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SPECIALTY SECTION

This article was submitted to
Plant Metabolism and Chemodiversity,
a section of the journal
Frontiers in Plant Science

RECEIVED 07 July 2022

ACCEPTED 08 August 2022

PUBLISHED 15 September 2022

CITATION

Liu J, Meng J, Li R, Jiang H, Fu L, Xu T,
Zhu G-Y, Zhang W, Gao J, Jiang Z-H,
Yang Z-F and Bai L-P (2022) Integrated
network pharmacology analysis,
molecular docking, LC-MS analysis
and bioassays revealed the potential
active ingredients and underlying
mechanism of *Scutellariae radix*
for COVID-19.
Front. Plant Sci. 13:988655.
doi: 10.3389/fpls.2022.988655

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Integrated network pharmacology analysis, molecular docking, LC-MS analysis and bioassays revealed the potential active ingredients and underlying mechanism of *Scutellariae radix* for COVID-19

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Scutellariae radix ("Huang-Qin" in Chinese) is a well-known traditional herbal medicine and popular dietary supplement in the world, extensively used in prescriptions of TCMs as adjuvant treatments for coronavirus pneumonia 2019 (COVID-19) patients in China. According to the differences in its appearance, *Scutellariae radix* can be classified into two kinds: ZiQin (1~3 year-old *S. baicalensis* with hard roots) and KuQin (more than 3 year-old *S. baicalensis* with withered pithy roots). In accordance with the clinical theory of TCM, KuQin is superior to ZiQin in cooling down the heat in the lung. However, the potential active ingredients and underlying mechanisms of *Scutellariae radix* for the treatment of COVID-19 remain largely unexplored. It is still not clear whether there is a difference in the curative effect of ZiQin and KuQin for the treatment of COVID-19. In this research, network pharmacology, LC-MS based plant metabolomics, and *in vitro* bioassays were integrated to explore both the potential active components and mechanism of *Scutellariae radix* for the treatment of COVID-19. As the results, network pharmacology combined with molecular docking analysis indicated that *Scutellariae radix* primarily regulates the MAPK and NF- κ B signaling pathways *via* active components such as baicalein and scutellarin, and blocks SARS-CoV-2 spike binding to human ACE2 receptors. *In vitro* bioassays showed that

baicalein and scutellarein exhibited more potent anti-inflammatory and anti-infectious effects than baicalin, the component with the highest content in *Scutellariae radix*. Moreover, baicalein inhibited SARS-CoV-2's entry into Vero E6 cells with an IC₅₀ value of 142.50 μM in a plaque formation assay. Taken together, baicalein was considered to be the most crucial active component of *Scutellariae radix* for the treatment of COVID-19 by integrative analysis. In addition, our bioassay study revealed that KuQin outperforms ZiQin in the treatment of COVID-19. Meanwhile, plant metabolomics revealed that baicalein was the compound with the most significant increase in KuQin compared to ZiQin, implying the primary reason for the superiority of KuQin over ZiQin in the treatment of COVID-19.

KEYWORDS

Scutellariae radix, LC-MS analysis, network pharmacology, COVID-19, anti-inflammatory, anti-infective

Introduction

Coronavirus disease 2019 (COVID-19) pandemic, caused by the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), has recently resulted in a global public health crisis, and posed a huge threat and challenge to the public healthcare system. In China, traditional Chinese medicines (TCMs) have made great contributions in the battle against COVID-19 (Li L. et al., 2021; Lyu et al., 2021; Zhao et al., 2021), in combination with Western medicines. Based on a statistical analysis of usage frequency in the compositions of the TCMs prescriptions for COVID-19 in 34 provincial areas of China, it was found that the dried root of *Scutellaria baicalensis Georgi* (named as “Huang-Qin” in Chinese) is one of the most commonly used TCMs for adjuvant treatment of COVID-19 (Figures 1A,B). Due to its anti-inflammatory and anti-infectious activities, *Scutellariae radix* has been used by Chinese people for the treatment of diseases including upper respiratory tract infection and enteritis over more than 3,000 years (Dinda et al., 2017; Liao et al., 2021). Moreover, *Scutellariae radix* is also a popular functional food and dietary supplement commonly used internationally (Delerue et al., 2021).

Based on the different appearance and the theory of traditional Chinese medicine, *Scutellariae radix* is divided into two types of commercial products specification. One is “ZiQin” with complete and hard xylem from 1 ~ 3 year-old *S. baicalensis*, and the other is “KuQin” with withered and rotten xylem from more than 3 year-old *S. baicalensis* (Figure 1C and Supplementary Figure 1) (Cao et al., 2019; Ren et al., 2021). According to the clinical experience of traditional Chinese medicine, KuQin is superior to ZiQin in the treatment of pneumonia, and ZiQin has a better effect than KuQin in the treatment of enteritis (Ren et al., 2021). Recently, there have been a few reports on the anti-COVID-19 effects of the extract

of *Scutellariae radix* (Song et al., 2020, 2021; Gao et al., 2021; Liu et al., 2021). However, it remains unexplored for the potential active ingredients and underlying mechanism of *Scutellariae radix* for the treatment of COVID-19. It is still unclear whether there is a difference in the curative effect of KuQin and ZiQin for the treatment of COVID-19. This study explored potential active components and anti-COVID-19 mechanism, anti-inflammatory and anti-infectious effects of both ZiQin and KuQin based on an integrated approach of network pharmacology analysis, molecular docking, LC-MS based plant metabolomics, and pharmacological validation (Deng et al., 2020; Pan et al., 2020; Yang et al., 2020; Li Z. T. et al., 2021; Zhou et al., 2021; Zhang et al., 2022). The chemical difference and potential anti-COVID-19 effect of ZiQin and KuQin were also compared. The findings of this study were aimed to provide a scientific support and valuable guidance to the proper usage of *Scutellariae radix* for COVID-19.

Materials and methods

Reagents and standards

Deionized water was prepared by using a Millipore water purification system (Merck, Hong Kong, China). LC-MS grade acetonitrile, methanol, and formic acid were bought from Anaqua Global International Inc., Limited (United States). LPS (*Escherichia coli* 0127: B8) and dexamethasone were obtained from Sigma Chemical Co. (St. Louis, MO, United States). Chemical standards for UHPLC-Q-TOF-MS analyses were purchased from Chengdu Must Bio-Technology Co., Ltd. (Apigenin ≥ 98%, Baicalein ≥ 98%, Chrysin ≥ 98%, scutellarein ≥ 98%, Wogonin ≥ 98%,

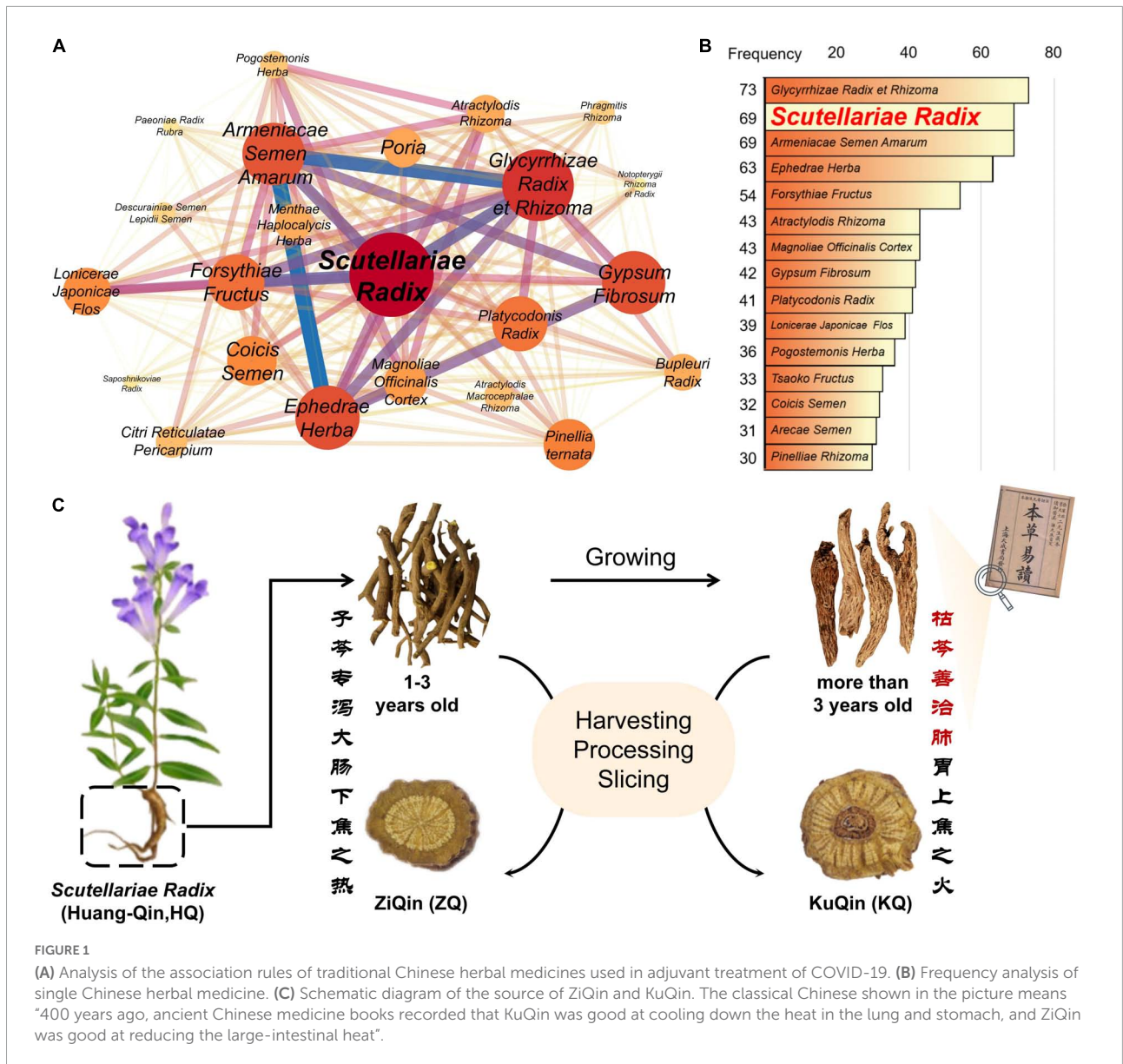


FIGURE 1
 (A) Analysis of the association rules of traditional Chinese herbal medicines used in adjuvant treatment of COVID-19. (B) Frequency analysis of single Chinese herbal medicine. (C) Schematic diagram of the source of ZiQin and KuQin. The classical Chinese shown in the picture means "400 years ago, ancient Chinese medicine books recorded that KuQin was good at cooling down the heat in the lung and stomach, and ZiQin was good at reducing the large-intestinal heat".

Oroxylin A ≥ 98%, and Baicalin ≥ 98% in purity), and Innochem (Isoscutellarein ≥ 95%, and Wogonoside ≥ 98% in purity), respectively.

at the State Key Laboratory of Quality Research in Chinese Medicine, Macau University of Science and Technology.

Sample collection of *Scutellariae radix*

The dried roots of *S. baicalensis Georgi* were collected from provinces of Inner Mongolia, Shanxi and Hebei of China during September in 2020 to March in 2021, and authenticated by Dr. Bai Li-Ping (State Key Laboratory of Quality Research in Chinese Medicine, Macau University of Science and Technology, Taipa, Macau, China). The detailed information of samples was shown in **Supplementary Figure 1** and **Supplementary Table 1**. The voucher specimens were deposited

Measurement of major components of ZiQin and KuQin by UHPLC-Q-TOF-MS analysis

According to the literature (Qiao et al., 2016), 10 mg of methanol extract powder was dissolved in 70% methanol (10 mL) with 30 min ultrasonication (40 kHz, 300 W). The sample was kept at room temperature for 5 min, and the supernatant was filtered through a 0.22 μm microporous membrane

before use. An aliquot of 1 μ L was injected into instrument for analysis.

An Agilent 6545 Accurate-Mass Q-TOF spectrometer coupled to an Agilent 1290 Infinity Binary LC system (UHPLC, Santa Clara, CA, United States) was employed. Samples were separated on an Agilent Eclipse Plus C18 column (1.8 μ m, 2.1 mm ID \times 150 mm). Mobile phase (A) was water containing formic acid (0.1%, v/v), and mobile phase (B) was acetonitrile and methanol (3:1, v/v). A gradient elution program was used as follows: 0–3 min, 80% A; 3–16 min, 80–64% A; 16–17 min, 64–40% A; 17–23 min, 40–20% A. The flow rate was 0.35 mL/min, and the column temperature was maintained at 55°C. The mass spectrometer was operated in the negative ESI mode. The parameters were as follows: Gas Temp: 325°C; Gas Flow: 11 L/min; Sheath Gas Temp: 350°C; Sheath Gas Flow: 12 L/min; scan range: m/z 50–1700.

The file format of the raw data was converted by Abf (Analysis Base File) Converter software, and then peak extraction, peak alignment, and normalization were processed by MS-DIAL (Tsugawa et al., 2015) software. SIMCA 17 software was utilized for metabolomics data analysis.

Active compound database construction for *Scutellariae radix*

The ingredients of *Scutellariae radix* were collected through the LC-MS identification (Table 1) and ETCM (Xu et al., 2019), TCMSP (Ru et al., 2014), and TCMID (Huang et al., 2018) database. The structures of the active compounds of *Scutellariae radix* were accustomed according to the PubChem database. The properties of absorption, distribution, metabolism, and excretion (ADME) were considered as important indicators for the effectiveness of drug candidates in modern drug discovery. The structures of the active ingredients from *Scutellariae radix* were then imported into “ADMET Descriptors” module in Discovery Studio 2021 (Dassault Systèmes BIOVIA, San Diego, CA) in SDF file format for the ADME prediction. Then these structures were put into the SwissADME (Daina et al., 2017) database for drug-like screening study.

Target prediction of *Scutellariae radix* and COVID-19-associated target screening

The potential targets of the pharmacodynamic chemical compositions of *Scutellariae radix* were collected by SwissTargetPrediction (Daina et al., 2019) and ETCM databases. In order to better understand the mechanism of the disease, the COVID-19 related genes were collected from two resources (Yao et al., 2020). One was from database mining that the

GeneCards¹ were employed to obtain COVID-19-related genes. The other was from analysis of microarray data that the differential expressed genes (DEGs) were obtained by analyzing GSE147507 (Blanco-Melo et al., 2020), which was obtained from the GEO database² via NetworkAnalyst online analysis with a threshold of adjust p-value ≤ 0.05 and $|\log_2 FC| \geq 1.0$. A gene library of the anti-COVID-19 targets of *Scutellariae radix* was established by comparing and analyzing the targets shared by the COVID-19-associated targets and the predicted targets of *Scutellariae radix* (Liu et al., 2020).

Bioinformatics analysis

To visualize and analyze the relationship between herbal medicine and disease, Cytoscape (version 3.7.2) (Shannon et al., 2003) was employed to construct the following three networks, “herb-compound-target-disease” network, the PPI network based on the importation of the gene symbols of the anti-COVID-19 targets of *Scutellariae radix* into the STRING (Szklarczyk et al., 2021) database, and target-signal pathway network.

The Metascape database³ (Zhou et al., 2019) is a web-based portal used for the analysis of GO (biological processes, molecular function, and cellular component) (Ashburner et al., 2000) and KEGG pathways (Kanehisa and Goto, 2000; Kanehisa et al., 2017). In this study, the gene symbols of the anti-COVID-19 targets of *Scutellariae radix* were imported into the Metascape platform, with Homo sapiens as the organism. The results of GO and KEGG pathway enrichment analysis were saved, and visualization was completed by the RStudio software.

Molecular docking

The crystal structures of proteins were downloaded from RCSB Protein Data Bank. In each crystal structure, the ligand and protein were remained and other molecules like water were removed by Discovery Studio 2021. The position of the active pocket is defined by the ligand on the protein crystal structure. Every protein was prepared by “Prepare Protein” module in Discovery Studio 2021. The chemical structures of compounds were extracted from PubChem, and then optimized by using “Prepare Ligands” module. Finally, each compound was docked into the protein active site by “CDOCKER” module in Discovery Studio 2021 (Gagnon et al., 2016; Xu et al., 2021). CDOCKER is a precise semi-flexible molecular docking technology that has long been recognized by peer-reviewed journals (Gagnon et al., 2016).

1 <https://www.genecards.org/>

2 <https://www.ncbi.nlm.nih.gov/>

3 <http://metascape.org/>

TABLE 1 Identification of compounds from *Scutellariae radix* extract by UHPLC-Q-TOF-MS.

No.	t_R (min)	[M-H] ⁻ measured	(m/z) predicted	Δ (ppm)	MS/MS	Formula	Identification
1	3.69	303.0507	303.0510	-0.99	125.0244, 149.0242, 177.0183, 259.0613, 285.0396	C ₁₅ H ₁₂ O ₇	(2R,3R)-3,5,7,2',6'- Pentahydroxyflavanone
2	5.33	547.1450	547.1457	-1.28	337.0721, 367.0821, 427.1012, 457.1129, 487.1224	C ₂₆ H ₂₈ O ₁₃	Chrysin 6-C-arabinoside 8-C-glucoside
3	6.13	623.1968	623.1981	-2.09	161.0238, 461.1636	C ₂₉ H ₃₆ O ₁₅	Acteoside
4	6.56	547.1448	547.1457	-1.64	337.0704, 367.0809, 427.1015, 457.1140	C ₂₆ H ₂₈ O ₁₃	Chrysin 6-C- β -D-glucoside-8-C- α -L- arabinoside
5	6.91	581.1857	581.1876	-3.27	299.0886, 329.1034, 461.1435	C ₂₇ H ₃₄ O ₁₄	2',4',6'-Trihydroxydihydrochalcone 3'-C- β -d glucoside-6'-O- β -D glucoside
6	7.16	547.1445	547.1457	-2.19	337.0687, 367.0807, 427.0995, 457.1117	C ₂₆ H ₂₈ O ₁₃	Chrysin 6-C- β -D-glucoside-8-C- β -L- arabinoside
7	8.41	285.0403	285.0405	-0.70	133.0291, 151.0040, 199.0357	C ₁₅ H ₁₀ O ₆	5,7,2',6'-Tetrahydroxyflavone
8	10.22	345.0614	345.0616	-0.58	287.0164, 315.0139, 330.0369	C ₁₇ H ₁₄ O ₈	Viscidulin III
9	10.45	287.0560	287.0561	-0.35	125.0245, 161.0240	C ₁₅ H ₁₂ O ₆	2',5,6',7-Tetrahydroxyflavanone
10	10.91	285.0401	285.0405	-1.40	117.0345, 137.0240, 166.9946, 267.0338	C ₁₅ H ₁₀ O ₆	Scutellarein
11	11.11	445.0774	445.0776	-0.45	175.0245, 269.0448	C ₂₁ H ₁₈ O ₁₁	Baicalin
12	11.66	505.0975	505.0988	-2.57	299.0143, 314.0407, 329.0657	C ₂₃ H ₂₂ O ₁₃	Viscidulin II 2'-O- β -D-glucuronide
13	12.09	447.0926	447.0933	-1.57	243.0662, 271.0605	C ₂₁ H ₂₀ O ₁₁	Dihydrobaicalin
14	12.75	651.2276	651.2294	-2.76	160.0166, 175.0402, 193.0507, 475.1790	C ₃₁ H ₄₀ O ₁₅	Cistanoside D
15	12.98	445.0770	445.0776	-1.35	174.9556, 269.0450	C ₂₁ H ₁₈ O ₁₁	Norwogonin 8-O- β -D-glucuronide
16	14.04	429.0819	429.0827	-1.86	176.8982, 253.0511	C ₂₁ H ₁₈ O ₁₀	Chrysin 7-O- β -D-glucuronide
17	14.38	459.0931	459.0933	-0.44	113.0242, 175.0243, 283.0602	C ₂₂ H ₂₀ O ₁₁	Oroxylin A 7-O- β -D-glucuronide
18	14.97	445.0760	445.0776	-3.59	174.9557, 269.0450	C ₂₁ H ₁₈ O ₁₁	Baicalein 6-O- β -D-glucuronide
19	15.34	459.0928	459.0933	-1.09	113.0245, 268.0371, 283.0607	C ₂₂ H ₂₀ O ₁₁	Wogonoside
20	15.47	461.1079	461.1089	-2.17	175.0239, 270.0447, 285.0672	C ₂₂ H ₂₂ O ₁₁	(2S)-5,7-Dihydroxy-6- methoxyflavanone 7-O- β -D-glucuronide
21	16.28	489.1025	489.1038	-2.66	216.9330, 283.0246, 298.0480, 313.0706	C ₂₃ H ₂₂ O ₁₂	5,7-Dihydroxy-8,2'- Dimethoxyflavone 7-O- β -D-glucuronide
22	16.98	299.0555	299.0561	-2.01	136.9865, 256.0363, 284.0321	C ₁₆ H ₁₂ O ₆	4'-Hydroxywogonin
23	17.62	269.0455	269.0455	0	171.0458, 197.0602, 225.0567	C ₁₅ H ₁₀ O ₅	5,7,2'-Trihydroxyflavone
24	17.75	299.0554	299.0561	-2.34	136.9700, 256.0331, 284.0319	C ₁₆ H ₁₂ O ₆	Scutevulin
25	17.85	329.0661	329.0667	-1.82	109.9983, 165.9892, 314.0418	C ₁₅ H ₁₀ O ₅	5,7,6'-Trihydroxy-8,2'- dimethoxyflavone
26	17.97	299.0556	299.0561	-1.67	153.9866, 284.0327	C ₁₆ H ₁₂ O ₆	5,7,2'-Trihydroxy-6-methoxyflavone
27	18.05	269.0454	269.0455	-0.37	111.0081, 223.0478, 241.0499	C ₁₅ H ₁₀ O ₅	Baicalein
28	18.20	329.0662	329.0667	-1.52	299.0185, 314.0425	C ₁₇ H ₁₄ O ₇	5,8,2'-Trihydroxy-6,7- dimethoxyflavone
29	18.95	343.0816	343.0823	-2.04	285.0606, 298.8586, 313.1408	C ₁₈ H ₁₆ O ₇	Skullcapflavone
30	19.16	283.0609	283.0612	-1.06	184.0520, 211.0384, 239.0317, 268.0367	C ₁₆ H ₁₂ O ₅	Wogonin
31	19.23	253.0506	253.0506	0	107.0137, 143.0116, 209.1528	C ₁₅ H ₁₀ O ₄	Chrysin
32	19.28	373.0922	373.0929	-1.88	300.0257, 328.0128, 343.0433	C ₁₉ H ₁₈ O ₈	Skullcapflavone II (5,6'-Dihydroxy- 6,7,8,2'-tetramethoxyflavone)
33	19.33	537.0810	537.0827	-3.17	245.0122, 391.0396	C ₃₀ H ₁₈ O ₁₀	8,8'-Bibaicalein
34	19.34	313.0712	313.0718	-1.92	283.0241, 298.0484	C ₁₇ H ₁₄ O ₆	5,8-Dihydroxy-6,7-dimethoxyflavone
35	19.48	283.0612	283.0612	0	239.0937, 268.0374	C ₁₆ H ₁₂ O ₅	Oroxylin A
36	19.89	343.0819	343.0823	-1.17	298.0075, 313.0370, 328.0593	C ₁₈ H ₁₆ O ₇	5,2'-Dihydroxy-6,7,8- trimethoxyflavone

Cell lines and SARS-CoV-2 virus

HEK-293T cells, RAW264.7 cells, and the African green monkey epithelial (Vero E6) cells were supplied by American Type Culture Collection (ATCC, Rockville, MD, United States). HEK-293T-ACE2^h cells were constructed by Sino Biological (Beijing, China). All the cell lines were cultured in Dulbecco Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum and 1% Gibco penicillin-streptomycin-glutamine (Thermo Fisher Scientific, Waltham, United States) at 37°C in a humidified atmosphere of 5% CO₂. Besides, 100 µg/mL hygromycin was used for the culture of HEK-293T-ACE2^h cells. SARS-CoV-2 (Genebank accession no. MT123290.1) was clinical isolate from the First Affiliated Hospital of Guangzhou Medical University. The viral titer was determined by 50% tissue culture infective dose (TCID₅₀) according to the cytopathic effect by use of Reed-Muench method. All the infection experiments of SARS-CoV-2 strain were performed in a biosafety level-3 laboratory.

Cell viability assay

The MTT assay was utilized to examine the cytotoxic effects of compounds on HEK-293T-ACE2^h cells, RAW264.7 cells, and Vero E6 cells. HEK-293T-ACE2^h cells were seeded into 96-well plate at a density of 1×10^4 cells per well and then treated with different concentrations of compounds for 4 h to measure the maximum non-toxic concentration (CC₀) of compounds. RAW264.7 cells (1×10^4 cells/well) or Vero E6 cells (5×10^4 cells/well) were seeded in 96-well plates for 24 h culture at 37°C in 5% CO₂ ambience. Next, the cells were incubated with extracts or compounds at various concentrations. After an incubation of RAW264.7 cells for 24 h and Vero E6 cells for 72 h, 10 µL MTT (5 mg/mL) was administered to each well and incubated for another 4 h. The supernatants were then removed, and the formed formazan crystals were dissolved in 100 µL DMSO. Finally, the absorbance value of each well was measured by a microplate reader at 570 nm. All data were obtained in triplicates and presented as means ± SD.

Quantitative detection of NO, tumor necrosis factor- α , and interleukin (IL)-6

RAW264.7 cells were seeded in 24-well (1×10^5 /well) plates and were pretreated with different concentrations of extracts or compounds for 1 h. The cells were then stimulated by LPS (100 ng/mL) for 18 h. The supernatant was then collected to determine NO production by using Griess reagent (Promega, Madison, WI, United States) according to the manufacturer's protocol. The levels of TNF- α and IL-6 in the supernatant were quantified by using the corresponding ELISA

kit (Cayman, Washtenau County, MI, United States) according to the manufacturer's instructions (Lio et al., 2020).

Biolayer interferometry binding assay

An Octet system (Octet RED96, ForteBio, United States) was employed for biolayer interferometry (BLI) to determine the binding kinetics and affinity of compounds or extracts with the proteins of SARS-CoV-2 spike RBD and human ACE2 (Sino Biological, Beijing, China). Either the recombinant His-tag human ACE2 (25 µg/mL aqueous solution) or recombinant His-tag SARS-CoV-2 spike RBD (25 µg/mL aqueous solution) was immobilized onto the surface of nitrilotriacetic acid (NNTA) biosensors (ForteBio, United States) that were dipped in a range of concentrations of compounds or extracts. Background signal was subtracted from all samples by using a reference biosensor loaded with either SARS-CoV-2 spike RBD or ACE2, which did not receive compounds or extracts under identical condition. The subtracted sensorgrams were then fitted to a 1:1 binding model to calculate the resulting equilibrium dissociation constant (K_D) value for this interaction. All experiments were repeated at least three times and K_D values were presented with mean ± SD.

Pseudovirus-based assay for the infection of HEK293T-ACE2^h cells

A SARS-CoV-2 spike pseudovirus was constructed according to our published protocol (Kong et al., 2021; Xu et al., 2022). This pseudovirus system contains both the SARS-CoV-2's spike protein and the Firefly luciferase reporter gene, which was employed to quantify compounds' capacity to inhibit virus's entry into host cells by measuring the luciferase's activity after infection of HEK-293T-ACE2^h cells. Briefly, 1×10^4 cells in 100 µL were seeded in 96-well plates and cultured for 24 h. Cells were first incubated in compound-containing media for 2 h at 37°C. After disposal of the media, 100 µL mixture solution of pseudovirus and compound (consisted of 30 µL pseudovirus, 20 µL media, and 50 µL two-fold concentration of compounds) was then transferred to HEK-293T-ACE2^h cells for 2 h incubation. After disposal of the above culture media, the cells were treated with fresh media for another 48 h of incubation at 37°C. The luciferase luminescence value was then read at 578 nm by a microplate reader (SpectraMax iD5 Multi-Mode Microplate Reader, Molecular Devices, United States) by using Firefly Luciferase Reporter Gene Assay Kit (China). The luciferase luminescence value of HEK-293T-ACE2^h cells infected with the SARS-CoV-2 spike pseudovirus was set as 100%. Pseudo virus inhibition rate (%) = $[1 - (\text{Sample fluorescence values} - \text{Average fluorescence value of the background}) / (\text{Average fluorescence value of virus control} -$

Average fluorescence value of the background)] \times 100%. The IC_{50} was determined by using the GraphPad Prism 8.0 software (La Jolla, CA, United States).

Plaque reduction assay

A pretreatment mode was used in plaque reduction assay to test the inhibitory effect of compounds on SARS-CoV-2's entry into host cells. The 50 plaque-forming units (PFU) of SARS-CoV-2 strains (Genebank accession no. MT123290.1) were pretreated with various concentrations of tested compounds at room temperature for 2 h. Then, the Vero E6 cell monolayers grown in 12-well plates were infected with the above mixture solution of both virus and compounds for 2 h incubation at 37°C. After removal of the above culture solution, the cell monolayers were covered with agar overlay (containing concentration: 0.6% agar and 2% FBS). The plates were subsequently incubated for 48 h at 37°C with 5% CO₂ ambience, and followed by fixation in 10% formalin for 30 min. Then, the agar overlays were removed, and the cell monolayers were stained with 1% crystal violet. Finally, the plaques were counted and photographed. All the plaque reduction assays were carried out in triplicate, and IC_{50} values were expressed with mean \pm SD.

Statistical analysis

The data were expressed as the mean \pm standard deviation (SD). Either t-test or non-parametric test was used to compare two groups. Statistical significance was set at $p < 0.05$. Analysis and graphing were performed by using GraphPad Prism 8.0 software (San Diego, CA, United States).

Result

Collection and screening of active compounds from *Scutellariae radix*

As shown in **Figure 2A**, 213 compounds of *Scutellariae radix* were firstly collected based on ETCM (Xu et al., 2019), TCMSP (Ru et al., 2014), and TCMID (Huang et al., 2018) database and LC-MS identification (**Table 1**). 109 potential active ingredients were then filtered out from the components of *Scutellariae radix* according to the criterion of ADME (absorption, distribution, metabolism and excretion) provided by the Discovery Studio 2021 software (**Figure 2B**). This study also used five drug-likeness filters, including Lipinski (Pfizer) (Lipinski et al., 2001), Muegge (Bayer) (Muegge et al., 2001), Ghose (Amgen) (Ghose et al., 1999), Veber (GlaxoSmithKline) (Veber et al., 2002), and Egan (Pharmacia) (Egan et al., 2000). Finally, the ADME and

drug-likeness screening yielded 61 potential active compounds (**Figure 2C** and **Supplementary Table 2**).

Herb-compound-target-disease network construction and network topological analysis

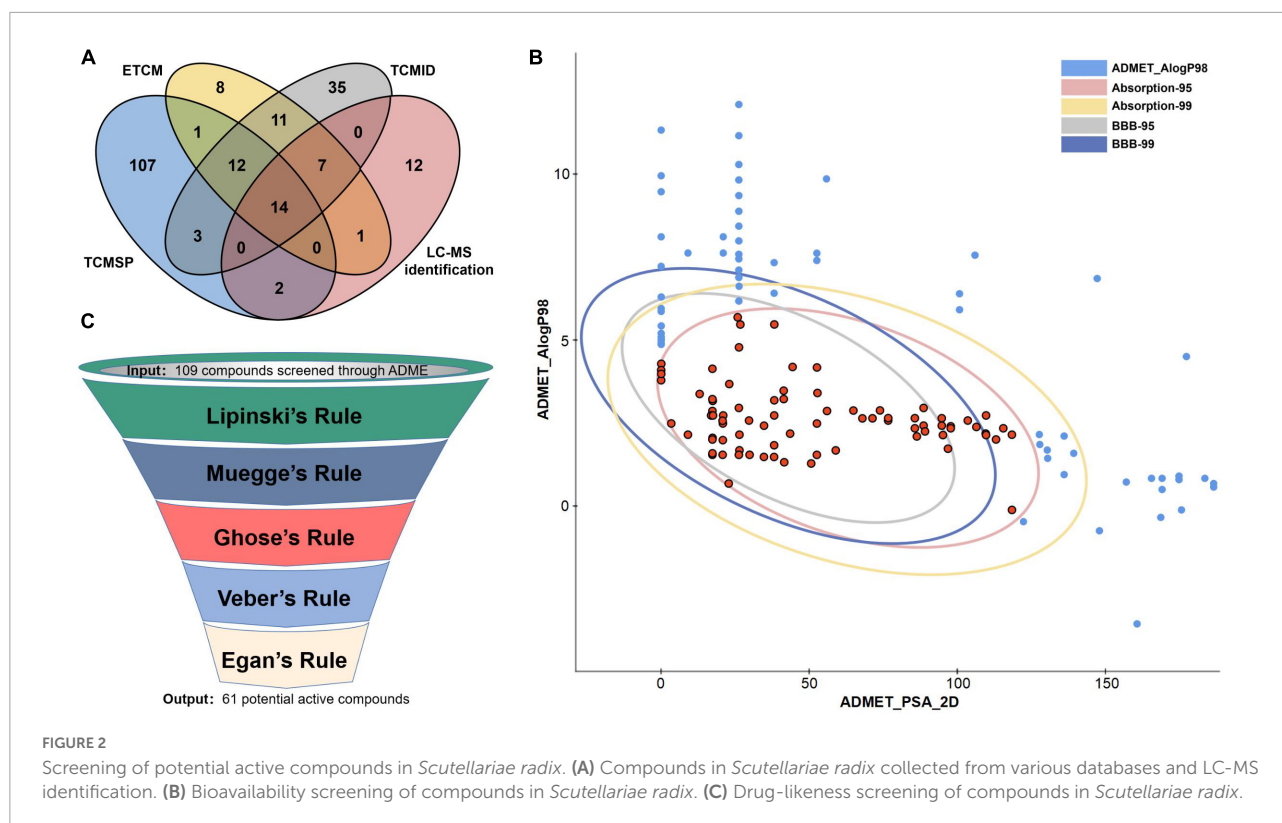
A total of 445 targets were predicted through two chemical similarity-based target search methods (Daina et al., 2019; Xu et al., 2019) by using 61 potential active compounds of *Scutellariae radix* (**Supplementary Tables 3, 4**).

COVID-19-associated genes were gathered from two different sources. First, the top 500 COVID-19-related genes were extracted from the GeneCards (Safran et al., 2010) database. Second, the transcriptomics analysis was used to investigate the genes associated with COVID-19 (**Supplementary Table 5**). The fold-changes between the two groups were depicted on the volcano plot (**Figures 3A,B**). As shown in **Figure 3C**, a total of 280 overlapping differentially expressed genes (DEGs) were screened out, including 243 up-regulated and 37 down-regulated DEGs (**Supplementary Table 6**). When the gene lists from the two sources were combined, 768 genes were thought to be potential targets for COVID-19 (**Figure 3D**).

The compound-target network and COVID-19 disease network were merged by Cytoscape software to construct a herb-compound-target-disease network that was composed of 108 nodes (1 herb, 50 compounds, 56 targets, and 1 disease) and 538 edges, as shown in **Figure 4A**. The composite network illuminated the relationship among the *Scutellariae radix*, candidate active compounds, potential therapeutic targets, and COVID-19. Based on the judgment criteria of previous network pharmacology research (Guo et al., 2022), it can be noticed that the top 10 active components in degree value (norwogonin, 4'-hydroxywogonin, 5,7,2',3'-tetrahydroxyflavone, isoscutellarein, baicalein, apigenin, 5,7,2'-trihydroxyflavone, chrysin, scutellarein, scutevulin, and 7,3',4'-trihydroxyflavone) are probably the main active ingredients of *Scutellariae radix* exerting anti-COVID-19 effect.

PPI network, GO and KEGG analyses of the anti-COVID-19 targets of *Scutellariae radix*

To explore the possible mechanism of *Scutellariae radix* as treatment against COVID-19, 56 potential anti-COVID-19 targets of *Scutellariae radix* were imported to the STRING (Szklarczyk et al., 2021) database in order to construct a PPI network (**Figure 4B**). Further cluster analysis was done by using the MCODE module of Cytoscape (Shannon et al., 2003) to generate the highly connected sub-network. It can be noticed



that the top 10 targets (VEGFA, AKT1, STAT3, TNF, MAPK3, MAPK1, EGFR, MAPK8, MTOR, and IL2) are potential hub genes of *Scutellariae radix* exerting anti-COVID-19 effect.

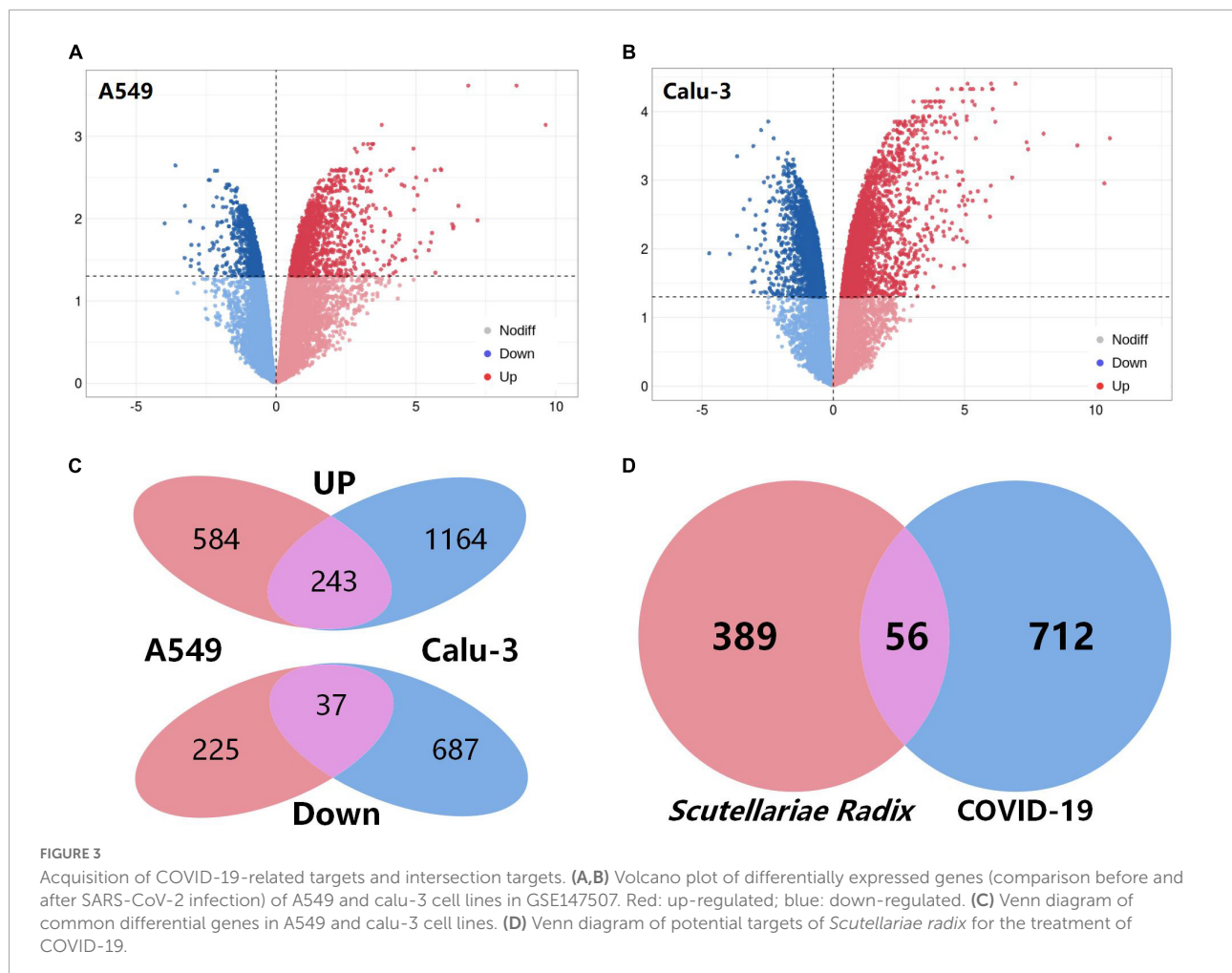
To further explore the various mechanisms of the anti-COVID-19 activity of *Scutellariae radix*, we performed a GO enrichment analysis (Ashburner et al., 2000) (biological processes, molecular function, and cellular component) of the 56 predicted targets. The top 10 enriched biological process terms, molecular function terms, and cellular component terms were determined (Figure 4C and Supplementary Table 7). The top five biological processes terms were peptidyl-serine phosphorylation, wound healing, positive regulation of kinase activity, positive regulation of cytokine production, and response to lipopolysaccharide. The top five molecular function terms were protein kinase activity, protein serine/threonine/tyrosine kinase activity, protein kinase binding, transcription factor binding, and cytokine receptor binding. The top five cellular component terms were membrane raft, vesicle lumen, perinuclear region of cytoplasm, postsynapse, and region of cytosol. Biological processes such as “wound healing, positive regulation of cytokine production, response to lipopolysaccharide, and inflammatory response” can be observed in the results of GO enrichment analysis. This result suggested that the anti-COVID-19 effect of *Scutellariae radix* was closely related to inflammation.

A KEGG pathway enrichment analysis (Kanehisa and Goto, 2000; Kanehisa et al., 2017) at $p < 0.01$ significance

level was performed for the 56 potential anti-COVID-19 targets of *Scutellariae radix* (Figure 4D and Supplementary Table 8). The top ten significantly enriched KEGG pathways were Pathways in cancer (hsa05200), AGE-RAGE signaling pathway in diabetic complications (hsa04933), MAPK signaling pathway (hsa04010), Insulin resistance (hsa04931), Platelet activation (hsa04611), Measles (hsa05162), NF-kappa B signaling pathway (hsa04064), Rheumatoid arthritis (hsa05323), Inflammatory bowel disease (IBD) (hsa05321) and Transcriptional misregulation in cancer (hsa05202). Notably, MAPK and NF- κ B signaling pathway are significant pathways regulating inflammatory cytokine storm. According to the results from GO and KEGG analyses, the potential mechanism of *Scutellariae radix* exerting anti-COVID-19 effect may be achieved by regulating MAPK and NF- κ B signaling pathways by the active ingredients of *Scutellariae radix* (Figure 4E).

Molecular docking predicted main active components of *Scutellariae radix* against COVID-19

Specific parameter setting information for molecular docking was presented in Supplementary Table 9. According to the results of network pharmacology, the targets in the MAPK and NF- κ B pathways, and the top 10 targets in the PPI network with degree value were considered as the main potential targets



of *Scutellariae radix* against COVID-19. Two additional targets (SARS-CoV-2 spike protein, and human ACE2 receptor) closely related to the direct infection of host cells by SARS-CoV-2 have also been included in this study, due to the reported anti-infectious ability of *Scutellariae radix* in other bacterial or viral diseases. A precise semi-flexible molecular docking method of CDocker (Gagnon et al., 2016; Xu et al., 2021) was used to further investigate the main components of *Scutellariae radix* for treating COVID-19. The top 10 compounds in network pharmacology analysis and another four constituents (wogonin, oroxylin A, baicalin, and wogonoside) that were often considered to be the active ingredients of *Scutellariae radix*, were docked into the active centers of the aforementioned targets. According to the result of molecular docking (Figure 5), the top five constituents with high docking scores against COVID-19 were scutellarein, isoscutellarein, 7,3',4'-trihydroxyflavone, 5,7,2',3'-tetrahydroxyflavone, and baicalein.

Lower binding energy indicated higher stability. Based on the binding energy score (Figure 5), IKBKB was identified as a key target of *Scutellariae radix* for the treatment of COVID-19 through anti-inflammatory effects. IKBKB plays an essential

role in the NF- κ B signaling pathway which is a key regulatory target in the activation process of NF- κ B (Cardinez et al., 2018). As displayed in Supplementary Figure 2, scutellarein formed four hydrogen bonds with Lys44, Glu97, Cys99, and Asp166 in IKBKB. In addition, two hydrogen bonds were formed between baicalein and IKBKB (Supplementary Figure 2). The binding energies of scutellarein and baicalein with IKBKB were - 43.61 and - 35.56 kJ/mol, respectively. The results indicated that the components could bind to the active sites of the targets. The surface envelops spike protein of SARS-CoV-2 is essential for its infection of host cells. The initial attachment of the SARS-CoV-2 to the host cell is initiated by interactions between the spike RBD and human receptor ACE2. According to the relevant literature (Lan et al., 2020; Teli et al., 2021), the key amino acid residues for spike to bind to ACE2 are Tyr449, Gln493, Gln498, Thr500, Asn501, Gly502, and Tyr505. As shown in Supplementary Figure 2C, scutellarein formed three hydrogen bonds with Arg403, Tyr453, and Ser494 in spike protein. Supplementary Figure 2D showed that baicalein could form stable hydrogen bond with spike protein Gln498, a key amino acid residue in the binding of spike protein to human ACE2 protein. In

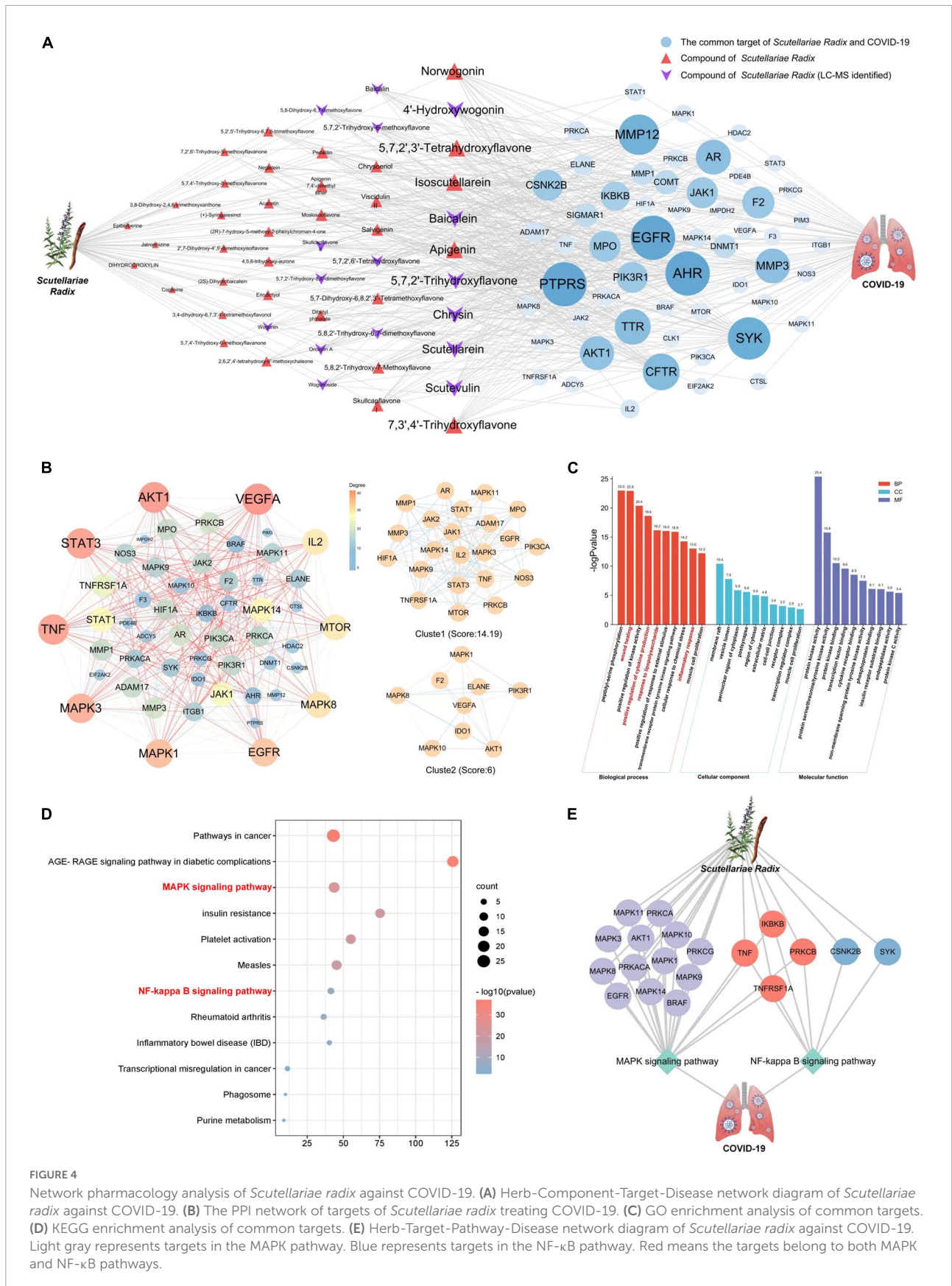
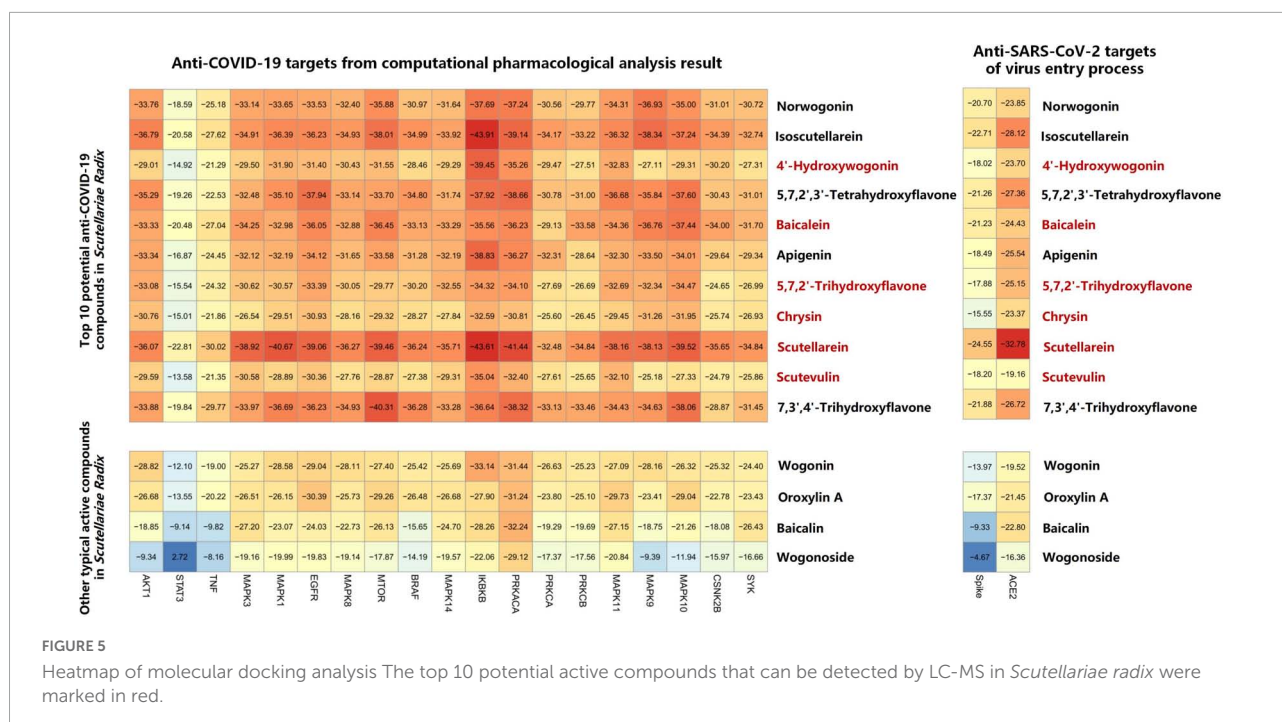


FIGURE 4 Network pharmacology analysis of *Scutellariae radix* against COVID-19. **(A)** Herb-Component-Target-Disease network diagram of *Scutellariae radix* against COVID-19. **(B)** The PPI network of targets of *Scutellariae radix* treating COVID-19. **(C)** GO enrichment analysis of common targets. **(D)** KEGG enrichment analysis of common targets. **(E)** Herb-Target-Pathway-Disease network diagram of *Scutellariae radix* against COVID-19. Light gray represents targets in the MAPK pathway. Blue represents targets in the NF-κB pathway. Red means the targets belong to both MAPK and NF-κB pathways.



addition, baicalein also formed hydrogen bonds with spike protein Ser494 and Gly496. According to molecular docking results, scutellarein and baicalein had the potential to prevent SARS-CoV-2 entry into host cells by blocking viral spike RBD.

Chemical analysis of *Scutellariae radix* by UHPLC-Q-TOF-MS

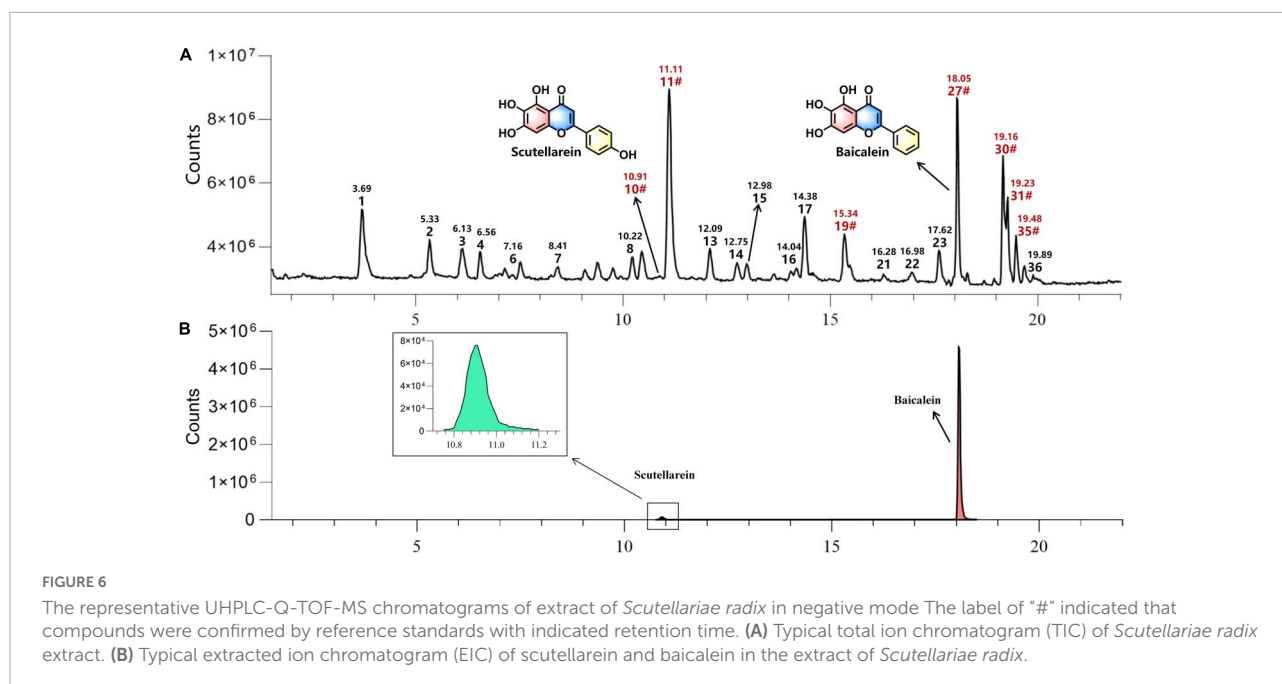
A total of 36 compounds were identified from two commercial products specification (ZiQin and KuQin) of *Scutellariae radix* by UHPLC-Q-TOF-MS analysis (Figure 6, Supplementary Figure 3, and Table 1). Only scutellarein and baicalein were found in the extract of *Scutellariae radix* out of the top 5 compounds revealed by molecular docking study. The other three constituents (isoscutellarein, 7,3',4'-trihydroxyflavone and 5,7,2',3'-tetrahydroxyflavone) were too trace to be detected from *Scutellariae radix* under our tested UHPLC-Q-TOF-MS condition. Therefore, scutellarein and baicalein were selected for subsequent biological verification by bioassays.

Anti-inflammatory and anti-infectious potential of *Scutellariae radix*

LPS-stimulated mouse macrophage RAW264.7 cells have been widely used as an inflammatory cell model to evaluate anti-inflammatory activity of drugs (Huang et al., 2013). Therefore, this study utilized this inflammation model to

simulate and explore the anti-inflammatory effect of the extracts of *Scutellariae radix* in COVID-19. The cytotoxicity of the extracts of ZiQin and KuQin in RAW264.7 was firstly examined by the MTT assay (Supplementary Figure 4). The extracts of both ZiQin and KuQin showed no significant cytotoxicity on RAW264.7 cells under the concentration of 400 $\mu\text{g/ml}$. There was no significant difference in the inhibitory effect of ZiQin extracts from three different origins (Inner Mongolia autonomous region, Hebei province and Shanxi province) on the NO release in the RAW264.7 inflammatory cell model, and so did KuQin (Figure 7A). Dose-dependent inhibition of NO release was observed for both ZiQin and KuQin extracts. ELISA assay was also used to determine the protein expression levels of TNF- α and IL-6, two inflammation cytokines in RAW264.7 inflammatory cell model. As the results, both ZiQin and KuQin extracts dose-dependently suppressed the release of TNF- α and IL-6 (Figures 7A–C). In addition, the inhibitory effects of KuQin extract on the release of NO, TNF- α and IL-6 were more potent than that of ZiQin extract ($p < 0.05$) at the concentration of 50 $\mu\text{g/ml}$ and 100 $\mu\text{g/ml}$ (Figures 7B,C). These results suggested that *Scutellariae radix* has anti-inflammatory potential, and the anti-inflammatory effect of KuQin extract was more potent than that of ZiQin extract.

SARS-CoV-2 enters the host cells by recognizing ACE2 receptor of human cells. Bi-layer interference (BLI) binding analysis was conducted to further evaluate the binding ability of extracts of *Scutellariae radix* with both the spike RBD and human ACE2 proteins, which could affect the recognition of SARS-CoV-2 spike RBD to human ACE2 receptor. As shown in Figures 7D,E,G,H, it was found that KuQin extract binds to the



spike RBD with an affinity (K_D value) of $112.84 \pm 29.40 \mu\text{g/mL}$, stronger than that ($331.88 \pm 55.24 \mu\text{g/mL}$) of ZiQin extract. KuQin extract binds to human ACE2 protein with a K_D value of $157.40 \pm 14.56 \mu\text{g/mL}$, while ZiQin gives a K_D value of $486.80 \pm 35.63 \mu\text{g/mL}$. In general, the binding affinity of KuQin with both SARS-CoV-2 spike RBD and human ACE2 proteins was significantly higher than that of ZiQin ($p < 0.01$). KuQin and ZiQin were further evaluated for their potential to inhibit viral entry of a SARS-CoV-2 spike pseudovirus bearing Firefly luciferase reporter into HEK-293T-ACE2^h host cells. As a consequence, the ability of SARS-CoV-2 spike pseudovirus entering into host cells was significantly reduced by the extracts of both KuQin and ZiQin, where IC_{50} value ($78.20 \pm 0.91 \mu\text{g/mL}$) of KuQin was approximately half that ($142.12 \pm 11.32 \mu\text{g/mL}$) of ZiQin ($p < 0.0001$) (Figures 7E,I), indicating a stronger antiviral activity of KuQin than ZiQin. Taken together, these results suggested that *Scutellariae radix* possesses both anti-inflammatory and anti-infectious potential. Furthermore, KuQin had superior anti-inflammatory and anti-SARS-CoV-2 activities to ZiQin.

Validation of main active components of *Scutellariae radix* for anti-inflammatory and anti-infectious effects

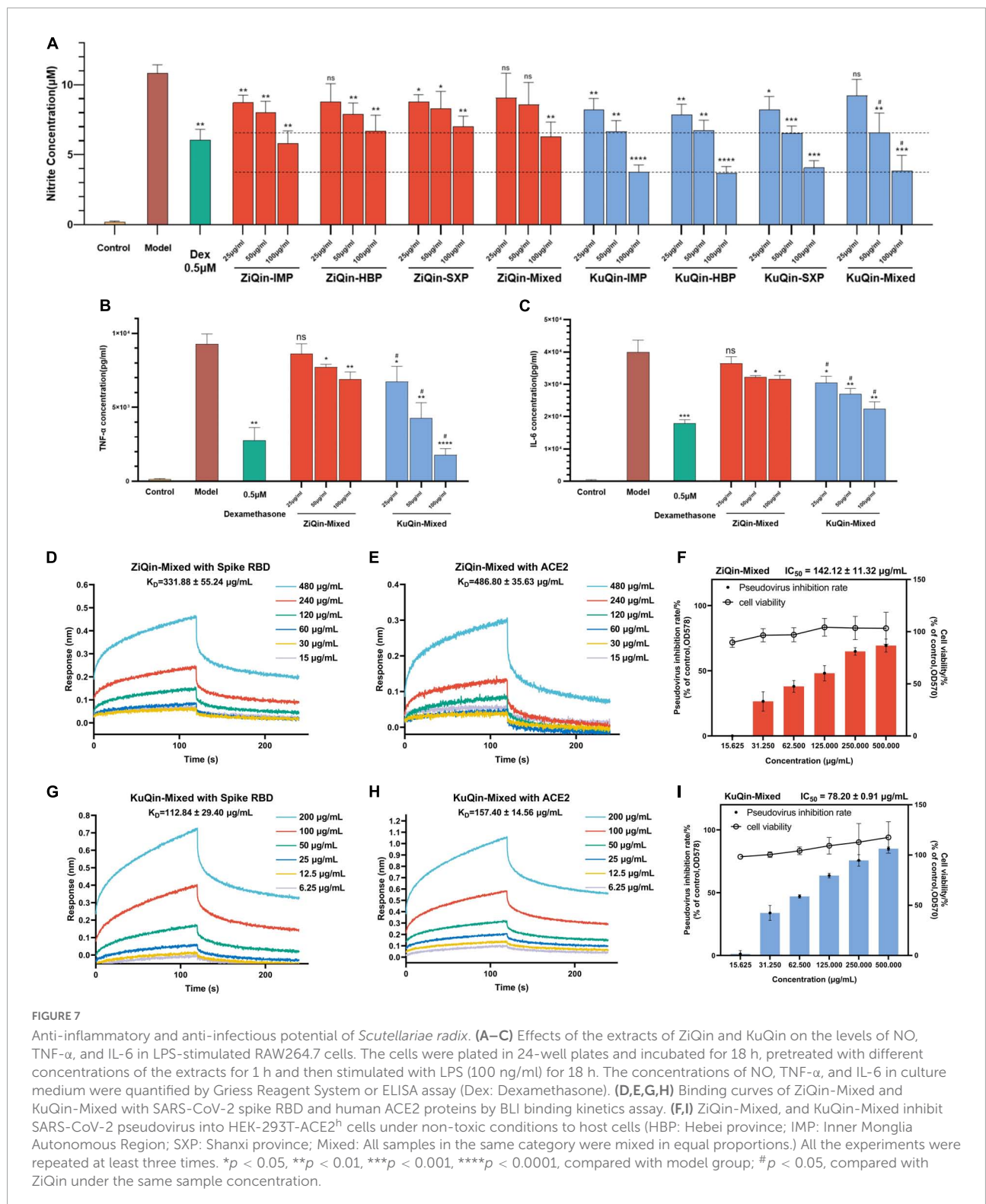
The effects of baicalein and scutellarein on the levels of NO, TNF- α , and IL-6 in LPS stimulated RAW264.7 cells were subsequently evaluated by using the same above bioassays. Baicalin, as the only chemical marker for quality control of

Scutellariae radix in Chinese Pharmacopoeia, was also evaluated for its activity. As shown in Figures 8A-C, it was found that baicalein and scutellarein demonstrated more potent inhibitory effects on the production of NO, TNF- α , and IL-6 than baicalin. Based on these results, it was considered that baicalein and scutellarein were active components with anti-inflammatory potential in *Scutellariae radix*.

As shown in Figures 8D,E,G,H,J,K, BLI binding assay revealed that baicalein, scutellarein and baicalin dose-dependently bind to both SARS-CoV-2 spike RBD and ACE2 proteins. The binding affinity of baicalein and scutellarein were 2.6 ~ 5.7 folds higher than that of baicalin with both proteins. Baicalein and scutellarein might more effectively prevent SARS-CoV-2 from binding to the host's ACE2 receptor by dual recognizing both spike RBD and human ACE2 proteins than baicalin. As shown in Figures 8F,I,L, baicalein, scutellarein and baicalin exhibited antiviral entry effect on SARS-CoV-2 spike pseudoviruses into host cells with IC_{50} values of 45.36, 37.86, and 101.83 μM , respectively. Based on both strong anti-inflammatory and anti-infectious potency, baicalein was selected for subsequent antiviral validation against SARS-CoV-2, and baicalin (the glycoside of baicalein) was also used for comparison.

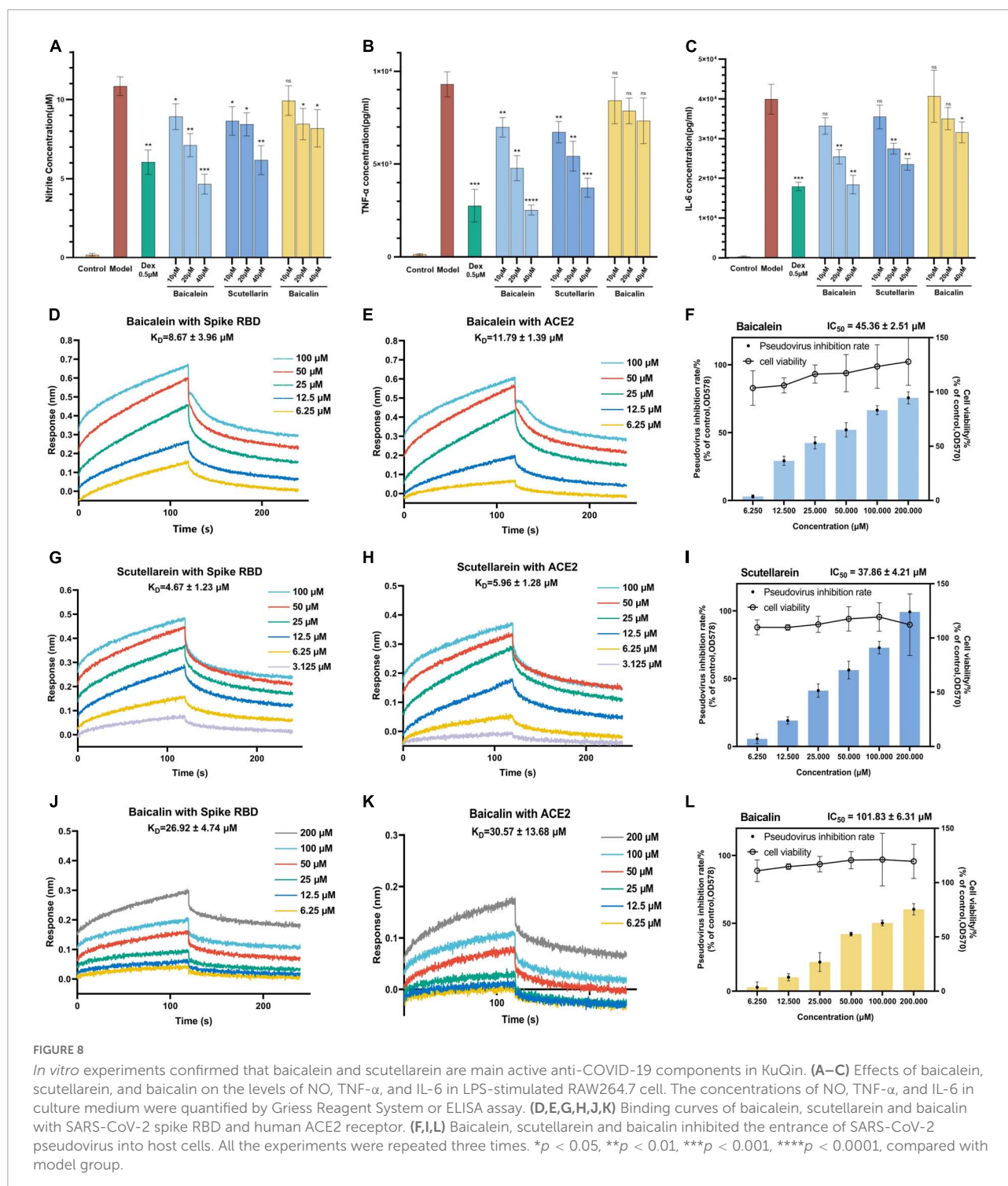
Antiviral effect of baicalein against SARS-CoV-2 by plaque formation assay

In order to confirm the above antiviral effect of baicalein against SARS-CoV-2, plaque formation assay was further carried out in a biosafety level 3 facility. Baicalein and baicalin were not



cytotoxic on Vero E6 cells at a tested concentration up to 250 and 500 μM , respectively. In a pretreatment mode, baicalin showed a dose-dependent inhibitory effect on SARS-CoV-2's entry into Vero E6 host cells with an IC_{50} value of 142.50 μM by

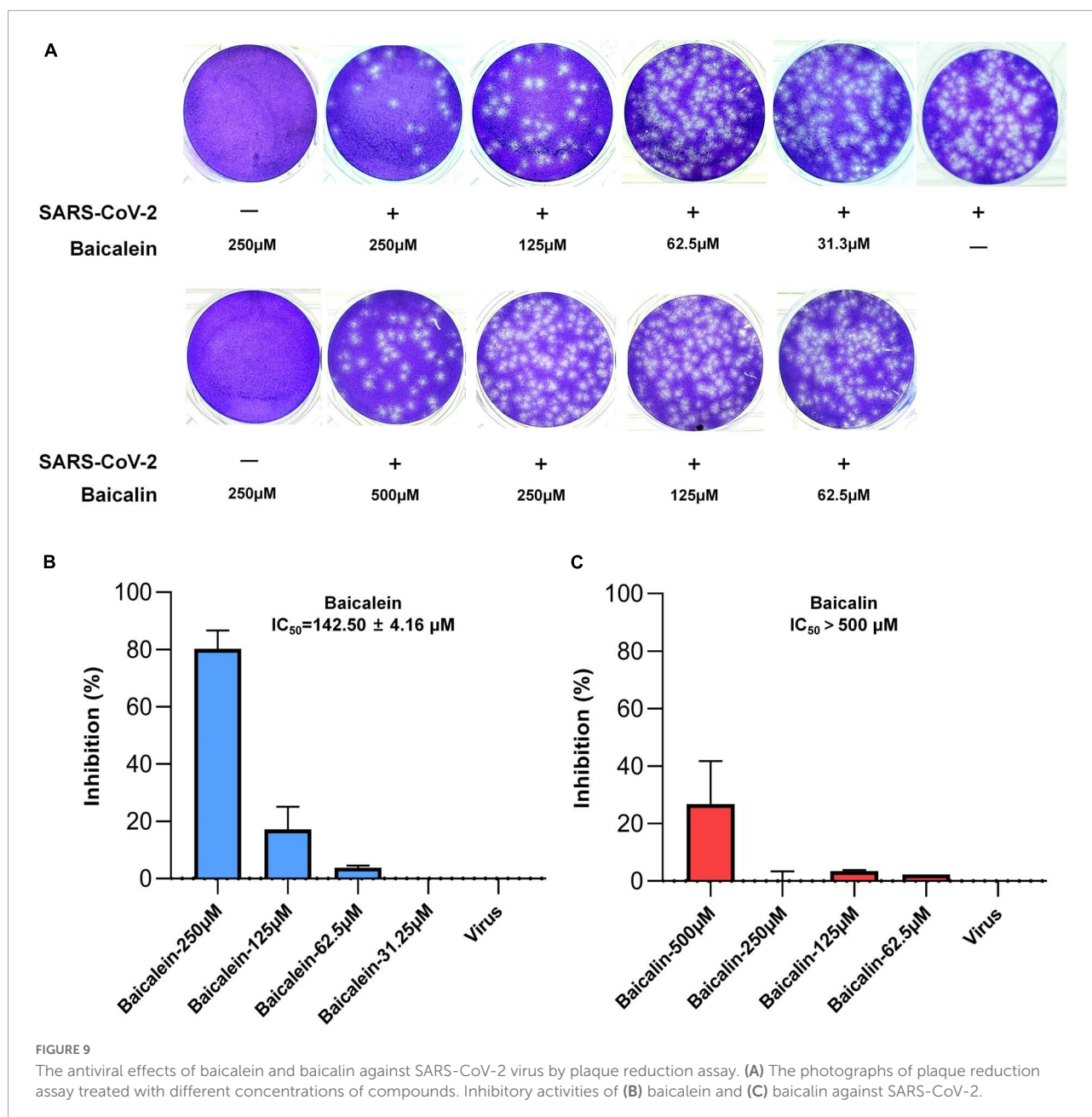
plaque formation assay, while baicalin displayed extremely weak antiviral activity even in the treatment concentration of 500 μM (Figure 9). The antiviral entry effect of baicalin against SARS-CoV-2 further confirmed the result from pseudovirus model.



KuQin's chemical basis for superior anti-COVID-19 potential to ZiQin

It was found that KuQin outperforms ZiQin in both anti-inflammatory and anti-infectious activity against SARS-CoV-2 based on the above results. The chemical difference of

KuQin and ZiQin was thus analyzed by UHPLC-UV instrument coupled with an ESI-TOF detector (Figures 10A–E). In order to avoid the influence of sample origin difference on the accuracy of this study, we have collected 3 batches of samples from three provinces (1 batch/province) for the chemical analysis of both ZiQin and KuQin. Figures 10A,B showed the typical



UHPLC-UV chromatograms of KuQin and ZiQin. KuQin showed a very similar chromatogram to that of ZiQin except for a remarkably higher content of baicalein (peak No. 27). Quantitation was performed by external standard method. The content of baicalin in KuQin was comparable to that in ZiQin (Figure 10C and Supplementary Table 10), but the content of baicalein in KuQin was 4.1-fold higher than that in ZiQin (Figure 10D and Supplementary Table 10). However, scutellarein was too trace to be detected at UV wavelength of 270 nm under our tested condition. This constituent can be detected by MS detector only in KuQin, but not in ZiQin (Figure 10E).

Principal Component Analysis (PCA) was then carried out to district the metabolic profiles between KuQin and ZiQin. According to the PCA score graph (Figure 10F), it can be seen intuitively that ZiQin and KuQin samples were divided into two categories, suggesting that regional differences did not have significant impact on the chemical composition of the two commercial products specification. OPLS-DA model was also used to further explore the specific chemical differences between ZiQin and KuQin. The established OPLS-DA model presented satisfied explanation and prediction according to the cross-validations analysis (Figure 10G), where $R^2X_{(cum)} = 0.906$, $R^2Y_{(cum)} = 0.996$ and $Q^2_{(cum)} = 0.954$. Permutation test with

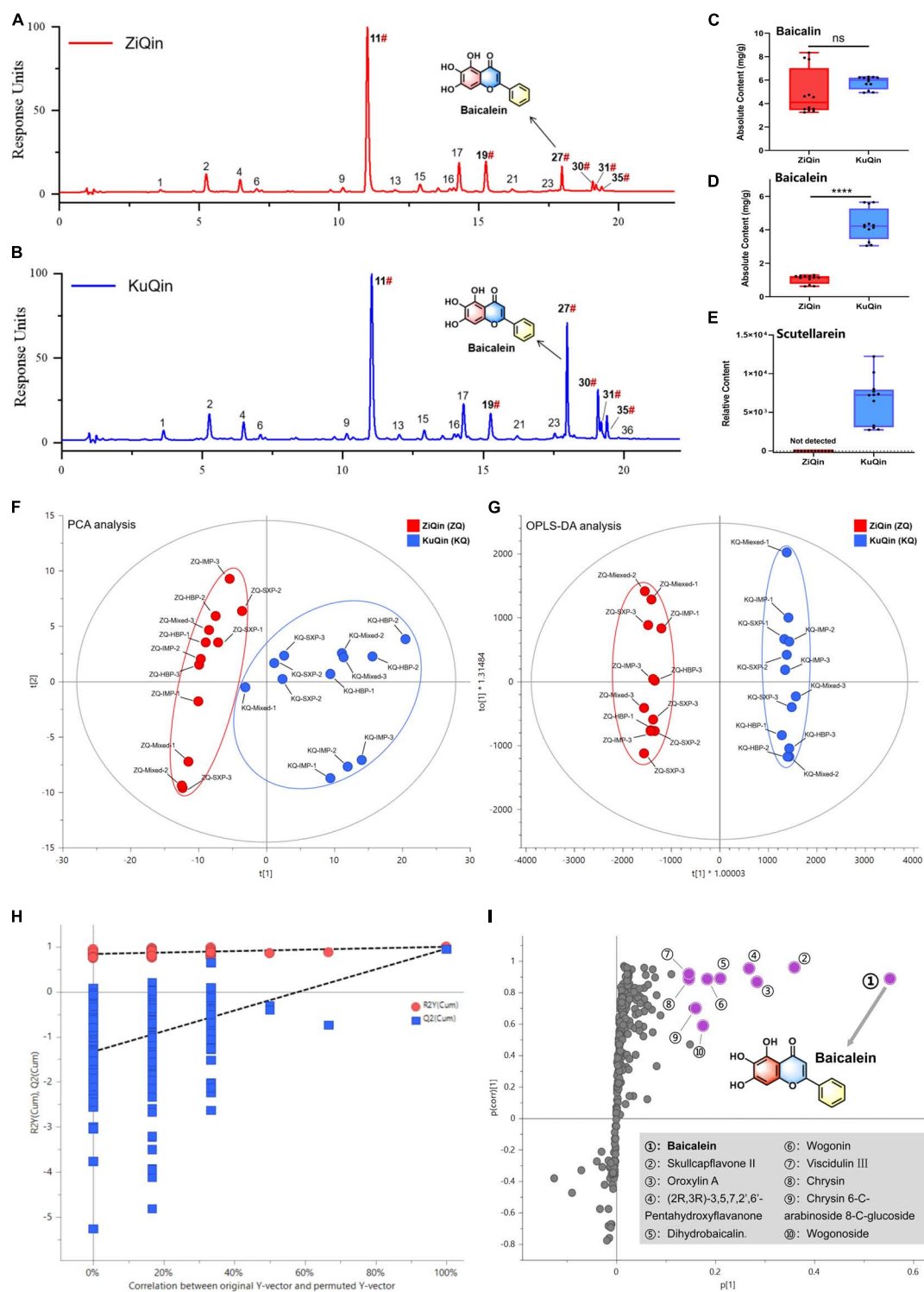


FIGURE 10 Chemical comparison between ZiQin and KuQin. **(A,B)** UHPLC-UV chromatograms of ZiQin and KuQin extract under UV wavelength of 270 nm. The label of “#” indicated that compounds were confirmed by reference standards. **(C,D)** Quantitative analysis of baicalin and baicalein in ZiQin (red) and KuQin (blue) extracts by UHPLC-UV with an external standard method, respectively. **(E)** The relative content of scutellarein under MS detection. **(F,G)** The PCA analysis and OPLS-DA analysis between ZiQin and KuQin. **(H)** Permutation test ($n = 200$) demonstrated no overfitting in the OPLS-DA model. **(I)** S-plot diagram from the OPLS-DA model between ZiQin and KuQin. The points highlighted in purple implied the compounds with significantly higher content in KuQin than those in ZiQin (HBP, Hebei province; IMP, Inner Mongolia Autonomous Region; SXP, Shanxi province; Mixed, All samples in the same category were mixed in equal proportions).

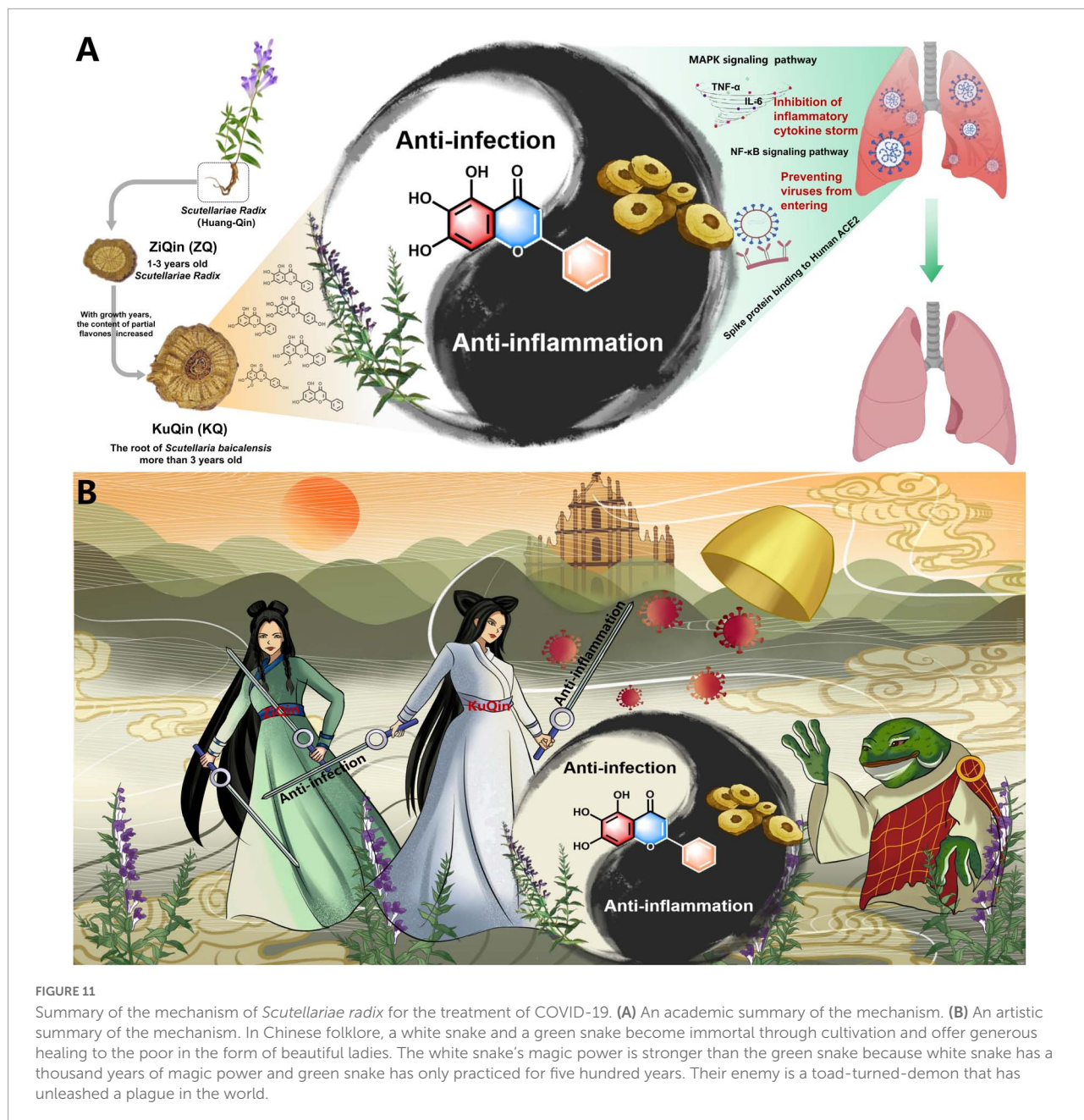


FIGURE 11 Summary of the mechanism of *Scutellariae radix* for the treatment of COVID-19. (A) An academic summary of the mechanism. (B) An artistic summary of the mechanism. In Chinese folklore, a white snake and a green snake become immortal through cultivation and offer generous healing to the poor in the form of beautiful ladies. The white snake’s magic power is stronger than the green snake because white snake has a thousand years of magic power and green snake has only practiced for five hundred years. Their enemy is a toad-turned-demon that has unleashed a plague in the world.

200 permutations also showed that Q^2 and R^2 value were lower than the original points, which suggested that the established model was deemed to be successful (Figure 10H). According to these two criteria, baicalein can be considered as the most significantly differential chemical marker of KuQin compared to ZiQin (Figure 10I).

Discussion

Globally, since the outbreak of COVID-19 in December 2019, WHO has reported 572 million confirmed cases of

COVID-19, including 6.39 million deaths. Until now, the prevention and treatment of COVID-19 remain a major global challenge. China mainly treats mild and ordinary patients with traditional Chinese medicine (TCM) to improve clinical symptoms such as fever, cough, anorexia, and fatigue, as well as to reduce the development of severe diseases. Through data mining of the official COVID-19 treatment guidelines of 34 provinces in China, we found that *Scutellariae radix* is extensively used in the treatment of non-severe patients. According to the theory of TCM, *Scutellariae radix* is divided into two specifications: ZiQin and KuQin. Ancient books of TCM have recorded that KuQin is better than ZiQin

in the treatment of pneumonia, but the opposite in the treatment of enteritis. Essentially, COVID-19 is a pneumonia caused by SARS-CoV-2 infection. It has not yet been reported whether the aforementioned TCM theories are applicable in the treatment of COVID-19, and what the mechanism is. However, due to the complex chemical components of *Scutellariae radix*, it remains difficult to elucidate its potential active compounds and precise pharmacological mechanisms in treating COVID-19.

The bioactive components and therapeutic mechanisms of *Scutellariae radix* were summarized in **Figure 11** for the treatment of COVID-19. In this research, network pharmacology and molecular docking analysis revealed that *Scutellariae radix* primarily regulated the MAPK and NF- κ B signaling pathways *via* active components such as scutellarein and baicalein, and blocked viral spike protein binding to ACE2 receptors to treat COVID-19. *In vitro* bioassays showed that scutellarein and baicalein were proved to possess more potent anti-inflammatory and anti-infectious activities than baicalin, the dominant principle with the highest content in *Scutellariae radix*. The biological activities of scutellarein and baicalein were comparable in the treatment of COVID-19. With respect to the relative content in *Scutellariae radix*, baicalein is one of the main constituents, while scutellarein is just a trace component. Moreover, baicalein was confirmed by plaque formation assay to inhibit SARS-CoV-2's entry into Vero E6 cells with an IC₅₀ value of 142.50 μ M. In summary, baicalein was considered to be the most important active component of *Scutellariae radix* for the treatment of COVID-19 by integrative analysis.

The *in vitro* bioassays demonstrated that KuQin outperforms ZiQin both in the anti-inflammatory and anti-infectious activities. Using non-targeted metabolomics technology, we discovered that the content of active aglycones in KuQin was significantly higher than that of ZiQin. It is worth mentioning that baicalein's VIP value (the value of variable important in projection) exceeded 1.58 times that of the second-ranked compound. This result suggested that baicalein was the compound with the most significant variation in KuQin compared to ZiQin. Quantitative analysis showed that the content of baicalein in KuQin was 4.1-fold higher than that in ZiQin. In summary, we considered that the significant increase of baicalein in KuQin is the main reason for the superiority of KuQin over ZiQin in the treatment of COVID-19. Based on the above quality evaluation of KuQin and ZiQin, this study also provided a scientific support for the traditional theory of TCM that KuQin was good at cooling down the heat in the lung. These findings suggested that KuQin could be used as a dietary supplement for COVID-19 patients.

In Chinese Pharmacopoeia (2020 version), baicalin was used as the only chemical marker for the quality control of *Scutellariae radix*. It was suggested that baicalein could be employed as an additional bioactive marker for the quality assurance of *Scutellariae radix* based on this study.

Conclusion

In summary, our study explored the potential bioactive components, underlying mechanism and quality of the widely used dietary supplement, *Scutellariae radix* for the treatment of COVID-19 by using network pharmacology, molecular docking, LC-MS analysis and *in vitro* biological evaluations. This investigation provided both theoretical and experimental supports for the treatment of COVID-19 with *Scutellariae radix*. Based on the findings of this study, KuQin was better than ZiQin for the prescriptions of TCMs treating COVID-19 due to its superior anti-inflammatory and anti-infectious effects to ZiQin. Baicalein was the most effective component with both anti-inflammatory and antiviral efficacy in *Scutellariae radix*. Therefore, baicalein was suggested to be used as an additional bioactive marker for the quality assurance of *Scutellariae radix* based on this study.

Data availability statement

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary material**.

Author contributions

L-PB and Z-HJ conceived and designed the whole study. Z-FY and RL designed the antiviral bioassay with SARS-CoV-2. JL performed the network pharmacology analysis, molecular docking, and LC-MS analysis and anti-inflammatory evaluations. JM, HJ, and RL performed all the antiviral experiments. LF and TX assisted the completion of some biological experiments. JL and JM carried out statistical analysis, chart drawing, and drafted the manuscript. L-PB and Z-HJ revised the manuscript. G-YZ and WZ gave comments on the revision of the manuscript. JG assisted the quantitative analysis of baicalein and baicalin in *Scutellariae radix*. L-PB and Z-HJ obtained the funding and supervised the whole study. All the authors approved the final version of the manuscript.

Funding

This research was financially supported by the Science and Technology Development Fund, Macau SAR (File no. 0023/2019/AKP, 0065/2020/A2, 0075/2019/AGJ, 0043/2020/AGJ, 0064/2020/A, and SKL-QRCM (MUST)-2020-2022). This work was also supported by the Emergency Key Program of Guangzhou Laboratory (Grant No. EKPG21-06). We also thank the Department of Science and Technology of Guangdong Province for the Support of GDST-FDCT Projects

(2021A0505080007) and Guangdong-Hong Kong-Macao Joint Laboratory of Respiratory Infectious Disease.

Acknowledgments

We thank the Department of Science and Technology of Guangdong Province for the Support of Guangdong-Hong Kong-Macao Joint Laboratory of Respiratory Infectious Disease.

Conflict of interest

Author JG was employed by Increasepharm (Hengqin) Institute Co., Ltd.

The remaining authors declare that the research was conducted in the absence of any commercial or financial

relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2022.988655/full#supplementary-material>

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