



# Small Intestinal Bacterial Overgrowth and Irritable Bowel Syndrome – An Update

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Small intestinal bacterial overgrowth (SIBO) is one manifestation of gut microbiome dysbiosis and is highly prevalent in IBS (Irritable Bowel Syndrome). SIBO can be diagnosed either by a small bowel aspirate culture showing  $\geq 10^3$  colony-forming units (CFU) per mL of aspirate, or a positive hydrogen lactulose or glucose breath test. Numerous pathogenic organisms have been shown to be increased in subjects with SIBO and IBS, including but not limited to *Enterococcus*, *Escherichia coli*, and *Klebsiella*. In addition, *Methanobrevibacter smithii*, the causal organism in a positive methane breath test, has been linked to constipation predominant irritable bowel syndrome (IBS-C). As *M. smithii* is an archaeon and can overgrow in areas outside of the small intestine, it was recently proposed that the term intestinal methanogen overgrowth (IMO) is more appropriate for the overgrowth of these organisms. Due to gut microbiome dysbiosis, patients with IBS may have increased intestinal permeability, dysmotility, chronic inflammation, autoimmunity, decreased absorption of bile salts, and even altered enteric and central neuronal activity. As a consequence, SIBO and IBS share a myriad of symptoms including abdominal pain, distention, diarrhea, and bloating. Furthermore, gut microbiome dysbiosis may be associated with select neuropsychological symptoms, although more research is needed to confirm this connection. This review will focus on the role of the gut microbiome and SIBO in IBS, as well as novel innovations that may help better characterize intestinal overgrowth and microbial dysbiosis.

**Keywords:** small intestinal bacterial overgrowth, irritable bowel syndrome, gut dysbiosis, breath test, methane, archaea, hydrogen, hydrogen sulfide

## INTRODUCTION

Irritable bowel syndrome (IBS) is a functional bowel disorder defined by recurrent abdominal pain for at least 1 day per week in the last 3 months that is associated with 2 or more of the following: related to defecation, associated with a change in stool form, or associated with a change in stool frequency (1). Symptom onset must occur at least 6 months prior to diagnosis, but many patients suffer long-term chronic symptoms as a result of this disorder. Patients may experience various comorbidities including, but not limited to, bloating, constipation, diarrhea, incontinence, and psychological disturbances.

Historically, IBS has been associated with stress and anxiety, and the brain-gut axis has been widely described as important in understanding IBS (2). Because of this, many treatments focus on antidepressants and neurobehavioral intervention (3). Although these treatments can be effective, newer studies have demonstrated more complex, organic etiologies specific to the gut that have led to novel therapeutic options. The complex network of etiologies likely represents various pathophysiological states that may be unique to the patient and include visceral hyperalgesia, intestinal permeability, immune activation, altered gastrointestinal motility, autoimmunity, and alteration of the gut microbiome. The last category, the gut microbiome, has seen an exponential growth in interest over the last few years. One manifestation of dysbiosis, small intestinal bacterial overgrowth (SIBO), is linked to IBS and will be the focus of this review.

## DEFINITION OF SIBO

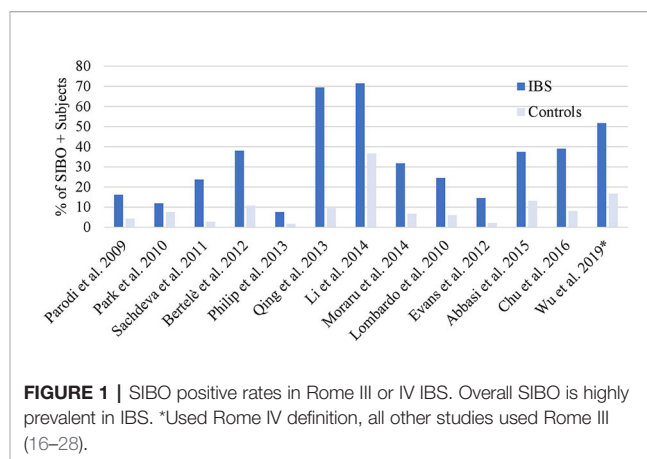
While a diagnosis of IBS is based on clinical symptoms, the gold standard for a diagnosis of SIBO is the presence of  $\geq 10^3$  colony forming units per milliliter (CFU/mL) of jejunal aspirate by culture (4, 5). However, aspiration is invasive and expensive, requiring a skilled gastroenterologist. Also, there may be sampling errors given that only a small segment of the small bowel can be aspirated, leaving the rest unexplored. Alternatively, a breath test can be used to assess microbial overgrowth in the gut. Hydrogen ( $H_2$ ) and methane ( $CH_4$ ) are exclusively produced by microbial metabolism and are exhaled on the breath (6–8). The North American consensus defines a rise in  $H_2 \geq 20$  parts per million (ppm) from baseline within 90 min of substrate ingestion as positive for the  $H_2$  breath test, and a  $CH_4$  level  $\geq 10$  ppm at any time is defined as positive for the  $CH_4$  breath test (4). Two of the most common carbohydrate substrates used for the breath test are glucose and lactulose. Glucose is a monosaccharide that is readily absorbed in the proximal small intestine, whereas lactulose is a disaccharide and has more limited absorbability (9). SIBO is associated with a myriad of symptoms including, but not limited to bloating, abdominal pain, nausea, constipation, and diarrhea. A positive  $H_2$  breath test is diagnostic of SIBO, which has been associated with diarrhea-predominant IBS (IBS-D) and IBS with mixed bowel habits (IBS-M) (10). A positive  $CH_4$  breath test is indicative of methanogen overgrowth, which has been associated with constipation predominant IBS (IBS-C) (5, 11, 12). Of note, the recent SIBO guidelines have reclassified  $CH_4$  positive breath test as intestinal methanogen overgrowth (IMO), as methanogenesis is likely not limited to the small intestine (5, 13–15). In addition, although measurement of  $CH_4$  is not always included, the North American consensus and the recent SIBO guidelines both recommend that  $CH_4$  be measured concurrently with  $H_2$  during breath testing (4, 5).

## PREVALENCE OF SIBO IN IBS

The relationship between SIBO and IBS was described in a 2020 meta-analysis of 25 case-control studies involving 3,192 IBS

subjects and 3,320 controls, that showed that the prevalence of SIBO in IBS was 31.0% (95% CI 29.4–32.6) with an OR of 3.7 (95% CI 2.3–6.0,  $p = 0.001$ ) compared to controls (16). This meta-analysis included studies with healthy controls and non-IBS patient controls. When comparing SIBO rates with only healthy controls the OR increased to 4.9 (95% CI 2.8–8.6,  $p = 0.001$ ). This meta-analysis included studies that had several definitions of SIBO, as well as several IBS diagnostic criteria. The Rome IV diagnostic criteria for IBS were published in 2016, and the meta-analysis did not include any studies utilizing the newer Rome IV criteria. A funnel plot evaluating publication bias showed some asymmetry, although after including only the top 15 high-quality studies with low risk of bias, the asymmetry was no longer observed. Interestingly, this increased the OR for SIBO in IBS with an OR of 4.1 (95% CI 3.0–5.6,  $p < 0.001$ ). Slightly higher rates of SIBO were seen in studies that utilized breath testing as opposed to small intestinal aspiration (35.5 vs. 33.5%, respectively). The two most common breath test utilized were glucose breath test ( $n = 9$ ) and lactulose breath test ( $n = 8$ ). The prevalence of SIBO in IBS subjects vs controls was 62.3 vs 33.5% for the lactulose breath test, and 20.7 vs 4.4% for glucose. The association between SIBO and IBS seemed to be the strongest for IBS-D vs IBS-C or IBS-M, at 35.5% (95% CI 32.7–40.3) vs 22.5% (95% CI 18.1–26.9) or 25.2% (95% CI 22.2–28.4), respectively. Since this meta-analysis, one study utilizing the newer Rome IV diagnostic criteria for IBS has shown that the prevalence of SIBO is increased in subjects with IBS (51.7 vs. 16.7%,  $p \leq .001$ ) (17). **Figure 1** summarizes the prevalence of SIBO in IBS vs controls from case-control studies ( $n > 80$ ) using Rome III and the latest Rome IV criteria for IBS (17–29). Our analysis of these 13 studies shows that the pooled SIBO rate is significantly higher in IBS subjects than in controls (30 vs 9%,  $n = 2,494$ ,  $p < 0.0001$ ). The number of healthy subjects with SIBO likely reflects the complex relationship between SIBO and symptoms. A balance of specific bacterial strains with different adaptive host factors may produce asymptomatic, or mildly symptomatic patients with SIBO.

In contrast to SIBO (based on a positive  $H_2$  breath test), which is more strongly associated with IBS-D, IMO has been linked to IBS-C (5, 11, 16, 30, 31). Unlike  $H_2$ , which has not been shown to directly induce diarrhea,  $CH_4$  has been shown to directly slow intestinal transit and causes constipation, both in



**FIGURE 1** | SIBO positive rates in Rome III or IV IBS. Overall SIBO is highly prevalent in IBS. \*Used Rome IV definition, all other studies used Rome III (16–28).

humans (32) and in animal models (33). Pimentel et al. showed that direct infusion of CH<sub>4</sub> in animals slows intestinal transit, and also showed that segments of guinea pig ileum exhibit increased contractile impulses when bathed in CH<sub>4</sub> (33). CH<sub>4</sub> appears to cause dysregulation in motility by amplifying neuronal activity in the intestine through the anticholinergic pathway (34). In humans, a study showed that the degree of constipation correlated with breath CH<sub>4</sub> levels in subjects with IBS (31). Of note, for patients with constipation without IBS, the percentage of CH<sub>4</sub> positive patients also correlated with constipation (35). In this study, those with decreased motility in the colon had higher levels of total and maximum CH<sub>4</sub> (35). Two strict anaerobic methanogens have been isolated from the human gut, *Methanospaera stadtmaniae* (36) and *Methanobrevibacter smithii* (37), with the latter being the predominant source of methanogenesis and associated with constipation (38). Supporting this, another study found that higher levels of *M. smithii* correlated with higher CH<sub>4</sub> on breath test (12). These studies shed light on a causal relationship between methanogen overgrowth and at least a subset of IBS-C. Interestingly, bile acids are a proposed etiology of functional diarrhea and IBS-D, and have been shown to decrease methanogenesis in human feces (39).

Only a few studies have attempted to characterize the small bowel microbiome in subjects with SIBO and IBS. A North American study found that SIBO subjects had a 7–8-fold increase in *Klebsiella* and *Escherichia/Shigella* compared to non SIBO patients (40). One Indian study, on jejunal aspirate of SIBO and IBS subjects, found that 40% of subjects had *Pseudomonas aeruginosa*, 6.7% had *Acinetobacter baumannii*, 13.3% had *Acinetobacter lwoffii*, 13.3% had *Staphylococcus spp.*, 6.7% had *Enterococcus faecalis*, 20% had *Escherichia coli*, 6.7% had *Enterococcus faecium*, 13.3% had *Klebsiella pneumoniae*, and 6.7% had *Streptococcus spp.* by culture (41). This study also noted an increase in *Acinetobacter lwoffii*, *Staphylococcus spp.*, *Enterococcus faecalis*, *Escherichia coli*, and *Klebsiella pneumoniae* in IBS subjects with SIBO vs. those without SIBO. Similarly, a Swedish study found that IBS subjects with SIBO had a high prevalence of Gram-negative *Bacilli* and *Enterobacter* on jejunal aspirate (42). Lastly, a study from Athens found high prevalence of *Escherichia coli*, *Enterococcus spp.*, and *Klebsiella pneumoniae* in duodenal aspirates from subjects with SIBO (43). Given the highly variable results of microbiome studies arising from different sampling locations, geography, and definitions of SIBO, caution should be exercised when generalizing these results.

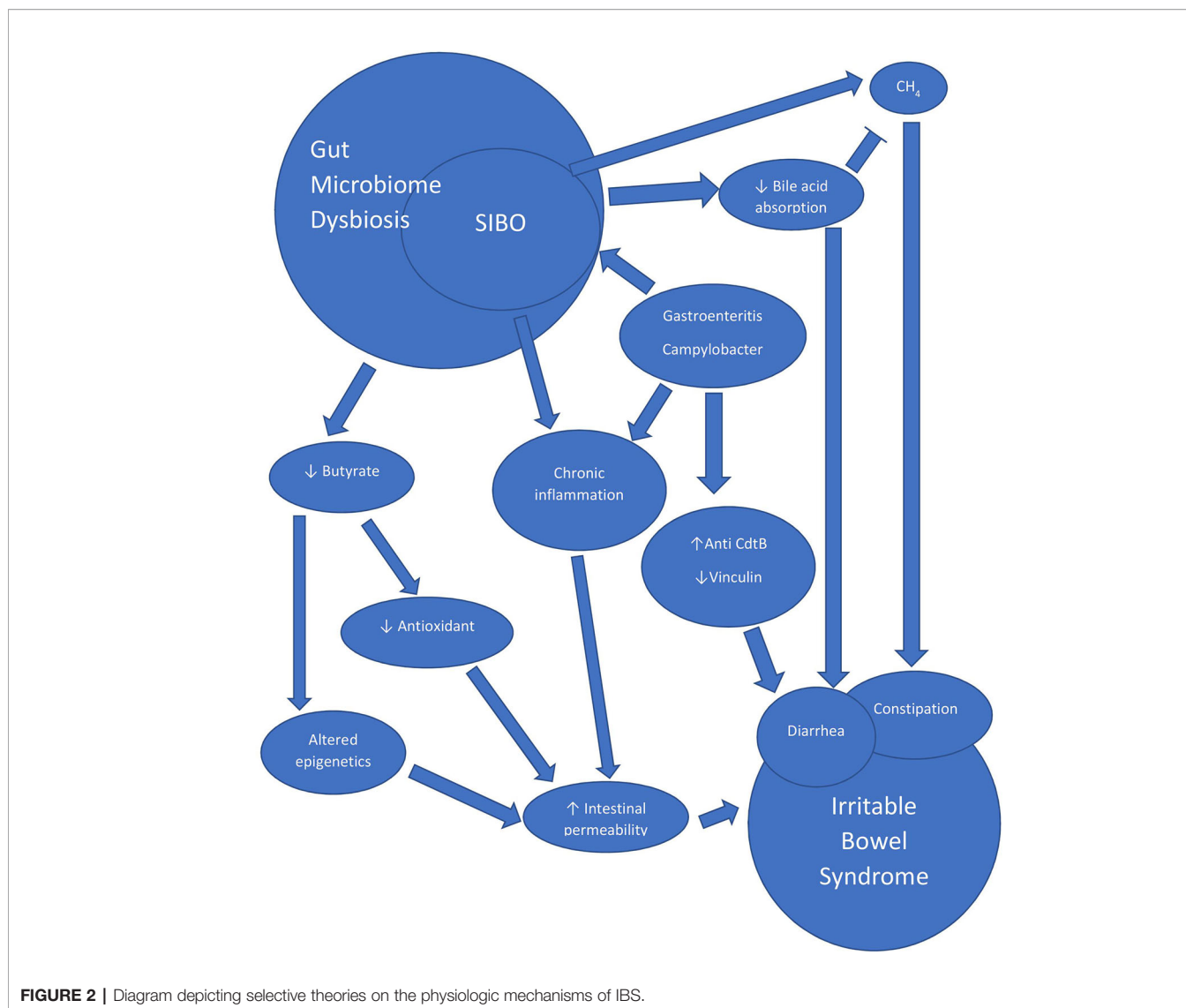
## GUT MICROBIOME DYSBIOSIS, IBS AND SIBO—FOCUS ON POST-INFECTIOUS IBS MODELS

SIBO only comprises a subset of gut microbiome dysbiosis, and this review would not be complete without discussing how microbial dysbiosis in general contributes to IBS. Previous

studies have shown that infectious etiologies such as infectious gastroenteritis (44, 45) and diverticulitis (46) are associated with the development of IBS, which have been termed post-infectious IBS (PI-IBS). A recent systemic review has shown that roughly 10% of patients with enteritis develop PI-IBS within the following year and the prevalence of PI-IBS seems to increase with time (47). These infections are thought to induce changes through long-lasting low-grade inflammation, an increase in intestinal permeability, and autoimmunity, ultimately leading to the symptoms of IBS (Figure 2). Given that roughly 10% of the population has IBS (48) and the that an estimated 10 million food-borne illnesses occur each year (49), it is possible that a significant portion of patients with IBS may have had gastroenteritis in the past that they cannot recall and may have PI-IBS. Interestingly, a mathematical model has shown that infectious gastroenteritis may contribute to a large proportion of IBS (50). Therefore, we hypothesize that studies of PI-IBS may be relevant to many subjects with IBS.

Infectious diarrhea has been known to cause intestinal permeability (51, 52), and a similar phenomenon is seen in patients with IBS (52, 53). This is thought to be partially mediated through bacterial effects on tight junctions (54). Although the mechanism(s) underlying how intestinal permeability persists after the acute infection is not entirely clear, there are many hypotheses centered on gut microbiome dysbiosis. Butyrate, a microbial metabolite, is a key player in maintaining a healthy epithelial barrier by regulating cell turnover, antioxidants, and energy maintenance in the gut lining (55, 56). Interestingly, one study found lower levels of butyrate-producing bacterial families such as Ruminococcaceae, an unknown family from the order Clostridiales, and Erysipelotrichaceae in subjects with IBS-D (57). Butyrate appears to mediate cell turnover through inhibition of histone deacetylase (HDAC), which causes apoptosis of the luminal cells (56). Interestingly, in a rodent model of IBS, inhibition of HDAC alleviated symptoms of visceral hypersensitivity (58). In addition, another candidate important in the maintenance of intestinal permeability has recently gained interest in IBS. A double-blind placebo-controlled randomized control trial in 2019 evaluated glutamine, a key amino acid that helps gut epithelial integrity (59), and found that an 8-week course significantly alleviated IBS symptoms (60). The improvement in symptoms was correlated with improvement in intestinal permeability. Interestingly, glutamine-enriched total parenteral nutrition did not improve intestinal permeability in malnourished surgical subjects undergoing gastrointestinal surgery (61), which may suggest additional mechanisms are needed to maintain the gut epithelial barrier which are not present in these subjects. Given that these subjects had poor oral intake coupled with severe gastrointestinal disease which necessitated surgery, their microbiome dysbiosis should be further investigated.

Secondly, inflammation seems to play a role in the pathophysiology of IBS. Previous studies have shown that rectal biopsies from subjects who had acute *Campylobacter enteritis* and diarrhea exhibit persistently elevated T-lymphocytes and



**FIGURE 2** | Diagram depicting selective theories on the physiologic mechanisms of IBS.

enteroendocrine cells, and that similar elevations are seen in subjects with post-infectious IBS (52). Another study found an increase in enterochromaffin cells and T-lymphocytes in subjects with new-onset post-infectious IBS following *Campylobacter* infection compared to both subjects who were asymptomatic following infection and to healthy controls (62). The increase in lymphocytes appears to be present in the general IBS population as well, as one study found a 1.8-fold increase in intraepithelial lymphocytes in IBS subjects as compared to asymptomatic controls (63). This is not surprising since a large proportion of IBS is suspected to be due to a previous infection (50). A mouse model of post-infectious IBS developed using *Trichinella spiralis* (64) found similar results, with enterochromaffin cells being persistently elevated in the bowel long after clearance of the acute infection, as well as an increase in intraepithelial cells and a decrease in intestinal cells of Cajal in mice that developed SIBO (65). In addition, a post-*Campylobacter* rat model of IBS exhibited persistent alterations in stool consistency and in rats that

exhibited SIBO, intraepithelial lymphocytes in the rectum and the left colon were also increased (66).

Autoimmunity may also play a significant role in the development of IBS as it relates to the gut microbiome. In a post-infectious IBS model, rats who were exposed to *Campylobacter* with an insertional deletion of the bacterial toxin cytolethal distending toxin B (CdtB) exhibited significantly fewer intraepithelial lymphocytes and less alterations in stool form (67). Antibodies to CdtB were found to have a high affinity to the neuronal protein vinculin, and a decrease in vinculin expression correlated with the number of times the rats were exposed to *Campylobacter* (68). Vinculin is a key cytoskeletal protein that plays an essential role in cell-matrix and cell-cell adhesion (69). In humans, numerous studies have found that these two biomarkers (antibodies to CdtB and vinculin) can differentiate between IBS-D/M (70–73) and other bowel diseases and conditions that cause diarrhea, although one study in Australia failed to show such relationship (74). A longitudinal observation of a single patient who developed IBS

after an episode of infectious diarrhea demonstrated a subsequent elevation in anti-CdtB, followed by an elevation in anti-vinculin, each coinciding with symptoms (44). A thorough evaluation failed to find an “organic” gastrointestinal disorder in this patient. In humans, this pathway seems to be specific to patients with IBS-D and IBS-M but not IBS-C (71), which is consistent with findings in the rat model.

Finally, SIBO and malabsorption has also been proposed as a mechanism underlying the development of diarrhea. It is thought that deconjugation of bile salts in the upper gut induces a decrease in absorption of fat and lipophilic vitamins, leading to the production of lithocholic acid, which is poorly absorbed and thought to be enterotoxic (75). In subjects with malabsorption syndrome with and without SIBO, subjects who had SIBO exhibited significantly elevated unconjugated bile salts, acetate, lactate, and formate than those without SIBO (76).

## MICROBIOME-TARGETED TREATMENT OF SIBO, IMO, AND IBS

There are treatments for both SIBO and IBS which target the microbiome, including treatment with a non-absorbable oral antibiotic, rifaximin. A meta-analysis which evaluated normalization of breath test in response to antibiotics for SIBO found that of the 10 studies included, rifaximin was the most common and was used in 8 studies. They found a pooled normalization rate of 49.5%, 95% CI 44.0–55.1 with rifaximin vs. 9.8%, 95% CI 4.6–17.8 with placebo (77). Rifaximin was compared against placebo in only 3 studies and had a favorable response with an effectiveness ratio of 1.97, 95% CI 0.93–4.17,  $p = 0.08$ , although this did not reach statistical significance. Four studies directly compared antibiotics to placebo and found an effectiveness ratio of 2.55, 95% CI 1.29–5.04,  $p = 0.03$  favoring antibiotics.

Similarly, multiple large randomized controlled trials have shown that rifaximin can improve symptoms in subjects with IBS without constipation (78, 79). Currently this is the only FDA-approved therapy to treat IBS by directly affecting the gut microbiome. Unlike other medications on the market for the treatment of IBS, the effects of this treatment outlasted the duration of treatment by weeks to months. This is thought to be due to the effects of rifaximin on the gut microbiome which persist after the treatment is over. Unfortunately, a subset of subjects experience a relapse of their symptoms, but repeat dosing with rifaximin appears to be safe and effective (79, 80). The mechanism of action for this drug is not entirely clear, but it appears to induce a small, transient reduction in Shannon index as well as a decrease in the relative abundance of 7 taxa including Enterobacteriaceae, Verrucomicrobiaceae, Peptostreptococcaceae, Pasteurellaceae, Synergistaceae, Eubacteriaceae, and Enterococcaceae (81), as determined from stool samples. These transient effects are consistent with clinical observations. Whether the reductions in these taxa are directly linked to the improvements in IBS symptoms is yet to be determined. Further studies are needed to also evaluate changes in small bowel and proximal colon.

A metaanalysis by Shah et al. found 7 studies which evaluated patients with SIBO in IBS and found that antibiotics relieve symptoms in 81.6% of patients. Only 5 studies reported eradication of SIBO and 93% of patients with a glucose breath test achieved normalization while 71.4% of patients who were diagnosed *via* small bowel aspirate culture reached normalization (16).

Furthermore, treatment with specific antibiotics results in decreased CH<sub>4</sub> levels that correlate with improvements in constipation (82, 83). Of note, while neomycin and rifaximin can each reduce constipation in IBS-C, using a combination of both appears to be most effective (84). Currently only a small number of studies with limited sample sizes have evaluated the use of antibiotics in the treatment of IMO, and larger, multicentered randomized control trials are needed to further characterize their efficacies and potential adverse events.

Perhaps the largest clinical indication of the overlap between SIBO and IBS was demonstrated by Rezaie et al, who showed that a positive lactulose breath test predicted a favorable response to rifaximin (59.7 vs 25.8%,  $p = 0.002$ ) (85) than patients who had a negative lactulose breath test. Given this finding, patients with IBS should undergo a lactulose breath test to rule out SIBO, as this may have treatment implications (86).

Finally, probiotics have also been evaluated in the treatment of IBS. A systemic review by Ford et al. (87) found that certain combinations of probiotics may be helpful in IBS, although there was significant heterogeneity between the studies with possible publication bias. In this study, combination probiotics LacClean Gold and VSL#3 had some efficacy with RR = 0.59; 95% CI 0.37–0.93 ( $n = 130$ ) and RR = 0.82; 95% CI 0.52–1.30 ( $n = 78$ ), respectively. Interpretation of metaanalyses of probiotic studies is difficult since different strains are studied in different combinations assessing various endpoints. Many studies also have small sample sizes, making it difficult to generalize the results.

## NEUROPSYCHOLOGICAL SYMPTOMS, SIBO, AND IBS

IBS patients can experience significant effects on their mental health with increased rates of comorbid psychological conditions such as stress and anxiety, as well as decreased energy and disruptions in sleep and functioning (88–90). Whether these conditions are part of the IBS-SIBO overlap remains a focus of research. A recent study by Rao et al. found that subjects with brain fog had increased rates of SIBO (91), and also found an association between brain fog and probiotic use as well as D-lactic acidosis, a condition found in patients with short bowel syndrome (92). Although the participants in this study were all referred from a gastrointestinal clinic, none were classified as having IBS, although the study did exclude subjects with “organic” gastrointestinal diseases. Furthermore, although this study was not placebo-controlled, an improvement was seen with initiation of antibiotics and the cessation of probiotics. Given this, probiotics may have subtle effects on health and despite their accessibility as an over-the-counter medication, should be taken cautiously. Similarly, in subjects with chronic fatigue

syndrome, another disease which carries an enigmatic pathophysiology, erythromycin was shown to have a significant effect on sleep duration but only in those that exhibited significant effects on the gut microbiome, specifically a decrease in *Streptococcus* levels (93). It is unclear whether the promotility effect of erythromycin or its antimicrobial effect played a greater role in these findings.

Finally, one study has demonstrated a possible connection from the gut microbiome to the brain *via* the vagus nerve. In this study, mice supplemented with *Lactobacillus rhamnosus* exhibited decreased stress-related behaviors and altered expression of gamma-aminobutyric acid (GABA) receptor in the brain (94). Interestingly, this effect was decreased in rodents who underwent vagotomy, suggesting a direct neuronal effect *via* the 10<sup>th</sup> cranial nerve. Unfortunately, many of these studies have methodological issues concerning blinding, open-labeled design, and small sample size, to name a few potential biases. Whether alterations in the gut microbiome can directly affect the central nervous system and whether further modification *via* antibiotics or probiotics can have beneficial effects on humans requires further research.

## FUTURE DIRECTIONS

Currently there is no consensus on the use of lactulose or glucose in breath testing (5). The lactulose breath test is limited by its potential false positive rates in patients with high motility, and the glucose breath test may not adequately detect SIBO in the distal bowel as it gets readily absorbed in the proximal small intestine (9). More validation studies are needed for standardization. As a potential solution, an ingestible capsule that can directly measure gas in the intestine is being developed. This may potentially improve the diagnostic accuracy over current methods which utilize breath testing.

There are several other innovations that may significantly influence the field of SIBO and IBS. A new study by Leite et al. has found that the use of dithiothreitol, a mucolytic, can significantly increase bacterial yield from small intestinal aspirates by approximately 3-fold (95). This will allow a more complete and thorough sampling of the small bowel which may lead to novel discoveries. As noted above, aspiration techniques

can vary and this may explain why one recent study did not find any correlation between symptoms and SIBO defined by CFU/mL in a duodenal aspirate (96). Further studies will be needed to optimize sampling methods for the intestinal microbiome but until then the breath test, which has potential clinical implications and can evaluate a larger section of the bowel (85), should be utilized in the diagnosis of SIBO (4).

Interestingly, another potential marker of bacterial overgrowth on a breath test, the exhaled gas hydrogen sulfide (H<sub>2</sub>S), has recently been explored. H<sub>2</sub>S appears to have implications for multiple gastrointestinal disorders, and can relax smooth muscle and have pro- and anti-inflammatory properties (97). Singer-Englar et al. described an association between diarrhea and levels of exhaled H<sub>2</sub>S (98). Whether there is also an association with IBS-D (the definition of which requires pain in addition to diarrhea) remains to be seen.

Lastly, given that dysbiosis does not necessarily mean an overall overgrowth in the number of CFU and that changes in the relative abundances of specific microbial strains or taxa can cause symptoms, a reliance on measuring the overall abundance of bacteria may not be suitable in the era of 16S rRNA sequencing and breath testing.

## CONCLUSION

IBS and SIBO appear to be intertwined. The methods for measuring SIBO still need further optimization. SIBO and other forms of gut microbiome dysbiosis are likely responsible for some symptoms in a subset of patients with IBS and can help guide treatment options. Given its therapeutic potential, further research in IBS and SIBO is needed.

## AUTHOR CONTRIBUTIONS

MP and WT contributed to the concept, design, and writing of the review article.

## REFERENCES

- Lacy BE, Mearin F, Chang L, Chey WD, Lembo AJ, Simren M, et al. Bowel Disorders. *Gastroenterology* (2016) 150:1393–1407.e5. doi: 10.1053/j.gastro.2016.02.031
- Drossman DA, Camilleri M, Mayer EA, Whitehead WE. AGA technical review on irritable bowel syndrome. *Gastroenterology* (2002) 123(6):2108–31. doi: 10.1053/gast.2002.37095
- Ford AC, Lacy BE, Harris LA, Quigley EMM, Moayyedi P. Effect of Antidepressants and Psychological Therapies in Irritable Bowel Syndrome: An Updated Systematic Review and Meta-Analysis. *Am J Gastroenterol* (2019) 114(1):21–39. doi: 10.1038/s41395-018-0222-5
- Rezaie A, Buresi M, Lembo A, Lin H, McCallum R, Rao S, et al. Hydrogen and Methane-Based Breath Testing in Gastrointestinal Disorders: The North American Consensus. *Am J Gastroenterol* (2017) 112(5):775–84. doi: 10.1038/ajg.2017.46
- Pimentel M, Saad RJ, Long MD, Rao SSC. ACG Clinical Guideline: Small Intestinal Bacterial Overgrowth. *Am J Gastroenterol* (2020) 115:165–78. doi: 10.14309/ajg.0000000000000501
- Levitt MD. Volume and composition of human intestinal gas determined by means of an intestinal washout technic. *N Engl J Med* (1971) 284(25):1394–8. doi: 10.1056/NEJM197106242842502
- Bond JH, Engel RR, Levitt MD. Factors influencing pulmonary methane excretion in man. An indirect method of studying the in situ metabolism of the methane-producing colonic bacteria. *J Exp Med* (1971) 133(3):572–88. doi: 10.1084/jem.133.3.572
- Christl SU, Murgatroyd PR, Gibson GR, Cummings JH. Production, metabolism, and excretion of hydrogen in the large intestine. *Gastroenterology* (1992) 102(4 Pt 1):1269–77. doi: 10.1016/0016-5085(92)90765-Q
- Romagnuolo J, Schiller D, Bailey RJ. Using breath tests wisely in a gastroenterology practice: an evidence-based review of indications and

- pitfalls in interpretation. *Am J Gastroenterol* (2002) 97(5):1113–26. doi: 10.1111/j.1572-0241.2002.05664.x
10. Chen B, Kim JJ, Zhang Y, Du L, Dai N. Prevalence and predictors of small intestinal bacterial overgrowth in irritable bowel syndrome: a systematic review and meta-analysis. *J Gastroenterol* (2018) 53(7):807–18. doi: 10.1007/s00535-018-1476-9
  11. Hwang L, Low K, Khoshini R, Melmed G, Sahakian A, Makhani M, et al. Evaluating breath methane as a diagnostic test for constipation-predominant IBS. *Dig Dis Sci* (2010) 55(2):398–403. doi: 10.1007/s10620-009-0778-4
  12. Ghoshal U, Shukla R, Srivastava D, Ghoshal UC. Irritable Bowel Syndrome, Particularly the Constipation-Predominant Form, Involves an Increase in Methanobrevibacter smithii, Which Is Associated with Higher Methane Production. *Gut Liver* (2016) 10(6):932–8. doi: 10.5009/gnl15588
  13. Takakura WR, Oh SJ, Singer-Englar T, Leite G, Pimentel M, Fridman A, et al. Patients with Appendectomy Have Significantly Less Methane in Their Exhaled Breath: is the Appendix a Vestigial Organ Or an Important Reservoir for Methanogens? *Gastroenterology* (2019) 156(6):S–1161. doi: 10.1016/S0016-5085(19)39870-1
  14. Weaver GA, Krause JA, Miller TL, Wolin MJ. Incidence of methanogenic bacteria in a sigmoidoscopy population: an association of methanogenic bacteria and diverticulosis. *Gut* (1986) 27(6):698–704. doi: 10.1136/gut.27.6.698
  15. Eckburg PB, Bik EM, Bernstein CN, Purdom E, Dethlefsen L, Sargent M, et al. Diversity of the human intestinal microbial flora. *Science* (2005) 308(5728):1635–8. doi: 10.1126/science.1110591
  16. Shah A, Talley NJ, Jones M, Kendall BJ, Koloski N, Walker MM, et al. Intestinal Bacterial Overgrowth in Irritable Bowel Syndrome: A Systematic Review and Meta-Analysis of Case-Control Studies. *Am J Gastroenterol* (2020) 115:190–201. doi: 10.14309/ajg.0000000000000504
  17. Wu KQ, Sun WJ, Li N, Chen YQ, Wei YL, Chen DF. Small intestinal bacterial overgrowth is associated with Diarrhea-predominant irritable bowel syndrome by increasing mainly Prevotella abundance. *Scand J Gastroenterol* (2019) 54:1–7. doi: 10.1080/00365521.2019.1694067
  18. Parodi A, Dulbecco P, Savarino E, Giannini EG, Bodini G, Corbo M, et al. Positive glucose breath testing is more prevalent in patients with IBS-like symptoms compared with controls of similar age and gender distribution. *J Clin Gastroenterol* (2009) 43(10):962–6. doi: 10.1097/MCG.0b013e3181a099a5
  19. Park JS, Yu JH, Lim HC, Kim JH, Yoon YH, Park HJ, et al. [Usefulness of lactulose breath test for the prediction of small intestinal bacterial overgrowth in irritable bowel syndrome]. *Korean J Gastroenterol* (2010) 56(4):242–8. doi: 10.4166/kjg.2010.56.4.242
  20. Sachdeva S, Rawat AK, Reddy RS, Puri AS. Small intestinal bacterial overgrowth (SIBO) in irritable bowel syndrome: frequency and predictors. *J Gastroenterol Hepatol* (2011) 26 Suppl 3:135–8. doi: 10.1111/j.1440-1746.2011.06654.x
  21. Bertelè A, Papadia C, Bosi S, Marcucci F, Corrente V, Ciarleglio A, et al. P.07.16 Bacterial Overgrowth In Irritable Bowel Syndrome: A Cohort Study. *Digest Liver Dis* (2012) 44:S132. doi: 10.1016/S1590-8658(12)60365-X
  22. Philip AJV, Mukunda M, Krishnadas D, Sobhana Devi R. SI-11 Prevalence of small intestinal bacterial overgrowth in patients with irritable bowel syndrome using lactulose hydrogen breath test. *Indian J Gastroenterol* (2013) 32(1):1–132. doi: 10.1007/s12664-013-0417-z
  23. Qing Y. Relationship between the irritable bowel syndrome and small intestinal bacterial. *J Gastroenterol Hepatol* (2013) 28(S3):23–693.
  24. Li N, Wang Z, Fei G, Zhu L, Chen W, Li X, et al. Measurement of methane and hydrogen on lactulose breath test detects small intestinal bacterial overgrowth in patients with irritable bowel syndrome. *Neurogastroenterol Motil* (2014) 26(s1):11–82.
  25. Moraru IG, Moraru AG, Andrei M, Iordache T, Drug V, Diculescu M, et al. Small intestinal bacterial overgrowth is associated to symptoms in irritable bowel syndrome. Evidence from a multicentre study in Romania. *Rom J Intern Med* (2014) 52(3):143–50.
  26. Lombardo L, Foti M, Ruggia O, Chiecchio A. Increased incidence of small intestinal bacterial overgrowth during proton pump inhibitor therapy. *Clin Gastroenterol Hepatol Off Clin Pract J Am Gastroenterol Assoc* (2010) 8(6):504–8. doi: 10.1016/j.cgh.2009.12.022
  27. Evans K, Lunn E, Raza S, Sanders DS, Higham S. PWE-050 Does small intestine bacterial overgrowth cause neurodysmotility in IBS and coeliac disease? *Gut* (2012) 61(Suppl 2):A317–A. doi: 10.1136/gutjnl-2012-302514d50
  28. Abbasi MH, Zahedi M, Darvish Moghadam S, Shafieipour S, HayatBakhsh Abbasi M. Small bowel bacterial overgrowth in patients with irritable bowel syndrome: the first study in iran. *Middle East J Dig Dis* (2015) 7(1):36–40.
  29. Chu H, Fox M, Zheng X, Deng Y, Long Y, Huang Z, et al. Small Intestinal Bacterial Overgrowth in Patients with Irritable Bowel Syndrome: Clinical Characteristics, Psychological Factors, and Peripheral Cytokines. *Gastroenterol Res Pract* (2016) 2016:3230859. doi: 10.1155/2016/3230859
  30. Pimentel M, Mayer AG, Park S, Chow EJ, Hasan A, Kong Y. Methane production during lactulose breath test is associated with gastrointestinal disease presentation. *Dig Dis Sci* (2003) 48(1):86–92. doi: 10.1023/A:1021738515885
  31. Chatterjee S, Park S, Low K, Kong Y, Pimentel M. The degree of breath methane production in IBS correlates with the severity of constipation. *Am J Gastroenterol* (2007) 102(4):837–41. doi: 10.1111/j.1572-0241.2007.01072.x
  32. Suri J, Kataria R, Malik Z, Parkman HP, Schey R. Elevated methane levels in small intestinal bacterial overgrowth suggests delayed small bowel and colonic transit. *Med (Baltimore)* (2018) 97(21):e10554. doi: 10.1097/MD.00000000000010554
  33. Pimentel M, Lin HC, Enayati P, van den Burg B, Lee HR, Chen JH, et al. Methane, a gas produced by enteric bacteria, slows intestinal transit and augments small intestinal contractile activity. *Am J Physiol Gastrointest Liver Physiol* (2006) 290(6):G1089–95. doi: 10.1152/ajpgi.00574.2004
  34. Park YM, Lee YJ, Hussain Z, Lee YH, Park H. The effects and mechanism of action of methane on ileal motor function. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2017) 29(9):e13077. doi: 10.1111/nmo.13077
  35. Attaluri A, Jackson M, Velestin J, Rao SS. Methanogenic flora is associated with altered colonic transit but not stool characteristics in constipation without IBS. *Am J Gastroenterol* (2010) 105(6):1407–11. doi: 10.1038/ajg.2009.655
  36. Miller TL, Wolin MJ. Methanosphaera stadtmaniae gen. nov., sp. nov.: a species that forms methane by reducing methanol with hydrogen. *Arch Microbiol* (1985) 141(2):116–22. doi: 10.1007/BF00423270
  37. Miller TL, Wolin MJ, Conway de Macario E, Macario AJ. Isolation of Methanobrevibacter smithii from human feces. *Appl Environ Microbiol* (1982) 43(1):227–32. doi: 10.1128/AEM.43.1.227-232.1982
  38. Kim G, Deepinder F, Morales W, Hwang L, Weitsman S, Chang C, et al. Methanobrevibacter smithii is the predominant methanogen in patients with constipation-predominant IBS and methane on breath. *Dig Dis Sci* (2012) 57(12):3213–8. doi: 10.1007/s10620-012-2197-1
  39. Florin TH, Woods HJ. Inhibition of methanogenesis by human bile. *Gut* (1995) 37(3):418–21. doi: 10.1136/gut.37.3.418
  40. Leite G, Morales W, Weitsman S, Celly S, Parodi G, Mathur R, et al. The microbiome in small intestinal bacterial overgrowth: the REIMAGINE study. *PLoS One* (2020).
  41. Ghoshal UC, Srivastava D, Ghoshal U, Misra A. Breath tests in the diagnosis of small intestinal bacterial overgrowth in patients with irritable bowel syndrome in comparison with quantitative upper gut aspirate culture. *Eur J Gastroenterol Hepatol* (2014) 26(7):753–60. doi: 10.1097/MEG.0000000000000122
  42. Posserud I, Stotzer PO, Björnsson ES, Abrahamsson H, Simren M. Small intestinal bacterial overgrowth in patients with irritable bowel syndrome. *Gut* (2007) 56(6):802–8. doi: 10.1136/gut.2006.108712
  43. Pylers E, Giamarellos-Bourboulis EJ, Tzivras D, Koussoulas V, Barbatzas C, Pimentel M. The prevalence of overgrowth by aerobic bacteria in the small intestine by small bowel culture: relationship with irritable bowel syndrome. *Dig Dis Sci* (2012) 57(5):1321–9. doi: 10.1007/s10620-012-2033-7
  44. Rezaie A, Pimentel M, Cohen E. Autoimmunity as a Potential Cause of Post-Infectious Gut Dysmotility: A Longitudinal Observation. *Am J Gastroenterol* (2017) 112(4):656–7. doi: 10.1038/ajg.2017.8
  45. Thabane M, Kottachchi DT, Marshall JK. Systematic review and meta-analysis: The incidence and prognosis of post-infectious irritable bowel syndrome. *Aliment Pharmacol Ther* (2007) 26(4):535–44. doi: 10.1111/j.1365-2036.2007.03399.x
  46. Cohen E, Fuller G, Bolus R, Modi R, Vu M, Shahedi K, et al. Increased risk for irritable bowel syndrome after acute diverticulitis. *Clin Gastroenterol Hepatol* (2013) 11(12):1614–9. doi: 10.1016/j.cgh.2013.03.007
  47. Klem F, Wadhwa A, Prokop LJ, Sundt WJ, Farrugia G, Camilleri M, et al. Prevalence, Risk Factors, and Outcomes of Irritable Bowel Syndrome After Infectious Enteritis: A Systematic Review and Meta-analysis. *Gastroenterology* (2017) 152(5):1042–54. doi: 10.1053/j.gastro.2016.12.039

48. Lovell RM, Ford AC. Global prevalence of and risk factors for irritable bowel syndrome: a meta-analysis. *Clin Gastroenterol Hepatol* (2012) 10(7):712–21. doi: 10.1016/j.cgh.2012.02.029
49. Scallan E, Hoekstra RM, Angulo FJ, Tauxe RV, Widdowson MA, Roy SL, et al. Foodborne illness acquired in the United States—major pathogens. *Emerg Infect Dis* (2011) 17(1):7–15. doi: 10.3201/eid1701.P11101
50. Shah ED, Riddle MS, Chang C, Pimentel M. Estimating the contribution of acute gastroenteritis to the overall prevalence of irritable bowel syndrome. *J Neurogastroenterol Motil* (2012) 18(2):200–4. doi: 10.5056/jnm.2012.18.2.200
51. Zuckerman MJ, Watts MT, Bhatt BD, Ho H. Intestinal permeability to [51Cr] EDTA in infectious diarrhea. *Dig Dis Sci* (1993) 38(9):1651–7. doi: 10.1007/BF01303174
52. Spiller RC, Jenkins D, Thornley JP, Hebden JM, Wright T, Skinner M, et al. Increased rectal mucosal enteroendocrine cells, T lymphocytes, and increased gut permeability following acute *Campylobacter* enteritis and in post-dysenteric irritable bowel syndrome. *Gut* (2000) 47(6):804–11. doi: 10.1136/gut.47.6.804
53. Dunlop SP, Hebden J, Campbell E, Naesdal J, Olbe L, Perkins AC, et al. Abnormal intestinal permeability in subgroups of diarrhea-predominant irritable bowel syndromes. *Am J Gastroenterol* (2006) 101(6):1288–94. doi: 10.1111/j.1572-0241.2006.00672.x
54. Zhang Q, Li Q, Wang C, Liu X, Li N, Li J. Enteropathogenic *Escherichia coli* changes distribution of occludin and ZO-1 in tight junction membrane microdomains in vivo. *Microb Pathog* (2010) 48(1):28–34. doi: 10.1016/j.micpath.2009.10.002
55. Hamer HM, Jonkers D, Venema K, Vanhoutvin S, Troost FJ, Brummer RJ. Review article: the role of butyrate on colonic function. *Aliment Pharmacol Ther* (2008) 27(2):104–19. doi: 10.1111/j.1365-2036.2007.03562.x
56. Hullar MA, Fu BC. Diet, the gut microbiome, and epigenetics. *Cancer J* (2014) 20(3):170–5. doi: 10.1097/PP0.0000000000000053
57. Pozuelo M, Panda S, Santiago A, Mendez S, Accarino A, Santos J, et al. Reduction of butyrate- and methane-producing microorganisms in patients with Irritable Bowel Syndrome. *Sci Rep* (2015) 5:12693. doi: 10.1038/srep12693
58. Moloney RD, Stilling RM, Dinan TG, Cryan JF. Early-life stress-induced visceral hypersensitivity and anxiety behavior is reversed by histone deacetylase inhibition. *Neurogastroenterol Motil Off J Eur Gastrointest Motil Soc* (2015) 27(12):1831–6. doi: 10.1111/nmo.12675
59. Rapin JR, Wiernsperger N. Possible links between intestinal permeability and food processing: A potential therapeutic niche for glutamine. *Clinics (Sao Paulo)* (2010) 65(6):635–43. doi: 10.1590/S1807-59322010000600012
60. Zhou Q, Verne ML, Fields JZ, Lefante JJ, Basra S, Salameh H, et al. Randomised placebo-controlled trial of dietary glutamine supplements for postinfectious irritable bowel syndrome. *Gut* (2019) 68(6):996–1002. doi: 10.1136/gutjnl-2017-315136
61. Hulsewe KW, van Acker BA, Hameeteman W, van der Hulst RR, Vainas T, Arends JW, et al. Does glutamine-enriched parenteral nutrition really affect intestinal morphology and gut permeability? *Clin Nutr* (2004) 23(5):1217–25. doi: 10.1016/j.clnu.2004.04.002
62. Dunlop SP, Jenkins D, Neal KR, Spiller RC. Relative importance of enterochromaffin cell hyperplasia, anxiety, and depression in postinfectious IBS. *Gastroenterology* (2003) 125(6):1651–9. doi: 10.1053/j.gastro.2003.09.028
63. Chadwick VS, Chen W, Shu D, Paulus B, Bethwaite P, Tie A, et al. Activation of the mucosal immune system in irritable bowel syndrome. *Gastroenterology* (2002) 122(7):1778–83. doi: 10.1053/gast.2002.33579
64. Bercik P, Wang L, Verdu EF, Mao YK, Blennerhassett P, Khan WI, et al. Visceral hyperalgesia and intestinal dysmotility in a mouse model of postinfective gut dysfunction. *Gastroenterology* (2004) 127(1):179–87. doi: 10.1053/j.gastro.2004.04.006
65. Chen B, Zhu S, Du L, He H, Kim JJ, Dai N. Reduced interstitial cells of Cajal and increased intraepithelial lymphocytes are associated with development of small intestinal bacterial overgrowth in post-infectious IBS mouse model. *Scand J Gastroenterol* (2017) 52(10):1065–71. doi: 10.1080/00365521.2017.1342141
66. Pimentel M, Chatterjee S, Chang C, Low K, Song Y, Liu C, et al. A new rat model links two contemporary theories in irritable bowel syndrome. *Dig Dis Sci* (2008) 53(4):982–9. doi: 10.1007/s10620-007-9977-z
67. Pokkunuri V, Pimentel M, Morales W, Jee SR, Alpern J, Weitsman S, et al. Role of Cytolethal Distending Toxin in Altered Stool Form and Bowel Phenotypes in a Rat Model of Post-infectious Irritable Bowel Syndrome. *J Neurogastroenterol Motil* (2012) 18(4):434–42. doi: 10.5056/jnm.2012.18.4.434
68. Pimentel M, Morales W, Pokkunuri V, Brikos C, Kim SM, Kim SE, et al. Autoimmunity Links Vinculin to the Pathophysiology of Chronic Functional Bowel Changes Following *Campylobacter jejuni* Infection in a Rat Model. *Dig Dis Sci* (2015) 60(5):1195–205. doi: 10.1007/s10620-014-3435-5
69. Bays JL, DeMali KA. Vinculin in cell-cell and cell-matrix adhesions. *Cell Mol Life Sci* (2017) 74(16):2999–3009. doi: 10.1007/s00018-017-2511-3
70. Schmulson M, Balbuena R, Corona de Law C. Clinical experience with the use of anti-CdtB and anti-vinculin antibodies in patients with diarrhea in Mexico. *Rev Gastroenterol Mex* (2016) 81(4):236–9. doi: 10.1016/j.rgmex.2016.07.002
71. Rezaie A, Park SC, Morales W, Marsh E, Lembo A, Kim JH, et al. Assessment of Anti-vinculin and Anti-cytolethal Distending Toxin B Antibodies in Subtypes of Irritable Bowel Syndrome. *Dig Dis Sci* (2017) 62(6):1480–5. doi: 10.1007/s10620-017-4585-z
72. Morales W, Rezaie A, Barlow G, Pimentel M. Second-Generation Biomarker Testing for Irritable Bowel Syndrome Using Plasma Anti-CdtB and Anti-Vinculin Levels. *Dig Dis Sci* (2019) 64(11):3115–21. doi: 10.1007/s10620-019-05684-6
73. Pike BL, Paden KA, Alcalá AN, Jaep KM, Gormley RP, Maue AC, et al. Immunological Biomarkers in Postinfectious Irritable Bowel Syndrome. *J Travel Med* (2015) 22(4):242–50. doi: 10.1111/jtm.12218
74. Talley NJ, Holtmann G, Walker MM, Burns G, Potter M, Shah A, et al. Circulating Anti-cytolethal Distending Toxin B and Anti-vinculin Antibodies as Biomarkers in Community and Healthcare Populations With Functional Dyspepsia and Irritable Bowel Syndrome. *Clin Transl Gastroenterol* (2019) 10(7):e00064. doi: 10.14309/ctg.0000000000000064
75. Gasbarrini A, Lauritano EC, Gabrielli M, Scarpellini E, Lupascu A, Ojetti V, et al. Small intestinal bacterial overgrowth: diagnosis and treatment. *Dig Dis* (2007) 25(3):237–40. doi: 10.1159/000103892
76. Bala L, Ghoshal UC, Ghoshal U, Tripathi P, Misra A, Gowda GA, et al. Malabsorption syndrome with and without small intestinal bacterial overgrowth: a study on upper-gut aspirate using 1H NMR spectroscopy. *Magn Reson Med* (2006) 56(4):738–44. doi: 10.1002/mrm.21041
77. Shah SC, Day LW, Somsouk M, Sewell JL. Meta-analysis: antibiotic therapy for small intestinal bacterial overgrowth. *Aliment Pharmacol Ther* (2013) 38(8):925–34. doi: 10.1111/apt.12479
78. Pimentel M, Lembo A, Chey WD, Zakko S, Ringel Y, Yu J, et al. Rifaximin therapy for patients with irritable bowel syndrome without constipation. *N Engl J Med* (2011) 364(1):22–32. doi: 10.1056/NEJMoa1004409
79. Lembo A, Pimentel M, Rao SS, Schoenfeld P, Cash B, Weinstock LB, et al. Repeat Treatment With Rifaximin Is Safe and Effective in Patients With Diarrhea-Predominant Irritable Bowel Syndrome. *Gastroenterology* (2016) 151(6):1113–21. doi: 10.1053/j.gastro.2016.08.003
80. Pimentel M, Cash BD, Lembo A, Wolf RA, Israel RJ, Schoenfeld P. Repeat Rifaximin for Irritable Bowel Syndrome: No Clinically Significant Changes in Stool Microbial Antibiotic Sensitivity. *Dig Dis Sci* (2017) 62(9):2455–63. doi: 10.1007/s10620-017-4598-7
81. Fodor AA, Pimentel M, Chey WD, Lembo A, Golden PL, Israel RJ, et al. Rifaximin is associated with modest, transient decreases in multiple taxa in the gut microbiota of patients with diarrhoea-predominant irritable bowel syndrome. *Gut Microbes* (2019) 10(1):22–33. doi: 10.1080/19490976.2018.1460013
82. Low K, Hwang L, Hua J, Zhu A, Morales W, Pimentel M. A combination of rifaximin and neomycin is most effective in treating irritable bowel syndrome patients with methane on lactulose breath test. *J Clin Gastroenterol* (2010) 44(8):547–50. doi: 10.1097/MCG.0b013e3181c64c90
83. Pimentel M, Chatterjee S, Chow EJ, Park S, Kong Y. Neomycin improves constipation-predominant irritable bowel syndrome in a fashion that is dependent on the presence of methane gas: subanalysis of a double-blind randomized controlled study. *Dig Dis Sci* (2006) 51(8):1297–301. doi: 10.1007/s10620-006-9104-6
84. Pimentel M, Chang C, Chua KS, Mirocha J, DiBaise J, Rao S, et al. Antibiotic treatment of constipation-predominant irritable bowel syndrome. *Dig Dis Sci* (2014) 59(6):1278–85. doi: 10.1007/s10620-014-3157-8
85. Rezaie A, Heimanson Z, McCallum R, Pimentel M. Lactulose Breath Testing as a Predictor of Response to Rifaximin in Patients With Irritable Bowel Syndrome With Diarrhea. *Am J Gastroenterol* (2019) 114(12):1886–93. doi: 10.14309/ajg.0000000000000444



86. Gupta A, Chey WD. Breath Testing for Small Intestinal Bacterial Overgrowth: A Means to Enrich Rifaximin Responders in IBS Patients? *Am J Gastroenterol* (2016) 111(3):305–6. doi: 10.1038/ajg.2016.32
87. Ford AC, Harris LA, Lacy BE, Quigley EMM, Moayyedi P. Systematic review with meta-analysis: the efficacy of prebiotics, probiotics, synbiotics and antibiotics in irritable bowel syndrome. *Aliment Pharmacol Ther* (2018) 48(10):1044–60. doi: 10.1111/apt.15001
88. Luscombe FA. Health-related quality of life and associated psychosocial factors in irritable bowel syndrome: a review. *Qual Life Res* (2000) 9(2):161–76. doi: 10.1023/A:1008970312068
89. Gaynes BN, Drossman DA. The role of psychosocial factors in irritable bowel syndrome. *Baillieres Best Pract Res Clin Gastroenterol* (1999) 13(3):437–52. doi: 10.1053/bega.1999.0038
90. Drossman DA, McKee DC, Sandler RS, Mitchell CM, Cramer EM, Lowman BC, et al. Psychosocial factors in the irritable bowel syndrome. A multivariate study of patients and nonpatients with irritable bowel syndrome. *Gastroenterology* (1988) 95(3):701–8. doi: 10.1016/S0016-5085(88)80017-9
91. Rao SSC, Rehman A, Yu S, Andino NM. Brain foginess, gas and bloating: a link between SIBO, probiotics and metabolic acidosis. *Clin Transl Gastroenterol* (2018) 9(6):162. doi: 10.1038/s41424-018-0030-7
92. Thompson JS, Rochling FA, Weseman RA, Mercer DF. Current management of short bowel syndrome. *Curr Probl Surg* (2012) 49(2):52–115. doi: 10.1067/j.cpsurg.2011.10.002
93. Jackson ML, Butt H, Ball M, Lewis DP, Bruck D. Sleep quality and the treatment of intestinal microbiota imbalance in Chronic Fatigue Syndrome: A pilot study. *Sleep Sci* (2015) 8(3):124–33. doi: 10.1016/j.slsci.2015.10.001
94. Bravo JA, Forsythe P, Chew MV, Escaravage E, Savignac HM, Dinan TG, et al. Ingestion of Lactobacillus strain regulates emotional behavior and central GABA receptor expression in a mouse via the vagus nerve. *Proc Natl Acad Sci U.S.A.* (2011) 108(38):16050–5. doi: 10.1073/pnas.1102999108
95. Leite GGS, Morales W, Weitsman S, Celly S, Parodi G, Mathur R, et al. Optimizing microbiome sequencing for small intestinal aspirates: validation of novel techniques through the REIMAGINE study. *BMC Microbiol* (2019) 19(1):239. doi: 10.1186/s12866-019-1617-1
96. Saffouri GB, Shields-Cutler RR, Chen J, Yang Y, Lekatz HR, Hale VL, et al. Small intestinal microbial dysbiosis underlies symptoms associated with functional gastrointestinal disorders. *Nat Commun* (2019) 10(1):2012. doi: 10.1038/s41467-019-09964-7
97. Linden DR. Hydrogen sulfide signaling in the gastrointestinal tract. *Antioxid Redox Signal* (2014) 20(5):818–30. doi: 10.1089/ars.2013.5312
98. Singer-Englar T, Rezaie A, Gupta K, Pichetshote N, Sedighi R, Lin E, et al. 182 - Competitive Hydrogen Gas Utilization by Methane- and Hydrogen Sulfide-Producing Microorganisms and Associated Symptoms: Results of a Novel 4-Gas Breath Test Machine. *Gastroenterology* (2018) 154(6):S-47. doi: 10.1016/S0016-5085(18)30625-5

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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