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# Histamine and histamine receptors: Roles in major depressive disorder

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Although the incidence of major depressive disorder (MDD) is high and its social impact is great, we still know very little about the pathophysiology of depression. The monoamine hypothesis of depression suggests that 5-HT, NE, and DA synergistically affect mood, which is the basis of current drug therapy for depression. However, histamine as a monoamine transmitter is rarely studied. Our review is the first time to illustrate the effect of histaminergic system on depression in order to find the way for the development of new antidepressant drugs. The brain neurotransmitter histamine is involved in MDD, and the brain histaminergic system operates through four receptors. Histamine and its receptors can also regulate the immune response to improve symptoms of depression. In addition, H3R can interact with other depression-related transmitters (including 5-HT, DA, GLU, and MCH); thus, histamine may participate in the occurrence of depression through other neural circuits. Notably, in rodent studies, several H3R and H1R antagonists were found to be safe and effective in alleviating depression-like behavior. To highlight the complex functions of histamine in depression, and reveals that histamine receptors can be used as new targets for antidepressant therapy.

## KEYWORDS

histamine, histamine receptors, immune regulation, major depressive disorder, therapy

## Introduction

Major depressive disorder (MDD) is a common and serious illness that affected more than 200 million people in 2017 and is estimated to have the second highest global disease burden by 2020 (1). Many factors, including heredity, neurotransmitters, immunity, oxidation, and the inflammatory system, are involved in the pathophysiology of MDD (2, 3). However, the pathological mechanism of MDD is not yet clear. In addition, current treatments for depression do not effectively or adequately reduce the

associated morbidity and mortality. In fact, as many as 50% of MDD patients do not experience complete remission after receiving antidepressant medication (4). These data highlight the importance of depressive disorder as a priority for public health projects and the importance of effective interventions to alleviate this burden.

Neuromodulation system dysfunction is common in depression. Based on the monoamine hypothesis of depression, monoamine reuptake inhibitors have been developed into antidepressants and have recently been widely used in the clinic (5). The relationship among serotonin, norepinephrine and dopamine (DA) in depression has been extensively studied, but the role of histamine, which is also a monoamine transmitter, is poorly understood (6). Histamine and its receptors were originally described as part of the immune and gastrointestinal systems, but their presence in the central nervous system (CNS) and importance in behavior are receiving increasing attention in CNS diseases, such as Alzheimer's disease, depression, sleep disorders, drug dependence disorders, and Parkinson's disease (6).

All effects of histamine are regulated by the G protein-coupled receptors the H1 receptor (H1R), H2R, H3R, and H4R (7). H1R, H2R, and H3R can be found in the CNS, where they act to regulate multiple physiological functions, such as energy homeostasis, cognition and attention, sensory and motor functions, and so on (8). Moreover, two of the histamine receptors are used in the clinical treatment of mental diseases. H1R is related to sleep, and H1R antagonists can be used to treat chronic insomnia (9). H3R is involved in cognition and attention, and H3R agonists can be used to treat alcohol addiction (10), Alzheimer's disease (11) and narcolepsy (12). To date, few studies have addressed the relationship between histamine and depression, and this direct relationship is unclear. This review discusses the role and involvement of histamine in MDD, with a focus on histamine receptors, and includes an examination of immune responses and the interaction between histamine and other transmitters. In addition, this review highlights the activity of histamine receptor agonists and antagonists as potential antidepressants in basic and preclinical studies. This review focuses primarily on recently published data; however, data from early studies are also offered to highlight the various developmental stages of the histamine theory in depression.

## The neurotransmitters related to depression

Stress plays a fundamental role in many causes of depression. Stress activates the HPA axis and causes the hypothalamus to release corticotropin-releasing

hormone, which acts on the anterior pituitary and releases adrenocorticotrophic hormone and finally triggers the adrenal cortex to release glucocorticoid and monoamine transmitters. MDD is associated with the deficiency of monoamines, especially 5-hydroxytryptamine (5-HT), noradrenaline (NA), and dopamine (DA), in the brain (13). The treatment of affective disorders is primarily based on the enhancement of the noradrenergic and serotonergic systems by selective or non-selective reuptake inhibitors. In the dopaminergic system, impaired neurotransmission is also associated with depression, and some preclinical studies have shown that agonists of dopaminergic receptors (D1R, D2R, and D3R, respectively) produce an antidepressant effect (14). Moreover, pure dopaminergic drugs such as pramipexole, DA precursors and DA reuptake inhibitors have been shown to have a therapeutic effect on depression (15).

Importantly, in addition to monoamines, other neurotransmitters, such as the glutamatergic or gamma-aminobutyric acid (GABA)ergic melanin-concentrating hormone (MCH), are involved in the pathophysiology of depression (16–20). Abnormal glutamatergic neurotransmission is involved in the development of mental illnesses, including schizophrenia, bipolar disorder and depression (16). Glutamate (GLU) receptors are classified as N-methyl-D-aspartate (NMDA), alpha-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) and metabotropic receptors (16). Among these classes, the NMDA receptor (NMDAR) is considered to be the main target for the development of a new generation of antidepressant drugs (16, 17). In fact, NMDAR antagonists such as ketamine can alleviate depressive symptoms in patients within hours of administration (18, 19). MCH is a 19-amino-acid cyclic peptide whose biological function is mediated by two G protein-coupled receptors called MCH receptor 1 (MCHR-1) and MCHR-2 (20). Preclinical studies suggest that the MCHergic system is involved in depression. Intra-locus coeruleus (LC) and intracerebroventricular injections of MCH produce depressive-like behavior, but this effect can be blocked by pretreatment with the MCH-R1 antagonist SNAP-94847 in the LC (21). In a clinical study, MCH serum levels were significantly decreased in patients with MDD after 4 weeks of antidepressant treatment (22). Overall, preclinical studies have demonstrated that antagonism of the MCHergic system may be a new treatment approach for antidepressant drugs.

Monoamine transmitters such as 5-HT, NA, and DA and non-monoamine transmitters such as GABA, NMDA, and MCH have been shown to be associated with the pathophysiology of depression. As a monoamine transmitter, histamine can interact with these transmitters and may also play an important role in depression, but research on this role is scarce.

## The function of histamine and the histamine receptor in the brain

Histamine plays an important role as a neurotransmitter in the brain, and its receptors have been shown to regulate many physiological functions that are affected in numerous CNS diseases (7, 8, 23) (Table 1).

The H1 receptor (H1R) is expressed in the CNS, lungs, and blood vessels (24). In the CNS, the main signaling pathway of H1R activates phospholipase C and protein kinase C (PKC) and catalyzes Ser/Thr phosphorylation of various downstream effectors, which subsequently induces intracellular  $Ca^{2+}$  release (25). H1R can activate most neurons in most brain regions, such as the brain stem, hypothalamus, hippocampus, thalamus, olfactory bulb, amygdala, septum, and cortex, by increasing  $Ca^{2+}$  (26). H1R participates in the regulation of motor function, mood, arousal, sleep and circadian rhythms, cognitive function, and pain perception (24). Moreover, H1R is involved in the modulation of several brain-controlled physiological functions, such as thermoregulation, food intake and energy consumption (24, 26). Blocking H1R with antagonists can increase the sensitivity of the CNS to seizures and sedation (24, 27). In addition, H1R antagonists can cross the blood-brain barrier (BBB) and cause drowsiness and are occasionally used to treat insomnia (9).

H2R is expressed in the stomach, CNS and heart (7, 24). Activation of H2R can stimulate the adenylyl cyclase, which in turn synthesizes cyclic adenosine monophosphate (cAMP). The cAMP then activates protein kinase A and the transcription factor cAMP response element-binding protein, which regulate neuronal physiology and plasticity (25, 28). In the human brain, H2R is widely distributed in the basal ganglia, hippocampus, amygdala, and cerebral cortex (29, 30). The function of H2R in the brain includes the modulation of circadian rhythm, cognitive processes, and food intake and glucose metabolism (24). Although H2R mediates important functions, and reports of CNS toxicity are infrequent, therapeutic applications for CNS-permeable ligands are relatively rare to date. Several studies have highlighted the potential of H2R antagonists to enhance the effects of opiate analgesics (31) and to exert a positive therapeutic effect on schizophrenia (32).

H3R is abundantly expressed in important areas of the brain, including the cerebral cortex, striatum, and hippocampus (29, 33, 34). H3R is characterized as an auto- and heteroreceptor that can regulate the synthesis and release of histamine or regulate the release of other neurotransmitters to influence the balance of different neurotransmitters (35, 36). Activation of Gi/o proteins by H3R results in the activation of mitogen-activated protein kinase (MAPK) pathways (36, 37). H3R regulates numerous behaviors, such as pain perception, locomotor activity, memory, food intake, circadian rhythms, anxiety, and cognition (38). Moreover, H3R antagonists can reduce alcohol intake in alcohol-preferring rats (10, 38), and inverse H3R agonists are

used in clinical trials for the treatment of Alzheimer's disease (11) and narcolepsy (12).

H4R was the last histamine receptor to be identified and is classified as a Gi/o-coupled GPCR (38, 39). H4R is mainly related to immune function and is expressed on immune cells, including eosinophils, mast cells and dendritic cells, where it induces calcium mobilization and chemotaxis (7, 40). However, the expression and function of H4R in the CNS remain controversial and require further research. In the human brain, H4R mRNA was detected in the amygdala, hippocampus, thalamus, cortex and spinal cord amygdala (41). By contrast, another report showed the opposite result, indicating that H4R may not exhibit functional expression on neurons in the CNS; thus, H4R may not be present on neurons in the brain but may be present on brain microglia, endothelial cells or epithelial cells (42).

H1R, H2R, and H3R are all expressed in the CNS and play physiological roles which are associated with depressive symptoms. The presence of H4R in the CNS remains controversial, but studies have linked it to immune function, and a growing body of research supports the immune inflammatory response to depression. Recent advances in our knowledge and understanding of histamine pharmacology, along with the identification and structural understanding of histamine receptors over the past decade, indicate the potential of new, improved antihistamines for future clinical development. Thus, there is still much to understand about the functional and pharmacological selectivity of these four receptors.

## The role of the histaminergic system in the pathophysiology of depression

### Histamine and the histamine receptor in brain areas relevant to depression

The histaminergic neurons in the mammalian brain are located in the tuberomammillary nucleus (TMN) of the posterior hypothalamus, which sends projection signals to all major parts of the brain (43), especially the areas important for cognitive function, such as the hippocampus, frontal cortex, basal forebrain and amygdala (44).

Brain histamine can confer biological protective effects against various unfavorable phenomena, such as denervation hypersensitivity, convulsions, stress susceptibility, ischemic injury and drug sensitization. A postmortem study of anterior cingulate cortex (ACC) and dorsolateral prefrontal cortex (DLPFC) tissues from individuals with depressive disorder bipolar disorder and control individuals evaluated the expression of histamine receptors and the enzymatic breakdown of histamine and histamine N-methyltransferase (HMT), indicating no change in the expression of these molecules

TABLE 1 Overview of histamine receptors in central nervous system (CNS).

	H1R	H2R	H3R	H4R
G protein coupling	Gaq	Gas	Gai/o	Gai/o
Signal	Activate PLC, PKC, Ser/Thr and calcium release (25)	Increase cAMP and activates PKA (25, 28)	Activate MAPK, PKB and calcium release (36)	Activate MAPK, PKB, calcium release and (7, 42)
Location	CNS, lungs, and blood vessels (24)	Stomach, CNS and heart (7, 24)	CNS (37)	Immune cells (eosinophils, mast cells and dendritic cells) (7, 40)
CNS areas	Brain stem, thalamus, hypothalamus, septum, hippocampus, olfactory bulb amygdala and cortex (26).	Basal ganglia, hippocampus, amygdala, and cerebral cortex (29, 30).	Cerebral cortex, striatum, and hippocampus (29, 33, 34)	Not exist or amygdala, thalamus, cortex, hippocampus, amygdala and spinal cord (41, 42)
CNS function	Motor function, mood, arousal, sleep, cognitive function, pain perception, circadian rhythm, food intake thermoregulation and energy consumption (24, 26)	Circadian rhythm, cognitive process, food intake and glucose metabolism (24)	Pain perception, locomotor activity, circadian rhythm, memory, anxiety, food intake and cognition (38)	Immune function

except for a significant decrease in HMT mRNA expression in the ACC of individuals with MDD (45). In addition, a clinical study showed that the histamine level in 184 first-episode adolescent patients was significantly higher than that in the controls (46). The inconsistent results between peripheral and central histamine studies are due to different measurement methods and BBB barrier effects. The BBB is the primary barrier to the CNS and prevents the influx of active substances from the surrounding circulatory system. The BBB is closely associated with MDD and other mental disorders, such as Alzheimer's and Parkinson's diseases. Histamine has been shown to be a major factor leading to high BBB permeability (47). Histamines generally do not pass through the BBB; however, histamine induces an increase in BBB permeability, which in turn promotes histamine entry into the CNS. Previous reports have supported this view by suggesting that people with various neurodegenerative diseases have increased levels of histamine in their cerebrospinal fluid and brain parenchyma (47, 48). However these studies may indicate a change in histaminergic system of patients with depression, but more clinical researches are needed to confirm the role of histamine in the development and treatment of depression.

Not only histamine, but also histamine receptors are differentially expressed in patients with depression. Positron emission tomography (PET) studies in humans have shown that histaminergic nervous system dysfunction, especially H1R dysfunction, is associated with depression (49). H1R binding in the prefrontal and frontal cortices and the cingulate gyrus was significantly decreased in patients with depression, and this reduction is associated with the severity of depression symptoms (50). H3R, which can regulate the release of histamine as an autoreceptor or regulate the release of other neurotransmitters as part of a heteroreceptor, is located on the dendrites and axons of histaminergic neurons (44). The H3 autoregulatory system seems to be linked to behavioral changes caused by stress,

decreased sleep, inflammation, and oxidative stress. *In vitro* experiments have suggested that some of the beneficial effects of H3R antagonists on mood and cognition may be mediated through H4R (51). A recent study showed that H4R knockout mice had depression-like symptoms and cognitive impairment, but the study did not provide any mechanistic insights (52). A recent study provided the first description of the behavioral phenotype of H4R-deficient (H4R knockout) mice, which finally showed that H4R modulates various symptoms of depression, such as locomotor activity, anxiety, depression and feeding behavior (52). Both human and animal studies have found that the expression of histamine receptor is different between depressed patients and normal control subjects and between animal models of depression and related control animals, which is related to depression symptoms. Further animal studies have found that depression-like symptoms can be caused by knocking down the H4 receptor, and be improved by using an H3 receptor antagonist (51, 52).

Although none of depression's symptoms are directly related to the histaminergic neuronal system, many brain regions show high levels of histamine and histamine receptors which can be used as indirect evidence that depression is related to histamine (45, 46, 49, 50).

## Interactions between histamine and other transmitters related to depression

Monoamine transmitters such as 5-hydroxytryptamine (5-HT), noradrenaline (NA), and dopamine (DA), and non-monoamine transmitters such as the GABA, N-methyl-D-aspartate (NMDA) and ergic MCH have been shown to be associated with the pathophysiology of depression (13, 16, 17, 21). As a monoamine transmitter, histamine interacts with these

transmitter systems to form a network that mediates sleep-wake regulation, circadian and eating rhythms, immunity, learning, attention, motor activity, and memory (53, 54) (Figure 1).

Histamine released from brain mast cells may modulate the mood state *via* interactions with other transmitters, including serotonin and NA. Mast cell-deficient mice exhibited higher anxiety and depression levels than wild-type mice, and the symptoms were inhibited by injection of imipramine, an inhibitor of serotonin and NA uptake (55). H3R inhibits 5-HT release in the substantia nigra pars reticulata (SNr); this effect is attributed to its direct action on 5-HT terminals (56). Histamine acting through H3R tonically hyperpolarizes 5-HT neurons and inhibits 5-HT signaling, and when this inhibition is relieved by H3R antagonism, 5-HT neuron excitability is increased (57). The observed H3R-mediated endogenous control of 5-HT neurons provides even more compelling evidence that H3R is a candidate target for antidepressant drugs (57).

H3R antagonists have been shown to regulate the dopaminergic signaling pathway, and in the striatum, DA receptor agonists increase DA receptor signaling in wild-type mice but not in H3R knockout mice (34). A recent study investigated the existence of D1R-H3R heteromers, which constitute unique devices that can direct dopaminergic and histaminergic signaling toward the MAPK pathway in a Gs-independent and Gi-dependent manner (58). An imbalance between the histamine regulatory function (decreased) and DA (increased *via* D1R) was demonstrated in the striatum of mice with congenital hyperammonemia, providing a theoretical basis for the development of aminergic wake-promoting therapeutics to treat hyperammonemic disorders (59). The D1R-H3R heteromer is expressed in the brain, and it can selectively link GABAergic neurons and MAPK signals in the direct striatal pathway. These results indicate that the D1R-H3R heteromer can integrate DA- and histamine-related signaling involved in the control of the direct striatal pathway in striatal neurons (36). Moreover, the formation of heteromers by D1R, H3R, and NR1A-NR2B NMDARs allows close contact between D1R, H3R and the two NMDAR subunits. Structural changes in the entire macromolecular complex by H3R antagonists appear to reduce NMDA or D1 receptor-mediated excitotoxic cell death in cortical cultures, and the results indicate that H3R antagonists have neuroprotective potential, and H3R in the D1R-H3R-NMDAR heterocomplex is expected to be a target for preventing neurodegeneration (60).

Melanin-concentrating hormone neurons express H3R but not H1R or H2R (61). Histamine directly inhibits MCH neurons through H3R by activating G protein-dependent inwardly rectifying potassium (GIRK) channels (61). This observation strongly suggests that histamine and MCH cooperate in the regulation of arousal and sleep. In addition, histamine can inhibit MCH through H3R (61). MCH is associated with depression because injecting MCH into the dorsal raphe

nucleus causes a depressive phenotype in rats, possibly because MCH inhibits serotonergic neurons, and MCHR-1 antagonists produce an antidepressant effect (62). Thus, high levels of histamine reduce sleep through the activation of H3R in the MCH system, and low levels of histamine may be associated with increased MCH activity and depressive behavior. A decrease in the histamine level may lead to depression by increasing MCH and histamine-derived functions such as physical activity, arousal, cognitive function, etc.

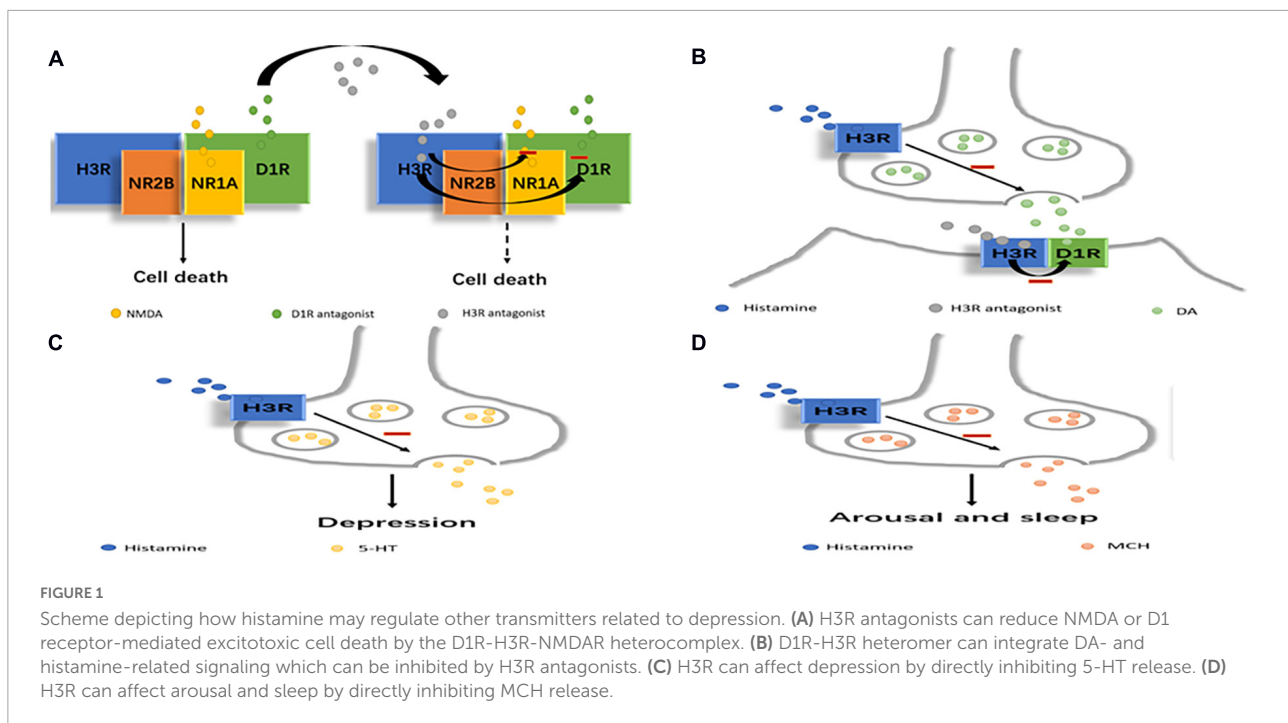
Since these H3R-expressing cells release many neurotransmitters regulated by H3R and the interactions among them, the role of H3R antagonists in depression may result from circuits involving many of these transmitters.

## Histamine regulates the immune system to ameliorate depression

Neuroimmune dysregulation is ubiquitous in different types of CNS diseases, and histamine is a pleiotropic monoamine that is involved in various neurophysiological functions and is a powerful regulator of the neuroimmune system (63, 64). An increasing amount of data have suggested that the immune system, including the cytokine network and innate immune response, plays a potential role in the etiology and pathophysiology of MDD (65–67). However, the detailed mechanism of immunity in depression has not been fully elucidated. A 6-year follow-up longitudinal study of 2,981 MDD patients (68) showed the associations of diagnosis and symptoms of depression with the interleukin-6 (IL-6) level, along with the ability of the IL-6 level to predict current MDD and greater symptom severity at follow-up. These results highlight the importance of investigating IL-6 as a potential marker. Twenty-five patients with unipolar MDD and 34 patients with schizophrenia, along with 50 healthy controls, were assessed for the presence of non-specific (auto-) immune antibodies, and significant differences were found, revealing the existence of a non-specific autoimmune disposition or reaction in at least a subgroup of patients with major depression and schizophrenia (69). Data from patients demonstrated a significant increase in peripheral urinary histamine levels in the group with depression compared with the control group, and the increased histamines further regulate the effects of inflammation (70).

Experiments with knockout mice revealed that H1R inhibits Th2 cell polarization, that H2R promotes Th1- and Th2-mediated immune responses, and that H4R affects innate and adaptive immunity *via* anti- or proinflammatory functions depending on the specific disease studied (40). In addition, H3R was involved in the inflammatory response only in the autoimmune encephalomyelitis model (71). Thus, the published studies highlight the complexity of histamine function in inflammation and the immune system.





Recently, all four histamine receptors have been shown to be expressed on immune cell microglia in the brain (72, 73). Histamine can stimulate microglial activation and the subsequent production of the proinflammatory factors tumor necrosis factor (TNF)-alpha and IL-6 *via* H1R and the H4R-MAPK and PI3K/AKT-NF-kappa B signaling pathways (72). Moreover, histamine induces microglial migration through H4R and inhibits lipopolysaccharide (LPS)-induced interleukin-1 beta (IL-1b) production (73). A selective and potent H3R inverse agonist suppressed microglial activation, chemotaxis and phagocytosis to ameliorate LPS-induced depression-like behavior in mice (74). Thus, histamine can indirectly affect neuronal function by influencing microglia and affecting inflammation. An H1R antagonist modulated the expression of the P2 × 7 receptor and suppressed microglial M1-like activation and production of proinflammatory cytokines, including IL-1b and TNF-a, in the hippocampus, thereby alleviating depressive-like behavior in mice (75). These results suggest that histamine plays an important role in neuroinflammation-related diseases, including depression, *via* microglia.

Interestingly, mast cells, which can promote inflammation and allergic reactions *via* the release of histamine, are also present in the brain. Mast cell degranulation and histamine release often promote awareness and positive behavior. For example, compared with wild-type mice, mast cell-deficient mice have increased delta power and reduced food seeking-motivated behavior; however, these mice also have more anxiety and depression than wild-type mice (55).

Recent research on histamine suggests that we need to reassess the role of histamine in the innate and adaptive immune responses in depression. However, the effect of differential expression or activation of histamine receptors on immune cells in depression has not been fully described, and further examination of this effective immunoregulatory network may lead to a better understanding of depression.

## Therapeutic prospects of histamine receptors antagonists for depression

Histamine and its receptors play an important role in the pathophysiology of depression. Recently, some studies have investigated antidepressant drugs related to histamine receptors (Table 2). Rodent studies revealed that acute and chronic treatment with an H3R antagonist can reduce depression-like conditions and that chronic H3R antagonism (*via* ciproxifan) alleviates depression-like conditions in mice *via* the modulation of stress-induced biochemical correlates such as BDNF, corticosterone, NUCB2/nesfatin-1, and CRH in the brain (51, 76, 77). The H3R antagonist clobenpropit reduces the activity of presynaptic H3Rs, increases histamine release, activates hippocampal H1R and H2R, and effectively ameliorates depression-like behavior in FSL mice. Its antidepressant effect can be blocked by H1R and H2R antagonists (H2R-ANTS) (77). In addition, systemic or hippocampal injection of clobenpropit reversed motor and cognitive impairment in a depressed rat model in a manner dependent on the release of histamine and the actions of H1R and H2R (78).

TABLE 2 Therapeutic prospect with histamine receptor for depression.

Ligand (Reference)	HR	Animal model	Administered (dose)	Function	Outcome
Ciproxifan (51, 76, 77)	H3R antagonist	Chronic unpredictable stress (CUS) model of depression in C57BL/6 J mice	Intraperitoneal injections (3 mg/kg.; for 3 weeks)	Modulate BDNF, corticosterone, NUCB2/nesfatin-1, and CRH in the brain	Alleviate depression like condition
Clobenpropit (77, 78)	H3R antagonist	Sprague-Dawley Rats or male FSL rats	Subcutaneously injections (5 mg/kg); hippocampal delivery, bilateral guide cannula (10 mM, 15 min);	Increases histamine release, activate hippocampal H1R and H2R	Improve depression-like behavior, reversed the motor and cognitive impairment
JNJ10181457 (JNJ) (74)	H3R inverse agonist	CX3C chemokine receptor 1 (CX3CR1)-green fluorescent protein	Intraperitoneal injections (10 mg/kg.; for 3 days)	Suppress microglial chemotaxis, phagocytosis	Improve depression-like behavior
3 h (3,5-dimethyl-isoxazole-4-carboxylic acid) (80)	H3R antagonist/inverse agonist	Male OF1 mice and male Sprague-Dawley rats	Oral (3 and 10 mg/kg)	Unknown	Improve depression-like behavior
Clemastine (75)	First-generation H1R antagonist	(GFP) mice were injected with LPS (1 mg/kg)	Intraperitoneal injections (10 mg/kg.; for 3 days)	Reduce inflammation	Alleviate stress-related depressive-like behavior

The H3R antagonist pitolisant can reduce olanzapine-induced depression-like symptoms in mice (79). The selective and potent H3R inverse agonist JNJ10181457 (JNJ) suppressed microglial chemotaxis and phagocytosis *in vivo* and suppressed microglial activation and improved depression-like behavior *ex vivo* in mice (74). The H3R antagonist/inverse agonist 3,5-dimethyl-isoxazole-4-carboxylic acid (3 h) was effective in the forced swimming test, suggesting its potential therapeutic effect as an antidepressant. After assessment of cardiovascular and neuropsychological/behavioral safety, the compound was widely considered a preclinical drug candidate (80). A quantitative structure–activity relationship (QSAR) study of the serotonin reuptake-inhibitory and H3-antagonistic activity of piperazine and diazepane amide derivatives provided a rational approach for combination treatment with histamine H3 antagonists and serotonin reuptake inhibitors, which may improve not only depressive mood but also cognitive impairment and fatigue in depression (81).

It was suggested that clemastine, a first-generation H1R antagonist, could impressively alleviate stress-related depressive-like behavior in mice by reducing inflammation (75). Interestingly, treatment with a 28-day course of paroxetine increased the mRNA expression of H1R and histidine decarboxylase in the frontal cortex of mice (82). Although further studies are needed, the literature suggests that histaminergic systems are partially responsible for the pathogenesis of sleep disturbance induced by treatment with selective serotonin reuptake inhibitors (SSRIs) or serotonin-norepinephrine reuptake inhibitors (SNRIs) in rodents (82).

Preclinical studies demonstrated that SSRIs, such as citalopram and paroxetine, and the bioactive lipid mediator

oleoylethanolamide (OEA) [but not the tricyclic antidepressant (TCA) imipramine or the selective NA reuptake inhibitor reboxetine] specifically require the brain histamine system to exert their antidepressant-like effects (83, 84). However, a case report indicated that cetirizine, a second-generation H1R antagonist, can induce depression and suicidal behavior, but the symptoms abated with the cessation of cetirizine (85). A meta-analysis of H2R antagonist treatment combined with antipsychotic treatment in patients with schizophrenia showed that H2R antagonist adjuvant therapy did not improve psychotic symptoms but increased the risk of depression (only two comparisons with nizatidine) (86). On the other hand, antihistamines, which are mainly used to treat chronic pruritus, may affect mood and sleep quality. Outpatients treated with cetirizine reported higher depression scores than outpatients treated with desloratadine, levocetirizine, and rupatadine (87).

In summary numerous animal studies have shown that HR is a new target for antidepressant therapy and has the advantage of improving not only emotional symptoms but also cognition. Moreover, a small number of studies have found that H1R antagonists have antidepressant effects. In addition, histamine was found to be involved in the effects of currently available antidepressants (SSRIs). There are several successful clinical treatments: (1) The H1R antagonist, doxepin, can be used for the treatment of insomnia (88). Doxepin can also act as norepinephrine, serotonin reuptake inhibitor and 5-HT<sub>2</sub>-receptor antagonist to exert antidepressant effect. Its action on histamine is one of its properties beside its effects of norepinephrine and serotonin. (2) The H3R-antagonist/inverse agonist, pitolisant, for the treatment of narcolepsy and obstructive sleep apnea (89–91). But so far there is no clinical study about depression.

This finding was surprising and suggested questions about the potential mechanisms and clinical applications of histamine. Therefore, more research is needed to analyze the role of the histaminergic system in depression and its therapeutic significance.

## Conclusion and future directions

Histamine as a monoamine transmitter is rarely studied for the effect on depression, our review is the first time to illustrate the effect of histaminergic system on depression in order to find the way for the development of new antidepressant drugs. We suggest that histaminergic system may play a pivotal role in the pathogenesis of depression. Histamine and its receptors can regulate the immune response to improve symptoms of depression, and H3R can interact with other transmitters such as NMDA, DA, 5-HT, and MCH which are related to depression. Moreover, H3R and H1R antagonist can alleviate depression in rodent studies, but the clinical potential of these drugs has not been tested. Such research is clearly the next step to open new areas of application.

## Author contributions

GW and HQ conceived the idea of the study and designed the study. All authors analyzed data,

wrote the manuscript, discussed the results, and revised the manuscript.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## References

1. GBD 2017 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the global burden of disease study 2017. *Lancet*. (2018) 392:1789–858. doi: 10.1016/S0140-6736(18)32279-7
2. Beijers L, Wardenaar KJ, van Loo HM, Schoevers RA. Data-driven biological subtypes of depression: systematic review of biological approaches to depression subtyping. *Mol Psychiatry*. (2019) 24:888–900. doi: 10.1038/s41380-019-0385-5
3. Fox ME, Lobo MK. The molecular and cellular mechanisms of depression: a focus on reward circuitry. *Mol Psychiatry*. (2019) 24:1798–815. doi: 10.1038/s41380-019-0415-3
4. Thase ME, Nierenberg AA, Vrijland P, van Oers HJ, Schutte AJ, Simmons JH. Remission with mirtazapine and selective serotonin reuptake inhibitors: a meta-analysis of individual patient data from 15 controlled trials of acute phase treatment of major depression. *Int Clin Psychopharmacol*. (2010) 25:189–98. doi: 10.1097/YIC.0b013e328330adb2
5. Perez-Caballero L, Torres-Sanchez S, Romero-López-Alberca C, González-Saiz F, Mico JA, Berrocoso E. Monoaminergic system and depression. *Cell Tissue Res*. (2019) 377:107–13. doi: 10.1007/s00441-018-2978-8
6. Baronio D, Gonchoroski T, Castro K, Zanatta G, Gottfried C, Riesgo R. Histaminergic system in brain disorders: lessons from the translational approach and future perspectives. *Ann Gen Psychiatry*. (2014) 13:34. doi: 10.1186/s12991-014-0034-y
7. Seifert R, Strasser A, Schneider EH, Neumann D, Dove S, Buschauer A. Molecular and cellular analysis of human histamine receptor subtypes. *Trends Pharmacol Sci*. (2013) 34:33–58. doi: 10.1016/j.tips.2012.11.001
8. Haas H, Panula P. The role of histamine and the tuberomammillary nucleus in the nervous system. *Nat Rev Neurosci*. (2003) 4:121–30. doi: 10.1038/nrn1034
9. Wang YQ, Takata Y, Li R, Zhang Z, Zhang MQ, Urade Y, et al. Doxepin and diphenhydramine increased non-rapid eye movement sleep through blockade of histamine H1 receptors. *Pharmacol Biochem Behav*. (2015) 129:56–64. doi: 10.1016/j.pbb.2014.12.002
10. Panula P. Histamine, histamine H<sub>3</sub> receptor, and alcohol use disorder. *Br J Pharmacol*. (2019) 177:634–41. doi: 10.1111/bph.14634
11. Grove RA, Harrington CM, Mahler A, Beresford I, Maruff P, Lowy MT, et al. A randomized, double-blind, placebo-controlled, 16-week study of the H3 receptor antagonist, GSK239512 as a monotherapy in subjects with mild-to-moderate Alzheimer's disease. *Curr Alzheimer Res*. (2014) 11:47–58. doi: 10.2174/1567205010666131212110148
12. Dauvilliers Y, Bassetti C, Lammers GJ, Arnulf I, Mayer G, Rodenbeck A, et al. Pitolisant versus placebo or modafinil in patients with narcolepsy: a double-blind, randomised trial. *Lancet Neurol*. (2013) 12:1068–75. doi: 10.1016/S1474-4422(13)70225-4
13. Yohn CN, Gergues MM, Samuels BA. The role of 5-HT receptors in depression. *Mol Brain*. (2017) 10:28. doi: 10.1186/s13041-017-0306-y
14. Pytka K, Podkowa K, Rapacz A, Podkowa A, Żmudzka E, Olczyk A, et al. The role of serotonergic, adrenergic and dopaminergic receptors in antidepressant-like effect. *Pharmacol Rep*. (2016) 68:263–74. doi: 10.1016/j.pharep.2015.08.007
15. Belujon P, Grace AA. Dopamine system dysregulation in major depressive disorders. *Int J Neuropsychopharmacol*. (2017) 20:1036–46. doi: 10.1093/ijnp/pyx056



16. Li CT, Yang KC, Lin WC. Glutamatergic dysfunction and glutamatergic compounds for major psychiatric disorders: evidence from clinical neuroimaging studies. *Front Psychiatry*. (2018) 9:767. doi: 10.3389/fpsy.2018.00767
17. Pochwat B, Nowak G, Szewczyk B. An update on NMDA antagonists in depression. *Expert Rev Neurother*. (2019) 19:1055–67. doi: 10.1080/14737175.2019.1643237
18. Witkin JM, Knutson DE, Rodriguez GJ, Shi S. Rapid-acting antidepressants. *Curr Pharm Des*. (2018) 24:2556–63. doi: 10.2174/1381612824666180730104707
19. Dutta A, McKie S, Deakin J. Ketamine and other potential glutamate antidepressants. *Psychiatry Res*. (2015) 225:1–13. doi: 10.1016/j.psychres.2014.10.028
20. Saito Y, Nagasaki H. The melanin-concentrating hormone system and its physiological functions. *Results Probl Cell Differ*. (2008) 46:159–79. doi: 10.1007/160\_2007\_052
21. Ye H, Cui X, Ding H, Cui S, Hu X, Liu Y, et al. Melanin-concentrating hormone (MCH) and MCH-R1 in the locus coeruleus may be involved in the regulation of depressive-like behavior. *Int J Neuropsychoph*. (2018) 21:1128–37. doi: 10.1093/ijnp/pyy088
22. Schmidt FM, Nowak C, Kratzsch J, Sander C, Hegerl U, Schonknecht P. Dynamics of melanin-concentrating hormone (MCH) serum levels in major depressive disorder during antidepressant treatment. *J Affect Disord*. (2015) 180:207–13. doi: 10.1016/j.jad.2015.03.039
23. Shan L, Swaab DF. Changes in histaminergic system in neuropsychiatric disorders and the potential treatment consequences. *Curr Neuropharmacol*. (2022) 20:403–11. doi: 10.2174/1570159X19666210909144930
24. Schneider EH, Neumann D, Seifert R. Modulation of behavior by the histaminergic system: lessons from H(1)R- and H(2)R-deficient mice. *Neurosci Biobehav Rev*. (2014) 42:252–66. doi: 10.1016/j.neubiorev.2014.03.009
25. Panula P, Chazot PL, Cowart M, Gutzmer R, Leurs R, Liu WL, et al. International union of basic and clinical pharmacology. XCVIII. Histamine receptors. *Pharmacol Rev*. (2015) 67:601–55. doi: 10.1124/pr.114.010249
26. Tabarean IV. Functional pharmacology of H1 histamine receptors expressed in mouse preoptic/anterior hypothalamic neurons. *Br J Pharmacol*. (2013) 170:415–25. doi: 10.1111/bph.12286
27. Simons FE, Simons KJ. Histamine and H1-antihistamines: celebrating a century of progress. *J Allergy Clin Immunol*. (2011) 128:1139–50. doi: 10.1016/j.jaci.2011.09.005
28. Atzori M, Lau D, Tansey EB, Chow A, Ozaita A, Rudy B, et al. H2 histamine receptor-phosphorylation of Kv3.2 modulates interneuron fast spiking. *Nat Neurosci*. (2000) 3:791–8. doi: 10.1038/77693
29. Jin CY, Panula P. The laminar histamine receptor system in human prefrontal cortex suggests multiple levels of histaminergic regulation. *Neuroscience*. (2005) 132:137–49. doi: 10.1016/j.neuroscience.2004.12.017
30. Traiffort E, Pollard H, Moreau J, Ruat M, Schwartz JC, Martinez-Mir MI, et al. Pharmacological characterization and autoradiographic localization of histamine H2 receptors in human brain identified with [125I]iodoaminopotentidine. *J Neurochem*. (1992) 59:290–9. doi: 10.1111/j.1471-4159.1992.tb08903.x
31. Hasanein P. Two histamine H2 receptor antagonists, zolantidine and cimetidine, modulate nociception in cholestatic rats. *J Psychopharmacol*. (2011) 25:281–8. doi: 10.1177/0269881109106912
32. Meskanen K, Ekelund H, Laitinen J, Neuvonen PJ, Haukka J, Panula P, et al. A randomized clinical trial of histamine 2 receptor antagonism in treatment-resistant schizophrenia. *J Clin Psychopharmacol*. (2013) 33:472–8. doi: 10.1097/JCP.0b013e3182970490
33. Anichtchik OV, Peitsaro N, Rinne JO, Kalimo H, Panula P. Distribution and modulation of histamine H(3) receptors in basal ganglia and frontal cortex of healthy controls and patients with Parkinson's disease. *Neurobiol Dis*. (2001) 8:707–16. doi: 10.1006/nbdi.2001.0413
34. Kononoff VJ, Nuutinen S, Tuominen M, Panula P. Histamine h3 receptor regulates sensorimotor gating and dopaminergic signaling in the striatum. *J Pharmacol Exp Ther*. (2016) 357:264–72. doi: 10.1124/jpet.115.230771
35. Drutel G, Peitsaro N, Karlstedt K, Wieland K, Smit MJ, Timmerman H, et al. Identification of rat H3 receptor isoforms with different brain expression and signaling properties. *Mol Pharmacol*. (2001) 59:1–8.
36. Moreno E, Hoffmann H, Gonzalez-Sepulveda M, Navarro G, Casado V, Cortes A, et al. Dopamine D1-histamine H3 receptor heteromers provide a selective link to MAPK signaling in GABAergic neurons of the direct striatal pathway. *J Biol Chem*. (2011) 286:5846–54. doi: 10.1074/jbc.M110.161489
37. Giovannini MG, Efooude M, Passani MB, Baldi E, Bucherelli C, Giachi F, et al. Improvement in fear memory by histamine-elicited ERK2 activation in hippocampal CA3 cells. *J Neurosci*. (2003) 23:9016–23. doi: 10.1523/JNEUROSCI.23-27-09016.2003
38. Schneider EH, Neumann D, Seifert R. Modulation of behavior by the histaminergic system: lessons from HDC-, H3R- and H4R-deficient mice. *Neurosci Biobehav Rev*. (2014) 47:101–21. doi: 10.1016/j.neubiorev.2014.07.020
39. Schneider EH, Schnell D, Papa D, Seifert R. High constitutive activity and a G-protein-independent high-affinity state of the human histamine H(4)-receptor. *Biochemistry-US*. (2009) 48:1424–38. doi: 10.1021/bi802050d
40. Neumann D, Schneider EH, Seifert R. Analysis of histamine receptor knockout mice in models of inflammation. *J Pharmacol Exp Ther*. (2014) 348:2–11. doi: 10.1124/jpet.113.204214
41. Strakhova MI, Nikkel AL, Manelli AM, Hsieh GC, Esbenshade TA, Brioni JD, et al. Localization of histamine H4 receptors in the central nervous system of human and rat. *Brain Res*. (2009) 1250:41–8. doi: 10.1016/j.brainres.2008.11.018
42. Schneider EH, Seifert R. The histamine H4-receptor and the central and peripheral nervous system: a critical analysis of the literature. *Neuropharmacology*. (2016) 106:116–28. doi: 10.1016/j.neuropharm.2015.05.004
43. Fujita A, Bonnavion P, Wilson MH, Mickelsen LE, Bloit J, de Lecea L, et al. Hypothalamic tuberomammillary nucleus neurons: electrophysiological diversity and essential role in arousal stability. *J Neurosci*. (2017) 37:9574–92. doi: 10.1523/JNEUROSCI.0580-17.2017
44. Panula P, Nuutinen S. The histaminergic network in the brain: basic organization and role in disease. *Nat Rev Neurosci*. (2013) 14:472–87. doi: 10.1038/nrn3526
45. Shan L, Qi XR, Balesar R, Swaab DF, Bao AM. Unaltered histaminergic system in depression: a postmortem study. *J Affect Disord*. (2013) 146:220–3. doi: 10.1016/j.jad.2012.09.008
46. Tao R, Fu Z, Xiao L. Chronic food antigen-specific IgG-mediated hypersensitivity reaction as a risk factor for adolescent depressive disorder. *Genomics Proteomics Bioinformatics*. (2019) 17:183–9. doi: 10.1016/j.gpb.2019.05.002
47. Sedeyn JC, Wu H, Hobbs RD, Levin EC, Nagele RG, Venkataraman V. Histamine induces Alzheimer's disease-like blood brain barrier breach and local cellular responses in mouse brain organotypic cultures. *Biomed Res Int*. (2015) 2015:937148. doi: 10.1155/2015/937148
48. Najjar S, Pearlman DM, Devinsky O, Najjar A, Zagzag D. Neurovascular unit dysfunction with blood-brain barrier hyperpermeability contributes to major depressive disorder: a review of clinical and experimental evidence. *J Neuroinflammation*. (2013) 10:142. doi: 10.1186/1742-2094-10-142
49. Yanai K, Tashiro M. The physiological and pathophysiological roles of neuronal histamine: an insight from human positron emission tomography studies. *Pharmacol Ther*. (2007) 113:1–15. doi: 10.1016/j.pharmthera.2006.06.008
50. Kano M, Fukudo S, Tashiro A, Utsumi A, Tamura D, Itoh M, et al. Decreased histamine H1 receptor binding in the brain of depressed patients. *Eur J Neurosci*. (2004) 20:803–10. doi: 10.1111/j.1460-9568.2004.03540.x
51. Kumar A, Dogra S, Sona C, Umrao D, Rashid M, Singh SK, et al. Chronic histamine 3 receptor antagonism alleviates depression like conditions in mice via modulation of brain-derived neurotrophic factor and hypothalamus-pituitary adrenal axis. *Psychoneuroendocrinology*. (2019) 101:128–37. doi: 10.1016/j.psyneuen.2018.11.007
52. Sanna MD, Ghelardini C, Thurmond RL, Masini E, Galeotti N. Behavioural phenotype of histamine H4 receptor knockout mice: focus on central neuronal functions. *Neuropharmacology*. (2017) 114:48–57. doi: 10.1016/j.neuropharm.2016.11.023
53. Haas HL, Sergeeva OA, Selbach O. Histamine in the nervous system. *Physiol Rev*. (2008) 88:1183–241. doi: 10.1152/physrev.00043.2007
54. Benarroch EE. Histamine in the CNS: multiple functions and potential neurologic implications. *Neurology*. (2010) 75:1472–9. doi: 10.1212/WNL.0b013e3181f884b1
55. Chikahisa S, Kodama T, Soya A, Sagawa Y, Ishimaru Y, Sei H, et al. Histamine from brain resident MAST cells promotes wakefulness and modulates behavioral states. *PLoS One*. (2013) 8:e78434. doi: 10.1371/journal.pone.0078434
56. Threlfell S, Cragg SJ, Kallio I, Turi GF, Coen CW, Greenfield SA. Histamine H3 receptors inhibit serotonin release in substantia nigra pars reticulata. *J Neurosci*. (2004) 24:8704–10. doi: 10.1523/JNEUROSCI.2690-04.2004
57. Spaethling JM, Piel D, Dueck H, Buckley PT, Morris JE, Fisher SA, et al. Serotonergic neuron regulation informed by in vivo single-cell transcriptomics. *FASEB J*. (2014) 28:771–80. doi: 10.1096/fj.13-240267
58. Ferrada C, Moreno E, Casado V, Bongers G, Cortes A, Mallol J, et al. Marked changes in signal transduction upon heteromerization of dopamine D1 and histamine H3 receptors. *Br J Pharmacol*. (2009) 157:64–75. doi: 10.1111/j.1476-5381.2009.00152.x
59. Sergeeva OA, Chepkova AN, Görg B, Rodrigues Almeida F, Bidmon HJ, Haas HL, et al. Histamine-induced plasticity and gene expression in corticostriatal

- pathway under hyperammonemia. *CNS Neurosci Ther.* (2019) 26:355–66. doi: 10.1111/cns.13223
60. Rodríguez-Ruiz M, Moreno E, Moreno-Delgado D, Navarro G, Mallol J, Cortés A, et al. Heteroreceptor complexes formed by dopamine d1, histamine h3, and N-Methyl-D-aspartate glutamate receptors as targets to prevent neuronal death in Alzheimer's disease. *Mol Neurobiol.* (2017) 54:4537–50. doi: 10.1007/s12035-016-9995-y
61. Parks GS, Olivas ND, Ikrar T, Sanathara NM, Wang L, Wang Z, et al. Histamine inhibits the melanin-concentrating hormone system: implications for sleep and arousal. *J Physiol.* (2014) 592:2183–96. doi: 10.1113/jphysiol.2013.268771
62. Lagos P, Urbanavicius J, Scorza MC, Miraballes R, Torterolo P. Depressive-like profile induced by MCH microinjections into the dorsal raphe nucleus evaluated in the forced swim test. *Behav Brain Res.* (2011) 218:259–66. doi: 10.1016/j.bbr.2010.10.035
63. Cacabelos R, Torrellas C, Fernández-Novoa L, López-Muñoz F. Histamine and immune biomarkers in CNS disorders. *Mediat Inflamm.* (2016) 2016:1–10. doi: 10.1155/2016/1924603
64. Cacabelos R, Torrellas C, Fernández-Novoa L, Aliev G. Neuroimmune crosstalk in CNS disorders: the histamine connection. *Curr Pharm Design.* (2016) 22:819. doi: 10.2174/1381612822666151209150954
65. Tang J, Yu W, Chen S, Gao Z, Xiao B. Microglia polarization and endoplasmic reticulum stress in chronic social defeat stress induced depression mouse. *Neurochem Res.* (2018) 43:985–94. doi: 10.1007/s11064-018-2504-0
66. Miller AH. Depression and immunity: a role for T cells? *Brain Behav Immun.* (2010) 24:1–8. doi: 10.1016/j.bbi.2009.09.009
67. Loftis JM, Huckans M, Morasco BJ. Neuroimmune mechanisms of cytokine-induced depression: current theories and novel treatment strategies. *Neurobiol Dis.* (2010) 37:519–33. doi: 10.1016/j.nbd.2009.11.015
68. Lamers F, Milaneschi Y, Smit JH, Schoevers RA, Wittenberg G, Penninx B. Longitudinal association between depression and inflammatory markers: results from the Netherlands study of depression and anxiety. *Biol Psychiatry.* (2019) 85:829–37. doi: 10.1016/j.biopsych.2018.12.020
69. Laske C, Zank M, Klein R, Stransky E, Batra A, Buchkremer G, et al. Autoantibody reactivity in serum of patients with major depression, schizophrenia and healthy controls. *Psychiatry Res.* (2008) 158:83–6. doi: 10.1016/j.psychres.2006.04.023
70. Peacock BN, Scheiderer DJ, Kellermann GH. Biomolecular aspects of depression: a retrospective analysis. *Compr Psychiatry.* (2017) 73:168–80. doi: 10.1016/j.comppsy.2016.11.002
71. Teuscher C, Subramanian M, Noubade R, Gao JF, Offner H, Zachary JF, et al. Central histamine H3 receptor signaling negatively regulates susceptibility to autoimmune inflammatory disease of the CNS. *Proc Natl Acad Sci USA.* (2007) 104:10146–51. doi: 10.1073/pnas.0702291104
72. Dong H, Zhang W, Zeng X, Hu G, Zhang H, He S, et al. Histamine induces upregulated expression of histamine receptors and increases release of inflammatory mediators from microglia. *Mol Neurobiol.* (2014) 49:1487–500. doi: 10.1007/s12035-014-8697-6
73. Ferreira R, Santos T, Goncalves J, Baltazar G, Ferreira L, Agasse F, et al. Histamine modulates microglia function. *J Neuroinflammation.* (2012) 9:90. doi: 10.1186/1742-2094-9-90
74. Iida T, Yoshikawa T, Kárpáti A, Matsuzawa T, Kitano H, Mogi A, et al. JNJ10181457, a histamine H3 receptor inverse agonist, regulates in vivo microglial functions and improves depression-like behaviours in mice. *Biochem Biophys Res Commun.* (2017) 488:534–40. doi: 10.1016/j.bbrc.2017.05.081
75. Su W, Zhang T, Jiang C, Wang W. Clemastine alleviates depressive-like behavior through reversing the imbalance of microglia-related pro-inflammatory state in mouse hippocampus. *Front Cell Neurosci.* (2018) 12:412. doi: 10.3389/fncel.2018.00412
76. Bahi A, Schwed JS, Walter M, Stark H, Sadek B. Anxiolytic and antidepressant-like activities of the novel and potent non-imidazole histamine H(3) receptor antagonist ST-1283. *Drug Des Devel Ther.* (2014) 8:627–37. doi: 10.2147/DDDT.S63088
77. Femenía T, Magara S, DuPont CM, Lindskog M. Hippocampal-dependent antidepressant action of the H3 receptor antagonist clobenpropit in a rat model of depression. *Int J Neuropsychoph.* (2015) 18:v32. doi: 10.1093/ijnp/pyv032
78. Femenía T, Magara S, DuPont CM, Lindskog M. Hippocampal-dependent antidepressant action of the H3 receptor antagonist clobenpropit in a rat model of depression. *Int J Neuropsychopharmacol.* (2015) 18:yv032. doi: 10.1093/ijnp/pyv032
79. Dudek M, Kuder K, Kołaczowski M, Olczyk A, Żmudzka E, Rak A, et al. H3 histamine receptor antagonist pitolisant reverses some subchronic disturbances induced by olanzapine in mice. *Metab Brain Dis.* (2016) 31:1023–9. doi: 10.1007/s11011-016-9840-z
80. Gao Z, Hurst WJ, Czechtizky W, Hall D, Moindrot N, Nagorny R, et al. Identification and profiling of 3,5-dimethyl-isoxazole-4-carboxylic acid [2-methyl-4-((2S,3'S)-2-methyl-[1,3']bipyrrolidinyl-1'-yl)phenyl] amide as histamine H3 receptor antagonist for the treatment of depression. *Bioorg Med Chem Lett.* (2013) 23:6269–73. doi: 10.1016/j.bmcl.2013.09.081
81. Sharma BK, Singh P, Shekhawat M, Sarbhai K, Prabhakar YS. Modelling of serotonin reuptake inhibitory and histamine H(3)antagonistic activity of piperazine and diazepane amides: QSAR rationales for co-optimization of the activity profiles. *SAR QSAR Environ Res.* (2011) 22:365–83. doi: 10.1080/1062936X.2011.569895
82. Rahmadi M, Narita M, Yamashita A, Imai S, Kuzumaki N, Suzuki T. Sleep disturbance associated with an enhanced orexinergic system induced by chronic treatment with paroxetine and milnacipran. *Synapse.* (2011) 65:652–7. doi: 10.1002/syn.20893
83. Munari L, Provensi G, Passani MB, Galeotti N, Cassano T, Benetti F, et al. Brain histamine is crucial for selective serotonin reuptake inhibitors' behavioral and neurochemical effects. *Int J Neuropsychopharmacol.* (2015) 18:v45. doi: 10.1093/ijnp/pyv045
84. Costa A, Cristiano C, Cassano T, Gallelli CA, Gaetani S, Ghelardini C, et al. Histamine-deficient mice do not respond to the antidepressant-like effects of oleylethanolamide. *Neuropharmacology.* (2018) 135:234–41. doi: 10.1016/j.neuropharm.2018.03.033
85. Garden BC, Francois D. Cetirizine-associated delusions and depression in an 18-year-old woman. *Clin Neuropharmacol.* (2013) 36:96–7. doi: 10.1097/WNF.0b013e318290b9b2
86. Kishi T, Iwata N. Efficacy and tolerability of histamine-2 receptor antagonist adjunct of antipsychotic treatment in schizophrenia: a meta-analysis of randomized placebo-controlled trials. *Pharmacopsychiatry.* (2015) 48:30–6. doi: 10.1055/s-0034-1390478
87. Ozdemir PG, Karadag AS, Selvi Y, Boysan M, Bilgili SG, Aydin A, et al. Assessment of the effects of antihistamine drugs on mood, sleep quality, sleepiness, and dream anxiety. *Int J Psychiatry Clin Pract.* (2014) 18:161–8. doi: 10.3109/13651501.2014.907919
88. Yeung WF, Chung KF, Yung KP, Ng TH. Doxepin for insomnia: a systematic review of randomized placebo-controlled trials. *Sleep Med Rev.* (2015) 19:75–83. doi: 10.1016/j.smrv.2014.06.001
89. Leher P. Efficacy of pitolisant 20 mg in reducing excessive daytime sleepiness and fatigue in patients with obstructive sleep apnoea syndrome: an individual patient data meta-analysis. *Clin Drug Investig.* (2022) 42:65–74. doi: 10.1007/s40261-021-01104-8
90. Krief S, Berrebi-Bertrand I, Nagmar I, Giret M, Belliard S, Perrin D, et al. Pitolisant, a wake-promoting agent devoid of psychostimulant properties: preclinical comparison with amphetamine, modafinil, and solriamfetol. *Pharmacol Res Perspect.* (2021) 9:e855. doi: 10.1002/prp2.855
91. Wang J, Li X, Yang S, Wang T, Xu Z, Xu J, et al. Pitolisant versus placebo for excessive daytime sleepiness in narcolepsy and obstructive sleep apnea: a meta-analysis from randomized controlled trials. *Pharmacol Res.* (2021) 167:105522. doi: 10.1016/j.phrs.2021.105522