



Identification, Characterization, and Functional Validation of Drought-responsive MicroRNAs in Subtropical Maize Inbreds

Jayaraman Aravind^{1,2}, Sharma Rinku^{1,3}, Banduni Pooja¹, Mittal Shikha¹, Shiriga Kaliyugam¹, Mallana Gowdra Mallikarjuna¹, Arun Kumar⁴, Atmakuri Ramakrishna Rao⁵ and Thirunavukkarasu Nepolean^{1*}

¹ Division of Genetics, Indian Agricultural Research Institute, New Delhi, India, ² Division of Germplasm Conservation, National Bureau of Plant Genetic Resources, New Delhi, India, ³ Department of Life Sciences, Shiv Nadar University, Gautam Buddha Nagar, India, ⁴ National Phytotron Facility, Indian Agricultural Research Institute, New Delhi, India, ⁵ Centre for Agricultural Bioinformatics, Indian Agricultural Statistics Research Institute, New Delhi, India

OPEN ACCESS

Edited by:

Purificación Lisón,
Universitat Politècnica de València,
Spain

Reviewed by:

Hao Li,
Northwest A&F University, China
Anil Kumar Singh,
ICAR-Indian Institute of Agricultural
Biotechnology, India

*Correspondence:

Thirunavukkarasu Nepolean
tnepolean@yahoo.com;
tnepolean@gmail.com

Specialty section:

This article was submitted to
Plant Physiology,
a section of the journal
Frontiers in Plant Science

Received: 20 February 2017

Accepted: 19 May 2017

Published: 02 June 2017

Citation:

Aravind J, Rinku S, Pooja B, Shikha M, Kaliyugam S, Mallikarjuna MG, Kumar A, Rao AR and Nepolean T (2017) Identification, Characterization, and Functional Validation of Drought-responsive MicroRNAs in Subtropical Maize Inbreds. *Front. Plant Sci.* 8:941. doi: 10.3389/fpls.2017.00941

MicroRNA-mediated gene regulation plays a crucial role in controlling drought tolerance. In the present investigation, 13 drought-associated miRNA families consisting of 65 members and regulating 42 unique target mRNAs were identified from drought-associated microarray expression data in maize and were subjected to structural and functional characterization. The largest number of members (14) was found in the zma-miR166 and zma-miR395 families, with several targets. However, zma-miR160, zma-miR390, zma-miR393, and zma-miR2275 each showed a single target. Twenty-three major drought-responsive *cis*-regulatory elements were found in the upstream regions of miRNAs. Many drought-related transcription factors, such as GAMYB, HD-Zip III, and NAC, were associated with the target mRNAs. Furthermore, two contrasting subtropical maize genotypes (tolerant: HKI-1532 and sensitive: V-372) were used to understand the miRNA-assisted regulation of target mRNA under drought stress. Approximately 35 and 31% of miRNAs were up-regulated in HKI-1532 and V-372, respectively. The up-regulation of target mRNAs was as high as 14.2% in HKI-1532 but was only 2.38% in V-372. The expression patterns of miRNA-target mRNA pairs were classified into four different types: Type I- up-regulation, Type II- down-regulation, Type III- neutral regulation, and Type IV- opposite regulation. HKI-1532 displayed 46 Type I, 13 Type II, and 23 Type III patterns, whereas V-372 had mostly Type IV interactions (151). A low level of negative regulations of miRNA associated with a higher level of mRNA activity in the tolerant genotype helped to maintain crucial biological functions such as ABA signaling, the auxin response pathway, the light-responsive pathway and endosperm expression under stress conditions, thereby leading to drought tolerance. Our study identified candidate miRNAs and mRNAs operating in important pathways under drought stress conditions, and these candidates will be useful in the development of drought-tolerant maize hybrids.

Keywords: drought, gene expression, maize, miRNA, mRNA, post-transcriptional changes

INTRODUCTION

Drought is one of the prevailing abiotic stresses that affect plant growth, development and grain yield (Ceccarelli and Grando, 1996). In particular, regions with insufficient water are more prone to drought due to uneven changes in weather conditions. Furthermore, shortages of water resources resulting from increasing human needs and growing climatic adversities exaggerate the effects of drought several fold (Rosegrant and Cline, 2003). Development of climate-resilient and drought-tolerant cultivars could help in sustaining food grain production in the present era of climate change. However, drought tolerance is a morphologically, physiologically, and genetically complex trait. Therefore, understanding the underlying molecular basis and regulation of drought tolerance could help in accelerating drought-tolerance breeding programs. Many reports have highlighted and proposed various genetic and molecular mechanisms of drought tolerance in different crops, including model plant systems (Ha et al., 2012; Shikha et al., 2017). Additionally, a functional understanding of stress responsive gene(s) and their regulation patterns can aid in devising new genetic tools (Langridge and Reynolds, 2015).

Post-transcriptional modification of RNAs is one of the major forms of regulation of gene expression and is primarily performed by microRNA (miRNA) molecules (Ding et al., 2009; Zhou et al., 2010). miRNAs belong to a non-coding family of RNAs and are known to play major roles in modulating gene expression to adjust plant metabolism to withstand stresses. The regulatory mechanism of miRNAs involves a change in self-concentration, targeting the mRNA quantity and modifying the mRNA expression *via* miRNA-protein complexes. These, in turn, change the ultimate expression of proteins upon exposure to stress (Ding et al., 2009; Wang B. et al., 2014). In plants, miRNA genes code for long pri-miRNA (primary miRNA) transcripts with imperfect stem-loop secondary structures, which are transcribed by RNA polymerase II (Lee et al., 2004; Xie et al., 2005). These transcripts are processed into ~70-nt pre-miRNAs and subsequently released as miRNA/miRNA duplexes by DCL-1 (dicer-like enzyme 1) in association with a dsRNA binding protein, HYL-1. Such duplexes are methylated by a dsRNA methylase, HEN1, and loaded into AGO1 (Kurihara and Watanabe, 2004; Vazquez et al., 2004; Yu et al., 2005). They are subsequently transported to the cytoplasm with the help of an exportin homolog, the HASTY protein (Bartel, 2004; Park et al., 2005), and cleaved into ~22-nt mature miRNAs. Mature miRNA strands are incorporated into a multiprotein complex, the RNA-induced silencing complex (RISC), where they guide the cleavage of complementary target mRNAs by AGO1, which possesses a PAZ domain and a catalytic PIWI domain (Vaucheret et al., 2004; Baumberger and Baulcombe, 2005).

The involvement of miRNAs in different abiotic stresses has been demonstrated in *Arabidopsis*. Overexpression of miR168, miR171 and miR396 under hypersalinity, mannitol, and cold stress was reported in *Arabidopsis* (Liu et al., 2008). Nevertheless, both the involvement of total miRNAs under drought stress and the systemic expression analysis of their

drought-related mechanism are still in progress and necessitate further exploration.

Maize is an important crop in the world, contributing significantly to food and nutritional security. However, the production of maize is most vulnerable to various abiotic stresses, especially drought. To date, functional genomics approaches have revealed large amounts of information on target mRNA control through miRNAs for various traits in maize. The expression of a class III homeodomain-leucine zipper (*HD-Zip III*) protein that functions in asymmetrical leaf development and that of a floral meristem transcription factor, *APETALA2*, responsible for meristem identity were found to be targeted by miR166 and miR172, respectively (Juarez et al., 2004). Similarly, miR156 is reported to target the expression of *tga1* (*Teosinte* glume architecture 1) (Chuck et al., 2007). Studies on differential expression of miRNAs have shed light on the regulatory roles of miRNAs in plant development (Kang et al., 2012; Liu et al., 2014) and stress responses in maize (Zhang et al., 2008; Ding et al., 2009; Wei et al., 2009; Zhai et al., 2013; Wu et al., 2014). Such investigations deciphering the regulatory control between miRNAs and target mRNAs will pave the way for a better understanding of the molecular mechanisms underlying drought stress responses. Differentially expressed genes (DEGs) have been identified in drought or low-moisture stress microarray studies (Yu, 2003; Yue et al., 2008; Hayano-Kanashiro et al., 2009; Li et al., 2009; Marino et al., 2009; Luo et al., 2010; Zheng et al., 2010; Lu et al., 2011; Hansen et al., 2013; Regulski et al., 2013). Such DEGs were found to be associated with drought-related miRNA targets.

Maize germplasm shows great genetic variability involving temperate, tropical, and subtropical groups, as well as dent, flint, semi flint and waxy types within germplasm groups. Efforts have been taken to characterize the drought-responsive miRNAs and target mRNAs in temperate maize germplasm (Li et al., 2013; Wang Y. G. et al., 2014). There have been no reports available on comprehensive characterization of drought-responsive miRNAs and target-mRNAs using subtropical maize germplasm. Therefore, in the present study, we identified putative regulatory miRNAs targeting drought-related mRNAs based on gene expression data from 12 drought/low-moisture stress experiments. The expression patterns of miRNA-target mRNA pairs were validated in the root and shoot tissues of two contrasting subtropical maize inbreds under drought stress. The functional annotation and the role of drought-related miRNA-target mRNA pairs were analyzed. Our study identified the differential interactions of miRNA-target mRNA pairs during drought stress in the tolerant genotype and explained their functional roles in drought tolerance.

MATERIALS AND METHODS

Plant Growth and Experimentation

Seeds of two contrasting subtropical maize inbreds, HKI-1532 (drought-tolerant) and V-372 (drought-susceptible), were grown in the National Phytotron Facility, Indian Agricultural Research Institute, New Delhi. Potting was done in triplicate

with sandy loam soil. The plants were maintained under controlled greenhouse conditions of 28/22°C (day/night) at a light intensity of 600 $\mu\text{mol m}^{-2}\text{s}^{-1}$ (16 h day/8 h night) with 50–55% relative humidity. Regular irrigation was provided for 15 days to the first set of plants (stress) after sowing and suspended for the next 5 days to induce severe drought stress (Kakumanu et al., 2012; Min et al., 2016; Nepolean et al., 2017). The second set of plants (control) was watered throughout the experiment. On the 21st day after sowing, leaf samples were collected for expression assays (Figure 1).

Prediction of miRNA-target mRNA Interactions

The entire set of maize mature miRNA sequences available (321) was downloaded from miRbase v19 (Griffiths-Jones et al., 2008; Kozomara and Griffiths-Jones, 2011). Four popular miRNA target prediction tools—RNAHybrid (Rehmsmeier et al., 2004), TargetFinder (Allen et al., 2005; <http://carringtonlab.org/resources/targetfinder>), TAPIR (FASTA search engine) (Bonnet et al., 2010), and psRNATarget (Dai and Zhao, 2011)—were used to identify targets for the miRNAs from among the 76,617 *Zea mays* B73 RegGen v2 mRNAs downloaded from NCBI (*National Center for Biotechnology Information*) Genome database identified by the GNOMON v2 gene prediction tool (Souvorov et al., 2010; ftp://ftp.ncbi.nlm.nih.gov/genomes/PLANTS/Zea_mays/Gnomon_v2/). Consensus predictions of miRNA-target mRNA interactions detected among the four tools were used for further analysis.

Identification of Putative Drought-Related miRNAs

DEGs identified in 12 microarray-based gene expression studies in maize under drought/low moisture stress and/or recovery irrigation conditions (Table S1) were selected for comparison with the target mRNAs identified in the consensus predictions of miRNA-target mRNA interactions. For the experiments where DEG lists were not available, the raw data from NCBI GEO (*Gene Expression Omnibus*) (Barrett et al., 2005) were reanalyzed to generate a list. The regulatory miRNAs of the DEG sequences that were also predicted to be miRNA targets were identified as putative drought-related miRNAs. For both the mature miRNA sequences and the target mRNA sequences with the respective target sites masked, multiple sequence alignment was done using the Clustal Omega program (Sievers et al., 2011). A distance matrix was constructed based on pairwise distances using an identity matrix with the “dist.alignment” function of the R package seqinR (Charif et al., 2008) and used to performing hierarchical clustering by complete linkage. The clusterings of miRNAs and their respective targets were compared visually by plotting a tanglegram using “cophyloplot” function of the R package ape (Paradis et al., 2004). Furthermore, the functional annotation of the corresponding target mRNAs was carried out by Blast2GO (Conesa et al., 2005).

Analysis of *cis*-Regulatory Elements (CREs) in miRNA Promoters in Maize

DNA sequences upstream of the starts of the precursor miRNA structures were identified and extracted using NCBI tools (<https://www.ncbi.nlm.nih.gov>). The transcription start sites (TSS) were predicted from the upstream regions of pri-miRNA sequences using softberry TSSP (Shahmuradov et al., 2005; <http://www.softberry.com/berry.phtml?topic=tss>). The promoter sequence, i.e., the 2-kb region upstream of the TSS, was retrieved using Perl Script. For families having >1 promoter, the sequence between the two promoters was considered for the identification of CREs using the PlantCARE (<http://bioinformatics.psb.ugent.be/webtools/plantcare/html/>) database.

RNA Isolation and qRT-PCR Assay

Total RNA was isolated from seedling leaf tissue using the Ultra Clean Plant RNA isolation kit (MO BIO Laboratories, USA). Primers specific to the target mRNA sequences were designed using BatchPrimer3 (You et al., 2008) and optimized to avoid primary and amplicon secondary structures using Beacon DesignerTM Free Edition (<http://www.premierbiosoft.com/qPCR/>) and IDT OligoAnalyzer (<http://eu.idtdna.com/analyzer/Applications/OligoAnalyzer/>), respectively (Table S2). Sequence specificity of the primers was assured by comparing their sequences with maize gene sequences using NCBI nucleotide BLAST (<http://blast.ncbi.nlm.nih.gov/Blast.cgi>). qRT-PCR for mature miRNAs was performed using a modified version of stem-loop qRT-PCR (Chen et al., 2005). The stem-loop primers, miRNA forward primers and universal reverse primers were designed according to Kramer (2011) (Table S3). The primer for the reference transcript U6snRNA was designed as previously described (Turner et al., 2013).

For target mRNAs, the first-strand cDNA synthesis was carried out with the RNA ProtoScript First Strand cDNA Synthesis Kit (New England Biolabs, Ipswich, Massachusetts) using 50 ng total RNA and 1 μM Oligo d(T)₂₃ VN primer. For miRNAs, reverse transcription reactions were carried out with 50 ng total RNA and 1 μM stem-loop RT primer. The reactions were incubated in a thermocycler for 30 min at 16°C, 30 min at 42°C and 5 min at 85°C and then held at 4°C.

For both mature miRNAs and their mRNA targets, real-time quantitative PCR with SYBR Green I was performed on an Agilent Technologies Mx3005P QPCR (Agilent Technologies, Santa Clara, California, United States) instrument. Briefly, each 25- μl PCR mixture contained approximately 100 ng cDNA, 9 ml Hotstart IT SYBR Green qPCR Master Mix (Affymetrix), ROX (Affymetrix) and 250 nM of each primer. The reactions were mixed gently and incubated at 94°C for 2 min (pre-heating), followed by 40 cycles of 94°C for 15 s (denaturation), 60°C for 30 s (primer annealing), and 72°C for 30 s (primer extension). Analysis was conducted on three biological replicates and two technical replicates for each treatment and genotype combination. U6 snRNA and 18S rRNA were used as internal controls for miRNA and mRNA, respectively. The $\Delta\Delta\text{Ct}$ method was used to determine the differences in expression levels among samples (Livak and Schmittgen, 2001).

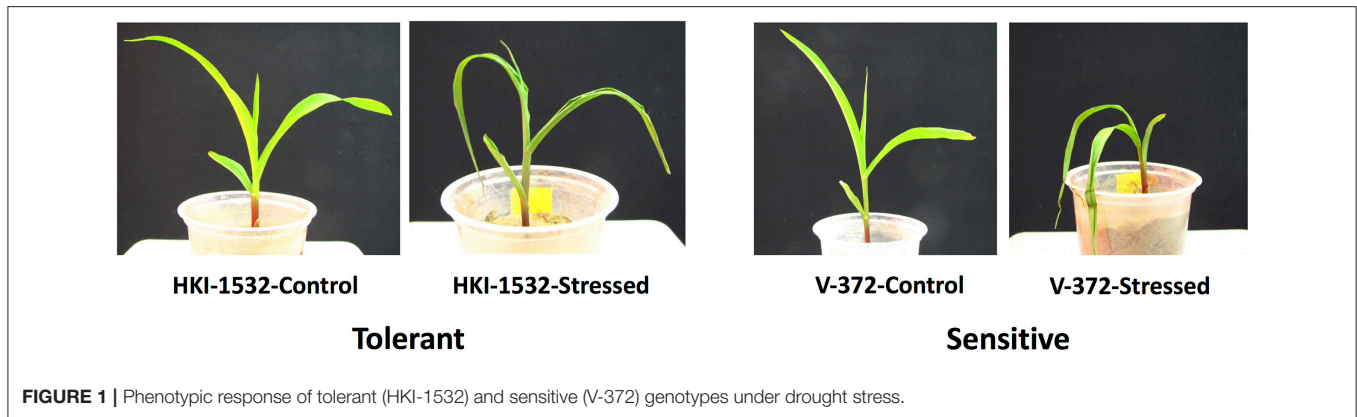


FIGURE 1 | Phenotypic response of tolerant (HKI-1532) and sensitive (V-372) genotypes under drought stress.

RESULTS

Identification of Putative Drought-Related miRNAs and Target mRNAs

Among the DEGs identified in 12 microarray-based gene expression studies under drought/low moisture stress and/or recovery irrigation conditions, 42 differentially expressed mRNA sequences were predicted to be putatively regulated by 65 miRNAs belonging to 13 families in 183 miRNA-target mRNA interactions (**Table 1**). These were considered for further expression analysis. The predicted interactions involved the same miRNAs targeting multiple mRNAs, as well as the same mRNA being targeted by multiple miRNAs. For most of the interactions, miRNAs with similar sequences targeted mRNA sequences that clustered together (**Figure 2**). The miRNA families with the largest numbers of members included zma-miR166, zma-miR395 and zma-miR156, with 14, 14, and 12 members, respectively. On the other hand, some miRNA families, such as zma-miR160, zma-miR399, zma-miR529, and zma-miR2275, had only one member each (**Figure 3**). The target distributions of miRNA families themselves were investigated, which showed that zma-miR166, zma-miR396, zma-miR529, zma-miR164, and zma-miR169 had the most targets, with 6, 6, 5, and 5 target mRNAs, respectively, while zma-miR160, zma-miR390, zma-miR393, and zma-miR2275 had the fewest target mRNAs, with a single target each (**Figure 3**).

Prediction of miRNA-Target mRNA Interactions

The four miRNA target prediction tools detected different but overlapping sets of miRNA-target mRNA pairs from among the 321 miRNAs and 76617 mRNAs of maize. Among these dissimilar sets, 594 consensus predictions of miRNA-target mRNA interactions could be identified. Compared to single prediction tool-based identification of miRNA-target mRNA interactions, we employed four prediction tools to choose the robust miRNA-target mRNA interactions. The total numbers of interactions supported by RNAHybrid, TargetFinder, TAPIR FASTA and psRNATarget were 31,262, 4,522, 3,406, and 3,072, respectively, including both unique and commonly identified interactions from each tool. The numbers of unique interactions

identified by RNAHybrid, TargetFinder, TAPIR FASTA and psRNATarget were 29869, 1264, 222, and 1293, respectively (**Figure 4**). The consensus predictions of miRNA-target mRNAs interactions involved 156 miRNAs belonging to 25 families, and 150 unique target mRNAs.

GO Enrichment Analysis of miRNA Target Genes

To gain insights into the functional roles of miRNAs, annotation of their predicted target mRNAs was carried out. GO terms were identified under three different functional categories: biological (57.14%), cellular (23.80%) and molecular (30.95%). In the biological process category, major functions included anatomical structure morphogenesis, multicellular organismal development and post-embryonic development. In the molecular process category, major functions included DNA binding, hydrolase activity and nucleotide binding (**Figure 5**). Furthermore, most of the target genes were confined to the nucleus.

Analysis of *cis*-Regulatory Elements

The promoter regions of 65 drought-related miRNAs were examined to detect the occurrence of conserved *cis*-regulatory elements. A total of 61 conserved CRE motifs were identified from the miRNA promoters. Among these, 41 were major *cis*-regulatory elements that were observed in >10 members, including the drought-related *cis*-elements ABRE (ABA-responsive element) and MBS (MYB-binding site) (**Table 2**). In miR169n of rice, the ABRE *cis*-acting element resided in the promoter region, implying an ABA-mediated response to stress (Zhao et al., 2007). An ABRE *cis*-element was also found in sorghum ABA-responsive genes (Buchanan et al., 2005). Similarly, in *Arabidopsis*, MBS was found in the upstream regions of all the miRNA genes in the rice shoot apices. Of the 65 miRNA members, miR164f showed the highest number of CREs (41), whereas miR156d had the lowest number (8).

Expression Patterns of miRNAs and Their Target mRNA(s)

To analyze the responses of contrasting genotypes to drought, transcriptional profiling of the identified putative drought-responsive miRNAs and their target-mRNAs was done at the

TABLE 1 | List of 13 drought-related miRNA families and their respective targets with annotations.

Family specific target mRNAs			
miRNA family	miRNA	Target mRNAs	Annotations
miR156	zma-miR156-a,c,d,e,f,g,h,i,j,k,l	gnl GNOMON 13750094.m	tpa: squamosa promoter-binding (sbp domain) transcription factor family protein
	zma-miR156-a,c,d,e,f,g,h,i,j,k,l	gnl GNOMON 66064033.m	Squamosa promoter-binding-like protein 13-like
miR159	zma-miR159e-3p	gnl GNOMON 11944063.m	Transcription factor gamyb
	zma-miR159e-5p	gnl GNOMON 22442014.m	Transcription factor gamyb
	zma-miR159-c,d,g,h,i	gnl GNOMON 57248043.m	Transcription factor
miR160	zma-miR160-f	gnl GNOMON 46030063.m	gdsI esterase lipase at5g45910-like
miR164	zma-miR164-f	gnl GNOMON 10380054.m	Hypothetical protein
	zma-miR164-f	gnl GNOMON 10384054.m	Hypothetical protein
	zma-miR164-f	gnl GNOMON 18192014.m	Hypothetical protein
	zma-miR164-b	gnl GNOMON 4218083.m	psbp domain-containing protein Chloroplastic-like
	zma-miR164-h	gnl GNOMON 46106013.m	Wound responsive protein
miR166	zma-miR166-a,b,c,d,e,f,g,h,i,j,k,l	gnl GNOMON 1168013.m	Rolled expressed
	zma-miR166-a,b,c,d,e,f,g,h,i,j,l	gnl GNOMON 15104054.m	Partial
	zma-miR166-a,b,c,d,e,f,g,h,i,j,k,m,n,l	gnl GNOMON 35860043.m	Tpa: homeobox lipid-binding domain family protein
	zma-miR166-a,b,c,d,e,f,g,h,i,j,k,m,n,l	gnl GNOMON 52446103.m	Rolled leaf1
	zma-miR166-a,b,c,d,e,f,g,h,i,j,k,m,n,l	gnl GNOMON 54238013.m	Tpa: homeobox lipid-binding domain family protein
	zma-miR166-a,b,c,d,e,f,g,h,i,j,k,m,n,l	gnl GNOMON 8472093.m	Homeobox-leucine zipper protein athb-15-like
miR169	zma-miR169-l,j,k,l	gnl GNOMON 74364063.m	Nuclear transcription factor y subunit a-3
	zma-miR169-l,j,k,l	gnl GNOMON 74366063.m	Nuclear transcription factor y subunit a-3
miR390	zma-miR390-a,b	gnl GNOMON 30954063.m	Activator of 90 kda heat shock protein atpase
miR393	zma-miR393-a,c	gnl GNOMON 39086093.m	Protein transport inhibitor response 1-like
	zma-miR393-a,c	gnl GNOMON 5722063.m	Atpsulfurylase
	zma-miR393-a,c	gnl GNOMON 92168013.m	Atpsulfurylase
miR395	zma-miR395-b,d,e,f,g,h,i,j,m,n,p,l	gnl GNOMON 5722063.m	Atpsulfurylase
		gnl GNOMON 92168013.m	Atpsulfurylase
miR396	zma-miR396-c,d	gnl GNOMON 3258103.m	Growth-regulating factor 1
	zma-miR396-c,d	gnl GNOMON 41140073.m	Growth-regulating factor 1
	zma-miR396-c,d	gnl GNOMON 54764053.m	Growth-regulating factor 1-like
	zma-miR396-c,d	gnl GNOMON 74420063.m	Growth-regulating factor 9
	zma-miR396-c,d	gnl GNOMON 8836063.m	Growth-regulating factor 8
	zma-miR396-c,d	gnl GNOMON 9726103.m	Growth-regulating factor
miR399	zma-miR399-e	gnl GNOMON 61030013.m	60s ribosomal protein l7a-like
	zma-miR399-e	gnl GNOMON 92930013.m	Heavy metal-associated domain containing expressed
miR529	zma-miR529	gnl GNOMON 24048063.m	Squamosa promoter-binding (sbp domain) transcription factor family protein isoform 1
	zma-miR529	gnl GNOMON 24052063.m	Squamosa promoter-binding (sbp domain) transcription factor family protein isoform 1
	zma-miR529	gnl GNOMON 24056063.m	Squamosa promoter-binding (sbp domain) transcription factor family protein isoform 1
	zma-miR529	gnl GNOMON 86696013.m	p8mtcp1
	zma-miR529	gnl GNOMON 86698013.m	p8mtcp1
	zma-miR529	gnl GNOMON 86706013.m	p8mtcp1
miR2275	zma-miR2275-a	gnl GNOMON 55702013.m	Mitochondrial protein

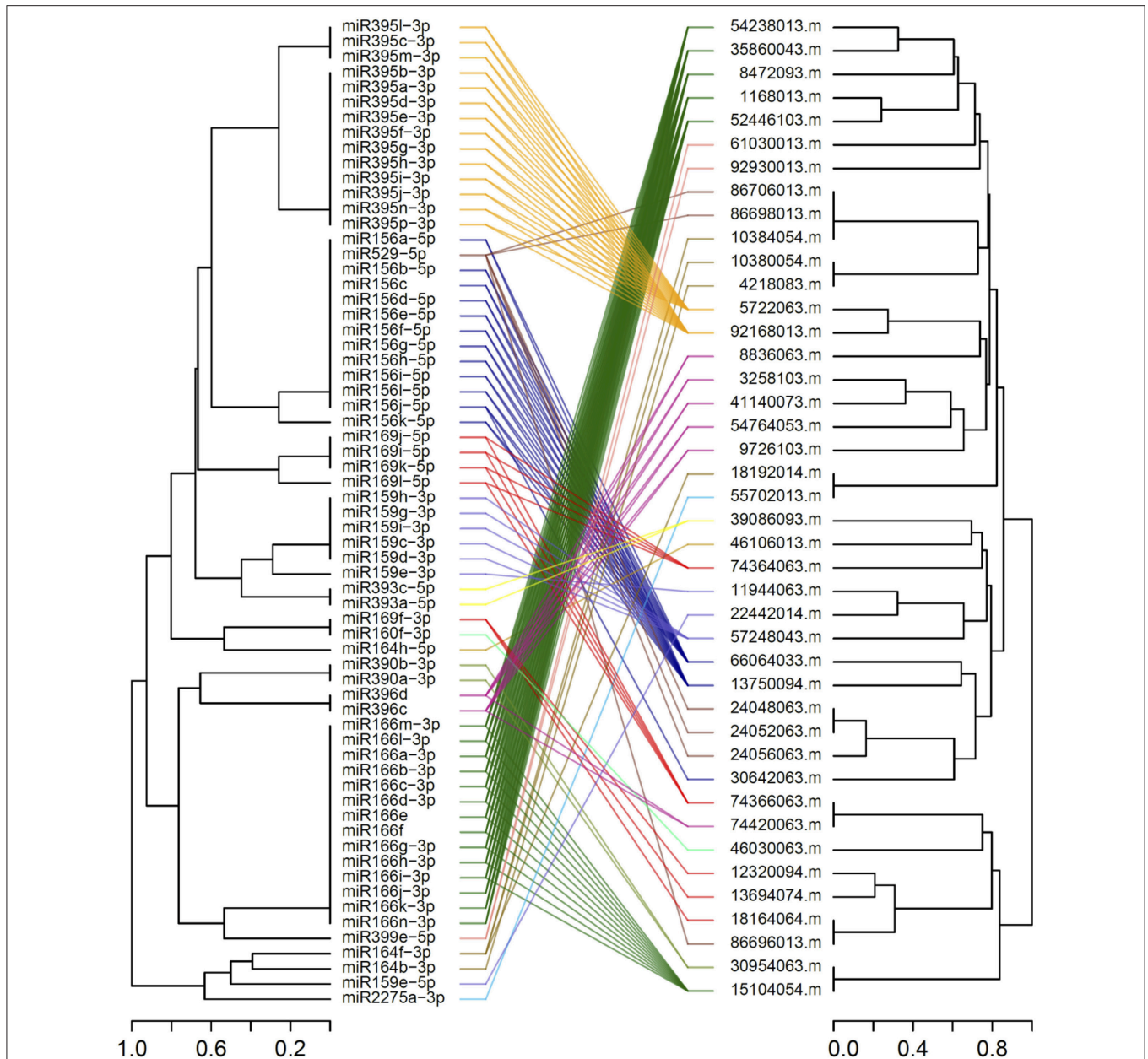
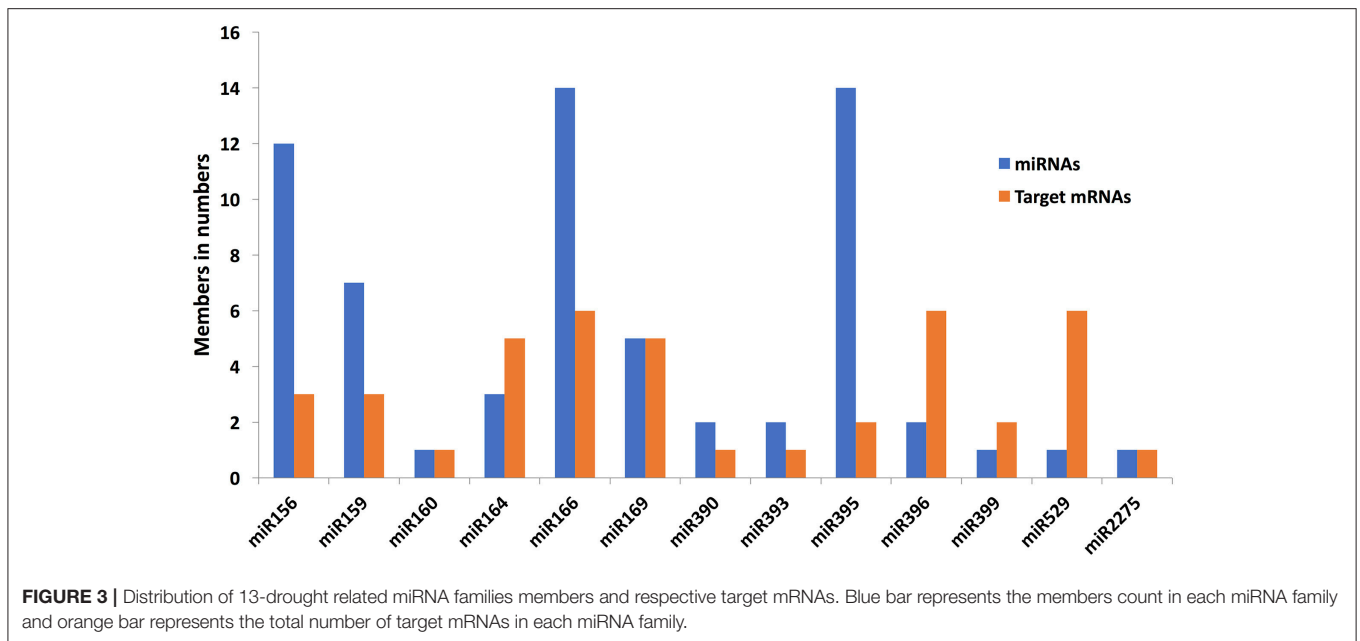


FIGURE 2 | Tanglegram of predicted mature miRNA and target mRNA sequences (with the target site masked) related to drought/low-moisture stress. Lines colored according to the miRNA family, connect the miRNA sequences to their respective target mRNAs.

seedling stage, revealing their interactions with each other. The significantly up-regulated miRNAs common to both genotypes (HKI-1532 and V-372) included *zma-miR164h*, *zma-miR169l*, *zma-miR396c*, *zma-miR396d*, and *zma-miR399e*. In tolerant genotype HKI-1532, 16 miRNAs belonging to the *zma-miR159*, *zma-miR160*, *zma-miR164*, *zma-miR166*, *zma-miR169*, *zma-miR390*, *zma-miR395*, *zma-miR396*, and *zma-miR399* families were significantly up-regulated. However, *zma-miR156* and *zma-miR159* were significantly down-regulated. Conversely, in V-372, members of 7 families—*zma-miR164*, *zma-miR169*, *zma-miR393*, *zma-miR396*, *zma-miR399*, *zma-miR529*, and *zma-miR2275*—were significantly up-regulated; and *zma-miR156*,

zma-miR159, *zma-miR166* and *zma-miR395* families were significantly down-regulated. These results indicate disparate patterns of regulation in the tolerant and susceptible genotypes (Figure 6).

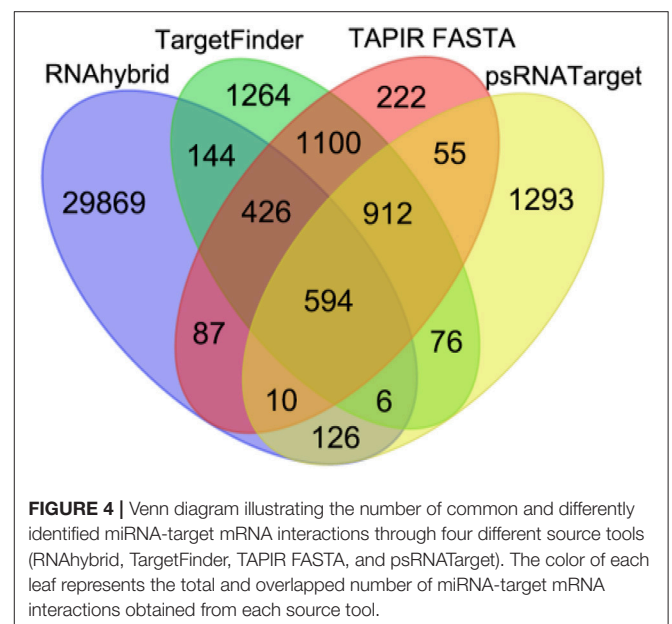
Furthermore, expression analysis of target mRNAs revealed six up-regulated, 18 down-regulated and 18 neutrally expressed mRNAs in HKI-1532. V-372, however, had one up-regulated, 10 down-regulated and 26 neutrally expressed mRNAs. The miRNA-target mRNA interactions were classified into four different groups: Type I, with miRNA-mRNA up-regulations; Type II, with miRNA-mRNA down-regulations; Type III, with neutral miRNA-mRNA expressions; and Type IV, with opposite



interactions. The tolerant genotype, HKI-1532, had 46 Type I, 13 Type II, 23 Type III, and 101 Type IV combinations. The susceptible genotype, V-372, had eight Type I, four Type II, 20 Type III, and 151 Type IV combinations (**Figure 6**).

DISCUSSION

Several investigations have revealed the role of miRNAs in modulating gene expression under abiotic stresses—cold (Lv et al., 2010), salt (Ding et al., 2009), aluminum tolerance (Kong et al., 2014), waterlogging (Zhai et al., 2013), and drought (Li et al., 2013; Wang Y. G. et al., 2014). Among the several abiotic stresses, drought is the most prominent stress in the sub-tropical maize production systems. A number of plant parameters, such as growth, yield, membrane integrity, pigment composition, osmotic relations and photosynthesis, are commonly affected by drought stress (Praba et al., 2009). The change in miRNA expression pattern under drought is an indication of stress responses in plants. Such findings provide an insight into drought tolerance mechanisms and can potentially aid in designing drought-tolerant cultivars (Zhao et al., 2007; Chen et al., 2012). The role of miRNAs in drought tolerance has been previously explored in maize using temperate germplasm. Wang Y. G. et al. (2014) identified 301 drought-responsive miRNAs in temperate maize germplasm and Li et al. (2013) detected differentially expressed 68 miRNAs falling under 29 miRNA families in a maize inbred R09. In the present investigation, 42 differentially expressed mRNA sequences were predicted to be putatively regulated by 65 miRNAs belonging to 13 families. Furthermore, a combination of 183 miRNA-target mRNA interactions were identified and validated in a contrasting pair of subtropical maize germplasm. Four prediction tools were employed to reveal high-confidence predictions of miRNA-target mRNA interactions due



to enhanced accuracy and site recognition efficacy as compared to previous single tool-based prediction (Ding et al., 2012; Li et al., 2013; Wang Y. G. et al., 2014). All 13 of the miRNA families were found to be conserved across taxonomic groups related to drought, ABA or oxidative stress response (Kim et al., 2003; Ding et al., 2013). These results suggested that specific microarray expression profiling data could be effectively used for identifying regulatory miRNAs.

miRNA genes originated from the preexisting genes through duplication events (Nozawa et al., 2012). miRNAs that are evolutionarily conserved are usually encoded by miRNA gene

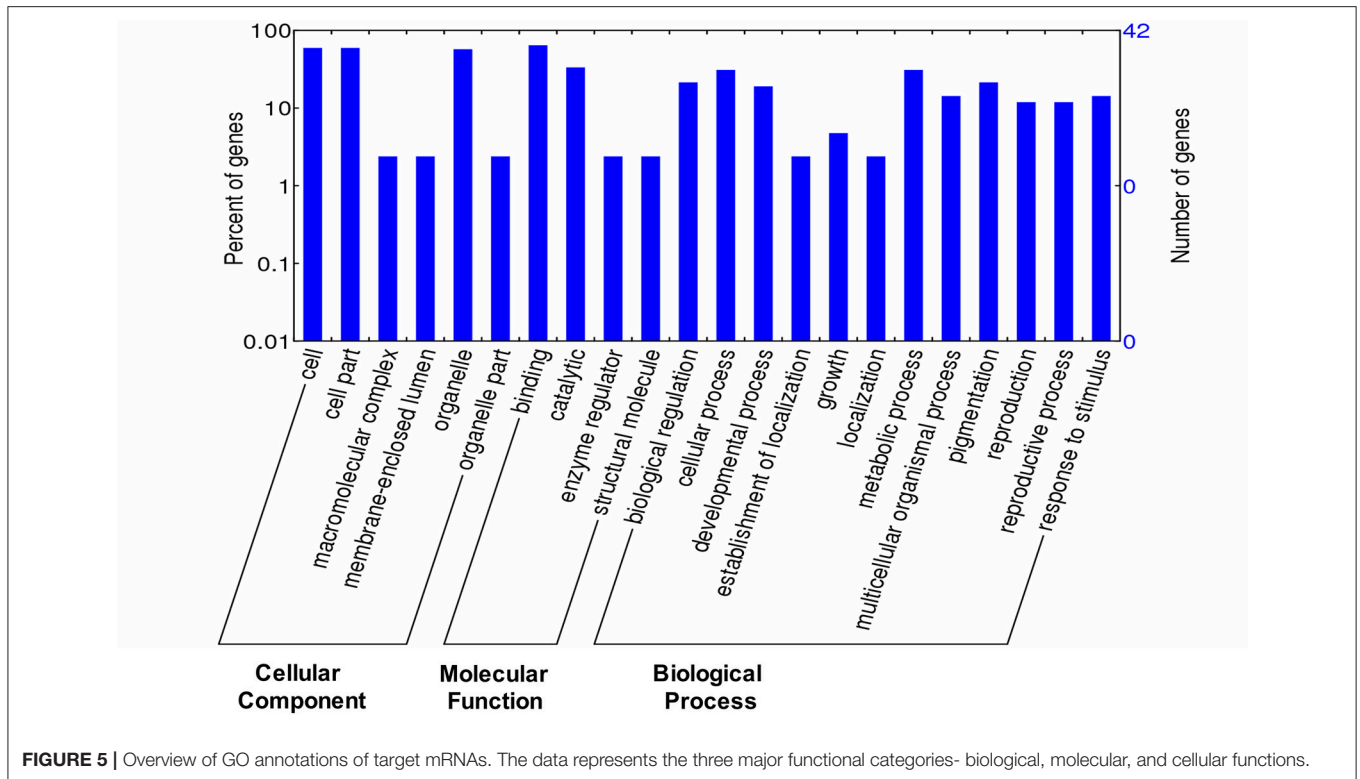


FIGURE 5 | Overview of GO annotations of target mRNAs. The data represents the three major functional categories- biological, molecular, and cellular functions.

families. This, coupled with the strong similarity requirements of plant miRNA - target site interactions, leads to overlapping functions of miRNAs belonging to the same families, buffering against loss of individual members (Jones-Rhoades et al., 2006). It was found that the families such as miR2118, miR395, and miR159 were highly conserved in maize and rice (Xu et al., 2017). In a few interactions, similar miRNA sequences belonging to the same families were found to target dissimilar target mRNA sequences, such as mature miRNAs from the 3' arm of miRNA family 166 and those from the 5' arm of miRNA family 169. In *Arabidopsis*, miR395 has been found to regulate an ATP sulfurylase and an unrelated sulfate transporter (Allen et al., 2005), and miR159 has been reported to target both *MYB101* and *MYB120* and two unrelated genes, *OPT1* and *ACS8* (Schwab et al., 2005). In maize, miR395 showed differential expression in response to salt stress, and was found to regulate *ATP sulfurylase*, *L-Isoaspartyl methyltransferase*, and *Beta-D-xylosidase* genes (Ding et al., 2009).

The promoter analysis of differentially expressed miRNAs indicated that they were predominantly involved in ABA signaling, the auxin response pathway, light-responsive pathways and endosperm expression. Among the 13 drought-related miRNA families, some families had common targets, i.e., zma-miR160, zma-miR390, and zma-miR393 were mostly related to the ARF (auxin response factor) transcription factor, which plays an active role in ABA and auxin mediated signaling under drought conditions (Ding et al., 2013). Similarly, miR164 targets the NAC transcription factor and *CUC* (cup shaped cotyledon) genes in *Arabidopsis* (Rhoades et al., 2002); these genes are

responsible for root and shoot development. Furthermore, zma-miR164 showed higher expression in crown roots but not in seminal roots of maize, suggesting that it could play a crucial role in development of crown roots (Kong et al., 2014).

Members of the miR166 family generally participate in regulation of their target, *HD-Zip III* (homeodomain-leucine zipper III), which is engaged in lateral root development, initiation of axillary leaf meristems and leaf polarity (Boualem et al., 2008) (Figure 7). In maize, it was reported that the leaf polarity was controlled by a subset of miR166 family members (Nogueira et al., 2009). The regulatory mechanism of miR390 is somewhat different, as it prompts the production of tasiRNA (TAS-3 derived small interfering RNA), which targets the ARFs (*ARF2*, *ARF3*, and *ARF4*) that function in lateral root emergence and organ polarity (Nogueira et al., 2009; Meng et al., 2010). miR399 is involved in regulation of a phosphate transporter that regulates the uptake of phosphate and its translocation (Pant et al., 2008). Our *in-silico* analysis identified various target mRNAs, including *SPL* (SQUAMOSA promoter-binding-like proteins), *GAMYB* (a gibberellin- and abscisic acid-related MYB), *ARF* (auxin response factor), *AST* (a sulfate transporter), and *GRFs* (growth regulating factors), among others. The majority of the identified targets are also conserved across other plant species, including model systems such as *Arabidopsis* (Adai et al., 2005) and rice (Wang et al., 2004; Luo et al., 2006).

The interactions of each miRNA with a specific target are of prime importance, as these interactions can help to discern the variation in drought-induced gene expression. For example, the miR156 family regulates the expression of *SPL*, leading

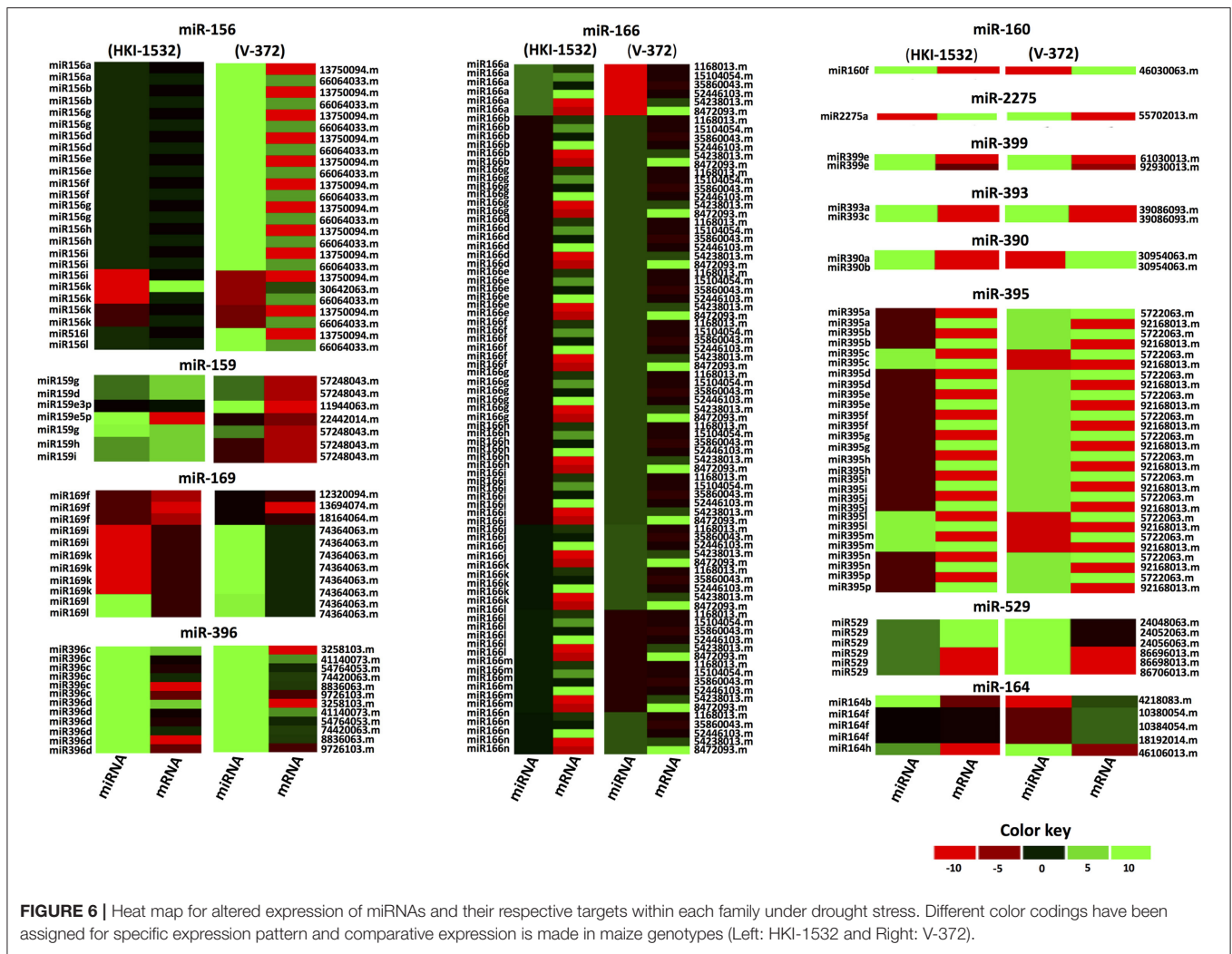
TABLE 2 | Putative *cis*-regulatory elements identified in the upstream region of drought responsive miRNA in maize.

<i>cis</i> -elements	Frequency	Sequence	Function
TATCCAT/C-motif	10	TATCCAT	<i>cis</i> -acting regulatory element; associated with G-box like motif; involved in sugar repression responsiveness
GA-motif	11	AAGGAAGA	Part of a light responsive element
LTR	12	CCGAAA	<i>cis</i> -acting element involved in low-temperature responsiveness
CCGTCC-box	12	CCGTCC	<i>cis</i> -acting regulatory element related to meristem specific activation
MNF1	12	GTGCCCTT	Light responsive element
ATCT-motif	12	AATCTAATCC	part of a conserved DNA module involved in light Responsiveness
GC-motif	14	CCCCCG	Enhancer-like element involved in anoxic specific inducibility
AE-box	14	AGAAACAT	Expression and repressed
GARE-motif	14	AAACAGA	Gibberellin-responsive element
TCT-motif	14	TCTTAC	Part of a light responsive element
RY-element	15	CATGCATG	<i>cis</i> -acting regulatory element involved in seed-specific regulation
5UTR Py-rich stretch	16	TTTCTTCTCT	<i>cis</i> -acting element conferring high transcription levels
TCA-element	16	CAGAAAAGGA	<i>cis</i> -acting element involved in salicylic acid responsiveness
ACE	17	GACACGTATG	<i>cis</i> -acting element involved in light responsiveness
Box I	17	TTTCAAA	Light responsive element
HSE	18	AAAAAATTC	<i>cis</i> -acting element involved in heat stress responsiveness
TGA-element	20	AACGAC	Auxin-responsive element
Box-W1	20	TTGACC	Fungal elicitor responsive element
W box	20	TTGACC	Elicitation; wounding and pathogen responsiveness. Binds WRKY type transcription factors
CCAAT-box	21	CAACGG	MYBHv1 binding site
CATT-motif	21	GCATTC	Part of a light responsive element
O2-site	22	GATGACATGG	<i>cis</i> -acting regulatory element involved in zein metabolism regulation
GCN4_motif	22	TGAGTCA	<i>cis</i> -regulatory element involved in endosperm expression
Box 4	22	ATTAAT	Part of a conserved DNA module involved in light responsiveness
CAT-box	24	GCCACT	<i>cis</i> -acting regulatory element related to meristem expression
GT1-motif	24	GGTTAA	Light responsive element
I-box	24	GATATGG	Part of a light responsive element
AAGAA-motif	26	GAAAGAA	-
TC-rich repeats	28	ATTTTCTTCA	<i>cis</i> -acting element involved in defense and stress responsiveness
GAG-motif	30	GAGAGAT	Part of a light responsive element
ABRE	31	GCAACGTGTC	<i>cis</i> -acting element involved in the abscisic acid responsiveness
circadian	31	CAANNNNATC	<i>cis</i> -acting regulatory element involved in circadian control
ARE	35	TGGTTT	<i>cis</i> -acting regulatory element essential for the anaerobic induction
CGTCA-motif	36	CGTCA	<i>cis</i> -acting regulatory element involved in the MeJA-responsiveness
TGACG-motif	36	TGACG	<i>cis</i> -acting regulatory element involved in the MeJA-responsiveness
Sp1	37	CC(G/A)CCC	Light responsive element
MBS	37	CAACTG	MYB binding site involved in drought-inducibility
G-Box	42	CACGTT	<i>cis</i> -acting regulatory element involved in light responsiveness
Skn-1_motif	45	GTCAT	<i>cis</i> -acting regulatory element required for endosperm expression
TATA-box	52	ATATAAT	Core promoter element around -30 of transcription start
CAAT-box	53	CCAAT	Common <i>cis</i> -acting element in promoter and enhancer regions

to plant developmental phase transitions (Wang et al., 2009; Chen et al., 2010) (**Figure 6**). Altered expression of this family during drought is evidence of its functional novelty in drought stress. Li et al. (2013) reported the up-regulation of miR156 at the early stage of drought stress in the maize seedlings. The regulation of SPL TFs through miR156 was also reported during somatic embryogenesis of maize (Chávez-Hernández et al., 2015). Additionally, Liu et al. (2014) reported that miR156 controlled several SPL genes during the juvenile-to-adult phase

transition in maize. Over-expression of miR156 encoding the maize *Cg1* gene showed to prevent the flowering, and improved the digestibility and starch content in switchgrass (Chuck et al., 2011). Similarly, the expression of *GRF*, which plays important role in leaf growth by transforming cell proliferation, is regulated by miR396c (Kim et al., 2003).

It was interesting to note that the expression patterns of some miRNAs were genotype-specific under drought stress. In this experiment, the Type IV (opposite) pattern

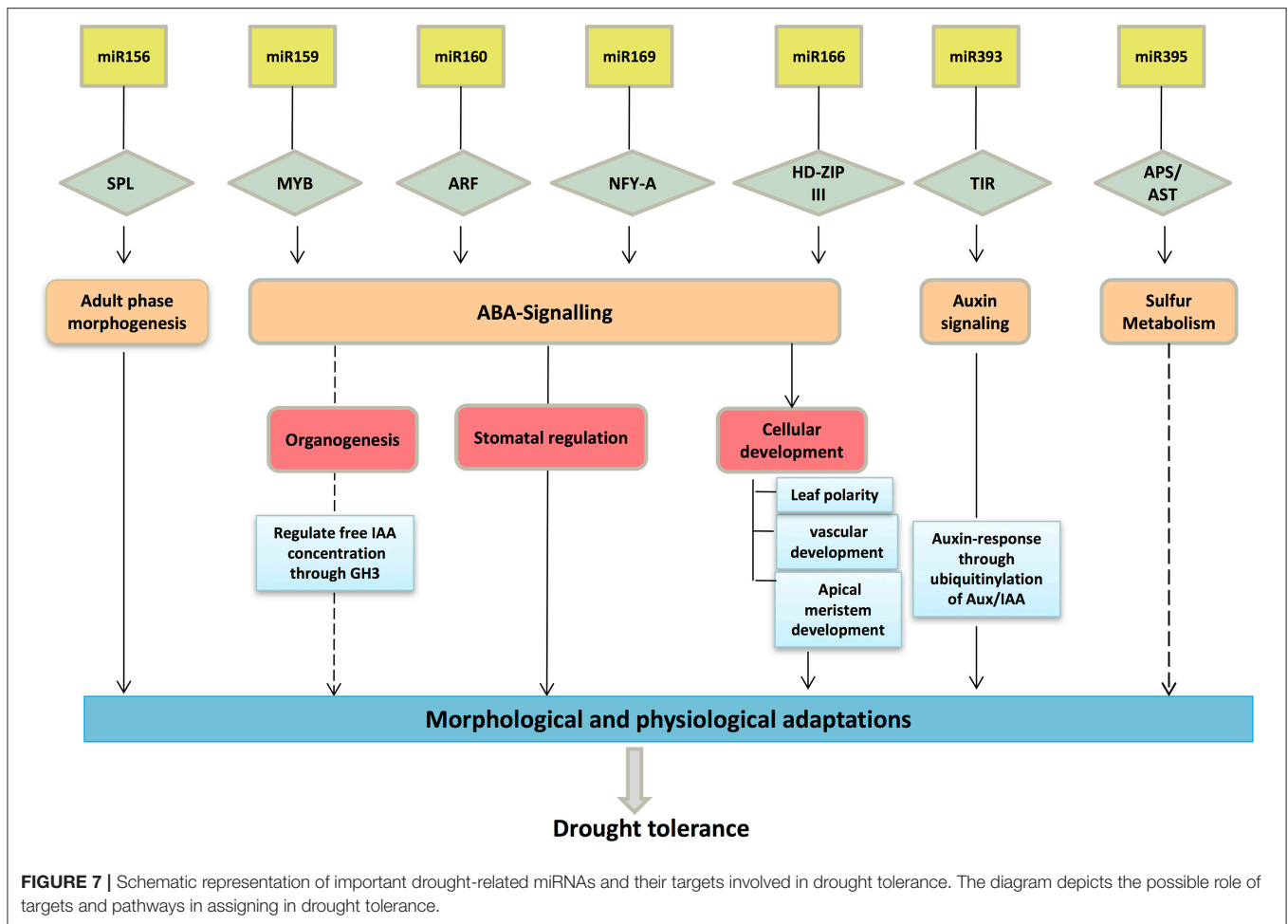


identified between miRNA and target mRNAs suggested negative regulation under drought stress. The up-regulation of miR396 in HKI-1532 suppressed the expression of its target, *GRF1*, whereas the up-regulation of miR396 in V-372 led to neutral expression of its target. This suggests that suppressed *GRF* expression under drought can provide tolerance by precluding leaf growth and function under stress (Figure 7). Our hypothesis is supported by the fact that *GRF8* reduces stomatal density in *Arabidopsis* (Liu et al., 2009). However, expression of miR396c and miR396d, targeting *GRFs*, was the same in both genotypes. It may be deduced that the small up-regulation of miRNAs in HKI-1532 contributes to drought tolerance. Similar results were obtained for miR396 in *Arabidopsis* (Liu et al., 2008) and tobacco (Frazier et al., 2011).

In the same way, Type I (up-regulated) miRNA-mRNA interaction was observed for miR169l, which targets transcription factor *NF-YA3* (nuclear transcription factor subunit A-3) in HKI1532. However, the same miRNA exhibited a Type IV interaction in V-372. Our hypothesis is supported by findings

in *Arabidopsis*, where the role of transcription factor *NF-YA* in drought tolerance is well documented (Liu et al., 2008). Similarly, Type IV interaction was also observed for the miR159 family, targeting the transcription factor *GAMYB* that is involved in flowering development and flowering time. Furthermore, overexpression of the miR159 family during stress aids in germination under stress conditions. In *Arabidopsis*, overexpression of *MYB* transcription factors (*MYB33* and *MYB101*) provides tolerance to drought by adapting ABA hypersensitivity (Reyes and Chua, 2007). Regulation of *MYB* transcription factors by the member of miR159 was also reported in temperate maize (Li et al., 2013) and was found to play a crucial role in stress tolerance (Wang Y. G. et al., 2014).

The miR393 family that targets the *TIR1* (transport inhibitor response 1) enzyme was up-regulated in V-372. This enzyme directly participates in the ubiquitinylation of inhibitors of the auxin response pathway (Dharmasiri and Estelle, 2002). miR160, representing a Type IV interaction in HKI-1532, plays a role in ABA-auxin interaction during drought (Liu et al., 2007).



Similarly, the Type IV interaction between up-regulated miR395 up-regulated and its down-regulated targets *ATP sulfurylase* and *HD-Zip* transcription factor (Buchner et al., 2004) were involved in seedling growth and seed germination in HKI-1532. This negative link indicates the role of miR395 in conferring drought tolerance. Our results are supported by the findings of studies conducted in *Arabidopsis* under drought stress, where overexpression of miR395 decreased seed growth and germination upon drought exposure (Kim et al., 2010). Similar to observations for miR156 family, Type II (down-regulated) interactions were found for *SPL* genes in both genotypes. *SPL* genes have been found to be involved in deferred blooming and the adult phase transition in *Arabidopsis* (Park et al., 2010). This can be correlated with the similar studies conducted in rice and maize (Wei et al., 2009; Zhou et al., 2010).

CONCLUSIONS

Drought-specific miRNAs and their target mRNAs were identified from existing drought-associated microarray expression profiling data, and their expression was assayed in two contrasting subtropical maize genotypes, HKI-1532 and V-372. The promoters of all drought-related miRNAs were

analyzed and confirmed the presence of important drought-responsive CREs. Eleven miRNAs belonging to nine families in HKI-1532 and seven miRNAs from three families in V-372 were differentially regulated in both genotypes. Among different miRNA-mRNA interactions, suppression of biologically important genes by miRNAs in V-372 suggests their weak performance under drought stress. It was noticed that the fact that some crucial genes being unaffected by regulatory miRNAs in tolerant genotype HKI1532 could have led to drought tolerance. Our experiment provided a clear understanding of miRNA regulation in drought response. To our knowledge, this is the first report on the role of miRNAs in regulation of drought tolerance in subtropical maize inbreds. Many of the identified candidate miRNAs and mRNAs from the present investigation could be used as potential candidates for development of drought-tolerant maize hybrids for the subtropical production system.

AUTHOR CONTRIBUTIONS

JA and TN: conceived and designed the experiments; JA, SK, AK, BP, and MGM: performed the experiments; SR, MS, and ARR: analyzed the data; All authors contributed to manuscript

preparation. All authors have read and approved the final manuscript.

ACKNOWLEDGMENTS

We thank the ICAR Network Projects on Transgenics in Crop Plants (Maize Functional Genomics Component: 21-22) and the Computational Biology and Agricultural Bioinformatics (Agril.Edn.14 (44)/2014-A&P) for funding

REFERENCES

- A dai, A., Johnson, C., Mlotshwa, S., Archer-Evans, S., Manocha, V., Vance, V., et al. (2005). Computational prediction of miRNAs in *Arabidopsis thaliana*. *Genome Res.* 15, 78–91. doi: 10.1101/gr.2908205
- Allen, E., Xie, Z., Gustafson, A. M., and Carrington, J. C. (2005). microRNA-directed phasing during trans-acting siRNA biogenesis in plants. *Cell* 121, 207–221. doi: 10.1016/j.cell.2005.04.004
- Barrett, T., Suzek, T. O., Troup, D. B., Wilhite, S. E., Ngau, W.-C., Ledoux, P., et al. (2005). NCBI GEO: mining millions of expression profiles-database and tools. *Nucleic Acids Res.* 33, D562–D566. doi: 10.1093/nar/gki022
- Bartel, D. P. (2004). MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 116, 281–297. doi: 10.1016/S0092-8674(04)00045-5
- Baumberger, N., and Baulcombe, D. C. (2005). Arabidopsis ARGONAUTE1 is an RNA Slicer that selectively recruits microRNAs and short interfering RNAs. *Proc. Natl. Acad. Sci. U.S.A.* 102, 11928–11933. doi: 10.1073/pnas.0505461102
- Bonnet, E., He, Y., Billiau, K., and van de Peer, Y. (2010). TAPIR, a web server for the prediction of plant microRNA targets, including target mimics. *Bioinformatics* 26, 1566–1568. doi: 10.1093/bioinformatics/btq233
- Boualem, A., Laporte, P., Jovanovic, M., Laffont, C., Plet, J., Combier, J. P., et al. (2008). MicroRNA166 controls root and nodule development in *Medicago truncatula*. *Plant J.* 54, 876–887. doi: 10.1111/j.1365-313X.2008.03448.x
- Buchanan, C. D., Lim, S., Salzman, R. A., Kagiampakis, I., Morishige, D. T., Weers, B. D., et al. (2005). *Sorghum bicolor*'s transcriptome response to dehydration, high salinity and ABA. *Plant Mol. Biol.* 58, 699–720. doi: 10.1007/s11103-005-7876-2
- Buchner, P., Takahashi, H., and Hawkesford, M. J. (2004). Plant sulphate transporters: co-ordination of uptake, intracellular and long-distance transport. *J. Exp. Bot.* 1765–1773. doi: 10.1093/jxb/erh206
- Ceccarelli, S., and Grandi, S. (1996). Drought as a challenge for the plant breeder. *Plant Growth Regul.* 20, 149–155. doi: 10.1007/BF00024011
- Charif, D., Humblot, L., Lobry, J. R., Necseulea, A., Palmeira, L., and Penel, S. (2008). *SeqinR 2.0-1: A Contributed Package to the R Project for Statistical Computing Devoted to Biological Sequences Retrieval and Analysis*. 268
- Chávez-Hernández, E. C., Alejandri-Ramírez, N. D., Juárez-González, V. T., and Dinkova, T. D. (2015). Maize miRNA and target regulation in response to hormone depletion and light exposure during somatic embryogenesis. *Front. Plant Sci.* 6:555. doi: 10.3389/fpls.2015.00555
- Chen, C., Ridzon, D. A., Broome, A. J., Zhou, Z., Lee, D. H., Nguyen, J. T., et al. (2005). Real-time quantification of microRNAs by stem-loop RT-PCR. *Nucleic Acids Res.* 33:e179. doi: 10.1093/nar/gni178
- Chen, H., Li, Z., and Xiong, L. (2012). A plant microRNA regulates the adaptation of roots to drought stress. *FEBS Lett.* 586, 1742–1747. doi: 10.1016/j.febslet.2012.05.013
- Chen, X., Zhang, Z., Liu, D., Zhang, K., Li, A., and Mao, L. (2010). SQUAMOSA promoter-binding protein-like transcription factors: star players for plant growth and development. *J. Integr. Plant Biol.* 52, 946–951. doi: 10.1111/j.1744-7909.2010.00987.x
- Chuck, G., Cigan, A. M., Saetern, K., and Hake, S. (2007). The heterochronic maize mutant *Corngrass1* results from overexpression of a tandem microRNA. *Nat. Genet.* 39, 544–549. doi: 10.1038/ng2001
- Chuck, G. S., Tobias, C., Sun, L., Kraemer, F., Li, C., Dibble, D., et al. (2011). Overexpression of the maize *Corngrass1* microRNA prevents flowering,

the study. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <http://journal.frontiersin.org/article/10.3389/fpls.2017.00941/full#supplementary-material>

- improves digestibility, and increases starch content of switchgrass. *Proc. Natl. Acad. Sci. U.S.A.* 108, 17550–17555. doi: 10.1073/pnas.1113971108
- Conesa, A., Götz, S., García-Gómez, J. M., Terol, J., Talón, M., and Robles, M. (2005). Blast2GO: A universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics* 21, 3674–3676. doi: 10.1093/bioinformatics/bti610
- Dai, X., and Zhao, P. X. (2011). PsRNATarget: a plant small RNA target analysis server. *Nucleic Acids Res.* 39, 155–159. doi: 10.1093/nar/gkr319
- Dharmasiri, S., and Estelle, M. (2002). The role of regulated protein degradation in auxin response. *Plant Mol. Biol.* 49, 401–409. doi: 10.1023/A:1015203013208
- Ding, D., Zhang, L., Wang, H., Liu, Z., Zhang, Z., and Zheng, Y. (2009). Differential expression of miRNAs in response to salt stress in maize roots. *Ann. Bot.* 103, 29–38. doi: 10.1093/aob/mcn205
- Ding, J., Zhou, S., and Guan, J. (2012). Finding MicroRNA targets in plants: current status and perspectives. *Genomics Proteomics Bioinformatics* 10, 264–275. doi: 10.1016/j.gpb.2012.09.003
- Ding, Y., Tao, Y., and Zhu, C. (2013). Emerging roles of microRNAs in the mediation of drought stress response in plants. *J. Exp. Bot.* 64, 3077–3086. doi: 10.1093/jxb/ert164
- Frazier, T. P., Sun, G., Burklew, C. E., and Zhang, B. (2011). Salt and drought stresses induce the aberrant expression of microRNA genes in tobacco. *Mol. Biotechnol.* 49, 159–165. doi: 10.1007/s12033-011-9387-5
- Griffiths-Jones, S., Saini, H. K., Van Dongen, S., and Enright, A. J. (2008). miRBase: tools for microRNA genomics. *Nucleic Acids Res.* 36, 154–158. doi: 10.1093/nar/gkm952
- Ha, S., Vankova, R., Yamaguchi-Shinozaki, K., Shinozaki, K., and Tran, L. S. P. (2012). Cytokinins: metabolism and function in plant adaptation to environmental stresses. *Trends Plant Sci.* 17, 172–179. doi: 10.1016/j.tplants.2011.12.005
- Hansen, S., Clay, S. A., Clay, D. E., Carlson, C. G., Reicks, G., Jarachi, Y., et al. (2013). Landscape features impact on soil available water, corn biomass, and gene expression during the late vegetative stage. *Plant Genome* 6, 1–9. doi: 10.3835/plantgenome2012.11.0029
- Hayano-Kanashiro, C., Calderón-Vásquez, C., Ibarra-Laclette, E., Herrera-Estrella, L., and Simpson, J. (2009). Analysis of gene expression and physiological responses in three Mexican maize landraces under drought stress and recovery irrigation. *PLoS ONE* 4:e7531. doi: 10.1371/journal.pone.0007531
- Jones-Rhoades, M. W., Bartel, D. P., and Bartel, B. (2006). MicroRNAs and their regulatory roles in plants. *Annu. Rev. Plant Biol.* 57, 19–53. doi: 10.1146/annurev.arplant.57.032905.105218
- Juarez, M. T., Kui, J. S., Thomas, J., Heller, B. A., and Timmermans, M. C. P. (2004). microRNA-mediated repression of rolled leaf1 specifies maize leaf polarity. *Nature* 428, 84–88. doi: 10.1038/nature02363
- Kakumanu, A., Ambavaram, M. M. R., Klumas, C., Krishnan, A., Batlang, U., Myers, E., et al. (2012). Effects of drought on gene expression in maize reproductive and leaf meristem tissue revealed by RNA-Seq. *Plant Physiol.* 160, 846–867. doi: 10.1104/pp.112.200444
- Kang, M., Zhao, Q., Zhu, D., and Yu, J. (2012). Characterization of microRNAs expression during maize seed development. *BMC Genomics* 13:360. doi: 10.1186/1471-2164-13-360
- Kim, J. H., Choi, D., and Kende, H. (2003). The AtGRF family of putative transcription factors is involved in leaf and cotyledon growth in *Arabidopsis*. *Plant J.* 36, 94–104. doi: 10.1046/j.1365-313X.2003.01862.x

- Kim, J. Y., Lee, H. J., Jung, H. J., Maruyama, K., Suzuki, N., and Kang, H. (2010). Overexpression of microRNA395c or 395e affects differently the seed germination of *Arabidopsis thaliana* under stress conditions. *Planta* 232, 1447–1454. doi: 10.1007/s00425-010-1267-x
- Kong, X., Zhang, M., Xu, X., Li, X., Li, C., and Ding, Z. (2014). System analysis of microRNAs in the development and aluminium stress responses of the maize root system. *Plant Biotechnol. J.* 12, 1108–1121. doi: 10.1111/pbi.1221
- Kozomara, A., and Griffiths-Jones, S. (2011). MiRBase: integrating microRNA annotation and deep-sequencing data. *Nucleic Acids Res.* 39, 152–157. doi: 10.1093/nar/gkq1027
- Kramer, M. F. (2011). Stem-loop RT-qPCR for miRNAs. *Curr. Protoc. Mol. Biol. Chapter 15:Unit 15.10*. doi: 10.1002/0471142727.mb1510s95
- Kurihara, Y., and Watanabe, Y. (2004). Arabidopsis micro-RNA biogenesis through Dicer-like 1 protein functions. *Proc. Natl. Acad. Sci. U.S.A.* 101, 12753–12758. doi: 10.1073/pnas.0403115101
- Langridge, P., and Reynolds, M. P. (2015). Genomic tools to assist breeding for drought tolerance. *Curr. Opin. Biotechnol.* 32, 130–135. doi: 10.1016/j.copbio.2014.11.027
- Lee, Y., Kim, M., Han, J., Yeom, K.-H., Lee, S., Baek, S. H., et al. (2004). MicroRNA genes are transcribed by RNA polymerase II. *EMBO J.* 23, 4051–4060. doi: 10.1038/sj.emboj.7600385
- Li, J. S., Fu, F. L., Ming, A. N., Zhou, S. F., She, Y. H., and Li, W. C. (2013). Differential expression of microRNAs in response to drought stress in maize. *J. Integr. Agricul.* 12, 1414–1422. doi: 10.1016/S2095-3119(13)60311-1
- Li, Y., Sun, C., Huang, Z., Pan, J., Wang, L., and Fan, X. (2009). Mechanisms of progressive water deficit tolerance and growth recovery of chinese maize foundation genotypes huangzao 4 and chang 7-2, which are proposed on the basis of comparison of physiological and transcriptomic responses. *Plant Cell Physiol.* 50, 2092–2111. doi: 10.1093/pcp/pcp145
- Liu, D., Song, Y., Chen, Z., and Yu, D. (2009). Ectopic expression of miR396 suppresses GRF target gene expression and alters leaf growth in Arabidopsis. *Physiol. Plant.* 136, 223–236. doi: 10.1111/j.1399-3054.2009.01229.x
- Liu, H.-H., Tian, X., Li, Y.-J., Wu, C.-A., and Zheng, C.-C. (2008). Microarray-based analysis of stress-regulated microRNAs in *Arabidopsis thaliana*. *RNA* 14, 836–843. doi: 10.1261/rna.895308
- Liu, H., Qin, C., Chen, Z., Zuo, T., Yang, X., Zhou, H., et al. (2014). Identification of miRNAs and their target genes in developing maize ears by combined small RNA and degradome sequencing. *BMC Genomics* 15:25. doi: 10.1186/1471-2164-15-25
- Liu, P. P., Montgomery, T. A., Fahlgren, N., Kasschau, K. D., Nonogaki, H., and Carrington, J. C. (2007). Repression of AUXIN RESPONSE FACTOR10 by microRNA160 is critical for seed germination and post-germination stages. *Plant J.* 52, 133–146. doi: 10.1111/j.1365-313X.2007.03218.x
- Livak, K. J., and Schmittgen, T. D. (2001). Analysis of relative gene expression data using real-time quantitative PCR and. *Methods* 25, 402–408. doi: 10.1006/meth.2001.1262
- Lu, H. F., Dong, H. T., Sun, C. B., Qing, D. J., Li, N., Wu, Z. K., et al. (2011). The panorama of physiological responses and gene expression of whole plant of maize inbred line YQ7-96 at the three-leaf stage under water deficit and re-watering. *Theor. Appl. Genet.* 123, 943–958. doi: 10.1007/s00122-011-1638-0
- Luo, M., Liu, J., Lee, R. D., Scully, B. T., and Guo, B. (2010). Monitoring the Expression of Maize Genes in Developing Kernels under Drought Stress using Oligo-microarray. *J. Integr. Plant Biol.* 52, 1059–1074. doi: 10.1111/j.1744-7909.2010.01000.x
- Luo, Y. C., Zhou, H., Li, Y., Chen, J. Y., Yang, J. H., Chen, Y. Q., et al. (2006). Rice embryogenic calli express a unique set of microRNAs, suggesting regulatory roles of microRNAs in plant post-embryonic development. *FEBS Lett.* 580, 5111–5116. doi: 10.1016/j.febslet.2006.08.046
- Lv, D. K., Bai, X., Li, Y., Ding, X. D., Ge, Y., Cai, H., et al. (2010). Profiling of cold-stress-responsive miRNAs in rice by microarrays. *Gene* 459, 39–47. doi: 10.1016/j.gene.2010.03.011
- Marino, R., Ponnaiah, M., Krajewski, P., Frova, C., Gianfranceschi, L., Pè, M. E., et al. (2009). Addressing drought tolerance in maize by transcriptional profiling and mapping. *Mol. Genet. Genomics* 281, 163–179. doi: 10.1007/s00438-008-0401-y
- Meng, Y., Ma, X., Chen, D., Wu, P., and Chen, M. (2010). MicroRNA-mediated signaling involved in plant root development. *Biochem. Biophys. Res. Commun.* 393, 345–349. doi: 10.1016/j.bbrc.2010.01.129
- Min, H., Chen, C., Wei, S., Shang, X., Sun, M., Xia, R., et al. (2016). Identification of drought tolerant mechanisms in maize seedlings based on transcriptome analysis of recombination inbred lines. *Front. Plant Sci.* 7:1080. doi: 10.3389/fpls.2016.01080
- Nepolean, T., Sharma, R., Singh, N., Shiriga, K., Mohan, S., Mittal, S., et al. (2017). Genome-wide expression and functional interactions of genes under drought stress in maize. *Int J. Genomics* 2017:2568706. doi: 10.1155/2017/2568706
- Nogueira, F. T., Chitwood, D. H., Madi, S., Ohtsu, K., Schnable, P. S., Scanlon, M. J., et al. (2009). Regulation of small RNA accumulation in the maize shoot apex. *PLoS Genet.* 5:e1000320. doi: 10.1371/journal.pgen.1000320
- Nozawa, M., Miura, S., and Nei, M. (2012). Origins and evolution of microRNA genes in plant species. *Genome Biol. Evol.* 4, 230–239. doi: 10.1093/gbe/evs002
- Pant, B. D., Buhtz, A., Kehr, J., and Scheible, W. R. (2008). MicroRNA399 is a long-distance signal for the regulation of plant phosphate homeostasis. *Plant J.* 53, 731–738. doi: 10.1111/j.1365-313X.2007.03363.x
- Paradis, E., Claude, J., and Strimmer, K. (2004). APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* 20, 289–290. doi: 10.1093/bioinformatics/btg412
- Park, M. Y., Wu, G., Gonzalez-Sulser, A., Vaucheret, H., and Poethig, R. S. (2005). Nuclear processing and export of microRNAs in Arabidopsis. *Proc. Natl. Acad. Sci. U.S.A.* 102, 3691–3696. doi: 10.1073/pnas.0405570102
- Park, W., Scheffler, B. E., Bauer, P. J., and Campbell, B. T. (2010). Identification of the family of aquaporin genes and their expression in upland cotton (*Gossypium hirsutum* L.). *BMC Plant Biol.* 10:142. doi: 10.1186/1471-2229-10-142
- Praba, M. L., Cairns, J. E., Babu, R. C., and Lafitte, H. R. (2009). Identification of physiological traits underlying cultivar differences in drought tolerance in rice and wheat. *J. Agron. Crop Sci.* 195, 30–46. doi: 10.1111/j.1439-037X.2008.00341.x
- Regulski, M., Lu, Z., Kendall, J., Donoghue, M. T. A., Reinders, J., Llaça, V., et al. (2013). The maize methylome influences mRNA splice sites and reveals widespread paramutation-like switches guided by small RNA. *Genome Res.* 23, 1651–1662. doi: 10.1101/gr.153510.112
- Rehmsmeier, M., Steffen, P., Höchsmann, M., Giegerich, R., and Ho, M. (2004). Fast and effective prediction of microRNA/target duplexes. *RNA* 10, 1507–1517. doi: 10.1261/rna.5248604
- Reyes, J. L., and Chua, N. H. (2007). ABA induction of miR159 controls transcript levels of two MYB factors during Arabidopsis seed germination. *Plant J.* 49, 592–606. doi: 10.1111/j.1365-313X.2006.02980.x
- Rhoades, M. W., Reinhart, B. J., Lim, L. P., Burge, C. B., Bartel, B., and Bartel, D. P. (2002). Prediction of plant microRNA targets. *Cell* 110, 513–520. doi: 10.1016/S0092-8674(02)00863-2
- Rosegrant, M. W., and Cline, S. A. (2003). Global food security: challenges and policies. *Science* 302, 1917–1919. doi: 10.1126/science.1092958
- Schwab, R., Palatnik, J. F., Riester, M., Schommer, C., Schmid, M., and Weigel, D. (2005). Specific effects of microRNAs on the plant transcriptome. *Dev. Cell* 8, 517–527. doi: 10.1016/j.devcel.2005.01.018
- Shahmuradov, I. A., Solovyev, V. V., and Gammerman, A. J. (2005). Plant promoter prediction with confidence estimation. *Nucleic Acids Res.* 33, 1069–1076. doi: 10.1093/nar/gki247
- Shikha, M., Kanika, A., Rao, A. R., Mallikarjuna, M. G., Gupta, H. S., and Nepolean, T. (2017). Genomic selection for drought tolerance using genome-wide SNPs in maize. *Front. Plant Sci.* 8:550. doi: 10.3389/fpls.2017.00550
- Sievers, F., Wilm, A., Dineen, D., Gibson, T. J., Karplus, K., Li, W., et al. (2011). Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Mol. Syst. Biol.* 7:539. doi: 10.1038/msb.2011.75
- Souvorov, A., Kapustin, Y., Kiryutin, B., Chetvernin, V., Tatusova, T., and Lipman, D. (2010). Gnomon-NCBI Eukaryotic Gene Prediction Tool. Bethesda, MD: National Center for Biotechnology Information.
- Turner, M., Adhikari, S., and Subramanian, S. (2013). Optimizing stem-loop qPCR assays through multiplexed cDNA synthesis of U6 and miRNAs. *Plant Signal. Behav.* 8:e24918. doi: 10.4161/psb.24918
- Vaucheret, H., Vazquez, F., Crété, P., and Bartel, D. P. (2004). The action of ARGONAUTE1 in the miRNA pathway and its regulation by the miRNA pathway are crucial for plant development. *Genes Dev.* 18, 1187–1197. doi: 10.1101/gad.1201404
- Vazquez, F., Gascioli, V., Crété, P., and Vaucheret, H. (2004). The nuclear dsRNA binding protein HYL1 is required for microRNA accumulation and plant

- development, but not posttranscriptional transgene silencing. *Curr. Biol.* 14, 346–351. doi: 10.1016/S0960-9822(04)00047-8
- Wang, B., Sun, Y. F., Song, N., Wei, J. P., Wang, X. J., Feng, H., et al. (2014). MicroRNAs involving in cold, wounding and salt stresses in *Triticum aestivum* L. *Plant Physiol. Biochem.* 80, 90–96. doi: 10.1016/j.plaphy.2014.03.020
- Wang, J. F., Zhou, H., Chen, Y. Q., Luo, Q. J., and Qu, L. H. (2004). Identification of 20 microRNAs from *Oryza sativa*. *Nucleic Acids Res.* 32, 1688–1695. doi: 10.1093/nar/gkh332
- Wang, J. W., Czech, B., and Weigel, D. (2009). miR156-regulated SPL transcription factors define an endogenous flowering pathway in *Arabidopsis thaliana*. *Cell* 138, 738–749. doi: 10.1016/j.cell.2009.06.014
- Wang, Y. G., An, M., Zhou, S. F., She, Y. H., Li, W. C., and Fu, F. L. (2014). Expression profile of maize microRNAs corresponding to their target genes under drought stress. *Biochem. Genet.* 52, 474–493. doi: 10.1007/s10535-016-0590-x
- Wei, L., Zhang, D., Xiang, F., and Zhang, Z. (2009). Differentially expressed miRNAs potentially involved in the regulation of defense mechanism to drought stress in maize seedlings. *Int. J. Plant Sci.* 170, 979–989. doi: 10.1086/605122
- Wu, F., Shu, J., and Jin, W. (2014). Identification and validation of miRNAs associated with the resistance of maize (*Zea mays* L.) to *Exserohilum turcicum*. *PLoS ONE* 9:e87251. doi: 10.1371/journal.pone.0087251
- Xie, Z., Allen, E., Fahlgren, N., Calamar, A., Givan, S. A., and Carrington, J. C. (2005). Expression of *Arabidopsis* MIRNA genes. *Plant Physiol.* 138, 2145–2154. doi: 10.1104/pp.105.062943
- Xu, J., Li, Y., Wang, Y., Liu, X., and Zhu, X. G. (2017). Altered expression profiles of microRNA families during de-etiolation of maize and rice leaves. *BMC Res. Notes* 10:108. doi: 10.1186/s13104-016-2367-x
- You, F. M., Huo, N., Gu, Y. Q., Luo, M.-C., Ma, Y., Hane, D., et al. (2008). BatchPrimer3: a high throughput web application for PCR and sequencing primer design. *BMC Bioinformatics* 9:253. doi: 10.1186/1471-2105-9-253
- Yu, B., Yang, Z. Y., Li, J. J., Minakhina, S., Yang, M. C., Padgett, R. W., et al. (2005). Methylation as a crucial step in plant microRNA biogenesis. *Science* 307, 932–935. doi: 10.1126/science.1107130
- Yu, L.-X. (2003). Comparative transcriptional profiling of placenta and endosperm in developing maize kernels in response to water deficit. *Plant Physiol.* 131, 568–582. doi: 10.1104/pp.014365
- Yue, G., Zhuang, Y., Li, Z., Sun, L., and Zhang, J. (2008). Differential gene expression analysis of maize leaf at heading stage in response to water-deficit stress. *Biosci. Rep.* 28, 125–134. doi: 10.1042/BSR20070023
- Zhai, L., Liu, Z., Zou, X., Jiang, Y., Qiu, F., Zheng, Y., et al. (2013). Genome-wide identification and analysis of microRNA responding to long-term waterlogging in crown roots of maize seedlings. *Physiol. Plant.* 147, 181–193. doi: 10.1111/j.13993054.2012.01653.x
- Zhang, Z., Wei, L., Zou, X., Tao, Y., Liu, Z., and Zheng, Y. (2008). Submergence-responsive microRNAs are potentially involved in the regulation of morphological and metabolic adaptations in maize root cells. *Ann. Bot.* 102, 509–519. doi: 10.1093/aob/mcn129
- Zhao, B., Liang, R., Ge, L., Li, W., Xiao, H., Lin, H., et al. (2007). Identification of drought-induced microRNAs in rice. *Biochem. Biophys. Res. Commun.* 354, 585–590. doi: 10.1016/j.bbrc.2007.01.022
- Zheng, J., Fu, J., Gou, M., Huai, J., Liu, Y., Jian, M., et al. (2010). Genome-wide transcriptome analysis of two maize inbred lines under drought stress. *Plant Mol. Biol.* 72, 407–421. doi: 10.1007/s11103-009-9579-6
- Zhou, L., Liu, Y., Liu, Z., Kong, D., Duan, M., and Luo, L. (2010). Genome-wide identification and analysis of drought-responsive microRNAs in *Oryza sativa*. *J. Exp. Bot.* 61, 4157–4168. doi: 10.1093/jxb/erq237

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2017 Aravind, Rinku, Pooja, Shikha, Kaliyugam, Mallikarjuna, Kumar, Rao and Nepolean. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.