



# **Corrigendum: Membrane Inlet Mass Spectrometry: A Powerful Tool for Algal Research**

Adrien Burlacot<sup>†</sup>, François Burlacot<sup>†</sup>, Yonghua Li-Beisson<sup>†</sup> and Gilles Peltier<sup>\*†</sup>

Aix Marseille Univ, Commissariat à l'énergie Atomique et aux énergies Alternatives (CEA), Centre National de la Recherche Scientifique (CNRS), Institut de Biosciences et Biotechnologies d'Aix- Marseille (BIAM), CEA Cadarache, Saint Paul-Lez-Durance, France

# **OPEN ACCESS**

#### Edited by:

Pietro Franceschi, Fondazione Edmund Mach. Italy

> **Reviewed by:** Giuseppe Pieraccini, University of Florence. Italy

> > \*Correspondence: Gilles Peltier gilles.peltier@cea.fr

#### <sup>†</sup>ORCID:

Adrien Burlacot orcid.org/0000-0001-7434-6416 François Burlacot orcid.org/0000-0001-9783-6848 Yonghua Li-Beisson orcid.org/0000-0003-1064-1816 Gilles Peltier orcid.org/0000-0002-2226-3931

#### Specialty section:

This article was submitted to Technical Advances in Plant Science, a section of the journal Frontiers in Plant Science

> Received: 20 November 2020 Accepted: 03 December 2020 Published: 22 December 2020

### Citation:

Burlacot A, Burlacot F, Li-Beisson Y and Peltier G (2020) Corrigendum: Membrane Inlet Mass Spectrometry: A Powerful Tool for Algal Research. Front. Plant Sci. 11:631667. doi: 10.3389/fpls.2020.631667 Keywords: gas exchange, photosynthesis, carbonic anhydrase,  $CO_2$  concentrating mechanism,  $O_2$  evolution,  $H_2$  production, microalgae, cyanobacteria

#### A Corrigendum on

## Membrane Inlet Mass Spectrometry: A Powerful Tool for Algal Research

by Burlacot, A., Burlacot, F., Li-Beisson, Y., and Peltier, G. (2020). Front. Plant Sci. 11:1302. doi: 10.3389/fpls.2020.01302

In the original article, in the paragraph on the "Assessment of Photosynthetic Oxygen Exchange," we defined the  $O_2$  uptake rate with a negative value when oxygen is consumed. Although it has been historically the first way to define it (Hoch and Kok, 1963), it makes more sense to use a positive value as adopted later by Radmer and Kok (1976) because a negative uptake would mean the usage of a double negative which implies production.

Therefore, Equations (1), (2), and (3) should be read;

$$O_2 \ Uptake = -v_{18}O_2 \times (1 + \frac{C_{16}O_2(t)}{C_{18}O_2(t)})$$
(1)

$$O_2 Evolution = v_{16}_{O_2} - v_{18}_{O_2} \times \frac{C_{16}_{O_2}(t)}{C_{18}_{O_2}(t)}$$
(2)

$$Net O_2 = O_2 Evolution - O_2 Uptake$$
(3)

As a consequence, the plots shown on the original **Figure 5** have been replaced by the attached **Figure 5**.

The authors would like to thank Dr. Duncan Fitzpatrick for highlighting this problem and for suggesting changes to increase clarity of the article.

The authors apologize for this error and state that this does not change the scientific conclusions of the article in any way. The original article has been updated.

# REFERENCES

- Hoch, G., Kok, B. (1963). A mass spectrometer inlet system for sampling gases dissolved in liquid phases. *Arch. Biochem. Biophys.* 101, 160–170. doi: 10.1016/0003-9861(63)90546-0
- Radmer, R. J., Kok, B. (1976). Photoreduction of O<sub>2</sub> pimes and replaces CO<sub>2</sub> assimilation. *Plant Physiol.* 58, 336–340. doi: 10.1104/pp.58.3.336

Copyright © 2020 Burlacot, Burlacot, Li-Beisson and Peltier. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.



**FIGURE 5** [*In vivo* measurements of photosynthetic  $O_2$  exchange in the presence of <sup>18</sup>O-labeled  $O_2$ . (**A**) Schematic view of oxygen exchange illustrated in *C. reinhardtii*. While photosystem II (PSII) produces unlabeled  $O_2$  from the photolysis of H<sub>2</sub>O, oxygen uptake mechanisms consume both <sup>18</sup>O-labeled and unlabeled  $O_2$  (**B**) <sup>16</sup>O<sub>2</sub> and <sup>18</sup>O<sub>2</sub> concentrations measured in *C. reinhardtii* cells during dark–light transients. (**C,D**) Calculated cumulated  $O_2$  exchanges (**C**) and the corresponding  $O_2$  exchange rates (**D**) for the same experiment. Cells were grown photoautotrophically in air, centrifuged and resuspended in fresh medium at a concentration of 20  $\mu$ g ChI ml<sup>-1</sup>. Upon addition of 5 mM HCO<sub>3</sub><sup>-</sup>, <sup>18</sup>O<sub>2</sub> was injected inside the cell suspension, and the reaction vessel was closed. After 5 min of dark adaptation, green light was turned on (500  $\mu$ mol photon m<sup>-2</sup> s<sup>-1</sup>) for 10 min. Levels of <sup>16</sup>O<sub>2</sub> and <sup>18</sup>O<sub>2</sub> were recorded at respective m/z = 32 and 36. O<sub>2</sub> Uptake (red), O<sub>2</sub> Evolution (blue), and Net O<sub>2</sub> production (black) were calculated as described; cumulated gas exchange were calculated by directly integrating obtained exchange rates. To limit noise on the exchange rates graphic, data shown in (**D**) are integrated with a sliding average of 30 s wide.