



## OPEN ACCESS

## EDITED BY

Junco S. Warren,  
Virginia Tech Carilion, United States

## REVIEWED BY

Tlili Barhoumi,  
King Abdullah International Medical Research  
Center (KAIMRC), Saudi Arabia  
Prabhu Mathiyalagan,  
Benthos Prime Central, United States

## \*CORRESPONDENCE

Zhong Wu,  
✉ wuzhong71@scu.edu.cn  
Wei-Tao Liang,  
✉ 1023444366@qq.com

<sup>†</sup>These authors have contributed equally to this work

RECEIVED 01 December 2023

ACCEPTED 25 July 2024

PUBLISHED 09 August 2024

## CITATION

Jiang D, Yue H, Liang W-T and Wu Z (2024),  
Developmental endothelial locus 1: the present  
and future of an endogenous factor in vessels.  
*Front. Physiol.* 15:1347888.  
doi: 10.3389/fphys.2024.1347888

## COPYRIGHT

© 2024 Jiang, Yue, Liang and Wu. This is an  
open-access article distributed under the terms  
of the [Creative Commons Attribution License  
\(CC BY\)](#). The use, distribution or reproduction in  
other forums is permitted, provided the original  
author(s) and the copyright owner(s) are  
credited and that the original publication in this  
journal is cited, in accordance with accepted  
academic practice. No use, distribution or  
reproduction is permitted which does not  
comply with these terms.

# Developmental endothelial locus 1: the present and future of an endogenous factor in vessels

Daisong Jiang<sup>†</sup>, Honghua Yue<sup>†</sup>, Wei-Tao Liang\* and Zhong Wu\*

Department of Cardiovascular Surgery, West China Hospital, Sichuan University, Chengdu, Sichuan, China

Developmental Endothelial Locus-1 (DEL-1), also known as EGF-like repeat and discoidin I-like domain-3 (EDIL3), is increasingly recognized for its multifaceted roles in immunoregulation and vascular biology. DEL-1 is a protein that is mainly produced by endothelial cells. It interacts with various integrins to regulate the behavior of immune cells, such as preventing unnecessary recruitment and inflammation. DEL-1 also helps in resolving inflammation by promoting efferocytosis, which is the process of clearing apoptotic cells. Its potential as a therapeutic target in immune-mediated blood disorders, cardiovascular diseases, and cancer metastasis has been spotlighted due to its wide-ranging implications in vascular integrity and pathology. However, there are still unanswered questions about DEL-1's precise functions and mechanisms. This review provides a comprehensive examination of DEL-1's activity across different vascular contexts and explores its potential clinical applications. It underscores the need for further research to resolve existing controversies and establish the therapeutic viability of DEL-1 modulation.

## KEYWORDS

developmental endothelial locus-1, integrin, inflammation, efferocytosis, vascular smooth muscle cell

## 1 Introduction

Increasing evidence suggests that local tissues, which were previously thought to be passive recipients of immunity, have equally active regulatory functions (Galli et al., 2011). The requirements of the tissues to which immune cells are recruited or reside are constantly changing, and the function of these cells is constantly being adapted (Galli et al., 2011; Matzinger and Kamala, 2011; Oyler-Yaniv et al., 2017). Immune plasticity can manifest at different levels, from the cellular to the molecular level (Galli et al., 2011). Cellular plasticity refers to the ability of cells to adapt and change their characteristics or functions in response to various stimuli or environmental cues (Oyler-Yaniv et al., 2017; Hajishengallis and Chavakis, 2019).

DEL-1 is a 52 kDa protein encoded by the EDIL3 gene (C Hidai 1, 1998). DEL-1 contains three epidermal growth factor (EGF)-like repeats at the N-terminus and the Arg-Gly-Asp (arginine-glycine-aspartate, RGD) module in the second EGF domain (EGF2) which combines with integrin to form a recognition system for cell adhesion (Ruoslahti, 1996; Schurpf et al., 2012). At the C-terminus, DEL-1 has two discoidal I-like domains capable of glycosaminoglycan and phosphatidylserine interactions (Hanayama et al., 2004; Hidai et al., 2007). DEL-1 is released from endothelial cells, mesenchymal stromal cells, and subsets of macrophages (Hajishengallis and Chavakis, 2019). DEL-1, due to its unique structure, can interact with a variety of integrin types, including Macrophage-1 antigen (Mac-1,  $\alpha$ M $\beta$ 2 integrin), lymphocyte function-associated antigen 1 (LFA-1,  $\alpha$ L $\beta$ 2 integrin),

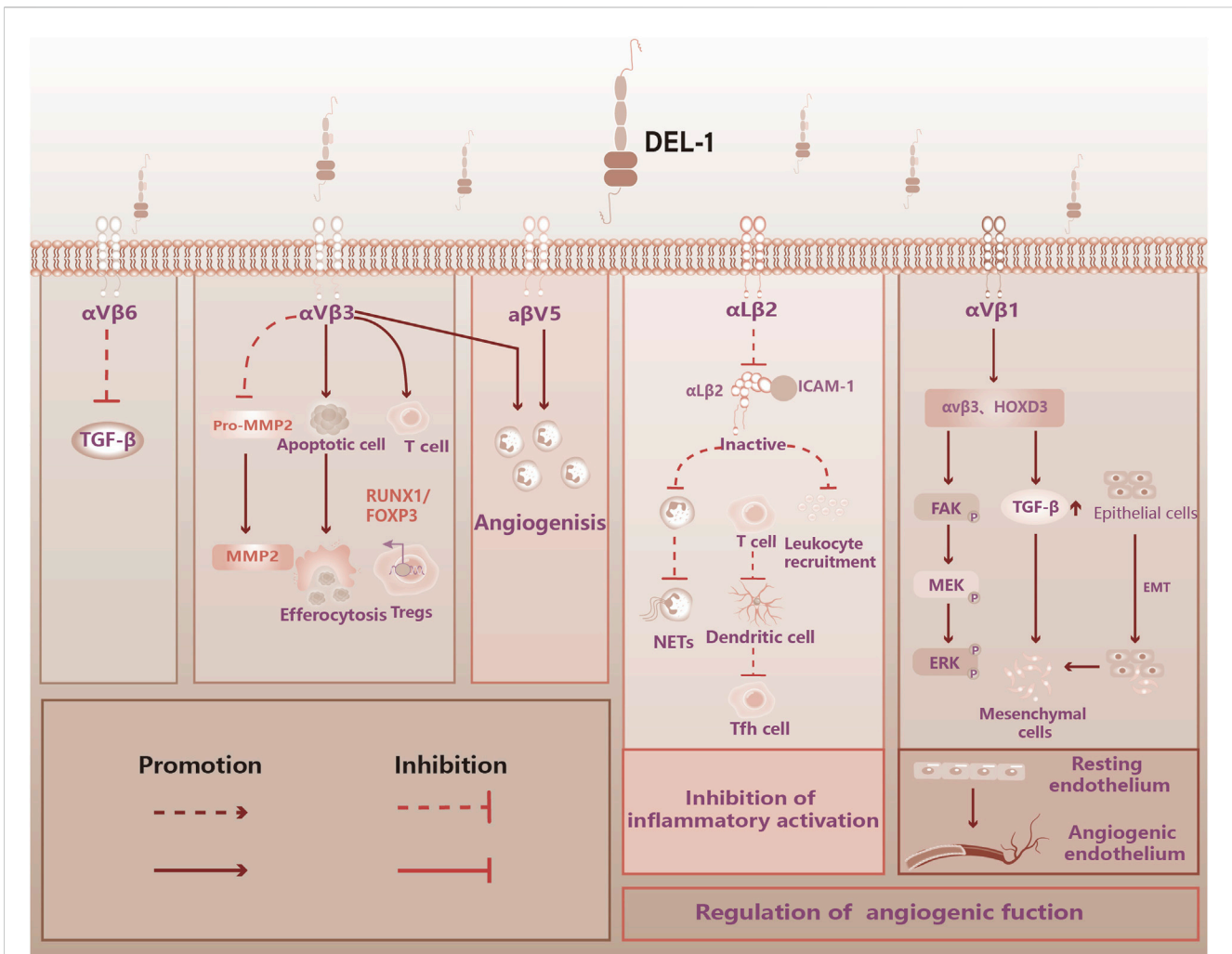
$\alpha v\beta 3$  integrin,  $\alpha v\beta 6$  integrin (Choi, 2009; Mitroulis et al., 2014; Kim et al., 2020; Li et al., 2021a). Its presence suggests that local factors regulate immune plasticity within tissues to maintain or reinstate tissue homeostasis (see Figure 1).

## 2 DEL-1 in immune modulation and inflammation

### 2.1 Structural and functional analysis of DEL-1

Integrins are versatile transmembrane receptors that function primarily as cell adhesion molecules, playing crucial roles in cellular

processes including signaling, migration, and tissue integrity (Rose et al., 2007). Integrins modulate a range of physiological and pathological processes, including by promoting or inhibiting the inflammatory response, contributing to thrombosis through platelet aggregation and endothelial interactions, and affecting tissue remodeling during fibrosis (Mezu-Ndubuisi and Maheshwari, 2021; Mrugacz et al., 2021; Van Hove et al., 2021). DEL-1 serves as a ligand for integrins and also interacts with phospholipids, playing a role in various integrin-mediated processes, modulating different stages of the host inflammatory response (Hajishengallis and Chavakis, 2019; Hajishengallis and Chavakis, 2021). DEL-1 regulates immune cell functions, including leukocytes and effector cells (such as T cells), through integrins (Hajishengallis et al., 2015; Wang et al., 2021) (see Table 1).



**FIGURE 1**  
 DEL-1 binds to integrin to perform immune functions in the vasculature. Figure 1 illustrates the physiological functions of DEL-1, which are known to bind to some of the integrins. DEL-1 binds to  $\alpha v\beta 3$  integrin to inhibit TGF- $\beta$  activation. DEL-1 binds to  $\alpha v\beta 3$  integrin: Inhibits the production of MMP2 by MMP2 precursors; Promotes phagocytosis of apoptotic cells by phagocytosis; Stabilized  $\alpha v\beta 3$  integrin-dependent Treg number; Promoting angiogenic effects. DEL-1 binding to  $\alpha v\beta 5$  integrin: Promoting angiogenic effects. DEL-1 binds to  $\alpha L\beta 2$  to inhibit inflammatory effects: Inhibition of Leukocyte aggregation; Inhibition of neutrophil release of neutrophil extracellular traps (NETs); Inhibition of T cell conversion to Tfh cell. DEL-1 binds to  $\alpha v\beta 1$ : regulation of the corresponding signaling pathway promotes the transformation of resting endothelium into angiogenic endothelium. TGF- $\beta$ : Transforming Growth Factor-beta, FOXP3: Forkhead Box Protein P3, NETs: Neutrophil Extracellular Traps, HOXD3: Homeobox D3, FAK: Focal Adhesion Kinase, MEK: Mitogen-Activated Protein Kinase Kinase, ERK: Extracellular Signal-Regulated Kinase, EMT: Epithelial-Mesenchymal Transition.

TABLE 1 Important studies on integrin-mediated downstream immune function by DEL-1.

Type of Integrin	First author	Year	Subject (cell, participants, animals)	Main effect
$\alpha_1\beta_2$ (LFA-1)	Wang H et al. (Wang et al., 2021)	2021	C57BL/6J mice	Inhibiting the induction of Tfh cells
	Yang N et al. (Yang et al., 2015)	2015	BMSCs / C57BL/6J mice	Transformation of Th17 cells to Treg
	Eskan MA et al. (Eskan et al., 2012)	2012	C57BL/6J mice	Inhibition of IL-17 mediated inflammation
	Yan S et al. (Yan et al., 2018)	2018	asthmatic patients, controls	
$\alpha M\beta 2$ (Mac-1)	Shin J et al. (Shin et al., 2015)	2015	C57BL/6J mice, cynomolgus monkeys	Inhibition the osteoclastogenesis
$\alpha v\beta 3$	Kourtzelis I et al. (Kourtzelis et al., 2019)	2019	HMDMs, BMDMs, peritoneal macrophages, neutrophils, C57BL/6 mice	Enhances efferocytic activity
	Li X et al. (Li et al., 2020a)	2020	Human peripheral Treg, C57BL/6 mice	Reducing monocyte/macrophage and T cell infiltration
$\alpha v\beta 3/\alpha v\beta 5$	Zhong J et al. (Zhong et al., 2003)	2003	HUVEC, C57BL/6 mice, New Zealand white rabbits	Promotes angiogenesis
$\alpha v\beta 6$	Kim DY et al. (Kim et al., 2020b)	2020	Primary lung fibroblasts, C57BL/6 mice	Hinders integrin-mediated TGF- $\beta$ activation

HMDMs: Human monocyte-derived macrophages, BMDMs; Mouse bone marrow-derived macrophages, HUVEC: Human umbilical vein endothelial cells.

## 2.2 DEL-1 combining integrins to regulate immune response and suppress inflammation

Endothelial cell-derived DEL-1 acts on integrins such as LFA-1, and Mac-1, interrupting the binding, to their respective ligands, thereby acting as an intrinsic regulator of leukocytes recruitment and a suppressor of early-stage inflammation (Mitroutis et al., 2014; Jia et al., 2023).

Previous studies have confirmed the anti-inflammatory properties of DEL-1 (Maekawa et al., 2020). Erythromycin-induced DEL-1 transcription attenuates LFA-1-mediated neutrophilic and mucosal inflammation (Maekawa et al., 2020). Additionally, DEL-1 transcription was mediated by Janus kinase 2 (JAK2) and the Phosphoinositide 3-kinase (PI3K)/Protein Kinase B (AKT) signaling pathway. The activation of these signaling pathways upregulates DEL-1 expression in lung microvascular endothelial cells. Targeting these signaling pathways to enhance DEL-1 expression could offer a novel approach to treating inflammatory and age-related diseases (Maekawa et al., 2020). In a mouse model of colon cancer, DEL-1 was shown to reduce neutrophil recruitment and infiltration while increasing the number of M1 macrophages as an efficient anticancer immunotherapy (Tian et al., 2022).

Modulating EDIL3 gene expression has been shown to influence immune cells and cytokines and may contribute to various pathologies, including acute lung injury and fibrosis (Choi et al., 2008; Kim et al., 2020; Li et al., 2022b). In several mouse models of hypertension, Filer et al. discovered that while DEL-1 upregulation did not lead to differential expression of pro-matrix metalloproteinase-2 (Pro-MMP-2) in the aorta and heart, it did inhibit  $\alpha v\beta 3$  integrin-dependent activation of Pro-MMP-2 in isolated mouse and human aortas. This inhibition prevented the transformation of Pro-MMP-2 to active MMP-2, significantly reducing MMP-2 expression (Failer et al., 2022).

Injection of recombinant DEL-1 protein has been observed to attenuate the increase in systolic blood pressure, likely through the prevention of adverse aortic remodeling, potentially by improving endothelial function and reducing endothelial permeability (Failer et al., 2022). Whereas DEL-1 deficiency has been correlated with heightened neutrophil infiltration and enhanced formation of neutrophil extracellular traps (NETs), thus exacerbating pressure-overload-induced heart failure. Conversely, *in vitro* experiments have revealed that DEL-1 can inhibit the formation of NETs in a Mac-1-dependent mechanism (Zhao et al., 2023).

## 2.3 DEL-1 regulates T cells' functions

### 2.3.1 DEL-1 interplay with Th17

IL-17, primarily released by Th17 cells, plays a crucial role in neutrophil recruitment, the production of the pro-inflammatory cytokines IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, and TNF, and host antimicrobial peptides, such as  $\beta$ -defensin 2 (Liang et al., 2006; Amatya et al., 2017; Altieri et al., 2018). IL-17 is involved in multiple autoimmune and cardiovascular diseases (Mills, 2023).

DEL-1 inhibits IL-17-induced immune cell recruitment and migration, effectively inhibiting inflammation associated with various diseases such as multiple sclerosis and bone loss (Eskan et al., 2012; Khader, 2012; Choi et al., 2015). Local administration of DEL-1 in the airways effectively curtails IL-17 production and diminishes neutrophil accumulation, reinforcing its role as an anti-inflammatory agent (Yan et al., 2018). DEL-1 exerts its inhibitory effects by targeting intercellular adhesion molecule 1 (ICAM-1)-dependent neutrophil recruitment, thereby mitigating airway inflammation (Choi et al., 2008). When the homeostatic regulation of DEL-1 and IL-17 is disrupted, it can drive associated inflammatory diseases when IL-17 is dominant (Hajishengallis and Chavakis, 2019). Previous studies have reported that IL-17 can downregulate DEL-1 in human endothelial cells by targeting the key transcription factor CCAAT/enhancer-binding protein (C/

EBP $\beta$ ) via glycogen synthase kinase-3 (GSK-3 $\beta$ ), and this effect can be reversed by Resolvin-D1 (RvD1) (Maekawa et al., 2015). Recent research indicates that DEL-1 and RvD1 collaboratively impede inflammation. The RvD1-DEL-1 axis inhibits IL-17-induced pathological inflammation in a DEL-1-dependent manner. Upon interaction with G-protein-coupled receptor 32 (GPR32) and lipoxin A4 receptor/formylpeptide receptor 2 (ALX/FPR2) on endothelial cells, RvD1 facilitates anti-inflammatory actions (Maekawa et al., 2015).

DEL-1 deficiency leads to the overproduction of IL-17 and neutrophil migration, resulting in increased endothelial permeability and damage to the blood-brain barrier (Choi et al., 2015). In vascular diseases, various stimuli upregulate IL-17 expression (Nishida et al., 2020). In Kawasaki disease, high DEL-1 antibody levels are associated with excessive IL-17-mediated inflammation, and DEL-1 deficiency can exacerbate the pro-inflammatory response and increase the likelihood of coronary atherogenesis (Consiglio et al., 2020). IL-17 can enhance WNT/ $\beta$ -catenin signaling, which exerts proliferative and anti-apoptotic effects on vascular smooth muscle cells (VSMCs) via this pathway (Zhang et al., 2018; Wang et al., 2019b). Th17/IL-17 can activate the downstream NF- $\kappa$ B pathway to induce endothelial cell senescence (Zhang et al., 2021). Given DEL-1's role in the downregulating IL-17 activity (Yan et al., 2018) its modulation presents a promising therapeutic avenue for the management of immune and inflammatory diseases characterized by aberrant IL-17 signaling.

### 2.3.2 DEL-1 promotes T cell phenotype maintenance

Regulatory T cells, also known as Treg cells, are a crucial type of immune cell that helps in preventing autoimmune diseases and maintaining immune tolerance (Kim, 2009). These cells suppress the function of effector T cells which can cause harm to the body. Although it is not yet clear how Treg cells protect the cardiovascular system, their positive effects suggest that they could be a promising treatment option (Meng et al., 2016). The bone marrow serves as a reservoir for Treg cells (Zou et al., 2004). Notably, glucocorticoid (GC)-induced leucine zipper (GILZ) expression in bone marrow mesenchymal stem cells (MSCs) increases the expression and secretion of DEL-1 (Yang et al., 2015b). Additionally, DEL-1 can facilitate the transformation of Th17 cells to Treg cells via GILZ, thus stimulating the generation of Treg cells (Yang et al., 2015a). T cells are a diverse group of immune cells found in atherosclerotic plaques, which are the buildup of fat and other substances in artery walls. T cells are distributed differently in the intimal layer of atherosclerotic plaques (Libby, 2012; Marchini et al., 2021). The forkhead box transcription factor (FOXP3) is a transcription factor that plays a key role in the development and function of Treg cells (Hori et al., 2003).

DEL-1 stabilized  $\alpha$ v $\beta$ 3 integrin-dependent Treg number, which reduced inflammatory cell recruitment and cytokine production in cardiovascular organs (Failer et al., 2022). *In vivo*, FoxP3<sup>+</sup> Treg cells can inhibit the development of atherosclerosis by exerting regulatory control over lipoprotein metabolism. Treg cell lineages demonstrate plasticity, and the downregulation or loss of FoxP3 expression in ApoB-reactive

(ApoB<sup>+</sup>) Treg cells was observed during lineage tracing in ApoE<sup>-/-</sup> mice (Wolf et al., 2020). Depletion of Tregs leads to impaired clearance of very low-density lipoprotein cholesterol (VLDL-C) and celiac remnants, and increased atherosclerosis (Klingenberg et al., 2013). This depletion of Tregs leads to the transformation of the Treg cell phenotype and the subsequent loss of their immunosuppressive function (Gaddis et al., 2018). Similarly, this regulatory mechanism may attenuate IL-17-mediated atherosclerotic disease (Roy et al., 2022). Undoubtedly, atherosclerosis is a multifactorial disease characterized by the accumulation of numerous insults and immune-related factors, which can disrupt the normal homeostatic properties of the endothelial monolayer (Libby et al., 2019; Libby, 2021). Local accumulation of cholesterol has an important impact on the development of atherosclerosis, especially low-density lipoprotein cholesterol (LDL-C), which triggers an immune response in CD4<sup>+</sup> T-helper cells (Th).

Cholesterol accumulation in cellular lipid rafts promotes the activation and proliferation of T cells as well as the differentiation of Th1 and Th17 cells (Chistiakov et al., 2019). HDL cholesterol was positively related to the relative frequency of conventional Tregs (CD25<sup>+</sup>CD127<sup>-</sup> cells in CD4<sup>+</sup>CD45RA<sup>-</sup> T cells). LDL-C levels were negatively correlated with the number of conventional Treg (Schmitz et al., 2023).

DEL-1 increases the expression of runt-related transcription factor-1 (RUNX1) and FOXP3 in conventional T cells, facilitating their conversion to induced Treg (Li et al., 2020). In the absence of exogenous TGF- $\beta$ 1, the stability of FOXP3 expression conferred upon Treg restimulation has been associated with reduced recruitment of inflammatory cells and decreased pro-inflammatory cytokine production, which is strongly linked to cardiovascular health (Li et al., 2020; Failer et al., 2022). Intriguingly, Treg cells can also transform proatherogenic subsets, including CD8<sup>+</sup> T cells and  $\gamma$  $\delta$  T cells. However, the mutual crosstalk between T-cell subsets and different types of immune cells has not been clarified (Saigusa et al., 2020). In addition, previous studies did not specifically regulate DEL-1 or further investigate the downstream pathways or mechanisms.

### 2.4 Macrophage-derived DEL-1 regulates efferocytosis

Previous studies have indicated that DEL-1 inhibits the production of macrophage migration inhibitor (MIF) in macrophages and also suppresses nuclear factor kappa-B (NF- $\kappa$ B)-mediated downstream inflammatory activation (Lee et al., 2014).

Efferocytosis, the process of removing dying cells, signifies the end of programmed cell death. Apoptotic cells are effectively phagocytosed and cleared through increased secretion of anti-inflammatory mediators and the activation of reparative transcriptional programs (Doran et al., 2020). Through the enzymatic action of scramblase, phosphatidylserine is exposed on the outer membrane of these dying cells, as a critical signal for phagocytes to recognize and engulf apoptotic cells (Leventis and

Grinstein, 2010; Segawa and Nagata, 2015). Efferocytosis is vital for maintaining vascular homeostasis and health, as it inhibits inflammation and attenuates the progression of lesions such as atherosclerosis and aortic dissection (Zhang et al., 2019; Doddapattar et al., 2022; De Meyer et al., 2024). During efferocytosis, apoptotic cell-derived nucleotides promote the proliferation of noninflammatory macrophages, thereby facilitating inflammation regression and establishing a regenerative feedback loop (Gerlach et al., 2021). Disruption of this positive feedback loop results in a decline in phagocytosis and secondary efferocytosis, causing secondary necrosis in apoptotic cells and the release of intracellular DAMPs (Nagata, 2018; Morioka et al., 2019). Deficient efferocytosis, as a sign of inadequate inflammation resolution, leads to the accumulation of secondarily necrotic macrophages (Kasikara et al., 2021). Enhancement of efferocytosis has potential therapeutic implications for the maintenance of vascular integrity (Lee et al., 2018).

Macrophage-derived DEL-1 aids in resolving inflammation by increasing efferocytosis. Specifically, the  $\alpha v \beta 3$  integrin on macrophages mediates the efferocytosis of apoptotic cells (Kourtzelis et al., 2019). Upon exposure to external pathogenic factors, macrophages may attenuate the DEL-1-induced efferocytosis and related downstream elements within macrophages themselves (Li et al., 2023a). DEL-1 has been implicated in several cellular signaling pathways related to efferocytosis. Sirtuins (SIRT) are a family of NAD<sup>+</sup>-dependent deacetylases, including histone deacetylases that regulate glucose homeostasis, inflammation, genome stability, and DNA repair (Chang and Guarente, 2014; Liu et al., 2021). SIRT6 regulates macrophage infiltration and inflammation resolution by targeting the miR-216/217 cluster in the DEL-1/CD36 axis and contributes to the resolution of inflammation in diabetic periodontitis (Zhao et al., 2021). DEL-1 also exhibits promise in the upstream regulation of efferocytosis, which is mediated by SIRT6 (Grootaert et al., 2021; Li et al., 2022c). It promotes efferocytosis to enhance the resolution of inflammation and has been shown to alleviate hypertension-induced fibrosis, medial thickening, and elastin loss in the aortic epicardium through its immunomodulatory effects (Dasgupta et al., 2012b; Failer et al., 2022).

To sum up, the expression of DEL-1 promotes macrophage efferocytosis and the elimination of apoptotic cells and prevents the activation of the immune response (Yatim et al., 2017). Efferocytosis promotes macrophage production of vascular endothelial growth factor C (VEGFC) and effectively inhibits pro-inflammatory macrophage activation (Glinton et al., 2022). Endothelial cells can produce vasoactive substances to alter or maintain hemodynamics (Kruger-Genge et al., 2019).

The protein PCSK9 is implicated in accelerating vascular aging by hindering efferocytosis in endothelial cells. This indicates that efferocytosis may play a crucial role in slowing down vascular aging (Liu et al., 2023). Although the role of DEL-1 in promoting efferocytosis in non-macrophagic cells has not been established, research is required to confirm whether DEL-1 is involved in efferocytosis in endothelial cells, as endothelial cells are one of the significant producers of DEL-1 (Hidai et al., 1998). This could help to identify the potential protective mechanisms of DEL-1.

### 3 The enigma of DEL-1 in atherosclerosis

DEL-1 may promote atherosclerosis, contrary to earlier descriptions of its anti-inflammatory and efferocytosis effects (Hajishengallis and Chavakis, 2019). Surprisingly, DEL-1 expression was predominantly localized to foam cells within the intimal layers of coronary arteries in patients with coronary heart disease (CHD) and sudden cardiac death (SCD) and was significantly elevated compared to that in healthy people. In ApoE<sup>-/-</sup> mice, a positive correlation was observed between macrophage-derived DEL-1 levels and various atherosclerosis-related factors, such as plaque area, luminal stenosis, plaque foam cell counts, and downstream inflammatory factors. Moreover, the inhibition of the upstream pathway resulted in the suppression of DEL-1 expression, thereby exacerbating atherosclerotic lesions (Lu et al., 2023). This finding underscores the potential significance of DEL-1 in the regulation of atherosclerosis and warrants further investigation of its role in atherosclerosis. Previous studies have indicated that DEL-1 inhibits the production of macrophage migration inhibitor (MIF) in macrophages and also suppresses nuclear factor kappa-B (NF- $\kappa$ B)-mediated downstream inflammatory activation. DEL-1 is implicated in cardiovascular pathology through contrasting roles; while it appears to confer protection against inflammatory processes and vascular damage, recent evidence shows it may be associated with the promotion of atherosclerosis (Lee et al., 2014).

Chronic local inflammatory dysregulation may disrupt the regulation of endothelial immunity, such as enhanced vascular permeability, which regulates the recruitment of white blood cells and the activation of immune cells, may be impaired (Lee et al., 2014; Wang et al., 2022a). Instead, macrophages exert their pro-inflammatory effects after foam cell formation during atherosclerosis (Rader and Puré, 2005; Li et al., 2022a). The study mentioned above did not investigate the effects of the specific regulation of DEL-1 on downstream signaling, and there may be non-redundant signals other than DEL-1 that play a cross-regulatory role, thus affecting the results. However, previous research has shown that DEL-1 can bind to the phospholipid structure of oxidized low-density lipoprotein (oxLDL) and can inhibit the uptake of oxLDL by human coronary artery endothelial cells (HCAECs) or macrophages in a dose-dependent manner (Kakino et al., 2016). DEL-1 also inhibits the oxLDL-induced expression of monocyte chemoattractant protein-1 (MCP-1) and ICAM-1, as well as the secretion of endothelin-1 by HCAECs, ultimately mitigating atherosclerosis (Bennett et al., 2016; Kakino et al., 2016). The impact of the mice's age on the experimental results has not yet been determined. There are methodological differences between the use of different drugs in animal models of atherosclerosis that may contribute to discrepancies in research findings related to the role of DEL-1 in atherosclerosis (Kakino et al., 2016; Emini Veseli et al., 2017). Given the variance in findings, it is critical to verify the physiological role of DEL-1 in atherosclerosis using a variety of animal models to ensure the consistency and reproducibility of results.

Additionally, overexpression of DEL-1 under atherosclerotic conditions may only be a feed-forward mechanism closely related to the complex inflammatory state of the disease and is not sufficient

to suggest that DEL-1 promotes atherosclerosis (Di Pietro et al., 2023). DEL-1 effectively regulates the recruitment and chemotaxis of leukocytes, thereby preventing excessive inflammation (Hajishengallis and Chavakis, 2019). This is an important mechanism of its potential therapeutic effect.

## 4 DEL-1 facilitates angiogenesis

Angiogenesis, the process of forming new blood vessels, is driven by signals originating from surrounding tissues (Ramasamy et al., 2015; Jeong et al., 2021). Endothelial cells not only participate in this process by releasing proangiogenic growth factors but also exhibit dramatic plasticity, regulating vascular shape and organization through autocrine and paracrine signaling mechanisms (Bentley et al., 2014; McCoy et al., 2021). They can release inflammatory mediators, cell adhesion molecules, and immunomodulatory molecules to regulate normal vascular physiology (Shao et al., 2020). When endothelial cells are exposed to pathogens or inflammatory stressors, they may undergo a cascade of maladaptive changes including disruption of glycolytic and lipoprotein pathways, thereby contributing to vascular diseases (Xu et al., 2021).

DEL-1 is notable for its role in promoting angiogenesis, which is the formation of new blood vessels. The protein is actively expressed by embryonic endothelial cells where it contributes to the intricate shaping and remodeling of the developing vasculature, illustrating its importance during early growth stages (Hidai et al., 1998). DEL-1-induced angiogenesis is dependent on the integrins  $\alpha v \beta 3$  and  $\alpha v \beta 5$ , in addition to some forms of embryonic and tumor-induced angiogenesis (Zhong et al., 2003). Studies have demonstrated that DEL-1 can interact with vascular endothelial growth factor (VEGF) to regulate its role in angiogenesis (Zou et al., 2009; Niu et al., 2023). This interaction may have the potential to prevent aortic dissection by increasing VEGF pathway inhibitors (Oshima et al., 2017).

Evidence suggests that DEL-1 (whether administered or overexpressed) contributes to angiogenesis, and a lack of endogenous DEL-1 has not been shown to impact normal angiogenesis in animal models (Ho et al., 2004). The aortic ring assay showed that neither endothelial angiogenesis nor developmental angiogenesis in the retina was affected by DEL-1 deficiency (Klotzsche-von Ameln et al., 2017). This suggests that constitutive DEL-1 expression does not promote physiological angiogenesis (Hsu et al., 2008). It has been suggested that DEL-1 prevents ischemia-driven neovascularization by inhibiting inflammatory activation downstream of  $\alpha L \beta 2$  (Klotzsche-von Ameln et al., 2017). The functional differences exhibited by DEL-1, may be influenced by environmental factors. In the disc angiogenesis system, DEL-1 was shown to upregulate the expression of fibroblast growth factor and significantly enhance fibrovascular growth (Ho et al., 2004). Clinical investigations into DEL-1's therapeutic applications have shown promise, with the administration of DEL-1 plasmids leading to improved clinical manifestations in patients suffering from peripheral arterial disease, though the improvements did not achieve statistical significance, which warrants further study to conclusively determine efficacy (Grossman et al., 2007).

An increase in endothelial expression of DEL-1 can promote angiogenesis and endothelial cell migration (Beckham et al., 2014; Cobb et al., 2021). Cohort studies of patients with uterine cancer have shown that an increase in EDIL3 in dermal mesenchymal stem cells (DMSCs) stimulates the expression of  $\alpha v \beta 3$  and  $\alpha 5 \beta 1$  from endothelial cells and promotes endothelium-associated angiogenesis (Niu et al., 2022a; Niu et al., 2022b). These changes were not limited to physiological conditions. DEL-1 expression levels in psoriatic DMSCs are 2.54-fold higher than those in healthy controls (Niu et al., 2016). This activation results in excessive angiogenesis and vasodilation, ultimately leading to an increase in both epidermal thickness and microvascular density (Niu et al., 2020; Niu et al., 2022b). DEL-1 can promote vascular repair and neovascularization, leading to adverse remodeling. DEL-1 also interacts with  $\alpha v \beta 6$  integrin, which is a known activator of transforming growth factor-beta (TGF- $\beta$ ), and effectively inhibits integrin-mediated TGF- $\beta$  activation. Interestingly, the DEL-1-mediated inhibition of TGF- $\beta$  signaling only occurs in response to increased levels of  $\alpha v$  integrins (Kim et al., 2020). Specifically, DEL-1 mediates the activation of TGF- $\beta$  and ERK signaling, promoting epithelial-mesenchymal transition (EMT) and angiogenesis, a critical step that can contribute to the distant metastasis of tumor cells in hepatocellular carcinoma (Xia et al., 2015; Jeong et al., 2017).

The angiogenic effects of DEL-1 promote distant metastasis of tumor cells. However, DEL-1 inhibits inflammatory cell migration, thereby reducing associated tumor complications (Hyun et al., 2020). Further molecular and clinical research is crucial to determine the potential role of DEL-1 in the pathophysiology and treatment of peripheral vascular disease. It is also important to investigate whether uncontrolled neovascularisation and adverse vascular remodeling might occur.

## 5 DEL-1 in the clearance of platelets

Serving as versatile effectors in both thrombosis and hemostasis, platelets orchestrate immune responses by releasing pro-inflammatory cytokines, including IL-1 $\beta$ , thereby enhancing endothelial permeability and guiding other immune cells to areas of injury or infection (Poher and Sessa, 2007; Koupnova et al., 2018; Margraf and Zarbock, 2019).

Platelet microparticles (PMPs) are tiny fragments of cells that are released from platelets, which can be triggered by different stimuli, such as the activation of platelets by thrombin, collagen, or other agonists. Platelet overactivation and PMPs play a key role in the pathogenesis of cardiovascular and autoimmune diseases (Lannan et al., 2014). The size of PMPs depends on the nature of cell activation stimuli (Ponomareva et al., 2017).

The adhesion of platelets to the activated endothelium is facilitated by increased expression of adhesion molecules. This process, along with the subsequent binding of platelets to leukocytes, is crucial for forming platelet-leukocyte aggregates. These aggregates are instrumental in atherogenesis, with ongoing platelet-leukocyte interactions being a central component of this pathology (May et al., 2007; Twarock et al., 2016; Pircher et al., 2019). In a high-glucose environment, PMPs markedly diminish endothelial nitric oxide (NO) bioavailability while concurrently

amplifying reactive oxygen species (ROS) synthesis. This dual action exacerbates endothelial dysfunction, culminating in heightened permeability and cellular damage (Wang et al., 2019a).

DEL-1 binds to platelet phosphatidylserine and mediates endothelial clearance of platelet particles. Due to redundant mechanisms, depletion of DEL-1 does not impact coagulation under normal physiological conditions but leads to enhanced coagulation in disease states characterized by pathological increases in particulate production (Dasgupta et al., 2012a). DEL-1 can selectively disrupt the interaction between Mac-1 integrins on leukocytes and platelets, thereby inhibiting the thrombotic inflammatory injury that can arise, for instance, during myocardial infarction when platelet-leukocyte aggregates contribute to thrombosis (Kourtzelis et al., 2016). Romanidou et al. reported that in pregnant patients with thrombotic microangiopathy, circulating DEL-1 levels are significantly diminished and may represent a significant biomarker (Romanidou et al., 2023). Although the specific mechanism by which the reduction in circulating DEL-1 contributes to this type of widespread systemic thrombosis has not been identified at this time, the clearance of PMPs by DEL-1 and inhibiting the associated inflammation and thrombosis may hold therapeutic potential in preventing thrombosis or vascular injury associated with the pathological state.

## 6 DEL-1 suppresses endoplasmic reticulum stress

The endoplasmic reticulum (ER) plays a key role in the synthesis, folding, and structural modification of proteins in cells (Schwarz and Blower, 2016). The endoplasmic reticulum stress (ER stress) response is a key regulator of inflammation and cell death. The capability of the ER to fold proteins varies greatly between cell types. When misfolded proteins accumulate in the endoplasmic reticulum, the ability of the ER to correctly fold and posttranslationally modify proteins can be compromised, leading to substantial cellular dysfunction which can contribute to both genetic mutations and vulnerability to environmental stressors (Wiseman et al., 2022). To maintain normal protein folding homeostasis, the endoplasmic reticulum activates the unfolded protein response (UPR) (Ron and Walter, 2007). However, excessive production of UPR signaling molecules can be detrimental. Persistent engagement of the UPR signaling cascade within the endoplasmic reticulum can also lead to an accumulation of misfolded proteins, resulting in ER stress (Oakes and Papa, 2015).

Prolonged ER stress can lead to an increased protein load and, if unmitigated, can ultimately result in cell death (Marciniak and Ron, 2006). Currently, two primary approaches target ER stress: one involves directly influencing the accumulation of misfolded proteins within the ER. While the other modulates the signaling of the unfolded protein response (UPR) via ER stress sensors or enzymes that regulate their downstream effects (Marciniak et al., 2022). The UPR is regulated by three endoplasmic reticulum transmembrane sensors: protein kinase R-like endoplasmic reticulum kinase (PERK), inositol-requiring enzyme 1 $\alpha$  (IRE1 $\alpha$ ), and activating transcription factor 6 (ATF6). These factors are crucial for mediating the UPR and coordinating the cellular response to ER stress (Yap et al., 2021). Overexpression of DEL-1

has been shown to attenuate LPS-induced oxidative stress (Li et al., 2022b). In addition, DEL-1 attenuated palmitate-induced and HFD-induced skeletal muscle ER stress and insulin resistance via SIRT1/SERCA2-mediated signaling (Sun et al., 2020).

The AMP-activated protein kinase (AMPK) and peroxisome proliferator-activated receptor delta (PPAR $\delta$ ) are key regulatory proteins within the cell (Kemmerer et al., 2015). Downregulation of this signaling could promote ER stress-induced vascular endothelial dysfunction in adult rat offspring exposed to maternal diabetes (Luo et al., 2022). DEL-1 counteracts ER stress by activating the AMPK pathway, which, in turn, induces autophagy—a crucial defense mechanism that helps preserve cellular function. Additionally, DEL-1 protects tendon cells from stress-induced apoptosis (Park et al., 2022) (see Figure 2).

Within the vasculature, vascular smooth muscle is situated in the middle layer of the vessel and controls vasoconstriction and diastole. However, deviations from normal physiological activity in vascular smooth muscle can induce phenotypic transformation or diminish VSMC populations. Such changes can concurrently injure the endothelium, undermining vascular integrity (Baumann et al., 1981; Kuang et al., 2016; Grewal et al., 2021). Emerging evidence highlights that ER stress in VSMCs contributes to the progression of various vascular diseases through the release of extracellular vesicles and the disruption of autophagy; these diseases include vascular calcification and aortic dissection (Clement et al., 2019; Li et al., 2021b; Furmanik et al., 2021). In response to pathogenic stimuli, VSMCs may undergo phenotypic transformation into macrophage-like cells, a process often driven by the accumulation of lipids within atherosclerotic plaques (Bennett et al., 2016).

Mild or transient ER stress caused by the accumulation of misfolded or unfolded proteins, binding immunoglobulin protein (BiP) dissociates from IRE1 $\alpha$  and ATF6 but remains bound to PERK. Then, IRE1 $\alpha$  dimerizes and is phosphorylated, leading to the splicing of X-box binding protein 1 (XBP-1) (Cubillos-Ruiz et al., 2017). Notably, this factor inhibits pathological processes, including oxidative stress in endothelial cells and autophagy in VSMCs (Zhao et al., 2013; Martin et al., 2014). Concurrently, BiP dissociation triggers the translocation and cleavage of ATF6. If ER stress persists, BiP will also dissociate from PERK. This action induces the phosphorylation of the downstream protein eukaryotic translation initiation factor 2 $\alpha$  (eIF2 $\alpha$ ), which is a crucial step in the ER stress response. This leads to the activation of the transcription factor C/EBP homologous protein (CHOP), which initiates the apoptotic pathway (Wu and Kaufman, 2006; Zhang et al., 2006; Cnop et al., 2017). In the case of atherosclerosis, prolonged and intensified ER stress contributes to sustained and enhanced UPR signaling, leading to cell death and exacerbating plaque formation in the arteries (Zhou and Tabas, 2013). Disturbed ER homeostasis triggers the progression of cardiovascular disease, which ultimately further exacerbates ER stress, creating a vicious circle (Ren et al., 2021).

Increased salivary gland CHOP expression in Sjögren's syndrome (SS) is implicated in the promotion of apoptosis and in enhancing leukocyte infiltration. This upregulation correlates with decreased DEL-1 levels, providing mechanistic insight into the relationship between CHOP and DEL-1 (Baban et al., 2013). Inhibition of unspliced XBP1 (XBP1u) increases vascular calcification and stimulates ER stress to promote vascular

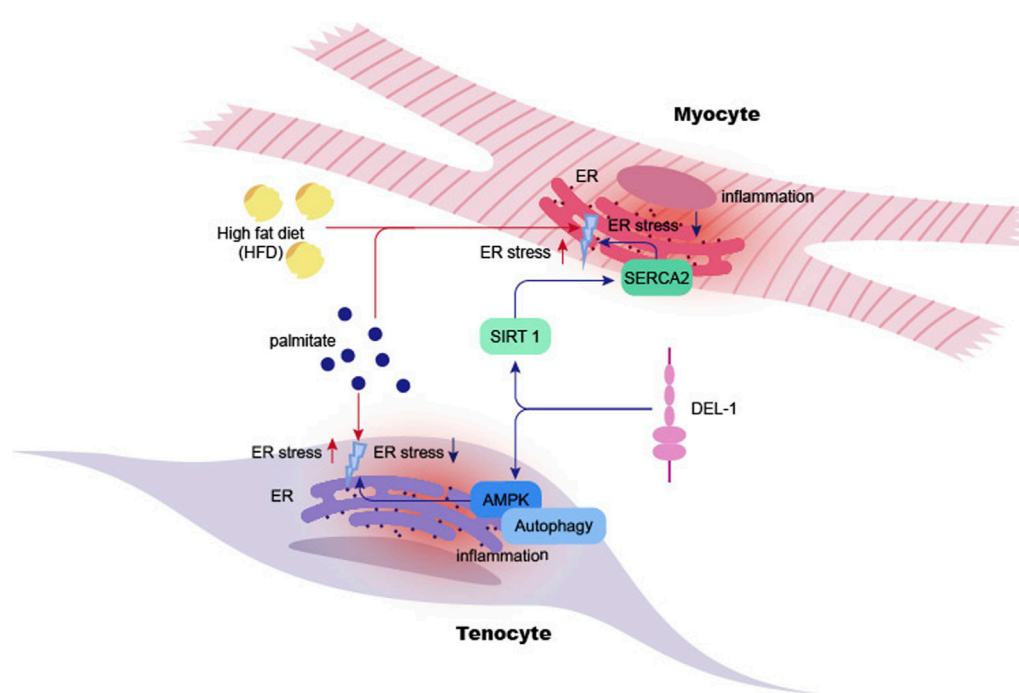


FIGURE 2

DEL-1 inhibits ER stress in myocyte and tenocyte. High-fat diet or palmitate significantly increases ER stress in myocyte cells in mice, leading to inflammation in myocytes and subsequent cellular damage. DEL-1 treatment enhances SIRT1 expression in myocytes in a dose-dependent manner, mitigating ER stress under palmitate treatment through the SIRT1/SERCA2 pathway. Additionally, DEL-1 enhances AMPK phosphorylation in tenocytes, initiating autophagy and reducing palmitate-induced inflammation. The figure indicates that DEL-1 helps alleviate muscle and tendon inflammation exacerbated by obesity. ER: Endoplasmic Reticulum, SIRT1: Sirtuin 1, SERCA2: Sarcoplasmic/Endoplasmic Reticulum Calcium ATPase 2, AMPK: AMP-activated protein kinase.

remodeling. XBP1u interacts with  $\beta$ -catenin and promotes its degradation via the ubiquitin-proteasome pathway. This interaction inhibits the transcriptional activity of the beta-catenin/TCF complex, which is responsible for regulating osteogenic markers such as runt-related transcription factor 2 (Runx2) and Msh homeobox 2 (Msx2), which play roles in vascular calcification (Yang et al., 2022). When the autophagy protein 5 (Atg5) gene is absent in VSMCs, autophagy is impaired, increasing the susceptibility of VSMCs to cell death. This impairment in autophagy enhances ER stress activation, subsequently promoting inflammation in VSMCs through an IRE-1 $\alpha$ -dependent mechanism (Clement et al., 2019). Cholesterol contributes to ER stress, which can trigger the conversion of VSMCs into cells exhibiting features characteristic of macrophages and fibroblasts, a process modulated by the UPR (Chattopadhyay et al., 2021).

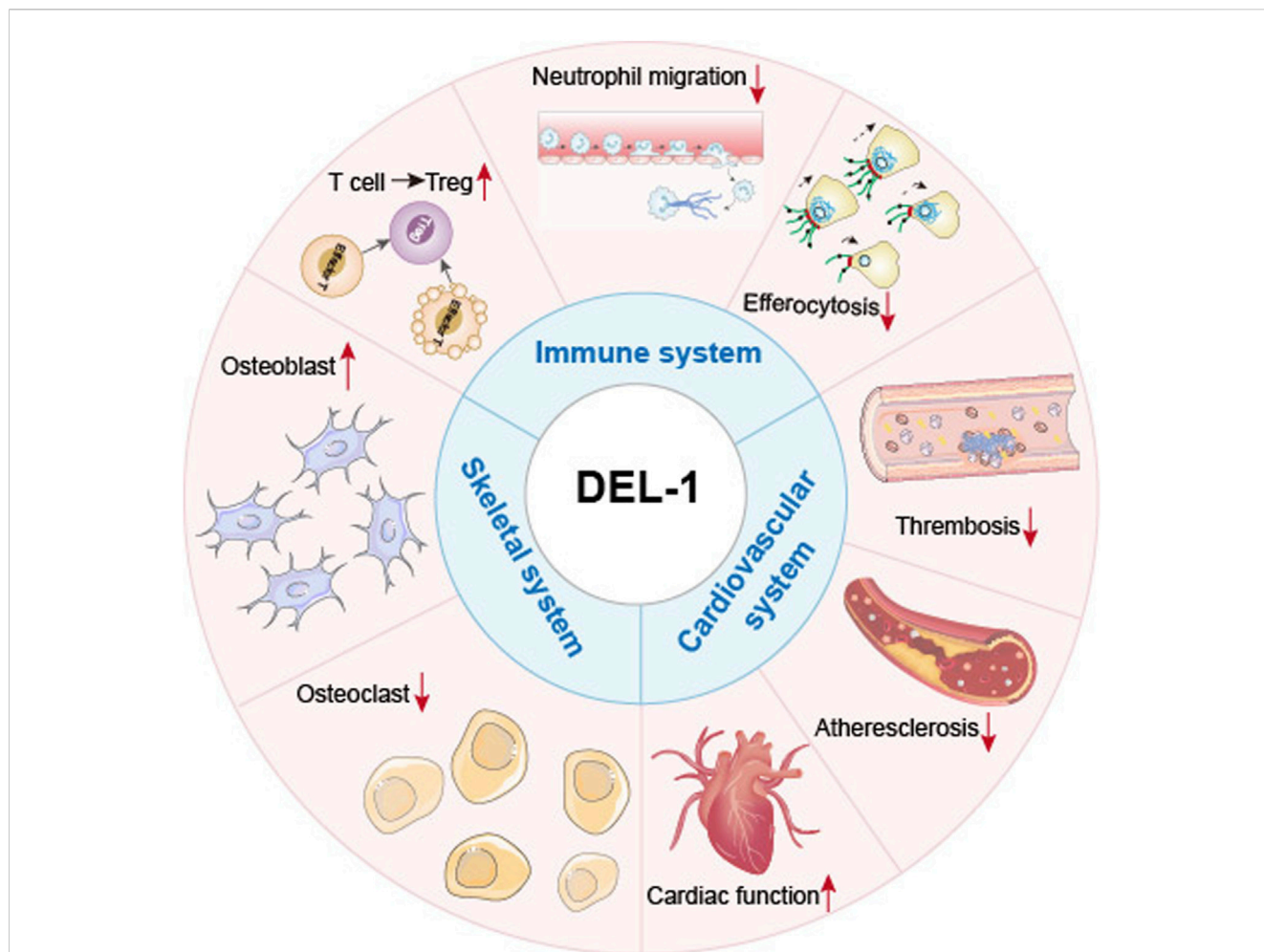
Available evidence suggests that DEL-1 contributes to the inhibition of ER stress-induced apoptosis (Park et al., 2022). Recently, DEL-1 levels have been observed to be elevated in response to exercise, by inhibiting ER stress. This inhibition can contribute to reduced inflammation and improved insulin sensitivity, as indicated by evidence (Kwon et al., 2020). However, to date, no specific research has explored the ability of DEL-1 to alleviate ER stress in VSMCs. It is important to note that the ER in skeletal muscle is referred to as the sarcoplasmic reticulum (SR). Skeletal muscle fibers contain the sarcoplasmic reticulum, a highly differentiated structure composed of tubular networks aligned with sarcomeres, reflecting distinct morphological and structural

characteristics compared to vascular smooth muscle (Cusimano et al., 2009; Sorrentino, 2011). In contrast, the ER in vascular smooth muscle, while lacking the specialized alignment of the SR, presents a reticulated network that pervades the entire cell. Investigating whether these structural differences influence the outcomes of DEL-1 intervention in VSMCs could provide valuable insights for therapeutic strategies. In addition to mitigating ER stress, DEL-1 may also protect against damage to VSMCs induced by other signaling pathways. For instance, signaling by the receptor activator of nuclear factor kappa-beta ligand (RANKL) and the cytoplasmic calcineurin-dependent nuclear factor of activated T-cells (NFATc1) has been shown to facilitate the upregulation of bone morphogenetic proteins (BMPs) (Panizo et al., 2009; Singh et al., 2012; Shiny et al., 2016). Furthermore, DEL-1 has been demonstrated to inhibit the RANK-NFATc1 signaling axis, thus impeding osteoblast activation (Shin et al., 2015). This interaction likely represents a crucial mechanism by which DEL-1 sustains vascular integrity and health. The paracrine signaling between endothelial cells (ECs) and vascular smooth muscle cells (VSMCs) is likely to play an instrumental role in mediating the vasoprotective effects of DEL-1.

## 7 DEL-1 as a potential therapeutic target and outlook in vessels

In various vascular diseases, inflammation is a crucial factor in promoting a spectrum of pathological changes (An et al.,





**FIGURE 3**  
 General roles of DEL-1 in various systems and cell types. Summary of the physiological roles of DEL-1 in various organs or cells within the cardiovascular, immune, and skeletal systems. In the cardiovascular system, DEL-1 improves cardiac function and inhibits thrombosis and the progression of atherosclerosis. In the immune system, DEL-1 exerts immunomodulatory functions through interactions with various integrins, including inhibiting immune cell activation processes such as neutrophil migration; enhancing the efferocytosis process of phagocytes to promote inflammation resolution; promoting the differentiation of other T cell types into Treg cells, and inhibiting the excessive activation and proliferation of effector T cells. In the skeletal system, DEL-1 promotes the maturation of osteoblasts, inhibits the maturation of osteoclasts, promotes bone regeneration, and reduces inflammatory responses.

2019). DEL-1 regulates the production of inflammatory factors and the activation of signaling pathways, thereby attenuating the impact of immune cells and their associated inflammation on the vasculature (Li et al., 2021a). The current evidence on the therapeutic potential of DEL-1 for cardiovascular disease remains a subject of debate. This controversy highlights the necessity for more rigorous clinical studies and mechanistic research to clarify the therapeutic benefits of DEL-1 in cardiovascular pathology (Zhao et al., 2022).

Further exploration of DEL-1 as a potential clinical therapeutic target or biomarker for vascular diseases is warranted. The differences in the expression and function of DEL-1 in different organs suggest that assessing its viability as a therapeutic target could be an important direction for future research (Figure 3). Amidst the ongoing debate on the use of DEL-1, its capacity to suppress inflammation and cytotoxicity should be acknowledged as a therapeutic option for

cardiovascular diseases, targeting multiple cell types including ECs and VSMCs (Wang et al., 2022b). Further research is required to develop treatment modalities or drug delivery methods. This would allow the investigation of crosstalk between DEL-1 and other regulators of angiogenesis, thereby advancing our understanding of the role of DEL-1 in cardiovascular disease treatment (Cabrera and Makino, 2022; Zhao et al., 2022).

Conditional Knockout of the EDIL3 gene using CRISPR/Cas9 gene-editing technology may help assess the effects of DEL-1 on downstream signaling molecules by reducing the interference from any compensatory mechanisms or indirect signaling pathways, assessing functional changes in the intima, media, and adventitia layers of the vasculature (Wang et al., 2022b; Li et al., 2023b). The immunomodulatory effects of DEL-1 vary among cell types and environments, highlighting its immune plasticity. Although a connection has been identified between DEL-1 and specific

vasoactive factors in angiogenesis (Niu et al., 2023), the interplay between DEL-1 and other molecules remains poorly understood. As vascular tissue engineering moves toward clinical applications, replicating vascular physiological function is one of the key barriers (Ciucurel and Sefton, 2014; Margolis et al., 2023). There have been limited studies exploring the use of DEL-1 in tissue engineering constructs to regulate endothelial cells (Ciucurel and Sefton, 2014). Looking forward, endogenous factors like DEL-1 may prove crucial for the fabrication of a multiscale vascular hierarchy under physiological conditions. While DEL-1 has shown promising safety results in a clinical study (Grossman et al., 2007), our incomplete understanding of its physiological functions underscores the need for refined animal experiments to comprehensively determine its effects on various systems before advancing to further clinical trials.

## Author contributions

DJ: Writing—original draft, Conceptualization. HY: Writing—review and editing, Visualization. W-TL: Writing—review and editing, Software. ZW: Supervision, Writing—review and editing, Project administration, Funding acquisition.

## References

- Altieri, A., Piyadasa, H., Recksiedler, B., Spicer, V., and Mookherjee, N. (2018). Cytokines IL-17, TNF and IFN- $\gamma$  alter the expression of antimicrobial peptides and proteins disparately: a targeted proteomics analysis using SOMAScan technology. *Vaccines (Basel)* 6, 51. doi:10.3390/vaccines6030051
- Amatya, N., Garg, A. V., and Gaffen, S. L. (2017). IL-17 signaling: the yin and the yang. *Trends Immunol.* 38, 310–322. doi:10.1016/j.it.2017.01.006
- An, Z., Li, J., Yu, J., Wang, X., Gao, H., Zhang, W., et al. (2019). Neutrophil extracellular traps induced by IL-8 aggravate atherosclerosis via activation NF- $\kappa$ B signaling in macrophages. *Cell. Cycle* 18, 2928–2938. doi:10.1080/15384101.2019.1662678
- Baban, B., Liu, J. Y., Abdelsayed, R., and Mozaffari, M. S. (2013). Reciprocal relation between GADD153 and Del-1 in regulation of salivary gland inflammation in Sjögren syndrome. *Exp. Mol. Pathol.* 95, 288–297. doi:10.1016/j.yexmp.2013.09.002
- Baumann, F. G., Catinella, F. P., Cunningham, J. N., JR., and Spencer, F. C. (1981). Vein contraction and smooth muscle cell extensions as causes of endothelial damage during graft preparation. *Ann. Surg.* 194, 199–211. doi:10.1097/0000658-198108000-00015
- Beckham, C. J., Olsen, J., Yin, P. N., Wu, C. H., Ting, H. J., Hagen, F. K., et al. (2014). Bladder cancer exosomes contain EDIL-3/Del1 and facilitate cancer progression. *J. Urol.* 192, 583–592. doi:10.1016/j.juro.2014.02.035
- Bennett, M. R., Sinha, S., and Owens, G. K. (2016). Vascular smooth muscle cells in atherosclerosis. *Circ. Res.* 118, 692–702. doi:10.1161/CIRCRESAHA.115.306361
- Bentley, K., Philippides, A., and Ravasz Regan, E. (2014). Do endothelial cells dream of eclectic shape? *Dev. Cell.* 29, 146–158. doi:10.1016/j.devcel.2014.03.019
- Cabrera, J. T. O., and Makino, A. (2022). Efferocytosis of vascular cells in cardiovascular disease. *Pharmacol. Ther.* 229, 107919. doi:10.1016/j.pharmthera.2021.107919
- Chang, H. C., and Guarente, L. (2014). SIRT1 and other sirtuins in metabolism. *Trends Endocrinol. Metab.* 25, 138–145. doi:10.1016/j.tem.2013.12.001
- Chattopadhyay, A., Kwartler, C. S., Kaw, K., Li, Y., Kaw, A., Chen, J., et al. (2021). Cholesterol-induced phenotypic modulation of smooth muscle cells to macrophage/fibroblast-like cells is driven by an unfolded protein response. *Arterioscler. Thromb. Vasc. Biol.* 41, 302–316. doi:10.1161/ATVBAHA.120.315164
- Chistiakov, D. A., Kashirskikh, D. A., Khotina, V. A., Grechko, A. V., and Orekhov, A. N. (2019). Immune-inflammatory responses in atherosclerosis: the role of myeloid cells. *J. Clin. Med.* 8, 1798. doi:10.3390/jcm8111798
- Choi, E. Y. (2009). Inhibition of leukocyte adhesion by developmental endothelial locus-1 (del-1). *Immune Netw.* 9, 153–157. doi:10.4110/in.2009.9.5.153

## Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This research was funded by the National Natural Science Foundation of China, Grant number 82370319; the National Natural Science Foundation of China, Grant number 82172060.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Choi, E. Y., Chavakis, E., Czabanka, M. A., Langer, H. F., Fraemohs, L., Economopoulou, M., et al. (2008). Del-1, an endogenous leukocyte-endothelial adhesion inhibitor, limits inflammatory cell recruitment. *Science* 322, 1101–1104. doi:10.1126/science.1165218

Choi, E. Y., Lim, J. H., Neuwirth, A., Economopoulou, M., Chatzigeorgiou, A., Chung, K. J., et al. (2015). Developmental endothelial locus-1 is a homeostatic factor in the central nervous system limiting neuroinflammation and demyelination. *Mol. Psychiatry* 20, 880–888. doi:10.1038/mp.2014.146

Ciucurel, E. C., and Sefton, M. V. (2014). Del-1 overexpression in endothelial cells increases vascular density in tissue-engineered implants containing endothelial cells and adipose-derived mesenchymal stromal cells. *Tissue Eng. Part A* 20, 1235–1252. doi:10.1089/ten.TEA.2013.0242

Clement, M., Chappell, J., Raffort, J., Lareyre, F., Vandestienne, M., Taylor, A. L., et al. (2019). Vascular smooth muscle cell plasticity and autophagy in dissecting aortic aneurysms. *Arterioscler. Thromb. Vasc. Biol.* 39, 1149–1159. doi:10.1161/ATVBAHA.118.311727

Cnop, M., Toivonen, S., Igoillo-Esteve, M., and Salpea, P. (2017). Endoplasmic reticulum stress and eIF2 $\alpha$  phosphorylation: the Achilles heel of pancreatic  $\beta$  cells. *Mol. Metab.* 6, 1024–1039. doi:10.1016/j.molmet.2017.06.001

Cobb, L. P., Siamakpour-Reihani, S., Zhang, D., Qin, X., Owzar, K., Zhou, C., et al. (2021). Obesity and altered angiogenic-related gene expression in endometrial cancer. *Gynecol. Oncol.* 163, 320–326. doi:10.1016/j.ygyno.2021.08.010

Consiglio, C. R., Cotugno, N., Sardh, F., Pou, C., Amodio, D., Rodriguez, L., et al. (2020). The immunology of multisystem inflammatory syndrome in children with COVID-19. *Cell.* 183, 968–981 e7. doi:10.1016/j.cell.2020.09.016

Cubillos-Ruiz, J. R., Bettigole, S. E., and Glimcher, L. H. (2017). Tumorigenic and immunosuppressive effects of endoplasmic reticulum stress in cancer. *Cell.* 168, 692–706. doi:10.1016/j.cell.2016.12.004

Cusimano, V., Pampinella, F., Giacomello, E., and Sorrentino, V. (2009). Assembly and dynamics of proteins of the longitudinal and junctional sarcoplasmic reticulum in skeletal muscle cells. *Proc. Natl. Acad. Sci. U. S. A.* 106, 4695–4700. doi:10.1073/pnas.0810243106

Dasgupta, S. K., Ie, A., Chavakis, T., Rumbaut, R. E., and Thiagarajan, P. (2012a). Developmental endothelial locus-1 (Del-1) mediates clearance of platelet microparticles by the endothelium. *Circulation* 125, 1664–1672. doi:10.1161/CIRCULATIONAHA.111.068833

Dasgupta, S. K., Ie, A., Chavakis, T., Rumbaut, R. E., and Thiagarajan, P. (2012b). Developmental endothelial locus-1 (Del-1) mediates clearance of platelet microparticles by the endothelium. *Circulation* 125, 1664–1672. doi:10.1161/CIRCULATIONAHA.111.068833

- de Meyer, G. R. Y., Zurek, M., Puylaert, P., and Martinet, W. (2024). Programmed death of macrophages in atherosclerosis: mechanisms and therapeutic targets. *Nat. Rev. Cardiol.* 21, 312–325. doi:10.1038/s41569-023-00957-0
- di Pietro, P., Vecchione, C., and Carrizzo, A. (2023). Targeting the CXCR4/DEL-1 axis to tackle atherosclerosis. *Int. J. Cardiol.* 380, 37–38. doi:10.1016/j.ijcard.2023.03.001
- Doddapattar, P., Dev, R., Ghatge, M., Patel, R. B., Jain, M., Dhanesha, N., et al. (2022). Myeloid cell PKM2 deletion enhances efferocytosis and reduces atherosclerosis. *Circ. Res.* 130, 1289–1305. doi:10.1161/CIRCRESAHA.121.320704
- Doran, A. C., Yurdagul, A., JR., and Tabas, I. (2020). Efferocytosis in health and disease. *Nat. Rev. Immunol.* 20, 254–267. doi:10.1038/s41577-019-0240-6
- Emini Veseli, B., Perrotta, P., de Meyer, G. R. A., Roth, L., van der Donckt, C., Martinet, W., et al. (2017). Animal models of atherosclerosis. *Eur. J. Pharmacol.* 816, 3–13. doi:10.1016/j.ejphar.2017.05.010
- Eskan, M. A., Jotwani, R., Abe, T., Chmelar, J., Lim, J. H., Liang, S., et al. (2012). The leukocyte integrin antagonist Del-1 inhibits IL-17-mediated inflammatory bone loss. *Nat. Immunol.* 13, 465–473. doi:10.1038/ni.2260
- Failor, T., Amponsah-Offeh, M., Neuwirth, A., Kourtzelis, I., Subramanian, P., Mirtschink, P., et al. (2022). Developmental endothelial locus-1 protects from hypertension-induced cardiovascular remodeling via immunomodulation. *J. Clin. Invest.* 132, e126155. doi:10.1172/JCI126155
- Furmanik, M., van Gorp, R., Whitehead, M., Ahmad, S., Bordoloi, J., Kapustin, A., et al. (2021). Endoplasmic reticulum stress mediates vascular smooth muscle cell calcification via increased release of Grp78 (Glucose-Regulated protein, 78 kDa)-Loaded extracellular vesicles. *Arterioscler. Thromb. Vasc. Biol.* 41, 898–914. doi:10.1161/ATVBAHA.120.315506
- Gaddis, D. E., Padgett, L. E., Wu, R., Mckimming, C., Romines, V., Taylor, A. M., et al. (2018). Apolipoprotein AI prevents regulatory to follicular helper T cell switching during atherosclerosis. *Nat. Commun.* 9, 1095. doi:10.1038/s41467-018-03493-5
- Galli, S. J., Borregaard, N., and Wynn, T. A. (2011). Phenotypic and functional plasticity of cells of innate immunity: macrophages, mast cells and neutrophils. *Nat. Immunol.* 12, 1035–1044. doi:10.1038/ni.2109
- Gerlach, B. D., Ampomah, P. B., Yurdagul, A., JR., Liu, C., Luring, M. C., Wang, X., et al. (2021). Efferocytosis induces macrophage proliferation to help resolve tissue injury. *Cell. Metab.* 33, 2445–2463.e8. doi:10.1016/j.cmet.2021.10.015
- Glinton, K. E., Ma, W., Lantz, C., Grigoryeva, L. S., Deberge, M., Liu, X., et al. (2022). Macrophage-produced VEGFC is induced by efferocytosis to ameliorate cardiac injury and inflammation. *J. Clin. Invest.* 132, e140685. doi:10.1172/JCI140685
- Grewal, N., Velders, B. J. J., Gittenberger-de Groot, A. C., Poelmann, R., Klautz, R. J. M., van Brakel, T. J., et al. (2021). A systematic histopathologic evaluation of type-A aortic dissections implies a uniform multiple-hit causation. *J. Cardiovasc. Dev. Dis.* 8, 12. doi:10.3390/jcdd8020012
- Grootaert, M. O. J., Finigan, A., Figg, N. L., Uryga, A. K., and Bennett, M. R. (2021). SIRT6 protects smooth muscle cells from senescence and reduces atherosclerosis. *Circ. Res.* 128, 474–491. doi:10.1161/CIRCRESAHA.120.318353
- Grossman, P. M., Mendelsohn, F., Henry, T. D., Hermiller, J. B., Litt, M., Saucedo, J. F., et al. (2007). Results from a phase II multicenter, double-blind placebo-controlled study of Del-1 (VLTS-589) for intermittent claudication in subjects with peripheral arterial disease. *Am. Heart J.* 153, 874–880. doi:10.1016/j.ahj.2007.01.038
- Hajishengallis, G., and Chavakis, T. (2019). DEL-1-Regulated immune plasticity and inflammatory disorders. *Trends Mol. Med.* 25, 444–459. doi:10.1016/j.molmed.2019.02.010
- Hajishengallis, G., and Chavakis, T. (2021). DEL-1: a potential therapeutic target in inflammatory and autoimmune disease? *Expert Rev. Clin. Immunol.* 17, 549–552. doi:10.1080/1744666X.2021.1915771
- Hajishengallis, G., Chavakis, T., Hajishengallis, E., and Lambris, J. D. (2015). Neutrophil homeostasis and inflammation: novel paradigms from studying periodontitis. *J. Leukoc. Biol.* 98, 539–548. doi:10.1189/jlb.3VMR1014-468R
- Hanayama, R., Tanaka, M., Miwa, K., and Nagata, S. (2004). Expression of developmental endothelial locus-1 in a subset of macrophages for engulfment of apoptotic cells. *J. Immunol.* 172, 3876–3882. doi:10.4049/jimmunol.172.6.3876
- Hidai, T. Z., Penta, K., Mikhail, A., Kawana, M., Quertermous, E. E., Aoka, Y., et al. (1998a). Cloning and characterization of developmental endothelial locus-1: an embryonic endothelial cell protein that binds the alphavbeta3 integrin receptor. *Genes. and Dev.* 12, 21–33. doi:10.1101/gad.12.1.21
- Hidai, C., Kawana, M., Kitano, H., and Kokubun, S. (2007). Discoidin domain of Del1 protein contributes to its deposition in the extracellular matrix. *Cell. Tissue Res.* 330, 83–95. doi:10.1007/s00441-007-0456-9
- Hidai, C., Zupancic, T., Penta, K., Mikhail, A., Kawana, M., Quertermous, E. E., et al. (1998b). Cloning and characterization of developmental endothelial locus-1: an embryonic endothelial cell protein that binds the alphavbeta3 integrin receptor. *Genes. Dev.* 12, 21–33. doi:10.1101/gad.12.1.21
- Ho, H. K., Jang, J. J., Kaji, S., Spector, G., Fong, A., Yang, P., et al. (2004). Developmental endothelial locus-1 (Del-1), a novel angiogenic protein: its role in ischemia. *Circulation* 109, 1314–1319. doi:10.1161/01.CIR.0000118465.36018.2D
- Hori, S., Nomura, T., and Sakaguchi, S. (2003). Control of regulatory T cell development by the transcription factor Foxp3. *Science* 299, 1057–1061. doi:10.1126/science.1079490
- Hsu, G. P., Mathy, J. A., Wang, Z., Xia, W., Sakamoto, G., Kundu, R., et al. (2008). Increased rate of hair regrowth in mice with constitutive overexpression of Del1. *J. Surg. Res.* 146, 73–80. doi:10.1016/j.jss.2007.02.024
- Hyun, Y. M., Seo, S. U., Choi, W. S., Kwon, H. J., Kim, D. Y., Jeong, S., et al. (2020). Endogenous DEL-1 restrains melanoma lung metastasis by limiting myeloid cell-associated lung inflammation. *Sci. Adv.* 6, eabc4882. doi:10.1126/sciadv.abc4882
- Jeong, J. H., Ojha, U., and Lee, Y. M. (2021). Pathological angiogenesis and inflammation in tissues. *Arch. Pharm. Res.* 44, 1–15. doi:10.1007/s12272-020-01287-2
- Jeong, D., Ban, S., Oh, S., Jin Lee, S., Yong Park, S., and Koh, Y. W. (2017). Prognostic significance of EDIL3 expression and correlation with mesenchymal phenotype and microvessel density in lung adenocarcinoma. *Sci. Rep.* 7, 8649. doi:10.1038/s41598-017-08851-9
- Jia, M., Fu, H., Jiang, X., Wang, L., Xu, J., Barnes, P. J., et al. (2023). DEL-1, as an anti-neutrophil transepithelial migration molecule, inhibits airway neutrophilic inflammation in asthma. *Allergy* 79, 1180–1194. doi:10.1111/all.15882
- Kakino, A., Fujita, Y., Nakano, A., Horiuchi, S., and Sawamura, T. (2016). Developmental endothelial locus-1 (Del-1) inhibits oxidized low-density lipoprotein activity by direct binding, and its overexpression attenuates atherosclerosis in mice. *Circ. J.* 80, 2541–2549. doi:10.1253/circj.CJ-16-0808
- Kasikara, C., Schilperoord, M., Gerlach, B., Xue, C., Wang, X., Zheng, Z., et al. (2021). Deficiency of macrophage PHACTR1 impairs efferocytosis and promotes atherosclerotic plaque necrosis. *J. Clin. Invest.* 131, e145275. doi:10.1172/JCI145275
- Kemmerer, M., Finkelnagel, F., Cavalcante, M. F., Abdalla, D. S., Müller, R., Brüne, B., et al. (2015). AMP-activated protein kinase interacts with the peroxisome proliferator-activated receptor delta to induce genes affecting fatty acid oxidation in human macrophages. *PLoS One* 10, e0130893. doi:10.1371/journal.pone.0130893
- Khader, S. A. (2012). Restraining IL-17: del-1 deals the blow. *Nat. Immunol.* 13, 433–435. doi:10.1038/ni.2290
- Kim, C. H. (2009). FOXP3 and its role in the immune system. *Adv. Exp. Med. Biol.* 665, 17–29. doi:10.1007/978-1-4419-1599-3\_2
- Kim, D. Y., Lee, S. H., Fu, Y., Jing, F., Kim, W. Y., Hong, S. B., et al. (2020). Del-1, an endogenous inhibitor of TGF- $\beta$  activation, attenuates fibrosis. *Front. Immunol.* 11, 68. doi:10.3389/fimmu.2020.00068
- Klingenberg, R., Gerdes, N., Badeau, R. M., Gisterà, A., Strothoff, D., Ketelhuth, D. F., et al. (2013). Depletion of FOXP3+ regulatory T cells promotes hypercholesterolemia and atherosclerosis. *J. Clin. Invest.* 123, 1323–1334. doi:10.1172/JCI63891
- Klotzsche-von Ameln, A., Cremer, S., Hoffmann, J., Schuster, P., Khedr, S., Korovina, I., et al. (2017). Endogenous developmental endothelial locus-1 limits ischaemia-related angiogenesis by blocking inflammation. *Thromb. Haemost.* 117, 1150–1163. doi:10.1160/TH16-05-0354
- Koupenova, M., Clancy, L., Corkrey, H. A., and Freedman, J. E. (2018). Circulating platelets as mediators of immunity, inflammation, and thrombosis. *Circ. Res.* 122, 337–351. doi:10.1161/CIRCRESAHA.117.310795
- Kourtzelis, I., Kotlabova, K., Lim, J. H., Mitroulis, I., Ferreira, A., Chen, L. S., et al. (2016). Developmental endothelial locus-1 modulates platelet-monocyte interactions and instant blood-mediated inflammatory reaction in islet transplantation. *Thromb. Haemost.* 115, 781–788. doi:10.1160/TH15-05-0429
- Kourtzelis, I., Li, X., Mitroulis, I., Grosser, D., Kajikawa, T., Wang, B., et al. (2019). DEL-1 promotes macrophage efferocytosis and clearance of inflammation. *Nat. Immunol.* 20, 40–49. doi:10.1038/s41590-018-0249-1
- Kruger-Genge, A., Blocki, A., Franke, R. P., and Jung, F. (2019). Vascular endothelial cell biology: an update. *Int. J. Mol. Sci.* 20, 4411. doi:10.3390/ijms20184411
- Kuang, S. Q., Medina-Martinez, O., Guo, D. C., Gong, L., Regalado, E. S., Reynolds, C. L., et al. (2016). FOXE3 mutations predispose to thoracic aortic aneurysms and dissections. *J. Clin. Invest.* 126, 948–961. doi:10.1172/JCI83778
- Kwon, C. H., Sun, J. L., Kim, M. J., Abd, A. M., Jeong, J. H., and Jung, T. W. (2020). Clinically confirmed DEL-1 as a myokine attenuates lipid-induced inflammation and insulin resistance in 3T3-L1 adipocytes via AMPK/HO-1 pathway. *Adipocyte* 9, 576–586. doi:10.1080/21623945.2020.1823140
- Lannan, K. L., Phipps, R. P., and White, R. J. (2014). Thrombosis, platelets, microparticles and PAH: more than a clot. *Drug Discov. Today* 19, 1230–1235. doi:10.1016/j.drudis.2014.04.001
- Lee, S. H., Kim, D. Y., Kang, Y. Y., Kim, H., Jang, J., Lee, M. N., et al. (2014). Developmental endothelial locus-1 inhibits MIF production through suppression of NF- $\kappa$ B in macrophages. *Int. J. Mol. Med.* 33, 919–924. doi:10.3892/ijmm.2014.1645
- Lee, W., Park, S. Y., Yoo, Y., Kim, S. Y., Kim, J. E., Kim, S. W., et al. (2018). Macrophagic stabilin-1 restored disruption of vascular integrity caused by sepsis. *Thromb. Haemost.* 118, 1776–1789. doi:10.1055/s-0038-1669477
- Leventis, P. A., and Grinstein, S. (2010). The distribution and function of phosphatidylerine in cellular membranes. *Annu. Rev. Biophys.* 39, 407–427. doi:10.1146/annurev.biophys.093008.131234

- Liang, S. C., Tan, X. Y., Luxenberg, D. P., Karim, R., Dunussi-Joannopoulos, K., Collins, M., et al. (2006). Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *J. Exp. Med.* 203, 2271–2279. doi:10.1084/jem.20061308
- Li, B., Xin, Z., Gao, S., Li, Y., Guo, S., Fu, Y., et al. (2023a). SIRT6-regulated macrophage efferocytosis epigenetically controls inflammation resolution of diabetic periodontitis. *Theranostics* 13, 231–249. doi:10.7150/thno.78878
- Libby, P. (2012). Inflammation in atherosclerosis. *Arterioscler. Thromb. Vasc. Biol.* 32, 2045–2051. doi:10.1161/ATVBAHA.108.179705
- Libby, P. (2021). The changing landscape of atherosclerosis. *Nature* 592, 524–533. doi:10.1038/s41586-021-03392-8
- Libby, P., Buring, J. E., Badimon, L., Hansson, G. K., Deanfield, J., Bittencourt, M. S., et al. (2019). Atherosclerosis. *Nat. Rev. Dis. Prim.* 5, 56. doi:10.1038/s41572-019-0106-z
- Li, H., Cao, Z., Wang, L., Liu, C., Lin, H., Tang, Y., et al. (2022a). Macrophage subsets and death are responsible for atherosclerotic plaque formation. *Front. Immunol.* 13, 843712. doi:10.3389/fimmu.2022.843712
- Li, M., Zhong, D., and Li, G. (2021a). Regulatory role of local tissue signal Del-1 in cancer and inflammation: a review. *Cell. Mol. Biol. Lett.* 26, 31. doi:10.1186/s11658-021-00274-9
- Li, R., Zeng, J., and Ren, T. (2022b). Expression of DEL-1 in alveolar epithelial cells prevents lipopolysaccharide-induced inflammation, oxidative stress, and eosinophil recruitment in acute lung injury. *Int. Immunopharmacol.* 110, 108961. doi:10.1016/j.intimp.2022.108961
- Li, T., Yang, Y., Qi, H., Cui, W., Zhang, L., Fu, X., et al. (2023b). CRISPR/Cas9 therapeutics: progress and prospects. *Signal Transduct. Target Ther.* 8, 36. doi:10.1038/s41392-023-01309-7
- Liu, G., Chen, H., Liu, H., Zhang, W., and Zhou, J. (2021). Emerging roles of SIRT6 in human diseases and its modulators. *Med. Res. Rev.* 41, 1089–1137. doi:10.1002/med.21753
- Liu, S., Wu, J., Stolarz, A., Zhang, H., Boerma, M., Byrum, S. D., et al. (2023). PCSK9 attenuates efferocytosis in endothelial cells and promotes vascular aging. *Theranostics* 13, 2914–2929. doi:10.7150/thno.83914
- Li, W., Feng, W., Su, X., Luo, D., Li, Z., Zhou, Y., et al. (2022c). SIRT6 protects vascular smooth muscle cells from osteogenic transdifferentiation via Runx2 in chronic kidney disease. *J. Clin. Investig.* 132, e150051. doi:10.1172/JCI150051
- Li, X., Colamatteo, A., Kalafati, L., Kajikawa, T., Wang, H., Lim, J. H., et al. (2020). The DEL-1/ $\beta$ 3 integrin axis promotes regulatory T cell responses during inflammation resolution. *J. Clin. Investig.* 130, 6261–6277. doi:10.1172/JCI137530
- Li, Y., Li, Y., Li, Y., Yang, Z., Geng, H., Liu, C., et al. (2021b). Inhibition of endoplasmic reticulum stress mediates the ameliorative effect of apelin on vascular calcification. *J. Mol. Cell. Cardiol.* 152, 17–28. doi:10.1016/j.yjmcc.2020.11.017
- Luo, H., Lan, C., Fan, C., Gong, X., Chen, C., Yu, C., et al. (2022). Down-regulation of AMPK/PPAR $\delta$  signalling promotes endoplasmic reticulum stress-induced endothelial dysfunction in adult rat offspring exposed to maternal diabetes. *Cardiovasc Res.* 118, 2304–2316. doi:10.1093/cvr/cvab280
- Lu, Y., Zhou, M., Peng, J., Li, F., Dai, J., Wan, C., et al. (2023). DEL-1 suppression attenuates atherosclerosis by modulating macrophagic GSK-3 $\beta$ /CEBP- $\beta$  signaling pathway. *Int. J. Cardiol.* 376, 115–124. doi:10.1016/j.ijcard.2023.01.068
- Maekawa, T., Hosur, K., Abe, T., Kantarci, A., Ziogas, A., Wang, B., et al. (2015). Antagonistic effects of IL-17 and D-resolvins on endothelial Del-1 expression through a GSK-3 $\beta$ -C/EBP $\beta$  pathway. *Nat. Commun.* 6, 8272. doi:10.1038/ncomms9272
- Maekawa, T., Tamura, H., Domon, H., Hiyoshi, T., Isono, T., Yonezawa, D., et al. (2020). Erythromycin inhibits neutrophilic inflammation and mucosal disease by upregulating DEL-1. *JCI Insight* 5, e136706. doi:10.1172/jci.insight.136706
- Marchini, T., Hansen, S., and Wolf, D. (2021). ApoB-specific CD4(+) T cells in mouse and human atherosclerosis. *Cells* 10, 446. doi:10.3390/cells10020446
- Marciniak, S. J., Chambers, J. E., and Ron, D. (2022). Pharmacological targeting of endoplasmic reticulum stress in disease. *Nat. Rev. Drug Discov.* 21, 115–140. doi:10.1038/s41573-021-00320-3
- Marciniak, S. J., and Ron, D. (2006). Endoplasmic reticulum stress signaling in disease. *Physiol. Rev.* 86, 1133–1149. doi:10.1152/physrev.00015.2006
- Margolis, E. A., Friend, N. E., Rolle, M. W., Alsberg, E., and Putnam, A. J. (2023). Manufacturing the multiscale vascular hierarchy: progress toward solving the grand challenge of tissue engineering. *Trends Biotechnol.* 41, 1400–1416. doi:10.1016/j.tibtech.2023.04.003
- Margraf, A., and Zarbock, A. (2019). Platelets in inflammation and resolution. *J. Immunol.* 203, 2357–2367. doi:10.4049/jimmunol.1900899
- Martin, D., Li, Y., Yang, J., Wang, G., Margariti, A., Jiang, Z., et al. (2014). Unspliced X-box-binding protein 1 (XBP1) protects endothelial cells from oxidative stress through interaction with histone deacetylase 3. *J. Biol. Chem.* 289, 30625–30634. doi:10.1074/jbc.M114.571984
- Matzinger, P., and Kamala, T. (2011). Tissue-based class control: the other side of tolerance. *Nat. Rev. Immunol.* 11, 221–230. doi:10.1038/nri2940
- May, A. E., Langer, H., Seizer, P., Bigalke, B., Lindemann, S., and Gawaz, M. (2007). Platelet-leukocyte interactions in inflammation and atherothrombosis. *Semin. Thromb. Hemost.* 33, 123–127. doi:10.1055/s-2007-969023
- Mccooy, M. G., Nascimento, D. W., Velepparambil, M., Murtazina, R., Gao, D., Tkachenko, S., et al. (2021). Endothelial TLR2 promotes proangiogenic immune cell recruitment and tumor angiogenesis. *Sci. Signal* 14, eabc5371. doi:10.1126/scisignal.abc5371
- Meng, X., Yang, J., Dong, M., Zhang, K., Tu, E., Gao, Q., et al. (2016). Regulatory T cells in cardiovascular diseases. *Nat. Rev. Cardiol.* 13, 167–179. doi:10.1038/nrcardio.2015.169
- Mezu-Ndubuisi, O. J., and Maheshwari, A. (2021). The role of integrins in inflammation and angiogenesis. *Pediatr. Res.* 89, 1619–1626. doi:10.1038/s41390-020-01177-9
- Mills, K. H. G. (2023). IL-17 and IL-17-producing cells in protection versus pathology. *Nat. Rev. Immunol.* 23, 38–54. doi:10.1038/s41577-022-00746-9
- Mitroulis, I., Kang, Y. Y., Gahmberg, C. G., Siegert, G., Hajishengallis, G., Chavakis, T., et al. (2014). Developmental endothelial locus-1 attenuates complement-dependent phagocytosis through inhibition of Mac-1-integrin. *Thromb. Haemost.* 111, 1004–1006. doi:10.1160/TH13-09-0794
- Morioka, S., Maueröder, C., and Ravichandran, K. S. (2019). Living on the edge: efferocytosis at the interface of homeostasis and pathology. *Immunity* 50, 1149–1162. doi:10.1016/j.immuni.2019.04.018
- Mrugacz, M., Bryl, A., Falkowski, M., and Zorena, K. (2021). Integrins: an important link between angiogenesis, inflammation and eye diseases. *Cells* 10, 1703. doi:10.3390/cells10071703
- Nagata, S. (2018). Apoptosis and clearance of apoptotic cells. *Annu. Rev. Immunol.* 36, 489–517. doi:10.1146/annurev-immunol-042617-053010
- Nishida, N., Aoki, H., Ohno-Urabe, S., Nishihara, M., Furusho, A., Hirakata, S., et al. (2020). High salt intake worsens aortic dissection in mice: involvement of IL (Interleukin)-17A-Dependent ECM (extracellular matrix) metabolism. *Arterioscler. Thromb. Vasc. Biol.* 40, 189–205. doi:10.1161/ATVBAHA.119.313336
- Niu, X., Chang, W., Liu, R., Hou, R., Li, J., Wang, C., et al. (2016). mRNA and protein expression of the angiogenesis-related genes EDIL3, AMOT and ECM1 in mesenchymal stem cells in psoriatic dermis. *Clin. Exp. Dermatol.* 41, 533–540. doi:10.1111/ced.12783
- Niu, X., Han, Q., Liu, Y., Li, J., Hou, R., Li, J., et al. (2020). Psoriasis-associated angiogenesis is mediated by EDIL3. *Microvasc. Res.* 132, 104056. doi:10.1016/j.mvr.2020.104056
- Niu, X., Han, Q., Li, X., Li, J., Liu, Y., Li, Y., et al. (2022a). Del-1 in psoriasis induced the expression of av $\beta$ 3 and  $\alpha$ 5 $\beta$ 1 in endothelial cells. *Curr. Mol. Med.* 22, 442–448. doi:10.2174/1566524021666210729112700
- Niu, X., Han, Q., Li, X., Li, J., Liu, Y., Li, Y., et al. (2022b). EDIL3 influenced the av $\beta$ 3-FAK/MEK/ERK axis of endothelial cells in psoriasis. *J. Cell. Mol. Med.* 26, 5202–5212. doi:10.1111/jcmm.17544
- Niu, X., Li, X., Feng, Z., Han, Q., Li, J., Liu, Y., et al. (2023). EDIL3 and VEGF synergistically affect angiogenesis in endothelial cells. *Clin. Cosmet. Investig. Dermatol* 16, 1269–1277. doi:10.2147/CCID.S411253
- Oakes, S. A., and Papa, F. R. (2015). The role of endoplasmic reticulum stress in human pathology. *Annu. Rev. Pathol.* 10, 173–194. doi:10.1146/annurev-pathol-012513-104649
- Oshima, Y., Tanimoto, T., Yuji, K., and Tojo, A. (2017). Association between aortic dissection and systemic exposure of vascular endothelial growth factor pathway inhibitors in the Japanese adverse drug event report database. *Circulation* 135, 815–817. doi:10.1161/CIRCULATIONAHA.116.025144
- Oyler-Yaniv, A., Oyler-Yaniv, J., Whitlock, B. M., Liu, Z., Germain, R. N., Huse, M., et al. (2017). A tunable diffusion-consumption mechanism of cytokine propagation enables plasticity in cell-to-cell communication in the immune system. *Immunity* 46, 609–620. doi:10.1016/j.immuni.2017.03.011
- Panizo, S., Cardus, A., Encinas, M., Parisi, E., Valcheva, P., LóPEZ-Ongil, S., et al. (2009). RANKL increases vascular smooth muscle cell calcification through a RANK-BMP4-dependent pathway. *Circ. Res.* 104, 1041–1048. doi:10.1161/CIRCRESAHA.108.189001
- Park, T. J., Park, S. Y., Cho, W., Oh, H., Lee, H. J., Abd el-Aty, A. M., et al. (2022). Developmental endothelial locus-1 attenuates palmitate-induced apoptosis in tenocytes through the AMPK/autophagy-mediated suppression of inflammation and endoplasmic reticulum stress. *Bone Jt. Res.* 11, 854–861. doi:10.1302/2046-3758.1112.BJR-2022-0077.R2
- Pircher, J., Engelmann, B., Massberg, S., and Schulz, C. (2019). Platelet-neutrophil crosstalk in atherothrombosis. *Thromb. Haemost.* 119, 1274–1282. doi:10.1055/s-0039-1692983

- Pober, J. S., and Sessa, W. C. (2007). Evolving functions of endothelial cells in inflammation. *Nat. Rev. Immunol.* 7, 803–815. doi:10.1038/nri2171
- Ponomareva, A. A., Nevzorova, T. A., Mordakhanova, E. R., Andrianova, I. A., Rauova, L., Litvinov, R. I., et al. (2017). Intracellular origin and ultrastructure of platelet-derived microparticles. *J. Thromb. Haemost.* 15, 1655–1667. doi:10.1111/jth.13745
- Rader, D. J., and Puré, E. (2005). Lipoproteins, macrophage function, and atherosclerosis: beyond the foam cell? *Cell. Metab.* 1, 223–230. doi:10.1016/j.cmet.2005.03.005
- Ramasamy, S. K., Kusumbe, A. P., and Adams, R. H. (2015). Regulation of tissue morphogenesis by endothelial cell-derived signals. *Trends Cell. Biol.* 25, 148–157. doi:10.1016/j.tcb.2014.11.007
- Ren, J., Bi, Y., Sowers, J. R., Hetz, C., and Zhang, Y. (2021). Endoplasmic reticulum stress and unfolded protein response in cardiovascular diseases. *Nat. Rev. Cardiol.* 18, 499–521. doi:10.1038/s41569-021-00511-w
- Romanidou, G., Konstantinidis, T. G., Natsi, A.-M., Kantartzi, K., Panopoulou, M., Kontomanolis, E., et al. (2023). Decreased levels of soluble developmental endothelial locus-1 are associated with thrombotic microangiopathy in pregnancy. *Int. J. Mol. Sci.* 24, 11762. doi:10.3390/ijms241411762
- Ron, D., and Walter, P. (2007). Signal integration in the endoplasmic reticulum unfolded protein response. *Nat. Rev. Mol. Cell. Biol.* 8, 519–529. doi:10.1038/nrm2199
- Rose, D. M., Alon, R., and Ginsberg, M. H. (2007). Integrin modulation and signaling in leukocyte adhesion and migration. *Immunol. Rev.* 218, 126–134. doi:10.1111/j.1600-065X.2007.00536.x
- Roy, P., Orecchioni, M., and Ley, K. (2022). How the immune system shapes atherosclerosis: roles of innate and adaptive immunity. *Nat. Rev. Immunol.* 22, 251–265. doi:10.1038/s41577-021-00584-1
- Ruoslahti, E. (1996). RGD and other recognition sequences for integrins. *Annu. Rev. Cell. Dev. Biol.* 12, 697–715. doi:10.1146/annurev.cellbio.12.1.697
- Saigusa, R., Winkels, H., and Ley, K. (2020). T cell subsets and functions in atherosclerosis. *Nat. Rev. Cardiol.* 17, 387–401. doi:10.1038/s41569-020-0352-5
- Schmitz, T., Freuer, D., Linseisen, J., and Meisinger, C. (2023). Associations between serum cholesterol and immunophenotypical characteristics of circulatory B cells and Tregs. *J. Lipid Res.* 64, 100399. doi:10.1016/j.jlr.2023.100399
- Schurpf, T., Chen, Q., Liu, J. H., Wang, R., Springer, T. A., and Wang, J. H. (2012). The RGD finger of Del-1 is a unique structural feature critical for integrin binding. *FASEB J.* 26, 3412–3420. doi:10.1096/fj.11-202036
- Schwarz, D. S., and Blower, M. D. (2016). The endoplasmic reticulum: structure, function and response to cellular signaling. *Cell. Mol. Life Sci.* 73, 79–94. doi:10.1007/s00118-015-2052-6
- Segawa, K., and Nagata, S. (2015). An apoptotic 'eat me' signal: phosphatidylserine exposure. *Trends Cell. Biol.* 25, 639–650. doi:10.1016/j.tcb.2015.08.003
- Shao, Y., Saredy, J., Yang, W. Y., Sun, Y., Lu, Y., Saaoud, F., et al. (2020). Vascular endothelial cells and innate immunity. *Arterioscler. Thromb. Vasc. Biol.* 40, e138–e152. doi:10.1161/ATVBAHA.120.314330
- Shin, J., Maekawa, T., Abe, T., Hajishengallis, E., Hosur, K., Pyram, K., et al. (2015). DEL-1 restrains osteoclastogenesis and inhibits inflammatory bone loss in nonhuman primates. *Sci. Transl. Med.* 7, 307ra155. doi:10.1126/scitranslmed.aac5380
- Shiny, A., Regin, B., Mohan, V., and Balasubramanyam, M. (2016). Coordinated augmentation of NFAT and NOD signaling mediates proliferative VSMC phenotype switch under hyperinsulinemia. *Atherosclerosis* 246, 257–266. doi:10.1016/j.atherosclerosis.2016.01.006
- Singh, N. K., Kundumani-Sridharan, V., Kumar, S., Verma, S. K., Kotla, S., Mukai, H., et al. (2012). Protein kinase N1 is a novel substrate of NFATc1-mediated cyclin D1-CDK6 activity and modulates vascular smooth muscle cell division and migration leading to inward blood vessel wall remodeling. *J. Biol. Chem.* 287, 36291–36304. doi:10.1074/jbc.M112.361220
- Sorrentino, V. (2011). Sarcoplasmic reticulum: structural determinants and protein dynamics. *Int. J. Biochem. Cell. Biol.* 43, 1075–1078. doi:10.1016/j.biocel.2011.04.004
- Sun, J. L., Park, J., Lee, T., Jeong, J. H., and Jung, T. W. (2020). DEL-1 ameliorates high-fat diet-induced insulin resistance in mouse skeletal muscle through SIRT1/SERCA2-mediated ER stress suppression. *Biochem. Pharmacol.* 171, 113730. doi:10.1016/j.bcp.2019.113730
- Tian, S., Lin, G., Piao, L., and Liu, X. (2022). Del-1 enhances therapeutic efficacy of bacterial cancer immunotherapy by blocking recruitment of tumor-infiltrating neutrophils. *Clin. Transl. Oncol.* 24, 244–253. doi:10.1007/s12094-021-02679-6
- Twarock, S., Bagheri, S., Bagheri, S., and Hohlfeld, T. (2016). Platelet-vessel wall interactions and drug effects. *Pharmacol. Ther.* 167, 74–84. doi:10.1016/j.pharmthera.2016.07.008
- van Hove, I., Hu, T. T., Beets, K., van Bergen, T., Etienne, I., Stitt, A. W., et al. (2021). Targeting RGD-binding integrins as an integrative therapy for diabetic retinopathy and neovascular age-related macular degeneration. *Prog. Retin Eye Res.* 85, 100966. doi:10.1016/j.preteyeres.2021.100966
- Wang, G. H., Ma, K. L., Zhang, Y., Hu, Z. B., Liu, L., Lu, J., et al. (2019a). Platelet microparticles contribute to aortic vascular endothelial injury in diabetes via the mTORC1 pathway. *Acta Pharmacol. Sin.* 40, 468–476. doi:10.1038/s41401-018-0186-4
- Wang, S. W., Gao, C., Zheng, Y. M., Yi, L., Lu, J. C., Huang, X. Y., et al. (2022b). Current applications and future perspective of CRISPR/Cas9 gene editing in cancer. *Mol. Cancer* 21, 57. doi:10.1186/s12943-022-01518-8
- Wang, H., Li, X., Kajikawa, T., Shin, J., Lim, J. H., Kourtzelis, I., et al. (2021). Stromal cell-derived DEL-1 inhibits Tfh cell activation and inflammatory arthritis. *J. Clin. Invest.* 131, e150578. doi:10.1172/JCI150578
- Wang, L., Liu, J., Wang, W., Qi, X., Wang, Y., Tian, B., et al. (2019b). Targeting IL-17 attenuates hypoxia-induced pulmonary hypertension through downregulation of  $\beta$ -catenin. *Thorax* 74, 564–578. doi:10.1136/thoraxjnl-2018-211846
- Wang, S., Wang, B., Shi, Y., MÖLLER, T., Stegmeyer, R. I., Strlic, B., et al. (2022a). Mechanosensation by endothelial PIEZO1 is required for leukocyte diapedesis. *Blood* 140, 171–183. doi:10.1182/blood.2021014614
- Wiseman, R. L., Mesgarzadeh, J. S., and Hendershot, L. M. (2022). Reshaping endoplasmic reticulum quality control through the unfolded protein response. *Mol. Cell.* 82, 1477–1491. doi:10.1016/j.molcel.2022.03.025
- Wolf, D., Gerhardt, T., Winkels, H., Michel, N. A., Pramod, A. B., Ghosheh, Y., et al. (2020). Pathogenic autoimmunity in atherosclerosis evolves from initially protective apolipoprotein B(100)-reactive CD4(+) T-regulatory cells. *Circulation* 142, 1279–1293. doi:10.1161/CIRCULATIONAHA.119.042863
- Wu, J., and Kaufman, R. J. (2006). From acute ER stress to physiological roles of the Unfolded Protein Response. *Cell. Death Differ.* 13, 374–384. doi:10.1038/sj.cdd.4401840
- Xia, H., Chen, J., Shi, M., Gao, H., Sekar, K., Seshachalam, V. P., et al. (2015). EDIL3 is a novel regulator of epithelial-mesenchymal transition controlling early recurrence of hepatocellular carcinoma. *J. Hepatol.* 63, 863–873. doi:10.1016/j.jhep.2015.05.005
- Xu, S., Ilyas, I., Little, P. J., Li, H., Kamato, D., Zheng, X., et al. (2021). Endothelial dysfunction in atherosclerotic cardiovascular diseases and beyond: from mechanism to pharmacotherapies. *Pharmacol. Rev.* 73, 924–967. doi:10.1124/pharmrev.120.000096
- Yang, L., Dai, R., Wu, H., Cai, Z., Xie, N., Zhang, X., et al. (2022). Unspliced XBP1 counteracts  $\beta$ -catenin to inhibit vascular calcification. *Circ. Res.* 130, 213–229. doi:10.1161/CIRCRESAHA.121.319745
- Yang, N., Baban, B., Isales, C. M., and Shi, X.-M. (2015a). Crosstalk between bone marrow-derived mesenchymal stem cells and regulatory T cells through a glucocorticoid-induced leucine zipper/developmental endothelial locus-1-dependent mechanism. *FASEB J.* 29, 3954–3963. doi:10.1096/fj.15-273664
- Yang, N., Baban, B., Isales, C. M., and Shi, X. M. (2015b). Crosstalk between bone marrow-derived mesenchymal stem cells and regulatory T cells through a glucocorticoid-induced leucine zipper/developmental endothelial locus-1-dependent mechanism. *Faseb J.* 29, 3954–3963. doi:10.1096/fj.15-273664
- Yan, S., Chen, L., Zhao, Q., Liu, Y. N., Hou, R., Yu, J., et al. (2018). Developmental endothelial locus-1 (Del-1) antagonizes Interleukin-17-mediated allergic asthma. *Immunol. Cell. Biol.* 96, 526–535. doi:10.1111/imcb.12023
- Yap, K. N., Yamada, K., Zikeli, S., Kiaris, H., and Hood, W. R. (2021). Evaluating endoplasmic reticulum stress and unfolded protein response through the lens of ecology and evolution. *Biol. Rev. Camb Philos. Soc.* 96, 541–556. doi:10.1111/brv.12667
- Yatim, N., Cullen, S., and Albert, M. L. (2017). Dying cells actively regulate adaptive immune responses. *Nat. Rev. Immunol.* 17, 262–275. doi:10.1038/nri.2017.9
- Zhang, K., Shen, X., Wu, J., Sakaki, K., Saunders, T., Rutkowski, D. T., et al. (2006). Endoplasmic reticulum stress activates cleavage of CREBH to induce a systemic inflammatory response. *Cell.* 124, 587–599. doi:10.1016/j.cell.2005.11.040
- Zhang, L., Cheng, H., Yue, Y., Li, S., Zhang, D., and He, R. (2018). H19 knockdown suppresses proliferation and induces apoptosis by regulating miR-148b/WNT/ $\beta$ -catenin in ox-LDL-stimulated vascular smooth muscle cells. *J. Biomed. Sci.* 25, 11. doi:10.1186/s12929-018-0418-4
- Zhang, L., Liu, M., Liu, W., Hu, C., Li, H., Deng, J., et al. (2021). Th17/IL-17 induces endothelial cell senescence via activation of NF- $\kappa$ B/p53/Rb signaling pathway. *Lab. Invest.* 101, 1418–1426. doi:10.1038/s41374-021-00629-y
- Zhang, Z., Jiang, Y., Zhou, Z., Huang, J., Chen, S., Zhou, W., et al. (2019). Scavenger receptor A1 attenuates aortic dissection via promoting efferocytosis in macrophages. *Biochem. Pharmacol.* 168, 392–403. doi:10.1016/j.bcp.2019.07.027
- Zhao, M., Zheng, Z., Li, C., Wan, J., and Wang, M. (2022). Developmental endothelial locus-1 in cardiovascular and metabolic diseases: a promising biomarker and therapeutic target. *Fountiers Immunol.* 13, 1053175. doi:10.3389/fimmu.2022.1053175
- Zhao, M., Zheng, Z., Yin, Z., Zhang, J., Peng, S., Liu, J., et al. (2023). DEL-1 deficiency aggravates pressure overload-induced heart failure by promoting neutrophil infiltration

and neutrophil extracellular traps formation. *Biochem. Pharmacol.* 218, 115912. doi:10.1016/j.bcp.2023.115912

Zhao, Y., Li, X., Cai, M. Y., Ma, K., Yang, J., Zhou, J., et al. (2013). XBP-1 $\alpha$  suppresses autophagy by promoting the degradation of FoxO1 in cancer cells. *Cell. Res.* 23, 491–507. doi:10.1038/cr.2013.2

Zhao, Y., Zhang, J., Zheng, Y., Zhang, Y., Zhang, X. J., Wang, H., et al. (2021). NAD<sup>+</sup> improves cognitive function and reduces neuroinflammation by ameliorating mitochondrial damage and decreasing ROS production in chronic cerebral hypoperfusion models through Sirt1/PGC-1 $\alpha$  pathway. *J. Neuroinflammation* 18, 207. doi:10.1186/s12974-021-02250-8

Zhong, J., Eliceiri, B., Stupack, D., Penta, K., Sakamoto, G., Quertermous, T., et al. (2003). Neovascularization of ischemic tissues by gene delivery of the

extracellular matrix protein Del-1. *J. Clin. Investig.* 112, 30–41. doi:10.1172/JCI17034

Zhou, A. X., and Tabas, I. (2013). The UPR in atherosclerosis. *Semin. Immunopathol.* 35, 321–332. doi:10.1007/s00281-013-0372-x

Zou, L., Barnett, B., Safah, H., Larussa, V. F., Evdemon-Hogan, M., Mottram, P., et al. (2004). Bone marrow is a reservoir for CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cells that traffic through CXCL12/CXCR4 signals. *Cancer Res.* 64, 8451–8455. doi:10.1158/0008-5472.CAN-04-1987

Zou, X., Qiao, H., Jiang, X., Dong, X., Jiang, H., and Sun, X. (2009). Downregulation of developmentally regulated endothelial cell locus-1 inhibits the growth of colon cancer. *J. Biomed. Sci.* 16, 33. doi:10.1186/1423-0127-16-33