-14) [19\)](#page-10-16). At birth, the mode of delivery directly determines the most abundant type of bacteria present in a newborn's intestine [\(17\)](#page-10-14). Gut microbiome of vaginally delivered children mainly consists of gram-negative bacteria capable of synthesizing lipopolysaccharides, which are known to be effective stimulators of innate immunity [\(20\)](#page-10-17). In contrast, the microbiome of cesarean section (C-section) delivered babies is mainly composed of opportunistic pathogens circulating in the hospital's environment [\(21\)](#page-10-18). These initial variations in microbial composition can strongly impact the innate lymphoid cell maturation. However, any defects in essential microbiota can be mitigated by immediate parent/caregiver-baby skin-on-skin contact or maternal vaginal/fecal microbiota transplantation to the baby, known as vaginal seeding [\(20–](#page-10-17)[22\)](#page-10-19). During lactation, breastfeeding provides specific prebiotics (such as inulin and human oligosaccharides) that stimulate the growth of commensal microbiota and thereby affect immune responses, particularly those of the innate arm [\(23\)](#page-10-20). Eventually, the gut microbiome matures due to social and livestock interactions, infections and possibly active immunization via vaccination, resulting in an increased microbial diversity, which

has a profound impact on both adaptive and mucosal immunity [\(17,](#page-10-14) [24\)](#page-10-21).

In this review, we summarize the current knowledge on childhood leukemia risk factors that are known to influence the immune system and the microbial constitution of the child's gut. We discuss evidence on how factors such as childbirth mode, breastfeeding, commercial milk substitutes, early life social and livestock contacts, and vaccination may influence microbiome composition and increase or reduce the risk of ALL development. We also discuss how parents or caregivers can compensate for lack of microbial seeding in newborns after birth. Based on the collected evidence, we eventually provide simple recommendations that can be communicated to parents and caregivers to reduce the individual risk of childhood leukemia.

2 Fetal stage

2.1 Maternal nutrition and ALL risk

Robust evidence indicates that microbiome colonization already starts *in utero* [\(19,](#page-10-16) [25\)](#page-10-22). Maternal diet influences fetal development by affecting epigenetic, DNA synthesis and repair processes. Furthermore, maternal diet affects fetal immune establishment and may potentially impact leukemia initiation [\(25\)](#page-10-22). Maternal diet may influence the infant gut microbiome composition through vertical microbial transmission via vaginal delivery and breastfeeding, contributing to the infant's immune

development [\(26\)](#page-11-0). A study of mother-infant couplets recruited in the New Hampshire Birth Cohort investigating the association between maternal diet components and fetal microbiome confirms the influence of maternal diet on the infant gut microbiome as stratified by the delivery mode [\(26\)](#page-11-0). The large multiethnic casecontrol California Childhood Leukemia Study (CCLS), concluded that higher maternal diet quality, rich in one-carbon nutrients and vitamin supplements before and during pregnancy correlated with a lower risk for ALL in offspring (OR = 0.88 , Cl 0.78– 0.98) and similarly a reduced risk for acute myeloid leukemia (AML) (OR = 0.76 , Cl $0.52-1.11$) [\(26\)](#page-11-0). A recent meta-analysis on maternal diet and ALL risk indicated an inverse relationship between ALL risk and maternal consumption of fruits (OR 0.71; 95% Cl 0.59–0.86), as opposed to coffee intake (OR 1.45; 95%, Cl, 1.12–1.89) [\(27\)](#page-11-1). This could be explained by the fact that fruits are a source of vitamins, minerals, and folate, all known to be implicated in DNA methylation and repair [\(25,](#page-10-22) [27\)](#page-11-1) adjusted for maternal educational attainment and gestational diabetes, but not for socio-economic or maternal health status. In this study, adjustments were done for maternal educational attainment and gestational diabetes, but not for maternal health conditions or socioeconomic status. Similarly, several studies associate an elevated childhood leukemia risk to a diet low in vitamin A and minerals (particularly selenium) and report a direct association of reduced ALL risk and maternal diet containing eggs, seafood, fish and poultry meat, with the exception of red or processed meat [\(28](#page-11-2)[–30\)](#page-11-3). Eggs, fish and poultry are respectively rich in choline, folate and omega-3 fatty acids,

which play an essential role in histone modification, antiinflammatory processes and/or DNA methylation, reducing the likelihood of epigenetic alterations that may lead to chromosomal aberrations predisposing for ALL. Caffeine intake may inhibit DNA topoisomerase II, potentially giving rise to chromosomal aberrations described in childhood ALL [\(22,](#page-10-19) [31,](#page-11-4) [32\)](#page-11-5). These findings apply to other food/drinks that contain DNA topoisomerase II inhibitors [\(Table 1\)](#page-5-0). Moreover, studies within the NewGeneris cohort have examined maternal caffeine intake and an increased frequency of micronuclei in neonatal blood, linking dietary caffeine exposure to chromosome instability, genome rearrangements, and mutagenesis. However, chromosomal aberrations have not been analyzed directly.

Herbal tea intake in pregnancy seems to uphold protective properties against ALL occurrence, potentially due to its low caffeine content and richness in flavonoids engaged in anti-proliferative and antioxidant processes [\(33,](#page-11-6) [34\)](#page-11-7). Tobacco and alcohol intake evidently remain a threat due to their interference respectively with caffeine and folate metabolism [\(25,](#page-10-22) [35\)](#page-11-8).

Furthermore, dietary carcinogens including nitrosamines, ingested by the mother can pass through the placenta and

expose the fetus to pre-leukemic transformative stimuli and increase the risk of childhood leukemia development [\(36\)](#page-11-9). The NewGeneris Cohort study assessed the transplacental transmission of biomarkers of dietary exposure to carcinogens, such as oxidative fat metabolites, acrylamide, PAHs and nitrosamines [\(Table 2\)](#page-5-1) in 1151 newborn cord blood samples. This was done by quantifying the levels of reactive metabolites bound to either hemoglobin or DNA [\(36\)](#page-11-9). This technique primarily focuses on detecting adducts, which are formed when reactive metabolites bind to hemoglobin or DNA, as biomarkers of exposure to genotoxic compounds. Hemoglobin and DNA adducts were analyzed using high resolution mass spectrometry, whereas dioxin was measured via a commercially available validated bioassay (Dioxin Responsive Chemical Activated LUciferase gene eXpression or short "DR CALUX" bioassay). Indeed, most of the newborns were exposed to (pre)carcinogens and a successive analysis of this study found a significant association between the numbers of micronuclei in the cord blood lymphocytes and the level of exposure to maternal dietary carcinogens [\(36\)](#page-11-9). Micronuclei are cytogenetic biomarkers whose frequency correlates to carcinogen-induced cancer risk. The study above highlights potential molecular

mechanisms that may contribute to *in-utero* carcinogen-induced leukemia [\(36\)](#page-11-9).

Taken together, ALL is a multifactorial disease, whose development relies on an interplay between genetic and several environmental factors, including maternal exposure to dietary carcinogens. Current studies suggest that reducing exposure to dietary carcinogens could potentially mitigate genetic predispositions to ALL [\(Table 2\)](#page-5-1). However, further research needs to be done to better understand the connection between maternal nutrition during pregnancy and the causative genetic factors. This should be analyzed in carefully controlled large mother-newborn cohorts, including for instance, dietary interventions. Investigating the impact of maternal antioxidant intake, or Mediterranean diet on ALL risk could provide valuable insights. In addition, mechanistic studies are necessary to clarify the role of maternal

folate and omega-3 fatty acids in regulating DNA methylation and maintaining normal epigenetic marks in developing fetal cells.

3 Birth to infancy

3.1 Mode of delivery, prebiotics, vaginal seeding, skin-to-skin care, breastfeeding, and ALL risk

Mode of birth delivery represents a major contributor of microbiome colonization in newborns directly after birth [\(37\)](#page-11-10). Vaginal delivery is considered as one of the most important early microbiome colonizing factors [\(38\)](#page-11-11). This is supported by the fact that children delivered vaginally show a higher T-cell reactivity (lasting up to age of 2 years) and a more diverse gut microbiota composition compared to C-section delivered children [\(39\)](#page-11-12). However, three large US case-control population-based studies of C-section delivery and childhood ALL reveal no strong association [\(40,](#page-11-13) [41\)](#page-11-14). A similar finding was reported by the United Kingdom Childhood Cancer Study (UKCCS) [\(42\)](#page-11-15). Other studies were able to identify a direct link between mode of delivery and overt childhood leukemia [\(38,](#page-11-11) [43\)](#page-11-16). In a Californian registry-based casecontrol study, stratification of cases was done according to the major leukemia subtypes [\(38\)](#page-11-11). Analysis of the correlation between C-section and childhood leukemia risk was based on a logistic regression model adjusted for accepted influencing leukemia risk factors such as breastfeeding, gestational age, household income and Hispanic ethnicity [\(38\)](#page-11-11). Further stratified analyses revealed that a strong association exists C-section delivery and childhood ALL among Hispanic mother and child dyads (OR, 2.34; 95% CI, 1.23–4.46) [\(38\)](#page-11-11).

TABLE 1 DNA topoisomerase II inhibitors present in food and environment [\(31\)](#page-11-4).

It is commonly accepted that vaginal delivery accounts for a large mother-to-neonate microbial transmission [\(44\)](#page-11-17). However, the conflicting evidence on C-section delivery and ALL risk makes a link between delivery-related microbiome alterations and ALL uncertain. Furthermore, the reduced share of maternal microbiome during C-section delivery can still be compensated for after birth by other mother-to-child microbial transmission routes like skin contact, breastfeeding or even prebiotic use and vaginal microbiome seeding [\(22,](#page-10-19) [23,](#page-10-20) [45\)](#page-11-18). In terms of childhood leukemia prevention, inulin and human milk oligosaccharides (HMO) are considered as useful prebiotics due to their beneficial effect on stimulating the growth of benign commensal gut bacteria. Benign commensal gut bacteria reduce oxidative stress and decrease gut colonization by *Fusobacterium*, a bacterium known to display procancer properties [\(23\)](#page-10-20).

Vaginal seeding, a newly emerged concept, consists of mainly oral administration of vaginal fluid to newborns delivered via C-section, aiming to compensate for the lack of microbiome exchange in absence of a vaginal birth [\(46\)](#page-11-19). This method of maternal bacteria transfer has been recommended by researchers due to a rising prevalence of C-section births [\(46\)](#page-11-19). In the first study of vaginal seeding, published in 2016, Dominguez-Bello et al., showed that exposing C-section born children to their mother's vaginal fluid could enrich the gut microbiome similarly to the vaginally delivered counterparts [\(46\)](#page-11-19). A more recent observational study including a larger number of newborns delivered via C-section mode that underwent vaginal seeding also reports a comparable microbiome composition between vaginally and C-section delivered babies [\(22\)](#page-10-19). So far, there are no reported adverse side effects due to this intervention, including transmitted infections. However, as with vaginal deliveries, vaginal seeding carries the risk of hepatitis B virus, hepatitis C virus, HIV or herpes simplex virus (HSV) transmission [\(47\)](#page-11-20). Considering these risks, we recommend more conventional alternatives of infant microbiome seeding such as skin-to-skin contact or breastfeeding.

TABLE 2 Newborn exposure to carcinogens via maternal dietary intake during pregnancy.

Data collected from NewGeneris database [\(36\)](#page-11-9). † 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) toxic equivalency quotients.

The share of maternal microbiome to the child across six maternal and four infant body sites was calculated employing a fast-expectation maximization microbial source tracking algorithm [\(45\)](#page-11-18). Breastfeeding was the predominant contributor of infant microbiome colonization (31.6%). However, a newborns' microbiota development benefitted from contact with other maternal sites including skin (25.7%), saliva (18.6%), nasopharynx (9.4%), feces (4.1%), and vagina (3.5%) [\(45\)](#page-11-18). Importantly, 58.5% of infant microbiota can originate from any of the maternal transmission sources and the limited maternal microbial share due to a C-section delivery can easily be compensated [\(45\)](#page-11-18).

Additional evidence attests to the essential role of skin-to-skin contact in building up a healthy newborn immunity by equipping the baby with beneficial microbes including *Staphylocococcus epidermis* which prevents potential pathogen colonization and exerts anti-inflammatory properties [\(48\)](#page-11-21). Newborns experiencing skin-to-skin contact during the 1st h after birth revealed a greater share of maternal microbiota and consequently a more diverse microbiome compared to neonates without skin-to-skin contact [\(49\)](#page-11-22). Taken together, immediate after birth skin-to-skin care, either with the infant's mother or a caregiver, is highly recommended [\(50\)](#page-11-23). Additionally, several studies report a likelihood for the mother to initiate and carry on the breastfeeding practice after having experienced immediate skin-to-skin contact with their babies [\(51\)](#page-11-24). That is associated with an elevated level of oxytocin, which is a vital lactation hormone that not only improves bonding and reduces stress levels, but also facilitates the process of milk release during breastfeeding [\(52\)](#page-11-25).

Maternal antibodies, passively transferred to neonates through breastfeeding, provide a crucial protection against pathogens during early life [\(53\)](#page-11-26). In a recent study it was found that antibody-mediated protective immunity can be obtained from the commensal microbiome of pregnant mice via breastfeeding [\(54\)](#page-11-27). Studies examining breast milk composition discovered that puerperium-stage milk (<1 week after birth) consists of ∼70% immune cells which drop down to 0–2% in the postpartum period (>2 weeks after birth) [\(55,](#page-11-28) [56\)](#page-11-29). Furthermore, profiling of mature-stage milk revealed three previously unknown and unique epithelial lactocyte subpopulations found to play a pivotal role in immune defense and intestinal development [\(55\)](#page-11-28). Bogeart et al., described not only decreased microbial transmission resulting from the C-section births but, a more significant impact of breastmilk related to a higher amount of *Rothia mucilaginosa* presence found in the fecal microbiota of C-section born children [\(45\)](#page-11-18). This bacterium has been shown to positively influence the gut microbiota by altering its composition. In mice studies, *R. mucilaginosa* increased the abundance of beneficial bacteria such as *Firmicutes* and *Lactobacillus* while reducing harmful bacteria like *Bacteroidetes* [\(57\)](#page-11-30). These changes enhance gut health by promoting nutrient absorption and metabolic balance. This metabolic activity also produces short-chain fatty acids and other metabolites that have systemic effects, influencing gut health and maintaining immune balance by interacting with immune cells and cytokines [\(57\)](#page-11-30). Preliminary studies suggest that some *R. mucilaginosa* exhibits anti-inflammatory properties. For instance, its abundance was found to be negatively correlated with proinflammatory markers such as interleukin IL-8 and IL-1β in

a cohort of adults with bronchiectasis [\(58\)](#page-11-31). This may help in modulating inflammation by interacting with Toll-like receptors (TLRs) and other immune pathways [\(58\)](#page-11-31). In addition, the findings from Bogeart et al., reiterate that breastfeeding may partially make up for the reduced infant microbial seeding upon C-section delivery and breastmilk represents the biggest contributor to newborn gut microbiome (31.6% of microbial content) compared to other afterbirth transmission routes [\(45\)](#page-11-18). Large meta-analysis studies from 2015 and 2021 suggest that continuation of breastfeeding for at least 6 months may result in a decrease of childhood leukemia incidence by 14–20% [\(59\)](#page-11-32). However, there is noticeable differences in childhood leukemia rates and breastfeeding practices between high-income (HIC) and medium-low income countries with lower breastfeeding rates in HIC. Clearly, maternal socioeconomic status influences the choice to breastfeed and its duration. Aiming to rule out the socioeconomic differences, an analysis was carried out focusing only on 12 studies conducted in the HIC countries. The result unveiled a statistically significant inverse correlation between childhood leukemia and breastfeeding for over 6 months (OR, 0.84; 95% CI, 0.78–0.91) [\(59\)](#page-11-32). Breastfeeding offers a low-cost and usually accessible public health measure for childhood leukemia prevention. However, breastfeeding practice faces many barriers, including lactation problems, infant behavior, early return to work, socioeconomic status, lack of social support and self-efficacy, and unsupportive childcare [\(60\)](#page-11-33). Hence, for the mothers unable to breastfeed, immediate skin-to-skin contact, early life social, and livestock contact, vaccination, as well as daycare attendance provide alternative and effective stimuli for an early priming of the infant's immunity.

It is calculated that infants receive a load of up to 1 million immune cells in every feeding [\(61\)](#page-11-34). Human milk, in comparison to commercial sources, consists of maternal immune cells and prebiotics (e.g., HMOs) [\(56\)](#page-11-29). For instance, HMOs offer an essential protection in the context of an immature immunity and represent the third largest component of breastmilk. By contrast, commercial milk only contains traces of this complex sugar [\(61\)](#page-11-34). For over a decade, market statistics show a drastic increase of babies fed with commercial milk formula instead of human milk [\(62,](#page-11-35) [63\)](#page-11-36). A three-paper series published in 2023 in Lancet raises the concern of challenged breastfeeding practice due to highly predatory tactics used by formula milk industry [\(64\)](#page-11-37). In one of the series, they outline the long-term benefits of breastfeeding in fighting disease for both mothers and the newborns by providing an immune boost that cannot be reproduced by commercial milk substitutes [\(65\)](#page-11-38). Examining the relationship between exclusive breast milk and pure milk powder in preventing leukemia, a large retrospective case-control study of children diagnosed with leukemia vs. healthy controls found that the consumption of commercial milk powder instead of breastmilk might significantly increase the incidence of overt childhood leukemia [\(66\)](#page-11-39). Whereas, analysis of the association between duration of breastfeeding and childhood leukemia incidence predictably indicated a slightly reduced leukemia risk upon >6 months of breastfeeding [\(67\)](#page-11-40). Although a better understanding of the biological mechanisms between breastfeeding and risk of childhood leukemia is needed, existing data indicates a protective effect of breastmilk against leukemia development.

4 Early childhood

4.1 Social and/or livestock contacts and ALL risk

Early life microbial colonization is essential for the maturation of immune system and originates from the maternal microbiota [\(37\)](#page-11-10). Although, microbial colonization might commence *in utero*, it is a persistent natural process and its largest share happens after birth. Hence, daycare attendance could help maintain a nurtured microbiome and facilitate immune system maturation through early exposure to common infections, which in turn may reduce the risk of ALL development [\(68\)](#page-11-41). In a Danish childcare database study the ALL risk for children attending childcare is estimated to be reduced by 32% [\(69\)](#page-11-42). However, these findings are not supported by a second recent Danish cohort study in which, childcare attendance shows no significant reduction in the context of ALL risk reduction [\(68\)](#page-11-41). Since the enrolment age was 2–14 years in the later study, it could be inferred that immune modulation following microbiome alterations is critical to ALL risk during the 1st years of life [\(68\)](#page-11-41).

The "delayed infection" theory postulates that early infections decrease the risk of childhood ALL. Early infections have shown to be beneficial in priming the immune system [\(3\)](#page-10-2). Epidemiological studies on exposure to infectious agents and immune challenges by proxy in infancy (<1 year of age) support a protective effect of early infections against ALL [\(3\)](#page-10-2). Taking this into account, we suggest early life livestock contacts, siblingship, or daycare attendance as they may provide the necessary after-birth microbiome colonization and subsequently protection against ALL.

In line with the "delayed infection" theory, a small ALL cluster was identified in Milan among seven children following AH1N1 swine flu [\(12\)](#page-10-11). No evidence of previous exposure to ionizing and non-ionizing radiation or other leukemic causative chemicals was uncovered. Since none of the children attended nursery during the 1st year of life and six out of seven were firstborn, it is probable that lack of exposure to infections during early life became the reason of these ALL cases [\(12,](#page-10-11) [70\)](#page-11-43). Interestingly, a German nationwide, populationbased assessment of the influence of COVID-19 pandemic on the incidence of BCP-ALL childhood cancers revealed a general increase in 2020, which dropped below average in 2021 [\(71\)](#page-11-44).

Additionally, a large-scale epidemiological study conducted after Germany's reunification showed a 25% higher childhood ALL incidence in the former East Germany as compared to only 1% increase in West Germany and the rest of Europe, over the same period of 6 years [\(72\)](#page-11-45). In contrast to West Germany, nearly all the babies in the eastern part had to attend state nursery centers, a practice which was discontinued after reunification [\(72\)](#page-11-45). Hence, cessation of universal daycare attendance after reunification may in part account for this shift [\(72\)](#page-11-45). Further supportive evidence on this stems from a recent meta-analysis of 7,847 leukemia cases and 11,667 controls by the Childhood Leukemia International Consortium [\(73\)](#page-12-0). The consortium demonstrated that regular contact of children (<1 year of age) with livestock, poultry and pets reduced the risk of ALL development significantly (livestock: $OR = 0.65$, 95% CI: 0.50, 0.85) [\(73\)](#page-12-0).

4.2 Vaccination and ALL risk

Acquiring infections during the 1st year of life strengthens the immune system due to antibody production [\(74\)](#page-12-1). "Trained immunity" during early life resulting from vaccination is a newly emerging concept which could impact childhood leukemia prevention as it primes the immune system and boosts immunity [\(75\)](#page-12-2). For instance, the Bacille Calmette-Guerin (BCG) vaccination is thought to shape the innate immune response by production of trained natural killer cells (NK), macrophages and monocytes [\(76\)](#page-12-3). So far, many epidemiological meta-analyses investigating the association between early life vaccination and ALL risk have been carried out, but only BCG vaccination has shown a protective effect in terms of childhood ALL prevention [\(74,](#page-12-1) [76\)](#page-12-3). In a retrospective study, R. Rosenthal was the first to describe the role of BCG vaccination in childhood ALL incidence declension in 1972 [\(77\)](#page-12-4). Since then, other observational studies suggest the valuable effect of BCG vaccination in reducing leukemia incidence [\(75,](#page-12-2) [78\)](#page-12-5). However, no firm conclusion can be made as the outcome of these studies were often inconsistent. A lack of assessment of other ALL influencing factors including environmental stimuli (microbes/etiological agents), social contacts or mother-to-child interaction may account for the variable outcome. Notably, countries with an active BCG vaccination policy report the lowest childhood leukemia incidence [\(74,](#page-12-1) [77\)](#page-12-4). The protective association between BCG vaccine and childhood leukemia rate was also observed while comparing ALL cases among children living in either East or West Germany before and after the reunification. In the former East, opposite to West Germany, BCG vaccination was mandatory (in addition to daycare attendance), until reunification [\(76\)](#page-12-3). Accordingly, data showed a lower childhood leukemia incidence in the former East Germany with 3/100,000 children as compared to 3.7/100,000 in former West Germany, but reunification eventually canceled out this disparity [\(76\)](#page-12-3). The influence of different vaccines, such as rubella, measles, mumps, diphtheria-tetanus-pertussis (DTaP) poliomyelitis, hepatitis B (HBV), and BCG in affecting the incidence of childhood ALL has also been explored in a large meta-analysis study [\(74\)](#page-12-1). Results showed no evidence of reduced leukemia risk for all the other vaccines, whereas analysis of early vaccination (<3 months of age) with the BCG vaccine revealed a statistically robust protection from ALL [\(74\)](#page-12-1). These associations are supported by numerous studies reporting on BCG vaccination of newborns and leukemia incidence in Austria, Chicago, Quebec and Germany [\(74\)](#page-12-1).

The hypothalamus-pituitary-adrenal axis hypothesis by Schmiegelow proposes that early life infections lead to an increase in plasma cortisol levels and may consequently facilitate the elimination of preleukemic cells [\(69\)](#page-11-42). Vaccine administration can also increase plasma cortisol, which might account for how vaccination may prevent childhood ALL development [\(79\)](#page-12-6). Additionally, in line with Greaves' delayed infection theory, exposure to vaccines' pathogen-associated molecular patterns

(PAMPs) stimulates innate immune response, which could mimic common infections acquired due to nursery attendance [\(74\)](#page-12-1). However, further research is necessary to elucidate this concept, as this does not explain why reduction of leukemia risk appears to be limited to only the BCG vaccine.

5 Discussion and recommendations for the target population

Microbiome-targeted interventions are an emerging area of research for preventing ALL in genetically predisposed children and several prevention strategies, including diet modulation and/or early-life microbial exposure are currently under investigation. Epidemiological and experimental studies demonstrate that a healthy gut microbiome holds great potential in protecting against childhood leukemia development [\(2,](#page-10-1) [10,](#page-10-9) [23,](#page-10-20) [25\)](#page-10-22). Several environmental factors (prenatal and especially postnatal) reviewed above may provide the adequate gut microbiome diversity that could aid in the protection against ALL development through the establishment of strong early life immunity [\(Figure 4\)](#page-8-0) [\(4,](#page-10-3) [17,](#page-10-14) [20,](#page-10-17) [23,](#page-10-20) [24,](#page-10-21) [46,](#page-11-19) [69,](#page-11-42) [72,](#page-11-45) [80\)](#page-12-7).

As reported by several epidemiological and experimental studies, diet during pregnancy may strongly influence maternal and fetal microbiome and provide the bacteria needed for "priming" of the offspring's immunity [\(25,](#page-10-22) [26,](#page-11-0) [29,](#page-11-46) [30\)](#page-11-3).

C-section births have generally increased over the past two decades, potentially reducing microbial exchange at birth. While interventions like vaginal/fecal microbiome seeding remain still debatable, we strongly advocate early skin-to-skin mother-toinfant/and, or guardian-to-infant contact to promote microbial

transfer and subsequent early immune priming, as supported by epidemiological studies. Early skin-to-skin contact is an important determinant in shaping a strong immunity. Therefore, an uninterrupted skin-to-skin mother or caregiver-infant contact for at least the 1st hour after birth, particularly for the non-vaginally born babies is recommended. Similarly, early social and livestock contact may offer an additional strategy toward childhood leukemia prevention [\(45,](#page-11-18) [46,](#page-11-19) [48](#page-11-21)[–50,](#page-11-23) [72,](#page-11-45) [73\)](#page-12-0).

Bogaert et al., demonstrates that breastmilk is the major source of newborn microbiota and can greatly compensate on its own for the lack of maternal microbial transmission after a C-section delivery. Hence, if feasible for the mother both physically and mentally, we encourage this practice as well as the implementation of breastfeeding-friendly measures at the workplace and focus on maternal mental wellbeing [\(45,](#page-11-18) [55,](#page-11-28) [56,](#page-11-29) [59\)](#page-11-32).

Additionally, epidemiological studies show that vaccination and early exposure to infections or pathogens have a protective role against childhood leukemia, as well as asthma and allergies. Hence, overtly sterile environments and limited interaction with siblings or pets may serve as a trigger for dysregulated immune responses later on and negatively influence the onset of leukemia among genetically predisposed children [\(74–](#page-12-1)[77,](#page-12-4) [79\)](#page-12-6).

To advance these prevention strategies (dietary adjustments and early-life microbial exposure) further studies are needed to clarify the mechanisms linking gut microbiome alterations to ALL. For families with a genetic predisposition, adopting a healthy diet, avoiding unnecessary antibiotic use, and exploring evidence-based probiotic therapies could be practical steps to support microbiome health and potentially reduce ALL risk [\(1,](#page-10-0) [10,](#page-10-9) [11,](#page-10-10) [23,](#page-10-20) [80,](#page-12-7) [81\)](#page-12-8). However, consulting healthcare professionals before starting any intervention is essential to ensure safety and efficacy.

6 Conclusions

Growing evidence suggests that early childhood ALL is in principle a preventable disease and the key to its prevention is nurturing early life gut microbiome and immune maturation that can be achieved effectively and safely through a healthy maternal diet, breastfeeding, vaginal mode of delivery, early skinto-skin mother/caregiver-to-infant contact, nursery attendance, vaccination, and early life social and livestock contacts to facilitate exposure to common infection agents [\(Figure 5\)](#page-9-0).

Author contributions

EK: Conceptualization, Data curation, Investigation, Visualization, Writing – original draft. VJ: Writing – review

& editing. PS: Writing – review & editing. NR: Writing – review & editing. RB: Writing – review & editing. LB: Writing – review & editing. FC: Writing – review & editing. EG: Writing – review & editing. ND: Writing – review & editing. SJ: Investigation, Writing – review & editing. AB: Funding acquisition, Investigation, Resources, Supervision, Writing – review & editing. AP: Investigation, Supervision, Writing – review & editing. GK: Funding acquisition, Resources, Supervision, Writing – review & editing. UF: Conceptualization, Funding acquisition, Investigation, Resources, Supervision, Validation, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by the Deutsche José-Carreras Leukämie-Stiftung (DJCLS 18R/2021), the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation)—grant no. 495318549, the Deutsche Krebshilfe (DKH, German Cancer Aid) within the "Cancer Prevention—Graduate School" (CPGS)—grant no. 70114736, the Deutsche Kinderkrebsstiftung (DKKS)—grant no.

References

1. Masetti R, Muratore E, Leardini D, Zama D, Turroni S, Brigidi P, et al. Gut microbiome in pediatric acute leukemia: from predisposition to cure. *Blood Adv.* (2021) 5:4619–29. doi: [10.1182/bloodadvances.2021005129](https://doi.org/10.1182/bloodadvances.2021005129)

2. Fischer U, Yang JJ, Ikawa T, Hein D, Vicente-Dueñas C, Borkhardt A, et al. Cell fate decisions: the role of transcription factors in early B-cell development and leukemia. *Blood Cancer Discov*. (2020) 2020:224–33. doi: [10.1158/2643-3230.BCD-20-0011](https://doi.org/10.1158/2643-3230.BCD-20-0011)

3. Greaves M. A causal mechanism for childhood acute lymphoblastic leukaemia. *Nat Rev Cancer.* (2018) 18:471–84. doi: [10.1038/s41568-018-0015-6](https://doi.org/10.1038/s41568-018-0015-6)

4. Andrés-Jensen L, Attarbaschi A, Bardi E, Barzilai-Birenboim S, Bhojwani D, Hagleitner MM, et al. Severe toxicity free survival: physician-derived definitions of unacceptable long-term toxicities following acute lymphocytic leukaemia. *Lancet Haematol.* (2021) 8:e513–e23. doi: [10.1016/S2352-3026\(21\)00136-8](https://doi.org/10.1016/S2352-3026(21)00136-8)

5. Kearns PR, Vassal G, Ladenstein R, Schrappe M, Biondi A, Blanc P, et al. A European paediatric cancer mission: aspiration or reality? *Lancet Oncol.* (2019) 20:1200–2. doi: [10.1016/S1470-2045\(19\)30487-5](https://doi.org/10.1016/S1470-2045(19)30487-5)

6. Fulbright JM, Raman S, McClellan WS, August KJ. Late effects of childhood leukemia therapy. *Curr Hematol Malig Rep.* (2011) 6:195– 205. doi: [10.1007/s11899-011-0094-x](https://doi.org/10.1007/s11899-011-0094-x)

7. Steliarova-Foucher E, Fidler MM, Colombet M, Lacour B, Kaatsch P, Piñeros M, et al. Changing geographical patterns and trends in cancer incidence in children and adolescents in Europe, 1991–2010 (Automated Childhood Cancer Information System): a population-based study. *Lancet Oncol.* (2018) 19:1159–69.

8. Wolfe I, Thompson M, Gill P, Tamburlini G, Blair M, van den Bruel A, et al. Health services for children in western Europe. *Lancet.* (2013) 381:1224– 34. doi: [10.1016/S0140-6736\(12\)62085-6](https://doi.org/10.1016/S0140-6736(12)62085-6)

9. Schafer D, Olsen M, Lahnemann D, Stanulla M, Slany R, Schmiegelow K, et al. Five percent of healthy newborns have an ETV6-RUNX1 fusion as revealed by DNA-based GIPFEL screening. *Blood.* (2018) 131:821–6. doi: [10.1182/blood-2017-09-808402](https://doi.org/10.1182/blood-2017-09-808402)

10. Peppas I, Ford AM, Furness CL, Greaves MF. Gut microbiome immaturity and childhood acute lymphoblastic leukaemia. *Nat Rev Cancer.* (2023) 23:565– 76. doi: [10.1038/s41568-023-00584-4](https://doi.org/10.1038/s41568-023-00584-4)

11. Greaves MF. Speculations on the cause of childhood acute lymphoblastic leukemia. *Leukemia.* (1988) 2:120–5.

12. Beneforti L, Dander E, Bresolin S, Bueno C, Acunzo D, Bertagna M, et al. Pro-inflammatory cytokines favor the emergence of ETV6-RUNX1-positive preleukemic cells in a model of mesenchymal niche. *Br J Haematol.* (2020) 190:262– 73. doi: [10.1111/bjh.16523](https://doi.org/10.1111/bjh.16523)

13. Heath CW Jr, Hasterlik RJ. Leukemia among children in a suburban community. *CA Cancer J Clin*. (1990) 40:27–50. doi: [10.3322/canjclin.40.1.29](https://doi.org/10.3322/canjclin.40.1.29)

A2023/31, the Bundesministerium für Bildung und Forschung (BMBF, the German Ministry for Education and Research)—grant no. 01KD2410A (EDI-4-ALL), the Bundesamt für Strahlenschutz (BfS)—grant no. 3622S32231, the Parents' initiative Löwenstern e.V., and the Katharina Hardt-Stiftung.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

14. Francis SS, Selvin S, Yang W, Buffler PA, Wiemels JL. Unusual space-time patterning of the Fallon, Nevada leukemia cluster: evidence of an infectious etiology. *Chem Biol Interact.* (2012) 196:102–9. doi: [10.1016/j.cbi.2011.02.019](https://doi.org/10.1016/j.cbi.2011.02.019)

15. Kroll ME, Draper GJ, Stiller CA, Murphy MF. Childhood leukemia incidence in Britain, 1974-2000: time trends and possible relation to influenza epidemics. *J Natl Cancer Inst.* (2006) 98:417–20. doi: [10.1093/jnci/djj095](https://doi.org/10.1093/jnci/djj095)

16. Christoph B, Ute F, Klaus K, Stefan MP, Michael G, Cai C, et al. Infection as a cause of childhood leukemia: virus detection employing whole genome sequencing. *Haematologica.* (2017) 102:e179–83. doi: [10.3324/haematol.2016.155382](https://doi.org/10.3324/haematol.2016.155382)

17. Kalbermatter C, Fernandez Trigo N, Christensen S, Ganal-Vonarburg SC. Maternal microbiota, early life colonization and breast milk drive immune development in the newborn. *Front Immunol.* (2021) 12:683022. doi: [10.3389/fimmu.2021.683022](https://doi.org/10.3389/fimmu.2021.683022)

18. Younge N, McCann JR, Ballard J, Plunkett C, Akhtar S, Araújo-Pérez F, et al. Fetal exposure to the maternal microbiota in humans and mice. *JCI Insight*. (2019) 4:127806. doi: [10.1172/jci.insight.127806](https://doi.org/10.1172/jci.insight.127806)

19. Li Y, Toothaker JM, Ben-Simon S, Ozeri L, Schweitzer R, McCourt BT, et al. *In utero* human intestine harbors unique metabolome, including bacterial metabolites. *JCI Insight*. (2020) 5:138751. doi: [10.1172/jci.insight.138751](https://doi.org/10.1172/jci.insight.138751)

20. Wampach L, Heintz-Buschart A, Fritz JV, Ramiro-Garcia J, Habier J, Herold M, et al. Birth mode is associated with earliest strain-conferred gut microbiome functions and immunostimulatory potential. *Nat Commun.* (2018) 9:5091. doi: [10.1038/s41467-018-07631-x](https://doi.org/10.1038/s41467-018-07631-x)

21. Shao Y, Forster SC, Tsaliki E, Vervier K, Strang A, Simpson N, et al. Stunted microbiota and opportunistic pathogen colonization in caesarean-section birth. *Nature.* (2019) 574:117–21. doi: [10.1038/s41586-019-1560-1](https://doi.org/10.1038/s41586-019-1560-1)

22. Song SJ, Wang J, Martino C, Jiang L, Thompson WK, Shenhav L, et al. Naturalization of the microbiota developmental trajectory of Cesarean-born neonates after vaginal seeding. *Medicine*[. \(2021\) 2:951–64.e5. doi: 10.1016/j.medj.2021.](https://doi.org/10.1016/j.medj.2021.05.003) 05.003

23. Martyniak A, Zakrzewska Z, Schab M, Zawartka A, Wedrychowicz A, Skoczen S, et al. Prevention and health benefits of prebiotics, probiotics and postbiotics in acute lymphoblastic leukemia. Microorganisms. (2023) and postbiotics in acute lymphoblastic leukemia. *Microorganisms*. (2023) 11:71775. doi: [10.3390/microorganisms11071775](https://doi.org/10.3390/microorganisms11071775)

24. Zimmermann P. The immunological interplay between vaccination and the intestinal microbiota. *NPJ Vaccines.* (2023) 8:24. doi: [10.1038/s41541-023-00627-9](https://doi.org/10.1038/s41541-023-00627-9)

25. Whitehead TP, Metayer C, Wiemels JL, Singer AW, Miller MD. Childhood leukemia and primary prevention. *Curr Probl Pediatr Adolesc Health Care.* (2016) 46:317–52. doi: [10.1016/j.cppeds.2016.08.004](https://doi.org/10.1016/j.cppeds.2016.08.004)

26. Lundgren SN, Madan JC, Emond JA, Morrison HG, Christensen BC, Karagas MR, et al. Maternal diet during pregnancy is related with the infant stool microbiome in a delivery mode-dependent manner. *Microbiome.* (2018) 6:109. doi: [10.1186/s40168-018-0490-8](https://doi.org/10.1186/s40168-018-0490-8)

27. Wiemels JL, Smith RN, Taylor GM, Eden OB, Alexander FE, Greaves MF. Methylenetetrahydrofolate reductase (MTHFR) polymorphisms and risk of molecularly defined subtypes of childhood acute leukemia. *Proc Natl Acad Sci USA.* (2001) 98:4004–9. doi: [10.1073/pnas.061408298](https://doi.org/10.1073/pnas.061408298)

28. Jensen CD, Block G, Buffler P, Ma X, Selvin S, Month S. Maternal dietary risk factors in childhood acute lymphoblastic leukemia (United States). *Cancer Causes Control.* (2004) 15:559–70. doi: [10.1023/B:CACO.0000036161.98734.17](https://doi.org/10.1023/B:CACO.0000036161.98734.17)

29. Kwan ML, Jensen CD, Block G, Hudes ML, Chu LW, Buffler PA. Maternal diet and risk of childhood acute lymphoblastic leukemia. *Public Health Rep.* (2009) 124:503–14. doi: [10.1177/003335490912400407](https://doi.org/10.1177/003335490912400407)

30. Bailey HD, Miller M, Langridge A, de Klerk NH, van Bockxmeer FM, Attia J, et al. Maternal dietary intake of folate and vitamins B6 and B12 during pregnancy and the risk of childhood acute lymphoblastic leukemia. *Nutr Cancer.* (2012) 64:1122– 30. doi: [10.1080/01635581.2012.707278](https://doi.org/10.1080/01635581.2012.707278)

31. Spector LG, Xie Y, Robison LL, Heerema NA, Hilden JM, Lange B, et al. Maternal diet and infant leukemia: the DNA topoisomerase II inhibitor hypothesis: a report from the children's oncology group. *Cancer Epidemiol Biomarkers Prev.* (2005) 14:651–5. doi: [10.1158/1055-9965.EPI-04-0602](https://doi.org/10.1158/1055-9965.EPI-04-0602)

32. Ross JA, Potter JD, Reaman GH, Pendergrass TW, Robison LL. Maternal exposure to potential inhibitors of DNA topoisomerase II and infant leukemia (United States): a report from the Children's Cancer Group. *Cancer Causes Control.* (1996) 7:581–90. doi: [10.1007/BF00051700](https://doi.org/10.1007/BF00051700)

33. Alam M, Ali S, Ashraf GM, Bilgrami AL, Yadav DK, Hassan MI. Epigallocatechin 3-gallate: from green tea to cancer therapeutics. *Food Chem.* (2022) 379:132135. doi: [10.1016/j.foodchem.2022.132135](https://doi.org/10.1016/j.foodchem.2022.132135)

34. Thomopoulos TP, Ntouvelis E, Diamantaras AA, Tzanoudaki M, Baka M, Hatzipantelis E, et al. Maternal and childhood consumption of coffee, tea and cola beverages in association with childhood leukemia: a meta-analysis. *Cancer Epidemiol.* (2015) 39:1047–59. doi: [10.1016/j.canep.2015.08.009](https://doi.org/10.1016/j.canep.2015.08.009)

35. Brown CR, Jacob P 3rd, Wilson M, Benowitz NL. Changes in rate and pattern of caffeine metabolism after cigarette abstinence. *Clin Pharmacol Ther.* (1988) 43:488– 91. doi: [10.1038/clpt.1988.63](https://doi.org/10.1038/clpt.1988.63)

36. Kleinjans J, Botsivali M, Kogevinas M, Merlo DF. Fetal exposure to dietary carcinogens and risk of childhood cancer: what the NewGeneris project tells us. *Br Med J.* (2015) 351:h4501. doi: [10.1136/bmj.h4501](https://doi.org/10.1136/bmj.h4501)

37. Zheng D, Liwinski T, Elinav E. Interaction between microbiota and immunity in health and disease. *Cell Res.* (2020) 30:492–506. doi: [10.1038/s41422-020-0332-7](https://doi.org/10.1038/s41422-020-0332-7)

38. Francis SS, Selvin S, Metayer C, Wallace AD, Crouse V, Moore TB, et al. Mode of delivery and risk of childhood leukemia. *Cancer Epidemiol Biomarkers Prev.* (2014) 23:876–81. doi: [10.1158/1055-9965.EPI-13-1098](https://doi.org/10.1158/1055-9965.EPI-13-1098)

39. Wiemels J, Ma X. Stressful exit from the womb and risk of childhood leukaemia. *Lancet Haematol.* (2016) 3:e155–6. doi: [10.1016/S2352-3026\(16\)00019-3](https://doi.org/10.1016/S2352-3026(16)00019-3)

40. Podvin D, Kuehn CM, Mueller BA, Williams M. Maternal and birth characteristics in relation to childhood leukaemia. *Paediatr Perinat Epidemiol.* (2006) 20:312–22. doi: [10.1111/j.1365-3016.2006.00731.x](https://doi.org/10.1111/j.1365-3016.2006.00731.x)

41. Lupatsch JE, Kreis C, Konstantinoudis G, Ansari M, Kuehni CE, Spycher BD. Birth characteristics and childhood leukemia in Switzerland: a register-based case-control study. *Cancer Causes Control.* (2021) 32:713–23. doi: [10.1007/s10552-021-01423-3](https://doi.org/10.1007/s10552-021-01423-3)

42. Bonaventure A, Simpson J, Ansell P, Roman E. Paediatric acute lymphoblastic leukaemia and caesarean section: a report from the United Kingdom Childhood Cancer Study (UKCCS). *Paediatr Perinat Epidemiol.* (2020) 34:344–9. doi: [10.1111/ppe.12662](https://doi.org/10.1111/ppe.12662)

43. Yang Y, Yu C, Fu R, Xia S, Ni H, He Y, et al. Association of cesarean section with risk of childhood leukemia: a meta-analysis from an observational study. *Hematol Oncol.* (2023) 41:182–91. doi: [10.1002/hon.3070](https://doi.org/10.1002/hon.3070)

44. Mortensen MS, Rasmussen MA, Stokholm J, Brejnrod AD, Balle C, Thorsen J, et al. Modeling transfer of vaginal microbiota from mother to infant in early life. *Elife*. (2021) 10:57051. doi: [10.7554/eLife.57051](https://doi.org/10.7554/eLife.57051)

45. Bogaert D, van Beveren GJ, de Koff EM, Lusarreta Parga P, Balcazar Lopez CE, Koppensteiner L, et al. Mother-to-infant microbiota transmission and infant microbiota development across multiple body sites. *Cell Host Microbe*. (2023) 31:447– 60.e6. doi: [10.1016/j.chom.2023.01.018](https://doi.org/10.1016/j.chom.2023.01.018)

46. Hourigan SK, Dominguez-Bello MG, Mueller NT. Can maternal-child microbial seeding interventions improve the health of infants delivered by Cesarean section? *Cell Host Microbe.* (2022) 30:607–11. doi: [10.1016/j.chom.2022.02.014](https://doi.org/10.1016/j.chom.2022.02.014)

47. Huynh J, Palasanthiran P, McMullan B. Potential transmission of herpes simplex virus via vaginal seeding. *Pediatr Infect Dis J.* (2018) 37:e278. doi: [10.1097/INF.0000000000001965](https://doi.org/10.1097/INF.0000000000001965)

48. Manus MB, Kuthyar S, Perroni-Marañón AG, Núñez-de la Mora A, Amato KR. Infant skin bacterial communities vary by skin site and infant age across populations in Mexico and the United States. *mSystems*. (2020) 5:20. doi: [10.1128/mSystems.00834-20](https://doi.org/10.1128/mSystems.00834-20)

49. Hendricks-Muñoz KD, Xu J, Parikh HI, Xu P, Fettweis JM, Kim Y, et al. Skin-toskin care and the development of the preterm infant oral microbiome. *Am J Perinatol.* (2015) 32:1205–16. doi: [10.1055/s-0035-1552941](https://doi.org/10.1055/s-0035-1552941)

50. Widström AM, Brimdyr K, Svensson K, Cadwell K, Nissen E. Skin-to-skin contact the first hour after birth, underlying implications and clinical practice. *Acta Paediatr.* (2019) 108:1192–204. doi: [10.1111/apa.14754](https://doi.org/10.1111/apa.14754)

51. Safari K, Saeed AA, Hasan SS, Moghaddam-Banaem L. The effect of mother and newborn early skin-to-skin contact on initiation of breastfeeding, newborn temperature and duration of third stage of labor. *Int Breastfeed J.* (2018) 13:32. doi: [10.1186/s13006-018-0174-9](https://doi.org/10.1186/s13006-018-0174-9)

52. Niwayama R, Nishitani S, Takamura T, Shinohara K, Honda S, Miyamura T, et al. Oxytocin mediates a calming effect on postpartum mood in primiparous mothers. *Breastfeed Med.* (2017) 12:103–9. doi: [10.1089/bfm.2016.0052](https://doi.org/10.1089/bfm.2016.0052)

53. Caballero-Flores G, Sakamoto K, Zeng MY, Wang Y, Hakim J, Matus-Acuña V, et al. Maternal immunization confers protection to the offspring against an attaching and effacing pathogen through delivery of IgG in breast milk. *Cell Host Microbe*. (2019) 25:313–23.e4. doi: [10.1016/j.chom.2018.12.015](https://doi.org/10.1016/j.chom.2018.12.015)

54. Zheng W, Zhao W, Wu M, Song X, Caro F, Sun X, et al. Microbiota-targeted maternal antibodies protect neonates from enteric infection. *Nature.* (2020) 577:543– 8. doi: [10.1038/s41586-019-1898-4](https://doi.org/10.1038/s41586-019-1898-4)

55. Gleeson JP, Chaudhary N, Fein KC, Doerfler R, Hredzak-Showalter P, Whitehead KA. Profiling of mature-stage human breast milk cells identifies six unique lactocyte subpopulations. *Sci Adv*. (2022) 8:eabm6865. doi: [10.1126/sciadv.abm6865](https://doi.org/10.1126/sciadv.abm6865)

56. Hassiotou F, Geddes DT, Hartmann PE. Cells in human milk: state of the science. *J Hum Lact.* (2013) 29:171–82. doi: [10.1177/0890334413477242](https://doi.org/10.1177/0890334413477242)

57. Rigauts C, Aizawa J, Taylor SL, Rogers GB, Govaerts M, Cos P, et al. R othia mucilaginosa is an anti-inflammatory bacterium in the respiratory tract of patients with chronic lung disease. *Eur Respirat J*. (2022) 59:1293. doi: [10.1183/13993003.01293-2021](https://doi.org/10.1183/13993003.01293-2021)

58. Song MJ, Kim DH, Kim S-Y, Kang N, Jhun BW. Comparison of the sputum microbiome between patients with stable nontuberculous mycobacterial pulmonary disease and patients requiring treatment. *BMC Microbiol.* (2024) 24:172. doi: [10.1186/s12866-024-03308-2](https://doi.org/10.1186/s12866-024-03308-2)

59. Amitay EL, Keinan-Boker L. Breastfeeding and childhood leukemia incidence: a meta-analysis and systematic review. *J Am Med Assoc Pediatr.* (2015) 169:e151025. doi: [10.1001/jamapediatrics.2015.1025](https://doi.org/10.1001/jamapediatrics.2015.1025)

60. Tomori C. Overcoming barriers to breastfeeding. *Best Pract Res Clin Obstet Gynaecol.* (2022) 83:60–71. doi: [10.1016/j.bpobgyn.2022.01.010](https://doi.org/10.1016/j.bpobgyn.2022.01.010)

61. Walsh C, Lane JA, van Sinderen D, Hickey RM. From lab bench to formulated ingredient: characterization, production, and commercialization of human milk oligosaccharides. *J Funct Foods.* (2020) 72:104052. doi: [10.1016/j.jff.2020.104052](https://doi.org/10.1016/j.jff.2020.104052)

62. Baker P, Smith J, Salmon L, Friel S, Kent G, Iellamo A, et al. Global trends and patterns of commercial milk-based formula sales: is an unprecedented infant and young child feeding transition underway? *Public Health Nutr.* (2016) 19:2540– 50. doi: [10.1017/S1368980016001117](https://doi.org/10.1017/S1368980016001117)

63. Rollins N, Piwoz E, Baker P, Kingston G, Mabaso KM, McCoy D, et al. Marketing of commercial milk formula: a system to capture parents, communities, science, and policy. *Lancet.* (2023) 401:486–502. doi: [10.1016/S0140-6736\(22\)01931-6](https://doi.org/10.1016/S0140-6736(22)01931-6)

64. The L. Unveiling the predatory tactics of the formula milk industry. *Lancet.* (2023) 401:409. doi: [10.1016/S0140-6736\(23\)00118-6](https://doi.org/10.1016/S0140-6736(23)00118-6)

65. Pérez-Escamilla R, Tomori C, Hernández-Cordero S, Baker P, Barros AJD, Bégin F, et al. Breastfeeding: crucially important, but increasingly challenged in a market-driven world. *Lancet.* (2023) 401:472–85. doi: [10.1016/S0140-6736\(22\)01932-8](https://doi.org/10.1016/S0140-6736(22)01932-8)

66. Gao Z, Wang R, Qin ZX, Dong A, Liu CB. Protective effect of breastfeeding against childhood leukemia in Zhejiang Province, PR China: a retrospective casecontrol study. *Libyan J Med.* (2018) 13:1508273. doi: [10.1080/19932820.2018.1508273](https://doi.org/10.1080/19932820.2018.1508273)

67. Güngör D, Nadaud P, LaPergola CC, Dreibelbis C, Wong YP, Terry N, et al. Infant milk-feeding practices and diabetes outcomes in offspring: a systematic review. *Am J Clin Nutr*. (2019) 109(Suppl.7):817s−37s. doi: [10.1093/ajcn/nqy311](https://doi.org/10.1093/ajcn/nqy311)

68. Søegaard SH, Rostgaard K, Kamper-Jørgensen M, Schmiegelow K, Hjalgrim H. Childcare attendance and risk of childhood acute lymphoblastic leukaemia: a register study based on the Danish childcare database. *Int J Cancer.* (2023) 152:1817– 26. doi: [10.1002/ijc.34413](https://doi.org/10.1002/ijc.34413)

69. Kamper-Jørgensen M, Woodward A, Wohlfahrt J, Benn CS, Simonsen J, Hjalgrim H, et al. Childcare in the first 2 years of life reduces the risk of childhood acute lymphoblastic leukemia. *Leukemia.* (2008) 22:189–93. doi: [10.1038/sj.leu.2404884](https://doi.org/10.1038/sj.leu.2404884)

70. Lillie K. Leukaemia and lockdown: the delayed infection model of childhood acute lymphoblastic leukaemia and the COVID-19 pandemic. *Pediatr Blood Cancer.* (2021) 68:e29194. doi: [10.1002/pbc.29194](https://doi.org/10.1002/pbc.29194)

71. Borkhardt A, Schüz J, Trübenbach C, Wellbrock M, Spix C, Erdmann F. Temporal changes of the incidence of childhood B-cell precursor acute lymphoblastic leukaemia in Germany during the COVID-19 pandemic. *Leukemia.* (2022) 36:2908– 11. doi: [10.1038/s41375-022-01730-x](https://doi.org/10.1038/s41375-022-01730-x)

72. Wellbrock M, Spix C, Grabow D, Borkhardt A, Zeeb H, Erdmann F. 28-year incidence and time trends of childhood leukaemia in former East Germany compared to West Germany after German reunification: a study

from the German Childhood Cancer Registry. *Cancer Epidemiol.* (2021) 73:101968. doi: [10.1016/j.canep.2021.101968](https://doi.org/10.1016/j.canep.2021.101968)

73. Orsi L, Magnani C, Petridou ET, Dockerty JD, Metayer C, Milne E, et al. Living on a farm, contact with farm animals and pets, and childhood acute lymphoblastic leukemia: pooled and meta-analyses from the Childhood Leukemia International Consortium. *Cancer Med.* (2018) 7:2665–81. doi: [10.1002/cam4.1466](https://doi.org/10.1002/cam4.1466)

74. Marron M, Brackmann LK, Kuhse P, Christianson L, Langner I, Haug U, et al. Vaccination and the risk of childhood cancer-a systematic review and meta-analysis. *Front Oncol.* (2020) 10:610843. doi: [10.3389/fonc.2020.610843](https://doi.org/10.3389/fonc.2020.610843)

75. Hauer J, Fischer U, Borkhardt A. Toward prevention of childhood ALL by early-life immune training. *Blood.* [\(2021\) 138:1412–28. doi: 10.1182/blood.20200](https://doi.org/10.1182/blood.2020009895) 09895

76. Hauer J, Fischer U, Auer F, Borkhardt A. Regional BCG vaccination policy in former East- and West Germany may impact on both severity of SARS-CoV-2 and incidence of childhood leukemia. *Leukemia.* (2020) 34:2217– 9. doi: [10.1038/s41375-020-0871-4](https://doi.org/10.1038/s41375-020-0871-4)

77. Rosenthal SR, Crispen RG, Thorne MG, Piekarski N, Raisys N, Rettig P, et al. vaccination and leukemia mortality. *Natl Cancer Inst Monogr.* (1973) 39:189–92.

78. Singh S, Kishore D, Singh RK. "Trained Immunity" from *Mycobacterium* spp. exposure (BCG vaccination and environmental) may have an impact on the incidence of early childhood leukemia. *Front Immunol.* (2023) 14:1193859. doi: [10.3389/fimmu.2023.1193859](https://doi.org/10.3389/fimmu.2023.1193859)

79. Azevedo-Silva F, Camargo B, Pombo-de-Oliveira MS. Implications of infectious diseases and the adrenal hypothesis for the etiology of childhood acute lymphoblastic leukemia. *Braz J Med Biol Res.* (2010) 43:226–9. doi: [10.1590/S0100-879X2010007500011](https://doi.org/10.1590/S0100-879X2010007500011)

80. Aronica TS, Carella M, Balistreri CR. Different levels of therapeutic strategies to recover the microbiome to prevent/delay acute lymphoblastic leukemia (ALL) or arrest its progression in children. *Int J Mol Sci.* (2024) 25:3928. doi: [10.3390/ijms25073928](https://doi.org/10.3390/ijms25073928)

81. Zhang Y, Zhao X, Zhang J, Zhang Y, Wei Y. Advancements in the impact of human microbiota and probiotics on leukemia. *Front Microbiol*. (2024) 15:1423838. doi: [10.3389/fmicb.2024.1423838](https://doi.org/10.3389/fmicb.2024.1423838)