



Examination of the Addictive and Behavioral Properties of Fatty Acid-Binding Protein Inhibitor SBFI26

Panayotis K. Thanos^{1*}, Brendan H. Clavin¹, John Hamilton¹, Joseph R. O'Rourke¹, Thomas Maher¹, Christopher Koumas¹, Erick Miao¹, Jessenia Lankop¹, Aya Elhage¹, Samir Haj-Dahmane¹, Dale Deutsch² and Martin Kaczocha³

¹Behavioral Neuropharmacology and Neuroimaging Laboratory on Addictions, Research Institute on Addictions, University at Buffalo, Buffalo, NY, USA, ²Department of Biochemistry, Stony Brook University, Stony Brook, NY, USA, ³Department of Anesthesiology, Stony Brook University, Stony Brook, NY, USA

OPEN ACCESS

Edited by:

Marjin Lijffijt,
Baylor College of Medicine, USA

Reviewed by:

Luigi Janiri,
Università Cattolica del Sacro Cuore,
Italy
Gaurav Bedse,
Vanderbilt University, USA

*Correspondence:

Panayotis K. Thanos
pthanos@ria.buffalo.edu

Specialty section:

This article was submitted to
Addictive Disorders,
a section of the journal
Frontiers in Psychiatry

Received: 11 December 2015

Accepted: 24 March 2016

Published: 05 April 2016

Citation:

Thanos PK, Clavin BH, Hamilton J, O'Rourke JR, Maher T, Koumas C, Miao E, Lankop J, Elhage A, Haj-Dahmane S, Deutsch D and Kaczocha M (2016) Examination of the Addictive and Behavioral Properties of Fatty Acid-Binding Protein Inhibitor SBFI26. *Front. Psychiatry* 7:54. doi: 10.3389/fpsy.2016.00054

The therapeutic properties of cannabinoids have been well demonstrated but are overshadowed by such adverse effects as cognitive and motor dysfunction, as well as their potential for addiction. Recent research on the natural lipid ligands of cannabinoid receptors, also known as endocannabinoids, has shed light on the mechanisms of intracellular transport of the endocannabinoid anandamide by fatty acid-binding proteins (FABPs) and subsequent catabolism by fatty acid amide hydrolase. These findings facilitated the recent development of SBFI26, a pharmacological inhibitor of epidermal- and brain-specific FABP5 and FABP7, which effectively increases anandamide signaling. The goal of this study was to examine this compound for any possible rewarding and addictive properties as well as effects on locomotor activity, working/recognition memory, and propensity for sociability and preference for social novelty (SN) given its recently reported anti-inflammatory and analgesic properties. Male C57BL mice were split into four treatment groups and conditioned with 5.0, 20.0, 40.0 mg/kg SBFI26, or vehicle during a conditioned place preference (CPP) paradigm. Following CPP, mice underwent a battery of behavioral tests [open field, novel object recognition (NOR), social interaction (SI), and SN] paired with acute SBFI26 administration. Results showed that SBFI26 did not produce CPP or conditioned place aversion regardless of dose and did not induce any differences in locomotor and exploratory activity during CPP- or SBFI26-paired open field activity. We also observed no differences between treatment groups in NOR, SI, and SN. In conclusion, as SBFI26 was shown previously by our group to have significant analgesic and anti-inflammatory properties, here we show that it does not pose a risk of dependence or motor and cognitive impairment under the conditions tested.

Keywords: endocannabinoid, FABP, anandamide, addiction, conditioned place preference, reward, memory, social behavior

INTRODUCTION

The endocannabinoid system (ECS), which includes the lipid-derived neurotransmitters arachidonylethanolamide (anandamide/AEA) and 2-arachidonoyl glycerol (2-AG) (1), has been widely studied (2) and implicated in numerous functions and diseases, including schizophrenia and Parkinson's (3), pain sensation (4), anxiety (5), learning and memory (6), emotion (7), obesity (8),

and drug addiction (9). In the central nervous system, 2-AG and AEA primarily bind to the CB1 receptor (10). This discovery has led to the development of several synthetic compounds that can modify both the anabolism and catabolism of these endogenous signaling molecules.

Arachidonylethanolamide is hydrolyzed by fatty acid amide hydrolase (FAAH), which is found in abundance throughout the CNS along with CB1 receptors and is the main enzyme responsible for AEA breakdown (11, 12). FAAH inhibitors, such as URB597, significantly elevate AEA levels and have been deemed effective in alleviating depression and anxiety symptoms (13) as well as pain and inflammation (14) in rodents, and at therapeutic doses, they lack the psychomotor deficits associated with direct CB1 receptor agonists (15). However, these compounds lack specificity (16). For example, FAAH is highly expressed in the liver (17), and systemic inhibition is linked with hyperglycemia and insulin resistance (18).

Due to the fact that AEA is a lipid ligand and that its catabolizing enzyme FAAH is localized on the endoplasmic reticulum and mitochondria, it has been deemed likely that a specific mechanism is responsible for the transport of AEA through the cellular cytosol for hydrolysis (19). Fatty acid-binding proteins (FABPs) have been proposed by Kaczocha et al. (20) as the means of transport for AEA. A particular focus has been on FABP3, 5, and 7 that are highly expressed within the brain and may provide a tissue-specific approach to elevating AEA. More recent findings have emerged supporting this claim and the possibility for this transport mechanism as a target for therapeutic modification (21–25). A recently developed compound, SBFI26, inhibits two of the most prevalent FABPs in the brain (FABP5 and FABP7) (21). Kaczocha et al. (26) have shown that SBFI26 inhibits visceral, inflammatory, and neuropathic pain by increasing AEA levels in rodent brain. FABP3 (heart-FABP) was not targeted (although it is also abundant in the CNS), because its deletion has been shown to induce age-related cardiac hypertrophy (27) and apoptosis and mitochondrial dysfunction in mouse embryonic cells (28).

The effect of increased AEA levels induced by SBFI26 has revealed a gateway for therapeutic potential beyond analgesia. The abuse of prescription and non-prescription opioids, such as heroin, has transitioned from a form of self-medication to a worldwide epidemic (29, 30). Given recent evidence of endocannabinoid modulation of pain and the abuse liability of many opioids, we hypothesize that manipulation of endogenous AEA signaling by SBFI26 might have important clinical potential. It is important first though to establish whether this compound may have abuse liability or exhibit some of the characteristic negative side effects of cannabinoid compounds.

In addition to the well-known “tetrad” of cannabinoid effects [analgesia, hypothermia, catalepsy, and sedation (31)], increased activation of CB1 can lead to aversive reactions and deficits in humans and animals, sometimes in a biphasic manner that contrasts the therapeutic effects of moderate increases in CB1 activation (32). For example, elevation of AEA *via* FAAH administration has produced anxiolytic effects (33), but high doses of CB1 agonists can produce anxiogenic responses (5).

Accordingly center time in the open field, which is characterized as a measure of anxiolytic response, is significantly reduced in WT C57 mice administered THC, as is total open field locomotor activity at high doses (34). However, low doses of the synthetic CB1 agonist WIN55,212-2, URB597, and the “AEA transport inhibitor” AM404, which also raises AEA levels, did not decrease open field center time (35).

Learning and memory are also impaired by administration of THC in clinical studies (6, 36, 37) along with a variety of other CB1 agonists in rodents (38, 39). Both URB597 and the stable synthetic AEA analog R-(+)-methanandamide cause deficits in object recognition and performance in a variety of other short-term memory tests in a CB1-dependent manner (40, 41), and it was also recently demonstrated that URB597 impairs LTP in hippocampal brain slices from wild-type, but not CB1^{-/-} mice (40). CB1 receptors are abundant in hippocampus (31), and AEA, but not 2-AG, has been deemed the ligand responsible for CB1-dependent memory deficits when in excess in this brain area (42). Given these findings, it is entirely possible that SBFI26 could potentially cause cognitive and learning deficits.

Similarly, recognition memory for social stimuli is also mediated at least in part by CB1 activity. Selective deletion of CB1 in specific neuronal populations has elicited similar effects on exploration in both the object recognition and social interaction (SI)/social novelty (SN) procedures, and the hippocampus has been primarily implicated in both (43). However, preference for SI and SN is mediated by a range of additional variables, such as olfactory, auditory, and ectocrine cues, as well as unshared neural correlates such as the involvement of the hormones oxytocin and vasopressin (44). In turn, although the tests for SI and SN can be used to operationalize aspects of learning and memory as well as anxiogenic responses (34), these paradigms are reflective of combinations of multiple behaviors, are considered a model for characterizing many different psychiatric disorders, and may reveal treatment effects not observed by testing recognition memory and exploratory drive alone (45).

Perhaps most importantly, pharmacological ECS manipulation has brought up the concern of addictive properties associated with the variety of compounds used to achieve this. Squirrel monkeys, for example, have been shown to self-administer AEA, an effect blocked by CB1 antagonism (46). Conversely, Scherma et al. (47) showed that while mice receiving intravenous injection of anandamide did not induce a conditioned place preference (CPP), the introduction of URB597 along with AEA resulted in conditioned place aversion. More recently, squirrel monkeys were also shown to self-administer AM404, and this compound also reinstated drug seeking for THC and cocaine (48). Similar to THC, it appears AEA might exhibit varying qualities under particular conditions that warrant further explanation. The present study examined the effects of the specific FABP5/7 inhibitor SBFI26 on CPP, a well-established model used to measure the rewarding properties of stimuli. Additionally, we studied a range of other behaviors previously shown to be affected by endocannabinoids in attempt to characterize any cannabimimetic side effects and potential clinical utility.

MATERIALS AND METHODS

Animals

Male C57BL/6 mice (22–30 g, 9–10 weeks old, Taconic Farms) were used for all experiments ($n = 66$). The animals were single housed at room temperature (22°C) and in controlled humidity conditions and kept on a 12-h inverted light cycle beginning at 0900 hours with *ad libitum* access to water and food. Food intake and body weights were monitored daily. The animals were habituated to the experimental room for 1 week before testing. The experiments conducted herein conform to the National Institutes of Health Guidelines for the Care and Use of Laboratory Animals and were approved by the University Institutional Animal Care and Use Committee. Mice were divided into four treatment groups (vehicle, 5.0, 20.0, or 40.0 mg/kg SBFI26).

Drugs

The FABP inhibitor SBFI26 was synthesized as in Berger et al. (21). The drug was dissolved in DMSO:cremophor-EL:saline (4% DMSO:10% Cremophor-EL) and administered intraperitoneally at a volume of 10 μ l/g body weight. Doses used were determined by our previous report with SBFI26 (26).

Equipment

The CPP apparatus used was a Habitest three-chamber model (Coulbourn Instruments; Allentown, PA, USA), as previously described (49). The present study used 1" black and white stripes on two opposite walls in the left compartment, which contained crisscrossed half-inch metal flooring, whereas the right compartment consisted of perforated stainless-steel flooring with round holes on staggered centers, and 1" black and white checkering on two opposite walls.

Locomotor and exploratory behavior were examined using an open field arena and the Tru Scan software photo beam tracking system (Coulbourn Instruments, Whitehall, PA, USA). The open field arena was a 16" \times 16", and the perimeter was designated as the area within 2.5" of the walls. The center consisted of the area within the perimeter.

Novel object recognition (NOR) testing also took place in the open field arena, and objects used were similar 5" pink buckets and white vases. SI and SN testing took place in a plexiglass three-chamber apparatus, as described in Ref. (50), and used 5" black wire cups to hold stimulus mice. Tests were recorded by digital video camera.

Procedures

Conditioned Place Preference

The CPP paradigm was performed, as described in Ref. (49, 51), with some modifications (60 min conditioning sessions and 15 min pretest/test). SBFI26 or vehicle was administered 50 min prior to each conditioning session, and animals were returned to their home cage between injection and testing. The present study determined preference by calculating the percentage of time spent in one compartment compared to the other. This percentage is regarded as *percent preference*. Compartment preference for CPP was evaluated by comparing the percent time (time in

one chamber divided by total time spent in both chambers) spent in the drug-paired chamber on preconditioning day to test day. Locomotor activity during CPP was also measured each conditioning day and binned for a time period of 1 h.

Open Field Activity

Locomotor activity and exploratory behavior were measured in an open field arena for 1 h, as previously described (52). Measurements included total distance traveled (FP Distance), time spent in center (Center Time), and time spent in a rearing position (VP Time). SBFI26 injections took place 50 min before the animal entered the open field.

Novel Object Recognition

Novel object recognition was carried out, as previously described (53). During the "familiar" phase, mice were placed in the center of the arena and allowed to explore two identical objects in either corner for 5 min. They were then returned to their homecage for 30 min, during which time one of the "familiar" objects was switched with a "novel" object. Subjects were then returned to the arena for the "novel" phase, and the proportion of time spent exploring the novel object over total exploration was scored for analysis. Buckets and vases were counterbalanced, as being considered the "familiar" and "novel" objects to avoid bias. SBFI26 injections occurred 50 min prior to the middle of the "familiar" run. Exploration of an object was considered to be occurring if the test mouse was within 1" of the circumference of the object and oriented toward it. Recorded videos were scored by two research assistants, and scored were compared with an inter-rater reliability score.

Social Interaction and Novelty

Social interaction and SN were performed in a three-chambered arena with open doors, as described in Ref. (50), with minor modifications. Briefly, the test began with a 5-min habituation period in which the entire arena with both empty cups could freely be explored. Water bottles were placed atop cups to prevent climbing. The experimenter then introduced a conspecific male stimulus mouse to one of the cups for a 10-min "familiar" run (SI), followed immediately by the introduction of another conspecific male in the other cup in the "novel" run (SN). After 10 more minutes of exploration, all mice were returned to their home cages. SBFI26 injections occurred 50 min prior to the middle of the "familiar" run. Exploration of a conspecific was considered to be occurring if the test mouse was within 1" of the circumference of the cup and oriented toward it. Recorded videos were rated with Top Scan software (Clever Sys Inc., VA, USA).

Statistical Analysis

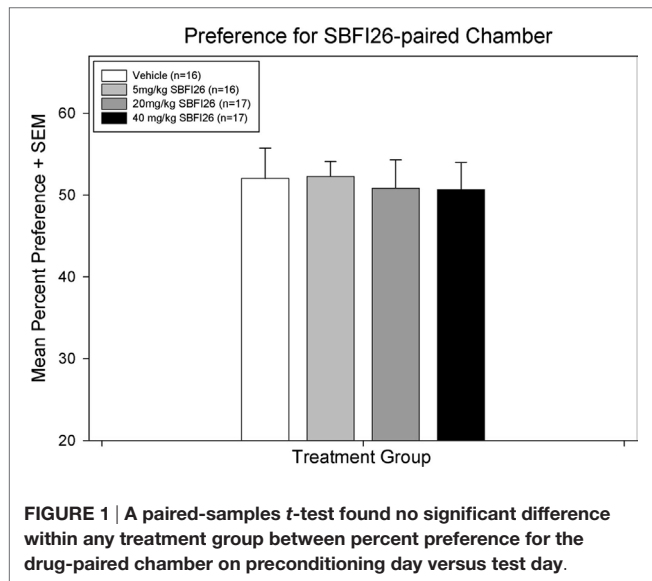
Paired-samples *t*-tests were used to calculate differences in preference between CPP preconditioning and test day for each treatment group, followed by a one-way ANOVA to detect differences in preference on test day between treatment groups. A two-way repeated measures (RM) ANOVA with SBFI26 dose and treatment (SBFI26 or vehicle) as factors was used to analyze locomotor activity throughout CPP conditioning. Open field parameters, NOR, SI, and SN were compared between treatment

groups using one-way ANOVAs. All statistics and graphing were performed using SigmaPlot 11.0 software (Systat Software Inc., San Jose, CA, USA).

RESULTS

Conditioned Place Preference

Paired-samples *t*-tests found no significant preference for the SBF126-paired chamber with any of the treatment doses administered (see **Figure 1**); vehicle [$t(12) = -1.18, p = 0.26$], 5.0 mg/kg [$t(15) = -1.87, p = 0.081$], 20.0 mg/kg [$t(15) = -1.79, p = 0.094$], and 40.0 mg/kg [$t(14) = -1.87, p = 0.082$]. There was also no difference between treatment groups in mean preference on test day [$F(3,56) = 0.060, p = 0.98$].



CPP – Locomotor Activity

A two-way RM ANOVA on average total activity between treatment groups and treatment days revealed no significant differences between groups [$F(3,58) = 1.81, p = 0.16$] or treatment day [$F(1,58) = 1.11, p = 0.30$; figure not shown].

Open Field

No differences were revealed between treatment groups with respect to FP distance, a secondary measure of locomotor activity [$F(3,53) = 0.36, p = 0.78$]. There were also no differences in time spent in the vertical plane [$F(3,53) = 1.63, p = 0.19$] and time spent in the center [$F(3,53) = 0.25, p = 0.86$], and no significant interaction detected (**Figure 2**).

Novel Object Recognition

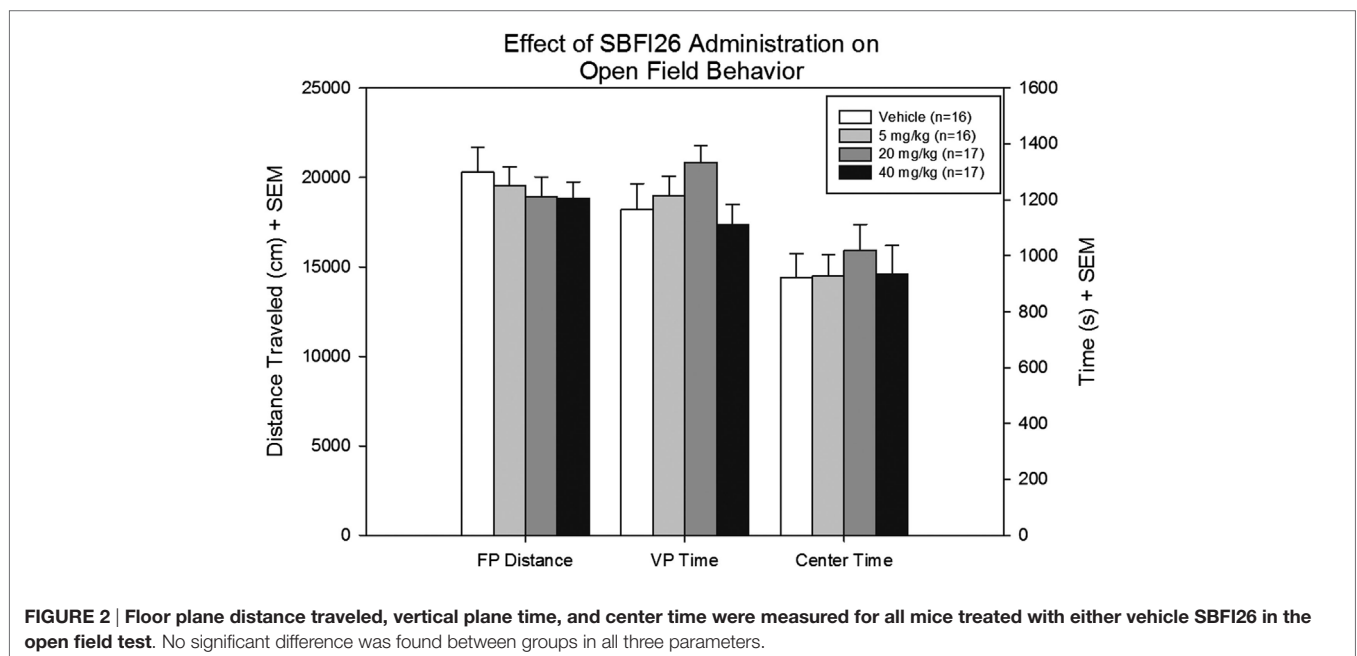
The proportion of time spent exploring the novel object did not differ with respect to SBF126 dose [$F(3,53) = 0.31, p = 0.82$]. A one-way ANOVA also did not find any significant differences in total object exploration time between treatment groups [$F(3,53) = 0.78, p = 0.51$; figure not shown].

Social Interaction and Novelty

Separate one-way ANOVAs were conducted to analyze the acute effects of SBF126 on social behavior – defined as SI and SN. No significant main effect was found for the treatment of SBF126 doses given [$F(3,38) = 2.07, p = 0.12$; figure not shown] on SI (interaction time with cup versus mouse). Similarly, no effect was found on SN (interaction time with familiar mouse versus novel mouse) for the SBF126 doses given [$F(3,38) = 1.579, p = 0.210$].

DISCUSSION

By targeting the intracellular anandamide transporters FABP5 and FABP7, this study examined the effects of SBF126 on CPP



reward behavior, locomotor and exploratory activity in the open field, NOR, SI, and SN. Results showed that SBFI26 did not induce CPP nor aversion. In addition, SBFI26 did not have any effects on locomotor activity measured during CPP conditioning or in the open field. We also found that SBFI26 did not have effects on other behaviors measured.

THC and other CB1 agonists have been shown to exhibit a biphasic effect on reward and locomotion (54) as well as anxiety (55), and elevate dopamine (DA) in the nucleus accumbens such as all known drugs of abuse (56). FAAH inhibitors vary in their ability to produce rewarding effects and increases in DA concentrations in the nucleus accumbens; these effects are specific to which compound is being administered (57), as well as route of administration (58). Therefore, it was unknown if SBFI26 would have rewarding effects. The present study demonstrates that SBFI26 does not elicit the acquisition of reward or CPP behavior or CPA (aversion). Given that we recently reported the analgesic and anti-inflammatory properties of SBFI26 (26), the use of SBFI26 for pain management may provide future promise.

In addition, SBFI26 was shown not to have any effects on multiple test parameters of locomotor and exploratory activity, working memory as measured by NOR, and propensity for SI and SN. Specifically, SBFI26 did not produce differences in activity during the CPP procedure or the open field parameters of distance traveled, time in the center, or exploratory or rearing behavior (as measured by time spent in the vertical plane). Proportion of time exploring the novel object in NOR, and time spent exploring the stimulus mouse in SI and SN also revealed no differences between SBFI26 treatment groups. Due to the previously mentioned biphasic action of cannabinoids, we chose to utilize an effective dose of SBFI26 (20.0 mg/kg) for these experiments, as well as a low and high dose to tease out potential side effects across different degrees of FABP5/7 inhibition. SBFI26 did not cause any behavioral changes in the tests conducted at any dose.

These findings are in contrast to previous studies showing deficits in NOR induced by acute doses of CB1 agonists and R-(+)-methanandamide (41), as well as URB597 (40, 42), and decreased SI time in control mice (34). The latter finding is particularly intriguing, given recent evidence that oxytocin elevates AEA in the nucleus accumbens to facilitate social reward (59). One possible explanation for the lack of significant differences in behavior in the present study is the degree to which SBFI26 raises AEA levels – URB597 has been shown to cause a fourfold increase of this endocannabinoid in mouse brain (5), whereas our group detected significant but smaller increases from SBFI26 (26). Therefore, SBFI26 may raise AEA concentrations to a degree that can still have therapeutic effects without detriments. Although we did not detect any anxiolytic effects of acute SBFI26 administration as measured by center time in the open field, this realm of research warrants further investigation given previous data of AEA elevation on anxiety (60).

These results are encouraging given the potential analgesic and anti-inflammatory properties of SBFI26, but more research is needed on the effects of SBFI26 testing on other drug abuse paradigms (i.e., self-administration, locomotor sensitization). Future research should also be directed at interactions with commonly abused drugs, such as cocaine, heroin, and marijuana.

AUTHOR CONTRIBUTIONS

All authors have significantly contributed to the research described in this article. Developed/planned the experiment: PT, SH-D, DD, and MK; performed the experiments: BC, JO, TM, CK, and EM; analyzed the data: PT, BC, TM, CK, AE, and JL; wrote the paper: PT, BC, and JH.

FUNDING

This research was funded by the Research Foundation of New York and NIH (1R01DA035923-01).

REFERENCES

- Porter AC, Sauer JM, Knierman MD, Becker GW, Berna MJ, Bao J, et al. Characterization of a novel endocannabinoid, virodhamine, with antagonist activity at the CB1 receptor. *J Pharmacol Exp Ther* (2002) **301**(3):1020–4. doi:10.1124/jpet.301.3.1020
- Hashimoto-dani Y, Ohno-Shosaku T, Kano M. Endocannabinoids and synaptic function in the CNS. *Neuroscientist* (2007) **13**(2):127–37. doi:10.1177/1073858406296716
- Piomelli D. The molecular logic of endocannabinoid signalling. *Nat Rev Neurosci* (2003) **4**(11):873–84. doi:10.1038/nrn1247
- Hohmann AG, Suplita RL II. Endocannabinoid mechanisms of pain modulation. *AAPS J* (2006) **8**(4):E693–708. doi:10.1208/aapsj080479
- Batista LA, Gobira PH, Viana TG, Aguiar DC, Moreira FA. Inhibition of endocannabinoid neuronal uptake and hydrolysis as strategies for developing anxiolytic drugs. *Behav Pharmacol* (2014) **25**(5–6):425–33. doi:10.1097/FBP.0000000000000073
- Puighermanal E, Busquets-Garcia A, Maldonado R, Ozaita A. Cellular and intracellular mechanisms involved in the cognitive impairment of cannabinoids. *Philos Trans R Soc Lond B Biol Sci* (2012) **367**(1607):3254–63. doi:10.1098/rstb.2011.0384
- Zanettini C, Panlilio LV, Aliczki M, Goldberg SR, Haller J, Yasar S. Effects of endocannabinoid system modulation on cognitive and emotional behavior. *Front Behav Neurosci* (2011) **5**:57. doi:10.3389/fnbeh.2011.00057
- Romero-Zerbo SY, Bermudez-Silva FJ. Cannabinoids, eating behaviour, and energy homeostasis. *Drug Test Anal* (2014) **6**(1–2):52–8. doi:10.1002/dta.1594
- Maldonado R, Valverde O, Berrendero F. Involvement of the endocannabinoid system in drug addiction. *Trends Neurosci* (2006) **29**(4):225–32. doi:10.1016/j.tins.2006.01.008
- Ueda N, Tsuboi K. Discrimination between two endocannabinoids. *Chem Biol* (2012) **19**(5):545–7. doi:10.1016/j.chembiol.2012.05.001
- Koutek B, Prestwich GD, Howlett AC, Chin SA, Salehani D, Akhavan N, et al. Inhibitors of arachidonoyl ethanolamide hydrolysis. *J Biol Chem* (1994) **269**(37):22937–40.
- Serrano A, Parsons LH. Endocannabinoid influence in drug reinforcement, dependence and addiction-related behaviors. *Pharmacol Ther* (2011) **132**(3):215–41. doi:10.1016/j.pharmthera.2011.06.005
- Cippitelli A, Cannella N, Braconi S, Duranti A, Tontini A, Bilbao A, et al. Increase of brain endocannabinoid anandamide levels by FAAH inhibition and alcohol abuse behaviours in the rat. *Psychopharmacology (Berl)* (2008) **198**(4):449–60. doi:10.1007/s00213-008-1104-0

14. Schlosburg JE, Kinsey SG, Lichtman AH. Targeting fatty acid amide hydrolase (FAAH) to treat pain and inflammation. *AAPS J* (2009) **11**(1):39–44. doi:10.1208/s12248-008-9075-y
15. Varvel SA, Wise LE, Niyuhire F, Cravatt BF, Lichtman AH. Inhibition of fatty-acid amide hydrolase accelerates acquisition and extinction rates in a spatial memory task. *Neuropsychopharmacology* (2007) **32**(5):1032–41. doi:10.1038/sj.npp.1301224
16. Rouzer CA. A new class of cyclooxygenase inhibitors with anti-anxiety activity. *Vanderbilt Inst Chem Biol* (2013). Available from: http://www.vanderbilt.edu/vicb/DiscoveriesArchives/cox-2_anti-anxiety.html
17. Vaithesvaran B, Yang L, Hartil K, Glaser S, Yazulla S, Bruce JE, et al. Peripheral effects of FAAH deficiency on fuel and energy homeostasis: role of dysregulated lysine acetylation. *PLoS One* (2012) **7**(3):e33717. doi:10.1371/journal.pone.0033717
18. Touriño C, Oveisi F, Lockney J, Piomelli D, Maldonado R. FAAH deficiency promotes energy storage and enhances the motivation for food. *Int J Obes (Lond)* (2010) **34**(3):557–68. doi:10.1038/ijo.2009.262
19. Ligresti A, Morera E, Van Der Stelt M, Monory K, Lutz B, Ortner G, et al. Further evidence for the existence of a specific process for the membrane transport of anandamide. *Biochem J* (2004) **380**(Pt 1):265–72. doi:10.1042/BJ20031812
20. Kaczocha M, Glaser ST, Deutsch DG. Identification of intracellular carriers for the endocannabinoid anandamide. *Proc Natl Acad Sci U S A* (2009) **106**(15):6375–80. doi:10.1073/pnas.0901515106
21. Berger WT, Ralph BP, Kaczocha M, Sun J, Balius TE, Rizzo RC, et al. Targeting fatty acid binding protein (FABP) anandamide transporters – a novel strategy for development of anti-inflammatory and anti-nociceptive drugs. *PLoS One* (2012) **7**(12):e50968. doi:10.1371/journal.pone.0050968
22. Fowler CJ. Transport of endocannabinoids across the plasma membrane and within the cell. *FEBS J* (2013) **280**(9):1895–904. doi:10.1111/febs.12212
23. Furuhashi M, Hotamisligil GS. Fatty acid-binding proteins: role in metabolic diseases and potential as drug targets. *Nat Rev Drug Discov* (2008) **7**(6):489–503. doi:10.1038/nrd2589
24. Glaser ST, Kaczocha M, Deutsch DG. Anandamide transport: a critical review. *Life Sci* (2005) **77**(14):1584–604. doi:10.1016/j.lfs.2005.05.007
25. Sanson B, Wang T, Sun J, Wang L, Kaczocha M, Ojima I, et al. Crystallographic study of FABP5 as an intracellular endocannabinoid transporter. *Acta Crystallogr D Biol Crystallogr* (2014) **70**(Pt 2):290–8. doi:10.1107/s1399004713026795
26. Kaczocha M, Rebecchi MJ, Ralph BP, Teng YH, Berger WT, Galbavy W, et al. Inhibition of fatty acid binding proteins elevates brain anandamide levels and produces analgesia. *PLoS One* (2014) **9**(4):e94200. doi:10.1371/journal.pone.0094200
27. Storch J, Thumser AE. Tissue-specific functions in the fatty acid-binding protein family. *J Biol Chem* (2010) **285**(43):32679–83. doi:10.1074/jbc.R110.135210
28. Liu YQ, Song GX, Liu HL, Wang XJ, Shen YH, Zhou LJ, et al. Silencing of FABP3 leads to apoptosis-induced mitochondrial dysfunction and stimulates Wnt signaling in zebrafish. *Mol Med Rep* (2013) **8**(3):806–12. doi:10.3892/mmr.2013.1586
29. Krashin D, Murinova N, Ballantyne J. Management of pain with comorbid substance abuse. *Curr Psychiatry Rep* (2012) **14**(5):462–8. doi:10.1007/s11920-012-0298-3
30. McCauley JL, Mercer MA, Barth KS, Brady KT, Back SE. Pain management perceptions among prescription opioid dependent individuals. *Drug Alcohol Depend* (2014) **142**:354–8. doi:10.1016/j.drugalcdep.2014.06.024
31. Mackie K. Understanding cannabinoid psychoactivity with mouse genetic models. *PLoS Biol* (2007) **5**(10):e280. doi:10.1371/journal.pbio.0050280
32. Moreira FA, Grieb M, Lutz B. Central side-effects of therapies based on CB1 cannabinoid receptor agonists and antagonists: focus on anxiety and depression. *Best Pract Res Clin Endocrinol Metab* (2009) **23**(1):133–44. doi:10.1016/j.beem.2008.09.003
33. Gunduz-Cinar O, Hill MN, McEwen BS, Holmes A. Amygdala FAAH and anandamide: mediating protection and recovery from stress. *Trends Pharmacol Sci* (2013) **34**(11):637–44. doi:10.1016/j.tips.2013.08.008
34. Boucher AA, Arnold JC, Duffy L, Schofield PR, Micheau J, Karl T. Heterozygous neuregulin 1 mice are more sensitive to the behavioural effects of Delta9-tetrahydrocannabinol. *Psychopharmacology (Berl)* (2007) **192**(3):325–36. doi:10.1007/s00213-007-0721-3
35. Gomes FV, Casarotto PC, Resstel LBM, Guimarães FS. Facilitation of CB1 receptor-mediated neurotransmission decreases marble burying behavior in mice. *Prog Neuropsychopharmacol Biol Psychiatry* (2011) **35**(2):434–8. doi:10.1016/j.pnpbp.2010.11.027
36. Curran HV, Brignell C, Fletcher S, Middleton P, Henry J. Cognitive and subjective dose-response effects of acute oral delta 9-tetrahydrocannabinol (THC) in infrequent cannabis users. *Psychopharmacology (Berl)* (2002) **164**(1):61–70. doi:10.1007/s00213-002-1169-0
37. Ranganathan M, D'Souza DC. The acute effects of cannabinoids on memory in humans: a review. *Psychopharmacology (Berl)* (2006) **188**(4):425–44. doi:10.1007/s00213-006-0508-y
38. Fadda P, Robinson L, Fratta W, Pertwee RG, Riedel G. Differential effects of THC- or CBD-rich cannabis extracts on working memory in rats. *Neuropharmacology* (2004) **47**(8):1170–9. doi:10.1016/j.neuropharm.2004.08.009
39. Varvel S, Hamm R, Martin B, Lichtman A. Differential effects of δ 9-THC on spatial reference and working memory in mice. *Psychopharmacology (Berl)* (2001) **157**(2):142–50. doi:10.1007/s002130100780
40. Basavarajappa BS, Nagre NN, Xie S, Subbanna S. Elevation of endogenous anandamide impairs LTP, learning and memory through CB1 receptor signaling in mice. *Hippocampus* (2014) **24**(7):808–18. doi:10.1002/hipo.22272
41. Kosiorek P, Hryniewicz A, Bialuk I, Zawadzka A, Winnicka MM. Cannabinoids alter recognition memory in rats. *Pol J Pharmacol* (2003) **55**(5):903–10.
42. Busquets-Garcia A, Puighermanal E, Pastor A, de la Torre R, Maldonado R, Ozaita A. Differential role of anandamide and 2-arachidonoylglycerol in memory and anxiety-like responses. *Biol Psychiatry* (2011) **70**(5):479–86. doi:10.1016/j.biopsych.2011.04.022
43. Haring M, Kaiser N, Monory K, Lutz B. Circuit specific functions of cannabinoid CB1 receptor in the balance of investigatory drive and exploration. *PLoS One* (2011) **6**(11):e26617. doi:10.1371/journal.pone.0026617
44. Bielsky IF, Young LJ. Oxytocin, vasopressin, and social recognition in mammals. *Peptides* (2004) **25**(9):1565–74. doi:10.1016/j.peptides.2004.05.019
45. Crawley JN. *What's Wrong with My Mouse?: Behavioral Phenotyping of Transgenic and Knockout Mice*. Hoboken, NJ: Wiley-Interscience (2007).
46. Justinova Z, Yasar S, Redhi GH, Goldberg SR. The endogenous cannabinoid 2-arachidonoylglycerol is intravenously self-administered by squirrel monkeys. *J Neurosci* (2011) **31**(19):7043–8. doi:10.1523/JNEUROSCI.6058-10.2011
47. Scherma M, Medalie J, Fratta W, Vadivel SK, Makriyannis A, Piomelli D, et al. The endogenous cannabinoid anandamide has effects on motivation and anxiety that are revealed by fatty acid amide hydrolase (FAAH) inhibition. *Neuropharmacology* (2008) **54**(1):129–40. doi:10.1016/j.neuropharm.2007.08.011
48. Schindler CW, Scherma M, Redhi GH, Vadivel SK, Makriyannis A, Goldberg SR, et al. Self-administration of the anandamide transport inhibitor AM404 by squirrel monkeys. *Psychopharmacology (Berl)* (2016). doi:10.1007/s00213-016-4211-3
49. Thanos PK, Kim R, Cho J, Michaelides M, Anderson BJ, Primeaux SD, et al. Obesity-resistant S5B rats showed greater cocaine conditioned place preference than the obesity-prone OM rats. *Physiol Behav* (2010) **101**(5):713–8. doi:10.1016/j.physbeh.2010.08.011
50. Moy SS, Nadler JJ, Perez A, Barbaro RP, Johns JM, Magnuson TR, et al. Sociability and preference for social novelty in five inbred strains: an approach to assess autistic-like behavior in mice. *Genes Brain Behav* (2004) **3**(5):287–302. doi:10.1111/j.1601-1848.2004.00076.x
51. Mantsch JR, Weyer A, Vranjkovic O, Beyer CE, Baker DA, Caretta H. Involvement of noradrenergic neurotransmission in the stress- but not cocaine-induced reinstatement of extinguished cocaine-induced conditioned place preference in mice: role for beta-2 adrenergic receptors. *Neuropsychopharmacology* (2010) **35**(11):2165–78. doi:10.1038/npp.2010.86
52. Delis F, Thanos PK, Rombola C, Rosko L, Grandy D, Wang GJ, et al. Chronic mild stress increases alcohol intake in mice with low dopamine D2 receptor levels. *Behav Neurosci* (2013) **127**(1):95–105. doi:10.1037/a0030750
53. Antunes M, Biala G. The novel object recognition memory: neurobiology, test procedure, and its modifications. *Cogn Process* (2012) **13**(2):93–110. doi:10.1007/s10339-011-0430-z

54. Katsidoni V, Kastellakis A, Panagis G. Biphasic effects of Delta9-tetrahydrocannabinol on brain stimulation reward and motor activity. *Int J Neuropsychopharmacol* (2013) **16**(10):2273–84. doi:10.1017/s1461145713000709
55. Kinsey SG, O'Neal ST, Long JZ, Cravatt BF, Lichtman AH. Inhibition of endocannabinoid catabolic enzymes elicits anxiolytic-like effects in the marble burying assay. *Pharmacol Biochem Behav* (2011) **98**(1):21–7. doi:10.1016/j.pbb.2010.12.002
56. Oleson EB, Cheer JF. A brain on cannabinoids: the role of dopamine release in reward seeking. *Cold Spring Harb Perspect Med* (2012) **2**(8):a012229. doi:10.1101/cshperspect.a012229
57. Justinova Z, Mascia P, Secci M, Redhi G, Piomelli D, Goldberg S. The FAAH inhibitor PF-04457845 has THC-like rewarding and reinstatement effects in squirrel monkeys and increases dopamine levels in the nucleus accumbens shell in rats (838.6). *FASEB J* (2014) **28**(1 Suppl). Available from: http://www.fasebj.org/content/28/1_Supplement/838.6.abstract
58. Solinas M, Justinova Z, Goldberg SR, Tanda G. Anandamide administration alone and after inhibition of fatty acid amide hydrolase (FAAH) increases dopamine levels in the nucleus accumbens shell in rats. *J Neurochem* (2006) **98**(2):408–19. doi:10.1111/j.1471-4159.2006.03880.x
59. Wei D, Lee D, Cox CD, Karsten CA, Penagarikano O, Geschwind DH, et al. Endocannabinoid signaling mediates oxytocin-driven social reward. *Proc Natl Acad Sci U S A* (2015) **112**(45):14084–9. doi:10.1073/pnas.1509795112
60. Gaetani S, Cuomo V, Piomelli D. Anandamide hydrolysis: a new target for anti-anxiety drugs? *Trends Mol Med* (2003) **9**(11):474–8. doi:10.1016/j.molmed.2003.09.005

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Thanos, Clavin, Hamilton, O'Rourke, Maher, Koumas, Miao, Lankop, Elhage, Haj-Dahmane, Deutsch and Kaczocha. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.