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Plants' [molecular behavior](https://www.frontiersin.org/articles/10.3389/fpls.2024.1423625/full) [to heavy metals: from](https://www.frontiersin.org/articles/10.3389/fpls.2024.1423625/full) [criticality to toxicity](https://www.frontiersin.org/articles/10.3389/fpls.2024.1423625/full)

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The contamination of soil and water with high levels of heavy metals (HMs) has emerged as a significant obstacle to agricultural productivity and overall crop quality. Certain HMs, although serving as essential micronutrients, are required in smaller quantities for plant growth. However, when present in higher concentrations, they become very toxic. Several studies have shown that to balance out the harmful effects of HMs, complex systems are needed at the molecular, physiological, biochemical, cellular, tissue, and whole plant levels. This could lead to more crops being grown. Our review focused on HMs' resources, occurrences, and agricultural implications. This review will also look at how plants react to HMs and how they affect seed performance as well as the benefits that HMs provide for plants. Furthermore, the review examines HMs' transport genes in plants and their molecular, biochemical, and metabolic responses to HMs. We have also examined the obstacles and potential for HMs in plants and their management strategies.

KEYWORDS

agricultural productivity, cross-tolerance, genotoxicity, hormesis, molecular responses, transport genes

1 Introduction

Plants, similar to other living species, are vulnerable to high levels of heavy metals (HMs) in the atmosphere, resulting from both human activities and environmental factors ([El-Sappah and Rather, 2022;](#page-15-0) [Li et al., 2023\)](#page-17-0). The poisoning of the environment with HMs is mostly caused by intensive mining operations, fast industrialization, and widespread agricultural activities ([Adnan et al., 2024\)](#page-14-0). The presence of high levels of HMs in soil and water is a notable illustration of human activities that have a substantial impact on the environment and constitute a considerable hazard ([El-Sappah et al., 2021a;](#page-15-0) [Hama Aziz](#page-16-0) [et al., 2023](#page-16-0)). HMs may be transferred over long distances in both gaseous and particle forms, leading to their rapid buildup in biological systems, water, and sediment [\(Mishra et al., 2017\)](#page-17-0).

A total of 53 elements have been classified as HMs based on their density, which exceeds 5 $g/cm³$ ([Ali and Khan, 2018](#page-14-0)). For the essential metabolic operations of plant cells, a total of 17 elements are required. However, only six of these elements are classified as HMs: copper (Cu), zinc (Zn), manganese (Mn), iron (Fe), molybdenum (Mo), and nickel (Ni). The macroelements are carbon (C), oxygen (O), hydrogen (H), magnesium (Mg), sulfur (S), nitrogen (N), calcium (Ca), phosphorus (P), and potassium (K), while the microelements are Cu, Zn, Mn, Mo, boron (B), and chlorine (Cl) [\(Fan et al., 2021](#page-15-0)). In plants, the macro- and microelements are essential for the regulation of numerous physiological and biochemical processes, such as chlorophyll formation, photosynthesis, nucleic acid metabolism, protein modification, intra-compartmental redox reactions, carbohydrate metabolism, and N fixation ([Dutta et al., 2018](#page-15-0); [Zayed et al., 2023\)](#page-19-0).

It is intriguing that while numerous HMs function as microelements, others, such as aluminum (Al), cadmium (Cd), chromium (Cr), lead (Pb), and mercury (Hg), have detrimental effects on plants. These consequences include impaired photosynthesis, chlorosis, decreased biomass output, disrupted water balance, and impaired nutrient absorption ([Angulo-](#page-14-0)[Bejarano et al., 2021](#page-14-0)). The unpreceded use of agrochemicals, long-term application of municipal sewage effluent, industrial waste disposal, waste incineration, and vehicle exhausts are the primary sources of HMs in agricultural soils [\(Mishra et al., 2017\)](#page-17-0).

Plants ingest and accumulate HMs in soil with high concentrations, which subsequently reach human nutrition through the food chain [\(Angon et al., 2024](#page-14-0)). The absorption of HMs by both underground and above-ground surfaces of plants can have a direct or indirect impact on plant health [\(Emamverdian](#page-15-0) [et al., 2015\)](#page-15-0). The inhibition of cytoplasmic enzymes and the injury to cell structures are the direct consequences of oxidative stress ([Jadia and Fulekar, 2009\)](#page-16-0). An often observed result of HM toxicity is the overabundance of reactive oxygen species (ROS) and methylglyoxal (MG), both of which can lead to lipid peroxidation, protein oxidation, enzyme deactivation, DNA damage, disruption of ionic balance in plant cells, and/or interaction with other essential components of plant cells ([Hossain et al., 2012](#page-16-0); [Jomova](#page-16-0) [et al., 2023\)](#page-16-0).

Some HMs indirectly impose oxidative stress through a variety of mechanisms, including the depletion of glutathione, the binding of sulfhydryl groups of proteins [\(Jaishankar et al., 2014](#page-16-0)), the inhibition of antioxidative enzymes, or the induction of ROSproducing enzymes such as NADPH oxidases [\(Bielen et al.,](#page-14-0) [2013\)](#page-14-0). Regardless of whether it is direct or indirect, plants that are exposed to high levels of HMs experience a reduction or even the complete cessation of all metabolic activities [\(Singh et al., 2015\)](#page-18-0).

Plant cells react to the toxicity caused by HMs via complex and interrelated systems that operate at many levels and include both immediate and long-lasting processes ([Dutta et al., 2018](#page-15-0)). The immediate or short-term reactions include the rapid modification of the rates at which hundreds or even thousands of genes are transcribed, followed by alterations in physiological and metabolic processes [\(Dutta et al., 2018\)](#page-15-0). On the other hand, genetic alterations and epigenetic modifications are associated with enduring reactions ([Ryu et al., 2015](#page-18-0)). The control of gene expression, which is an essential part of the plant's response to stress, usually involves making changes to the levels of stress-responsive genes in a way that is both common to all plants and specific to each individual plant ([Gallo-Franco et al., 2020](#page-15-0)).

Therefore, it is logical to expect that plants will react to HMs' toxicity, which causes both oxidative and genotoxic effects, by organizing and combining different elements of stress perception and signaling networks, with the possibility of communication at different stages, depending on the circumstances [\(Dutta et al.,](#page-15-0) [2018](#page-15-0)). The environmental, ecological, and genetic effects of HMs on plants, as well as their resources, occurrence, and agroecological ramifications, were discussed in the current review. The challenges and prospects of HMs' impacts on plants, as well as the methods for mitigating them, have also been the subject of discussion.

2 HMs in plants: resources, occurrence, and agroecological ramifications

The presence of HMs in soil may have negative effects on human and animal health as well as on soil quality, fertility, and agricultural productivity [\(Rashid et al., 2023](#page-18-0)). Given the fast-paced changes in the economy and culture, many hazardous materials found in polluted soil constitute a risk to both the general people and the environment ([Hajam et al., 2023\)](#page-16-0). Cd, Hg, Cu, Zn, Ni, Pb, Cr, and arsenic (As) are often detected as contaminants in soil settings ([Priya et al., 2023](#page-18-0)). This kind of pollution poses a significant biological risk, is widely spread, and is prevalent in the soil environment ([Adnan et al., 2024\)](#page-14-0). The concentration of hazardous materials in the soil is beyond the acceptable threshold in five million areas worldwide (Rodri[guez Eugenio et al., 2018\)](#page-18-0).

According to the Environmental Protection Agency (EPA) ([Goyer et al., 2004\)](#page-16-0), Hg, Pb, Cd, and As are the most dangerous metals/metalloids in the environment. Human activities, such as the use of fertilizers in agriculture, the manufacturing of compounds, and the extraction of minerals, are the main causes of the creation of hazardous substances in soil [\(Tang et al., 2019\)](#page-19-0). Multiple studies have shown that natural sources of HMs in the environment are often of lesser importance when compared to human activities ([Dixit et al., 2015](#page-15-0)). There are two main origins of HMs: natural and anthropogenic [\(Angon et al., 2024](#page-14-0)). The HMs are mostly derived from volcanic and sedimentary minerals, making them the most abundant natural sources ([Alengebawy et al., 2021](#page-14-0)).

The main origin of HMs in soils is the parent material from which they were first generated ([Angon et al., 2024\)](#page-14-0). Sedimentary rocks make up around 5% of the Earth's mantle, whereas igneous elements make up 95% [\(Sarwar et al., 2017](#page-18-0)). On the other hand, the phrase "anthropogenic" usually refers to sources that are created by humans. Anthropogenic activities, such as burning fossil fuels for electricity, disposing of municipal waste, applying fertilizer, using pesticides, and irrigating with effluent, increase the levels of HMs in agricultural soil settings ([Angon et al., 2024](#page-14-0)).

El-Sappah et al. [10.3389/fpls.2024.1423625](https://doi.org/10.3389/fpls.2024.1423625)

Effective soil management is a crucial aspect of sustainable agriculture, with soil biology playing a vital role in this context ([Srivastava et al., 2017\)](#page-18-0). Soil microorganisms are essential components of the ecosystem ([Jiang and Li, 2020\)](#page-16-0). Microorganisms play a crucial role in maintaining soil fertility by breaking down organic matter and cycling nutrients ([Wu et al.,](#page-19-0) [2024](#page-19-0)). However, stressors such as excessive temperature, pH, salinity, and chemical pollution might have a negative impact on them [\(Paz-Ferreiro and Fu, 2016](#page-17-0)). As the quantity of HMs grows, the capacity of microorganisms to survive declines ([Igiri](#page-16-0) [et al., 2018\)](#page-16-0).

The addition of Pb–Cu slurry, Pb–Cu dust, Pb–Zn dust, and Cd–Pb–Zn to forest soil resulted in a reduction in the number of colony-forming units (CFUs) of bacteria and fungi [\(Srivastava et al.,](#page-18-0) [2017\)](#page-18-0). Generally, the impact of low levels of HMs on soil respiration is minimal [\(Verma et al., 2010](#page-19-0)). However, when HMs' pollution or toxicity intensifies, this effect becomes less significant. The introduction of HMs may either enhance or hinder Nmineralization, which can be related to differences in the experimental approach, variances in soil parameters, and substrate concentrations [\(Dai et al., 2004](#page-15-0)). HMs' pollution generally has a negative impact on N transformation processes, which, in turn, affects N-mineralization ([Dai et al., 2004;](#page-15-0) [Hamsa](#page-16-0) [et al., 2017](#page-16-0)). The bioavailability of metals in soils is influenced by factors such as metal concentrations, soil pH, organic matter, and sediment content ([Rieuwerts et al., 1998\)](#page-18-0). HMs play a crucial role in controlling the activities of various soil enzymes, such as arylsulfatase, alkaline phosphatase, b-glucosidase, cellulase, dehydrogenase, invertase, protease, and urease [\(Aponte et al.,](#page-14-0) [2020\)](#page-14-0). [Figure 1](#page-3-0) depicts the many sources of HMs ([Angon](#page-14-0) [et al., 2024\)](#page-14-0).

3 Plant response to HM exposure

The persistence of toxic HMs in the soil ecosystem is a considerable hazard for living animals and plants [\(Kraemer, 2009;](#page-16-0) [Abd Elnabi et al., 2023\)](#page-14-0). The plant roots serve as the main points of contact for terrestrial plants to be exposed to harmful HMs [\(Podar](#page-18-0) [and Maathuis, 2022](#page-18-0)). Plants have developed new, adaptive, and precise methods to withstand the harmful effects of HM stress ([Tiwari and Lata, 2018](#page-19-0)). This mechanism involves various strategies such as immobilization, exclusion outside the plasma membrane, restriction of absorption and transport, synthesis of specific HM transporters, induction of stress proteins, and chelation and sequestration by specific ligands [\(Clemens, 2001;](#page-15-0) [DalCorso et al.,](#page-15-0) [2008;](#page-15-0) [Adrees et al., 2015](#page-14-0)).

To maintain a low concentration of metal ions in the cytoplasm, it is feasible to block the transport of hazardous metals across the plasma membrane, which is the cellular mechanism for HMs' tolerance ([Hall, 2002\)](#page-16-0). Here are two direct approaches. The objective may be achieved by either augmenting the attachment of metal ions to the cell wall or expelling the metal from the cell using active efflux pumps. Another approach to detoxification includes the process of chelation or modifying the concentration of harmful metal ions to a lower level, thereby rendering them inactive [\(Tong et al., 2004](#page-19-0)). Several factors, such as plant structure, plant life cycle, plant vigor, soil pH, root system depth, temperature, partial oxygen pressure, carbohydrate level, respiration rate, nutrient interface, and microbial presence, have a significant impact on the accumulation of metals in plants ([Chen et al., 2006\)](#page-15-0).

Plants have the ability to cause HMs to form negatively charged particles by changing the pH of the soil around their roots or by releasing negatively charged ions such as PO_4^3 ⁻. During the process of adsorption, the surface of the root has the ability to bind a substantial amount of HMs. The accumulation of these HMs [Cd, Ni, strontium (sr), and Pb] in plant root tissues happens quickly ([Hossain et al., 2012](#page-16-0)). The plants have been categorized into three categories based on their survival strategies under adverse conditions: accumulators, excluders, and indicators ([Baker, 1981\)](#page-14-0). Plants undergo hyperaccumulation of HMs, resulting in the accumulation of metals exceeding 0.1%–1% of the dry weight. The term "hyperaccumulator" was used by Baker and Brooks (1989) to refer to plants that have a leaf nickel concentration above 1,000 mg/g. Hyperaccumulator species refer to plants that have the ability to collect more than 100 mg of Cd per kilogram or more than 500 mg of Cr per kilogram in dried plant leaf tissue ([Kumar et al., 2019](#page-17-0)). A plant with a hyperaccumulator trait is capable of accumulating and enduring significant levels of metal pollution.

Some plant species have the ability to flourish in soil that is polluted with HMs and may collect substantial levels of metals in such soil [\(Lombi et al., 2002\)](#page-17-0). The primary methods involved in the hyperaccumulation of toxic metals in plants include bio-activation of HMs in the rhizosphere through root microbe interfaces, enhanced activity of metal conveyor proteins in cell membranes, detoxification of metals by restricting them to apoplasts, chelation of HMs in the cytoplasm by multiple ligands, and sequestration of metals into the vacuole by multiple ligands ([Kumar et al., 2019](#page-17-0)).

4 The performance of seeds and seedlings under HM stress

While the seed coat initially offers some defense against metal stress before germination, it will gradually rupture or become more porous throughout the germination process [\(Kranner and Colville,](#page-17-0) [2011](#page-17-0)). Current data suggests that metals have two distinct impacts on seed germination: their overall toxicity and their ability to hinder the uptake of water ([Osman and Fadhlallah, 2023](#page-17-0)). Pb significantly affects the physical and biological characteristics of seeds, hindering their ability to sprout, the roots to grow, the seedlings to develop, the plants to grow, water to be transported, chlorophyll to be produced, and protein to be synthesized ([Collin et al., 2022\)](#page-15-0).

Pb also hampers the production of ATP, causes the oxidation of lipids, and leads to DNA damage, culminating in an accumulation of ROS [\(Pourrut et al., 2011;](#page-18-0) [Ur Rahman et al., 2024](#page-19-0)). Soils contaminated with Pb hinder the growth of seedlings by causing an increase in lipid peroxidation and the activation of enzymes such as superoxide dismutase, guaiacol peroxidase, ascorbate peroxidase,

Various origins of heavy metals. Coal combustion, mining, refining, soil erosion, and volcanic eruptions are all sources of antimony. Sources of arsenic include smelting, mining, atmospheric deposition, pesticides, and geological sedimentation. Volcanic dust and coal and hydrocarbon combustion are sources of beryllium as well. Sources of cadmium include plastic, fertilizer, pesticides, refining, and welding. Sources of chromium include textiles, dyeing, electroplating, paint manufacturing, steel fabrication, and tanning. Copper is obtained through mining, refining, painting, plating, and printing. Coal combustion, electroplating, battery manufacturing, mining, paint, and pigments are all lead sources. Batteries, coal combustion, geothermal activities, mining, paint and paper industries, volcanic eruptions, and geological weathering are all sources of mercury. Sources of nickel include porcelain enameling, electroplating, non-ferrous metals, and pigments. Sources of selenium include coal combustion and mining. Sources of silver include the production of batteries, mining, photographic processing, and smelting. Production of cement, combustion of fossil fuels, metal smelting, and hydrocarbon refining are all sources of thallium. Brass manufacturing, mining, hydrocarbon refining, and plumbing are all sources of zinc. This figure was made using BioRender.

and glutathione (GSH)–ascorbate cycle enzymes ([Sethy and](#page-18-0) [Ghosh, 2013](#page-18-0)).

Cd is known for its capacity to inhibit seed germination via several methods. It has a negative impact on metabolic reactivation by decreasing the number of hydrolyzing enzymes, hindering starch mobilization, and preventing seed imbibition. It may also influence signaling via Ca, mitogen-activated protein kinases (MAPKs), and transcription factors (TFs) as well as the levels of phytohormones such abscisic acid (ABA), auxin (AUX), gibberellic acid (GA), and ethylene (ET) [\(Rahoui et al., 2010;](#page-18-0) [Vijayaragavan et al., 2011](#page-19-0)). Cd toxicity also induces the upregulation of glutathione peroxidase (Gpx) expression and decreases the activity of glutathione reductase ([Branca et al., 2020\)](#page-14-0). Additionally, Cd toxicity hinders the proper functioning of mitochondria ([Genchi et al., 2020](#page-16-0)).

On the other hand, Co triggers DNA methylation in Vicia faba seeds ([Rancelis et al., 2012\)](#page-18-0) while Cu is harmful to young sunflower

plants, causing oxidative stress by producing ROS and reducing catalase activity (Garcí[a et al., 1999](#page-16-0); [Pena et al., 2011](#page-17-0)). Under stress conditions, the germination rate is decreased and there is a stimulation of biomass mobilization, which hinders the breakdown of starch and sucrose in reserve tissue ([Sethy and](#page-18-0) [Ghosh, 2013\)](#page-18-0).

Cu poisoning induces oxidative stress by increasing the expression of antioxidant and stress-related proteins, hence altering metabolic processes [\(Liu et al., 2020](#page-17-0)). Ni is a noxious agent that impacts plant species by altering enzyme function and hindering seed germination and growth (Pandolfi[ni et al., 2006;](#page-17-0) [Sethy and Ghosh, 2013\)](#page-18-0). It impacts the process of breaking down and moving food reserves in plants, resulting in decreased plant height, root length, fresh and dry weight, chlorophyll content, enzyme carbonic anhydrase activity, malondialdehyde content, electrolyte leakage, and photosynthetic pigments ([Alam et al.,](#page-14-0) [2007\)](#page-14-0). Ni stress has a detrimental impact on Brassica nigra seeds, resulting in a substantial decrease in growth, leaf water potential, pigments, and photosynthetic machinery ([Yusuf et al., 2012\)](#page-19-0).

5 The beneficial roles of metals in plants

Plants need six HMs, namely, Cu, Zn, Mn, Mo, Fe, and possibly Ni. Cu is a metallic element that is essential for the process of photosynthesis and is present in numerous enzyme systems [\(Festa](#page-15-0) [and Thiele, 2011\)](#page-15-0). Cu also improves the flavor and color of fruits, vegetables, and flowers by increasing the sugar content in plants (Ló[pez-Vargas et al., 2018](#page-17-0)). Furthermore, Cu is essential for the respiration of plants and is involved in the production and formation of seeds ([Chen et al., 2022\)](#page-15-0). Zn, on the other hand, is a component of the enzymatic system and plant metabolism ([Hamzah Saleem et al., 2022](#page-16-0)). It is essential for the synthesis of RNA and protein as well as the production of chlorophyll and carbohydrates [\(Umair Hassan et al., 2020;](#page-19-0) [Costa et al., 2023\)](#page-15-0). Additionally, it is involved in the production of growth hormones, which are responsible for the regulation of plant growth and stem elongation [\(Saboor et al., 2021](#page-18-0)).

In addition, Zn permits plants to endure frigid temperatures ([Kudo et al., 2023\)](#page-17-0). Mn is essential for photosynthesis and respiration [\(Alejandro et al., 2020](#page-14-0)). The availability of N, P, and Ca to the plant is enhanced by Mn, which facilitates their decomposition ([Yang et al., 2021b](#page-19-0)). Additionally, Mn activates numerous enzyme systems, some of which are responsible for safeguarding plants from specific environmental stressors, such as drought, winter cold, salt damage, and ozone damage as well as specific soil-borne diseases and fungal leaf diseases [\(Alejandro et al.,](#page-14-0) [2020](#page-14-0)). Additionally, it facilitates pollen tube development and pollen germination [\(Sawidis et al., 2021\)](#page-18-0). It is also essential for the production of chlorophyll and protein [\(Mousavi et al., 2011\)](#page-17-0).

Conversely, the plant necessitates Mo to convert nitrates into ammonia, a form that it can assimilate ([Liu et al., 2022\)](#page-17-0). Mo is also indispensable for specific microorganisms, such as rhizobia, which have a symbiotic relationship with legumes and contribute to the fixation of atmospheric N in legumes [\(Bursakov et al., 2023](#page-15-0)). Mo also facilitates the conversion of inorganic forms of phosphorus into organic forms that are able to be absorbed by the plant ([Seeda et al.,](#page-18-0) [2020\)](#page-18-0). In contrast, Fe is critical to the plant's development and health. It is vital in metabolic activities such as DNA synthesis, energy transmission, photosynthesis, and respiration ([Rai et al.,](#page-18-0) [2021;](#page-18-0) [Ning et al., 2023](#page-17-0)). Finally, Ni is essential for the biological fixation of atmospheric N in legumes and the metabolism of N in plants [\(Mendes et al., 2023](#page-17-0)). It is involved in the metabolism, iron assimilation, senescence, and disease resistance of plants ([Begum](#page-14-0) [et al., 2022\)](#page-14-0). The beneficial effects and toxicity of critical HMs in various plants are reviewed in [Figure 2.](#page-5-0)

Metal ions can cause hormetic reactions in plants [\(Salinitro](#page-18-0) [et al., 2021\)](#page-18-0). Hormesis is a biphasic reaction to diverse chemicals in living organisms that likely produce an adaptive stress response ([Mattson, 2008\)](#page-17-0). It is likely an adaptive stress response induced by a disturbance of homeostasis caused by low levels of biotic or abiotic stimuli [\(Vargas-Hernandez et al., 2017\)](#page-19-0). HMs stimulate plant hormetic responses, which may impact nutrient absorption, activate particular defense processes, create ROS, activate antioxidant responses, and improve photosynthetic system efficiency, leading to increased biomass. This mechanism is thought to be an adaptive reaction to stress [\(Salinitro et al., 2021\)](#page-18-0).

On the other hand, plants that efficiently protect themselves against one kind of stress may increase their tolerance to other types of stress ([Perincherry et al., 2021\)](#page-17-0). Cross-tolerance is a phenomena that highlights plants' capacity to swiftly adjust to changing environments, exhibiting their well-developed and durable defensive regulatory networks ([Foyer et al., 2016](#page-15-0)). HMs may endanger herbivores, and HMs have been shown in studies to have an effect on fungi and insects, particularly aphids [\(Chen et al.,](#page-15-0) [2020a](#page-15-0)). Air and soil pollution, especially HMs, may alter plant– insect or plant–disease connections. This pollution may be harmful, causing a hormesis effect and influencing these species' behavior and metabolism ([Perincherry et al., 2021\)](#page-17-0).

6 HMs–Transporters in plants

Metal ions are transported by a diverse array of transporters in organelles. Recent advancements in molecular and genetics research have found many crucial gene families that play a role in metal transport [\(Drew et al., 2021\)](#page-15-0) [\(Table 1](#page-6-0)). These gene families have the potential to greatly contribute to HMs' tolerance in hyperaccumulator plants. So far, scientists have discovered many different types of metal transporter proteins in plants ([Singh et al.,](#page-18-0) [2015](#page-18-0)). These transporters are responsible for the safe storage of ions and the assistance of plants in recovering from the adverse effects of metal stress. These proteins have important roles in absorbing, transporting, sequestering, and storing metals in particular parts of the cell. Additionally, they have a noteworthy impact on the regulation of metal levels inside plant cells ([Drew et al., 2021\)](#page-15-0).

Metal transporters can be classified into six main groups: natural resistant-associated macrophage protein (NRAMP) ([Li](#page-17-0) [et al., 2024](#page-17-0)), Zn-regulated, Fe-regulated transporter-like proteins (ZIP) ([Shuting et al., 2022](#page-18-0)), cation diffusion facilitator (CDF) transporters ([Kolaj-Robin et al., 2015](#page-16-0); [El-Sappah et al., 2021b](#page-15-0); [El-](#page-15-0)[Sappah et al., 2023](#page-15-0)), yellow stripe-like (YSL) proteins [\(Islam et al.,](#page-16-0) [2020](#page-16-0)), and P1B-type HMs ATPases (HMAs) ([Batool et al., 2023\)](#page-14-0).

Cell organelles include specialized compartments dedicated to certain processes, including photosynthesis, respiration, phytohormone production, and metal detoxification [\(Jogawat et al.,](#page-16-0) [2021\)](#page-16-0). Among these transporters, the vacuole plays a crucial role in the accumulation and compartmentalization of metals for their detoxification. This is a significant strategy for lowering metal stress-related ailments in plants [\(Emamverdian et al., 2015](#page-15-0); [Sharma](#page-18-0) [et al., 2016\)](#page-18-0). Chloroplasts and mitochondria need transition metals to carry out essential operations such as photosynthesis, electron transport system, photoprotection, and other processes. Maintaining metal homeostasis is crucial for the optimal functioning and structural integrity of chloroplasts and mitochondria ([Nouet et al., 2011\)](#page-17-0).

Metal and nonmetal ions are primarily stored in vacuoles within plant cells. Additionally, the neutralization and mitigation of the detrimental effects of metal ions are contingent upon their storage in the vacuole [\(Hall, 2002\)](#page-16-0). Tonoplasts (vacuolar membrane) contain a variety of transport proteins, such as MTPs, ABC transporters (ABCCs, ABCGs), HMAs, Ca^{2+} exchangers (CAXs), and NRAMPs. These proteins either remove metals from the cytosol or deposit them in the vacuolar area ([Zhang et al., 2018a\)](#page-19-0).

Chloroplasts have a crucial role in hosting transition metals due to their involvement in the process of photosynthesis and the breakdown of water molecules ([Schmidt et al., 2020\)](#page-18-0). Chloroplasts play a crucial role in reducing the harmful effects of metal toxicity by capturing metal ions to facilitate metal detoxification. The double membrane structure of the cellular envelope allows for the protection of its integrity and the detoxification of metals in the intermembrane gap. Various transporters located on the inner and outer membranes are necessary to maintain the homeostasis of chloroplastic metals ([Nouet et al., 2011\)](#page-17-0).

Mitochondria are a crucial cellular organelle occasionally referred to as the cell's powerhouse due to their substantial involvement in chemosmosis. This is because certain metals are essential for the correct functioning of mitochondria, as they serve as cofactors for critical enzymes and are also involved in the composition of electron transport molecules [\(Jogawat et al.,](#page-16-0) [2021](#page-16-0)). Therefore, it is imperative to maintain the equilibrium of metals within the mitochondria. ATMs and mitochondrial iron transporters (MITs) are the two primary categories of transporters that modulate metal levels in mitochondria ([Nouet et al., 2011](#page-17-0)).

Golgi apparatus is an essential component of the endomembrane system, which plays a crucial role in directing membrane-bound proteins (such as transporters) to either the plasma membrane or organelles [\(Jogawat et al., 2021](#page-16-0)). The major purpose of this is to regulate the balance of metals in the body. Metal transporters are also found in the Golgi apparatus to detect and import metals for their arrangement ([Bressler et al., 2007\)](#page-14-0). When exposed to metal stress, the Golgi apparatus system responds by reorganizing the endomembrane system and storing excess metals in vesicles [\(De Caroli et al., 2020\)](#page-15-0). This process involves reducing the number of metal transporters at the plasma membrane. This substantially reduces the detrimental impacts of metals ([De Caroli et al., 2020](#page-15-0)).

TABLE 1 List of gene families that play a role in metal transport.

(Continued)

Finally, endoplasmic reticulum (ER) is also part of the endomembrane system and has a vast lumen to store and utilize metals for different purposes [\(De Caroli et al., 2020\)](#page-15-0). Broadspecificity transporters for Cd, Cu, and Zn are identified on the ER membrane and are sometimes found to be localized on the plasma membrane. This common localization might be due to the continuum of the endomembrane system as part of the secretory pathway ([Jogawat et al., 2021\)](#page-16-0).

7 Molecular, biochemical, and metabolic plant responses toward HMs

7.1 Negative impact of metals on plants

Exposure to HMs causes several reactions in plants, including physiological, biochemical, and agricultural production responses ([Singh et al., 2020\)](#page-18-0). The toxicity of HMs is influenced by a variety of factors, such as the plant species, the concentration of the individual metal, its chemical structure, soil composition, and pH level [\(Abd](#page-14-0) [Elnabi et al., 2023](#page-14-0)). Certain HMs, including Cu and Zn, are essential for the vegetative plant growth ([Arif et al., 2016\)](#page-14-0). HMs can participate in enzyme processes by forming complexes with enzymes and substrates, acting as cofactors and activators ([Witkowska et al., 2021\)](#page-19-0). Trace metal nutrients have a vital role in redox reactions, electron transportation, and structural functions in nucleic acid processing ([Sunda, 2012](#page-18-0)).

Additionally, HMs have various effects on the functioning of the photosynthetic system at different levels of organization ([Ventrella](#page-19-0) [et al., 2009\)](#page-19-0). HMs directly affect plants by interfering with the PS I and PS II processes and indirectly affect photosynthesis, growth, and yield [\(Singh et al., 2015](#page-18-0)). Additionally, certain HMs, including Cd and Hg, possess phytotoxic properties that impede metalsensitive enzymes, resulting in growth retardation and the mortality of organisms [\(Alengebawy et al., 2021\)](#page-14-0). HMs can be classified into two categories based on their ability to undertake redox reactions: redox-active and redox-inactive [\(Ercal et al., 2001\)](#page-15-0). The redox reaction within cells is facilitated by redox-active transition metals, including Fe, Cu, Cr, and Co ([Kostenkova et al.,](#page-16-0) [2022\)](#page-16-0). This process leads to the production of superoxide $(O_2^{\bullet -})$, hydrogen peroxide (H_2O_2) , and hydroxyl radicals $(\bullet$ OH) ([Collin,](#page-15-0) [2019\)](#page-15-0). The oxidative stress is induced by the indirect interactions

with the antioxidant defense system, disruption of the electron transport chain, and induction of lipid peroxidation, which are the results of exposure to redox-inactive HMs ([Bhattacharyya](#page-14-0) [et al., 2014](#page-14-0)).

In plants, HMs are involved in the production and release offree radicals through chemical reactions, metabolic pathways, and physiological processes [\(Emamverdian et al., 2015](#page-15-0)). The ROS are generated by biological systems through the production of radicals that are centered on oxygen, S, N, and carbon ([Phaniendra et al.,](#page-17-0) [2015](#page-17-0)). The lipid content of thylakoid membranes is altered in plants that are exposed to HMs stress, which leads to membrane degradation and lipid peroxidation ([Emamverdian et al., 2015\)](#page-15-0). The primary site of lipid peroxidation is polyunsaturated fatty acids, and it is comprised of three distinct phases: initiation, advancement, and termination. The functionality of PS II is impeded by HMs, which leads to an increase in the formation of O_2 ⁻⁻ in leaves and an increase in lipid peroxidation ([Hasanuzzaman et al., 2020\)](#page-16-0). Recent research has revealed that HMs can damage numerous physiological systems by generating ROS that induce lipid peroxidation [\(Shahid et al., 2014](#page-18-0)). Furthermore, the rate of photosynthesis and PS II can be significantly affected by the byproducts of lipid peroxidation (Pospiš[il and Yamamoto, 2017](#page-18-0)).

Chlorophylls (Chl) and carotenoids are essential pigments that are involved in the conversion of solar energy to chemical energy during photosynthesis ([Hashimoto et al., 2016](#page-16-0)). The production of photosynthetic compounds is specifically influenced by HMs ([Ventrella et al., 2009\)](#page-19-0). Chlorosis and plant growth retardation are frequently observed in metal-contaminated environments. These findings suggest that the biosynthesis of photosynthetic compounds has been disrupted [\(Yadav, 2010](#page-19-0)). Consequently, these variables influence the proliferation of plastids, the efficiency of photosynthesis, and the overall metabolism. Additionally, HMs inhibit the accumulation of photosynthetic compounds ([Ventrella et al., 2009\)](#page-19-0).

7.2 The mechanism of HM uptake and tolerance

The development of plants as phytoremediation agents is contingent upon an understanding of the genetic basis and interrelated network of physiological and molecular mechanisms that govern plant tolerance to specific HMs [\(Hossain et al., 2012\)](#page-16-0). Different plant species may have developed distinct mechanisms to tolerate excessive HMs, and even within a single plant species, multiple mechanisms may be in operation ([Asiminicesei et al.,](#page-14-0) [2024\)](#page-14-0). To endure excessive HMs, plants possess both constitutive and adaptive mechanisms ([Hasanuzzaman et al., 2013\)](#page-16-0). To identify the underlying mechanisms of HMs' accumulation, tolerance, and adaptive mechanisms to contend with HM stress, physiological, biochemical, and molecular approaches are still being employed ([Mashabela et al., 2023](#page-17-0)).

Among the adaptive mechanisms that tolerant plants have evolved are the synthesis of particular phytochelatins (PCs) and metallothioneins (MTs), induction of mechanisms opposing the effects of ROS and MG, induction of stress proteins, the biosynthesis of proline (Pro), polyamines (PAs), and signaling molecules like salicylic acid (SA) and nitric oxide (NO) ([Hossain](#page-16-0) [et al., 2012](#page-16-0); [Hasanuzzaman et al., 2019](#page-16-0)). Figure 3 demonstrates the process of HMs' sequestration in plant cells, specifically within the vacuoles. HMs are absorbed by plants through root interception, entrance into roots, and translocation to the stem [\(Khan et al.,](#page-16-0) [2023\)](#page-16-0). The entrance of HMs into the organism is contingent upon the sort of HM [\(Yimer et al., 2024\)](#page-19-0).

The prevention of superfluous HMs from infiltrating the plant is one method of reducing or preventing the toxic effects of HMs.

Plants can accomplish this by precipitating or complexing HMs in the root environment ([Riyazuddin et al., 2021](#page-18-0)). Plants can precipitate HMs by either increasing the pH of the rhizosphere or excreting anions, such as phosphate ([Hinsinger et al., 2003;](#page-16-0) [Chen](#page-15-0) [et al., 2017](#page-15-0)). In response to Al stress, root exudation of phosphate has been observed in maize (Calderón-Vá[zquez et al., 2011\)](#page-15-0). Additionally, malate exudation from the roots of sorghum and citrate exudation from the roots of maize have also been documented in response to Cd stress ([Piñeros et al., 2002\)](#page-18-0). These results lend credence to the hypothesis that the HM-binding capabilities of root exudates may serve as a critical mechanism for stabilizing HMs in the vicinity of the root, thereby rendering them unavailable to the plant and reducing the toxicity encountered by the plant [\(Hossain et al., 2012](#page-16-0)).

There must also be other processes since certain tolerant and hyperaccumulator plants really absorb more HMs than sensitive plants. An essential adaptive strategy for HM tolerance in plants is the cellular exclusion of HMs ([Riyazuddin et al., 2021\)](#page-18-0). The apoplastic space is the location of a significant proportion of HMs in plant roots, which implies an exclusion mechanism ([Sattelmacher, 2001](#page-18-0)). The cell wall–plasma membrane interface has the potential to serve as a site of HM tolerance, as it accumulates substantial amounts of HMs [\(Danouche et al., 2021\)](#page-15-0). The plant

FIGURE 3

Sequestration of heavy metals in plant cells within the vacuoles. The uptake of heavy metal (HMs) ions is facilitated by a variety of transporters, such as the cation diffusion facilitator (CDF) family, the heavy-metal-transporting ATPase (HM ATPase), the copper transporter (CTR), the zinc-regulated, iron-regulated transporter-like proteins (ZIP), and the natural resistance-associated macrophage protein (NRAMP). For instance, HMs, such as Cd² enter the cytosol and initiate the production of phytochelatins (PCs) after being transported by members of the ZIP family. PCs are produced through a transpeptidation reaction from reduced glutathione (GSH) in a non-translational manner. The primary function of PCs is to bind cytosolic HMs, which results in the formation of the HM–PC complex. In the case of Cd²⁺ ions, these bind with low-molecular-weight (LMW) complex and form the LMW Cd-complex. This complex is subsequently transported into the vacuole by a tonoplast-localized ATP-binding-cassette (ABC) transporter. The LMW Cd-complex is collected and converted into a high-molecular-weight (HMW) complex within the vacuole. This complex contains supplementary Cd²⁺ ions. The tonoplast-localized cation/proton exchanger (CAX) transporters facilitate the direct interaction between these HMW complexes and protons, thereby enabling them to access the vacuole. Metal tolerance proteins (MTPs) and NRAMPs are transporters that are present in the tonoplast. They are accountable for the migration of metal ions to facilitate compartmentalization or remobilization. Organic acids, amino acids, and metallothioneins (MTs) are among the chelators that contribute to the regulation of metal levels in the cytosol to a safe and low level. ROS, reactive oxygen species. This figure was made using BioRender.

cation exchange capacity (CECs) of sensitive wheat cultivars is substantially lower than that of tolerant cultivars ([Masion and](#page-17-0) [Bertsch, 1997](#page-17-0)). This implies that tolerant cultivars utilize a high CEC to complex HMs at the cell wall and obstruct their entry into the cell [\(Bortoloti and Baron, 2022](#page-14-0)).

Once HMs infiltrate the cell, plants employ a variety of strategies to mitigate their toxicity [\(Pande et al., 2022](#page-17-0)). One approach is to transport or sequester HMs into the vacuole, which serves as an appropriate storage reservoir for excessively accumulated HMs [\(Peng and Gong, 2014\)](#page-17-0). Vacuolar assimilation of the majority of solutes is stimulated by two vacuolar proton pumps: a vacuolar proton-ATPase (V-ATPase) and a vacuolar proton pyrophosphatase (V-Ppase) [\(Hossain et al., 2012\)](#page-16-0). Grasses have the capacity to actively transport Zn into vacuoles, with more tolerant clones being able to maintain the process at higher external Zn levels than sensitive clones ([Brookes et al., 1981\)](#page-15-0). Either channels or transporters can facilitate uptake. Genetic and molecular techniques have identified a variety of genes ([Table 2\)](#page-10-0) that are involved in the uptake of transition HM ions into cells, the sequestration of HMs in the vacuole, the remobilization of HMs from the vacuole, the loading of HMs in the xylem, and the discharge of HMs ([Gill et al., 2021](#page-16-0)).

Zn-regulated transporter (ZRT), Fe-regulated transporter (IRT)-like protein ZIP family, ATP-binding cassette (ABC) transporters, the P-type metal ATPases, NRAMP family, multidrug resistance-associated proteins (MRP), CDF family of proteins, copper transporter (COPT) family proteins, pleiotropic drug resistance (PDR) transporters, YSL transporter, and CAX are among the well-characterized HM transporter proteins [\(Toyoda](#page-19-0) [et al., 2008;](#page-19-0) [Pittman and Hirschi, 2016;](#page-18-0) [Wang et al., 2021a;](#page-19-0) [Pacheco](#page-17-0) [et al., 2023](#page-17-0)). MTs and PCs are two forms of peptide metal-binding ligands that are essential for the detoxification and tolerance of HMs in plants that are subjected to HM stress [\(Faizan et al., 2024\)](#page-15-0).

PCs are synthesized from GSH and are induced by a variety of HMs, including Cd, Hg, Ag, Cu, Ni, Au, Pb, As, and Zn ([Faizan et al.,](#page-15-0) [2024](#page-15-0)). The activity of ABC transporters accumulates them in the vacuole, thereby restricting the circulation of free Cd^{2+} within the cytosol, and they complex Cd ions through the thiolic group (–SH) of cysteine ([Salbitani et al., 2023](#page-18-0)). PCs are produced by both HMresistant and HM-sensitive plants; however, certain reports have concluded that PCs are not the primary cause of the hyperaccumulation of Zn, Ni, or Pb [\(Hossain et al., 2012\)](#page-16-0). The chelation of HM ions is not the sole mechanism of the HM detoxification process ([Gulcin and Alwasel, 2022](#page-16-0)). The HM ion complex is transported to the vacuole and stabilized, thereby forming a complex with sulfides or organic acid, following the activation of PC synthase by HM ions and HM chelation by the synthesized PCs ([Faizan et al., 2024](#page-15-0)). Nevertheless, the HM specificity or species specificity of hyperaccumulation is not adequately elucidated by the formation of HM complexes. Consequently, the precise function of PCs in the HM tolerance mechanism at the cellular level is yet to be ascertained [\(Emamverdian et al., 2015\)](#page-15-0).

Plant MTs are polypeptides that are cysteine-rich, low molecular weight, and capable of engaging HMs through their cysteine residues [\(Freisinger, 2011](#page-15-0)). Their physiological functions encompass the protection against intracellular oxidative damage,

the sequestration of toxic HMs, and the maintenance of essential transition HM homeostasis ([Subramanian Vignesh and Deepe,](#page-18-0) [2017\)](#page-18-0). The cysteine residue arrangement has resulted in the division of plant MTs into three classes, which are diverse [\(Guo](#page-16-0) [et al., 2003](#page-16-0)). The organization of cysteine residues confers distinct MT isoforms and their capacity to bind and sequester distinct HM ions for the purposes of detoxification and homeostasis ([Ruttkay-](#page-18-0)[Nedecky et al., 2013\)](#page-18-0).

Factors such as hormones, cytotoxic agents, and HMs induce MT biosynthesis, which is regulated at the transcriptional level ([Thirumoorthy et al., 2007](#page-19-0)). Gene expression studies have demonstrated that MT genes are differentially regulated in response to a variety of HM stresses ([Qu et al., 2024](#page-18-0)). The role of MTs in HM detoxification and homeostasis has been demonstrated by a variety of data [\(Ruttkay-Nedecky et al., 2013\)](#page-18-0). However, the metal-inducibility of plant MTs has not always been demonstrated. Additional information regarding the structures and properties of MTs could provide a more comprehensive understanding of their functions and mechanism(s) of action [\(Hossain et al., 2012\)](#page-16-0). The molecular mechanisms of HM transport, trafficking, tolerance, and homeostasis in plants are likely to be further elucidated through the use of a model system and a model hyperaccumulator, such as Arabidopsis halleri and particularly Thlaspi species [\(Pasricha](#page-17-0) [et al., 2021](#page-17-0)).

In plants, metal chelation can be classified into two categories: internal tolerance and external exclusion. During the external detoxification process, organic acids expelled from plant roots may combine with HM ions to create stable HM–ligand complexes, which may change the HM ions' mobility and bioavailability ([Sabreena et al., 2022\)](#page-18-0). This obstructs the entry of HM ions into plants and prevents their accumulation in sensitive root sites. Organic acids may chelate with HM in the cytosol during internal HM detoxification, resulting in the transformation of the ions into a less toxic or nontoxic form [\(Gasic and Korban, 2006\)](#page-16-0). Plants generate a variety of ligands for Al, Cd, Cu, Ni, Co, and Zn. Potential ligands for HMs include carboxylic and amino acids, including citrate, malate, and oxalate, histidine and nicotianamine, and phosphate derivatives (phytate), which are involved in detoxification and tolerance ([Hossain et al., 2012\)](#page-16-0). Citrate has a significant affinity for chelating HM ions, and other HMs, including Cd, Ni, Co, and Zn, also exhibit a high affinity for citrate ([Gasic and](#page-16-0) [Korban, 2006;](#page-16-0) [Hossain et al., 2012\)](#page-16-0).

At low Cd concentrations, citric acid is a significant ligand and contributes to the accumulation and tolerance of Zn [\(Najeeb et al.,](#page-17-0) [2011](#page-17-0)). HMs such as Al are also detoxified, and oxalate is secreted by the roots [\(Zheng et al., 2005\)](#page-19-0). In response to Al stress, buckwheat (Fagopyrum esculentum Moench.) secretes oxalic acid from the roots and accumulates nontoxic-Al-oxalate in the leaves ([Feng Ma](#page-15-0) [et al., 1998\)](#page-15-0). Consequently, detoxification occurs both internally and externally [\(Hall, 2002](#page-16-0)). Histidine and nicotianamine are also involved in the chelation of HM ions in the xylem fluid and within plant cells ([Zhakypbek et al., 2024\)](#page-19-0). Nicotianamine is a nonproteinogenic amino acid that is mobile within the plant and has been identified in phloem fluid as well as in root and leaf cells ([Klatte et al., 2009](#page-16-0)). It is suggested that it may be involved in the regulation of HM transfer within plant cells [\(Takahashi et al., 2003\)](#page-19-0).

TABLE 2 Compilation of genes exhibiting variable expression in response to various heavy metals.

(Continued)

TABLE 2 Continued

Plants that are HM-tolerant frequently prevent HMs from being transmitted from root to stem by either detoxifying them through chelation or storage or retaining them in root cells ([Singh et al.,](#page-18-0) [2015\)](#page-18-0). Nevertheless, a unique group of plants known as

hyperaccumulators effectively transport HMs to the shoot through the xylem, a process that is likely facilitated by transpiration. They are capable of accumulating HMs from modest external concentrations, with the majority of them being translocated to the shoot ([Yang et al., 2005](#page-19-0)). Hyperaccumulators exhibit an unusually high uptake of HM at the root membrane level, which may be attributed to the presence of a high expression of an HM transporter in the plasma membrane [\(Skuza et al., 2022\)](#page-18-0). Efficient intracellular compartmentalization and chelation may be the cause of this high HM tolerance.

A complex network of biochemical adaptive strategies, known as the antioxidant system, is present in plants to detoxify a variety of ROS (Dumanović [et al., 2020\)](#page-15-0). In general, this system can be divided into two categories. The first group comprises enzymes, including superoxide dismutase, catalase, ascorbate peroxidase, and glutathione reductase, that eliminate oxygen radicals and their metabolites [\(Rajput et al., 2021](#page-18-0)). Nonenzymatic compounds such as glutathione, ascorbate, and phenolics comprise the second group. These compounds have the ability to neutralize ROS without transforming into deleterious radicals themselves [\(Zandi and](#page-19-0) [Schnug, 2022\)](#page-19-0).

The presence of the two HMA proteins, hma2/hma4, is essential for the uptake of Cd in the shoot via the xylem ([Kraemer, 2009\)](#page-16-0). When cultivated on regular soil, the shoot of the hma2/hma4 double-mutant shows severe signs of Zn shortage, as previously shown [\(Haydon and Cobbett, 2007\)](#page-16-0). This suggests that both HMA2 and HMA4 are required for the movement of Zn from the roots to the shoots in regular soils ([Claus et al., 2013](#page-15-0)). On the other hand, the heat shock proteins (HSPs) are molecular chaperones that are essential for the protection and repair of proteins under stress conditions as well as for the folding and assembly of proteins [\(El-](#page-15-0)[Sappah et al., 2017](#page-15-0); [Abbas et al., 2022\)](#page-14-0). In response to Cd stress, they can enhance the accumulation of large HSPs, such as HSP70, and are induced by transition metals (Zn, Cu, Cd, Hg, Al, and Cr) ([Hasan et al., 2017\)](#page-16-0).

Additionally, HSPs can prevent irreversible protein denaturation as a result of oxidative stress or facilitate proteolytic degradation [\(Kumar et al., 2022](#page-17-0)). Nevertheless, the extent of their involvement in HM tolerance is still mainly obscure. Enzymes that modify metal oxidation states or facilitate the incorporation of HMs into organic molecules are known as metal-modifying enzymes ([Kaczmarek et al., 2009\)](#page-16-0). After being supplied with Cr(VI) in nutrient culture, Eichhornia crassipes, a water hyacinth, accumulated innocuous Cr(III) in its root and branch tissues ([Giri and Patel, 2011](#page-16-0)). This implies that E. crassipes detoxified Cr (VI) during root assimilation and transported a portion of the detoxified Cr to leaf tissues. A reductase at the root cell membrane in dicots reduces Fe and potentially Cu prior to assimilation ([Cohen](#page-15-0) [et al., 1997\)](#page-15-0).

Plants may reduce harmful substances (e.g., HMs) through a process called in situ reduction, which can be advantageous for phytoremediation by helping to detoxify the environment [\(Ka](#page-16-0)fle [et al., 2022](#page-16-0)). Metal-responsive transcription factor 1 (MTF-1) plays a crucial role in the cellular response and tolerance to HM stress by activating genes important for HM uptake, transport, and detoxification [\(Wang et al., 2004\)](#page-19-0). The TFs involved in the reaction to HM stress and tolerance have been reported in several plant species ([Niekerk et al., 2024\)](#page-17-0). On the other hand, oxidative stress and antioxidative defense systems are induced by HM stress ([Mansoor et al., 2023](#page-17-0)). These systems are constituted offree-radical-

scavenging molecules, such as ascorbate (AsA) and GSH, and the enzymes involved in their biosynthesis and reduction ([Rajput](#page-18-0) [et al., 2021](#page-18-0)).

During times of stress, specifically HM stress, SA interacts with many plant hormones, including AUX, ABA, and GA, to promote the synthesis of antioxidant chemicals and enzymes. This interaction serves to notify and assist plants treated with HM, helping to alleviate the stress caused by HM [\(Sharma et al., 2020\)](#page-18-0). SA is a natural signal molecule that is crucial for the regulation of physiological and biochemical processes, thereby enhancing the resistance of plants to biotic and abiotic stresses ([Mishra](#page-17-0) [et al., 2024](#page-17-0)).

On the other hand, Pro accumulation in response to HM stress has also been extensively documented ([Hayat et al., 2012\)](#page-16-0). Enhanced protection against Cd stress is provided by increased Pro levels in microalgae ([Siripornadulsil et al., 2002](#page-18-0)). Pro plays a vital role in mitigating the harmful effects of Cd stress by protecting against damage caused by free radicals and maintaining a controlled reducing environment within the cell rather than just isolating Cd ([Hayat et al., 2012\)](#page-16-0). However, PAs are organic cations that exist naturally and possess nonenzymatic antioxidant characteristics. They are thought to function as second messengers in regulating plant growth and development processes ([Raychaudhuri](#page-18-0) [et al., 2021](#page-18-0)).

PAs and Pro are components of the "general adaptation syndrome" (GAS) response to environmental adversities, including nutrient scarcity, HMs, and low temperatures [\(Gill](#page-16-0) [et al., 2012\)](#page-16-0). Engineered plants that overexpress genes involved in the biosynthesis of PAs exhibit an enhanced ability to withstand a range of environmental stressors, including HMs [\(Kajla et al., 2023\)](#page-16-0). By modulating the level and toxicity of ROS and hormones, NO, a ubiquitous bioactive signaling molecule, serves a critical function in a wide range of physiological processes in plants ([Jomova et al.,](#page-16-0) [2023](#page-16-0)). By governing the general mechanisms for cellular redox homeostasis and promoting the transformation of O_2 ⁺⁻ to H_2O_2 and O2, NO safeguards plants from oxidation damage [\(Huang](#page-16-0) [et al., 2019](#page-16-0)).

NO may also safeguard cells from oxidative processes by promoting the synthesis of GSH ([Lu, 2013](#page-17-0)), in addition to its direct ROS scavenging activity and the modulation of lipid peroxidation through lipoxygenase (LOX) inhibition. Under HM stress, exogenous NO can effectively induce tomato seedlings to modify their physiological and biochemical mechanisms to protect against Cu toxicity, thereby preserving their metabolic capacity and normal growth capabilities ([Singh et al., 2015\)](#page-18-0).

8 Challenges and prospective

It is crucial to examine the interactions between different HMs in plants. Certain HMs may have synergistic effects, whereby their collective presence amplifies toxicity beyond what would be anticipated based on individual doses ([Angon et al., 2024\)](#page-14-0). Conversely, some combinations may have antagonistic effects, where one metal reduces the toxicity of another. In order to assess and control risks, it is important to acknowledge these linkages. Subsequent research should strive to measure the combined impacts of HMs on the physiological processes, growth, and reproductive capabilities of plants. In order to do this, researchers may examine the correlations between dosage and response, the patterns of bioaccumulation, and the effects, particularly on different tissues. There has been a scarcity of research on agricultural genetic diversity and the mechanisms of plant adaptation. The sensitivity and tolerance of plants to HMs might vary depending on the species and genotypes [\(Emamverdian](#page-15-0) [et al., 2015\)](#page-15-0).

Studying the way various plant species and genotypes react to environmental difficulties might impede the progress of creating metal-tolerant agricultural cultivars. It is crucial to bridge this gap in order to comprehend the ability of plants to withstand challenges, provide guidance for agricultural activities in polluted areas, and assist in environmental remediation efforts. Furthermore, investigating the transfer of HMs to crops is crucial for the development of sustainable agriculture. Gaining insight into the mechanisms by which HMs persist and migrate throughout successive plant generations may provide valuable guidance for addressing pollution in affected areas. This knowledge can be particularly useful in developing phytoremediation systems that effectively eliminate toxins from contaminated soil.

The current scientific investigations have mostly concentrated on the detection and analysis of microplastics in soil. Nevertheless, there is an increasing curiosity in comprehending the mechanisms by which these minute plastic particles might carry HMs and influence their dispersion and accessibility. It is crucial to enhance several techniques that aid in reducing and managing the stress caused by harmful substances in plants. Using a single technique is ultimately unrealistic and inadequate for effectively restoring soil that has been polluted with HMs [\(Priya et al., 2023\)](#page-18-0). There have been numerous methods developed to mitigate or prevent HM pollution and to reestablish vegetation in polluted soil ([Priya et al., 2023](#page-18-0)).

Restoring the flora of soil that has been contaminated with HMs is a highly promising approach known as phytoremediation [\(Yan](#page-19-0) [et al., 2020\)](#page-19-0). Public acceptance has been achieved, and it offers numerous advantages over other physicochemical treatments ([Mitra et al., 2022\)](#page-17-0).

The most effective and cost-efficient approach is the recent emergence of the introduction of nanoparticles (NPs) into plants to increase their tolerance to HM toxicity and facilitate the cleansing of these toxic elements [\(Zhou et al., 2020](#page-19-0)). Genetic engineering is a valuable method for modifying plants to manifest specific characteristics, including rapid growth, high biomass output, strong tolerance and accumulation of HMs, and adaptability to a variety of climatic and geological conditions ([Yan et al., 2020\)](#page-19-0). As a result, it will be essential to have a thorough understanding of the processes by which plants absorb, transport, and eliminate HMs as well as the identification and analysis of a variety of molecules and signaling pathways in order to create genetically engineered plant species that are optimal for phytoremediation ([Priya et al., 2023\)](#page-18-0). Enhanced tolerance or accumulation of HMs in plants may be achieved by manipulating genes associated with HM absorption,

translocation, sequestration, and tolerance. In addition, the bioavailability of HMs may be improved by the use of chelating compounds and microorganisms, which, in turn, facilitates their accumulation in plants [\(Olaniran et al., 2013](#page-17-0)). In addition, they can be employed to improve the health of the soil and to further encourage the growth and fitness of plants. Several hyperaccumulator plants have been identified, and the most direct method for phytoremediation is the use of HM hyperaccumulators ([Skuza et al., 2022\)](#page-18-0). However, there are certain limitations that impede the utilization of these natural hyperaccumulators in phytoremediation [\(Yan et al., 2020\)](#page-19-0).

Due to their capacity to penetrate plants extensively, exhibit superior adsorption, and deliver targeted effects, they may be instrumental in the regulation of photosynthesis and the detoxification of ROS [\(Rasheed et al., 2022\)](#page-18-0). Subsequently, they can substantially improve the germination, growth, and yield of plant seeds (Kornarzyń[ski et al., 2020\)](#page-16-0). NPs also facilitate plant growth by modulating the movement and distribution of both mobile and immobile forms of HMs [\(Zhou et al., 2020](#page-19-0)). The potential of NPs to significantly improve the remediation of metal-contaminated soils in the future is suggested by the positive results observed in the use of NPs, particularly in the enhancement of plants' resistance to HMs and the facilitation of their development.

9 Conclusion

HMs are defined as metals with densities more than 5 g cm^3 . HMs account for 53 of the almost 90 elements found in nature. Plant nutrition is thought to need a grand total of 18 components. Some of its parts are thought to have positive effects. HMs in the soil can have a negative impact on human and animal health, soil quality, fertility, and agricultural productivity. Plants have developed ways to deal with HM stress, such as immobilization, exclusion outside the plasma membrane, limited absorption and transport, production of specific HM transporters, activation of stress proteins, and chelation and sequestration by specific ligands.

Metals have two distinct impacts on seed germination: their overall toxicity and their ability to hinder water uptake. Plants absorb and store HMs using concentration gradients and selective absorption. These chemicals influence enzymes, cellular metabolism, and the production of nucleic acids, proteins, and pigments for photosynthesis. Recent molecular and genetic research has identified gene families that play a role in metal transport, contributing to metal tolerance in hyperaccumulator plants. Understanding the interactions between different HMs in plants is crucial for assessing and controlling risks.

Research on agricultural genetic diversity and plant adaptation mechanisms is also essential for developing metal-tolerant agricultural cultivars and reducing the harmful impact of contaminated soil on crop growth and productivity. Finally, the current review examined the locations in plants where HMs are found. It discussed the benefits of HMs to plants, the consequences on seed performance, and the initial plant reaction to HM exposure. This review also looks at the existence of HM transport genes in plants and how plants react to HMs on a molecular, biochemical, and metabolic level, respectively. Methods for managing HMs in plants, together with the associated challenges and opportunities, have been also examined.

Author contributions

AE-S: Conceptualization, Funding acquisition, Validation, Visualization, Writing – original draft, Writing – review & editing. YZ: Visualization, Writing – review & editing. QH: Funding acquisition, Visualization, Writing – review & editing. BC: Writing – review & editing. SS: Writing – review & editing. MA: Writing – review & editing. KY: Conceptualization, Writing – review & editing. JL: Conceptualization, Funding acquisition, Writing – review & editing. KE-T: Conceptualization, Funding acquisition, Validation, Visualization, Writing – review & editing.

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References

Abbas, M., Li, Y., Elbaiomy, R. G., Yan, K., Ragauskas, A. J., Yadav, V., et al. (2022). Genome-wide analysis and expression profiling of SlHsp70 gene family in Solanum lycopersicum revealed higher expression of SlHsp70-11 in roots under Cd^{2+} stress. Front. Biosci. 27, 186. doi: [10.31083/j.fbl2706186](https://doi.org/10.31083/j.fbl2706186)

Abd Elnabi, M. K., Elkaliny, N. E., Elyazied, M. M., Azab, S. H., Elkhalifa, S. A., Elmasry, S., et al. (2023). Toxicity of heavy metals and recent advances in their removal: a review. Toxics 11, 580. doi: [10.3390/toxics11070580](https://doi.org/10.3390/toxics11070580)

Adnan, M., Xiao, B., Ali, M. U., Xiao, P., Zhao, P., Wang, H., et al. (2024). Heavy metals pollution from smelting activities: a threat to soil and groundwater. Ecotoxicol. Environ. Saf. 274, 116189. doi: [10.1016/j.ecoenv.2024.116189](https://doi.org/10.1016/j.ecoenv.2024.116189)

Adrees, M., Ali, S., Rizwan, M., Ibrahim, M., Abbas, F., Farid, M., et al. (2015). The effect of excess copper on growth and physiology of important food crops: a review. Environ. Sci. pollut. Res. 22, 8148–8162doi: [10.1007/s11356-015-4496-5](https://doi.org/10.1007/s11356-015-4496-5)

Alam, M. M., Hayat, S., Ali, B., and Ahmad, A. (2007). Effect of 28-homobrassinolide treatment on nickel toxicity in Brassica juncea. Photosynthetica 45, 139–142. doi: [10.1007/s11099-007-0022-4](https://doi.org/10.1007/s11099-007-0022-4)

Alejandro, S., Höller, S., Meier, B., and Peiter, E. (2020). Manganese in plants: from acquisition to subcellular allocation. Front. Plant Sci. 11, 300. doi: [10.3389/](https://doi.org/10.3389/fpls.2020.00300) [fpls.2020.00300](https://doi.org/10.3389/fpls.2020.00300)

Alengebawy, A., Abdelkhalek, S. T., Qureshi, S. R., and Wang, M. Q. (2021). Heavy metals and pesticides toxicity in agricultural soil and plants: ecological risks and human health implications. Toxics 9, 42. doi: [10.3390/toxics9030042](https://doi.org/10.3390/toxics9030042)

Ali, H., and Khan, E. (2018). What are heavy metals? Long-standing controversy over the scientific use of the term 'heavy metals' – proposal of a comprehensive definition. Toxicol. Environ. Chem. 100, 6–19. doi: [10.1080/02772248.2017.1413652](https://doi.org/10.1080/02772248.2017.1413652)

Angon, P. B., Islam, M. S., Kc, S., Das, A., Anjum, N., and Poudel, A. (2024). Sources, effects and present perspectives of heavy metals contamination: soil, plants and human food chain. Heliyon 10, e28357. doi: [10.1016/j.heliyon.2024.e28357](https://doi.org/10.1016/j.heliyon.2024.e28357)

Angulo-Bejarano, P. I., Puente-Rivera, J., and Cruz-Ortega, R. (2021). Metal and metalloid toxicity in plants: an overview on molecular aspects. Plants 10, 635. doi: [10.3390/plants10040635](https://doi.org/10.3390/plants10040635)

Aponte, H., Meli, P., Butler, B., Paolini, J., Matus, F., Merino, C., et al. (2020). Metaanalysis of heavy metal effects on soil enzyme activities. Sci. Total Environ. 737, 139744. doi: [10.1016/j.scitotenv.2020.139744](https://doi.org/10.1016/j.scitotenv.2020.139744)

Arif, N., Yadav, V., Singh, S., Singh, S., Ahmad, P., Mishra, R. K., et al. (2016). Influence of high and low levels of plant-beneficial heavy metal ions on plant growth and development. Front. Environ. Sci. 4, 69. doi: [10.3389/fenvs.2016.00069](https://doi.org/10.3389/fenvs.2016.00069)

Arrivault, S., Senger, T., and Krämer, U. (2006). The Arabidopsis metal tolerance protein AtMTP3 maintains metal homeostasis by mediating Zn exclusion from the shoot under Fe deficiency and Zn oversupply. Plant J. 46, 861–879. doi: [10.1111/j.1365-](https://doi.org/10.1111/j.1365-313X.2006.02746.x) [313X.2006.02746.x](https://doi.org/10.1111/j.1365-313X.2006.02746.x)

Asiminicesei, D. M., Fertu, D. I., and Gavrilescu, M. (2024). Impact of heavy metal pollution in the environment on the metabolic profile of medicinal plants and their therapeutic potential. Plants 13, 913. doi: [10.3390/plants13060913](https://doi.org/10.3390/plants13060913)

Baker, A. J. M. (1981). Accumulators and excluders strategies in response of plants to heavy metals. J. Plant Nutr. 3, 643–654. doi: [10.1080/01904168109362867](https://doi.org/10.1080/01904168109362867)

Batool, T. S., Aslam, R., Gul, A., Paracha, R. Z., Ilyas, M., De Abreu, K., et al. (2023). Genome-wide analysis of heavy metal ATPases (HMAs) in Poaceae species and their potential role against copper stress in Triticum aestivum. Sci. Rep. 13, 7551. doi: [10.1038/s41598-023-32023-7](https://doi.org/10.1038/s41598-023-32023-7)

Begum, W., Rai, S., Banerjee, S., Bhattacharjee, S., Mondal, M. H., Bhattarai, A., et al. (2022). A comprehensive review on the sources, essentiality and toxicological profile of nickel. RSC. Adv. 12, 9139–9153. doi: [10.1039/D2RA00378C](https://doi.org/10.1039/D2RA00378C)

Bhati, K. K., Sharma, S., Aggarwal, S., Kaur, M., Shukla, V., Kaur, J., et al. (2015). Genome-wide identification and expression characterization of ABCC-MRP transporters in hexaploid wheat. Front. Plant Sci. 6, 488. doi: [10.3389/fpls.2015.00488](https://doi.org/10.3389/fpls.2015.00488)

Bhattacharjee, B., Ali, A., Tuteja, N., Gill, S., and Pattanayak, A. (2023). Identification and expression pattern of aluminum-responsive genes in roots of rice genotype with reference to Al-sensitivity. Sci. Rep. 13, 12184. doi: [10.1038/s41598-023-39238-8](https://doi.org/10.1038/s41598-023-39238-8)

Bhattacharyya, A., Chattopadhyay, R., Mitra, S., and Crowe, S. E. (2014). Oxidative stress: an essential factor in the pathogenesis of gastrointestinal mucosal diseases. Physiol. Rev. 94, 329–354. doi: [10.1152/physrev.00040.2012](https://doi.org/10.1152/physrev.00040.2012)

Bielen, A., Remans, T., Vangronsveld, J., and Cuypers, A. (2013). The influence of metal stress on the availability and redox state of ascorbate, and possible interference with its cellular functions. Int. J. Mol. Sci. 14, 6382–6413. doi: [10.3390/ijms14036382](https://doi.org/10.3390/ijms14036382)

Bortoloti, G. A., and Baron, D. (2022). Phytoremediation of toxic heavy metals by Brassica plants: a biochemical and physiological approach. Environ. Adv. 8, 100204. doi: [10.1016/j.envadv.2022.100204](https://doi.org/10.1016/j.envadv.2022.100204)

Branca, J. J. V., Fiorillo, C., Carrino, D., Paternostro, F., Taddei, N., Gulisano, M., et al. (2020). Cadmium-induced oxidative stress: focus on the central nervous system. Antioxidants 9, 492. doi: [10.3390/antiox9060492](https://doi.org/10.3390/antiox9060492)

Bressler, J. P., Olivi, L., Cheong, J. H., Kim, Y., Maerten, A., and Bannon, D. (2007). Metal transporters in intestine and brain: their involvement in metal-associated neurotoxicities. Hum. Exp. Toxicol. 26, 221-229. doi: [10.1177/0960327107070573](https://doi.org/10.1177/0960327107070573)

Brookes, A., Collins, J. C., and Thurman, D. A. (1981). The mechanism of zinc tolerance in grasses. J. Plant Nutr. 3, 695–705. doi: [10.1080/01904168109362872](https://doi.org/10.1080/01904168109362872)

Brunetti, P., Zanella, L., De Paolis, A., Di Litta, D., Cecchetti, V., Falasca, G., et al. (2015). Cadmium-inducible expression of the ABC-type transporter AtABCC3 increases phytochelatin-mediated cadmium tolerance in Arabidopsis. J. Exp. Bot. 66, 3815–3829. doi: [10.1093/jxb/erv185](https://doi.org/10.1093/jxb/erv185)

Bursakov, S. A., Kroupin, P. Y., Karlov, G. I., and Divashuk, M. G. (2023). Tracing the element: the molecular bases of molybdenum homeostasis in legumes. Agronomy 13, 2300. doi: [10.3390/agronomy13092300](https://doi.org/10.3390/agronomy13092300)

Calderón-Vázquez, C., Sawers, R. J., and Herrera-Estrella, L. (2011). Phosphate deprivation in maize: genetics and genomics. Plant Physiol. 156, 1067–1077. doi: [10.1104/pp.111.174987](https://doi.org/10.1104/pp.111.174987)

Castrillo, G., Sánchez-Bermejo, E., de Lorenzo, L., Crevillén, P., Fraile-Escanciano, A., Tc, M., et al. (2013). WRKY6 transcription factor restricts arsenate uptake and transposon activation in Arabidopsis. Plant Cell 25, 2944–2957. doi: [10.1105/](https://doi.org/10.1105/tpc.113.114009) [tpc.113.114009](https://doi.org/10.1105/tpc.113.114009)

Chen, G., Li, J., Han, H., Du, R., and Wang, X. (2022). Physiological and molecular mechanisms of plant responses to copper stress. Int. J. Mol. Sci. 23, 12950. doi: [10.3390/](https://doi.org/10.3390/ijms232112950) iims232112950

Chen, J., Wang, J. W., and Shu, Y. H. (2020a). Review on the effects of heavy metal pollution on herbivorous insects. Ying Yong Sheng Tai Xue Bao 31, 1773–1782. doi: [10.13287/j.1001-9332.202005.035](https://doi.org/10.13287/j.1001-9332.202005.035)

Chao, L. M., Liu, Y. Q., Chen, D. Y., Xue, X. Y., Mao, Y. B., and Chen, X. Y. (2017). Arabidopsis transcription factors SPL1 and SPL12 confer plant thermotolerance at reproductive stage. Mol. Plant 10, 735–748. doi: [10.1016/j.molp.2017.03.010](https://doi.org/10.1016/j.molp.2017.03.010)

Chen, P., Chen, T., Li, Z., Jia, R., Luo, D., Tang, M., et al. (2020b). Transcriptome analysis revealed key genes and pathways related to cadmium-stress tolerance in Kenaf (Hibiscus cannabinus L.). Ind. Crop Prod. 158, 112970. doi: [10.1016/j.indcrop.2020.112970](https://doi.org/10.1016/j.indcrop.2020.112970)

Chen, Y., Wang, Y., Wu, W., Lin, Q., and Xue, S. (2006). Impacts of chelate-assisted phytoremediation on microbial community composition in the rhizosphere of a copper accumulator and non-accumulator. Sci. Total Environ. 356, 247–255. doi: [10.1016/](https://doi.org/10.1016/j.scitotenv.2005.04.028) [j.scitotenv.2005.04.028](https://doi.org/10.1016/j.scitotenv.2005.04.028)

Chen, Y. T., Wang, Y., and Yeh, K. C. (2017). Role of root exudates in metal acquisition and tolerance. Curr. Opin. Plant Biol. 39, 66–72. doi: [10.1016/j.pbi.2017.06.004](https://doi.org/10.1016/j.pbi.2017.06.004)

Chen, J., Yang, L., Gu, J., Bai, X., Ren, Y., Fan, T., et al. (2015). MAN 3 gene regulates cadmium tolerance through the glutathione-dependent pathway in Arabidopsis thaliana. New Phytol. 205, 570–582. doi: [10.1111/nph.13101](https://doi.org/10.1111/nph.13101)

Chiang, H. C., Lo, J. C., and Yeh, K. C. (2006). Genes associated with heavy metal tolerance and accumulation in Zn/Cd hyperaccumulator Arabidopsis halleri: a genomic survey with cDNA microarray. Environ. Sci. Technol. 40, 6792–6798. doi: [10.1021/](https://doi.org/10.1021/es061432y) [es061432y](https://doi.org/10.1021/es061432y)

Claus, J., Bohmann, A., and Chavarría-Krauser, A. (2013). Zinc uptake and radial transport in roots of Arabidopsis thaliana: a modelling approach to understand accumulation. Ann. Bot. 112, 369–380. doi: [10.1093/aob/mcs263](https://doi.org/10.1093/aob/mcs263)

Clemens, S. (2001). Molecular mechanisms of plant metal tolerance and homeostasis. Planta 212, 475–486. doi: [10.1007/s004250000458](https://doi.org/10.1007/s004250000458)

Cohen, C. K., Norvell, W. A., and Kochian, L. V. (1997). Induction of the root cell plasma membrane ferric reductase (an exclusive role for Fe and Cu). Plant Physiol. 114, 1061–1069. doi: [10.1104/pp.114.3.1061](https://doi.org/10.1104/pp.114.3.1061)

Collin, F. (2019). Chemical basis of reactive oxygen species reactivity and involvement in neurodegenerative diseases. Int. J. Mol. Sci. 20 (10), 2407. doi: [10.3390/ijms20102407](https://doi.org/10.3390/ijms20102407)

Collin, S., Baskar, A., Geevarghese, D. M., Ali, M. N. V. S., Bahubali, P., Choudhary, R., et al. (2022). Bioaccumulation of lead (Pb) and its effects in plants: a review. J. Hazard. Mater. Lett. 3, 100064. doi: [10.1016/j.hazl.2022.100064](https://doi.org/10.1016/j.hazl.2022.100064)

Costa, M. I., Sarmento-Ribeiro, A. B., and Gonçalves, A. C. (2023). Zinc: from biological functions to therapeutic potential. *Int. J. Mol. Sci.* 24 (5), 4822. doi: [10.3390/](https://doi.org/10.3390/ijms24054822) [ijms24054822](https://doi.org/10.3390/ijms24054822)

Dai, J., Becquer, T., Rouiller, J. H., Reversat, G., Bernhard-Reversat, F., and Lavelle, P. (2004). Influence of heavy metals on C and N mineralization and microbial biomass in Zn-, Pb-, Cu-, and Cd-contaminated soils. Appl. Soil Ecol. 25, 99–109. doi: [10.1016/](https://doi.org/10.1016/j.apsoil.2003.09.003) [j.apsoil.2003.09.003](https://doi.org/10.1016/j.apsoil.2003.09.003)

DalCorso, G., Farinati, S., Maistri, S., and Furini, A. (2008). How plants cope with cadmium: staking all on metabolism and gene expression. J. Integr. Plant Biol. 50, 1268–1280. doi: [10.1111/j.1744-7909.2008.00737.x](https://doi.org/10.1111/j.1744-7909.2008.00737.x)

Danouche, M., El Ghachtouli, N., and El Arroussi, H. (2021). Phycoremediation mechanisms of heavy metals using living green microalgae: physicochemical and molecular approaches for enhancing selectivity and removal capacity. Heliyon 7, e07609. doi: [10.1016/j.heliyon.2021.e07609](https://doi.org/10.1016/j.heliyon.2021.e07609)

Daspute, A. A., Sadhukhan, A., Tokizawa, M., Kobayashi, Y., Panda, S. K., and Koyama, H. (2017). Transcriptional regulation of aluminum-tolerance genes in higher plants: clarifying the underlying molecular mechanisms. Front. Plant Sci. 8, 1358. doi: [10.3389/fpls.2017.01358](https://doi.org/10.3389/fpls.2017.01358)

De Caroli, M., Furini, A., DalCorso, G., Rojas, M., and Di Sansebastiano, G. P. (2020). Endomembrane reorganization induced by heavy metals. Plants 9, 482. doi: [10.3390/](https://doi.org/10.3390/plants9040482) [plants9040482](https://doi.org/10.3390/plants9040482)

Delhaize, E., Gruber, B. D., Pittman, J. K., White, R. G., Leung, H., Miao, Y., et al. (2007). A role for the AtMTP11 gene of Arabidopsis in manganese transport and tolerance. Plant J. 51, 198–210. doi: [10.1111/j.1365-313X.2007.03138.x](https://doi.org/10.1111/j.1365-313X.2007.03138.x)

Desbrosses-Fonrouge, A. G., Voigt, K., Schröder, A., Arrivault, S., Thomine, S., and Krämer, U. (2005). Arabidopsis thaliana MTP1 is a Zn transporter in the vacuolar membrane which mediates Zn detoxification and drives leaf Zn accumulation. FEBS. Lett. 579, 4165–4174. doi: [10.1016/j.febslet.2005.06.046](https://doi.org/10.1016/j.febslet.2005.06.046)

Dixit, R., Wasiullah, X., Malaviya, D., Pandiyan, K., Singh, U. B., Sahu, A., et al. (2015). Bioremediation of heavy metals from soil and aquatic environment: an overview of principles and criteria of fundamental processes. Sustainability 7, 2189– 2212. doi: [10.3390/su7022189](https://doi.org/10.3390/su7022189)

Dräger, D. B., Desbrosses-Fonrouge, A. G., Krach, C., Chardonnens, A. N., Meyer, R. C., Saumitou-Laprade, P., et al. (2004). Two genes encoding Arabidopsis halleri MTP1 metal transport proteins co-segregate with zinc tolerance and account for high MTP1 transcript levels. Plant J. 39, 425–439. doi: [10.1111/j.1365-313X.2004.02143.x](https://doi.org/10.1111/j.1365-313X.2004.02143.x)

Drew, D., North, R. A., Nagarathinam, K., and Tanabe, M. (2021). Structures and general transport mechanisms by the major facilitator superfamily (MFS). Chem. Rev. 121, 5289–5335. doi: [10.1021/acs.chemrev.0c00983](https://doi.org/10.1021/acs.chemrev.0c00983)

Du, Z. Y., Chen, M. X., Chen, Q. F., Gu, J. D., and Chye, M. L. (2015). Expression of Arabidopsis acyl-CoA-binding proteins AtACBP 1 and AtACBP 4 confers P b (II) accumulation in Brassica juncea roots. Plant Cell Environ. 38, 101–117. doi: [10.1111/](https://doi.org/10.1111/pce.12382) [pce.12382](https://doi.org/10.1111/pce.12382)

Dumanović, J., Nepovimova, E., Natić, M., Kuča, K., and Jaćević, V. (2020). The significance of reactive oxygen species and antioxidant defense system in plants: a concise overview. Front Plant Sci. 11, 552969. doi: [10.3389/fpls.2020.552969](https://doi.org/10.3389/fpls.2020.552969)

Dutta, S., Mitra, M., Agarwal, P., Mahapatra, K., De, S., Sett, U., et al. (2018). Oxidative and genotoxic damages in plants in response to heavy metal stress and maintenance of genome stability. Plant Signal. Behav. 13, e1460048. doi: [10.1080/](https://doi.org/10.1080/15592324.2018.1460048) [15592324.2018.1460048](https://doi.org/10.1080/15592324.2018.1460048)

El-Sappah, A. H., Abbas, M., Rather, S. A., Wani, S. H., Soaud, N., Noor, Z., et al. (2023). Genome-wide identification and expression analysis of metal tolerance protein (MTP) gene family in soybean (Glycine max) under heavy metal stress. Mol. Biol. Rep. 502975, 2990doi: [10.1007/s11033-022-08100-x](https://doi.org/10.1007/s11033-022-08100-x)

El-Sappah, A. H., Elbaiomy, R. G., Elrys, A. S., Wang, Y., Zhu, Y., Huang, Q., et al. (2021a). Genome-wide identification and expression analysis of metal tolerance protein gene family in Medicago truncatula under a broad range of heavy metal stress. Front. Genet. 12, 713224. doi: [10.3389/fgene.2021.713224](https://doi.org/10.3389/fgene.2021.713224)

El-Sappah, A. H., Elrys, A. S., Desoky, E. M., Zhao, X., Bingwen, W., El-Sappah, H. H., et al. (2021b). Comprehensive genome wide identification and expression analysis of MTP gene family in tomato (Solanum lycopersicum) under multiple heavy metal stress. Saudi J. Biol. Sci. 28, 6946–6956. doi: [10.1016/j.sjbs.2021.07.073](https://doi.org/10.1016/j.sjbs.2021.07.073)

El-Sappah, A. H., and Rather, S. A. (2022). "Genomics approaches to study abiotic stress tolerance in plants," in Plant abiotic stress physiology. Eds. T. Aftab and K. R. Hakeem (CRC Press, Boca Raton, FL, USA), 25–46. doi: [10.1201/9781003180579](https://doi.org/10.1201/9781003180579)

El-Sappah, A. H., Shawky, A. S. H., Sayed-Ahmad, M. S., and Youssef, M. A. H. (2017). Estimation of heat shock protein 70 (hsp 70) gene expression in Nile tilapia (Oreochromis niloticus) using quantitative Real-Time PCR. Zagazig J.Agric. Res. 44, 1003–1015. doi: [10.21608/zjar.2017.52300](https://doi.org/10.21608/zjar.2017.52300)

Emamverdian, A., Ding, Y., Mokhberdoran, F., and Xie, Y. (2015). Heavy metal stress and some mechanisms of plant defense response. Sci. World J. 2015, 756120. doi: [10.1155/2015/756120](https://doi.org/10.1155/2015/756120)

Ercal, N., Gurer-Orhan, H., and Aykin-Burns, N. (2001). Toxic metals and oxidative stress part I: mechanisms involved in metal-induced oxidative damage. Curr. Top. Med. Chem. 1, 529–539. doi: [10.2174/1568026013394831](https://doi.org/10.2174/1568026013394831)

Ezaki, B., Gardner, R. C., Ezaki, Y., and Matsumoto, H. (2000). Expression of aluminum-induced genes in transgenic Arabidopsis plants can ameliorate aluminum stress and/or oxidative stress. Plant Physiol. 122, 657–666. doi: [10.1104/pp.122.3.657](https://doi.org/10.1104/pp.122.3.657)

Faizan, M., Alam, P., Hussain, A., Karabulut, F., Tonny, S. H., Cheng, S. H., et al. (2024). Phytochelatins: key regulator against heavy metal toxicity in plants. Plant Stress 11, 100355. doi: [10.1016/j.stress.2024.100355](https://doi.org/10.1016/j.stress.2024.100355)

Fan, X., Zhou, X., Chen, H., Tang, M., and Xie, X. (2021). Cross-talks between macro- and micronutrient uptake and signaling in plants. Front. Plant Sci. 12, 663477. doi: [10.3389/fpls.2021.663477](https://doi.org/10.3389/fpls.2021.663477)

Feng Ma, J., Hiradate, S., and Matsumoto, H. (1998). High aluminum resistance in buckwheat. Ii. Oxalic acid detoxifies aluminum internally. Plant Physiol. 117, 753–759. doi: [10.1104/pp.117.3.753](https://doi.org/10.1104/pp.117.3.753)

Festa, R. A., and Thiele, D. J. (2011). Copper: an essential metal in biology. Curr. Biol. 21, R877–R883. doi: [10.1016/j.cub.2011.09.040](https://doi.org/10.1016/j.cub.2011.09.040)

Foyer, C. H., Rasool, B., Davey, J. W., and Hancock, R. D. (2016). Cross-tolerance to biotic and abiotic stresses in plants: a focus on resistance to aphid infestation. J. Exp. Bot. 67, 2025–2037. doi: [10.1093/jxb/erw079](https://doi.org/10.1093/jxb/erw079)

Freisinger, E. (2011). Structural features specific to plant metallothioneins. J. Biol. Inorg. Chem. 16, 1035–1045. doi: [10.1007/s00775-011-0801-z](https://doi.org/10.1007/s00775-011-0801-z)

Gallo-Franco, J. J., Sosa, C. C., Ghneim-Herrera, T., and Quimbaya, M. (2020). Epigenetic control of plant response to heavy metal stress: a new view on aluminum tolerance. Front. Plant Sci. 11, 602625. doi: [10.3389/fpls.2020.602625](https://doi.org/10.3389/fpls.2020.602625)

García, A., Baquedano, F. J., Navarro, P., and Castillo, F. J. (1999). Oxidative stress induced by copper in sunflower plants. Free Radic. Res. 31, S45–S50. doi: [10.1080/](https://doi.org/10.1080/10715769900301311) [10715769900301311](https://doi.org/10.1080/10715769900301311)

Garcia-Molina, A., Andrés-Colás, N., Perea-García, A., Del Valle-Tascón, S., Peñarrubia, L., and Puig, S. (2011). The intracellular Arabidopsis COPT5 transport protein is required for photosynthetic electron transport under severe copper
deficiency. *Plant J.* 65, 848–860. doi: [10.1111/j.1365-313X.2010.04472.x](https://doi.org/10.1111/j.1365-313X.2010.04472.x)

Garcia-Molina, A., Andrés-Colás, N., Perea-García, A., Neumann, U., Dodani, S. C., Huijser, P., et al. (2013). The Arabidopsis COPT6 transport protein functions in copper distribution under copper-deficient conditions. Plant Cell Physiol. 54, 1378–1390. doi: [10.1093/pcp/pct088](https://doi.org/10.1093/pcp/pct088)

Gasic, K., and Korban, S. S. (2006). "Heavy metal stress," in Physiology and molecular biology of stress tolerance in plants, vol. pp . Eds. M. Rao, K. Raghavendra, A. Janardhan and K. Reddy (Springer, Dordrecht. Netherlands), 219–254. doi: [10.1007/1-4020-4225-](https://doi.org/10.1007/1-4020-4225-6_8219-254) [6_8219-254](https://doi.org/10.1007/1-4020-4225-6_8219-254)

Genchi, G., Sinicropi, M. S., Lauria, G., Carocci, A., and Catalano, A. (2020). The effects of cadmium toxicity. Int. J. Environ. Res. Public Health 17 (11), 3782. doi: [10.3390/ijerph17113782](https://doi.org/10.3390/ijerph17113782)

Gill, R. A., Ahmar, S., Ali, B., Saleem, M. H., Khan, M. U., Zhou, W., et al. (2021). The role of membrane transporters in plant growth and development, and abiotic stress tolerance. Int. J. Mol. Sci. 22, 12792. doi: [10.3390/ijms222312792](https://doi.org/10.3390/ijms222312792)

Gill, S. S., Khan, N. A., and Tuteja, N. (2012). Cadmium at high dose perturbs growth, photosynthesis and nitrogen metabolism while at low dose it up regulates sulfur assimilation and antioxidant machinery in garden cress (Lepidium sativum L.). Plant Sci. 182, 112–120. doi: [10.1016/j.plantsci.2011.04.018](https://doi.org/10.1016/j.plantsci.2011.04.018)

Giri, A. K., and Patel, R. K. (2011). Toxicity and bioaccumulation potential of Cr (VI) and Hg (II) on differential concentration by Eichhornia crassipes in hydroponic culture. Water Sci. Technol. 63, 899–907. doi: [10.2166/wst.2011.268](https://doi.org/10.2166/wst.2011.268)

Goyer, R., Golub, M., Choudhury, H., Hughes, M., Kenyon, E., and Stifelman, M. (2004). "Issue paper on the human health effects of metals," in US environmental protection agency risk assessment forum (ERG, Lexington, KY, USA), 1200.

Grotz, N., Fox, T., Connolly, E., Park, W., Guerinot, M. L., and Eide, D. (1998). Identification of a family of zinc transporter genes from Arabidopsis that respond to zinc deficiency. Proc. Natl. Acad. Sci. U.S.A. 95, 7220–7224. doi: [10.1073/](https://doi.org/10.1073/pnas.95.12.7220) [pnas.95.12.7220](https://doi.org/10.1073/pnas.95.12.7220)

Guerinot, M. L. (2000). The ZIP family of metal transporters. Biochim. Biophys. Acta 1465, 190–198. doi: [10.1016/S0005-2736\(00\)00138-3](https://doi.org/10.1016/S0005-2736(00)00138-3)

Gulcin, İ., and Alwasel, S. (2022). Metal ions, metal chelators and metal chelating assay as antioxidant method. Processes 10, 132. doi: [10.3390/pr10010132](https://doi.org/10.3390/pr10010132)

Guo, W. J., Bundithya, W., and Goldsbrough, P. B. (2003). Characterization of the Arabidopsis metallothionein gene family: tissue-specific expression and induction during senescence and in response to copper. New Phytol. 159, 369–381. doi: [10.1046/j.1469-8137.2003.00813.x](https://doi.org/10.1046/j.1469-8137.2003.00813.x)

Hajam, Y. A., Kumar, R., and Kumar, A. (2023). Environmental waste management strategies and vermi transformation for sustainable development. Environ. Challen. 13, 100747. doi: [10.1016/j.envc.2023.100747](https://doi.org/10.1016/j.envc.2023.100747)

Hall, J. L. (2002). Cellular mechanisms for heavy metal detoxification and tolerance. J. Exp. Bot. 53, 1–11. doi: [10.1093/jexbot/53.366.1](https://doi.org/10.1093/jexbot/53.366.1)

Hama Aziz, K. H., Mustafa, F. S., Omer, K. M., Hama, S., Hamarawf, R. F., and Rahman, K. O. (2023). Heavy metal pollution in the aquatic environment: efficient and low-cost removal approaches to eliminate their toxicity: a review. RSC. Adv. 13, 17595– 17610. doi: [10.1039/D3RA00723E](https://doi.org/10.1039/D3RA00723E)

Hamsa, N., Yogesh, G. S., Koushik, U., and Patil, L. (2017). Nitrogen transformation in soil: effect of heavy metals. Int. J. Curr. Microbiol. Appl. Sci. 6, 816–832. doi: [10.20546/ijcmas.2017.605.092](https://doi.org/10.20546/ijcmas.2017.605.092)

Hamzah Saleem, M., Usman, K., Rizwan, M., Al Jabri, H., and Alsafran, M. (2022). Functions and strategies for enhancing zinc availability in plants for sustainable agriculture. Front. Plant Sci. 13, 1033092. doi: [10.3389/fpls.2022.1033092](https://doi.org/10.3389/fpls.2022.1033092)

Hasan, M. K., Cheng, Y., Kanwar, M. K., Chu, X. Y., Ahammed, G. J., and Qi, Z. Y. (2017). Responses of plant proteins to heavy metal stress-a review. Front.Plant Sci. 8, 1492. doi: [10.3389/fpls.2017.01492](https://doi.org/10.3389/fpls.2017.01492)

Hasanuzzaman, M., Alhaithloul, H. A. S., Parvin, K., Bhuyan, M. B., Tanveer, M., Mohsin, S. M., et al. (2019). Polyamine action under metal/metalloid stress: regulation of biosynthesis, metabolism, and molecular interactions. Int. J. Mol. Sci. 20 (13), 3215. doi: [10.3390/ijms20133215](https://doi.org/10.3390/ijms20133215)

Hasanuzzaman, M., Bhuyan, M. B., Zulfiqar, F., Raza, A., Mohsin, S. M., Mahmud, J. A., et al. (2020). Reactive oxygen species and antioxidant defense in plants under abiotic stress: revisiting the crucial role of a universal defense regulator. Antioxidants 9, 681. doi: [10.3390/antiox9080681](https://doi.org/10.3390/antiox9080681)

Hasanuzzaman, M., Nahar, K., Alam, M. M., Roychowdhury, R., and Fujita, M. (2013). Physiological, biochemical, and molecular mechanisms of heat stress tolerance in plants. Int. J. Mol. Sci. 14, 9643–9684. doi: [10.3390/ijms14059643](https://doi.org/10.3390/ijms14059643)

Hashimoto, H., Uragami, C., and Cogdell, R. J. (2016). Carotenoids and photosynthesis. Subcell. Biochem. 79, 111–139. doi: [10.1007/978-3-319-39126-7_4](https://doi.org/10.1007/978-3-319-39126-7_4)

Hayat, S., Hayat, Q., AlYemeni, M. N., Wani, A. S., Pichtel, J., and Ahmad, A. (2012). Role of proline under changing environments: a review. Plant Signal. Behav. 7, 1456– 1466. doi: [10.4161/psb.21949](https://doi.org/10.4161/psb.21949)

Haydon, M. J., and Cobbett, C. S. (2007). A novel major facilitator superfamily protein at the tonoplast influences zinc tolerance and accumulation in Arabidopsis. Plant Physiol. 143, 1705–1719. doi: [10.1104/pp.106.092015](https://doi.org/10.1104/pp.106.092015)

Hinsinger, P., Plassard, C., Tang, C., and Jaillard, B. (2003). Origins of root-mediated pH changes in the rhizosphere and their responses to environmental constraints: a review. Plant Soil. 248, 43–59. doi: [10.1023/A:1022371130939](https://doi.org/10.1023/A:1022371130939)

Hossain, M. A., Piyatida, P., da Silva, J. A. T., and Fujita, M. (2012). Molecular mechanism of heavy metal toxicity and tolerance in plants: central role of glutathione in detoxification of reactive oxygen species and methylglyoxal and in heavy metal chelation. J. Bot. 2012, 872875. doi: [10.1155/2012/872875](https://doi.org/10.1155/2012/872875)

Hu, S., Yu, Y., Chen, Q., Mu, G., Shen, Z., and Zheng, L. (2017). OsMYB45 plays an important role in rice resistance to cadmium stress. Plant Sci. 264, 1–8. doi: [10.1016/](https://doi.org/10.1016/j.plantsci.2017.08.002) [j.plantsci.2017.08.002](https://doi.org/10.1016/j.plantsci.2017.08.002)

Huang, Y., Adeleye, A. S., Zhao, L., Minakova, A. S., Anumol, T., and Keller, A. A. (2019). Antioxidant response of cucumber (Cucumis sativus) exposed to nano copper pesticide: quantitative determination via LC-MS/MS. Food Chem. 270, 47–52. doi: [10.1016/j.foodchem.2018.07.069](https://doi.org/10.1016/j.foodchem.2018.07.069)

Huang, D., Huo, J., and Liao, W. (2021). Hydrogen sulfide: roles in plant abiotic stress response and crosstalk with other signals. Plant Sci. 302, 110733. doi: [10.1016/](https://doi.org/10.1016/j.plantsci.2020.110733) [j.plantsci.2020.110733](https://doi.org/10.1016/j.plantsci.2020.110733)

Igiri, B. E., Okoduwa, S. I. R., Idoko, G. O., Akabuogu, E. P., Adeyi, A. O., and Ejiogu, I. K. (2018). Toxicity and bioremediation of heavy metals contaminated ecosystem from tannery wastewater: a review. J. Toxicol. 2018, 2568038. doi: [10.1155/2018/](https://doi.org/10.1155/2018/2568038) [2568038](https://doi.org/10.1155/2018/2568038)

Islam, M. A., Guo, J., Peng, H., Tian, S., Bai, X., Zhu, H., et al. (2020). TaYS1A, a yellow stripe-like transporter gene, is required for wheat resistance to Puccinia striiformis f.sp. Tritici. Genes 11 (12), 1452. doi: [10.3390/genes11121452](https://doi.org/10.3390/genes11121452)

Jadia, C. D., and Fulekar, M. H. (2009). Phytoremediation of heavy metals: recent techniques. Afr. J. Biotechnol. 8, 921–928.

Jaishankar, M., Tseten, T., Anbalagan, N., Mathew, B. B., and Beeregowda, K. N. (2014). Toxicity, mechanism and health effects of some heavy metals. Interdiscip. Toxicol. 7, 60–72. doi: [10.2478/intox-2014-0009](https://doi.org/10.2478/intox-2014-0009)

Jiang, X., and Li, M. (2020). "Chapter 5 - Ecological safety hazards of wastewater," in High-risk pollutants in wastewater. Eds. H. Ren and X. Zhang. (Elsevier, Amsterdam, Netherlands), 101–123.

Jogawat, A., Chhaya, Y. B., and Narayan, O. P. (2021). Metal transporters in organelles and their roles in heavy metal transportation and sequestration mechanisms in plants. Physiol. Plant 173, 259–275. doi: [10.1111/ppl.13370](https://doi.org/10.1111/ppl.13370)

Jomova, K., Raptova, R., Alomar, S. Y., Alwasel, S. H., Nepovimova, E., Kuca, K., et al. (2023). Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. Arch. Toxicol. 97, 2499–2574. doi: [10.1007/s00204-023-03562-9](https://doi.org/10.1007/s00204-023-03562-9)

Kaczmarek, M., Cachau, R. E., Topol, I. A., Kasprzak, K. S., Ghio, A., and Salnikow, K. (2009). Metal ions-stimulated iron oxidation in hydroxylases facilitates stabilization of HIF-1 alpha protein. Toxicol. Sci. 107, 394–403. doi: [10.1093/toxsci/kfn251](https://doi.org/10.1093/toxsci/kfn251)

Kafle, A., Timilsina, A., Gautam, A., Adhikari, K., Bhattarai, A., and Aryal, N. (2022). Phytoremediation: mechanisms, plant selection and enhancement by natural and synthetic agents. Environ. Adv. 8, 100203. doi: [10.1016/j.envadv.2022.100203](https://doi.org/10.1016/j.envadv.2022.100203)

Kajla, M., Roy, A., Singh, I. K., and Singh, A. (2023). Regulation of the regulators: transcription factors controlling biosynthesis of plant secondary metabolites during biotic stresses and their regulation by miRNAs. Front. Plant Sci. 14, 1126567. doi: [10.3389/fpls.2023.1126567](https://doi.org/10.3389/fpls.2023.1126567)

Khan, F., Siddique, A. B., Shabala, S., Zhou, M., and Zhao, C. (2023). Phosphorus plays key roles in regulating plants' physiological responses to abiotic stresses. Plants 12 (15), 2861. doi: [10.3390/plants12152861](https://doi.org/10.3390/plants12152861)

Klatte, M., Schuler, M., Wirtz, M., Fink-Straube, C., Hell, R., and Bauer, P. (2009). The analysis of Arabidopsis nicotianamine synthase mutants reveals functions for nicotianamine in seed iron loading and iron deficiency responses. Plant Physiol. 150, 257–271. doi: [10.1104/pp.109.136374](https://doi.org/10.1104/pp.109.136374)

Kobae, Y., Uemura, T., Sato, M. H., Ohnishi, M., Mimura, T., Nakagawa, T., et al. (2004). Zinc transporter of Arabidopsis thaliana AtMTP1 is localized to vacuolar membranes and implicated in zinc homeostasis. Plant Cell Physiol. 45, 1749–1758. doi: [10.1093/pcp/pci015](https://doi.org/10.1093/pcp/pci015)

Kolaj-Robin, O., Russell, D., Hayes, K. A., Pembroke, J. T., and Soulimane, T. (2015). Cation diffusion facilitator family: structure and function. FEBS. Letters. 589, 1283– 1295. doi: [10.1016/j.febslet.2015.04.007](https://doi.org/10.1016/j.febslet.2015.04.007)

Koren'Kov, V., Park, S., Cheng, N. H., Sreevidya, C., Lachmansingh, J., Morris, J., et al. (2007). Enhanced Cd^{2+} -selective root-tonoplast-transport in tobaccos expressing Arabidopsis cation exchangers. Planta 225, 403–411. doi: [10.1007/s00425-006-0352-7](https://doi.org/10.1007/s00425-006-0352-7)

Kornarzyński, K., Sujak, A., Czernel, G., and Wiacek, D. (2020). Effect of Fe₃O₄ nanoparticles on germination of seeds and concentration of elements in Helianthus annuus L. under constant magnetic field. Sci. Rep. 10, 8068. doi: [10.1038/s41598-020-](https://doi.org/10.1038/s41598-020-64849-w) [64849-w](https://doi.org/10.1038/s41598-020-64849-w)

Kostenkova, K., Scalese, G., Gambino, D., and Crans, D. C. (2022). Highlighting the roles of transition metals and speciation in chemical biology. Curr. Opin. Chem. Biol. 69, 102155. doi: [10.1016/j.cbpa.2022.102155](https://doi.org/10.1016/j.cbpa.2022.102155)

Kraemer, U. (2009). The dilemma of controlling heavy metal accumulation in plants. New Phytol. 181, 3–5. doi: [10.1111/j.1469-8137.2008.02699.x](https://doi.org/10.1111/j.1469-8137.2008.02699.x)

Kranner, I., and Colville, L. (2011). Metals and seeds: biochemical and molecular implications and their significance for seed germination. Environ. Exp. Bot. 72, 93–105. doi: [10.1016/j.envexpbot.2010.05.005](https://doi.org/10.1016/j.envexpbot.2010.05.005)

Kudo, H., Qian, Z., Inoue, C., and Chien, M. F. (2023). Temperature dependence of metals accumulation and removal kinetics by Arabidopsis halleri ssp. gemmifera. Plants 12, 877. doi: [10.3390/plants12040877](https://doi.org/10.3390/plants12040877)

Kumar, V., Roy, S., Behera, B. K., and Das, B. K. (2022). Heat shock proteins (Hsps) in cellular homeostasis: a promising tool for health management in crustacean aquaculture. Life 12 (11), 1777. doi: [10.3390/life12111777](https://doi.org/10.3390/life12111777)

Kumar, V., Singh, J., and Kumar, P. (2019). "Heavy metals accumulation in crop plants: Sources, response mechanisms, stress tolerance and their effects, Contaminants in agriculture and environment: health risks and remediation. Eds. V. Kumar, R. Kumar, J. Singh and P. Kumar (Agro Environ Media, Haridwar, India), 38– 57. doi: [10.26832/AESA-2019-CAE-0161-04](https://doi.org/10.26832/AESA-2019-CAE-0161-04)

Li, J., Abbas, M., Desoky, E. S. M., Zafar, S., Soaud, S. A., Hussain, S. S., et al. (2023). Analysis of metal tolerance protein (MTP) family in sunflower (Helianthus annus L.)
and role of HaMTP10 as cadmium antiporter under moringa seed extract. Ind. Crops Prod. 202, 117023. doi: [10.1016/j.indcrop.2023.117023](https://doi.org/10.1016/j.indcrop.2023.117023)

Li, Z., Cao, Z., Ma, X., Cao, D., Zhao, K., Zhao, K., et al. (2024). Natural resistanceassociated macrophage proteins are involved in tolerance to heavy metal $\rm Cd2^{+}$ toxicity and resistance to bacterial wilt of peanut (Arachis hypogaea L.). Plant Physiol. Biochem. 207, 108411. doi: [10.1016/j.plaphy.2024.108411](https://doi.org/10.1016/j.plaphy.2024.108411)

Li, S., Zhou, X., Li, H., Liu, Y., Zhu, L., Guo, J., et al. (2015). Overexpression of ZmIRT1 and ZmZIP3 enhances iron and zinc accumulation in transgenic Arabidopsis. PLOS One 10, e0136647. doi: [10.1371/journal.pone.0136647](https://doi.org/10.1371/journal.pone.0136647)

Lin, Y., Gritsenko, D., Feng, S., Teh, Y. C., Lu, X., and Xu, J. (2016). Detection of heavy metal by paper-based microfluidics. Biosens. Bioelectron. 83, 256–266. doi: [10.1016/j.bios.2016.04.061](https://doi.org/10.1016/j.bios.2016.04.061)

Lin, Y. F., Severing, E. I., Te Lintel Hekkert, B., Schijlen, E., and Aarts, M. G. (2014). A comprehensive set of transcript sequences of the heavy metal hyperaccumulator Noccaea caerulescens. Front. Plant Sci. 5, 261. doi: [10.3389/fpls.2014.00261](https://doi.org/10.3389/fpls.2014.00261)

Liu, L., An, M. M., Li, X. J., Han, Z., Li, S. X., and Li, B. (2022). Molybdenum-induced effects on nitrogen absorption and utilization under different nitrogen sources in Vitis vinifera. J. Plant Interact. 17, 756–765. doi: [10.1080/17429145.2022.2089752](https://doi.org/10.1080/17429145.2022.2089752)

Liu, H., Guo, H., Jian, Z., Cui, H., Fang, J., Zuo, Z., et al. (2020). Copper induces oxidative stress and apoptosis in the mouse liver. Oxid. Med. Cell. Longev. 2020, 1359164. doi: [10.1155/2020/1359164](https://doi.org/10.1155/2020/1359164)

Lombi, E., Zhao, F. J., Wieshammer, G., Zhang, G., and McGrath, S. P. (2002). In situ fixation of metals in soils using bauxite residue: biological effects. Environ. Pollut. 118, 445–452. doi: [10.1016/S0269-7491\(01\)00295-0](https://doi.org/10.1016/S0269-7491(01)00295-0)

López-Vargas, E. R., Ortega-Ortíz, H., Cadenas-Pliego, G., de Alba Romenus, K., Cabrera de la Fuente, M., Benavides-Mendoza, A., et al. (2018). Foliar application of copper nanoparticles increases the fruit quality and the content of bioactive compounds in tomatoes. Appl. Sci. 8 (7), 1020. doi: [10.3390/app8071020](https://doi.org/10.3390/app8071020)

Lu, S. C. (2013). Glutathione synthesis. Biochim. Biophys. Acta 1830, 3143–3153. doi: [10.1016/j.bbagen.2012.09.008](https://doi.org/10.1016/j.bbagen.2012.09.008)

Mansoor, S., Ali, A., Kour, N., Bornhorst, J., AlHarbi, K., Rinklebe, J., et al. (2023). Heavy metal induced oxidative stress mitigation and ROS scavenging in plants. Plants 12 (16), 3003. doi: [10.3390/plants12163003](https://doi.org/10.3390/plants12163003)

Mashabela, M. D., Masamba, P., and Kappo, A. P. (2023). Applications of metabolomics for the elucidation of abiotic stress tolerance in plants: a special focus on osmotic stress and heavy metal toxicity. Plants 12, 269. doi: [10.3390/plants12020269](https://doi.org/10.3390/plants12020269)

Masion, A., and Bertsch, P. M. (1997). Aluminum speciation in the presence of wheat root cell walls: a wet chemical study. Plant Cell Environ. 20, 504–512. doi: [10.1046/](https://doi.org/10.1046/j.1365-3040.1997.d01-86.x) [j.1365-3040.1997.d01-86.x](https://doi.org/10.1046/j.1365-3040.1997.d01-86.x)

Mattson, M. P. (2008). Hormesis defined. Ageing Res. Rev. 7, 1–7. doi: [10.1016/](https://doi.org/10.1016/j.arr.2007.08.007) [j.arr.2007.08.007](https://doi.org/10.1016/j.arr.2007.08.007)

Mei, H., Cheng, N. H., Zhao, J., Park, S., Escareno, R. A., Pittman, J. K., et al. (2009). Root development under metal stress in Arabidopsis thaliana requires the H⁺/cation antiporter CAX4. New Phytol. 183, 95–105. doi: [10.1111/j.1469-8137.2009.02831.x](https://doi.org/10.1111/j.1469-8137.2009.02831.x)

Mendes, N. A. C., Cunha, M. L. O., Bosse, M. A., Silva, V. M., Moro, A. L., Agathokleous, E., et al. (2023). Physiological and biochemical role of nickel in
nodulation and biological nitrogen fixation in *Vigna unguiculata* L. Walp. *Plant* Physiol. Biochem. 201, 107869. doi: [10.1016/j.plaphy.2023.107869](https://doi.org/10.1016/j.plaphy.2023.107869)

Menguer, P. K., Farthing, E., Peaston, K. A., Ricachenevsky, F. K., Fett, J. P., and Williams, L. E. (2013). Functional analysis of the rice vacuolar zinc transporter OsMTP1. J. Exp. Bot. 64, 2871–2883. doi: [10.1093/jxb/ert136](https://doi.org/10.1093/jxb/ert136)

Mishra, S., Roychowdhury, R., Ray, S., Hada, A., Kumar, A., Sarker, U., et al. (2024). Salicylic acid (SA)-mediated plant immunity against biotic stresses: an insight on molecular components and signaling mechanism. Plant Stress 11, 100427. doi: [10.1016/](https://doi.org/10.1016/j.stress.2024.100427) [j.stress.2024.100427](https://doi.org/10.1016/j.stress.2024.100427)

Mishra, J., Singh, R., and Arora, N. K. (2017). Alleviation of heavy metal stress in plants and remediation of soil by rhizosphere microorganisms. Front. Microbiol. 8, 1706. doi: [10.3389/fmicb.2017.01706](https://doi.org/10.3389/fmicb.2017.01706)

Mitra, S., Chakraborty, A. J., Tareq, A. M., Emran, T. B., Nainu, F., Khusro, A., et al. (2022). Impact of heavy metals on the environment and human health: Novel therapeutic insights to counter the toxicity. J. King Saud. Univ. Sci. 34, 101865. doi: [10.1016/j.jksus.2022.101865](https://doi.org/10.1016/j.jksus.2022.101865)

Mousavi, S. R., Shahsavari, M., and Rezaei, M. (2011). A general overview on Manganese (Mn) importance for crops production. Aust. J. Basic Appl. Sci. 5, 1799– 1803.

Muneer, S., Jeong, B. R., Kim, T. H., Lee, J. H., and Soundararajan, P. (2014). Transcriptional and physiological changes in relation to Fe uptake under conditions of Fe-deficiency and Cd-toxicity in roots of Vigna radiata L. J. Plant Res. 127, 731–742. doi: [10.1007/s10265-014-0660-0](https://doi.org/10.1007/s10265-014-0660-0)

Najeeb, U., Jilani, G., Ali, S., Sarwar, M., Xu, L., and Zhou, W. (2011). Insights into cadmium induced physiological and ultra-structural disorders in Juncus effusus L. and its remediation through exogenous citric acid. J. Hazard. Mater. 186, 565–574. doi: [10.1016/j.jhazmat.2010.11.037](https://doi.org/10.1016/j.jhazmat.2010.11.037)

Nakanishi, H., Ogawa, I., Ishimaru, Y., Mori, S., and Nishizawa, N. (2006). Iron deficiency enhances cadmium uptake and translocation mediated by the Fe
transporters OsIRT1 and OSIRT2 in rice. Soil Sci. Plant Nutr. 52, 464-469. doi: [10.1111/j.1747-0765.2006.00055.x](https://doi.org/10.1111/j.1747-0765.2006.00055.x)

Niekerk, L. A., Gokul, A., Basson, G., Badiwe, M., Nkomo, M., Klein, A., et al. (2024). Heavy metal stress and mitogen activated kinase transcription factors in plants: Exploring heavy metal-ROS influences on plant signaling pathways. Plant Cell Environ. 47 (8), 2793–2810. doi: [10.1111/pce.14926](https://doi.org/10.1111/pce.14926)

Ning, X., Lin, M., Huang, G., Mao, J., Gao, Z., and Wang, X. (2023). Research progress on iron absorption, transport, and molecular regulation strategy in plants. Front. Plant Sci. 14, 1190768. doi: [10.3389/fpls.2023.1190768](https://doi.org/10.3389/fpls.2023.1190768)

Nouet, C., Motte, P., and Hanikenne, M. (2011). Chloroplastic and mitochondrial metal homeostasis. Trends. Plant Sci. 16, 395–404. doi: [10.1016/j.tplants.2011.03.005](https://doi.org/10.1016/j.tplants.2011.03.005)

Oda, K., Otani, M., Uraguchi, S., Akihiro, T., and Fujiwara, T. (2011). Rice ABCG43 is Cd inducible and confers Cd tolerance on yeast. Biosci. Biotechnol. Biochem. 75, 1211–1213. doi: [10.1271/bbb.110193](https://doi.org/10.1271/bbb.110193)

Olaniran, A. O., Balgobind, A., and Pillay, B. (2013). Bioavailability of heavy metals in soil: impact on microbial biodegradation of organic compounds and possible improvement strategies. Int. J. Mol. Sci. 14, 10197–10228. doi: [10.3390/ijms140510197](https://doi.org/10.3390/ijms140510197)

Osman, H. E., and Fadhlallah, R. S. (2023). Impact of lead on seed germination, seedling growth, chemical composition, and forage quality of different varieties of Sorghum. J. Umm Al-Qura Univ. Appl. Sci. 9, 77–86. doi: [10.1007/s43994-022-00022-5](https://doi.org/10.1007/s43994-022-00022-5)

Pacheco, D. D. R., Santana, B. C. G., Pirovani, C. P., and de Almeida, A. F. (2023). Zinc/iron-regulated transporter-like protein gene family in Theobroma cacao L: Characteristics, evolution, function and 3D structure analysis. Front. Plant Sci. 14, 1098401. doi: [10.3389/fpls.2023.1098401](https://doi.org/10.3389/fpls.2023.1098401)

Pan, D., Yi, J., Li, F., Li, X., Liu, C., Wu, W., et al. (2020a). Dynamics of gene expression associated with arsenic uptake and transport in rice during the whole growth period. BMC Plant Biol. 20, 133. doi: [10.1186/s12870-020-02343-1](https://doi.org/10.1186/s12870-020-02343-1)

Pan, G., Zhao, L., Li, J., Huang, S., Tang, H., Chang, L., et al. (2020b). Physiological responses and tolerance of flax (Linum usitatissimum L.) to lead stress. Acta Physiol. Plant. 42, 1–9. doi: [10.1007/s11738-020-03103-2](https://doi.org/10.1007/s11738-020-03103-2)

Pande, V., Pandey, S. C., Sati, D., Bhatt, P., and Samant, M. (2022). Microbial interventions in bioremediation of heavy metal contaminants in agroecosystem. Front. Microbiol. 13, 824084. doi: [10.3389/fmicb.2022.824084](https://doi.org/10.3389/fmicb.2022.824084)

Pandolfini, T., Gabbrielli, R., and Comparini, C. (2006). Nickel toxicity and peroxidase activity in seedlings of Triticum aestivum. Plant Cell Environ. 15, 719-725. doi: [10.1111/j.1365-3040.1992.tb01014.x](https://doi.org/10.1111/j.1365-3040.1992.tb01014.x)

Park, J., Song, W. Y., Ko, D., Eom, Y., Hansen, T. H., Schiller, M., et al. (2012). The phytochelatin transporters AtABCC1 and AtABCC2 mediate tolerance to cadmium and mercury. Plant J. 69, 278–288. doi: [10.1111/j.1365-313X.2011.04789.x](https://doi.org/10.1111/j.1365-313X.2011.04789.x)

Pasricha, S., Mathur, V., Garg, A., Lenka, S., Verma, K., and Agarwal, S. (2021). Molecular mechanisms underlying heavy metal uptake, translocation and tolerance in hyperaccumulators-an analysis: Heavy metal tolerance in hyperaccumulators. Environ. Challenges 4, 100197. doi: [10.1016/j.envc.2021.100197](https://doi.org/10.1016/j.envc.2021.100197)

Paz-Ferreiro, J., and Fu, S. (2016). Biological indices for soil quality evaluation: perspectives and limitations. Land Degrad. Dev. 27, 14–25. doi: [10.1002/ldr.2262](https://doi.org/10.1002/ldr.2262)

Peiter, E., Montanini, B., Gobert, A., Pedas, P., Husted, S., Maathuis, F. J., et al. (2007). A secretory pathway-localized cation diffusion facilitator confers plant manganese tolerance. Proc. Natl. Acad. Sci. U.S.A. 104, 8532–8537. doi: [10.1073/](https://doi.org/10.1073/pnas.0609507104) [pnas.0609507104](https://doi.org/10.1073/pnas.0609507104)

Pena, L. B., Azpilicueta, C. E., and Gallego, S. M. (2011). Sunflower cotyledons cope with copper stress by inducing catalase subunits less sensitive to oxidation. J. Trace. Elem. Med. Biol. 25, 125–129. doi: [10.1016/j.jtemb.2011.05.001](https://doi.org/10.1016/j.jtemb.2011.05.001)

Peng, J. S., and Gong, J. M. (2014). Vacuolar sequestration capacity and long-distance metal transport in plants. Front. Plant Sci. 5, 19. doi: [10.3389/fpls.2014.00019](https://doi.org/10.3389/fpls.2014.00019)

Perea-García, A., Garcia-Molina, A., Andrés-Colás, N., Vera-Sirera, F., Pérez-Amador, M. A., Puig, S., et al. (2013). Arabidopsis copper transport protein COPT2 participates in the cross-talk between iron deficiency responses and low-phosphate signaling. Plant Physiol. 162, 180-194. doi: [10.1104/pp.112.212407](https://doi.org/10.1104/pp.112.212407)

Perincherry, L., Stępień, Ł., and Vasudevan, S. E. (2021). Cross-tolerance and autoimmunity as missing links in abiotic and biotic stress responses in plants: a perspective toward secondary metabolic engineering. Int. J. Mol. Sci. 22, 11945. doi: [10.3390/ijms222111945](https://doi.org/10.3390/ijms222111945)

Phaniendra, A., Jestadi, D. B., and Periyasamy, L. (2015). Free radicals: properties, sources, targets, and their implication in various diseases. Indian J. Clin. Biochem. 30, 11-26. doi: [10.1007/s12291-014-0446-0](https://doi.org/10.1007/s12291-014-0446-0)

Piñeros, M. A., Magalhaes, J. V., Carvalho Alves, V. M., and Kochian, L. V. (2002). The physiology and biophysics of an aluminum tolerance mechanism based on root citrate exudation in maize. Plant Physiol. 129, 1194–1206. doi: [10.1104/pp.002295](https://doi.org/10.1104/pp.002295)

Pittman, J. K., and Hirschi, K. D. (2016). CAX-ing a wide net: Cation/H(+) transporters in metal remediation and abiotic stress signaling. Plant Biol. 18, 741– 749. doi: [10.1111/plb.12460](https://doi.org/10.1111/plb.12460)

Podar, D., and Maathuis, F. J. M. (2022). The role of roots and rhizosphere in providing tolerance to toxic metals and metalloids. Plant Cell Environ. 45, 719–736. doi: [10.1111/pce.14188](https://doi.org/10.1111/pce.14188)

Pospišil, P., and Yamamoto, Y. (2017). Damage to photosystem II by lipid peroxidation products. Biochim. Biophys. Acta Gen. Subj. 1861, 457–466. doi: [10.1016/j.bbagen.2016.10.005](https://doi.org/10.1016/j.bbagen.2016.10.005)

Pourrut, B., Shahid, M., Dumat, C., Winterton, P., and Pinelli, E. (2011). Lead uptake, toxicity, and detoxification in plants. Rev. Environ. Contam. Toxicol. 213, 113–136. doi: [10.1007/978-1-4419-9860-6_4](https://doi.org/10.1007/978-1-4419-9860-6_4)

Priya, A. K., Muruganandam, M., Ali, S. S., and Kornaros, M. (2023). Clean-up of heavy metals from contaminated soil by phytoremediation: a multidisciplinary and eco-friendly approach. Toxics 11, 422. doi: [10.3390/toxics11050422](https://doi.org/10.3390/toxics11050422)

Qu, T., Ma, Y., Yun, M., and Zhao, C. (2024). Transcriptome analysis revealed the possible reasons for the change of Ni resistance in Rhus typhina after spraying melatonin. Plants 13 (10), 1287. doi: [10.3390/plants13101287](https://doi.org/10.3390/plants13101287)

Rahoui, S., Chaoui, A., and Ferjani, E. (2010). Membrane damage and solute leakage from germinating pea seed under cadmium stress. J. Hazard. Mater. 178, 1128–1131. doi: [10.1016/j.jhazmat.2010.01.115](https://doi.org/10.1016/j.jhazmat.2010.01.115)

Rai, S., Singh, P. K., Mankotia, S., Swain, J., and Satbhai, S. B. (2021). Iron homeostasis in plants and its crosstalk with copper, zinc, and manganese. Plant Stress 1, 100008. doi: [10.1016/j.stress.2021.100008](https://doi.org/10.1016/j.stress.2021.100008)

Rajput, V. D., Singh, R. K., Verma, K. K., Sharma, L., Quiroz-Figueroa, F. R., et al. (2021). Recent developments in enzymatic antioxidant defense mechanism in plants with special reference to abiotic stress. Biology 10 (4), 267. doi: [10.3390/](https://doi.org/10.3390/biology10040267) [biology10040267](https://doi.org/10.3390/biology10040267)

Rancelis, V., Cesniene, T., Kleizaite, V., Zvingila, D., and Balciuniene, L. (2012). Influence of cobalt uptake by Vicia faba seeds on chlorophyll morphosis induction, SOD polymorphism, and DNA methylation. Environ. Toxicol. 27, 32–41. doi: [10.1002/](https://doi.org/10.1002/tox.20609) [tox.20609](https://doi.org/10.1002/tox.20609)

Rasheed, A., Li, H., Tahir, M. M., Mahmood, A., Nawaz, M., Shah, A. N., et al. (2022). The role of nanoparticles in plant biochemical, physiological, and molecular responses under drought stress: a review. Front. Plant Sci. 13, 976179. doi: [10.3389/](https://doi.org/10.3389/fpls.2022.976179) [fpls.2022.976179](https://doi.org/10.3389/fpls.2022.976179)

Rashid, A., Schutte, B. J., Ulery, A., Deyholos, M. K., Sanogo, S., Lehnhoff, E. A., et al. (2023). Heavy metal contamination in agricultural soil: environmental pollutants affecting crop health. Agronomy 13, 1521. doi: [10.3390/agronomy13061521](https://doi.org/10.3390/agronomy13061521)

Raychaudhuri, S. S., Pramanick, P., Talukder, P., and Basak, A. (2021). "Polyamines, metallothioneins, and phytochelatins—Natural defense of plants to mitigate heavy metals," in Bioactive natural products. Ed. F. R. S. Atta-ur-Rahman (Elsevier, Amsterdam, Netherlands), 227–261. doi: [10.1016/B978-0-12-819487-4.00006-9](https://doi.org/10.1016/B978-0-12-819487-4.00006-9)

Ricachenevsky, F. K., Menguer, P. K., Sperotto, R. A., Williams, L. E., and Fett, J. P. (2013). Roles of plant metal tolerance proteins (MTP) in metal storage and potential use in biofortification strategies. Front. Plant Sci. 4, 144. doi: [10.3389/fpls.2013.00144](https://doi.org/10.3389/fpls.2013.00144)

Rieuwerts, J. S., Thornton, I., Farago, M. E., and Ashmore, M. R. (1998). Factors influencing metal bioavailability in soils: preliminary investigations for the development of a critical loads approach for metals. Chem. Speciation Bioavailability 10, 61–75. doi: [10.3184/095422998782775835](https://doi.org/10.3184/095422998782775835)

Riyazuddin, R., Nisha, N., Ejaz, B., Khan, M. I. R., Kumar, M., Ramteke, P. W., et al. (2021). A comprehensive review on the heavy metal toxicity and sequestration in plants. Biomolecules 12, 43. doi: [10.3390/biom12010043](https://doi.org/10.3390/biom12010043)

Rodríguez Eugenio, N., McLaughlin, M. J., Pennock, D. J.Land and Water Development Div. Food and Agriculture Organization of the United Nations (2018). Soil pollution: a hidden reality (Rome (Italy: FAO).

Ruttkay-Nedecky, B., Nejdl, L., Gumulec, J., Zitka, O., Masarik, M., Eckschlager, T., et al. (2013). The role of metallothionein in oxidative stress. Int. J. Mol. Sci. 14, 6044– 6066. doi: [10.3390/ijms14036044](https://doi.org/10.3390/ijms14036044)

Ryu, H. W., Lee, D. H., Won, H. R., Kim, K. H., Seong, Y. J., and Kwon, S. H. (2015). Influence of toxicologically relevant metals on human epigenetic regulation. Toxicol. Res. 31, 1–9. doi: [10.5487/TR.2015.31.1.001](https://doi.org/10.5487/TR.2015.31.1.001)

Saboor, A., Ali, M. A., Hussain, S., El Enshasy, H. A., Hussain, S., Ahmed, N., et al. (2021). Zinc nutrition and arbuscular mycorrhizal symbiosis effects on maize (Zea mays L.) growth and productivity. Saudi J. Biol. Sci. 28, 6339–6351. doi: [10.1016/](https://doi.org/10.1016/j.sjbs.2021.06.096) [j.sjbs.2021.06.096](https://doi.org/10.1016/j.sjbs.2021.06.096)

Sabreena, S., Bhat, S. A., Kumar, V., and Ganai, B. A. (2022). Phytoremediation of heavy metals: an indispensable contrivance in green remediation technology. Plants 11 (9), 1255. doi: [10.3390/plants11091255](https://doi.org/10.3390/plants11091255)

Salbitani, G., Maresca, V., Cianciullo, P., Bossa, R., Carfagna, S., and Basile, A. (2023). Non-protein thiol compounds and antioxidant responses involved in bryophyte heavymetal tolerance. Int. J. Mol. Sci. 24 (6), 5302. doi: [10.3390/ijms24065302](https://doi.org/10.3390/ijms24065302)

Salinitro, M., Mattarello, G., Guardigli, G., Odajiu, M., and Tassoni, A. (2021). Induction of hormesis in plants by urban trace metal pollution. Sci. Rep. 11, 20329. doi: [10.1038/s41598-021-99657-3](https://doi.org/10.1038/s41598-021-99657-3)

Sancenón, V., Puig, S., Mira, H., Thiele, D. J., and Peñarrubia, L. (2003). Identification of a copper transporter family in Arabidopsis thaliana. Plant Mol. Biol. 51, 577–587. doi: [10.1023/a:1022345507112](https://doi.org/10.1023/a:1022345507112)

Sarwar, N., Imran, M., Shaheen, M. R., Ishaque, W., Kamran, M. A., Matloob, A., et al. (2017). Phytoremediation strategies for soils contaminated with heavy metals: modifications and future perspectives. Chemosphere 171, 710–721. doi: [10.1016/](https://doi.org/10.1016/j.chemosphere.2016.12.116) [j.chemosphere.2016.12.116](https://doi.org/10.1016/j.chemosphere.2016.12.116)

Sasaki, A., Yamaji, N., and Ma, J. F. (2014). Overexpression of OsHMA3 enhances Cd tolerance and expression of Zn transporter genes in rice. J. Exp. Bot. 65, 6013–6021. doi: [10.1093/jxb/eru340](https://doi.org/10.1093/jxb/eru340)

Sattelmacher, B. (2001). The apoplast and its significance for plant mineral nutrition. New Phytol. 149, 167–192. doi: [10.1046/j.1469-8137.2001.00034.x](https://doi.org/10.1046/j.1469-8137.2001.00034.x)

Sawidis, T., Bayçu, G., Weryszko-Chmielewska, E., and Sulborska, A. (2021). Impact of manganese on pollen germination and tube growth in lily. Acta. Agrobot. 74, 1–17. doi: [10.5586/aa.746](https://doi.org/10.5586/aa.746)

Schmidt, S. B., Eisenhut, M., and Schneider, A. (2020). Chloroplast transition metal regulation for efficient photosynthesis. Trends. Plant Sci. 25, 817–828. doi: [10.1016/](https://doi.org/10.1016/j.tplants.2020.03.003) [j.tplants.2020.03.003](https://doi.org/10.1016/j.tplants.2020.03.003)

Seeda, A., Abou El-Nour, E.-Z., and Zaghloul, S. (2020). Importance of molybdenum and it diverse role in plant physiology: a review. Middle East J. Appl. Sci. 10, 228–249. doi: [10.36632/mejas/2020.10.2.23](https://doi.org/10.36632/mejas/2020.10.2.23)

Sethy, S. K., and Ghosh, S. (2013). Effect of heavy metals on germination of seeds. J. Nat. Sci. Biol. Med. 4, 272–275. doi: [10.4103/0976-9668.116964](https://doi.org/10.4103/0976-9668.116964)

Shahid, M., Pourrut, B., Dumat, C., Nadeem, M., Aslam, M., and Pinelli, E. (2014). Heavy-metal-induced reactive oxygen species: phytotoxicity and physicochemical changes in plants. Rev. Environ. Contam. Toxicol. 232, 1–44. doi: [10.1007/978-3-319-](https://doi.org/10.1007/978-3-319-06746-9_1) [06746-9_1](https://doi.org/10.1007/978-3-319-06746-9_1)

Shahzad, Z., Gosti, F., Frérot, H., Lacombe, E., Roosens, N., Saumitou-Laprade, P., et al. (2010). The five AhMTP1 zinc transporters undergo different evolutionary fates towards adaptive evolution to zinc tolerance in Arabidopsis halleri. PLOS Genet. 6, e1000911. doi: [10.1371/journal.pgen.1000911](https://doi.org/10.1371/journal.pgen.1000911)

Sharma, S. S., Dietz, K.-J., and Mimura, T. (2016). Vacuolar compartmentalization as indispensable component of heavy metal detoxification in plants. Plant Cell Environ. 39, 1112–1126. doi: [10.1111/pce.12706](https://doi.org/10.1111/pce.12706)

Sharma, A., Sidhu, G. P. S., Araniti, F., Bali, A. S., Shahzad, B., Tripathi, D. K., et al. (2020). The role of salicylic acid in plants exposed to heavy metals. Molecules 25, 540. doi: [10.3390/molecules25030540](https://doi.org/10.3390/molecules25030540)

Shi, M., Wang, S., Zhang, Y., Wang, S., Zhao, J., Feng, H., et al. (2020). Genome-wide characterization and expression analysis of ATP-binding cassette (ABC) transporters in strawberry reveal the role of FvABCC11 in cadmium tolerance. Sci. Hortic. 271, 109464. doi: [10.1016/j.scienta.2020.109464](https://doi.org/10.1016/j.scienta.2020.109464)

Shingu, Y., Kudo, T., Ohsato, S., Kimura, M., Ono, Y., Yamaguchi, I., et al. (2005). Characterization of genes encoding metal tolerance proteins isolated from Nicotiana glauca and Nicotiana tabacum. Biochem. Biophys. Res. Commun. 331, 675–680. doi: [10.1016/j.bbrc.2005.04.010](https://doi.org/10.1016/j.bbrc.2005.04.010)

Shuting, Z., Hongwei, D., Qing, M., Rui, H., Huarong, T., and Lianyu, Y. (2022). Identification and expression analysis of the ZRT, IRT-like protein (ZIP) gene family in Camellia sinensis (L.) O. Kuntze. Plant Physiol. Biochem. 172, 87–100. doi: [10.1016/](https://doi.org/10.1016/j.plaphy.2022.01.008) [j.plaphy.2022.01.008](https://doi.org/10.1016/j.plaphy.2022.01.008)

Singh, S., Parihar, P., Singh, R., Singh, V. P., and Prasad, S. M. (2015). Heavy metal tolerance in plants: role of transcriptomics, proteomics, metabolomics, and ionomics. Front. Plant Sci. 6, 1143. doi: [10.3389/fpls.2015.01143](https://doi.org/10.3389/fpls.2015.01143)

Singh, S., Yadav, V., Arif, N., Singh, V. P., Dubey, N. K., Ramawat, N., et al. (2020). "Heavy metal stress and plant life: uptake mechanisms, toxicity, and alleviation," in Plant life under changing environment. Eds. D. K. Tripathi, V. P. Singh, V. ,. D. K. Chauhan, S. Sharma, S. M. Prasad, N. K. Dubey and N. Ramawat (Academic Press, Cambridge, Massachusetts, USA), 271–287. doi: [10.1016/B978-0-12-818204-8.00001-1](https://doi.org/10.1016/B978-0-12-818204-8.00001-1)

Siripornadulsil, S., Traina, S., Verma, D. P., and Sayre, R. T. (2002). Molecular mechanisms of proline-mediated tolerance to toxic heavy metals in transgenic microalgae. Plant Cell 14, 2837–2847. doi: [10.1105/tpc.004853](https://doi.org/10.1105/tpc.004853)

Skuza, L., Szućko-Kociuba, I., Filip, E., and Bożek, I. (2022). Natural molecular mechanisms of plant hyperaccumulation and hypertolerance towards heavy metals. Int. J. Mol. Sci. 23, 9335. doi: [10.3390/ijms23169335](https://doi.org/10.3390/ijms23169335)

Song, W. Y., Mendoza-Cózatl, D. G., Lee, Y., Schroeder, J. I., Ahn, S. N., Lee, H. S., et al. (2014). Phytochelatin–metal (loid) transport into vacuoles shows different substrate preferences in barley and Arabidopsis. Plant Cell Environ. 37, 1192–1201. doi: [10.1111/pce.12227](https://doi.org/10.1111/pce.12227)

Srivastava, V., Sarkar, A., Singh, S., Singh, P., de Araujo, A. S. F., and Singh, R. P. (2017). Agroecological responses of heavy metal pollution with special emphasis on soil health and plant performances. Front. Environ. Sci. 5, 64. doi: [10.3389/fenvs.2017.00064](https://doi.org/10.3389/fenvs.2017.00064)

Su, N., Ling, F., Xing, A., Zhao, H., Zhu, Y., Wang, Y., et al. (2020). Lignin synthesis mediated by CCoAOMT enzymes is required for the tolerance against excess Cu in Oryza sativa. Environ. Exp. Bot. 175, 104059. doi: [10.1016/j.envexpbot.2020.104059](https://doi.org/10.1016/j.envexpbot.2020.104059)

Subramanian Vignesh, K., and Deepe, G. S. Jr. (2017). Metallothioneins: emerging modulators in immunity and infection. Int. J. Mol. Sci. 18 (10), 2197. doi: [10.3390/](https://doi.org/10.3390/ijms18102197) [ijms18102197](https://doi.org/10.3390/ijms18102197)

Sunda, W. G. (2012). Feedback interactions between trace metal nutrients and phytoplankton in the ocean. Front. Microbiol. 3, 204. doi: [10.3389/fmicb.2012.00204](https://doi.org/10.3389/fmicb.2012.00204)

Sung, D. Y., Kim, T. H., Komives, E. A., Mendoza-Cózatl, D. G., and Schroeder, J. I. (2009). ARS5 is a component of the 26S proteasome complex, and negatively regulates thiol biosynthesis and arsenic tolerance in Arabidopsis. Plant J. 59, 802-813. doi: [10.1111/j.1365-313X.2009.03914.x](https://doi.org/10.1111/j.1365-313X.2009.03914.x)

Sunkar, R., Kaplan, B., Bouché, N., Arazi, T., Dolev, D., Talke, I. N., et al. (2000). Expression of a truncated tobacco NtCBP4 channel in transgenic plants and disruption
of the homologous *Arabidopsis* CNGC1 gene confer Pb²⁺ tolerance. *Plant J.* 24, 533– 542. doi: [10.1046/j.1365-313x.2000.00901.x](https://doi.org/10.1046/j.1365-313x.2000.00901.x)

Takahashi, M., Terada, Y., Nakai, I., Nakanishi, H., Yoshimura, E., Mori, S., et al. (2003). Role of nicotianamine in the intracellular delivery of metals and plant reproductive development. Plant Cell 15, 1263–1280. doi: [10.1105/tpc.010256](https://doi.org/10.1105/tpc.010256)

Tang, J., Zhang, J., Ren, L., Zhou, Y., Gao, J., Luo, L., et al. (2019). Diagnosis of soil contamination using microbiological indices: a review on heavy metal pollution. J.Environ. Manage. 242, 121–130. doi: [10.1016/j.jenvman.2019.04.061](https://doi.org/10.1016/j.jenvman.2019.04.061)

Thirumoorthy, N., Manisenthil Kumar, K. T., Shyam Sundar, A., Panayappan, L., and Chatterjee, M. (2007). Metallothionein: an overview. World J. Gastroenterol. 13, 993–996. doi: [10.3748/wjg.v13.i7.993](https://doi.org/10.3748/wjg.v13.i7.993)

Tiwari, S., and Lata, C. (2018). Heavy metal stress, signaling, and tolerance due to plant-associated microbes: an overview. Front. Plant Sci. 9, 452. doi: [10.3389/](https://doi.org/10.3389/fpls.2018.00452) [fpls.2018.00452](https://doi.org/10.3389/fpls.2018.00452)

Tong, Y. P., Kneer, R., and Zhu, Y. G. (2004). Vacuolar compartmentalization: a second-generation approach to engineering plants for phytoremediation. *Trends. Plant*
*Sci. 9, 7–*9. doi: [10.1016/j.tplants.2003.11.009](https://doi.org/10.1016/j.tplants.2003.11.009)

Toyoda, Y., Hagiya, Y., Adachi, T., Hoshijima, K., Kuo, M. T., and Ishikawa, T. (2008). MRP class of human ATP binding cassette (ABC) transporters: historical background and new research directions. Xenobiotica 38, 833–862. doi: [10.1080/](https://doi.org/10.1080/00498250701883514) [00498250701883514](https://doi.org/10.1080/00498250701883514)

Tyagi, S., Singh, K., and Upadhyay, S. K. (2021). Molecular characterization revealed the role of catalases under abiotic and arsenic stress in bread wheat (Triticum aestivum L.). J. Hazard. Mater. 403, 123585. doi: [10.1016/j.jhazmat.2020.123585](https://doi.org/10.1016/j.jhazmat.2020.123585)

Umair Hassan, M., Aamer, M., Umer Chattha, M., Haiying, T., Shahzad, B., Barbanti, L., et al. (2020). The critical role of zinc in plants facing the drought stress. Agriculture 10, 396. doi: [10.3390/agriculture10090396](https://doi.org/10.3390/agriculture10090396)

Ur Rahman, S., Qin, A., Zain, M., Mushtaq, Z., Mehmood, F., Riaz, L., et al. (2024). Pb uptake, accumulation, and translocation in plants: plant physiological, biochemical, and molecular response: a review. Heliyon 10, e27724. doi: [10.1016/j.heliyon.2024.e27724](https://doi.org/10.1016/j.heliyon.2024.e27724)

van der Zaal, B. J., Neuteboom, L. W., Pinas, J. E., Chardonnens, A. N., Schat, H., Verkleij, J. A., et al. (1999). Overexpression of a novel Arabidopsis gene related to putative zinc-transporter genes from animals can lead to enhanced zinc resistance and accumulation. Plant Physiol. 119, 1047–1056. doi: [10.1104/pp.119.3.1047](https://doi.org/10.1104/pp.119.3.1047)

Vargas-Hernandez, M., Macias-Bobadilla, I., Guevara-Gonzalez, R. G., Romero-Gomez, S. J., Rico-Garcia, E., Ocampo-Velazquez, R. V., et al. (2017). Plant hormesis management with biostimulants of biotic origin in agriculture. Front. Plant Sci. 8, 1762. doi: [10.3389/fpls.2017.01762](https://doi.org/10.3389/fpls.2017.01762)

Ventrella, A., Catucci, L., Piletska, E., Piletsky, S., and Agostiano, A. (2009). Interactions between heavy metals and photosynthetic materials studied by optical techniques. Bioelectrochemistry 77, 19–25. doi: [10.1016/j.bioelechem.2009.05.002](https://doi.org/10.1016/j.bioelechem.2009.05.002)

Verma, R., Yadav, D., Singh, C., Suman, A., and Gaur, A. (2010). Effect of heavy metals on soil respiration during decomposition of sugarcane (Saccharum officinarum L.) trash in different soils. Plant Soil Environ. 56, 76–81. doi: [10.17221/1773-PSE](https://doi.org/10.17221/1773-PSE)

Vidal, C., Larama, G., Riveros, A., Meneses, C., and Cornejo, P. (2021). Main molecular pathways associated with copper tolerance response in Imperata cylindrica by de novo transcriptome assembly. Plants 10, 357. doi: [10.3390/plants10020357](https://doi.org/10.3390/plants10020357)

Vijayaragavan, M., Prabhahar, C., Sureshkumar, J., Natarajan, A., Vijayarengan, P., and Sharavanan, S. (2011). Toxic effect of cadmium on seed germination, growth and biochemical contents of cowpea (Vigna unguiculata L.) plants. Int. Multidiscip. Res. J. 1, 1–6. Available at: [https://api.semanticscholar.org/CorpusID:83038868.](https://api.semanticscholar.org/CorpusID:83038868)

Wang, Y., Meng, Y., Mu, S., Yan, D., Xu, X., Zhang, L., et al. (2021b). Changes in phenotype and gene expression under lead stress revealed key genetic responses to lead tolerance in Medicago sativa L. Gene 791, 145714. doi: [10.1016/j.gene.2021.145714](https://doi.org/10.1016/j.gene.2021.145714)

Wang, Q., Wei, N., Jin, X., Min, X., Ma, Y., and Liu, W. (2021a). Molecular characterization of the COPT/Ctr-type copper transporter family under heavy metal stress
in alfalfa. *Int. J. Biol. Macromol.* 181, 644–652. doi: [10.1016/j.ijbiomac.2021.03.173](https://doi.org/10.1016/j.ijbiomac.2021.03.173)

Wang, Y., Wimmer, U., Lichtlen, P., Inderbitzin, D., Stieger, B., Meier, P. J., et al. (2004). Metal-responsive transcription factor-1 (MTF-1) is essential for embryonic liver development and heavy metal detoxification in the adult liver. Faseb. J. 18, 1071– 1079. doi: [10.1096/fj.03-1282com](https://doi.org/10.1096/fj.03-1282com)

Witkowska, D., Słowik, J., and Chilicka, K. (2021). Heavy metals and human health: ossible exposure pathways and the competition for protein binding sites. Molecules 26 (19), 6060. doi: [10.3390/molecules26196060](https://doi.org/10.3390/molecules26196060)

Wu, X., Chen, Q., Chen, L., Tian, F., Chen, X., Han, C., et al. (2022). A WRKY transcription factor, PyWRKY75, enhanced cadmium accumulation and tolerance in
poplar. *Ecotoxicol. Environ. Saf.* 239, 113630. doi: [10.1016/j.ecoenv.2022.113630](https://doi.org/10.1016/j.ecoenv.2022.113630)

Wu, H., Cui, H., Fu, C., Li, R., Qi, F., Liu, Z., et al. (2024). Unveiling the crucial role of soil microorganisms in carbon cycling: a review. Sci. Total Environ. 909, 168627. doi: [10.1016/j.scitotenv.2023.168627](https://doi.org/10.1016/j.scitotenv.2023.168627)

Xia, Y., Qi, Y., Yuan, Y., Wang, G., Cui, J., Chen, Y., et al. (2012). Overexpression of Elsholtzia haichowensis metallothionein 1 (EhMT1) in tobacco plants enhances copper tolerance and accumulation in root cytoplasm and decreases hydrogen peroxide production. J. Hazard. Mater. 233, 65–71. doi: [10.1016/j.jhazmat.2012.06.047](https://doi.org/10.1016/j.jhazmat.2012.06.047)

Xiao, S., Gao, W., Chen, Q. F., Ramalingam, S., and Chye, M. L. (2008). Overexpression of membrane-associated acyl-CoA-binding protein ACBP1 enhances lead tolerance in Arabidopsis. Plant J. 54, 141–151. doi: [10.1111/j.1365-313X.2008.03402.x](https://doi.org/10.1111/j.1365-313X.2008.03402.x)

Xu, J., Yin, H., Liu, X., and Li, X. (2010). Salt affects plant Cd-stress responses by modulating growth and Cd accumulation. Planta 231, 449-459. doi: [10.1007/s00425-](https://doi.org/10.1007/s00425-009-1070-8) $009 - 1070 - 8$

Xu, Y., Zhang, S., Guo, H., Wang, S., Xu, L., Li, C., et al. (2014). OsABCB14 functions in auxin transport and iron homeostasis in rice (Oryza sativa L.). Plant J. 79, 106–117. doi: [10.1111/tpj.12544](https://doi.org/10.1111/tpj.12544)

Yadav, S. K. (2010). Heavy metals toxicity in plants: an overview on the role of glutathione and phytochelatins in heavy metal stress tolerance of plants. South Afr. J. Bot. 76, 167–179. doi: [10.1016/j.sajb.2009.10.007](https://doi.org/10.1016/j.sajb.2009.10.007)

Yan, A., Wang, Y., Tan, S. N., Mohd Yusof, M. L., Ghosh, S., and Chen, Z. (2020). Phytoremediation: a promising approach for revegetation of heavy metal-polluted land. Front. Plant Sci. 11, 359. doi: [10.3389/fpls.2020.00359](https://doi.org/10.3389/fpls.2020.00359)

Yang, X., Feng, Y., He, Z., and Stoffella, P. J. (2005). Molecular mechanisms of heavy metal hyperaccumulation and phytoremediation. J. Trace Elem. Med. Biol. 18, 339–353. doi: [10.1016/j.jtemb.2005.02.007](https://doi.org/10.1016/j.jtemb.2005.02.007)

Yang, G., Fu, S., Huang, J., Li, L., Long, Y., Wei, Q., et al. (2021a). The tonoplast-localized transporter OsABCC9 is involved in cadmium tolerance and accumulation in rice. Plant Sci. 307, 110894. doi: [10.1016/j.plantsci.2021.110894](https://doi.org/10.1016/j.plantsci.2021.110894)

Yang, Y. R., Hou, S. L., Zhang, Z. W., Hu, Y. Y., Ding, C., Yang, G. J., et al. (2021b). Effects of nitrogen addition on plant manganese nutrition in a temperate steppe. J. Plant Nutr. Soil. Sci. 184, 688–695. doi: [10.1002/jpln.202100144](https://doi.org/10.1002/jpln.202100144)

Yao, J., Sun, J., Chen, Y., Shi, L., Yang, L., and Wang, Y. (2020). The molecular mechanism underlying cadmium resistance in NHX1 transgenic Lemna turonifera was studied by comparative transcriptome analysis. Plant Cell Tiss. Organ. Cult. 143, 189– 200. doi: [10.1007/s11240-020-01909-z](https://doi.org/10.1007/s11240-020-01909-z)

Yimer, M., Ansari, S. N., Berehe, B. A., Gudimella, K. K., Gedda, G., Girma, W. M., et al. (2024). Adsorptive removal of heavy metals from wastewater using cobaltdiphenylamine (Co-DPA) complex. BMC Chem. 18, 23. doi: [10.1186/s13065-024-](https://doi.org/10.1186/s13065-024-01128-z) $01128-z$

Yuan, M., Li, X., Xiao, J., and Wang, S. (2011). Molecular and functional analyses of COPT/Ctr-type copper transporter-like gene family in rice. BMC Plant Biol. 11, 69. doi: [10.1186/1471-2229-11-69](https://doi.org/10.1186/1471-2229-11-69)

Yuan, L., Yang, S., Liu, B., Zhang, M., and Wu, K. (2012). Molecular characterization of a rice metal tolerance protein, OsMTP1. Plant Cell Rep. 31, 67–79. doi: [10.1007/](https://doi.org/10.1007/s00299-011-1140-9) [s00299-011-1140-9](https://doi.org/10.1007/s00299-011-1140-9)

Yusuf, M., Fariduddin, Q., Varshney, P., and Ahmad, A. (2012). Salicylic acid minimizes nickel and/or salinity-induced toxicity in Indian mustard (Brassica juncea) through an improved antioxidant system. Environ. Sci. pollut. Res. 19, 8–18. doi: [10.1007/s11356-011-0531-3](https://doi.org/10.1007/s11356-011-0531-3)

Zandi, P., and Schnug, E. (2022). Reactive oxygen species, antioxidant responses and implications from a microbial modulation perspective. *Biology* 11, 155. doi: [10.3390/](https://doi.org/10.3390/biology11020155) [biology11020155](https://doi.org/10.3390/biology11020155)

Zayed, O., Hewedy, O. A., Abdelmoteleb, A., Ali, M., Youssef, M. S., Roumia, A. F., et al. (2023). Nitrogen journey in plants: from uptake to metabolism, stress response, and microbe interaction. Biomolecules 13 (10), 1443. doi: [10.3390/biom13101443](https://doi.org/10.3390/biom13101443)

Zhakypbek, Y., Kossalbayev, B. D., Belkozhayev, A. M., Murat, T., Tursbekov, S., Abdalimov, E., et al. (2024). Reducing heavy metal contamination in soil and water using phytoremediation. Plants 13 (11), 1534. doi: [10.3390/plants13111534](https://doi.org/10.3390/plants13111534)

Zhan, J., Zou, W., Li, S., Tang, J., Lu, X., Meng, L., et al. (2022). OsNAC15 regulates tolerance to zinc deficiency and cadmium by binding to OsZIP7 and OsZIP10 in rice. Int. J. Mol. Sci. 23, 11771. doi: [10.3390/ijms231911771](https://doi.org/10.3390/ijms231911771)

Zhang, Y., Deng, B., and Li, Z. (2018b). Inhibition of NADPH oxidase increases defense enzyme activities and improves maize seed germination under Pb stress. Ecotoxicol. Environ. Saf. 158, 187–192. doi: [10.1016/j.ecoenv.2018.04.028](https://doi.org/10.1016/j.ecoenv.2018.04.028)

Zhang, J., Martinoia, E., and Lee, Y. (2018a). Vacuolar transporters for cadmium and arsenic in plants and their applications in phytoremediation and crop development. Plant Cell Physiol. 59, 1317–1325. doi: [10.1093/pcp/pcy006](https://doi.org/10.1093/pcp/pcy006)

Zheng, S. J., Yang, J. L., He, Y. F., Yu, X. H., Zhang, L., You, J. F., et al. (2005). Immobilization of aluminum with phosphorus in roots is associated with high aluminum resistance in buckwheat. Plant Physiol. 138, 297–303. doi: [10.1104/pp.105.059667](https://doi.org/10.1104/pp.105.059667)

Zhou, P., Adeel, M., Shakoor, N., Guo, M., Hao, Y., Azeem, I., et al. (2020). Application of nanoparticles alleviates heavy metals stress and promotes plant growth: an overview. Nanomaterials 11, 26. doi: [10.3390/nano11010026](https://doi.org/10.3390/nano11010026)

Zhou, B., Yao, W., Wang, S., Wang, X., and Jiang, T. (2014). The metallothionein gene, TaMT3, from *Tamarix androssowii* confers Cd^{2+} tolerance in tobacco. *Int. J. Mol.* Sci. 15, 10398–10409. doi: [10.3390/ijms150610398](https://doi.org/10.3390/ijms150610398)

Zhu, C., Yu, J., Cao, S., Wu, X., Meng, W., and Hou, X. (2022). Transcriptomicsbased analysis of genes related to lead stress and their expression in the roots of Pogonatherum crinitum. Front. Plant Sci. 13, 1066329. doi: [10.3389/fpls.2022.1066329](https://doi.org/10.3389/fpls.2022.1066329)