



The Mating System of the Wild-to-Domesticated Complex of *Gossypium hirsutum* L. Is Mixed

Rebeca Velázquez-López^{1*†}, Ana Wegier^{1*†}, Valeria Alavez¹, Javier Pérez-López¹, Valeria Vázquez-Barrios¹, Denise Arroyo-Lambaer¹, Alejandro Ponce-Mendoza² and William E. Kunin³

¹ Laboratorio de Genética de la Conservación, Jardín Botánico, Instituto de Biología, Universidad Nacional Autónoma de México, Mexico City, Mexico, ² Comisión Nacional para el Conocimiento y Uso de la Biodiversidad, Mexico City, Mexico, ³ Department of Ecology and Evolution, Faculty of Biological Sciences, University of Leeds, Leeds, United Kingdom

OPEN ACCESS

Edited by:

Charles Roland Clement,
National Institute of Amazonian
Research, Brazil

Reviewed by:

Sevan Suni,
Harvard University, United States
Alexandre Magno Sebbenn,
Instituto Florestal, Brazil
Fernanda Amato Gaiotto,
Universidade Estadual de Santa Cruz,
Brazil

*Correspondence:

Rebeca Velázquez-López
rebecavelazquezl@gmail.com
Ana Wegier
awegier@ib.unam.mx;
awegier@gmail.com

[†]These authors have contributed
equally to this work.

Specialty section:

This article was submitted to
Evolutionary and Population Genetics,
a section of the journal
Frontiers in Plant Science

Received: 18 August 2017

Accepted: 13 April 2018

Published: 09 May 2018

Citation:

Velázquez-López R, Wegier A,
Alavez V, Pérez-López J,
Vázquez-Barrios V,
Arroyo-Lambaer D,
Ponce-Mendoza A and Kunin WE
(2018) The Mating System of the
Wild-to-Domesticated Complex
of *Gossypium hirsutum* L. Is Mixed.
Front. Plant Sci. 9:574.
doi: 10.3389/fpls.2018.00574

The domestication syndrome of many plants includes changes in their mating systems. The evolution of the latter is shaped by ecological and genetic factors that are particular to an area. Thus, the reproductive biology of wild relatives must be studied in their natural distribution to understand the mating system of a crop species as a whole. *Gossypium hirsutum* (upland cotton) includes both domesticated varieties and wild populations of the same species. Most studies on mating systems describe cultivated cotton as self-pollinated, while studies on pollen dispersal report outcrossing; however, the mating system of upland cotton has not been described as mixed and little is known about its wild relatives. In this study we selected two wild metapopulations for comparison with domesticated plants and one metapopulation with evidence of recent gene flow between wild relatives and the crop to evaluate the mating system of cotton's wild-to-domesticated complex. Using classic reproductive biology methods, our data demonstrate that upland cotton presents a mixed mating system throughout the complex. Given cotton's capacity for outcrossing, differences caused by the domestication process in cultivated individuals can have consequences for its wild relatives. This characterization of the diversity of the wild relatives in their natural distribution, as well as their interactions with the crop, will be useful to design and implement adequate strategies for conservation and biosecurity.

Keywords: cotton, crop wild relatives, mating system, reproductive success, xenogamy, autogamy, domestication process, introgression

INTRODUCTION

Plant domestication is a complex and continuing process (Casas et al., 2007; Vaughan et al., 2007). For 10,000 years, humans have selected attributes of interest in a range of economically valuable plants through their management and utilization (Gepts, 2004); consequently, different techniques, trait preferences, environments, and selection intensities have shaped the degree of domestication of each species (Meyer and Purugganan, 2013). Today we can find: (1) crop populations that are highly domesticated and depend on human intervention for survival; (2) semi-domesticated populations with recognizable traits of the domestication syndrome, but able to survive in the wild if human intervention ceases; (3) incipiently domesticated populations whose selected traits

have not yet diverged markedly from those found in wild populations; (4) incidentally co-evolved populations that adapt to human disturbed environments, but without direct human selection; (5) feral populations derived from 2, 3, or 4; and (6) wild relatives (Clement, 1999). Given these diverse scenarios, the biological diversity contained in wild-to-domesticated complexes should be considered in studies about crop ecology and evolution (Warwick and Stewart, 2005; Casas et al., 2007).

In plants, one of the key life history traits is the mating system (Vaughan et al., 2007). This feature helps determine the genetic composition of populations and, therefore, has a crucial role in the evolution of species (Charlesworth, 2006); additionally, it explains who is mating with whom, which is a fundamental issue for conservation biology (Barrett and Harder, 1996). The mating system often changes during domestication (Meyer et al., 2012), and wild relatives contain the plesiomorphic state of this trait (Ellstrand et al., 1999; Doebley et al., 2006; Andersson and de Vicente, 2010). A shift from the ancestral system toward a new one can be selected until fixation; for instance, there are some crops that are unable to reproduce without human intervention (Ellstrand et al., 1999), such as vegetatively propagated sycamore fig and other fruit trees (Zohary and Spiegel-Roy, 1975). Some crops have multiple mating systems, such as domesticated 'Maradol' (*Carica papaya*), which is hermaphroditic, while native varieties and wild papayas are dioecious (Carvalho and Renner, 2012). Importantly, the characterization of the mating system of many plant species has been biased toward the domesticated counterparts, because only a sub-sample of the wild-to-domesticated complex was used (e.g., *Carica papaya* (Damasceno et al., 2009), *Persea americana* (Ish-Am et al., 1999), *Piper nigrum* (Thangaselvabal et al., 2008)). This bias may have profound consequences for the conservation of conspecific wild relatives, especially because conclusions drawn from studies with domesticated varieties are extrapolated to the whole species, failing to consider the genotypic and phenotypic diversity that wild relatives possess. The conservation of this diversity is fundamental, because it is a genetic reservoir that includes a wider range of adaptive traits that may be of additional agricultural relevance, such as resistance to pests and pathogens and tolerance to abiotic stresses (Warschefsky et al., 2014).

Upland cotton, *Gossypium hirsutum*, is an economically important plant species, particularly known for being the leading source of natural fiber. Worldwide, over 90% of cotton production comes from cultivars of *G. hirsutum* and in 2014 the species ranked eighth in the world's harvested area, reaching almost 35 million hectares (Crop production, FAOSTAT, 2017). Given the economic importance of the species, its mating system has been the focus of several studies since 1903 (Simpson, 1954); however, the majority of them concentrated on domesticated cotton and have described it as predominantly autogamous and self-pollinated (see Supplementary Material 1). On the other hand, studies on pollen dispersal of *G. hirsutum*, from the beginning of its modern breeding as a crop to the present day, refer to cotton's ability to produce offspring by crossing (1944–2016; see Supplementary Material 1 for a review); however, the mating system is not described as mixed (Loden and Richmond, 1951; Richmond, 1951; Simpson, 1954; Imam and Allard, 1965;

Meredith and Bridge, 1973). A specific study on the mating system of wild populations in their natural distribution is lacking.

In Mesoamerica, *G. hirsutum* exists as a complex of wild to domesticated forms (Brubaker and Wendel, 1994); hence, it is an ideal region to characterize the mating system of this upland cotton complex, identify possible differences, and to integrate this information into regional management plans. In Mexico - its center of origin, diversity and domestication (Ulloa et al., 2005; Burgeff et al., 2014; Pérez-Mendoza et al., 2016) - the complex includes cultivated and highly improved varieties, genetically modified varieties, traditionally managed landraces, feral, and wild populations. All of them belong to the primary gene pool of the species (Andersson and de Vicente, 2010) and gene flow among them occurs, even over long distances (Wegier et al., 2011). Moreover, eight wild *G. hirsutum* metapopulations have been recognized, based on geographic, ecologic, and genetic differences (Wegier et al., 2011; Bauer-Panskus et al., 2013). Wegier et al. (2011) demonstrated that recent gene flow, followed by introgressive hybridization, occurs between a number of wild populations distributed in the north and south of Mexico, and commercial cotton cultivars in the northern states of the country. Our study provides the first data on the mating system of wild *G. hirsutum in situ* within the natural distribution of the species in Mexico. In order to assess this, we evaluated the capacity of domesticated cotton, wild cotton, and wild cotton with evidence of introgression, to produce offspring by either xenogamy (cross-pollination between different genets) or autogamy (self-fertilization).

MATERIALS AND METHODS

Study System

Upland cotton, *G. hirsutum* L., is a species with wild, feral, and semi-domesticated populations (Brubaker and Wendel, 1994). All cultivated forms, including the highly improved varieties or genetically engineered varieties, cannot be considered as fully domesticated, because they are able to survive even if human intervention stops.

Wild *G. hirsutum* flowers all year round. Flowers are white, hermaphrodite, cup shaped, with a single central style surrounded at the bottom by stamens (Meade, 1918; Smith and Cothren, 1999). Some plants exhibit flowers with a colored disk inside of the base of the cup that ranges from deep red to light yellow (Tan et al., 2013). Flowers remain open between 8 and 11 h; at the start of the day, they are all white and when they close the sepals start turning pink at the base (Smith and Cothren, 1999). Anthesis takes place in the morning, as soon as the flower completely opens, and the stamens start to release pollen soon afterward (Smith and Cothren, 1999). Flowers produce both pollen and nectar as a reward for visitors (Wäckers and Bonifay, 2004).

Study Sites

Sampling was performed in coastal dunes and dry forests of Mexico, in three of the eight wild cotton metapopulations defined genetically, geographically, and ecologically by

Wegier et al. (2011), namely: Central Pacific Metapopulation (CPM), Yucatan Peninsula Metapopulation (YPM), and South Pacific Metapopulation (SPM). SPM is of particular interest due to evidence of recent introgressive hybridization with domesticated plants (Wegier et al., 2011). Given the distinctive extinction-colonization dynamic observed in cotton metapopulations (Wegier, 2013), the full extent of CPM, YPM, and SPM was surveyed to find *G. hirsutum* patches with enough flowers (Figure 1). Hand-pollination treatments (Tate and Simpson, 2004; Machado and Sazima, 2008; Hernández-Montero and Sosa, 2016) were carried out during the dry season, between November 2012 and May 2013: CPM in November 2012, YPM in December 2012, and SPM in February 2013. Sites were revisited for fruit collection after 86 days, on average. In addition, domesticated cotton plants, bought in local markets, were kept under greenhouse conditions in Mexico City to maintain a suitable temperature (Figure 1).

Mating System

In order to execute the hand-pollination treatments to test for different mating systems, a search was conducted for flower buds before anthesis (Tate and Simpson, 2004; Machado and Sazima, 2008; Hernández-Montero and Sosa, 2016). In each metapopulation, 40 replicates of the five pollination treatments were set up (Table 1), anticipating the risk of collecting too few fruits afterward: assisted self-pollination, automatic self-pollination, assisted cross-pollination (Kearns and Inouye, 1993), emasculated control (to avoid automatic self-pollination), and control (open-pollination). Multiple treatments were placed on the same plant where possible to control for individual variation; however, due to the variability of the number of flowers, not all of the plants held the same type or number of treatments. Moreover, when flowers were scarce, treatments were placed daily within each study site, in up to four patches per metapopulation, until the 40 replicates per treatment were completed. Special care was taken to avoid changing or altering the environment (i.e., without introducing new genotypes or changing plant abundances or distributions). The same experimental design was applied for domesticated plants in a greenhouse that allowed the entry of local insects. Some treatments required mesh bags to exclude any pollinator access that could alter the results (Table 1). The treatments that did not include bagging before anthesis were bagged after flower closure to help control for mechanical damage from bagging.

Reproductive Success

Fruit-set was calculated as the percentage of recorded fruits produced by each treatment in each metapopulation (Dafni, 1992). In each study site, 20 flowers not involved in the pollination-treatments were collected and brought back to the laboratory in separate sealed containers with 70% alcohol. Each flower was dissected to count the number of ovules present. An average number of ovules was calculated for each wild metapopulation and for domesticated plants. Afterward, seed-set (Schoper et al., 1987; Burd, 1994) was calculated as the percentage of seeds obtained from each fruit for each pollination treatment in relation to the average number of ovules of the study population

to which these fruits belonged. Additionally, all seeds were weighed individually to estimate the seed weight per treatment in each study site. Later, all the seeds were germinated individually. Each seed was washed with 2% Captan (PESTANAL®, Merck) solution and covered with a damp cotton swab; tissue culture lids were used. Seeds were checked daily until all reached emergence of the radicle. While some studies on seed germination consider only a set of seeds (Schemske, 1983; Gil and López, 2015; Raphael et al., 2017; Farooq et al., 2018), we took into account all of the collected seeds for the analysis.

Outcrossing Rate

The outcrossing rate (Te) was calculated for each study site following Barrett et al. (1996):

$$Te = 1 - S \quad (1)$$

where S is the selfing rate, estimated with the fruit-set results from our selfing (W_s) and outcrossing (W_x) treatments, i.e., automatic self-pollination and emasculated control, respectively. For CPM, W_x was obtained with the fruit-set from the assisted cross-pollination treatment, because none of the emasculated control results were found when revisiting the metapopulation for fruit collection.

$$S = \left(\frac{w_s}{w_s + w_x} \right) \quad (2)$$

Statistical Analyses

To test if there were significant differences in seed-set and seed weight among treatments, a Generalized Linear Mixed Model GLMM (Zuur et al., 2009) was used considering the plant as a random factor, because the pollination treatments were not equally represented in each plant (as explained in section 2.3). For GLMM analyses, a Quasi-Poisson distribution was considered for seed-set and a Gaussian distribution for seed weight (Cayuela, 2009). Afterward, a Tukey *post hoc* test was performed to evaluate the significance of the results. To compare germination frequencies and percentage of fruit-set, a chi square test was used with the *post hoc* standardized residue test for each one. Outliers were identified using the method described by Viechtbauer and Cheung (2010); to summarize, a multivariate detection method (Cook distance) was used to calculate the distance among all data points, and those that were not included in the general model were identified as “influential data points” or outlier values. Germination was calculated as the number of germinated seeds in relation to the total number of seeds (Gómez, 2004; Gil and López, 2015). All tests were carried out with the *lme4*, *multcomp*, *stats*, and *ggplot2* packages of R version 3.4.3 (R Core Team, 2017). The scripts utilized for the analyses are available online at <https://github.com/conservationgenetics/BiologiaReproductiva.git>.

RESULTS

Fruit-Set, Seed-Set, and Seed Weight

All treatments produced fruits regardless of the metapopulation (Table 2). The CPM open-pollinated control showed the highest

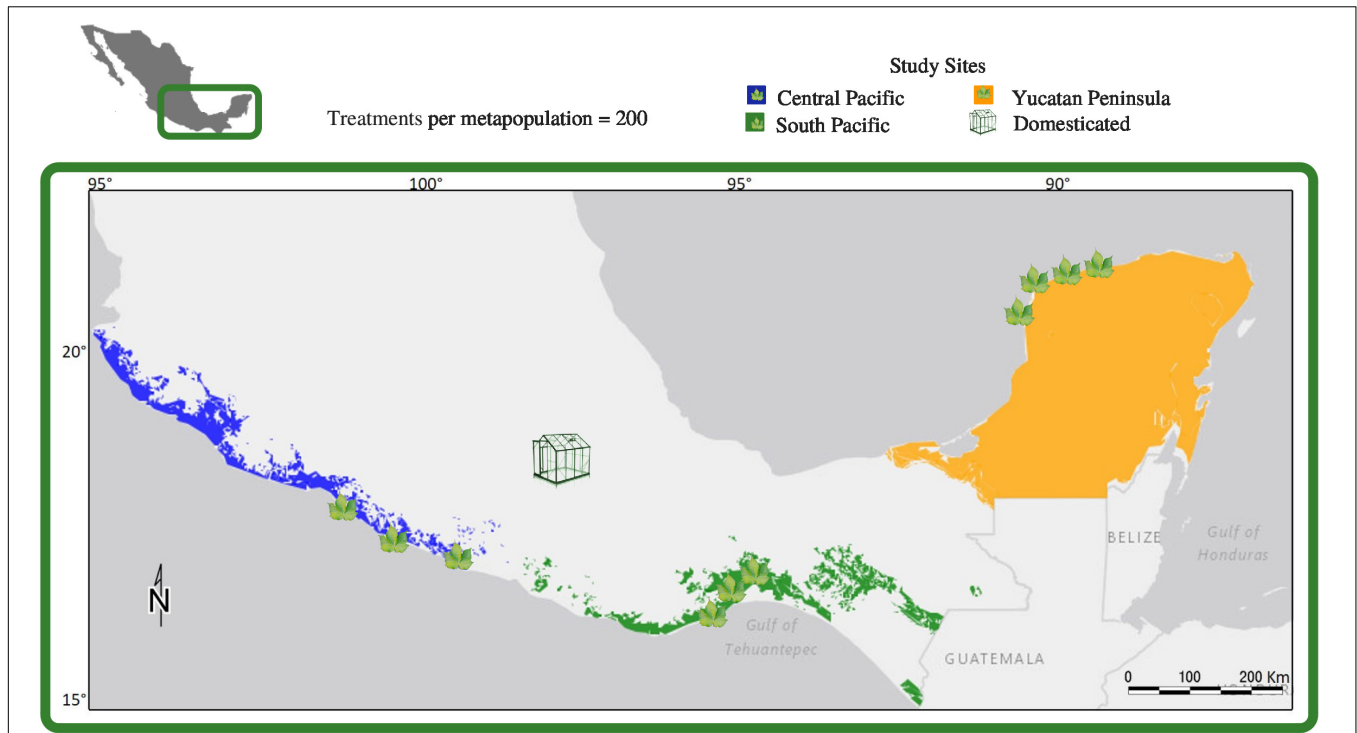


FIGURE 1 | Approximate distributions of the Central Pacific (blue), Southern Pacific (green) and Yucatan Peninsula (orange) upland cotton metapopulations studied in Mexico, with locations of field study sites (leaves) and the greenhouse in Mexico City where cultivated upland cotton was grown out for this study. The Central Pacific and Yucatan Peninsula metapopulations are wild and the Southern Pacific metapopulation contains populations with evidence of recent introgression with cultivated upland cotton.

TABLE 1 | Characteristics of each of the pollination treatments applied to *G. hirsutum* flowers, following Dafni (1992), in three field wild metapopulations and cultivated cotton in the greenhouse.

Hand Treatment	Pollination type	Emasculation	Bag	Pollen source
Automatic self-pollination	Autogamy	No	Yes	Bagged before anthesis and no hand pollination provided.
Assisted self-pollination	Autogamy	No	Yes	Hand pollinated from the anthers to the stigma of the same flower; bagged before anthesis, and after hand self-pollination.
Assisted cross-pollination	Xenogamy	Yes	Yes	Pollen from a different plant was transferred to the stigma of the emasculated focal flower before bag placement.
Emasculated Control	Allogamy (xenogamy and geitonogamy)	Yes	No	Flower emasculated and no hand pollination done; bagged only after closing.
Open-pollination Control	Autogamy and allogamy	No	No	No hand pollination or emasculatation of the flower; bagged only after closing.

value of all treatments among all groups; YPM showed the highest fruit-set produced by outcrossing, and the lowest by automatic self-pollination and control treatment. On the other hand, the highest fruit-set was observed for all the treatments in SPM, with exception of the open-pollination control.

Seeds were produced both by outcrossing and selfing treatments in all metapopulations (Figure 2). The average number of seeds per fruit was 15.9 in SPM, 15.4 in domesticated, 12.3 in CPM, and 10.0 in YPM, while the average number of ovules was 16.3 in SPM, 28.6 in domesticated, 15.6 in CPM, and 13.4 in YPM. Regarding seed-set, the control treatment of

domesticated cotton was lower than that of wild and introgressed plants [$P(\chi^2) = 4.13 \times 10^{-4}$, $df = 3$] (Figure 3). When evaluating the results of each metapopulation individually, CPM presented seed-set differences between the control and the rest of the treatments ($P(\chi^2) = 0.1 \times 10^{-4}$, $df = 3$), in SPM the differences were found between cross-pollination and all treatments, except emasculated control [$P(\chi^2) = 0.001$, $df = 4$], while in YPM and the domesticated there were no significant differences among treatments (Figure 2). On the other hand, seed weight only presented differences in SPM, between cross-pollination and both assisted and automatic self-pollination [$P(\chi^2) = 0.026$,

TABLE 2 | Fruit-set percentage in five pollination treatments applied to *G. hirsutum* and Chi-square test per treatment.

Sites	Open-pollination: control	Emasculated control	Assisted self-pollination	Automatic self-pollination	Cross-pollination
YPM	20.93*	20.00	32.69*	13.73*	51.85*
CPM	60.00*	ND	20.69*	25.93	24.14
SPM	48.84	78.26	73.33*	69.23*	54.05
Domesticated	43.48	10.00	60.00*	34.09	19.05
Chi-square test	$\chi^2 = 18.70, df = 3,$ $p = 3.1 \times 10^{-4}$	NA	$\chi^2 = 37.68, df = 3,$ $p = 3.3 \times 10^{-8}$	$\chi^2 = 47.69, df = 3,$ $p = 2.46 \times 10^{-10}$	$\chi^2 = 26.78, df = 3,$ $p = 6.5 \times 10^{-6}$

*Categories significantly different using adjusted standardized residuals greater than 2.0 and less than -2.0 (χ^2 test). ND: Fruit-set could not be determined because none of the treated flowers were found when revisiting the metapopulation for fruit collection. NA: the chi-square test could not be determined because one of the treatments was not collected.

$df = 4$] (Figure 2). In YPM, CPM, and the domesticated cotton, no significant differences among treatments were observed (Figure 2).

Germination

Less than 20% of the seeds from wild metapopulations CPM and YPM germinated. Regarding the domesticated group, 40–63% of the seeds germinated, except for the seeds produced by cross-pollination that only reached 28%. The seeds from the five treatments assessed at SPM showed germination percentages above 83%, with cross-pollination reaching the highest value of 96% (Table 3).

With regard to the germination rate of the seeds that germinated (Figure 4), the slope of the curve suggests that wild upland cotton presents some kind of inhibition to the completion of germination, whereas domesticated populations do not display this behavior. As shown in Figure 4A, domesticated seeds germinated faster, within the first 6 days, whereas seeds presenting evidence of introgressive hybridization (SPM) reached 95% of germination within the first 7 days and continued germinating for 48 days. Unlike domesticated and SPM seeds, the seeds of wild plants germinated over the course of 73 days (Figure 4A). Concerning the pollination treatments from all study sites, 50% of the seeds of all treatments germinated within the first 5 days; however, after the 5th day the difference in germination rate is evident between autogamy and the rest of the treatments (Figure 4B).

Outcrossing Rate

All study groups presented outcrossing rates different from 0 and 1 (i.e., $1 > Te > 0$), which is indicative of a mixed mating system. Wild metapopulations (YPM and CPM) recorded a higher outcrossing rate (0.72 and 0.71, respectively) than SPM (0.40) and domesticated (0.65) (Supplementary Material 1).

DISCUSSION

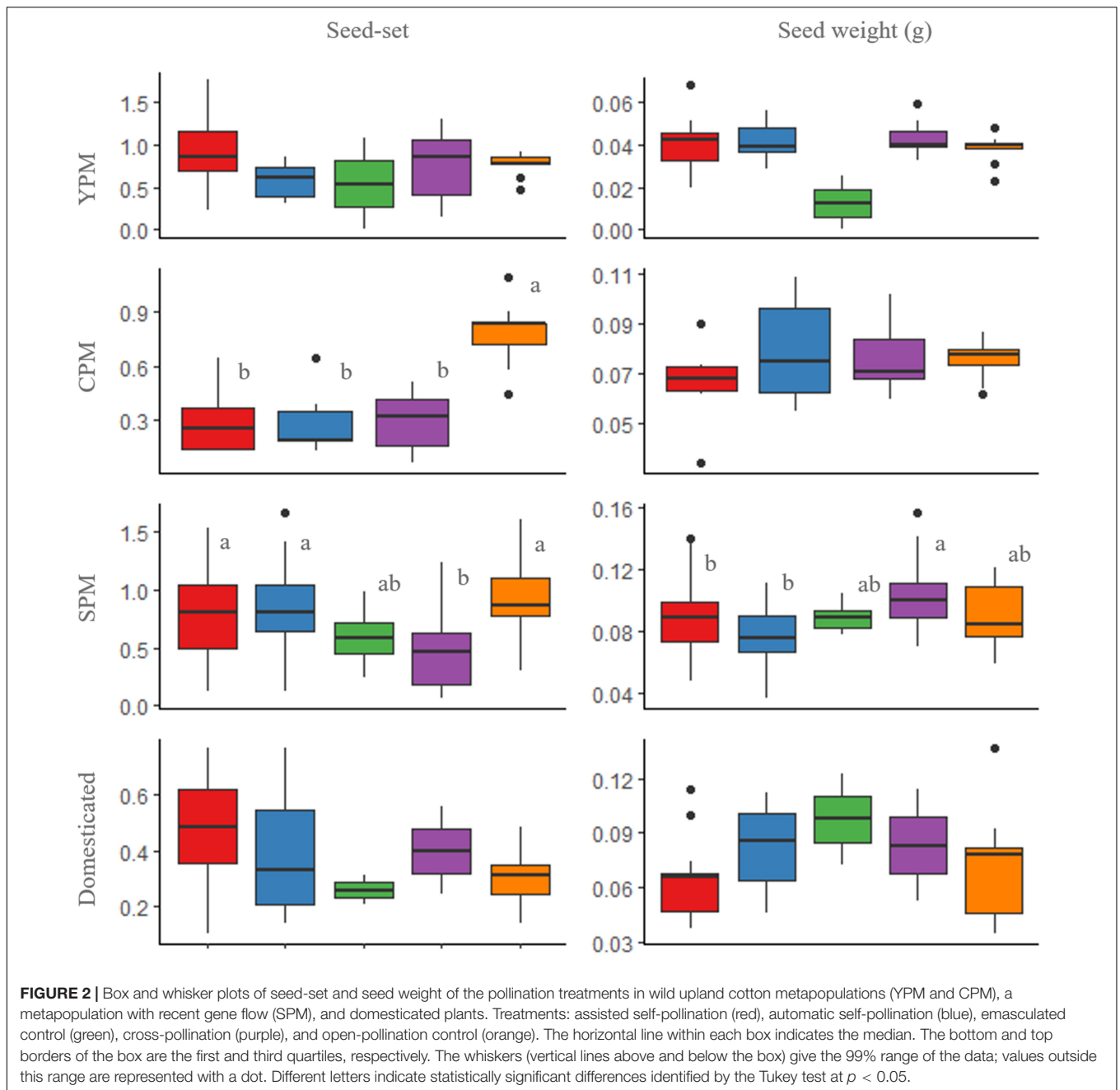
Mating System of Upland Cotton's Wild-Domesticated Complex

Richards (1997) defined autogamy as within-flower or self-pollination, and allogamy as the pollination between pollen and ovules of different flowers; moreover, he further divided

allogamy into geitonogamy (i.e., pollination between different flowers on the same genet) and xenogamy (i.e., pollination between pollen and ovules of different genets). Our results show that wild and domesticated cotton produce offspring in all pollination treatments (Figures 2, 4B and Tables 2, 3); thus, the analyzed plants have the capacity to produce progeny by both autogamy and xenogamy. To discard geitonogamy, it is necessary to perform a molecular genetic analysis of paternity. However, since autogamy is common in our system, there is no need to discard this type of allogamy. Furthermore, previous studies (see Supplementary Material 1), together with our own, indicate that the *G. hirsutum* wild-domesticated complex has a mixed mating system. This result is particularly relevant in upland cotton's center of origin, because of its significance on strategies for long-term conservation of genetic diversity in the event of gene flow between wild and domesticated relatives (Ellstrand, 1992).

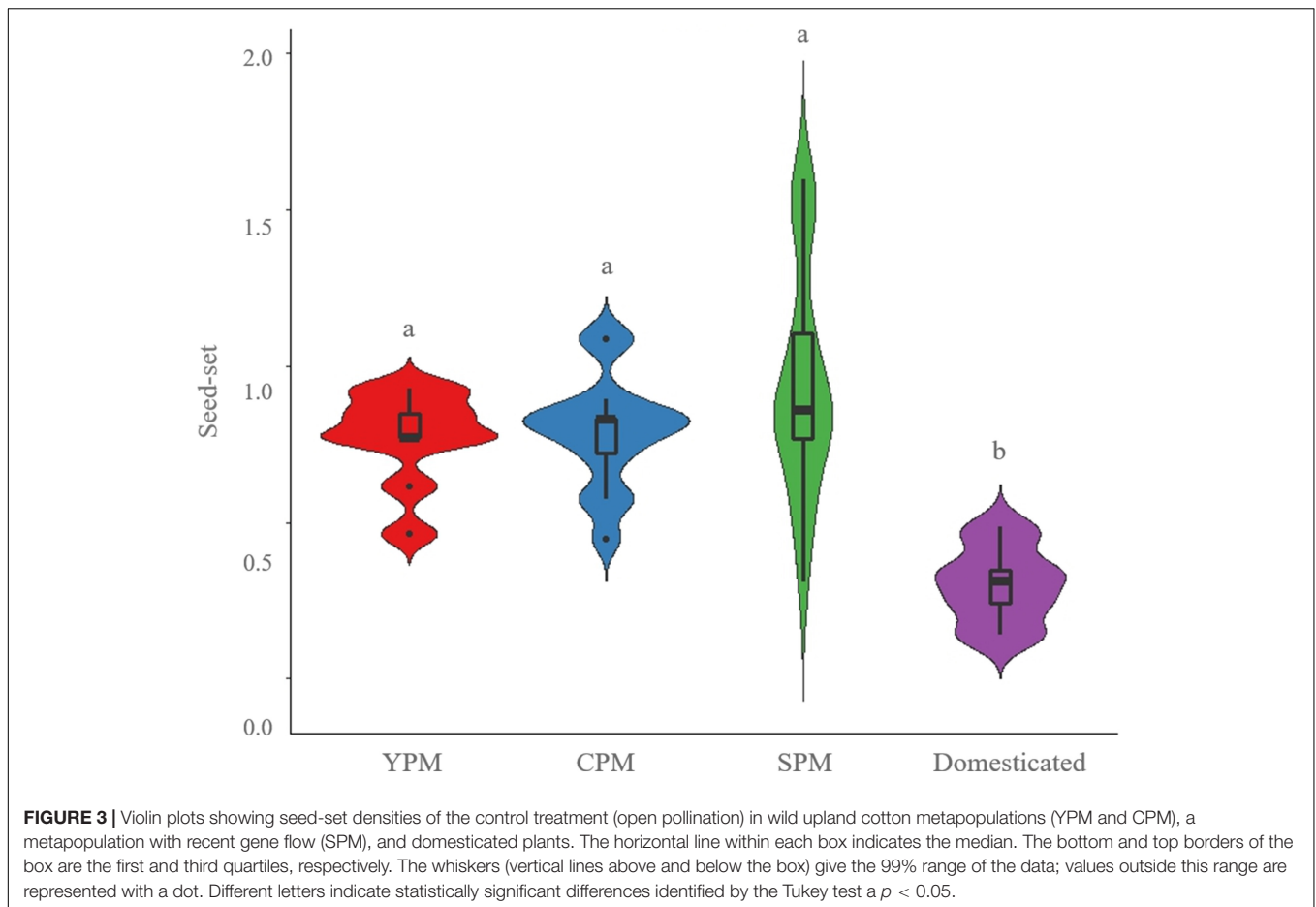
Barrett and Eckert (1990) and Barrett et al. (1996) described the outcrossing rate (Te), which indicates that when the value is 0.5 the mating system is equally balanced between self and cross-pollination. Any value different from 0 (completely self-pollinated) or 1 (completely cross-pollinated) implies a mixed mating system; when $Te > 0.5$, the system is predominantly allogamous-xenogamous, whereas when $Te < 0.5$, the system is predominantly autogamous. Our observed rates vary from $Te > 0.5$, e.g., 0.71 (CPM), 0.72 (YPM) and 0.65 (domesticated), to $Te < 0.5$, e.g., 0.40 (SPM). Domesticated plants, and wild CPM and YPM, have a greater contribution of seeds from cross-pollination in the next generation, although the contribution of self-pollination is high and important, and it contributed to maintenance of genetic structure. The high contribution of self-pollinated seeds in SPM is striking, far from being similar or intermediate between wild and domesticated; local factors may be affecting the result and should be addressed in a future study.

To further explore the mixed mating system of the species, we compared the germination rate of seeds produced by different pollination treatments. We found that within the first 5 days the seeds for all treatments reach 50% of germination. After the 10th day, a notorious difference on germination rate (<15%) among autogamy and the other treatments is observed. Such discrepancy is due to the difference in number of seeds produced in each treatment (Figure 4B). As suggested theoretically, when germination does not differ among treatments, self-pollination



is not the cause of inbreeding depression (Charlesworth and Willis, 2009). The mating system described in our study coincides with Baker's law of reproductive assurance (Pannell and Spencer, 1998), where species that migrate long distances colonize or recolonize patches initially by self-fertilization; then, because of its perennial nature, generations overlap in the same area and plants are pollinated by close relatives or by themselves in the absence of pollinators (Kalisz et al., 2004). The information described here, agrees with the ecological and genetic evidence that describes the metapopulation dynamics of *G. hirsutum*, along with the ability to migrate long distances, historically and currently (Wegier et al., 2011).

In addition, Wegier et al. (2011) reported high values of gene flow among metapopulations in the same study area, which could homogenize genetic variation, but their data exhibit population structure ($k = 8$) and high F_{ST} . Self-pollination and cross-pollination seem to maintain the genetic diversity of the species in the wild, although crossings with domesticated members of the complex (Wendel et al., 1992; Wegier et al., 2011) or even domesticated plants of *Gossypium barbadense* (Brubaker et al., 1993; Brubaker and Wendel, 1994; Ellstrand et al., 1999; Ellstrand, 2014) might be contributing to these results. In addition, gene flow with feral cotton can also take place (Rache Cardenal et al., 2013; de Menezes et al., 2015).



Finally, one of the fitness components measured in plants is seed weight (Primack and Kang, 1989), due to the fact that larger seeds perform better because of the higher amount of resources they possess (Armstrong and Westoby, 1993; Westoby et al., 1996). In our research, seed weight showed no significant difference between treatments within metapopulations (Figure 2).

Differences of Reproductive Traits Within *G. hirsutum*'s Wild to Domesticated Complex

Our analyses show differences in characters linked to some of the reproductive structures of upland cotton, which can be associated with the domestication syndrome and will be discussed below.

Ovule Number

There are significant differences in ovule number [$P(\chi^2) = 0.001$, $df = 2$], which was initially estimated to obtain the seed-set in each population (Supplementary Material 2). Wild plants produce on average 14.5 ovules per flower, while cultivated plants produce twice as many. Several authors have described a change in ovule number as a consequence of evolutionary processes. For instance, Pasquet (1998) found that ovule number supports the physiological division of cultivated cowpeas [*Vigna*

unguiculata (L.) Walp.] into two different groups: cultivars able to flower early under inductive conditions, with ovule number lower than 17 (*Biflora* and *Melanophthalmus*) and cultivars not able to do so, with ovule number higher than 17 (*Unguiculata* and *Sesquipedalis*). Moreover, Andargie et al. (2014) reported a pair of quantitative trait loci (QTLs; *qon1* and *qon3*) that regulate ovule number in cowpea; the alleles from the wild parent increase this trait as opposed to the cultivated, which reveals a feature of cowpea's domestication syndrome. In the case of climbing common bean (*Phaseolus vulgaris* L.), among the changes that occurred during the domestication process is the modification on the number of ovules, which changed from 5–8 to 2–9 ovules (Gepts and Debouck, 1991).

Seed-Set

As shown in Figure 3, there are significant differences in the seed-set of wild and domesticated populations. From a much larger number of ovules, domesticated plants (open pollination controls) produce, proportionally, a lower quantity of seeds, which implies that they are not efficiently using the resources invested on ovule production (Cilas et al., 2010). Variation in seed number per boll is produced by the interplay of the plant genetics and the environment, which in turn generates either the lack of seed fertilization or completion of embryo

TABLE 3 | Germination of seeds obtained from each upland cotton metapopulation and treatment.

Metapopulation	Traits	Germinated	Not germinated	Percentage of germination per treatment	Percentage of germination per metapopulation
CPM+ $\chi^2 = 2.07$, $df = 3$, $P = 0.55$	Assisted self-pollination	2	26	7.14	
	Automatic self-pollination	4	28	12.50	11.95
	Cross pollination	2	30	6.25	
	Open-pollination: control	25	159	13.59	
YPM $\chi^2 = 6.68$, $df = 4$, $p = 0.15$	Assisted self-pollination	35	168	17.24	
	Automatic self-pollination	7	45	13.46	13.45
	Cross pollination	13	126	9.35	
	Open-pollination: Control	12	78	13.33	
	Emasculated control	0	14	0.00	
SPM $\chi^2 = 19.29$, $df = 4$, $p = 0.0006$	Assisted self-pollination	393	45	89.73	
	Automatic self-pollination	298	60*	83.24	94.12
	Cross pollination	149	6*	96.13	
	Open-pollination: Control	292	44	86.90	
	Emasculated control	150	25	85.71	
Domesticated $\chi^2 = 23.68$, $df = 4$, $p = 9.24 \times 10^{-5}$	Assisted self-pollination	101	114	46.98	
	Automatic self-pollination	145	84*	63.32	53.57
	Cross pollination	10*	26*	27.78	
	Open-pollination: control	53	40	56.99	
	Emasculated control	6	9	40.00	

+All the repetitions of the emasculated control were placed, but none of them were found when revisiting the metapopulation for fruit collection. *Categories significantly different using adjusted standardized residuals greater than 2.0 and less than -2.0 (χ^2 test).

growth post-fertilization (Davidonis et al., 1996); therefore, our results are influenced by the experimental design and, in the future, a common garden experiment will provide insight into the effect of the environment. In comparison, wild plants are more efficient, producing seeds from nearly all of their ovules, although the net number of seeds is smaller than that produced by domesticated fruits. Many features associated with domestication are not advantageous in terms of reproduction and survival of following generations lacking human intervention (Gepts, 2004), because the selective pressures by which they have evolved are determined by humans (see categories 1–4 of the classification proposed by Clement, 1999). As a result, gene flow between wild relatives and cultivated plants could have negative consequences (Andersson and de Vicente, 2010), however, it could also give rise to *in situ* reservoirs of domesticated genes for the future (Ellstrand, 2018). Each domesticated cotton plant develops 50% more descendant plants than the wild plants do within their natural distribution, so the ecological-evolutionary consequences of this result will depend on the evolutionary process and the agro-ecological or ecosystem context in which plants are developed.

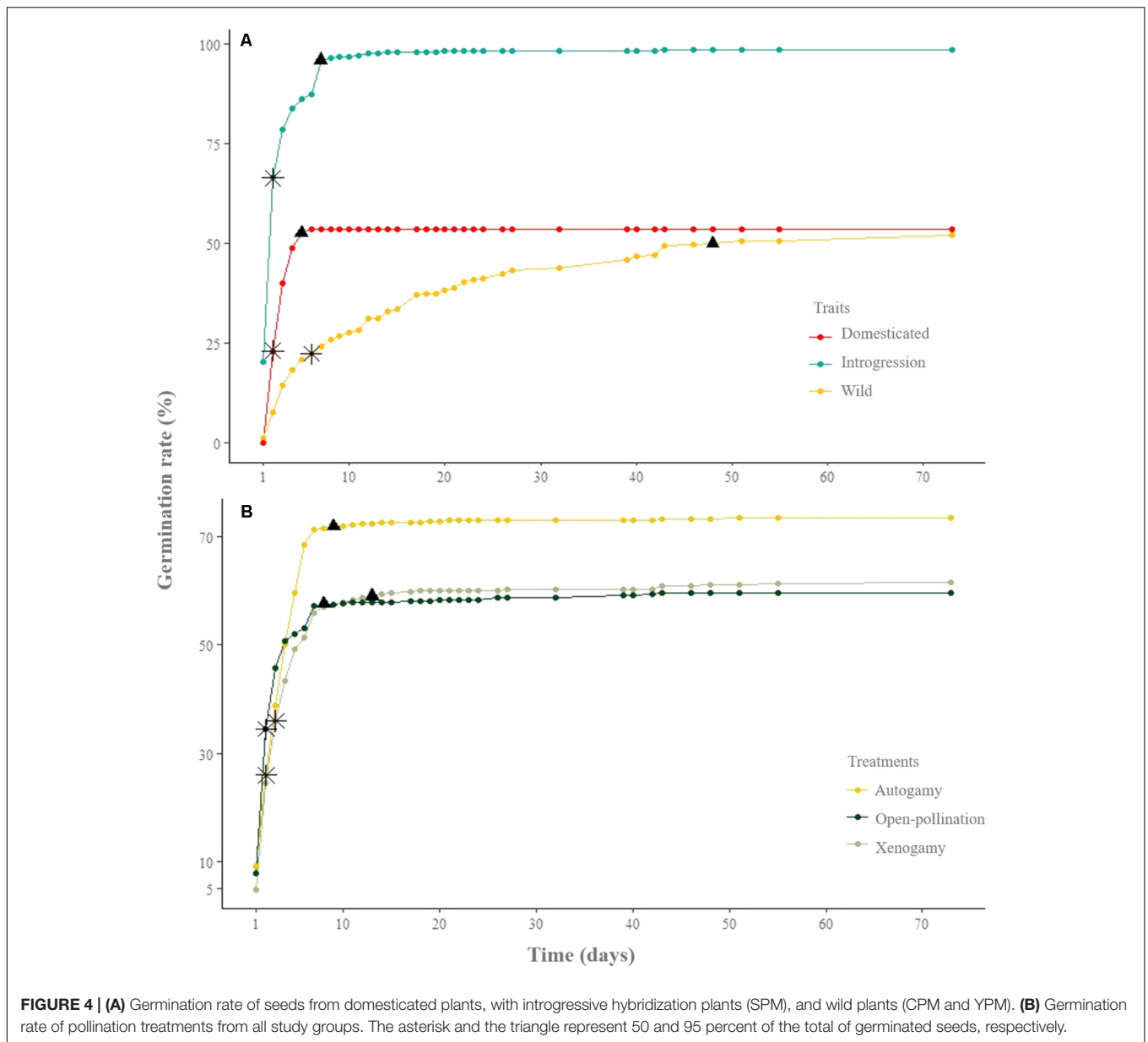
Germination

One of the traits selected for during domestication is rapid germination (Frary and Doganlar, 2003), as this helps crops to start to grow at the same time and contributes to synchronous fruiting. Over time, this trait contributes to harvesting efforts and, therefore, unconsciously selects for loss of dormancy. In natural habitats, conditions are less predictable, and dormancy will contribute to different seeds germinating in different environmental conditions (Long et al., 2015). Our results on seeds

that reached germination agree with what has been described for other domesticated plants that have undergone similar evolutionary processes (Fuller and Allaby, 2009; Abbo et al., 2014; Hernández et al., 2017): domesticated seeds germinate faster and practically simultaneously, whereas their wild relatives display dormancy (Figure 4A).

Distinctive Traits of SPM

With respect to SPM (selected for study because of evidence of recent introgression with domesticated plants; Wegier et al., 2011), the reproductive system is mixed, as it is in wild populations without introgression and in domesticated populations. However, some of the traits that determine reproductive success are unique to this population: the variability in seed-set values is markedly different (Figures 2, 3); its fruits produce more seeds than the other populations (similar to domesticated fruits, but from half the number of ovules, which makes them very efficient) (Supplementary Material 2); and these seeds have a higher percentage of germination than the other populations (Table 3). These characteristics can have demographic consequences in the short term, unless there are other factors that regulate this growth. On the other hand, contrary to what was expected for SPM, their resemblance to domesticated seed germination is higher than with the wild ones. The behavior is also dissimilar for introgressed seeds, which took longer to complete germination than domesticated and wild seeds (Figure 4A). This last phase displays a very slow response in SPM, probably associated with the loss of physiological responses, resembling domesticated plants. It is important that these analyses are repeated in subsequent years, to confirm if there is an eco-genetic trend (Price and Waser, 1979;



Edmands, 2007; Ellstrand and Rieseberg, 2016), or if it was the result of local conditions.

Conservation and Biosecurity Implications

Many nations want to defend the rights of the next generations to enjoy and decide about biodiversity and its services, aware that decisions made today will have an impact on the natural resources available in the future (Bennett et al., 2015; Steffen et al., 2015; Morales et al., 2017). Upland cotton is a remarkably important plant for humanity, not only due to the versatile uses of its fiber, but for many other applications (Wegier et al., 2016). It follows that cotton's wild-to-domesticated complex and its environment should be a conservation priority. Mesoamerican

dry forests and coastal dunes contain the ecosystems and evolutionary processes that originated, mold, and maintain wild cotton diversity and its interactions. These evolutionary services (Faith et al., 2010; Bailey, 2011; Rudman et al., 2017) are essential for species conservation, because preserving this genetic diversity allows the capacity to adapt to environmental changes (Ellstrand, 1992; Hartl, 2000). However, the factors that mold each part of the wild-to-domesticated complex are different; for example, the conservation of native traditional varieties depends to a great extent on the communities that cultivate them, their management techniques, and interests (Zhang et al., 2007). Hence, the parts of the complex that could be used for crop improvement will depend on the objectives of the new processes of domestication and breeding (Ellstrand, 2018; Mastretta-Yanes et al., 2018).

Gene flow between crops and wild relatives should be examined on a case-by-case basis (Stewart et al., 2003), especially when genetically modified organisms (GMO) are involved, because the consequences depend on the nature of the transferred genes and their regulatory mechanisms (Ellstrand, 2003). For instance, a recent study has demonstrated that genetic modifications can affect fitness traits in the long-term (Hernández-Terán et al., 2017). An important issue to keep in mind is that for gene flow to occur, a crop must be within pollination distance of a compatible population (Ellstrand and Hoffman, 1990), but in the case of domesticated plants the distances can be shortened by human activities (Dyer et al., 2009; Wegier et al., 2011). Several studies have documented hybridization events between crops and their wild relatives; for instance, in the United Kingdom, one-third of the 36 species analyzed by Raybould and Gray (1993) hybridize with at least one element of the local flora; in the Netherlands, a quarter of 42 species does (de Vries et al., 1992); and all but one of the 13 crops reviewed by Ellstrand et al. (1999) hybridize naturally with their wild relatives in some part of their agricultural distribution (including *G. hirsutum* and other species of subgenus *Karpas*). These hybridization events could lead to a decline in wild genetic diversity, as opposed to native semi-domesticated varieties in traditional Mesoamerican systems where there is evidence that domesticated genomes have formed not only by selection under domestication, but also by gene flow with other closely related populations and species (Rendón-Anaya et al., 2017). For this reason, the wild-to-domesticated dynamics in terms of genetic diversity, reproductive biology, and gene flow should be well understood in the natural distribution of the species of interest, because extrapolating conclusions based on external or incomplete information about species complexes is inconsistent with the objectives of conservation and biosafety (Beebe et al., 1997; Acevedo et al., 2016).

In this study we found that the reproductive capacity of introgressed cotton is greater than that of wild and domesticated plants. This reveals a scenario that de Wet (1968), de Wet and Harlan (1975) and Keeler et al. (1996) had already described, where wild relatives of some introgressed crops can become weeds that are difficult to control. The wide genetic diversity of *G. hirsutum*, along with factors modified by traditional genetic improvement and modern genetic engineering, will be problematic for agroecosystems (Altieri, 2000) and ecosystem conservation if they increase cotton's weediness or invasiveness (Schafer et al., 2011). Cotton has already been reported to persist in a few tropical regions, such as the north of Australia, Vietnam, México, the continental United States and Hawaii (Hawkins et al., 2005; Andersson and de Vicente, 2010; USDA, 2018), so it will be necessary to monitor these changes in wild populations given the species great capacity for long distance migration by natural and anthropogenic means.

Finally, local conditions can influence the results of reproductive biology studies (Ellstrand and Foster, 1983; Hucl, 1996; Murray et al., 2002); hence, it was essential to assess the mating system of *G. hirsutum* within its natural distribution. Some of the factors that have an effect on the results can be associated with the environment (pollen viability,

nectar production, and pollinator activity due to environmental conditions; Ahrent and Caviness, 1994; Ibarra-Pérez et al., 1997; Chaves-Barrantes et al., 2014), ecological interactions (foraging rate, floral consistency, efficiency of pollen deposition, interactions with arthropodofauna, and composition of pollinator species; Rudgers, 2004; Kessler et al., 2012; Johnson et al., 2015), as well as the landscape (species abundance and surrounding species distributions; Murray et al., 2002). In this study, the results of automatic self-pollination and the emasculated control provide evidence that autogamy and allogamy occur naturally in upland cotton's natural distribution. The occurrence of the latter highlights the importance of native pollinators on the reproductive biology of *G. hirsutum* and, consequently, conservation strategies should take this key interaction into consideration.

CONCLUSION

This study found that upland cotton's wild-to-domesticated complex presents a mixed mating system. This information is new for wild, domesticated, and introgressed *G. hirsutum* in its natural distribution, but it is in agreement with previous studies in populations of domesticated cotton (Supplementary Material 3). Consequently, *G. hirsutum* should be considered as having a mixed reproductive strategy throughout its whole complex, rather than being primarily autogamous. Management strategies and policies meant to conserve the diversity of cotton's wild-to-domesticated complex must take this into account.

Furthermore, physiological differences were found between cultivated cotton and its wild relatives, especially in traits such as the number of ovules per flower, number of viable seeds per fruit, and their germination behavior. Given the evidence of gene flow and introgression, these traits should be monitored systematically in wild populations and agroecosystems of interest for conservation, as well as the impact on ecological interactions, such as pollination. On the other hand, the diversity contained in the wild-to-domesticated complex must be included in long-term conservation strategies, so that future generations can have access to genetic resources with greater chances of surviving the changing environments.

AUTHOR CONTRIBUTIONS

AW, VA, and RV-L designed the research, participated in fieldwork, performed the analyses, and wrote the manuscript. AW coordinated the study. WK designed the fieldwork and revised the analyses. AP-M participated in fieldwork and performed the analyses. JP-L, VV-B, and DA-L conducted the analyses. All authors analyzed the results and wrote the manuscript.

FUNDING

This work was financially supported by the project "Program for the conservation of wild populations of *Gossypium hirsutum*

in Mexico”, DGAP003/WN003/18 funded by the DGSPNR (*Dirección General del Sector Primario y Recursos Naturales Renovables*) that belongs to SEMARNAT and CONABIO; complemented with the support of CONACYT scholarships (213557, 609346) and projects CONACYT-PN247672, UNAM PAPIIT No. IV200117.

ACKNOWLEDGMENTS

We are grateful for the support of all the people in the communities where our work was done. We express our gratitude to the reviewers, whose comments and suggestions greatly improved our manuscript, and especially thank Charles R. Clement for his recommendations and advice. Also, we wish to thank Mariana Benítez, Alejandro Correa-Metrio, Ana Elena Escalante, Ulises Rosas, and Kristy Walker for their valuable comments on our work. In addition, we would like to acknowledge the support of the colleagues who assisted with sampling and laboratory work: Néstor Chavarría, José Luis Caldú,

Erick Tovar, Atsiry López, Adriana Uscanga, Cirene Gutiérrez, Luis Barba, Adriana Calderón, Lislie Solis, Florencia García-Campusano, Haven López-Sánchez, Brian Urbano, and especially Diana Peña. Pamela Rodríguez collaborated with the historical analysis. We would also like to thank Professor Zenón Cano for his teachings that have contributed to our professional development through generations. DA-L is carrying out a postdoctorate funded by CONACYT-PN247672. JP-L and VV-B thank CONACYT for grants 609346 and 477713, respectively as well as the Maestría en Ciencias Biológicas, UNAM. Finally, we are especially grateful to Nancy Corona and the CARB-CONABIO team, who have provided support and wisdom through our entire journey.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2018.00574/full#supplementary-material>

REFERENCES

- Abbo, S., Pinhasi van-Oss, R., Gopher, A., Saranga, Y., Ofner, I., and Peleg, Z. (2014). Plant domestication versus crop evolution: a conceptual framework for cereals and grain legumes. *Trends Plant Sci.* 19, 351–360. doi: 10.1016/j.tplants.2013.12.002
- Acevedo, F., Huerta, E., and Burgeff, C. (2016). “Biosafety and environmental releases of GM crops in Mesoamerica: context does matter,” in *Ethnobotany of Mexico: Interactions of People and Plants in Mesoamerica*, eds R. Lira, A. Casas, and J. Blancas (New York, NY: Springer), 491–505.
- Ahrent, D. K., and Caviness, C. E. (1994). Natural cross-pollination of twelve soybean cultivars in Arkansas. *Crop Sci.* 34, 376–378. doi: 10.2135/cropsci1994.0011183X003400020013x
- Altieri, M. A. (2000). The ecological impacts of transgenic crops on agroecosystem health. *Ecosyst. Health* 6, 13–23.
- Andargie, M., Pasquet, R. S., Gowda, B. S., Muluvi, G. M., and Timko, M. P. (2014). Molecular mapping of QTLs for domestication-related traits in cowpea (*V. unguiculata* (L.) Walp.). *Euphytica* 200, 401–412. doi: 10.1007/s10681-014-1170-9
- Andersson, M. S., and de Vicente, M. C. (2010). *Gene Flow between Crops and their Wild Relatives*. Baltimore, MD: John Hopkins University Press.
- Armstrong, D. P., and Westoby, M. (1993). Seedlings from large seeds tolerated defoliation better: a test using phylogenetically independent contrasts. *Ecology* 74, 1092–1100. doi: 10.2307/1940479
- Bailey, J. K. (2011). From genes to ecosystems: a genetic basis to ecosystem services. *Popul. Ecol.* 53, 47–52. doi: 10.1007/s10144-010-0251-4
- Barrett, S. C. H., and Eckert, C. G. (1990). “Variation and evolution of mating systems in seed plants,” in *Biological Approaches and Evolutionary Trends in Plants*, ed. S. Kawano (Tokyo: Academic Press), 229–254. doi: 10.1016/B978-0-12-402960-6.50019-6
- Barrett, S. C. H., and Harder, L. D. (1996). Ecology and evolution of plant mating. *Trends Ecol. Evol.* 11, 73–79. doi: 10.1016/0169-5347(96)81046-9
- Barrett, S. C. H., Harder, L. D., and Worley, A. C. (1996). The comparative biology of pollination and mating in flowering plants. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 351, 1271–1280. doi: 10.1098/rstb.1996.0110
- Bauer-Panskus, A., Breckling, B., Hamberger, S., and Then, C. (2013). Cultivation-independent establishment of genetically engineered plants in natural populations: current evidence and implications for EU regulation. *Environ. Sci. Eur.* 25:34. doi: 10.1186/2190-4715-25-34
- Beebe, S., Toro Ch, O., González, A. V., Chacón, M. I., and Debouck, D. G. (1997). Wild-weed-crop complexes of common bean (*Phaseolus vulgaris* L., Fabaceae) in the Andes of Peru and Colombia, and their implications for conservation and breeding. *Genet. Resour. Crop Evol.* 44, 73–91. doi: 10.1023/a:1008621632680
- Bennett, E. M., Cramer, W., Begossi, A., Cundill, G., Díaz, S., Egoh, B. N., et al. (2015). Linking biodiversity, ecosystem services, and human well-being: three challenges for designing research for sustainability. *Curr. Opin. Environ. Sustain.* 14, 76–85. doi: 10.1016/j.cosust.2015.03.007
- Brubaker, C. L., Koontz, J. A., and Wendel, J. F. (1993). Bidirectional cytoplasmic and nuclear introgression in the New World cottons, *Gossypium barbadense* and *G. hirsutum* (Malvaceae). *Am. J. Bot.* 80, 1203–1208. doi: 10.2307/244549
- Brubaker, C. L., and Wendel, J. F. (1994). Reevaluating the origin of domesticated cotton (*Gossypium hirsutum*; Malvaceae) using nuclear restriction fragment length polymorphisms (RFLPs). *Am. J. Bot.* 81, 1309–1326. doi: 10.2307/2445407
- Burd, M. (1994). Bateman’s principle and plant reproduction: the role of pollen limitation in fruit and seed set. *Bot. Rev.* 60, 83–139. doi: 10.1007/BF02856594
- Burgeff, C., Huerta, E., Acevedo, F., and Sarukhán, J. (2014). *How Much can GMO and Non-GMO Cultivars Coexist in a Megadiverse Country*. Available at: <http://www.agbioforum.org/v17n1/v17n1a10-burgeff.htm>
- Carvalho, F. A., and Renner, S. S. (2012). A dated phylogeny of the papaya family (Caricaceae) reveals the crop’s closest relatives and the family’s biogeographic history. *Mol. Phylogenet. Evol.* 65, 46–53. doi: 10.1016/j.ympev.2012.05.019
- Casas, A., Otero-Arnaiz, A., Pérez-Negrón, E., and Valiente-Banuet, A. (2007). *In situ* management and domestication of plants in Mesoamerica. *Ann. Bot.* 100, 1101–1115. doi: 10.1093/aob/mcm126
- Cayuela, L. (2009). *Modelos Lineales Generalizados (GLM)*. Junta de Andalucía, Granada: Centro Andaluz de Medio Ambiente. Granada: Universidad de Granada.
- Charlesworth, D. (2006). Evolution of plant breeding systems. *Curr. Biol.* 16, R726–R735. doi: 10.1016/j.cub.2006.07.068
- Charlesworth, D., and Willis, J. H. (2009). The genetics of inbreeding depression. *Nat. Rev. Genet.* 10, 783–796. doi: 10.1038/nrg2664
- Chaves-Barrantes, N. F., Araya-Villalobos, R., and Debouck, D. G. (2014). Cruzamiento natural en frijol común en Costa Rica. *Agron. Mesoam.* 25, 23–33. doi: 10.15517/am.v25i1.14490
- Cilas, C., Machado, R., and Motamayor, J.-C. (2010). Relations between several traits linked to sexual plant reproduction in *Theobroma cacao* L.: number

- of ovules per ovary, number of seeds per pod, and seed weight. *Tree Genet. Genomes* 6, 219–226. doi: 10.1007/s11295-009-0242-9
- Clement, C. R. (1999). 1492 and the loss of amazonian crop genetic resources. I. The relation between domestication and human population decline. *Econ. Bot.* 53, 188–202. doi: 10.1007/bf02866498
- R Core Team (2017). *R: A Language and Environment for Statistical Computing*. Vienna: The R Foundation for Statistical Computing.
- Dafni, A. (1992). *Pollination Ecology: A Practical Approach*. Oxford: Oxford University Press.
- Damasceno, P. C., Pereira, T. N. S., Pereira, M. G., Silva, F. F. D., Souza, M. D. M., and Nicoli, R. G. (2009). Preferential reproduction mode of hermaphrodite papaya plant (*Carica papaya* L.; Caricaceae). *Rev. Bras. Frutic.* 31, 182–189. doi: 10.1590/S0100-29452009000100025
- Davidonis, G. H., Johnson, A., Landivar, J., and Hinojosa, O. (1996). Influence of low-weight seeds and motes on the fiber properties of other cotton seeds. *Field Crops Res.* 48, 141–153. doi: 10.1016/S0378-4290(96)01018-0
- de Menezes, I. P. P., da Silva, J. O., Malafaia, G., Silveira, R. D. D., and Barroso, P. A. V. (2015). Natural hybridization between *Gossypium mustelinum* and exotic allotetraploid cotton species. *Genet. Mol. Res.* 14, 14177–14180. doi: 10.4238/2015.October.29.40
- de Vries, F. T., van der Meijden, R., and Branden-burg, W. A. (1992). Botanical files: a study of the real chances for spontaneous gene flow from cultivated plants to the wild flora of the Netherlands. *Gorteria* 1(Suppl.), 1–100.
- de Wet, J. M. J. (1968). The origin of weediness in plants. *Okla. Acad. Sci.* 47, 14–17.
- de Wet, J. M. J., and Harlan, J. R. (1975). Weeds and domesticates: evolution in the man-made habitat. *Econ. Bot.* 29, 99–108. doi: 10.1007/bf02863309
- Doebly, J. F., Gaut, B. S., and Smith, B. D. (2006). The molecular genetics of crop domestication. *Cell* 127, 1309–1321. doi: 10.1016/j.cell.2006.12.006
- Dyer, G. A., Serratos-Hernández, J. A., Perales, H. R., Gepts, P., Piñeyro-Nelson, A., Chávez, A., et al. (2009). Dispersal of transgenes through maize seed systems in Mexico. *PLoS One* 4:e5734. doi: 10.1371/journal.pone.0005734
- Edmands, S. (2007). Between a rock and a hard place: evaluating the relative risks of inbreeding and outbreeding for conservation and management. *Mol. Ecol.* 16, 463–475. doi: 10.1111/j.1365-294X.2006.03148.x
- Ellstrand, N. C. (1992). Gene flow by pollen: implications for plant conservation genetics. *Oikos* 63, 77–86. doi: 10.2307/3545517
- Ellstrand, N. C. (2003). Current knowledge of gene flow in plants: implications for transgene flow. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 358, 1163–1170. doi: 10.1098/rstb.2003.1299
- Ellstrand, N. C. (2014). Is gene flow the most important evolutionary force in plants? *Am. J. Bot.* 101, 737–753. doi: 10.3732/ajb.1400024
- Ellstrand, N. C. (2018). Does introgression of crop alleles into wild and weedy living populations create cryptic in situ germplasm banks? *Mol. Ecol.* 27, 38–40. doi: 10.1111/mec.14390
- Ellstrand, N. C., and Foster, K. W. (1983). Impact of population structure on the apparent outcrossing rate of grain sorghum (*Sorghum bicolor*). *Theor. Appl. Genet.* 66, 323–327. doi: 10.1007/bf00251167
- Ellstrand, N. C., and Hoffman, C. A. (1990). Hybridization as an avenue of escape for engineered genes. *Bioscience* 40, 438–442. doi: 10.2307/1311390
- Ellstrand, N. C., Prentice, H. C., and Hancock, J. F. (1999). Gene flow and introgression from domesticated plants into their wild relatives. *Annu. Rev. Ecol. Syst.* 30, 539–563. doi: 10.1146/annurev.ecolsys.30.1.539
- Ellstrand, N. C., and Rieseberg, L. H. (2016). When gene flow really matters: gene flow in applied evolutionary biology. *Evol. Appl.* 9, 833–836. doi: 10.1111/eva.12402
- Faith, D. P., Magallón, S., Hendry, A. P., Conti, E., Yahara, T., and Donoghue, M. J. (2010). Ecosystem services: an evolutionary perspective on the links between biodiversity and human well-being. *Curr. Opin. Environ. Sustain.* 2, 66–74. doi: 10.1016/j.cosust.2010.04.002
- FAOSTAT (2017). *FAO (Food and Agriculture Organization of the United Nations). FAOSTAT Crops Production*. Available at: <http://www.fao.org/faostat/en/#data/QC> [accessed July 2017].
- Farooq, J., Farooq, A., Riaz, M., Ahmad, S., Amin, M., and Hussain, A. (2018). Effect of pickings on germination percentage and fiber quality traits in *Gossypium hirsutum* L. *J. Nat. Fibers* 15, 273–286. doi: 10.1080/15440478.2017.1325429
- Frary, A., and Doganlar, S. (2003). Comparative genetics of crop plant domestication and evolution. *Turk. J. Agric. For.* 27, 59–69.
- Fuller, D. Q., and Allaby, R. (2009). “Seed dispersal and crop domestication: shattering, germination and seasonality in evolution under cultivation,” in *Annual Plant Reviews Fruit Development and Seed Dispersal*, Vol. 38, ed. L. Østergaard (Oxford: Wiley-Blackwell), 238–295.
- Gepts, P. (2004). “Crop domestication as a long-term selection experiment,” in *Plant Breeding Reviews*, ed. J. Janick (Oxford: John Wiley & Sons, Inc), 1–44.
- Gepts, P., and Debouck, D. G. (1991). “Origin, domestication, and evolution of the common bean (*Phaseolus vulgaris* L.),” in *Common Beans: Research for Crop Improvement*, eds A. van Schoonhoven and O. Voysest (Wallingford: Commonwealth Agricultural Bureaux International), 7–53.
- Gil, R. A., and López, M. E. (2015). Características germinativas de semillas del algodón nativo, *Gossypium* sp., de fibra verde, lila y marrón. *REBIOL* 35, 39–46.
- Gómez, R. M. L. (2004). Estimación de la capacidad germinativa y el vigor de las semillas de diomate (*Astronium graveolens* Jacq.) sometidas a diferentes tratamientos y condiciones de almacenamiento. *Rev. Fac. Nac. Agron. Medellín* 57, 2215–2227.
- Hartl, D. L. (2000). *A Primer of Population Genetics*. Sunderland, MA: Sinauer Associates.
- Hawkins, J. S., Pleasants, J., and Wendel, J. F. (2005). Identification of AFLP markers that discriminate between cultivated cotton and the Hawaiian island endemic, *Gossypium tomentosum* Nuttall ex Seeman. *Genet. Resour. Crop Evol.* 52, 1069–1078. doi: 10.1007/s10722-004-6115-z
- Hernández, F., Lindström, L. I., Parodi, E., Poverene, M., and Presotto, A. (2017). The role of domestication and maternal effects on seed traits of crop-wild sunflower hybrids (*Helianthus annuus*). *Ann. Appl. Biol.* 171, 237–251. doi: 10.1111/aab.12368
- Hernández-Montero, J. R., and Sosa, V. J. (2016). Reproductive biology of *Pachira aquatica* Aubl. (Malvaceae: Bombacoideae): a tropical tree pollinated by bats, sphingid moths and honey bees. *Plant Species Biol.* 31, 125–134. doi: 10.1111/1442-1984.12096
- Hernández-Terán, A., Wegier, A., Benítez, M., Lira, R., and Escalante, A. E. (2017). Domesticated, genetically engineered, and wild plant relatives exhibit unintended phenotypic differences: a comparative meta-analysis profiling rice, canola, maize, sunflower, and pumpkin. *Front. Plant Sci.* 8:2030. doi: 10.3389/fpls.2017.02030
- Hucl, P. (1996). Out-crossing rates for 10 Canadian spring wheat cultivars. *Can. J. Plant Sci.* 76, 423–427. doi: 10.4141/cjps96-075
- Ibarra-Pérez, F. J., Ehdaie, B., and Waines, J. G. (1997). Estimation of outcrossing rate in common bean. *Crop Sci.* 37, 60–65. doi: 10.2135/cropsci1997.0011183X003700010009x
- Imam, A. G., and Allard, R. W. (1965). Population studies in predominantly self-pollinated species. VI. Genetic variability between and within natural populations of wild oats from differing habitats in California. *Genetics* 51, 49–62.
- Ish-Am, G., Barrientos-Priego, A., Castañeda-Vildozola, S., and Gazit, S. (1999). Avocado (*Persea americana* Mill.) pollinators in its region of origin. *Rev. Chapingo Ser. Hortic.* 5, 137–143.
- Johnson, M. T., Campbell, S. A., and Barrett, S. C. (2015). Evolutionary interactions between plant reproduction and defense against herbivores. *Annu. Rev. Ecol. Syst.* 46, 191–213. doi: 10.1146/annurev-ecolsys-112414-054215
- Kalish, S., Vogler, D. W., and Hanley, K. M. (2004). Context-dependent autonomous self-fertilization yields reproductive assurance and mixed mating. *Nature* 430, 884–887. doi: 10.1038/nature02776
- Kearns, C. A., and Inouye, D. W. (1993). *Techniques for Pollination Biologists*. Boulder, CO: University Press of Colorado.
- Keeler, K. H., Turner, C. E., and Bolick, M. R. (1996). “Movement of crop transgenes into wild plants,” in *Herbicide-Resistant Crops. Agricultural, Environmental, Economic, Regulatory, and Technical Aspects*, ed. S. O. Duke (Boca Raton, FL: Lewis Publishers), 303–330.

- Kessler, D., Bhattacharya, S., Diezel, C., Rothe, E., Gase, K., Schöttner, M., et al. (2012). Unpredictability of nectar nicotine promotes outcrossing by hummingbirds in *Nicotiana attenuata*. *Plant J.* 71, 529–538. doi: 10.1111/j.1365-3113.2012.05008.x
- Loden, H. D., and Richmond, T. R. (1951). Hybrid vigor in cotton -cytogenetic aspects and practical applications. *Econ. Bot.* 5, 387–408. doi: 10.1007/BF02984805
- Long, R. L., Gorecki, M. J., Renton, M., Scott, J. K., Colville, L., Goggin, D. E., et al. (2015). The ecophysiology of seed persistence: a mechanistic view of the journey to germination or demise. *Biol. Rev. Camb. Philos. Soc.* 90, 31–59. doi: 10.1111/brv.12095
- Machado, I. C., and Sazima, M. (2008). Pollination and breeding system of *Melochia tomentosa* L. (Malvaceae), a keystone floral resource in the Brazilian Caatinga. *Flora* 203, 484–490. doi: 10.1016/j.flora.2007.09.003
- Mastretta-Yanes, A., Acevedo Gasman, F., Burgeff, C., Cano Ramírez, M., Piñero, D., and Sarukhán, J. (2018). An initiative for the study and use of genetic diversity of domesticated plants and their wild relatives. *Front. Plant Sci.* 9:209. doi: 10.3389/fpls.2018.00209
- Meade, R. M. (1918). Bee keeping may increase the cotton crop. *J. Hered.* 9, 282–285. doi: 10.1093/oxfordjournals.jhered.a111968
- Meredith, W. R., and Bridge, R. R. (1973). Natural crossing in cotton (*Gossypium hirsutum* L.) in the Delta of Mississippi. *Crop Sci.* 13, 551–552. doi: 10.2135/cropsci1973.0011183X001300050016x
- Meyer, R. S., DuVal, A. E., and Jensen, H. R. (2012). Patterns and processes in crop domestication: an historical review and quantitative analysis of 203 global food crops. *New Phytol.* 196, 29–48. doi: 10.1111/j.1469-8137.2012.04253.x
- Meyer, R. S., and Purugganan, M. D. (2013). Evolution of crop species: genetics of domestication and diversification. *Nat. Rev. Genet.* 14, 840–852. doi: 10.1038/nrg3605
- Morales, D. V., Molaes, S., and Ladio, A. H. (2017). Firewood resource management in different landscapes in NW Patagonia. *Front. Ecol. Evol.* 5:111. doi: 10.3389/fevo.2017.00111
- Murray, B. G., Morrison, I. N., Friesen, L. F., Murray, B. G., Morrison, I. N., and Friesen, L. F. (2002). Pollen-mediated gene flow in wild oat. *Weed Sci.* 50, 321–325. doi: 10.1614/0043-1745(2002)050[0321:PMGFIW]2.0.CO;2
- Pannell, J. R., and Spencer, C. H. B. (1998). Baker's law revisited: reproductive assurance in a metapopulation. *Evolution* 52, 657–668. doi: 10.2307/2411261
- Pasquet, R. S. (1998). Morphological study of cultivated cowpea *Vigna unguiculata* (L.) Walp. Importance of ovule number and definition of cv gr *Melanophthalmus*. *Agronomie* 18, 61–70. doi: 10.1139/gen-2013-0028
- Pérez-Mendoza, C., Tovar Gómez, M. D. R., Obispo Gonzalez, Q., Legorreta Padilla, F. D. J., and Ruiz Corral, J. (2016). Recursos genéticos del algodón en México: conservación ex situ, *in situ* y su utilización. *Rev. Mex. Cienc. Agric.* 7, 5–16.
- Price, M. V., and Waser, N. M. (1979). Pollen dispersal and optimal outcrossing in *Delphinium nelsoni*. *Nature* 277, 294–297. doi: 10.1038/277294a0
- Primack, R. B., and Kang, H. (1989). Measuring fitness and natural selection in wild plant populations. *Annu. Rev. Ecol. Syst.* 20, 367–396. doi: 10.1146/annurev.es.20.110189.002055
- Rache Cardenal, L. Y., Mora-Oberlander, J., and Chaparro-Giraldo, A. (2013). Study of gene flow from GM cotton (*Gossypium hirsutum*) varieties in “El Espina” (Tolima, Colombia). *Acta Biol. Colomb.* 18, 489–498.
- Raphael, J. P. A., Gazola, B., Nunes, J. G. S., Macedo, G. C., and Rosolem, C. A. (2017). Cotton germination and emergence under high diurnal temperatures. *Crop Sci.* 57, 2761–2769. doi: 10.2135/cropsci2017.03.0182
- Raybould, A. F., and Gray, A. J. (1993). Genetically modified crops and hybridization with wild relatives: a UK perspective. *J. Appl. Ecol.* 30, 199–219. doi: 10.2307/2404623
- Rendón-Anaya, M., Montero-Vargas, J. M., Saburido-Álvarez, S., Vlasova, A., Capella-Gutiérrez, S., Ordaz-Ortiz, J. J., et al. (2017). Genomic history of the origin and domestication of common bean unveils its closest sister species. *Genome Biol.* 18:60. doi: 10.1186/s13059-017-1190-6
- Richards, A. J. (1997). *Plant Breeding Systems*. London: Garland Science. doi: 10.1007/978-1-4899-3043-9
- Richmond, T. R. (1951). Procedures and methods of cotton breeding with special reference to American cultivated species. *Adv. Genet.* 4, 213–245. doi: 10.1016/S0065-2660(08)60236-9
- Rudgers, J. A. (2004). Enemies of herbivores can shape plant traits: selection in a facultative ant-plant mutualism. *Ecology* 85, 192–205. doi: 10.1890/02-0625
- Rudman, S. M., Kreitzman, M., Chan, K. M. A., and Schluter, D. (2017). Ecosystem services: rapid evolution and the provision of ecosystem services. *Trends Ecol. Evol.* 32, 403–415. doi: 10.1016/j.tree.2017.02.019
- Schafer, M. G., Ross, A. A., Londo, J. P., Burdick, C. A., Lee, E. H., Travers, S. E., et al. (2011). The establishment of genetically engineered canola populations in the U.S. *PLoS One* 6:e25736. doi: 10.1371/journal.pone.0025736
- Schemske, D. W. (1983). Breeding system and habitat effects on fitness components in three Neotropical *Costus* (Zingiberaceae). *Evolution* 37, 523–539. doi: 10.2307/2408265
- Schooper, J. B., Lambert, R. J., Vasilas, B. L., and Westgate, M. E. (1987). Plant factors controlling seed set in maize the influence of silk, pollen, and ear-leaf water status and tassel heat treatment at pollination. *Plant Physiol.* 83, 121–125. doi: 10.1104/pp.83.1.121
- Simpson, D. M. (1954). *Natural Cross-Pollination in Cotton*. Technical Bulletin 1094. Washington, DC: United States Department of Agriculture.
- Smith, C. W., and Cothren, J. T. (1999). *Cotton: Origin, History, Technology, and Production*. New York, NY: John Wiley & Sons.
- Steffen, W., Richardson, K., Rockström, J., Cornell, S. E., Fetzer, I., Bennett, E. M., et al. (2015). Planetary boundaries: guiding human development on a changing planet. *Science* 347:1259855. doi: 10.1126/science.1259855
- Stewart, C. N., Halfhill, M. D., and Warwick, S. I. (2003). Transgene introgression from genetically modified crops to their wild relatives. *Nat. Rev. Genet.* 4, 806–817. doi: 10.1038/nrg1179
- Tan, J., Wang, M., Tu, L., Nie, Y., Lin, Y., and Zhang, X. (2013). The flavonoid pathway regulates the petal colors of cotton flower. *PLoS One* 8:e72364. doi: 10.1371/journal.pone.0072364
- Tate, J. A., and Simpson, B. B. (2004). Breeding system evolution in *Tarasa* (Malvaceae) and selection for reduced pollen grain size in the polyploid species. *Am. J. Bot.* 91, 207–213. doi: 10.3732/ajb.91.2.207
- Thangaselval, T., Gailce Leo Justin, C., and Leelamathi, M. (2008). Black pepper (*Piper nigrum* L.) “The king of species” - A review. *Agric. Rev.* 29, 89–98.
- Ulloa, M., Saha, S., Jenkins, J. N., Meredith, J. W. R., McCarty, J. J. C., and Stelly, D. M. (2005). Chromosomal assignment of RFLP linkage groups harboring important QTLs on an intraspecific cotton (*Gossypium hirsutum* L.) Joinmap. *J. Hered.* 96, 132–144. doi: 10.1093/jhered/esi020
- USDA (2018). *The PLANTS Database*. Available at: <http://plants.usda.gov> [accessed February 21, 2018].
- Vaughan, D. A., Balázs, E., and Heslop-Harrison, J. S. (2007). From crop domestication to super-domestication. *Ann. Bot.* 100, 893–901. doi: 10.1093/aob/mcm224
- Viechtbauer, W., and Cheung, M. W. L. (2010). Outlier and influence diagnostics for meta-analysis. *Res. Synth. Methods* 1, 112–125. doi: 10.1002/jrsm.11
- Wäckers, F. L., and Bonifay, C. (2004). How to be sweet? Extrafloral nectar allocation by *Gossypium hirsutum* fits optimal defense theory predictions. *Ecology* 85, 1512–1518. doi: 10.1890/03-0422
- Warschewsky, E., Penmetza, R. V., Cook, D. R., and von Wettberg, E. J. B. (2014). Back to the wilds: tapping evolutionary adaptations for resilient crops through systematic hybridization with crop wild relatives. *Am. J. Bot.* 101, 1791–1800. doi: 10.3732/ajb.1400116
- Warwick, S. I., and Stewart, C. N. (2005). “Crops come from wild plants? how domestication, transgenes, and linkage together shape fertility,” in *Crop Fertility and Volunteerism*, ed. J. Gressel (Boca Raton, FL: CRC Press), 9–30.
- Wegier, A. (2013). *Diversidad genética y conservación de Gossypium hirsutum silvestre y cultivado en México*. Ph.D. thesis, Universidad Nacional Autónoma de México, Mexico City.
- Wegier, A., Alavez, V., and Piñero, D. (2016). “Cotton: traditional and modern uses,” in *Ethnobotany of Mexico: Interactions of People and Plants in Mesoamerica*, eds R. Lira, A. Casas, and J. Blancas (New York, NY: Springer-Verlag), 439–456.

- Wegier, A., Piñeyro-Nelson, A., Alarcón, J., Gálvez-Mariscal, A., Alvarez-Buylla, E. R., and Piñero, D. (2011). Recent long-distance transgene flow into wild populations conforms to historical patterns of gene flow in cotton (*Gossypium hirsutum*) at its centre of origin. *Mol. Ecol.* 20, 4182–4194. doi: 10.1111/j.1365-294X.2011.05258.x
- Wendel, J. F., Brubaker, C. L., and Percival, A. E. (1992). Genetic diversity in *Gossypium hirsutum* and the origin of upland cotton. *Am. J. Bot.* 79, 1291–1310. doi: 10.2307/2445058
- Westoby, M., Leishman, M., Lord, J., Poorter, H., and Schoen, D. J. (1996). Comparative ecology of seed size and dispersal [and discussion]. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 351, 1309–1318. doi: 10.1098/rstb.1996.0114
- Zhang, W., Ricketts, T. H., Kremen, C., Carney, K., and Swinton, S. M. (2007). Ecosystem services and dis-services to agriculture. *Ecol. Econ.* 64, 253–260. doi: 10.1016/j.ecolecon.2007.02.024
- Zohary, D., and Spiegel-Roy, P. (1975). Beginnings of fruit growing in the Old World. *Science* 187, 319–327. doi: 10.1126/science.187.4174.319
- Zuur, A. F., Ieno, E. N., Walker, N. J., Saveliev, A. A., and Smith, G. M. (eds). (2009). “GLMM and GAMM,” in *Mixed Effects Models and Extensions in Ecology with R* (New York, NY: Springer), 323–341.

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Velázquez-López, Wegier, Alavez, Pérez-López, Vázquez-Barrios, Arroyo-Lambaer, Ponce-Mendoza and Kunin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.