



Role of Exogenous and Endogenous Hydrogen Sulfide (H₂S) on Functional Traits of Plants Under Heavy Metal Stresses: A Recent Perspective

Muhammad Saleem Arif¹, Tahira Yasmeen^{1*}, Zohaib Abbas¹, Shafaqat Ali^{1,2*}, Muhammad Rizwan¹, Nada H. Aljarba³, Saad Alkahtani⁴ and Mohamed M. Abdel-Daim^{4,5}

¹ Department of Environmental Science and Engineering, Government College University Faisalabad, Faisalabad, Pakistan, ² Department of Biological Sciences and Technology, China Medical University, Taichung, Taiwan, ³ Department of Biology, College of Science, Princess Nourah Bint Abdulrahman University, Riyadh, Saudi Arabia, ⁴ Department of Zoology, College of Science, King Saud University, Riyadh, Saudi Arabia, ⁵ Pharmacology Department, Faculty of Veterinary Medicine, Suez Canal University, Ismailia, Egypt

OPEN ACCESS

Edited by:

Mukesh Kumar Kanwar, Zhejiang University, China

Reviewed by:

Elke Bloem, Julius Kühn-Institut, Germany Syed Tahir Ata-UI-Karim, The University of Tokyo, Japan

*Correspondence:

Tahira Yasmeen rida_akash@hotmail.com; tahirayasmeen@gcuf.edu.pk Shafaqat Ali shafaqataligill@yahoo.com; shafaqataligill@gcuf.edu.pk

Specialty section:

This article was submitted to Plant Nutrition, a section of the journal Frontiers in Plant Science

Received: 25 March 2020 Accepted: 27 November 2020 Published: 07 January 2021

Citation:

Arif MS, Yasmeen T, Abbas Z, Ali S, Rizwan M, Aljarba NH, Alkahtani S and Abdel-Daim MM (2021) Role of Exogenous and Endogenous Hydrogen Sulfide (H₂S) on Functional Traits of Plants Under Heavy Metal Stresses: A Recent Perspective. Front. Plant Sci. 11:545453. doi: 10.3389/fpls.2020.545453 Improving growth and productivity of plants that are vulnerable to environmental stresses, such as heavy metals, is of significant importance for meeting global food and energy demands. Because heavy metal toxicity not only causes impaired plant growth, it has also posed many concerns related to human well-being, so mitigation of heavy metal pollution is a necessary priority for a cleaner environment and healthier world. Hydrogen sulfide (H₂S), a gaseous signaling molecule, is involved in metalrelated oxidative stress mitigation and increased stress tolerance in plants. It performs multifunctional roles in plant growth regulation while reducing the adverse effects of abiotic stress. Most effective function of H₂S in plants is to eliminate metal-related oxidative toxicity by regulating several key physiobiochemical processes. Soil pollution by heavy metals presents significant environmental challenge due to the absence of vegetation cover and the resulting depletion of key soil functions. However, the use of stress alleviators, such as H₂S, along with suitable crop plants, has considerable potential for an effective management of these contaminated soils. Overall, the present review examines the imperative role of exogenous application of different H₂S donors in reducing HMs toxicity, by promoting plant growth, stabilizing their physiobiochemical processes, and upregulating antioxidative metabolic activities. In addition, crosstalk of different growth regulators with endogenous H_2S and their contribution to the mitigation of metal phytotoxicity have also been explored.

Keywords: biochemical properties, physiological activities, heavy metal stress, hydrogen sulfide, signaling molecule, oxidative impairment

INTRODUCTION

Heavy metals are a group of metal elements having peculiar physical and chemical properties, which are also known to possess higher specific gravity > 4 g cm⁻³ in nature (Grant and Grant, 1987; Duffus, 2002). Environmental occurrence of heavy metal can be of both natural and anthropogenic origin; however, unprecedented release and their strong ecological persistence have now become a

1

serious toxicological and public health challenge worldwide (Arif et al., 2019; Zheng et al., 2020). Under natural conditions, heavy metals are the intrinsic component of earth crust and are often dispersed in soil, water, and atmosphere as a result of many geological processes, i.e., forest fire and volcanic eruption (Lado et al., 2008). A range of anthropogenic activities, such as intensive pesticides, as well as fertilizers use, vehicular emissions, mining activities, and industrial wastes, are the main contributors of heavy metal pollution across various domains of environment. Besides this, heavy environmental loading of metal toxicants also emanates from different wastewater sources. Globally, large volume of untreated wastewater is being discharged directly into waterways and soil, where they pose serious concerns for ecosystem stability (Mataka et al., 2006).

In soil, level of individual heavy metal concentration is a primary indicator often used to determine the degree of ecotoxicological effects on plants. For instance, some metals, such as nickel (Ni), molybdenum (Mo), zinc (Zn), and copper (Cu), are plant micronutrients and are phytotoxic only if their concentration is higher in soil (Lasat, 2002), whereas few other metallic elements, in particular chromium (Cr), lead (Pb), and cadmium (Cd), are hazardous to plants even at low soil concentrations. Heavy metals can either be found in dissolved or immobilized form; however, high immobilization rates can cause stronger detrimental effects on plants because of their in situ persistence and concentration buildup over time (Alloway, 1995, 2013). Consequently, they tend to impair various growth attributes of plants (Ahmad et al., 2012). Furthermore, some of these metals, such as Cr, Cd, Mn, and Zn, are also recognized to exert hormetic responses, which are reflected by a positive growth response at low concentrations and by phytotoxicity at higher metal concentrations (Azevedo and Lea, 2005). Detrimental effects of heavy metals on plants may involve oxidative stress, stunted growth, and toxicity-induced metabolic anomalies.

After nitric oxide and carbon monoxide, hydrogen sulfide (H₂S) is the third most important naturally occurring gaseous molecule known for its cellular signaling in biology (Yang et al., 2008). In plants, synthesis and release of H₂S typically occur during different stages of metabolic activities. It is generally formed in cut branches, tissue cultures, and leaf discs, whereas it is discharged into the surrounding environment from green cells of the plants (Rennenberg et al., 1990). Under normal growth conditions, numerous plants such as soybean, pumpkin, cotton, cantaloupe, squash, corn, and cucumber were found to release H₂S from leaf into the exterior environment (Wilson et al., 1978). Besides increasing enzymatic activity of vigorous cysteine (Cys), H₂S secretion is also recognized to enhance sulfite and sulfate metabolic activities (Rennenberg, 1983, 1984). However, numerous reports have observed paradoxical functions of H₂S in the regulation of plants physiological and biochemical traits. For instance, H₂S has been established to act as sulfur source at lower concentration, while promoting phytotoxic effects on plant growth at elevated level (De Kok et al., 2002; Li, 2013; Hancock and Whiteman, 2016; Li et al., 2016; Hancock, 2017; Huo et al., 2018). In most plants, elongated exposure to higher H₂S level eventually caused leaf removal after developing leaf injury, which can lead to overall deterioration of plant growth.

Likewise, H_2S triggered O_2 obstruction and subsequent impaired nutrient acquisition in rice seedlings were reported by Wang et al. (2012). Positive influence of H_2S on various plants has also been reported in literature. Shoot deposited sulfur (S) from H_2S appears to provide major site-specific S regulation for plants to improve plant growth, particularly under sulfur-deprived conditions. In many plants, H_2S is reported to be involved in the regulation of key physiological and growth functions, such as formation of adventitious root in cucumber, stomatal conductance in *Arabidopsis thaliana*, enhanced tolerance against salinity in alfalfa during seed germination, and regulation of thiol levels in *A. thaliana* (Riemenschneider et al., 2005a,b; Lisjak et al., 2010; Lin et al., 2012).

The H_2S is a convenient stress signaling molecule, as its biosynthesis process can take place in various cellular components once plants experience stress such as heavy metal exposure (Zulfiqar and Hancock, 2020). Upon its production, H_2S can be highly mobile across plant membranes and can either be influxed in or effluxed out of the plant system as a way out against heavy metal stress (Shivaraj et al., 2020). Plant stress adaptation against heavy metal in H_2S -treated plant is initiated via antioxidant activities (Kushwaha and Singh, 2019), accumulation of osmoregulators (Tian et al., 2016), cell signaling protein (He et al., 2019), and by different gene expressions (Pandey and Gautam, 2019). Overall, it enables plants to combat against stress factor such as heavy metals via effective removal of reactive oxygen species (ROS) by adjusting intracellular redox balance.

A plant experiment with nickel spiking has demonstrated that H_2S can enhance rice nickel tolerance prompted mainly by preventing chloroplast damage as a result of improved N metabolism under excessive nickel contamination (Rizwan et al., 2019). However, the role of H_2S as a signaling molecule in plants is still not fully understood despite the fact that the release of H_2S has been demonstrated in many plant species. Notably, desulfhydrases (H_2S -releasing enzymes) have functionally been endorsed as key H_2S volatile in plants (Riemenschneider et al., 2005b). In another case, enhanced L-Cys desulfhydrase (LCD) activity under biotic stress further reaffirms its significant potency as an adaptive defense approach under stress agriculture (Rausch and Wachter, 2005). H_2S was also involved in a promotional role of superior root organogenesis in *Ipomoea batatas, Salix matsudana*, and *Glycine max* L. (Zhang et al., 2009).

The antioxidant enzymes are another type of stress mediator in plants, which are often activated as a key defense response after recognition of given stressor, including heavy metals (Zhang et al., 2010a,b). Foliar application of sodium hydrosulfide (NaHS), an H₂S donor, led to an upscale induction response of different antioxidant enzymes and reduced the concentration of H₂O₂ in wheat seedlings to enhance resistance against heat stress. Also, Zhang et al. (2011) emphasized that H₂S can have an effective role in plant protection against different types of oxidative stress. Water-soluble antioxidants, such as ascorbic acid and glutathione, were expressed at enhanced level upon fumigation with H₂S, which consequently delivered higher water stress tolerance in wheat plants (Shan et al., 2011). As NaHS is characterized as key antioxidant inducer; therefore, plant growth promotive activities including root organogenesis (Zhang et al., 2009), stomatal regulation (Lisjak et al., 2010), and seed germination (Zhang et al., 2010a,b) were substantially improved in response to heavy metal stresses. Interestingly, exogenously applied H_2S (100 ppb) has shown striking effect on plant growth improvement of beet, alfalfa, and lettuce (Thompson and Kats, 1978). Furthermore, H_2S foliar spraying has also improved the accumulation of vital nutrients in plant (Wang et al., 2012).

As a secondary messenger, nitric oxide and H₂S can collectively trigger signal transduction resulting in higher intracellular buildup of these molecules, which are indeed a plant's necessity to cope with metal-induced oxidative stress. In this way, use of H₂S with nitric oxide could exert a regulatory response to transporters and antioxidant systems to alleviate phytotoxic effects of heavy metals (Li et al., 2012; Wang et al., 2019). Considering all the background information about the role of H₂S in biological system and, most importantly, its interaction with crop plants under stress agriculture, we aimed to get an advanced overview of H₂S-related growth promotive effects on crop plants. Soil contamination by heavy metals imposes greater ecological concern because of lack of plant cover and resultant land degradation. However, use of stress alleviators, such as H₂S, along with suitable crop plant holds a great potential for the successful restoration of these contaminated soils. We also focus on how the use of H₂S and other precursor compounds interacts with plants to counteract heavy metal toxicity. Moreover, our discussion also focuses H₂S-mediated response mechanisms exhibited by plants toward heavy metal toxicity.

CHEMISTRY OF H₂S

 H_2S is a weak acid with good water solubility and commonly exists in neutral molecular form (H_2S). Although HS^- is a major ionic form of H_2S involved in most of its biological reactions, S^{2-} also exists in minor concentration due to higher dissociation constant for second ionization (Filipovic et al., 2018). Despite being a highly water-soluble compound, H_2S tends to be unstable under natural conditions as it slowly oxidizes to elemental sulfur having a weak solubility in aqueous solution. Moreover, the volatile nature of H_2S exacerbates its experimental use in the environment. For instance, nearly half-dose of H_2S could be lost in 5 min from open cell culture wells (DeLeon et al., 2012). Consequently, H_2S handling imposes greater challenge of its precise measurement under field conditions (Wang et al., 2014; Peng and Xian, 2015).

$$H_2S \rightleftharpoons HS^- + H^+ \rightleftharpoons S^{2-} + 2H^+$$
 (in aqueous solution)

OCCURRENCE AND BIOSYNTHESIS OF H₂S IN PLANTS

There has been a proposition long ago that plants can produce and release H_2S themselves, particularly when exposed to external sulfur (S) stimuli, i.e., Cys, sulfate, sulfite, or SO₂ (Wilson et al., 1978; Sekiya et al., 1982a,b). This was thought to be a mechanism for regulating sulfur homeostasis (Calderwood and Kopriva, 2014). However, mechanistic understanding of H₂S generation in plants and its interaction with other cellular components remains elusive. In higher plants, H₂S biosynthesis pathway emerges in different subcellular compartments, where main enzymes linked to sulfur metabolism have the potential to initiate H₂S biogenesis (Corpas et al., 2019b; Chen et al., 2020). In plants, most common enzymes involved in the H₂S biosynthesis enzymes include LCD, D-Cys desulfhydrase (DCD), 1-3-cyanoalaninesynthase, sulfite reductase, and Cys synthase (Yamasaki and Cohen, 2016). Among various plant cellular organelles, chloroplast serves as the major H₂S production site due to localization of sulfite reductase enzyme, which catalyzes the reduction of sulfite to sulfide during sulfate assimilation pathway. In addition to this, cytosol can also generate H₂S by the action of DCD and LCD, accompanied by ammonia and pyruvate production. In chloroplast, sulfide concentration is two times greater than that found into the cytosol (Krueger et al., 2009). However, this sulfide is dissociated into its ionized forms due to the basic physiological conditions and therefore unable to pass through the membranes to the cytosol (Kabil and Banerjee, 2010). In mitochondria, the synthesis of H_2S can be regulated by β-cyanoalanine synthase (a pyridoxal phosphatedependent enzyme), which transforms both cyanide and L-Cys into β -cyanoalanine and H₂S in order to degrade toxic cyanogen (Gotor et al., 2019). Recently, Corpas et al. (2019a) also provided an evidence of H₂S generation in the peroxisomes of Arabidopsis; however, it is still unclear whether its generation pathway is endogenous or recruited from other cellular compartments (e.g., cytosol). Additionally, the expression of plant cellular proteins [L-Cys desulfurase such as O-acetylserine thiol lyase (OAS-TL) and Nifs-like proteins] also processes the H₂S synthesis in different cellular organelles (Gotor et al., 2019). These enzymes, with their varied expression, are therefore involved in controlling the production of H₂S across various cellular compartments of the plant. On the other hand, occurrence of H₂S in these cellular organelles with strong lipophilic characteristic promotes its translocation in the lipid bilayer of cell membranes (Cuevasanta et al., 2017).

H₂S is a flammable-toxic gaseous molecule that is often distinguished by stinky rotten eggs smell. It has also shown strong concentration dependent affinity for reactions as it can disrupt mitochondrial cytochrome activity and even reduce mitochondrial respiration (Mancardi et al., 2009). Surprisingly, research in recent years has unraveled the significance of H₂S as a gasotransmitter that promotes plant growth and development at various stages of plant life cycle (Xuan et al., 2020). Other than algae, fungi, and few prokaryotes, plants are known to take leverage of taking up the naturally occurring sulfate (SO_4^{2-}) source of S from soil and incorporate it into organic forms (Takahashi et al., 2011). In S assimilation process, sulfate taken up by plant roots is initially reduced to H₂S by catalytic activity of adenosine 5'-phosphosulfate reductase and sulfite reductase and eventually transformed into Cys via O-OAS-TL. Therefore, H₂S is an extremely important intermediate in the thiometabolism pathway.

The use of H_2S as signaling molecule has now become very common; thus, basic mechanism of its functional activities has been decrypted (Mustafa et al., 2009; Aroca et al., 2015). Some recent proteomic analyses have described a new posttranslational modification of proteins, where reactive Cys residues (as an H_2S signal) can modify protein function by converting the thiol group (-SH) into a persulfide group (-SSH) known as persulfidation. In most cases, persulfide adducts exhibit higher nucleophilicity relative to the thiol group, and as a result, modified Cys displayed highly complex reactivity (Paul and Snyder, 2012). This might be the rationale of widespread persulfidation in nature, which largely affects protein over O₂ and N species (Ida et al., 2014) (**Figure 1**). Overall, complex functional interactions of H_2S based on its donor, concentration gradient, and plant section tend to describe actions of specific protein after translational modification.

The nature of H₂S-mediated specific cellular modifications still lacks clarity, because thermodynamic reaction involving H₂S and a thiol is unfavorable. Sulfane sulfur is a sulfur atom that has the peculiar ability to bind reversibly to other sulfur atoms to form hydropersulfides (R-S-SH) and polysulfides (-S-Sn-S-). These polysulfides tend to be far more efficient in persulfidation, as they are more nucleophilic than H₂S (Toohey, 2011). Recently, new molecular weight persulfides were identified as possible mediator of sulfide signaling. In this relation, Cys-persulfide (Cys-SSH), glutathione persulfide, and its persulfurated species Cys-SSnH and GSSnH have been designated as redox regulators (Kasamatsu et al., 2016; Kimura et al., 2017). Recently, the endogenous Cys-SSH production synthetized by prokaryotic and mammalian cysteinyl-tRNA synthetases using L-Cys as substrate has been described. The Cys polysulfides bound to tRNA are incorporated into polypeptides that are synthesized de novo in the ribosomes, suggesting that these enzymes are the principal Cys persulfide synthases in vivo (Akaike et al., 2017).

PROMOTIVE ROLE OF H₂S IN REGULATING PLANT HEAVY METAL STRESS

Of various abiotic stresses in environment, heavy metal-triggered stress always has very serious repercussions for plant growth and productivity. Like other abiotic stress, heavy metal stress is also associated with unregulated overproduction of ROS, which can influence plant metabolism and physiological activities by inflicting range of oxidative stress damages. Among the signaling molecules, H_2S is now an established regulator of growth in plants exposed to plethora of abiotic stresses, including heavy metal stress (Rather et al., 2020).

EXOGENOUS H₂S APPLICATION: A PRECURSOR OF HEAVY METAL STRESS ALLEVIATION IN PLANTS

Given a suite of heavy metal mitigation approaches competing for an effective stress management in agriculture, it is extremely pertinent to consider only those measures that can provide plant benefits only in an eco-friendly way. Although H_2S is toxic for many living organisms as evidenced by mitochondrion inhibition (Wang et al., 2019), its central role in key plant physiological functions and therapeutic use in various human ailments have been well developed in recent times (Aroca et al., 2020). The H_2S in gaseous form is the simplest method of its usage in the laboratory; however, it is not realistic practice due to nontargeted ecotoxicological consequences both for the humans and environment (Rubright et al., 2017). In most cases, NaHS and/or sodium sulfide (Na₂S) are used as H_2S donor molecules specifically due to their higher dissolution, resulting in a short albeit sustained pulse of H_2S (**Table 1**).



H ₂ S source	H ₂ S conc.	Plant species	Experiment	Duration/ growth stage	Heavy metal/source	Exposure level	Tolerance mechanisms involved	References
Sodium hydrosulfide (NaHS)	100 μΜ	Alfalfa (Medicago sativa L.)	Potted soil	60 days/ young plant	Lead, cadmium	217 mg kg ⁻¹ of Pb 4.95 mg kg ⁻¹ of Cd	Upscaled antioxidant activity (POD, CAT, APX, SOD) and reduced oxidative damage (MDA, H_2O_2 , O^2) decreased the absorption of metal ions	Fang et al., 2020
Sodium hydrosulfide (NaHS)	100 μΜ	Zucchini (<i>Cucurbita pepo</i> L.)	Potted soil	14 days/ young seedlings	Nickel: Ni (NO ₃) ₂	50 mg of Ni	Decreased metal ions accumulation, restored ionic homeostasis and averted oxidative membrane damages by upregulation of phenolic and flavonoid metabolites	Valivand and Amooaghaie, 2020
Sodium sulfite (Na ₂ SO ₃)	0.5 mM	Foxtail millet (Setaria italica L.)	Hydroponic culture	12 days/ young seedlings	Cadmium	100 μM	Suppressed nitrate reductase and nitric oxide synthase-dependent endogenous nitric oxide (NO), which further enhanced the enzyme activities, i.e., SOD and POD, leading to the rescue of Cd related root growth inhibition	Han et al., 2020
Sodium hydrosulfide (NaHS)	200 μΜ	Cauliflower (<i>Brassica</i> oleracea L.)	Potted soil	42 days/ transplants	Chromium: K ₂ Cr ₂ O ₇	10–100 μM	Restricted oxidative stress damages (EL, MDA, H ₂ O ₂), increased antioxidant activity (SOD, CAT, APX, POD) in leaves and root tissues	Ahmad et al., 2020
Sodium hydrosulfide (NaHS)	150 μΜ	Woad plant (Isatis indigotica L.)	Hydroponic culture	28 days/ young seedlings	Cadmium	22.5 μM	Decreased intracellular metal ion toxicity and induced production of metallothioneins (metal-binding protein) restricting root-to-shoot Cd translocation	Jia et al., 2020
Sodium hydrosulfide (NaHS)	0.2 mM	Cauliflower (Brassica oleracea L.)	Hydroponic culture	24 days/ young seedlings	Lead: Pb (CH ₃ COO) ₂ ·3H ₂ O	0.5 mM	Scavenging of ROS through antioxidant activity and Pb ion chelation by non-protein thiol and total glutathione	Chen et al., 2018
Sodium hydrosulfide (NaHS)	0.2 mM	Pepper (Capsicum annuum L.)	Sand, peat, and perlite mixed pots	70 days/ mature plants	Zinc: ZnSO ₄ · 7H ₂ O	0.5 mM	Reduced metal ions uptake, improved N, P, and Fe uptake and promoted antioxidant activities for limiting membrane oxidative stress damage	Kaya et al., 2018
Sodium hydrosulfide (NaHS)	500 μΜ	Maize (Zea mays L.)	Hydroponic culture	19 days/ young seedlings	Chromium	200 μΜ	Alleviated Cr toxicity by reducing the production of cyto-toxic methylglyoxal via glutathione-S-transferase (GST) detoxification	Kharbech et al., 2020
Sodium hydrosulfide (NaHS)	50 μΜ	Oilseed rape (<i>Brassica</i> napus L.)	Hydroponic culture	18 days/ young seedlings	Cadmium: CdCl ₂	20 µM	Prompted higher L-cysteine desulfhydrase (LCD) for S-metabolism, minimized Cd translocation to shoots-leaves and prevented chlorosis	Yu et al., 2019

TABLE 1 Effects of exogenous application of	different H ₂ S sources on plant gro	with regulations under heavy metals stress.
---	---	---

(Continued)

TABLE 1 | Continued

H ₂ S source	H_2S conc.	Plant species	Experiment	Duration/ growth stage	Heavy metal/source	Exposure level	Tolerance mechanisms involved	References
Sodium hydrosulfide (NaHS)	100 μM	Coriander (Coriandrum sativum L.)	Hydroponic culture	18 days/ young seedlings	Copper: CuSO ₄	100 μΜ	Reduced electrolyte leakage by regulating antioxidant enzyme activity via ascorbate-dlutathione cycle	Karam and Keramat, 2017
Sodium hydrosulfide (NaHS)	200 μM	Black night shade (Solanum nigrum L.)	Hydroponic culture	21 days/ young seedlings	Zinc: ZnCl ₂	400 μΜ	Reduced free cytosolic metal content in roots by upregulated metallothioneins mediated Zn-chelation and antioxidant response mechanism	Liu et al., 2016
Sodium hydrosulfide (NaHS)	0.8 mM	Wheat (<i>Triticum</i> aestivum L.)	Plastic trays/ sand- vermiculite	14 days/ young seedlings	Copper: CuSO4	100 μM	Ascorbate and glutathione metabolism minimized Cu toxicity by reducing malondialdehyde content and limiting electrolyte leakage	Shan et al., 2012
Sodium hydrosulfide (NaHS)	200 μM	Cotton (Gossypium hirsutum L.)	Hydroponic culture	28 days/ young seedlings	Lead: Pb (NO ₃) ₂	50 μM	Improved photosynthetic pigmentation, enhanced antioxidant activities and eventually decreasing malondialdehyde (MDA), electrolyte leakage, and H ₂ O ₂ production	Bharwana et al., 2014
Sodium hydrosulfide (NaHS)	100 μM	Pea (Pisum sativum L.)	Hydroponic culture	30 days/ young seedlings	Arsenate: Na ₂ HAsO ₄ × 7H ₂ O	50 μΜ	Revitalized redox cell status against arsenate toxicity by promoting ascorbate-glutathione metabolism and counteract ROS induced membrane damage	Singh et al., 2015
Sodium hydrosulfide (NaHS)	100 μΜ	Rice (Oryza sativa L.)	Hydroponic culture	35 days/ young seedlings	Nickel: NiSO4 · 6H20	200 μM	Enhanced chloroplast biogenesis and regulate nitrogen (N) metabolism via enzymes, i.e., nitrate reductase, nitrite reductase, glutamate synthase, glutamate oxaloacetate transaminase, glutamine synthetase, and glutamate pyruvate transaminase, and boost plant tolerance under Ni stress	Rizwan et al., 2019
Sodium hydrosulfide (NaHS)	200 µM	Rice (Oryza sativa L.)	Hydroponic culture	28 days/ young seedlings	Mercury: HgCl ₂	100 μΜ	Sequestered metal ions in roots and prevented plant oxidative damages by maintaining low MDA and H ₂ O ₂	Chen et al., 2016

POD, peroxidase; SOD, superoxide dismutase; CAT, catalase; APX, ascorbate peroxidase; MDA, malondialdehyde; H₂O₂, hydrogen peroxide, EL, electrolyte leakage.

Production of ROS, by-product of physiological metabolism, is a typical plant response under abiotic stress including heavy metal stress. Nevertheless, plants vary in their response against the different oxidative stresses linked to ROS in species-cultivar-specific ways. In plants, a robust antioxidative defense system (enzymatic and non-enzymatic) has been evolved to scavenge the excessive ROS accumulation, which can in turn counteract the harmful impacts of oxidative stress. Increasing evidence demonstrated that NaHS treatment can ameliorate and repair oxidative stress caused by heavy metal toxicity (Luo et al., 2020). Exogenous application of H_2S can suppress the burst of ROS by activating enzymatic and non-enzymatic defense components of ascorbate-glutathione cycle and eventually avert oxidative stress damage to plants.

Legume plants are of considerable significance for the remediation of heavy metal-polluted soil due to the unique symbiotic assemblage of N-fixing bacteria (rhizobia) with leguminous host plant (Reichman, 2007; Shen et al., 2019). In alfalfa, exogenous application of NaHS (100 μ M) has mitigated the compounding effects of dual metal stress (Pb/Cd) on legume-rhizobium symbiosis (**Table 1**; Fang et al., 2020). Application of H₂S donor molecule boosted up the survival rate of rhizobia by enhancing soil enzyme activity, facilitating

nutrient transformations, and shifting both composition and diversity of soil microbial population. This study concludes that H_2S -mediated symbiosis has resulted in development of greater plant resistance to metal-induced toxicity as evident by increased antioxidant enzyme activity and reduced metal ion absorption. Similar to these findings, Mostofa et al. (2015) investigated a rice–cadmium interaction model, where H_2S provided further evidence of being an efficient growth regulator to mitigate Cd-related growth suppression and reduction in biomass. Moreover, rice growth revitalization performance was primarily triggered by a three-way Cd alleviation process, including low Cd uptake/accumulation, mineral nutrient upregulation, and photosynthetic functions and timely induction of antioxidant response.

Heavy metals, such as Ni, are known to cause disruption in the absorption and utilization of key mineral elements in plants (Sharma and Dhiman, 2013). Importantly, Ni metal ions demonstrate a strong competitive affinity for bivalent cations, i.e., Ca, Mg, Mn, Fe, Zn, which could reduce uptake of these essential elements and obstruct normal plant growth and development (Ahmad et al., 2011). In hydroponic culture, NaHS (100 µM)treated young seedlings of Zucchini plant showed a decrease in Ni accumulation by reviving essential mineral homeostasis (Valivand and Amooaghaie, 2020). Furthermore, this study validates the role of H₂S in osmotic adjustment, as indicated by the proline and sugar content of plant exposed to heavy metal stress. Clearly, NaHS facilitated an increase in flavonoid and phenolic secretions likely to reduce the oxidative damage due to ROS scavenging, which in turn led to the improved Ni tolerance in Zucchini seedlings.

Phenolic and flavonoid are not the solitary metabolites those are exuded by heavy metal-stressed plants. Ahmad et al. (2020) demonstrated that NaHS treatment can upregulate antioxidant enzyme activity in Cr-exposed cauliflower seedlings, which is central to the amelioration of Cr-related oxidative damages, as well as reduction in metal ion translocation, to aerial plant parts. Root growth is an important predictor of plant productivity in agriculture, while impaired root growth has often been distinguished as one of the most common and earliest symptoms of plant exposure to metal toxicity (Liu et al., 2018). In foxtail millet, Cd-induced root growth inhibition was reversed by an SO₂ derivative compound Na₂SO₃ (Han et al., 2020). This study concludes the existence of crosstalk between SO₂ and nitric oxide (NO) for nitrate reductase (NR)-nitric oxide synthase-dependent endogenous NO signaling that prompted an upregulated antioxidant enzyme activity and suppressed genes associated with the Cd uptake (SiNRAMP1, SiNRAMP6, SiIRT1, and SiIRT2).

Plant cell wall is a first architectural barrier to avert transmembrane movements of toxic materials, such as heavy metals (Krzesłowska, 2011), as it can allow cellular compartmentation of the metal ions and reduce the phytotoxic effects of heavy metal exposure (Lai, 2015). Metal-tolerant proteins such as metallothionein and phytochelatins are crucial metal-binding ligand that regulates cationic homeostasis of plant cell wall (Yu et al., 2018; Zhi et al., 2020). In Woad plants, Jia et al. (2020) identified strict connection between Cd chelation and S metabolic products based on the weakening of metal ion translocation from root to shoot. The study confirmed that NaHS stimulated the endogenous metal binding proteins, i.e., metallothionein 1A and phytochelatins, which in turn promoted Cd accumulation in the cell wall by modifying its contents, thus reducing intracellular metal ion mobility for detoxification. Previously, Jia et al. (2016) established an intertwining effect of H₂S and Cys in A. thaliana L. They found that sulfur metabolism has key role to play in the growth and development of plants exposed to Cd toxicity. Collectively, plant stress alleviators such as H₂S and Cys have been shown to resurrect root growth, as well as to increase plant Cd tolerance via S metabolite feedback loop. An active synergy between H₂S and proline pulls together millet plant from negative effects of Cd toxicity, as evidenced by induced Cd tolerance and stimulated biomass production (Tian et al., 2016). Apart from higher proline accumulation, as well as proline dehydrogenase (PDH) and proline-5-carboxylate reductase (P5CR) activities, the transcript levels of PDH and P5CR were also enhanced after H₂S treatment.

Also, Yu et al. (2019) have shown that exogenous NaHS application can increase Cd retention into the root cell wall of oilseed rape by stimulating LCD activity. Furthermore, NaHS led to increase in cellular pectin, and root methylesterase activity validates higher metal-binding capacity of root cell that repressed Cd translocation to aerial plant sections. In another study, downregulated metal homeostasis and uptake were recorded when young seedlings of black night shade plant were treated with NaHS under Zn stress (Liu et al., 2016). It has been noted that expression of the metallothionein was increased, leading to an improvement in plant Zn tolerance as shown by the chelation of excessive Zn in the cytoplasm. Moreover, elevated expression of antioxidant enzyme, CAT2, also prevented oxidative stress damages in metal-stressed plants as a result of H_2S treatment.

Seed viability is one of the crucial determinants of healthy plant growth and increased production. Seed germination and its emergence and subsequent seedling establishment contribute proportionally to achieve sustainability, growth, and productivity. Exogenous NaHS application has improved both seed germination and seedling emergence in cauliflower by scavenging Pb-induced ROS (Chen et al., 2018). Interestingly, H₂S-led exhibition of plant protection against Pb stress was comparable to ROS scavengers, i.e., 4,5-dihydroxy-1,3-benzene disulfonic acid and N, N'-dimethylthiourea. They concluded that non-enzymatic antioxidants, such as non-protein thiol and total glutathione, were upregulated by H₂S and improved Pb tolerance by ROS-scavenging and/or directly chelating metal ions. Plant exposure to metal ions often represses the activity of transporters, resulting in ionic imbalance and reduced nutrients assimilation in plants (Vaculík et al., 2020). In Zn-exposed pepper plants, exogenous NaHS treatment reduced Zn plant accumulation and enhanced the absorption of key mineral elements, i.e., Fe, N, P (Kaya et al., 2020). Moreover, mitigating effects of NaHS have been further up-scaled by antioxidant activity and osmotic adjustment to minimize membrane oxidative damage. Methylglyoxal (MG), a cytotoxic metabolic by-product of glycolytic pathways, usually interacts with macromolecules to trigger protein inactivation and induces oxidative damages

to plant under abiotic stress, such as heavy metals (Hoque et al., 2016; Bhuyan et al., 2020). In maize, Cr tolerance of young seedlings was linked to the suppression of NADPH oxidase activity, resulting in restricted ROS accumulation following exogenous application of NaHS (Kharbech et al., 2020). This study illustrated the potential function of glutathione in minimizing Cr toxicity by reducing MG content while preserving glutathione-ascorbate homeostasis for additional S metabolism, as demonstrated by the activity of glutathione S-transferase and reductase enzymes. Similarly, NaHS pretreatment also reduced the lipid peroxidation and electrolyte leakage in coriander seedling exposed to Cu toxicity (Karam and Keramat, 2017). These results further substantiate that NaHS-led changes in endogenous H₂S were presumably involved in the prevention of oxidative damages via cellular ascorbate-glutathione cycle. Similarly, mitigation of Cu toxicity in wheat has been linked to H₂S-related ascorbate-glutathione cycle (Shan et al., 2012). Moreover, decrease in lipid peroxidation and electrolyte leakage further implies a systemic defense response activated by NaHS application.

Photosynthesis is a key physiological process for plant productivity and directly provides energy for plant growth. Photosynthetic pigments are very sensitive to various abiotic stress including heavy metals, which can negatively affect rate of photosynthesis via chlorophyll and carotenoid degradation (Amari et al., 2017). RuBISCO (a multi-meric photosynthetic enzyme), reflects potential for plant productivity and the efficiency of resource use by its net C assimilation rate. There is some substantial evidence in literature that described about H₂S role of being a key regulator of plant photosynthetic apparatus (Carmo-Silva et al., 2015; Cummins et al., 2018). In spinach, Chen et al. (2011) found that NaHS-treated plants had significant increase in chlorophyll content alongside higher soluble protein content and biomass yield. This possibly highlights the significance of RuBISCO activity, which promotes chloroplast biogenesis, photosynthetic enzyme expression, and thiol redox alterations. Also, Bharwana et al. (2014) reported that NaHS application could regulate the photosynthetic activity of cotton seedlings under Pb toxicity. They concluded that higher photosynthesis is obviously a defense mechanism intended to minimize metal toxicity by accelerating rate of photosynthesis, which ultimately contribute to cope with predictable oxidative stress. Further results showed that H₂S also promoted the reversal of electrolyte leakage in cotton seedlings caused by Pb toxicity, as shown by reduced H₂O₂ and MDA contents. In another study, Rizwan et al. (2019) reported NaHS-induced upregulation of chloroplast biogenesis and N metabolism in rice plant exposed to Ni stress. The NaHS application has been found to increase the activities of various N-related enzymes, i.e., NR, nitrite reductase, glutamate synthase, glutamate oxaloacetate transaminase, glutamine synthetase, and glutamate pyruvate transaminase. Furthermore, key involvement of H₂S in Ni stress regulation of rice plant was also validated by the expression of genes abundance associated with N metabolism. Also, Chen et al. (2016) revealed the molecular basis of rice stress adaptation against mercury (Hg) contamination. In their study, H₂S pretreatment extended membrane transcriptional expression of

bZIP60 and OsMT-1, which were involved in Hg localization in roots. In addition, Hg-related plant membrane damages were attenuated by scavenging ROS and downregulation of H_2O_2 and MDA, which eventually led to growth promotion of rice seedlings. The redox status of plant cell can be disrupted by accumulation of ROS associated with metal toxicity, resulting in a cascade of retarded physiological and morphological functions (Schutzendubel and Polle, 2002). In pea seedlings, H_2S application restored the cellular redox status of pea suffering from arsenate toxicity (Singh et al., 2015). It appears that H_2S mediated recovery of ascorbate–glutathione enzymes pool was a pivotal contributor to plant defense, as depicted by suppression of oxygen free radicals and membrane damage.

STRESS ALLEVIATORS AND ENDOGENOUS H₂S: A COMBATING TOOL TOWARD METAL DETOXIFICATION

In some recent reports, enrolment of different plant growth regulators has also unveiled endogenous H₂S synthesis in plant that eventually provide a protective role in crop plants under stress (Table 2). NO has been deemed to be an important signaling compound that can stimulate plant growth and development in agriculture. A plethora of studies has investigated the key involvement of NO in activating plant defense response to heavy metal stress (Bai et al., 2014; Rizwan et al., 2018; Hu et al., 2019; Sharma et al., 2020a). A strong synergic association between exogenously applied NO and resultant endogenous synthesis of H₂S was involved in enhanced Cr resistance of tomato seedlings (Alamri et al., 2020). It appears that increased S assimilation and related enzyme metabolic activities have mitigated the depressing effect of metal ions in both cellular and molecular levels. Salicylic acid (SA), a phenolic compound, has been known to control a broad range of physiological and biochemical functions in plants to combat stressful conditions, including heavy metals (Hasanuzzaman et al., 2019; Sharma et al., 2020b). The exogenous application of Na₂SiO₃, a silicon derivative compound, endorsed a functional relationship between NO and H₂S in regulating the Cd stress of pepper plant (Kaya et al., 2020). It was evident that upregulation of endogenous NO and H₂S was central to metal stress alleviation as confirmed by lower Cd content of leaves. In addition, improvement in plant antioxidant activities, nutrients uptake (e.g., Ca, K), photosynthesis activity, and water relations upscaled the plant metal tolerance. In pepper experiment, SA prompted 66% higher buildup of endogenous H₂S in Pbexposed leaves over untreated control (Kaya, 2020). It was further elucidated that efficient crosstalk between SA and H₂S averted induced phytotoxicity effect by minimizing Cd accumulation in leaves via upregulated metabolisms of the enzymes involved in ascorbate-glutathione cycle. Furthermore, SA and H₂S augmented leaf relative water, water potential, and proline content, which has key contribution in restoring the TABLE 2 | Use of different stress alleviators and their interactions with endogenous H_2S for higher metal tolerance in plants.

Stress alleviator	Exogenous H ₂ S	Plant species	Experiment	Duration/ growth stage	Heavy metal/source	Exposure level	Endogenous H_2S	Stress alleviating symptoms	References
Silicon (Na ₂ SiO ₃) 2.0 mM	NO	Pepper (Capsicum annuum L.)	Potted soil	35 days/ young seedlings	Cadmium: CdCl ₂	0.1 mM	Upregulated $\sim 10 \ \mu mol \ g^{-1}$ fresh weight	Reduced leaf Cd content and oxidative stress Improved K and Ca uptake	Kaya et al., 2020
Nitric oxide (NO) 50 μM	NO	Tomato (Solanum Iycopersicum L.)	Pot filled with vermiculite- perlite	21 days/ young seedlings	Chromium: K ₂ Cr ₂ O ₇	100 μM Cr (VI)	Upregulated ~98 nmol g ⁻¹ fresh weight	Increased S assimilation and kinked enzyme metabolism averted DNA and oxidative damages.	Alamri et al., 2020
Thiamine (THI) 50 mgL ⁻¹	0.2 mM sodium hydrosulfide (NaHS)	Strawberry (<i>Fragaria</i> × <i>ananassa</i> Duch)	Hydroponic culture	28 days	Cadmium: CdCl ₂	0.1 mM	Upregulated $\sim 15 \ \mu mol \ g^{-1}$ fresh weight	Upregulated endogenous NO and hydrogen sulfide (H ₂ S); Enhanced Ca and K uptake	Kaya and Aslan, 2020
0.5 mM salicylic acid (SA)	0.2 mM sodium hydrosulfide (NaHS)	Pepper (Capsicum annuum L.)	Perlite pot	28 days/ young seedlings	Lead; PbCl ₂	0.1 mM	Upregulated ~23 μmol g ⁻¹ fresh weight	Pb induced phytotoxicity was alleviated by SA + NaHS related upregulation in ascorbate–glutathione cycle	Kaya, 2020
Calcium (CaCl ₂) 15 mM	100 μM sodium hydrosulfide (NaHS)	Zucchini (<i>Cucurbita pepo</i> L.)	Hydroponic culture	14 days/ young seedlings	Nickel: Ni (NO ₃) ₂	50 mg L ^{−1}	Upregulated ~323 nmol g ⁻¹ fresh weight	Both extracellular (Ca ²⁺⁾ and intracellular (CaM) complex activated crosstalk of Ca ²⁺ and H ₂ S to revert plant oxidative damages	Valivand et al., 2019
0.5 mM salicylic acid (SA)	0.5 mM sodium hydrosulfide (NaHS	Maize (Zea mays L.)	Potted sand-perlite	9 days/ young seedlings	Lead: Pb (NO ₃) ₂	2.5 mM	Not detected	Regulated metal toxicity by increasing glycine betaine and NO at the expense of S-amino acids metabolism, i.e., arginine, methionine	Zanganeh et al., 2018
0.5 mM salicylic acid (SA)	NO	Maize (Zea mays L.)	Potted sand-perlite	9 days/ young seedlings	Lead: Pb (NO ₃) ₂	2.5 mM	Upregulated ~3.5 μmol g ⁻¹ fresh weight	Minimized chlorophyll related damages, built-up ascorbic acid glutathione and upregulation of antioxidant enzyme activity	Zanganeh et al., 2019
1 μM methyl jasmonate (MeJA)	50 μM sodium hydrosulfide (NaHS)	Foxtail millet (Se <i>taria italica</i> L.)	Hydroponic culture	14 days/ young seedlings	Cadmium chloride (CdCl ₂)	200 μΜ	Upregulated 75–120 nm g ^{–1} fresh weight	Alleviated growth retardations by decreasing H_2O_2 and malondialdehyde content, also repressed Cd accumulation in seedlings	Tian et al., 2017

phenotypic appearance of Pb-stressed plants. In another study with maize exposed to Pb stress, Zanganeh et al. (2018) pointed out the synergic effect of SA and H_2S , which contributed to the relegation of metal-induced phytotoxicity by augmenting glycine-betaine and NO signaling. It was found that SA and NO signaling play an instrumental role in plant growth regulation as they boosted the S-assimilation via arginine-methionine metabolism and thus prevented Pb-induced oxidative stress injury in plants.

Thiamine (THI) is another unique biomolecules that can effectively control plant growth by facilitating the synthesis of carbohydrate, nucleic acids, adenosine triphosphate, and nicotinamide adenine dinucleotide phosphate (Nosaka, 2006). At the same time, it can also activate defensive responses in plants as a non-cofactor (Bettendorff and Wins, 2013; Yusof et al., 2015). Recent work utilizing THI as growth regulators in strawberry transplants exposed to Cd stress has revealed a 1.7-fold increase in endogenous H₂S levels and displayed enhanced metal tolerance (Kaya and Aslan, 2020). They found that endogenous NO levels have also demonstrated the similar rise following THI treatment. The increase in leaf H₂S concentration caused the upregulation of MDA and H₂O₂, while antioxidant enzyme activities were downregulated to overcome Cd toxicity. Furthermore, endogenous H₂S improved mechanical stability and physiological functions of strawberry plants by markedly increasing the uptake of key nutrient elements, i.e., calcium and potassium. Although calcium (Ca) is typically an essential macronutrient required for normal plant growth, it can act as a universal messenger to establish systemic defense response and tolerance acquisition under stressful conditions (Jalmi et al., 2018). Several studies confirmed that exogenous Ca application can lead to higher stress tolerance in plants upon heavy metal exposure (Gonzalez et al., 2012; Ahmad et al., 2015; Aziz et al., 2015). According to Valivand et al. (2019), two-sided crosstalk between exogenous Ca and endogenous H₂S appears to mediate defense response in Zucchini plant exposed to Ni stress. The Ca signaling cascade from roots to leaves has been involved in endogenous H₂S synthesis that contributes toward a systemic acclimation against Ni stress. At the same time, antioxidant enzyme activities and genetic expression of calmodulin (CaM) protein further strengthen the metal ion tolerance of young seedlings, as demonstrated by reduced electrolyte leakage and oxidative injury. This study further supports the fact that both intracellular and extracellular Ca-based complexes are important for the synthesis of endogenous H₂S to improve plant metal tolerance. Ascorbate-glutathione metabolism involves antioxidant defense system that perceives stress and regulates plant growth by coordinating the activities of detoxification of ROS by its key enzymes-ascorbate peroxidase, monodehydroascorbate reductase, dehydroascorbate reductase, and glutathione reductase (Hasanuzzaman et al., 2017). In maize, Zanganeh et al. (2019) showed that endogenous H₂S positively regulated early growth of young seedlings via SA-mediated Pb detoxification. Plant exposure to metal ion

significantly deteriorated chlorophyll content and restricted nutrients uptake, whereas H₂S-SA crosstalk was instrumental in reversal of selected plant physiological and biochemical attributes, indicated by higher plant expression of ascorbic acid-glutathione metabolic activities. Methyl jasmonate (MeJA), a volatile derivative of jasmonic acid, serves as main cell signaling molecule mediating various key plant processes and also triggers plant defense in response to wide array of abiotic stresses (Raza et al., 2020). In foxtail millet, exogenous MeJA and NaHS showed an increase in endogenous H_2S (75–120 nm g FW^{-1}) and restore seedling growth under Cd stress (Tian et al., 2017). The results suggest that endogenous H₂S buildup was a pivotal component of ROS mitigation, thus hindering the accumulation of Cd in young seedlings. In addition, positive interplay between MeJA and H₂S augmented Cd-induced expression of the homeostasis-related genes (MTP1, MTP12, CAX2, and ZIP4).

CONCLUSION AND FUTURE PROSPECTIVE

In agriculture, abiotic stress involving heavy metals contributes to major production losses globally. It is broadly acknowledged that H_2S can mitigate the abiotic stresses including heavy metal stress. This review provides insight into beneficial aspects of H_2S on plant biochemical and physiological responses against heavy metal stress. After a thorough review of the available literature, we found that heavy metals inhibit plant growth, which unfortunately disturbs food production and eventually leads to food shortfall. Application of H_2S has shown effective mitigation of heavy metals related by strengthening the biochemical and physiological functions of plants. Treatment of H_2S devotedly leads to the enhancement in plant growth, photosynthetic pigments, biomass, nutrient uptake, gas exchange parameters, and antioxidant enzymes of plants.

Taking due account of the above findings and studies, it is strongly concluded that H₂S treatment effectively reduces the harmful effects of many heavy metals (Al, Cd, B, Pb, Cr, and Cu) by obstructing the accumulation of heavy metals in various plants. Such results strongly indicate that H₂S treatment could be used effectively as a signal molecule to inhibit the oxidative stress caused by heavy metal contamination. Recently, interaction of certain signal molecules and growth regulators with H₂S application has been investigated in order to regulate plant growth against heavy metal stress. Nonetheless, future work on other emerging signaling molecules and phytohormones is highly desirable, which can provide us a better understanding of these signaling molecules that affect the concentration of plant hormones and thus control the toxicity of metals in plants. Moreover, further investigations at genomics, transcriptomic, and metabolomics scale are required to explore the particular H_2S -generated tolerance mechanism in various plants against heavy metal stress. On the other hand, effective, widespread, and ongoing field trials using organic amendments and/or metal-tolerant microbial inoculant can also be tested for their protective role against the exposed contaminants, which can also upscale the effectiveness of H_2S in heavy metal-contaminated degraded land.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

REFERENCES

- Ahmad, I., Akhtar, M. J., Zahir, Z. A., and Jamil, A. (2012). Effect of cadmium on seed germination and seedling growth of four wheat (*Triticum aestivum* L.) cultivars. *Pak. J. Bot.* 5, 1569–1574.
- Ahmad, M. S., Ashraf, M., and Hussain, M. (2011). Phytotoxic effects of nickel on yield and concentration of macro- and micro-nutrients in sunflower (*Helianthus annuus* L.) achenes. *J. Hazard. Mater.* 185, 1295–1303. doi: 10. 1016/j.jhazmat.2010.10.045
- Ahmad, P., Sarwat, M., Bhat, N. A., Wani, M. R., Kazi, A. G., Tran, L. S., et al. (2015). Alleviation of cadmium toxicity in *Brassica juncea L.* by calcium application involves various physiological and biochemical strategies. *PLoS One* 10:e0114571. doi: 10.1371/journal.pone.0114571
- Ahmad, R., Ali, S., Rizwan, M., Dawood, M., Farid, M., Hussain, A., et al. (2020). Hydrogen sulfide alleviates chromium stress on cauliflower by restricting its uptake and enhancing antioxidative system. *Physiol. Plant.* 168, 289–300.
- Akaike, T., Ida, T., Wei, F.-Y., Nishida, M., Kumagai, Y., Alam, M. M., et al. (2017). Cysteinyl-tRNA synthetase governs cysteine polysulfidation and mitochondrial bioenergetics. *Nat. Commun.* 8:1177.
- Alamri, S., Ali, H. M., Khan, M. I. R., Singh, V. P., and Siddiqui, M. H. (2020). Exogenous nitric oxide requires endogenous hydrogen sulfide to induced the resilience through sulfur assimilation in tomato seedlings under hexavalent chromium toxicity. *Plant Physiol. Biochem.* 155, 20–34. doi: 10.1016/j.plaphy. 2020.07.003
- Alloway, B. J. (1995). *Heavy Metals in Soils*. Glasgow: Blackie Academic and Professional.
- Alloway, B. J. (2013). *Heavy Metals in Soils: Trace Metals and Metalloids in Soils and Their Bioavailability*, 3rd Edn. Glasgow: Blackie Academic and Professional.
- Amari, T., Ghnaya, T., and Abdelly, C. (2017). Nickel, cadmium and lead phytotoxicity and potential of halophytic plants in heavy metal extraction. *South Afr. J. Bot.* 111, 99–110. doi: 10.1016/j.sajb.2017.03.011
- Arif, M. S., Yasmeen, T., Shahzad, S. M., Riaz, M., Rizwan, M., Iqbal, S., et al. (2019). Lead toxicity induced phytotoxic effects on mung bean can be relegated by lead tolerant *Bacillus subtilis* (PbRB3). *Chemosphere* 234, 70–80. doi: 10.1016/j. chemosphere.2019.06.024
- Aroca, A., Gotor, C., Bassham, D. C., and Romero, L. C. (2020). Hydrogen sulfide: from a toxic molecule to a key molecule of cell life. *Antioxidants*. 9:621. doi: 10.3390/antiox9070621
- Aroca, A., Serna, A., Gotor, C., and Romero, L. C. (2015). S-sulfhydration: a cysteine posttranslational modification in plant systems. *Plant Physiol.* 168, 334–342. doi: 10.1104/pp.15.00009
- Azevedo, R. A., and Lea, P. J. (2005). Toxic metals in plants. Br. J. Plant Physiol. 17:1. doi: 10.1590/s1677-04202005000100001
- Aziz, H., Sabir, M., Ahmad, H. R., Aziz, T., Zia-ur-Rehman, M., Hakeem, K. R., et al. (2015). Alleviating effect of calcium on nickel toxicity in rice. *Clean Soil Air Water* 43, 787–866.
- Bai, X. Y., Dong, Y. J., Wang, Q. H., Xu, L. L., Kong, J., and Liu, S. (2014). Effects of lead and nitric oxide on photosynthesis, antioxidative ability, and mineral element content of perennial ryegrass. *Biol. Plant.* 59, 163–170. doi: 10.1007/s10535-014-0476-8

FUNDING

This research was funded by the Deanship of Scientific Research at Princess Nourah Bint Abdulrahman University through the Fast-track Research Funding Program.

ACKNOWLEDGMENTS

The authors highly acknowledge the Government College University, Faisalabad, Pakistan for its support. This work was financially supported by Fast-track Research Funding Program under the Deanship of Scientific Research at Princess Nourah Bint Abdulrahman University.

- Bettendorff, L., and Wins, P. (2013). Thiamine triphosphatase and the CYTH superfamily of proteins. *FEBS J.* 280, 6443–6455. doi: 10.1111/febs.12498
- Bharwana, S. A., Ali, S., Farooq, M. A., Ali, B., Iqbal, N., Abbas, F., et al. (2014). Hydrogen sulfide ameliorates lead-induced morphological, photosynthetic, oxidative damages and biochemical changes in cotton. *Environ. Sci. Pollut. Res.* 21, 717–731. doi: 10.1007/s11356-013-1920-6
- Bhuyan, M. H. M. B., Parvin, K., Mohsin, S. M., Mahmud, J. A., Hasanuzzaman, M., and Fujita, M. (2020). Modulation of cadmium tolerance in rice: insight into vanillic acid-induced upregulation of antioxidant defense and glyoxalase systems. *Plants* 9:188. doi: 10.3390/plants9020188
- Calderwood, A., and Kopriva, S. (2014). Hydrogen sulfide in plants: from dissipation of excess sulfur to signaling molecule. *Nitric Oxide* 41, 72–78. doi: 10.1016/j.niox.2014.02.005
- Carmo-Silva, E., Scales, J. C., Madgwick, P. J., and Parry, M. A. J. (2015). Optimizing Rubisco and its regulation for greater resource use efficiency. *Plant Cell Environ.* 38, 1817–1832. doi: 10.1111/pce.12425
- Chen, J., Wu, F. H., Wang, W. H., Zheng, C. J., Lin, G. H., Dong, X. J., et al. (2011). Hydrogen sulfide enhances photosynthesis through promoting chloroplast biogenesis, photosynthetic enzyme expression, and thiol redox modification in *Spinacia oleracea* seedlings. J. Exp. Bot. 62, 4481–4493. doi: 10.1093/jxb/err145
- Chen, T., Tian, M., and Han, Y. (2020). Hydrogen sulfide: a multi-tasking signal molecule in the regulation of oxidative stress responses. J. Exp. Bot. 71, 2862– 2869. doi: 10.1093/jxb/eraa093
- Chen, Z., Chen, M., and Jiang, M. (2016). Hydrogen sulfide alleviates mercury toxicity by sequestering it in roots or regulating reactive oxygen species productions in rice seedlings. *Plant Physiol. Biochem.* 111, 179–192. doi: 10. 1016/j.plaphy.2016.11.027
- Chen, Z., Yang, B., Hao, Z., Zhu, J., Zhang, Y., and Xu, T. (2018). Exogenous hydrogen sulfide ameliorates seed germination and seedling growth of cauliflower under lead stress and its antioxidant role. *J. Plant Growth Regul.* 37, 5–15. doi: 10.1007/s00344-017-9704-8
- Corpas, F. J., Barroso, J. B., Gonzalez-Gordo, S., Munoz-Vargas, M. A., and Palma, J. M. (2019a). Hydrogen sulfide: a novel component in *Arabidopsis* peroxisomes which triggers catalase inhibition. *J. Integr. Plant Biol.* 61, 871–883.
- Corpas, F. J., González-Gordo, S., Cañas, A., and Palma, J. M. (2019b). Nitric oxide and hydrogen sulfide in plants: which comes first? J. Exp. Bot. 70, 4391–4404. doi: 10.1093/jxb/erz031
- Cuevasanta, E., Moller, M. N., and Alvarez, B. (2017). Biological chemistry of hydrogen sulfide and persulfides. Arch. Biochem. Biophys. 617, 9–25. doi: 10. 1016/j.abb.2016.09.018
- Cummins, P. L., Kannappan, B., and Gready, J. E. (2018). Directions for optimization of photosynthetic carbon fixation: RuBisCO's efficiency may not be so constrained after all. *Front. Plant Sci.* 9:183.
- De Kok, L. J., Castro, A., Durenkamp, M., Stuiver, C. E. E., Westerman, S., Yang, L., et al. (2002). "Sulphur in plant physiology," in *Proceedings No 500, The International Fertiliser Society*, New York, NY. doi: 10.1016/j.plaphy.2016.12. 024
- DeLeon, E. R., Stoy, G. F., and Olson, K. R. (2012). Passive loss of hydrogen sulfide in biological experiments. *Anal. Biochem.* 421, 203–207. doi: 10.1016/j.ab.2011. 10.016

- Duffus, J. H. (2002). "Heavy metals" a meaningless term? (IUPAC Technical Report). Pure appl. Chem. 74, 793–807. doi: 10.1351/pac200274050793
- Fang, L., Ju, W., Yang, C., Jin, X., Liu, D., Li, M., et al. (2020). Exogenous application of signaling molecules to enhance the resistance of legume-rhizobium symbiosis in Pb/Cd-contaminated soils. *Environ. Pollut.* 265:114744. doi: 10.1016/j. envpol.2020.114744
- Filipovic, M. R., Zivanovic, J., Alvarez, B., and Banerjee, R. (2018). Chemical biology of H₂S signaling through persulfidation. *Chem. Rev.* 118, 1253–1337. doi: 10.1021/acs.chemrev.7b00205
- Gonzalez, A., Cabrera, M., de, L., Henriquez, M. J., Contreras, R. A., Morales, B., et al. (2012). Cross talk among calcium, hydrogen peroxide, and nitric oxide and activation of gene expression involving calmodulins and calcium-dependent protein kinases in *Ulva compressa* exposed to copper excess. *Plant Physiol*. 158, 1451–1462. doi: 10.1104/pp.111.191759
- Gotor, C., García, I., Aroca, Á, Laureano-Marín, A. M., Arenas-Alfonseca, L., Jurado-Flores, A., et al. (2019). Signaling by hydrogen sulfide and cyanide through post-translational modification. *J. Exp.Bot.* 70, 4251–4265. doi: 10. 1093/jxb/erz225
- Grant, R., and Grant, C. (1987). Grant and Hackh's Chemical Dictionary. New York, NY: McGraw-Hill.
- Han, Y., Yin, Y., and Yi, H. (2020). Decreased endogenous nitric oxide contributes to sulfur dioxide derivative alleviated cadmium toxicity in *Foxtail millet* roots. *Environ. Exp. Bot.* 177:104144. doi: 10.1016/j.envexpbot.2020.104144
- Hancock, J. T. (2017). Harnessing evolutionary toxins for signaling: reactive oxygen species, nitric oxide and hydrogen sulfide in plant cell regulation. *Front. Plant. Sci.* 8:189.
- Hancock, J. T., and Whiteman, M. (2016). Hydrogen sulfide signaling: interactions with nitric oxide and reactive oxygen species. Ann. N. Y. Acad. Sci. 1365, 5–14. doi: 10.1111/nyas.12733
- Hasanuzzaman, M., Matin, M. A., Fardus, J., Hasanuzzaman, M., Hossain, M. S., and Parvin, K. (2019). Foliar application of salicylic acid improves growth and yield attributes by upregulating the antioxidant defense system in Brassica campestris plants grown in lead-amended soils. *Acta Agro. Bot.* 72:1765.
- Hasanuzzaman, M., Nahar, K., Anee, T. I., and Fujita, M. (2017). Exogenous silicon attenuates cadmium-induced oxidative stress in *Brassica napus* L. by modulating AsA-GSH pathway and glyoxalase system. *Front. Plant Sci.* 8:1061.
- He, H., Li, Y., and He, L. F. (2019). Role of nitric oxide and hydrogen sulfide in plant aluminum tolerance. *Biometals* 32, 1–9. doi: 10.1007/s10534-018-0156-9
- Hoque, T. S., Hossain, M. A., Mostofa, M. G., Burritt, D. J., Fujita, M., and Tran, L. S. P. (2016). Methylglyoxal: an emerging signaling molecule in plant abiotic stress responses and tolerance. *Front. Plant Sci.* 7:1341.
- Hu, Y., Lu, L., Tian, S., Li, S., Liu, X., Gao, X., et al. (2019). Cadmium-induced nitric oxide burst enhances Cd tolerance at early stage in roots of a hyperaccumulator *Sedum alfredii* partially by altering glutathione metabolism. *Sci. Total Environ.* 650, 2761–2770. doi: 10.1016/j.scitotenv.2018.09.269
- Huo, J., Huang, D., Zhang, J., Fang, H., Wang, B., Wang, C., et al. (2018). Hydrogen sulfide: a gaseous molecule in postharvest freshness. *Front. Plant Sci.* 27:1172.
- Ida, T., Sawa, T., Ihara, H., Tsuchiya, Y., Watanabe, Y., Kumagai, Y., et al. (2014). Reactive cysteine persulfides and S-polythiolation regulate oxidative stress and redox signaling. *Proc. Natl. Acad. Sci. U.S.A.* 111, 7606–7611. doi: 10.1073/pnas. 1321232111
- Jalmi, S. K., Bhagat, P. K., Verma, D., Noryang, S., Tayyeba, S., Singh, K., et al. (2018). Traversing the links between heavy metal stress and plant signaling. *Front. Plant Sci.* 9:12.
- Jia, H., Wang, X., Dou, Y., Liu, D., Si, W., Fang, H., et al. (2016). Hydrogen sulfide - cysteine cycle system enhances cadmium tolerance through alleviating cadmium-induced oxidative stress and ion toxicity in *Arabidopsis* roots. *Sci. Rep.* 6:39702.
- Jia, H., Wang, X., Shi, C., Guo, J., Ma, P., Wei, T., et al. (2020). Hydrogen sulfide decreases Cd translocation from root to shoot through increasing Cd accumulation in cell wall and decreasing Cd²⁺ influx in *Isatis indigotica*. *Plant Physiol. Biochem.* 155, 605–612. doi: 10.1016/j.plaphy.2020.08.033
- Kabil, O., and Banerjee, R. (2010). Redox biochemistry of hydrogen sulfide. J. Biol. Chem. 285, 21903–21907. doi: 10.1074/jbc.r110.12 8363
- Karam, E. A., and Keramat, B. (2017). Hydrogen sulfide protects coriander seedlings against copper stress by regulating the ascorbate-glutathione cycle in leaves. J. Plant Process Funct. 5, 59–64.

- Kasamatsu, S., Nishimura, A., Morita, M., Matsunaga, T., Abdul Hamid, H., and Akaike, T. (2016). Redox signaling regulated by cysteine persulfide and protein polysulfidation. *Molecules* 21:E1721.
- Kaya, C. (2020). Salicylic acid-induced hydrogen sulphide improves lead stress tolerance in pepper plants by upraising the ascorbate-glutathione cycle. *Physiol. Plant* [Epub ahead of print]. doi: 10.1111/ppl.13159
- Kaya, C., Akram, N. A., Ashraf, M., Alyemeni, M. N., and Ahmad, P. (2020). Exogenously supplied silicon (Si) improves cadmium tolerance in pepper (*Capsicum annuum* L.) by up-regulating the synthesis of nitric oxide and hydrogen sulfide. J. Biotechnol. 316, 35–45. doi: 10.1016/j.jbiotec.2020.04.008
- Kaya, C., Ashraf, M., and Akram, N. A. (2018). Hydrogen sulfide regulates the levels of key metabolites and antioxidant defense system to counteract oxidative stress in pepper (*Capsicum annuum* L.) plants exposed to high zinc regime. *Environ. Sci. Pollut. Res.* 25, 12612–12618. doi: 10.1007/s11356-018-1510-8
- Kaya, C., and Aslan, M. (2020). Hydrogen sulphide partly involves in thiamineinduced tolerance to cadmium toxicity in strawberry (Fragaria x ananassa Duch) plants. *Environ. Sci. Pollut. Res.* 27, 941–953. doi: 10.1007/s11356-019-07056-z
- Kharbech, O., Massoud, M. B., Sakouhi, L., Djebali, W., Mur, L. A. J., and Chaoui, A. (2020). Exogenous application of hydrogen sulfide reduces chromium toxicity in maize seedlings by suppressing NADPH oxidase activities and methylglyoxal accumulation. *Plant Physiol. Biochem.* 154, 646–656. doi: 10. 1016/j.plaphy.2020.06.002
- Kimura, Y., Koike, S., Shibuya, N., Lefer, D., Ogasawara, Y., and Kimura, H. (2017). 3-Mercaptopyruvate sulfurtransferase produces potential redox regulators cysteine- and glutathione-persulfide (Cys-SSH and GSSH) together with signaling molecules H₂S₂, H₂S₃ and H₂S. *Sci. Rep.* 7:10459.
- Krueger, S., Niehl, A., Martin, M. C., Steinhauser, D., Donath, A., Hildebrandt, T., et al. (2009). Analysis of cytosolic and plastidic serine acetyltransferase mutants and subcellular metabolite distributions suggests interplay of the cellular compartments for cysteine biosynthesis in *Arabidopsis. Plant Cell Environ.* 32, 349–367. doi: 10.1111/j.1365-3040.2009.01928.x
- Krzesłowska, M. (2011). The cell wall in plant cell response to trace metals: polysaccharide remodeling and its role in defense strategy. *Acta Physiol. Plant.* 33, 35–51. doi: 10.1007/s11738-010-0581-z
- Kushwaha, B. K., and Singh, V. P. (2019). Glutathione and hydrogen sulfide are required for sulfur-mediated mitigation of Cr (VI) toxicity in tomato, pea and brinjal seedlings. *Physiol. Plant* 168, 406–421.
- Lado, L. R., Hengl, T., and Reuter, H. I. (2008). Heavy metals in European soils: a geostatistical analysis of the FOREGS geochemical database. *Geoderma* 148, 189–199. doi: 10.1016/j.geoderma.2008.09.020
- Lai, H. Y. (2015). Subcellular distribution and chemical forms of cadmium in *Impatiens walleriana* in relation to its phytoextraction potential. *Chemosphere* 138, 370–376. doi: 10.1016/j.chemosphere.2015.06.047
- Lasat, M. M. (2002). Phytoextraction of toxic metals A review of biological mechanism. J Environ. Qual. 31, 109–120. doi: 10.2134/jeq2002.1090
- Li, L., Wang, Y., and Shen, W. (2012). Roles of hydrogen sulphide and nitric oxide in the alleviation of cadmium-induced oxidative damage in alfalfa seedling roots. *Biometals* 25, 617–631. doi: 10.1007/s10534-012-9551-9
- Li, Z. G. (2013). Hydrogen sulfide: a multifunctional gaseous molecule in plants. Russ. J. Plant Physiol. 60, 733–740. doi: 10.1134/s1021443713060058
- Li, Z. G., Min, X., and Zhou, Z. H. (2016). Hydrogen sulfide: a signal molecule in plant cross-adaptation. *Front. Plant Sci.* 7:1621.
- Lin, Y. T., Li, M. Y., Cui, W. T., Lu, W., and Shen, W. B. (2012). Haem oxygenase-1 is involved in hydrogen sulfide-induced cucumber adventitious root formation. *J. Plant Growth Regul.* 3, 519–528. doi: 10.1007/s00344-012-9262-z
- Lisjak, M., Srivastava, N., Teklic, T., Civale, L., Lewandowski, K., Wilson, I., et al. (2010). A novel hydrogen sulfide donor causes stomatal opening and reduces nitric oxide accumulation. *Plant Physiol. Biochem.* 48, 931–935. doi: 10.1016/j. plaphy.2010.09.016
- Liu, T. J., Liu, X. N., Liu, M. L., and Wu, L. (2018). Evaluating heavy metal stress levels in rice based on remote sensing phenology. *Sensors* 18:860. doi: 10.3390/s18030860
- Liu, X., Chen, J., Wang, G.-H., Wang, W.-H., Shen, Z.-J., Luo, M.-R., et al. (2016). Hydrogen sulfide alleviates zinc toxicity by reducing zinc uptake and regulating genes expression of antioxidative enzymes and metallothioneins in roots of the cadmium/zinc hyperaccumulator *Solanum nigrum* L. *Plant Soil* 400, 177–192. doi: 10.1007/s11104-015-2719-7

- Luo, S., Calderón-Urrea, A., Yu, J., Liao, W., Xie, J., Lv, J., et al. (2020). The role of hydrogen sulfide in plant alleviates heavy metal stress. *Plant Soil* 449, 1–10. doi: 10.1007/s11104-020-04471-x
- Mancardi, D., Penna, C., Merlino, A., Del Soldato, P., Wink, D. A., and Pagliaro, P. (2009). Physiological and pharmacological features of the novel gasotransmitter: hydrogen sulfide. *BBA Bioenerget*. 1787, 864–872. doi: 10. 1016/j.bbabio.2009.03.005
- Mataka, L. M., Henry, E. M. T., Masamba, W. R. L., and Sajidu, S. M. (2006). Lead remediation of contaminated water using *Moringa stenopetala* sp., and *Moringa oleifera* sp., seed powder. *Inter. J. Environ. Sci. Tech.* 2, 131–139. doi: 10.1007/bf03325916
- Mostofa, M. G., Rahman, A., Ansary, M. M. U., Watanabe, A., Fujita, M., and Tran, L. S. P. (2015). Hydrogen sulfide modulates cadmium-induced physiological and biochemical responses to alleviate cadmium toxicity in rice. *Sci. Rep.* 5:14078.
- Mustafa, A. K., Gadalla, M. M., Sen, N., Kim, S., Mu, W., Gazi, S. K., et al. (2009). H₂S signals through protein S-sulfhydration. *Sci. Signal.* 2:ra72. doi: 10.1126/scisignal.2000464
- Nosaka, K. (2006). Recent progress in understanding thiamin biosynthesis and its genetic regulation in *Saccharomyces cerevisiae*. *Appl. Microbiol. Biotechnol.* 72, 30–40. doi: 10.1007/s00253-006-0464-9
- Pandey, A. K., and Gautam, A. (2019). Stress responsive gene regulation in relation to hydrogen sulfide in plants under abiotic stress. *Physiol. Plant* 168, 511–525.
- Paul, B. D., and Snyder, S. H. (2012). H2S signalling through protein sulfhydration and beyond. Nat. Rev. Mol. Cell Biol. 13, 499–507. doi: 10.1038/nrm3391
- Peng, B., and Xian, M. (2015). Hydrogen sulfide detection using nucleophilic substitution-cyclization-based fluorescent probes. *Methods Enzymol.* 554, 47– 62. doi: 10.1016/bs.mie.2014.11.030
- Rather, B. A., Mir, I. R., Sehar, Z., Anjum, N. A., Masood, A., and Khan, N. A. (2020). The outcomes of the functional interplay of nitric oxide and hydrogen sulfide in metal stress tolerance in plants. *Plant Physiol. Biochem.* 155, 523–534. doi: 10.1016/j.plaphy.2020.08.005
- Rausch, T., and Wachter, A. (2005). Sulfur metabolism: a versatile platform for launching defence operations. *Trends Plant Sci.* 10, 503–509. doi: 10.1016/j. tplants.2005.08.006
- Raza, A., Charagh, S., Zahid, Z., Mubarik, M. S., Javed, R., Siddiqui, M. H., et al. (2020). Jasmonic acid: a key frontier in conferring a biotic stress tolerance in plants. *Plant Cell Rep.* [Epub ahead of print]. doi: 10.1007/s00299-020-02614-z
- Reichman, S. M. (2007). The potential use of the legume-rhizobium symbiosis for the remediation of arsenic contaminated sites. *Soil Biol. Biochem.* 39, 2587–2593. doi: 10.1016/j.soilbio.2007.04.030
- Rennenberg, H. (1983). Role of O-acetylserine in hydrogen sulfide emission from pumpkin leaves in response to sulfate. *Plant Physiol.* 73, 560–565. doi: 10.1104/ pp.73.3.560
- Rennenberg, H. (1984). The fate excess of sulfur in higher plants. *Annu. Rev. Plant Physiol.* 35, 121–153. doi: 10.1146/annurev.pp.35.060184.001005
- Rennenberg, H., Huber, B., Schroder, P., Stahl, K., Haunold, W., Georgii, H. W., et al. (1990). Emission of volatile sulfur compounds from spruce trees. *Plant Physiol.* 92, 560–564. doi: 10.1104/pp.92.3.560
- Riemenschneider, A., Nikiforova, V., Hoefgen, R., De Kok, L. J., and Papenbrock, J. (2005a). Impact of elevated H₂S on metabolite levels, activity of enzymes and expression of genes involved in cysteine metabolism. *Plant Physiol. Biochem.* 43, 473–448. doi: 10.1016/j.plaphy.2005.04.001
- Riemenschneider, A., Riedel, K., Hoefgen, R., Papenbrok, J., and Hesse, H. (2005b). Impact of reduced O-acetylserine (thiol) lyase iso-form contents on potato plant metabolism. *Plant Physiol.* 137, 892–900. doi: 10.1104/pp.104.057125
- Rizwan, M., Mostofa, M. G., Ahmad, M. Z., Imtiaz, M., Mehmood, S., Adeel, M., et al. (2018). Nitric oxide induces rice tolerance to excessive nickel by regulating nickel uptake, reactive oxygen species detoxification and defense- related gene expression. *Chemosphere* 191, 23–35. doi: 10.1016/j.chemosphere.2017.09.068
- Rizwan, M., Mostofa, M. G., Ahmad, M. Z., Zhou, Y., Adeel, M., Mehmood, S., et al. (2019). Hydrogen sulfide enhances rice tolerance to nickel through the prevention of chloroplast damage and the improvement of nitrogen metabolism under excessive nickel. *Plant Physiol. Biochem.* 138, 100–111. doi: 10.1016/j. plaphy.2019.02.023
- Rubright, S. L. M., Pearce, L. L., and Peterson, J. (2017). Environmental toxicology of hydrogen sulfide Nitric Oxide. *Biol. Chem.* 71:1. doi: 10.1016/j.niox.2017.09. 011

- Schutzendubel, A., and Polle, A. (2002). Plant responses to abiotic stresses: heavy metal-induced oxidative stress and protection by mycorrhization. J. Exp. Bot. 53, 1351–1365. doi: 10.1093/jexbot/53.372.1351
- Sekiya, J., Schmidt, A., Wilson, L. G., and Filner, P. (1982a). Emission of hydrogen sulfide by leaf tissue in response to L-cysteine. *Plant Physiol.* 70, 430–436. doi: 10.1104/pp.70.2.430
- Sekiya, J., Wilson, L. G., and Filner, P. (1982b). Resistance to injury by sulfur dioxide: correlation with its reduction to, and emission of, hydrogen sulfide in Cucurbitaceae. *Plant Physiol.* 70, 437–441. doi: 10.1104/pp.70.2.437
- Shan, C., Dai, H., and Sun, Y. (2012). Hydrogen sulfide protects wheat seedlings against copper stress by regulating the ascorbate and glutathione metabolism in leaves. *Aust. J. Crop Sci.* 6, 248–254.
- Shan, C. J., Zhang, S. L., Li, D. F., Zhao, Y. Z., Tian, X. L., Zhao, X. L., et al. (2011). Effects of exogenous hydrogen sulfide on the ascorbate and glutathione metabolism in wheat seedlings leaves under water stress. *Acta. Physiol. Plant.* 33, 2533–2540. doi: 10.1007/s11738-011-0746-4
- Sharma, A., and Dhiman, A. (2013). Nickel and cadmium toxicity in plants. J. Pharm. Sci. Innov. 2, 20–24. doi: 10.7897/2277-4572.02213
- Sharma, A., Sidhu, G. P. S., Araniti, F., Bali, A. S., Shahzad, B., Tripathi, D. K., et al. (2020b). The role of salicylic acid in plants exposed to heavy metals. *Molecules* 25:540. doi: 10.3390/molecules25030540
- Sharma, A., Soares, C., Sousa, B., Martins, M., Kumar, V., Shahzad, B., et al. (2020a). Nitric oxide-mediated regulation of oxidative stress in plants under metal stress: a review on molecular and biochemical aspects. *Physiol. Planta.* 168, 318–344. doi: 10.1111/ppl.13004
- Shen, G. T., Ju, W. L., Liu, Y. Q., Guo, X. B., Zhao, W., and Fang, L. C. (2019). Impact of urea addition and rhizobium inoculation on plant resistance in metal contaminated soil. *Int. J. Environ. Res. Publ. Health* 16:1955. doi: 10.3390/ ijerph16111955
- Shivaraj, S. M., Vats, S., Bhat, J. A., Dhakte, P., Goyal, V., Khatri, P., et al. (2020). Nitric oxide and hydrogen sulfide crosstalk during heavy metal stress in plants. *Physiol. Planta* 168, 437–455.
- Singh, V. P., Singh, S., Kumar, J., and Prasad, S. M. (2015). Hydrogen sulfide alleviates toxic effects of arsenate in pea seedlings through up-regulation of the ascorbate-glutathione cycle: possible involvement of nitric oxide. *J. Plant Physiol.* 181, 20–29. doi: 10.1016/j.jplph.2015.03.015
- Takahashi, H., Kopriva, S., Giordano, M., Saito, K., and Hell, R. (2011). Sulfur assimilation in photosynthetic organisms: molecular functions and regulations of transporters and assimilatory enzymes. *Annu. Rev. Plant Biol.* 62, 157–184. doi: 10.1146/annurev-arplant-042110-103921
- Thompson, C. R., and Kats, G. (1978). Effects of continuous hydrogen sulphide fumigation on crop and forest plants. *Environ. Sci. Technol.* 12, 550–553. doi: 10.1021/es60141a001
- Tian, B., Qiao, Z., Zhang, L., Li, H., and Pei, Y. (2016). Hydrogen sulfide and proline coperate to alleviate cadmium stress in foxtail millet seedlings. *Plant Physiol. Biochem.* 109, 293–299. doi: 10.1016/j.plaphy.2016.10.006
- Tian, B., Zhang, Y., Jin, Z., Liu, Z., and Pei, Y. (2017). Role of hydrogen sulfide in the methyl jasmonate response to cadmium stress in *Foxtail millet. Front. Biosci. Landmark* 225:30–538.
- Toohey, J. I. (2011). Sulfur signaling: is the agent sulfide or sulfane? *Anal. Biochem.* 413, 1–7. doi: 10.1016/j.ab.2011.01.044
- Vaculík, M., Lukačová, Z., Bokor, B., Martinka, M., Tripathi, D. K., and Lux, A. (2020). Alleviation mechanisms of metal(loid) stress in plants by silicon: a review. J. Exp. Bot. 71, 6744–6757. doi: 10.1093/jxb/eraa288
- Valivand, M., and Amooaghaie, R. (2020). Sodium hydrosulfide modulates membrane integrity, cation homeostasis, and accumulation of phenolics and osmolytes in Zucchini under nickel stress. *J. Plant Growth Regul.* [Epub ahead of print]. doi: 10.1007/s00344-020-10101-8
- Valivand, M., Amooaghaie, R., and Ahadi, A. M. (2019). Interplay between hydrogen sulfide and calcium/calmodulin enhances systemic acquired acclimation and antioxidative defense against nickel toxicity in zucchini. *Environ. Exp. Bot.* 158, 40–50. doi: 10.1016/j.envexpbot.2018.11.006
- Wang, H., Ji, F., Zhang, Y., Hou, J., Liu, W., Huang, J., et al. (2019). Interactions between hydrogen sulphide and nitric oxide regulate two soybean citrate transporters during the alleviation of aluminium toxicity. *Plant Cell Environ*. 42, 2340–2356. doi: 10.1111/pce.13555
- Wang, K., Peng, H., and Wang, B. (2014). Recent advances in thiol and sulfide reactive probes. J. Cell Biochem. 115, 1007–1022. doi: 10.1002/jcb.24762

- Wang, S., Chi, Q., Hu, X., Cong, Y., and Li, S. (2019). Hydrogen sulfide-induced oxidative stress leads to excessive mitochondrial fission to activate apoptosis in broiler myocardia. *Ecotoxicol. Environ. Saf.* 183, 1–9.
- Wang, Y. Q., Li, L., Cui, W. T., Xu, S., Shen, W. B., and Wang, R. (2012). Hydrogen sulfide enhances alfalfa (*Medicago sativa*) tolerance against salinity during seed germination by nitric oxide pathway. *Plant Soil* 351, 107–119. doi: 10.1007/ s11104-011-0936-2
- Wilson, L. G., Bressan, R. A., and Filner, P. (1978). Light-dependent emission of hydrogen sulfide from plants. *Plant Physiol.* 61, 184–189. doi: 10.1104/pp.61.2. 184
- Xuan, L., Li, J., Wang, X., and Wang, C. (2020). Crosstalk between hydrogen sulfide and other signal molecules regulates plant growth and development. *Int. J. Mol. Sci.* 21:4593. doi: 10.3390/ijms21134593
- Yamasaki, H., and Cohen, M. F. (2016). Biological consilience of hydrogen sulfide and nitric oxide in plants: gases of primordial earth linking plant, microbial and animal physiologies. *Nitric Oxide* 55, 91–100. doi: 10.1016/j.niox.2016.04.002
- Yang, G., Wu, L., Jiang, B., Yang, W., Qi, J., Cao, K., et al. (2008). H₂S as a physiologic vasorelaxant, hypertension in mice with deletion of cystathionine γ-lyase. *Science* 322, 587–590. doi: 10.1126/science.1162667
- Yu, X. Z., Ling, Q. L., Li, Y. H., and Lin, Y. J. (2018). mRNA analysis of genes encoded with *Phytochelatin Synthase* (PCS) in rice seedlings exposed to chromium: the role of phytochelatins in Cr detoxification. *Bull. Environ. Contam. Toxicol.* 101, 257–261. doi: 10.1007/s00128-018-2362-0
- Yu, Y., Zhou, X. Y., Zhu, Z. H., and Zhou, K. J. (2019). Sodium hydrosulfide mitigates cadmium toxicity by promoting cadmium retention and inhibiting its translocation from roots to shoots in *Brassica napus. J. Agric. Food Chem.* 67, 433–440. doi: 10.1021/acs.jafc.8b04622
- Yusof, Z. N. B., Borhan, F. P., Mohamad, F. A., and Rusli, M. H. (2015). The effect of *Ganoderma boninense* infection on the expressions of thiamine (vitamin B1) biosynthesis genes in oil palm. J. Oil Palm Res. 27, 12–18.
- Zanganeh, R., Jamei, R., and Rahmani, F. (2018). Impacts of seed priming with salicylic acid and sodium hydrosulfide on possible metabolic pathway of two amino acids in maize plant under lead stress. *Mol. Biol. Res. Comm.* 7, 83–87.
- Zanganeh, R., Jamei, R., and Rahmani, F. (2019). Role of salicylic acid and hydrogen sulfide in promoting lead stress tolerance and regulating free amino acid composition in *Zea mays L. Acta Physiol. Plant.* 41:94.

- Zhang, H., Hua, S. L., Zhang, Z. J., Hua, L. Y., Jiang, C. X., Wei, Z. J., et al. (2011). Hydrogen sulfide acts as a regulator of flower senescence in plants. *Postharvest Biol. Technol.* 60, 251–257. doi: 10.1016/j.postharvbio.2011. 01.006
- Zhang, H., Hu, L. Y., Hu, K. D., Jiang, C. X., and Luo, J. P. (2010a). Hydrogen sulfide alleviated chromium toxicity in wheat. *Biol. Plant.* 54, 743–747. doi: 10.1007/s10535-010-0133-9
- Zhang, H., Tan, Z. Q., Hu, L. Y., Wang, S. H., Luo, J. P., and Jones, R. L. (2010b). Hydrogen sulfide alleviates aluminum toxicity in germinating wheat seedlings. *J. Integr. Plant Biol.* 52, 556–567. doi: 10.1111/j.1744-7909.2010.00946.x
- Zhang, H., Tang, J., Liu, X. P., Wang, Y., Yu, W., Peng, W. Y., et al. (2009). Hydrogen sulfide promotes root organogenesis in Ipomoea batatas, Salix matsudana and Glycine max. J. Integr. Plant Biol. 51, 1084–1092.
- Zheng, S., Wang, Q., Yuan, Y., and Sun, W. (2020). Human health risk assessment of heavy metals in soil and food crops in the Pearl River Delta urban agglomeration of China. *Food Chem.* 316:126213. doi: 10.1016/j.foodchem. 2020.126213
- Zhi, J., Liu, X., Yin, P., Yang, R., Liu, J., and Xu, J. (2020). Overexpression of the metallothionein gene PaMT3-1 from *Phytolacca americana* enhances plant tolerance to cadmium. *Plant Cell Tissue Organ. Cult.* 143, 211–218. doi: 10. 1007/s11240-020-01914-2
- Zulfiqar, F., and Hancock, J. T. (2020). Hydrogen sulfide in horticulture: emerging roles in the era of climate change. *Plant Physiol. Biochem.* 155, 667–675. doi: 10.1016/j.plaphy.2020.08.010

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Arif, Yasmeen, Abbas, Ali, Rizwan, Aljarba, Alkahtani and Abdel-Daim. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.