



OPEN ACCESS

EDITED AND REVIEWED BY
Mark Blyth,
University of East Anglia, United Kingdom

*CORRESPONDENCE
Zhi-Yan Du
✉ duz@hawaii.edu

RECEIVED 17 July 2023
ACCEPTED 22 August 2023
PUBLISHED 05 September 2023

CITATION
Du Z-Y, Bhat WW, Kai G, Khozin-Goldberg I, Yu X-H, Zienkiewicz A and Zienkiewicz K (2023) Editorial: Metabolic engineering of valuable compounds in photosynthetic organisms.
Front. Plant Sci. 14:1260454.
doi: 10.3389/fpls.2023.1260454

COPYRIGHT
© 2023 Du, Bhat, Kai, Khozin-Goldberg, Yu, Zienkiewicz and Zienkiewicz. This is an open-access article distributed under the terms of the [Creative Commons Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Editorial: Metabolic engineering of valuable compounds in photosynthetic organisms

Zhi-Yan Du^{1*}, Wajid Waheed Bhat², Guoyin Kai³,
Inna Khozin-Goldberg⁴, Xiao-Hong Yu⁵,
Agnieszka Zienkiewicz⁶ and Krzysztof Zienkiewicz⁶

¹Department of Molecular Biosciences and Bioengineering, University of Hawaii at Manoa, Honolulu, HI, United States, ²Plant Biotechnology and Agrotechnology Division, Council of Scientific and Industrial Research-Indian Institute of Integrative Medicine (CSIR-IIIM), Jammu, India, ³Laboratory of Medicinal Plant Biotechnology, College of Pharmacy, Zhejiang Chinese Medical University, Hangzhou, Zhejiang, China, ⁴French Associates Institute for Agriculture and Biotechnology of Drylands, Jacob Blaustein Institutes for Desert Research, Ben-Gurion University of the Negev, Sede Boqer, Israel, ⁵Biology Department, Brookhaven National Laboratory, Upton, NY, United States, ⁶Centre for Modern Interdisciplinary Technologies, Nicolaus Copernicus University in Toruń, Toruń, Poland

KEYWORDS

metabolic engineering, synthetic biology, plants and algae, natural products, secondary metabolites, genetic modification, breeding

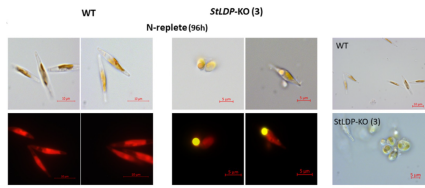
Editorial on the Research Topic

Metabolic engineering of valuable compounds in photosynthetic organisms

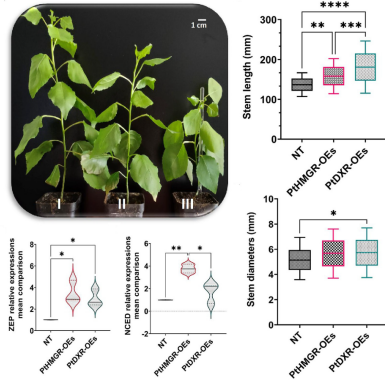
Photosynthetic organisms, including plants and algae, possess a remarkable ability to harness carbon dioxide and solar energy, enabling them to produce a vast array of complex compounds such as phenolic acids (Zhou et al., 2021), terpenes (Miller et al., 2020), unsaturated fatty acids (Kokabi et al., 2020; Gan et al., 2022), and other lipid products (Zienkiewicz and Zienkiewicz, 2020). This inherent capability positions them as highly promising platforms for the sustainable production of valuable biomolecules. While the industrial application of photosynthetic organisms in synthetic biology is not as advanced as that of model heterotrophs or mammalian systems, their significance as primary contributors to global biomass can be further developed. In fact, they are increasingly emerging as key players in the booming field of synthetic bioproducts, driven by advancements in genome editing tools and other innovative technologies. As we explore and exploit the potential of photosynthetic organisms, we open up exciting possibilities for the production of environmentally friendly and renewable biomaterials that can address pressing societal and ecological challenges.

This Research Topic includes eight original research and two review articles, with a special focus on the metabolic engineering of valuable biomaterials in plants and algae. Taparia et al., developed modular CRISPR/Cas9 constructs for the model diatom *Phaeodactylum tricornerutum* that allow the multiplexed targeting and creation of marker-free genome-edited lines. The system was used to knock out *StLDP*, the gene encoding Stramenopile-type lipid droplet protein essential for lipid droplet biogenesis (Figure 1). Mellor et al. expressed human P450s in tobacco chloroplasts to produce indican, suggesting a strategy for producing high-value chemicals or drug metabolites in photosynthetic organisms (Figure 1). Another research article investigated the biosynthesis of isoprenoids in poplar, and revealed that the 3-hydroxy-3-methylglutaryl-CoA reductase (HMGR) and 1-deoxy-D-

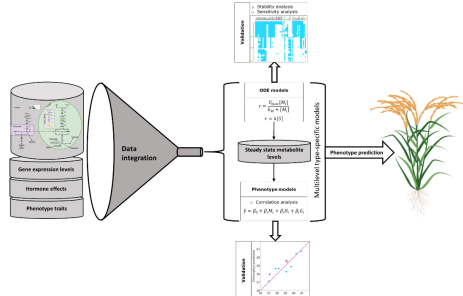
A CRISPR/Cas9 editing in *Phaeodactylum tricornutum* targeting the *StLDP* gene



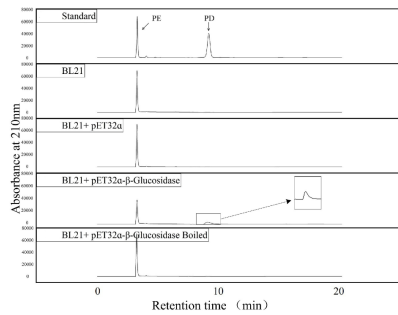
C Phenotypic changes by overexpressing *PtHMGR* and *PtDXR* in *Populus trichocarpa*



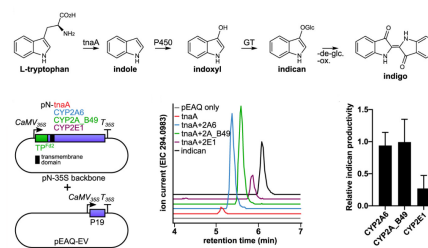
E Pullulanase and isoamylase in starch catabolism of *Manihot esculenta* Crantz



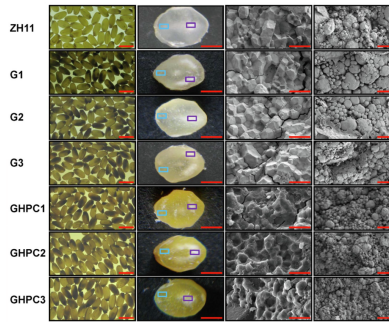
G *Platycodon grandifloras* β -glucosidase converts glycosylated platycoside E to Platycodin D



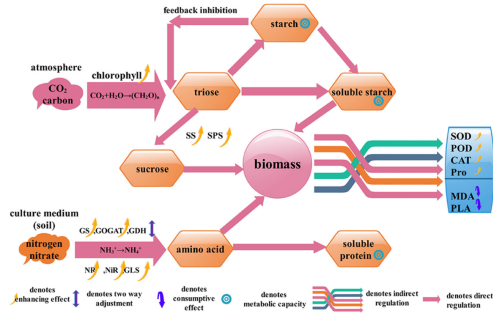
B Indigo pathway introduced into tobacco (*Nicotiana benthamiana*) chloroplasts



D Endosperm phenotypes of transgenic rice grains



F Regulation of biomass accumulation in *Salvia miltiorrhiza*



H Overexpressing *Onobrychis viciifolia* MYBPA2 in *Medicago sativa* hairy roots



FIGURE 1

Overview of the original research articles in this Synthetic Biology Research Topic. Asterisks indicate statistically significant differences, * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$

xylulose5-phosphate reductoisomerase (DXR) play important roles in regulating the genes in methylerythritol phosphate (MEP) and mevalonic acid (MVA) pathways and isoprenoids made from the MEP and MVA pathways (Figure 1) (Movahedi et al.). Li et al. produced carotenoids in rice (*Oryza sativa*) endosperm by

overexpressing rice GOLDEN2-LIKE (*OsGLK*) transcription factor and *OsGLK* with three other carotenogenic genes, *tHMG1* (truncated *Saccharomyces cerevisiae* 3-hydroxy-3-methylglutaryl-CoA reductase), *ZmPSY1* (*Zea mays* L. phytoene synthase), and *PaCrtI* (*Pantoea ananatis* phytoene desaturase), to improve the nutritional

composition of rice (Figure 1). Another research article in rice developed models by multilevel mathematical modeling using the data from rice lines with genome modification in MVA pathways, providing tools that can help prioritize metabolic engineering strategies for specific metabolic goals through exogenous pathways (Figure 1) (Basallo et al.). In perennial herbs, Wang et al. identified physiological/biochemical indicators, such as enzyme activities of glutamine synthetase (GS), glutamate synthase (GLS), glutamate dehydrogenase (GDH), peroxidase (POD), and catalase (CAT), were related to biomass accumulation in *Salvia miltiorrhiza* (Figure 1); Su et al. characterized the β -glucosidase in *Platycodon grandifloras*, which can convert glycosylated platycoside E to Platycodin D *in vitro* (Figure 1); Jin et al. identified an MYB transcription factor OvMYBPA2 in *Onobrychis viciifolia* by transcriptome analyses and confirmed its function in the regulation of proanthocyanidins in transgenic *Medicago sativa* (Figure 1). Strand and Walker reviewed bioengineering from an energetics perspective using photosynthetic organisms for bioproducts of interest (Figure 1). Another review article discussed the recent progress in engineering fatty acids and storage lipids in various plant species and tissues and summarized an inventory of specific lipogenic factors for plant lipid products (Figure 1) (Cai et al.).

Author contributions

Z-YD: Funding acquisition, Resources, Visualization, Writing – original draft, Writing – review & editing. WB: Writing – review & editing. GK: Writing – review & editing. IK-G: Writing – review & editing. X-HY: Writing – review & editing. AZ: Writing – review & editing. KZ: Writing – review & editing.

References

- Gan, L., Park, K., Chai, J., Updike, E. M., Kim, H., Voshall, A., et al. (2022). Divergent evolution of extreme production of variant plant monounsaturated fatty acids. *Proc. Natl. Acad. Sci. U.S.A.* 119, e2201160119. doi: 10.1073/pnas.2201160119
- Kokabi, K., Gorelova, O., Zorin, B., Didi-Cohen, S., Itkin, M., Malitsky, S., et al. (2020). Lipidome remodeling and autophagic response in the arachidonic-acid-rich microalga *lobosphaera incisa* under nitrogen and phosphorous deprivation. *Front. Plant Sci.* 11, 614846. doi: 10.3389/fpls.2020.614846
- Miller, G. P., Bhat, W. W., Lanier, E. R., Johnson, S. R., Mathieu, D. T., and Hamberger, B. (2020). The biosynthesis of the anti-microbial diterpenoid leubethanol in *Leucophyllum frutescens* proceeds via an all- *cis* prenyl intermediate. *Plant J.* 104, 693–705. doi: 10.1111/tpj.14957
- Zhou, W., Li, S., Maoz, I., Wang, Q., Xu, M., Feng, Y., et al. (2021). SmJRB1 positively regulates the accumulation of phenolic acid in *Salvia miltiorrhiza*. *Ind. Crops Prod* 164, 113417. doi: 10.1016/j.indcrop.2021.113417
- Zienkiewicz, K., and Zienkiewicz, A. (2020). Degradation of lipid droplets in plants and algae—Right time, many paths, one goal. *Front. Plant Sci* 11. doi: 10.3389/fpls.2020.579019

Funding

Research of the Topic Editors is supported by the funds from NSF 2121410 (-YD), the DOE Office of Science, Office of BER, DE-SC0021369 (X-HY).

Acknowledgments

The editors would like to thank all reviewers who evaluated manuscripts and contributors to this Research Topic.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The authors declared that they were an editorial board member of Frontiers, at the time of submission. This had no impact on the peer review process and the final decision.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.