Check for updates

OPEN ACCESS

EDITED BY Ying Sun, Cornell University, United States

REVIEWED BY Karansher Singh Sandhu, Bayer Crop Science, United States Jacob Washburn, United States Department of Agriculture, United States

*CORRESPONDENCE Chen-Tuo Liao ctliao@ntu.edu.tw

SPECIALTY SECTION This article was submitted to Plant Breeding, a section of the journal Frontiers in Plant Science

RECEIVED 03 May 2022 ACCEPTED 01 July 2022 PUBLISHED 27 July 2022

CITATION

Chung P-Y and Liao C-T (2022) Selection of parental lines for plant breeding *via* genomic prediction. *Front. Plant Sci.* 13:934767. doi: [10.3389/fpls.2022.934767](https://doi.org/10.3389/fpls.2022.934767)

COPYRIGHT

© 2022 Chung and Liao. This is an open-access article distributed under the terms of the [Creative Commons](http://creativecommons.org/licenses/by/4.0/) [Attribution License \(CC BY\).](http://creativecommons.org/licenses/by/4.0/) The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

[Selection of parental lines for](https://www.frontiersin.org/articles/10.3389/fpls.2022.934767/full) plant breeding *via* genomic prediction

Ping-Yuan Chung^{1,2} and Chen-Tuo Liao^{1*}

¹Department of Agronomy, National Taiwan University, Taipei, Taiwan, ²Institute of Statistical Science, Academia Sinica, Taipei, Taiwan

A set of superior parental lines is imperative for the development of highperforming inbred lines in any biparental crossing program for crops. The main objectives of this study are to (a) develop a genomic prediction approach to identify superior parental lines for multi-trait selection, and (b) generate a software package for users to execute the proposed approach before conducting field experiments. According to different breeding goals of the target traits, a novel selection index integrating information from genomicestimated breeding values (GEBVs) of candidate accessions was proposed to evaluate the composite performance of simulated progeny populations. Two rice (*Oryza sativa* L.) genome datasets were analyzed to illustrate the potential applications of the proposed approach. One dataset applied to the parental selection for producing inbred lines with satisfactory performance in primary and secondary traits simultaneously. The other one applied to demonstrate the application of producing inbred lines with high adaptability to different environments. Overall, the results showed that incorporating GEBV and genomic diversity into a selection strategy based on the proposed selection index could assist in selecting superior parents to meet the desired breeding goals and increasing long-term genetic gain. An R package, called IPLGP, was generated to facilitate the widespread application of the approach.

KEYWORDS

genetic gain, genome-wide markers, mixed models, multiple-trait selection, selection index

Introduction

Parental line selection in plant breeding usually has two differing goals: (i) identify suitable parents for commercial hybrid varieties and (ii) identify suitable parents to develop inbred lines for subsequent breeding cycles [\(Gaynor et al., 2017\)](#page-11-0). For goal (i), the selection is based on an evaluation of hybrid performance [\(Wu et al., 2019\)](#page-12-0); however, for goal (ii), the selection is based on the performance of progeny populations [\(Chung and Liao, 2020\)](#page-11-1). In this study, we focus on the latter, i.e., parental line selection for the development of high-performing inbred lines using a biparental crossing scheme. Genomic selection based on the statistical method of genomic prediction (GP) has emerged as a promising approach to improving quantitative traits. The main concept of GP is to capture all the effects of quantitative trait loci by using highdensity DNA markers over an entire genome [\(Meuwissen et al., 2001\)](#page-11-2). Marker effects

are estimated using a GP model built from phenotypic and genotypic data of a training population. After model training, genomic estimated breeding values (GEBVs) for candidate accessions are estimated from their genotypic data alone. Genomic selection is then performed based on these resulting GEBVs [\(Heffner et al., 2010\)](#page-11-3).

Because genomic selection was specifically designed to predict complex traits such as the grain yield (YLD) of a crop, most published genomic selection studies have focused on single-trait approaches without exploiting information from multiple correlated agronomic traits [\(Schulthess et al.,](#page-11-4) [2016\)](#page-11-4). Yet, attaining most breeding goals usually requires the improvement of multiple traits. Besides a high YLD, an ideal cultivar is also expected to perform well in some secondary traits [\(Guo et al., 2020;](#page-11-5) [Sandhu et al., 2021\)](#page-11-6). For example, in rice, a low plant height (PH) can reduce the incidence of lodging and an early flowering time (FT) can reduce the cultivation period; therefore, simultaneously inheriting these traits of high YLD, low PH, and early FT is often sought by rice breeders. In practice, it would be desirable to develop appropriate GP approaches for multi-trait genomic selection (MTGS). In studies by [Jia and Jannink \(2012\)](#page-11-7), [Hayashi and Iwata \(2013\)](#page-11-8), and Guo et al. [\(2014\)](#page-11-9), they reported that prediction accuracy for a target trait with low heritability could be substantially improved when a correlated indicator trait with higher heritability was also included in the GP model. There are three types of multi-trait GP models commonly used for MTGS, including (i) linear mixed models [\(VanRaden, 2008;](#page-12-1) [Endelman, 2011\)](#page-11-10), (ii) Bayesian models [\(Perez and de los Campos, 2014;](#page-11-11) [Montesinos-Lopez et al., 2016\)](#page-11-12), and (iii) machine- and deep-learning models [\(Smith et al., 2013;](#page-11-13) [Lecun et al., 2015\)](#page-11-14). Recently, [Sandhu et al. \(2021\)](#page-11-6) compared the performance of the above models based on the prediction for grain yield and grain protein content in wheat (Triticum aestivum L.). The results of this article showed that multitrait machine- and deep-learning models were able to increase prediction accuracy and should be employed in large-scale breeding programs. To harness the benefits of MTGS for plant breeding, [Schulthess et al. \(2016\)](#page-11-4), [Fernandes et al. \(2018\)](#page-11-15), Ward et al. [\(2019\)](#page-12-2), and [Guo et al. \(2020\)](#page-11-5) used multi-trait prediction models to augment quantitative traits in various crops.

A selection index is often used in multi-trait breeding programs because it combines information from multiple traits, and incorporates the capacity of favorable levels of some traits to compensate for unfavorable levels in other traits (Dolan et al., [1996\)](#page-11-16). Several different selection indices can be used in MTGS, including those based on economic values, phenotypic correlations, genotypic correlations, and enhancing some traits while limiting other traits [\(Baker, 1986;](#page-11-17) Ceron-Rojas and Crossa, [2021\)](#page-11-18). [Schulthess et al. \(2016\)](#page-11-4) compared the prediction accuracies of different selection indices using various prediction methods and recommended implementing a single-trait GP model by treating a selection index itself as a new single trait. [Covarrubias-Pazaran et al. \(2018\)](#page-11-19) demonstrated that the use

of multi-trait genomic best linear unbiased prediction (multitrait GBLUP) models could improve selection accuracy and subsequently lead to more reliable selection indices. Notably, those studies cited above focused on prediction accuracy for target traits or selection indices. Alternatively, Lehermeier et al. [\(2017\)](#page-11-20) emphasized that genetic gain can be increased considerably when the crosses are selected based on their genomic usefulness function compared to selection based on mean GEBVs. In this respect, [Yao et al. \(2018\)](#page-12-3) combined GP with Monte Carlo simulations to select superior parents for wheat breeding. The authors applied a selection index to incorporate YLD and two crop quality-related traits, and calculated a usefulness function based on the selection index values of simulated progeny populations. Their findings also showed that utilizing the usefulness function for parental selection is capable of providing higher genetic gain than the use of a mid-parent GEBV.

Both [Lehermeier et al. \(2017\)](#page-11-20) and [Yao et al. \(2018\)](#page-12-3) cautioned that parental selection strategies should not focus solely upon truncation selection that selects the top fraction of candidate accessions with the top GEBVs. To preserve genetic variation to maximize selection responses in progeny populations, plant breeders should avoid selecting closely related parental lines in the base population. Accordingly, [Chung and Liao \(2020\)](#page-11-1) proposed strategies whereby both GEBV and genomic diversity (GD) were taken into account for single-trait selection. However, such single-trait selection strategies can result in different choices of parental lines for different target traits, and this may cause confusion in practical applications. The improvement of genetic stocks usually warrants considering multiple traits at once, because economic value and net genetic merits depend on almost all the traits responsible for the desired crop phenotype [\(Falconer and Mackay, 1996\)](#page-11-21).

In this study, our aim was to develop and validate a useful genomic prediction approach to select parental lines for producing progeny populations with superior performance in multiple target traits. To do this, a multi-trait GBLUP model was used to simultaneously predict normalized GEBVs of the multiple target traits. A new selection index integrating information from the normalized GEBVs was then proposed to evaluate the composite performance of simulated progeny populations. Three different strategies considering GEBV and/or GD were compared through a stochastic simulation approach for producing progeny populations. Finally, an R package called IPLGP [\(Chung and Liao, 2022\)](#page-11-22) was generated in conducting this study.

Materials and methods

Tropical rice genome dataset: The rice (Oryza sativa L.) genome dataset presented in [Spindel et al. \(2015\)](#page-11-23)

was analyzed first. This dataset contains 73,147 singlenucleotide polymorphism (SNP) markers and 363 elite breeding lines belonging to indica or indica-admixed groups. The phenotypic data include 4 years (from 2009 to 2012), two seasons per year (dry and wet), and YLD, PH, and FT for each season. Unfortunately, PH data for the 2009 wet season were not available. Phenotypic values of 35 of the 363 breeding lines were also missing; hence, adjusted means derived from 328 breeding lines were used in our study. The adjusted means were obtained using the residuals derived separately for each trait by the following model:

$$
y_{ijk} = \mu + A_i + S_j + (AS)_{ij} + B_k + e_{ijk}
$$
 (1)

where y_{ijk} is the phenotypic value of the trait at year *i*, season j and block k ; A_i is the fixed effect of year i ; S_j is the fixed effect of season j; $(AS)_{ij}$ is the interaction effect between year *i* and season *j*; B_k is the fixed effect of block k ; and e_{ijk} is the residual. One SNP marker was randomly chosen per 0.1-cM interval over each chromosome because [Spindel et al. \(2015\)](#page-11-23) had shown that the subset of the full markers was efficient enough for genomic selection for this collection of rice germplasm. This resulted in 10,772 out of the 73,147 SNP markers being used for this example. The SNP genotype at each locus was coded as −1, 0, or 1, where 1 indicates homozygosity for the major allele, −1 indicates homozygosity for the minor allele, and 0 indicates heterozygosity. After the SNP coding, any missing loci were imputed as 1.

44k rice genome dataset: The rice genome dataset is presented in [Zhao et al. \(2011\)](#page-12-4). It was originally collected for a genome-wide association study and was reanalyzed here. It contains 44,100 SNP markers and 36 traits of 413 accessions, and this dataset features a strong subpopulation structure. All SNP markers with a missing rate > 0.05 and a minor allele frequency < 0.05 were first removed from the dataset. This left 34,233 SNP markers. To avoid redundant SNP markers in calculating the genomic relationship between individuals, about one-third of these SNP markers (11,043 out of the 34,233) evenly distributed over each chromosome were selected. Their SNP coding was performed as described above for the tropical rice dataset. Only those 300 of the 413 accessions with no missing FT data from all the three locations— Arkansas (FT-Ark), Faridpur (FT-Far), and Aberdeen (FT-Abe)—were used here for building the required multi-trait GBLUP model. To simulate the genotypic data of progeny populations for both the rice datasets, the Gramene Annotated Nipponbare Sequence provided by [Youens-Clark et al. \(2011\)](#page-12-5) was used to estimate recombination rates between two adjacent SNP markers.

The multi-trait GBLUP model for fitting normalized phenotypic values

The target traits of interest were classified into three types according to their breeding goals. The larger-the-better: the larger phenotypic value is desirable; the smaller-the-better: the smaller phenotypic value is desirable; and the nominal-the-best: the nominal value is the best because it is the one that satisfies the target set by the plant breeder. Therefore, a given phenotypic value that falls around the nominal value is desirable for this last type. For example, FT may be set to a specific time for balancing the duration of cultivation and the vegetative growth period. Accordingly, the vectors of phenotypic values for traits were first normalized as follows. Let $w_i = (y_i - \delta \mathbf{1}_n)/s_i$, where δ is set to the sample mean of the phenotypic values for both the larger-thebetter and the smaller-the-better types, and to the desired target value for the nominal-the-best type; s_i is the sample standard deviation of those phenotypic values; I_n is the vector of order *n* with all elements equal to 1; and $y_i = [y_{i1}, \ldots, y_{in}]^T$ is the vector of phenotypic values for $i = 1, 2, ..., t$. Here *n* is the number of individuals in the training population, and t is the number of target traits.

Let

$$
w = \begin{bmatrix} w_1 \\ \vdots \\ w_t \end{bmatrix}; \mu = \begin{bmatrix} \mu_1 \\ \vdots \\ \mu_t \end{bmatrix}; \ g = \begin{bmatrix} g_1 \\ \vdots \\ g_t \end{bmatrix}; \text{ and } e = \begin{bmatrix} e_1 \\ \vdots \\ e_t \end{bmatrix}
$$

where μ_i , g_i , and e_i , respectively, denote the general mean, the vector of genomic values, and the vector of random errors for trait i. The additive effects multi-trait GBLUP model is given as:

$$
w = \mu \otimes I_n + g + e \tag{2}
$$

where ⊗ denotes the Kronecker product [\(Searle, 1982\)](#page-11-24). It is assumed that **g** and **e** are mutually independent and separately follow a multivariate normal distribution, as denoted by

$$
g \sim \text{MVN} (0, \Sigma_A \otimes K)
$$

and

$$
e \sim \text{MVN} (0, \Sigma_e \otimes I_n)
$$

where **0** is a zero vector, Σ_A is the genetic variance-covariance matrix for additive effects among the t target traits, K is a genomic relationship matrix for additive effects among the *n* individuals, Σ_e is the residual variance-covariance matrix among the t target traits, and I_n is the identity matrix of order *n*. Here, Σ_A and Σ_e can be represented as

$$
\Sigma_A = \begin{bmatrix} \sigma_{A_1}^2 & \cdots & \sigma_{A_{1t}} \\ \vdots & \ddots & \vdots \\ \sigma_{A_{t1}} & \cdots & \sigma_{A_t}^2 \end{bmatrix} \text{ and } \Sigma_e = \begin{bmatrix} \sigma_{e_1}^2 & \cdots & \sigma_{e_{1t}} \\ \vdots & \ddots & \vdots \\ \sigma_{e_{1t}} & \cdots & \sigma_{e_t}^2 \end{bmatrix}
$$

where $\sigma_{A_i}^2$ and $\sigma_{e_i}^2$ are the respective variances for the additive effects and the random errors for trait *i*, and $\sigma_{A_{ii}}$ and $\sigma_{e_{ii}}$ are the corresponding covariances between traits i and j . The genomic relationship matrix was calculated as $K = MM^T/p$, where M is the marker coding matrix regarding the additive effects, and p is the number of markers.

Let $\hat{\mu}$ be the best linear unbiased estimate (BLUE) for μ , and \hat{g} be the best linear unbiased predictor (BLUP) for **g**, then $\hat{\mu}$ and \hat{g} can be obtained from the following linear mixed model equations [\(Henderson, 1975\)](#page-11-25):

$$
\begin{bmatrix} nI_t & I_t \otimes I_n^T \\ I_t \otimes I_n & I_{nt} + (\Sigma_e \Sigma_A^{-1}) \otimes K^{-1} \end{bmatrix} \begin{bmatrix} \hat{\mu} \\ \hat{\mathbf{g}} \end{bmatrix} = \begin{bmatrix} (I_t \otimes I_n^T)w \\ w \end{bmatrix} . (3)
$$

The restricted maximum likelihood estimates (REMLs) for Σ_A and Σ_e were plugged into Eq. (3) to generate $\hat{\mu}$ and \hat{g} . The R package sommer [\(Covarrubias-Pazaran, 2016\)](#page-11-26) was used to calculate these estimates from training data.

Predicting GEBVs for simulated progeny populations

The performance of a set of parental lines was evaluated based on the GEBVs of their progeny populations. Genotypic data of the progeny populations were generated using the simulation approach of [Chung and Liao \(2020\)](#page-11-1). This was mainly based on the mapping function of recombination rate on linkage distance between two adjacent markers as presented in [Haldane](#page-11-27) [\(1919\)](#page-11-27). The required GEBVs were then predicted using the multi-trait GBLUP model of (2). Let h_i denote the vector of genomic values for trait i in a simulated progeny population, and **K**pt denote the genomic relationship matrix between the simulated progeny population and the training population. From [Henderson \(1977\)](#page-11-28), the BLUP for **h**i is given by

$$
\hat{\boldsymbol{h}}_i = \boldsymbol{K}_{pt} \boldsymbol{K}^{-1} \, \hat{\boldsymbol{g}}_i \tag{4}
$$

where \hat{g}_i is the BLUP for the vector of genomic values of trait i obtained from Eq. (3). The GEBVs for the simulated progeny population are then predicted by $\hat{\mu}_i \bm{{1}}_n\!+\!\hat{\bm{h}}_i$, where $\hat{\mu}_i$ is the BLUE of μ_i obtained from Eq. (3), for $i = 1, 2, \ldots, t$.

The selection index

For a particular individual, the selection index below was used to integrate its normalized GEBVs for the multiple target traits:

$$
SI = \sum_{i=1}^{t} w_i Z_i \tag{5}
$$

where w_i is a pre-specified weight for trait i subject to the constraint that $\sum_{i=1}^{t} w_i = 1$; and Z_i is designated as $GEBV_i$ for the larger-the-better case, as $-GEBV_i$ for the smaller-the-better case, and as $-|GEBV_i|$ (the absolute value of $GEBV_i$) for the nominal-the-best case. The selection index conveys an overall performance score for the individual. Note that the normalized $GEBV_i$ are scalars with no measuring units. The larger the selection index, the better the composite performance.

Procedure for selecting superior parental lines

For the tropical rice dataset, the aim was to select a set of parental lines whose progeny populations had high YLD, low PH, and low FT. For the 44k rice dataset, the breeding goal was assumed to identify a set of superior accessions that would produce inbred lines with an FT as close as possible to the nominal value set as 80 days at all three locations (FT-Ark, FT-Far, and FT-Abe). The resulting inbred lines would be anticipated to have high adaptability to the three different locations. The selection procedure can be described as follows.

Step 1

All available phenotypic values in each dataset were normalized as described above. The ensuing normalized data were used to build the multi-trait GBLUP model given by Eq. (2). The trained multi-trait GBLUP model predicted the normalized GEBVs of the target traits for each dataset; then the corresponding selection index values were obtained for all the accessions in the candidate population.

The selection index integrating the normalized GEBVs of the three target traits in the tropical rice dataset was defined this way:

$$
SI(tropical) = w_1 GEBV_{YLD} - w_2 GEBV_{PH} - w_3 GEBV_{FT} (6)
$$

where w_1 , w_2 , and w_3 are pre-specified index weights. Note the minus signs applied in the equation for PH and FT because smaller values are preferable for these two traits. The index weights w_1 , w_2 , and w_3 were respectively specified as 0.6, 0.2, and 0.2. For the sake of contrast, another setting of 1, 0, and 0 was used that corresponded to the single-trait selection for YLD. To compare the improvement of the strategy changed from single-trait selection to multi-trait selection, an index was defined as follows:

$$
IR = [GEBV of (0.6, 0.2, 0.2) - GEBV of (1, 0, 0)]
$$

$$
\div GEBV of (1, 0, 0) \times 100\%.
$$
 (7)

The selection index for the 44k rice dataset was defined as:

$$
SI (44k) = -w_1|GEBV_{FT-Ark}| - w_2|GEBV_{FT-Far}|
$$

$$
-w_3|GEBV_{FT-Abe}|
$$
 (8)

where the index weights w_1 , w_2 , and w_3 were equally set to be 1/3.

GEBV averages of the best individuals at each generation for the tropical rice dataset. GEBV-O, Subset of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal *D*-scores chosen from the candidate set S^c ; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of \mathcal{S}_{c} . YLD, grain yield; PH, plant height; FT, flowering time.

Step 2

Based on the normalized GEBVs of the candidate accessions obtained from Step 1, three strategies were implemented to select a subset of 10 parental lines from the candidate population. (i) The GEBV only (GEBV-O) strategy, which selected the top 10 accessions with the highest selection index values. (ii) The GD only (GD-O) strategy, which searched for an optimal subset of 10 accessions from S_c that is the set composed of those accessions whose selection index values were above average. This resulting optimal subset achieved the maximal D-score, where the D-score is the determinant of the genomic relationship matrix corresponding to the selected accessions, and it was used to measure the genomic diversity of the selected accessions [\(Chung and Liao, 2020\)](#page-11-1). (iii) The GEBV-GD strategy, which considered both GEBV and GD. This strategy retained the top two accessions with the highest selection index values, and then searched for another eight accessions among the remainder of S_c . The resulting 10 accessions achieved the maximal D-score.

Step 3

For each subset of 10 parental lines generated from Step 2, any two parental lines were crossed to produce $45 F₁$ hybrids, and then each F_1 hybrid produced 60 individuals by selfpollinating and applying the simulation approach of Chung and Liao [\(2020\)](#page-11-1); hence, a total of $45 \times 60 = 2,700$ F₂ individuals. Again, the GEBVs of F_2 individuals were calculated via the trained multi-trait GBLUP models in Step 1. The individuals with the top 45 selection index values in the F₂ generation were selected, and those produced 2,700 F_3 individuals (each F_2 individual produced 60 F₃ individuals).

Step 4

The procedure of generating genotypic data, predicting the GEBVs, and selecting the top 45 individuals was performed repeatedly, to produce 2,700 F10 individuals presumed to constitute a fixed population of inbred lines.

Step 5

For the final 2,700 F_{10} individuals generated from each subset of 10 parental lines, the best F_{10} inbred line with the highest index value was identified. The above analysis procedure was repeated 30 times to obtain the best F_{10} inbred lines from each repetition per strategy. To evaluate improvements in both the larger-the-better and the smaller-the-better target traits attained by each strategy, the genetic gain was calculated this way:

$$
genetic gain = \overline{GEBV}_{F_{10}} - \overline{GEBV}_{P}
$$
 (9)

where $\overline{GEBV}_{F_{10}}$ is the GEBV average among the 2,700 F₁₀ individuals, and \overline{GEBV} is the GEBV average among the 10

TABLE 1 GEBV averages of the best F_{10} inbred lines for the tropical rice dataset.

¹GEBV-O, Subset of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal D-scores chosen from the candidate set S_c ; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c .

²YLD, grain yield; PH, plant height; FT, flowering time.

Different lowercase letters indicate significant differences between the strategies for a given trait ($p < 0.01$; LSD test).

 $IR = [GEBV of (0.6, 0.2, 0.2) - GEBV of (1, 0, 0)] \div GEBV of (1, 0, 0) \times 100\%$.

selected parental lines. For the nominal-the-best target trait, genetic gain was defined as follows:

$$
\text{genetic gain (nominal)} = \text{mean } (|\text{GEBV}_{F_{10}} - \delta|)
$$
\n
$$
-\text{mean } (|\text{GEBV}_{p} - \delta|) \tag{10}
$$

where *mean* ($|GEBV_{F_{10}} - \delta|$) is the averaged deviation of the F₁₀ inbred lines from the nominal value δ , and *mean* ($|GEBV_p - \delta|$) is the averaged deviation of the parental lines from δ .

The GEBVs of the target traits on their original measuring units were obtained via back-transformation from their normalized GEBVs. Furthermore, pairwise comparisons among the GEBV averages for each trait were carried out using the least significant difference (LSD) test.

Results

Hereafter, GEBVs are reported for the target traits based on their original scales.

Tropical rice genome dataset

The GEBV averages of the best individuals from the 30 repetitions per generation are displayed in [Figure 1.](#page-4-0) Evidently, the sought-after GEBV average decreased from parental generation to F_1 generation. In contrast to YLD, which improved from F_1 down through the F_{10} generation under every

TABLE 2 GEBV average for parental lines, GEBV average for F_{10} inbred lines, and genetic gain for the tropical rice dataset.

¹GEBV-O, Subset of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal D-scores chosen from the candidate set S.; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c .

²Different lowercase letters indicate significant differences among the strategies for a given trait ($p < 0.01$; LSD test).

strategy tested, the desirability in the GEBV average for both PH and FT improved going from the F_1 to F_3 or F_4 generation, but gradually declined in later generations. A strategy with the index weight of 0.2 provided the best F_{10} inbred lines, these having a better PH and FT than those generated from the same strategy whose index weight was 0.

The end-point of the GEBV averages for the best F_{10} individuals from the 30 repetitions and the improvement of Eq. (7) are presented in [Table 1.](#page-5-0) For any of the three strategies GEBV-O, GD-O, and GEBV-GD, using selection index weights of 1, 0, 0 (i.e., the single-trait selection for YLD) always outperformed 0.6, 0.2, and 0.2 for YLD in terms of statistical significance. Conversely, any strategy with selection index weights of 0.6, 0.2, and 0.2 led to greater improvement in the two secondary target traits PH and FT than when using 1, 0, and 0 instead. The GEBV-GD had the largest GEBV average for YLD among the three strategies when using index weights of 1, 0, and 0, and the GD-O had the largest one when 0.6, 0.2, and 0.2 index weights were used. However, for YLD, there was no significant difference among the three strategies when applying either set of index weights. The GD-O performed worst with respect to GEBV averages for both PH and FT among the three strategies, with index weights of 0.6, 0.2, and 0.2. For the GEBV-O strategy, the secondary traits of PH and FT were respectively improved by 5.27 and 6.36%, but the primary trait of YLD fell by 2.83%,

when the index weights 1, 0, and 0 were changed to 0.6, 0.2, and 0.2. The improvements obtained under GD-O amounted to 5.90% (gain in PH), 3.12% (gain in FT), and 2.85% (loss in YLD). Moreover, corresponding percentages for the GEBV-GD were 4.62% (gain in PH), 5.21% (gain in FT), and 3.13% (loss in YLD). Consequently, for this dataset, either GEBV-GD or GEBV-O with index weights of 0.6, 0.2, or 0.2 may be used to select a suitable set of parental lines for producing highperforming inbred lines simultaneously featuring high YLD, low PH, and low FT traits.

The average genetic gain for a given target trait, as calculated by Eq. (9), from the 30 repetitions appears in [Table 2.](#page-6-0) Based on these results, the \overline{GEBV} can be ranked as $GEBV-O >$ GEBV-GD > GD-O in descending desirability when using selection index weights of 0.6, 0.2, 0.2 or 1, 0, 0. The ranking of genetic gains is reversed to GD-O > GEBV-GD > GEBV-O with any of the two index weights used for all the three target traits, except that GEBV-GD > GD-O > GEBV-O ensues with index weights of 1, 0, and 0 for PH. Nevertheless, the genetic gain in PH from GEBV-GD (−8.25 cm) did not differ significantly from GD-O (-7.73 cm). Any strategy applied had a greater genetic gain in YLD with index weights of 1, 0, and 0 than 0.6, 0.2, and 0.2. Conversely, any strategy had a greater genetic gain on both PH and FT with index weights of 0.6, 0.2, and 0.2 than 1, 0, and 0, except in the case of

the candidate set S_c; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c . FT-Ark, flowering time in Arkansas; FT-Far, flowering time in Faridpur; FT-Abe, flowering time in Aberdeen.

GEBV-O upon PH (the former had −3.27 cm and the latter had −5.59 cm).

44k rice genome dataset

The GEBV averages of the best individuals from the 30 repetitions per generation are shown in [Figure 2,](#page-7-0) for which the two GEBV averages of the parental and F_{10} generations are in [Table 3.](#page-8-0) From [Figure 2,](#page-7-0) it is interesting to see that all the curves approached the nominal value of 80 days. From [Table 3,](#page-8-0) the three end-point values of $\overline{GEBV}_{F_{10}}$ for $\overline{GEBV}_{F_{10}}$ for FT-Far

TABLE 3 GEBV averages of the best F_{10} inbred lines for the 44k rice dataset based on the index weights of 1/3, 1/3, and 1/3.

Train^1	Strategy ²	$\overline{GEBV}p$	$\overline{GEBV}_{F_{10}}$ ³
FT-Ark	GEBV-O	82.141	79.983 ^c
	$GD-O$	84.131	79.979 ^c
	GEBV-GD	84.256	79.969°
FT-Far	GEBV-O	72.776	74.497 ^d
	$GD-O$	73.252	74.588 ^d
	GEBV-GD	73.474	74.718 ^d
FT-Abe	GEBV-O	85.435	81.797 ^b
	$GD-O$	92.561	81.928 ^b
	GEBV-GD	91.931	82.569 ^a

¹FT-Ark, flowering time in Arkansas; FT-Far, flowering time in Faridpur; FT-Abe, flowering time in Aberdeen.

²GEBV-O, Subset of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal D-scores chosen from the candidate set S_c ; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c .

³Different lowercase letters indicate significant differences between the strategies for a given trait (p < 0.01; LSD test).

are about 74 days, this slightly less $\overline{GEBV}_{F_{10}}$ values of FT-Far differed significantly from those of FT-Ark and FT-Abe, and the FT-Far seems less improved than either FT-Ark or FT-Abe. At these three locations, based on the LSD testing for $\overline{GEBV}_{F_{10}}$, the GEBV-O, or GD-O implemented with equal index weights can be used to select a set of parental lines for producing inbred lines with FT being close to the nominal value of 80 days.

The genetic gains for these nominal-the-best traits calculated using Eq. (10) are displayed in [Table 4.](#page-8-1) Evidently, FT-Far undergoes a relatively smaller genetic gain among the three target traits, and the GD-O and GEBV-GD strategies lead to greater genetic gain than does GEBV-O.

Discussion

As suggested in [Figure 1](#page-4-0) for the tropical rice dataset, it is reasonable to infer that the secondary traits of PH and FT did not improve with the primary trait of YLD when using the single-trait selection strategies for YLD; i.e., those with the index weights of 1, 0, 0. Conversely, employing multi-trait selection strategies that used index weights of 0.6, 0.2, and 0.2 resulted in some improvement for both PH and FT. However, these results do not generally imply that grain yield will be improved more by not selecting for other traits simultaneously. There were several studies revealing that grain yield can be improved by multitrait instead of single-trait selection. For example, selecting for grain yield and nutation content in cereal crops simultaneously [\(Jia and Jannink, 2012;](#page-11-7) [Schulthess et al., 2016\)](#page-11-4), and selecting for grain yield and yield-related traits such as harvest index, spike fertility, and thousand grain weight in wheat simultaneously [\(Guo et al., 2020\)](#page-11-5).

The FT at three different locations in the 44k rice dataset was used to demonstrate that the proposed approach can be applied to select parents for producing inbred lines with

TABLE 4 GEBV average for parental lines, GEBV average for F_{10} inbred lines, and genetic gain for the 44k rice dataset based on the index weights of 1/3, 1/3, and 1/3.

Train^1	Strategy ²	Mean of $ \overline{GEBV}_P - \delta $	Mean of $ \overline{GEBV}_{F_{10}} - \delta $	Genetic gain ³ (nominal)
FT-Ark	GEBV-O	2.224	0.044^a	$-2.180^{\rm d}$
	$GD-O$	4.137	0.085^{a}	$-4.053c$
	GEBV-GD	4.256	0.073 ^a	-4.184 ^c
FT-Far	GEBV-O	7.224	5.503 ^d	-1.721 ^{de}
	$GD-O$	6.748	5.412 ^d	-1.336^e
	GEBV-GD	6.526	5.282 ^d	-1.244 ^e
FT-Abe	GEBV-O	5.435	1.797 ^b	-3.638°
	$GD-O$	12.561	1.956 ^c	-10.606^a
	GEBV-GD	11.931	2.569 ^c	$-9.361^{\rm b}$

 $^1\rm{FT-Ark},$ flowering time in Arkansas; FT-Far, flowering time in Faridpur; FT-Abe, flowering time in Aberdeen.

²GEBV-O, Subset of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal D-scores chosen from the candidate set S_c ; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c .

³Different lowercase letters indicate significant differences between the strategies for a given trait ($p < 0.01$; LSD test).

of the 10 accessions with the highest selection index values; GD-O, Subset of the 10 accessions with the maximal *D*-scores chosen from the candidate set S_c ; GEBV-GD, Subset of the top two accessions with the highest selection index values, and another eight accessions chosen from the reminder of S_c. FT-Ark, flowering time in Arkansas; FT-Far, flowering time in Faridpur; FT-Abe, flowering time in Aberdeen.

high adaptability to different environments. As suggested in [Figure 2,](#page-7-0) the performance of FT-Far may be further improved by increasing its index weight. Hence, we modified the index weights to 0.3, 0.4, and 0.3, and re-ran the procedure; these results are displayed in [Figure 3.](#page-9-0) Evidently, the curve for FT-Far got a little closer to the target value of 80 days compared with [Figure 2;](#page-7-0) however, the other two traits were also affected, incurring some diminished improvement, particularly for FT-Abe. There is no golden standard for assigning the index weights to the traits because those traits can change from time to time or vary from one location to another in breeding programs (Ceron-Rojas and Crossa, [2021\)](#page-11-18). Fortunately, the user can fine-tune the index weights and re-run the procedure easily using our R package, until the simulated progeny populations satisfy the desired breeding goals.

Incorporating the multi-trait GBLUP model and the selection index into the framework for single-trait selection presented in our previous article [\(Chung and Liao, 2020\)](#page-11-1), we extended this multi-trait selection approach. Our proposed multi-trait approach was also able to conduct single-trait selection by assigning the index weight as 1 for the trait of interest, and 0 for the remaining traits. This multi-trait model-based approach is advantageous over those selected for an independent trait because it takes into account the information among the correlated traits. Moreover, our proposed approach has merit over the approach promoted by [Schulthess et al. \(2016\)](#page-11-4), in which the selection index was treated as a new trait using a single-trait model for selection. That is, our proposed approach enabled us to assess the performance of each target trait in the progeny populations.

When conducting a breeding program to improve several quantitative traits at once, selection using a selection index is long known to be more efficient than that relying on independent culling levels or tandem selection [\(Hazel and Lush,](#page-11-29) [1942\)](#page-11-29). Recently, [Ceron-Rojas and Crossa \(2021\)](#page-11-18) provided a review on the statistical theory of linear selection index from phenotypic to genomic selection, in which a linear selection index was defined as a linear combination of unobservable individual traits' breeding values, weighted by the trait economic values. The proposed selection index in this study basically meets the requirements of the definition. Overall, this proposed selection index is arguably a straightforward and easy way to evaluate the composite performance of individuals. The GD quantified by the D-score is kind of different from the genetic diversity quantified by the genetic variances of traits. The former used in our study was calculated from genotypic data of individuals alone, but the latter used in the genomic usefulness function [\(Lehermeier et al., 2017;](#page-11-20) [Yao et al., 2018\)](#page-12-3) was estimated from both phenotypic and genotyped data. This means that the GD measures the genomic information for

a set of individuals and is independent of the traits under investigation. [Anderson et al. \(1998\)](#page-11-29) found that the introduction of the dominance variance has only a small positive effect on the selection response. As discussed in [Ceron-Rojas and Crossa](#page-11-18) [\(2021\)](#page-11-18), the multi-trait GBLUP model used in our study assuming that only additive effects are transmitted from generation to generation seems acceptable.

We generated the R package IPLGP to facilitate the wider application of the proposed approach. A user can install the package from the R official repository CRAN or GitHub. IPLGP provides the required R functions to replicate the results of this study. Note that a user needs to provide the linkage distances between SNP markers when running the procedure for her/his dataset. A training population consisting of both phenotypic and genotypic data is needed to build the required multi-trait GBLUP model. If historical phenotypic data are not available, a pilot experiment is recommended to phenotype a set of individuals, which can be determined using an optimization algorithm [\(Ou and Liao, 2019\)](#page-11-30).

We addressed the crucial issue of how adequately incorporating genomic diversity into conventional truncation selection could improve the likelihood of identifying superior parental lines for multiple traits in plant breeding efforts. More importantly, we have shown that combining GP with simulated progeny populations could help breeders to discover superior parental lines before conducting field experiments. However, the phenotypic value of a trait is affected by the genotype (G), environment (E), and their $G \times E$ interaction. In reality, the local environment can significantly influence the performance of progeny populations during the growth period of each generation until they reach the F_{10} generation. As such, parental lines selected from our simulation study may not necessarily perform as expected. Therefore, conducting more field experiments with various plant species to validate our study's key findings would be worthwhile.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author.

Author contributions

P-YC: data curation, investigation, software, and validation. C-TL: conceptualization, project administration, supervision, writing the original draft, and review and editing. All authors contributed to the article and approved the submitted version.

Funding

This research was supported by the Ministry of Science and Technology, Taiwan (grant number MOST 110-2118-M-002- 002-MY2).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships

References

Anderson, E., Spanos, K., Mullin, T., and Lindgren, D. (1998). Phenotypic selection compared to restricted combined index selection for many generations. Silva Fennica 32, 111–120. doi: [10.14214/sf.689](https://doi.org/10.14214/sf.689)

aker, R. J. (1986). Selection Indices in Plant Breeding. Boca Raton: CRC Press.

Ceron-Rojas, J. J., and Crossa, J. (2021). The statistical theory of linear selection indices from phenotypic to genomic selection. Crop. Sci. 62, 537-563. doi: [10.1002/csc2.20676](https://doi.org/10.1002/csc2.20676)

Chung, P. Y., and Liao, C. T. (2020). Identification of superior parental lines for biparental crossing via genomic prediction. PLoS ONE 15, e0243159. doi: [10.1371/journal.pone.0243159](https://doi.org/10.1371/journal.pone.0243159)

Chung, P. Y., and Liao, C. T. (2022). IPLGP: Identification of Parental Lines via Genomic Prediction. R package version 2.0.2. Available online at: [http://CRAN.R](http://CRAN.R-project.org/package=IPLGP)[project.org/package=IPLGP](http://CRAN.R-project.org/package=IPLGP)

Covarrubias-Pazaran, G. (2016). Genome-assisted prediction of quantitative traits using the R package sommer. PLOS ONE 11, e0156744. doi: [10.1371/journal.pone.0156744](https://doi.org/10.1371/journal.pone.0156744)

Covarrubias-Pazaran, G., Schlautman, B., Diaz-Garcia, L., Grygleski, E., Polashock, J., Johnson-Cicalese, J., et al. (2018). Multivariate GBLUP improves accuracy of genomic selection for yield and fruit weight in biparental populations of Vaccinium macrocarpon Ait. Front. Plant Sci. 9, 1310. doi: [10.3389/fpls.2018.01310](https://doi.org/10.3389/fpls.2018.01310)

Dolan, D. J., Stuthman, D. D., Kolb, F. L., and Hewings, A. D. (1996). Multiple trait selection in a recurrent selection population in oat (*Avena sativa* L.). *Crop Sci.*
36, 1207–1211. doi: [10.2135/cropsci1996.0011183X003600050023x](https://doi.org/10.2135/cropsci1996.0011183X003600050023x)

Endelman, J. B. (2011). Ridge regression and other kernels for genomic slection with R package rrBLUP. Plant Genome 4, 250–255. doi: [10.3835/plantgenome2011.08.0024](https://doi.org/10.3835/plantgenome2011.08.0024)

Falconer, D. S., and Mackay, T. F. C. (1996). Introduction to Quantitative Genetics. 4th ed. San Francisco: Benjamin-Cummings Pub Co.

Fernandes, S. B., Dias, K. O. G., Ferreira, D. F., and Brown, P. J. (2018). Efficiency of multi-trait, indirect, and trait-assisted genomic selection for improvement of biomass sorghum. Theor. Appl. Genet. 131, 747–755. doi: [10.1007/s00122-017-3033-y](https://doi.org/10.1007/s00122-017-3033-y)

Gaynor, R. C., Gorjanc, G., Bentley, A. R., Ober, E. S., Howell, P., Jackson, R., et al. (2017). A two-part strategy for using genomic selection to develop inbred lines. Crop Sci. 57, 2372–2386. doi: [10.2135/cropsci2016.09.0742](https://doi.org/10.2135/cropsci2016.09.0742)

Guo, G., Zhao, F., Wang, Y., Zhang, Y., Du, L., and Su, G. (2014). Comparison of single-trait and multiple-trait genomic prediction models. BMC Genet. 15, 30. doi: [10.1186/1471-2156-15-30](https://doi.org/10.1186/1471-2156-15-30)

Guo, J., Khan, J., Pradhan, S., Shahi, D., Khan, N., Avci, M., et al. (2020). Multitrait genomic prediction of yield-related traits in US soft wheat under variable water regimes. Genes 11, 1270. doi: [10.3390/genes11111270](https://doi.org/10.3390/genes11111270)

Haldane, J. B. S. (1919). The combination of linkage values and the calculation of distance between the loci for linked factors. Genetics 8, 299–309. doi: [10.1007/BF02983270](https://doi.org/10.1007/BF02983270)

Hayashi, T., and Iwata, H. (2013). A Bayesian method and its variational approximation for prediction of genomic breeding values in multiple traits. BMC Bioinformatics 14, 1–14. doi: [10.1186/1471-2105-14-34](https://doi.org/10.1186/1471-2105-14-34)

that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Hazel, L. N., and Lush, J. L. (1942). The efficiency of three methods of selection. J. Hered. 33, 393–399. doi: [10.1093/oxfordjournals.jhered.a105102](https://doi.org/10.1093/oxfordjournals.jhered.a105102)

Heffner, E. L., Lorenz, A. J., Jannink, J. L., and Sorrells, M. E. (2010). Plant breeding with genomic selection: gain per unit time and cost. Crop Sci. 50, 1681–1690. doi: [10.2135/cropsci2009.11.0662](https://doi.org/10.2135/cropsci2009.11.0662)

Henderson, C. R. (1975). Best linear unbiased estimation and prediction under a selection model. Biometrics 31, 423–447. doi: [10.2307/2529430](https://doi.org/10.2307/2529430)

Henderson, C. R. (1977). Best linear unbiased prediction of breeding values not in the model for records. J. Dairy Sci. 60, 783-787. doi: [10.3168/jds.S0022-0302\(77\)83935-0](https://doi.org/10.3168/jds.S0022-0302(77)83935-0)

Jia, Y., and Jannink, J. L. (2012). Multiple-trait genomic selection methods increase genetic value prediction accuracy. Genetics 192, 1513–1522. doi: [10.1534/genetics.112.144246](https://doi.org/10.1534/genetics.112.144246)

Lecun, Y., Bengio, Y., and Hinton, G. (2015). Deep learning. Nature 521, 436–444. doi: [10.1038/nature14539](https://doi.org/10.1038/nature14539)

Lehermeier, C., Teyssedre, S., and Schon, C. C. (2017). Genetic gain increases by applying the usefulness criterion with improved variance prediction in selection of crosses. Genetics 207, 1651–1661. doi: [10.1534/genetics.117.300403](https://doi.org/10.1534/genetics.117.300403)

Meuwissen, T. H. E., Hayes, B. J., and Goddard, M. E. (2001). Prediction of total genetic value using genome-wide dense marker maps. Genetics 157, 1819–1829. doi: [10.1093/genetics/157.4.1819](https://doi.org/10.1093/genetics/157.4.1819)

Montesinos-Lopez, O. A., Montesinos-Lopez, A., Crossa, J., Toledo, F. H., Perez-Hernandez, O., Eskridge, K. M., et al. (2016). A genomic Bayesian multitrait and multi-environment model. G3 Genes Genomes Genet. 6, 2725–2774. doi: [10.1534/g3.116.032359](https://doi.org/10.1534/g3.116.032359)

Ou, J. H., and Liao, C. T. (2019). Training set determination for genomic selection. Theor. App. Genet. 132, 2781–2792. doi: [10.1007/s00122-019-03387-0](https://doi.org/10.1007/s00122-019-03387-0)

Perez, P., and de los Campos, G. (2014). Genome-wide regression and prediction with the BGLR statistical package. Genetics 198, 483–495. doi: [10.1534/genetics.114.164442](https://doi.org/10.1534/genetics.114.164442)

Sandhu, K., Patil, S. S., Pumphrey, M., and Carter, A. (2021). Multitrait machineand deep-learning models for genomic selection usng spectral information in a
wheat breeding program. *Plant Genome* 14, e20119. doi: [10.1002/tpg2.20119](https://doi.org/10.1002/tpg2.20119)

Schulthess, A. W., Wang, Y., Miedaner, T., Wilde, P., Reif, J. C., and Zhao, Y. (2016). Multiple-trait- and selection indices-genomic predictions for grain yield and protein content in rye for feeding purposes. Theor. Appl. Genet. 129, 273–287. doi: [10.1007/s00122-015-2626-6](https://doi.org/10.1007/s00122-015-2626-6)

Searle, S. R. (1982). Matrix algebra useful for statistics. New York: John Wiley and Sons.

Smith, P. F., Ganesh, S., and Liu, P. (2013). A comparison of random forest regression and multiple linear regression for prediction in neuroscience. J. Neurosci. Methods 220, 85–91. doi: [10.1016/j.jneumeth.2013.08.024](https://doi.org/10.1016/j.jneumeth.2013.08.024)

Spindel, J., Begum, H., Akdemir, D., Virk, P., Collard, B., Redona, E., et al. (2015). Genomic selection and association mapping in rice (Oryza sativa): Effect of trait genetic architecture, training population composition, marker number and statistical model on accuracy of rice genomic selection in elite, tropical rice breeding lines. PLoS Genet[. 11, e1005350. doi: 10.1371/journal.pgen.](https://doi.org/10.1371/journal.pgen.1005350) 1005350

VanRaden, P. M. (2008). Efficient methods to compute genomic predictions. J. Dairy Sci. 91, 4414–4423. doi: [10.3168/jds.2007-0980](https://doi.org/10.3168/jds.2007-0980)

Ward, B. P., Brown-Guedira, G., Tyagi, P., Kolb, F. L., Van
Sanford, D. A., Sneller, C. H., et al. (2019). Multienvironment and
multitrait genomic selection models in unbalanced early-generation
wheat yield trials. Crop Sc 0189

Wu, P. Y., Tung, C. W., Lee, C. Y., and Liao, C. T. (2019). Genomic prediction of pumpkin hybrid performance. Plant Genome 12, 180082. doi: [10.3835/plantgenome2018.10.0082](https://doi.org/10.3835/plantgenome2018.10.0082)

Yao, J., Zhao, D., Chen, X., Zhang, Y., and Wang, J. (2018). Use of genomic selection and breeding simulation in cross prediction for improvement of yield and quality in wheat (Triticum aestivum L.). Crop J. 6, 353–365. doi: [10.1016/j.cj.2018.05.003](https://doi.org/10.1016/j.cj.2018.05.003)

Youens-Clark, K., Buckler, E., Casstevens, T., Chen, C., DeClerck, G., Derwen, P., et al. (2011). Gramene database in 2010: updates and extensions. Nucleic Acid Res. 39, D1085–D1094. doi: [10.1093/nar/gkq1148](https://doi.org/10.1093/nar/gkq1148)

Zhao, K., Tung, C. W., Eizenga, G. C., Wright, M. H., Ali, M. L., Price, A. H., et al. (2011). Genome-wide association mapping reveals a rich genetic architecture of complex traits in Oryza sativa. Nat. Commun. 2, 467. doi: [10.1038/ncomms1467](https://doi.org/10.1038/ncomms1467)