



Effects of Antimony Stress on Photosynthesis and Growth of *Acorus calamus*

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This study was aimed to explore that effects of Sb on physiological parameters of *Acorus calamus* and the possibility of using *A. calamus* as a remediation plant. *A. calamus* potted experiments were conducted using different concentrations (0, 250, 500, 1000, and 2000 mg/kg) of antimony potassium tartrate (Sb^{3+}) (marked as CK, T₁, T₂, T₃, and T₄, respectively) and potassium pyroantimonate (Sb^{5+}) (marked as CK, T'₁, T'₂, T'₃, and T'₄, respectively). The effects of Sb stress (Sb^{3+} and Sb^{5+}) on leaf photosynthetic pigments, biomass, photosynthetic characteristics and chlorophyll fluorescence parameters of potted *A. calamus* were studied. With the rise of Sb^{3+} concentration from T₁ to T₄, the leaf pigment contents (chlorophyll a, b, carotenoid), plant height, dry weight, net photosynthetic rate (Pn), stomatal conductance (Gs), evaporation rate (E), PSII maximum photochemical efficiency (Fv/Fm), and PSII electron transfer quantum yield rate (ΦPSII) of *A. calamus* all reduced, while intercellular CO₂ concentration (Ci) significantly increased. The reduction of Pn was mainly induced by non-stomatal limitation. Chlorophyll a/b ratio increased significantly versus the control, while carotenoid/chlorophyll ratio (Car/Chl) first decreased and then increased. The leaf Chl a, Chl b, Car, plant height, dry weight, Pn, Gs, E, Fv/Fm, and ΦPSII all maximized in T'₁ (250 mg/kg), but were not significantly different from the control. As the Sb^{5+} concentration increased from T'₂ to T'₄, the above indices all decreased and were significantly different from the control. Moreover, intercellular CO₂ concentration (Ci) decreased significantly. The reduction of Pn was caused by non-stomatal limitation, indicating the mesophyll cells were damaged. The Car/Chl ratio was stable within 0–500 mg/kg Sb, but decreased in T₃ and T₄, and rose in T'₃ and T'₄. After Sb^{3+} and Sb^{5+} treatments, translocation factor varied 19.44–27.8 and 19.44–24.86%, respectively. In conclusion, different from Sb^{3+} treatment, Sb^{5+} treatment showed a Hormesi effect, as low-concentration treatment promoted *A. calamus* growth, but high-concentration treatment inhibited its growth. The two forms of Sb both caused unfavorable effects on *A. calamus*, but the seedlings did not die and were modestly adaptive and Sb-accumulative. *A. calamus*, which is easily maintained and cultivated, can serve as a good candidate for phytoremediation of water contaminated with Sb.

Keywords: antimony, *Acorus calamus*, photosynthetic pigment, photosynthesis, biomass, phytoremediation

INTRODUCTION

Antimony (Sb) is a ubiquitous trace element in the environment and a global pollutant and has been included as a priority pollutant by United States Environmental Protection Agency owing to its latent toxicity and carcinogenicity (Liu et al., 2013; Corrales et al., 2014). Some pollutants of heavy metal come from a variety of natural and man-made sources (Sytar et al., 2016). However, extensive use of Sb compounds has discharged numerous Sb into waters, soils and air, causing severe Sb pollution. For instance, the soil Sb concentrations in a Sb mining area of Italy were 19–4400 mg·kg⁻¹, and the root Sb concentrations of *Pistacia lentiscus* were 0.46–22.3 mg/kg (Cidu et al., 2014). The soil and plant Sb concentrations in a tin deposit mountain of Hunan, China, were 10–2159 and 143.7 mg·kg⁻¹, respectively (Wang et al., 2010; He et al., 2012), which interfered with plant growth and human health (Liu et al., 2010; Fu et al., 2011; Li et al., 2017). Thus, Sb pollution cannot be ignored.

Sb mainly exists as Sb⁺³ and Sb⁺⁵ in the environment, and its toxicity is related with the concentration and existing form (Huang et al., 2012). Generally, Sb⁺³ is more toxic than Sb⁺⁵. Sb is not a necessary element for plant growth (Filella et al., 2002, 2009), but can yet be absorbed by plants. The Sb absorbing ability differs among plant species (Shtangeeva et al., 2012). The Sb compounds affect the electron transfer, photosystem II carbonic anhydrase activity and the chloroplast glutathione reductase during photosynthesis (Karacan et al., 2016).

Sb existing as solutions in soils can be easily absorbed by plants, which would induce toxic effects to plants, including growth retardation, photosynthesis repression, and reduced synthesis of some metabolites (Feng et al., 2013). As reported, transport proteins are critical in the Sb absorption by plants. Sb⁺³ is absorbed by plants through transport proteins (NIP1 group) (Kamiya and Fujiwara, 2009). Under Sb⁺⁵ stress, the biomass of maize was significantly reduced (Cai et al., 2016). Ji et al. (2017) used a synchrotron of X-ray absorption near-edge structure to analyze the forms of Sb in ryegrass *in vivo*. In a soil-plant system, *Arbuscular mycorrhizal fungi* can relieve the Sb toxicity to plants (Wei et al., 2016; Pierart et al., 2017). Owing to the similar chemical properties between Sb and As, the Sb absorption mechanisms by plants can be deduced from those of As. However, the response mechanism to Sb stress is still unclear.

Acorus calamus is an adaptive and wet/drought-resistant tall perennial herb capable of purifying wastewater and relieving water eutrophication. *A. calamus* has been widely used in urban ecological park construction, and has been adopted in recent 20 years to absorb N, P and heavy metals from polluted waters, which shows its high eco-environmental values.

The action of Sb on plants would induce complex effects. Sb at low concentration could modestly promote the growth of some plant species, and its toxicity only acts at high concentrations. However, the mechanisms how *A. calamus* responds to different forms of Sb are yet unclear. This study was aimed to explore (i) the effects of Sb on physiological parameters of *A. calamus*, including photosynthesis efficiency, intercellular CO₂ concentration, and photosystem action sites; (ii) the poisoning mechanism of Sb to *A. calamus*, especially the photosynthesis

system; (iii) the possibility of using *A. calamus* to repair heavy Sb pollution areas. This study will scientifically underlie the introduction and application of Sb-resistant plants, and offer theoretical references for further exploring the ecological values of *A. calamus*.

MATERIALS AND METHODS

Materials

The study site was located in a greenhouse of Huaibei Normal University, Anhui province, China. The indoor temperature (18–28°C) was favorable for spring sowing and the growth of winter crops. *A. calamus* seedlings were purchased from a commercial supplier. *L-Antimony Potassium Tartrate* (III) (C₈H₄K₂O₁₂Sb₂) and *Potassium acid pyroantimonate* (V) (K₂H₂Sb₂O₇·4H₂O) were both analytically pure (Shanghai Sinopharm Group Co., Ltd.). *A. calamus* was planted in 30 round pots (upper and lower inside diameters = 20 and 18 cm, respectively; height = 16 cm).

Methods

Each pot was each filled with 10 kg of air-dried soil (3-mm sieve), which was mixed evenly. Then the pots were added with different concentrations (0, 250, 500, 1000, and 2000 mg/kg) of C₈H₄K₂O₁₂Sb₂ (Sb⁺⁵, marked as CK, T₁, T₂, T₃, and T₄, respectively) or K₂H₂Sb₂O₇·4H₂O (Sb⁺³, marked as CK, T'₁, T'₂, T'₃, and T'₄, respectively). Totally 10 treatments were set. All experiments were performed in triplicate. The growing plants were irrigated with distilled water if necessary. After 60 days, samples were collected, crushed and sent for measurement of Sb concentrations.

Data Measurement and Calculation

The net photosynthetic rate (P_n), stomatal conductance (G_s), evaporation rate (E), and intercellular CO₂ concentration (C_i) per unit leaf area in *calamus* leaves under different treatments were measured on a portable Licor-6400XT photosynthesis tester (United States). Photosynthesis was tested in open air and at light intensity 800 mol·m⁻²·s⁻¹, CO₂ concentration = 360 μmol·mol⁻¹ on sunny days during 9:00–11:30 am. After 30 min of dark adaptation, a saturation pulse induces maximal fluorescence yield(Y), and Y is given by a portable adjustable chlorophyll fluorescence device (Mini-Pam, Germany). Fluorescence induction curves were detected using the built-in automatic light source. The fiber optics are held at short distance (ca. 10 mm) to a leaf of *A. calamus*, and the START-key is pressed. The real quantum yield of PSII (ΦPSII) is proceeding automatically within seconds.

The leaves with the same positions and maturity degrees between photosynthesis and chlorophyll fluorescence were collected and their chlorophyll concentrations were detected on a UV-2550 ultraviolet-visible (UV-vis) spectrophotometer (Shimadzu, Japan) in the laboratory. Each experiment involved three to five leaves, and each leaf was measured three times. The optical densities (ODs) at 663, 645, and 470 nm were measured. Chlorophyll concentrations were expressed as the mean in the unit of mg·g⁻¹·FW. Before measurement of biomass

and Sb concentrations, green was removed by oven-drying the seedlings at 105°C for 30 min, followed by drying at 70°C and weighing. The Sb concentrations in the underground and aboveground parts were measured on a WF-210 atomic absorption spectrophotometer (Beifen-Ruili Instrument Co., Ltd., China). The Sb translocation factor (TF%) was computed as aboveground Sb concentration / underground Sb concentration.

Statistic Analysis

All data were expressed as the average of three repeated measurements. In one-way analysis of variance (ANOVA), the homogeneity of variance was compared multiple times using the least significant difference (LSD) method. In case of heterogeneity, Dunnett's T3 was used to test the difference of the same index among different treatments. Statistical analysis and plotting were performed on SPSS 20.0 and SigmaPlot 12.5, respectively. Data were expressed as mean \pm standard deviation (SD).

RESULTS AND ANALYSIS

Effects of Sb Stress on Leaf Pigment Concentrations

The leaf photosynthesis pigment concentrations after different treatments (Sb³⁺ and Sb⁵⁺) are listed in **Table 1**. Compared with CK, the contents of leaf chlorophylls a, b, and a + b (Chl a, Chl b, and Chl a+b), and carotenoid (Car) all significantly decreased after Sb³⁺ treatments (T₁, T₂, T₃, and T₄) and were all negatively correlated with the Sb³⁺ concentrations ($R^2 = -0.8076, -0.8044, -0.8119$ and -0.8116 , respectively; all $P < 0.01$). However, Chlorophyll a/b ratios (Chl a/b) all increased significantly, while the carotenoid/chlorophyll ratios (Chl/Car) decreased from 0.23 to 0.18. The Chl a, Chl b and Car contents in group T₄ versus CK decreased by 77.17, 81.16, and 83.05%, respectively. One-way ANOVA showed the Chl a, Chl b, Chl a+b, and Car contents after Sb³⁺ treatments were significantly different from the CK ($F = 874, 166.5, 561.2, 75$, respectively; all $P < 0.001$). Multiple comparison results expressed in a-d (**Tables 1, 2** and **Figures 1–3**) showed the data of different letters were significantly different at the 95% confidence interval (CI). while the data of the same letter were not significantly different at 95% CI.

The leaf Chl a, Chl b, Chl a+b, and Car contents in group T'₁ all insignificantly increased from the CK ($P > 0.05$). The contents of these pigments all significantly decreased in groups T'₂, T'₃, and T'₄ compared with the CK ($P < 0.05$) and were negatively correlated with the Sb⁵⁺ concentration. The Chl a/b minimized to 2.52 in group T'₁ and maximized to 3.24 in group T'₂, while the Chl/Car ratio rose from 0.23 to 0.30. One-way ANOVA showed the Chl a, Chl b, Chl a+b, and Car contents after Sb⁵⁺ treatments were significantly different from the CK ($F = 760.9, 67.9, 688.1, 26$, respectively; all $P < 0.001$).

Effects of Sb Stress on Seedling Biomass

The growing parameters of *A. calamus* after different treatments (Sb³⁺ and Sb⁵⁺) are listed in **Figure 1**. The growing conditions

of *A. calamus* can be reflected by plant height and plant dry weight. The plant heights, dry aboveground weights and dry underground weights all decreased significantly with the rise of Sb³⁺ concentration (T₁, T₂, and T₃) in a negatively correlated way. ($R^2 = -0.922, -0.935, \text{ and } -0.942$, respectively; all $P < 0.01$). The three parameters of group T₄ versus CK decreased by 35, 36, and 32.4%, respectively. One-way ANOVA showed the plant heights, dry aboveground weights and dry underground weights after Sb³⁺ treatments were significantly different from the CK ($F = 14, 7.8, \text{ and } 6.8$, respectively; all $P < 0.01$).

As for Sb⁵⁺ treatments, the three parameters all maximized in group T'₁, but were not significantly different from the CK ($P > 0.05$). The three parameters all significantly decreased in groups T'₂, T'₃, and T'₄ compared the CK ($P < 0.05$). One-way ANOVA showed the plant heights, dry aboveground weights and dry underground weights after Sb⁵⁺ treatments were significantly different from the CK ($F = 15.8, 7.6, \text{ and } 6.2$, respectively; all $P < 0.01$).

Effects of Sb Stress on Aboveground or Underground Sb Accumulation

The mass fractions of Sb in underground and aboveground tissues after different treatments (Sb³⁺ and Sb⁵⁺) are listed in **Table 2**. Clearly, after Sb³⁺ treatments (T₁, T₂, T₃, and T₄), the underground and aboveground Sb concentrations fell within 39.72–79.68 and 9.28–22.15 mg/kg, respectively, showing significant differences among treatments ($P < 0.05$); TF% varied between 19.44 and 27.8%. One-way ANOVA showed the Sb mass fractions in underground and aboveground tissues after Sb³⁺ treatments were both significantly different from the CK ($F = 2422.9, F = 378.4$, both $P < 0.001$).

After Sb⁵⁺ treatments (T'₁, T'₂, T'₃, and T'₄), the underground and aboveground Sb concentrations fell within 37.85–78.8 and 8.5–19.59 mg/kg, respectively, showing significant differences among treatments ($P < 0.001$); TF% varied between 19.44 and 24.86%. One-way ANOVA showed the Sb mass fractions in underground and aboveground tissues after Sb⁵⁺ treatments were both significantly different from CK ($F = 1939.3, F = 243.8$, both $P < 0.001$).

Effects of Sb Stress on Leaf Gas Exchange Parameters of *A. calamus*

The leaf gas exchange parameters of *A. calamus* after different treatments (Sb³⁺ and Sb⁵⁺) are listed in **Figure 2**. Clearly, the Pn, Gs, and E were all negatively correlated with the increasing Sb³⁺ concentration ($R^2 = -0.8498, -0.8638, \text{ and } -0.8453$, respectively; all $P < 0.01$). While Ci was positively correlated with Sb³⁺ concentration ($R^2 = 0.8824, P < 0.01$). The Pn, Gs, and E of group T₄ versus CK decreased by 38.2, 65.8 and 55.8%, respectively. One-way ANOVA showed the leaf Pn, Gs, and E after Sb³⁺ treatments were all significantly different from the CK ($F = 5.6, 26.3, \text{ and } 30.8$, respectively; all $P < 0.05$), but Ci was not significantly different ($F = 2.5, P > 0.05$).

After Sb⁵⁺ treatments, the leaf Pn, Gs, and E of group T'₁ all increased insignificantly compared with the CK ($P > 0.05$);

TABLE 1 | Effects of antimony stress on leaf photosynthetic pigments in *A. calamus* plants.

| Sb treatment (mg·kg ⁻¹ soil) | | Chl a (mg·g ⁻¹ FW) | Chl b (mg·g ⁻¹ FW) | Chla+b (mg·g ⁻¹ FW) | Car (mg·g ⁻¹ FW) | Chl a/b | Car/Chl |
|---|-----------------------|-------------------------------|-------------------------------|--------------------------------|-----------------------------|--------------|--------------|
| Sb ³⁺ | CK(0) | 1.84 ± 0.04a | 0.69 ± 0.05a | 2.53 ± 0.09a | 0.59 ± 0.05a | 2.67 ± 0.14a | 0.23 ± 0.01a |
| | T ₁ (250) | 1.72 ± 0.02b | 0.54 ± 0.01b | 2.26 ± 0.02b | 0.52 ± 0.04a | 3.19 ± 0.04b | 0.23 ± 0.02a |
| | T ₂ (500) | 1.54 ± 0.05c | 0.47 ± 0.04c | 2.01 ± 0.09c | 0.44 ± 0.05b | 3.28 ± 0.14b | 0.22 ± 0.02a |
| | T ₃ (1000) | 0.46 ± 0.04d | 0.14 ± 0.03d | 0.60 ± 0.06d | 0.11 ± 0.01c | 3.29 ± 0.32b | 0.18 ± 0.01b |
| | T ₄ (2000) | 0.43 ± 0.05d | 0.13 ± 0.02d | 0.56 ± 0.06d | 0.10 ± 0.02c | 3.31 ± 0.04b | 0.18 ± 0.02b |
| ANOVA | F | 874*** | 166.5*** | 561.2*** | 75*** | 7.1*** | 23.5*** |
| Sb ⁵⁺ | CK(0) | 1.84 ± 0.04a | 0.69 ± 0.05a | 2.53 ± 0.09a | 0.59 ± 0.05a | 2.67 ± 0.14b | 0.23 ± 0.01b |
| | T ₁ (250) | 1.89 ± 0.03a | 0.75 ± 0.06a | 2.64 ± 0.09a | 0.62 ± 0.05ab | 2.52 ± 0.15b | 0.23 ± 0.01b |
| | T ₂ (500) | 1.75 ± 0.03b | 0.54 ± 0.05b | 2.29 ± 0.08b | 0.53 ± 0.04b | 3.24 ± 0.25a | 0.23 ± 0.01b |
| | T ₃ (1000) | 0.77 ± 0.04c | 0.27 ± 0.04c | 1.04 ± 0.08c | 0.31 ± 0.02c | 2.85 ± 0.24a | 0.30 ± 0.01a |
| | T ₄ (2000) | 0.72 ± 0.04c | 0.25 ± 0.05c | 0.97 ± 0.09c | 0.27 ± 0.02c | 2.88 ± 0.35a | 0.28 ± 0.02a |
| ANOVA | F | 760.9*** | 67.9*** | 288.1*** | 26*** | 4*** | 152.4*** |

The data with same letter(s) in the same column indicate no significant differences at $P < 0.05$. The data followed with different letter(s) in the same column show significant differences at $P < 0.05$. *** $P < 0.001$. The same in the following tables and figures.

TABLE 2 | Effects of antimony stress on Sb content of *A. calamus*.

| Sb treatment (mg·kg ⁻¹ soil) | | Net Sb content in plant tissues (mg·kg ⁻¹ DW biomass) | | Translocation factor from underground to aboveground parts (TF%) |
|---|-----------------------|--|------------------|--|
| | | Underground part | Aboveground part | |
| Sb ³⁺ | CK(0) | 0.36 ± 0.02d | 0.07 ± 0.01c | 19.44 |
| | T ₁ (250) | 39.72 ± 1.0c | 9.28 ± 0.51b | 23.36 |
| | T ₂ (500) | 42.59 ± 1.17c | 10.65 ± 0.53b | 25.01 |
| | T ₃ (1000) | 73.27 ± 1.32b | 19.34 ± 0.89a | 26.40 |
| | T ₄ (2000) | 79.68 ± 1.45a | 22.15 ± 0.88a | 27.80 |
| ANOVA | F | 2422.9*** | 378.4*** | |
| Sb ⁵⁺ | CK(0) | 0.36 ± 0.02d | 0.07 ± 0.01c | 19.44 |
| | T ₁ (250) | 37.85 ± 1.38c | 8.50 ± 3.70b | 22.46 |
| | T ₂ (500) | 41.07 ± 1.17c | 9.63 ± 0.43b | 23.44 |
| | T ₃ (1000) | 71.54 ± 1.30b | 17.25 ± 0.79a | 24.11 |
| | T ₄ (2000) | 78.80 ± 1.61a | 19.59 ± 0.73a | 24.86 |
| ANOVA | F | 1939.3*** | 243.8*** | |

the three parameters in groups T₂, T₃, and T₄ all significantly decreased (all $P < 0.05$). One-way ANOVA showed the leaf Pn, Gs, E, and Ci after Sb⁵⁺ treatments were all significantly different from the CK ($F = 4.7, 33.1, 28.3, F = 3.6$, respectively; all $P < 0.05$).

Effects of Sb Stress on Leaf Chlorophyll Fluorescence Parameters of *A. calamus*

The leaf chlorophyll fluorescence parameters of *A. calamus* after different treatments (Sb³⁺ and Sb⁵⁺) are listed in Figure 3. The F_v/F_m and Φ_{PSII} both decreased significantly with the rising Sb³⁺ concentration in a negatively correlated way ($R^2 = -0.9306, -0.9619$; both $P < 0.01$). The F_v/F_m and Φ_{PSII} of group T₄ versus the CK decreased significantly by 31.6 and 34.5%, respectively (both $P < 0.05$). One-way ANOVA showed the F_v/F_m and Φ_{PSII} after Sb³⁺ treatments, were significantly different from the CK ($F = 102, F = 82$, both $P < 0.001$).

The F_v/F_m and Φ_{PSII} of group T₂, T₃, and T₄ versus the CK all decreased significantly (all $P < 0.05$), but increased insignificantly in group T₁ (both $P > 0.05$). One-way ANOVA showed the F_v/F_m and Φ_{PSII} after Sb⁵⁺ treatments were significantly different from the CK ($F = 96, F = 46$, both $P < 0.001$).

DISCUSSION AND CONCLUSIONS

Chlorophylls are an important type of pigments needed by plants for photosynthesis and participate in optical energy absorption, transfer and conversion during photosynthesis. Chlorophylls are pivotal in photosynthesis and their content variations reflect the degree of photosynthesis (Bot et al., 1990; Zhang et al., 2011). However, heavy metals (e.g., Ni, Cu) could induce toxic effects on chlorophylls, such as synthesis inhibition and

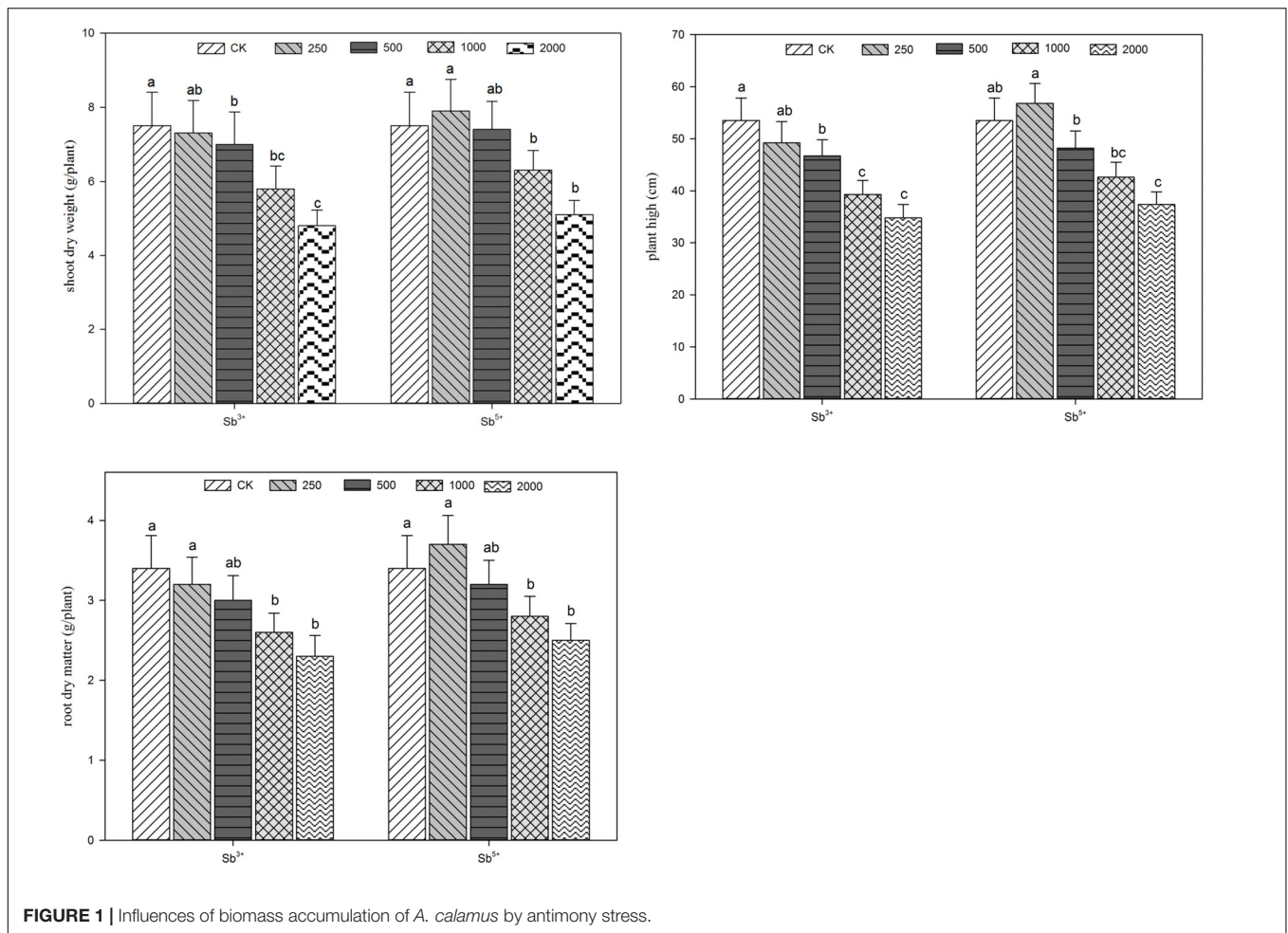


FIGURE 1 | Influences of biomass accumulation of *A. calamus* by antimony stress.

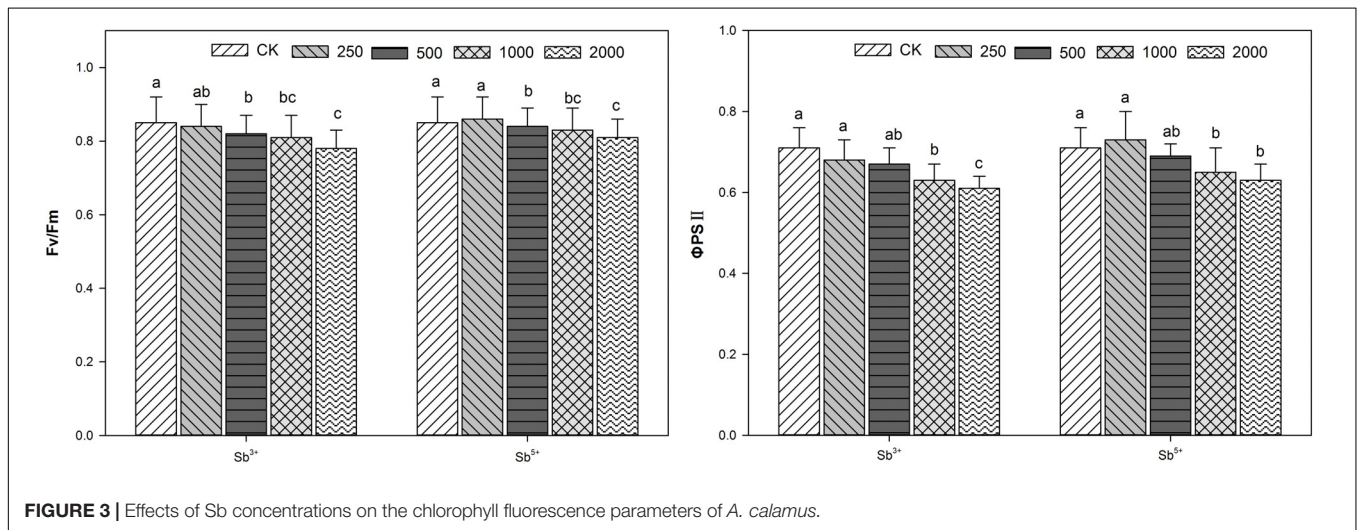
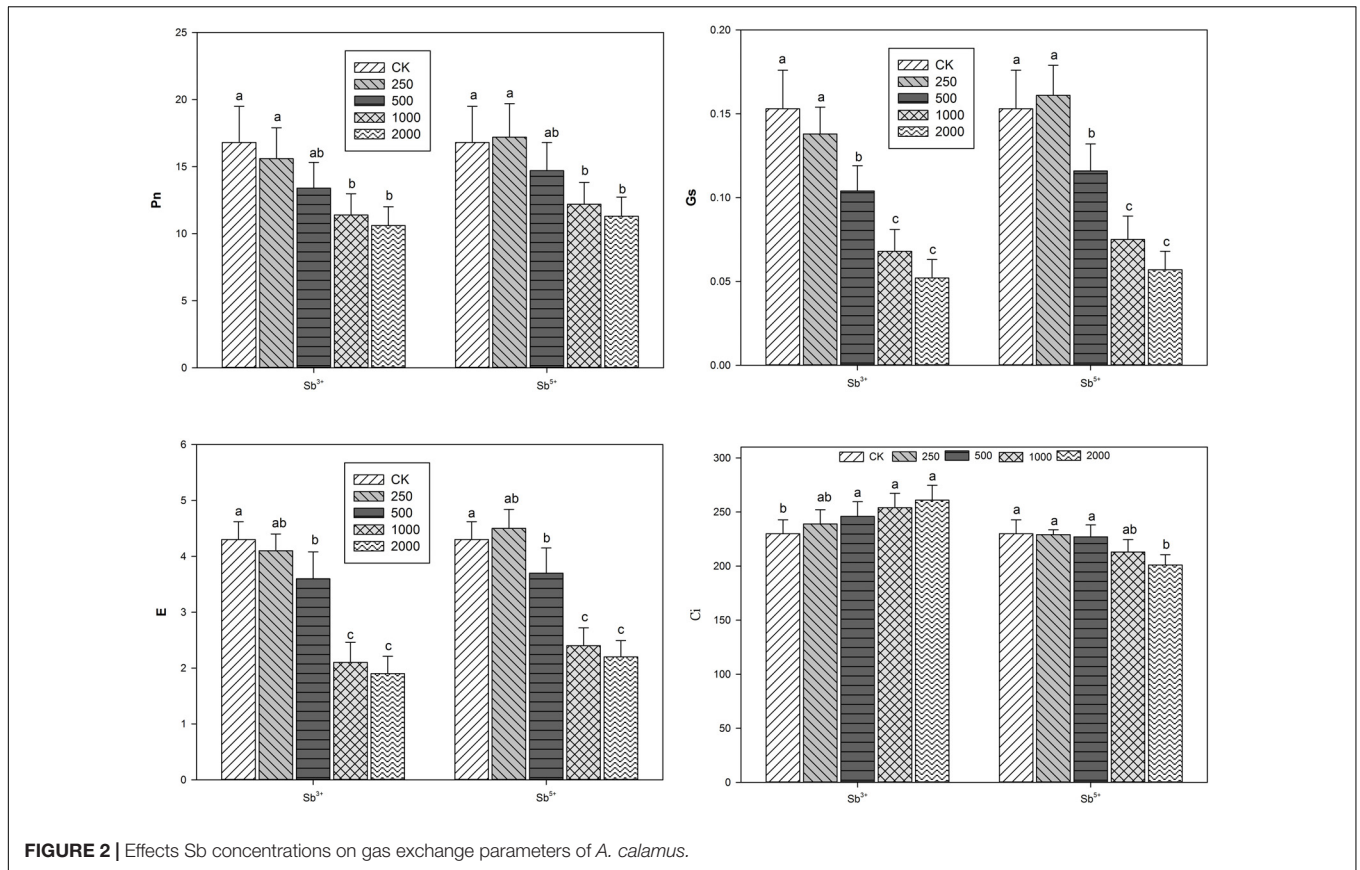
structural destruction (Sreekanth et al., 2013; Llagostera et al., 2016).

In this study, the pigment contents all decreased under high-concentration Sb stress (500–2000 mg/kg). The possible reason is that though Sb is not necessary to the growth of *A. calamus*, when the environmental Sb concentration rises, more Sb would enter plant cells to bind with the sulfhydryl group of chloroplast proteins, destroying the structures and functions of chloroplasts and disturbing the balanced chlorophyll enzymatic activity ratios. As a result, the accelerated chlorophyll decomposition and broken physiological balance would decrease chlorophyll contents, photosynthesis, growth, and biomass *in vivo*. Moreover, under Sb stress, Sb accumulates largely in roots (Table 2), which may interfere with the cation absorption and chlorophyll synthesis of plants, and block the transportation of some nutrient elements into leaves.

The Chl a/b ratios in groups T₁, T₂, T₃, and T₄ basically rose with the increasing Sb³⁺ concentration, but changed in different ways from Chl a or Chl b concentrations. The Chl a/b ratio reflects the ratio between the appressed and non-appressed membrane regions, and is inversely proportional to the degree of thylakoid oppression (Anderson and Aro, 1994), a way where the plants adaptively use optical energy to the largest

extent. Firstly, thylakoid oppression makes light-harvesting chlorophyll-protein complexes (LHCII) more tightly connected and improves the light-harvesting ability and energy transfer efficiency (Dutta, 1986). Secondly, thylakoid oppression enhances the photoinduced electron/proton transfer ability of the PSII and cytochrome bf complex, and thereby most largely accelerates the linear electron transfer (Chow, 1999). The rise of Chl a/b ratio induced by Sb stress may also indicate the partial thylakoid unoppression and the occurrence of photoinhibition in leaves. The reduction of thylakoid oppression contributes to dissipation of excessive excitation energy and is an active manifestation of *A. calamus* in adaptation to environmental stress. Chl b is more prone to Sb-induced than Chl a, and the reduction of Chl b and Chl a contents reflects the leaf senescence. The reduction of relative Chl b content would reduce optical energy capture, active O generation, and protein degradation, which improve the Sb-tolerance of *A. calamus*.

The Chl a/b ratios in groups T'₁, T'₂, T'₃, and T'₄ changed in very complicatedly ways, probably because the Sb absorption led to a metabolic disorder in *A. calamus*. The Car/Chl ratio was basically stable with the presence of 0–500 mg/kg Sb, but significantly decreased in groups T₃ and T₄ compared with the CK ($P < 0.01$), and significantly rose in groups



T'_3 and T'_4 ($P < 0.01$), indicating the effects of Sb on carotenoids of *A. calamus* are complicated when different forms are considered. Carotenoids can absorb visible light and transfer the absorbed optical energy to chlorophyll a, and can antioxidant and dissipate excessive optical energy. The Sb stress causes carotenoid degradation, and thereby structural change of thylakoid membranes, reduction of electron transfer and PSII activity. However, Sb stress destroys the ability of

carotenoids to clear away oxygen free radicals *in vivo* in *A. calamus*, destabilizing thylakoid membranes and harming the plants.

Plant height reflects the effects of Sb stress on the growth and development of *A. calamus*. The toxicity of Sb would make leaves thinner, smaller, less developed and the plants shorter. Beyond a certain concentration, Sb stress would inhibit the growth of *A. calamus* and reduce physiological activity. *A. calamus* can

adopt antioxidation enzyme systems (e.g., superoxide dismutase SOD, peroxidase POD) *in vivo* to efficiently relieve the harms of low Sb stress, but the Sb concentration exceeding a certain level could weaken the stress-resistant ability and inhibits plant growth, which is manifested as a reduction of plant height, dry weight and biomass.

The translocation factor (TF%) is the concentration ratio of an element in the aboveground part to the underground part of a plant. TF% reflects the ability of a heavy metal to transfer from the underground part to the aboveground part (e.g., leaves, stem) (Islam et al., 2013). In *A. calamus*, the antimony concentration is about four times higher in underground part than in aboveground part, and TF% is from 19.44% to 27.8%. As reported, the Sb accumulation in maize was promoted by the rise of soil Sb concentration, and Sb easily transferred from roots to shoots, with the largest TF% of 2.05 (Dwivedi et al., 2015). The high soil Sb concentrations significantly reduced the growth and biomass of maize, and inhibited the activities of POD and SOD (Dwivedi et al., 2015). Thus, the TF% of Sb may be related to the plant species and Sb concentration (Pan et al., 2011). The root Sb concentration of the arsenic hyperaccumulator *Pteris vittata* reached 12000 mg/kg and accounted for 99% of hosted Sb in the plants (Tisarum et al., 2014). The enrichment ability of sunflower (*Helianthus annuus* L.) under Sb stress was larger in roots than in leaves, but Sb stress inhibited plant growth; the Sb accumulation in the aboveground part significantly altered the physical conditions of sunflower, reduced intercellular gaps and made leaf tissues closer (Vaculik et al., 2015; Ortega et al., 2017). According to the report, root is the main organ for taking up metal nanoparticles from water are badly influenced in comparison with shoot in some plants (Tripathi et al., 2016; Anshu et al., 2017).

Acorus calamus is a Sb non-hyperaccumulator, and the root-absorbed Sb is mostly hosted in roots, which avoids the transfer to the aboveground part and the occurrence of harms. This detoxification mechanism is very significant in eliminating the biological toxicity of Sb from the aboveground part. A larger TF% indicates the aboveground part is more able to accumulate a heavy metal and transfer the majority to the aboveground part, where the heavy metal can be removed during harvest. With the increase of Sb concentration in the growing medium, the TF% rises in significantly ($P > 0.05$; **Table 2**), indicating the Sb absorbed by *A. calamus* is mostly enriched in the underground part, which is favorable for plant growth and enhances the stress resistance. In comparison, the change of the total concentration of antimony in the growth substrate is closely accords with that of roots Sb concentration of *A. calamus* ($R^2 = 0.82$). Phytoremediation technology in which plants take up the contaminants from the soil/water and keep them in the root system is categorized as phytostabilization. According to the remediation mechanisms, phytoremediation falls into five general categories: phytofiltration, phytoextraction, phytodegradation, phytostabilization, and phytovolatilization (Sytar et al., 2016).

Background value of antimony in terrestrial vascular plants was less than 0.05 mg/kg dry weight (Baroni et al., 2000), and antimony accumulated to 5–10 mg/kg in plant tissue would be

toxic to plant (Kabata-Pendias and Pendias, 2010). But in the present study, Sb concentrations in the highest 79.68 mg/kg dry weight didn't cause the death of *A. calamus*, so *A. calamus* had some resistance to antimony.

Sb level in the underground of *Plantago lanceolata* were 1150mg/kg dry weight, yet *Plantago lanceolata* are not used to deal with antimony polluted water (Baroni et al., 2000). Sb levels in shoots of *Typha latifolia*, *Scirpus sylvaticus*, and *Phragmites australis* were 15, 19, and 15 mg/kg dry weight, respectively; the accumulation ability of Sb in *A. calamus* (shoots, 22.15mg/kg dry weight) is better than three aquatic plants.

Phytoremediation is a new cheap and eco-friendly technique that depends plants to clean the environmental pollution by heavy metals (Sytar et al., 2016). In the present study, the aquatic plant *A. calamus* was tested for its ability to accumulate Sb from contaminated water in laboratory experiments. The results showed that *A. calamus* serves as a good candidate for phytoremediation of water contaminated with Sb.

Photosynthesis, an important evaluation criterion of plant productivity, is a physiological process very sensitive to heavy metal stress and directly provides energy for plant growth and development. The photosynthesis ability of *A. calamus* as a wetland plant is related with its ability to absorb pollutants from water. Generally, stomatal factors mainly include the number, size and opening degree of stomas, while non-stomatal factors include enzymatic activity and photosynthesis components. Under environmental stresses, both stomatal limitation and non-stomatal limitation could lead to the reduction of Pn (Lal et al., 1996; Wu et al., 2014) and can be basically discriminated by Ci. When Pn and Ci both decrease, the decrease of photosynthesis ability is induced by stomatal limitation; when Pn declines but Ci increases, the cause is non-stomatal limitation (Farquhar and Sharkey, 1982).

In addition to the stoma closure, the reduction of Pn is also related to the decrease of mesophyll cell photosynthetic activity. In our study, Pn declined but Ci rose in groups T₁, T₂, T₃, and T₄, indicating the reduction of Pn was mainly caused by non-stomatal factors and that the mesophyll cells of *A. calamus* were injured and the photosynthesis activity was weakened, leading to the reduction of CO₂ fixation ability and the accumulation of intercellular CO₂. The photosynthesis parameters all maximized with the presence of 250 mg/kg Sb⁵⁺, but the inhibitory effects of high concentrations (500–2000 mg/kg) on photosynthesis were gradually enhanced. Pn and Ci decline simultaneously, indicating the partial stoma closure is the main cause of photosynthetic rate reduction.

Chlorophyll fluorescence is an ideal probe to study the relationship between photosynthesis and environmental stress and provides rich information for research on PSII and electron transfer. The chlorophyll fluorescence properties reflect the relationship between photosynthesis physiology and environmental stress. F_v/F_m is the major index of PSII photochemical efficiency and reflects the initial optical energy conversion efficiency. F_v/F_m indicates the optical energy using ability of PSII and is closely related to the photosynthesis inhibition degree. The presence of stress would reduce F_v/F_m

and destroy the PSII reaction centers. In our study, with the increase of Sb concentration, the F_v/F_m declined in groups T₁, T₂, T₃, and T₄, indicating the PSII reaction centers were destroyed. Φ_{PSII} indicates the real photochemical efficiency upon the partial closure of reaction centers when the PSII reaction centers are stressed environmentally. PSII was confirmed as an important site of photoinhibition, but its light use efficiency can be reduced by drought/flooding, heavy metals, saline or other stress factors, and there by the plants were injured by photoinhibition. As reported, the chlorophyll fluorescence parameters of Ramie (*Boehmeria nivea* L.) including F_v/F_m and Φ_{PSII} were not significantly changed under Sb stress (20–200 mg·L⁻¹) (Chai et al., 2016), but were significantly reduced under salty or drought stress (Huang et al., 2013). The maximum photochemical quantum yield (F_v/F_m) of maize under Sb stress (50–1000 mg·kg⁻¹ soil) declined (Zhang et al., 2017). *A. utriculata* is a nickel hyperaccumulator, and its F_v/F_m was basically unchanged after high concentration treatment and its photosynthetic performance was still high (Roccotiello et al., 2016). Excessive cadmium inhibits the effects of photosynthesis by means of disrupting the PSII functions, however, different from Cd, lead (Pb) stress inhibits the effects of photosynthesis by disrupting chloroplast ultrastructure and thylakoid membrane lipid composition (Kalaji et al., 2016).

In our study, F_v/F_m and Φ_{PSII} both significantly decreased after treatments T₂, T₃, and T₄ compared with the control, but the declining amplitudes were smaller than in treatments T₁, T₂, T₃, and T₄, indicating Sb⁵⁺ and Sb³⁺, after reaching certain levels, both could damage the PSII reaction centers.

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AUTHOR CONTRIBUTIONS

XZ conducted the experimental design and statistic analysis. CS and PZ analyzed the data. FL wrote the paper with assistance from XZ. All authors commented on an earlier draft of manuscript.

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