



OPEN ACCESS

EDITED BY

Robin Mesnage,
King's College London, United Kingdom

REVIEWED BY

Svetlana Stanišić,
Singidunum University, Serbia
Shuva Bhowmik,
University of Otago, New Zealand

*CORRESPONDENCE

Chia-Te Kung
✉ a0953283092@gmail.com

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 14 March 2023

ACCEPTED 08 August 2023

PUBLISHED 30 August 2023

CITATION

Cheng F-J, Wang C-H, Pan H-Y, Chen C-C, Huang W-T, Li S-H, Wang L-J, Wang C-C, Lee W-C, Tsai K-F, Ou Y-C and Kung C-T (2023) Levels of organophosphate flame retardants and their metabolites among 391 volunteers in Taiwan: difference between adults and children. *Front. Public Health* 11:1186561. doi: 10.3389/fpubh.2023.1186561

COPYRIGHT

© 2023 Cheng, Wang, Pan, Chen, Huang, Li, Wang, Wang, Lee, Tsai, Ou and Kung. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Levels of organophosphate flame retardants and their metabolites among 391 volunteers in Taiwan: difference between adults and children

Fu-Jen Cheng^{1†}, Chih-Hwa Wang^{1†}, Hsiu-Yung Pan¹, Chih-Cheng Chen², Wan-Ting Huang³, Shau-Hsuan Li⁴, Liang-Jen Wang⁵, Chin-Chou Wang⁶, Wen-Chin Lee⁷, Kai-Fan Tsai⁷, Yu-Che Ou⁸ and Chia-Te Kung^{1*}

¹Department of Emergency Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ²Section of Neonatology, Pediatrics Department, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ³Department of Laboratory Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ⁴Division of Hematology-Oncology, Department of Internal Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ⁵Department of Child and Adolescent Psychiatry, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ⁶Department of Occupational Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ⁷Division of Nephrology, Department of Internal Medicine, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan, ⁸Department of Obstetrics and Gynecology, Kaohsiung Chang Gung Memorial Hospital, Chang Gung University College of Medicine, Kaohsiung, Taiwan

Background: Organophosphate flame retardants (OPFRs) are ubiquitous in the environment. The compositions and concentrations of different OPFRs metabolites vary in different environments depending on different human activities. The objective of the present study was to evaluate the exposure of different age groups to OPFRs in Taiwan.

Methods: Volunteers provided urine samples and responded to questionnaires including demographic factors, underlying disease, lifestyle information, and occupation from October 2021 to January 2022. OPFR measurements were performed using a Waters Acquity Ultra-Performance Liquid Chromatography system coupled with a Waters Xevo TQ-XS mass spectrometer.

Results: A total of 391 volunteers (74 children and 317 adults) were enrolled in this study. The concentrations (presented as $\mu\text{g/g}$ creatinine) of bis(1,3-dichloro-2-propyl) phosphate (BDCPP, $p = 0.029$) and tri-n-butyl phosphate (TNBP, $p = 0.008$) were higher in the adult group, while the concentrations of bis-2-chloroethyl phosphate (BCEP, $p = 0.024$), diphenyl phosphate (DPHP, $p < 0.001$), tris(1,3-dichloro-2-propyl) phosphate (TDCPP, $p = 0.009$), and Tris(2-butoxyethyl) phosphate (TBEP, $p = 0.007$) were higher in the child group. Compared with school age children (>6 years), the concentration of di(2-n-butoxyethyl) phthalate (DBEP, 1.14 vs. 0.20 $\mu\text{g/g}$ creatinine, $p = 0.001$), DPHP (1.23 vs. 0.54 $\mu\text{g/g}$ creatinine, $p = 0.036$), TBEP (1.63 vs. 0.29 $\mu\text{g/g}$ creatinine, $p < 0.001$), and the sum of OPFR metabolites (ΣOPFRs , 6.58 vs. 2.04 $\mu\text{g/g}$ creatinine, $p < 0.001$) were statistically higher in preschool-aged children. After adjusting for confounding factors, preschool age [odds ratio (OR): 4.579, 95% confidence interval (CI): 1.389–13.115] and

current smoker (OR: 5.328, 95%CI: 1.858–14.955) were independently associated with the risk of Σ OPFRs higher than 90 percentile.

Conclusion: This study revealed the distribution of different OPFRs metabolites in children and adults. DBEP, DPHP, TBEP, and Σ OPFR were higher in preschool-aged children. Pre-school age and current smoking status were independent risk factors for Σ OPFRs higher than 90 percentile.

KEYWORDS

organophosphate flame retardant, OPFR, OPFR metabolites, Tris(2-butoxyethyl) phosphate, Taiwan

1. Introduction

Organophosphate flame retardants (OPFRs) were introduced in the early twentieth century and became the major flame retardant in the market after polybrominated diphenyl ethers (PBDEs) were banned and phased-out. OPFRs are incorporated by physical addition, therefore, they are easily released into the environment through abrasion, leaching, wearing, and volatilization (1, 2). In fact, OPFRs can be emitted into the environment during production, transport, and application (3). OPFRs have been detected in various environments, such as air, dust, water (including lakes, rivers, oceans, drinking water, etc.), soil, and sediments, as well as in human samples and biota (4–7). Adverse health effects have been reported with some OPFRs. For instance, tris (2-carboxylethyl) phosphine (TCEP) has been listed as a carcinogen by the California Environmental Protection Agency since 1992 (8) and classified as a Category 2 carcinogen by the European Union (9). Tris(2-butoxyethyl) phosphate (TBEP) has been shown to induce developmental toxicity in zebrafish (10) and abnormal sperm morphology and testicular pathology in rats (11). Tri-n-butyl phosphate (TNBP) and triphenyl phosphate (TPHP) have been reported to be neurotoxic to zebrafish larvae and rats (12, 13). The dominant compounds of the OPFRs detected in different environmental media varied between different regions and might be influenced by human activity. For example, tris (1-chloro-2-propyl) phosphate (TCPP) (Australia, France, Germany, UK, USA), TBEP (Austria, Italy, Spain), and TCEP (China) are the three dominant compounds found in surface water. The concentrations of organophosphate esters (OPEs), one type of OPFRs, in dust are higher in more industrialized countries (such as the US, Canada, Norway, Sweden, the Netherlands, South Korea, and Australia) than in less industrialized countries (such as Colombia, Romania, Pakistan, Saudi Arabia, Kuwait, Nepal, Philippines, India, and Vietnam). Humans are exposed to OPFRs via various pathways, including air inhalation, dust ingestion, diet ingestion, and dermal contact with diet ingestion, which are reported to be the most significant pathways (14). In one study, the median estimated daily intakes (EDI) of OPEs through dust ingestion were around 0.29–64.8 ng/kg body weight (bw)/day for children and 0.07–14.9 ng/kg bw/day for adults (15).

The difference between adults and children might be related to differences in the major pathways of exposure. Human exposure to OPFRs has been confirmed by the frequent detection of OPFR metabolites in urine. For examples, bis(1-chloro-2-propyl) 1-hydroxy-2-propyl phosphate (BCIPHIPP) and diphenyl phosphate (DPHP) were reported to be the two most abundant compounds in urine in one study where they accounted for 46 and 39% of the total amount

of OPFRs (16). The highest level of DPHP was found in Australia, with geometric means (GM) of 24,400 pg./mL and 63,500 pg./mL from the two sample groups, respectively (17). High level of DPHP (GM=9,000 pg./mL) was observed in the urine samples collected from US gymnasts after training. Among three different studies of populations in North Carolina, a similar range of bis(1,3-dichloro-2-propyl) phosphate (BDCPP) was found, with GMs of 1,800 pg./mL in pregnant women (18), 2,300 pg./mL in infants (19), and 2,320 pg./mL in the general population (20). 2-ethylhexyl diphenyl phosphate (EHDPHP), tris(2-chloropropyl) phosphate (TCPP), and TBEP were reported to be the three major OPFRs that participants had been exposed to in other studies (6, 7). The varied urine levels and compositions of OPFRs probably imply differences in exposure sources, pathways, and intensities among different countries and populations. Currently, only limited data regarding human exposure to OPFRs in Taiwan is available (21). It is also unclear whether there is a difference in the exposure to OPFRs between adults and children. Our study thus aims to analyze urine samples from volunteers to identify the composition of OPFRs in human bodies of people in Taiwan and to compare the difference in exposure and composition between adults and children.

2. Materials and methods

2.1. Study population

From October 2021 to January 2022, 391 volunteers were recruited to investigate OPFR metabolites in their urine samples. The volunteers were relatively healthy without a history of major comorbidities, such as renal disease, stroke, malignancy, or myocardial infarction. After agreeing to the study design, each volunteer provided written informed consent and completed a face-to-face questionnaire. Demographic factors, such as age, sex, and underlying diseases, and lifestyle information, such as alcohol consumption, smoking, and occupation, were collected through the questionnaire. Urine samples were collected in polypropylene tubes, sub-aliquoted into 1.5 mL tubes, and then stored at -80°C until extraction.

2.2. Determination of OPFR metabolites and urine creatinine

OPFR measurements were performed using a Waters Acquity Ultra Performance Liquid Chromatography (UPLC) system (Milford)

TABLE 1 Demographic factors of 391 volunteers.

Demographic characteristics of volunteers (<i>n</i> = 391)	<i>n</i> (%)
Age (mean ± SD)	35.7 ± 16.7
Male	192 (49.1)
Diabetes	19 (4.9)
Hypertension	35 (9.1)
Dyslipidemia	13 (3.4)
Liver disease	12 (3.1)
Current smoker	32 (8.3)
Alcohol consumption	71 (18.5)
Manufacturing	28 (7.3)
Service industry	228 (59.4)
Student	68 (17.7)
Unemployed	60 (15.6)

TABLE 2 Detection frequency of the 10 OPFRs and their metabolites in adults (≥17 years) and children (<17 years) groups.

All	Number = 391 (%)		
Demographic characteristics of volunteers	Child (<i>n</i> = 74)	Adult (<i>n</i> = 317)	<i>p</i>
Age (mean ± SD)	8.4 ± 1.1	42.1 ± 11.3	<0.001
Male	40	152	0.344
Detection frequency			
BDCPP	11	90	0.012
BCEP	41	119	0.005
DBEP	37	134	0.228
DNBP	6	32	0.603
DPHP	71	252	0.001
TDCPP	22	55	0.016
TCEP	8	42	0.572
TBEP	50	173	0.042
TNBP	49	251	0.018
TPHP	30	108	0.294

coupled with a Waters Xevo TQ-XS mass spectrometer (Milford), operated in either negative or positive electrospray ionization (ESI) mode. The methods of OPFRs analysis have been described in a previous study (21). Briefly, all OPFRs and metabolites were separated using a Waters Acquity UPLC BEH Phenyl column preceded by a Waters XBridge BEH C18 Direct Connect HP-isolated column. The mobile phases were 0.5% formic acid in water (A) and 0.5% formic acid in methanol (B). The gradient was increased from initial 5 to 50% of solvent (B) linearly within 0.75 min. Then the mobile phase (B) was increased to 100% in another 3 min and held for 4.5 min. Finally, the gradient was decreased to the initial 5% of solvent (B) for a 2 min re-equilibrium. The target analytes were identified according to the retention time and ratio of the two selected precursor-ion-produced ion transitions compared with those of the standards. The

concentrations of OPFRs were quantified using 12-point calibration curves (ranged from 0.02 ppb to 50 ppb with a two-fold dilution), the limits of quantitation (LOQs) were 0.02 ng/mL (TNBP, DNBP, TBEP, DEBP, TPHP, DPHP, TDCPP, BDCPP, TCEP) and 0.05 ng/mL (BCEP) and the limit of detection (LOD) was shown in Supplementary Table S1. Their recoveries were performed using the internal standard method based on individual isotope-labeled internal standards were also shown in Supplementary Table S1.

Urine creatinine was measured by colorimetric Jaffe method using the MeDiPRO creatinine kinase test (Formosa Biomedical Technology, Taipei, Taiwan). The assay was performed using an automatic biochemical analyzer Hitachi LABOSPECT 008 (Hitachi, Yokohama, Japan). Creatinine standard solution was purchased from Wako Pure Chemical Corporation (Osaka, Japan). Urine quality control material was purchased from Bio-Rad (Hercules, California, United States). The method was standardized according to the isotope dilution mass spectrometry (IDMS) traceable National Institute of Standards and Technology Standard Reference Material (NIST SRM) 967. This study was approved by the Institutional Review Board of Chang Gung Memorial Hospital (IRB no:202001031A3 and NO:202001028A3) and was conducted in accordance with the Code of Ethics of the World Medical Association (Declaration of Helsinki).

2.3. Statistical analysis

The OPFR metabolite concentrations were presented in two ways: micrograms per liter [µg/L] and micrograms per gram of creatinine [µg/g creatinine]. Normally distributed continuous data are presented as mean ± standard deviation (SD), and non-normally distributed continuous data are presented as the mean with 95% confidence intervals (CIs). The independent *t*-test and Mann-Whitney U test were used to examine the differences in the distribution of continuous variables. Chi-square tests were used to assess the differences in categorical variables. Binary logistic regression model was used to examine the likelihood of having concentrations above the 90 percentile for major OPFR metabolites and calculate odds ratios (ORs), 95% CIs, and *p*-values. Statistical significance was set at *p* < 0.05. All statistical analyses were performed using SPSS version 25.0 (IBM Corp., Armonk, NY, United States).

3. Results

3.1. Demographic factors of volunteers

Table 1 shows the demographic characteristics of the 391 participants. The mean age of the study participants was 35.7 years. 192 (49.1%) volunteers were male, 35 (9.1%) had hypertension, 32 (8.3%) volunteers were current smokers, and 228 (59.4%) engaged in service industry.

3.2. Distribution of OPFRs and its metabolites in adults and children

Table 2 shows the detection frequency of the 10 OPFR metabolites in the adults (≥17 years) and children (<17 years) groups. The

TABLE 3 Concentrations of OPFRs and their metabolites in adult and child groups.

	Child (=74)	Adult (=317)	<i>p</i>
Male	40	152	0.344
Age	8.4 ± 4.1	42.1 ± 11.3	<0.001
OPFRs metabolites (mean, 95% CI)			
BDCPP (µg/L)	0.28 (0.09–0.46)	0.69 (0.48–0.89)	0.016
BDCPP (µg/g creatinine)	0.30 (0.11–0.48)	1.00 (0.70–1.32)	0.029
BCEP (µg/L)	0.44 (0.30–0.58)	0.37 (0.29–0.45)	0.02
BCEP (µg/g creatinine)	0.67 (0.34–1.00)	0.64 (0.38–0.92)	0.024
DBEP (µg/L)	0.25 (0.14–0.36)	0.33 (0.17–0.49)	0.209
DBEP (µg/g creatinine)	0.45 (0.18–0.72)	0.50 (0.26–0.74)	0.119
DNBP (µg/L)	0.00 (0.00–0.00)	0.01 (0.00–0.02)	0.561
DNBP (µg/g creatinine)	0.00 (0.00–0.01)	0.02 (0.01–0.05)	0.463
DPHP (µg/L)	0.52 (0.33–0.71)	0.92 (–0.44–2.29)	<0.001
DPHP (µg/g creatinine)	0.73 (0.46–1.00)	1.31 (–0.47–3.10)	<0.001
TDCPP (µg/L)	0.03 (0.01–0.05)	0.02 (0.01–0.03)	0.016
TDCPP (µg/g creatinine)	0.04 (0.02–0.07)	0.04 (0.01–0.08)	0.009
TCEP (µg/L)	0.25 (0.08–0.73)	0.29 (0.18–0.40)	0.63
TCEP (µg/g creatinine)	0.31 (0.07–0.54)	0.46 (0.27–0.67)	0.512
TBEP (µg/L)	0.37 (0.23–0.51)	0.22 (0.17–0.27)	0.013
TBEP (µg/g creatinine)	0.65 (0.30–1.00)	0.41 (0.26–0.57)	0.007
TNBP (µg/L)	0.03 (0.02–0.04)	0.06 (0.05–0.08)	0.009
TNBP (µg/g creatinine)	0.08 (0.02–0.14)	0.10 (0.08–0.13)	0.008
TPHP (µg/L)	0.02 (0.01–0.03)	0.02 (0.01–0.02)	0.322
TPHP (µg/g creatinine)	0.03 (0.01–0.05)	0.03 (0.02–0.05)	0.274
Sum of OPFRs (µg/L)	2.19 (1.75–2.62)	2.93 (1.50–4.35)	0.175
Sum of OPFRs (µg/g creatinine)	3.27 (2.34–4.19)	4.50 (2.59–6.43)	0.254

detection rates of BDCPP ($p=0.012$) and TNBP ($p=0.018$) were higher in the adult group, while bis(2-chloroethyl) phosphate (BCEP) ($p=0.005$), DPHP ($p=0.001$), tris (1,3-dichloro-2-propyl) phosphate (TDCPP) ($p=0.016$), and TBEP ($p=0.042$) were higher in the children group.

Table 3 shows the demographic factors of volunteers and concentrations of OPFR metabolites in the adult and child groups, reported in both µg/L and µg/g creatinine. The concentrations of BDCPP ($p=0.016$ and 0.029) and TNBP ($p=0.009$ and 0.008 , respectively) were significantly higher in the adult group. The concentrations of BCEP ($p=0.020$ and 0.024), DPHP ($p<0.001$ and <0.001), TDCPP ($p=0.016$ and 0.009), and TBEP ($p=0.013$ and 0.007) were significantly higher in the children group. The sum of OPFR metabolites (Σ OPFRs) was not significantly different between the adult and child groups ($p=0.175$ and 0.254 , respectively).

The mean percentage of each OPFR metabolite in the Σ OPFRs (µg/g creatinine) is shown in Figure 1. DPHP ($21.9 \pm 28.7\%$), BDCPP ($16.8 \pm 29.8\%$), BCEP ($15.1 \pm 25.1\%$), and TBEP ($13.4 \pm 20.3\%$) were the leading components of the Σ OPFRs in adults (Figure 1A). DPHP ($30.1 \pm 29.9\%$), BCEP ($22.8 \pm 25.8\%$), TBEP ($16.1 \pm 2.3\%$), and di(2-n-butoxyethyl) phthalate (DBEP) ($10.8 \pm 14.9\%$) were the leading components of the Σ OPFRs in children (Figure 1B).

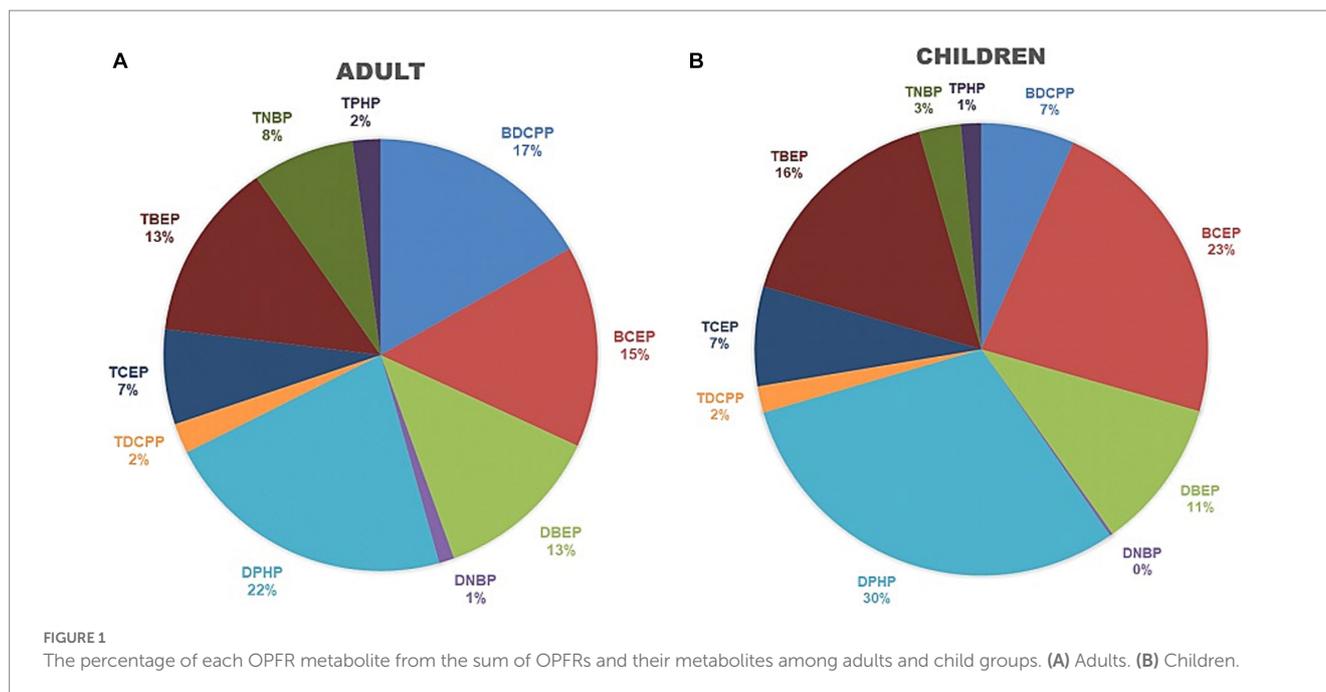
3.3. OPFR metabolites distribution among different age groups

Table 4 shows the concentrations of OPFR metabolites among preschool-aged children (≤ 6 years) and school-aged children (> 6 years), represented in µg/L and µg/g creatinine. The concentrations of DBEP (µg/g creatinine, $p=0.001$), DPHP (µg/g creatinine, $p=0.036$), TDCPP (µg/g creatinine, $p=0.044$), TBEP ($p=0.004$ and $p<0.001$), TNBP ($p=0.032$ and 0.004), and Σ OPFRs ($p=0.043$ and <0.001) were significantly higher in the preschool-aged children.

Table 5 shows the results of OPFR metabolites in younger adults (≤ 40 years) and older adults (> 40 years). The difference in all OPFR metabolites between these two groups was not statistically significant. The mean level of Σ OPFRs metabolites was $2.94 \mu\text{g/L}$ ($6.58 \mu\text{g/g creatinine}$) in the younger adult group and $1.91 \mu\text{g/L}$ ($2.04 \mu\text{g/g creatinine}$) in the older adult group.

3.4. Risk factors for concentrations of each OPFRs above the 90th percentile

The characteristics and comorbidities of the volunteers above and under the 90th percentile of major OPFRs, including BDCPP, BCEP,



DBEP, DPHP, TBEP, and Σ OPFRs, are listed in [Supplementary Tables S2–S7](#). A binary logistic regression model was used to analyze the independent factors associated with the risk of major OPFRs metabolites and Σ OPFRs higher than 90 percentile. As shown in [Table 6](#), preschool age ($p=0.015$) and current smoking status ($p=0.002$) were independently associated with the risk of Σ OPFRs higher than 90 percentile. Being a student was an independent factor associated with lower risk of BDCPP higher than 90 percentile (OR=0.154, 95% CI: 0.008–0.9, $p=0.036$). Pre-school age ($p=0.005$) and current smoking ($p=0.017$) were independently associated with the risk of BCEP higher than 90 percentile. Pre-school age ($p=0.015$), current smoking status ($p=0.047$), and diabetes ($p=0.023$) were independently associated with a risk of DBEP higher than 90 percentile. Pre-school age ($p=0.002$) and diabetes ($p=0.013$) were independently associated with a risk of DPHP higher than 90 percentile. Pre-school age ($p=0.009$) was independently associated with TBEP higher than 90 percentile.

4. Discussion

The present study is the first to evaluate the composition of major OPFRs and their metabolites in humans in Taiwan. We found a difference in the distribution among children and adults. BDCPP and TNBP were higher in adults, while BCEP, DPHP, TDCPP, and TBEP were higher in children. The BCEP, DBEP, DPHP, TBEP, TNBP, and Σ OPFRs were higher in preschool-aged children. Preschool age was an independent risk factor for Σ OPFRs, DPHP, BCEP, DBEP, DPHP, and TBEP more than 90 percentile. Current smoking was an independent risk factor for Σ OPFR, BCEP, and DBEP more than 90 percentile.

OPFRs were detected in the air, indoor dust, soil, sediment, water, and sludge. A study based on Global Atmospheric Passive sampling (GPAS) network revealed that TCEP, TBEP, TCPP, TDCPP, TPHP, triethyl phosphate (TEP), and TNBP were most frequently detected

OPEs in outdoor air, with detected frequencies of 88.2, 84.0, 81.7, 75.7, 69.2, 53, and 40.2%, respectively (22). TCPP, TCEP, TBEP, and TPHP were four dominant OPEs, which accounted for 45, 18, 17, and 9% of the total concentrations of OPEs (Σ_{18} OPEs). The total concentrations of OPRs in indoor air were higher than those found in outdoor air. For example, the level of Σ OPEs in indoor air was eight times higher than that in outdoor air (median:40.2 ng/m³ vs. 5.13 ng/m³) in Germany (23) and more than 100 times higher in Stockholm, Sweden (340 ng/m³ vs. 3.1 ng/m³) (24). The composition of OPFRs in air varies between different countries and across indoor microenvironments, possibly because of their diverse applications in building materials and consumer products (24). Overall, TCPP, tri-iso-butylphosphate (TiBP), TNBP, and TBEP are the most abundant OPEs in indoor air. TBEP, TCPP, TCEP, TDCPP, and TPHP were the five compounds most frequently discovered in home dust at high concentrations. Higher levels of OPFRs are found in more industrialized countries (such as the US, Canada, South Korea, the Netherlands, and Australia) than that in less industrialized countries (such as Pakistan, Nepal, Vietnam, etc.). TBEP was dominated in countries with high levels of Σ OPFRs such as Japan (25), Australia (26), Canada (27), and South Korea (15) with the contributions ranging from 30 to 97%. Data on the OPFRs concentrations and compositions in different environmental media in Taiwan are lacking. We speculated that TDCPP, TCEP, TPHP, and TBEP might be the main OPFRs in our surroundings, since their metabolites were most frequently identified in our study.

Human exposure to OPFS occurs through various pathways, including air inhalation, dust ingestion, dietary intake, and dermal adsorption. In a study conducted, it was determined that the primary route of exposure for heavier organic phosphate esters (OPEs) such as TBEP and TPHP was through ingestion of dust. On the other hand, volatile OPEs like TCEP (tris(2-chloroethyl) phosphate) and TCPP (tris(2-chloro-1-methylethyl) phosphate) were predominantly absorbed through inhalation of air (6). Indoor dust from 12 countries was investigated in another study, and the reported median estimated

TABLE 4 Concentrations of OPFRs and their metabolites among pre-school-age children (≤ 6 years) and school-age children (>6 years).

Demographic characteristics of volunteers	≤ 6 years ($n = 20$)	>6 years ($n = 54$)	p
Age (mean \pm SD)	4.8 \pm 2.2	11.5 \pm 2.4	<0.001
Male	18	22	
OPFRs	Mean (95% CI)	Mean (95% CI)	
BDCPP ($\mu\text{g/L}$)	0.26 (−0.01–0.54)	0.28 (0.04–0.52)	0.529
BDCPP ($\mu\text{g/g creatinine}$)	0.44 (−0.05–0.92)	0.24 (0.06–0.43)	0.443
BCEP ($\mu\text{g/L}$)	0.65 (0.26–1.03)	0.36 (0.24–0.49)	0.221
BCEP ($\mu\text{g/g creatinine}$)	1.47 (0.31–2.63)	0.38 (0.23–0.53)	0.1
DBEP ($\mu\text{g/L}$)	0.46 (0.12–0.80)	0.18 (0.09–0.26)	0.01
DBEP ($\mu\text{g/g creatinine}$)	1.14 (0.18–2.09)	0.20 (0.10–0.30)	0.001
DNBP ($\mu\text{g/L}$)	0.00 (0.00–0.00)	0.00 (0.00–0.00)	0.58
DNBP ($\mu\text{g/g creatinine}$)	0.00 (0.00–0.01)	0.00 (0.00–0.01)	0.598
DPHP ($\mu\text{g/L}$)	0.60 (0.19–1.00)	0.49 (0.27–0.71)	0.425
DPHP ($\mu\text{g/g creatinine}$)	1.23 (0.42–2.04)	0.54 (0.32–0.76)	0.036
TDCPP ($\mu\text{g/L}$)	0.04 (0.00–0.08)	0.03 (0.01–0.05)	0.089
TDCPP ($\mu\text{g/g creatinine}$)	0.08 (0.02–0.14)	0.03 (0.01–0.06)	0.044
TCEP ($\mu\text{g/L}$)	0.25 (−0.11–0.62)	0.24 (0.05–0.44)	0.937
TCEP ($\mu\text{g/g creatinine}$)	0.34 (−0.17–0.85)	0.29 (0.03–0.56)	0.91
TBEP ($\mu\text{g/L}$)	0.62 (0.23–1.02)	0.27 (0.14–0.40)	0.004
TBEP ($\mu\text{g/g creatinine}$)	1.63 (0.41–2.85)	0.29 (0.16–0.42)	<0.001
TNBP ($\mu\text{g/L}$)	0.04 (0.03–0.06)	0.03 (0.02–0.03)	0.032
TNBP ($\mu\text{g/g creatinine}$)	0.2 (−0.01–0.41)	0.03 (0.02–0.05)	0.004
TPHP ($\mu\text{g/L}$)	0.01 (0.00–0.03)	0.02 (0.01–0.04)	0.967
TPHP ($\mu\text{g/g creatinine}$)	0.05 (0.00–0.01)	0.02 (0.01–0.03)	0.612
Sum of OPFRs ($\mu\text{g/L}$)	2.94 (1.91–3.98)	1.91 (1.45–2.36)	0.043
Sum of OPFRs ($\mu\text{g/g creatinine}$)	6.58 (3.67–9.49)	2.04 (1.61–2.47)	<0.001

daily intake (EDIs) of OPEs through dust ingestion were in the range of 0.29–64.8 ng/Kg body weight (bw)/day for children and 0.07–14.9 ng/kg bw/day for adults (15). He et al. (28) compared the EDIs of nine OPEs from air, dust, and food to discover that TBEP (accounting from the total exposure: 57%) was mainly from indoor dust (ingestion and dermal contact), while tris(2-chloroisopropyl) phosphate (TCIPP) (77%), TCEP (84%), and TNBP (93%) were mainly from dietary intake. Additionally, for young children, several unique exposure pathways exist that could contribute to their exposure to OPFRs. These pathways include the use of infant products and a higher frequency of hand-to-mouth behavior, which may increase their susceptibility to OPFR exposure (19). The hand-to-mouth pathway, rapid ventilation rates in infants and toddlers, and use of infant products might result in increased OPFRs exposure through dermal contact, dust ingestion, and air inhalation, thus explaining why levels of TBEP, DPHP, and BCEP were higher in children, especially for young children in the present study. Due to the potential inhalation and dermal exposure through dust and air, increasing the frequency of vacuuming and handwashing may be effective in reducing children's exposure to Organophosphate Flame Retardants (OPFRs). These measures could help minimize the risk associated with OPFR exposure in children (28).

Adults are inevitably exposed to OPFRs in daily life since OPFRs are ubiquitous in the environment. Increased exposure to OPFRs might result from special behaviors, such as tobacco use, cigarette use, and occupational exposure. Higher detection rates and higher concentrations of OPFRs metabolites were reported in the urine of firefighters, which might be related to a higher incidence of OPFRs exposure from work (29–31). Higher detection frequencies of DPHP in spot urine samples from nail salon workers have also been reported (32, 33), and the plausible reasons might be increased TPHP exposure from the workplace of nail polishing via air inhalation, dermal absorption, and dust ingestion. People who smoke cigarettes are supposed to be exposed to higher amounts of TPHP because the adjusted GM of DPHP for cigarette smokers is higher than that of non-smokers (34). Sun et al. (35) noticed that the urine concentrations of bis(1-chloro-2-propyl) phosphate (BCIPP) in smokers and ex-smokers were higher (GM 42.3 pg/mL and GM 0.9 pg/mL, respectively) than those in non-smokers [GM 19.7 pg/mL, <limit of quantification (LOQ)], suggesting that tobacco smoking might lead to altered metabolism which results in the formation of di(methylphenyl) phosphate (DMPP) and BCIPP from their corresponding parent compounds. Our study discovered that the detection rate and urine concentration of BDCPP and TNBP were significantly higher in the

TABLE 5 OPFRs and their metabolites in younger adults (≤ 40 years) group and older adults (> 40 years) group.

All	Number=	%	
Demographic characteristics of volunteers	≤ 40 years ($n = 152$)	> 40 ($n = 165$)	<i>p</i>
Age (mean \pm SD)	33.0 \pm 5.4	50.5 \pm 8.4	<0.001
Male	65	87	0.076
OPFRs	Mean (95% CI)	Mean (95% CI)	
BDCPP ($\mu\text{g/L}$)	0.53 (0.36–0.71)	0.83 (0.46–1.19)	0.509
BDCPP ($\mu\text{g/g creatinine}$)	0.89 (0.45, 1.33)	1.55 (0, 0.97)	0.478
BCEP ($\mu\text{g/L}$)	0.32 (0.22–0.42)	0.42 (0.29–0.54)	0.872
BCEP ($\mu\text{g/g creatinine}$)	0.42 (0.03, 0.80)	0.85 (0.48, 1.22)	0.907
DBEP ($\mu\text{g/L}$)	0.32 (0.11–0.54)	0.33 (0.09–0.57)	0.398
DBEP ($\mu\text{g/g creatinine}$)	0.53 (0.19, 0.87)	0.47 (0.14, 0.79)	0.406
DNBP ($\mu\text{g/L}$)	0.02 (0.00–0.03)	0.01 (0.00–0.02)	0.169
DNBP ($\mu\text{g/g creatinine}$)	0.03 (0.01, 0.06)	0.01 (0.01, 0.03)	0.167
DPHP ($\mu\text{g/L}$)	0.21 (0.14–0.27)	1.58 (–1.05–4.22)	0.337
DPHP ($\mu\text{g/g creatinine}$)	0.33 (–2.14, 2.90)	2.21 (–0.26, 4.68)	0.253
TDCPP ($\mu\text{g/L}$)	0.02 (0.01–0.04)	0.01 (0.00–0.02)	0.812
TDCPP ($\mu\text{g/g creatinine}$)	0.05 (0.01, 0.10)	0.02 (–0.02, 0.06)	0.854
TCEP ($\mu\text{g/L}$)	0.24 (0.13–0.36)	0.34 (0.16–0.52)	0.64
TCEP ($\mu\text{g/g creatinine}$)	0.33 (0.05, 0.61)	0.59 (0.32, 0.86)	0.667
TBEP ($\mu\text{g/L}$)	0.21 (0.14–0.28)	0.24 (0.16–0.31)	0.473
TBEP ($\mu\text{g/g creatinine}$)	0.41 (0.19, 0.62)	0.40 (0.20, 0.61)	0.294
TNBP ($\mu\text{g/L}$)	0.06 (0.05–0.07)	0.06 (0.04–0.07)	0.636
TNBP ($\mu\text{g/g creatinine}$)	0.11 (0.07, 0.14)	0.09 (0.06, 0.13)	0.624
TPHP ($\mu\text{g/L}$)	0.02 (0.01–0.03)	0.02 (0.01–0.02)	0.797
TPHP ($\mu\text{g/g creatinine}$)	0.03 (0.01, 0.04)	0.03 (0.01, 0.04)	0.627
Sum of OPFRs ($\mu\text{g/L}$)	1.96 (1.60–2.31)	3.82 (1.10–6.55)	0.3
Sum of OPFRs ($\mu\text{g/g creatinine}$)	3.27 (0.50, 6.04)	5.87 (3.20, 8.53)	0.253

adult groups, and that current smoking was an independent risk factor for Σ OPFRs, BCEP, and DBEP more than 90 percentile.

The assessment of human OPFRs exposure through biomonitoring presents certain challenges due to the relatively short half-lives (ranging from hours to days) of OPFRs in biota (5). Considering the rapid metabolism of Organophosphate Flame Retardants (OPFRs), the analysis and assessment of exposure to OPFRs may be better focused on the excreted metabolites, as they provide a more appropriate target for analysis (17, 36). BDCPP and DPHP were the most frequently detected OPFRs in spot urine samples among the nine OPFRs [BDCPP, BCIPP, BCEP, DPHP, di-*p*-cresylphosphate (DPCP), di-*o*-cresylphosphate (DOCP), dibutyl phosphate (DNBP), dibenzyl phosphate (DBZP), tetrabromobenzoic acid (TBBA)] studied in the National Health and Nutrition Examination Survey (NHANES) in the United States during 2013–2014 (37). In this study, BDCPP and DPHP were found in approximately 92% of the samples, followed by BCEP, DNBP, and BCIPP, with the detection frequencies of 89, 81, and 61%, respectively. Moreover, the concentration was highest for DPHP (ranged <0.16 – $193 \mu\text{g/L}$), followed by BDCPP (<0.11 – $169 \mu\text{g/L}$), and BCEP (<0.08 – $110 \mu\text{g/L}$) (37). Wang et al. (38) analyzed eight types of OPFRs [DHPH, BDCPP, DOCP+DPCP, bis(2-butoxyethyl) phosphate (BBOEP), BCIPHIPP, 4-hydroxyphenyl-diphenyl phosphate

(4-HO-DPHP), dibutyl phosphate (DBP), and TCEP] in the urine samples collected from healthy volunteers and found that DPHP, DOCP + DPCP, and BCIPHIPP were detected in more than 90% of the spot and first morning voids. Several studies have documented high detection rates of BDCPP and DPHP in pregnant women and children (36, 39, 40).

In our study, DPHP, TBEP, and TNBP were the most frequently detected OPFRs in urine samples collected from children, with detection frequencies of 95.9, 67.6, and 66.2%, respectively. The concentration was highest for DPHP (ranged 0.33 – $0.71 \mu\text{g/L}$), followed by BCEP (0.30 – $0.58 \mu\text{g/L}$), and TBEP (0.23 – $0.51 \mu\text{g/L}$). In the adult group, DPHP was detected in approximately 79.5% of the samples, followed by TNBP (79.2%), and TBEP (54.6%). The concentration was highest for DPHP (ranged 0.44 – $2.29 \mu\text{g/L}$), followed by BDCPP (0.48 – $0.89 \mu\text{g/L}$), BCEP (0.29 – $0.45 \mu\text{g/L}$), and DBEP (0.17 – $0.49 \mu\text{g/L}$). The detection frequencies of OPFRs metabolites in urine samples in our study were consistent with the results reported by Ospina et al. (37), with DPHP, TNBP, and bis(*n*-butyl) phosphate (BNBP) (the metabolite of TNBP) being the most frequently detected. In particular, the detection rate of TBEP was significantly higher in the study group. TBEP is a plasticizer and antifoaming agent used in paints and floor finishes (41). Besides, our

TABLE 6 Binary logistic regression of each independent factor associated with the risk of major OPFRs metabolite and Σ OPFRs higher than 90 percentile.

	Adjusted odds ratios for Σ OPFRs >90 percentile					Adjusted odds ratios for BDCPP >90 percentile			
	OR	95% CI		p		OR	95% CI		p
Pre-school age (<6 years)	4.579	1.389	13.115	0.015	Pre-school age (<6 years)	1.149	0.060	7.063	0.901
Male sex	0.747	0.326	1.643	0.471	Male sex	0.762	0.348	1.604	0.478
Current smoker	5.328	1.858	14.955	0.002	Current smoker	1.748	0.519	5.188	0.347
					Service industry	1.381	0.623	3.395	0.439
					Student	0.154	0.008	0.9	0.036
	Adjusted odds ratios for BCEP >90 percentile					Adjusted odds ratios for DBEP >90 percentile			
	OR	95% CI		p		OR	95% CI		p
Pre-school age (<6 years)	5.139	1.700	14.075	0.005	Pre-school age (<6 years)	4.549	1.377	13.057	0.015
Male sex	0.789	0.364	1.660	0.535	Male sex	0.754	0.340	1.613	0.470
Current smoker	3.791	1.293	10.502	0.017	Current smoker	3.222	1.016	9.430	0.047
					Diabetes	4.141	1.246	11.96	0.023
	Adjusted odds ratios for DPHP >90 percentile					Adjusted odds ratios for TBEP >90 percentile			
	OR	95% CI		p		OR	95% CI		p
Pre-school age (<6 years)	6.439	2.086	18.187	0.002	Pre-school age (<6 years)	4.636	1.517	12.852	0.009
Male sex	0.589	0.260	1.262	0.176	Male sex	2.046	0.967	4.446	0.061
Current smoker	2.603	0.714	8.400	0.139	Current smoker	1.552	0.475	4.361	0.442
Diabetes	4.583	1.422	13.339	0.013	Manufacturing	0.292	0.016	1.486	0.16
Hypertension	2.117	0.723	5.478	0.162					

findings revealed that hypertension and diabetes are independent factors with a DPHP (diphenyl phosphate) levels exceeding the 90th percentile. However, our results could not clarify whether these associations are attributed to the diseases themselves or the medications used.

TBEP was detected in dust from solid waste, e-waste dumping sites, landfills, and wastewater treatment facilities (42). Previous studies from Japan revealed that TBEP was the most prevalent OPFRs in houses (43, 44). Studies utilizing zebrafish as an animal model reported that TBEP induced neurotoxicity and developmental impairments (45–47). Additionally, it was found to reduce body length, heart rate, survival rate, and hatching rate (48), as well as modify motor behavior and oxidative stress in zebrafish (49). TBEP was also shown to induce abnormal sperm morphology and testicular pathology in rats (11). The higher detection frequency of TBEP might have resulted from a higher amount of TBEP exposure from indoor air and home dust. Further work is needed to explore the concentrations of these OPFRs in different environmental media in Taiwan to elucidate the major sources of TBEP exposure.

BDCPP is unique to TDCPP and is the most appropriate metabolite to study TDCPP exposure. BCEP and DPHP were the corresponding metabolites of TCEP and TPHP, respectively. Reports has shown that TDCPP and TCEP increase apoptosis, decrease cell growth, alter morphology, and induce changes in the mRNA and protein expression levels of GAP43, tubulin, and NF-H (50). TCEP has been classified as a Category 2 carcinogen

by the European Union (9) and has been listed as a carcinogen by the California Environmental Protection Agency since 1992 (8). The European Commission proposed restricting TCEP and TDCPP in toys intended for children (under 3 years) or those that can be put toys into their mouth (>3 years) (50). The application of TCEP was effectively banned after it was listed on the Annex XVI authorization by the European Union. The addition of halogenated OPFRs to children's products, mattresses, furniture, and electronic enclosures was prohibited in 2017 by the United States Consumer Product Safety Commission (CPSC). TPHP is a potent monocyte carboxylase inhibitor in human blood (51), and it induces contact allergy (52). TPHP disrupts reproductive performance in zebrafish (53), and it interferes with the endocrine (54–56) and metabolic systems (57). TDCPP and TPHP found in dust are associated with alterations in male hormone levels and decreased sperm quality (58). Adverse health effects following OPFRs exposure have been previously reported. The importance of surveillance and close monitoring of OPFRs exposure, in addition to the regulation of OPFRs applications, should be emphasized.

5. Limitations

First, our study was conducted in a single region and a single hospital with recruitment through volunteered enrollment; therefore,

the results might not be generalizable to other populations, especially for different regions or countries. Second, a single measurement of OPFRs and their metabolites in a spot urine sample was used; however, weekly or monthly variations might exist. Third, we only examined the OPFRs and their metabolites in urine samples and analyzed their compositions and concentrations in the surrounding environment. Exposure assessment was not performed either. The direct relationship between environmental exposure and our results could not be determined.

6. Conclusion

In conclusion, our study identified different OPFRs and their metabolite distributions in children and adults. We found that BDCPP and TNBP were higher in adults, while BCEP, DPHP, TDCPP, and TBEP were higher in children. The BCEP, DBEP, DPHP, TBEP, TNBP, and Σ OPFRs were higher in preschool-aged children. Preschool age was an independent risk factor for Σ OPFRs, DPHP, BCEP, DBEP, DPHP, and TBEP more than 90 percentile. Current smoking was an independent risk factor for Σ OPFR, BCEP, and DBEP more than 90 percentile. Since children are more vulnerable to OPFRs exposure, more works are needed to better elucidate the relationship from environmental exposure and health impacts. We would collect more samples from children and also from the surrounding environments in Taiwan in the future. We hope that the results of this study will motivate the government to pay closer attention to managing the environmental pollution caused by OPFRs and their metabolites.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by Institutional Review Board of Kaohsiung Chang Gung Memorial Hospital (IRB NO: 202001031A3 and NO: 202001028A3). The studies were conducted in accordance with the local legislation and institutional

requirements. Written informed consent for participation in this study was provided by the participants' legal guardians/next of kin.

Author contributions

F-JC and C-TK contributed to conception and design of the study. F-JC organized the database. H-YP performed the statistical analysis. C-HW and H-YP wrote the first draft of the manuscript. All authors wrote sections of the manuscript. All authors contributed to the article and approved the submitted version.

Funding

This study was supported in part by research grants from Kaohsiung Chang Gung Memorial Hospital (Grant number CMRPG8K1301-3). The sponsor played no role in the study design, data collection, analysis, interpretation, writing of the report, or the decision to submit the article for publication.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpubh.2023.1186561/full#supplementary-material>

References

1. Marklund A, Andersson B, Haglund P. Screening of organophosphorus compounds and their distribution in various indoor environments. *Chemosphere*. (2003) 53:1137–46. doi: 10.1016/S0045-6535(03)00666-0
2. Lai S, Xie Z, Song T, Tang J, Zhang Y, Mi W, et al. Occurrence and dry deposition of organophosphate esters in atmospheric particles over the northern South China Sea. *Chemosphere*. (2015) 127:195–200. doi: 10.1016/j.chemosphere.2015.02.015
3. Bekele TG, Zhao H, Wang Q, Chen J. Bioaccumulation and trophic transfer of emerging organophosphate flame retardants in the marine food webs of Laizhou Bay. *North China Environ Sci Technol*. (2019) 53:13417–26. doi: 10.1021/acs.est.9b03687
4. Brandsma SH, Leonards PE, Leslie HA, de Boer J. Tracing organophosphorus and brominated flame retardants and plasticizers in an estuarine food web. *Sci Total Environ*. (2015) 505:22–31. doi: 10.1016/j.scitotenv.2014.08.072
5. van der Veen I, de Boer J. Phosphorus flame retardants: properties, production, environmental occurrence, toxicity and analysis. *Chemosphere*. (2012) 88:1119–53. doi: 10.1016/j.chemosphere.2012.03.067
6. Xu F, Giovanoulis G, van Waas S, Padilla-Sanchez JA, Papadopoulou E, Magnér J, et al. Comprehensive study of human external exposure to organophosphate flame retardants via air, dust, and hand wipes: the importance of sampling and assessment strategy. *Environ Sci Technol*. (2016) 50:7752–60. doi: 10.1021/acs.est.6b00246
7. Xu F, Tay JH, Covaci A, Padilla-Sanchez JA, Papadopoulou E, Haug LS, et al. Assessment of dietary exposure to organohalogen contaminants, legacy and emerging flame retardants in a Norwegian cohort. *Environ Int*. (2017) 102:236–43. doi: 10.1016/j.envint.2017.03.009
8. Stubbings WA, Kajiwara N, Takigami H, Harrad S. Leaching behaviour of hexabromocyclododecane from treated curtains. *Chemosphere*. (2016) 144:2091–6. doi: 10.1016/j.chemosphere.2015.10.121
9. European Union. (2009). Available at: <https://echa.europa.eu/substanceinformation/-/substanceinfo/100.003.744>
10. Han Z, Wang Q, Fu J, Chen H, Zhao Y, Zhou B, et al. Multiple bio-analytical methods to reveal possible molecular mechanisms of developmental toxicity in zebrafish

- embryos/larvae exposed to tris(2-butoxyethyl) phosphate. *Aquat Toxicol.* (2014) 150:175–81. doi: 10.1016/j.aquatox.2014.03.013
11. Pan HY, Cheng FJ, Huang KC, Kung CT, Huang WT, You HL, et al. Exposure to tris(2-butoxyethyl) phosphate induces abnormal sperm morphology and testicular histopathology in male rats. *Ecotoxicol Environ Saf.* (2022) 241:113718. doi: 10.1016/j.ecoenv.2022.113718
12. Behl M, Hsieh JH, Shafer TJ, Mundy WR, Rice JR, Boyd WA, et al. Use of alternative assays to identify and prioritize organophosphorus flame retardants for potential developmental and neurotoxicity. *Neurotoxicol Teratol.* (2015) 52:181–93. doi: 10.1016/j.ntt.2015.09.003
13. Shi Q, Wang M, Shi F, Yang L, Guo Y, Feng C, et al. Developmental neurotoxicity of triphenyl phosphate in zebrafish larvae. *Aquat Toxicol.* (2018) 203:80–7. doi: 10.1016/j.aquatox.2018.08.001
14. Ding J, Deng T, Xu M, Wang S, Yang F. Residuals of organophosphate esters in foodstuffs and implication for human exposure. *Environ Pollut.* (2018) 233:986–91. doi: 10.1016/j.envpol.2017.09.092
15. Li W, Wang Y, Asimakopoulos AG, Covaci A, Gevaio B, Johnson-Restrepo B, et al. Organophosphate esters in indoor dust from 12 countries: concentrations, composition profiles, and human exposure. *Environ Int.* (2019) 133:105178. doi: 10.1016/j.envint.2019.105178
16. Xu F, Eulaers I, Alves A, Papadopoulou E, Padilla-Sanchez JA, Lai FY, et al. Human exposure pathways to organophosphate flame retardants: associations between human biomonitoring and external exposure. *Environ Int.* (2019) 127:462–72. doi: 10.1016/j.envint.2019.03.053
17. Van den Eede N, Erratico C, Exarchou V, Maho W, Neels H, Covaci A. *In vitro* biotransformation of tris(2-butoxyethyl) phosphate (TBOEP) in human liver and serum. *Toxicol Appl Pharmacol.* (2015) 284:246–53. doi: 10.1016/j.taap.2015.01.021
18. Hoffman K, Lorenzo A, Butt CM, Adair L, Herring AH, Stapleton HM, et al. Predictors of urinary flame retardant concentration among pregnant women. *Environ Int.* (2017) 98:96–101. doi: 10.1016/j.envint.2016.10.007
19. Hoffman K, Butt CM, Chen A, Limkakeng AT Jr, Stapleton HM. High exposure to organophosphate flame retardants in infants: associations with baby products. *Environ Sci Technol.* (2015) 49:14554–9. doi: 10.1021/acs.est.5b03577
20. Hoffman K, Butt CM, Webster TF, Preston EV, Hammel SC, Makey C, et al. Temporal trends in exposure to organophosphate flame retardants in the United States. *Environ Sci Technol Lett.* (2017) 4:112–8. doi: 10.1021/acs.estlett.6b00475
21. Tsai KE, Cheng FJ, Huang WT, Kung CT, Lee CT, Cheng BC, et al. The associations between renal disease severity and exposure to organophosphate flame retardants in patients with chronic kidney disease. *Environ Int.* (2022) 170:107573. doi: 10.1016/j.envint.2022.107573
22. Rauer C, Schuster JK, Eng A, Harner T. Global atmospheric concentrations of brominated and chlorinated flame retardants and organophosphate esters. *Environ Sci Technol.* (2018) 52:2777–89. doi: 10.1021/acs.est.7b06239
23. Zhou L, Hiltcher M, Gruber D, Puttmann W. Organophosphate flame retardants (OPFRs) in indoor and outdoor air in the Rhine/Main area, Germany: comparison of concentrations and distribution profiles in different microenvironments. *Environ Sci Pollut Res Int.* (2017) 24:10992–1005. doi: 10.1007/s11356-016-6902-z
24. Wong F, de Wit CA, Newton SR. Concentrations and variability of organophosphate esters, halogenated flame retardants, and polybrominated diphenyl ethers in indoor and outdoor air in Stockholm. *Sweden Environ Pollut.* (2018) 240:514–22. doi: 10.1016/j.envpol.2018.04.086
25. Mizouchi S, Ichiba M, Takigami H, Kajiwara N, Takamuku T, Miyajima T, et al. Exposure assessment of organophosphorus and organobromine flame retardants via indoor dust from elementary schools and domestic houses. *Chemosphere.* (2015) 123:17–25. doi: 10.1016/j.chemosphere.2014.11.028
26. He C, Wang X, Thai P, Baduel C, Gallen C, Banks A, et al. Organophosphate and brominated flame retardants in Australian indoor environments: levels, sources, and preliminary assessment of human exposure. *Environ Pollut.* (2018) 235:670–9. doi: 10.1016/j.envpol.2017.12.017
27. Fan X, Kubwabo C, Rasmussen PE, Wu F. Simultaneous determination of thirteen organophosphate esters in settled indoor house dust and a comparison between two sampling techniques. *Sci Total Environ.* (2014) 491–492:80–6. doi: 10.1016/j.scitotenv.2013.12.127
28. He C, Wang X, Tang S, Thai P, Li Z, Baduel C, et al. Concentrations of organophosphate esters and their specific metabolites in food in Southeast Queensland, Australia: is dietary exposure an important pathway of organophosphate esters and their metabolites? *Environ Sci Technol.* (2018) 52:12765–73. doi: 10.1021/acs.est.8b03043
29. Trowbridge J, Gerona R, McMaster M, Ona K, Clarity C, Bessonneau V, et al. Organophosphate and Organohalogen flame-retardant exposure and thyroid hormone disruption in a cross-sectional study of female firefighters and office workers from San Francisco. *Environ Sci Technol.* (2022) 56:440–50. doi: 10.1021/acs.est.1c05140
30. Clarity C, Trowbridge J, Gerona R, Ona K, McMaster M, Bessonneau V, et al. Associations between polyfluoroalkyl substance and organophosphate flame retardant exposures and telomere length in a cohort of women firefighters and office workers in San Francisco. *Environ Health.* (2021) 20:97. doi: 10.1186/s12940-021-00778-z
31. Mayer AC, Fent KW, Chen IC, Sammons D, Toennis C, Robertson S, et al. Characterizing exposures to flame retardants, dioxins, and furans among firefighters responding to controlled residential fires. *Int J Hyg Environ Health.* (2021) 236:113782. doi: 10.1016/j.ijheh.2021.113782
32. Estill CF, Mayer A, Slone J, Chen IC, Zhou M, La Guardia MJ, et al. Assessment of triphenyl phosphate (TPhP) exposure to nail salon workers by air, hand wipe, and urine analysis. *Int J Hyg Environ Health.* (2021) 231:113630. doi: 10.1016/j.ijheh.2020.113630
33. Kim UJ, Wang Y, Li W, Kannan K. Occurrence of and human exposure to organophosphate flame retardants/plasticizers in indoor air and dust from various microenvironments in the United States. *Environ Int.* (2019) 125:342–9. doi: 10.1016/j.envint.2019.01.065
34. Wei B, Goniewicz ML, O'Connor RJ, Travers MJ, Hyland AJ. Urinary metabolite levels of flame retardants in electronic cigarette users: a study using the data from NHANES 2013–2014. *Int J Environ Res Public Health.* (2018) 15:201. doi: 10.3390/ijerph15020201
35. Sun Y, Gong X, Lin W, Liu Y, Wang Y, Wu M, et al. Metabolites of organophosphate ester flame retardants in urine from Shanghai. *China Environ Res.* (2018) 164:507–15. doi: 10.1016/j.envres.2018.03.031
36. Butt CM, Congleton J, Hoffman K, Fang M, Stapleton HM. Metabolites of organophosphate flame retardants and 2-ethylhexyl tetrabromobenzoate in urine from paired mothers and toddlers. *Environ Sci Technol.* (2014) 48:10432–8. doi: 10.1021/es5025299
37. Ospina M, Jayatilaka NK, Wong LY, Restrepo P, Calafat AM. Exposure to organophosphate flame retardant chemicals in the U.S. general population: data from the 2013–2014 National Health and nutrition examination survey. *Environ Int.* (2018) 110:32–41. doi: 10.1016/j.envint.2017.10.001
38. Wang LM, Luo D, Li X, Hu LQ, Chen JX, Tu ZZ, et al. Temporal variability of organophosphate flame retardant metabolites in spot, first morning, and 24-h urine samples among healthy adults. *Environ Res.* (2021) 196:110373. doi: 10.1016/j.envres.2020.110373
39. Thomas MB, Stapleton HM, Dills RL, Violette HD, Christakis DA, Sathyanarayana S. Demographic and dietary risk factors in relation to urinary metabolites of organophosphate flame retardants in toddlers. *Chemosphere.* (2017) 185:918–25. doi: 10.1016/j.chemosphere.2017.07.015
40. He C, English K, Baduel C, Thai P, Jagals P, Ware RS, et al. Concentrations of organophosphate flame retardants and plasticizers in urine from young children in Queensland, Australia and associations with environmental and behavioural factors. *Environ Res.* (2018) 164:262–70. doi: 10.1016/j.envres.2018.02.040
41. Wei GL, Li DQ, Zhuo MN, Liao YS, Xie ZY, Guo TL, et al. Organophosphorus flame retardants and plasticizers: sources, occurrence, toxicity and human exposure. *Environ Pollut.* (2015) 196:29–46. doi: 10.1016/j.envpol.2014.09.012
42. Saquib Q, Al-Salem AM, Siddiqui MA, Ansari SM, Zhang X, Al-Khedhairi AA. Tris(2-butoxyethyl) phosphate (TBEP): a flame retardant in solid waste display hepatotoxic and carcinogenic risks for humans. *Chemosphere.* (2022) 296:133977. doi: 10.1016/j.chemosphere.2022.133977
43. Tajima S, Araki A, Kawai T, Tsuboi T, Ait Bamai Y, Yoshioka E, et al. Detection and intake assessment of organophosphate flame retardants in house dust in Japanese dwellings. *Sci Total Environ.* (2014) 478:190–9. doi: 10.1016/j.scitotenv.2013.12.121
44. Brandsma SH, de Boer J, van Velzen MJ, Leonards PE. Organophosphorus flame retardants (PFRs) and plasticizers in house and car dust and the influence of electronic equipment. *Chemosphere.* (2014) 116:3–9. doi: 10.1016/j.chemosphere.2014.02.036
45. Kwon B, Shin H, Moon HB, Ji K, Kim KT. Effects of tris(2-butoxyethyl) phosphate exposure on endocrine systems and reproduction of zebrafish (*Danio rerio*). *Environ Pollut.* (2016) 214:568–74. doi: 10.1016/j.envpol.2016.04.049
46. Ma Z, Tang S, Su G, Miao Y, Liu H, Xie Y, et al. Effects of tris (2-butoxyethyl) phosphate (TBOEP) on endocrine axes during development of early life stages of zebrafish (*Danio rerio*). *Chemosphere.* (2016) 144:1920–7. doi: 10.1016/j.chemosphere.2015.10.049
47. Huang Y, Liu J, Yu L, Liu C, Wang J. Gonadal impairment and parental transfer of tris (2-butoxyethyl) phosphate in zebrafish after long-term exposure to environmentally relevant concentrations. *Chemosphere.* (2019) 218:449–57. doi: 10.1016/j.chemosphere.2018.11.139
48. Xiong H, Huang Y, Mao Y, Liu C, Wang J. Inhibition in growth and cardiotoxicity of tris (2-butoxyethyl) phosphate through down-regulating Wnt signaling pathway in early developmental stage of zebrafish (*Danio rerio*). *Ecotoxicol Environ Saf.* (2021) 208:111431. doi: 10.1016/j.ecoenv.2020.111431
49. Jiang F, Liu J, Zeng X, Yu L, Liu C, Wang J. Tris (2-butoxyethyl) phosphate affects motor behavior and axonal growth in zebrafish (*Danio rerio*) larvae. *Aquat Toxicol.* (2018) 198:215–23. doi: 10.1016/j.aquatox.2018.03.012
50. European Commission (EC). (2014). Available at: <https://eur-lex.europa.eu/legal-content/EN/TXT/PDF/?uri=CELEX:32014L0079&from=CS>
51. Ta N, Li C, Fang Y, Liu H, Lin B, Jin H, et al. Toxicity of TDCPP and TCEP on PC12 cell: changes in CAMKII, GAP43, tubulin and NE-H gene and protein levels. *Toxicol Lett.* (2014) 227:164–71. doi: 10.1016/j.toxlet.2014.03.023

52. Saboori AM, Lang DM, Newcombe DS. Structural requirements for the inhibition of human monocyte carboxylesterase by organophosphorus compounds. *Chem Biol Interact.* (1991) 80:327–38. doi: 10.1016/0009-2797(91)90092-1
53. Camarasa JG, Serra-Baldrich E. Allergic contact dermatitis from triphenyl phosphate. *Contact Dermatitis.* (1992) 26:264–5. doi: 10.1111/j.1600-0536.1992.tb00241.x
54. Liu X, Ji K, Jo A, Moon HB, Choi K. Effects of TDCPP or TPP on gene transcriptions and hormones of HPG axis, and their consequences on reproduction in adult zebrafish (*Danio rerio*). *Aquat Toxicol.* (2013) 134–135:104–11. doi: 10.1016/j.aquatox.2013.03.013
55. Kojima H, Takeuchi S, Itoh T, Iida M, Kobayashi S, Yoshida T. *In vitro* endocrine disruption potential of organophosphate flame retardants via human nuclear receptors. *Toxicology.* (2013) 314:76–83. doi: 10.1016/j.tox.2013.09.004
56. Zhang Q, Lu M, Dong X, Wang C, Zhang C, Liu W, et al. Potential estrogenic effects of phosphorus-containing flame retardants. *Environ Sci Technol.* (2014) 48:6995–7001. doi: 10.1021/es5007862
57. Du Z, Zhang Y, Wang G, Peng J, Wang Z, Gao S. TPhP exposure disturbs carbohydrate metabolism, lipid metabolism, and the DNA damage repair system in zebrafish liver. *Sci Rep.* (2016) 6:21827. doi: 10.1038/srep21827
58. Meeker JD, Stapleton HM. House dust concentrations of organophosphate flame retardants in relation to hormone levels and semen quality parameters. *Environ Health Perspect.* (2010) 118:318–23. doi: 10.1289/ehp.0901332