Check for updates

OPEN ACCESS

EDITED BY Eberval Gadelha Figueiredo, University of São Paulo, Brazil

REVIEWED BY Roland Nau, University Medical Center Göttingen, Germany Tatjana Pekmezovic, University of Belgrade, Serbia

*CORRESPONDENCE Hannah E. Brindle ⊠ hbrindle@liverpool.ac.uk

RECEIVED 06 March 2024 ACCEPTED 04 December 2024 PUBLISHED 31 January 2025

CITATION

Brindle HE, Choisy M, Christley R, French N, Griffiths M, Thai PQ, van Doorn HR and Nadjm B (2025) Review of the aetiologies of central nervous system infections in Vietnam. *Front. Public Health* 12:1396915. doi: 10.3389/fpubh.2024.1396915

COPYRIGHT

© 2025 Brindle, Choisy, Christley, French, Griffiths, Thai, van Doorn and Nadjm. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Review of the aetiologies of central nervous system infections in Vietnam

Hannah E. Brindle^{1,2*}, Marc Choisy^{3,4}, Robert Christley¹, Neil French¹, Michael Griffiths¹, Pham Quang Thai⁵, H. Rogier van Doorn^{2,4} and Behzad Nadjm^{2,6}

¹Institute of Infection, Veterinary and Ecological Sciences, Faculty of Health and Life Sciences, University of Liverpool, Liverpool, United Kingdom, ²Oxford University Clinical Research Unit, Hanoi, Vietnam, ³Oxford University Clinical Research Unit, Ho Chi Minh City, Vietnam, ⁴Centre for Tropical Medicine and Global Health, University of Oxford, Oxford, United Kingdom, ⁵National Institute of Hygiene and Epidemiology, Hanoi, Vietnam, ⁶The Medical Research Council, The Gambia at London School of Hygiene and Tropical Medicine, Fajara, Gambia

Central nervous system (CNS) infections are an important cause of morbidity and mortality in Vietnam, with many studies conducted to determine the aetiology. However, the cause remains unknown in a large proportion of cases. Although a systematic review of the aetiologies of CNS infections was conducted in the Mekong region, there are no known published reviews of the studies specifically in Vietnam. Here, we review the cause of CNS infections in Vietnam while also considering the potential aetiologies where a cause was not identified, based on the literature from the region. In particular, we focus on the most common pathogens in adults and children including Streptococcus suis which is associated with the consumption of raw pig products, and Japanese encephalitis virus, a mosquito-borne pathogen. We also discuss pathogens less commonly known to cause CNS infections in Vietnam but have been detected in neighbouring countries such as Orientia tsutsugamushi, Rickettsia typhi and Leptospira species and how these may contribute to the unknown causes in Vietnam. We anticipate that this review may help guide future public health measures to reduce the burden of known pathogens and broaden testing to help identify additional aetiologies.

KEYWORDS

central nervous system infection (CNSi), Vietnam, Japanese encephalitis, *Streptococcus suis*, meningitis, encephalitis

Introduction

Central nervous system (CNS) infections are an important cause of morbidity and mortality worldwide with the highest burden in low and middle-income countries. In 2016, encephalitis resulted in 103,000 deaths and 670.4 million Disability-Adjusted Life Years (DALYS) and meningitis, 318,000 deaths and 21.9 million DALYS (1). Vietnam is located in Southeast Asia, neighbouring or in proximity to Cambodia, Laos, Thailand, Myanmar and China. This subregion is a 'hotspot' for emerging and zoonotic infections due to increasing urbanisation, cross-border population movements, farming practices and animal trade, and climate change (2).

In a systematic review of the aetiologies of encephalitis in the Mekong region (defined as Cambodia, Laos, Vietnam, Thailand and Yunnan province in China), Japanese encephalitis virus (JEV) was the most commonly referenced pathogen, accounting for 47% (n = 35) of cases (3). However, there are no known published reviews of the studies conducted only in Vietnam.

This review will describe the prevalence, the epidemiology and the risk factors of the common causes of encephalitis and meningitis in Vietnam based on publications since 1998 (Supplementary Tables S1, S2) (4–15). It will also discuss the rarer causes of CNS infections in the wider region which may be contributing to cases of unknown aetiology in the country.

Bacteria

Streptococcus suis

Streptococcus suis, a Gram-positive coccus, was first detected in Vietnam in 1997 (16, 17) and became a notifiable disease in 2011 (18). The pathogen is found in the gastro-intestinal, genital and respiratory tract of pigs who can develop complications including encephalitis and arthritis (19, 20). Human become infected after consuming or being exposed via skin abrasions, to undercooked pig blood or products (21–23). The incidence of *S. suis* in humans is highest in Southeast Asia where contact with pig products is more common (24). *S. suis* was first detected in Vietnam in 1997 and became a notifiable disease in 2011 (18). The most recent available national surveillance data are from 2017 when there were 178 reported cases (25).

Male gender, older age, having diabetes mellitus and excessive alcohol consumption are all associated with increased risk of infection (21, 26, 27)

and, although it can cause septicaemia, the most common presentation is meningitis (17). Infections in children are very rare as they as less likely to have contact with the raw or uncooked pig products (21). In northern Vietnam, the incidence of human S. suis was shown to be correlated with pig density (26) and it has been suggested that infections of S. suis in humans may correspond with outbreaks of Porcine Respiratory and Reproductive Syndrome (PRRS) which is associated with secondary bacterial infections in pigs (23, 26). No vaccines currently exist against S. suis for humans although candidates are in the pipeline (28). While there are no commercial porcine vaccines, autologous vaccines are used in some circumstances (29). Of the studies outlined in Supplementary Table S1, S. suis was the most common pathogen in adults accounting for 17.6% (n = 448) of pathogens. However, it is much less common in children accounting for only 0.5% (n = 10) of pathogens (Supplementary Tables S3, S4; Figure 1).

In a study of 450 patients admitted to the Hospital for Tropical Diseases in Ho Chi Minh City between 1997 and 2004 with suspected bacterial meningitis, the most common symptoms were headache and neck stiffness, both accounting for 94% (n = 142) of the cases, followed by vomiting which accounted for 66.2% (n = 100) of the cases. Forty-seven (31.1%) of the cases had a Glasgow Coma Score of less than 11. While only four cases died, 40 cases still had sequelae at 6 months after discharge with 14 suffering from severe hearing loss (30). In a more recent study of the outcomes of patients with *S. suis* admitted to the National Hospital for Tropical Diseases in Hanoi between November 2014 and October 2015,



of whom 97.4% (n = 76) had meningitis, the prevalence ratio of vestibular dysfunction at 9 months after discharge in those recruited prospectively (n = 47) compared to controls was 1.8 (95%CI 1.1–2.5) compared to controls (31).

Streptococcus pneumoniae

Streptococcus pneumoniae, a Gram-positive coccus, colonises the upper respiratory tract and is transmitted between humans via contact with infected secretions (32). Children, older people and those with immunosuppression are at greater risk of developing invasive disease such as pneumonia, meningitis and sepsis (33). In many countries, *S. pneumoniae* is the most common cause of bacterial meningitis (34). In Vietnam, it was reported as the second most common pathogen causing CNS infection in both adults (5.4%, n = 138) and children (5.6%, n = 109); (Supplementary Tables S3, S4; Figure 1). In some countries in the global north, where the pneumococcal vaccine has been rolled-out, a reduction in the incidence of pneumococcal meningitis has been seen (34). The pneumococcal vaccine has not yet been introduced in Vietnam, although studies have been conducted to help determine the method of introduction and dosing schedules (35, 36) and it is due to be included in the Expanded Programme for Immunisation (EPI) in 2025.

Mycobacterium tuberculosis

The intracellular mycobacterium Mycobacterium tuberculosis (MTB) is transmitted via aerosolised droplets and can result in pulmonary or less, commonly, extrapulmonary disease (37). It is estimated that 1% of infections result in tuberculous meningitis (TBM) (38) and in a study conducted in Ho Chi Minh City between 2009 and 2011, the majority of cases of TBM occurred in young children (39). Although data from Vietnam in 2022 showed that coverage of the Bacillus Calmette-Guérin (BCG) among those aged one-year in Vietnam was 88% (43), it is possible that TBM remains under-diagnosed including in recent studies where the percentage of cases was less than 3% in both adults and children (Figure 1; Supplementary Tables S3, S4). Laboratory diagnosis of TBM is challenging. A separate study conducted in Ho Chi Minh City (HCMC), Vietnam suggested that at least 6 mL of cerebrospinal fluid (CSF) is needed to culture M. tuberculosis and microscopy using the Ziehl-Neelsen (ZN) method needs to be performed for at least 30 minutes to identify acid-fast bacilli (40). GeneXpert MTB/RIF Ultra is now recommended by the World Health Organisation as the first line diagnostic test for TBM although this had lower negative predictive value in studies where a higher proportion of cases were human immunodeficiency virus (HIV) negative, including in Vietnam (41, 42). Given the challenges with differentiating between bacterial meningitis and TBM solely on clinical features, a diagnostic aid was developed based on age, duration of illness and laboratory features from 251 adults admitted to the Clinical Research Unit at the Centre for Tropical Diseases in Ho Chi Minh City between 1997 and 2000. This tool had a sensitivity of 86% and a specificity of 79% when applied prospectively to 75 adults with either tuberculous or bacterial meningitis (44).

The prognosis for TBM remains poor. In a study conducted in Ho Chi Minh City, 15% (n = 15) of children with TBM died and 33% (n = 27) had a disability at the end of treatment (39). Much research has been conducted in Vietnam on the use of corticosteroids for the treatment of TBM. A randomised, double-blind controlled trial of 545

patients over 14 years of age with TBM conducted in Ho Chi Minh City, showed that the use of dexamethasone reduced the risk of death compared to a placebo (relative risk 0.69; 95% confidence interval (CI) 0.52–0.92, p = 0.01) (45). However, a separate trial also conducted in Ho Chi Minh City found that there was no benefit using dexamethasone compared to placebo among HIV positive adults (46).

Haemophilus influenzae

Haemophilus influenzae, is a Gram negative coccobacillus which can colonise the nasopharynx, especially in children, and can be transmitted via nasal secretions (47, 48). In some cases, it can cause a localised infection such as sinusitis or otitis media and less commonly, pneumonia and meningitis. Invasive disease is mainly due to capsular type b (Hib). The expansion of the vaccine is likely to have contributed to the global decline in cases of meningitis due to Hib from 435,000 cases in 1990 to 187,000 cases in 2019 (a 56.9% reduction), and deaths from 38,000 in 1990 to 11,100 (a 70.8% reduction) (49). In a study conducted in Ho Chi Minh City between 1993 and 1996, cases of Hib accounted for 35% of cases of meningitis in children (4). However, the Hib conjugate vaccine was introduced into the Vietnam National Expanded Programme on Immunisation (EPI) in June 2010 (9) and as a result, cases of Hib cases in children decreased from 34.9% (n = 30) in a study starting in 1995 (4) to 0.5% (n = 1/203) in a study starting 2014 (Figure 1; Supplementary Tables S3, S4) (14).

Neisseria meningitidis

Neisseria meningitidis, a Gram negative coccus, can colonise the nasopharynx and is transmitted via aerosol or secretions (50, 51). Invasive disease can manifest as meningitis, septicaemia, or both (52). In southern Vietnam, the incidence rate between 2011 and 2021 was 0.02 per 100,000 persons/year with serogroup B accounting for 90% of cases (53). However, N. meningitidis it is one of the less common pathogens reported in Vietnam accounting for less than 2% of pathogens in both adults and children (Figure 1; Supplementary Tables S3, S4) (21-23). A recent study showed that antimicrobial resistance, most frequently to chloramphenicol and ciprofloxacin, was common among isolates of invasive meningococcal disease (53). As the availability of meningococcal vaccines increased globally, due to the increase in availability of the meningococcal vaccines, global case numbers of N. meningitidis decreased from 744,000 in 1990 to 433,000 in 2019 (a 41.7% reduction) and deaths from 80,900 to 32,100 (a 60.2% reduction) (49) Vaccination in Vietnam is only available for groups at high-risk and is not part of the national immunisation programme (54). However, in 2022, an expert advisory group recommended that the four component meningococcal B vaccine (4CMenB) start as soon as possible in children (55).

Orientia tsutsugamushi (scrub typhus)

The Gram-negative obligate intracellular bacterium *Orientia tsutsugamushi*, the cause of scrub typhus, is transmitted by the bite of an infected *Leptotrombidium delicense* (trombiculid) mite or 'chigger' which is found in damp soil or detritus in wild, overgrown areas (56–59). A study conducted in Hanoi found that 55.7% (n = 140) of the

patients admitted to hospital with scrub typhus from 2001 to 2003 were farmers (60). Those with scrub typhus develop an eschar, an area of skin necrosis at the site of the bite and localised lymphadenopathy. Clinical symptoms of scrub typhus may then range from fever, cough, headache and myalgia to meningitis and/or multi-organ failure in severe cases (56, 61, 62).

The diagnosis of scrub typhus is challenging as *Orientia tsutsugamushi* is not easily cultured, the PCR detection of DNA in blood has a low sensitivity, and the gold standard indirect immunofluorescence using paired sera (60) is rarely available and requires high levels of expertise. It is therefore possible, that cases of CNS infection due to *Orientia tsutsugamushi* are under-reported, particularly in adults. In recent studies of the aetiology of CNS infections in Vietnam, only one case was detected (in a child) (14). Despite this, in a study of patients of all ages with altered consciousness or neurological findings in Laos, 2.9% (n = 31) had *O. tsutsugamushi* (63) and in a study of patients aged two months to 78 years with acute encephalitis syndrome (AES) in Lucknow, India, 8.7% (n = 357) were diagnosed with the pathogen (64).

Rickettsia typhi

Murine typhus caused by *Rickettsia typhi*, is transmitted when the faeces of infected fleas, which live on rodents, enter the body via bites, abrasions or mucous membranes (65, 66) with cases normally occurring in urban areas (61, 67). Most cases present with a mild illness however, murine typhus can result in severe disease affecting the pulmonary, renal, gastrointestinal and neurological systems (68). Reporting of CNS infections in Vietnam due to *R. typhi* is however, rare. Gabor et al., 2022, found three cases of *Rickettsia* species in adults with clinically suspected CNS infections in Hanoi, which were co-infections with other pathogens (13) and in the study by Pommier et al., 2022, there was one case of *Rickettsia* species in a child in Hanoi who had encephalitis (14). This compares with 2.7% (n = 28) of cases of *Rickettsia typhi* among cases of suspected CNS infection in the study in Laos (63).

Leptospira species

Leptospirosis is caused by the spirochaete Leptospira and is transmitted to humans via the urine of infected mammals, particularly rodents (69). Typically, those with leptospirosis experience fever, myalgia and headache however, in severe cases, renal failure, hepatic failure and pulmonary haemorrhagic may occur (69, 70). On occasion, there may be CNS manifestations (71, 72). Testing for leptospirosis can be challenging as culture and PCR both have a low sensitivity, the former due to the long incubation period required. The microscopic agglutination test is considered the gold standard diagnostic (73), which is technically challenging and often unavailable, likely resulting in under-diagnosis. Perhaps for these reasons, in recent studies of the aetiologies of CNS infections in Vietnam, either diagnostics were either not performed or no cases of leptospirosis were detected (11-15) (Supplementary Table S2). For comparison, Leptospira species were detected in the CSF 2.9% (n = 31) of patients with a suspected CNS infection in the study in Laos (63).

Viruses

Japanese encephalitis virus

Japanese encephalitis virus (JEV), an RNA *Flavivirus* of which there are five genotypes, is transmitted in an enzootic cycle between *Culex* mosquitoes, animals such as wading birds and pigs, and humans with the latter acting as dead-end hosts (74–82). The seroprevalence of JEV in pigs in Vietnam ranges from 60 to 100% depending on where the pigs were farmed and the age of the pig (83–85). Less than 1% of humans infected with JEV develop symptoms ranging from a mild flu-like illness to encephalitis (74, 86).

Japanese encephalitis (JE) has been a notifiable disease in Vietnam since 2017, based on the case definition for viral encephalitis of 'fever greater than 38°C and a change in mental status, seizures, abnormal movements, tremor or spastic paralysis'. Case confirmation is based on the presence of anti-IgM antibodies in the CSF (87). While surveillance is largely passive, cases are also detected through sentinel site surveillance for meningoencephalitis in children aged up to 15 years. The incidence of JE is highest in the northern region of the country, where peaks are seen in the summer months of May–July. In the southern, central and highland regions, the incidence is relatively constant throughout the year (87, 88).

JE is seen most commonly in children. However, it may also be seen in adults as evidenced in studies in Vietnam (6, 9, 12). Cases in adults are more likely to be seen where there is no pre-existing immunity for example, in areas which have only recently experienced epidemics (87).

Vaccination against JE in Vietnam commenced in 1997, starting in high-risk provinces before expanding to all by 2014. Two doses of an inactivated mouse brain-derived vaccine are given to children aged 1-5 years, 1-2 weeks apart followed by a booster one year later (87). As a result of the vaccination programme, the national proportion of cases of AES which are due to JE has reduced (87, 89). However, the case incidence of JE still remains high in some provinces, particularly those in the northwest (90). As a result of this Vietnam conducts regular vaccination catch-up campaigns. However, it was recommended in the WHO 2006 position paper on Japanese Encephalitis Vaccines that the mouse brain-derived vaccines should gradually be replaced with the new generation JE vaccines which have a better safety profile (91). Japan is an example of the successful control of JE due to human and porcine vaccination, improved living conditions and vector control (92, 93). In Japan, more than 1,000 cases were reported per year prior to 1960 (94). However, since 2000, less than 10 cases have been reported per year with the exception of 2016 when 11 cases were reported (95).

Dengue virus

Dengue virus (DENV) is a *Flavivirus* with all four serotypes found in Vietnam (96, 97). Transmission occurs primarily via the bite of an infected *Aedes aegypti* and less commonly an *Aedes albopictus* mosquito (98). In southern and central Vietnam, which report the majority of cases, transmission occurs throughout the year with a peak during the rainy season from July to September. Most cases in northern Vietnam occur in the city of Hanoi during

10.3389/fpubh.2024.1396915

the autumn months, with few cases seen during the winter. This is despite the ability of the mosquitoes to survive the cooler temperatures by residing in concrete tanks with broken lids (99). Dengue is a notifiable disease in Vietnam however, it is likely that there is significant under-reporting as many cases are not hospitalised (98). Approximately every 10 years there were large outbreaks including in 1987 (100, 101), 1998 (100, 102), 2009 (100, 103) 2017 (104). However, more recently, the time between outbreaks reduced with one occurring in 2022 (138). Some outbreaks have been reported to coincide with increased activity of the El Niño and La Niña weather patterns (103).

Of those infected with DENV who develop symptoms, most have a mild illness including headache, fever, myalgia and rash however, secondary infection with a heterologous serotype can increase the risk of severe dengue which can result in haemodynamic shock and multi-organ failure (96, 105). Some of those infected may develop CNS manifestations including meningitis, encephalitis or meningoencephalitis (106–108). DENV accounted for 2.0% (n = 50) of cases in adults and 1.5% (n = 30) of cases in children (Supplementary Tables S3, S4; Figure 1). In 2024, the Ministry of Health for Vietnam approved the tetravalent live attenuated Qdenga vaccine however, it is not part of the EPI (109).

Enterovirus

Enteroviruses, belonging to the family Picornavirdae, are transmitted via the faecal-oral and respiratory routes, can cause a range of clinical manifestations from a mild flu-like illness to hand, foot and mouth disease (HFMD) with oral ulcers and a rash on the hands, feet and buttocks (89, 110) and meningitis or encephalitis if the virus crosses the blood-brain barrier (111, 112). The genotype enterovirus A71 (EV-A71) has a particular ability to result in severe neurological manifestations (112) however, it was not isolated in Vietnam until 2003 (89). Enteroviruses accounted for a slightly higher proportion of cases of CNS infections in children (3.6%, n = 71) compared to adults (2.0%, *n* = 50; Supplementary Tables S3, S4; Figure 1). Monovalent vaccines based on the EV-A71 sub-genotype C4 are licenced in China with vaccine effectiveness against HFMD caused by EV-A71 reported to be 63.4% for complete vaccination (113). However, these are not available in Vietnam.

Herpes simplex virus

Herpes simplex virus (HSV) belongs to the family *Herpesviridae* (111). HSV type 1 (HSV-1) is transmitted via oral contact and HSV type 2 (HSV-2), via sexual contact. Most people who are infected with HSV-1 or HSV-2 have mild or no symptoms. However, in some, HSV type 1 (HSV-1) causes oral or genital blisters/ulcers ('cold sores') whereas HSV-2 causes genital blisters/ulcers (114). Herpes Simplex Encephalitis (HSE) is most commonly caused by the reactivation of latent herpes simplex virus type 1 (HSV-1). Meningitis, is however, more likely to be caused by herpes simplex virus type 2 (HSV-2) (111). HSE is the most common cause of viral encephalitis in adults in countries in Europe and North America (115, 116) however, vaccines

against both HSV-1 and HSV-2 remain in development (117). Although it was also the most common viral aetiology among adults in Vietnam, the proportion of cases was still relatively low at 3.0% (n = 75; Supplementary Tables S3, S4; Figure 1).

Parasites and fungi

Compared to countries in other parts of the world, particularly those in sub-Saharan Africa, the prevalence of HIV in Vietnam is low. The 2023 estimate of HIV among adults aged 15 to 49 years was 0.3% (118). Due to this, the proportion of cases of CNS infections caused by parasites and fungi which are most often associated with immunosuppression, is much lower than in areas with high HIV prevalence. However, data from 2020 estimated that in Vietnam, 3.1% of those who were newly diagnosed with acquired-immunodeficiency syndrome (AIDS) or were living with HIV and drug resistance to antiretroviral therapy, were positive for cryptococcal antigen and of these, 70% developed into cryptococcal meningitis (119). Additionally, of the studies in Supplementary Table S1, Cryptococcus neoformans accounted for 0.5% (n = 13) of cases of CNS infections among adults (Supplementary Tables S3, S4; Figure 1). C. neoformans is a yeast which is found in the environment including soil and avian guano. Those who are immunosuppressed, particularly those living with HIV who have a low CD4 count, are at risk of cryptococcosis, with the most common presentation being cryptococcal meningitis (120).

Similarly, malaria incidence in Vietnam is low. In 2022, there were 455 cases of malaria (approximately 0.47 cases per 100,000 population) with the country aiming to eliminate all species by 2030 (121). As a result of this, unlike in for example, sub-Saharan Africa, cases of CNS infection due to malaria are rarely reported in Vietnam. In the study of childhood encephalitis by Pommier et al., *Plasmodium falciparum* accounted for less than 1% of causes of CNS infections (14).

Although not reported in the studies in Supplementary Tables S1–S4, neurocysticercosis is a known parasitic cause of CNS infections in Vietnam however, diagnosis is based on magnetic resonance imaging (MRI). In a study of the safety and efficacy of praziquantel in the treatment of patients with neurocysticercosis in Hanoi between 2017 and 2020, 104 patients were diagnosed with the condition (122). Similarly, other parasitic causes of CNS infections which may have been missed in the studies may include *Angiostrongylus cantonensis* which accounted for 67.3% of 55 cases of eosinophilic meningitis between 2008 and 2014 in Ho Chi Minh City (123).

Autoimmune

Autoimmune causes of encephalitis are associated with a variety of autoantibodies against the neuronal cell surface or synapse with the common being the anti-N-methyl-_D-aspartic acid (anti-NDMA) receptor antibody and the anti-leucine rich, glioma-inactivated 1 [anti-LGI1; voltage-gated potassium channel (VGKC) antibody] (124). In a study conducted in the Hospital for Tropical Diseases, Ho Chi Minh City over a period

of 18 months between 2015 and 2016, of the 99 patients admitted with encephalitis, nine tested positive for anti-NMDA receptor encephalitis (125). Two case reported of patients treated in Hanoi, have also been published (126) and, in a recent study of the aetiology of encephalitis in children in Hanoi, Pommier et al., reported six cases of anti-NDMA receptor encephalitis (14). A rare case of alpha-amino-3-hydroxy-5-methyl-4-isoxazole propionic acid (AMPA) receptor autoimmune encephalitis was also recently described in Ho Chi Minh City (127).

Lychee toxicity

In both Vietnam and India, it was noticed that outbreaks of encephalitis coincided with the lychee harvesting season. A retrospective analysis conducted between of 239 paediatric patients with encephalitis from Bac Giang province, northern Vietnam, between 2004 and 2009, found an independent association between the surface proportion of litchi plantation and the incidence of acute encephalitis (128). A case-control study conducted in northern India in 2014, found that among 104 cases and the same number of controls, those who had eaten lychees were at higher odds of having an encephalopathy compared to those who had not (odds ratio 7.8 (95%CI 3.6-24). The encephalopathy was thought to be due to the effect of the hypoglycin A or methylenecyclopropylglycine (MCPG) toxins (129). Following this publication, it was reported that 247 cases of CNS disease occurring between 1960 and 2017 in China, had developed following the consumption of lychees (130) however, there have been no known further reports in Vietnam.

Unknown aetiologies

In all studies of the aetiology of CNS infections in Vietnam, the cause continues to remain unknown in a large proportion of cases despite extensive diagnostics (11–15). This is not unique to Vietnam and, globally, unknown aetiologies account for a large proportion of CNS infections (131). However, those for which diagnostics are more challenging such as *Orientia tsutsugamushi*, or require imaging such as neurocysticercosis, may be under-reported, particularly the latter where the seroprevalence of *Taenia solium* is relatively high in at-risk populations (132). Other helminths such as *Angiostrongylus cantonensis* should be considered in those with a CSF eosinophilic pleocytosis and accounting for as many as 3% of cases of CNS infections (123, 133); and in returning travellers with gram negative meningitis, chronic infection with Strongyloides should be considered (134).

Following analysis of the spatio-temporal distribution of cases of acute encephalitis syndrome in Vietnam, which showed that the highest incidence was in the summer months in the northern provinces and a positive association with temperature, it was hypothesised that a number of cases were due to vector-borne diseases (135). Therefore, surveillance for vector-borne diseases, which are not necessarily routinely tested in patients with CNS infections, but for which Vietnam hosts competent vectors, such as chikungunya virus (CHIKV) and Zika virus (ZIKV) could be improved. Nipah virus (NiV), although never detected in human cases in Vietnam, may be considered in the differential diagnosis in any future outbreaks of encephalitis. NiV has been detected by reverse transcriptase polymerase chain reaction (RT-PCR) in urine samples of *Pteropus lylei* bats in Cambodia (136) and nearly half of the serum samples obtained from *Rousettus leschenaulti* bats in Hoa Binh province, Vietnam were positive for anti-NiV IgG by enzyme-linked immunosorbent assay (ELISA) (137).

Conclusion

While the heterogeneity in testing strategies in research studies makes comparisons and trends difficult to assess and care should be taken in future studies employ comprehensive testing to provide comparable data, the majority of cases of CNS infections in Vietnam were caused by S. suis and JEV. We would therefore recommend continuing to provide risk communication messaging around the consumption and preparation of raw pig products as well as improving awareness of S. suis among healthcare workers, particularly where bacterial cultures may be negative due to the prior use of antibiotics. Surveillance for JE should be continued with catch-up vaccination campaigns where required, while vaccination should be considered for S. pneumoniae and N. meningitis. Despite the challenges in determining the unknown aetiologies of many CNS infections, we would recommend considering pathogens such as Orientia tsutsugamushi as well as arboviruses such as CHIKV or ZIKV in the differential diagnosis.

Author contributions

HB: Conceptualization, Data curation, Funding acquisition, Methodology, Writing – original draft, Writing – review & editing. MC: Conceptualization, Supervision, Writing – review & editing. RC: Supervision, Writing – review & editing. NF: Supervision, Writing – review & editing. MG: Supervision, Writing – review & editing. PT: Writing – review & editing. HD: Conceptualization, Supervision, Writing – review & editing. BN: Conceptualization, Supervision, Writing – review & editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. HB was funded by a Wellcome Trust Clinical PhD Programme (grant number: 102465). For the purpose of Open Access, the author has applied a CC BY public copyright licence to any Author Accepted Manuscript version arising from this submission.

Acknowledgments

The original literature review of central nervous system infections in Vietnam was published in the PhD thesis by Hannah Brindle 'The Epidemiology of and Risk Factors for Central Nervous System Infections in Vietnam' (https://livrepository.liverpool.ac.uk/3110575/).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

1. Feigin VL, Nichols E, Alam T, Bannick MS, Beghi E, Blake N, et al. Global, regional, and national burden of neurological disorders, 1990–2016: a systematic analysis for the global burden of disease study 2016. *Lancet Neurol.* (2019) 18:459–80. doi: 10.1016/S1474-4422(18)30499-X

2. Coker RJ, Hunter BM, Rudge JW, Liverani M, Hanvoravongchai P. Emerging infectious diseases in Southeast Asia: regional challenges to control. *Lancet.* (2011) 377:599–609. doi: 10.1016/S0140-6736(10)62004-1

3. Tarantola A, Goutard F, Newton P, De LX, Lortholary O, Cappelle J, et al. Estimating the burden of Japanese encephalitis virus and other Encephalitides in countries of the Mekong region. *PLoS Negl Trop Dis.* (2014) 8:e2533. doi: 10.1371/journal.pntd.0002533

4. Tran TT, Le QT, Tran TN, Nguyen NT, Pedersen FK, Schlumberger M. The etiology of bacterial pneumonia and meningitis in Vietnam. *Pediatr Infect Dis J*. (1998) 17:S192–4. doi: 10.1097/00006454-199809001-00023

5. Nguyen THM, Tran THC, Thwaites G, Ly VC, Dinh XS, Ho Dang TN, et al. Dexamethasone in Vietnamese adolescents and adults with bacterial meningitis. *N Engl J Med.* (2007) 357:2431-40. doi: 10.1056/NEJMoa070852

6. Tan LV, Thai LH, Phu NH, Nghia HDT, Chuong LV, Sinh DX, et al. Viral Aetiology of central nervous system infections in adults admitted to a tertiary referral Hospital in Southern Vietnam over 12 years. *PLoS Negl Trop Dis.* (2014) 8:e3127. doi: 10.1371/journal.pntd.0003127

7. Tan LV, Qui PT, Ha DQ, Hue NB, Bao LQ, Cam BV, et al. Viral etiology of encephalitis in children in southern Vietnam: results of a one-year prospective descriptive study. *PLoS Negl Trop Dis.* (2010) 4:e854. doi: 10.1371/journal.pntd.0000854

8. Taylor WR, Nguyen K, Nguyen D, Nguyen H, Horby P, Nguyen HL, et al. The Spectrum of central nervous system infections in an adult referral Hospital in Hanoi. *Vietnam PLOS ONE.* (2012) 7:e42099. doi: 10.1371/journal.pone.0042099

9. Ho Dang Trung N, Le Thi PT, Wolbers M, Nguyen Van Minh H, Nguyen Thanh V, Van MP, et al. Aetiologies of central nervous system infection in Viet Nam: A prospective provincial hospital-based descriptive surveillance study. *PLoS One.* (2012) 7. doi: 10.1371/journal.pone.0037825

10. An PN, Huong TTT. Etiological structure of acute encephalitis in children and immunization program concerns at The National Hospital of PaediatricS in Vietnam. *Southeast Asian J Trop Med Public Health.* (2014) 45:37–9.

11. Ngo CC, Katoh S, Hasebe F, Dhoubhadel BG, Hiraoka T, Hamaguchi S, et al. Characteristics and biomarkers of patients with central nervous system infection admitted to a referral hospital in northern Vietnam. *Trop Med Health*. (2021) 49:42. doi: 10.1186/s41182-021-00322-2

12. Brindle HE, Nadjm B, Choisy M, Christley R, Griffiths M, Baker S, et al. Aetiology and potential animal exposure in central nervous system infections in Vietnam. *EcoHealth.* (2022). 19:463–474. doi: 10.1007/s10393-022-01618-3

13. Gabor JJ, Anh CX, Sy BT, Hoan PQ, Quyen DT, The NT, et al. Aetiologies and clinical presentation of central nervous system infections in Vietnamese patients: a prospective study. *Sci Rep.* (2022) 12:18065. doi: 10.1038/s41598-022-23007-0

 Pommier JD, Gorman C, Crabol Y, Bleakley K, Sothy H, Santy K, et al. Childhood encephalitis in the greater Mekong region (the SouthEast Asia encephalitis project): a multicentre prospective study. *Lancet Glob Health*. (2022) 10:e989–e1002. doi: 10.1016/ S2214-109X(22)00174-7

15. Pallerla SR, Van Dong D, Linh LTK, Van Son T, Quyen DT, Hoan PQ, et al. Diagnosis of pathogens causing bacterial meningitis using Nanopore sequencing in a resource-limited setting. *Ann Clin Microbiol Antimicrob.* (2022) 21:39. doi: 10.1186/s12941-022-00530-6

 Horby P, Wertheim H, Ha NH, Trung NV, Trinh DT, Taylor W, et al. Stimulating the development of national *Streptococcus suis* guidelines in Viet Nam through a strategic research partnership. *Bull World Health Organ.* (2010) 88:458–61. doi: 10.2471/BLT.09.067959

17. Hughes JM, Wilson ME, Wertheim HFL, Nghia HDT, Taylor W, Schultsz C. *Streptococcus suis*: An emerging human pathogen. *Clin Infect Dis.* (2009) 48:617–25. doi: 10.1086/596763

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpubh.2024.1396915/ full#supplementary-material

18. Huong VTL, Turner HC, Kinh NV, Thai PQ, Hoa NT, Horby P, et al. Burden of disease and economic impact of human *Streptococcus suis* infection in Viet Nam. *Trans R Soc Trop Med Hyg.* (2019) 113:341–50. doi: 10.1093/trstmh/trz004

19. Besung INK, Suarjana IGK, Agustina KK, Winaya IBO, Soeharsono H, Suwiti NK, et al. Isolation and identification of *Streptococcus suis* from sick pigs in Bali, Indonesia. *BMC Res Notes.* (2019) 12:795. doi: 10.1186/s13104-019-4826-7

20. Ferrando ML, Schultsz C. A hypothetical model of host-pathogen interaction of *Streptococcus suis* in the gastro-intestinal tract. *Gut Microbes*. (2016) 7:154–62. doi: 10.1080/19490976.2016.1144008

21. Nghia HDT, Ho DTN, Tu LTP, Le TPT, Wolbers M, Thai CQ, et al. Risk factors of *Streptococcus suis* infection in Vietnam. *A case-control study PloS One*. (2011) 6:e17604. doi: 10.1371/journal.pone.0017604

22. Huong VTL, Hoa NT, Horby P, Bryant JE, Van Kinh N, Toan TK, et al. Raw pig blood consumption and potential risk for *Streptococcus suis* infection. *Vietnam Emerg Infect Dis.* (2014) 20:1895–8. doi: 10.3201/eid2011.140915

23. Huong VTL, Thanh LV, Phu VD, Trinh DT, Inui K, Tung N, et al. Temporal and spatial association of *Streptococcus suis* infection in humans and porcine reproductive and respiratory syndrome outbreaks in pigs in northern Vietnam. *Epidemiol Infect.* (2016) 144:35–44. doi: 10.1017/S0950268815000990

24. Van SA, Brouwer MC, Schultsz C, Van Der EA, Van De BD. *Streptococcus suis* meningitis: A systematic review and Meta-analysis. *PLoS Negl Trop Dis.* (2015) 9:e0004191. doi: 10.1371/journal.pntd.0004191

25. Choisy M, Contamin L. Gdpm: data from the general Department of Preventive Medicine (GDPM) of Vietnam. R package version 0.1.0. (2019).

26. Wertheim HFL, Nguyen HN, Taylor W, Lien TTM, Ngo HT, Nguyen TQ, et al. *Streptococcus suis*, an important cause of adult bacterial meningitis in northern Vietnam. *PLoS One*. (2009) 4:e5973. doi: 10.1371/journal.pone.0005973

27. Huong VTL, Ha N, Huy NT, Horby P, Nghia HDT, Thiem VD, et al. Epidemiology, clinical manifestations, and outcomes of *Streptococcus suis* infection in humans. *Emerg Infect Dis.* (2014) 20:1105–14. doi: 10.3201/eid2007.131594

28. Liang S, Zhang S, Bao Y, Zhang Y, Liu X, Yao H, et al. Combined Immunoinformatics to design and evaluate a multi-epitope vaccine candidate against *Streptococcus suis* infection. *Vaccine*. (2024) 12:137. doi: 10.3390/vaccines12020137

29. Jeffery A, Gilbert M, Corsaut L, Gaudreau A, Obradovic MR, Cloutier S, et al. Immune response induced by a *Streptococcus suis* multi-serotype autogenous vaccine used in sows to protect post-weaned piglets. *Vet Res.* (2024) 55:57. doi: 10.1186/s13567-024-01313-x

30. Mai NTH, Hoa NT, Nga TVT, Linh LD, Chau TTH, Sinh DX, et al. *Streptococcus suis* meningitis in adults in Vietnam. *Clin Infect Dis Off Publ Infect Dis Soc Am.* (2008) 46:659–67. doi: 10.1086/527385

31. Huong VTL, Long HB, Kinh NV, Ngan TTD, Dung VTV, Nadjm B, et al. Longterm outcomes of patients with *Streptococcus suis* infection in Viet Nam: A case-control study. *J Inf Secur.* (2018) 76:159–67. doi: 10.1016/j.jinf.2017.09.019

32. Kadioglu A, Weiser JN, Paton JC, Andrew PW. The role of *Streptococcus pneumoniae* virulence factors in host respiratory colonization and disease. *Nat Rev Microbiol.* (2008) 6:288–301. doi: 10.1038/nrmicro1871

33. Bogaert D, De Groot R, Hermans PWM. Streptococcus pneumoniae colonisation: the key to pneumococcal disease. Lancet Infect Dis. (2004) 4:144–54. doi: 10.1016/S1473-3099(04)00938-7

34. McGill F, Heyderman RS, Panagiotou S, Tunkel AR, Solomon T. Acute bacterial meningitis in adults. *Lancet*. (2016) 388:3036–47. doi: 10.1016/S0140-6736(16)30654-7

35. Le Polain De Waroux O, Edmunds WJ, Takahashi K, Ariyoshi K, Mulholland EK, Goldblatt D, et al. Predicting the impact of pneumococcal conjugate vaccine programme options in Vietnam. *Hum Vaccin Immunother*. (2018) 14:1939–47. doi: 10.1080/21645515.2018.1467201

36. Licciardi PV, Temple B, Dai VTT, Toan NT, Uyen D, Nguyen CD, et al. Immunogenicity of alternative ten-valent pneumococcal conjugate vaccine schedules in infants in Ho Chi Minh City, Vietnam: results from a single-blind, parallel-group, openlabel, randomised, controlled trial. *Lancet Infect Dis.* (2021) 21:1415–28. doi: 10.1016/ S1473-3099(20)30775-1

37. Pai M, Behr MA, Dowdy D, Dheda K, Divangahi M, Boehme CC, et al. Tuberculosis. *Nat Rev Dis Primers*. (2016) 2:16076. doi: 10.1038/nrdp.2016.76

38. Thwaites GE, Van TR, Schoeman J. Tuberculous meningitis: more questions, still too few answers. *Lancet Neurol.* (2013) 12:999–1010. doi: 10.1016/S1474-4422(13)70168-6

39. Bang ND, Caws M, Truc TT, Duong TN, Dung NH, Ha DTM, et al. Clinical presentations, diagnosis, mortality and prognostic markers of tuberculous meningitis in Vietnamese children: a prospective descriptive study. *BMC Infect Dis.* (2016) 16:573. doi: 10.1186/s12879-016-1923-2

40. Thwaites GE, Chau TTH, Farrar JJ. Improving the bacteriological diagnosis of tuberculous meningitis. *J Clin Microbiol.* (2004) 42:378–9. doi: 10.1128/ JCM.42.1.378-379.2004

41. Bahr NC, Nuwagira E, Evans EE, Cresswell FV, Bystrom PV, Byamukama A, et al. Diagnostic accuracy of Xpert MTB/RIF ultra for tuberculous meningitis in HIV-infected adults: a prospective cohort study. *Lancet Infect Dis.* (2018) 18:68–75. doi: 10.1016/S1473-3099(17)30474-7

42. Donovan J, Cresswell FV, Thuong NTT, Boulware DR, Thwaites GE, Bahr NC. Xpert MTB/RIF ultra for the diagnosis of tuberculous meningitis: A small step forward. *Clin Infect Dis Off Publ Infect Dis Soc Am*. (2020) 71:2002–5. doi: 10.1093/cid/ciaa473

43. WHO. World Health Organization. GHO | by category | BCG - immunization coverage estimates by country. (2023). Available from: https://apps.who.int/gho/data/ view.main.80500?lang=en (Accessed 11 December 2024).

44. Thwaites G, Chau T, Stepniewska K, Phu N, Chuong L, Sinh D, et al. Diagnosis of adult tuberculous meningitis by use of clinical and laboratory features. *Lancet.* (2002) 360:1287–92. doi: 10.1016/S0140-6736(02)11318-3

45. Thwaites GE, Nguyen DB, Nguyen HD, Hoang TQ, Do TTO, Nguyen TCT, et al. Dexamethasone for the treatment of tuberculous meningitis in adolescents and adults. *N Engl J Med.* (2004) 351:1741–51. doi: 10.1056/NEJMoa040573

46. Donovan J, Bang ND, Imran D, Nghia HDT, Burhan E, Huong DTT, et al. Adjunctive dexamethasone for tuberculous meningitis in HIV-positive adults. *N Engl J Med.* (2023) 389:1357–67. doi: 10.1056/NEJMoa2216218

47. World Health Organization. *Haemophilus influenzae*: vaccine preventable diseases surveillance standards. (2018). Available from: https://www.who.int/publications/m/ item/vaccine-preventable-diseases-surveillance-standards-haemophilus-influenzae (Accessed 11 December 2024).

48. World Health Organization. *Haemophilus influenzae* type b (Hib). (2023) Available at: https://www.who.int/teams/health-product-policy-and-standards/ standards-and-specifications/norms-and-standards/vaccine-standardization/hib (Accessed 11 December 2024).

49. Wunrow HY, Bender RG, Vongpradith A, Sirota SB, Swetschinski LR, Novotney A, et al. Global, regional, and national burden of meningitis and its aetiologies, 1990–2019: a systematic analysis for the global burden of disease study 2019. *Lancet Neurol.* (2023) 22:685–711. doi: 10.1016/S1474-4422(23)00195-3

50. Rosenstein NE, Perkins BA, Stephens DS, Popovic T, Hughes JM. Meningococcal disease. N Engl J Med. (2001) 344:1378–88. doi: 10.1056/NEJM200105033441807

51. World Health Organization. Meningococcal meningitis. (2023). Available from: https://www.who.int/teams/health-product-policy-and-standards/standards-and-specifications/norms-and-standards/vaccine-standardization/meningococcal-meningitis (Accessed 11 December 2024).

52. World Health Organization. Meningococcus: vaccine preventable diseases surveillance standards. (2023). Available from: https://www.who.int/publications/m/ item/vaccine-preventable-diseases-surveillance-standards-meningococcus (Accessed 11 December 2024).

53. Phan TV, Vo DTT, Nguyen HTK, Ho TNL, Pham QD, Luong QC, et al. Characterizing *Neisseria meningitidis* in southern Vietnam between 2012 and 2021: A predominance of the chloramphenicol-resistant ST-1576 lineage. *IJID Reg.* (2024) 10:52–9. doi: 10.1016/j.ijregi.2023.11.013

54. Thisyakorn U, Carlos J, Chotpitayasunondh T, Dien TM, Gonzales MLAM, Huong NTL, et al. Invasive meningococcal disease in Malaysia, Philippines, Thailand, and Vietnam: An Asia-Pacific expert group perspective on current epidemiology and vaccination policies. *Hum Vaccin Immunother*. (2022) 18:2110759. doi: 10.1080/21645515.2022.2110759

55. Nguyen PNT, Thai PQ, Dien TM, Hai DT, Dai VTT, Luan NH, et al. 4CMenB vaccination to prevent meningococcal B disease in Vietnam: expert review and opinion. *Infect Dis Ther*. (2024) 13:423–37. doi: 10.1007/s40121-023-00905-y

56. Paris DH, Shelite TR, Day NP, Walker DH. Unresolved problems related to scrub typhus: A seriously neglected life-threatening disease. *Am J Trop Med Hyg.* (2013) 89:301–7. doi: 10.4269/ajtmh.13-0064

57. Xu G, Walker DH, Jupiter D, Melby PC, Arcari CM. A review of the global epidemiology of scrub typhus. *PLoS Negl Trop Dis.* (2017) 11:e0006062. doi: 10.1371/journal.pntd.0006062

58. Traub R, Wisseman CL. The ecology of chigger-borne rickettsiosis (scrub typhus). J Med Entomol. (1974) 11:237–303. doi: 10.1093/jmedent/11.3.237

59. Seong SY, Choi MS, Kim IS. Orientia tsutsuganushi infection: overview and immune responses. Microbes Infect. (2001) 3:11–21. doi: 10.1016/S1286-4579(00)01352-6

60. Nadjm B, Thuy PT, Trang VD, Dang Ha L, Kinh NV, Wertheim HF. Scrub typhus in the northern provinces of Vietnam: an observational study of admissions to a national referral hospital. *Trans R Soc Trop Med Hyg.* (2014) 108:739–40. doi: 10.1093/trstmh/tru145

61. Vallée J, Thaojaikong T, Moore CE, Phetsouvanh R, Richards AL, Souris M, et al. Contrasting spatial distribution and risk factors for past infection with scrub typhus and murine typhus in Vientiane City, Lao PDR. *PLoS Negl Trop Dis.* (2010) 4:e909. doi: 10.1371/journal.pntd.0000909

62. Viswanathan S, Muthu V, Iqbal N, Remalayam B, George T. Scrub typhus meningitis in South India — A retrospective study. *PLoS One*. (2013) 8:e66595. doi: 10.1371/journal.pone.0066595

63. Dittrich S, Rattanavong S, Lee SJ, Panyanivong P, Craig SB, Tulsiani SM, et al. Orientia, rickettsia, and leptospira pathogens as causes of CNS infections in Laos: a prospective study. *Lancet Glob Health*. (2015) 3:e104–12. doi: 10.1016/S2214-109X(14)70289-X

64. Jain P, Prakash S, Tripathi PK, Chauhan A, Gupta S, Sharma U, et al. Emergence of *Orientia tsutsugamushi* as an important cause of acute encephalitis syndrome in India. *PLoS Negl Trop Dis.* (2018) 12:e0006346. doi: 10.1371/journal.pntd.0006346

65. Azad AF. Epidemiology of murine typhus. Annu Rev Entomol. (1990) 35:553–70. doi: 10.1146/annurev.en.35.010190.003005

66. Gasem MH, Wagenaar JFP, Goris MGA, Adi MS, Isbandrio BB, Hartskeerl RA, et al. Murine typhus and leptospirosis as causes of acute undifferentiated fever. *Indonesia Emerg Infect Dis.* (2009) 15:975–7. doi: 10.3201/eid1506.081405

67. Hamaguchi S, Cuong NC, Tra DT, Doan YH, Shimizu K, Tuan NQ, et al. Clinical and epidemiological characteristics of scrub typhus and murine typhus among hospitalized patients with acute undifferentiated fever in northern Vietnam. *Am J Trop Med Hyg.* (2015) 92:972–8. doi: 10.4269/ajtmh.14-0806

68. Doppler JF, Newton PN. A systematic review of the untreated mortality of murine typhus. *PLoS Negl Trop Dis.* (2020) 14:e0008641. doi: 10.1371/journal.pntd.0008641

69. Adler B, de la Peña MA. Leptospira and leptospirosis. Vet Microbiol. (2010) 140:287-96. doi: 10.1016/j.vetmic.2009.03.012

70. Costa F, Hagan JE, Calcagno J, Kane M, Torgerson P, Martinez-Silveira MS, et al. Global morbidity and mortality of leptospirosis: A systematic review. *PLoS Negl Trop Dis.* (2015) 9:e0003898. doi: 10.1371/journal.pntd.0003898

71. Romero EC, Blanco RM, Yasuda PH. Aseptic meningitis caused by Leptospira spp diagnosed by polymerase chain reaction. *Mem Inst Oswaldo Cruz*. (2010) 105:988–92. doi: 10.1590/S0074-02762010000800007

72. Pappachan J, Mathew S, Kp A, Khader A, Bharghavan P, Kareem M, et al. Risk factors for mortality in patients with leptospirosis during an epidemic in northern Kerala. *Natl Med J India*. (2003) 17:e0006346:240–2.

73. Warnasekara J, Srimantha S, Kappagoda C, Jayasundara D, Senevirathna I, Matthias M, et al. Diagnostic method-based underestimation of leptospirosis in clinical and research settings; an experience from a large prospective study in a high endemic setting. *PLoS Negl Trop Dis.* (2022) 16:e0010331. doi: 10.1371/journal.pntd.0010331

74. Solomon T, Dung NM, Kneen R, Gainsborough M, Vaughn DW, Khanh VT. Japanese encephalitis. *J Neurol Neurosurg Psychiatry*. (2000) 68:405–15. doi: 10.1136/jnnp.68.4.405

75. Scherer WF, Buescher EL, Flemings MB, Noguchi A, Scanlon J. Ecologic studies of Japanese encephalitis virus in Japan. III. Mosquito factors. Zootropism and vertical flight of Culex tritaeniorhynchus with observations on variations in collections from animal-baited traps in different habitats. *Am J Trop Med Hyg.* (1959) 8:665–77. doi: 10.4269/ajtmh.1959.8.665

76. Buescher EL, Scherer WF, McCLURE HE, Moyer JT, Rosenberg MZ, Yoshii M, et al. Ecologic studies of Japanese encephalitis virus in Japan. IV. Avian infection. *Am J Trop Med Hyg.* (1959) 8:678–88. doi: 10.4269/ajtmh.1959.8.678

77. Scherer WF, Moyer JT, Izumi T, Gresser I. Ecologic studies of Japanese encephalitis virus in Japan. VI. Swine infection. *Am J Trop Med Hyg.* (1959) 8:698–706. doi: 10.4269/ ajtmh.1959.8.698

78. Longbottom J, Browne AJ, Pigott DM, Sinka ME, Golding N, Hay SI, et al. Mapping the spatial distribution of the Japanese encephalitis vector, Culex tritaeniorhynchus Giles, 1901 (Diptera: Culicidae) within areas of Japanese encephalitis risk. *Parasit Vectors*. (2017) 10:148. doi: 10.1186/s13071-017-2086-8

79. Keiser J, Maltese MF, Erlanger TE, Bos R, Tanner M, Singer BH, et al. Effect of irrigated rice agriculture on Japanese encephalitis, including challenges and opportunities for integrated vector management. *Acta Trop.* (2005) 95:40–57. doi: 10.1016/j.actatropica.2005.04.012

80. Self LS, Shin HK, Kim KH, Lee KW, Chow CY, Hong HK. Ecological studies on Culex tritaeniorhynchus as a vector of Japanese encephalitis. *Bull World Health Organ.* (1973) 49:41–7.

81. Li MH, Fu SH, Chen WX, Wang HY, Guo YH, Liu QY, et al. Genotype V Japanese encephalitis virus is emerging. *PLoS Negl Trop Dis.* (2011) 5:e1231. doi: 10.1371/journal. pntd.0001231

82. Do LP, Bui TM, Hasebe F, Morita K, Phan NT. Molecular epidemiology of Japanese encephalitis in northern Vietnam, 1964–2011: genotype replacement. *Virol J.* (2015) 12:51. doi: 10.1186/s12985-015-0278-4

83. Lindahl J, Boqvist S, Ståhl K, Thu HTV, Magnusson U. Reproductive performance in sows in relation to Japanese encephalitis virus seropositivity in an endemic area. *Trop Anim Health Prod.* (2012) 44:239–45. doi: 10.1007/s11250-011-0005-0

84. Lindahl JF, Ståhl K, Chirico J, Boqvist S, Thu HTV, Magnusson U. Circulation of Japanese encephalitis virus in pigs and mosquito vectors within can Tho City. *Vietnam PLoS Negl Trop Dis.* (2013) 7:e2153. doi: 10.1371/journal.pntd.0002153

85. Ruget AS, Beck C, Gabassi A, Trevennec K, Lecollinet S, Chevalier V, et al. Japanese encephalitis circulation pattern in swine of northern Vietnam and consequences for swine's vaccination recommendations. *Transbound Emerg Dis.* (2018) 65:1485–92. doi: 10.1111/tbed.12885

86. Campbell GL, Hills SL, Fischer M, Jacobson JA, Hoke CH, Hombach JM, et al. Estimated global incidence of Japanese encephalitis: a systematic review. *Bull World Health Organ.* (2011) 89:766–74. doi: 10.2471/BLT.10.085233

87. Yen NT, Duffy MR, Hong NM, Hien NT, Fischer M, Hills SL. Surveillance for Japanese encephalitis in Vietnam, 1998–2007. *Am J Trop Med Hyg.* (2010) 83:816–9. doi: 10.4269/ajtmh.2010.10-0262

88. Cuc Y, Phòng Tế Dự, General Department of Preventive Medicine. (2021). Available from: https://vncdc.gov.vn/en (Accessed 11 December 2024).

89. Van Tu P, Thao NTT, Perera D, Truong KH, Tien NTK, Thuong TC, et al. Epidemiologic and Virologic investigation of hand, foot, and mouth disease, southern Vietnam, 2005. *Emerg Infect Dis.* (2007) 13:1733–41. doi: 10.3201/eid1311.070632

90. Choisy M, Contamin L. Gso: data from the general statistical office (GSO) of Vietnam. R package version 0.1.0. (2019).

91. World Health Organization. Japanese encephalitis vaccines: WHO position paper -February 2015. (2015). Available from: https://iris.who.int/bitstream/handle/10665/242325/ WER9009_69-88.PDF?sequence=1 (Accessed 11 December 2024).

92. Igarashi A. Control of Japanese encephalitis in Japan: immunization of humans and animals, and vector control. *Curr Top Microbiol Immunol.* (2002) 267:139–52. doi: 10.1007/978-3-642-59403-8_7

93. Erlanger TE, Weiss S, Keiser J, Utzinger J, Wiedenmayer K. Past, present, and future of Japanese encephalitis. *Emerg Infect Dis.* (2009) 15:1–7. doi: 10.3201/eid1501.080311

94. Kuwayama M, Ito M, Takao S, Shimazu Y, Fukuda S, Miyazaki K, et al. Japanese encephalitis virus in meningitis patients, Japan. *Emerg Infect Dis.* (2005) 11:471–3. doi: 10.3201/eid1103.040285

95. World Health Organization. Immunization data Japanese encephalitis (JE) reported cases and incidence. (2023). Available at: https://immunizationdata.who.int/global/wiise-detail-page/japanese-encephalitis-(je)-reported-cases-and-incidence?CODE=Global&YEAR=) (Accessed 11 December 2024).

96. Simmons CP, Farrar JJ, Nguyen van VC, Wills B. Dengue. N Engl J Med. (2012) 366:1423–32. doi: 10.1056/NEJMra1110265

97. Nguyen-Tien T, Lundkvist Å, Lindahl J. Urban transmission of mosquito-borne flaviviruses – a review of the risk for humans in Vietnam. *Infect Ecol Epidemiol.* (2019) 9:1660129. doi: 10.1080/20008686.2019.1660129

98. Colón-González FJ, Bastos LS, Hofmann B, Hopkin A, Harpham Q, Crocker T, et al. Probabilistic seasonal dengue forecasting in Vietnam: A modelling study using superensembles. *PLoS Med.* (2021) 18:e1003542. doi: 10.1371/journal.pmed.1003542

99. Tsunoda T, Cuong TC, Dong TD, Yen NT, Le NH PTV, et al. Winter refuge for Aedes aegypti and ae. Albopictus mosquitoes in Hanoi during winter. *PLoS One*. (2014) 9:e95606. doi: 10.1371/journal.pone.0095606

100. Lee HS, Nguyen-Viet H, Nam VS, Lee M, Won S, Duc PP, et al. Seasonal patterns of dengue fever and associated climate factors in 4 provinces in Vietnam from 1994 to 2013. *BMC Infect Dis.* (2017) 17:218. doi: 10.1186/s12879-017-2326-8

101. Tran HP, Adams J, Jeffery JAL, Nguyen YT, Vu NS, Kutcher SC, et al. Householder perspectives and preferences on water storage and use, with reference to dengue, in the Mekong Delta, southern Vietnam. *Int Health.* (2010) 2:136–42. doi: 10.1016/j. inhe.2009.12.007

102. Ha DQ, Tien NT, Huong VT, Loan HT, Thang CM. Dengue epidemic in southern Vietnam, 1998. *Emerg Infect Dis.* (2000) 6:422–5. doi: 10.3201/eid0604.000421

103. Minh An DT, Rocklöv J. Epidemiology of dengue fever in Hanoi from 2002 to 2010 and its meteorological determinants. *Glob Health Action*. (2014) 7:23074. doi: 10.3402/gha.v7.23074

104. Sun G, Trung NV, Matsui T, Ishibashi K, Kirimoto T, Furukawa H, et al. Field evaluation of an infectious disease/fever screening radar system during the 2017 dengue fever outbreak in Hanoi, Vietnam: a preliminary report. *J Inf Secur.* (2017) 75:593–5. doi: 10.1016/j.jinf.2017.10.005

105. World Health Organization. Dengue and severe dengue. (2024). Available from: https://www.who.int/news-room/fact-sheets/detail/dengue-and-severe-dengue (Accessed 11 December 2024).

106. Carod-Artal FJ, Wichmann O, Farrar J, Gascón J. Neurological complications of dengue virus infection. *Lancet Neurol.* (2013) 12:906–19. doi: 10.1016/S1474-4422(13)70150-9

107. Araújo F, Nogueira R, Araújo MS, Perdigão A, Cavalcanti L, Brilhante R, et al. Dengue in patients with central nervous system manifestations, Brazil. *Emerg Infect Dis.* (2012) 18:677–9. doi: 10.3201/eid1804.111522

108. Domingues RB, Kuster GW, Onuki-Castro FL, Souza VA, Levi JE, Pannuti CS. Involvement of the central nervous system in patients with dengue virus infection. *J Neurol Sci.* (2008) 267:36–40. doi: 10.1016/j.jns.2007.09.040

109. vietnamnews.vn. Vaccine provides immunity against dengue virus. (2024). Available at: https://vietnamnews.vn/life-style/1666237/vaccine-provides-immunityagainst-dengue-virus.html (Accessed 11 December 2024).

110. World Health Organization. Enterovirus 71. Available at: https://www.who.int/ teams/health-product-policy-and-standards/standards-and-specifications/norms-andstandards/vaccine-standardization/enterovirus-71

111. Ludlow M, Kortekaas J, Herden C, Hoffmann B, Tappe D, Trebst C, et al. Neurotropic virus infections as the cause of immediate and delayed neuropathology. *Acta Neuropathol (Berl)*. (2016) 131:159–84. doi: 10.1007/s00401-015-1511-3

112. Chen BS, Lee HC, Lee KM, Gong YN, Shih SR. Enterovirus and encephalitis Front Microbiol. (2020) 11:261. doi: 10.3389/fmicb.2020.00261

113. Duan X, Zhang L, Ding L, Zhang C, Chen Z, Cheng Y, et al. Effectiveness of enterovirus A71 vaccine against pediatric HFMD and disease profile of post-vaccination infection. *Vaccine*. (2024) 42:2317–25. doi: 10.1016/j. vaccine.2024.02.026

114. World Health Organization. Herpes simplex virus. (2024). Available from: https://www.who.int/news-room/fact-sheets/detail/herpes-simplex-virus

115. Granerod J, Ambrose HE, Davies NW, Clewley JP, Walsh AL, Morgan D, et al. Causes of encephalitis and differences in their clinical presentations in England: a multicentre, population-based prospective study. *Lancet Infect Dis.* (2010) 10:835–44. doi: 10.1016/S1473-3099(10)70222-X

116. Glaser CA, Honarmand S, Anderson LJ, Schnurr DP, Forghani B, Cossen CK, et al. Beyond viruses: clinical profiles and etiologies associated with encephalitis. *Clin Infect Dis.* (2006) 43:1565–77. doi: 10.1086/509330

117. Krishnan R, Stuart PM. Developments in vaccination for herpes simplex virus. *Front Microbiol.* (2021) 12:798927. doi: 10.3389/fmicb.2021.798927

118. UNAIDS. Country factsheets Viet Nam. (2023). Available from: https://www. unaids.org/en/regionscountries/countries/vietnam (Accessed 11 December 2024).

119. Duong TMN, Le MH, Beardsley J, Denning DW, Le NH, Nguyen BNT. Updated estimation of the burden of fungal disease in Vietnam. *Mycoses*. (2023) 66:346–53. doi: 10.1111/myc.13559

120. Sloan DJ, Parris V. Cryptococcal meningitis: epidemiology and therapeutic options. *Clin Epidemiol.* (2014) 6:169–82. doi: 10.2147/CLEP.S38850

121. Nguyen XT, Win HOo, Win H, Ngo TVA, Ngo TT, Dinh SH, et al. Fowkes - Facilitators, barriers and acceptability of malaria reactive surveillance and response strategies in Vietnam: a mixed-methods study: BMJ Public Health. (2024) 2:e000961.

122. Thang TD, Phuc TQ, Thanh DT, Thieu NQ, Tho TH, Loi CB, et al. Efficacy and safety of praziquantel in the treatment of neurocysticercosis in Vietnam. *Trop Biomed.* (2022) 39:462–6. doi: 10.47665/tb.39.3.018

123. McBride A, Chau TTH, Hong NTT, Mai NTH, Anh NT, Thanh TT, et al. *Angiostrongylus cantonensis* is an important cause of eosinophilic meningitis in southern Vietnam. *Clin Infect Dis Off Publ Infect Dis Soc Am*. (2017) 64:1784–7. doi: 10.1093/ cid/cix118

124. Newman MP, Blum S, Wong RCW, Scott JG, Prain K, Wilson RJ, et al. Autoimmune encephalitis. Intern Med J. (2016) 46:148-57. doi: 10.1111/imj.12974

125. Nguyen Thi Hoang M, Nguyen Hoan P, Le Van T, McBride A, Ho Dang Trung N, Tran Tan T, et al. First reported cases of anti-NMDA receptor encephalitis in Vietnamese adolescents and adults. *J Neurol Sci.* (2017) 373:250–3. doi: 10.1016/j. jns.2017.01.004

126. Tu NP, Nha PB, Hung ND, Minh NH, Anh HN, Dinh TC. Treatment of anti-NMDA receptor encephalitis with ovarian Teratoma removal: A literature review and two case reports. *Open Access Maced J Med Sci.* (2019) 7:4378–82. doi: 10.3889/ oamjms.2019.839

127. Trung Hieu NL, Minh Duc N, Tra My TT, Hieu Anh B, Tan Lien Bang M, Minh TP. First reported case of anti-Ampa receptor encephalitis in a Vietnamese adolescent. *Clin Med Insights Case Rep.* (2021) 14:11795476211037782. doi: 10.1177/11795476211037782

128. Paireau J, Tuan NH, Lefrançois R, Buckwalter MR, Nghia ND, Hien NT, et al. Litchi–associated acute encephalitis in children, northern Vietnam, 2004–2009. *Emerg Infect Dis.* (2012) 18:1817–24. doi: 10.3201/eid1811.111761

129. Shrivastava A, Kumar A, Thomas JD, Laserson KF, Bhushan G, Carter MD, et al. Association of acute toxic encephalopathy with litchi consumption in an outbreak in Muzaffarpur, India, 2014: a case-control study. *Lancet Glob Health*. (2017) 5:e458–66. doi: 10.1016/S2214-109X(17)30035-9

130. Zhang LJ, Fontaine RE. Lychee-associated encephalopathy in China and its reduction since 2000. *Lancet Glob Health*. (2017) 5:e865. doi: 10.1016/S2214-109X(17)30291-7

131. Granerod J, Tam CC, Crowcroft NS, NWS D, Borchert M, Thomas SL. Challenge of the unknown. A systematic review of acute encephalitis in non-outbreak situations. *Neurology.* (2010) 75:924–32. doi: 10.1212/WNL.0b013e3181f11d65

132. Trung DD, Praet N, Cam TDT, Lam BVT, Manh HN, Gabriël S, et al. Assessing the burden of human cysticercosis in Vietnam. *Trop Med Int Health*. (2013) 18:352–6. doi: 10.1111/tmi.12043

133. Hiraoka T, Cuong NC, Hamaguchi S, Kikuchi M, Katoh S, Anh LK, et al. Meningitis patients with *Angiostrongylus cantonensis* may present without eosinophilia in the cerebrospinal fluid in northern Vietnam. *PLoS Negl Trop Dis.* (2020) 14:e0008937. doi: 10.1371/journal.pntd.0008937

134. Shimasaki T, Chung H, Shiiki S. Five cases of recurrent meningitis associated with chronic Strongyloidiasis. Am J Trop Med Hyg. (2015) 92:601–4. doi: 10.4269/ajtmh.14-0564

135. Brindle HE, Bastos LS, Christley R, Contamin L, Dang LH, Anh DD, et al. The spatio-temporal distribution of acute encephalitis syndrome and its association with climate and landcover in Vietnam. *BMC Infect Dis.* (2023) 23:403. doi: 10.1186/s12879-023-08300-1

136. Cappelle J, Hoem T, Hul V, Furey N, Nguon K, Prigent S, et al. Nipah virus circulation at human–bat interfaces. *Cambodia Bull World Health Organ.* (2020) 98:539–47. doi: 10.2471/BLT.20.254227

137. Hasebe F, Thuy NTT, Inoue S, Yu F, Kaku Y, Watanabe S, et al. Serologic evidence of Nipah virus infection in bats, Vietnam - volume 18, number 3—march 2012 - emerging infectious diseases journal - CDC. (2023). Available from: https://wwwnc.cdc. gov/eid/article/18/3/11-1121_article (Accessed 11 December 2024).

138. Nabeshima T, Ngwe Tun MM, Thuy NTT, Hang NLK, Mai LTQ, Hasebe F, et al. An outbreak of a novel lineage of dengue virus 2 in Vietnam in 2022. Nagasaki University Vietnam Research Group. *J Med Virol*. (2023) 95:e29255.