



Analyzing spatial patterns linked to the ecology of herbivores and their natural enemies in the soil

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Modern agricultural systems can benefit from the application of concepts and models from applied ecology. When understood, multitrophic interactions among plants, pests, diseases and their natural enemies can be exploited to increase crop production and reduce undesirable environmental impacts. Although the understanding of subterranean ecology is rudimentary compared to the perspective aboveground, technologies today vastly reduce traditional obstacles to studying cryptic communities. Here we emphasize advantages to integrating as much as possible the use of these methods in order to leverage the information gained from studying communities of soil organisms. PCR-based approaches to identify and quantify species (real time qPCR and next generation sequencing) greatly expand the ability to investigate food web interactions because there is less need for wide taxonomic expertise within research programs. Improved methods to capture and measure volatiles in the soil atmosphere *in situ* make it possible to detect and study chemical cues that are critical to communication across trophic levels. The application of SADIE to directly assess rather than infer spatial patterns in belowground agroecosystems has improved the ability to characterize relationships between organisms in space and time. We review selected methodology and use of these tools and describe some of the ways they were integrated to study soil food webs in Florida citrus orchards with the goal of developing new biocontrol approaches.

Keywords: PCR-based molecular methods, soil food webs, herbivore-induced plant volatiles, SADIE analysis, biological control

INTRODUCTION

Challenges for modern agriculture include producing enough food while simultaneously reducing negative impacts on the environment and using our resources in sustainable ways. A major challenge in ecology is understanding where multitrophic interactions unfold, and how to characterize and interpret them. An objective common to both realms is the development of a more holistic understanding of interacting organisms that affect plants directly and through ancillary processes such as soil fertility and levels of pests and diseases. An additional aspect consistent in both agriculture and ecology is that plant relationships with members of the belowground community surrounding roots receive far less attention than their aboveground counterparts (Hunter, 2001). Although they are an undoubtedly important and vital facet of plant health, these rhizosphere communities are often overlooked either by convention or because of the difficulty associated with observing, processing, and quantifying cryptic organisms. By identifying organisms that contribute to the complex multitrophic interactions in the soil it becomes possible to infer the existence of underlying processes, such as intraspecific (reproduction, dispersal, mortality) and interspecific (competition, predation) interactions among organisms or responses to environmental heterogeneity (Perry and Dixon, 2002; Ings et al., 2009). Such research allows for advances in both sustainable

agricultural practices and the foundation of plant-based interactions in ecology.

Recent methodological advances in biology, chemistry, and statistics have resulted in unprecedented opportunities in agroecology. The growing availability of molecular genetics methods, public datasets, and free software via online platforms has fostered the use of PCR-based techniques for accurately exploring the diversity of organisms, their relationships, and their functions in ecosystems. Advances in chemical ecology techniques are facilitating the study of belowground signals and cues which play intricate roles between organisms and draw linkages between species and their responses to one another across trophic levels. The characterization and analysis of the spatial patterns of organisms involved in chemical-mediated interactions is one approach to identifying their linkages with one another and with physical attributes of the soil environment. These areas when overlaid allow for the quantification of cryptic organisms and the interpretation of communication between them in different environments.

In this paper, we shall consider selected approaches that can facilitate studies of the subterranean plant environment and emphasize how they can be integrated to enhance biological control of crop pests (Ali et al., 2013; Campos-Herrera et al., 2013). The use of molecular tools to identify and quantify organisms at

multiple trophic levels is providing opportunities to characterize targeted species in complex and cryptic soil systems with precision (Campos-Herrera et al., 2011a,b, 2012). Next generation sequencing (NGS) systems and high-throughput tools can reveal previously unknown organisms and novel functions of soil communities (He et al., 2010; Hackl et al., 2012). In a similar way, soil organisms perceive the environment and communicate with each other by using different physical and chemical signals (Torr et al., 2004; Rasmann et al., 2005; Dillman et al., 2012). In particular, chemical cues soluble in the soil matrix are considered of great interest for the biocontrol potential use in agroecosystems. Improved methods to more accurately characterize flux in mixtures of volatile gases in the soil pores presents the opportunity to understand semiochemical signaling that regulates interactions between plants and organisms in their rhizospheres (Ali et al., 2012, 2013; Robert et al., 2013). Spatial and temporal linkages between soil organisms and their relationships with the abiotic environment are most commonly made with multivariate statistical or geostatistical methods. Spatial analysis by distance indices (SADIE) is a less used method, described here because it uniquely employs spatial and temporal data in a way that also provides fiducial limits to inferences regarding spatial aggregation and the spatial or temporal association between variables. We conclude the paper with an example of how these methods are being used to study how soil food webs might modulate the rates of an herbivore-disease complex across different habitats where citrus is grown in Florida.

MOLECULAR TECHNIQUES FOR CHARACTERIZING SOIL COMMUNITIES AND FOOD WEBS

PCR-based techniques are employed to identify and quantify numerous organisms in complex matrices such as soil, gut content or fecal samples. They are also used with RNA to characterize gene expression in communities (e.g., Mengoni et al., 2005), an application reviewed by Bustin et al. (2009) and van Pelt-Verkuil et al. (2010). Because the same methods are employed to measure all targeted organisms, these techniques have greatly extended our ability to study cryptic species, associations and processes in nature (Carreón-Martínez and Heath, 2010; Campos-Herrera et al., 2011b). PCR methods are used routinely to accurately monitor bacteria, fungi, nematodes, insects, and associated cryptic organisms (Hoogendoorn and Heimpel, 2001; Atkins et al., 2005; Garipey et al., 2007; Wang et al., 2009; Campos-Herrera et al., 2011b; Griffiths et al., 2012; Pathak et al., 2012; Hilton et al., 2013). They are an important alternative to traditional morphological identification, especially in their ability to detect and identify juvenile stages, very small quantities of tissues and even degraded material (Chen et al., 2010; Pathak et al., 2012). The limited taxonomic expertise required is especially important for studies of complex food webs involving organisms at several trophic levels. PCR-based methods also permit direct measurement of interacting organisms, eliminating the need for artificial culturing that can confound population estimates (Chen et al., 2010; Pathak et al., 2012).

Following are descriptions of some PCR-based methods being used to study subterranean communities, along with examples used to illustrate their comparative advantages or disadvantages.

PCR-BASED METHODS USED FOR BELOWGROUND STUDIES IN AGROECOSYSTEMS

Aboveground predator-prey interactions were first studied with PCR-based methods in the early part of this century. These studies employed conventional PCR protocols to investigate interactions between diverse groups such as scarabids and slugs (Dodd et al., 2003) and spiders, hemipterans and lepidopterans (Ma et al., 2005). The methods soon evolved to permit the use of several specific primers simultaneously in the same multiplex reaction (Agustí et al., 2003; Juen and Traugott, 2006, 2007; Traugott et al., 2006). For example, several belowground invertebrates were identified by multiplex PCR as members of a guild that preys on the garden chafer *Phyllopertha horticola* (Coleoptera, Scarabaeidae) (Juen and Traugott, 2007). Chafer content in the predator gut was detectable for up to 24 h post-feeding. Chafer eggs and larvae served as food for these predators, but appeared to be secondary resources. Although conventional PCR protocols have revealed important insights into ecological interactions aboveground in agroecosystems (e.g., Gagnon et al., 2011; Hatteland et al., 2011; Pumariño et al., 2011; Moreno-Ripoll et al., 2012; Romeu-Dalmau et al., 2012), new techniques such as quantitative real time PCR (qPCR) and NGS are naturally displacing the use of conventional PCR-based methods.

The application of PCR techniques to belowground systems required the development of protocols and systems that separated the target organisms from the soil matrix and avoided the co-extraction of certain chemical compounds such as humic acids that can confound with the PCR reaction (van Pelt-Verkuil et al., 2010). Early studies of belowground community structure used denaturing/thermal gradient gel electrophoresis (PCR-DGGE/TGGE) and terminal restriction fragment length polymorphism (PCR-TRFLP). Both techniques have technical similarities and the common objective of detecting differences in DNA sequences without requiring *a priori* sequencing or background information. They use gel resolution to assess differences among samples. Both amplify the target area using primers complementary to the flanking region (universal or more specific). They depict the biodiversity in a sample by producing a profile pattern of target organisms. Nematode, fungal and bacterial community structures in agroecosystems have been characterized using these techniques (Hagn et al., 2003; Nunan et al., 2005; Sato and Toyota, 2006; Dickie and FitzJohn, 2007; Donn et al., 2007; Wang et al., 2009; Griffiths et al., 2012; Bissett et al., 2013; Hilton et al., 2013) (Table 1). The main advantage of TRFLP with respect the DGGE/TGGE technique is the opportunity to compare data from different runs, whereas cryptic bands and intraspecific variation make the comparison of results between gels difficult (Nunan et al., 2005; Pompanon et al., 2012); however, an important advantage of DGGE is the ability to excise and sequence gel fragments. In addition to these methods, conventional PCR-based methods have also been proposed to study belowground predator-prey interactions (Read et al., 2006; Waldner et al., 2013; Wallinger et al., 2013) including protocols for multiplexing (Harper et al., 2005; Eitzinger and Traugott, 2011). However, as in aboveground systems, the use of qPCR and NGS are replacing these previous techniques.

Table 1 | Examples of belowground studies using PCR-based methods for agricultural systems.

Agricultural area/field experiment/material employed	Ecological measurement	Organisms involved	PCR-based method	References
Winter wheat in an arable field (Germany)	Community structure	Fungi	DGGE	Hagn et al., 2003
Grazed grassland (UK)	Community structure	Bacteria	TRFLP and DGGE	Nunan et al., 2005
Arable soil (UK)	Community structure	Nematode	Clone and sequencing	Griffiths et al., 2006
Grassland (Kansas, USA)	Community structure	Free-living bacterivorous nematodes	Multiplex qPCR	Jones et al., 2006a,b
—	Community structure	Nematode	DGGE	Sato and Toyota, 2006
Arable soil, dune sand, coniferous forest, pasture, and moorland (UK)	Community structure	Nematode	TRFLP	Donn et al., 2007
Maize crops (USA)	Predator-prey	Insects	qPCR	Lundgren et al., 2009
Natural restoration and cropping management (China)	Community structure	Bacteria	DGGE	Wang et al., 2009
Grazer pasture and forest (South Carolina, USA)	Community structure	Bacteria	Clone and sequencing	Hamilton et al., 2009
Agroecosystems	Predator-prey	Beetles as predators; earthworms as prey	qPCR	King et al., 2010
Agroecosystems	Predator-prey	Mite as predator; nematode as prey	qPCR	Heidemann et al., 2011
Maize crops (USA)	Community structure	Insects (and plant damage linked)		Lundgren and Fergen, 2011
Citrus groves (Florida, USA)	Predator-prey; food webs	Entomopathogenic nematodes, nematophagous fungi, ectoparasitic bacteria, free-living nematodes	qPCR and nested	Campos-Herrera et al., 2011b, 2012; Pathak et al., 2012
Tillage trial (UK)	Community structure	Nematode	d-TRFLP	Griffiths et al., 2012
Agricultural field (Japan)	Community structure	Nematode	Next generation sequencing	Morise et al., 2012
Abandon field (Netherlands)	Community structure	Nematode	qPCR	Vervoort et al., 2012
Tillage and nutrient additions in wheat cropping experiment (Australia)	Community structure	Bacteria	TRFLP	Bissett et al., 2013
Oilseed rape field trial (UK)	Community structure	Fungi and bacteria	TRFLP	Hilton et al., 2013

Note that we have excluded references corresponding to the development of new molecular methods for particular species, without establishing relationship among other (structure or food web).

Quantitative real-time PCR (qPCR) has been extensively developed for belowground systems (Table 1). As with conventional PCR protocols, qPCR uses species-specific molecular markers to identify and quantify target species. The advantage of qPCR compared to conventional PCR is that the order of magnitude increase in the amplification potential of tiny quantities or even degraded material. This method has been successfully employed to detect cryptic organisms and assessed diversity and interactions among bacteria, fungi, nematodes, insect and plants from above and belowground systems (Atkins et al., 2005; Jones et al., 2006a,b; Zhang et al., 2006; Lundgren et al., 2009; King et al., 2010; Campos-Herrera et al., 2011a,b, 2012; Heidemann et al., 2011; Lundgren and Fergen, 2011; Pathak et al., 2012; Vervoort et al., 2012). Nevertheless, qPCR also has important limitations. For example, not all species contain

sequences in the genes currently used to discriminate taxa that are suitable for valid primers, or in some cases probes. Some groups, such as free living nematodes, frequently lack adequate numbers of published sequences for comparison when designing molecular probes, so this method can finally be used for some undescribed, regional species (Jones et al., 2006a,b). In such cases, it may still be useful to use qPCR to measure broader taxa with similar ancestry/behavior rather than individual species (Campos-Herrera et al., 2012). The worldwide tendency to increasingly publish sequences in public domains is rapidly increasing the opportunity to design species-specific primers/probe combinations for a diverse array of organisms. Another current issue for qPCR technology is the reproducibility and viability of multiplexing in order to reduce the expense of these protocols. Often, the use of several species-specific

markers in the same reaction reduces the amplification of target organisms if the initial quantities are minimal. Although some multiplex reaction have been described (Jones et al., 2006a; Berry et al., 2008), it is advisable to perform single reactions for precise estimation because competition between target DNAs or development of interacting primers (primer-dimers) can impede or produce a false signal (Zijlstra and van Hoof, 2006; Berry et al., 2008). This approach is particularly appropriate when evaluating samples with quarantine requirements.

DNA barcoding is a PCR-based method used extensively in studies of belowground systems, especially to assess community structure (Table 1). Two main approaches have been used in the last decade: clone-sequencing (i.e., insert the PCR product, usually in a plasmid, and get the sequence) and NGS, also called high-throughput sequencing, which does not require cloning. These two techniques obtain the full sequence of the products derived with selected primers and both are quantitative and species specific. Barcoding approaches are rapidly expanding the ability to study concomitant organisms in soil samples, for example, to compare community structure under different management programs. Important limitations include high reaction and equipment costs (that are rapidly declining) and the complexity of the amplicons that are generated. A great deal of time and experience are required to organize and assess the validity of hundreds or thousands of generated amplicons. Increasingly, new bioinformatic skills are used as substitutes for zoological-taxonomical expertise in order to provide an ecological context for the molecular operational taxonomic units (MOTUs) that are identified from multiple-alignments and classified by their associations with one another (Blaxter et al., 2005). Of course, if taxa defined by MOTU methods are used for ecological purposes, they must eventually be associated with sequences from specimens identified by Linnaean taxonomy.

Other new PCR-based systems emerging during the last 5 years are some that use microfluidic systems (also microfluidic droplet PCR) (Zhu et al., 2012; Chang et al., 2013). Although these new protocols were developed for human clinical diagnostics, their application in agroecological studies is only a question of time. This technology incorporate chips with microfluidic technology that allows nucleic acid amplification in a compartmentalized reaction in a minimal volume. Their critical advantages are reduced PCR processing time and reaction volume, and substantially reduced cost (Zhu et al., 2012; Chang et al., 2013).

FUNDAMENTALS OF DEVELOPING qPCR, DNA BARCODING AND NEXT GENERATION SEQUENCING PROTOCOLS

The specificity of qPCR is a consequence of the species-specific primers (and probes, if used) that are developed for target species. The biochemistry involves linking fluorescent products to double stranded DNA (e.g., SYBR Green®) or designing specific fluorescent probes (e.g., TaqMan® probes) that provide greater specificity to the primers. Both systems produce increased fluorescence that can be quantified during PCR cycling when used with the targeted species.

In the case of NGS, there are international efforts to define target areas for barcoding standards (i.e., CboL, <http://www.barcodeoflife.org>); however, exceptions may be desirable depending on objectives (Valentini et al., 2009; Pompanon et al., 2012).

The clone-sequencing method has provided outstanding results in studies of distribution, diversity and community structure (i.e., Griffiths et al., 2006; Hamilton et al., 2009), but the technique is more time consuming and costly than new methods of direct amplifications in NGS (Pompanon et al., 2012; van der Heijden and Wagg, 2013). Extensively used NGS platforms include: 454 GS FLX (Life Sciences, Roche), HiSeq 2000 by Illumina (Solexa Technology), and AB SOLiDv4 (Agencourt technology). Each technology is based on a different sequencing approach: 454 GS FLX uses pyrosequencing, HiSeq 2000 sequences by synthesis and AB SOLiDv4 by ligation and two-base coding, and all of these methods produce reads of >98% accuracy (Liu et al., 2012). The maximum output of data is generated by HiSeq 2000 (600 Gb) and also is compatible with reads until 3 G for the cheapest cost, estimated as \$0.07 per million bases. However, 454 GS FLX has the advantage of a fast read time and can run in just 24 h (Liu et al., 2012). Therefore, the selection of the optimum platform depends on the experimental conditions and requirements (see Glenn, 2011 and Liu et al., 2012).

Several concepts need to be considered when PCR-based experiments are designed for a new organism or taxonomic group. Because real time qPCR and DNA barcoding using NGS are currently the most widely used methods for the study of belowground interactions in agroecosystems, important differences in the requirements for both methods are summarized in Table 2. Technical aspects about the development of these protocols were extensively detailed in Bustin et al. (2009); Campos-Herrera et al. (2010); van Pelt-Verkuil et al. (2010) and Pompanon et al. (2012). The first step is selecting the most appropriate method to address the question. Both techniques require relatively costly reagents. By contrast, while many institutions are able to invest in qPCR equipment used in individual programs, NGS facilities are much more expensive and tend to be supported by multiple users in an institution. The type of data generated by both systems is different, although both can be perfectly complementary. Real time qPCR can detect and quantify (expressed as numbers, copies, quantities, depending on the standard curve units) target organisms accurately. The limitation of this method is that you only find what you are looking for, so it is not possible to detect species for which the appropriate primers (and probes) have not been designed and used. On the other hand, NGS systems provide all the sequences derived from primers selected to amplify a target taxonomic group. Multiple sequences, MOTUs, are then identified that correspond to known and unknown species, in theory. NGS approaches require special attention to avoid “chimera” sequences. Some tools such as QIIME can analyze the data while detecting chimera sequences (Caporaso et al., 2010). If the effects of different agroecosystems on the biodiversity of a target group are of interest, NGS approaches can provide meaningful information. They are also useful when there is no available information regarding the key players in an agroecosystem (Pompanon et al., 2012). However, when there are limited numbers of known organisms interacting in a somewhat predictable manner, qPCR protocols can produce the relevant data in a short time and at the least cost.

Table 2 | General considerations for the development of PCR-based methods (from Bustin et al., 2009; Campos-Herrera et al., 2010; van Pelt-Verkuil et al., 2010; Pompanon et al., 2012) and comparison between the two most extended methods in belowground studies: real time qPCR and DNA barcoding using next generation sequencing.

Experimental steps	Concept	Common considerations	Real time qPCR	Next generation sequencing
Design " <i>in silico</i> "	Selection of the target sequence	Adequate and meaningful for the study; check availability in the system to be able to compare with known species	ITS rDNA, D2D3, COI	SSU, LSU, sometimes ITS, customized
	Coverage and resolution	Range of taxa susceptible to be amplified	Species-specific	Generalistic, amplify broad taxonomic groups
	Primer and amplicon equilibrium	The length and composition of the primers will affect the specificity of the amplification; secondary structures should be avoided; the PCR product, the amplicon, should be into the range for optimal amplification	80–250 bp	200–600 bp, depending on the platform
	Amplification efficiency	The efficiency might depend on the quality of the DNA (degraded, inhibitors presented, etc.)	Might be improved by adding some reagents (i.e., BSA or DMSO) or by diluting the DNA	
Sample preparation	Design and sampling strategy	Include biological and technical replicates; tagging and multiplexing approaches available	Different dyes for multiplexing	Different molecular tags to separate treatments
	DNA extraction	Multiple kits available; desirable, verify the quality and quantity by electrophoresis or spectrophotometric systems (nano-drop)	–	–
Optimize reaction	PCR conditions	Experimental establishment of annealing temperature, time for extension, number of cycles; check for possible inhibitors	Important the number of cycles in nested qPCR experiments	–
	Sensitivity and specificity	Check the lowest number of amplicons detected of the target species/taxonomic group (dynamic range)	Important for quantification. Serial dilutions of the target DNA will serve for defining the limit of accurate detection for our standard curve	Important to establish the minimum taxonomic unit detected
Data analysis and validation of the experiments	Type of generated data	Units or type of quantification	Detection and quantification of the target organisms. Absolute quantification is possible if a standard curve is included in the run; relative quantification is possible among target species	Molecular operational taxonomic units (MOTUs). Special care need to be taken for the detection of "chimera" sequences, as a subproduct of amplification that provide a non real sequence
	Taxa assignment	Identification with species or taxonomic group with known identity and possible defined ecological traits	Amplifications are compared with the positive control, the DNA from the known target organisms; additionally, postsequencing analysis can be performed and comparison with reference database	Comparison with reference database (i.e., GenBank, IBOL, EMBL, DDBJ or customized for specific studies)
	Repeatability, reproducibility, and accuracy	Measurement of the intra-assay variance, inter-assay variance and difference between experimental measurement and actual values, respectively	Critical to compare measurements from a run to another	Desirable, although costly

Although molecular methods are alternatives to conventional methods that rely on morphological identification, they should often complement rather than replace traditional methods. For example, Campos-Herrera et al. (2011a) found a strong positive correlation between known quantities of nematodes and numbers measure by qPCR, whereas Griffiths et al. (2006) suggest that molecular techniques underestimated certain groups compared to observations using morphology. Gibb et al. (2008) suggested that congruence of morphology and molecular characterization can be confounded by steps that reduce the recovery efficiency or the PCR amplification. The study of the arbuscular mycorrhizal fungi (AMF) provides another good example of why both approaches complement each other, because the biological species concept of AMF is still unresolved (Sharmah et al., 2010).

METHODS FOR DETECTING HERBIVORE-INDUCED CUES BELOWGROUND

Plant volatile organic compounds (VOCs), which mainly comprise terpenoids, fatty acid derivatives, phenylpropanoids and benzenoids (Dudareva et al., 2004) have been the center of intensive studies of plant-herbivore-predator interactions for more than two decades (Dicke and Sabelis, 1988; Turlings et al., 1990). VOCs blends can be complex, comprising hundreds of compounds, some of which are not produced by intact or mechanically damaged plants but are synthesized *de novo* in response to herbivore attack (Turlings and Wäckers, 2004; Mumm and Dicke, 2010). Indirect defense is described as a phenomenon of plant defense strategy where plants attract enemies of the herbivore when “attacked” (Dicke and Sabelis, 1988; Turlings et al., 1990). Although a diversity of root derived exudates can play a role in the rhizosphere community (Badri and Vivanco, 2009), root volatiles can and have recently been shown to play a role in contexts that might change the abundance, location, and diversity of organisms in the soil in a manner similar to signals aboveground (Ali et al., 2013).

Research on plant VOCs produced after insect herbivory has been dominated by studies aboveground (see Kessler and Morrell, 2010), probably due to methodological constraints related to subterranean investigations. More recently, the number of studies showing that herbivore induced belowground volatiles trigger predator attraction in the soil has increased. Belowground investigations began with two key studies which demonstrated for the first time that unknown cues were responsible for attracting entomopathogenic nematodes to insect damaged roots (Boff et al., 2001; van Tol et al., 2001). Tritrophic interactions with belowground herbivore-induced plant volatiles (HIPV) signaling have been described both in agricultural systems (Rasmann et al., 2005; Ali et al., 2010, 2011) and in a natural context (Rasmann et al., 2011). Because of the extensive literature focused on aboveground plant volatiles and methods, here we describe the techniques used for belowground assessment, focusing our attention on the advances related to agroecosystems and their application in the control of insect pests.

Factors in addition to VOCs are known to influence both behavior of herbivores and their natural enemies belowground. Root exudates also influence spatial distribution of root feeders (Johnson and Gregory, 2006; Johnson and Nielsen, 2012; Gfeller

et al., 2013). Some recent techniques were developed to measure root exudates that influence herbivore behavior using LESA (liquid extraction surface analysis) (Robert et al., 2013). Although these studies are sound and contribute to belowground interactions we shall focus here on cues that have been used in agricultural/biological control contexts and have been detected with non-destructive techniques, sampling/manipulating behavior in a manner that allows for large scale patterns in field research. Thus, far VOCs have been a major candidate for use in biological control and also to explain multitrophic cascades both above and below ground (Kaplan, 2012a,b; Poelman et al., 2012; Ali et al., 2013).

METHODS FOR DETECTING BELOWGROUND HERBIVORE-INDUCED VOLATILES

The technical difficulties associated with dynamics of the soil ecosystem have been a major limitation in studying belowground multitrophic interactions. Soil is a complex, tri-phasic medium making the analysis of individual factors and their interactions considerably difficult, and hence, most research has been based on *in vitro* analysis of individual factors (Rasmann et al., 2012a). Researchers willing to study factors associated with roots signals are often focused on the study of (i) root diffusates (often used to convey non-volatile substances diffusing through the soil and establishing a gradient), (ii) root leachates (method of obtaining an extract from the roots, more than it does to the solution itself), and or (iii) root exudates (most often restricted to liquids that gradually “ooze” from its source, but can be applied to volatiles as well). New approaches to evaluate root volatiles have only recently been developed and applied in researches on chemical and evolutionary ecology. As mentioned earlier, all of these can serve as signals and cues for herbivores and their natural enemies. However, in this review we focus on HIPVs as they are often shown to be involved in multitrophic interactions that include natural enemies and have been shown to be detectable from intact plants. Although methods exist for the evaluation of additional exudates from intact plant roots (i.e., polydimethylsiloxane (PDMS) for Solid phase root zone extraction (SPRE), see Mohny et al., 2009), thus far they have not been used to evaluate induction related to root herbivores.

Rasmann et al. (2005) provided for the first time the evaluation and identification of indirect volatile defenses of maize roots using solid phase micro-extraction (SPME), which is a method of sampling volatiles without the use of solvents. Briefly, an adsorbent-coated fused silica fiber (with properties similar to a gas chromatography column) can collect volatile compounds from the headspace of a sample. The volatile compounds once fixed to the SPME fiber can then be thermally desorbed in an injection port of a gas chromatograph. Then, these can be further analyzed and/or identified when coupled with known standards or libraries of mass spectroscopy. Rasmann et al. (2005) crushed flash-frozen roots samples, either fed-upon or non-fed-upon into a fine powder to analyse the effects root herbivory had on the plant produced VOCs. This powder was then exposed to the SPME fiber, allowing all the volatiles that had accumulated in either treatment to be sampled and compared with Gas Chromatography-Mass Spectrometry (GC-MS). SPME is a rapid and simple extraction method that does not require the use of

solvents and its detection limits can reach parts per trillion (ppt) levels for certain compounds (Pawliszyn, 1997). However, one of the limitations of this technique is that it is a destructive method of sampling root material: both the plant and herbivore must be separated and volatiles from this interaction can only be examined after harvesting and crushing the plant tissues.

Ali et al. (2010) were able to non-destructively sample belowground herbivore induced volatiles from citrus roots using another technique, a flow-through dynamic sampling coupled with adsorbent traps. In this case, volatiles can be collected and extracted by elution of an adsorbent with low boiling point solvents. Adsorbent traps are usually made of glass tubes filled with the granulated adsorbent, held in place by stainless steel mesh, glass wool plugs, or Teflon fitted rings. By connecting the adsorbent trap to a vacuum pump and pulling air through glass chambers (containing intact citrus plants either with or without feeding larvae) Ali et al. (2010) were able to sample volatiles associated with belowground herbivory *in situ* using a non-destructive approach. The volatiles collected on this trap are rinsed using a non-polar (e.g., methanol, hexane, dichloromethane) solvent and analyzed with GC-MS. This methodology allows for the sample to be retained in a solvent, which can be analyzed multiple times. Additionally, the solvents containing root volatiles were also tested in sand-filled two-choice bioassays chambers. By using this approach, Ali et al. (2010, 2011) found evidence for entomopathogenic nematode attraction to volatiles from infested citrus roots. Moreover, a soil probe was recently used to sample soil volatiles in Florida citrus groves at a depth of 20 cm (Figure 1) (Ali et al., 2012), which constitute an important advance in the study of volatiles *in situ*. This soil probe was developed to collect the compound on adsorbent traps by using a vacuum pump to pull air from the soil. Recently, a non-destructive method similar to the one used by Ali et al. (2010) to collect root volatiles was also successfully used to detect (*E*)- β -caryophyllene from maize roots (Robert et al., 2013).

Proton-Transfer-Reaction Mass-Spectrometry (PTR-MS) is an additional non-destructive analysis that has recently been applied to the detection of belowground trace HIPVS in real-time (Danner et al., 2012). Briefly, the PTR-MS technology pumps air to be analyzed continuously through a drift tube reactor, and a

fraction of the VOCs is ionized in proton-transfer reactions with hydronium ions (H_3O^+). The ionized molecules usually form a protonated molecular ion $[\text{M}+\text{H}]^+$, where “M” is the molecular mass of the parent molecule (for detailed descriptions of the PTR-MS technology see Hansel et al., 1995; de Gouw et al., 2003; Boamfa et al., 2004). By using this technique, organic compounds such as ketones, aldehydes, alcohols, oxygenated aromatic and aliphatic compounds will be readily protonated (Danner et al., 2012). Proton transfer utilizes a soft ionization method, so it generally leads to less overall fragmentation of the product ions, which is a tremendous advantage, and also the mass of the product ion equals the VOC mass plus one (de Gouw and Warneke, 2007; Danner et al., 2012). At the end of the drift tube, the reagent and product ions are measured by a quadrupole mass spectrometer. The product ion signal is proportional to the VOC mixing ratio. PTR-MS can monitorize numerous VOCs with a high sensitivity (10–100 pptv) and rapid response time (1–10 s) (de Gouw and Warneke, 2007), although regular calibration with the gas mixture is required for accurate and reproducible quantification of trace gases (Danner et al., 2012).

There are many advantages of the PTR-MS method. The more common methods of collecting plant VOCs on to filters with polymer adsorbents makes it necessary to preconcentrate the sample before analysis by collecting volatiles during a range of minutes to hours; this reduces the ability to resolve the timing of VOC emission measurements (Danner et al., 2012). Furthermore, solvents used to elute VOCs from the adsorbents may introduce contamination before GC analysis. A major disadvantage is that PTR-MS only determines the mass of product ions, which is, of course, not a unique indicator of the VOC identity. It is clear that isomers cannot be distinguished, and the interpretation of mass spectra is further complicated by the formation of cluster ions and the fragmentation of product ions. A significant experimental effort has therefore been made to characterize the specificity of PTR-MS for different VOCs.

Early studies using PTR-MS already has shown how it can be successfully applied to analyze VOCs produced by aboveground (wounded) plant parts (Fall et al., 1999). Two recent examples show its promise as a technique to evaluate belowground interactions in preliminary PTR-MS results on herbivore-induced root responses in *Brassica* species. Danner et al. (2012) monitored VOCs emanating from roots of potted turnip plants (*Brassica rapa* subsp. *rapa* var. Nancy) during infestation with a belowground herbivore, *Delia radicum*, the larvae of the cabbage root fly. The resulting mass scans found that the intensities of several molecular masses were enhanced in root fly infested *B. rapa* roots (Danner et al., 2012). In a second example, Danner et al. (2012) monitored the induction of VOCs in *Brassica juncea* roots after infestation with *Delia radicum* in real-time and compared it to a control treatment. Initially, they detected only a low emission rate of compounds, which steadily increased with longer feeding times of the root flies. In control plants, the VOC emissions remained at a very low level, allowing a clear distinction between control and infested plants within a few hours after infestation (Danner et al., 2012).

In general, these techniques are informative and effective in different manners. For example, the non-destructive sampling

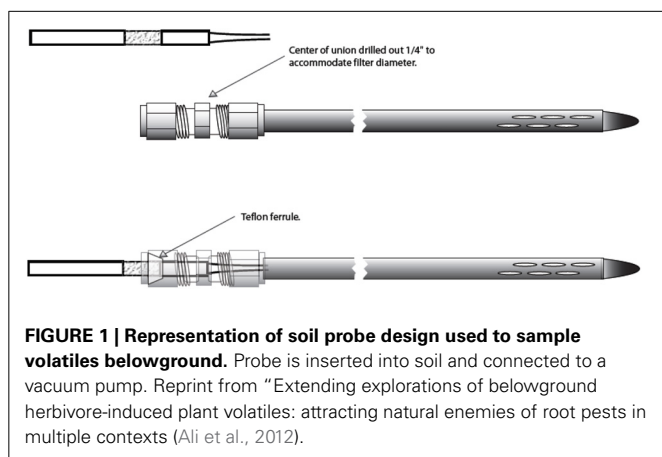


FIGURE 1 | Representation of soil probe design used to sample volatiles belowground. Probe is inserted into soil and connected to a vacuum pump. Reprint from “Extending explorations of belowground herbivore-induced plant volatiles: attracting natural enemies of root pests in multiple contexts (Ali et al., 2012).

techniques are useful in evaluating belowground interactions *in situ*. Also, they may potentially prove useful in additional contexts. However, the properties of the surrounding soil might interfere and, hence, make resolution difficult with the potential for significant background. In this case, SPME can be considered, since eliminates such background, although it can introduce complications from tissue maceration (enzymes or oxidation can rapidly change the chemical profile) and may not accurately represent the blend released from intact living organisms (Tollsten and Bergström, 1988; Heath and Manukian, 1992). The combination of techniques and refinements of approaches might produce the best resolution for the target individual system.

EXAMPLES OF BELOWGROUND HERBIVORE INDUCED PLANT VOLATILES IN AGROECOSYSTEMS

The considerable advances in research on molecular mechanisms and ecological signaling of insect herbivore induced VOCs launch promising prospects of manipulating the release of these compounds in order to enhance crop protection. Encouraging examples from both laboratory and field experiments support this approach to develop novel ecologically drive crop protection strategies. On the agricultural side, at the moment, the best-known belowground tritrophic interaction is the maize system, described for the first time by Rasmann et al. (2005). When the larvae of the western corn rootworm, *Diabrotica virgifera virgifera*, attack the roots, European maize varieties emit in soil the sesquiterpene (*E*)- β -caryophyllene (E β C) (Rasmann et al., 2005; Kollner et al., 2008). This compound is a highly attractive HIPV to the entomopathogenic nematode *Heterorhabditis megidis* in the laboratory as well as in the field (Rasmann et al., 2005; Kollner et al., 2008; Hiltbold et al., 2010a). Several laboratory and field experiments with various corn lines and synthetic compounds have shown that E β C is an ideal compound to diffuse through the complex belowground soil matrix. In fact, it is among the less costly terpenoid that could be travelling within the soil (Hiltbold and Turlings, 2008), and that is under selection (Kollner et al., 2004, 2008).

Another example of highly complex volatile blends in agroecosystems was described by using the roots of cotton (*Gossypium herbaceum*) and the larvae of the chrysomelid beetle *Diabrotica balteata*. After feeding by this generalist root feeder, cotton plants were scored to emit >10 compounds, where at least 7 terpenoid volatiles were observed (Rasmann and Turlings, 2008). The sesquiterpenoid aristolene was discussed as being a good candidate for playing a major role in *Heterorhabditis megidis* nematode attraction, although future studies might confirmed it (Rasmann and Turlings, 2008). These authors also tested the nematode preference against damaged roots of cowpea (*Vigna unguiculata*) plants. In contrast to the other crops, corn and cotton, cowpea plants emitted almost undetectable amounts of volatiles that also resulted in lower nematode attractions (Rasmann and Turlings, 2008).

More recently, Ali et al. (2010) have demonstrated that citrus roots upon feeding by the root weevil *Diaprepes abbreviatus* emit several terpenes in the surrounding soil. Using belowground olfactometers Ali et al. (2010, 2011) found that the entomopathogenic nematodes were significantly more attracted

to citrus roots infested with the insect larvae than by roots mechanically damaged or pots containing only soil. However, Ali et al. (2011) observed that insect induced roots of citrus tree could also attract the plant-parasitic nematode *Tylenchulus semipenetrans*. Consequently, this may reduce the exploitation of citrus induced VOCs emission in biological control strategies targeting *Diaprepes abbreviatus* if rootstocks are not truly resistant to *T. semipenetrans*. Most recently, Ali et al. (2012, 2013) has combined qPCR methods with the field application of root larvae induced volatiles to evaluate attraction of multiple nematode species and nematophagous fungi (NF). This research shows that multiple species of naturally occurring entomopathogenic nematodes as well as “hyperparasites” of entomopathogenic nematode killed cadavers are responding to this cue (Ali et al., 2013).

APPLICATION IN AGRICULTURE: MANIPULATION AND ENHANCEMENT OF BELOWGROUND SIGNALS

At this moment, there are very few published examples of cue-based behavioral manipulation in belowground contexts. The use of natural products to enhance biocontrol is typically compatible with integrated pest management; deploying HIPVs aboveground by controlled release dispensers has been shown to increase plant recruitment and retention of beneficial parasites or predators (Thaler, 1999; James and Grasswitz, 2005). In an analogous belowground investigation, entomopathogenic nematode infection of *Diabrotica virgifera virgifera* larvae was increased by spiking soil surrounding maize roots with the HIPV, E β C (Rasmann et al., 2005). Ali et al. (2012) has recently increased mortality of root pests in the field by enhancing host location of naturally occurring entomopathogenic nematodes in citrus and blueberry crops with the application of the citrus root volatile, 1, 5-dimethylcyclodeca-1, 5, 7-triene (pregiejere).

After identification of herbivore-induced compound attractive to natural enemies, it is possible genetically manipulate the plant in order to (i) make a plant more attractive to beneficial predators or parasitoids or (ii) to restore the phenotype that was lost due to natural or human selection (Rasmann et al., 2012a). The first examples of such an approach are aboveground (Kappers et al., 2005). More recently, the terpene synthase gene TPS23, which is responsible for the synthesis of E β C, has been identified in maize (Kollner et al., 2008). Most of the European maize varieties and Teosinte produce this sesquiterpene whereas American varieties do not (Rasmann et al., 2005; Kollner et al., 2008). This indicates a shift in the gene activity through breeding selection (Kollner et al., 2008). Degenhardt et al. (2009) restored the ability of maize to recruit entomopathogenic nematodes by inserting a TPS23 gene from *Origanum vulgare* into a non-producing maize line, demonstrating in a field experiment that the transformed maize line was significantly more attractive for the entomopathogenic nematode *Heterorhabditis megidis* compared to the wild type, leading to increased protection for transformed plants. This constituted the first demonstration in the field that plant genotype engineering could enhance biological control.

Entomopathogenic nematodes appear as good candidates for an inundative biological control strategy. At the moment, there are knowledge of key attractants for specific entomopathogenic nematodes species (Hallem et al., 2011). (Hiltbold et al., 2010a)

evaluated whether selection for enhanced responsiveness to the crucial root signal E β C could improve the efficiency of these nematodes in controlling the larvae of the chrysomelid beetle *Diabrotica virgifera virgifera*. Using root-zone olfactometers, a population of the nematode *Heterorhabditis bacteriophora* was successfully selected. Originally, this nematode population did not respond to attraction effect by the emission of E β C (Hiltpold et al., 2010b) even though its effectiveness in controlling *D. v. virgifera* larvae is high (Kurzt et al., 2009). After the selection process, the new population responded much better to E β C in laboratory experiments and also was able to control the pest in the field significantly better in presence of the belowground signal (Hiltpold et al., 2010b). Interestingly, the establishment and the persistence in the field were not influenced by the selection process (Hiltpold et al., 2010b). Here we see the great potential of selecting beneficial organisms for a better and faster response, which also resulted in higher infection rates. Even with some constraints, such as knowledge of key compound/blends, and the laborious selection process, selecting for specific nematode populations could be combined with selection of more attractive plant genotypes, making biological control of insect pests a success (Rasmann et al., 2012a,b).

Recent advances are showing the potential of belowground organisms to interact with aboveground food webs. In this regard, Pineda et al. (2013) have evaluated the mechanism involved in interactions between a beneficial rhizobacterium (*Pseudomonas fluorescens*) and a parasitoid (*Diaeretiella rapae*) by using the model system *Arabidopsis thaliana*-*Myzus persicae*. In laboratory studies, they combined ecological, molecular, and chemical approaches to study how the rhizobacterial colonization modified the complex composition of the HIPV. They observed that the compounds produced by the rhizobacteria-aphids-plant treatment negatively influenced the behavior of the aphid-parasitoid when compared with the effect of the blend induced by just aphid-plant. These authors have demonstrated that the non-pathogenic rhizobacteria effect on parasitoid activity is mediated by jasmonic acid pathways that associate with plant volatile production. In contrast, work by Robert et al. (2013) has found that the production of an HIPV which is beneficial to the protection of the plant's roots, was simultaneously found to attract aboveground herbivores. Thus, manipulation of these blends and studies in more natural conditions might provide additional insight on the complex multitrophic interactions occurring above- and belowground, and it should be acknowledged the effects and responses in one subsystem of the plant can have very different roles on other systems.

METHODS TO MEASURE SPATIAL STRUCTURE OF ORGANISMS IN FIELD AGROECOSYSTEMS

During the past several decades, new technologies and spatial statistics offer a number of tools for point pattern analysis, which provide improved detection and characterization of spatial heterogeneity such as gradient or clustering (Perry et al., 2002). Earlier two-dimensional maps of spatial patterns were developed for plants and for relatively immobile organisms (Diggle, 1983). Meanwhile, the spatial information of mobile organisms was restricted to counts in traps at specific locations (Perry, 1995).

Initially, the methods used to describe spatial patterns focused on the intensity of aggregation and were based on the relationship between the sample mean and variance (Taylor, 1961; Iwao, 1968). Spatial information provided by these methods was criticized by Perry and Hewitt (1991) for not considering geographic location of each sample unit, and in consequence, these indices could not be used for comparing or mapping the spatial patterns of populations.

Currently available technologies (i.e., such as the Stanford Geostatistical Modeling Software (S-GeMS) (<http://sgems.sourceforge.net>) or the TerraSeer Space-time Information System (STIS) (which public domain software is: <http://www.terraseer.com/products/atlasviewer.html>) combine the use of geostatistics, as a statistical procedure that uses sample values and locations simultaneously for characterizing spatial patterns and estimating values at unsampled locations (Clark, 2001) and global positioning systems (GPS) to determine the location of each sample unit (Goovaerts, 1997, 1999). Notwithstanding the utility of geostatistical procedures to characterize spatial patterns, they do not provide tests to assess the statistical significance of the estimated patterns. To overcome this limitation, Perry and Hewitt (1991) developed a new method, SADIE, which uses the spatial information in the sample in ways that permit inference testing in order to make the information more understandable from a theoretical biological perspective. Despite this advantage, SADIE is used infrequently compared to geostatistical methods for community studies. Accordingly, we describe the method here with some examples of its application to agroecology.

SADIE: BASIC CONCEPT AND EXTENSIONS

SADIE is computed with free software, SADIESshell 1.22 (Kelvin F. Conrad and IACR-Rothamsted, 2001) available for download at <http://www.rothamsted.bbsrc.ac.uk/pie/sadie/>. The method evolved from a spatial analysis based on a single index to an analysis of count data that are spatially-referenced with two coordinates (x, y) that can be irregularly spaced and not necessarily on a grid (Perry, 1995, 1998). The development of new SADIE indices and maps, increased the ability to characterize the spatial information in a sample (Perry et al., 1999) and to test the association or dissociation of spatial patterns from two sets of data (Perry and Dixon, 2002) (see chronological development, a brief description of indices and graphical displays provided by this spatial analysis in Table 3).

SADIE uses a transportation algorithm to move individuals from their initial positions to new ones, using the minimal possible total distance or "distance to regularity" (D), in order to convert a given spatial pattern to one of regularity (Perry, 1995). The flows of individuals from areas of relatively high population density to areas where population density is relatively low could be visual observed using an "initial and final plot" (IAF) (Perry, 1995). To assess the magnitude of aggregation, the observed D is compared to those for large numbers of randomly distributed permutations of the counts observed among the sample units. The ratio of the observed D to the mean of D s from the simulated distributions provides an "index of aggregation" (Ia). A value of $Ia = 1$ suggests a spatially random pattern, $Ia > 1$ suggests an aggregated pattern and $Ia < 1$ indicates a regular pattern

Table 3 | Summary of the main indices and graphical displays provided by SADIE analysis.

SADIE indices	Definition	Graphical display	References
Distance to regularity (D)	Measures the minimum effort that the individuals in the sample would need to expend to move to an arrangement where there was an equal number in each sample unit	Initial and final plot (IAF)	Perry, 1995
Aggregation index (Ia)	The ratio of D to the mean of the simulated distribution		Perry, 1995, 1998
Cluster indices (vi and vj)	vi measures the degree to which the unit contributes to clustering as a member of a patch vj measures the degree to which the unit contributes to clustering as a member of a gap	“Red-blue plots” • Contour map • Vector flow plot • Empirical distribution function plot of ranked average outflow/inflow distances (EDF)	Perry et al., 1999
Local and global association index (X)	Local association index is calculated by comparing the cluster index (vi or vj) for each set of data at the same sample unit The global index (X) is calculated as a mean of the local indices	Map of local association and dissociation	Perry and Dixon, 2002

(Perry, 1998). A significance test (Pa) for the probability that the observed data is no more aggregated than expected from a random permutation of the counts is provided by determining the proportion of simulated D s with values less than that of the observed D (Perry, 1998).

Perry et al. (1999) described two forms in which aggregation of count data occur: (i) as patch clusters when there is a high density of counts near to one another or (ii) as gap clusters comprising of relatively small or zero neighboring counts. The SADIE analysis provides a local index of clustering (vi or vj) for each sample point, by employing randomizations in which the observed counts are permuted amongst the sample units. Sample units with counts greater than the overall mean are assigned a patch cluster index (vi), which by convention is positive; while units with counts less than the mean are assigned a gap cluster index (vj), which by convention is negative. Additionally, SADIE provides for patches an overall index vi and associated probability Pi and for gaps an overall index vj and associated probability Pj . Heuristic threshold fixed values of 1.5 and -1.5 for vi and vj , respectively, discriminate sampling units associated with index values >1.5 (patches) from sampling units with index values <1.5 (gaps) (Perry et al., 1999). Each local clustering index (vi and vj) may be contoured by interpolation and mapped using the “red-blue plot” methodology, which enables the characterization of clusters with respect to type (patch or gap), number, position and size (i.e., % or m^2 of each cluster type) for a given site (Perry et al., 1999). For convention, patches are represented on maps by contoured areas shaded red, where all neighboring sample units have cluster indices $vi > 1.5$; gap contour areas where all of the units have indices $vj < 1.5$ are shaded in blue. Unshaded areas represent locations where counts are arranged effectively at random. SADIE software also provides information to plot other graphical displays such as “vector flow” plots and “empirical distribution function plots of ranked average outflow/inflow distances” (EDF) (Perry et al., 1999), although these are less represented in applied studies.

A very useful extension of SADIE measures the spatial association between patterns for two sets of count data, when they are sampled at the same spatially-referenced units (Perry and Dixon, 2002). This method is based on the correlation between the cluster indices of the two data sets at each sample unit, which is used to obtain an overall index of association (X), calculated as a mean of the local clustering index (vi or vj) of each sample unit. The X index describes the degree of association or dissociation between the two populations; these also provide tests (P) (Dutilleul, 1993) that may be mapped. In cases when the clustering indices of two populations compared at the same sampling site are both vi or both vj , the SADIE program assigns a positive index, indicating a local association for the sample site. Conversely, if the index of one population is vi and the other is vj , then the SADIE index will be negative, indicative of local dissociation. This test is a useful tool to identify interspecific interactions or responses to environmental heterogeneity (Perry and Dixon, 2002). A temporal sequence of “red-blue plots” (for a single set of data) and SADIE maps of local association (for two sets of data), can also provide information about temporal changes, when the same species was/were sampled at the same sample units on successive occasions.

SADIE LIMITATIONS AND ALTERNATIVES

Although SADIE is used increasingly for agricultural studies (see examples below), the methodology has some drawbacks. One of these is the analysis of dispersal of individuals from a single focus (P). To solve this problem, Korie et al. (1998) suggested the use of other statistics that allow the description of movement. Previously, for datasets in which spread was known to have originated from a central source, Perry (1995) used the SADIE (Ia) as well as the statistic δ defined as the distance between the centroid (C) (representing the average position of individuals in the experimental plot) and the known focus. Thus, δ is used to quantify the displacement of the entire organism population and represents the degree to which the counts occupy the edge rather

than the center of the arrangement. To quantify the spread of the population about its displaced position, they defined the statistic Φ as the average of the squared distances of the position of each individual to the centroid (C). The statistic Δ was defined as the average distance of individual positions from the central release focus (P) and was used to quantify the total distance moved from P . Similarly, to characterize movement using aphid data, they used the statistics γ , to quantify the relative degree of movement between and within rows of the plot. When the insects moved entirely along a row, the value of γ would be 1; if their movements were entirely perpendicular to the rows, then γ would be 0, and if there was completely random movement the γ -value would be 0.5. These improved indices were used to successfully analyze dispersal from a single focus of different datasets for insects and plants (Korie et al., 1998; Smyrnioudis et al., 2001; Diaz et al., 2012). However, for datasets with unknown focus, which represent an invasion from the edge of a sample area, Korie et al. (1998) discussed the backtracking algorithm proposed by Perry (1995) to estimate the focus of a given cluster of individuals.

Another limitation detected in SADIE analysis discussed by Xu and Madden (2003) concerns the edge effect and the detection of small clusters in elongated areas. They demonstrated that this problem occurs because the local clustering indices (vi and vj) are mathematically related to the (Ia). A new method named MAPCOMP (MAP COMParison) was proposed by Lavigne et al. (2010) to analyse the spatial patterns of count data, and results were compared to those of the red-blue plot analysis of SADIE. MAPCOMP is based on permutation tests, as in the red-blue SADIE method, but uses the Hellinger distance between the density map of counts and the density map of sampling effort. This approach had better theoretical properties than the SADIE method to detect spatial heterogeneity when clusters were located on square or elongated domains and more or less close to the edges. Also, it is able to detect cluster patterns for small samples size and clusters with small radius. These advantages of MAPCOMP were demonstrated by Lavigne et al. (2010) in an analysis of the spatial distribution of codling moth, *Cydia pomonella*, diapausing larvae in eight orchards in France, suggesting that this method could be useful in cases of conservation biology of rare species or for agricultural pests where population densities are expected to be low and habitats may be geometrically intricate. However, further studies are needed to compare these new indices with conventional ones for different types of data.

SADIE CONTRIBUTIONS TO THE SPATIAL ANALYSIS OF AGROECOSYSTEMS

The methods developed for SADIE are increasingly used to describe and understand many ecological processes. However, as shown by Perry et al. (2002), no single analytical method can identify all the spatial characteristics of data. Consequently, the use and comparison of more than one method for spatial analysis of ecological data is recommended.

Winder et al. (2001) provide the only report to date that used SADIE to analyze spatial patterns of both surface and subterranean organisms. Indeed, literature concerning belowground patterns is scarce compared to that concerning aboveground spatial structure. For example, aboveground studies using SADIE

have analyzed the spatial patterns of vegetation (Maestre, 2003), seed banks (González-Andújar et al., 1999), epidemics (Moreno et al., 2007), and various taxa of invertebrates (Winder et al., 1999; Rossi, 2003; Archard et al., 2004), to address theoretical and applied questions regarding agroecosystems. The studies to date primarily provide an understanding of interspecific interactions or responses to environmental heterogeneity (Perry and Dixon, 2002; Diaz et al., 2010; Nachappa et al., 2011). Several studies have employed SADIE methodology to enhance the manipulation of multiple natural enemies in ephemeral habitats, mainly in annual crops, which are characterized by temporal and spatial discontinuity for herbivorous pests and their natural enemies that exploit such habitats. Diaz et al. (2010) reported that the spatial-temporal congruence between two natural enemies, the fungus *Pandora neoaphidis* infecting aphids and the aphidophagous hoverflies larvae only occurred near to the time of lettuce harvest, reducing the risk of intraguild predation between these two natural enemies at field scale. As a consequence, both natural enemies could be good candidates to control aphids on lettuce in a conservation biological control strategy as part of an integrated pest management approach. In addition, this study highlighted the encouraging early temporal congruence of both natural enemies and their aphid prey through manipulation of external habitats that can provide refuge for both *P. neoaphidis* and hoverflies (Diaz et al., 2010). SADIE was used by Costamagna and Landis (2011) to demonstrate that the naturally occurring community of generalist predators exerts strong top-down suppression of *Aphis glycines* populations at multiple scales, and no evidence was found that the presence of prey refugia at these scales can lead to population outbreaks. Another recent example is the explicit use of SADIE spatial maps to investigate benefits from adjacent woody vegetation on predators and parasitoids within vineyards, which concluded with recommendations for management of the non-crop areas adjacent to farms (Thomson and Hoffmann, 2013).

We find fewer examples of the use of SADIE to study belowground organisms, although several authors have employed it to characterize the spatial distribution of entomopathogenic nematodes. Wilson et al. (2003) used the spatial information provided by SADIE to study the persistence of the EPN species *Heterorhabditis bacteriophora*, when this nematode was used as an inundative biocontrol agent to control several species of scarab beetle larvae in crops. In this study, nematodes were applied following a uniform distribution, in one central circular patch and in individual patches. They observed that nematodes applied in patches moved from their initial application sites and became more evenly distributed, whereas the distribution of nematodes in plots with uniform application became patchier as nematodes died. Neither application method affected the persistence or efficacy of the nematodes. An elegant study by Spiridonov et al. (2007) identified aggregated and highly associated spatial patterns of *Steinernema feltiae* and *Steinernema affine* using Lloyd's index of patchiness and SADIE, when both species were measured at a fine (5 × 5 cm) scale. They characterized infective juvenile (IJ) physiological age (I–IV groups) based on the retention of the sheath and a visible lipid content. Lloyds index identify group I as being the most aggregated and SADIE identify group II as

the most aggregated. Both methods revealed random distribution for older individuals (groups III and IV). A discrepancy between the two aggregation indices when considering larval age groups was likely due to sampling scale, reinforcing the importance of using multiple methods to study spatial patterns. More recently, SADIE analysis was used to characterize the spatial patterns of entomopathogenic nematodes and other members of their soil food web such as NF and some free-living nematodes that compete with EPN for the cadaver (**Figure 2**) (Campos-Herrera et al., 2012). Positive correlations between the three guilds supported Lindford's hypothesis that introducing organic matter to soil (EPN-killed insects) promotes population growth by free living nematodes and natural enemies of nematodes (Lindford et al., 1938).

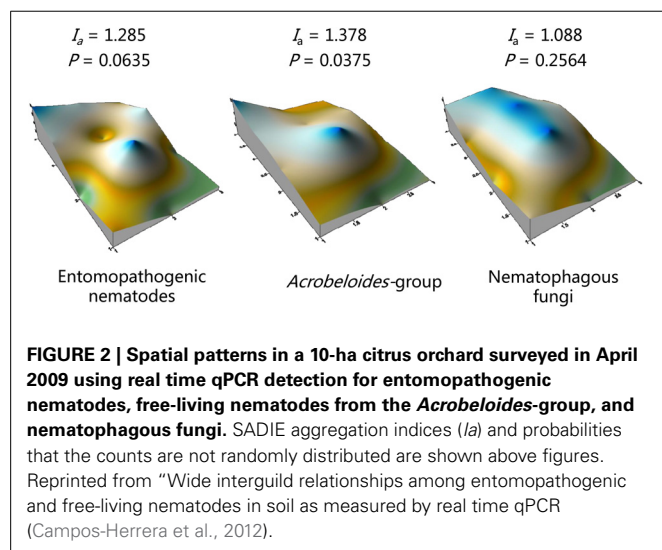
Earthworms are recognized as soil engineers (Jones et al., 2006c) because the population or community distributions of these macroinvertebrates affect many ecosystem processes related to organic matter mineralization, aggregate formation (Richard et al., 2012) and resource availability in the soil (Jiménez et al., 2011). Spatial structure in earthworm communities has been related mainly to environmental heterogeneity posed by soil and vegetation patchiness and to environmental gradients at larger scale (Decaëns et al., 2009; Jiménez et al., 2011). Hernández et al. (2007) used geostatistical and multivariate analysis to explain the relationships between the horizontal distribution of earthworm communities and some soil factors in grassland in Madrid Spain. Jiménez et al. (2011) combined SADIE analysis to detect gap and patch clusters and association/dissociation between earthworm species with geostatistics, to investigate the spatial distribution of an earthworm community together with the heterogeneity of selected soil properties in a gallery forest of the Colombian "Llanos." The degree of autocorrelation of spatial pattern was assessed with semi-variogram and they used the partial Mantel test to explain the relationship between the spatial pattern of earthworm density and soil environmental variables, concluding that the earthworm community of this gallery forest showed a random structure in a spatially clumped soil environment. Another study using SADIE provided insights into earthworm

assembly rules on pastures of Northwestern France, by showing evidences for the driving role of local factors on earthworm spatial distribution and community assembly (Richard et al., 2012).

Multiyear and multilocation experiments used SADIE to study the corn rootworm *Diabrotica* spp. (Coleoptera: Chrysomelidae) to determine its spatial pattern and investigate spatial associations with environment factors in cornfields which provided an understanding of such spatial interrelationships has a potential to reduce sampling and management costs to reduce rootworm populations (Park and Tollefson, 2006). A recent study revealed the belowground distribution of *Diabrotica virgifera virgifera* using SADIE, demonstrating that larvae had an aggregated distribution throughout their development and that pattern changes of feeding larvae were linked with the feeding preference of the three larval instars for different types of roots (Schumann and Vidal, 2012).

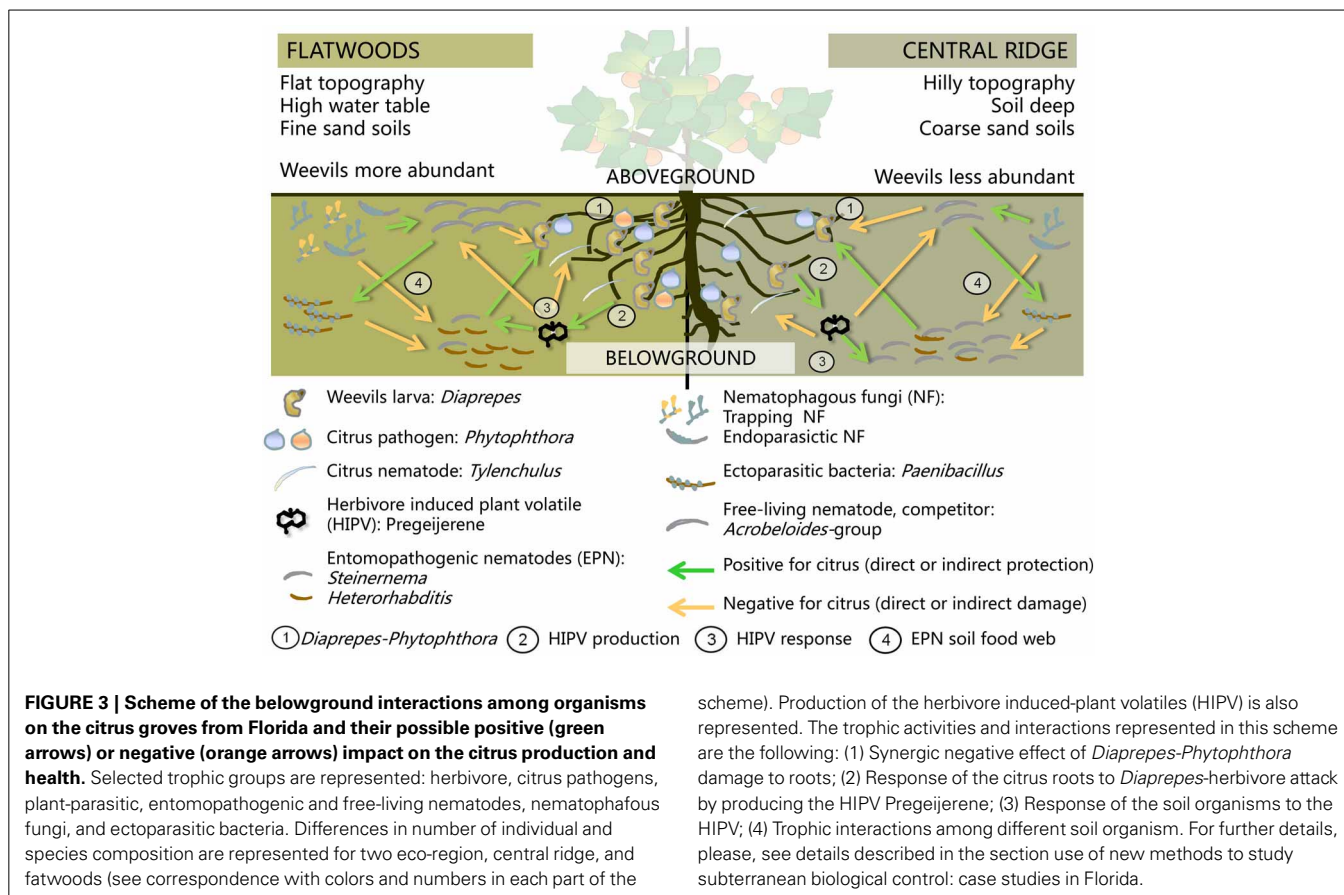
The soil environment also constitutes an important reservoir for the diversity of entomopathogenic fungi, belonging to the orders Hypocreales and Entomophthorales, which are able to cause natural epizootics, regulating populations of above and belowground insects. The comparison of SADIE and Geographical Information System (GIS) was used to evaluate the validity of GIS to describe the spatial patterns of several soil borne entomopathogenic fungi, such as *Beauveria bassiana* and *Paecilomyces fumosoroseus* within agriculture and hedgerow ecosystems, respectively (Meyling and Eilenberg, 2006). They demonstrated the suitability of GIS for identifying distribution patterns of soil borne entomopathogenic fungi and the importance of large sample sizes to describe local biodiversity of the fungi in the soil environment.

The use and application of these statistical tools are increasing in scope, with tremendous potential to link above with belowground systems. In this regard, an interesting recent study developed by Eisenhauer et al. (2011) linked the impact of plant diversity and the presence above-belowground invertebrates to the stability of plant community productivity in space and time in experimental grassland communities. They concluded that changes in plant diversity at one trophic level are not reflected by changes in multitrophic interrelationships and that both above- and belowground invertebrates decouple the positive relationship between spatial and temporal stability of plant community productivity.



USE OF NEW METHODS TO STUDY SUBTERRANEAN BIOLOGICAL CONTROL: CASE STUDIES IN FLORIDA

The measurement and analytical methods described in this paper were used in recent studies (Duncan and Stelinski, 2013) designed to help manage a subterranean weevil herbivore in Florida citrus orchards (**Figure 3**). *Diaprepes abbreviatus* is a serious citrus pest whose larvae cause feeding damage to the tree roots and facilitate root infection by plant pathogenic *Phytophthora* spp. oomycetes (**Figure 3**, interaction 1) (Graham et al., 2003; Dolinski et al., 2012). Duncan et al. (2003) suggested the possible involvement of natural enemies in the *D. abbreviatus* spatial pattern across different eco-regions of the Florida peninsula. Because weevils are more abundant in flatwoods regions than on the central ridge, they hypothesized that EPN species diversity, richness or



abundance might be higher in the central ridge. Duncan et al. (2007) used conventional methods to characterize multitrophic links involved in the plant-herbivore-EPN food web in a Florida citrus orchard. In order to improve the ability to study relationships among weevils, EPNs, natural enemies of EPNs and abiotic soil conditions across the regions, molecular primers, and probes were developed to quantify selected soil organisms at multiple trophic levels from a single sample of DNA (Figure 3, interaction 4). Real time qPCR was then used to identify and quantify (1) 13 species of EPN (Campos-Herrera et al., 2011a,b, 2012, 2013), (2) 7 NF (Atkins et al., 2005; Zhang et al., 2006; Pathak et al., 2012), (3) 2 ectoparasitic bacteria in the genus *Paenibacillus* that limit the mobility of the EPN infective juveniles (IJs) (Campos-Herrera et al., 2011b), (4) *Acroboloides*-group of free-living bacteriophagous nematodes (FLBNs), reported to be competitors of EPNs in the cadaver (Campos-Herrera et al., 2012), and (v) a citrus pathogen associated with *D. abbreviatus* root damage (Huang et al., 2010).

Nematodes were extracted from soil in 53 orchards in a geospatial survey across the two regions. DNA from the nematode samples was then probed to quantify EPNs and organisms associated with nematodes in each sample. Spatial patterns from the data and relationships between patterns were then characterized using redundancy (RDA) and SADIE analyses (Campos-Herrera et al., 2013). A preliminary study to validate the methods from samples in a single orchard revealed significant aggregation of EPNs and of FLBNs, but not NF as measured by SADIE,

and significant associations between all three guilds when measured by correlation analysis (Figure 2; Campos-Herrera et al., 2012). Across the two regions of the geospatial survey, three of the four most frequently encountered EPN species were significantly aggregated as measured by the SADIE *Ia* (Campos-Herrera et al., 2013). The spatial patterns of *Steinernema diaprepesi* and *Heterorhabditis zealandica* were significantly associated with that of the central ridge eco-region, supporting the possibility that EPNs influence the *D. abbreviatus* spatial pattern, because *S. diaprepesi* is reported to be more virulent to this weevil than other native EPN species (El-Borai et al., 2012). Variables that might affect EPN patterns were identified from redundancy analyses. Several variables that affect soil water potential (depth to groundwater, water holding capacity, content of clay and organic matter) were significant predictors of the soil community composition. These variables explained more than 40% of the variability of *S. diaprepesi* in the survey and suggested that the nematode is encountered most frequently and in greatest abundance in soil with lower rather than higher water potential. A long-term field experiment provided some support for the possibility that native EPNs regulate weevils more effectively in drier soil conditions. Duncan et al. (2013) substituted sand for native soil in tree planting holes. Fewer weevils emerged from sand and trees grew larger than in the native loamy sand soil. Sentinel weevil larvae buried in the plots were killed at higher rates than in native soil; however, the abundance of EPNs measured by qPCR was not different in either soil. Thus, greater regulation of weevils by native EPNs

in soil with low water potential may result because such soils favor parasitism by some species, rather than by increasing EPN abundance.

A semiochemical basis for multitrophic relationships (citrus plant-herbivore-biological control agent-natural enemies) has also been studied (Ali et al., 2010, 2011, 2012, 2013). Molecular probes were used together with *in situ* recovery of HIPV to demonstrate that naturally occurring EPNs as well as free living, bacterivorous nematodes that sometimes compete with EPNs were attracted by the volatile compound pregeijerene (1,5-dimethylcyclodeca-1,5,7-triene) that emanates from citrus roots fed upon by weevils (Ali et al., 2012, 2013) (Figure 3, interactions 2 and 3). The net effect of the FLBNs on EPN efficacy in this system is unknown and ultimately the effects of these interactions on plant production/yield need to be measured to understand whether manipulating indirect plant defenses is practical for sustainable pest control.

FUTURE PROSPECTS

Rapidly evolving techniques to measure and analyze spatial and temporal dynamics of community structure, interguild relationships, and plant communication present ever greater opportunities for interdisciplinary studies to fundamentally advance our understanding of how plants interact with other organisms and their environment. For example, stable isotope analysis continues to be widely used to assess prey-predator links and ecosystem functioning such as feedback in prey availability and predator hunting style (Wimp et al., 2013). However, molecular techniques such as those described here are also helping to reveal and characterize trophic interactions between many organisms, such as predation rates in the field by various species of NF (Pathak et al., 2012) or the plant preferences of generalist vs. specialist herbivores and their effects on the natural resources (Hereward

and Walter, 2012). A method to analyze invertebrate regurgitate, made possible by refined PCR techniques, has eliminated the need to destructively sample rare specimens for predator-prey assessment (Waldner and Traugott, 2012). The development of multiplex systems will further encourage the use of PCR methods to provide new insights into predator-prey relationships that serve to develop new approaches for biological control of pests. Linking indirect plant defense and predator-prey behavior, in time and space, by using PCR-methods to identify and quantify organisms and their relationships with plant signaling has obvious potential for pest management. We have described reports of this phenomenon for EPNs in citrus orchards and maize fields. PCR was also recently used to identify important arthropod and acarid predators of *D. virgifera virgifera* (Lundgren et al., 2009; Lundgren and Fergen, 2011). Linking knowledge of how maize semiochemicals function to affect a diverse predator/pathogen guild would be an important step toward a holistic understanding of food web dynamics and how they can be manipulated to enhance sustainable pest management. Clearly, the daunting challenges to characterizing the functions of soil communities posed by their taxonomic and behavioral complexity are being overcome by rapid methodological advances on many fronts.

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REFERENCES

- Agustí, N., Shayler, S. P., Harwood, J. D., Vaughan, I. P., Sunderland, K. D., and Symondson, W. O. C. (2003). Collembola as alternative prey sustaining spiders in arable ecosystems: prey detection within predators using molecular markers. *Mol. Ecol.* 12, 3467–3475. doi: 10.1046/j.1365-294X.2003.02014.x
- Ali, J. G., Alborn, H. T., and Stelinski, L. L. (2010). Subterranean herbivore-induced volatiles released by *Citrus* roots upon feeding by *Diaprepes abbreviatus* recruit entomopathogenic nematodes. *J. Chem. Ecol.* 36, 361–368. doi: 10.1007/s10886-010-9773-7
- Ali, J. G., Alborn, H. T., and Stelinski, L. L. (2011). Constitutive and induced subterranean plant volatiles attract both entomopathogenic and plant parasitic nematodes. *J. Ecol.* 99, 26–35. doi: 10.1111/j.1365-2745.2010.01758.x
- Ali, G. J., Alborn, H. T., Campos-Herrera, R., Kaplan, F., Duncan, L. W., Rodríguez-Sanoa, C., et al. (2012). Extending explorations of belowground herbivore-induced plant volatiles: attracting natural enemies of root pests in multiple contexts. *PLoS ONE* 7:e38146. doi: 10.1371/journal.pone.0038146
- Ali, A. G., Campos-Herrera, R., Alborn, H. T., Duncan, L. W., and Stelinski, L. L. (2013). Sending mixed messages: a trophic cascade produced by a belowground herbivore induced signal. *J. Chem. Ecol.* 39, 1140–1147. doi: 10.1007/s10886-10013-0332-x
- Archard, G. A., Bohan, D. A., Hughes, L., and Wiltshire, C. W. (2004). Spatial sampling to detect slug abundance in an arable field. *Ann. Appl. Biol.* 145, 165–173. doi: 10.1111/j.1744-7348.2004.tb00372.x
- Atkins, S. D., Clark, I. M., Pande, S., Hirsch, P. R., and Kerry, B. R. (2005). The use of real-time PCR and species-specific primers for the identification and monitoring of *Paecilomyces lilacinus*. *FEMS Microbiol. Ecol.* 51, 257–264. doi: 10.1016/j.femsec.2004.09.002
- Badri, D. V., and Vivanco, J. M. (2009). Regulation and function of root exudates. *Plant Cell Environ.* 32, 666–681. doi: 10.1111/j.1365-3040.2009.01926.x
- Berry, S. D., Fargette, M., Spaul, V. W., Morand, S., and Cadet, P. (2008). Detection and quantification of root-knot nematode (*Meloidogyne javanica*), lesion nematode (*Pratylenchus zaei*) and dagger nematode (*Xiphinema elongatum*) parasites of sugarcane using real-time PCR. *Mol. Cell. Probes* 22, 168–176. doi: 10.1016/j.mcp.2008.01.003
- Bissett, A., Richardson, A. E., Baker, G., Kirkegaard, J., and Thrall, P. H. (2013). Bacterial community response to tillage and nutrient additions in a long-term wheat cropping experiment. *Soil Biol. Biochem.* 58, 281–292. doi: 10.1016/j.soilbio.2012.12.002
- Blaxter, M., Mann, J., Chapman, T., Thomas, F., Whitton, C., Floyd, R., et al. (2005). Defining operational taxonomic units using DNA barcode data. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 360, 1935–1943. doi: 10.1098/rstb.2005.1725
- Boamfa, E. I., Steeghs, M. M. L., Cristescu, S. M., and Harren, F. J. M. (2004). Trace gas detection from fermentation processes in apples; an intercomparison study between proton-transfer-reaction mass spectrometry and laser photoacoustics. *Int. J. Mass Spectrom.* 239, 193–201. doi: 10.1016/j.ijms.2004.07.017
- Boff, M. I. C., Zoon, F. C., and Smits, P. H. (2001). Orientation of *Heterorhabditis megidis* to insect hosts and plant roots in a Y-tube sand olfactometer. *Entomol. Exp. Appl.* 98, 329–337. doi: 10.1046/j.1570-7458.2001.00789.x
- Bustin, S. A., Benes, V., Garson, J. A., Hellems, J., Huggett, J., Kubista, M., et al. (2009). The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments.

- Clin. Chem.* 55, 611–622. doi: 10.1373/clinchem.2008.112797
- Campos-Herrera, R., Stuart, R. J., El-Borai, F. E., Gutiérrez, C., and Duncan, L. W. (2010). “Entomopathogenic nematode ecology and biological control in Florida citrus,” in *Integrated Management of Arthropod Pests and Insect Borne Diseases*, Vol. 5, eds A. Ciancio and K. G. Mukerji (New York, NY: Springer Science + Business Media B.V.), 101–130.
- Campos-Herrera, R., Johnson, E. G., El-Borai, F. E., Stuart, R. J., Graham, J. H., and Duncan, L. W. (2011a). Long-term stability of entomopathogenic nematode spatial patterns measured by sentinel insects and real-time PCR assays. *Ann. Appl. Biol.* 158, 55–68. doi: 10.1111/j.1744-7348.2010.00433.x
- Campos-Herrera, R., El-Borai, F. E., Stuart, R. J., Graham, J. H., and Duncan, L. W. (2011b). Entomopathogenic nematodes, phoretic *Paenibacillus* spp., and the use of real time quantitative PCR to explore soil food webs in Florida citrus groves. *J. Invertebr. Pathol.* 108, 30–39. doi: 10.1016/j.jip.2011.06.005
- Campos-Herrera, R., El-Borai, F. E., and Duncan, L. W. (2012). Wide interguild relationships among entomopathogenic and free-living nematodes in soil as measured by real time qPCR. *J. Invertebr. Pathol.* 111, 126–135. doi: 10.1016/j.jip.2012.07.006
- Campos-Herrera, R., Pathak, E., El-Borai, F. E., Stuart, R. J., Gutiérrez, C., Rodríguez-Martín, J. A., et al. (2013). Geospatial patterns of soil properties and the biological control potential of entomopathogenic nematodes in Florida citrus groves. *Soil Biol. Biochem.* 66, 163–174. doi: 10.1016/j.soilbio.2013.07.011
- Caporaso, J. G., Kuczynski, J., Stombaugh, J., Bittinger, K., Bushman, F. D., Costello, E. K., et al. (2010). QIIME allows analysis of highthroughput community sequencing data. *Nat. Methods* 7, 335–336. doi: 10.1038/nmeth.f.303
- Carreón-Martínez, L., and Heath, D. D. (2010). Revolution in food web analysis and trophic ecology: diet analysis by DNA and stable isotope analysis. *Mol. Ecol.* 19, 25–27. doi: 10.1111/j.1365-294X.2009.04412.x
- Chang, C.-M., Chang, W.-H., Wang, C.-H., Wang, J.-H., Maid, J. D., and Lee, G.-B. (2013). Nucleic acid amplification using microfluidic systems. *Lab Chip* 13, 1225–1242. doi: 10.1039/c3lc41097h
- Chen, X. Y., Daniell, T. J., Neilson, R., O’Flaherty, V., and Griffiths, B. S. (2010). A comparison of molecular methods for monitoring soil nematodes and their use as biological indicators. *Eur. J. Soil Biol.* 46, 319–324. doi: 10.1016/j.ejsobi.2010.05.002
- Clark, I. (2001). *Practical Geostatistics*. Available online at: www.uni-tuebingen.de/egpinfo/Pedometrie/PracticalGeostatistics1979.pdf
- Costamagna, A. C., and Landis, D. A. (2011). Lack of strong refuges allows top-down control of soybean aphid by generalist natural enemies. *Biol. Control* 57, 184–192. doi: 10.1016/j.biocontrol.2011.03.006
- Danner, H., Samudrala, D., Cristescu, S. M., and Van Dam, N. M. (2012). Tracing hidden herbivores: time-resolved non-invasive analysis of belowground volatiles by Proton-Transfer-Reaction Mass Spectrometry (PTR-MS). *J. Chem. Ecol.* 38, 785–794. doi: 10.1007/s10886-012-0129-3
- de Gouw, J., and Warneke, C. (2007). Measurements of volatile organic compounds in the earth’s atmosphere using proton-transfer-reaction mass spectrometry. *Mass Spectrom. Rev.* 26, 223–257. doi: 10.1002/mas.20119
- de Gouw, J., Warneke, C., Karl, T., Eerdeken, G., van der Veen, C., and Fall, R. (2003). Sensitivity and specificity of atmospheric trace gas detection by proton-transfer-reaction mass spectrometry. *Int. J. Mass Spectrom.* 223, 365–382. doi: 10.1016/S1387-3806(02)00926-0
- Decaens, T., Jiménez, J. J., and Rossi, J. P. (2009). A null model analysis of the spatio-temporal distribution of earthworm species assemblages in Colombia grasslands. *J. Trop. Ecol.* 25, 415–427. doi: 10.1017/S0266467409006075
- Degenhardt, J., Hiltbold, I., Kollner, T. G., Frey, M., Gierl, A., Gershenson, J., et al. (2009). Restoring a maize root signal that attracts insect-killing nematodes to control a major pest. *Proc. Natl. Acad. Sci. U.S.A.* 106, 13213–13218. doi: 10.1073/pnas.0906365106
- Díaz, B. M., Legarrea, S., Marcos-García, M. A., and Fereres, A. (2010). The spatio-temporal relationships among aphids, the entomopathogenic fungus *Pandora neoaphidis* and aphidophagous hoverflies in outdoor lettuce. *Biol. Control* 53, 304–311. doi: 10.1016/j.biocontrol.2009.12.002
- Díaz, B. M., Barrios, L., and Fereres, A. (2012). Interplant movement and spatial distribution of alate and apterous morphs of *Nasonovia ribisnigri* (Homoptera: Aphididae) on lettuce. *Bull. Entomol. Res.* 102, 406–414. doi: 10.1017/S0007485311000745
- Dicke, M., and Sabelis, M. W. (1988). How plants obtain predatory mites as bodyguards. *Neth. J. Zool.* 38, 148–165. doi: 10.1163/156854288X00111
- Dickie, I. A., and FitzJohn, R. G. (2007). Using terminal restriction fragment length polymorphism (T-RFLP) to identify mycorrhizal fungi: a methods review. *Mycorrhiza* 17, 259–270. doi: 10.1007/s00572-007-0129-2
- Diggle, P. J. (1983). *Statistical Analysis of Spatial Point Patterns*. London, UK: Academic press.
- Dillman, A. R., Guillermin, M. L., Lee, J. H., Kim, B., Sternberg, P. W., and Hallem, E. A. (2012). Olfaction shapes host-parasite interactions in parasitic nematodes. *Proc. Natl. Acad. Sci. U.S.A.* 109, 13896–13897. doi: 10.1073/pnas.1211436109
- Dodd, C. S., Bruford, M. W., Symondson, W. O. C., and Glen, D. M. (2003). “Detection of slug DNA within carabid predators using prey-specific PCR primers,” in *Slug and Snail Pests: Agricultural, Veterinary and Environmental Perspectives*, ed G. B. J. Dussard (Alton, UK: British Crop Protection Council), 13–20.
- Dolinski, C., Choo, H. Y., and Duncan, L. W. (2012). Grower acceptance of entomopathogenic nematodes: case studies on three continents. *J. Nematol.* 44, 226–235.
- Donn, S., Griffiths, B. S., Neilson, R., and Daniell, T. J. (2007). DNA extraction from soil nematodes for multi-sample community studies. *Appl. Soil Ecol.* 38, 20–26. doi: 10.1016/j.apsoil.2007.08.006
- Dudareva, N., Pichersky, E., and Gershenson, J. (2004). Biochemistry of plant volatiles. *Plant Physiol.* 135, 1893–1902. doi: 10.1104/pp.104.049981
- Duncan, L. W., and Stelinski, L. L. (2013). A call to arms: protecting and summoning native nematodes to kill *Diaprepes* root weevil. *Citrus Ind. Mag.* (in press).
- Duncan, L. W., Graham, J. H., Dunn, D. C., Zellers, J., McCoy, C. W., and Nguyen, K. (2003). Incidence of endemic entomopathogenic nematodes following application of *Steinernema riobrave* for control of *Diaprepes abbreviatus*. *J. Nematol.* 35, 178–186.
- Duncan, L. W., Graham, J. H., Zellers, J., Bright, D., Dunn, D. C., El-Borai, F. E., et al. (2007). Food web responses to augmenting the entomopathogenic nematodes in bare and animal manure-mulched soil. *J. Nematol.* 39, 176–189.
- Duncan, L. W., Stuart, R. J., El-Borai, F. E., Campos-Herrera, R., Pathak, E., and Graham, J. H. (2013). Modifying orchard planting sites conserves entomopathogenic nematodes, reduces weevil herbivory and increases citrus tree growth, survival and fruit yield. *Biol. Control* 64, 26–36. doi: 10.1016/j.biocontrol.2012.09.006
- Dutilleul, P. (1993). Modifying the *t*-test for assessing the correlation between two spatial processes. *Biometrics* 49, 305–314. doi: 10.2307/2532625
- Eisenhauer, N., Milcu, A., Allan, E., Nitschke, N., Scherber, C., Temperton, V., et al. (2011). Impact of above- and below-ground invertebrates on temporal and spatial stability of grassland of different diversity. *J. Ecol.* 99, 572–582. doi: 10.1111/j.1365-2745.2010.01783.x
- Eitzinger, B., and Traugott, M. (2011). Which prey sustains cold-adapted invertebrate generalist predators in arable land? Examining prey choices by molecular gut content analysis. *J. Appl. Ecol.* 48, 591–599. doi: 10.1111/j.1365-2664.2010.01947.x
- El-Borai, F. E., Stuart, R. J., Campos-Herrera, R., Pathak, E., and Duncan, L. W. (2012). Entomopathogenic nematodes, root weevil larvae, and dynamic interactions among soil texture, plant growth, herbivory, and predation. *J. Invertebr. Pathol.* 109, 134–142. doi: 10.1016/j.jip.2011.10.012
- Fall, R., Karl, T., Hansel, A., Jordan, A., and Lindinger, W. (1999). Volatile organic compounds emitted after leaf wounding: on-line analysis by proton-transfer-reaction mass spectrometry. *J. Geophys. Res.* 104, 15963–15974. doi: 10.1029/1999JD900144
- Gagnon, A.-É., Doyon, J., Heimpel, G. E., and Brodeur, J. (2011). Prey DNA detection success following digestion by intraguild predators: influence of prey and predator species. *Mol. Ecol. Resour.* 11, 1022–1032. doi: 10.1111/j.1755-0998.2011.03047.x
- Garipey, T. D., Kuhlmann, U., Gillott, C., and Erlandson, M. (2007). Parasitoids, predators and PCR: the use of diagnostic molecular markers in biological control of arthropods. *J. Appl. Entomol.* 131, 225–240. doi: 10.1111/j.1439-0418.2007.01145.x
- Gfeller, A., Laloux, M., Barsics, F., Kati, D. E., Haubruge, E., du Jardin, P., et al. (2013). Characterization of

- volatile organic compounds emitted by barley (*Hordeum vulgare* L.) roots and their attractiveness to wireworms. *J. Chem. Ecol.* 39, 1129–1139. doi: 10.1007/s10886-10013-0302-3
- Gibb, K., Beard, J., O'Reagain, P., Christian, K., Torok, V., and Keller, K. O. (2008). Assessing the relationship between patch type and soil mites: a molecular approach. *Pedobiologia* 51, 445–461. doi: 10.1016/j.pedobi.2008.01.004
- Glenn, T. C. (2011). Field guide to next-generation DNA sequencers. *Mol. Ecol. Res.* 11, 759–769. doi: 10.1111/j.1755-0998.2011.03024.x
- González-Andújar, J. L., Perry, J. N., and Moss, S. R. (1999). Modeling effects of spatial pattern on the seed bank dynamics of *Alopecurus Myosuroides*. *Weed Sci.* 47, 697–705.
- Goovaerts, P. (1997). *Geostatistics for Natural Resources Evaluation*. New York, NY: Oxford University Press, 483.
- Goovaerts, P. (1999). Geostatistics in soil science: state-of-the-art and perspectives. *Geoderma* 89, 1–45. doi: 10.1016/S0016-7061(98)00078-0
- Graham, J. H., Bright, D. B., and McCoy, C. W. (2003). Phytophthora-Diaprepes weevil complex: *Phytophthora* spp. relationship with citrus rootstocks. *Plant Dis.* 87, 85–90. doi: 10.1094/PDIS.2003.87.1.85
- Griffiths, B. S., Donn, S., Neilson, R., and Daniell, T. J. (2006). Molecular sequencing and morphological analysis of a nematode community. *Appl. Soil Ecol.* 32, 325–337. doi: 10.1016/j.apsoil.2005.07.006
- Griffiths, B. S., Daniell, T. J., Donn, S., and Neilson, R. (2012). Bioindication potential of using molecular characterisation of the nematode community: response to tillage. *Eur. J. Soil Biol.* 49, 92–97. doi: 10.1016/j.ejsobi.2011.09.002
- Hackl, E., Schloter, M., Szukics, U., Bodrossy, L., and Sessitsch, A. (2012). “From single genes to microbial networks,” in *Soil Ecology and Ecosystem Services*, ed D. H. Wall (Oxford, UK: Oxford University Press), 65–81.
- Hagn, A., Pritsch, K., Schloter, M., and Munch, J. C. (2003). Fungal diversity in agricultural soil under different farming management systems, with special reference to biocontrol strains of *Trichoderma* spp. *Biol. Fertil. Soils* 38, 236–244. doi: 10.1007/s00374-003-0651-0
- Hallem, E. A., Dillman, A. R., Hong, A. V., Zhang, Y. J., Yano, J. M., DeMarco, S. F., et al. (2011). A sensory code for host seeking in parasitic nematodes. *Curr. Biol.* 21, 377–383. doi: 10.1016/j.cub.2011.01.048
- Hamilton, H. C., Strickland, M. S., Wickings, K., Bradford, M. A., and Fierer, N. (2009). Surveying soil faunal communities using a direct molecular approach. *Soil Biol. Biochem.* 41, 1311–1314. doi: 10.1016/j.soilbio.2009.03.021
- Hansel, A., Jordan, A., Holzinger, R., Prazeller, P., Vogel, W., and Linderger, W. (1995). Proton transfer reaction mass spectrometry: on-line trace gas analysis at the ppb level. *Int. J. Mass Spectrom.* 149/150, 609–619. doi: 10.1016/0168-1176(95)04294-U
- Harper, G. L., King, R. A., Dodd, C. S., Harwood, J. D., Glen, D. M., Bruford, M. W., et al. (2005). Rapid screening of invertebrate predators for multiple prey DNA targets. *Mol. Ecol.* 14, 819–827. doi: 10.1111/j.1365-294X.2005.02442.x
- Hatteland, B. A., Symondson, W. O. C., King, R. A., Skage, M., Schander, C., and Solhøy, T. (2011). Molecular analysis of predation by carabid beetles (Carabidae) on the invasive Iberian slug *Arion lusitanicus*. *Bull. Entomol. Res.* 23, 1–12.
- He, Z., Deng, Y., van Nostrand, J. D., Tu, Q., Xu, M., Hemme, C. L., et al. (2010). GeoChip 3.0 as a high-throughput tool for analyzing microbial community composition, structure and functional activity. *ISME J.* 4, 1167–1179. doi: 10.1038/ismej.2010.46
- Heath, R. R., and Manukian, A. (1992). Development and evaluation of systems to collect volatile semiochemicals from insects and plants using a charcoal-infused medium for air purification. *J. Chem. Ecol.* 18, 1209–1226. doi: 10.1007/BF00980075
- Heidemann, K., Scheu, S., Ruess, L., and Maraun, M. (2011). Molecular detection of nematode predation and scavenging in oribatid mites: laboratory and field experiments. *Soil Biol. Biochem.* 43, 2229–2236. doi: 10.1016/j.soilbio.2011.07.015
- Hernández, P., Fernández, R., Novo, M., Trigo, D., and Cosín, J. D. (2007). Geostatistical and multivariate analysis of the horizontal distribution of a fan earthworm community in El Molar (Madrid, Spain). *Pedobiologia* 51, 13–21. doi: 10.1016/j.pedobi.2006.11.002
- Hereward, J. P., and Walter, G. H. (2012). Molecular interrogation of the feeding behaviour of field captured individual insects for interpretation of multiple host plant use. *PLoS ONE* 7:e44435. doi: 10.1371/journal.pone.0044435
- Hilton, S., Bennett, A. J., Keane, G., Bending, G. D., Chandler, D., Stobart, R., et al. (2013). Impact of shortened crop rotation of oilseed rape on soil and rhizosphere microbial diversity in relation to yield decline. *PLoS ONE* 8:e59859. doi: 10.1371/journal.pone.0059859
- Hiltbold, I., and Turlings, T. C. J. (2008). Belowground chemical signaling in maize: when simplicity rhymes with efficiency. *J. Chem. Ecol.* 34, 628–635. doi: 10.1007/s10886-008-9467-6
- Hiltbold, I., Toepfer, S., Kuhlmann, U., and Turlings, T. C. J. (2010a). How maize root volatiles affect the efficacy of entomopathogenic nematodes in controlling the western corn rootworm? *Chemoeology* 20, 155–162. doi: 10.1007/s00049-009-0034-6
- Hiltbold, I., Baroni, M., Toepfer, S., Kuhlmann, U., and Turlings, T. C. J. (2010b). Selection of entomopathogenic nematodes for enhanced responsiveness to a volatile root signal helps to control a major root pest. *J. Exp. Biol.* 213, 2417–2423. doi: 10.1242/jeb.041301
- Hoogendoorn, M., and Heimpel, G. E. (2001). PCR-based gut content analysis of insect predators: using ribosomal ITS-1 fragments from prey to estimate predation frequency. *Mol. Ecol.* 10, 2059–2067. doi: 10.1046/j.1365-294X.2001.01316.x
- Huang, J., Li, C., Xiao, C., and Wang, G. (2010). Detection of Phytophthora nicotianae in soil with real-time quantitative PCR. *J. Phytopathol.* 158, 15–21. doi: 10.1111/j.1439-0434.2009.01554.x
- Hunter, M. D. (2001). Out of sight, out of mind: the impacts of root-feeding insects in natural and managed systems. *Agric. For. Entomol.* 3, 3–9. doi: 10.1046/j.1461-9563.2001.00083.x
- Ings, T. C., Montoya, J. M., Bascompte, J., Blüthgen, N., Brown, L., Dormann, C. F., et al. (2009). Ecological networks – beyond food webs. *J. Anim. Ecol.* 78, 253–269. doi: 10.1111/j.1365-2656.2008.01460.x
- Iwao, S. (1968). A new regression method for analyzing the aggregation pattern of animal populations. *Res. Popul. Ecol.* 10, 1–20. doi: 10.1007/BF02514729
- James, D., and Grasswitz, T. (2005). Synthetic herbivore-induced plant volatiles increase field captures of parasitic wasps. *Biocontrol* 50, 871–880. doi: 10.1007/s10526-005-3313-3
- Jiménez, J. J., Decaëns, T., Amézquita, E., Rao, I., Thomas, R. J., and Lavelle, P. (2011). Short-range spatial variability of soil physico-chemical variables related to earthworm clustering in a neotropical gallery forest. *Soil Biol. Biochem.* 43, 1071–1080. doi: 10.1016/j.soilbio.2011.01.028
- Johnson, S. N., and Gregory, P. J. (2006). Chemically-mediated host-plant location and selection by root-feeding insects. *Physiol. Entomol.* 31, 1–13. doi: 10.1111/j.1365-3032.2005.00487.x
- Johnson, S. N., and Nielsen, U. N. (2012). Foraging in the dark – chemically mediated host plant location by belowground insect herbivores. *J. Chem. Ecol.* 38, 604–614. doi: 10.1007/s10886-012-0106-x
- Jones, K. L., Todd, T. C., and Herman, M. A. (2006a). Development of taxo-specific markers for high-throughput screening of microbial-feeding nematodes. *Mol. Ecol. Notes* 6, 712–714. doi: 10.1111/j.1471-8286.2006.01319.x
- Jones, K. L., Todd, T. C., Wall-Beam, J. L., Koolen, J. D., Blair, J. M., and Herman, M. A. (2006b). Molecular approach for assessing responses of microbial-feeding nematodes to burning and chronic nitrogen enrichment in a native grassland. *Mol. Ecol.* 15, 2601–2609. doi: 10.1111/j.1365-294X.2006.02971.x
- Jones, C. G., Gutiérrez, J. I., Groffman, P. M., and Shachack, M. (2006c). Linking ecosystem engineers to soil processes: a framework using the Jenny State Factor Equation. *Eur. J. Soil Biol.* 42, 539–553. doi: 10.1016/j.ejsobi.2006.07.017
- Juen, A., and Traugott, M. (2006). Amplification facilitators and multiplex PCR: tools to overcome PCR-inhibition in DNA-gut content analysis of soil-living invertebrates. *Soil Biol. Biochem.* 38, 1872–1879. doi: 10.1016/j.soilbio.2005.11.034
- Juen, A., and Traugott, M. (2007). Revealing species-specific trophic links in soil food webs: molecular identification of scarab predators. *Mol. Ecol.* 16, 1545–1557. doi: 10.1111/j.1365-294X.2007.03238.x
- Kaplan, I. (2012a). Attracting carnivorous arthropods with plant volatiles: the future of biocontrol or playing with fire? *Biol. Control* 60, 77–89. doi: 10.1016/j.biocontrol.2011.10.017
- Kaplan, I. (2012b). Trophic complexity and the adaptive value of damage-induced plant volatiles.

- PLoS Biol.* 10:e1001437. doi: 10.1371/journal.pbio.1001437
- Kappers, I. F., Aharoni, A., van Herpen, T., Luckerhoff, L. L. P., Dicke, M., and Bouwmeester, H. J. (2005). Genetic engineering of terpenoid metabolism attracts bodyguards to Arabidopsis. *Science* 309, 2070–2072. doi: 10.1126/science.1116232
- Kessler, A., and Morrell, K. (2010). “Plant volatile signalling: multitrophic interactions in the headspace,” in *The Chemistry and Biology of Volatiles*, ed A. Herrmann (Chichester: Wiley), 95–122. doi: 10.1002/9780470669532.ch4
- King, R. A., Vaughan, I. P., Bell, J. R., Bohan, D. A., and Symondson, W. O. C. (2010). Prey choice by carabid beetles feeding on an earthworm community analysed using species- and lineage-specific PCR primers. *Mol. Ecol.* 19, 1721–1732. doi: 10.1111/j.1365-294X.2010.04602.x
- Kollner, T. G., Schnee, C., Gershenzon, J., and Degenhardt, J. (2004). The variability of sesquiterpenes cultivars is controlled by allelic emitted from two *Zea mays* variation of two terpene synthase genes encoding stereoselective multiple product enzymes. *Plant Cell* 16, 1115–1131. doi: 10.1105/tpc.019877
- Kollner, T. G., Held, M., Lenk, C., Hiltbold, I., Turlings, T. C. J., Gershenzon, J., et al. (2008). A maize (E)-beta-caryophyllene synthase implicated in indirect defense responses against herbivores is not expressed in most American maize varieties. *Plant Cell* 20, 482–494. doi: 10.1105/tpc.107.051672
- Korie, S., Clark, S. J., Perry, J. N., Muggleston, M. A., Barlett, P. W., Marshall, E. J. P., et al. (1998). Analyzing maps of dispersal around a single focus. *Environ. Ecol. Stat.* 5, 317–344. doi: 10.1023/A:1009603804998
- Kurtz, B., Hiltbold, I., Turlings, T. C. J., Kuhlmann, U., and Toepfer, S. (2009). Comparative susceptibility of larval instars and pupae of the western corn rootworm to infection by three entomopathogenic nematodes. *Biocontrol* 54, 255–262. doi: 10.1007/s10526-008-9156-y
- Lavigne, C., Ricci, B., Franck, P., and Senoussi, R. (2010). Spatial analysis of ecological count data: a density map comparison approach. *Basic Appl. Ecol.* 11, 734–742. doi: 10.1016/j.baec.2010.08.011
- Linford, M. B., Yap, F., and Oliveira, J. M. (1938). Reduction of soil populations of the root knot nematode during decomposition of organic matter. *Soil Sci.* 45, 127–141. doi: 10.1097/00010694-193802000-00004
- Liu, L., Li, Y., Li, S., Hu, N., He, Y., Pong, R., et al. (2012). Comparison of next-generation sequencing systems. *J. Biomed. Biotech.* 2012:251364 doi: 10.1155/2012/251364
- Lundgren, J. G., Ellsbury, M. E., and Prischmann, D. A. (2009). Analysis of the predator community of a subterranean herbivorous insect based on polymerase chain reaction. *Ecol. Appl.* 19, 2157–2166. doi: 10.1890/08-1882.1
- Lundgren, J. G., and Fergen, J. K. (2011). Enhancing predation of a subterranean insect pest: a conservation benefit of winter vegetation in agroecosystems. *Appl. Soil Ecol.* 51, 9–16. doi: 10.1016/j.apsoil.2011.08.005
- Ma, J., Li, D. D., Keller, M., Schmidt, O., and Feng, X. (2005). A DNA marker to identify predation of *Plutella xylostella* (Lep., Plutellidae) by *Nabis kinbergii* (Hem., Nabidae) and *Lycosa* sp. (Araneae, Lycosidae). *J. Appl. Entomol.* 129, 330–335. doi: 10.1111/j.1439-0418.2005.00977.x
- Maestre, F. T. (2003). Small-scale spatial patterns of two soil lichens in semi-arid Mediterranean steppe. *Lichenologist* 35, 71–81. doi: 10.1006/lich.2002.0425
- Mengoni, A., Tatti, E., Decorosi, F., Viti, C., Bazzicalupo, M., and Giovannetti, L. (2005). Comparison of 16S rRNA and 16S rDNA T-RFLP approaches to study bacterial communities in soil microcosms treated with chromate as perturbing agent. *Microb. Ecol.* 50, 375–384. doi: 10.1007/s00248-004-0222-4
- Meyling, N. V., and Eilenberg, J. (2006). Occurrence and distribution of soil borne entomopathogenic fungi within a single organic agroecosystem. *Agr. Ecosyst. Environ.* 113, 336–341. doi: 10.1016/j.agee.2005.10.011
- Mohney, B., Matz, T., LaMoreaux, J., Wilcox, D., Gimsing, A., Mayer, P., et al. (2009). *In situ* silicone tube microextraction: a new method for undisturbed sampling of root-exuded thiophenes from marigold (*Tagetes erecta* L.) in soil. *J. Chem. Ecol.* 1279–1287. doi: 10.1007/s10886-009-9711-8
- Moreno, A., Nebreda, M., Diaz, B. M., García, M., Salas, F., and Fereres, A. (2007). Temporal and spatial spread of lettuce mosaic virus (LMV) in lettuce crops in central Spain: factors involved in LMV epidemics. *Ann. Appl. Biol.* 150, 351–360. doi: 10.1111/j.1744-7348.2007.00135.x
- Moreno-Ripoll, R., Gabarra, R., Symondson, W. O. C., King, R. A., and Agustí, N. (2012). Trophic relationships between predators, whiteflies and their parasitoids in tomato greenhouses: a molecular approach. *Bull. Entomol. Res.* 7, 1–9.
- Morise, H., Miyazaki, E., Yoshimitsu, S., and Eki, T. (2012). Profiling nematode communities in unmanaged flowerbed and agricultural field soils in Japan by DNA barcode sequencing. *PLoS ONE* 7:e51785. doi: 10.1371/journal.pone.0051785
- Mumm, R., and Dicke, M. (2010). Variation in natural plant products and the attraction of bodyguards involved in indirect plant defense. *Can. J. Zool.* 88, 628–667. doi: 10.1139/Z10-032
- Nachappa, P., Margolies, D. C., Nechols, J. R., and Campbell, J. F. (2011). Variation in predator foraging behaviour changes predator-prey spatio-temporal dynamics. *Funct. Ecol.* 25, 1309–1317. doi: 10.1111/j.1365-2435.2011.01892.x
- Nunan, N., Daniell, T. J., Singh, B. K., Papert, A., McNicol, J. W., and Prosser, J. I. (2005). Links between plant and rhizoplane bacterial communities in grassland soils, characterized using molecular techniques. *Appl. Environ. Microbiol.* 71, 6784–6792. doi: 10.1128/AEM.71.11.6784-6792.2005
- Park, Y.-L., and Tollefson, J. J. (2006). Spatio-temporal dynamics of corn rootworm, *Diabrotica* spp., adults and their spatial association with environment. *Entomol. Exp. Appl.* 120, 105–112. doi: 10.1111/j.1570-7458.2006.00428.x
- Pathak, E., El-Borai, F. E., Campos-Herrera, R., Jonhson, E. G., Stuart, R. J., Graham, J. H., et al. (2012). Use of real-time PCR to discriminate predatory and saprophagous behavior by nematophagous fungi. *Fungal Biol.* 116, 563–573. doi: 10.1016/j.funbio.2012.02.005
- Pawliszyn, J. (1997). *Solid Phase Microextraction – Theory and Practice*. 1st Edn. New York, NY: Wiley-VCH.
- Perry, J. N., and Hewitt, M. (1991). A new index of aggregation for animal counts. *Biometrics* 47, 1505–1518. doi: 10.2307/2532402
- Perry, J. N. (1995). Spatial analysis by distance indices. *J. Anim. Ecol.* 64, 303–314. doi: 10.2307/5892
- Perry, J. N. (1998). Measures of spatial pattern for counts. *Ecology* 79, 1008–1017. doi: 10.1890/0012-9658(1998)079[1008:MOSPFC]2.0.CO;2
- Perry, J. N., and Dixon, P. M. (2002). A new method to measure spatial association for ecological count data. *Ecoscience* 9, 133–141.
- Perry, J. N., Winder, L., Holand, J. M., and Alston, R. D. (1999). Red-blue plots for clusters in count data. *Ecol. Lett.* 2, 106–113. doi: 10.1046/j.1461-0248.1999.22057.x
- Perry, J. N., Liebhold, A. M., Rosenberg, M. S., Dungan, J., Miriri, M., Jakomulka, A., et al. (2002). Illustrations and guidelines for selecting statistical methods for quantifying spatial patterns in ecological data. *Ecography* 25, 578–600. doi: 10.1034/j.1600-0587.2002.250507.x
- Pineda, A., Soler, R., Weddergeris, B. T., Shimwela, M. M., van Loon, J. J. A., and Dicke, M. (2013). Non-pathogenic rhizobacteria interfere with the attraction of parasitoids to aphid-induced plant volatiles via jasmonic acid signalling. *Plant Cell Environ.* 36, 393–404. doi: 10.1111/j.1365-3040.2012.02581.x
- Poelman, E. H., Bruinsma, M., Zhu, F., Weddergeris, B. T., Boursault, A. E., de Jongema, Y., et al. (2012). Hyperparasitoids use herbivore-induced plant volatiles to locate their parasitoid host. *PLoS Biol.* 10:e1001435. doi: 10.1371/journal.pbio.1001435
- Pompanon, F., Deagle, B. E., Symondson, W. O. C., Brown, D. S., Jarman, S. N., and Taberlet, P. (2012). Who is eating what: diet assessment using next generation sequencing. *Mol. Ecol.* 21, 1931–1950. doi: 10.1111/j.1365-294X.2011.05403.x
- Pumariño, L., Alomar, O., and Agustí, N. (2011). Development of specific ITS markers for plant DNA identification within herbivorous insects. *Bull. Entomol. Res.* 101, 271–276. doi: 10.1017/S0007485310000465
- Rasmann, S., and Turlings, T. C. J. (2008). First insights into specificity of belowground tritrophic interactions. *Oikos* 117, 362–369. doi: 10.1111/j.2007.0030-1299.16204.x
- Rasmann, S., Kollner, T. G., Degenhardt, J., Hiltbold, I., Toepfer, S., Kuhlmann, U., et al. (2005). Recruitment of entomopathogenic nematodes by insect-damaged maize roots. *Nature* 434, 732–737. doi: 10.1038/nature03451
- Rasmann, S., Bauerle, T. L., Poveda, K., and Vannette, R. (2011). Predicting root defence against herbivores during succession.

- Funct. Ecol.* 25, 368–379. doi: 10.1111/j.1365-2435.2010.01811.x
- Rasmann, S., Hiltbold, I., and Ali, J. G. (2012a). “The role of root-produced volatile secondary metabolites in mediating soil interactions,” in *Advances in Selected Plant Physiology Aspects*, eds G. Montanaro and B. Cichio (Croatia: InTech Open Access Publisher), 269–290.
- Rasmann, S., Ali, J. G., Helder, J., and van der Putten, W. H. (2012b). Ecology and evolution of soil nematode chemotaxis. *J. Chem. Ecol.* 38, 615–628. doi: 10.1007/s10886-012-0118-6
- Read, D. S., Sheppard, S. K., Bruford, M. W., Glen, D. M., and Symondson, W. O. C. (2006). Molecular detection of predation by soil micro-arthropods on nematodes. *Mol. Ecol.* 15, 1963–1972. doi: 10.1111/j.1365-294X.2006.02901.x
- Richard, B., Legras, M., Margarie, P., Mathieu, J., Barot, S., Caro, G., et al. (2012). Spatial organization of earthworm assemblages in pastures of northwestern France. *Eur. J. Soil Biol.* 53, 62–69. doi: 10.1016/j.ejsobi.2012.08.005
- Robert, C. A. M., Erb, M., Hiltbold, I., Hibbard, B., Gaillard, M., Bilat, J., et al. (2013). Genetically engineered maize plants reveal distinct costs and benefits of constitutive volatile emissions in the field. *Plant Biotechnol. J.* 11, 628–639. doi: 10.1111/pbi.12053
- Romeu-Dalmau, C., Piñol, J., and Agustí, N. (2012). Detecting aphid predation by earwigs in organic citrus orchards using molecular markers. *Bull. Entomol. Res.* 14, 1–7.
- Rossi, J. P. (2003). Clusters in earthworm spatial distribution. *Pedobiologia* 47, 490–496. doi: 10.1078/0031-4056-00218
- Sato, E., and Toyota, K. (2006). Application of PCR-DGGE into community structure analysis of soil nematodes. *Jpn. J. Soil Sci. Plant Nutr.* 77, 157–163 (in Japanese with English abstract).
- Schumann, M., and Vidal, S. (2012). Dispersal and spatial distribution of western corn rootworm larvae in relation to root phenology. *Agr. For. Entomol.* 14, 331–339. doi: 10.1111/j.1461-9563.2012.00573.x
- Sharmah, D., Jha, D. K., and Pandey, R. R. (2010). Molecular approaches in arbuscular mycorrhizal research: a review. *J. Phytol.* 2, 75–90.
- Smyrnioudis, I. N., Harrington, R., Clark, S. J., and Katis, N. (2001). The effect of natural enemies on the spread of barley yellow dwarf virus (BYDV) by *Rhopalosiphum padi* (Hemiptera: Aphididae). *Bull. Entomol. Res.* 91, 301–306. doi: 10.1079/BER2001110
- Spiridonov, S. E., Moens, M., and Wilson, M. J. (2007). Fine scale spatial distributions of two entomopathogenic nematodes in a grassland soil. *Appl. Soil Ecol.* 37, 192–201. doi: 10.1016/j.apsoil.2007.06.009
- Taylor, L. R. (1961). Aggregation, variance and the mean. *Nature* 189, 732–735. doi: 10.1038/189732a0
- Thaler, J. S. (1999). Jasmonate-inducible plant defences cause increased parasitism of herbivores. *Nature* 399, 686–688. doi: 10.1038/21420
- Thomson, L. J., and Hoffmann, A. A. (2013). Spatial scale of benefits from adjacent woody vegetation on natural enemies within vineyards. *Biol. Control* 64, 57–65. doi: 10.1016/j.biocontrol.2012.09.019
- Tollsten, L., and Bergström, G. (1988). Headspace volatiles of whole plants and macerated plant parts of *Brassica sinapis*. *Phytochemistry* 27, 4013–4018. doi: 10.1016/0031-9422(88)83085-1
- Torr, P., Heritage, S., and Wilson, M. J. (2004). Vibrations as a novel signal for host location by parasitic nematodes. *Int. J. Parasitol.* 34, 997–999. doi: 10.1016/j.ijpara.2004.05.003
- Traugott, M., Zangerl, P., Juen, A., Schallhart, N., and Pfiffner, L. (2006). Detecting key parasitoids of lepidopteran pests by multiplex PCR. *Biol. Control* 39, 39–46. doi: 10.1016/j.biocontrol.2006.03.001
- Turlings, T. C. J., Tumlinson, J. H., and Lewis, W. J. (1990). Exploitation of herbivore-induced plant odors by host-seeking parasitic wasps. *Science* 250, 1251–1253. doi: 10.1126/science.250.4985.1251
- Turlings, T. C. J., and Wäckers, F. (2004). “Recruitment of predators and parasitoids by herbivore-injured plants,” in *Advances in Insect Chemical Ecology*, eds R. T. Cardé and J. G. Millar (Cambridge, UK: Cambridge University Press), 21–75.
- Valentini, A., Pompanon, F., and Taberlet, P. (2009). DNA barcoding for ecologists. *Trends Ecol. Evol.* 24, 110–117. doi: 10.1016/j.tree.2008.09.011
- van der Heijden, M. G. A., and Wagg, C. (2013). Soil microbial diversity and agro-ecosystem functioning. *Plant Soil* 363, 1–5. doi: 10.1007/s11104-012-1545-4
- van Pelt-Verkuil, E., van Belkum, A., and Hays, J. P. (2010). *Principles and Technical Aspects of PCR Amplification*. Springer Science + Business Media B.V., 330.
- van Tol, R. W. H. M., van der Sommen, A. T. C., Boff, M. I. C., van Bezooijen, J., Sabelis, M. W., and Smits, P. H. (2001). Plants protect their roots by alerting the enemies of grubs. *Ecol. Lett.* 4, 292–294. doi: 10.1046/j.1461-0248.2001.00227.x
- Vervoort, M. T. W., Vonk, J. A., Mooijman, P. J. W., van den Elsen, S. J. J., van Megen, H. H. B., Veenhuizen, P., et al. (2012). SSU ribosomal DNA-based monitoring of nematode assemblages reveals distinct seasonal fluctuations within evolutionary heterogeneous feeding guilds. *PLoS ONE* 7:e47555. doi: 10.1371/journal.pone.0047555
- Waldner, T., and Traugott, M. (2012). DNA-based analysis of regurgitates: a noninvasive approach to examine the diet of invertebrate consumers. *Mol. Ecol. Res.* 12, 669–675. doi: 10.1111/j.1755-0998.2012.03135.x
- Waldner, T., Sint, D., Juen, A., and Traugott, M. (2013). The effect of predator identity on post-feeding prey DNA detection success in soil-dwelling macro-invertebrates. *Soil Biol. Biochem.* 63, 116–123. doi: 10.1016/j.soilbio.2013.03.030
- Wallinger, C., Staudacher, K., Schallhart, N., Peter, E., Dresch, P., Juen, A., et al. (2013). The effect of plant identity and the level of plant decay on molecular gut content analysis in a herbivorous soil insect. *Mol. Ecol. Res.* 13, 75–83. doi: 10.1111/1755-0998.12032
- Wang, G.-H., Jin, J., Liu, J.-J., Chen, X.-L., Liu, J.-D., and Liu, X.-B. (2009). Bacterial community structure in a mollisol under long-term natural restoration, cropping, and bare fallow history estimated by PCR-DGGE. *Pedosphere* 19, 156–165. doi: 10.1016/S1002-0160(09)60105-3
- Wimp, G. M., Murphy, S. M., Lewis, D., Douglas, M. R., Ambikapathi, R., van-Tull, L. A., et al. (2013). Predator hunting mode influences patterns of prey use from grazing and epigeic food webs. *Oecologia* 171, 505–515. doi: 10.1007/s00442-012-2435-4
- Winder, L., Perry, J. N., and Holland, J. M. (1999). The spatial and temporal distribution of the grain aphid *Sitobion avenae* in winter wheat. *Entomol. Exp. Appl.* 93, 277–290. doi: 10.1046/j.1570-7458.1999.00588.x
- Winder, L., Alexander, C. J., Holland, J. M., Woolley, C., and Perry, J. N. (2001). Modelling the dynamic spatio-temporal response of predators to transient prey patches in the field. *Ecol. Lett.* 4, 568–576. doi: 10.1046/j.1461-0248.2001.00269.x
- Wilson, M. J., Lewis, E. E., Yoder, F., and Gaugler, R. (2003). Application pattern and persistence of the entomopathogenic nematode *Heterorhabditis bacteriophora*. *Biol. Control* 26, 180–188. doi: 10.1016/S1049-9644(02)00125-1
- Xu, X. M., and Madden, L. V. (2003). Considerations for the use of SADIE statistics to quantify spatial patterns. *Ecography* 26, 821–830. doi: 10.1111/j.0906-7590.2003.03667.x
- Zhang, L. M., Liu, X. Z., Zhu, S. F., and Chen, S. Y. (2006). Detection of the nematophagous fungus *Hirsutiella rhossiliensis* in soil by real-time PCR and parasitism bioassay. *Biol. Control* 36, 316–323. doi: 10.1016/j.biocontrol.2005.08.002
- Zhu, Z., Jenkins, G., Zhang, W., Zhang, M., Guan, Z., and James Yang, C. (2012). Single-molecule emulsion PCR in microfluidic droplets. *Anal. Bioanal. Chem.* 403, 2127–2143. doi: 10.1007/s00216-012-5914-x
- Zijlstra, C., and van Hoof, R. A. (2006). A multiplex real-time polymerase chain reaction (TaqMan) assay for the simultaneous detection of *Meloidogyne chiwoodi* and *M. fallax*. *Phytopathology* 96, 1255–1262. doi: 10.1094/PHYTO-96-1255

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