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# Editorial: Transcription factors and arrhythmogenesis

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## Editorial on the Research Topic Transcription factors and arrhythmogenesis

The present Research Topic, entitled “Transcription Factors and Arrhythmogenesis” aims at highlighting the functional role of gene regulatory networks of cardiac enriched transcription factors and their target genes in the pathophysiology of cardiac arrhythmias. Generally speaking, transcription factors bind to the promoter regions of target genes and regulate their expression (Ciccarelli et al., 2011; Larsen et al., 2020; Henley and Koehler, 2021; Tang et al., 2021; de Mattos et al., 2022; Xiao et al., 2022; Jiang and Qian, 2023; Yu et al., 2023; Zhao et al., 2023). Transcription factors can regulate arrhythmogenesis by modulating the expression of genes involved in the cardiac conduction system and/or myocardial structures, as well as through indirect effects on ionic currents, calcium homeostasis, inflammation, oxidative stress, and cardiac remodeling (Liang et al., 2015; Lu et al., 2015; Crespo-Garcia et al., 2022).

Tom McDonald and collaborators elegantly describe the phenotypic variability in iPSC-induced cardiomyocytes and cardiac fibroblasts carrying diverse mutations in the LMNA gene, encoding lamin A/C (Yang et al.). These mutations are known to lead to familial arrhythmogenic cardiomyopathy with a high penetrance; however, there is a phenotypic variability in terms of disease onset, severity, and rate of progression. Recent evidence has shown that induced pluripotent stem cell (iPSC) represent a reliable strategy to create disease-in-a-dish models (Varzideh et al., 2022). Thus, the authors generated seven patient-specific iPSC lines with different LMNA mutations and successfully differentiated them in cardiomyocytes and cardiac fibroblasts. They observed electrophysiological aberrations, sarcomere disarray, and increased apoptosis in cardiomyocytes, and detected several irregularities of the nuclear membrane morphology in cardiac fibroblasts. Intriguingly, co-culture assays of cardiomyocytes and fibroblasts carrying LMNA mutations show exaggerated electrical disturbances (Yang et al.), suggesting that conduction properties of cardiomyocytes might be adversely

affected after coculture with fibroblasts in *LMNA* mutation-associated dilated cardiomyopathy. Additionally, patient- or mutation-specific iPSC may serve as an ideal platform for predicting and testing new effective therapeutics.

The *ZFHX3* gene is one of the most studied genes associated with atrial fibrillation (AFib) (Benjamin et al., 2009; Tomomori et al., 2018; Cheng et al., 2019). In a single-center, retrospective, observational cohort study conducted in 1,782 patients who underwent AFib catheter ablation, Hwang et al. found an association between genetic variants of *ZFHX3* and extra-pulmonary vein (PV) triggers. Extra-PV triggers are main causes in AFib recurrence post catheter ablation. *ZFHX3* knockdown in HL-1 atrial myocytes was found to induce electrical remodeling and increase metabolic stress (Cheng et al., 2019; Lkhagva et al., 2021). These pieces of evidence might explain how extra-PV triggers occur in *ZFHX3* genetic variants after ablation.

Qin et al. examined the effects of Gluconolactone (D-glucono-1,5-lactone, GDL) in cardiac ischemia/reperfusion (I/R) injury both *in vivo* (in mice) and *in vitro* (in neonatal cardiomyocytes). GDL is a food additive (E-number: E575) present in several dietary products including bread, cheese, wine, yogurt, and tofu (Romo-Rodriguez et al., 2015). The authors observed that GDL attenuated I/R injury and reduced reperfusion-induced arrhythmias and oxidative stress. They also provide a mechanism explaining these findings, showing that GDL acts as a potent activator of PKC $\epsilon$ -mediated ERK signaling (Qin et al.). PKC $\epsilon$  may provide cardioprotection in I/R injury *via* activating mitochondrial ALDH2 to scavenge toxic aldehyde-lipid peroxidation products, opening of mitochondrial K<sub>ATP</sub> channels to decrease ROS production and calcium overload, and inhibiting the activation of L-type calcium channels (Inagaki et al., 2006; Ferreira et al., 2012). Moreover, although not directly investigated in the Qin's paper, PKC $\epsilon$  is a known activator of the transcription factors c-Jun and STAT3 (signal transducer and activator of transcription 3) (Batarseh et al., 2010; Wang et al., 2018). Accordingly, GDL was suggested to have cardioprotective potential against I/R injury.

In conclusion, the research area investigating how transcription factors are linked to the pathophysiology of cardiac arrhythmias is

quite active, and further studies are warranted to identify novel molecular mechanisms and eventually therapeutic targets to tackle this major issue in cardiovascular medicine.

## Author contributions

GS wrote the first draft; All the authors edited the manuscript and approved its final version.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## References

- Batarseh, A., Li, J., and Papadopoulos, V. (2010). Protein kinase C epsilon regulation of translocator protein (18 kDa) Tspo gene expression is mediated through a MAPK pathway targeting STAT3 and c-Jun transcription factors. *Biochemistry* 49 (23), 4766–4778. doi:10.1021/bi100020e
- Benjamin, E. J., Rice, K. M., Arking, D. E., Pfeufer, A., van Noord, C., Smith, A. V., et al. (2009). Variants in *ZFHX3* are associated with atrial fibrillation in individuals of European ancestry. *Nat. Genet.* 41 (8), 879–881. doi:10.1038/ng.416
- Cheng, W. L., Kao, Y. H., Chao, T. F., Lin, Y. K., Chen, S. A., and Chen, Y. J. (2019). MicroRNA-133 suppresses *ZFHX3*-dependent atrial remodeling and arrhythmia. *Acta Physiol. (Oxf)*. 227 (3), e13322. doi:10.1111/apha.13322
- Ciccarelli, M., Sorriento, D., Cipolletta, E., Santulli, G., Fusco, A., Zhou, R. H., et al. (2011). Impaired neoangiogenesis in  $\beta_2$ -adrenoceptor gene-deficient mice: Restoration by intravascular human  $\beta_2$ -adrenoceptor gene transfer and role of NF $\kappa$ B and CREB transcription factors. *Br. J. Pharmacol.* 162 (3), 712–721. doi:10.1111/j.1476-5381.2010.01078.x
- Crespo-Garcia, T., Camara-Checa, A., Dago, M., Rubio-Alarcon, M., Rapun, J., Tamargo, J., et al. (2022). Regulation of cardiac ion channels by transcription factors: Looking for new opportunities of druggable targets for the treatment of arrhythmias. *Biochem. Pharmacol.* 204, 115206. doi:10.1016/j.bcp.2022.115206
- de Mattos, K., Viger, R. S., and Tremblay, J. J. (2022). Transcription factors in the regulation of leydig cell gene expression and function. *Front. Endocrinol. (Lausanne)*. 13, 881309. doi:10.3389/fendo.2022.881309
- Ferreira, J. C., Mochly-Rosen, D., and Boutjdir, M. (2012). Regulation of cardiac excitability by protein kinase C isozymes. *Front. Biosci. Sch. Ed.* 4 (2), 532–546. doi:10.2741/283
- Henley, M. J., and Koehler, A. N. (2021). Advances in targeting 'undruggable' transcription factors with small molecules. *Nat. Rev. Drug Discov.* 20 (9), 669–688. doi:10.1038/s41573-021-00199-0
- Inagaki, K., Churchill, E., and Mochly-Rosen, D. (2006). Epsilon protein kinase C as a potential therapeutic target for the ischemic heart. *Cardiovasc. Res.* 70 (2), 222–230. doi:10.1016/j.cardiores.2006.02.015
- Jiang, Y., and Qian, H. Y. (2023). Transcription factors: Key regulatory targets of vascular smooth muscle cell in atherosclerosis. *Mol. Med.* 29 (1), 2. doi:10.1186/s10020-022-00586-2
- Larsen, A. I., Valborgland, T., Ogne, C., Lindal, S., Halvorsen, B., Munk, P. S., et al. (2020). Plasma tumour necrosis factor correlates with mRNA expression of tumour necrosis factor and mitochondrial transcription factors in skeletal muscle in patients with chronic heart failure treated with cardiac resynchronization therapy: Potential role in myopathy. *Eur. J. Prev. Cardiol.* 27 (19), 2362–2366. doi:10.1177/2047487319855796
- Liang, X., Zhang, Q., Cattaneo, P., Zhuang, S., Gong, X., Spann, N. J., et al. (2015). Transcription factor ISL1 is essential for pacemaker development and function. *J. Clin. Invest.* 125 (8), 3256–3268. doi:10.1172/JCI68257
- Lkhagva, B., Lin, Y. K., Chen, Y. C., Cheng, W. L., Higa, S., Kao, Y. H., et al. (2021). *ZFHX3* knockdown dysregulates mitochondrial adaptations to tachypacing in atrial

- myocytes through enhanced oxidative stress and calcium overload. *Acta Physiol. (Oxf)* 231 (4), e13604. doi:10.1111/apha.13604
- Lu, L., Sirish, P., Zhang, Z., Woltz, R. L., Li, N., Timofeyev, V., et al. (2015). Regulation of gene transcription by voltage-gated L-type calcium channel, Cav1.3. *J. Biol. Chem.* 290 (8), 4663–4676. doi:10.1074/jbc.M114.586883
- Romo-Rodriguez, P., Acevedo-Aguilar, F. J., Lopez-Torres, A., Wrobel, K., Wrobel, K., and Gutierrez-Corona, J. F. (2015). Cr(VI) reduction by gluconolactone and hydrogen peroxide, the reaction products of fungal glucose oxidase: Cooperative interaction with organic acids in the biotransformation of Cr(VI). *Chemosphere* 134, 563–570. doi:10.1016/j.chemosphere.2014.12.009
- Tang, X. H., Gambardella, J., Jankauskas, S., Wang, X., Santulli, G., Gudas, L. J., et al. (2021). A retinoic acid receptor beta (2) agonist improves cardiac function in a heart failure model. *J. Pharmacol. Exp. Ther.* 379 (2), 182–190. doi:10.1124/jpet.121.000806
- Tomomori, S., Nakano, Y., Ochi, H., Onohara, Y., Sairaku, A., Tokuyama, T., et al. (2018). Maintenance of low inflammation level by the ZFH3 SNP rs2106261 minor allele contributes to reduced atrial fibrillation recurrence after pulmonary vein isolation. *PLoS One* 13 (9), e0203281. doi:10.1371/journal.pone.0203281
- Varzideh, F., Mone, P., and Santulli, G. (2022). Bioengineering strategies to create 3D cardiac constructs from human induced pluripotent stem cells. *Bioengineering* 9 (4), 168. doi:10.3390/bioengineering9040168
- Wang, C., Li, H., Wang, S., Mao, X., Yan, D., Wong, S. S., et al. (2018). Repeated non-invasive limb ischemic preconditioning confers cardioprotection through PKC- $\epsilon$ /STAT3 signaling in diabetic rats. *Cell Physiol. Biochem.* 45 (5), 2107–2121. doi:10.1159/000488047
- Xiao, D., Caldow, M., Kim, H. J., Blazev, R., Koopman, R., Manandi, D., et al. (2022). Time-resolved phosphoproteome and proteome analysis reveals kinase signaling on master transcription factors during myogenesis. *iScience* 25 (6), 104489. doi:10.1016/j.isci.2022.104489
- Yu, C., Li, X., Zhao, Y., and Hu, Y. (2023). The role of FOXA family transcription factors in glucolipid metabolism and NAFLD. *Front. Endocrinol. (Lausanne)* 14, 1081500. doi:10.3389/fendo.2023.1081500
- Zhao, J., Yu, L., Xue, X., Xu, Y., Huang, T., Xu, D., et al. (2023). Diminished  $\alpha 7$  nicotinic acetylcholine receptor ( $\alpha 7$ nAChR) rescues amyloid- $\beta$  induced atrial remodeling by oxi-CaMKII/MAPK/AP-1 axis-mediated mitochondrial oxidative stress. *Redox Biol.* 59, 102594. doi:10.1016/j.redox.2022.102594