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# Combining proximal and remote sensing to assess 'Calatina' olive water status

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Developing an efficient and sustainable precision irrigation strategy is crucial in contemporary agriculture. This study aimed to combine proximal and remote sensing techniques to show the benefits of using both monitoring methods, simultaneously assessing the water status and response of 'Calatina' olive under two distinct irrigation levels: full irrigation (FI), and drought stress (DS, -3 to -4 MPa). Stem water potential ( $\Psi_{stem}$ ) and stomatal conductance (g<sub>s</sub>) were monitored weekly as reference indicators of plant water status. Crop water stress index (CWSI) and stomatal conductance index (Ig) were calculated through ground-based infrared thermography. Fruit gauges were used to monitor continuously fruit growth and data were converted in fruit daily weight fluctuations ( $\Delta W$ ) and relative growth rate (RGR). Normalized difference vegetation index (NDVI), normalized difference RedEdge index (NDRE), green normalized difference vegetation index (GNDVI), chlorophyll vegetation index (CVI), modified soil-adjusted vegetation index (MSAVI), water index (WI), normalized difference greenness index (NDGI) and green index (GI) were calculated from data collected by UAV-mounted multispectral camera. Data obtained from proximal sensing were correlated with both  $\Psi_{\text{stem}}$  and  $g_{\text{s}}$ , while remote sensing data were correlated only with  $\Psi_{\text{stem}}$ . Regression analysis showed that both CWSI and Ig proved to be reliable indicators of  $\Psi_{\text{stem}}$  and  $g_{\text{s}}$ . Of the two fruit growth parameters,  $\Delta W$  exhibited a stronger relationship, primarily with  $\Psi_{\text{stem}}$  . Finally, NDVI, GNDVI, WI and NDRE emerged as the vegetation indices that correlated most strongly with  $\Psi_{stem}$ , achieving high R<sup>2</sup> values. Combining proximal and remote sensing indices suggested two valid approaches: a more simplified one involving the use of CWSI and either NDVI or WI, and a more comprehensive one involving CWSI and  $\Delta W$  as proximal indices, along with WI as a multispectral index. Further studies on combining proximal and remote sensing data will be necessary in order to find strategic combinations of sensors and establish intervention thresholds.

#### KEYWORDS

precision irrigation, drought stress, *Olea europaea* L., crop water stress index, normalized difference vegetation index, stem water potential, UAVs

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# **1** Introduction

In recent years, climate change has created serious problems regarding the availability and use of water in agriculture, especially in areas characterized by scarce rainfall and high temperatures during the summer (Konapala et al., 2020; Pokhrel et al., 2021). These conditions of severe drought are occurring in the Mediterranean basin, creating significant challenges for local agriculture (Carella et al., 2023).

Deficit irrigation strategies have been demonstrated to be effective methods for achieving efficient water use in regions facing water scarcity (Nikolaou et al., 2020). The objective of implementing a deficit irrigation strategy is to achieve satisfactory crop yields by supplying the crop with a reduced irrigation volume compared to its potential water requirements (Tong et al., 2022). Growing crops that are well-adapted to water deficit conditions (deficit irrigation) is one of the strategies to achieve satisfactory productivity without the overuse of water resources (Gómez-Bellot et al., 2024). In this context, the olive tree (*Olea europaea* L.) emerges as a highly resilient species to water stress, with its production being influenced not only by climatic factors but also by management practices (Massenti et al., 2022b).

Another important aspect of current olive cultivation is the trend toward adopting high-density (HD) planting systems and mechanizing cultural operations. Combining the advantages of comprehensive mechanization, precision agriculture techniques, and the use of local cultivars that well adapt to the climate and soil can greatly reduce production costs associated with labor expenses (Lo Bianco et al., 2021). An olive cultivar that fits well modern growing systems (i.e., high planting density and level of mechanization) is 'Calatina', a minor Sicilian cultivar recently rediscovered thanks to its low vigor and high yield efficiency. 'Calatina' also possesses a degree of shoot bending and branching density suitable for HD and super-high-density (SHD) planting systems along with high harvesting efficiency (large fruits), demonstrating similar or better productive performance than 'Arbequina', the most widespread cultivar for HD and SHD systems (Caruso et al., 2021; Carella et al., 2022; Massenti et al., 2022a). Under HD and SHD systems, water needs and irrigation management become crucial to reach optimum yields and high quality final products. However, there are no studies on 'Calatina' water status assessment and irrigation management.

Studying how olive orchards respond to external influences presents significant challenges due to the diverse range of environments across the Mediterranean and the wide variety of planting systems employed. Prolonged periods of drought stress can trigger various physiological responses in olive trees. These include the closure of stomata (Mairech et al., 2021; Carella et al., 2023), limitation of photosynthesis (Melaouhi et al., 2021), decreased gas exchange (Pierantozzi et al., 2020), and osmotic adjustments (Scalisi et al., 2020; Abboud et al., 2021; Marino et al., 2021). When favorable water conditions return after a period of water deficit, olive trees undergo partial recovery in processes such as transpiration, photosynthesis, chlorophyll fluorescence indices, and osmotic potential (Connor, 2005; Fernandez, 2014). This implies that while tissue water content may rapidly increase, leaf function may take several days to fully restore, depending on the severity of the stress and cultivar-specific restoring capacity (Fernandez, 2014; Scalisi et al., 2020; Wahab et al., 2022).

The timing of irrigation events can be supported by data related to either soil moisture content or plant water status (Bazzi et al., 2019). The established conventional method for assessing plant water status is by measuring  $\Psi_{stem}$  using Scholander's pressure chamber. However, this method is labor-intensive and timeconsuming, and requires high skills by operators. In recent years, technological advancements have enabled the testing and introduction of various plant-based sensors for assessing plant water status and improve irrigation water management in orchard systems. While many of these sensors do not directly measure plant water status, they monitor specific physiological processes that demonstrate varying correlations with plant water status (Carella et al., 2024). However, due to the multitude of factors influencing tree physiological responses, such as tree phenological stage, environmental conditions, and genotype-specific traits, the development of simplified and standardized water management protocols using these sensors remains complex (Cocozza et al., 2015; Scalisi et al., 2020). In the meantime, remote and proximal sensing technologies for assessing field variability are becoming increasingly common in precision agriculture. This trend is propelled by their relatively lower costs and non-invasive nature compared to conventional methods (Caruso et al., 2022a).

The use of proximal sensing techniques, such as infrared thermography (Ben-Gal et al., 2009) and daily fruit diameter variation by fruit gauges (Morandi et al., 2007; Boini et al., 2019; Giovannini et al., 2022; Khosravi et al., 2022; Carella et al., 2023), are potentially effective and continuous methods to assess plant water status. Scalisi et al. (2020) observed that coupling  $\Psi_{stem}$  with fruit gauges and leaf patch clamp pressure probes (LPCP probes) data can be a reliable tool for evaluating fruit tree water status and smarter irrigation management. Moreover, García-Tejero et al. (2018) determined that the infrared thermography approach has a great advantage due to the large amount of information provided. The use of infrared thermography has proved to be a suitable technique for monitoring the water status of fruit trees (Egea et al., 2016; García-Tejero et al., 2018; Blanco et al., 2023). In details, the most utilized method for plant water status assessment involves the normalization of canopy temperature through the calculation of some indices, e.g., crop water stress index (CWSI) and stomatal conductance index (Ig) (Jackson et al., 1981; Idso, 1982; Jones et al., 2002).

Remote sensing techniques enable the rapid detection of spatial variability of tree water status over large areas using thermal, multispectral and hyperspectral cameras (Carella et al., 2024). Among the most common platforms used in remote sensing for detailed scale irrigation management are Unmanned Aerial Vehicles (UAVs) (Roma and Catania, 2022). Several studies have found that remote sensing over olive orchards can provide an accurate estimation of tree water status. Most studies emphasize that remote sensing with thermal cameras is highly correlated with plant water status (Ben-Gal et al., 2009; Egea et al., 2016; Caruso et al., 2022b). However, these techniques have some limitations, mainly related to the calculation method used to determine the CWSI (Berni et al., 2009). Nonetheless, detection of spectral plant

condition also allows for an estimation of plant water status (Marino et al., 2014; Rallo et al., 2014; Marques et al., 2023). Indeed, several studies have obtained correlations between various vegetation indices and plant water conditions. Among the most used indices, we find normalized difference vegetation index (NDVI, Rouse et al., 1974), normalized difference RedEdge (NDRE, Maccioni et al., 2001), and indices that utilize the Short Wave InfraRed (SWIR) band. Although the latter indices are very efficient, they cannot be obtained from normal multispectral cameras (Herrmann et al., 2010). For this reason, water stress conditions are increasingly being investigated and detected with normal multispectral cameras, as they are cheaper and easier to use.

The combined use of proximal and remote sensing techniques can offer a more comprehensive and accurate indication of plant water status and irrigation requirements. Proximal sensors provide accurate and continuous real-time data for individual plants, while data from UAVs or satellites can extend coverage across the field (Matese et al., 2018; Jin et al., 2022; Carella et al., 2024). Remote sensing data provide valuable information on spatial variability through effective field mapping, allowing the strategic positioning of proximal sensors only in certain areas of the field (Alexopoulos et al., 2023; Roma et al., 2023). Data collected by soil electrical conductivity sensors and UAVs were used by Caruso et al. (2022) to delineate homogeneous zones within a densely irrigated olive orchard. They observed that tree water use efficiency (WUE) varies according to the placement within the orchard. Furthermore, they found that tree vigor emerges as a predominant factor affecting the final fruit yield under optimal water availability.

On these bases, this study aims to combine proximal and remote sensing techniques to show the benefits of using both monitoring methods, while simultaneously assessing the water status and response of 'Calatina' olive under two distinct irrigation regimes.

# 2 Materials and methods

The experiment was conducted from summer to fall 2023, in a high-density olive orchard (6 x 2 spacing, 833 trees/ha) located near Sciacca (37°29' N and 13°12' E, 138 m a.s.l., Figure 1A). Eleven-year-old own-rooted trees were trained to "free palmette" along North-South-oriented hedgerows. The Sicilian cultivar Calatina was selected as potentially suitable for new high-density plantings mainly due to its low vigor, high productivity and good olive oil quality (Massenti et al., 2022a). The soil is a sandy-clay-loam (60% sand, 18% silt, 22% clay), with pH 7.7 and low active carbonates (<5%). Trees were regularly fertilized and pruned according to ordinary practices.

Air temperature (T) and relative humidity (RH) were measured at one-hour interval using an Elitech RC-51H data-logger (Elitech, London, UK), placed in the orchard within the experimental plot. RH (%) and T (°C) data were used to calculate vapor pressure deficit (VPD, kPa) with the following equation (Chazarreta et al., 2023):

$$VPD = VP_s - VP_a$$

where:

$$VP_{s}(saturatedvaporpressure) = 0.6108^{\frac{17.271}{T+237.3}}$$

$$VP_a(actualvaporpressure) = \frac{RH}{100 VP_s}$$

Trees were irrigated weekly using two self-compensating in-line drippers per tree, each delivering 16 L/h. Two irrigation levels were imposed, selected based on two ranges of stem water potential: Full Irrigation (FI), maintaining the tree water potential at approximately -1.5 to -2.5 MPa (conventional farm management), and Drought Stress (DS), maintaining the tree water potential within the range of -3.5 to -4 MPa, according to the thresholds



#### FIGURE 1

Experimental site location (A); Experimental plot with Crop Surface Model (CSM) of the olive orchard, and full irrigation (FI) and drought stress (DS) treatments design (B).

defined in Marra et al. (2016) and Marino et al. (2018). Nine plants per treatment were selected and divided into three blocks of three plants each (Figure 1B). A buffer plant separates each block to avoid influence between treatments.

Measurements were carried out at stages III (cell expansion) and IV (maturation) of fruit development.

## 2.1 Plant water status

Midday stem water potential ( $\Psi_{stem}$ ; MPa) was the main reference for evaluation of plant water status.  $\Psi_{stem}$  was measured with a Scholander's pressure chamber (PMS 1000, Instrument Company, Albany, OR, USA). Measurements were taken at around 13:00 on four shoots per plant, each covered with aluminum foil 1 hour before measurement (Moriana et al., 2012). Leaf stomatal conductance ( $g_s$ ; mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-2</sup>) and net photosynthesis ( $P_n$ ; µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-2</sup>) were measured weekly using a portable gas exchange system CIRAS-2 (PP Systems<sup>®</sup>, Hitchin, UK) on two sun-exposed leaves per plant (Marino et al., 2018). Both parameters were measured once a week, specifically on 14 July, 1, 8, 16, 22 and 29 August; 6, 11, 19 and 26 September; and 3 and 10 October.

## 2.2 Proximal sensing measurements

### 2.2.1 Thermal imaging

Infrared thermal images were taken with a FLIR i7 hand-held thermal camera (FLIR systems, Wilsonville, Oregon, USA), with a resolution of 140 x 140 pixels and a spectral range of 7.5-13 µm. The camera has a thermal sensitivity of 0.1°C and an accuracy of  $\pm$  2%. Emissivity was set at 0.98, according to Rubio et al. (1997). Thermal images were taken weekly, at the same time of the other measurements. Each image included the canopy of each individual tree, along with a fully transpiring reference (Tweet as a non-water stressed baseline) and a non-transpiring reference (T<sub>dry</sub>). The references were obtained following the methodology proposed by Jones et al., 2002. Twee was determined using leaves sprayed with water and a drop of detergent (0.01% v/v) (Fuentes et al., 2012) 1 minute before taking the thermal image as wet references. For T<sub>dry</sub>, leaves were covered with petroleum jelly about 1 h before measurement to artificially close stomata and inhibit transpiration. Crop water stress index (CWSI) was calculated using the following equation:

$$CWSI = \frac{T_c - T_{wet}}{T_{dry} - T_{wet}}$$

 $T_c$  represented the actual temperature of the canopy,  $T_{wet}$  and  $T_{dry}$  were the lower and upper limits of canopy temperature. Thermal data were extracted using the ThermaCAM Researcher Pro 2.10 software (FLIR systems, Wilsonville, Oregon, USA) by manually selecting the regions of interest (ROIs), avoiding the empty spaces, and the wet and dry references. All temperature values corresponded to the average temperature of pixels within the selected area. In addition, according to

Jones et al. (2002) a further index related to stomatal conductance (stomatal conductance index - Ig) was calculated by using the same references of CWSI, i.e., with the equation:

$$Ig = \frac{T_{dry} - T_c}{T_c - T_{wet}}$$

 $T_{c}$ ,  $T_{dry}$  and  $T_{wet}$  were the same parameters utilized in CWSI equation. Ig may also represent a reliable index for assessing plant water status, as it is theoretically proportional to  $g_s$ , as observed in several studies (Jones et al., 2002; Costa et al., 2012; Belfiore et al., 2019). Thermal images were taken on 1, 8, 16, 22 and 29 August; 6, 11, 19 and 26 September; and 3 and 10 October.

### 2.2.2 Fruit-based sensing

Fruit diameter was continuously monitored during the entire period, at 15-min intervals, with the fruit gauges described by Morandi et al. (2007), wired to a CR-1000 data logger (Campbell Scientific, Inc., Logan, UT, USA). Drupes were continuously measured using 10 fruit gauges, with one sensor per plant for a total of five plants per treatment. Fruit gauges were placed on sun exposed fruits at about 1.5 m from the ground (corresponding to medium canopy height). At the end of the measurement period, the fruit equatorial diameter was converted to fruit weight, as suggested by (Morandi et al., 2007). The following equation was used for the conversion:

$$W(g) = a \times D(mm)^{b}$$

where W was the fruit weight and D the diameter. For our fruit in the experiment, a and b were 0.003 (± 0.0004 S.E.) and 2.59 (± 0.052 S.E.), respectively, with  $R^2 = 0.960$ . This equation was obtained by regressing diameter and weight data of 200 fruits randomly sampled from plants exposed to the two irrigation levels throughout the trial period. The stages of fruit growth were identified by monitoring the diameter and weight (Rallo and Rapoport, 2001; Trentacoste et al., 2010). Subsequently, fruit daily weight fluctuations ( $\Delta W$ , g) and relative growth rate (RGR, mg g<sup>-1</sup> min<sup>-1</sup>), were calculated.  $\Delta W$  was obtained from the subtraction between the maximum and the minimum daily weight averaged for all monitored fruits. RGR was calculated from the absolute growth rate (AGR, g min<sup>-1</sup>) of individual fruits. In details, AGR and RGR were calculated as follows:

and

$$RGR = \frac{AGR}{W_0}$$

 $AGR = \frac{W_1 - W_0}{t_1 - t_0}$ 

In the equations,  $W_1$  and  $W_0$  were the fruit weights at time  $t_1$  and  $t_0$ , respectively. RGR provides an indication of dry mass accumulation in the fruit, while  $\Delta W$  is primarily related to fruit water exchanges through the xylem and transpiration (Carella et al., 2021). Such parameters were correlated with  $\Psi_{stem}$  and  $g_s$  in order to evaluate how the fruits responded to changes in plant water status, in terms of water exchanges and carbohydrates uptake.

# 2.3 Remote sensing measurements

# 2.3.1 Flight scheduling and multispectral data acquisition

A DJI Phantom 4 Multispectral UAV (DJI, Shenzhen, China) equipped with a multispectral camera was used to obtain the reflectance in different narrow bands of the electromagnetic spectrum of each plant. Specifically, the camera has six 1/2.9" CMOS sensors, i.e. one RGB sensor and five monochrome sensors with band centers in Blue (B, 450 ± 16nm), Green (G, 560 ± 16nm), Red (R, 650 ± 16nm), RedEdge (RE, 730 ± 16nm) and Near InfraRed (NIR, 840 ± 26nm). Two flights were carried out on 19 September and 10 October with automatic flight configuration using the way-points and RTK mode for correcting the GNSS signal. The flights were performed at about 13:00 at an altitude of 70 m, generating a Ground Surface Distance (GSD) of 3.6 cm. Before each flight, a calibration panel and 10 Ground Control Points (GCP) were positioned and georeferenced with the Stonex S7-G instrument according to Roma et al. (2023). The image acquisition was made in stop-and-go mode with 70% front and side overlap ratio, while the gimbal pitch was set at 90° (downwards).

## 2.3.2 Image processing

The photogrammetric reconstruction was carried out with Agisoft Photoscan Professional 1.7.3. (Agisoft Metashape, Saint Petersburg, Russia) using structure-from-motion (SfM) algorithms to obtain the multi-band orthomosaic and Digital Elevation Model (DEM). The OBIA (Object-Based Image Analysis) of geo-spatial and multispectral data was performed in the open-source software QGIS 3.2 (QGIS Geographic Information System). Specifically, orthomosaic segmentation and classification to separate the canopy from the background were carried out using the machine learning algorithm implemented in the Orfeo Tool Box (OTB tool). Once the canopies were obtained, the spectral information for each tree was extracted using Statistical Zone tools (Stateras and Kalivas, 2020; Roma et al., 2023). To obtain the information of each tree, a centroid of plants was identified, and a sub-plot of  $6 \ge 2$  m was built for each one (Figure 2). The digital numbers (DN) inside of each sub-plot were used to evaluate the spectral and geometric features per plant.

The spectral information was related to the determination of several water sensitive vegetation indices used in the literature (Table 1).

# 2.4 Statistical analysis

The means of  $\Psi_{\text{stem}}$ ,  $g_s$  and  $P_n$  of FI and DS plants were compared by using repeated measure ANOVA at the 0.05 significance level using Jamovi 2.4.14 procedures (The Jamovi Project, 2023). Linear and nonlinear regression analysis were performed to relate the parameters obtained from proximal and remote sensing with  $\Psi_{\text{stem}}$  and  $g_s$  using Sigmaplot 14.0 (Systat Software Inc., Chicago, IL, USA) procedures. The remote sensing data were analyzed with two-way analysis of variance using date and irrigation levels as main factors, followed by Tukey's *post hoc* test.

# **3** Results and discussion

# 3.1 Climate and irrigation data

As expected, the highest values of VPD were recorded in July. Specifically, the highest VPD was recorded on 18 July. In contrast,



Vegetation index	Acronym	Equation	Reference	
Chlorophyll Vegetation Index	CVI	(pNIR/pG) * (pR+pG)	Vincini et al. (2008)	
Green Index	GI	ρG/pR	Zarco-Tejada et al. (2005)	
Green Normalized Difference Vegetation Index	GNDVI	(pNIR–pG)/ (pNIR+pG)	Gitelson et al. (1996)	
Modified Soil- Adjusted Vegetation Index	MSAVI	$\begin{array}{l} 0.5 * \{ 2 \ \rho \text{NIR} + 1 \ - \\ \text{SQRT}[(2 \ \rho \text{NIR} + 1)^2 \ - \ 8 \\ (\rho \text{NIR} \ - \ \rho \text{R})] \} \end{array}$	Qi et al. (1994)	
Normalized Differential Greenness Index	NDGI	(ρG–ρR)/(ρG+ρR)	Pôças et al. (2015)	
Normalized Difference RedEdge Index	NDRE	(pNIR–pRedEdge)/ (pNIR+pRedEdge)	Maccioni et al. (2001)	
Normalized Difference Vegetation Index	NDVI	(pNIR–pR)/(pNIR+pR)	Rouse et al. (1974)	
Water Index	WI	ρR/pNIR	Peñuelas et al. (1993)	

TABLE 1 Vegetation indices considered to determine tree water status, using wavelengths within the VIS-NIR range.

G, Green; R, Red; NIRm, Near Infrared spectral bands;  $\rho$ , reflectance.

the lowest level was reached on 16 October (Figure 3A). Seven rainfall events occurred during the trial, totaling 51.2 mm of rain. The most intense rainfall events occurred on 8 and 23 September (22 mm and 12 mm, respectively). In FI trees, irrigation was carried

out weekly with a total of 40 mm (Figure 3B), for a total of 571.2 mm (Table 1) (including rainfall). No irrigation was applied between 6 September and 19 September due to a district water shortage. In DS trees, emergency irrigation was carried out when  $\Psi_{\text{stem}}$  went below -4 MPa, and near fruit ripening, totaling 291.2 mm (including rainfall).

## 3.2 Tree water status and gas exchange

At the beginning of the trial, all trees received the same amount of irrigation water; indeed, no significant differences were found between FI and DS trees in terms of  $\Psi_{stem}$ , g<sub>s</sub>, and P<sub>n</sub> (Figure 4). Regarding  $\Psi_{stem}$ , significant differences were observed between irrigation treatments from 1 August until 19 September (Figure 4A). No significant differences were observed from 26 September until the end of the experiment (10 October). On 26 September,  $\Psi_{stem}$  values were above -2 MPa in both FI and DS trees, due to both rewatering and a rainy event on 23 September. A slight increase in  $\Psi_{\text{stem}}$  in DS plants was observed on 29 August, following a light rainfall event. The lowest  $\Psi_{stem}$  level was recorded on 19 September in DS plants; however, a low value (< 2.5 MPa) was also observed in FI plants on the same date. This was due to a temporary water network failure, so no irrigation was carried out on 18 September. For this reason, irrigation was postponed to 20 September.

The trend of  $g_s$  over time was consistent with that of  $\Psi_{stem}$  for almost the entire experiment (Figure 4B). On 1 August, despite the differences in  $\Psi_{stem}$ , no significant differences of  $g_s$  were observed between the two irrigation treatments. Most likely, hydration levels were not sufficiently low for the trees to exhibit different stomatal



Daily mean vapor pressure deficit (VPD, (A) and water supply (irrigation and rainfall, (B) to full irrigated (FI), and drought stressed (DS) 'Calatina olive trees.

FIGURE 3

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FIGURE 4

Trends of midday stem water potential ( $\Psi_{stem}$ , **A**), stomatal conductance (g<sub>s</sub>, **B**) and net photosynthesis (P<sub>n</sub>, **C**) in 'Calatina' olive trees from 14 July to 10 October 2023. Black and white dots represent full irrigated (FI) and drought stressed (DS) trees, respectively. Error bars indicate standard errors of means. n.s., no significantly different; \*, significantly different for P < 0.01; \*\*\*, significantly different for P < 0.01; means of the threshold between cell expansion and maturation stage, corresponding to about 23 September 2023.

opening behavior. In subsequent dates, significant differences in  $g_s$  between irrigation treatments were observed until 19 September. After this date, despite the rainfall events and rewatering, 'Calatina' trees seemed to keep memory of the watering differences. Indeed, DS significantly reduced  $g_s$  on 3 October. This behavior may have been mediated by chemical signals like abscisic acid, possibly accumulated in the roots and transferred to the shoots where it causes a  $g_s$  reduction (Brito et al., 2020).

Regarding  $P_n$ , no significant differences between the two irrigation treatments were observed until 8 August (Figure 4C). On this date, despite differences in  $g_s$ ,  $P_n$  of FI and DS trees was not statistically different. In other words, despite the reduction in stomatal closure, similar levels of  $CO_2$  were assimilated, suggesting an increase water use efficiency. This usually happens when stomatal closure is only partial and it decreases water loss more than  $CO_2$  intake due to the different partial pressures of the two gasses (Lawson and Blatt, 2014). On 15 and 22 August,  $P_n$  was significantly reduced by DS. As in  $\Psi_{stem}$ , no significant differences of  $P_n$  were observed from 26 September until the end of the experiment.

Within the observed  $\Psi_{stem}$  range (from -4.7 to -1.6), the relationship with  $g_s$  was described by a direct exponential model (Figure 5A). Specifically, the model shows a strong direct relationship from -1.6 to -2.7 MPa, exhibiting a continuous decrease in  $g_s$  sensitivity as  $\Psi_{stem}$  decreases. In other words, as water stress progresses, stomatal response loses sensitivity and the stomatal closure is gradually lost; as a result, minimal transpiration is maintained. This behavior has been already reported in olive (Moriana et al., 2012; Marino et al., 2018).

 $P_n$  ranged from 4.9 to 18.3 µmol CO<sub>2</sub> m<sup>-2</sup> s<sup>-1</sup>. In the observed  $\Psi_{stem}$  range, the relationship between  $\Psi_{stem}$  and  $P_n$  was also best represented by a direct exponential model. As  $\Psi_{stem}$  decreased,  $P_n$  decreased more slowly, gradually reducing sensitivity to  $\Psi_{stem}$  (Figure 5B). This kind of relationship was also observed in 'Arbequina' olive (Marino et al., 2018; Ahumada-Orellana et al., 2019).



FIGURE 5

Relationships between stem water potential ( $\Psi_{stem}$ ) and stomatal conductance [g<sub>s</sub>, **(A)**] and net photosynthesis [P<sub>n</sub>, **(B)**] in 'Calatina' olive trees from 1 August to 10 October 2023.

## 3.3 Proximal sensing

### 3.3.1 Thermal imaging

A negative linear relationship between CWSI and  $\Psi_{stem}$  was observed (Figure 6A). CWSI ranged from approximately 0.35 to 0.8 MPa, while  $\Psi_{stem}$  ranged from about -1.6 to -4.7 MPa. This indicates that canopy temperature normalized with CWSI is proportional to changes in the water status of 'Calatina' trees. Therefore, CWSI calculated using thermal imagery proved to be a useful parameter for plant water stress assessment, serving as an alternative to the more laborious measurement of  $\Psi_{stem}$  by using the pressure chamber. Similar ranges and relationships between  $\Psi_{\text{stem}}$  and CWSI have been documented in 'Arbequina' (Egea et al., 2016) and in 'Barnea' and 'Cobrançosa' (Ben-Gal et al., 2009; Marques et al., 2023) olive. Caruso et al. (2022b) found a significant relationship between remotely sensed CWSI and  $\Psi_{stem}$ in 'Frantoio' and 'Leccino' olive, with values comparable to those observed in this study. Linear relationships between the two parameters have also been documented in other fruit species such as cherry (Blaya-Ros et al., 2020), apple (Mohamed et al., 2021), peach (Ramírez-Cuesta et al., 2022) and grapevine (Pou et al., 2014).

On the other hand, the relationship between CWSI and  $g_s$  followed an inverse exponential model (Figure 6B) similar to the one between  $\Psi_{stem}$  and  $g_s$ . In this case, CWSI increased almost linearly as  $g_s$  decreased up to approximately 200 mmol H<sub>2</sub>O m<sup>-2</sup> s<sup>-1</sup>. Below this value, CWSI was more sensitive to  $g_s$  changes, and increased more rapidly. To date, mainly linear relationships between CWSI and  $g_s$  have been documented in olive (García-Tejero et al., 2017; Marques et al., 2023). However, due to the small

number of olive cultivars on which this relationship has been studied, it is likely that the latter may vary depending on the cultivar.

The relationship between  $g_s$  and  $\Psi_{stem}$  also followed a direct exponential function (Figure 5A). Since Ig was developed as thermal index for estimation of stomatal conductance, the relationship between Ig and  $\Psi_{stem}$  (Figure 6C) was similar to the one between  $\Psi_{stem}$  and  $g_s$ . Specifically, from -1.6 toward -4.7 MPa there was a tendency of Ig to lose sensitivity to changes in  $\Psi_{stem}$  as the latter decreased.

A positive linear relationship between Ig and  $g_s$  was observed (Figure 6D), confirming that Ig is a more direct estimator of stomatal conductance. Similar relationships were found in several studies with other species (Jones et al., 2002; Reinert et al., 2012; Yu et al., 2015).

## 3.3.2 Fruit based-sensing

For associations with  $\Delta W$  and RGR only the period of cell expansion was taken into account, as the fruit exhibits minimal response to changes in water status during the maturation stage. In fact, during maturation stage, internal and external changes in the fruit texture, flavor, and color prevail over water exchanges (Giovannoni, 2001; Carella et al., 2021). On the other hand, when the fruit is in the stage of cell division, phloem and xylem contribution are similar (Morandi et al., 2007). During this phase, carbohydrate intake is essential. They are mainly sourced from actively photosynthesizing leaves and transported into the fruit through the phloem (Génard et al., 2008). On the contrary, during pit hardening stage, fruit water flows are very limited. At this stage,



#### FIGURE 6

Relationships between stem water potential ( $\Psi_{stem}$ ) and crop water stress index [CWSI, (**A**)], stomatal conductance ( $g_s$ ) and CWSI (**B**),  $\Psi_{stem}$  and stomatal conductance index [Ig, (**C**)], and  $g_s$  and Ig (**D**) in 'Calatina' olive trees from 1 August to 10 October 2023.

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water deficit do not affect fruit growth (Goldhamer, 1997; Moriana et al., 2003; Corell et al., 2022). At fruit cell expansion stage, a direct exponential relationship between  $\Delta W$  and  $\Psi_{stem}$  was observed (Figure 7A). Specifically, as  $\Psi_{stem}$  increased, fruit weight fluctuations become more and more pronounced, increasing its response sensitivity. At  $\Psi_{\text{stem}}$  values below -2.5 MPa,  $\Delta W$  began to lose response sensitivity, suggesting that below this threshold fruit water flows tended to stabilize. This relationship between  $\Delta W$  and  $\Psi_{\text{stem}}$  suggests that below this  $\Psi_{\text{stem}}$  value, the drupe may promptly respond to water stress by enhancing its water retention capacity. Specifically, a gradual fruit stomatal closure may be occurred, decreasing fruit transpiration rate and maintaining appropriate tissue hydration (Lescourret et al., 2001; Morandi et al., 2010). Secondly, a fruit osmotic adjustment may be occurred, leading to the accumulation of solutes capable of decreasing the osmotic potential and contributing to maintain tissues hydration. In olive, fruit osmotic adjustment was documented by Girón et al. (2015) in cv. Manzanillo. To date, few studies have correlated fruit  $\Delta W$  and  $\Psi_{\text{stem}}$ . In 'Arbequina' olive, Fernandes et al. (2018) observed a linear relationship between the two parameters in full irrigated plants, while no relationship was observed in trees under water deficit. Furthermore, it is worth noting that the 'Calatina' fruit maintained a high water exchange capacity even when the  $\Psi_{stem}$  ranged from about -3 to -2.5 MPa (mild water stress condition).

A positive linear relationship was observed between  $\Delta W$  and  $g_s$  (Figure 7B). However, such relationship proved to be less tight than the relationship between  $\Psi_{stem}$  and  $\Delta W$ . A direct exponential relationship was also observed between RGR and  $\Psi_{stem}$  (Figure 7C). It is interesting to note that RGR is markedly higher at  $\Psi_{stem}$  levels between -2 and -1.6 MPa, further suggesting that

maintaining plants within that range ensures optimal fruit growth rates. Conversely, RGR began to approach zero at  $\Psi_{stem}$  of about -3 MPa, losing sensitivity to further decreases of  $\Psi_{stem}$ , and reaching slightly negative values at  $\Psi_{stem}$  below -3 MPa. In other studies, mainly linear relationships between fruit growth rate and  $\Psi_{stem}$  were reported (Boini et al., 2019; Scalisi et al., 2020; Marino et al., 2021). However, the range of  $\Psi_{stem}$  examined was narrower than in this study.

No significant relationship was found between RGR and gs, suggesting that fruit growth mainly responded to changes in  $\Psi_{stem}$ rather than g<sub>s</sub> (Figure 7D). In 'Gala' apple, Boini et al. (2019) studied the correlations between fruit growth parameters,  $\Psi_{stem}$  and gas exchanges. The Authors found that the fruit growth rate exhibited the strongest correlation with  $\Psi_{stem}$ . Interestingly, the fruit daily size fluctuations, in contrast to this study, showed a stronger relationship with  $g_s$  rather than with  $\Psi_{stem}\!.$  This discrepancy may stem from measurements taken during the late stages of apple cell expansion, where factors influencing fruit external and internal changes may differ from olive. For example, in apples, simple sugar accumulation prevails, while in olives, oil accumulation and oleuropein degradation occur (Tombesi, 1993; Tijero et al., 2021). As expected, no significant relationships between fruit growth parameters and  $\Psi_{stem}$  or  $g_s$  were identified after the cell expansion stage in our study.

## 3.4 Remote sensing

The experiments allowed the investigation of the spectral conditions of each plant as a function of  $\Psi_{stem}$ . Specifically,



FIGURE 7

Relationships between fruit daily weight fluctuations ( $\Delta W$ ) and stem water potential [ $\Psi_{stem}$ , (**A**)],  $\Delta W$  and stomatal conductance [ $g_s$ , (**B**)], fruit relative growth rate [RGR, (**C**)] and  $\Psi_{stem}$ , and RGR and  $g_s$  (**D**) in 'Calatina' olive trees from 1 August to 26 September 2023.

different behavior was observed in the different bands and consequently also in the vegetation indices (Figure 8). In the VIS and NIR zones, the average reflectance of the vegetation showed the typical plant trend. Indeed, in the visible zone of the electromagnetic spectrum (400 - 700 nm), a higher reflectance of DS plants compared to FI plants was observed (Figure 8A). In the NIR zone (700 - 900 nm), the opposite trend was observed. Specifically, the reflectance in the bands of blue, green, red, RedEdge and NIR were 3.8% 8.4%, 5.6%, 25.3% and 38%, respectively in the FI trees, while they were 4.1%, 9.02%, 6.3%, 25.4% and 37% in the DS trees. Similar reflectance patterns have also been observed in other studies conducted on olive trees (Rallo et al., 2014) as well as other crops (Pôças et al., 2017). NDVI is considered the reference vegetation index for plant vigor and health. In this regard, the differences between FI and DS plants are evident only on September 19 (Figure 8B), where the  $\Psi_{stem}$  range was wider (Figure 4A).

All the indices were higher in FI than in DS plants, except for the WI (Table 2). Indeed, while all the other indices are used to estimate plant growth and health conditions (Noori and Panda, 2016; Jorge et al., 2019; Jurado et al., 2020; Stateras and Kalivas, 2020), the WI is an index directly related to water stress detection (Peñuelas et al., 1993; Pôças et al., 2017). Furthermore, an increase of all the indices is observed as the season progresses. All indices were able to differentiate between the two irrigation treatments at the time of greater water stress (19 September), lower  $\Psi_{\text{stem}}$ . Furthermore, only NDRE, GNDVI, WI, NDGI, and GI were able to differentiate the two treatments also in October. The linear relationships between indices calculated from UAV multispectral camera and  $\Psi_{stem}$  on 19 September and 10 October were split by date because they showed significantly different slopes as a function of date (Table 3). In detail, significant differences between slopes were found in all the vegetation indices taken into account. This can be explained because on 10 October, trees were more hydrated, which reduced the measured range of  $\Psi_{stem}$ , making the changes in indices not easily appreciable. Indeed, on 10 October, the regressions slopes were less steep than those on 19 September.

A strong and significant positive linear relationship was found between NDVI and  $\Psi_{stem}$  on 19 September (Figure 9A), while a weaker but significant linear relationship was observed on 10 October. NDVI is a vegetation index closely dependent on chlorophyll content and leaf cell structures. Specifically, chlorophyll has a strong absorption peak in the red region, while leaf mesophyll constituents and canopy structure are the factors that positively influence canopy NIR reflectance (Gitelson, 2004; Caruso et al., 2023). Indeed, increases in NIR reflectance correlate with increases in leaf thickness (Castro and Sanchez-Azofeifa, 2008). Thus NDVI is mainly used for the assessment of changes in canopy biophysical properties (leaf area index, fraction of absorbed photosynthetically active radiation, chlorophyll content, etc.) (Gitelson, 2004). These characteristics are closely related to the water status of the plant, as water is the main component of metabolic processes (e.g., photosynthesis) that determine chlorophyll content and canopy structure (Hailemichael et al., 2016). Since these parameters are closely related to plant water



Representation of canopy NDVI along a tree row for full irrigation (FI) and drought stress (DS) treatments. The frequency histogram (A) concerns the first acquisition date (19 September) while the box-plot (B) shows the NDVI values on 19 September and 10 October 2023 for both treatments.

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TABLE 2 Vegetation indices for 'Calatina' olive trees on 19 September and 10 October.

	NDVI <sup>a</sup>	NDRE	GNDVI	CVI	MSAVI	WI	NDGI	GI	
19 September									
DS	0.692 ± 0.012	0.191 ± 0.005	0.613 ± 0.014	2.891 ± 0.088	0.495 ± 0.031	0.171 ± 0.011	$0.184 \pm 0.014$	1.441 ± 0.022	
FI	0.733 ± 0.014	0.201 ± 0.006	$0.639 \pm 0.014$	3.038 ± 0.121	0.521 ± 0.022	$0.149 \pm 0.009$	$0.202 \pm 0.011$	$1.488 \pm 0.029$	
	*	**	**	*	*	**	*	**	
10 October									
DS	0.733 ± 0.022	$0.210 \pm 0.007$	0.639 ± 0.013	3.051 ± 0.140	$0.505 \pm 0.040$	$0.147 \pm 0.009$	0.190 ± 0.023	1.481 ± 0.038	
FI	0.752 ± 0.019	0.233 ± 0.012	$0.660 \pm 0.020$	3.131 ± 0.121	0.553 ± 0.038	$0.129 \pm 0.022$	$0.220 \pm 0.034$	$1.564 \pm 0.073$	
	n.s.	**	*	n.s.	n.s.	*	*	*	

When present, letters denote significant differences between full irrigated (FI) and drought stressed (DS) trees (P < 0.05). n.s., no significantly different; \*, significantly different for P < 0.05; \*\*, significantly different for P < 0.01. NDVI, normalized difference vegetation index; NDRE, normalized difference RedEdge index; GNDVI, green normalized difference vegetation index; CVI, chlorophyll vegetation index; MSAVI, modified soil-adjusted vegetation index; WI, water index; NDGI, normalized difference greenness index; GI, green index.

status, NDVI serves as a reliable indirect indicator of plant water status. Similar relationships in olive were found by Marino et al. (2014) and Rallo et al. (2014).

Also MSAVI was linearly related to  $\Psi_{stem}$  on both dates, but reported the lowest R<sup>2</sup> on 19 September and one of the highest R<sup>2</sup> on 10 October (Figure 9B). To date, mixed results have been found in the literature on the effectiveness of the MSAVI and similar soiladjusted indexes for assessing plant water status. In grapevine, Romero et al. (2018) reported a stronger correlation between MSAVI and  $\Psi_{stem}$  compared to NDVI. Conversely, Conesa et al. (2019) observed a weaker relationship in nectarine. Since this is an index that tends to remove the soil effect, the effectiveness of removing this effect may depend on different soil characteristics, as the spectral signature of the soil background varies with color, moisture, texture, etc (Baumgardner et al., 1986). Ren and Feng (2015) compared MSAVI with two soil unadjusted vegetation indices (NDVI and simple ratio index, SR), finding that the performance of MSAVI for estimating above-ground vegetation is lower than NDVI and SR.

TABLE 3 Comparison of the regression slopes for the relationships between vegetation indices and stem water potential ( $\Psi_{stem}$ ) on 19 September and 10 October in 'Calatina' olive trees (t-test, P < 0.05).

	Slc	P-value	
	19 September	10 October	
NDVI	0.018	0.106	0.008
MSAVI	0.016	0.126	0.004
NDRE	0.008	0.047	0.014
NDGI	0.01	0.107	0.005
GNDVI	0.017	0.088	0.007
WI	-0.012	-0.072	0.003
CVI	0.106	0.397	0.033
GI	0.031	0.348	0.005

Relationships of  $\Psi_{\text{stem}}$  with NDRE were similar to those with NDVI (Figure 9C). Since NDRE is calculated using the RedEdge band, it turns out to be a sensitive indicator of changes in chlorophyll content, thus indirectly related to plant water status (Boiarskii and Hasegawa, 2019; Roma et al., 2023). Similar relationships have been observed in the literature in other tree species such as sweet cherry (Blanco et al., 2020) and grapevine (Tang et al., 2022).

Relationships with NDGI showed relatively low  $R^2$  values on both dates (Figure 9D). This result partly disagrees with results found in other studies, which instead emphasize its reliability (Rallo et al., 2014; Pôças et al., 2015). However, since it is a greenness index, it may be influenced by factors like canopy exposure, e.g., paraheliotropism, as suggested by Rallo et al. (2014). In addition, the NIR band, which is highly sensitive to changes in the plant water status, is not used to calculate NDGI (Raz et al., 2003).

A strong significant linear relationship was found between GNDVI and  $\Psi_{stem}$  on both 19 September and 10 October (Figure 9E), suggesting that this vegetation index may be one of the most accurate in estimating 'Calatina' tree water status. GNDVI, taking into account the Green band, is positively correlated with anthocyanin content and negatively correlated with chlorophyll content, making it very sensitive to abiotic stresses (Viña and Gitelson, 2010). Since GNDVI is very sensitive to variations in chlorophyll content, the stronger correlation observed on 10 October compared to other indices can be attributed to the pronounced impact of light changes on chlorophyll content. Indeed, photosynthetically active radiation (PAR) on 10 October was  $1633.55 \pm 195.11 \ \mu mol \ m^{-2} \ s^{-1}$  (in contrast to  $1121.15 \pm 205.50$ µmol m<sup>-2</sup> s<sup>-1</sup> recorded on 19 September). To date, there are not many studies on the assessment of olive tree water status using GNDVI. Rallo et al. (2014) found a significant relationship between GNDVI and  $\Psi_{stem}$  in 'Nocellara del Belice' olive, however, the coefficient of determination  $(R^2 = 0.41)$  was lower than in this study. Contrarily, numerous studies on grapevines have consistently demonstrated that GNDVI is one of the most reliable indirect predictors of  $\Psi_{\text{stem}}$  (Helman et al., 2018; Cogato et al., 2022; Caruso and Palai, 2023).



#### FIGURE 9

Linear relationships between stem water potential ( $\Psi_{stem}$ ) and normalized difference vegetation index [NDVI, (**A**)], modified soil-adjusted vegetation index [MSAVI, (**B**)], normalized difference RedEdge index [NDRE, (**C**)], normalized difference greenness index [NDGI, (**D**)], green normalized difference vegetation index [GNDVI, (**E**)], green index [GI, (**F**)], chlorophyll vegetation index [CVI, (**G**)], water index [WI, (**H**)] on 19 September (black dots) and 10 October (white dots) 2023.

Finally, CVI and GI were indices that correlated weakly with  $\Psi_{\text{stem}}$ , both on 19 September and 10 October (Figures 9G, H). Similarly to NDGI, they are closely related to greenness, so they can be easily affected by canopy exposure (Vincini et al., 2008; Rallo et al., 2014). Although CVI is a good indicator of chlorophyll status, it has been little used for plant water status assessment. Andrade Junior et al. (2022) found a significant linear relationship between CVI and  $\Psi_{stem}$  in soybean, but weaker than NDVI and optimized soil adjusted VI (OSAVI). GI, on the other hand, has been used in several experiments. In olive, Rallo et al. observed a relationship with similar coefficient of determination ( $R^2 = 0.45$ ). In 'Cabernet Sauvignon' grapevine, the relationship between GI and  $\Psi_{stem}$  was

weak (Romero et al., 2018), while in Grenache and Shiraz grapevines, it proved to be a good indicator of drought stress (Cogato et al., 2022).

WI was one of the indices that showed a strong, but negative relationship with  $\Psi_{stem}$  on both dates (Figure 9F). This occurred since it is given by the ratio of the Red to NIR reflectance. Thus, as water stress increases, Red reflectance increased and NIR reflectance decreased. Since this is an index closely dependent on NIR reflectance, it is often correlated with leaf relative water content (RWC) (Peñuelas et al., 1994). The reflectance in NIR is associated with plant cell structures, particularly the cell wall. Consequently, alterations in plant water status involving changes in cell turgor affect cell wall structure and ultimately NIR reflectance (Peñuelas et al., 1993). However, it is important to note that under prolonged water stress conditions, plants may adjust osmotically, which can affect the correlation between WI and  $\Psi_{stem}$ . Similar results in olive were found by Asgari et al. (2023) and Marino et al. (2014). On the other hand, Rallo et al. (2014) observed a weaker relationship between WI and  $\Psi_{stem}$ . WI was also found to be a good predictor of  $\Psi_{\text{stem}}$  in grapevine (Serrano et al., 2010) and 'Satsuma' mandarin (Dzikiti et al., 2010).

In summary, from highest to lowest R<sup>2</sup>, NDVI, GNDVI, WI, and NDRE were the indices that correlated best with  $\Psi_{stem}$ , especially when the range of hydration considered was wide. In contrast, CVI, NDGI, GI and MSAVI were worse predictors of  $\Psi_{stem}$  than the aforementioned (for a more detailed report on the regressions, see Supplementary Table 1). Considering the higher reliability of  $\Psi_{stem}$  as the primary parameter for assessing plant water status, correlations between all the vegetation indices and  $g_s$ were not reported.

# 3.5 Combination of proximal and remote sensing

Combination of proximal and remote sensing data was explored by correlation analysis only on September 19 (the date when a wide range of  $\Psi_{stem}$  was covered) (Table 4).

Regarding thermal indices, the tightest correlations were observed between CWSI and NDVI and between CWSI and WI, indicating that NDVI and WI are the best vegetation indices for combining proximal CWSI and multispectral remote sensing indices. A strong inverse correlation was also found between CWSI and NDRE and between CWSI and GNDVI, suggesting that these indices may be a valid alternative to NDVI and WI for assessing 'Calatina' olive water status. This confirms the results reported in section 3.4.

Significant correlations were also observed between Ig and NDVI, GNDVI, WI, NDRE, and CVI (Table 4). In contrast, correlations between Ig and MSAVI, NDGI, and GI were not significant. Overall, these findings indicate that CWSI is a reliable indicator of 'Calatina' olive water status when combined with various multispectral vegetation indices.

As for  $\Delta W$ , the only significant close correlations were found with WI and NDGI (Table 4), showing that WI is a recurrent and solid index for estimating the water status of 'Calatina' olive.

In summary, results suggested two possible combinations. The first involved the use of CWSI and NDVI or WI. This represents a more simplified solution, as it uses only two parameters, but is probably less accurate since these indices are related only to canopies. The other possibility involved CWSI and  $\Delta W$  as proximal indices, and WI as a multispectral index. Indeed, WI correlates best with both CWSI and  $\Delta W$ .

Although this approach uses multiple parameters, it would provide more comprehensive information about tree water status. Specifically, this combination integrates spatial and temporal scale information related to both canopy and fruit. Since these results were obtained under limited conditions (one cultivar, on a specific date, and in a specific geographic area), further studies are needed to validate this combined system.

# 4 Conclusions

The following trial aimed to combine proximal and remote sensing techniques to show the benefits of using both monitoring

		NDVI	NDRE	GNDVI	WI	CVI	MSAVI	NDGI	GI
CWSI	r	-0.827	-0.730	-0.761	0.813	-0.559	-0.678	-0.698	-0.694
	P-value	0.000	0.005	0.003	0.001	0.047	0.011	0.008	0.009
Ig	r	0.743	0.624	0.682	-0.680	0.596	0.533	0.505	0.502
	P-value	0.006	0.030	0.015	0.015	0.041	n.s.	n.s.	n.s.
ΔW	r	0.593	0.614	0.670	-0.716	0.508	0.457	0.692	0.690
	P-value	n.s.	n.s.	n.s.	0.041	n.s.	n.s.	0.050	n.s.

TABLE 4 Pearson's correlation analysis between proximal and remote sensing indices.

CWSI, crop water stress index; Ig, stomatal conductance index; ΔW, fruit daily weight fluctuation; NDVI, normalized difference vegetation index; NDRE, normalized difference RedEdge index; GNDVI, green normalized difference vegetation index; CVI, chlorophyll vegetation index; MSAVI, modified soil-adjusted vegetation index; WI, water index; NDGI, normalized difference greenness index; GI. green index.

Correlations are significant for P < 0.05; n.s., non-significant.

methods, while simultaneously assessing the water status and response of 'Calatina' olive under two distinct irrigation regimes.

Indices obtained from thermal imaging, continuous fruit monitoring and remote sensing showed significant relationships in most cases. In detail, CWSI calculated using the direct method proved to be a reliable indicator of both  $\psi_{stem}$  and  $g_s$ . While Ig correlated better with g<sub>s</sub>, thus proving to be a good indicator of stomatal conductance. As for the  $\Delta W$  and RGR of the fruit, calculated by continuous monitoring of fruit gauges, both parameters increase exponentially as  $\psi_{stem}$  increases. In particular,  $\Delta W$  was found to be the index that correlates most strongly with  $\psi_{stem}$  during the trial period. These data can provide valuable insights into the temporal dynamics of fruit growth, helping to identify the onset of water stress in the drupe. This could represent a probable time when to irrigate. Regarding vegetation indices obtained from multispectral data, NDVI, GNDVI, WI and NDRE were found to be the vegetation indices that correlate best with  $\psi_{stem}$ , achieving high levels of accuracy. Useful information on the spatial variability of the olive orchard related to plant water status can be obtained with these vegetation indices. This information can subsequently serve both for strategic placement of proximal sensors and for managing irrigation differently within the orchard considering the spatial variability. Finally, the combination of proximal and remote sensing data identified either the combined use of CWSI and NDVI or WI as a more simplified possibility, or the combination of CWSI,  $\Delta W$ , and WI as a more comprehensive and informative system. Further studies on combining proximal and remote sensing data will be necessary to validate the system and establish intervention thresholds. This should also include monitoring other plant organs (leaf, trunk) and providing quantitative indication on irrigation volume. Incorporating sensors to measure sap flow or leaf transpiration could offer valuable information about plant water usage.

# Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

# Author contributions

AC: Conceptualization, Data curation, Formal analysis, Investigation, Validation, Visualization, Writing – original draft.

# References

Ahumada-Orellana, L., Ortega-Farías, S., Poblete-Echeverría, C., and Searles, P. S. (2019). Estimation of stomatal conductance and stem water potential threshold values

RM: Conceptualization, Data curation, Investigation, Resources, Validation, Visualization, Writing – original draft. FM: Conceptualization, Resources, Supervision, Writing – review & editing. PC: Conceptualization, Resources, Supervision, Writing – review & editing. ER: Conceptualization, Data curation, Formal analysis, Investigation, Validation, Visualization, Writing – original draft. RL: Conceptualization, Formal analysis, Resources, Supervision, Writing – review & editing.

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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# Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2024.1448656/ full#supplementary-material

Abboud, S., Vives-Peris, V., Dbara, S., Gómez-Cadenas, A., Pérez-Clemente, R. M., Abidi, W., et al. (2021). Water status, biochemical and hormonal changes involved in the response of *Olea europaea* L. @ to water deficit induced by partial root-zone drying irrigation (PRD). *Sci. Hortic.* 276, 109737. doi: 10.1016/j.scienta.2020.109737

for water stress in olive trees (cv. Arbequina). Irrig. Sci. 37, 461-467. doi: 10.1007/s00271-019-00623-9

Alexopoulos, A., Koutras, K., Ali, S. B., Puccio, S., Carella, A., Ottaviano, R., et al. (2023). Complementary use of ground-based proximal sensing and airborne/ spaceborne remote sensing techniques in precision agriculture: A systematic review. *Agronomy* 13, 1942. doi: 10.3390/agronomy13071942

Andrade Junior, A. S., de, da Silva, S. P., Setúbal, I. S., de Souza, H. A., and Vieira, P. F. (2022). Remote detection of water and nutritional status of soybeans using UAV-based images. *Eng. Agric.* 42, e20210177. doi: 10.1590/1809-4430-eng.agric.v42n2e20210177/ 2022

Asgari, A., Hooshmand, A., Broumand-Nasab, S., and Zivdar, S. (2023). Potential application of spectral indices for olive water status assessment in (semi-) arid regions: A case study in Khuzestan Province, Iran. *Plant Direct* 7, e494. doi: 10.1002/pld3.494

Baumgardner, M. F., Silva, L. F., Biehl, L. L., and Stoner, E. R. (1986). Reflectance properties of soils. Adv. Agron. 38, 1-44. doi: 10.1016/S0065-2113(08)60672-0

Bazzi, C. L., Schenatto, K., Upadhyaya, S., Rojo, F., Kizer, E., and Ko-Madden, C. (2019). Optimal placement of proximal sensors for precision irrigation in tree crops. *Precis. Agric.* 20, 663–674. doi: 10.1007/s11119-018-9604-3

Belfiore, N., Vinti, R., Lovat, L., Chitarra, W., Tomasi, D., de Bei, R., et al. (2019). Infrared thermography to estimate vine water status: Optimizing canopy measurements and thermal indices for the varieties Merlot and Moscato in northern Italy. *Agronomy* 9, 821. doi: 10.3390/agronomy9120821

Ben-Gal, A., Agam, N., Alchanatis, V., Cohen, Y., Yermiyahu, U., Zipori, I., et al. (2009). Evaluating water stress in irrigated olives: correlation of soil water status, tree water status, and thermal imagery. *Irrig. Sci.* 27, 367–376. doi: 10.1007/s00271-009-0150-7

Berni, J., Zarco-Tejada, P., Sepulcre-Cantó, G., Fereres, E., and Villalobos, F. (2009). Mapping canopy conductance and CWSI in olive orchards using high resolution thermal remote sensing imagery. *Remote Sens. Environ.* 113, 2380–2388. doi: 10.1016/ j.rse.2009.06.018

Blanco, V., Blaya-Ros, P. J., Castillo, C., Soto-Vallés, F., Torres-Sánchez, R., and Domingo, R. (2020). Potential of UAS-based remote sensing for estimating tree water status and yield in sweet cherry trees. *Remote Sens.* 12, 2359. doi: 10.3390/rs12152359

Blanco, V., Willsea, N., Campbell, T., Howe, O., and Kalcsits, L. (2023). Combining thermal imaging and soil water content sensors to assess tree water status in pear trees. *Front. Plant Sci.* 14, 1197437. doi: 10.3389/fpls.2023.1197437

Blaya-Ros, P. J., Blanco, V., Domingo, R., Soto-Valles, F., and Torres-Sánchez, R. (2020). Feasibility of low-cost thermal imaging for monitoring water stress in young and mature sweet cherry trees. *Appl. Sci.* 10, 5461. doi: 10.3390/app10165461

Boiarskii, B., and Hasegawa, H. (2019). Comparison of NDVI and NDRE indices to detect differences in vegetation and chlorophyll content. *J. Mech. Contin Math Sci.* 4, 20–29. doi: 10.26782/jmcms

Boini, A., Manfrini, L., Bortolotti, G., Corelli-Grappadelli, L., and Morandi, B. (2019). Monitoring fruit daily growth indicates the onset of mild drought stress in apple. *Sci. Hortic.* 256, 108520. doi: 10.1016/j.scienta.2019.05.047

Brito, C., Dinis, L.-T., Ferreira, H., Moutinho-Pereira, J., and Correia, C. M. (2020). Foliar pre-treatment with abscisic acid enhances olive tree drought adaptability. *Plants* 9, 341. doi: 10.3390/plants9030341

Carella, A., Bulacio Fischer, P. T., Massenti, R., and Lo Bianco, R. (2024). Continuous plant-based and remote sensing for determination of fruit tree water status. *Horticulturae* 10, 516. doi: 10.3390/horticulturae10050516

Carella, A., Gianguzzi, G., Scalisi, A., Farina, V., Inglese, P., and Bianco, R. L. (2021). Fruit growth stage transitions in two mango cultivars grown in a Mediterranean environment. *Plants* 10, 1332. doi: 10.3390/plants10071332

Carella, A., Massenti, R., and Lo Bianco, R. (2023). Testing effects of vapor pressure deficit on fruit growth: a comparative approach using peach, mango, olive, orange, and loquat. *Front. Plant Sci.* 14. doi: 10.3389/fpls.2023.1294195

Carella, A., Massenti, R., Milazzo, G., Caruso, T., and Lo Bianco, R. (2022). Fruiting, morphology, and architecture of 'Arbequina' and 'Calatina' Olive branches. *Horticulturae* 8, 109. doi: 10.3390/horticulturae8020109

Caruso, G., and Palai, G. (2023). Assessing grapevine water status using Sentinel-2 images. *Italus Hortus* 30, 70–79. doi: 10.26353/j.itahort/2023.3.7079

Caruso, G., Palai, G., Gucci, R., and Priori, S. (2022a). Remote and proximal sensing techniques for site-specific irrigation management in the olive orchard. *Appl. Sci.* 12, 1309. doi: 10.3390/app12031309

Caruso, G., Palai, G., Marra, F. P., and Caruso, T. (2021). High-resolution UAV imagery for field olive (*Olea europaea* L.) phenotyping. *Horticulturae* 7, 258. doi: 10.3390/horticulturae7080258

Caruso, G., Palai, G., Tozzini, L., D'Onofrio, C., and Gucci, R. (2023). The role of LAI and leaf chlorophyll on NDVI estimated by UAV in grapevine canopies. *Sci. Hortic.* 322, 112398. doi: 10.1016/j.scienta.2023.112398

Caruso, G., Palai, G., Tozzini, L., and Gucci, R. (2022b). Using visible and thermal images by an unmanned aerial vehicle to monitor the plant water status, canopy growth and yield of olive trees (cvs. Frantoio and Leccino) under different irrigation regimes. *Agronomy* 12, 1904. doi: 10.3390/agronomy12081904

Castro, K. L., and Sanchez-Azofeifa, G. A. (2008). Changes in spectral properties, chlorophyll content and internal mesophyll structure of senescing Populus balsamifera and Populus tremuloides leaves. *sensors* 8, 51–69. doi: 10.3390/s8010051

Chazarreta, Y. D., Carcedo, A. J., Prado, S. A., Massigoge, I., Amas, J. I., Fernandez, J. A., et al. (2023). Enhancing maize grain dry-down predictive models. *Agric. For. Meteorol.* 334, 109427. doi: 10.1016/j.agrformet.2023.109427

Cocozza, C., Marino, G., Giovannelli, A., Cantini, C., Centritto, M., and Tognetti, R. (2015). Simultaneous measurements of stem radius variation and sap flux density

reveal synchronisation of water storage and transpiration dynamics in olive trees. *Ecohydrology* 8, 33-45. doi: 10.1002/eco.1483

Cogato, A., Jewan, S. Y. Y., Wu, L., Marinello, F., Meggio, F., Sivilotti, P., et al. (2022). Water stress impacts on grapevines (*Vitis vinifera* L.) in hot environments: physiological and spectral responses. *Agronomy* 12, 1819. doi: 10.3390/agronomy12081819

Conesa, M. R., Conejero, W., Vera, J., Ramírez-Cuesta, J. M., and Ruiz-Sánchez, M. C. (2019). Terrestrial and remote indexes to assess moderate deficit irrigation in earlymaturing nectarine trees. *Agronomy* 9, 630. doi: 10.3390/agronomy9100630

Connor, D. J. (2005). Adaptation of olive (Olea europaea L.) to water-limited environments. Aust. J. Agric. Res. 56, 1181–1189. doi: 10.1071/AR05169

Corell, M., Pérez-López, D., Andreu, L., Recena, R., Centeno, A., Galindo, A., et al. (2022). Yield response of a mature hedgerow oil olive orchard to different levels of water stress during pit hardening. *Agric. Water Manage.* 261, 107374. doi: 10.1016/j.agwat.2021.107374

Costa, J. M., Ortuño, M. F., Lopes, C. M., and Chaves, M. M. (2012). Grapevine varieties exhibiting differences in stomatal response to water deficit. *Funct. Plant Biol.* 39, 179–189. doi: 10.1071/FP11156

Dzikiti, S., Verreynne, J., Stuckens, J., Strever, A., Verstraeten, W., Swennen, R., et al. (2010). Determining the water status of Satsuma mandarin trees [Citrus Unshiu Marcovitch] using spectral indices and by combining hyperspectral and physiological data. *Agric. For. Meteorol.* 150, 369–379. doi: 10.1016/j.agrformet.2009.12.005

Egea, G., Padilla-Díaz, C. M., Martínez, J., Fernández, J. E., and Pérez-Ruiz, M. (2016). Use of aerial thermal imaging to assess water status variability in hedgerow olive orchards. *Agric. Water Manage.* 187, 210–221.

Fernandes, R. D. M., Cuevas, M. V., Diaz-Espejo, A., and Hernandez-Santana, V. (2018). Effects of water stress on fruit growth and water relations between fruits and leaves in a hedgerow olive orchard. *Agric. Water Manage.* 210, 32–40. doi: 10.1016/j.agwat.2018.07.028

Fernandez, J.-E. (2014). Understanding olive adaptation to abiotic stresses as a tool to increase crop performance. *Environ. Exp. Bot.* 103, 158–179. doi: 10.1016/j.envexpbot.2013.12.003

Fuentes, S., De Bei, R., Pech, J., and Tyerman, S. (2012). Computational water stress indices obtained from thermal image analysis of grapevine canopies. *Irrig. Sci.* 30, 523–536. doi: 10.1007/s00271-012-0375-8

García-Tejero, I., Hernández, A., Padilla-Díaz, C., Diaz-Espejo, A., and Fernández, J. (2017). Assessing plant water status in a hedgerow olive orchard from thermography at plant level. *Agric. Water Manage.* 188, 50–60. doi: 10.1016/j.agwat.2017.04.004

García-Tejero, I. F., Rubio, A. E., Viñuela, I., Hernández, A., Gutiérrez-Gordillo, S., Rodríguez-Pleguezuelo, C. R., et al. (2018). Thermal imaging at plant level to assess the crop-water status in almond trees (*cv.* Guara) under deficit irrigation strategies. *Agric. Water Manage.* 208, 176–186. doi: 10.1016/j.agwat.2018.06.002

Génard, M., Dauzat, J., Franck, N., Lescourret, F., Moitrier, N., Vaast, P., et al. (2008). Carbon allocation in fruit trees: from theory to modelling. *Trees* 22, 269–282. doi: 10.1007/s00468-007-0176-5

Giovannini, A., Venturi, M., Gutiérrez-Gordillo, S., Manfrini, L., Corelli-Grappadelli, L., and Morandi, B. (2022). Vascular and transpiration flows affecting apricot (Prunus Armeniaca L.) fruit growth. *Agronomy* 12, 989. doi: 10.3390/agronomy12050989

Giovannoni, J. (2001). Molecular biology of fruit maturation and ripening. Annu. Rev. Plant Biol. 52, 725–749. doi: 10.1146/annurev.arplant.52.1.725

Girón, I., Corell, M., Galindo, A., Torrecillas, E., Morales, D., Dell'Amico, J., et al. (2015). Changes in the physiological response between leaves and fruits during a moderate water stress in table olive trees. *Agric. Water Manage.* 148, 280–286. doi: 10.1016/j.agwat.2014.10.024

Gitelson, A. A. (2004). Wide dynamic range vegetation index for remote quantification of biophysical characteristics of vegetation. *J. Plant Physiol.* 161, 165–173. doi: 10.1078/0176-1617-01176

Gitelson, A. A., Kaufman, Y. J., and Merzlyak, M. N. (1996). Use of a green channel in remote sensing of global vegetation from EOS-MODIS. *Remote Sens. Environ.* 58, 289–298. doi: 10.1016/S0034-4257(96)00072-7

Goldhamer, D. A. (1997). Regulated deficit irrigation for California canning olives. *III Int Symposium on Olive Growing*. 474, 369–372. doi: 10.17660/ActaHortic. 1999.474.76

Gómez-Bellot, M. J., Parra, A., Nortes, P., Alarcón, J. J., and Ortuño, M. F. (2024). Searching for a deficit irrigation strategy to save water and improve fruit quality without compromising pomegranate production. *Sci. Hortic.* 324, 112631. doi: 10.1016/ j.scienta.2023.112631

Hailemichael, G., Catalina, A., González, M., and Martin, P. (2016). Relationships between water status, leaf chlorophyll content and photosynthetic performance in Tempranillo vineyards. *South Afr. J. Enol. Vitic.* 37, 149–156. doi: 10.21548/37-2-1004

Helman, D., Bahat, I., Netzer, Y., Ben-Gal, A., Alchanatis, V., Peeters, A., et al. (2018). Using time series of high-resolution planet satellite images to monitor grapevine stem water potential in commercial vineyards. *Remote Sens.* 10, 1615. doi: 10.3390/ rs10101615

Herrmann, I., Karnieli, A., Bonfil, D., Cohen, Y., and Alchanatis, V. (2010). SWIRbased spectral indices for assessing nitrogen content in potato fields. *Int. J. Remote Sens.* 31, 5127–5143. doi: 10.1080/01431160903283892 Idso, S. B. (1982). Non-water-stressed baselines: a key to measuring and interpreting plant water stress. *Agric. Meteorol.* 27, 59–70. doi: 10.1016/0002-1571(82)90020-6

Jackson, R. D., Idso, S., Reginato, R., and Pinter, J. P. (1981). Canopy temperature as a crop water stress indicator. *Water Resour. Res.* 17, 1133–1138. doi: 10.1029/WR017i004p01133

Jin, J., Huang, N., Huang, Y., Yan, Y., Zhao, X., and Wu, M. (2022). Proximal remote sensing-based vegetation indices for monitoring mango tree stem sap flux density. *Remote Sens.* 14, 1483. doi: 10.3390/rs14061483

Jones, H. G., Stoll, M., Santos, T., de Sousa, C., Chaves, M. M., and Grant, O. M. (2002). Use of infrared thermography for monitoring stomatal closure in the field: application to grapevine. *J. Exp. Bot.* 53, 2249–2260. doi: 10.1093/jxb/erf083

Jorge, J., Vallbé, M., and Soler, J. A. (2019). Detection of irrigation inhomogeneities in an olive grove using the NDRE vegetation index obtained from UAV images. *Eur. J. Remote Sens.* 52, 169–177. doi: 10.1080/22797254.2019.1572459

Jurado, J. M., Ortega, L., Cubillas, J. J., and Feito, F. (2020). Multispectral mapping on 3D models and multi-temporal monitoring for individual characterization of olive trees. *Remote Sens.* 12, 1106. doi: 10.3390/rs12071106

Khosravi, A., Zucchini, M., Mancini, A., and Neri, D. (2022). Continuous third phase fruit monitoring in olive with regulated deficit irrigation to set a quantitative index of water stress. *Horticulturae* 8, 1221. doi: 10.3390/horticulturae8121221

Konapala, G., Mishra, A. K., Wada, Y., and Mann, M. E. (2020). Climate change will affect global water availability through compounding changes in seasonal precipitation and evaporation. *Nat. Commun.* 11, 3044. doi: 10.1038/s41467-020-16757-w

Lawson, T., and Blatt, M. R. (2014). Stomatal size, speed, and responsiveness impact on photosynthesis and water use efficiency. *Plant Physiol*. 164, 1556–1570. doi: 10.1104/ pp.114.237107

Lescourret, F., Génard, M., Habib, R., and Fishman, S. (2001). Variation in surface conductance to water vapor diffusion in peach fruit and its effects on fruit growth assessed by a simulation model. *Tree Physiol.* 21, 735–741. doi: 10.1093/treephys/ 21.11.735

Lo Bianco, R., Proietti, P., Regni, L., and Caruso, T. (2021). Planting systems for modern olive growing: strengths and weaknesses. *Agriculture* 11, 494. doi: 10.3390/agriculture11060494

Maccioni, A., Agati, G., and Mazzinghi, P. (2001). New vegetation indices for remote measurement of chlorophylls based on leaf directional reflectance spectra. *J. Photochem. Photobiol. B* 61, 52–61. doi: 10.1016/S1011-1344(01)00145-2

Mairech, H., López-Bernal, Á., Moriondo, M., Dibari, C., Regni, L., Proietti, P., et al. (2021). Sustainability of olive growing in the Mediterranean area under future climate scenarios: Exploring the effects of intensification and deficit irrigation. *Eur. J. Agron.* 129, 126319. doi: 10.1016/j.eja.2021.126319

Marino, G., Caruso, T., Ferguson, L., and Marra, F. P. (2018). Gas exchanges and stem water potential define stress thresholds for efficient irrigation management in olive (Olea europea L.). *Water* 10, 342. doi: 10.3390/w10030342

Marino, G., Pallozzi, E., Cocozza, C., Tognetti, R., Giovannelli, A., Cantini, C., et al. (2014). Assessing gas exchange, sap flow and water relations using tree canopy spectral reflectance indices in irrigated and rainfed *Olea europaea L. Environ. Exp. Bot.* 99, 43–52. doi: 10.1016/j.envexpbot.2013.10.008

Marino, G., Scalisi, A., Guzmán-Delgado, P., Caruso, T., Marra, F. P., and Lo Bianco, R. (2021). Detecting mild water stress in olive with multiple plant-based continuous sensors. *Plants* 10, 131. doi: 10.3390/plants10010131

Marques, P., Pádua, L., Sousa, J. J., and Fernandes-Silva, A. (2023). Assessing the water status and leaf pigment content of olive trees: evaluating the potential and feasibility of unmanned aerial vehicle multispectral and thermal data for estimation purposes. *Remote Sens.* 15, 4777. doi: 10.3390/rs15194777

Marra, F., Marino, G., Marchese, A., and Caruso, T. (2016). Effects of different irrigation regimes on a super-high-density olive grove cv. "Arbequina": Vegetative growth, productivity and polyphenol content of the oil. *Irrig. Sci.* 34, 313–325. doi: 10.1007/s00271-016-0505-9

Massenti, R., Ioppolo, A., Veneziani, G., Selvaggini, R., Servili, M., Lo Bianco, R., et al. (2022a). Low tree vigor, free palmette training form, and high planting density increase olive and oil yield efficiency in dry, sloping areas of Mediterranean regions. *Horticulturae* 8, 817. doi: 10.3390/horticulturae8090817

Massenti, R., Scalisi, A., Marra, F. P., Caruso, T., Marino, G., and Lo Bianco, R. (2022b). Physiological and structural responses to prolonged water deficit in young trees of two olive cultivars. *Plants* 11, 1695. doi: 10.3390/plants11131695

Matese, A., Baraldi, R., Berton, A., Cesaraccio, C., Di Gennaro, S., Duce, P., et al. (2018). Estimation of water stress in grapevines using proximal and remote sensing methods. *Remote Sens.* 10, 114. doi: 10.3390/rs10010114

Melaouhi, A., Baraza, E., Escalona, J. M., El-AouOuad, H., Mahjoub, I., Bchir, A., et al. (2021). Physiological and biochemical responses to water deficit and recovery of two olive cultivars (*Olea europaea* L., Arbequina and Empeltre cvs.) under Mediterranean conditions. *Theor. Exp. Plant Physiol.* 33, 369–383. doi: 10.1007/s40626-021-00219-9

Mohamed, A. Z., Osroosh, Y., Peters, R. T., Bates, T., Campbell, C. S., and Ferrer-Alegre, F. (2021). Monitoring water status in apple trees using a sensitive morning crop water stress index. *Irrig. Drain.* 70, 27–41. doi: 10.1002/ird.2528

Morandi, B., Manfrini, L., Losciale, P., Zibordi, M., and Corelli-Grappadelli, L. (2010). The positive effect of skin transpiration in peach fruit growth. *J. Plant Physiol.* 167, 1033–1037. doi: 10.1016/j.jplph.2010.02.015

Morandi, B., Rieger, M., and Grappadelli, L. C. (2007). Vascular flows and transpiration affect peach (Prunus persica Batsch.) fruit daily growth. *J. Exp. Bot.* 58, 3941–3947. doi: 10.1093/jxb/erm248

Moriana, A., Orgaz, F., Pastor, M., and Fereres, E. (2003). Yield responses of a mature olive orchard to water deficits. *J. Am. Soc. Hortic. Sci.* 128, 425–431. doi: 10.21273/JASHS.128.3.0425

Moriana, A., Pérez-López, D., Prieto, M., Ramírez-Santa-Pau, M., and Pérez-Rodriguez, J. (2012). Midday stem water potential as a useful tool for estimating irrigation requirements in olive trees. *Agric. Water Manage.* 112, 43–54. doi: 10.1016/j.agwat.2012.06.003

Nikolaou, G., Neocleous, D., Christou, A., Kitta, E., and Katsoulas, N. (2020). Implementing sustainable irrigation in water-scarce regions under the impact of climate change. *Agronomy* 10, 1120. doi: 10.3390/agronomy10081120

Noori, O., and Panda, S. S. (2016). Site-specific management of common olive: Remote sensing, geospatial, and advanced image processing applications. *Comput. Electron. Agric.* 127, 680–689. doi: 10.1016/j.compag.2016.07.031

Peñuelas, J., Filella, I., Biel, C., Serrano, L., and Save, R. (1993). The reflectance at the 950–970 nm region as an indicator of plant water status. *Int. J. Remote Sens.* 14, 1887–1905. doi: 10.1080/01431169308954010

Peñuelas, J., Gamon, J., Fredeen, A., Merino, J., and Field, C. (1994). Reflectance indices associated with physiological changes in nitrogen-and water-limited sunflower leaves. *Remote Sens. Environ.* 48, 135–146. doi: 10.1016/0034-4257(94)90136-8

Pierantozzi, P., Torres, M., Tivani, M., Contreras, C., Gentili, L., Parera, C., et al. (2020). Spring deficit irrigation in olive (cv. Genovesa) growing under arid continental climate: Effects on vegetative growth and productive parameters. *Agric. Water Manage.* 238, 106212. doi: 10.1016/j.agwat.2020.106212

Pôças, I., Gonçalves, J., Costa, P. M., Gonçalves, I., Pereira, L. S., and Cunha, M. (2017). Hyperspectral-based predictive modelling of grapevine water status in the Portuguese Douro wine region. *Int. J. Appl. Earth Obs. Geoinformation* 58, 177–190. doi: 10.1016/j.jag.2017.02.013

Pôças, I., Rodrigues, A., Gonçalves, S., Costa, P. M., Gonçalves, I., Pereira, L. S., et al. (2015). Predicting grapevine water status based on hyperspectral reflectance vegetation indices. *Remote Sens.* 7, 16460–16479. doi: 10.3390/rs71215835

Pokhrel, Y., Felfelani, F., Satoh, Y., Boulange, J., Burek, P., Gädeke, A., et al. (2021). Global terrestrial water storage and drought severity under climate change. *Nat. Clim. Change* 11, 226–233. doi: 10.1038/s41558-020-00972-w

Pou, A., Diago, M. P., Medrano, H., Baluja, J., and Tardaguila, J. (2014). Validation of thermal indices for water status identification in grapevine. *Agric. Water Manage.* 134, 60–72. doi: 10.1016/j.agwat.2013.11.010

Qi, J., Chehbouni, A., Huete, A. R., Kerr, Y. H., and Sorooshian, S. (1994). A modified soil adjusted vegetation index. *Remote Sens. Environ.* 48, 119–126. doi: 10.1016/0034-4257(94)90134-1

Rallo, G., Minacapilli, M., Ciraolo, G., and Provenzano, G. (2014). Detecting crop water status in mature olive groves using vegetation spectral measurements. *Biosyst. Eng.* 128, 52–68. doi: 10.1016/j.biosystemseng.2014.08.012

Rallo, P., and Rapoport, H. (2001). Early growth and development of the olive fruit mesocarp. J. Hortic. Sci. Biotechnol. 76, 408–412. doi: 10.1080/14620316.2001.11511385

Ramírez-Cuesta, J. M., Ortuño, M., Gonzalez-Dugo, V., Zarco-Tejada, P. J., Parra, M., Rubio-Asensio, J. S., et al. (2022). Assessment of peach trees water status and leaf gas exchange using on-the-ground versus airborne-based thermal imagery. *Agric. Water Manage*. 267, 107628. doi: 10.1016/j.agwat.2022.107628

Raz, M., Karnieli, A., and Bonfil, D. (2003). Characterization of wheat grow conditions by visible and NIR reflectance. In: *Remote Sensing for Agriculture, Ecosystems, and Hydrology IV* (SPIE). Available online at: https://www.spiedigitallibrary.org/conference-proceedings-of-spie/4879/0000/Characterization-of-wheat-grow-conditions-by-visible-and-NIR-reflectance/10.1117/12.462481.short (Accessed July 4, 2024).

Reinert, S., Bögelein, R., and Thomas, F. M. (2012). Use of thermal imaging to determine leaf conductance along a canopy gradient in European beech (Fagus sylvatica). *Tree Physiol.* 32, 294–302. doi: 10.1093/treephys/tps017

Ren, H., and Feng, G. (2015). Are soil-adjusted vegetation indices better than soilunadjusted vegetation indices for above-ground green biomass estimation in arid and semi-arid grasslands? *Grass Forage Sci.* 70, 611–619. doi: 10.1111/gfs.12152

Roma, E., and Catania, P. (2022). Precision oliviculture: research topics, challenges, and opportunities—A review. *Remote Sens.* 14, 1668. doi: 10.3390/rs14071668

Roma, E., Catania, P., Vallone, M., and Orlando, S. (2023). Unmanned aerial vehicle and proximal sensing of vegetation indices in olive tree (*Olea europaea*). *J. Agric. Eng.* 54. doi: 10.4081/jae.2023.1536

Romero, M., Luo, Y., Su, B., and Fuentes, S. (2018). Vineyard water status estimation using multispectral imagery from an UAV platform and machine learning algorithms for irrigation scheduling management. *Comput. Electron. Agric.* 147, 109–117. doi: 10.1016/j.compag.2018.02.013

Rouse, J. W., Haas, R. H., Schell, J. A., Deering, D. W., and Harlan, J. C. (1974). Monitoring the vernal advancement and retrogradation (green wave effect) of natural vegetation. *NASAGSFC Type III Final Rep. Greenbelt Md* 371. Scalisi, A., Marino, G., Marra, F. P., Caruso, T., and Lo Bianco, R. (2020). A cultivarsensitive approach for the continuous monitoring of olive (*Olea europaea* L.) tree water status by fruit and leaf sensing. *Front. Plant Sci.* 11, 340. doi: 10.3389/fpls.2020.00340

Serrano, L., González-Flor, C., and Gorchs, G. (2010). Assessing vineyard water status using the reflectance based water index. *Agric. Ecosyst. Environ.* 139, 490–499. doi: 10.1016/j.agee.2010.09.007

Stateras, D., and Kalivas, D. (2020). Assessment of olive tree canopy characteristics and yield forecast model using high resolution UAV imagery. *Agriculture* 10, 385. doi: 10.3390/agriculture10090385

Tang, Z., Jin, Y., Alsina, M. M., McElrone, A. J., Bambach, N., and Kustas, W. P. (2022). Vine water status mapping with multispectral UAV imagery and machine learning. *Irrig. Sci.* 40, 715–730. doi: 10.1007/s00271-022-00788-w

Tijero, V., Girardi, F., and Botton, A. (2021). Fruit development and primary metabolism in apple. *Agronomy* 11, 1160. doi: 10.3390/agronomy11061160

Tombesi, A. (1993). Olive fruit growth and metabolism. II Int. Symposium Olive Growing 356, 225-232. doi: 10.17660/ActaHortic.1994.356.49

Tong, X., Wu, P., Liu, X., Zhang, L., Zhou, W., and Wang, Z. (2022). A global metaanalysis of fruit tree yield and water use efficiency under deficit irrigation. *Agric. Water Manage.* 260, 107321. doi: 10.1016/j.agwat.2021.107321 Trentacoste, E. R., Puertas, C. M., and Sadras, V. O. (2010). Effect of fruit load on oil yield components and dynamics of fruit growth and oil accumulation in olive (*Olea europaea L.*). *Eur. J. Agron.* 32, 249–254. doi: 10.1016/j.eja.2010.01.002

Viña, A., and Gitelson, A. A. (2010). Sensitivity to foliar anthocyanin content of vegetation indices using green reflectance. *IEEE Geosci. Remote Sens. Lett.* 8, 464–468. doi: 10.1109/LGRS.2010.2086430

Vincini, M., Frazzi, E., and D'Alessio, P. (2008). A broad-band leaf chlorophyll vegetation index at the canopy scale. *Precis. Agric.* 9, 303–319. doi: 10.1007/s11119-008-9075-z

Wahab, A., Abdi, G., Saleem, M. H., Ali, B., Ullah, S., Shah, W., et al. (2022). Plants' Physio-biochemical and phyto-hormonal responses to alleviate the adverse effects of drought stress: A comprehensive review. *Plants* 11, 1620. doi: 10.3390/plants11131620

Yu, M.-H., Ding, G.-D., Gao, G.-L., Zhao, Y.-Y., Yan, L., and Sai, K. (2015). Using plant temperature to evaluate the response of stomatal conductance to soil moisture deficit. *Forests* 6, 3748–3762. doi: 10.3390/f6103748

Zarco-Tejada, P. J., Berjón, A., Lopez-Lozano, R., Miller, J. R., Martín, P., Cachorro, V., et al. (2005). Assessing vineyard condition with hyperspectral indices: Leaf and canopy reflectance simulation in a row-structured discontinuous canopy. *Remote Sens. Environ.* 99, 271–287. doi: 10.1016/j.rse.2005.09.002