



Increased Predictive Accuracy of Multi-Environment Genomic Prediction Model for Yield and Related Traits in Spring Wheat (*Triticum aestivum* L.)

Vipin Tomar^{1,2,3*}, Daljit Singh^{4†}, Guriqbal Singh Dhillon⁵, Yong Suk Chung⁶, Jesse Poland⁴, Ravi Prakash Singh⁷, Arun Kumar Joshi^{1,3,7}, Yogesh Gautam¹, Budhi Sagar Tiwari² and Uttam Kumar^{1,3,7*}

OPEN ACCESS

Edited by:

Valentin Wimmer, KWS Saat, Germany

Reviewed by:

Pedro José Martínez-García, Spanish National Research Council, Spain Mian Abdur Rehman Arif, Nuclear Institute for Agriculture and Biology, Pakistan

*Correspondence:

Vipin Tomar viomics@gmail.com Uttam Kumar u.kumar@cgiar.org

[†]Present address:

Daljit Singh, The Climate Corporation, Bayer Crop Science, Creve Coeur, MO, United States

Specialty section:

This article was submitted to Plant Breeding, a section of the journal Frontiers in Plant Science

Received: 03 June 2021 Accepted: 03 September 2021 Published: 08 October 2021

Citation:

Tomar V, Singh D, Dhillon GS, Chung YS, Poland J, Singh RP, Joshi AK, Gautam Y, Tiwari BS and Kumar U (2021) Increased Predictive Accuracy of Multi-Environment Genomic Prediction Model for Yield and Related Traits in Spring Wheat (Triticum aestivum L.). Front. Plant Sci. 12:720123. doi: 10.3389/fpls.2021.720123 ¹ Borlaug Institute for South Asia, Ludhiana, India, ² Department of Biological Sciences and Biotechnology, Institute of Advanced Research, Gandhinagar, India, ³ International Maize and Wheat Improvement Center, New Delhi, India, ⁴ Department of Plant Pathology, Kansas State University, Manhattan, KS, United States, ⁶ Department of Biotechnology, Thapar Institute of Engineering & Technology, Patiala, India, ⁶ Department of Plant Resources and Environment, Jeju National University, Jeju-si, South Korea, ⁷ Global Wheat Program, International Maize and Wheat Improvement Center, Texcoco, Mexico

Genomic selection (GS) has the potential to improve the selection gain for complex traits in crop breeding programs from resource-poor countries. The GS model performance in multi-environment (ME) trials was assessed for 141 advanced breeding lines under four field environments via cross-predictions. We compared prediction accuracy (PA) of two GS models with or without accounting for the environmental variation on four quantitative traits of significant importance, i.e., grain yield (GRYLD), thousand-grain weight, days to heading, and days to maturity, under North and Central Indian conditions. For each trait, we generated PA using the following two different ME cross-validation (CV) schemes representing actual breeding scenarios: (1) predicting untested lines in tested environments through the ME model (ME_CV1) and (2) predicting tested lines in untested environments through the ME model (ME CV2). The ME predictions were compared with the baseline single-environment (SE) GS model (SE CV1) representing a breeding scenario, where relationships and interactions are not leveraged across environments. Our results suggested that the ME models provide a clear advantage over SE models in terms of robust trait predictions. Both ME models provided 2-3 times higher prediction accuracies for all four traits across the four tested environments, highlighting the importance of accounting environmental variance in GS models. While the improvement in PA from SE to ME models was significant, the CV1 and CV2 schemes did not show any clear differences within ME, indicating the ME model was able to predict the untested environments and lines equally well. Overall, our results provide an important insight into the impact of environmental variation on GS in smaller breeding programs where these programs can potentially increase the rate of genetic gain by leveraging the ME wheat breeding trials.

Keywords: single-environment, multi-environments, genotyping by sequencing, genomic selection (GS), genomics predictions, best linear unbiased predictions, wheat

1

INTRODUCTION

Wheat (*Triticum aestivum* L.) is an essential cereal to secure global food security (Curtis and Halford, 2014). Significant efforts are needed to accelerate high-yielding varieties to fulfill future global wheat demand by 2050 (Hellin et al., 2012). Hence, the enhancement of grain yield (GRYLD) is a foremost target for wheat breeders. GRYLD is a complex trait administered by many small-effect loci with significant loci × loci interactions (Arzani and Ashraf, 2017; Sehgal et al., 2017). Moreover, the GRYLD trait is associated with strong genotype × environment interaction, which makes its genetic enhancement a difficult work.

Genomic selection (GS) integrates genome-wide dense markers and, as presented by Meuwissen et al. (2001), is a different marker-assisted selection approach. GS proves to be a powerful tool to improve the selection accuracy and prediction for quantitative traits in crop breeding (Crossa et al., 2017). GS utilizes a large set of, usually unidentified markers, spread over the whole genome in the same way as every quantitative trait locus (QTL) is in linkage disequilibrium (LD). GS is particularly beneficial for traits that cannot be evaluated on a few plants and for traits that are hard to estimate. It is still a vital issue for plant breeders to upsurge the accuracy of genomic prediction for selecting the advanced breeding lines.

The GS has been widely used in wheat breeding to predict various traits, such as yield, disease resistance, grain weight, heading, iron and zinc contents, end-use quality, and physiological traits (Charmet et al., 2014; Velu et al., 2016; Hayes et al., 2017; Juliana et al., 2017a,b; Norman et al., 2017; Lozada et al., 2019; Tomar et al., 2021). As such, GS embraces the prospects for the genomic enhancement of qualitative and quantitative traits. Many available GS models have been tested on various breeding and trait scenarios. Earlier numerous comparative studies of the GS model predictions in wheat showed that Random Forest and Reproducing Kernel Hilbert Space models performed better for traits of interest. However, any single GS model could not outperform other models (Pérez-Rodríguez et al., 2012; Charmet et al., 2014). Earlier studies have stated that many interconnected factors impact the overall model performance (Jannink et al., 2010; Heslot et al., 2012), such as heritability, population structure, statistical models, i.e., parametric and nonparametric models, cross-validation (CV) approaches, the genetics of traits, training population size and composition, marker density, and LD among markers and QTLs (Jannink et al., 2010; Pérez-Rodríguez et al., 2012; Crossa et al., 2017; Norman et al., 2018; Lozada et al., 2019).

The GS delivers the promise to accelerate genetic gain by increasing precision in selecting and shortening the breeding cycles. However, GS is a relatively new and advanced method for smaller and low-resource South Asian wheat breeding programs. Previously, substantial progress has been made in testing and validating various models for GRYLD and related traits in wheat in South Asia, albeit on larger breeding populations (De los Campos et al., 2009; Crossa et al., 2010, 2011, 2016; Heffner et al., 2011; Burgueño et al., 2012; Pérez-Rodríguez et al., 2012; Rutkoski et al., 2015; Juliana et al., 2017a,b, 2019; González-Camacho et al., 2018). These studies have highlighted the impact of environment and genotype \times environment on the GS model performance. Therefore, to optimize the genetic gain from GS, the group of field-testing environments can be leveraged.

In this study, high-yielding, advanced wheat breeding lines from The International Maize and Wheat Improvement Center (CIMMYT) were evaluated for two consecutive wheat seasons (2017 and 2018) to adapt to the diverse environments of North and Central India. To evaluate the performance of multienvironment (ME) GS models on our specific set of selection environments, we tested different GS CV schemes mimicking the breeding schemes where untested lines and environmental performance are highly valuable to achieve the desired selection gains. This study is highly relevant particularly in the South Asian context where trial sizes are relatively small and broadly adapted wheat lines are sought after.

MATERIALS AND METHODS

Plant Material

A set of 141 South Asian spring wheat lines (*T. aestivum* L.) were selected from the International Yield Trial of CIMMYT International Nurseries (elite germplasm). These lines constitute a diverse association panel. The seeds of 141 genotypes were obtained from the Germplasm Resource Unit, CIMMYT (Mexico). Detailed information with a pedigree for each genotype is given in **Supplementary Table 1**.

Field Trials and Phenotypic Evaluation

The panel of selected lines was evaluated in field trials at the Borlaug Institute for South Asia (India) at Jabalpur (JBL) (23°14′00.6N and 80°04′40.7E) and Ludhiana (LDH) (30°59′28.74N and 75°44′10.87E), locations during the crop season for 2 years (2017 and 2018), genotypes were phenotyped and evaluated across all trials for four traits [days to maturity (DAYSMT), days to heading (DTHD), GRYLD, and thousandgrain weight (TGW)] (**Supplementary Table 2**). The experiment was conducted in two replications following the Randomized Block Design (RBD). The normal agronomic practice was followed for trial management. The row-to-row distance was maintained at 20 cm.

Genotyping-by-Sequencing and SNP Filtering

Genomic DNA was extracted from the fresh leaves of seedling wheat using the modified cetyltrimethylammonium bromide (CTAB) method (Dreisigacker et al., 2016). Genotyping-bysequencing (GBS) was performed in Illumina HiSeq 2500 using a protocol suggested by Poland et al. (2012). Single nucleotide polymorphism (SNP) calling was performed using TASSEL version 5.2.43 (Bradbury et al., 2007) using the TASSEL-GBSv2 pipeline. Using Beagle version 4.1, the missing data were imputed with default settings. After quality control (filter criteria: sample call rate > 0.8, Minor allele frequency (MAF) \geq 0.05, SNP call rate > 0.7), 14,563 polymorphic SNPs and 141 genotypes were selected for the follow-up analysis (**Supplementary Table 3**). Among polymorphic SNP markers, 40.66, 50.66, and 8.68% were from the A, B, and D genomes, respectively. With a genomic coverage of 13.9 GB and 14,563 markers across the genome, the average marker density was one marker per 0.95 Mb. The highest marker density with one marker per 0.54 Mb of chromosome 2B and the lowest marker density with one marker per 6.854 Mb at chromosome 4D were observed.

Statistical Analysis of Phenotypes

Each location-year combination is treated as a distinct environment for analysis purposes. Broad-sense heritability for each trait/environment combination was estimated at the plot level, and raw phenotypic values were adjusted to derive the best linear unbiased predictions (BLUPs) (**Supplementary Table 4**) for each trait at each environment using META-R (Alvarado et al., 2020) by using the following formula:

$$Y_{ik} = \mu + Rep_i + Gen_k + \epsilon_{ik} \text{(within environments)}$$

$$Y_{ijk} = \mu + Env_i + Rep_j (Env_i) + Gen_k + Env_i \times Gen_k$$

$$+ \epsilon_{ijk} \text{(across environments)}$$

where Y_{ik} is the trait of interest, μ is the mean effect, Rep_i is the effect of the *i*th replicate, Gen_k is the effect of the *k*th genotype, ϵ_{ik} is the error associated with the *i*th replication and the *k*th genotype, which is assumed to be normally and independently distributed, with mean 0 and homoscedastic variance. For across environments, Y_{ijk} is the trait response and the *i*th environment, $Rep_i(Env_i)$ is the effect of *j*th Rep in the *i*th environment, and $Env_i \times Gen_k$ is the environment \times genotype interaction. The resulting analysis produced the adjusted trait phenotypic values in the form of BLUPs within and across environments. The BLUPs model considers genotypes as random effects, minimizing the effect of screening time and other environmental effects.

In addition, the components of the phenotypic variance of a given trait at an individual environment and across environments were also extracted to calculate the broad-sense heritability using the formula as follows:

$$H^{2} = \frac{\sigma_{g}^{2}}{\sigma_{g}^{2} + \frac{\sigma_{e}^{2}}{nReps}}$$
(within environments)
$$H^{2} = \frac{\sigma_{g}^{2}}{\sigma_{g}^{2} + \frac{\sigma_{ge}^{2}}{nEnvs} + \frac{\sigma_{e}^{2}}{(nEnvs \times nReps)}}$$
(across environments)

where σ_g^2 and σ_e^2 are the genotype and error variance components, respectively, σ_{ge}^2 is genotype × environment interaction variance, nEnvs is the number of environments, and nReps is the number of replicates. All effects are considered random for calculating the BLUPs (**Supplementary Table 4**) and the broad-sense heritability. The BLUPs phenotypic distributions of GRYLD and other traits at each environment were plotted to check normality assumptions. Phenotypic and genetic correlations were calculated for each trait and environment combination in R software version 4.0.2. (R Core Team, 2019) using FactoMineR version 2.4 (Lê et al., 2008) and factoextra version 1.0.7 (Kassambara and Mundt, 2020).

Baseline Single-Environment (SE) Genomic BLUP Model (GBLUP), CV Schemes, and Predictive Ability

The baseline SE genomic prediction analysis was implemented in the BWGS program (Charmet et al., 2020). BWGS performs a GBLUP analysis using a marker-based relationship matrix. CV delivers an unbiased evaluation for the performance of a GS model; therefore, a 5-fold CV approach was implemented for reducing the unwanted bias (Kohavi, 1995), where the genotypes (for each trait separately) were randomly split into five equalsized folds. SE_CV1 model was fitted with missing phenotypic values for the tested individuals. Prediction accuracy (PA) was subsequently calculated as the correlation of predicted breeding values with the observed phenotypic values for the missing genotypes. This step was repeated for each environment and fold separately. The genomic PA was then calculated by iteratively assigning 1-fold as the validation set and the remaining folds as the training set. This five-fold validation process was repeated 50 times to randomly shuffle the lines in each fold. The accuracy of the genomic predictions was measured as the Pearson's correlation between the predicted and actual trait BLUPs.

A mixed model of the simplified form was fitted for genomic predictions as follows:

$$y = Xb + Zg + e$$

where *y* is a vector of adjusted phenotypes, X is a design matrix relating the fixed effects to each genotype, *b* is a vector of fixed effects, Z is a design matrix connecting records to genetic values, *g* is a vector of additive genetic effects for a genotype, and *e* is a vector of random normal deviates with variance δ_e^2 .

Advanced ME GBLUP Model, CV Schemes, and Predictive Ability

The advanced ME genomic prediction analysis was implemented in Solving Mixed Model Equations in the R (sommer) package (Covarrubias-Pazaran, 2016). Two types of ME_CV schemes representing actual breeding scenarios were implemented. The first scenario represents a use case where some genotypes are missing across all environments (ME_CV1). ME_CV1 was fitted by masking the phenotypic values of genotypes belonging to the test set across all environments. PA was calculated as the correlation of predicted and observed phenotypic values for the missing genotypes at each environment separately. In the second scenario, the entire trial or all genotypes are missing at one of the environments (ME_CV2). ME_CV2 was fitted by masking the phenotypic values of all lines in an SE. The trained model was then used to predict the breeding values of lines in the missing environment. PA was calculated as the correlation of predicted and observed phenotypic values of the tested lines. The CV schemes are illustrated in Figure 1.

In ME genomic predictions, the SE model was rewritten and implemented as follows:

$$y_{ij} = g_j + E_i + gE_{ij} + e_{ij}$$

where y_{ij} represents response of *j*th line in the *i*th environment ($i = 1, 2, \dots, i, j = 1, 2, \dots, j; g_j$ is the effect of *j*th line with

				Training	Validation	
SE_CV1	k1	k2	k3	k4	k5	
Env						
ME_CV1	k1	k2	k3	k4	k5	
JBL17						
JBL18						
LDH17						
LDH18						
			1.0			
ME_CV2	k1	k2	k3	k4	k5	
JBL17						
JBL18						

LDH18			
LDH17			
JBL18			

FIGURE 1 | Prediction scheme for the single-environment (SE) and multi-environment (ME) genomic prediction models with two cross-validation schemes (CV1 and CV2) used in this study. SE_CV1 model: the SE prediction model with CV scheme 1 where a trait [e.g., grain yield (GRYLD)] is predicted at a time; we used 80% of individuals as the training set (phenotyped and genotyped, light green) and 20% of the individuals as the testing set (genotyped only, light gray with validation code for the trait to be predicted, yield as an example here). ME_CV1 model: the ME prediction model with CV scheme 1 for new un-phenotyped individuals; we used 80% of individuals as the training set (phenotyped for all traits and genotyped; light green) and 20% of the individuals as the validation set (genotyped but not phenotyped for any trait; light gray with validation code for the trait to be predicted, GRYLD as an example here). ME_CV2 model: the ME prediction model with CV scheme 2 where 100% of the information from other traits are available for the individuals to be predicted; we used 80% of individuals as the training set (phenotyped for all traits and genotyped; light green) and 20% of individuals as the training set (phenotyped for all traits and genotyped; light green) and 20% of the individuals to be predicted; we used 80% of individuals as the training set (phenotyped for all traits and genotyped; light green) and 20% of individuals as the validation set (phenotyped for associated traits but not for the targeted traits, and genotyped; light gray with predication code for the trait to be predicted, yield as an example here). Rectangles represent genotypes, and colors represent whether the phenotypic information was used (light green) or not (light gray with validation code for the trait to be predicted, GRYLD as an example) for the population. A similar scheme was applied for predicting days to heading (DTHD), days to maturity (DAYSMT), and thousand-grain weight (TGW).

 $g = (g_{1,...,g_j})T \sim N(0, \delta_1^2 Gg), \delta_1^2$ is the genomic variance, Gg is the genomic relationship matrix. E_i represents the effect of the *i*th environment. gE_{ij} is the random term that takes into account the interaction between the genomic effect of *j*th line and the *i*th environment with $gE = (g_1 \ldots \ldots g_j)T \sim N (0, \delta_2^2 I_I \otimes G)$, where δ_2^2 is the interaction variance. E_{ij} is a random residual effect of the *j*th line in the *i*th environment $[N (0, \delta_2^2)]$, where δ_2^2 is the residual variance.

RESULTS

Heritability, Correlations, and Trait Characterization

A range of variation was detected for GRYLD and other related traits across environments/years (LDH17 and LDH18 and JBL17 and JBL18). The highest averaged GRYLD over environments/years was observed at JBL18 (9.4 ton/ha),

Loc [#]	Env	Trait ^{##}	H ²	G Var	R Var	G Mean	LSD	с٧	G Sig
JBL	JBL17	DTHD	0.84	10.98	4.04	81.96	3.65	2.45	0
		DAYSMT	0.86	5.86	1.89	124.82	2.52	1.10	0
		GRYLD	0.48	0.29	0.63	7.87	1.08	10.09	0.000151
		TGW	0.86	26.59	8.92	54.66	5.47	5.47	0
	JBL18	DTHD	0.78	12.79	7.30	79.26	4.71	3.41	0
	DAYSMT	0.71	4.89	3.96	124.67	3.32	1.60	9.08E-13	
	GRYLD	0.47	0.15	0.34	8.76	0.79	6.67	0.000172	
		TGW	0.80	12.53	6.34	46.22	4.45	5.45	0
LDH	LDH17	DTHD	0.96	12.61	1.19	94.85	2.11	1.15	0
	DAYSMT	0.74	4.79	3.29	148.73	3.09	1.22	6.88E-15	
	GRYLD	0.74	0.21	0.15	7.06	0.66	5.55	2.73E-14	
		TGW	0.81	15.42	7.03	45.48	4.73	5.83	0
	LDH18	DTHD	0.88	8.58	2.44	103.71	2.89	1.51	0
	DAYSMT	0.88	8.18	2.25	144.52	2.80	1.04	0	
		GRYLD	0.62	0.16	0.20	7.26	0.69	6.11	1.92E-08
		TGW	0.83	14.66	6.13	44.30	4.47	5.59	0

TABLE 1 Variability analysis of various yield-related agronomic traits for four environments at two locations.

[#]Loc, location; Env, Environment; H², heritability; G Var, genotypic variance; R Var, residual variance; LSD, least significant difference; CV, critical variance; G Sig, genotypic significance; LDH, Ludhiana; JBL, Jabalpur.

##DTHD, days to heading; DAYSMT, days to maturity; GRYLD, grain yield; TGW, thousand-grain weight.

TABLE 2 Variability analysis of various yield-related agronomic traits for four environments at two locations.											
Traits	H ²	G Var	$\mathbf{G} \times \mathbf{E} \mathbf{Var}$	R Var	G Mean	LSD	CV	n Rep	n Env	G Sig	G × E Sig
DTHD	0.90	8.94	2.29	3.74	89.94	2.69	2.15	2	4	8.93E-73	1.16E-18
DAYSMT	0.83	4.00	1.94	2.83	135.68	2.32	1.24	2	4	4.34E-44	2.01E-21
GRYLD	0.38	0.05	0.15	0.33	7.74	0.49	7.43	2	4	0.0003	3.69E-13
TGW	0.78	9.90	7.41	7.10	47.67	4.07	5.59	2	4	1.13E-33	4.23E-35

H², heritability; G Var, genotypic variance; R Var, residual variance; LSD, least significant difference; CV, critical variance; G Sig, genotypic significance; DTHD, days to heading; DAYSMT, days to maturity; GRYLD, grain yield; TGW, thousand-grain weight.

followed by JBL17 (8.7 ton/ha), LDH17 (8.2 ton/ha), and LDH18 (7.9 ton/ha). Similarly, the TGW trait also showed variation across environments. The highest averaged TGW over environments/years was observed at JBL17 (69g), followed by JBL18 (59.5g), LDH17 (58.4g), and LDH18 (53.5g). We observed significant G \times E interaction for the GRYLD and DAYSMT in JBL18 and LDH17 (**Tables 1**, **2**). For all traits, the broad-sense heritability ranged from 0.47 to 0.96. The broad-sense heritability of DTHD was the highest (0.96) in LDH17, while GRYLD, the lowest (0.47) was in JBL18, and the highest (0.74) was in LDH17. TGW had the highest stability and relatively high heritability (0.80–0.86) across environments.

The phenological traits DTHD and DAYSMT displayed the strongest positive correlation (0.88), followed by a weak positive correlation TGW-GRYLD (0.15), while GRYLD and DTHD (-0.73) demonstrated negative correlations. The lowest correlation was observed between GRYLD and DAYSMT (-0.76) traits. The principal component analysis (PCA) of multivariate analysis enables the easier understanding of effects and networks among different traits and elucidates genotypic difference among a set of given traits, i.e., the first two PCs explained 92% of the total variation. The PC1 explained 70.3% of the total variance and PC2 explained 21.7% of the total variance (**Figure 2**).

Baseline SE Model: Performance of Untested Lines in the Same Environment

A GS scenario representing SE breeding programs was tested. In this model, the PAs of the GS models for each of the four traits were separately generated for all four tested environments. In other words, the environments were treated as independent. Overall, the PA of the SE model was significantly lower among the three tested GS scenarios (**Table 4**; **Figure 3**). PA was the highest for TGW (0.34) and the lowest for GRYLD (0.18) traits. A relatively low moderate PA ranging between 0.24 and 0.25 was observed for DAYSMT and DTHD traits. Among the tested environments, JBL18 had the lowest overall PA (0.01–0.02) compared to the rest of the three environments for DTHD and

Frontiers in Plant Science | www.frontiersin.org



Advanced ME Model: Performance of Tested Lines in Untested Environments and Untested Lines in Tested Environments

The inclusion of environmental information in ME models significantly improved the PA over SE models across all traits and environments (**Figure 3**). A very high and consistent PA ranging from 0.69 to 0.85 was observed for all traits and environments for both ME models (ME_CV1 and ME_CV2). The most considerable improvement in PA due to ME was observed for the GRYLD trait, where PA increased from 0.18 to 0.73 for SE and ME models (**Table 4**). Interestingly, identical trait rankings were also observed for two ME models, where the DTHD ranked the highest (0.85) and GRYLD ranked the lowest (0.69–0.73) among all four traits. While the ME models

had identical trait rankings, the environments ranked slightly differently for the two models for all traits. For instance, both years (2017 and 2018) at the LDH location had higher overall PA compared to JBL for all traits.

DISCUSSION

Crop breeders regularly evaluate the performance of genotypes and collect multiple traits data in various environments. The genotype-based selection on phenotypic and GBS marker information using genomic prediction models is gradually acquiring acceptance in breeding with the initiation of economical next-generation sequencing (NGS) technologies (Poland and Rife, 2012). Limited study has been conducted using the multi-environment genomic prediction (ME-GP) methods due to the complexity and higher computing requirements (Oakey et al., 2016; Rincent et al., 2017; Montesinos-López et al., 2018; Roorkiwal et al., 2018; Bhandari et al., 2019; Tolhurst et al., 2019; Pandey et al., 2020).





TABLE 3 Genetic and phenotypic correlations in agronomic important tra
--

		Genetic correlations				ns	
Traits	DTHD	DAYSMT	GRYLD	Traits	DTHD	DAYSMT	GRYLD
DAYSMT	0.94			DAYSMT	0.83		
GRYLD	-0.30	-0.29		GRYLD	-0.22	-0.08	
TGW	-0.33	-0.26	0.22	TGW	-0.29	-0.24	0.07

DTHD, days to heading; DAYSMT, days to maturity; GRYLD, grain yield; TGW, thousand-grain weight.

Trait Correlation and Characterization: A Vital Factor for Improving Accuracy in ME-GP

In this study, advanced breeding lines as part of the bread wheat program of CIMMYT were evaluated under irrigated conditions at two locations (JBL and LDH) for 2 years (2017 and 2018) (i.e., four environments). This study evaluated four traits (i.e., DTHD, DAYSMT, GRYLD, and TGW) for use in an ME trait GP model. GRYLD and related traits were positively correlated to each other in two sets (i.e., 1: DAYSMT and DTHD; and 2: GRYLD and TGW) (**Figure 4**). This positive correlation of GRYLD with TGW in this study points out that the GRYLD was mainly distinct by

TABLE 4 | Genomic prediction accuracies averaged across four environments for four traits and three modeling scenarios (a) single-environment CV1 (SE_CV1), (b) multi-environment CV1 (ME_CV1), and (c) multi-environment CV2 (ME_CV2).

Traits	A	verage prediction acc	uracy
	SE_CV1	ME_CV1	ME_CV2
DAYSMT	0.24	0.78	0.78
DTHD	0.25	0.85	0.85
GRYLD	0.18	0.69	0.73
TGW	0.34	0.82	0.83

DTHD, days to heading; DAYSMT, days to maturity; GRYLD, grain yield; TGW, thousandgrain weight.

the TGW factor. The negative relationship between GRYLD and DTHD indicates that the early-headed genotypes play a vital role in the stability of advanced breeding line yield during grain filling and finally affecting the yield component (Sharma and Smith, 1986).

Yield and Related Trait Heritability Difference Among Environments

Our results showed that the heritability of the traits ranged from moderate (i.e., GRYLD) to high (i.e., DAYSMT, DTHD, and TGW). Among the four traits, the phenological traits (i.e., DTHD and DAYSMT) and TGW particularly showed high stable broad-sense heritability ranging from 0.71 to 0.96. It suggests the high quality of the phenotypic measurements and significant predictive potential of the traits. GRYLD, a highly quantitative and environmentally sensitive trait (Maphosa et al., 2014; Würschum et al., 2018), showed considerable fluctuation across environments with JBL environment having relatively lower heritability (0.47-0.48) compared to LDH (0.62-0.74). The variance explained by agronomic traits was significant (Table 1) and indicating a large $G \times E$ impact on GRYLD resulted in a lower heritability compared to other traits. Hence, lower heritability estimates for GRYLD were expected as numerous genes govern it. The low heritability and yield variances also could be the possible effect of the smaller plot size and lower sowing density (Rode et al., 2011; Sallam et al., 2015; Thorwarth et al., 2017; Bhatta et al., 2018) (Tables 1, 2). The climate in these two environments is considerably different. While the growing season length is relatively shorter in JBL due to the high overall temperature, the LDH location has a moderately colder climate and longer growing season (Mondal et al., 2016). On the one hand, these highly variable environments do underscore a highly challenging phenotypic landscape; it also presents a significant opportunity to leverage the ME trial framework for trait improvement (Lillemo et al., 2005; Braun et al., 2010). The presence of significant genetic and environmental correlations (i.e., positive correlation in TGW and GRYLD, and DAYSMT and DTHD) in our experiments led us to hypothesize that the correlated traits and environmental relationships can be leveraged to improve the selection accuracy through markerbased ME-GS models (Figure 4). Therefore, we proceeded with applying the ME model to test this hypothesis on our selected set of lines (**Table 3**).

SE and ME Genomic Prediction Across Years and Sites and ME Model Utilities in Crop Breeding

While weak predictive capability continues to be a major issue in successfully applying GS (Crossa et al., 2013), numerous studies have demonstrated that GS could be beneficial for quantitative traits such as GRYLD with low heritability and also on how GS can be utilized in a breeding program by using even low to moderate GP in early generation selection (Belamkar et al., 2018; Lado et al., 2018; Michel et al., 2018). There are several aspects influencing the PA of GP models. Some of the crucial aspects associated with this study of ME were the genetic relationship between the testing and training sets, the size of the training set, heritability and trait architecture, and correlations among traits and environments (Asoro et al., 2011; Crossa et al., 2013; Heslot et al., 2013; Sallam et al., 2015; Zhang et al., 2015; Duangjit et al., 2016; Lado et al., 2016; Wang et al., 2016; Thorwarth et al., 2017; Akdemir and Isidro-Sánchez, 2019; Olatoye et al., 2020). Even though the size of the population was small in our study, the GP using correlated traits in the ME_CV1 and ME_CV2 schemes had higher PA, indicating that correlated traits up to some extent could balance the impact on the sizes of small population.

Models that leverage E and G \times E components have been shown to improve the genomic prediction accuracies for highly quantitative traits such as phenology and GRYLD (Burgueño et al., 2012; Dias et al., 2018). To evaluate the potential of genomic predictions in highly productive but variable environments of JBL and LDH, we simulated three different genomic prediction scenarios representing actual breeding programs. A comparison of single and ME models showed a 2- to 3-fold improvement in model performance for all traits (Table 4; Figure 3). Among the four traits, GRYLD showed the highest (3.8X) absolute increase in PA from SE to ME models, highlighting the significance of ME modeling in GRYLD predictions. For the SE model, TGW had the most consistent PA across four environments (0.32-0.34), which was in agreement with the highly stable heritability and a lower fraction of $G \times E$ observed for this trait (**Table 2**; **Figure 3**). Interestingly, the PA of the two ME models (CV1 and CV2) showed no significant change, suggesting that the ME model was able to predict well the untested environments and lines equally. A model can be highly predictive of untested environments in scenarios where environments are highly correlated (Malosetti et al., 2016; Jarquín et al., 2017), which seems to be the case for our environments as reflected by the low $G \times E$ and high heritability (Table 1; Figure 3). Similarly, a remarkable improvement in the predictive performance of ME_CV1 can be partially attributed to the fact that our sampled set of lines came from the same breeding program and the sample size of 141 lines was relatively moderate. From the perspective of a breeding program, the strong performance of the two ME models suggests that our breeding program can increase the overall population size without losing any significant predictive power through sparse testing at these two environments (Cullis et al.,



FIGURE 4 | Distributions, scatter plots, and correlations between agronomic traits using best linear unbiased predictions from combining and four experiments [Ludhiana (LDH)17, LDH18, Jabalpur (JBL)17, and JBL18]. The distribution of DTHD, DAYSMT, GRYLD, and TGW values is displayed on the diagonal with environments indicated by colors. The top row represents the distribution of traits as boxplots. The upper right triangle shows pairwise correlation values as overall correlation in black color while other colors are represented individually as explained earlier. The correlations among environments are displayed as scatter plots in the lower triangular area and as the Pearson's correlation coefficients in the upper triangular area. Numbers indicate a correlation that is significantly different from 0 at an alpha level of 0.05. DTHD, DAYSM, GRYLD, and TGW. ## level of significance; ***p < 0.001, **p < 0.01, *p < 0.1, and p < 0.15.

2020; Jarquin et al., 2020). A high population size from the sparse testing framework here can deliver a high selection gain through increased selection intensity.

CONCLUSION

Breeding for quantitative traits is challenging due to the complex genetic architecture of traits that are highly affected by the complex $G \times E$ interactions in field trials. A suitable genomic prediction modeling strategy can potentially address

this challenge through ME genomic prediction models. In this study, we evaluated genomic prediction accuracies of advanced spring wheat lines under four diverse environments in two wheat-growing regions in India. The ME-GS models showed significant improvement over SE models in terms of prediction accuracies. Our results suggest that ME can be leveraged to improve the breeding selection efficiency for major agronomic and phonological traits. Over the years, CIMMYT has established an extensive network of field-testing sites in South Asian countries including India, Pakistan, Bangladesh, and Nepal. Our results suggest that the wheat breeding programs in these countries can greatly benefit from GS through better modeling of environmental variance and sparse testing of a larger cohort of breeding lines. Future research efforts will be directed toward including high-throughput phenotyping traits such as plant height, Normalized Difference Vegetation Index (NDVI), and senescence into the genomic prediction framework to improve the selection efficiency of spring wheat in the South Asian breeding programs.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

AUTHOR CONTRIBUTIONS

VT and DS drafted the manuscript. VT, DS, GD, and YC analyzed the data. UK, JP, and RS designed the field trials, conducted genotyping, and provided breeding lines. VT and YG collected field data. UK, BT, JP, RS, and AJ supervised the overall study. All authors contributed to the article and approved the submitted version.

REFERENCES

- Akdemir, D., and Isidro-Sánchez, J. (2019). Design of training populations for selective phenotyping in genomic prediction. *Sci. Rep.* 9, 1–15. doi: 10.1038/s41598-018-38081-6
- Alvarado, G., Rodríguez, F. M., Pacheco, A., Burgueño, J., Crossa, J., Vargas, M., et al. (2020). META-R: a software to analyze data from multienvironment plant breeding trials. *Crop J.* 8, 745–756. doi: 10.1016/j.cj.2020. 03.010
- Arzani, A., and Ashraf, M. (2017). Cultivated ancient wheats (*triticum spp.*): a potential source of health-beneficial food products. *Compr. Rev. Food Sci. Food Saf.* 16, 477–488. doi: 10.1111/1541-4337.12262
- Asoro, F. G., Newell, M. A., Beavis, W. D., Scott, M. P., and Jannink, J. (2011). Accuracy and training population design for genomic selection on quantitative traits in elite north american oats. *Plant Genom.* 4:007. doi: 10.3835/plantgenome2011.02.0007
- Belamkar, V., Guttieri, M. J., Hussain, W., Jarquín, D., El-basyoni, I., Poland, J., et al. (2018). Genomic selection in preliminary yield trials in a winter wheat breeding program. G3 Genes, Genomes, Genet. 8, 2735–2747. doi: 10.1534/g3.118.200415
- Bhandari, A., Bartholom, J., Cao-Hamadoun, T.-V., Kumari, N., Frouin, J., Kumar, A., et al. (2019). Selection of trait-specific markers and multi-environment models improve genomic predictive ability in rice. *PLoS ONE* 14:e0208871. doi: 10.1371/journal.pone.0208 871
- Bhatta, M., Morgounov, A., Belamkar, V., and Baenziger, P. S. (2018). Genome-Wide association study reveals novel genomic regions for grain yield and yield-related traits in drought-stressed synthetic hexaploid wheat. *Int. J. Mol. Sci.* 19:3011. doi: 10.3390/ijms191 03011

FUNDING

This study was supported by the United States Agency for International Development (USAID), Feed the Future Innovation Lab for Applied Wheat Genomics (Cooperative Agreement No. AID-OAA-A-13-00051), and CGIAR Research Program on Wheat (CRP) Partner Grant to BISA (Grant Code: A5017.09.64).

ACKNOWLEDGMENTS

We would like to thank the field staff at the field sites of the Borlaug Institute of South Asia at Jabalpur and Ludhiana for their assistance with the data collection.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2021. 720123/full#supplementary-material

Supplementary Figure 1 | Weather information of LDH17 and LDH18.

Supplementary Figure 2 | Weather information of JBL17 and JBL18.

Supplementary Table 1 | List of 141 genotypes with pedigree information used in this study.

Supplementary Table 2 | List of traits that were evaluated during this study in the field trials.

Supplementary Table 3 | GBS HapMap data used in this study.

Supplementary Table 4 | Best linear unbiased predictions (BLUPs) data used in this study.

- Bradbury, P. J., Zhang, Z., Kroon, D. E., Casstevens, T. M., Ramdoss, Y., and Buckler, E. S. (2007). TASSEL: software for association mapping of complex traits in diverse samples. *Bioinformatics* 23, 2633–2635. doi:10.1093/bioinformatics/btm308
- Braun, H. J., Atlin, G., and Payne, T. (2010). "Multi-location testing as a tool to identify plant response to global climate change," in *Climate Change and Crop Production*, ed M. P. Reynolds (CABI International), 115–138.
- Burgueño, J., Campos, G., de los, Weigel, K., and Crossa, J. (2012). Genomic prediction of breeding values when modeling genotype × environment interaction using pedigree and dense molecular markers. *Crop Sci.* 52, 707–719. doi: 10.2135/cropsci2011.06.0299
- Charmet, G., Storlie, E., Oury, F. X., Laurent, V., Beghin, D., Chevarin, L., et al. (2014). Genome-wide prediction of three important traits in bread wheat. *Mol. Breed.* 34, 1843–1852. doi: 10.1007/s11032-014-0143-y
- Charmet, G., Tran, L.-G., Auzanneau, J., Rincent, R., and Bouchet, S. (2020). BWGS: A R package for genomic selection and its application to a wheat breeding programme. *PLoS ONE* 15:e0222733. doi:10.1371/journal.pone.0222733
- Covarrubias-Pazaran, G. (2016). Genome-Assisted prediction of quantitative traits using the R package sommer. *PLoS ONE* 11:e0156744. doi: 10.1371/journal.pone.0156744
- Crossa, J., Campos, G., de los, Pérez, P., Gianola, D., Burgueño, J., Araus, J. L., et al. (2010). Prediction of genetic values of quantitative traits in plant breeding using pedigree and molecular markers. *Genetics* 186, 713–724. doi: 10.1534/genetics.110.118521
- Crossa, J., Jarquín, D., Franco, J., Pérez-Rodríguez, P., Burgueño, J., Saint-Pierre, C., et al. (2016). Genomic prediction of gene bank wheat landraces. G3 Genes|Genomes|Genetics 6:1819. doi: 10.1534/g3.116.029637
- Crossa, J., Pérez, P., Campos, G., de los, Mahuku, G., Dreisigacker, S., and Magorokosho, C. (2011). Genomic selection and prediction in plant

breeding. J. Crop Improv. 25, 239-261. doi: 10.1080/15427528.2011. 558767

- Crossa, J., Pérez, P., Hickey, J., Burgueño, J., Ornella, L., Cerón-Rojas, J., et al. (2013). Genomic prediction in CIMMYT maize and wheat breeding programs. *Hered* 112, 48–60. doi: 10.1038/hdy.2013.16
- Crossa, J., Pérez-Rodríguez, P., Cuevas, J., Montesinos-López, O., Jarquín, D., de los Campos, G., et al. (2017). Genomic selection in plant breeding: methods, models, and perspectives. *Trends Plant Sci.* 22, 961–975. doi: 10.1016/j.tplants.2017.08.011
- Cullis, B. R., Smith, A. B., Cocks, N. A., and Butler, D. G. (2020). The design of early-stage plant breeding trials using genetic relatedness. J. Agric. Biol. Environ. Stat. 25, 553–578. doi: 10.1007/s13253-020-00403-5
- Curtis, T., and Halford, N. G. (2014). Food security: the challenge of increasing wheat yield and the importance of not compromising food safety. *Ann. Appl. Biol.* 164, 354–372. doi: 10.1111/aab.12108
- De los Campos, G., Naya, H., Gianola, D., Crossa, J., Legarra, A., Manfredi, E., et al. (2009). Predicting quantitative traits with regression models for dense molecular markers and pedigree. *Genetics* 182, 375–385. doi: 10.1534/genetics.109.101501
- Dias, K. O. D. G., Gezan, S. A., Guimarães, C. T., Nazarian, A., da Costa e Silva, L., Parentoni, S. N., et al. (2018). Improving accuracies of genomic predictions for drought tolerance in maize by joint modeling of additive and dominance effects in multi-environment trials. *Hered* 121, 24–37. doi: 10.1038/s41437-018-0053-6
- Dreisigacker, S., Deepmala, S., Jaimez, R. A., Luna-Garrid, B., Muñoz-Zavala, S., Núñez-Ríos, C., et al. (2016). *CIMMYT Wheat Molecular Genetics: Laboratory Protocols and Applications to Wheat Breeding.* Mexico, DF: CIMMYT.
- Duangjit, J., Causse, M., and Sauvage, C. (2016). Efficiency of genomic selection for tomato fruit quality. *Mol. Breed.* 36, 1–16. doi: 10.1007/s11032-016-0453-3
- González-Camacho, J. M., Ornella, L., Pérez-Rodríguez, P., Gianola, D., Dreisigacker, S., and Crossa, J. (2018). Applications of machine learning methods to genomic selection in breeding wheat for rust resistance. *Plant Genome* 11:170104. doi: 10.3835/plantgenome2017.11.0104
- Hayes, B. J., Panozzo, J., Walker, C. K., Choy, A. L., Kant, S., Wong, D., et al. (2017). Accelerating wheat breeding for end-use quality with multi-trait genomic predictions incorporating near infrared and nuclear magnetic resonance-derived phenotypes. *Theor. Appl. Genet.* 130, 2505–2519. doi: 10.1007/s00122-017-2972-7
- Heffner, E. L., Jannink, J.-L., Iwata, H., Souza, E., and Sorrells, M. E. (2011). Genomic selection accuracy for grain quality traits in biparental wheat populations. *Crop Sci.* 51, 2597–2606. doi: 10.2135/cropsci2011.05.0253
- Hellin, J., Shiferaw, B., Cairns, J. E., Reynolds, M., Ortiz-Monasterio, I., Banziger, M., et al. (2012). Climate change and food security in the developing world: Potential of maize and wheat research to expand options for adaptation and mitigation. J. Dev. Agric. Econ. 4, 311–321. doi: 10.5897/JDAE11.112
- Heslot, N., Akdemir, D., Sorrells, M. E., and Jannink, J. L. (2013). Integrating environmental covariates and crop modeling into the genomic selection framework to predict genotype by environment interactions. *Theor. Appl. Genet.* 127, 463–480. doi: 10.1007/s00122-013-2231-5
- Heslot, N., Yang, H.-P., Sorrells, M. E., and Jannink, J.-L. (2012). Genomic selection in plant breeding: a comparison of models. *Crop Sci.* 52, 146–160. doi: 10.2135/cropsci2011.06.0297
- Jannink, J.-L., Lorenz, A. J., and Iwata, H. (2010). Genomic selection in plant breeding: from theory to practice. *Brief. Funct. Genomics.* 9, 166–177. doi: 10.1093/bfgp/elq001
- Jarquin, D., Howard, R., Crossa, J., Beyene, Y., Gowda, M., Martini, J. W. R., et al. (2020). Genomic prediction enhanced sparse testing for multi-environment trials. G3 Genes Genomes Genet. 10, 2725–2739. doi: 10.1534/g3.120. 401349
- Jarquín, D., Silva, C. L., da, Gaynor, R. C., Poland, J., Fritz, A., Howard, R., et al. (2017). Increasing genomic-enabled prediction accuracy by modeling genotype × environment interactions in kansas wheat. *Plant Genome* 10:0130. doi: 10.3835/plantgenome2016.12.0130
- Juliana, P., Montesinos-López, O. A., Crossa, J., Mondal, S., González Pérez, L., Poland, J., et al. (2019). Integrating genomic-enabled prediction and highthroughput phenotyping in breeding for climate-resilient bread wheat. *Theor. Appl. Genet.* 132, 177–194. doi: 10.1007/s00122-018-3206-3
- Juliana, P., Singh, R. P., Singh, P. K., Crossa, J., Huerta-Espino, J., Lan, C., et al. (2017a). Genomic and pedigree-based prediction for leaf, stem,

and stripe rust resistance in wheat. Theor. Appl. Genet. 130, 1415–1430. doi: 10.1007/s00122-017-2897-1

- Juliana, P., Singh, R. P., Singh, P. K., Crossa, J., Rutkoski, J. E., Poland, J. A., et al. (2017b). Comparison of models and whole-genome profiling approaches for genomic-enabled prediction of *Septoria Tritici Blotch, Stagonospora Nodorum Blotch*, and Tan Spot resistance in wheat. *Plant Genome* 10:0082. doi: 10.3835/plantgenome2016.08.0082
- Kassambara, A., and Mundt, F. (2020). Factoextra: Extract and Visualize the Results of Multivariate Data Analyses. Available online at: https://cran.r-project.org/ packagefactoextra (accessed May 05, 2020).
- Kohavi, R. (1995). A study of cross-validation and bootstrap for accuracy estimation and model selection. *Proceedings of IJCAI'95* 2, 1137–1143.
- Lado, B., Barrios, P. G., Quincke, M., Silva, P., and Gutiérrez, L. (2016). Modeling genotype × environment interaction for genomic selection with unbalanced data from a wheat breeding program. *Crop Sci.* 56, 2165–2179. doi: 10.2135/cropsci2015.04.0207
- Lado, B., Vázquez, D., Quincke, M., Silva, P., Aguilar, I., and Gutiérrez, L. (2018). Resource allocation optimization with multi-trait genomic prediction for bread wheat (*Triticum aestivum* L.) baking quality. *Theor. Appl. Genet.* 131, 2719–2731. doi: 10.1007/s,00122-018-3186-3
- Lê, S., Josse, J., and Husson, F. (2008). FactoMineR: an R package for multivariate analysis. J. Stat. Softw. 25, 1–18. doi: 10.18637/jss.v025.i01
- Lillemo, M., Ginkel, M., van, Trethowan, R. M., Hernandez, E., and Crossa, J. (2005). Differential adaptation of CIMMYT bread wheat to global high temperature environments. *Crop Sci.* 45, 2443–2453. doi: 10.2135/cropsci2004.0663
- Lozada, D. N., Mason, R. E., Sarinelli, J. M., and Brown-Guedira, G. (2019). Accuracy of genomic selection for grain yield and agronomic traits in soft red winter wheat. *BMC Genet.* 20, 1–12. doi: 10.1186/s12863-019-0785-1
- Malosetti, M., Bustos-Korts, D., Boer, M. P., and Eeuwijk, F. A., van (2016). Predicting responses in multiple environments: issues in relation to genotype × environment interactions. *Crop Sci.* 56, 2210–2222. doi: 10.2135/cropsci2015.05.0311
- Maphosa, L., Langridge, P., Taylor, H., Parent, B., Emebiri, L. C., Kuchel, H., et al. (2014). Genetic control of grain yield and grain physical characteristics in a bread wheat population grown under a range of environmental conditions. *Theor. Appl. Genet.* 7 127, 1607–1624. doi: 10.1007/s00122-014-2322-y
- Meuwissen, T. H. E., Hayes, B. J., and Goddard, M. E. (2001). Prediction of total genetic value using genome-wide dense marker maps. *Genetics*. 157. 4 1819–1829. doi: 10.1093/genetics/157.4.1819
- Michel, S., Kummer, C., Gallee, M., Hellinger, J., Ametz, C., Akgöl, B., et al. (2018). Improving the baking quality of bread wheat by genomic selection in early generations. *Theor. Appl. Genet.* 131, 477–493. doi: 10.1007/s00122-017-2998-x
- Mondal, S., Singh, R. P., Mason, E. R., Huerta-Espino, J., Autrique, E., and Joshi, A. K. (2016). Grain yield, adaptation and progress in breeding for earlymaturing and heat-tolerant wheat lines in South Asia. F. Crop. Res. 192, 78–85. doi: 10.1016/j.fcr.2016.04.017
- Montesinos-López, A., Montesinos-López, O. A., Gianola, D., Crossa, J., and Hernández-Suárez, C. M. (2018). Multi-environment genomic prediction of plant traits using deep learners with dense architecture. G3 Genes|Genomes|Genet. 8, 3813–3828. doi: 10.1534/g3.118.200740
- Norman, A., Taylor, J., Edwards, J., and Kuchel, H. (2018). Optimising genomic selection in wheat: effect of marker density, population size and population structure on prediction accuracy. G3 Genes|Genomes|Genetics 8, 2889–2899. doi: 10.1534/g3.118.200311
- Norman, A., Taylor, J., Tanaka, E., Telfer, P., Edwards, J., Martinant, J.-P., et al. (2017). Increased genomic prediction accuracy in wheat breeding using a large Australian panel. *Theor. Appl. Genet.* 130, 2543–2555. doi: 10.1007/s00122-017-2975-4
- Oakey, H., Cullis, B., Thompson, R., Comadran, J., Halpin, C., and Waugh, R. (2016). Genomic selection in multi-environment crop trials. G3 Genes[Genomes]Genet. 6, 1313–1326. doi: 10.1534/g3.116.027524
- Olatoye, M. O., Clark, L. V., Labonte, N. R., Dong, H., Dwiyanti, M. S., Anzoua, K. G., et al. (2020). Training population optimization for genomic selection in miscanthus. G3 Genes Genees Genet. 10, 2465–2476. doi: 10.1534/g3.120.401402

- Pandey, M. K., Chaudhari, S., Jarquin, D., Janila, P., Crossa, J., Patil, S. C., et al. (2020). Genome-based trait prediction in multi- environment breeding trials in groundnut. Theor. Appl. Genet. 133, 3101-3117. doi: 10.1007/s00122-020-03658-1
- Pérez-Rodríguez, P., Gianola, D., González-Camacho, J. M., Crossa, J., Manès, Y., and Dreisigacker, S. (2012). Comparison between linear and nonparametric regression models for genome-enabled prediction in wheat. G3 Genes|Genomes|Genetics 2, 1595-1605. doi: 10.1534/g3.112.003665
- Poland, J. A., Brown, P. J., Sorrells, M. E., and Jannink, J.-L. (2012). Development of high-density genetic maps for barley and wheat using a novel two-enzyme Genotyping-by-Sequencing approach. PLoS ONE 7:e32253. doi: 10.1371/journal.pone.0032253
- Poland, J. A., and Rife, T. W. (2012). Genotyping-by-Sequencing for plant breeding and genetics. Plant Genome 5:005. doi: 10.3835/plantgenome,2012.05.0005
- R Core Team (2019). R: A Language and Environment for Statistical Computing. R Found. Stat. Comput. Avaialble online at: https://www.R-project.org/
- Rincent, R., Kuhn, E., Monod, H., Oury, F.-X., Rousset, M., Allard, V., et al. (2017). Optimization of multi-environment trials for genomic selection based on crop models. Theor. Appl. Genet. 130, 1735-1752. doi: 10.1007/s00122-017-29 22-4
- Rode, J., Ahlemeyer, J., Friedt, W., and Ordon, F. (2011). Identification of markertrait associations in the German winter barley breeding gene pool (Hordeum vulgare L.). Mol. Breed. 30, 831-843. doi: 10.1007/s11032-011-9667-6
- Roorkiwal, M., Jarquin, D., Singh, M. K., Gaur, P. M., Bharadwaj, C., Rathore, A., et al. (2018). Genomic-enabled prediction models using multienvironment trials to estimate the effect of genotype × environment interaction on prediction accuracy in chickpea. Sci. Rep. 8, 1-11. doi: 10.1038/s41598-018-30027-2
- Rutkoski, J., Singh, R. P., Huerta-Espino, J., Bhavani, S., Poland, J., Jannink, J. L., et al. (2015). Genetic gain from phenotypic and genomic selection for quantitative resistance to stem rust of wheat. Plant Genome 8:74. doi: 10.3835/plantgenome2014.10.0074
- Sallam, A. H., Endelman, J. B., Jannink, J.-L., and Smith, K. P. (2015). Assessing genomic selection prediction accuracy in a dynamic barley breeding population. Plant Genome 8:20. doi: 10.3835/plantgenome2014.05.0020
- Sehgal, D., Autrique, E., Singh, R., Ellis, M., Singh, S., and Dreisigacker, S. (2017). Identification of genomic regions for grain yield and yield stability and their epistatic interactions. Sci. Rep. 7, 1-12. doi: 10.1038/srep41578
- Sharma, R. C., and Smith, E. L. (1986). Selection for high and low harvest index in three winter wheat populations1. Crop Sci. 26, 1147-1150. doi: 10.2135/cropsci1986.0011183X002600060013x
- Thorwarth, P., Ahlemeyer, J., Bochard, A.-M., Krumnacker, K., Blümel, H., Laubach, E., et al. (2017). Genomic prediction ability for yield-related traits

in German winter barley elite material. Theor. Appl. Genet. 130, 1669-1683. doi: 10.1007/s00122-017-2917-1

- Tolhurst, D. J., Mathews, K. L., Smith, A. B., and Cullis, B. R. (2019). Genomic selection in multi-environment plant breeding trials using a factor analytic linear mixed model. J. Anim. Breed. Genet. 136, 279-300. doi: 10.1111/jbg.12404
- Tomar, V., Dhillon, G. S., Singh, D., Singh, R. P., Poland, J., Chaudhary A. A., et al. (2021). Evaluations of genomic prediction and identification of new loci for resistance to stripe rust disease in wheat (Triticum aestivum L.) Front. Genet. 12:710485 (in press). doi: 10.3389/fgene.2021.710485
- Velu, G., Crossa, J., Singh, R. P., Hao, Y., Dreisigacker, S., Perez-Rodriguez, P., et al. (2016). Genomic prediction for grain zinc and iron concentrations in spring wheat. Theor. Appl. Genet. 129, 1595-1605. doi: 10.1007/s00122-016-2726-y
- Wang, X., Li, L., Yang, Z., Zheng, X., Yu, S., Xu, C., et al. (2016). Predicting rice hybrid performance using univariate and multivariate GBLUP models based on North Carolina mating design II. Hered 118, 302-310. doi: 10.1038/hdy.2016.87
- Würschum, T., Leiser, W. L., Langer, S. M., Tucker, M. R., and Longin, C. F. H. (2018). Phenotypic and genetic analysis of spike and kernel characteristics in wheat reveals long-term genetic trends of grain yield components. Theor. Appl. Genet. 131, 2071-2084. doi: 10.1007/s00122-018-3133-3
- Zhang, J., Song, Q., Cregan, P. B., and Jiang, G.-L. (2015). Genome-wide association study, genomic prediction and marker-assisted selection for seed weight in soybean (Glycine max). Theor. Appl. Genet. 129, 117-130. doi: 10.1007/s00122-015-2614-x

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Tomar, Singh, Dhillon, Chung, Poland, Singh, Joshi, Gautam, Tiwari and Kumar. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Tomar et al