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Setting a stage: Inflammation during preeclampsia and postpartum

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Preeclampsia (PE) is a leading cause of maternal and fetal mortality worldwide. The immune system plays a critical role in normal pregnancy progression; however, inappropriate inflammatory responses have been consistently linked with PE pathophysiology. This inflammatory phenotype consists of activation of the innate immune system, adaptive immune system, and increased inflammatory mediators in circulation. Moreover, recent studies have shown that the inflammatory profile seen in PE persists into the postpartum period. This manuscript aims to highlight recent advances in research relating to inflammation in PE as well as the inflammation that persists postpartum in women after a PE pregnancy. With the advent of the COVID-19 pandemic, there has been an increase in obstetric disorders associated with COVID-19 infection during pregnancy. This manuscript also aims to shed light on the relationship between COVID-19 infection during pregnancy and the increased incidence of PE in these women.

KEYWORDS

preecclampsia, immune cells, chronic inflammation, pregnancy, postpartum

Introduction

Preeclampsia (PE) is a multisystem obstetric disorder that presents as new onset of hypertension in conjunction with evidence of end-organ dysfunction beyond the 20th week of gestation (ACOG, 2022). PE is a leading cause of maternal-perinatal morbidity and mortality each year and occurs in about 7% of pregnancies (English et al., 2015). In conjunction with hypertension, PE patients may present with proteinuria, HELLP (Hemolysis, Elevated Liver enzymes, and Low Platelets), or visual disturbances (Roos et al., 2012; ACOG, 2022). The only cure for PE is delivery of the fetal-placental unit, making PE a leading cause of premature birth (Staff et al., 2022). Moreover, the offspring of PE pregnancies are at higher risks for stillbirth, FGR (Fetal Growth Restriction), and additional neonatal complications (Fox et al., 2019). Therefore, the management of PE pregnancies is a delicate balance between maternal and fetal health, which could be attributed to the lack of any major developments or changes in treatments for PE in the last 50 years (Bell, 2010).

PE is a state of chronic inflammation with activation of antigen presenting cells (APC's), T helper (Th) cells, B cells, and Natural killer cells which contribute to the presentation of PE symptoms during pregnancy (LaMarca et al., 2016; Aneman et al., 2020). Th cells secrete cytokines that activate innate immune cells, induce production of anti-angiogenic factors, and increase sodium conductance in the nephron (LaMarca et al., 2016). B cells produce agonistic antibodies against the angiotensin II type 1 receptor (AT1-AA) which activate the



fetal antigens to Th cells and polarize them towards a regulatory Th cell phenotype (Tregs). These Tregs then quell immune responses in the placenta and maintain an anti-inflammatory environment through the second trimester. As a normal pregnancy reaches its end, maternal immune cells produce inflammatory signals that help signal the uterus for delivery. In the postpartum period, maternal immune cells retain memory of fetal antigens to help in future pregnancies. In early-onset preeclampsia, inflammatory macrophages, dendritic cells, NK cells, and Th cells disrupt spiral artery formation leading to placental hypoxia. Hypoxia leads to cell death which leads to placental dysfunction and release of fetal antigens. Antigen Presenting Cells present these fetal antigens and polarize Th cells toward an inflammatory phenotype. Inflammatory Th cells then induce production of inflammatory and anti-angiogenic factors from the placenta and other immune cells which lead to placental, vascular, and renal dysfunction contributing to hypertension in pregnancy. These inflammatory cells from preeclampsia retain memory postpartum, and increase the risk for developing PE in subsequent pregnancies. In late-onset preeclampsia, the spiral artery formation occurs normally, however eventually the needs of the placenta out-weigh the perfusion capacity of the uterus, leading to placental hypoxia and a similar inflammatory environment as seen in early-onset preeclampsia.

Renin-Angiotensin system and activate killer cells to target AT1-AA marked cells (Campbell et al., 2018). Moreover, innate immune cells contribute to the inflammatory cytokine milieu while contributing to tissue damage in the kidney and placenta (Aneman et al., 2020). Over the COVID-19 pandemic, there has been an increase in the incidence of PE in women that contracted COVID-19 during their pregnancy (Jamieson and Rasmussen, 2022). COVID-19 infection during pregnancy leads to an immune reaction in the decidua; however, this immune reaction appears to change depending on the trimester in which the COVID-19 infection occurs (Juttukonda et al., 2022). Furthermore, studies in women that contract COVID-19 during pregnancy have shown that there are increased markers of placental dysfunction or placental damage (Jaiswal et al., 2021; Schwartz et al., 2022). Inflammation and placental damage following COVID-19 infection could be contributing factors toward the increased incidence of PE in women with a history of COVID-19 infection during their pregnancies (Villar et al., 2021).

While symptoms of PE usually end upon delivery of the placenta, the inflammation persists into the mother's postpartum period. Research has demonstrated that patients have an increased risk of recurrent PE,

hypertension, coronary heart disease and stroke following a PE pregnancy (Wu et al., 2017; Opichka et al., 2021), thereby illustrating the long-term cardiovascular complications caused by PE. Furthermore, the immune system and inflammation has been repeatedly implicated in the pathophysiology of cardiovascular disease during pregnancy and outside of pregnancy (Harrison et al., 2021; Stefanska et al., 2021). Therefore, this review aims to provide insight into the potential role of immune cells and immune mediators in contributing to PE and cardiovascular disease postpartum of PE. This review aims to highlight clinical studies that have elucidated contributions of the immune system in physiological pregnancy and PE; while also presenting basic science studies that have provided mechanistic insight into how immune mediators impact pregnancy (Figure 1).

Placental ischemia—The initiating event in preeclampsia pathophysiology

During normal pregnancies, fetal trophoblasts invade the maternal myometrium with the aid of uterine immune cells (Staff

et al., 2022). Trophoblasts and immune cells secrete proteases and angiogenic factors to progressively remodel uterine spiral arteries into high-capacity, low-resistance vessels (Albrecht and Pepe, 2020). In PE pregnancies, there is a breakdown in placental perfusion which ultimately results in placental ischemia. Placental ischemia induces the production of inflammatory modulators and antiangiogenic factors (Zhang, 2018) which contribute to vascular dysfunction in the placenta and peripheral vessels. Furthermore, these circulating placental factors cause endothelial dysfunction and oxidative stress which contribute to hypertension and multi-organ dysfunction as seen in PE (Rana et al., 2019).

PE presents as systolic pressure ≥140 mmHg or diastolic pressure ≥90 mmHg with an accompanying organ dysfunction including elevated 24-h urine protein value of ≥300 mg, urine protein to creatine ratio of ≥0.3, thrombocytopenia, impaired liver function, pulmonary edema, or new-onset headache with or without visual disturbances (Karrar and Hong, 2022). PE can be stratified by the timing of symptom presentation. While these categories of PE are unified by placental ischemia, they are believed to stem from different etiologies. PE that develops prior to 34 weeks of gestation is considered early-onset (EO) PE, PE that develops after 34 weeks of gestation is considered late-onset (LO) PE, and while PE that develops after delivery of the fetal-placental unit is considered postpartum PE (PP-PE) (Raymond and Peterson, 2011; Hauspurg and Jeyabalan, 2022). EOPE is characterized by insufficient invasion of trophoblasts into the endometrium resulting in poor spiral artery remodeling leading to placental ischemia and PE phenotype (Tamas et al., 2022). EOPE is considered the most severe form of PE and causes the highest rates of morbidity and mortality in mother and neonate compared of the PE subsets (Madazli et al., 2014). EOPE mothers are the most likely to have placental pathologies and higher PE recurrence rates compared to LOPE mothers (Li et al., 2014; Fillion et al., 2021). Mothers with both placental pathologies (syncytial knots, placental infarcts, etc.) and severe PE symptoms are more likely to have strokes or develop cardiovascular disease postpartum (Veerbeek et al., 2015). LOPE is associated with normal placentation in early pregnancy, with placental dysfunction occurring later in pregnancy. LOPE is projected to occur when the needs of the placenta are above the perfusion capacity of the uterus leading to placental ischemia and the symptoms of PE (Staff et al., 2022). Even though LOPE is less associated with neonatal morbidity (Staff et al., 2016; Wadhwani et al., 2020), LOPE still predisposes mothers and offspring to develop cardiovascular disease later in life. Moreover, children born of LOPE pregnancies are less likely to be growth restricted. However, they still have increased incidence of cardiovascular disease and stroke in later life compared to children born of normal pregnancies (Kajantie et al., 2009; Touwslager et al., 2012), thereby suggesting that cardiovascular disease in PE offspring is caused by more than just FGR. LOPE is often associated with increased maternal BMI, high weight gain during pregnancy, and other metabolic disorders (Robillard et al., 2019; Robillard et al., 2022). Obesity and metabolic disorders are considered states of chronic inflammation (Esposito and Giugliano, 2004; Khanna et al., 2022), and these metabolic and inflammatory signals may contribute to the increase risk of cardiovascular disease in offspring of women with LOPE. Lastly, there are rare cases where patients develop a PE phenotype after delivery, which is known as PP-PE. Patients with PP-PE have similar placental vasculopathies that resemble LOPE by demonstrating a similar role for placenta dysfunction (Ditisheim et al., 2020; Hauspurg and Jeyabalan, 2022). But, more studies are needed to elucidate the mechanisms that induce postpartum PE.

Dynamic shifts in the immune system during normal pregnancy

Implantation and the first trimester of pregnancy

In a normal pregnancy, the maternal immune system must balance immune tolerance of the semi-allogenic fetus with defending the mother from disease. However, disruption of this balance has been associated with developing PE. During pregnancy, women undergo time-dependent shifts in activation of the innate and adaptive immune systems, but dysregulation of the immune system can cause adverse outcomes in both the mother and fetus (Cornelius et al., 2019). In the first trimester, macrophages (M ϕ) and Natural Killer (NK) cells are two of the most prevalent cells in the uterus (Jena et al., 2019). Uterine subsets of both NK (uNK) cells Mø $(uM\phi)$ are pivotal in uterine remodeling during pregnancy (Gustafsson et al., 2008; Jetten et al., 2014) (Bazhenov et al., 2019). The uNK cell population have high concentrations of intracellular killer granules, but do not release cytolytic granules and are not efficient in cell killing compared to conventional NK cells (Faas and de Vos, 2017). uNK cells have higher expression of Killer cell Immunoglobulin-like Receptors than conventional NK cells; however, uNK cells lack FcyRIII (CD16), which may contribute to their reduced killing capacity in normal pregnancies (Koopman et al., 2003). CD16 recognizes the FC region of IgG antibodies and is responsible for antibody induced cellular cytotoxicity (Yeap et al., 2017). uMq are unique compared to peripheral macrophages (Houser et al., 2011); however, uMq resemble alternatively activated M2s phenotype which produces lower levels of the inflammatory IL-1 while producing higher levels of the anti-inflammatory IL-10 compared to inflammatory M1s (Mizuno et al., 1994; Gough et al., 2001). These uM ϕ are also important in anti-microbial defense in the uterus and clearing cellular debris during pregnancy (Thiruchelvam et al., 2013).

During embryo implantation, the uterine immune cells help to induce vascular remodeling while initiating processes to promote fetal tolerance. uNK cells produce copious amounts of interferon gamma (IFN-y) which destabilizes vascular endothelial cells and initiates uterine artery remodeling (Ashkar et al., 2000). This uNK cell derived IFN- γ also activates uM ϕ s which allow them to assist in vascular remodeling. These uNK cells and Møs migrate and localize to fetal trophoblasts (Helige et al., 2014). The uNK cells and Møs then produce proteases to destabilize vascular smooth muscle (Naruse et al., 2009; Smith et al., 2009) and angiogenic factors (Ferrante et al., 2013; El-Azzamy et al., 2018) that promote growth of new blood vessels and maturation of existing blood vessels. During this critical stage of implantation, uterine stromal cells and uterine dendritic cells (uDCs) work to prevent excessive vascularization during implantation. Uterine stromal cells are also important cytokine producers and cross-talk with uDCs during pregnancy. Uterine stromal cells produce granulocyte-colony stimulating factor

(G-CSF) which appears to increase IL-1 β from uDCs (Shao et al., 2020). In the short term, this may contribute to uDCs producing soluble Flt1, which binds soluble vascular endothelial growth factor (VEGF) and serves as a control for angiogenesis during pregnancy (Plaks et al., 2008).

IFN- γ from uNK cells also induces uM ϕ 's to express HLA-II, which is an important in inducing T helper (Th) cell activation (Xie et al., 2005). T helper (Th) cells are CD4+ T cells that are crucial for both directing immune responses and helping to orchestrate transitions in pregnancy. Th1 cells are characterized by the expression of the transcription factor Tbet, and help with uterine remodeling during the first trimester (Faas and De Vos, 2018). Th1s and uNK cells also produce Tumor Necrosis Factor (TNF)-a which prevents excessive invasion of trophoblasts into the uterus through activating the NF-kB pathway (Todt et al., 1996; Torchinsky et al., 2003). Inhibiting IFN-γ in early pregnancy prevents successful uterine artery remodeling therefore causing fetal demise (Ashkar et al., 2000), suggesting roles for Th1s and uNK cells to promote vascular remodeling in pregnancy through IFN-y production. Overall, uNK cells, uMqs, uDCs, and Th1s cells carefully collaborate to promote the formation of the maternal-fetal interface during the first trimester. Moreover, inflammatory and anti-inflammatory mechanisms are important during implantation in the first trimester.

Second trimester of pregnancy

The transition from early to mid-pregnancy is accompanied by a shift from a pro-inflammatory to an anti-inflammatory uterine environment. DCs are regulate vascularization during the first trimester however DCs are also professional APC's that internalize antigens and present them on maior histocompatibility complex-II in order to activate Th cells. In pregnancy, DCs help transition from the early pregnancy proinflammatory Th1 environment to the middle pregnancy antiinflammatory Th2 environment. Cross-talk between uterine stromal cells and uDCs eventually promotes uDCs to express co-stimulators of Th cell activation CD80 and CD86 (Shao et al., 2020). Moreover, these uDCs polarize Th cells towards a Th2 phenotype in vitro. Negishi et al. (2012) found that DCs that express Dendritic Cell Inhibitory Receptor 2 (33D1+) DCs slowly increase in number during the first trimester and these 33D1+ DCs promote the switch from Th1s to Th2s in middle pregnancy. Further supporting a role for uDCs to induce the pivot from the inflammatory environment needed for implantation towards the anti-inflammatory Th2 environment needed for pregnancy tolerance. Møs are another important APC that help with clearing cellular debris and help polarize Th cells towards the appropriate Th subtype to promote fetal tolerance in pregnancy. Specifically, T cell immunoglobulin mucin-3 (Tim- 3^+) uM ϕ help polarize Th cells towards anti-inflammatory Th2 and Treg phenotypes (Li et al., 2022). Tim-3⁺ blockade lead to decreased fetal viability and fetal growth restriction, further implicating Mq in promoting fetal health during pregnancy. Together these studies show that uDCs and uMq help to promote an anti-inflammatory environment in the transition towards the second trimester of pregnancy.

Th2 cells begin to increase near the end of the first trimester and continue to predominate over Th1 cells through the rest of the pregnancy (Saito et al., 1999). Th2 cells are producers of the cytokine IL-4, which prevents Th1 cells proliferation and promotes a Th2 profile in normal pregnancies (Lazarski et al., 2013). IL-4 also inhibits IFN- γ production, which reduces NK cell activation and promotes fetal tolerance (Zissler et al., 2016). Th2 cells also promote B cells to produce protective "asymmetric" antibodies during pregnancy (Zenclussen et al., 2001). These asymmetric antibodies bind fetal antigens on the placenta, but do not activate killer cells. Asymmetric antibodies effectively block the fetal antigens from being found by the maternal immune system. Therefore asymmetric antibodies protect paternal antigens and the fetus from killer immune cells while promoting immune tolerance in pregnancy (Gutierrez et al., 2005).

In normal pregnancy, paternal antigen-specific Treg cells expand in circulation and in the uterus to promote fetal tolerance (Kahn and Baltimore, 2010; Shima et al., 2015; Huang et al., 2020). Tregs are known as important producers of IL-10 during pregnancy (Cheng and Sharma, 2015). IL-10 reduces antigen presentation co-stimulatory molecules by APCs which downregulates the ability of APCs to produce IL-12, which is an important stimulator of NK cells and Th1s (Moore et al., 2001; Zundler and Neurath, 2015). Treg cells also constitutively express CTLA4 (Cytotoxic T-Lymphocyte Associated-protein), which reduces the ability of APCs to activate new Th cells and prevent new inflammatory responses (Qureshi et al., 2011). Therefore, Tregs use IL-10 and CTLA4 to prevent new Th1 polarization. IL-10 also protects nitric oxide production pathways (Gunnett et al., 2002) and prevents vascular dysfunction from endogenous vasoconstrictors (Didion et al., 2009).

Third trimester of pregnancy and parturition

As pregnancy reaches its end, parturition is characterized by a drastic shift towards an inflammatory phenotype (Leimert et al., 2021). Two to four weeks prior to parturition are characterized by a shift from pregnancy maintenance to preparation for labor (Stelzer et al., 2021). During this period, IL-6 and IL-1 β promote the production of prostaglandins, endothelin-1, and cyclooxygenases which induce uterine contractions during labor (Ashkar et al., 2000). This inflammatory phenotype may in part be due to the progressive decrease in circulating progesterone and Progesterone Induced Blocking Factor (PIBF) that occurs as normal pregnancies progress to parturition (Polgar et al., 2004; Hudic et al., 2016). Progesterone and PIBF both inhibit lymphocyte proliferation and inflammatory cytokine production, which potentially implicates them as important anti-inflammatory agents in the earlier portion of the pregnancy (Fedotcheva et al., 2022). Near the 37th week of pregnancy, PIBF decreases triggering increases in IL-1 β and IL-6 along with decreases in the anti-inflammatory cytokines IL-1Ra and IL-9 (Polgar et al., 2004; Jarmund et al., 2021). Uterine levels of IL-6 appear to be important to induce the onset of labor, as IL-6 deficient mice experience delayed labor compared to controls (Gomez-Lopez et al., 2016). As IL-6 is produced by many cell types, multiple cellular sources of IL-6 likely contribute to the IL-6 needed for parturition. Among major IL-6 producers are M φ s which are sensitive toward changes in estrogen and progesterone during pregnancy (Mendoza-Cabrera et al., 2020). At term, uM φ s invade the decidua and cervix and produce reactive oxygen species and TNF- α which lead promote cervical ripening (Hamilton et al., 2012). At this time, uterine levels of the inflammatory Th9 and Th17 subsets increase (Gomez-Lopez et al., 2016). However there has not been a direct link drawn between uM φ s and the increase in inflammatory Th subsets. Moreover, the precise mechanisms that induce the shift towards parturition have not been fully elucidated and more studies are needed to tease at the role of the immune system during parturition.

The chronic inflammatory environment in preeclampsia

The immune system during PE is in a state of chronic inflammation (Cornelius et al., 2019; Aneman et al., 2020; Stefanska et al., 2021). Patients with PE have activation of both the innate and adaptive arms of the immune system which induce a feed-forward mechanism for inflammation (Murray et al., 2021). Cytolytic NK cells, macrophages, dendritic cells induce tissue damage and vascular dysfunction in PE while activating Th cells through antigen presentation (Deer et al., 2023). Moreover, patients with PE have activated Th cells and activated B cells producing agonistic antibodies against the angiotensin II type 1 receptor (AT1-AA) which lead to antigen specific mechanisms of cell destruction and tissue damage (LaMarca et al., 2016). The communication between immune cells during pregnancy in preeclampsia may also lead to immune memory which could contribute to inflammation postpartum (van Rijn et al., 2016; Kieffer et al., 2017; Brien et al., 2019). This prolonged inflammatory state contributes to hypertension, endothelial dysfunction, and fetal complications (Murray et al., 2021). Because PE presents after the 20th week of gestation, most studies that have investigated inflammation in PE have been limited to studying middle gestation and late gestation periods.

Dendritic cells and macrophages in preeclampsia

While the initiating factor for uterine inflammation in PE is not fully understood, $M\phi$ and DCs play a role to expand the inflammatory environment in PE. PE placentas have increased invading DCs and macrophages compared to normal placentas (Huang et al., 2008). These uMq also produce high levels of TNF- α and IFN- γ which contribute to apoptosis of fetal trophoblasts in pregnant mice (Blois et al., 2004). Excessive debris from dying trophoblasts could lead to more internalization and presentation of fetal antigens by $M\phi$ and DCs leading to inflammatory Th subsets in PE. Moreover, PE placentas have shown increased chemokines CCL2, CCL4, CCL7, and CCL20 which promote recruitment of innate immune cells and T cells which could contribute to placental dysfunction in PE. There has been some dispute in the phenotype of uMqs in PE, with some studies showing lower levels of M2s in the placenta while others show higher levels of M2s during PE (Reister et al., 1999; Katabuchi et al., 2003; Schonkeren et al., 2011). These discrepancies have been equated to location of M φ s, citing less localization M2s near trophoblasts and spiral arteries in PE (Faas et al., 2014). Illustrating an important distinction in macrophage location during the pathophysiology of PE. However, there have been few studies to investigate the role of M φ s in the induction of improper spiral artery remodeling in PE; therefore, much more research is needed to better understand how M φ s impact the progression of PE pregnancies.

DCs have been similarly underserved in research of PE pregnancies. In PE there is a higher conventional myeloid DCs to plasmacytoid DC ratio in the circulation (Darmochwal-Kolarz et al., 2003; Wang et al., 2013; Li et al., 2019). These studies also associated the rise in myeloid DCs with increased circulating Th1 and Th17 populations, postulating that myeloid DCs are related to the increased inflammatory Th cell phenotype of PE. However, these studies do not provide as much insight into the placental DC populations in PE. A study by Zhang et al. (2017) showed that there were increased mature dendritic cells in the decidua of PE women, and these DCs had higher expression of a DC specific long non-coding RNA strand that assists in DC maturation by phosphorylating STAT-3. Moreover, a study by Panda et al. (2012) showed that circulating DCs expressed higher levels of TLR3, TLR4, and TLR9 compared to DCs from control patients. These DCs also secreted higher levels of TNF-a, IFN-a, IL-6, and IL-12 measured by flow cytometry. The authors also found that PE DCs were less able to mount inflammatory responses to TLR activation, suggesting that the dysregulation of TLR signaling also impaired DC responses to pathogens. Interestingly, women with PE have increased circulating memory T cells during pregnancy (Chaiworapongsa et al., 2002). Which could suggest that antigen presentation to Th cells leads to development of Th1's and Th17s during PE but also leads to memory T cell production that persists after a PE pregnancy. However, there is limited research available investigating how DCs become altered in PE and how they are involved in placental dysfunction in PE.

T helper cells and B cells in preeclampsia

Patients with PE have an increase in Th1/Th2 ratio and an increase Th17/Treg ratio in the placenta and in the circulation (Eghbal-Fard et al., 2019; Romao-Veiga et al., 2022) (Wallace et al., 2014). The location of where Th cells become activated is unknown, but Th cells induce dysfunction at the placental level as well as in other organs and the vasculature (Murray et al., 2021). Th cells are important mediators to spread immune signals throughout the body; and, Th1 cells and Th17 subsets are potent inducers of inflammatory actions and promote cell mediated cytotoxicity (Wang et al., 2020). Interestingly, adoptive transfer of placental Th cells from placentas of PE women induce hypertension in pregnant athymic nude rats (Harmon et al., 2019). Th cell adoptive transfer also lead to increased plasma sflt-1, IL-17, and TNF-a, while also leading to increased expression of pre-proendothelin-1 mRNA which is the precursor to the potent vasoconstrictor endothelin-1. This study suggested that placental Th cells contribute to vascular and renal dysfunction during PE. Animal models of PE have been important to help in delineating the

effects of T cells on the placenta, the vascular, and the kidney in PE. Adoptive transfer of Th1 cells from the Reduced Uterine Perfusion Pressure (RUPP) model of PE induce hypertension and AT1-AA, placental mitochondrial oxidative stress and FGR, as well as renal oxidative stress during pregnancy (Zenclussen, 2006; LaMarca et al., 2008a). Together these studies illustrate that Th cells activated in response to placental ischemia contribute to dysfunction in the vasculature, placenta, and the kidney during pregnancy. Moreover, adoptive transfer of Th17 cells induce hypertension, placental NK cell activation, circulating NK cell activation, and FGR in pregnant rats (Shields et al., 2018). Further showing that Th17 cells are also able to contribute to placental and vascular dysfunctions in pregnancy with the help of NK cells.

Our lab has shown that Th cells stimulated by placental ischemia from rats or humans induce hypertension, mitochondrial dysfunction, sflt-1, AT1-AA, and other features of PE in pregnant rats (Deer et al., 2021; Reeve et al., 2022). However, blockade of communication between Th cells and B cells prevents the features of PE induced by Th cell adoptive transfer (Cornelius et al., 2015). This indicates that Th cell-B cell communication could be an important feature contributing to the pathophysiology of PE. This is attributed to T cell-B cell communication as an integral component in stimulating B cells to transform into Memory B cells which retain antigen memory long after antigen exposure (Palm and Henry, 2019). There has been much less investigation into the role of B cells in the pathophysiology of PE. B cells can be divided into B1 and B2 subsets. B1 cells are innate-like B cells that are responsible for T cell independent antibody responses while B2 cells are classical B cells that are responsible for T cell dependent antibody responses (Mahajan et al., 2021). In the context of PE, Jensen et al. implicated B1 cells as potential producers of AT1-AA in vitro after exposing isolated placental B1 cells to patient serum (Jensen et al., 2012). Our lab recently showed that adoptive transfer of RUPP B2 cells into pregnant rats were able to induce hypertension, AT1-AA production, and NK cell activation during pregnancy (Herrock et al., 2022), therefore, implicating both B1 and B2 subsets in the pathophysiology of PE.

Soluble immune factors in preeclampsia

Th1 cells are important producers of inflammatory cytokines IFN- γ and TNF- α which are both increased in women with PE (Sheibak et al., 2020). Inhibiting TNF- α in RUPP rats attenuated hypertension, Endothelin-1, oxidative stress, and NK cell activation (LaMarca et al., 2008b; Cunningham et al., 2020). Other studies have infused TNF-a into pregnant animals to see the effects of total TNFα during pregnancy (Bobek et al., 2015; Ampey et al., 2019; Jayaram et al., 2021). TNF- α alone induces symptoms of PE in animal models including hypertension, FGR, oxidative stress, Endothelin-1 and AT1-AA in pregnancy (LaMarca et al., 2005; Jayaram et al., 2021). But inhibition of TNF-a in multiple models of PE can attenuate hypertension, maternal inflammation, and fetal morbidity (Irani et al., 2010; Travis et al., 2021). PE patients also have increased Th17s and IL-17 (Ding et al., 2019; El Shahaway et al., 2019). IL-17 can cause more Th17 cells which make more IL-17 in a positivefeedback loop which contributes to the inflammation in PE (Travis et al., 2020). Th17s and IL-17 have been connected with oxidative stress, NK cell activation, anti-angiogenic factors, and AT1-AA production in pregnancy, all of which contribute to hypertension and FGR (Cornelius et al., 2013; Travis et al., 2020). Alternatively, IL-17 inhibition prevents hypertension and oxidative stress in placental ischemic rats (Travis et al., 2020) and elicits natural killer cell activation and hypertension further supporting a roll for Th17s and IL-17 in PE (Travis et al., 2019).

Women with PE have activated B cells producing AT1-AA which causes AT1 receptor activation leading to vasoconstriction in afferent arterioles, renal dysfunction and hypertension during pregnancy (Wallukat et al., 1999). AT1-AA acts by binding the AT1 receptor and stimulating it similar to angiotensin II but also work synergistically with endogenous angiotensin II to further increase blood pressure, oxidative stress, production of sflt-1 and Endothelin, indicating that it may exacerbate the activity of endogenous ANGII in PE patients (Brewer et al., 2013). AT1-AA also causes natural killer cell activation, which may be one mechanism responsible for multi-organ dysfunction in PE (Cunningham et al., 2018; Zhai et al., 2022). AT1-AA is also able to induce oxidative stress through NADPH oxidase and activate NF-kB pathway which contribute to vascular stress in PE (Dechend et al., 2003). Moreover, AT1-AA is also increased in the RUPP model of placental ischemia and our lab has shown that directly inhibiting AT1-AA can prevent the PE-like phenotype associated with placental ischemia (Cunningham et al., 2018).

Immune memory after normal pregnancy

Adaptive immune memory

After the immune adaptations that happen in pregnancy, the maternal immune system retains memory of paternal antigens in order to facilitate future pregnancies. Following activation, T cells and B cells transform into effector cells and memory cells. While the effector cells are responsible for the short-term immune response, the memory cells retain long-term antigen memory in case of future infections. Paternal antigen memory Treg cells remain after a healthy pregnancy, and persist at least one-year postpartum (Kieffer et al., 2017). Upon initiation of another pregnancy with the same partner, memory Tregs rapidly expand in the circulation upon initiation of a new pregnancy (Tilburgs et al., 2008). These memory Tregs then progressively migrate towards the placenta and promote tolerance of the new fetus (Rowe et al., 2012; Gomez-Lopez et al., 2020). It has been suggested that insufficient memory Treg expansion could contribute to infertility (Jasper et al., 2006) suggesting an important role for Tregs in pregnancy success. Moreover, first-time pregnancies have higher incidence of PE (ACOG, 2022), lower Treg expansion (Rowe et al., 2012), and increased sflt-1 (Bdolah et al., 2014) compared to second-time pregnancies. Furthermore, if a subsequent pregnancy is with a different father than the first, there is no Treg induced protection against PE (Need, 1975). These trends could be explained by the lack of paternal-antigen Tregs, implicating paternal-antigen Tregs to promote fetal tolerance in pregnancy.

Innate trained immunity

"Trained Immunity" is a relatively new concept in immunology and appears to be unique to the innate immune system. In simple terms, innate immune cells can make epigenetic modifications after encountering an antigen (Netea et al., 2016). This style of memory is in contrast to adaptive immunity, which involves gene recombination events to retain long-term antigen memory. A study by Novakovic et al. (2016) showed that murine Mqs retain epigenetic modifications following *in vitro* exposure to LPS. This study also importantly showed that these epigenetic modifications may be reverse, showing the increased volatility of trained immunity compared to adaptive immunity. Trained immunity is associated with a three-month to one-year lifetime, but there have been cases of trained immunity lasting up to five years (Nankabirwa et al., 2015).

In the context of pregnancy, trained immunity is still in its infancy in terms of investigation.Gamliel et al. (2018) showed that a subset of "pregnancy trained" uNK cells were more readily able to produce IFN- γ and VEGF compared to untrained uNK cells. As stated above, both IFN- γ and VEGF are crucial in implantation and establishing the maternal-fetal interface. Therefore, this study provides crucial insight into direct applications of trained immunity during physiological pregnancy. However, this is one of the only studies to show direct mechanisms of trained immunity in human pregnancy.

Immune memory and inflammation after a preeclamptic pregnancy

Adaptive immune memory

A recent study also showed that women with PP-PE have increased inflammatory markers and placental pathologies (Brien et al., 2019). Brien et al. found that patients with PE or PP-PE had increased CD4+ and CD8+ T cells in circulation at time of PE presentation. However, only PP-PE patients had increased circulating Natural Killer T (NKT) cells. NKT cells are an innatelike lymphoid cells that stem from the thymus but share markers of T cells and NK cells (Hashemi et al., 2017). While NKT cells have been implicated in the pathophysiology of PE, this study further implicates NKT cells and inflammation in the pathophysiology of PP-PE. Histological analysis of placentas revealed increased syncytial knots, a morphological marker of dysfunction, suggesting that placental dysfunction is important in PP-PE. This study shows that inflammation may be important in the pathophysiology of PP-PE as well. Therefore, therapies targeting inflammatory mediators may be helpful in treating PE in pregnancy or postpartum.

Following a PE pregnancy, there is a persistent inflammatory profile. Vitoratos et al. (2010) found that PE women had increased plasma TNF- α during pregnancy and this increase in TNF- α persisted three months postpartum. Interestingly, they found that plasma IL-6 was not changed during pregnancy, but plasma IL-6 increased postpartum in PE women compared to NP indicating that inflammation may worsen after a PE pregnancy. It was later found that previously PE women have an exaggerated inflammatory response to the flu vaccine which included increased C reactive protein and IL-18 which are both involved in Th1 responses (van Rijn et al., 2016). This indicates that there are alterations to the normal inflammatory response in women that previously had PE that lasts well into their postpartum period. However, circulating memory Th cells are decreased in patients with a previous PE pregnancy (Kieffer et al., 2019), suggesting that Th dysfunction continues into the postpartum period after PE. Studies of Treg cells in idiopathic pre-term labor patients revealed that decreased functional Tregs is associated with recurrent pregnancy loss (Gomez-Lopez et al., 2020), which could be caused by a sustained inflammatory environment. This is mirrored in subsequent pregnancies following PE, where Tregs do not undergo peripheral expansion nor do they migrate to the uterus (Tsuda et al., 2018). This suggests that dysfunctional memory Tregs may be important in PE recurrence. Interestingly, there is an inverse relationship between memory Tregs and Memory B cells in previously PE patients where lower Tregs is associated with higher Memory B cells (Zeng et al., 2013). Memory B cells from a previously PE pregnancy could continue to secrete AT1-AA and contribute to PE in subsequent pregnancies. Rieber-Mohn et al. found that AT1-AA was increased in serum eight years postpartum of a PE pregnancy (Rieber-Mohn et al., 2018), supporting a role for Memory B cells and AT1-AA not only in PE pathophysiology but also in consequences observed in postpartum PE. Future studies in humans or animal models of PE could investigate the role of memory immune cells to contribute to PE in future pregnancies.

Innate trained immunity

Unfortunately, there is even less information investigating trained immunity in PE than normal pregnancy. However, an elegant study by Huang et al. showed that both uneventful and PE pregnancies lead to methylation changes in both Th and NK cell populations (Huang et al., 2021). These DNA methylation events are distinct between physiological pregnancies or PE pregnancies, further implicating immunological memory in the increased risk to develop PE following a PE pregnancy. Please refer to the cited article by Huang et al. to get a more thorough understanding of this fascinating immunological adaption to pregnancy.

Adverse outcomes associated with risk factors for preeclampsia and COVID-19

There are many factors that predispose patients to develop PE in pregnancy. Cardiometabolic disease like obesity, type 2 diabetes (T2D), and hypertension increase the risk for adverse outcomes of pregnancy, including PE (Alston et al., 2022). Each of these diseases have adverse effects on the health of pregnancy, but importantly they are all recognized as states of chronic inflammation (Harrison et al., 2021). Moreover, patients with autoimmune disease like Systemic Lupus Erythematosus, Rheumatoid Arthritis, and Type 1 Diabetes are at an increased risk for developing PE (Tamas et al., 2022). There is also an association between infectious diseases in pregnancy and increased incidence of PE, such as seen in those diagnosed with COVID-19 (Nourollahpour Shiadeh et al., 2017; Lokki et al., 2018).

SARS-CoV-2 infection has been shown to significantly increase complications of pregnancy including PE (Jamieson and Rasmussen, 2022). In 2019, the novel, zoonotic virus SARS-CoV-2 became a global pandemic (Sun et al., 2020). SARS-CoV-2 acts by binding ACE-2 (Angiotensin Converting Enzyme-2) in order to invade host cells (Seyed Hosseini et al., 2020). The virus takes over the host cell, replicates, and eventually spreads to other cells in the body. While its uncertain if pregnant patients have higher risk of contracting COVID (Jamieson and Rasmussen, 2022), pregnant patients with COVID have higher rates of morbidity and mortality (Sayad et al., 2022). Importantly, pregnant patients with COVID have higher rates PE, eclampsia, and preterm birth compared to control (Villar et al., 2021; Wei et al., 2021). Placental samples from a cohort of women with COVID in pregnancy and fetal demise have showed copious placental fibrosis, histiocytic intervillositis, and trophoblast necrosis which has been attributed to COVID induced placental destruction (Jaiswal et al., 2021; Schwartz et al., 2022). Strangely, COVID patients also have increased serum levels of AT1-AA (Rodriguez-Perez et al., 2021). AT1-AA in COVID patients could contribute to post-senescence hypertension in some former COVID patients (Angeli et al., 2022); but, AT1-AA could also contribute to the increased risk for PE in pregnant patients. Moreover, the Renin Angiotensin Aldosterone System (RAAS) must compensate for increased fluid loads during pregnancy (Irani and Xia, 2011; Xia and Kellems, 2011), but dysregulation of RAAS by COVID infection could also contribute to hypertension during pregnancy. Additionally, patients diagnosed with COVID during pregnancy should be monitored postpartum to examine the effects of their uniquely challenging pregnancy.

Conclusion

In conclusion, developing PE is a major contributor to cardiovascular and immune complications in pregnancy and postpartum. After a PE pregnancy, patients have increased incidence of cardiovascular disease, stroke, and PE in a subsequent pregnancy which could be caused by inflammatory mediators. Moreover, developing severe PE is associated with post-traumatic stress disorders that may discourage patients from

References

ACOG (2022). Preeclampsia and high blood pressure during pregnancy. Philadelphia, PA: ACOG. [cited 2022 06.07.2022]; Available from: https://www.acog.org/womens-health/faqs/preeclampsia-and-high-blood-pressure-during-pregnancy.

Albrecht, E. D., and Pepe, G. J. (2020). Regulation of uterine spiral artery remodeling: A review. *Reprod. Sci.* 27 (10), 1932–1942. doi:10.1007/s43032-020-00212-8

Alston, M. C., Redman, L. M., and Sones, J. L. (2022). An overview of obesity, cholesterol, and systemic inflammation in preeclampsia. *Nutrients* 14 (10), 2087. doi:10. 3390/nu14102087

Ampey, A. C., Boeldt, D. S., Clemente, L., Grummer, M. A., Yi, F., Magness, R. R., et al. (2019). TNF-alpha inhibits pregnancy-adapted Ca(2+) signaling in uterine artery endothelial cells. *Mol. Cell. Endocrinol.* 488, 14–24. doi:10.1016/j.mce.2019.02.008

Aneman, I., Pienaar, D., Suvakov, S., Simic, T. P., Garovic, V. D., and McClements, L. (2020). Mechanisms of key innate immune cells in early- and late-onset preeclampsia. *Front. Immunol.* 11, 1864. doi:10.3389/fimmu.2020.01864

Angeli, F., Reboldi, G., Trapasso, M., and Verdecchia, P. (2022). Hypertension after COVID-19 vaccination. G. Ital. Cardiol. (Rome) 23 (1), 10–14. doi:10.1714/3715.37055

Ashkar, A. A., Di Santo, J. P., and Croy, B. A. (2000). Interferon gamma contributes to initiation of uterine vascular modification, decidual integrity, and uterine natural killer

having future desired pregnancies (Hoedjes et al., 2011). Recent evidence after the COVID pandemic has shed more light on the relationship between placental ischemia, inflammation, and PE suggesting that immune dysregulation could be a major contributor to PE development. Yet, more studies are needed to investigate the mechanisms involved to cause cardiovascular disease or PE recurrence in previously PE patients. Immune memory may contribute to cardiovascular disease or PE recurrence following a PE pregnancy, but more studies could investigate mechanisms of inflammation to induce these features in animal models of PE. Modulators of inflammation have shown promise to lessen maternal disease in models of PE, but longitudinal studies could show efficacy of these agents in the postpartum period. Importantly, investigators must continue to reveal important pathophysiological components that lead to the onset of PE so that we may better serve patients stricken by this terrible disease.

Author contributions

OH: Conceptualization, Writing—Original Draft; ED: Writing—Reviewing and Editing; BL: Conceptualization, Supervision, Writing—Reviewing and Editing.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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cell maturation during normal murine pregnancy. J. Exp. Med. 192 (2), 259–270. doi:10. 1084/jem.192.2.259

Bazhenov, D. O., Khokhlova, E. V., Viazmina, L. P., Furaeva, K. N., Mikhailova, V. A., Kostin, N. A., et al. (2019). Characteristics of natural killer cell interaction with trophoblast cells during pregnancy. *Curr. Mol. Med.* 20, 202–219. doi:10.2174/1566524019666190808103227

Bdolah, Y., Elchalal, U., Natanson-Yaron, S., Yechiam, H., Bdolah-Abram, T., Greenfield, C., et al. (2014). Relationship between nulliparity and preeclampsia may be explained by altered circulating soluble fms-like tyrosine kinase 1. *Hypertens. Pregnancy* 33 (2), 250–259. doi:10.3109/10641955.2013.858745

Bell, M. J. (2010). A historical overview of preeclampsia-eclampsia. J. Obstet. Gynecol. Neonatal Nurs. 39 (5), 510–518. doi:10.1111/j.1552-6909.2010.01172.x

Blois, S. M., Alba Soto, C. D., Tometten, M., Klapp, B. F., Margni, R. A., and Arck, P. C. (2004). Lineage, maturity, and phenotype of uterine murine dendritic cells throughout gestation indicate a protective role in maintaining pregnancy. *Biol. Reprod.* 70 (4), 1018–1023. doi:10.1095/biolreprod.103.022640

Bobek, G., Surmon, L., Mirabito, K. M., Makris, A., and Hennessy, A. (2015). Placental regulation of inflammation and hypoxia after TNF-alpha infusion in mice. *Am. J. Reprod. Immunol.* 74 (5), 407–418. doi:10.1111/aji.12417

Brewer, J., Liu, R., Lu, Y., Scott, J., Wallace, K., Wallukat, G., et al. (2013). Endothelin-1, oxidative stress, and endogenous angiotensin II: Mechanisms of angiotensin II type I receptor autoantibody-enhanced renal and blood pressure response during pregnancy. *Hypertension* 62 (5), 886–892. doi:10.1161/ HYPERTENSIONAHA.113.01648

Brien, M. E., Boufaied, I., Soglio, D. D., Rey, E., Leduc, L., and Girard, S. (2019). Distinct inflammatory profile in preeclampsia and postpartum preeclampsia reveal unique mechanisms. *Biol. Reprod.* 100 (1), 187–194. doi:10.1093/biolre/ioy164

Campbell, N., LaMarca, B., and Cunningham, M. W., Jr. (2018). The role of agonistic autoantibodies to the angiotensin II type 1 receptor (AT1-AA) in pathophysiology of preeclampsia. *Curr. Pharm. Biotechnol.* 19 (10), 781–785. doi:10.2174/ 1389201019666180925121254

Chaiworapongsa, T., Gervasi, M. T., Refuerzo, J., Espinoza, J., Yoshimatsu, J., Berman, S., et al. (2002). Maternal lymphocyte subpopulations (CD45RA+ and CD45RO+) in preeclampsia. *Am. J. Obstet. Gynecol.* 187 (4), 889–893. doi:10.1067/mob.2002.127309

Cheng, S. B., and Sharma, S. (2015). Interleukin-10: A pleiotropic regulator in pregnancy. Am. J. Reprod. Immunol. 73 (6), 487-500. doi:10.1111/aji.12329

Cornelius, D. C., Hogg, J. P., Scott, J., Wallace, K., Herse, F., Moseley, J., et al. (2013). Administration of interleukin-17 soluble receptor C suppresses TH17 cells, oxidative stress, and hypertension in response to placental ischemia during pregnancy. *Hypertension* 62 (6), 1068–1073. doi:10.1161/HYPERTENSIONAHA.113.01514

Cornelius, D. C., Castillo, J., Porter, J., Amaral, L. M., Campbell, N., Paige, A., et al. (2015). Blockade of CD40 ligand for intercellular communication reduces hypertension, placental oxidative stress, and AT1-AA in response to adoptive transfer of CD4+ T lymphocytes from RUPP rats. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 309 (10), R1243–R1250. doi:10.1152/ajpregu.00273.2015

Cornelius, D. C., Cottrell, J., Amaral, L. M., and LaMarca, B. (2019). Inflammatory mediators: A causal link to hypertension during preeclampsia. *Br. J. Pharmacol.* 176 (12), 1914–1921. doi:10.1111/bph.14466

Cunningham, M. W., Jayaram, A., Deer, E., Amaral, L. M., Vaka, V. R., Ibrahim, T., et al. (2020). Tumor necrosis factor alpha (TNF-alpha) blockade improves natural killer cell (NK) activation, hypertension, and mitochondrial oxidative stress in a preclinical rat model of preeclampsia. *Hypertens. Pregnancy* 39 (4), 399–404. doi:10.1080/10641955. 2020.1793999

Cunningham, M. W., Jr., Castillo, J., Ibrahim, T., Cornelius, D. C., Campbell, N., Amaral, L., et al. (2018). AT1-AA (angiotensin II type 1 receptor agonistic autoantibody) blockade prevents preeclamptic symptoms in placental ischemic rats. *Hypertension* 71 (5), 886–893. doi:10.1161/HYPERTENSIONAHA.117.10681

Darmochwal-Kolarz, D., Rolinski, J., Tabarkiewicz, J., Leszczynska-Gorzelak, B., Buczkowski, J., Wojas, K., et al. (2003). Myeloid and lymphoid dendritic cells in normal pregnancy and pre-eclampsia. *Clin. Exp. Immunol.* 132 (2), 339–344. doi:10. 1046/j.1365-2249.2003.02136.x

Dechend, R., Viedt, C., Muller, D. N., Ugele, B., Brandes, R. P., Wallukat, G., et al. (2003). AT1 receptor agonistic antibodies from preeclamptic patients stimulate NADPH oxidase. *Circulation* 107 (12), 1632–1639. doi:10.1161/01.CIR.0000058200. 90059.B1

Deer, E., Reeve, K. E., Amaral, L., Vaka, V. R., Franks, M., Campbell, N., et al. (2021). CD4+ T cells cause renal and placental mitochondrial oxidative stress as mechanisms of hypertension in response to placental ischemia. *Am. J. Physiol. Ren. Physiol.* 320 (1), F47–F54. doi:10.1152/ajprenal.00398.2020

Deer, E., Herrock, O., Campbell, N., Cornelius, D., Fitzgerald, S., Amaral, L. M., et al. (2023). The role of immune cells and mediators in preeclampsia. *Nat. Rev. Nephrol.* doi:10.1038/s41581-022-00670-0

Didion, S. P., Kinzenbaw, D. A., Schrader, L. I., Chu, Y., and Faraci, F. M. (2009). Endogenous interleukin-10 inhibits angiotensin II-induced vascular dysfunction. *Hypertension* 54 (3), 619–624. doi:10.1161/HYPERTENSIONAHA.109.137158

Ding, H., Dai, Y., Lei, Y., Wang, Z., Liu, D., Li, R., et al. (2019). Upregulation of CD81 in trophoblasts induces an imbalance of Treg/Th17 cells by promoting IL-6 expression in preeclampsia. *Cell. Mol. Immunol.* 16 (1), 302–312. doi:10.1038/s41423-018-0186-9

Ditisheim, A., Sibai, B., and Tatevian, N. (2020). Placental findings in postpartum preeclampsia: A comparative retrospective study. *Am. J. Perinatol.* 37 (12), 1217–1222. doi:10.1055/s-0039-1692716

Eghbal-Fard, S., Yousefi, M., Heydarlou, H., Ahmadi, M., Taghavi, S., Movasaghpour, A., et al. (2019). The imbalance of Th17/Treg axis involved in the pathogenesis of preeclampsia. *J. Cell. Physiol.* 234 (4), 5106–5116. doi:10.1002/jcp.27315

El Shahaway, A. A., Abd Elhady, R. R., Abdelrhman, A. A., and Yahia, S. (2019). Role of maternal serum interleukin 17 in preeclampsia: Diagnosis and prognosis. *J. Inflamm. Res.* 12, 175–180. doi:10.2147/JIR.S206800

El-Azzamy, H., Dambaeva, S. V., Katukurundage, D., Salazar Garcia, M. D., Skariah, A., Hussein, Y., et al. (2018). Dysregulated uterine natural killer cells and vascular remodeling in women with recurrent pregnancy losses. *Am. J. Reprod. Immunol.* 80 (4), e13024. doi:10.1111/aji.13024

English, F. A., Kenny, L. C., and McCarthy, F. P. (2015). Risk factors and effective management of preeclampsia. *Integr. Blood Press Control* 8, 7–12. doi:10.2147/IBPC. S50641

Esposito, K., and Giugliano, D. (2004). The metabolic syndrome and inflammation: Association or causation? *Nutr. Metab. Cardiovasc Dis.* 14 (5), 228–232. doi:10.1016/s0939-4753(04)80048-6

Faas, M. M., and de Vos, P. (2017). Uterine NK cells and macrophages in pregnancy. *Placenta* 56, 44–52. doi:10.1016/j.placenta.2017.03.001

Faas, M. M., and De Vos, P. (2018). Innate immune cells in the placental bed in healthy pregnancy and preeclampsia. *Placenta* 69, 125–133. doi:10.1016/j.placenta. 2018.04.012

Faas, M. M., Spaans, F., and De Vos, P. (2014). Monocytes and macrophages in pregnancy and pre-eclampsia. *Front. Immunol.* 5, 298. doi:10.3389/fimmu.2014.00298

Fedotcheva, T. A., Fedotcheva, N. I., and Shimanovsky, N. L. (2022). Progesterone as an anti-inflammatory drug and immunomodulator: New aspects in hormonal regulation of the inflammation. *Biomolecules* 12 (9), 1299. doi:10.3390/biom12091299

Ferrante, C. J., Pinhal-Enfield, G., Elson, G., Cronstein, B. N., Hasko, G., Outram, S., et al. (2013). The adenosine-dependent angiogenic switch of macrophages to an M2-like phenotype is independent of interleukin-4 receptor alpha (IL-4R α) signaling. *Inflammation* 36 (4), 921–931. doi:10.1007/s10753-013-9621-3

Fillion, A., Guerby, P., Menzies, D., Lachance, C., Comeau, M. P., Bussieres, M. C., et al. (2021). Pathological investigation of placentas in preeclampsia (the PEARL study). *Hypertens. Pregnancy* 40 (1), 56–62. doi:10.1080/10641955.2020.1866008

Fox, R., Kitt, J., Leeson, P., Aye, C. Y. L., and Lewandowski, A. J. (2019). Preeclampsia: Risk factors, diagnosis, management, and the cardiovascular impact on the offspring. *J. Clin. Med.* 8 (10), 1625. doi:10.3390/jcm8101625

Gamliel, M., Goldman-Wohl, D., Isaacson, B., Gur, C., Stein, N., Yamin, R., et al. (2018). Trained memory of human uterine NK cells enhances their function in subsequent pregnancies. *Immunity* 48 (5), 951–962 e5. doi:10.1016/j.immuni.2018. 03.030

Gomez-Lopez, N., Olson, D. M., and Robertson, S. A. (2016). Interleukin-6 controls uterine Th9 cells and CD8(+) T regulatory cells to accelerate parturition in mice. *Immunol. Cell. Biol.* 94 (1), 79–89. doi:10.1038/icb.2015.63

Gomez-Lopez, N., Arenas-Hernandez, M., Romero, R., Miller, D., Garcia-Flores, V., Leng, Y., et al. (2020). Regulatory T cells play a role in a subset of idiopathic preterm labor/birth and adverse neonatal outcomes. *Cell. Rep.* 32 (1), 107874. doi:10.1016/j. celrep.2020.107874

Gough, M. J., Melcher, A. A., Ahmed, A., Crittenden, M. R., Riddle, D. S., Linardakis, E., et al. (2001). Macrophages orchestrate the immune response to tumor cell death. *Cancer Res.* 61 (19), 7240–7247. https://aacrjournals.org/cancerres

Gunnett, C. A., Heistad, D. D., and Faraci, F. M. (2002). Interleukin-10 protects nitric oxide-dependent relaxation during diabetes: Role of superoxide. *Diabetes* 51 (6), 1931–1937. doi:10.2337/diabetes.51.6.1931

Gustafsson, C., Mjosberg, J., Matussek, A., Geffers, R., Matthiesen, L., Berg, G., et al. (2008). Gene expression profiling of human decidual macrophages: Evidence for immunosuppressive phenotype. *PLoS One* 3 (4), e2078. doi:10.1371/journal.pone. 0002078

Gutierrez, G., Gentile, T., Miranda, S., and Margni, R. A. (2005). Asymmetric antibodies: A protective arm in pregnancy. *Chem. Immunol. Allergy* 89, 158–168. doi:10.1159/000087964

Hamilton, S., Oomomian, Y., Stephen, G., Shynlova, O., Tower, C. L., Garrod, A., et al. (2012). Macrophages infiltrate the human and rat decidua during term and preterm labor: Evidence that decidual inflammation precedes labor. *Biol. Reprod.* 86 (2), 39. doi:10.1095/biolreprod.111.095505

Harmon, A. C., Ibrahim, T., Cornelius, D. C., Amaral, L. M., Cunningham, M. W., Jr, Wallace, K., et al. (2019). Placental CD4(+) T cells isolated from preeclamptic women cause preeclampsia-like symptoms in pregnant nude-athymic rats. *Pregnancy Hypertens*. 15, 7–11. doi:10.1016/j.preghy.2018.10.007

Harrison, D. G., Coffman, T. M., and Wilcox, C. S. (2021). Pathophysiology of hypertension: The mosaic theory and beyond. *Circ. Res.* 128 (7), 847–863. doi:10.1161/CIRCRESAHA.121.318082

Hashemi, V., Dolati, S., Hosseini, A., Gharibi, T., Danaii, S., and Yousefi, M. (2017). Natural killer T cells in Preeclampsia: An updated review. *Biomed. Pharmacother.* 95, 412–418. doi:10.1016/j.biopha.2017.08.077

Hauspurg, A., and Jeyabalan, A. (2022). Postpartum preeclampsia or eclampsia: Defining its place and management among the hypertensive disorders of pregnancy. *Am. J. Obstet. Gynecol.* 226 (2S), S1211–S1221. doi:10.1016/j.ajog.2020.10.027

Helige, C., Ahammer, H., Moser, G., Hammer, A., Dohr, G., Huppertz, B., et al. (2014). Distribution of decidual natural killer cells and macrophages in the neighbourhood of the trophoblast invasion front: A quantitative evaluation. *Hum. Reprod.* 29 (1), 8–17. doi:10.1093/humrep/det353

Herrock, O. T., Deer, E., Amaral, L. M., Campbell, N., Lemon, J., Ingram, N., et al. (2022). B2 cells contribute to the pathophysiology of hypertension in response to placental ischemia. *Am. J. Physiol. Ren. Physiol.* 324, F179–F192. doi:10.1152/ajprenal. 00190.2022

Hoedjes, M., Berks, D., Vogel, I., Franx, A., Visser, W., Duvekot, J. J., et al. (2011). Symptoms of post-traumatic stress after preeclampsia. J. Psychosom. Obstet. Gynaecol. 32 (3), 126–134. doi:10.3109/0167482X.2011.599460 Houser, B. L., Tilburgs, T., Hill, J., Nicotra, M. L., and Strominger, J. L. (2011). Two unique human decidual macrophage populations. *J. Immunol.* 186 (4), 2633–2642. doi:10.4049/jimmunol.1003153

Huang, S. J., Chen, C. P., Schatz, F., Rahman, M., Abrahams, V. M., and Lockwood, C. J. (2008). Pre-eclampsia is associated with dendritic cell recruitment into the uterine decidua. *J. Pathol.* 214 (3), 328–336. doi:10.1002/path.2257

Huang, N., Chi, H., and Qiao, J. (2020). Role of regulatory T cells in regulating fetalmaternal immune tolerance in healthy pregnancies and reproductive diseases. *Front. Immunol.* 11, 1023. doi:10.3389/fimmu.2020.01023

Huang, X., Wang, L., Zhao, S., Liu, H., Chen, S., Wu, L., et al. (2021). Pregnancy induces an immunological memory characterized by maternal immune alterations through specific genes methylation. *Front. Immunol.* 12, 686676. doi:10.3389/fimmu. 2021.686676

Hudic, I., Szekeres-Bartho, J., Stray-Pedersen, B., Fatušić, Z., Polgar, B., and Ećim-Zlojutro, V. (2016). Lower urinary and serum progesterone-induced blocking factor in women with preterm birth. *J. Reprod. Immunol.* 117, 66–69. doi:10.1016/j.jri.2016. 07.003

Irani, R. A., and Xia, Y. (2011). Renin angiotensin signaling in normal pregnancy and preeclampsia. *Semin. Nephrol.* 31 (1), 47–58. doi:10.1016/j.semnephrol.2010.10.005

Irani, R. A., Zhang, Y., Zhou, C. C., Blackwell, S. C., Hicks, M. J., Ramin, S. M., et al. (2010). Autoantibody-mediated angiotensin receptor activation contributes to preeclampsia through tumor necrosis factor-alpha signaling. *Hypertension* 55 (5), 1246–1253. doi:10.1161/HYPERTENSIONAHA.110.150540

Jaiswal, N., Puri, M., Agarwal, K., Singh, S., Yadav, R., Tiwary, N., et al. (2021). COVID-19 as an independent risk factor for subclinical placental dysfunction. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 259, 7–11. doi:10.1016/j.ejogrb.2021.01.049

Jamieson, D. J., and Rasmussen, S. A. (2022). An update on COVID-19 and pregnancy. Am. J. Obstet. Gynecol. 226 (2), 177-186. doi:10.1016/j.ajog.2021.08.054

Jarmund, A. H., Giskeodegard, G. F., Ryssdal, M., Steinkjer, B., Stokkeland, L. M. T., Madssen, T. S., et al. (2021). Cytokine patterns in maternal serum from first trimester to term and beyond. *Front. Immunol.* 12, 752660. doi:10.3389/fimmu.2021.752660

Jasper, M. J., Tremellen, K. P., and Robertson, S. A. (2006). Primary unexplained infertility is associated with reduced expression of the T-regulatory cell transcription factor Foxp3 in endometrial tissue. *Mol. Hum. Reprod.* 12 (5), 301–308. doi:10.1093/molehr/gal032

Jayaram, A., Deer, E., Amaral, L. M., Campbell, N., Vaka, V. R., Cunningham, M., et al. (2021). The role of tumor necrosis factor in triggering activation of natural killer cell, multi-organ mitochondrial dysfunction and hypertension during pregnancy. *Pregnancy Hypertens.* 24, 65–72. doi:10.1016/j.preghy.2021.02.006

Jena, M. K., Nayak, N., Chen, K., and Nayak, N. R. (2019). Role of macrophages in pregnancy and related complications. *Arch. Immunol. Ther. Exp. Warsz.* 67 (5), 295–309. doi:10.1007/s00005-019-00552-7

Jensen, F., Wallukat, G., Herse, F., Budner, O., El-Mousleh, T., Costa, S. D., et al. (2012). CD19+CD5+ cells as indicators of preeclampsia. *Hypertension* 59 (4), 861–868. doi:10.1161/HYPERTENSIONAHA.111.188276

Jetten, N., Verbruggen, S., Gijbels, M. J., Post, M. J., De Winther, M. P. J., and Donners, M. M. P. C. (2014). Anti-inflammatory M2, but not pro-inflammatory M1 macrophages promote angiogenesis *in vivo*. *Angiogenesis* 17 (1), 109–118. doi:10.1007/s10456-013-9381-6

Juttukonda, L. J., Wachman, E. M., Boateng, J., Jain, M., Benarroch, Y., and Taglauer, E. S. (2022). Decidual immune response following COVID-19 during pregnancy varies by timing of maternal SARS-CoV-2 infection. *J. Reprod. Immunol.* 151, 103501. doi:10. 1016/j.jri.2022.103501

Kahn, D. A., and Baltimore, D. (2010). Pregnancy induces a fetal antigen-specific maternal T regulatory cell response that contributes to tolerance. *Proc. Natl. Acad. Sci. U. S. A.* 107 (20), 9299–9304. doi:10.1073/pnas.1003909107

Kajantie, E., Eriksson, J. G., Osmond, C., Thornburg, K., and Barker, D. J. P. (2009). Pre-eclampsia is associated with increased risk of stroke in the adult offspring: The helsinki birth cohort study. *Stroke* 40 (4), 1176–1180. doi:10.1161/STROKEAHA.108. 538025

Karrar, S. A., and Hong, P. L. (2022). Preeclampsia. Treasure Island (FL): StatPearls.

Katabuchi, H., Yih, S., Ohba, T., Matsui, K., Takahashi, K., Takeya, M., et al. (2003). Characterization of macrophages in the decidual atherotic spiral artery with special reference to the cytology of foam cells. *Med. Electron Microsc.* 36 (4), 253–262. doi:10. 1007/s00795-003-0223-2

Khanna, D., Khanna, S., Khanna, P., Kahar, P., and Patel, B. M. (2022). Obesity: A chronic low-grade inflammation and its markers. *Cureus* 14 (2), e22711. doi:10.7759/ cureus.22711

Kieffer, T. E., Faas, M. M., Scherjon, S. A., and Prins, J. R. (2017). Pregnancy persistently affects memory T cell populations. *J. Reprod. Immunol.* 119, 1–8. doi:10. 1016/j.jri.2016.11.004

Kieffer, T. E. C., Scherjon, S. A., Faas, M. M., and Prins, J. R. (2019). Lower activation of CD4(+) memory T cells in preeclampsia compared to healthy pregnancies persists postpartum. *J. Reprod. Immunol.* 136, 102613. doi:10.1016/j.jri.2019.102613

Koopman, L. A., Kopcow, H. D., Rybalov, B., Boyson, J. E., Orange, J. S., Schatz, F., et al. (2003). Human decidual natural killer cells are a unique NK cell subset with immunomodulatory potential. *J. Exp. Med.* 198 (8), 1201–1212. doi:10.1084/jem. 20030305

LaMarca, B. B., Cockrell, K., Sullivan, E., Bennett, W., and Granger, J. P. (2005). Role of endothelin in mediating tumor necrosis factor-induced hypertension in pregnant rats. *Hypertension* 46 (1), 82–86. doi:10.1161/01.HYP.0000169152.59854.36

LaMarca, B., Wallukat, G., Llinas, M., Herse, F., Dechend, R., and Granger, J. P. (2008). Autoantibodies to the angiotensin type I receptor in response to placental ischemia and tumor necrosis factor alpha in pregnant rats. *Hypertension* 52 (6), 1168–1172. doi:10.1161/HYPERTENSIONAHA.108.120576

LaMarca, B., Speed, J., Fournier, L., Babcock, S. A., Berry, H., Cockrell, K., et al. (2008). Hypertension in response to chronic reductions in uterine perfusion in pregnant rats: Effect of tumor necrosis factor-alpha blockade. *Hypertension* 52 (6), 1161–1167. doi:10. 1161/HYPERTENSIONAHA.108.120881

LaMarca, B., Cornelius, D. C., Harmon, A. C., Amaral, L. M., Cunningham, M. W., Faulkner, J. L., et al. (2016). Identifying immune mechanisms mediating the hypertension during preeclampsia. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 311 (1), R1–R9. doi:10.1152/ajpregu.00052.2016

Lazarski, C. A., Ford, J., Katzman, S. D., Rosenberg, A. F., and Fowell, D. J. (2013). IL-4 attenuates Th1-associated chemokine expression and Th1 trafficking to inflamed tissues and limits pathogen clearance. *PLoS One* 8 (8), e71949. doi:10.1371/journal.pone. 0071949

Leimert, K. B., Xu, W., Princ, M. M., Chemtob, S., and Olson, D. M. (2021). Inflammatory amplification: A central tenet of uterine transition for labor. *Front. Cell. Infect. Microbiol.* 11, 660983. doi:10.3389/fcimb.2021.660983

Li, X. L., Chen, T. T., Dong, X., Gou, W. L., Lau, S., Stone, P., et al. (2014). Early onset preeclampsia in subsequent pregnancies correlates with early onset preeclampsia in first pregnancy. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 177, 94–99. doi:10.1016/j.ejogrb.2014. 03.043

Li, J., Huang, L., Wang, S., and Zhang, Z. (2019). The prevalence of regulatory T and dendritic cells is altered in peripheral blood of women with pre-eclampsia. *Pregnancy Hypertens.* 17, 233–240. doi:10.1016/j.preghy.2019.07.003

Li, M., Sun, F., Xu, Y., Chen, L., Chen, C., Cui, L., et al. (2022). Tim-3⁺ decidual Møs induced Th2 and Treg bias in decidual CD4⁺T cells and promoted pregnancy maintenance via CD132. *Cell. Death Dis.* 13 (5), 454. doi:10.1038/s41419-022-04899-2

Lokki, A. I., Heikkinen-Eloranta, J. K., and Laivuori, H. (2018). The immunogenetic conundrum of preeclampsia. *Front. Immunol.* 9, 2630. doi:10. 3389/fimmu.2018.02630

Madazli, R., Yuksel, M. A., Imamoglu, M., Tuten, A., Oncul, M., Aydin, B., et al. (2014). Comparison of clinical and perinatal outcomes in early- and late-onset preeclampsia. Arch. Gynecol. Obstet. 290 (1), 53–57. doi:10.1007/s00404-014-3176-x

Mahajan, V. S., Mattoo, H., Sun, N., Viswanadham, V., Yuen, G. J., Allard-Chamard, H., et al. (2021). B1a and B2 cells are characterized by distinct CpG modification states at DNMT3A-maintained enhancers. *Nat. Commun.* 12 (1), 2208. doi:10.1038/s41467-021-22458-9

Mendoza-Cabrera, M. I., Navarro-Hernandez, R. E., Santerre, A., Ortiz-Lazareno, P. C., Pereira-Suarez, A. L., and Estrada-Chavez, C. (2020). Effect of pregnancy hormone mixtures on cytokine production and surface marker expression in naive and LPS-activated THP-1 differentiated monocytes/macrophages. *Innate Immun.* 26 (2), 84–96. doi:10.1177/1753425919864658

Mizuno, M., Aoki, K., and Kimbara, T. (1994). Functions of macrophages in human decidual tissue in early pregnancy. *Am. J. Reprod. Immunol.* 31 (4), 180–188. doi:10. 1111/j.1600-0897.1994.tb00865.x

Moore, K. W., de Waal Malefyt, R., Coffman, R. L., and O'Garra, A. (2001). Interleukin-10 and the interleukin-10 receptor. *Annu. Rev. Immunol.* 19, 683–765. doi:10.1146/annurev.immunol.19.1.683

Murray, E. J., Gumusoglu, S. B., Santillan, D. A., and Santillan, M. K. (2021). Manipulating CD4+ T cell pathways to prevent preeclampsia. *Front. Bioeng. Biotechnol.* 9, 811417. doi:10.3389/fbioe.2021.811417

Nankabirwa, V., Tumwine, J. K., Mugaba, P. M., Tylleskar, T., and Sommerfelt, H.PROMISE- EBF Study Group (2015). Child survival and BCG vaccination: A community based prospective cohort study in Uganda. *BMC Public Health* 15, 175. doi:10.1186/s12889-015-1497-8

Naruse, K., Lash, G. E., Innes, B. A., Otun, H. A., Searle, R. F., Robson, S. C., et al. (2009). Localization of matrix metalloproteinase (MMP)-2, MMP-9 and tissue inhibitors for MMPs (TIMPs) in uterine natural killer cells in early human pregnancy. *Hum. Reprod.* 24 (3), 553–561. doi:10.1093/humrep/den408

Need, J. A. (1975). Pre-eclampsia in pregnancies by different fathers: Immunological studies. *Br. Med. J.* 1 (5957), 548–549. doi:10.1136/bmj.1.5957.548

Negishi, Y., Wakabayashi, A., Shimizu, M., Ichikawa, T., Kumagai, Y., Takeshita, T., et al. (2012). Disruption of maternal immune balance maintained by innate DC subsets results in spontaneous pregnancy loss in mice. *Immunobiology* 217 (10), 951–961. doi:10.1016/j.imbio.2012.01.011

Netea, M. G., Joosten, L. A. B., Latz, E., Mills, K. H. G., Natoli, G., Stunnenberg, H. G., et al. (2016). Trained immunity: A program of innate immune memory in health and disease. *Science* 352 (6284), aaf1098. doi:10.1126/science.aaf1098

Nourollahpour Shiadeh, M., Behboodi Moghadam, Z., Adam, I., Saber, V., Bagheri, M., and Rostami, A. (2017). Human infectious diseases and risk of preeclampsia: An updated review of the literature. *Infection* 45 (5), 589–600. doi:10.1007/s15010-017-1031-2

Novakovic, B., Habibi, E., Wang, S. Y., Arts, R. J. W., Davar, R., Megchelenbrink, W., et al. (2016). β -Glucan reverses the epigenetic state of LPS-induced immunological tolerance. *Cell.* 167 (5), 1354–1368 e14. doi:10.1016/j.cell.2016.09.034

Opichka, M. A., Rappelt, M. W., Gutterman, D. D., Grobe, J. L., and McIntosh, J. J. (2021). Vascular dysfunction in preeclampsia. *Cells* 10 (11), 3055. doi:10.3390/ cells10113055

Palm, A. E., and Henry, C. (2019). Remembrance of things past: Long-term B cell memory after infection and vaccination. *Front. Immunol.* 10, 1787. doi:10.3389/fimmu. 2019.01787

Panda, B., Panda, A., Ueda, I., Abrahams, V. M., Norwitz, E. R., Stanic, A. K., et al. (2012). Dendritic cells in the circulation of women with preeclampsia demonstrate a pro-inflammatory bias secondary to dysregulation of TLR receptors. *J. Reprod. Immunol.* 94 (2), 210–215. doi:10.1016/j.jri.2012.01.008

Plaks, V., Birnberg, T., Berkutzki, T., Sela, S., BenYashar, A., Kalchenko, V., et al. (2008). Uterine DCs are crucial for decidua formation during embryo implantation in mice. *J. Clin. Invest.* 118 (12), 3954–3965. doi:10.1172/JCI36682

Polgar, B., Nagy, E., Miko, E., Varga, P., and Szekeres-Bartho, J. (2004). Urinary progesterone-induced blocking factor concentration is related to pregnancy outcome. *Biol. Reprod.* 71 (5), 1699–1705. doi:10.1095/biolreprod.104.030437

Qureshi, O. S., Zheng, Y., Nakamura, K., Attridge, K., Manzotti, C., Schmidt, E. M., et al. (2011). Trans-endocytosis of CD80 and CD86: A molecular basis for the cellextrinsic function of CTLA-4. *Science* 332 (6029), 600–603. doi:10.1126/science. 1202947

Rana, S., Lemoine, E., Granger, J. P., and Karumanchi, S. A. (2019). Preeclampsia: Pathophysiology, challenges, and perspectives. *Circ. Res.* 124 (7), 1094–1112. doi:10. 1161/CIRCRESAHA.118.313276

Raymond, D., and Peterson, E. (2011). A critical review of early-onset and late-onset preeclampsia. *Obstet. Gynecol. Surv.* 66 (8), 497–506. doi:10.1097/OGX. 0b013e3182331028

Reeve, K. E., Deer, E., Amaral, L. M., Cornelius, D. C., Herrock, O., Harmon, A. C., et al. (2022). Placental CD4+ T cells from preeclamptic patients cause autoantibodies to the angiotensin II type I receptor and hypertension in a pregnant rat model of preeclampsia. *Explor. Med.*, 99–111. doi:10.37349/emed.2022.00077

Reister, F., Frank, H. G., Heyl, W., Kosanke, G., Huppertz, B., Schroder, W., et al. (1999). The distribution of macrophages in spiral arteries of the placental bed in preeclampsia differs from that in healthy patients. *Placenta* 20 (2-3), 229–233. doi:10.1053/ plac.1998.0373

Rieber-Mohn, A. B., Sugulle, M., Wallukat, G., Alnaes-Katjavivi, P., Leite Storvold, G., Bolstad, N., et al. (2018). Auto-antibodies against the angiotensin II type I receptor in women with uteroplacental acute atherosis and preeclampsia at delivery and several years postpartum. *J. Reprod. Immunol.* 128, 23–29. doi:10.1016/j.jri.2018. 05.008

Robillard, P. Y., Dekker, G., Scioscia, M., Bonsante, F., Iacobelli, S., Boukerrou, M., et al. (2019). Increased BMI has a linear association with late-onset preeclampsia: A population-based study. *PLoS One* 14 (10), e0223888. doi:10.1371/journal.pone. 0223888

Robillard, P. Y., Dekker, G., Scioscia, M., and Saito, S. (2022). Progress in the understanding of the pathophysiology of immunologic maladaptation related to early-onset preeclampsia and metabolic syndrome related to late-onset preeclampsia. *Am. J. Obstet. Gynecol.* 226 (2S), S867–S875. doi:10.1016/j.ajog.2021.11.019

Rodriguez-Perez, A. I., Labandeira, C. M., Pedrosa, M. A., Valenzuela, R., Suarez-Quintanilla, J. A., Cortes-Ayaso, M., et al. (2021). Autoantibodies against ACE2 and angiotensin type-1 receptors increase severity of COVID-19. *J. Autoimmun.* 122, 102683. doi:10.1016/j.jaut.2021.102683

Romao-Veiga, M., Ribeiro, V. R., Matias, M. L., Nunes, P. R., Romagnoli, G. G., Peracoli, J. C., et al. (2022). DAMPs are able to skew CD4(+) T cell subsets and increase the inflammatory profile in pregnant women with preeclampsia. *J. Reprod. Immunol.* 149, 103470. doi:10.1016/j.jri.2021.103470

Roos, N. M., Wiegman, M. J., Jansonius, N. M., and Zeeman, G. G. (2012). Visual disturbances in (pre)eclampsia. *Obstet. Gynecol. Surv.* 67 (4), 242–250. doi:10.1097/OGX.0b013e318250a457

Rowe, J. H., Ertelt, J. M., Xin, L., and Way, S. S. (2012). Pregnancy imprints regulatory memory that sustains anergy to fetal antigen. *Nature* 490 (7418), 102–106. doi:10.1038/ nature11462

Saito, S., Sakai, M., Sasaki, Y., Tanebe, K., Tsuda, H., and Michimata, T. (1999). Quantitative analysis of peripheral blood Th0, Th1, Th2 and the Th1:Th2 cell ratio during normal human pregnancy and preeclampsia. *Clin. Exp. Immunol.* 117 (3), 550–555. doi:10.1046/j.1365-2249.1999.00997.x Sayad, B., Mohseni Afshar, Z., Mansouri, F., Salimi, M., Miladi, R., Rahimi, S., et al. (2022). Pregnancy, preeclampsia, and COVID-19: Susceptibility and mechanisms: A review study. *Int. J. Fertil. Steril.* 16 (2), 64–69. doi:10.22074/IJFS.2022.539768.1194

Schonkeren, D., van der Hoorn, M. L., Khedoe, P., Swings, G., van Beelen, E., Claas, F., et al. (2011). Differential distribution and phenotype of decidual macrophages in preeclamptic versus control pregnancies. *Am. J. Pathol.* 178 (2), 709–717. doi:10. 1016/j.ajpath.2010.10.011

Schwartz, D. A., Avvad-Portari, E., Babal, P., Baldewijns, M., Blomberg, M., Bouachba, A., et al. (2022). Placental tissue destruction and insufficiency from COVID-19 causes stillbirth and neonatal death from hypoxic-ischemic injury: A study of 68 cases with SARS-CoV-2 placentitis from 12 countries. *Arch. Pathol. Lab. Med.* 146, 660–676. doi:10.5858/arpa.2022-0029-SA

Seyed Hosseini, E., Riahi Kashani, N., Nikzad, H., Azadbakht, J., Hassani Bafrani, H., and Haddad Kashani, H. (2020). The novel coronavirus Disease-2019 (COVID-19): Mechanism of action, detection and recent therapeutic strategies. *Virology* 551, 1–9. doi:10.1016/j.virol.2020.08.011

Shao, Q., Liu, X., Huang, Y., Chen, X., and Wang, H. (2020). Human decidual stromal cells in early pregnancy induce functional Re-programming of monocyte-derived dendritic cells *via* crosstalk between G-CSF and IL-1β. *Front. Immunol.* 11, 574270. doi:10.3389/fimmu.2020.574270

Sheibak, N., Mahmoudzadeh-Sagheb, H., Moudi, B., and Heidari, Z. (2020). Elevated immunoexpression of interferon-gamma in placenta tissue samples from pregnancies complicated with preeclampsia compared to the placenta previa. *Pregnancy Hypertens.* 22, 175–180. doi:10.1016/j.preghy.2020.08.003

Shields, C. A., McCalmon, M., Ibrahim, T., White, D. L., Williams, J. M., LaMarca, B., et al. (2018). Placental ischemia-stimulated T-helper 17 cells induce preeclampsiaassociated cytolytic natural killer cells during pregnancy. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 315 (2), R336–R343. doi:10.1152/ajpregu.00061.2018

Shima, T., Inada, K., Nakashima, A., Ushijima, A., Ito, M., Yoshino, O., et al. (2015). Paternal antigen-specific proliferating regulatory T cells are increased in uterinedraining lymph nodes just before implantation and in pregnant uterus just after implantation by seminal plasma-priming in allogeneic mouse pregnancy. *J. Reprod. Immunol.* 108, 72–82. doi:10.1016/j.jiri.2015.02.005

Smith, S. D., Dunk, C. E., Aplin, J. D., Harris, L. K., and Jones, R. L. (2009). Evidence for immune cell involvement in decidual spiral arteriole remodeling in early human pregnancy. *Am. J. Pathol.* 174 (5), 1959–1971. doi:10.2353/ajpath. 2009.080995

Staff, A. C., Redman, C. W. G., Williams, D., Leeson, P., Moe, K., Thilaganathan, B., et al. (2016). Pregnancy and long-term maternal cardiovascular health: Progress through harmonization of research cohorts and biobanks. *Hypertension* 67 (2), 251–260. doi:10.1161/HYPERTENSIONAHA.115.06357

Staff, A. C., Fjeldstad, H. E., Fosheim, I. K., Moe, K., Turowski, G., Johnsen, G. M., et al. (2022). Failure of physiological transformation and spiral artery atherosis: Their roles in preeclampsia. *Am. J. Obstet. Gynecol.* 226 (2S), S895–S906. doi:10.1016/j.ajog. 2020.09.026

Stefanska, K., Zieliński, M., Jankowiak, M., Zamkowska, D., Sakowska, J., Adamski, P., et al. (2021). Cytokine imprint in preeclampsia. *Front. Immunol.* 12, 667841. doi:10. 3389/fimmu.2021.667841

Stelzer, I. A., Ghaemi, M. S., Han, X., Ando, K., Hedou, J. J., Feyaerts, D., et al. (2021). Integrated trajectories of the maternal metabolome, proteome, and immunome predict labor onset. *Sci. Transl. Med.* 13 (592), eabd9898. doi:10.1126/scitranslmed.abd9898

Sun, P., Lu, X., Xu, C., Sun, W., and Pan, B. (2020). Understanding of COVID-19 based on current evidence. J. Med. Virol. 92 (6), 548–551. doi:10.1002/jmv.25722

Tamas, P., Kovacs, K., Varnagy, A., Farkas, B., Alemu Wami, G., and Bodis, J. (2022). Preeclampsia subtypes: Clinical aspects regarding pathogenesis, signs, and management with special attention to diuretic administration. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 274, 175–181. doi:10.1016/j.ejogrb.2022.05.033

Thiruchelvam, U., Dransfield, I., Saunders, P. T. K., and Critchley, H. O. D. (2013). The importance of the macrophage within the human endometrium. *J. Leukoc. Biol.* 93 (2), 217–225. doi:10.1189/jlb.0712327

Tilburgs, T., Roelen, D. L., van der Mast, B. J., de Groot-Swings, G. M., Kleijburg, C., Scherjon, S. A., et al. (2008). Evidence for a selective migration of fetus-specific CD4+CD25bright regulatory T cells from the peripheral blood to the decidua in human pregnancy. *J. Immunol.* 180 (8), 5737–5745. doi:10.4049/jimmunol.180.8. 5737

Todt, J. C., Yang, Y., Lei, J., Lauria, M. R., Sorokin, Y., Cotton, D. B., et al. (1996). Effects of tumor necrosis factor-alpha on human trophoblast cell adhesion and motility. *Am. J. Reprod. Immunol.* 36 (2), 65–71. doi:10.1111/j.1600-0897.1996.tb00141.x

Torchinsky, A., Shepshelovich, J., Orenstein, H., Zaslavsky, Z., Savion, S., Carp, H., et al. (2003). TNF-alpha protects embryos exposed to developmental toxicants. *Am. J. Reprod. Immunol.* 49 (3), 159–168. doi:10.1034/j.1600-0897.2003.01174.x

Touwslager, R. N., Houben, A. J. H. M., Gielen, M., Zeegers, M. P., Stehouwer, C. D. A., Zimmermann, L. J., et al. (2012). Endothelial vasodilatation in newborns is related to body size and maternal hypertension. *J. Hypertens.* 30 (1), 124–131. doi:10.1097/HJH. 0b013e32834d75c6

Travis, O. K., White, D., Pierce, W. A., Ge, Y., Stubbs, C. Y., Spradley, F. T., et al. (2019). Chronic infusion of interleukin-17 promotes hypertension, activation of cytolytic natural killer cells, and vascular dysfunction in pregnant rats. *Physiol. Rep.* 7 (7), e14038. doi:10.14814/phy2.14038

Travis, O. K., White, D., Baik, C., Giachelli, C., Thompson, W., Stubbs, C., et al. (2020). Interleukin-17 signaling mediates cytolytic natural killer cell activation in response to placental ischemia. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 318 (6), R1036–R1046. doi:10.1152/ajpregu.00285.2019

Travis, O. K., Tardo, G. A., Giachelli, C., Siddiq, S., Nguyen, H. T., Crosby, M. T., et al. (2021). Tumor necrosis factor-alpha blockade improves uterine artery resistance, maternal blood pressure, and fetal growth in placental ischemic rats. *Pregnancy Hypertens.* 25, 39–47. doi:10.1016/j.preghy.2021.05.002

Tsuda, S., Zhang, X., Hamana, H., Shima, T., Ushijima, A., Tsuda, K., et al. (2018). Clonally expanded decidual effector regulatory T cells increase in late gestation of normal pregnancy, but not in preeclampsia, in humans. *Front. Immunol.* 9, 1934. doi:10. 3389/fmmu.2018.01934

van Rijn, B. B., Bruinse, H. W., Veerbeek, J. H., Post Uiterweer, E. D., Koenen, S. V., van der Bom, J. G., et al. (2016). Postpartum circulating markers of inflammation and the systemic acute-phase response after early-onset preeclampsia. *Hypertension* 67 (2), 404–414. doi:10.1161/HYPERTENSIONAHA.115.06455

Veerbeek, J. H., Hermes, W., Breimer, A. Y., van Rijn, B. B., Koenen, S. V., Mol, B. W., et al. (2015). Cardiovascular disease risk factors after early-onset preeclampsia, lateonset preeclampsia, and pregnancy-induced hypertension. *Hypertension* 65 (3), 600–606. doi:10.1161/HYPERTENSIONAHA.114.04850

Villar, J., Ariff, S., Gunier, R. B., Thiruvengadam, R., Rauch, S., Kholin, A., et al. (2021). Maternal and neonatal morbidity and mortality among pregnant women with and without COVID-19 infection: The INTERCOVID multinational cohort study. *JAMA Pediatr.* 175 (8), 817–826. doi:10.1001/jamapediatrics.2021.1050

Vitoratos, N., Economou, E., Iavazzo, C., Panoulis, K., and Creatsas, G. (2010). Maternal serum levels of TNF-alpha and IL-6 long after delivery in preeclamptic and normotensive pregnant women. *Mediat. Inflamm.* 2010, 908649. doi:10.1155/2010/ 908649

Wadhwani, P., Saha, P. K., Kalra, J. K., Gainder, S., and Sundaram, V. (2020). A study to compare maternal and perinatal outcome in early vs. late onset preeclampsia. *Obstet. Gynecol. Sci.* 63 (3), 270–277. doi:10.5468/ogs.2020.63.3.270

Wallace, K., Cornelius, D. C., Scott, J., Heath, J., Moseley, J., Chatman, K., et al. (2014). CD4+ T cells are important mediators of oxidative stress that cause hypertension in response to placental ischemia. *Hypertension* 64 (5), 1151–1158. doi:10.1161/ HYPERTENSIONAHA.114.03590

Wallukat, G., Homuth, V., Fischer, T., Lindschau, C., Horstkamp, B., Jupner, A., et al. (1999). Patients with preeclampsia develop agonistic autoantibodies against the angiotensin AT1 receptor. *J. Clin. Invest.* 103 (7), 945–952. doi:10.1172/JCI4106

Wang, J., Su, L., Zhu, T., and Shen, M. (2013). Changes in the subsets of dendritic cells and T cells in peripheral blood of patients with preeclampsia. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi* 29 (1), 72–75. Wang, W., Sung, N., Gilman-Sachs, A., and Kwak-Kim, J. (2020). T helper (Th) cell profiles in pregnancy and recurrent pregnancy losses: Th1/Th2/Th9/Th17/Th22/Tfh cells. *Front. Immunol.* 11, 2025. doi:10.3389/fimmu.2020.02025

Wei, S. Q., Bilodeau-Bertrand, M., Liu, S., and Auger, N. (2021). The impact of COVID-19 on pregnancy outcomes: A systematic review and meta-analysis. *CMAJ* 193 (16), E540–E548. doi:10.1503/cmaj.202604

Wu, P., Haththotuwa, R., Kwok, C. S., Babu, A., Kotronias, R. A., Rushton, C., et al. (2017). Preeclampsia and future cardiovascular health: A systematic review and metaanalysis. *Circ. Cardiovasc Qual. Outcomes* 10 (2), e003497. doi:10.1161/ CIRCOUTCOMES.116.003497

Xia, Y., and Kellems, R. E. (2011). Receptor-activating autoantibodies and disease: Preeclampsia and beyond. *Expert Rev. Clin. Immunol.* 7 (5), 659–674. doi:10.1586/eci. 11.56

Xie, X., He, H., Colonna, M., Seya, T., Takai, T., and Croy, B. A. (2005). Pathways participating in activation of mouse uterine natural killer cells during pregnancy. *Biol. Reprod.* 73 (3), 510–518. doi:10.1095/biolreprod.104.033951

Yeap, W. H., Wong, K. L., Shimasaki, N., Teo, E. C. Y., Quek, J. K. S., Yong, H. X., et al. (2017). Corrigendum: CD16 is indispensable for antibody-dependent cellular cytotoxicity by human monocytes. *Sci. Rep.* 7, 46202. doi:10.1038/srep46202

Zenclussen, A. C. (2006). A novel mouse model for preeclampsia by transferring activated th1 cells into normal pregnant mice. *Methods Mol. Med.* 122, 401–412. doi:10. 1385/1-59259-989-3:401

Zenclussen, A. C., Gentile, T., Kortebani, G., Mazzolli, A., and Margni, R. (2001). Asymmetric antibodies and pregnancy. *Am. J. Reprod. Immunol.* 45 (5), 289–294. doi:10.1111/j.8755-8920.2001.450504.x

Zeng, B., Kwak-Kim, J., Liu, Y., and Liao, A. H. (2013). Treg cells are negatively correlated with increased memory B cells in pre-eclampsia while maintaining suppressive function on autologous B-cell proliferation. *Am. J. Reprod. Immunol.* 70 (6), 454–463. doi:10.1111/aji.12154

Zhai, R., Liu, Y., Tong, J., Yu, Y., Yang, L., Gu, Y., et al. (2022). Empagliflozin ameliorates preeclampsia and reduces postpartum susceptibility to adriamycin in a mouse model induced by angiotensin receptor agonistic autoantibodies. *Front. Pharmacol.* 13, 826792. doi:10.3389/fphar.2022.826792

Zhang, P. (2018). Decidual vasculopathy in preeclampsia and spiral artery remodeling revisited: Shallow invasion versus failure of involution. *AJP Rep.* 8 (4), e241–e246. doi:10.1055/s-0038-1675348

Zhang, W., Zhou, Y., and Ding, Y. (2017). Lnc-DC mediates the over-maturation of decidual dendritic cells and induces the increase in Th1 cells in preeclampsia. *Am. J. Reprod. Immunol.* 77 (6), e12647. doi:10.1111/aji.12647

Zissler, U. M., Chaker, A. M., Effner, R., Ulrich, M., Guerth, F., Piontek, G., et al. (2016). Interleukin-4 and interferon-gamma orchestrate an epithelial polarization in the airways. *Mucosal Immunol.* 9 (4), 917–926. doi:10.1038/mi.2015.110

Zundler, S., and Neurath, M. F. (2015). Interleukin-12: Functional activities and implications for disease. *Cytokine Growth Factor Rev.* 26 (5), 559–568. doi:10.1016/j. cytogfr.2015.07.003