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RECEIVED 14 November 2023 ACCEPTED 15 February 2024 PUBLISHED 20 March 2024

CITATION

Hao J, Zhao Z, Fu X, Zhao Y, Ateeq M, Mou L, Han Y, Liu Y, Yin Y, Zotova L, Serikbay D, Fan C, Hu Y-G and Chen L (2024) Effect of a novel dwarfing mutant site on chromosome 4B on agronomic traits in common wheat. *Front. Plant Sci.* 15:1338425. doi: 10.3389/fpls.2024.1338425

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Effect of a novel dwarfing mutant site on chromosome 4B on agronomic traits in common wheat

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The introduction of dwarfing genes triggered a wave of "green revolution". A number of wheats dwarfing genes have been reported in previous studies, and only a small fraction of these have been applied to production practices. Therefore, the development of novel dwarfing genes for wheat is of great value. In this study, a novel dwarfing site, Rht-yz, identified in the Yanzhan mutation, is located on chromosome 4B (30-33MB) and its mechanism of action is different from that of Rht-B1b (C-T mutation), but whether it affects the Rht-B1a (TraesCS4B02G043100) or other genes is unclear. Exogenously applied GA₃ experiments showed that Rht-yz is one of the gibberellin-insensitive dwarf genes. The effects of the dwarf gene Rht-yz on agronomic traits in wheat were evaluated in the field using Yanzhan, Yanzhan mutations, $F_{2:3}$ and $F_{3:4}$ lines. The results showed that Rht-yz improved lodging resistance by reducing plant height, increasing diameter, wall thickness and mechanical strength of the basal stem. In terms of yield traits, Rht-yz had negative effects on tiller number plant⁻¹, biomass plant⁻¹ and yield plant⁻¹, but had no significant effect on harvest index, 1000-kernel weight and spike traits. In addition, Rht-yz significantly increased crude protein, wet gluten and starch content. Therefore, the rational use of the new dwarfing site Rht-yz has potential and value in dwarf wheat breeding.

KEYWORDS

dwarf gene, plant height, lodging resistance, wheat, agronomic traits

Introduction

Wheat (Triticum aestivum L.) is an important cereal crop playing outstanding role in feeding the hunger world and global food security. Plant height is one of the main agronomic traits for morphogenesis affecting wheat yield. In the "Green Revolution" of late 20th century, Rht-B1b and Rht-D1b significantly reduced plant height in wheat and increased wheat production (Hedden, 2003). Wheat varieties carrying dwarf genes Rht-B1b and Rht-D1b (Chihoumu and Dharma) rapidly expanded the planting area, and the wheat dwarf gene was gradually used in wheat breeding. To date, more than 25 dwarf genes have been discovered. The mainly used wheat dwarf genes were Rht-B1b, Rht-D1b and Rht8, which limited the improvement in wheat yield potential. Rht-B1b/Rht-D1b reduced plant height by 23%, and Rht-B1b+Rht-D1b reduced plant height by 47% (Richards, 1992), but Rht-B1b and Rht-D1b had certain adverse effects on seedling traits. The dwarf gene Rht-B1b and Rht8 also had a significant effect on the tillers number plant⁻¹ in wheat. Rht-B1b reduced 1000-kernel weight but increased yield to eliminate the negative effect of small biomass (Hayat et al., 2019). Meanwhile, Rht-B1b and Rht-D1b associated with reduction in seed size and protein content (Casebow et al., 2016).

Furthermore, the dwarf gene *Rht3* has a three times greater effect on plant height than *Rht-B1b*, but plants that are too short are not conducive to yield formation (Gale et al., 1985; Flintham et al., 2000; Derkx et al., 2017). *Rht5* reduced plant height by 25.84%, but had a negative effect on spike development (Daoura et al., 2014). *Rht13* reduces plant height by 21.5% but has a negative effect on internode diameter (Wang et al., 2014). Therefore, many dwarf genes are difficult to be applied in wheat breeding.

Statistics show that about 70% of the wheat varieties released worldwide contain *Rht-B1b* or/and *Rht-D1b* (Evans, 2008). During 2012-2018, *Rht-D1b* was present at a frequency of more than 80% in Uniform Southern Soft Red Winter Wheat Nursery (Hayat et al., 2019). In addition, more than half of breeding lines in SRWW in the southern United States carry *Rht-D1b*, and approximately 31% of breeding lines carry *Rht-B1b* (Guedira et al., 2010). *Rht-B1b* and *Rht-D1b* were also widely distributed in Chinese wheat, the genetic frequencies of *Rht-D1b*, *Rht8* and *Rht-B1b* were 45.5%, 46.8% and 24.5%, respectively (Zhang et al., 2006).

However, the widespread distribution of single dwarf source is not beneficial to the sustainable development of wheat breeding, which reduces the diversity of dwarf sources and the safety of wheat production. Wheat dwarf breeding requires diversified dwarf genes. Therefore, it is important to identify new wheat dwarf genes and determine their application value for wheat dwarf breeding. The aim of this study was to identify the novel wheat dwarfing site *Rht-yz*, and to clarify its effects on wheat agronomic traits by measuring plant height, internode traits, lodging resistance traits, yield and grain traits.

Materials and methods

Materials and field experiment

The Yanzhan 4110 (followed as Yanzhan) and natural dwarf mutants of Yanzhan 4110 (followed as Yanzhan mutation) provided by Dong Puhui's research group at Henan University of Science and Technology (HAUST), were crossed to produce $F_{2:3}$ and $F_{3:4}$ populations, including 15 dwarf lines and 15 tall lines. Field trials were conducted in 2019-2020 and 2020-2021 at the experimental farm of the Institute of Water-Saving Agriculture for Arid Areas of China, Northwest A&F University, Yangling, Shaanxi, China. The experimental design was randomized complete block design (RCBD), and each line was planted in 2.5m² sized plots (five rows of 2 m long, 25 cm between two rows, and 6.7 cm between two plants). Each plot was repeated three times.

The GA₃ treatment was applied separately on both Yanzhan and Yanzhan mutation, and repeated three times. When wheat plants reached at stem elongation (Z31), early heading (Z51), early flowering (Z61), and grain filling (Z71) stages, GA₃ solution (100 μ M) was sprayed on the surfaces of leaves, culms and young panicles (Zadoks et al., 1974), and the control was applied with distilled water of the same volume. GA₃ treatments in field were performed referred to Duan et al. 2020a.

Genomic sequence clone

Genomic DNA was extracted by CTAB method (Clark, 1997). The genomic sequence of *Rht-B1b* was cloned with primers in Supplementary Table 1, using PCR conditions described. PCR was performed in a Peltier Thermal cycler with a 20µl volume used ApexHF HS DNA Polymerase FL (Accurate Biology, China), and the PCR program was set according to the instructions (temperature was 60°C, extension time was 3min). PCR products were distinguished by 2% agarose gel electrophoresis.

The PCR target band was purified using Universal DNA Purification Kit (Tiangen, China). After adding 'A' base at the end, the recovered product was cloned into the T vector (TAKARA pMD18-T Vector Cloning Kit). The vector was transferred into E. coli DH5 α competent cells by the 42°C heat shock method, and 950 μ l of SOC liquid medium was added to the 37°C shaking culture for 45 minutes. The product was plated in AMP-containing LB medium. After overnight incubation at 37°C, individual colonies were detected and sequenced. Primer synthesis and clone sequencing were performed at Tsingke Biology Technique (Beijing, China).

DNA extraction based on bulked segregation analysis

Yanzhan and Yanzhan mutation F_2 population used to create tall and dwarf mixed pools. Genomic DNA was extracted from the second leaf below the spike of 20 tall and 20 dwarf plants at the filling stage. Equal amounts of tall plant genomic DNA were mixed into tall mixed pools, and equal amounts of dwarf plants genomic DNA were mixed into dwarf mixed pools. Combined with 660K chip (Affymetrix), SNP typing was detected (CapitalBio Technology, Beijing, China).

Plant height and internode length traits

Plant height and the internode length were investigated at the mature stage. Internode diameter and wall thickness were measured with Vernier scale during the grain filling stage (Du et al., 2018), with five replicates.

With reference to the method of Zhao et al. (2021), the internodes strength tester was used to measure the mechanical strength of the 4th and 5th basal internodes. The center of gravity height and fresh weight of the main stem were also measured. The formula for calculating the lodging resistance index is as follows:

$$LRI = MS \div (CGH \times FW)$$

LRI: Lodging Resistance Index; MS: Mechanical Strength (N); CGH: Center of Gravity Height (m); FW: Fresh Weight(g).

Yield traits

The standard and method for yield characters were referred to "Physiological Breeding II: A field Guide to Wheat Phenotyping" (Reynolds, 2012). At maturity, tillers plant⁻¹, spike length, spikelet number spike⁻¹, grains number spike⁻¹, 1000-kernel weight, biomass plant⁻¹, yield plant⁻¹ and harvest index were measured using a random sample of 20 individual plants.

Grain traits

After threshing and drying, grain length, width and diameter were measured using the SC-G automatic seed analyzer (Wanshen, China). Grain quality traits, including water content, crude protein, wet gluten, water absorption, starch, stable time and formation time were measured using a Diode Array 7250 near-infrared reflectance (NIR) spectrometer (Perten Instrument AB, Sweden).

Data analysis

The formula for the difference between dwarf and tall plants as follow:

Difference (%) = $(Value_{Dwarf} - Value_{Tall}) \div Value_{Tall} \times 100\%$

Data was analyzed using analysis of variance (ANOVA) for a ttest of the difference between the dwarf and tall lines using IBM SPSS Statistics 23.0. * means significant at p < 0.05. ** means significant at p < 0.01, none means not significant at p > 0.05. Oligo 7 was used for primer design. Origin 2018 and Microsoft Office Excel 2016 were used to complete the figures and tables.

Results

Discovery of a novel dwarfing site

Exogenous GA₃ treatment of Yanzhan and Yanzhan mutation

Compared to Yanzhan, the height of the mutant was reduced by 22.5 cm (29.8%). At the jointing stage, Yanzhan and Yanzhan mutants were treated with exogenous GA_3 . The results showed that exogenous GA_3 had no significant effect on plant height, length, diameter and wall thickness of the fourth and fifth stems of Yanzhan and Yanzhan mutants (Table 1). This indicated that the type of Yanzhan mutation was gibberellin insensitive and the mutation was not responsive to exogenous gibberellin.

BSA (bulked segregation analysis) of F_2 population

The homozygous differential SNPs of dwarf and tall bulks were analyzed using BSA and 660K SNP array. The results showed that 92 high quality SNPs were detected, of which 36 (39.1%) were found on chromosome 4B, and the rest were distributed on other chromosomes. And the differential SNPs on chromosome 4B were mainly enriched on the short arm, of which 28 (77.8%) were within 30-33 Mb (Figure 1A). This indicated that the mutation causing dwarf plant height was likely to be located in this interval.

Genomic sequence clone of Rht-B1b

The reported dwarfing gene *Rht-B1b* was found within this interval by searching for high confidence genes in 4BS: 30-33 MB according to

TABLE 1 Effect of exogenous GA3 treatment on plant height and internodes traits of YZ and M.

Phenotypes		YZ	YZ-GA ₃	Difference	М	M-GA ₃	Difference
Plant height (cm)	Plant height Range (95%)	75.4±0.49 74.3~76.6	74.6±0.43 73.6~75.6	-0.8 (-1.1%)	52.9±0.45 51.9~53.9	54.7±0.45 53.7~55.8	1.8 (3.4%)
Internode length (cm)	I4 Range (95%)	7.5±0.12 7.2~7.8	7.5±0.13 7.0~7.9	0.0	4.8±0.26 4.1~5.4	5.4±0.27 4.6~6.3	0.6 (12.5%)
	15 Range (95%)	7.1±0.31 6.3~7.8	8.2±0.06 7.9~8.4	1.1 (15.5%)	3.1±0.14 2.7~3.5	3.5±0.25 3.2~4.7	0.4 (12.9%)
Internode diameter (mm)	I4 Range (95%)	4.3±0.03 4.3~4.4	4.3±0.04 4.2~4.4	0.0	4.6±0.04 4.5~4.7	4.7±0.05 4.6~4.8	0.1 (2.2%)
	15 Range (95%)	4.5±0.03 4.4~4.5	4.3±0.03 4.3~4.4	-0.2 (-4.4%)	4.5±0.09 4.1~4.4	4.4±0.09 4.3~4.6	-0.1 (-2.3%)
Internode thickness (mm)	I4 Range (95%)	0.8±0.03 0.7~0.8	0.7±0.02 0.7~0.8	-0.1 (-12.5%)	1.0±0.05 0.9~1.1	1.0±0.03 0.9~1.1	0.0
	I5 Range (95%)	0.8±0.03 0.7~0.9	0.8±0.02 0.7~0.8	0.0	1.1±0.05 0.9~1.1	1.1±0.04 1.2~1.3	0.0

YZ, Yanzhan; M, Yanzhan mutation. The data is mean ± SE (standard error). Different is effect of exogenous GA₃. Difference = (Treatment – Control)/Control*100%.



Bulked segregation analysis (BSA) results and sequencing of *Rht-yz*. (A) Results of BSA analysis in F_2 population. The 4B chromosome has 36 SNPs, which concentrates on 30-33MB. (B) Results of *Rht-B1b* sequence. The * pointss the mutant site of *Rht-B1b*, which had no different between Yanzhan and mutation lines. YZ, Yanzhan; M, Yanzhan mutation.

IWGSC RefSeq v1.1, in combination with gene annotation (Table 2). In order to determine the genomic sequence of *Rht-B1b*, primers were designed (Supplementary Table 1). The coding region sequence was cloned using PCR technology and monoclonal sequencing. The sequence alignment revealed that the mutation site of *Rht-B1b* was the same in Yanzhan and Yanzhan mutation, this indicated that the mechanism of plant height reduction of this dwarfing site may be different from that of *Rht-B1b* (C-T mutation) (Figure 1B).

Effect of *Rht-yz* on plant height and internodes traits

Rht-yz significantly reduced plant height. The height of Yanzhan was approximately 75.4 cm, and the height of the Yanzhan mutant carrying Rht-yz was reduced by 22.5 cm (29.8%). In addition, Rht-yz reduced plant height by 36.4% and 33.4% in the F_{2:3} and F_{3:4} populations, respectively (Figure 2). Rht-yz also had significant effects on internode length, diameter and thickness in wheat. Both Yanzhan and Yanzhan mutations had 5 internodes, and the Yanzhan mutations had significantly shorter internode lengths compared with Yanzhan (Figures 3A, B), in which the peduncle was significantly shortened by 8.0 cm (31.3%), I2, I3, I4 and I5 were significantly shortened by 3.9 cm (25.8%), 4.5 cm (38.8%), 2.7 cm (36.0%) and 4.0 cm (56.3%), respectively. In addition, compared with Yanzhan the diameters of I2, I3, I4, and I5 with Yanzhan mutations were significantly increased by 0.4 mm (9.8%), 0.4 mm (9.1%), 0.3 mm (7.0%), and 0.2 mm (4.4%), respectively, the thickness of I3, I4 and I5 was significantly increased by 0.2 mm (33.3%), 0.2 mm (25.0%) and 0.3 mm (37.5%) respectively (Figure 3C and Table 3).

Similar results were observed in the $F_{2:3}$ and $F_{3:4}$ populations, where *Rht-yz* significantly reduced plant height and internode length. *Rht-yz* also significantly increased the diameter of I2 and I5 and the wall thickness of I3, I4 and I5 (Table 3).

Effect of Rht-yz on lodging resistant traits

This study focused on the lodging resistance index of the basal stem (I4 and I5), which are most relevant for lodging resistance TABLE 2 Names of dwarf genes and positional information of chromosomes.

Name	Chromosome position
Rht1(RhtB1b)	4BS
Rht2(RhtD1b)	4DS
Rht3(RhtB1c)	4BS
Rht4	2BL
Rht5	3BS
Rht6	4D
Rht7	2AS
Rht8	2DS
Rht9	5AL
Rht10 (RhtD1c)	4DS
Rht11 (RhtB1e)	4BS
Rht12	5AL
Rht13	7BS
Rht14	6AS
Rht15	6A
Rht16	6AS
Rht17 (RhtB1p)	4BS
Rht18	6AS
Rht19	_
Rht20	_
Rht21	2A
Rht22	7AS
Rht23	5DL
Rht24	6AL
Rht25	6AS



FIGURE 2

Plant height of Yanzhan mutation population. (A) Difference of plant height between dwarf and tall lines. (B) The mature plant height of Yanzhan and Yanzhan mutation. ** significant at p<0.01.



FIGURE 3

Internodes traits of Yanzhan and Yanzhan mutation at the filling stage. (A) The main stem. (B) Difference of internode length. (C) Cross section of the middle of each internode. ** significant at p<0.01. P, peduncle. 12, 13, 14 and 15, the second internode under spike, the third internode under spike, the fourth internode under spike, the fifth internode under spike.

Internode traits	9	М	ΥZ	Difference	F _{2:3} dwarf	F _{2:3} tall	Difference	F _{3:4} dwarf	F _{3:4} tall	Difference
Length (cm)	Р	17.7±0.58	25.7±0.42	-8.0 (-31.1%)**	16.9±0.47	27.7±0.73	-10.8 (-39.0%)**	18.0±0.69	28.1±1.03	-10.1 (-35.9%) **
	12	11.2±0.28	15.1±0.44	-3.9 (-25.8%)**	8.8±0.18	17.4±0.22	-8.6 (-49.4%)**	9.7±0.33	14.3±0.35	-4.6 (-32.2%) **
	13	7.1±0.10	11.6±0.11	-4.5 (-38.8%)**	6.3±0.08	13.1±0.35	-6.8 (-51.8%)**	6.4±0.26	13.9±0.51	-7.5 (-54.0%) **
	I4	4.8±0.26	7.5±0.12	-2.7 (-36.0%)**	4.6±0.23	8.4±0.44	-3.8 (-45.2%)**	4.5±0.37	8.1±0.22	-3.6 (-44.4%) **
	15	3.1±0.14	7.1±0.31	-4.0 (-56.3%)**	3.9±0.24	6.4±0.22	-2.5 (-38.4%)**	4.0±0.19	6.5±0.44	-2.5 (-38.5%) **
Diameter	Р	3.3±0.03	3.4±0.03	-0.1 (-2.9%)	2.9±0.04	2.7±0.03	0.2 (7.4%)**	2.7±0.03	2.6±0.04	0.1 (3.8%)
(cm)	12	4.5±0.05	4.1±0.05	0.4 (9.8%)*	3.8±0.04	3.6±0.04	0.2 (5.6%)**	4.0±0.05	3.7±0.05	0.3 (8.1%)*
	13	4.8±0.04	4.4±0.05	0.4 (9.1%)*	4.1±0.04	4.1±0.05	0.0	4.3±0.03	4.1±0.05	0.2 (4.9%)
	I4	4.6±0.04	4.3±0.03	0.3 (7.0%)*	4.1±0.03	4.0±0.05	0.1 (2.5%)	4.2±0.06	4.0±0.04	0.2 (5.0%)*
	15	4.5±0.09	4.3±0.03	0.2 (4.4%)*	4.1±0.05	3.9±0.05	0.2 (5.1%)*	4.0±0.05	3.8±0.05	0.2 (5.3%)*
Thickness	Р	0.4±0.01	0.4±0.01	0.0	0.3±0.01	0.3±0.01	0.0	0.3±0.03	0.3±0.01	0.0
(cm)	12	0.5±0.02	0.5±0.02	0.0	0.4±0.01	0.4±0.02	0.0	0.4±0.02	0.4±0.01	0.0
	13	0.8±0.02	0.6±0.03	0.2 (33.3%)**	0.5±0.01	0.4±0.01	0.1 (25.0%) **	0.5±0.01	0.4±0.02	0.1 (25.0%)**
	I4	1.0±0.05	0.8±0.03	0.2 (25.0%)**	0.6±0.02	0.5±0.01	0.1 (20.0%) **	0.6±0.01	0.5±0.02	0.1 (20.0%)**
	15	1.1±0.05	0.8±0.03	0.3 (37.5%)**	0.7±0.02	0.5±0.01	0.2 (40.0%) **	0.6±0.03	0.5±0.03	0.1 (20.0%)**

TABLE 3 Effect of Rht-yz on internode traits.

YZ, Yanzhan; M, Yanzhan mutation. The data is mean ± SE (Standard Error). * significant at p<0.05; ** significant at p<0.01. P, peduncle. I2, I3, I4 and I5, the second internode under spike, the third internode under spike, the forth internode under spike, the fifth internode under spike.

traits (Figure 4). The results showed that Rht-yz significantly increased the mechanical strength of Yanzhan mutants I4 and I5 by 7.4% and 15.5% respectively. Meanwhile, Rht-yz significantly increased the lodging resistance index of I4 (72.7%) and I5 (90.6%) (Table 4).

Effect of *Rht-yz* on yield and yield component

In the parental materials, $F_{2:3}$ and $F_{3:4}$ populations, *Rht-yz* significantly reduced the tillers plant⁻¹, biomass plant⁻¹ and yield plant⁻¹. In addition, 1000-kernel weight and harvest index of the Yanzhan mutant and *Rht-yz* dwarf lines were also significantly reduced in the $F_{2:3}$ population. For spike traits, *Rht-yz* increased spike length, spikelet number spike⁻¹ and grain number spike⁻¹, but the difference was not significant (Table 5).

Effect of Rht-yz on grain size and quality

Grain length, width and diameter of the Yanzhan mutation were not significantly different from those of Yanzhan, but *Rht-yz* reduced grain width by 6.5% and also had a negative effect on grain plumpness (Figure 5).

Rht-yz significantly increased grain crude protein (12.8%), wet gluten (10.0%), starch (6.9%), dough stability time (70.9%) and

dough developing time (7.4%), but had no significant effect on water content and water absorption (Table 6).

Discussion

The dwarfing gene solved the lodging problem in wheat breeding and greatly increased wheat yield per unit area (Flintham et al., 2000; Hedden, 2003). However, the negative effects of many dwarfing genes on yield and other traits in wheat have led to difficulties in their application. In this study, a new dwarf wheat mutant was identified and found to carry a new dwarf gene, provisionally named *Rht-yz*. To clarify the genetic effects of this gene, the plant height and internode length, lodging resistance, yield and grain traits were investigated from Yanzhan, Yanzhan mutations, $F_{2:3}$ and $F_{3:4}$ populations.

Discovery of a novel dwarfing site

Rht-B1a is wild-type and encodes a normal DELLA protein. The *Rht-B1b* and *Rht-D1b* have single base mutations in the coding region of the DELLA protein (*Rht-B1b* has a C-T mutation and *Rht-D1b* has a G-T mutation in the coding region), which ultimately leads to inactivity of the DELLA protein and reduced plant height (Peng et al., 1999; Pearce et al., 2011). The *Rht-B1c* transcript has a 90bp insertion, while



Rht-B1d and *Rht-B1e* are prematurely terminated by single base (AT) mutations (Pearce et al., 2011), while *Rht-B1k* has displacement mutations in the VHIID domain to produce abnormal DELLA proteins. In this study, it was found that the number of differential SNPs enriched in the chromosome 4B was higher (36) and that there were 28 differential SNPs in the 30-33 MB interval, forming a differential SNPs enrichment peak. However, the number of differential SNPs was less and the distribution was scattered on other chromosomes, which did not form an obvious differential SNPs enrichment peak. Therefore, we can only obtain one interval (4B: 30-33MB) that regulates plant height, which contains a dwarf gene, *Rht-B1b*. The mutation site of the *Rht-B1b* did not differ between the Yanzhan and Yanzhan

TABLE 4	Effect of	Rht-yz on	internodes	lodging	resistance	index	of
basal inte	rnodes.						

Materials	Brea streng	aking gth (N)	Internodes lodging resistance index			
	14	15	14	15		
М	24.8 ± 2.51	32.9 ± 2.63	7.6 ± 0.62	10.1 ± 0.83		
YZ	23.1 ± 2.24	27.7 ± 3.14	4.4 ± 0.30	5.3 ± 0.42		
Difference	1.7(7.4%) *	4.3(15.5%) *	3.2(72.7%) **	4.8(90.6%) **		

The data is mean ± SE (Standard Error). * significant at p< 0.05; ** significant at p<0.01.

mutations. Therefore, *Rht-yz* is a new dwarfing site that regulates plant height and its mechanism of action is different from that of *Rht-B1b* (C-T mutation), but it is not clear whether it affects the *Rht-B1a* (*TraesCS4B02G043100*) or other genes.

Rht-yz improved the lodging resistance in wheat

Improving lodging resistance in wheat is one of the most important factors in increasing wheat yields. Many pre-Green Revolution wheat varieties were replaced due to lodging resistance defects. The wheat dwarf genes *Rht-B1b* and *Rht-D1b* reduced plant height by 23%, *Rht-B1b+Rht-D1b* reduced plant height by 50% (Butler et al., 2005). The dwarfing genes *Rht8* and *Rht12* reduced plant height by 8.0% and 40%, respectively (Tang et al., 2009; Chen et al., 2013). In this study, *Rht-yz* reduced plant height by approximately 29.8%, which was a moderate strength effect.

Studies have shown that the length of I4 and I5 is significantly correlated with lodging resistance in wheat (Kashiwagi et al., 2005), and lodging resistance is positively correlated with internode diameter, thickness and fullness (Zhao et al., 2021). Studies have shown that *Rht18* increased basal internode strength (Yang et al., 2015), while *Rht13* decreased basal internode diameter (Wang et al., 2014). In addition, the height of center of gravity is also an important factor in the lodging resistance index. In this study, *Rht-yz* mainly improved the lodging resistance by reducing the plant height and increasing the diameter, wall thickness and mechanical strength of the basal internode, which has important application value for dwarf breeding in wheat.

Rht-yz had negative effect on part of the yield components

The study reported that *Rht3* increased tiller number (Zhang et al., 1995), *Rht5* reduced spikelet number and grain number spike⁻¹ in two different genetic backgrounds (Daoura et al., 2014), and *Rht8* had little effect on yield traits (Rebetzke et al., 2012). In this study, *Rht-yz* had a significant negative effect on tiller number plant⁻¹, biomass plant⁻¹ and yield plant⁻¹, but no significant effect on harvest index.

An unexpected disease (wheat stripe rust) during the 2019-2020 growing season led to a yield reduction in the $F_{2:3}$ population. During

Genotype	м	ΥZ	Difference	F _{2:3} dwarf	F _{2:3} tall	Difference	F _{3:4} dwarf	$F_{3:4}$ tall	Difference
Tillers plant ⁻¹	6.5±0.43	9.0±0.82	-2.5 (-27.7%) **	5.8±0.83	7.6±1.43	-1.8 (-23.6%) *	7.9±0.45	11.4±0.45	-3.5 (-30.7%) **
Spike Length (cm)	9.7±0.35	9.4±0.21	0.3 (3.2%)	9.9±0.17	9.7±0.20	0.2 (2.4%)	9.7±0.51	10.0±0.27	-0.3 (-3.0%)
Spikelet number plant ⁻¹	23.9±0.93	21.2 ± 1.60	2.7 (12.7%)	19.6±0.45	17.6±0.40	2.0 (11.4%)	22.9±0.18	24.6±0.34	-1.7 (-6.9%)
Grain number plant ⁻¹	38.5±0.96	34.0±1.69	4.5 (13.2%)	33.5±1.40	34.2±1.15	-0.7 (-2.2%)	37.5±1.80	38.0±1.92	-0.5 (-1.3%)
1000-kernel weight (g)	39.2±3.50	43.3±4.26	-4.1 (-9.5%)	32.3±1.54	40.4±3.81	-8.1 (-20.1%)	43.9±0.53	48.2±1.22	-4.3 (-8.9%)
Biomass plant ⁻¹ (g)	20.2±1.05	34.8±3.24	-14.6 (-42.0%) **	22.7±0.60	36.2±2.09	-13.5 (-37.2%) **	30.3±2.68	42.4±4.23	-12.1 (-28.5%) **
Yield plant ⁻¹ (g)	7.1±0.60	12.5±1.55	-5.4 (-43.2%) **	6.4±0.37	13.3±1.00	-6.9 (-52.0%) **	9.3±1.15	13.8±1.46	-4.5 (-32.6%) **
Harvest index	0.35±0.02	0.39±0.01	-0.04 (-10.25%)	0.26±0.01	0.37±0.02	-0.1 (-29.6%) **	0.31±0.03	0.33±0.02	-0.02 (-6.1%)

TABLE 5 Effect of Rht-yz on yield traits.

The data is mean ± SE (Standard Error). * significant at p< 0.05; ** significant at p<0.01.

the 2020-2021 growing season, no signs of disease were observed. Therefore, the effect of $F_{3:4}$ on yield traits is closer to the actual effect of *Rht-yz*. The growth period of the Yanzhan mutation was 5-7 days later than that of Yanzhan, suggesting that *Rht-yz* may be involved in regulating the growth and development of wheat. Studies have also shown that *Rht4+Rht-B1b* has higher yield and harvest index under different moisture conditions (Liu et al., 2017). *Rht5* reduced wheat yield, but *Ppd-D1* effectively eliminated the negative effects of *Rht5* on yield and growth by advancing the growth period (Chen et al., 2018). These studies remind us that the negative effects of *Rht-yz* on yield can also be eliminated by introducing other new genes.

Rht-yz improved wheat grain quality

Protein content, starch content and gluten content in wheat have an important impact on the promotion and application of wheat varieties. The quality of flour is mainly determined by protein and starch content (Ahmed, 2015). The dwarf gene *Rht-B1c* significantly increased the protein content, *Rht14* had a negative effect on quality traits (Duan et al., 2020b), and *Rht15* increased the protein and wet gluten content (Zhao et al., 2021). In this study, *Rht-yz* significantly increased crude protein, wet gluten and starch content. Therefore, *Rht-yz* is of great significance for improving the grain quality in wheat.



Traits	М	YZ	Difference
Water content (%)	11.4±0.01	11.4±0.01	0.0
Crude protein (%)	15.0±0.17	13.3±0.06	1.7(12.4%)**
Wet gluten (%)	31.9±0.08	29.0±0.05	3.0(10.2%)**
Water absorption (%)	67.4±0.06	66.3±0.03	1.1(1.7%)
Starch (%)	60.4±0.31	56.5±0.20	3.8(6.8%)**
Dough stability time (min)	13.5±0.09	7.9±0.09	5.6(71.2%)**
Dough developing time (min)	2.9±0.03	2.7±0.03	0.2(8.2%)**

TABLE 6 Effect of *Rht-yz* on grain quality.

The data is mean ± SE (Standard Error). ** significant at p<0.01.

Conclusions

In this study, a new dwarfing site, *Rht-yz*, was identified in wild mutants that could improve lodging resistance by reducing plant height, increasing basal internode diameter, wall thickness and mechanical strength, and had a positive effect on grain quality traits. In addition, *Rht-yz* had a negative effect on some yield traits. Therefore, the rational use of the dwarf gene *Rht-yz* has some utility for lodging resistance breeding in wheat, but attention should be paid to improving its negative effects.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

Author contributions

JH: Investigation, Writing – original draft, Writing – review & editing. ZZ: Investigation, Writing – review & editing. XF: Investigation, Writing – review & editing. YZ: Investigation, Writing – review & editing. MA: Writing – review & editing. LM: Data curation, Writing – review & editing. YH: Data curation, Writing – review & editing. YL: Data curation, Writing – review & editing. YZ: Data curation, Writing – review & editing. LZ: Data curation, Writing – review & editing. DS: Data curation, Writing –

References

Ahmed, M. (2015). Response of spring wheat (*Triticum aestivum* L.) quality traits and yield to sowing date. *PloS One* 10 (4), e0126097. doi: 10.1371/journal.pone.0126097

Butler, J. D., Byrne, P. F., Mohammadi, V., Chapman, P. L., and Haley, S. D. (2005). Agronomic performance of alleles in a spring wheat population across a range of moisture levels. *Crop Sci.* 45, 939. doi: 10.2135/cropsci2004.0323

Casebow, R., Hadley, C., Uppal, R., Addisu, M., and Gooding, M. (2016). Reduced height (Rht) alleles affect wheat grain quality. *PloS One* 11, e0156056. doi: 10.1371/journal.pone.0156056

review & editing. CF: Writing – review & editing, Data curation. Y-GH: Funding acquisition, Supervision, Writing – review & editing. LC: Funding acquisition, Supervision, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was financially supported by the Basic Research Program of natural sciences of Shaanxi province (2023-JC-ZD-08), ANSO Joint Research and cooperation project (ANSO-CR-KP-2022-05), the National Natural Science Foundation of China (32171991), Key Research and Development Program of Shaanxi (2022NY-189).

Conflict of interest

Author YH was employed by the company Jiushenghe Seed Industry Co., Ltd. Author YL was employed by the company Yangling Digital Agricultural Technology Co., Ltd. Author YY was employed by the company Shaanxi Grain and Agriculture Group Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2024.1338425/ full#supplementary-material

Chen, L., Phillips, A. L., Condon, A. G., Parry, M. A. J., and Hu, Y.-G. (2013). GAresponsive dwarfing gene *Rht12* affects the developmental and agronomic traits in common bread wheat. *PloS One* 8, e62285. doi: 10.1371/journal.pone.0062285

Chen, L., Yang, Y., Cui, C., Lu, Q., Du, Y., Su, R., et al. (2018). Effects of Vrn-B1 and Ppd-D1 on developmental and agronomic traits in Rht5 dwarf plants of bread wheat. Field Crop Res. 219, 24–32. doi: 10.1016/j.fcr.2018.01.022

Clark, M. S. (1997). Plant Molecular Biology — A Laboratory Manual || Genomic DNA Isolation, Southern Blotting and Hybridization. Springer, Berlin, Heidelberg: Springer, 3– 53. doi: 10.1007/978-3-642-87873-2 Daoura, B. G., Chen, L., Du, Y., and Hu, Y. G. (2014). Genetic effects of dwarfing gene *Rht5* on agronomic traits in common wheat (Triticum aestivum L.) and QTL analysis on its linked traits. *Field Crops Res.* 156, 22–29. doi: 10.1016/j.fcr.2013.10.007

Derkx, A. P., Harding, C., Miraghazadeh, A., and Chandler, P. M. (2017). Overgrowth (Della) mutants of wheat: development, growth and yield of intragenic suppressors of the Rht-B1c dwarfing gene. *Funct. Plant Biol.* 44, 525. doi: 10.1071/ FP16262

Du, Y., Liang, C., Wang, Y., Yang, Z., Saeed, I., Daoura, B. G., et al. (2018). The combination of dwarfing genes *Rht4* and *Rht8* reduced plant height, improved yield traits of rainfed bread wheat (Triticum aestivum L.). *Field Crops Res.* 215, 149–155. doi: 10.1016/j.fcr.2017.10.015

Duan, S., Zhao, Z., Qiao, Y., Cue, C., Condon, A. G., Chen, L., et al. (2020a). Vigorous responsiveness of dwarf gene *Rht14* to exogenous GA3 evaluated on important morphological and agronomic traits in durum wheat. *Agron. J.* 112 (6), 5033–5044. doi: 10.1002/agj2.20409

Duan, S., Zhao, Z., Qiao, Y., Cui, C., Morgunov, A., Condon, A. G., et al. (2020b). GAR dwarf gene Rht14 reduced plant height and affected agronomic traits in durum wheat (Triticum durum) - ScienceDirect. *Field Crops. Res.* 248. doi: 10.1016/ j.fcr.2020.107721

Evans, A. (2008). Feeding the ten billion. World Today 64, 4-6.

Flintham, J. E., Angus, W. J., and Gale, M. (2000). Heterosis, overdominance for grain yield, and alpha-amylase activity in F1 hybrids between near-isogenic Rht dwarf and tall wheats. *J. Agric. Sci.* 129, 371–378. doi: 10.1017/S0021859697004899

Gale, M. D., Youssefian, S., and Russell, G. E. (1985). "Dwarfing genes in wheat." In *Progress in Plant Breeding-1* (Elsevier), pp. 1–35. doi: 10.1016/b978-0-407-00780-2.50005-9

Guedira, M., Brown-Guedira, G., Sanford, D. V., Sneller, C., Souza, E., and Marshall, D. (2010). Distribution of rht genes in modern and historic winter wheat cultivars from the eastern and central USA. *Crop Sci.* 50, 1811–1822. doi: 10.2135/cropsci2009.10.0626

Hayat, H., Mason, R. E., Lozada, D. N., Acuna, A., and Miller, R. G. (2019). Effects of allelic variation at *Rht-B1* and *Rht-D1* on grain yield and agronomic traits of southern US soft red winter wheat. *Euphytica* 215, 172. doi: 10.1007/s10681-019-2478-2

Hedden, P. (2003). The genes of the green revolution. Trends Genet. 19, 5-9. doi: 10.1016/s0168-9525(02)00009-4

Kashiwagi, T., Sasaki, H., and Ishimaru, K. (2005). Factors responsible for decreasing sturdiness of the lower part in lodging of rice (*Oryza sativa* L.). *Plant Product. Sci.* 8, 166–172. doi: 10.1626/pps.8.166

Liu, Y., Zhang, J., Hu, Y. G., and Chen, J. (2017). Dwarfing genes *Rht4* and *Rht-B1b* affect plant height and key agronomic traits in common wheat under two water regimes. *Field Crops Res.* 204, 242–248. doi: 10.1016/j.fcr.2017.01.020

Pearce, S., Saville, R., Vaughan, S. P., Chandler, P. M., Wilhelm, E. P., Korolev, A., et al. (2011). Molecular characterisation of *Rht1* dwarfing genes in hexaploid wheat (Triticum aestivum). *Plant Physiol.* 157 (4), 1820–31. doi: 10.1104/pp.111.183657

Peng, J., Richards, D. E., Hartley, N. M., Murphy, G. P., Devos, K. M., Flintham, J. E., et al. (1999). "Green revolution" genes encode mutant gibberellin response modulators. *Nature* 400, 256. doi: 10.1038/22307

Rebetzke, G. J., Ellis, M. H., Bonnett, D. G., Mickelson, B., Condon, A. G., and Richards, R. A. (2012). Height reduction and agronomic performance for selected gibberellin-responsive dwarfing genes in bread wheat (Triticum aestivum L.). *Field Crops Res.* 126, 87–96. doi: 10.1016/j.fcr.2011.09.022

Reynolds, M. (2012). Physiological Breeding II: A field Guide to Wheat Phenotyping, Physiological Breeding II: A field Guide to Wheat Phenotyping. Mexico: CIMMYT.

Richards, R. A. (1992). The effect of dwarfing genes in spring wheat in dry environments. I. Agronomic characteristics. *Aust. J. Agric. Res.* 43, 517-527. doi: 10.1071/AR9920517

Tang, N., Jiang, Y., He, B.-R., and Hu, Y.-G. (2009). The effects of dwarfing genes (*Rht-B1b*, rht-D1b, and *rht8*) with different sensitivity to GA_3 on the coleoptile length and plant height of wheat. *Agric. Sci. China.* 9, 11. doi: 10.1016/S1671-2927(08)60310-7

Wang, Y., Liang, C., Du, Y., Yang, Z., Condon, A. G., and Hu, Y. G. (2014). Genetic effect of dwarfing gene *Rht13* compared with *Rht-D1b* on plant height and some agronomic traits in common wheat (Triticum aestivum L.). *Field Crops Res.* 162, 39–47. doi: 10.1016/j.fcr.2014.03.014

Yang, Z.Y., Zheng, J.C., Liu, C.Y., Wang, Y.S., Condon, A.G., Chen, Y.F., et al. (2015). Effects of the GA-responsive dwarfing gene Rht18 from tetraploid wheat on agronomic traits of common wheat. *Field Crops Res.* 183, 92–101. doi: 10.1016/j.fcr.2015.07.028

Zadoks, J. C., Chang, T. T., and Konzak, F. C. (1974). A decimal code for the growth stages of cereals. *Weed Res.* 14, 415–421. doi: 10.1111/j.1365-3180.1974.tb01084.x

Zhang, R., Lu, J., Ma, G., Wu, Y., Min, F., and Zhao, Y. (1995). Effects of *rht3* gene and 4B-chromosome with *rht3* on photosynthetic carbon-assimilation of wheat. *Acta GENE. SINICA.* 22, 264–271.

Zhang, X., Yang, S., Zhou, Y., He, Z., and Xia, X. (2006). Distribution of the *Rht-B1b*, *Rht-D1b* and *Rht8* reduced height genes in autumn-sown Chinese wheats detected by molecular markers. *Euphytica* 152, 109–116. doi: 10.1007/s10681-006-9184-6

Zhao, Z., Duan, S., Hao, J., Cui, C., and Chen, L. (2021). The dwarf gene *Rht15* improved lodging resistance but differentially affected agronomic and quality traits in durum wheat. *Field Crops Res.* 263, 108058. doi: 10.1016/j.fcr.2021.108058