Check for updates

OPEN ACCESS

EDITED BY Krystyna Pierzchala-Koziec, University of Agriculture in Krakow, Poland

REVIEWED BY Keziban Atacan, Sakarya University of Applied Sciences, Turkey

*CORRESPONDENCE

Vladimir G. Vertiprakhov, vertiprakhov63@mail.ru Natalya V. Ovchinnikova, natalya.090@mail.ru

SPECIALTY SECTION

This article was submitted to Avian Physiology, a section of the journal Frontiers in Physiology

RECEIVED 12 February 2022 ACCEPTED 13 October 2022 PUBLISHED 25 October 2022

CITATION

Vertiprakhov VG and Ovchinnikova NV (2022), The activity of trypsin in the pancreatic juice and blood of poultry increases simultaneously in the postprandial period. *Front. Physiol.* 13:874664. doi: 10.3389/fphys.2022.874664

COPYRIGHT

© 2022 Vertiprakhov and Ovchinnikova. This is an open-access article distributed under the terms of the Creative Commons Attribution ticense (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original

author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The activity of trypsin in the pancreatic juice and blood of poultry increases simultaneously in the postprandial period

Vladimir G. Vertiprakhov¹* and Natalya V. Ovchinnikova (D)^{1,2}*

¹Timiryazev Russian State Agrarian University Moscow Agrarian Academy, Moscow, Russia, ²Physiology of Motivation Laboratory, Anokhin Research Institute of Normal Physiology, Moscow, Russia

Modern literature data indicate that the role of trypsin goes far beyond its digestive function. Once in the blood, trypsin is involved as part of the kallikreinkinin system in the regulation of blood pressure, regulates pancreatic function by activating PAR receptors, and influences inflammation and immunity in the cell. The interaction of trypsin in the intestine and serum in the living healthy organism has been insufficiently studied. On the basis of our own studies and literature data, we concluded that after overnight fasting the increase of trypsin activity in pancreatic juice and blood serum in the postprandial period occurs in parallel, which determines not only digestion of food protein but also the level of metabolism. Consequently, determining the optimal amount of crude protein in the diet during the morning meal is a paramount task for physiologists.

KEYWORDS

trypsin, pancreatic juice, broiler chickens, adaptation to diet composition, postprandial secretion

Introduction

Trypsin is secreted by the pancreas as trypsinogen, is activated by enterokinase in the duodenum, and hydrolyzes protein molecules to peptides and amino acids. Trypsin is an optimal marker for detecting changes in the physiological state of the pancreas because it is specific for this organ. It has been established that trypsin administration into the blood decreases enzymes output with pancreatic juice, and on the contrary, trypsin inhibitor administration is accompanied by enzymes secretion increase (Sheida et al., 2022).

The role of trypsin goes far beyond its digestive function. It was found that trypsin is involved in the activation of kallikrein, which regulates blood pressure (Chia et al., 2011). In addition, trypsin is an activator of PAR-receptors, which provide transmission of information to the cell during inflammatory processes and immunological reactions (Yarovaya et al., 2009). PAR2 activators have been shown to affect the pancreas and regulate the secretory function of the pancreas, stomach and salivary glands. There is evidence that PAR receptors activated directly by trypsin indicate a role for the enzyme in the pathogenesis of neurodegenerative diseases of the brain (Luo et al., 2007; Wang et al., 2008; Lohman et al., 2009; Almonte et al., 2013). Unlike other members of the PAR family,

which are effectively activated by thrombin, PAR2 represents a class of receptors activated by trypsin (Nieman, 2016).

Knowledge of the mechanisms of trypsin involvement in the norm and in the development of pathological processes is relevant because of pancreatic diseases, as there is a growing trend in the frequency of this disease worldwide (Forsmark, 2013; De la Iglesia-Garcia et al., 2018; Zhang et al., 2018). Knowledge of the interaction between trypsin-like enzymes and alpha-1proteinase inhibitor (α 1-PI) is particularly relevant because the role of α 1-PI in regulating viral tropism to human cells has been established. Patients with genetic deficiency of α 1-PI show increased susceptibility to SARS-CoV-2 virus and their infection has the most unfavorable form (Akbasheva et al., 2022).

Based on the literature and our own data, we showed pancreatic regulation during food intake and the role of trypsin as a trigger mechanism in metabolic regulation by examining trypsin activity in the duodenum and serum in poultry during the postprandial period.

Opinion is pancreatic secretion parallel or non-parallel?

In the world gastroenterological literature, the intensity of discussions about the expedient, adaptive secretion of the major digestive glands has not faded for more than a century. From our point of view, academician I.P. Pavlov's idea of the adaptive secretion of digestive enzymes, born in his laboratory the century before last, has lost its relevance in its original formulation and new research using modern approaches are required. For more than 70 years, it has been persistently studied exclusively at the organ level.

Most researchers adhere to the idea of I.P. Pavlov that pancreatic enzymes are secreted based on diet composition (Pavlov, 1951). The results of research in the second half of the 20th century supplemented and developed the theory of adaptation of pancreatic secretion to diet composition (Ugolev, 1961; Klimov and Fokina, 1987; Case, 1998; Korotko, 2005).

The situation changed radically after the results obtained by Palade, (1975) in the study of the processes of protein biosynthesis in the pancreatic cell. This work was awarded the Nobel Prize in 1974 (Palade, 1975). The authors found that zymogen granules contain all the enzymes produced by the gland. It was shown that all acinar cells of the pancreas are homogeneous, all of them, according to the same mechanisms of biosynthesis of any protein molecules, synthesize in one ratio the entire set of enzymes intended for export. In the process of ribosomal synthesis and subsequent intracellular transport, all these enzymes are completed, packed into separate secretory granules, and appear in the excretory pancreatic duct in this form. Each secretory granule has a strictly constant (depending on the type of animal and its age), unchanged composition in terms of the quantitative composition of all enzymes exported by the cell. The existing mechanisms that function according to the laws of biochemical logic do not leave the slightest possibility of influencing both the intensity of the biosynthesis of individual digestive enzymes and the rate of their secretion. Therefore, in contrast to Pavlov's adaptive, expedient theory of biosynthesis and secretion, Palade's hypothesis was called the theory of parallel secretion, i.e. strictly linked simultaneous and identical secretion of all enzymes synthesized and intended for export to the digestive tract.

At the same time, in the 70s of the last century, Russian scientists (Permyakovet al., 1973) established ultrastructural changes in acinar cells for each diet. With mixed or protein diet, changes in cell ultrastructures are similar and unfold cyclically in the cellular conveyor in full accordance with the biorhythm of acinar cells. A fat diet causes a restructuring of the acinar cell, affecting all its elements. There is hyperplasia and increased degranulation of the rough endoplasmic reticulum; transformation of rough vacuoles into smooth ones; hypertrophy of mitochondria and the Golgi complex. A carbohydrate diet is characterized by significant vacuolization of the endoplasmic reticulum with its subsequent reduction; decreased activity of the Golgi complex, mitochondrial hypertrophy. The physiological meaning of this structural rearrangement is the adaptation of the acinar cell to the predominant production of enzymes necessary for the processing of this type of food: lipases - with a fat diet, amylases - with a carbohydrate diet, proteases-with a protein diet (Di Magno and Layer, 1993; Case, 1998; Chandra and Liddle, 2015). It is the universality of the cellular "conveyor" of the acinar cell, adapted to the simultaneous synthesis of many enzymes, that allows it to be restructured when changing the diet. In addition, it was shown that there are peculiar bipolar islet-acinar cells in the pancreas, one-half of which, oriented towards the centroacinous duct, synthesizes zymogen granules, the other half, in close contact with the capillary network, produces granules similar in size, shape and degree of osmiophilia to those in endocrine cells. The emerging data on the presence of pancreatic enzymes in the blood have expanded the understanding of proteases role in metabolic processes in the body.

How enzymes are formed in the pancreas

It is known that the exosecretions of the digestive glands contain two pools of enzymes: newly synthesized and recreated. The rate of enzyme synthesis does not keep pace with exosecretion, which was shown when the enzyme secreting activity of the pancreas was taken into account (Rothman et al., 2002). Therefore, enzyme synthesis deficiency is compensated by their recreation. There are three ways enzymes enter the blood: endosecretion is the first way of transporting enzymes from glandulocytes to the interstitium, and then from it to the lymph and the bloodstream. The second way of transporting enzymes into the bloodstream is the enzymes resorption from the ducts of digestive glands (salivary, pancreatic and gastric) - "avoidance" of enzymes and the third way-enzymes enter the blood in the small intestine, mainly in the ileum (Korotko, 2006, 2007, 2011). Enzyme homeostasis is maintained by renal and extrarenal excretion of enzymes from the body, enzymes degradation by serine proteinases, and enzymes inactivation by specific inhibitors. The latter is relevant to serine proteinases - trypsin and chymotrypsin. Their main inhibitors in blood plasma are a1-proteinase inhibitor and $\alpha 2\text{-macroglobulin}.$ The former completely inactivates pancreatic proteinases, while the latter only limits their ability to break down high molecular weight proteins (Gőtze and Rothman, 1975; Liebow and Rothman, 1975; Heinrich et al., 1979; Korotko et al., 1986; Dosenko et al., 1999; Veremeenko et al., 2000, 2005). This complex has substrate specificity only for some low molecular weight proteins. It is not sensitive to other inhibitors of blood plasma proteinases, does not undergo autolysis, does not show any antigenic properties, but is recognized by cell receptors, and causes the formation of physiologically active substances in some cells (Dosenko et al., 1999).

Enzymes enter glandulocytes from the bloodstream, where they were transported by endosecretion, resorption from ducts reservoirs and small intestine, where they have a stimulating or inhibitory effect on enzymes secretion and, where they are recreated by glandulocytes together with their "own" enzymes. At this level of the secretory cycle, the signaling role of enzymes in the formation of the final enzyme spectrum of exosecretion is implemented using the principle of negative feedback at the level of the intracellular process, which was shown in experiments in vitro (Rothman, 1980). This principle is also used in the selfregulation of pancreatic secretion through reflex and paracrine mechanisms. Postprandially, portions of the secret deposited in the ducts are first transported into the cavity of the digestive tract, then portions of the secret with recreated enzymes, and, finally, the secret with recreated and newly synthesized enzymes is transported.

The concept of two enzyme pools of digestive glands exosecretions removes the question of the quantitative discrepancy between secreted and urgently synthesized enzymes by the digestive glands, since exosecretions always make up the sum of these two pools of enzymes. The ratios between the pools can change in the dynamics of exosecretion due to their different mobility in the postprandial period of glandular secretion. The recretory component of exosecretion is largely determined by the transport of enzymes into the bloodstream and the content of enzymes in it, changing in normal and pathological conditions. These processes are regulated by nervous and humoral mechanisms, apparently, with the participation of trypsin.

In the laboratory of Pavlov, it was shown that when fistula dogs lose pancreatic juice, hypersecretion of the gland is observed

at first, and intraduodenal administration of juice inhibits its secretion. This reverse or recurrent inhibition of secretion has been confirmed in experiments on rats, guinea pigs, cats, dogs, productive animals and humans (Korotko, 1992; Korotko and Voskanyan, 2001, 2005; Liddle, 2006). It was found that the effect of inhibition of pancreatic secretion is caused by intraduodenal introduction of proteases (trypsin, chymotrypsin and their zymogens), and this inhibition is removed by a trypsin inhibitor (Korotko and Baibekova, 1989). The specificity and high selectivity of the reverse inhibition of trypsin secretion was demonstrated in the experiments when an artificial trypsinspecific substrate, BAPNA (benzoyl-arginine-p-nitroanilide), was introduced into the duodenum, and it increased the activity of only trypsin (Korotko and Baibekova, 1989).

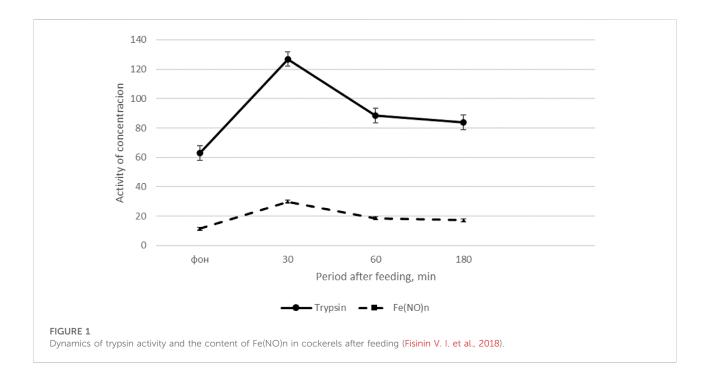
The results of the study on cockerels (Fisinin V. I. et al., 2018), presented in Figure 1 show that feed intake after a 14-h fast causes the most significant changes in trypsin activity. The activity of amylase and lipase in the blood plasma after feed intake does not change significantly. Moreover, the interval after feed intake is important: the greatest increase in trypsin activity in the blood is observed 30 min after feeding (by 66.0–67.1%) compared with the basal level of activity. 180 min after feeding trypsin activity in the blood decreases by 24.8% compared to trypsin activity 30 min after feed intake (see Figure 1).

On the x-axis - the period after feeding, min; on the y-axis, units of trypsin activity (U/L) and Fe(NO)n, μ M/l. The solid line is the dynamics of trypsin activity, the dotted line is the content of Fe(NO)n.

Thus, feed intake significantly increases trypsin activity and the content of NO donors in the blood plasma of roosters maximum 30 min after feeding, and then the indicators gradually decrease, approaching the initial level 180 min after feed intake.

In addition for the hypothesis, there is an inverse relationship between trypsin in the blood and pancreatic juice

The results of the experiment on poultry differ significantly from the data obtained on rats (Laporte and Tremolieres, 1971; Korotko and Baibekova, 1989), which showed an inverse relationship between trypsin in the blood and pancreatic juice. In experiments on chickens, when intravenous solutions of trypsin or pancreatic juice were administered to them, reverse inhibition and a decrease in the activity of duodenal proteases by 25.7–50.0% 30 min after injection were noted, which is apparently associated with the effect of trypsin on the control center of digestive system (medulla oblongata) (Vertiprakhov and Grozina, 2020). Therefore, an increase in trypsin activity in pancreatic juice and blood in the first 30 min after feeding can be explained by reflex effects on the secretory function of the pancreas, and subsequent regulation and adaptation to food composition is achieved by humoral factors, including participation of trypsin.



Analyzing the experimental data of pancreatic secretion in farm animals (Fisinin V. I. et al., 2018, 2019; Lebedev et al., 2019), it can be stated that in experiments with chronic pancreatic fistula, there are changes in pancreatic secretion during food intake, as well as those associated with the diet composition. The peculiar bipolar islet-acinar cells, which have a secretory and endocrine function, found during the ultrastructural analysis of the secretory cycle (Permyakov et al., 1973), suggest the possibility of simultaneous entry of trypsinogen both into the pancreatic duct and the bloodstream.

Discussion

The flow of appetitive pancreatic juice (rich in enzymes) into the duodenum begins immediately after a meal, at the same time the activity of trypsin in blood serum increases, and the function of the whole digestive conveyor throughout the day depends on this process. The work contributes to the issues of pancreatic secretion regulation by clarifying the hypothesis of inverse dependence of pancreatic enzymes on enzymes in the blood of animals by stating that in the postprandial phase the enzyme activity increases in parallel, simultaneously regulating the processes of hydrolysis of nutrients and their assimilation. Taking into account the formation of a conditioned reflex to individual components of the diet, the maximum level of assimilation of nutrients is achieved. Consequently, the morning meal is decisive in the daily diet and therefore further research should be aimed at studying the optimal level of protein in the morning diet.

Further study of protease stimulation during the morning meal, taking into account the role of trypsin as a hormone-like substance, suggests that this direction is promising for further research.

Author contributions

VV: article conception and preparation; NO: article conception and critical review; VV article supervision; and VV supervised the final form.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Akbasheva, O. E., Spirina, L. V., Dyakov, D. A., and Masunova, N. V. (2022). Proteolysis and α 1-proteinase inhibitor deficiency in SARS-CoV-2 infection. *Biomed. Chem.* 68 (3), 157–176. doi:10.18097/PBMC20226803157

Almonte, A. G., Qadri, L. H., Sultan, F. A., Watson, J. A., Mount, D. J., Rumbaugh, G., et al. (2013). Protease-activated receptor-1 modulates hippocampal memory formation and synaptic plasticity. *J. Neurochem.* 124 (1), 109–122. doi:10.1111/jnc.12075

Case, M. R. (1998). in *Pancreatic exocrine secretion: Mechanisms and control the pancreas/*. (Oxford: Blackwell Science Ltd), 63–100.

Chandra, R., and Liddle, R. A. (2015). Regulation of pancreatic secretion [electronic resource]//the pancreapedia : Exocrine pancreas knowledge base. doi:10.39.98/panc.2015.38

Chia, E., Kagota, S., Wijekoon, E. P., and McGuire, J. J. (2011). Protection of protease-activated receptor 2 mediated vasodilatation against angiotensin II-induced vascular dysfunction in mice. *BMC Pharmacol.* 28, 10. doi:10.1186/1471-2210-11-10

De la Iglesia-Garcia, D., Vallejo-Senra, N., Iglesias-Garcia, J., Lopez-Lopez, A., Nieto, L., and Dominguez-Munoz, J. E. (2018). Increased risk of mortality associated with pancreatic exocrine insufficiency in patients with chronic pancreatitis. *J. Clin. Gastroenterol.* 52 (8), e63–e72. doi:10.1097/MCG. 000000000000917

Di Magno, E. P., and Layer, P. (1993). "Human exocrine pancreatic enzyme secretion/E. P. Di Magno, P," in *Layer//the pancreas: Biology, pathobiology and diseases/*. Editor V. L. Go, et al. (New York: Raven), 275–300.

Dosenko, V. E., Veremeenko, K. N., and Kizim, A. I. (1999). Modern ideas about the mechanisms of proteolytic enzymes absorption in the gastrointestinal tract. *Probl. Med.* 7–8, 6–12.

Fisinin, V. I., Vertiprakhov, V. G., and Grozina, A. A. (2018b). The exocrine pancreatic function in chicken (Gallus gallus L.) fed diets containing different ingredients. S-h. Biol. 53 (4), 811–819. doi:10.15389/agrobiology.2018.4.811eng

Fisinin, V. I., Vertiprakhov, V. G., Kharitonov, E. L., and Grozina, A. A. (2019). Adaptation of pancreatic secretion and metabolism in animals with different types of digestion when replacing the protein component of the diet. *S-h. Biol.* 54 (6), 1122–1134. doi:10.15389/agrobiology.2019.6.1122eng

Fisinin, V. I., Vertiprakhov, V. G., Titov, V. Yu., and Grozina, A. A. (2018a). The postprandial dynamics of activity of the digestive enzymes and concentration of depot of nitric oxide in blood serum of cockerels. *Russ. J. physiology* 104, 976–983. doi:10.7868/S0869813918070080

Forsmark, C. E. (2013). Management of chronic pancreatitis. *Gastroenterology* 144, 1282–1291. doi:10.1053/j.gastro.2013.02.008

Gőtze, H., and Rothman, S. S. (1975). Enteropancreatic circulation of digestive enzyme as a conservation mechanism. *Nature* 257, 607–609. doi:10.1038/257607a0

Heinrich, H. C., Gabbe, E. E., Briiggeman, L., IcagicF.and ClassenM. (1979). Enteropancreatic circulation of trypsin in man. *Klin. Wochenschr.* 5723, 1295–1297. doi:10.1007/BF01492985

Klimov, P. K., and Fokina, A. A. (1987). *Physiology of the pancreas: Regulation of exocrine secretory function*. Leningrad: Nauka, 151.

Korotko, G. F., and Baibekova, G. D. (1989). Inhibition of pancreatic secretion by trypsin and trypsinogen. Fiziol. Zh. SSSR Im. I. M. Sechenova 74 (9), 1309–1315.

Korotko, G. F. (2007). Gastric digestion. Krasnodar: Publishing house Ltd B "Group B, 256.

Korotko, G. F., Kurzanov, A. N., and Lemeshkina, G. S. (1986). On the possibility of intestinal resorption of pancreatic hydrolases. Membrane digestion and absorption. Riga: Riga. Zinatne, 61–63.

Korotko, G. F. (2011). Recirculation of digestive enzymes. Krasnodar: Publishing House 'EDVI', 114.

Korotko, G. F. (2005). Secretion of the pancreas. 2nd add. Krasnodar: Publishing house Kub. Med. Univers., 312.

Korotko, G. F. (2006). Secretion of the salivary glands and elements of saliva diagnostics. Moscow: Publishing house "Academy of Natural History, 192.

Korotko, G. F. (1992). [The mechanism of the intestinal phase of pancreatic secretion]. Fiziol. Zh. SSSR Im. I. M. Sechenova 78 (9), 106–112.

Korotko, G. F., and Voskanyan, S. E. (2001). Regulation and self-regulation of pancreatic secretion. Adv. physiology 32 (4), 36–59.

Laporte, J. C., and Tremolieres, J. (1971). [Hormonal regulation of the enzymatic secretion of the exocrine pancreas]. C. R. Acad. Hebd. Seances Acad. Sci. D. 273, 1205–1207.

Lebedev, S. V., Sheida, E. V., Vertiprakhov, V. G., Gavrish, I. A., Kvan, O. V., Gubaidullina, I. Z., et al. (2019). A study of the exocrinous function of the cattle pancreas after the introduction of feed with a various protein source in rations. *Biosci. Res.* 16 (3), 2553–2562.

Liddle, R. A. (2006). in *Regulation of pancreatic secretion*. *Physiology of the gastrointestinal tract*. Editor L. R. Johnson. Fouth Edition (Academic Press is an imprint of Elsevier), 1397–1435. Chap. 55.

Liebow, C., and Rothman, S. (1975). Enteropancreatic circulation of digestive enzymes. *Science* 189, 472-474. doi:10.1126/science.1154022

Lohman, R. J., Jones, N. C., O'Brien, T. J., and Cocks, T. M. (2009). A regulatory role for protease- activated receptor-2 in motivational learning in rats. *Neurobiol. Learn. Mem.* 92 (3), 301–309. doi:10.1016/j.nlm.2009.03.010

Luo, W., Wang, Y., and Reiser, G. (2007). Protease-activated receptors in the brain: Receptor expression, activation, and functions in neurodegeneration and neuroprotection. *Brain Res. Rev.* 56 (2), 331–345. doi:10.1016/j.brainresrev.2007. 08.002

Nieman, M. T. (2016). Protease-activated receptors in hemostasis. Blood 128, 169-177. doi:10.1182/blood-2015-11-636472

Palade, G. (1975). Intracellular aspects of the process of protein synthesis. *Science* 189, 867. doi:10.1126/science.189.4206.867-b

Pavlov, I. P. (1951). Lectures on the work of the main digestive glands. Moscow, Leningrad, 11–215. Complete Works.

Permyakov, N. K., Podolsky, A. E., and Titova, G. P. (1973). Ultrastructural analysis of the pancreas secretory cycle. Moscow: Medicine, 239.

Rothman, S., Liebow, C., and Isenman, L. C. (2002). Conservation of digestive enzymes. *Physiol. Rev.* 82, 1–18. doi:10.1152/physrev.00022.2001

Rothman, S. S. (1980). Passage of proteins through membranes – old assumptions and new perspectives. *Am. J. Physiol.* 238, 391–402. doi:10.1152/ajpgi.1980.238.5. G391

Sheida, E. V., Lebedev, S. V., Miroshnikov, S. A., Grechkina, V. V., and Shoshina, O. V. (2022). Adaptive responses of the digestive system of cattle under the influence of dietary iron ultrafine particles in combination with fatty diets. *S-h. Biol.* 57 (2), 328–342. doi:10.15389/agrobiology.2022.2.328eng

Ugolev, A. M. (1961). Digestion and its adaptive evolution. Moscow: Higher School, 221p.

Veremeenko, K. N., Dosenko, V. E., Kizim, A. I., and Terzov, A. I. (2000). [The mechanisms of the curative action of systemic enzyme therapy]. Lik. Sprava 2, 3–11.

Veremeenko, K. N., Kizim, A. I., and Terzov, A. I. (2005). On the mechanisms of the therapeutic action of polyenzyme drugs. *art Heal*. 4 (20), 1.

Vertiprakhov, V. G., and Grozina, A. A. (2020). *Method for normalizing digestion in animals by administering parenteral pancreatic enzymes*. Patent for invention 2738930 C1, 12/18/2020. Application No. 2019139198 dated 12/03/2019.

Wang, Y., Luo, W., and Reiser, G. (2008). Trypsin and trypsin-like proteases in the brain: Proteolysis and cellular functions. *Cell. Mol. Life Sci.* 65 (2), 237–252. doi:10.1007/s00018-007-7288-3

Yarovaya, G. A., Blokhina, T. B., and Neshkova, E. A. (2009). Proteinase-activated receptors (PARs) are a signaling pathway initiated by limited proteolysis. *Lab. Med.* 10, 23–34.

Zhang, L., Sanagapalli, S., and Stoita, A. (2018). Challenges in diagnosis of pancreatic cancer. *World J. Gastroenterol.* 24 (19), 2047–2060. doi:10.3748/wjg.v24. i19.2047