



Plasticity of plasma membrane compartmentalization during plant immune responses

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Plasma membranes require high levels of plasticity to modulate the perception and transduction of extracellular and intracellular signals. Dynamic lateral assembly of protein complexes combined with an independent compositional lipid patterning in both membrane leaflets provide cells the opportunity to decorate this interface with specific proteins in an organized but dynamic manner. Such ability to dynamically reorganize the protein content of the plasma membrane is essential for the regulation of processes such as polarity of transport, development, and microbial infection. While the plant cell wall represents the first physical and mostly unspecific barrier for invading microbes, the plasma membrane is at the forefront of microbial recognition and initiation of defense responses. Accumulating evidence indicating dynamic compartmentalization of plasma membranes in response to environmental cues has increased the interest in the compositional heterogeneity of this bilayer. Here, we elucidate the recruitment of specific proteins into defined membrane structures that ensure functional compartmentalization of the bilayer during infection processes.

Keywords: heterogeneity, compartmentalization, membrane domains, membrane rafts, plant immunity, receptor-ligand interactions

INTRODUCTION

The interface between the cytoplasm and the outer environment in plant cells is comprised of the cell wall and the plasma membrane. In their fluid mosaic model, Singer and Nicolson (1972) proposed that biological membranes, such as the plasma membrane, consist of a phospholipid bilayered matrix that is randomly interspersed with integral proteins. Recent research on plasma membrane components has significantly refined this view. Accumulating evidence indicates that the plasma membrane consists of a phospholipid bilayer that contains dynamic membrane domains, some of which are enriched in sphingolipids, sterols and specific proteins and called membrane rafts (Brown and Rose, 1992; Simons and Ikonen, 1997; Xu et al., 2001). These membrane rafts are able to cluster into more stable (signaling) platforms upon the perception of certain stimuli and crosslinking (Kusumi et al., 2004; Hammond et al., 2005; Lingwood et al., 2008; Hogue et al., 2011). Most of the available knowledge and hypotheses on the function and formation of membrane rafts originates from studies in mammalian and yeast cells (Simson et al., 1995; Sheets et al., 1997; Simons and Ikonen, 1997; Malinska et al., 2003; Kusumi et al., 2004; Fan et al., 2010), where it has among others been shown that membrane rafts play an important role during microbial infection processes (Duncan et al., 2002; Lafont and van der Goot, 2005; van der Meer-Janssen et al., 2010). There is increasing evidence that membrane domains may be similarly involved in the interaction between microbes and plants (Zappel and Panstruga, 2008). Here we discuss the current view on plant plasma membrane remodeling in response to both symbiotic and pathogenic microbes.

DYNAMIC MEMBRANE DOMAINS IN EUKARYOTIC CELLS

Since the *Keystone Symposium on Lipid Rafts and Cell Function*, the consensus definition of membrane rafts is that they are small (10–200 nm), heterogeneous, highly dynamic, sterol- and sphingolipid-enriched domains that compartmentalize cellular processes (Pike, 2006; Simons and Gerl, 2010). These rather elusive membrane rafts, due to their nanoscale size and dynamic behavior, can be stabilized through protein–protein, protein–lipid, and lipid–lipid interactions to form larger platforms that allow visualization by conventional light microscopy. Two main compartmentalizing forces seem to form membrane domains. The lipid raft model describes how sphingolipids can laterally associate with sterol molecules into close-packed assemblies (liquid-ordered phases; Simons and Ikonen, 1997; Lingwood and Simons, 2010). These close-packed, highly saturated rafts have different properties compared with the surrounding, less ordered and highly unsaturated phospholipid bilayer. Due to these different properties certain transmembrane and membrane-associated proteins preferentially insert into these rafts. In mammalian cells post-translational modifications such as myristoylation and palmitoylation were shown to target proteins preferentially to membrane rafts in the cytoplasmic membrane leaflet, while the addition of a glycosylphosphatidylinositol (GPI) moiety anchors proteins predominantly to membrane rafts at the outer leaflet (Schroeder et al., 1994; Varma and Mayor, 1998; Morrow et al., 2002; Zacharias et al., 2002). Evidence for lipid-dependency of membrane raft formation has been obtained from both artificial membrane models and living cells (Pralle et al., 2000; Hammond et al., 2005; Baumgart et al., 2007; Beck et al., 2007; Roche et al., 2008;

Kaiser et al., 2009). It was also demonstrated that the cortical actin-based cytoskeleton plays a role in the compartmentalization of plasma membranes (Suzuki and Sheetz, 2001; Fujiwara et al., 2002; Kusumi et al., 2004; Lenne et al., 2006). Such impact, however, cannot be tested in artificial membrane models. In the membrane-skeleton fence and anchored-protein picket models the effect of the cytoskeleton on the lateral movement of membrane components is described (Kusumi and Sako, 1996; Sako et al., 1998; Fujiwara et al., 2002; Kusumi et al., 2004). The membrane-skeleton fence model proposes that the cytoplasmic domains of transmembrane and membrane-associated proteins can collide with the cytoskeletal filaments close to the membrane, thereby confining movement of these proteins to compartments that can vary in size from 30 to 250 nm. In the anchored-protein picket model, transmembrane proteins anchor to and line up along the cytoskeletal meshwork as pickets. Due to steric hindrance and hydrodynamic friction-like effects these proteins consequently limit diffusion of other membrane molecules in both membrane leaflets. Since both models affect membrane proteins as well as membrane lipids it can be assumed that dynamic compartmentalization of plasma membranes relies on an interplay between cytoskeleton-mediated and raft-derived effects. Indeed “hop-diffusion” where movement of proteins is temporarily confined by a cytoskeleton fence until the restrictive presence of the actin filaments is loosened by temporary breakdown of the filaments or an increase in the distance between the membrane and the cytoskeletal meshwork, has been described for a number of membrane raft proteins such as the human transferrin receptor (Sako and Kusumi, 1995; Kusumi et al., 2005). Membrane molecules can also move to neighboring compartments when they have gained sufficient kinetic energy to break through the restraining actin filament. The protein FORMIN-1 in *Arabidopsis thaliana* (thale cress) seems to be such a transmembrane picket protein, since it is able to anchor the actin cytoskeleton through the plasma membrane to the cell wall (Martiniere et al., 2011). However, experiments using the fluorescence recovery after photobleaching (FRAP) technique on fluorescently labeled membrane proteins in protoplasts that were re-growing their cell wall, demonstrate that in plants the constraining influence of the cell wall on the mobility of plasma membrane proteins is much greater than the influence of the cytoskeleton (Martiniere et al., 2012).

A frequently used, but much debated method in membrane raft research makes use of the differential packing of molecules in membranes to isolate close-packed assemblies from the plasma membrane through their greater resistance to detergents such as Triton (Heerklotz, 2002; Munro, 2003; Brown, 2006; Kierszniewska et al., 2009; Mongrand et al., 2010; Pathak and London, 2011; Tanner et al., 2011). Isolation of so called detergent-insoluble membranes (DIMs), or detergent-resistant membranes (DRMs), from *Nicotiana tabacum* (tobacco) demonstrated the enrichment of the sphingolipid glycosylceramide and the sterols stigmasterol, 24-methyl cholesterol, sitosterol, and cholesterol compared to detergent-soluble membrane (DSM) fractions. In contrast, relative amounts of phospholipids such as phosphatidylcholine and phosphatidylethanolamine are reduced in DIMs (Mongrand et al., 2004). Similar findings have been reported for DIMs prepared from *A. thaliana*, *Medicago truncatula* (barrel clover),

Phaseolus vulgaris (common bean), and *Zea mays* (maize; Borner et al., 2005; Lefebvre et al., 2007; Furt et al., 2010; Carmona-Salazar et al., 2011). Proteome analyses of membrane fractions from the above mentioned plant species and additionally from *Solanum tuberosum* (potato), *Sinapis alba* (white mustard), and the monocotyledonous plant species *Avena sativa* (oat), *Secale cereale* (rye), and *Oryza sativa* (rice) demonstrate enrichment of proteins involved in different types of processes in DIMs, including signal transduction and abiotic and biotic stress responses (Shahollari et al., 2004; Morel et al., 2006; Fujiwara et al., 2009; Stanislas et al., 2009; Keinath et al., 2010; Krügel et al., 2012; Takahashi et al., 2012). There seems to be a preference for palmitoylation and myristoylation in proteins involved in signaling processes and stress responses enriched in DIMs, possibly targeting them to membrane rafts in the cytoplasmic membrane leaflet (Morel et al., 2006). Since DIM fractions may only partially, if at all, reflect the composition of individual membrane raft classes, localization of these putative raft-resident molecules in *in planta* membrane domains has to be validated by other techniques to allay concerns about artifactual membrane raft formation and co-purification of non-raft constituents. Additional imaging approaches of putative membrane raft constituents and testing whether clustering of these putative raft-localized lipids, sterols or proteins is influenced by chemicals such as the sterol extracting methyl- β -cyclodextrin, support the proposed presence of such structures *in planta* (Krügel et al., 2008; Raffaele et al., 2009; Boutte et al., 2011). Furthermore, newly developed imaging techniques such as total internal reflection fluorescence microscopy (TIRFM) or variable-angle epifluorescence microscopy (VAEM) can be successfully applied to cell wall enclosed cells and allow the visualization of mobile proteins located in and around the plasma membrane with very high signal-to-background ratio (Owen et al., 2007; Konopka and Bednarek, 2008; Spira et al., 2012). A recent study showed TIRFM to also be applicable to study single molecule trafficking of PM-resident proteins in plant cells (Martiniere et al., 2012). These techniques will help to analyze domain structures in the plasma membrane in more detail in the future. Whether the plasma membrane domains formed in plant cells can always be categorized as membrane rafts remains to be fully elucidated.

THE ROLE OF MEMBRANE DOMAINS DURING INFECTION PROCESSES

The integral membrane proteins FLOTILLIN-1 and FLOTILLIN-2 (synonymously called REGGIE-2 and REGGIE-1) are frequently used as markers for membrane rafts in the cytoplasmic leaflet of mammalian cells (Lang et al., 1998; Morrow et al., 2002; Blonder et al., 2004; Nixon et al., 2011). In plants and animals, members of this protein family were described to be involved in clathrin-independent endocytosis and are present on host-derived membranes surrounding intracellular microorganisms, indicating raft-mediated endocytosis as a possible entry point for microorganisms (Panter et al., 2000; Dermine et al., 2001; Saalbach et al., 2002; Glebov et al., 2006; Murphy et al., 2007; Li et al., 2008, 2012; Korhonen et al., 2012). Furthermore, evidence that flotillins are interacting and/or co-localizing with many signaling components, such as receptors and mitogen-activated protein kinases (MAPK),

suggests that these proteins may act as scaffolds for a number of signaling processes (Langhorst et al., 2005; Haney et al., 2011; Amaddii et al., 2012). In *A. thaliana* a flotillin homolog was originally identified in DIMs and named AtFLOTILLIN-1 (AtFLOT1; Borner et al., 2005). Recently, a combination of confocal laser scanning microscopy, electron microscopy, and VAEM demonstrated that AtFLOT1 is present in distinct membrane domains, in clathrin-independent invaginations in the plasma membrane, and in endocytic vesicles with a size of approximately 100 nm (Li et al., 2012). Using transgenic plants expressing an artificial microRNA against AtFLOT1, the same study demonstrated that AtFLOT1 is required for meristem and seedling development. In *M. truncatula* MtFLOT2 and especially MtFLOT4 were found to be required for initiation of symbiotic infection structures (infection threads) and their elongation during interactions with the nitrogen-fixating bacterial symbiont *Sinorhizobium meliloti* (Haney and Long, 2010). Upon inoculation of *M. truncatula* roots with *S. meliloti* bacteria the evenly distributed fluorescently labeled MtFLOT4 punctae in the plasma membrane of a root hair cell start to accumulate at the root hair tip. These punctate structures might represent small clusters of membrane rafts that coalesce into a membrane raft platform at the root hair tip to facilitate the entry of the symbiont. Interestingly, fluorophore-tagged MtLYSIN-MOTIF-RECEPTOR-LIKE-KINASE-3 (MtLYK3), an RLK required for bacterial entry into the root hairs, localizes to mobile punctae in root hairs (Figure 1A) that become immobilized when the roots are inoculated with *S. meliloti* (Haney et al., 2011). Interestingly, fluorescently labeled MtFLOT4 and MtLYK3 punctae co-localize upon *S. meliloti* inoculation. In addition, the density of MtFLOT4 domains is decreased in the kinase-inactive MtLYK3 *hair curling-1* (*hcl-1*) mutant (Haney et al., 2011). These results suggest that these two proteins assemble into the same membrane domain upon perception of the symbiotic bacteria and possibly even interact with each other. Interestingly, the remorin protein *M. truncatula* SYMBIOTIC-REMORIN-1 (MtSYMREM1), a

confirmed interactor of the LYK3 receptor, localizes to distinct domains when over-expressed in transgenic *M. truncatula* roots (Figure 1B). Such membrane patterns have also been found on nodular infection threads (Figure 1C), at bacterial release sites, and in symbiosome membranes as demonstrated by immunolocalizations and immunogold-labeling electron microscopy (Lefebvre et al., 2010).

Over the years, the plant-specific remorin proteins have become the most widely accepted raft marker proteins in plants. They are highly enriched in DIMs from several plant species (Mongrand et al., 2004; Shahollari et al., 2004; Morel et al., 2006; Lefebvre et al., 2007; Keinath et al., 2010; Krügel et al., 2012; Takahashi et al., 2012) and localize to methyl- β -cyclodextrin-sensitive domains of around 75 nm in the cytoplasmic membrane leaflet in *N. tabacum* cells, as demonstrated by statistical analysis of electron microscopy data (Raffaele et al., 2009). Over-expression of the potato remorin StREM1.3 leads to the labeling of large, immobile membrane domains that might resemble small clusters of membrane rafts (Raffaele et al., 2009) similar to those labeled by MtSYMREM1 (Figure 1B). While the biological function of remorins still remains unknown, members of this highly diverse multi-gene family have been shown to regulate viral spreading in leaves (Raffaele et al., 2009), interactions between *M. truncatula* and *S. meliloti* during root nodule symbiosis (Lefebvre et al., 2010), and to serve potential roles during plant–pathogen interactions (Jarsch and Ott, 2011). Additionally, their ability to oligomerize and to interact with signaling proteins such as the symbiotic receptors and the negative regulator of immune responses RPM1-INTERACTING-PROTEIN-4 (RIN4), suggests functions as scaffolding proteins during signal transduction (Liu et al., 2009; Lefebvre et al., 2010; Toth et al., 2012). In *A. thaliana* suspension cells the group 1 remorin AtREM1.3 is phosphorylated in the intrinsically disordered N-terminal domain within 10 min of incubation with flg22, a conserved 22 amino acid peptide from the bacterial-derived elicitor flagellin (Benschop et al., 2007;

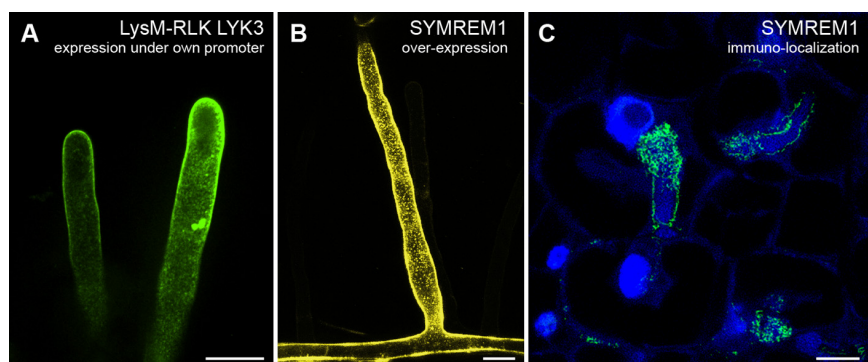


FIGURE 1 | Examples of membrane domain patterning in *Medicago truncatula*. (A) Root hair cells expressing GFP-tagged LysM-RLK MtLYK3 under the control of the native *LYK3* promoter in the kinase-inactive *LYK3* mutant line *hcl-1*. Membrane domains as described earlier (Haney et al., 2011) can be observed with spinning-disk confocal microscopy. (B) Over-expression of the YFP-tagged remorin protein MtSYMREM1 in root hair cells results in the labeling of immobile domains in the plasma membrane that can be imaged with confocal laser scanning microscopy. (C) As originally reported

(Lefebvre et al., 2010) similar domains, although smaller in size, can also be observed when immuno-localizing the MtSYMREM1 protein in the infection zone of mature root nodules using a specific MtSYMREM1 antibody. The MtSYMREM1 protein (green) resides on the infection thread membrane that surrounds invading rhizobial bacteria and accumulates at sites where these symbionts are released into the host cell. The plant and bacterial DNA is stained with DAPI (blue). Scale bars indicate 20 μ m (A,B) and 1 μ m (C).

Marín and Ott, 2012), suggesting close functional dependency of AtREM1.3 on the functional receptor-like kinase AtFLAGELLIN-SENSITIVE-2 (FLS2). In analogy to MtLYK3, flg22-dependent reduction in the lateral mobility of fluorescently tagged AtFLS2 was observed in *A. thaliana* protoplasts when analyzed by FRAP (Ali et al., 2007). These data suggest that AtFLS2 becomes part of a larger, less mobile complex upon ligand-binding and/or becomes confined to membrane rafts upon ligand-binding. The concept of ligand-dependent recruitment to membrane rafts is supported by the fact that FLS2 is highly recruited to DIMs upon flg22 treatment of *A. thaliana* suspension cells, as demonstrated by quantitative mass spectrometric analyses (Keinath et al., 2010). These findings together with FLS2 being endocytosed upon flg22 treatment in young leaves (Robatzek et al., 2006), support possible links between localization of signaling proteins in membrane domains and endocytic events.

The functional importance of membrane domains during infection processes is also underlined by results from a number of studies on interactions of plants with pathogenic oomycetes or fungi. The NADPH oxidase RESPIRATORY-BURST-OXIDASE-HOMOLOG-D (RBOHD)-mediated production of reactive oxygen species is one of the first signaling responses initiated upon the perception of the oomycete-derived elicitor cryptogein (Simon-Plas et al., 2002). NtRBOHD-mediated H₂O₂-generation was detected in small patches along the plasma membrane in *N. tabacum* cells using transmission electron microscopy after staining with CeCl₃ (Lherminier et al., 2009). A possible membrane raft localization of NtRBOHD is also supported by its identification in DIMs from *N. tabacum* after treatment with cryptogein, together with its negative regulator RAC/ROP GTPase NtRAC5 (Mongrand et al., 2004; Morel et al., 2006). *In planta* evidence for dynamic compartmentalization of membrane proteins was also reported upon host cell infection of *Hordeum vulgare* (barley) and *A. thaliana* by the powdery mildew fungus *Blumeria graminis*. Focal accumulations around fungal entry sites were observed for three otherwise evenly distributed fluorescently labeled proteins all involved in powdery mildew penetration resistance, namely MILDEW-RESISTANCE-LOCUS-O (MLO), the syntaxin HvREQUIRED-FOR-MLO-SPECIFIED-RESISTANCE-2 (HvROR2) and its ortholog AtPENETRATION-1 (AtPEN1; Bhat et al., 2005). These focal accumulations may represent membrane raft platforms, especially since they coincide with higher levels of sterols as demonstrated by filipin staining. The host-derived extrahaustorial membrane encasing the haustorial feeding structure of successfully entered fungi or oomycetes is clearly distinct from invaginated extensions of the plasma membrane, as demonstrated by the exclusion of several plasma membrane markers (Koh et al., 2005; Micali et al., 2011; Lu et al., 2012). However, differential presence of plasma membrane proteins such as AtFLS2, AtPEN1, and StREM1.3 in extrahaustorial membranes implies a mechanism to be in place that can actively determine whether proteins are included into the extrahaustorial membrane (Lu et al., 2012). Membrane reorganization may also be triggered and/or enhanced by changes in the lipid patterning of plasma membranes, as the polyphosphoinositide phosphatidylinositol-3-phosphate (PI3P) can be directly targeted by certain fungal and oomycete effectors during plant cell entry

(Kale et al., 2010). Polyphosphoinositides have been shown to be enriched in *N. tabacum* DIMs and electron microscopy of immunogold-labeled phosphatidylinositol-4,5-bisphosphate (PI4,5P₂) revealed the presence of distinct domains of approximately 25 nm in the plasma membrane (Furt et al., 2010). However, these domains are not methyl- β -cyclodextrin-sensitive, indicating that these domains are not membrane rafts according to the consensus definition.

CONCLUSIONS AND PERSPECTIVE

Although the concept of membrane rafts still remains to be unequivocally proven in plants, several lines of research clearly demonstrate that the plant plasma membrane is dynamically compartmentalized during biological processes such as infection of host cells by microorganisms. Differently sized domains that could resemble single membrane rafts, small clusters of membrane rafts, large membrane raft platforms, or other types of membrane domains beyond the membrane raft concept, can be observed in the plasma membrane with electron and/or conventional light microscopy. Current data suggest that there is not one but multiple types of membrane domains, that host multimeric signaling complexes which specifically assemble within the plasma membrane in a stimulus-dependent manner. Minor changes in lipid or protein aggregation, such as ligand-receptor complex formation, could induce the assembly of an appropriate membrane domain that can transduce the changing environmental conditions. The presence of signaling components in membrane rafts or other distinct domains may potentially modulate their activity and enhance the interactions between domain-resident components, while reducing interactions with non-domain components. A clear correlation between endocytosis and signaling is apparent in many of the discussed examples, arguing for the concept of membrane rafts serving as signaling hubs that can be exploited by (facultative) intracellular microbes to successfully establish themselves in their host. To gain further knowledge on the behavior of these dynamic plasma membrane domains *in planta*, imaging techniques with high spatial and/or temporal resolution are needed that go beyond conventional light microscopy. For example, electron microscopy and super-resolution fluorescent microscopy could be used to determine the size of plasma membrane domains in fixed samples, Förster resonance energy transfer (FRET)-based live-cell imaging to determine interactions between residents of these domains, and TIRFM or VAEM to visualize the dynamics of these domains in living cells with intact cell wall.

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REFERENCES

- Ali, G. S., Prasad, K. V. S. K., Day, I., and Reddy, A. S. N. (2007). Ligand-dependent reduction in the membrane mobility of FLAGELLIN SENSITIVE2, an *Arabidopsis* receptor-like kinase. *Plant Cell Physiol.* 48, 1601–1611.
- Amaddii, M., Meister, M., Banning, A., Tomasovic, A., Mooz, J., Rajalingam, K., and Tikkanen, R. (2012). Flotillin-1/Reggie-2 protein plays dual role in activation of receptor-tyrosine kinase/mitogen-activated protein kinase signaling. *J. Biol. Chem.* 287, 7265–7278.
- Baumgart, T., Hammond, A. T., Sengupta, P., Hess, S. T., Holowka, D. A., Baird, B. A., and Webb, W. W. (2007). Large-scale fluid/fluid phase separation of proteins and lipids in giant plasma membrane vesicles. *Proc. Natl. Acad. Sci. U.S.A.* 104, 3165–3170.
- Beck, J. G., Mathieu, D., Loudet, C., Buchoux, S., and Dufourc, E. J. (2007). Plant sterols in “rafts”: a better way to regulate membrane thermal shocks. *FASEB J.* 21, 1714–1723.
- Benschop, J. J., Mohammed, S., O’Flaherty, M., Heck, A. J. R., Slijper, M., and Menke, F. L. H. (2007). Quantitative phosphoproteomics of early elicitor signaling in *Arabidopsis*. *Mol. Cell. Proteomics* 6, 1198–1214.
- Bhat, R. A., Miklis, M., Schmelzer, E., Schulze-Lefert, P., and Panstruga, R. (2005). Recruitment and interaction dynamics of plant penetration resistance components in a plasma membrane microdomain. *Proc. Natl. Acad. Sci. U.S.A.* 102, 3135–3140.
- Blonder, J., Hale, M. L., Lucas, D. A., Schaefer, C. F., Yu, L. R., Conrads, T. R., Issaq, H. J., Stiles, B. G., and Veenstra, T. D. (2004). Proteomic analysis of detergent-resistant membrane rafts. *Electrophoresis* 25, 1307–1318.
- Borner, G. H. H., Sherrier, D. J., Weimar, T., Michaelson, L. V., Hawkins, N. D., Macaskill, A., Napier, J. A., Beale, M. H., Lilley, K. S., and Dupree, P. (2005). Analysis of detergent-resistant membranes in *Arabidopsis*. Evidence for plasma membrane lipid rafts. *Plant Physiol.* 137, 104–116.
- Boutte, Y., Men, S. Z., and Grebe, M. (2011). Fluorescent in situ visualization of sterols in *Arabidopsis* roots. *Nat. Protoc.* 6, 446–456.
- Brown, D. A. (2006). Lipid rafts, detergent-resistant membranes, and raft targeting signals. *Physiology* 21, 430–439.
- Brown, D. A., and Rose, J. K. (1992). Sorting of GPI-anchored proteins to glycolipid-enriched membrane subdomains during transport to the apical cell-surface. *Cell* 68, 533–544.
- Carmona-Salazar, L., El Hafidi, M., Enriquez-Arredondo, C., Vazquez-Vazquez, C., De La Vara, L. E. G., and Gavilanes-Ruiz, M. (2011). Isolation of detergent-resistant membranes from plant photosynthetic and non-photosynthetic tissues. *Anal. Biochem.* 417, 220–227.
- Dermine, J. F., Duclos, S., Garin, J., St-Louis, F., Rea, S., Parton, R. G., and Desjardins, M. (2001). Flotillin-1-enriched lipid raft domains accumulate on maturing phagosomes. *J. Biol. Chem.* 276, 18507–18512.
- Duncan, M. J., Shin, J. S., and Abraham, S. N. (2002). Microbial entry through caveolae: variations on a theme. *Cell. Microbiol.* 4, 783–791.
- Fan, J., Sammalkorpi, M., and Haataja, M. (2010). Formation and regulation of lipid microdomains in cell membranes: theory, modeling, and speculation. *FEBS Lett.* 584, 1678–1684.
- Fujiwara, M., Hamada, S., Hiratsuka, M., Fukao, Y., Kawasaki, T., and Shimamoto, K. (2009). Proteome analysis of detergent-resistant membranes (DRMs) associated with OsRac1-mediated innate immunity in rice. *Plant Cell Physiol.* 50, 1191–1200.
- Fujiwara, T., Ritchie, K., Murakoshi, H., Jacobson, K., and Kusumi, A. (2002). Phospholipids undergo hop diffusion in compartmentalized cell membrane. *J. Cell Biol.* 157, 1071–1081.
- Furt, F., Konig, S., Bessoule, J. J., Sargueil, F., Zallot, R., Stanislas, T., Noiro, E., Lherminier, J., Simon-Plas, F., Heilmann, I., and Mongrand, S. (2010). Polyphosphoinositides are enriched in plant membrane rafts and form microdomains in the plasma membrane. *Plant Physiol.* 152, 2173–2187.
- Glebov, O. O., Bright, N. A., and Nichols, B. J. (2006). Flotillin-1 defines a clathrin-independent endocytic pathway in mammalian cells. *Nat. Cell Biol.* 8, 46–54.
- Hammond, A. T., Heberle, F. A., Baumgart, T., Holowka, D., Baird, B., and Feigenson, G. W. (2005). Crosslinking a lipid raft component triggers liquid ordered-liquid disordered phase separation in model plasma membranes. *Proc. Natl. Acad. Sci. U.S.A.* 102, 6320–6325.
- Haney, C. H., and Long, S. R. (2010). Plant flotillins are required for infection by nitrogen-fixing bacteria. *Proc. Natl. Acad. Sci. U.S.A.* 107, 478–483.
- Haney, C. H., Riely, B. K., Tricoli, D. M., Cook, D. R., Ehrhardt, D. W., and Long, S. R. (2011). Symbiotic rhizobia bacteria trigger a change in localization and dynamics of the *Medicago truncatula* receptor kinase LYK3. *Plant Cell* 23, 2774–2787.
- Heerklotz, H. (2002). Triton promotes domain formation in lipid raft mixtures. *Biophys. J.* 83, 2693–2701.
- Hogue, I. B., Grover, J. R., Soheilian, F., Nagashima, K., and Ono, A. (2011). Gag induces the coalescence of clustered lipid rafts and tetraspanin-enriched microdomains at HIV-1 assembly sites on the plasma membrane. *J. Virol.* 85, 9749–9766.
- Jarsch, I. K., and Ott, T. (2011). Perspectives on remorin proteins, membrane rafts, and their role during plant-microbe interactions. *Mol. Plant Microbe Interact.* 24, 7–12.
- Kaiser, H. J., Lingwood, D., Levental, I., Sampaio, J. L., Kalvodova, L., Rajendran, L., and Simons, K. (2009). Order of lipid phases in model and plasma membranes. *Proc. Natl. Acad. Sci. U.S.A.* 106, 16645–16650.
- Kale, S. D., Gu, B. A., Capelluto, D. G. S., Dou, D. L., Feldman, E., Rumore, A., Arredondo, F. D., Hanlon, R., Fudal, I., Rouxel, T., Lawrence, C. B., Shan, W. X., and Tyler, B. M. (2010). External lipid PI3P mediates entry of eukaryotic pathogen effectors into plant and animal host cells. *Cell* 142, 284–295.
- Keinath, N. F., Kierszniowska, S., Lorek, J., Bourdais, G., Kessler, S. A., Shimosato-Asano, H., Grossniklaus, U., Schulze, W. X., Robatzek, S., and Panstruga, R. (2010). PAMP (pathogen-associated molecular pattern)-induced changes in plasma membrane compartmentalization reveal novel components of plant immunity. *J. Biol. Chem.* 285, 39140–39149.
- Kierszniowska, S., Seiwert, B., and Schulze, W. X. (2009). Definition of *Arabidopsis* sterol-rich membrane microdomains by differential treatment with methyl-beta-cyclodextrin and quantitative proteomics. *Mol. Cell. Proteomics* 8, 612–623.
- Koh, S., Andre, A., Edwards, H., Ehrhardt, D., and Somerville, S. (2005). *Arabidopsis thaliana* subcellular responses to compatible *Erysiphe cichoracearum* infections. *Plant J.* 44, 516–529.
- Konopka, C. A., and Bednarek, S. Y. (2008). Variable-angle epifluorescence microscopy: a new way to look at protein dynamics in the plant cell cortex. *Plant J.* 53, 186–196.
- Korhonen, J. T., Puolakkainen, M., Haivala, R., Penttila, T., Haveri, A., Markkula, E., and Lahesmaa, R. (2012). Flotillin-1 (Reggie-2) contributes to *Chlamydia pneumoniae* growth and is associated with bacterial inclusion. *Infect. Immun.* 80, 1072–1078.
- Krügel, U., He, H. X., Gier, K., Reins, J., Chincinska, I., Grimm, B., Schulze, W. X., and Kuhn, C. (2012). The potato sucrose transporter StSUT1 interacts with a DRM-associated protein disulfide isomerase. *Mol. Plant* 5, 43–62.
- Krügel, U., Veenhoff, L. M., Langbein, J., Wiederhold, E., Liesche, J., Friedrich, T., Grimm, B., Martinoia, E., Poolman, B., and Kuhn, C. (2008). Transport and sorting of the *Solanum tuberosum* sucrose transporter SUT1 is affected by posttranslational modification. *Plant Cell* 20, 2497–2513.
- Kusumi, A., Koyama-Honda, I., and Suzuki, K. (2004). Molecular dynamics and interactions for creation of stimulation-induced stabilized rafts from small unstable steady-state rafts. *Traffic* 5, 213–230.
- Kusumi, A., Nakada, C., Ritchie, K., Murase, K., Suzuki, K., Murakoshi, H., Kasai, R. S., Kondo, J., and Fujiwara, T. (2005). Paradigm shift of the plasma membrane concept from the two-dimensional continuum fluid to the partitioned fluid: high-speed single-molecule tracking of membrane molecules. *Annu. Rev. Biophys. Biomol. Struct.* 34, 351–378.
- Kusumi, A., and Sako, Y. (1996). Cell surface organization by the membrane skeleton. *Curr. Opin. Cell Biol.* 8, 566–574.
- Lafont, F., and van der Goot, F. G. (2005). Bacterial invasion via lipid rafts. *Cell. Microbiol.* 7, 613–620.
- Lang, D. M., Lommel, S., Jung, M., Ankerhold, R., Petrusch, B., Laessing, U., Wiechers, M. F., Platner, H., and Stuermer, C. A. (1998). Identification of reggie-1 and reggie-2 as plasmamembrane-associated proteins which cocluster with activated GPI-anchored cell adhesion molecules in non-caveolar micropatches in neurons. *J. Neurobiol.* 37, 502–523.
- Langhorst, M. F., Reuter, A., and Stuermer, C. A. (2005). Scaffolding microdomains and beyond: the function of reggie/flotillin proteins. *Cell. Mol. Life Sci.* 62, 2228–2240.
- Lefebvre, B., Furt, F., Hartmann, M. A., Michaelson, L. V., Carde, J. P., Sargueil-Boiron, F., Rossignol, M., Napier, J. A., Cullimore, J., Bessoule, J. J., and Mongrand, S. (2007). Characterization of lipid rafts from *Medicago truncatula* root plasma membranes: a proteomic study reveals the presence

- of a raft-associated redox system. *Plant Physiol.* 144, 402–418.
- Lefebvre, B., Timmers, T., Mbengue, M., Moreau, S., Herve, C., Toth, K., Bittencourt-Silvestre, J., Klaus, D., Deslandes, L., Godiard, L., Murray, J. D., Udvardi, M. K., Raffaele, S., Mongrand, S., Cullimore, J., Gamas, P., Niebel, A., and Ott, T. (2010). A remorin protein interacts with symbiotic receptors and regulates bacterial infection. *Proc. Natl. Acad. Sci. U.S.A.* 107, 2343–2348.
- Lenne, P. F., Wawrezynieck, L., Conchonaud, E., Wurtz, O., Boned, A., Guo, X. J., Rigneault, H., He, H. T., and Marguet, D. (2006). Dynamic molecular confinement in the plasma membrane by microdomains and the cytoskeleton meshwork. *EMBO J.* 25, 3245–3256.
- Lherminier, J., Elmayan, T., Fromentin, J., Elaroui, K. T., Vesa, S., Morel, J., Verrier, J. L., Cailleteau, B., Blein, J. P., and Simon-Plas, F. (2009). NADPH oxidase-mediated reactive oxygen species production: subcellular localization and reassessment of its role in plant defense. *Mol. Plant Microbe Interact.* 22, 868–881.
- Li, Q. R., Zhang, Q., Wang, C. Y., Li, N., and Li, J. S. (2008). Invasion of enteropathogenic *Escherichia coli* into host cells through epithelial tight junctions. *FEBS J.* 275, 6022–6032.
- Li, R., Liu, P., Wan, Y., Chen, T., Wang, Q., Mettbaach, U., Baluska, F., Samaj, J., Fang, X., Lucas, W. J., and Lin, J. (2012). A membrane microdomain-associated protein, *Arabidopsis* Flot1, is involved in a clathrin-independent endocytic pathway and is required for seedling development. *Plant Cell* 24, 2105–2122.
- Lingwood, D., Ries, J., Schwille, P., and Simons, K. (2008). Plasma membranes are poised for activation of raft phase coalescence at physiological temperature. *Proc. Natl. Acad. Sci. U.S.A.* 105, 10005–10010.
- Lingwood, D., and Simons, K. (2010). Lipid rafts as a membrane-organizing principle. *Science* 327, 46–50.
- Liu, J., Elmore, J. M., Fuglsang, A. T., Palmgren, M. G., Staskawicz, B. J., and Coaker, G. (2009). RIN4 functions with plasma membrane H⁺-ATPases to regulate stomatal apertures during pathogen attack. *PLoS Biol.* 7, e1000139. doi: 10.1371/journal.pbio.1000139
- Lu, Y. J., Schornack, S., Spallek, T., Geldner, N., Chory, J., Schellmann, S., Schumacher, K., Kamoun, S., and Robatzek, S. (2012). Patterns of plant subcellular responses to successful oomycete infections reveal differences in host cell reprogramming and endocytic trafficking. *Cell. Microbiol.* 14, 682–697.
- Malinska, K., Malinsky, J., Opekarova, M., and Tanner, W. (2003). Visualization of protein compartmentation within the plasma membrane of living yeast cells. *Mol. Biol. Cell* 14, 4427–4436.
- Marin, M., and Ott, T. (2012). Phosphorylation of intrinsically disordered regions in remorin proteins. *Front. Plant Sci.* 3:86. doi: 10.3389/fpls.2012.00086
- Martiniere, A., Gayral, P., Hawes, C., and Runions, J. (2011). Building bridges: formin1 of *Arabidopsis* forms a connection between the cell wall and the actin cytoskeleton. *Plant J.* 66, 354–365.
- Martiniere, A., Lavagi, I., Nageswaran, G., Rolfe, D. J., Maneta-Peyret, L., Luu, D. T., Botchway, S. W., Webb, S. E., Mongrand, S., Maurel, C., Martin-Fernandez, M. L., Kleine-Vehn, J., Friml, J., Moreau, P., and Runions, J. (2012). Cell wall constrains lateral diffusion of plant plasma-membrane proteins. *Proc. Natl. Acad. Sci. U.S.A.* doi: 10.1073/pnas.1202040109 [Epub ahead of print].
- Micali, C. O., Neumann, U., Grunewald, D., Panstruga, R., and O'Connell, R. (2011). Genesis of a specialized plant–fungal interface during host cell internalization of *Golovinomyces orontii* haustoria. *Cell. Microbiol.* 13, 210–226.
- Mongrand, S., Morel, J., Laroche, J., Claverol, S., Carde, J. P., Hartmann, M. A., Bonneau, M., Simon-Plas, F., Lessire, R., and Bessoule, J. J. (2004). Lipid rafts in higher plant cells – purification and characterization of triton X-100-insoluble microdomains from tobacco plasma membrane. *J. Biol. Chem.* 279, 36277–36286.
- Mongrand, S., Stanislas, T., Bayer, E. M., Lherminier, J., and Simon-Plas, F. (2010). Membrane rafts in plant cells. *Trends Plant Sci.* 15, 656–663.
- Morel, J., Claverol, S., Mongrand, S., Furt, F., Fromentin, J., Bessoule, J. J., Blein, J. P., and Simon-Plas, F. (2006). Proteomics of plant detergent-resistant membranes. *Mol. Cell. Proteomics* 5, 1396–1411.
- Morrow, I. C., Rea, S., Martin, S., Prior, I. A., Prohaska, R., Hancock, J. F., James, D. E., and Parton, R. G. (2002). Flotillin-1/Reggie-2 traffics to surface raft domains via a novel Golgi-independent pathway – identification of a novel membrane targeting domain and a role for palmitoylation. *J. Biol. Chem.* 277, 48834–48841.
- Munro, S. (2003). Lipid rafts: elusive or illusive? *Cell* 115, 377–388.
- Murphy, S. C., Fernandez-Pol, S., Chung, P. H., Murthy, S. N. P., Milne, S. B., Salomao, M., Brown, H. A., Lomasney, J. W., Mohandas, N., and Haldar, K. (2007). Cytoplasmic remodeling of erythrocyte raft lipids during infection by the human malaria parasite *Plasmodium falciparum*. *Blood* 110, 2132–2139.
- Nixon, B., Mitchell, L. A., Anderson, A. L., McLaughlin, E. A., O'Bryan, M. K., and Aitken, R. J. (2011). Proteomic and functional analysis of human sperm detergent resistant membranes. *J. Cell. Physiol.* 226, 2651–2665.
- Owen, D. M., Neil, M. A., French, P. M., and Magee, A. I. (2007). Optical techniques for imaging membrane lipid microdomains in living cells. *Semin. Cell Dev. Biol.* 18, 591–598.
- Panter, S., Thomson, R., De Bruxelles, G., Laver, D., Trevaskis, B., and Udvardi, M. (2000). Identification with proteomics of novel proteins associated with the peribacteroid membrane of soybean root nodules. *Mol. Plant Microbe Interact.* 13, 325–333.
- Pathak, P., and London, E. (2011). Measurement of lipid nanodomain (raft) formation and size in sphingomyelin/POPC/cholesterol vesicles shows TX-100 and transmembrane helices increase domain size by coalescing preexisting nanodomains but do not induce domain formation. *Biophys. J.* 101, 2417–2425.
- Pike, L. J. (2006). Rafts defined: a report on the keystone symposium on lipid rafts and cell function. *J. Lipid Res.* 47, 1597–1598.
- Pralle, A., Keller, P., Florin, E. L., Simons, K., and Horber, J. K. H. (2000). Sphingolipid-cholesterol rafts diffuse as small entities in the plasma membrane of mammalian cells. *J. Cell Biol.* 148, 997–1007.
- Raffaele, S., Bayer, E., Lafarge, D., Cluzet, S., Retana, S. G., Boubekeur, T., Leborgne-Castel, N., Carde, J. P., Lherminier, J., Noirot, E., Satiat-Jeunemaitre, B., Laroche-Traineau, J., Moreau, P., Ott, T., Maule, A. J., Raymond, P., Simon-Plas, F., Farmer, E. E., Bessoule, J. J., and Mongrand, S. (2009). Remorin, a Solanaceae protein resident in membrane rafts and plasmodesmata, impairs potato virus X movement. *Plant Cell* 21, 1541–1555.
- Robatzek, S., Chinchilla, D., and Boller, T. (2006). Ligand-induced endocytosis of the pattern recognition receptor FLS2 in *Arabidopsis*. *Genes Dev.* 20, 537–542.
- Roche, Y., Gerbeau-Pissot, P., Buhot, B., Thomas, D., Bonneau, L., Gresti, J., Mongrand, S., Perrier-Cornet, J. M., and Simon-Plas, F. (2008). Depletion of phytosterols from the plant plasma membrane provides evidence for disruption of lipid rafts. *FASEB J.* 22, 3980–3991.
- Saalbach, G., Erik, P., and Wienkoop, S. (2002). Characterisation by proteomics of peribacteroid space and peribacteroid membrane preparations from pea (*Pisum sativum*) symbiosomes. *Proteomics* 2, 325–337.
- Sako, Y., and Kusumi, A. (1995). Barriers for lateral diffusion of transferrin receptor in the plasma membrane as characterized by receptor dragging by laser tweezers: fence versus tether. *J. Cell Biol.* 129, 1559–1574.
- Sako, Y., Nagafuchi, A., Tsukita, S., Takeichi, M., and Kusumi, A. (1998). Cytoplasmic regulation of the movement of E-cadherin on the free cell surface as studied by optical tweezers and single particle tracking: corraling and tethering by the membrane skeleton. *J. Cell Biol.* 140, 1227–1240.
- Schroeder, R., London, E., and Brown, D. (1994). Interactions between saturated acyl chains confer detergent resistance on lipids and glycosylphosphatidylinositol (GPI)-anchored proteins – GPI-anchored proteins in liposomes and cells show similar behavior. *Proc. Natl. Acad. Sci. U.S.A.* 91, 12130–12134.
- Shahollari, B., Peskan-Berghofer, T., and Oelmüller, R. (2004). Receptor kinases with leucine-rich repeats are enriched in Triton X-100 insoluble plasma membrane microdomains from plants. *Physiol. Plant.* 122, 397–403.
- Sheets, E. D., Lee, G. M., Simson, R., and Jacobson, K. (1997). Transient confinement of a glycosylphosphatidylinositol-anchored protein in the plasma membrane. *Biochemistry* 36, 12449–12458.
- Simon-Plas, F., Elmayan, T., and Blein, J. P. (2002). The plasma membrane oxidase NtrbohD is responsible for AOS production in elicited tobacco cells. *Plant J.* 31, 137–147.
- Simons, K., and Gerl, M. J. (2010). Revitalizing membrane rafts: new tools and insights. *Nat. Rev. Mol. Cell Biol.* 11, 688–699.
- Simons, K., and Ikonen, E. (1997). Functional rafts in cell membranes. *Nature* 387, 569–572.
- Simon, R., Sheets, E. D., and Jacobson, K. (1995). Detection of temporary lateral confinement

- of membrane-proteins using single-particle tracking analysis. *Biophys. J.* 69, 989–993.
- Singer, S. J., and Nicolson, G. L. (1972). Fluid mosaic model of structure of cell-membranes. *Science* 175, 720–731.
- Spira, F., Mueller, N. S., Beck, G., Von Olshausen, P., Beig, J., and Wedlich-Soldner, R. (2012). Patchwork organization of the yeast plasma membrane into numerous coexisting domains. *Nat. Cell Biol.* 14, 640–648.
- Stanislas, T., Bouyssié, D., Rossignol, M., Vesa, S., Fromentin, J., Morel, J., Pichereaux, C., Monsarrat, B., and Simon-Plas, F. (2009). Quantitative proteomics reveals a dynamic association of proteins to detergent-resistant membranes upon elicitor signaling in tobacco. *Mol. Cell. Proteomics* 8, 2186–2198.
- Suzuki, K., and Sheetz, M. P. (2001). Binding of cross-linked glycosylphosphatidylinositol-anchored proteins to discrete actin-associated sites and cholesterol-dependent domains. *Biophys. J.* 81, 2181–2189.
- Takahashi, D., Kawamura, Y., Yamashita, T., and Uemura, M. (2012). Detergent-resistant plasma membrane proteome in oat and rye: similarities and dissimilarities between two monocotyledonous plants. *J. Proteome Res.* 11, 1654–1665.
- Tanner, W., Malinsky, J., and Opekarova, M. (2011). In plant and animal cells, detergent-resistant membranes do not define functional membrane rafts. *Plant Cell* 23, 1191–1193.
- Toth, K., Stratil, T. F., Madsen, E. B., Ye, J. Y., Popp, C., Antolin-Llovera, M., Grossmann, C., Jensen, O. N., Schussler, A., Parniske, M., and Ott, T. (2012). Functional domain analysis of the remorin protein LjSYM-REM1 in *Lotus japonicus*. *PLoS ONE* 7, e30817. doi: 10.1371/journal.pone.0030817
- van der Meer-Janssen, Y. P. M., Van Galen, J., Batenburg, J. J., and Helms, J. B. (2010). Lipids in host–pathogen interactions: pathogens exploit the complexity of the host cell lipidome. *Prog. Lipid Res.* 49, 1–26.
- Varma, R., and Mayor, S. (1998). GPI-anchored proteins are organized in submicron domains at the cell surface. *Nature* 394, 798–801.
- Xu, X. L., Bittman, R., Dupontail, G., Heissler, D., Vilcheze, C., and London, E. (2001). Effect of the structure of natural sterols and sphingolipids on the formation of ordered sphingolipid/sterol domains (rafts). *J. Biol. Chem.* 276, 33540–33546.
- Zacharias, D. A., Violin, J. D., Newton, A. C., and Tsien, R. Y. (2002). Partitioning of lipid-modified monomeric GFPs into membrane microdomains of live cells. *Science* 296, 913–916.
- Zappel, N. F., and Panstruga, R. (2008). Heterogeneity and lateral compartmentalization of plant plasma membranes. *Curr. Opin. Plant Biol.* 11, 632–640.
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