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# Genetic insights into non-syndromic Tetralogy of Fallot

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Congenital heart defects (CHD) include structural abnormalities of the heart or/ and great vessels that are present at birth. CHD affects around 1% of all newborns worldwide. Tetralogy of Fallot (TOF) is the most prevalent cyanotic congenital cardiac abnormality, affecting three out of every 10,000 live infants with a prevalence rate of 5–10% of all congenital cardiac defects. The four hallmark characteristics of TOF are: right ventricular hypertrophy, pulmonary stenosis, ventricular septal defect, and overriding aorta. Approximately 20% of cases of TOF are associated with a known disease or chromosomal abnormality, with the remaining 80% of TOF cases being non-syndromic, with no known aetiology. Relatively few TOF patients have been studied, and little is known about critical causative genes for nonsyndromic TOF. However, rare genetic variants have been identified as significant risk factors for CHD, and are likely to cause some cases of TOF. Therefore, this review aims to provide an update on well-characterized genes and the most recent variants identified for non-syndromic TOF.

#### KEYWORDS

Tetralogy of Fallot, non-syndromic, genetics, congenital heart disease, NOTCH1, FLT4, transcription factors

# Introduction

Congenital heart defects (CHD) are structural malformations of the heart or/and great vessels which affect about 1% of live born infants (Hoffman and Kaplan, 2002). Globally, CHDs are the greatest cause of death from non-communicable diseases in individuals under 30 years of age, and are also the leading cause of mortality from congenital abnormalities (Su et al., 2022). In a study conducted between 2000 and 2005 in Europe, it was reported that approximately 88% of CHD cases are not associated with chromosomal alterations (Dolk et al., 2011), suggesting other genetic defects are responsible for the majority of CHD in this population. Within CHD, Tetralogy of Fallot (TOF) is the most common cyanotic congenital heart defect, affecting 3 per 10,000 live births, and comprising 5–10% of all congenital heart abnormalities (Suzuki et al., 1996; Bailliard and Anderson, 2009; Postoev et al., 2014); and (Park and Thayer, 2014). The hallmark characteristics of TOF include four structural alterations to the heart and great vessels



(Figure 1), which are: pulmonary stenosis, ventricular septal defect (VSD), overriding aorta, and right ventricular (RV) hypertrophy (Shinebourne et al., 2006; Bailliard and Anderson, 2009); and reviewed in (Bedair and Iriart, 2019).

### Pathophysiology of TOF

The TOF heart anatomy permits blood to be mixed between the pulmonary and systemic circulations. This mixture commonly happens at the VSD, where a right-to-left shunt introduces deoxygenated blood into the systemic circulation, resulting in cyanosis. The differential pressure gradient between the RV and LV determines the right-to-left shunt across the VSD. The degree of the right ventricular outflow tract obstruction (or pulmonary stenosis) determines the quantity of pulmonary blood flow (the RV stroke volume) (Wilson et al., 2019). Understanding the dynamical variables that aggravate or improve the right-to-left shunt is critical to understanding the essential care of disease or preoperative neonates with TOF (Wilson et al., 2019).

An irregular blood flow pattern, chamber hypoplasia, obstructive lesions and aberrant venous and arterial connections found in individuals with CHD results in some of the most interesting and complicated abnormalities in normal heart physiology (Škorić-Milosavljević et al., 2021). The physiological changes surrounding hyper cyanotic incidents consist of either a reduction in systemic vascular resistance or a rise in pulmonary resistance, both of which paticipate to a right-to-left shunt across the VDS (Mcleod et al., 2018; Ho et al., 2019); and (Senst et al., 2022). Due to the right ventricle to left ventricle flow shunt and high flow levels through the overriding aorta, the endocardium encompassing the VSD faces higher wall shear stresses. In addition, the TOF right ventricle and left ventricle have higher pressures than normal hearts, but only the TOF right ventricle has higher wall shear stresses and thickening (Wiputra et al., 2018). Understanding the blood flow dynamics in TOF hearts, most likely due to anatomical differences and their different illnesses, could aid in future minimally invasive foetal heart therapies.

#### TABLE 1 Non-syndromic TOF genetic variants.

Name of the gene	Variant or genetic alteration	Cardiac phenotype	Role to TOF	References
NOTCH1	p.R448X, p.W1638X and p.Q1733X p.G115fsX6, p.N147fsX128 p.C1322fsX121 (c.5385-1delC) p.G200V p.C292Y p.Q1495K, p.C1549Y p.N1875S and several other variants	TOF	These variants cause loss of protein function. Some are premature stop codons, single base pair deletions causing frameshifts and premature termination, or a single nucleotide deletion in a consensus splice site sequence	Page et al. (2019)
	p.C283G, p.Q415K, p.E606K, p.N623K, p.C682Y, p.D710N, p.C815Y, p.C1536R	TOF	Eight ultra-rare missense NOTCH1variants would support aberrant vascular development and related signalling as probable causes in TOF.	Manshaei et al. (2020)
FLT4	p.30RfsX3, p.R82X, T168RfsX18, Y361X, P363PfsX64, Q736X, L935PfsX82, P948PfsX54, Q999X	TOF and/or PS, APVS, RAA, PA	Loss of <i>FLT4</i> function in TOF with evidence of incomplete penetrance	Jin et al. (2017)
	p.Y361X, p.Y369X, p.E896X, p.Q920X, p.R1031X, p.Q1126X p.P363fsX25, p.Q423fsX3, p.L636fsX3, p.Y853fsX20, p.N905fsX20 and p.Y1337fsX19 (c.3002-1C>T, c. 3002-2T>C, c.2300C>G and c.2849del21		These variants were loss of function, which include premature stop codons,frameshifts and premature truncation and splice variants	Page et al. (2019)
NKX2.5	c.73C>T, p.R25C, c.63A>G, pE21E	TOF/ASD	c.73C>T is considered the most recurrent <i>NKX2.5</i> variant in CHD patients (Kalayinia et al., 2021). The results suggested a potential impact of these variants on splicing	EL Bouchikhi et al. (2021)
	Several variants were reviewed in (Chung and Rajakumar, 2016)	TOF/ASD/Atresia/Stenosis	The most widely accepted hypothesis is that all molecules involved in cardiac development are reduced. However, additional research is needed to determine how the novel mutations identified in <i>NKX2.5</i> affect transcription factor function and thus heart development	Chung and Rajakumar (2016)
GATA4	c.1220C>A, p.P407Q; c.1138G>A, p.V380 M	TOF and patent foramen ovale (PFO)	Variant p.V380M was found in GATA4's second transcription activation TAD domain. p.P407Q was identified as pathogenic in the ClinVar and Human Genome Mutation Database (HGMD)	Qian et al. (2017)
GATA5	c.830C>T; p.P277L	TOF; overriding aorta, loss of the ventricular septum and aortic anterior wall, indicating the presence of a VSD in the foetus	This variant is pathogenic due to inhibiting <i>GATA5</i> nuclear translocation	Jin et al. (2022)
	c. 943T>A, p.8315T, c.274G>T, p.A928	TOF and/or PFO	These novel variants were found in the N- and C-termini of <i>GATA5</i> , as well as the functional domains, and may increase the risk of cardiac abnormalities	Qian et al. (2017)
	there are at least 31 identified GATA5 variations in TOF patients, containing 29 missense variants and two non - protein coding variants	TOF and/or VSD, BAV, aortic stenosis	Reduce <i>GATA5</i> transcription factor activity leading to valve morphogenesis defects and BAV.	Wei et al. (2013), Qian et al. (2017), Shi et al. (2014), Kassab et al. (2015)
GATA6	c.331G>A, p.D111N, c.972C>G, p.H324Q	TOF	Those two novel variants were observed in the TAD domains and might raise cardiac malformations	Qian et al. (2017)
TBX1	p.P43_G61del PPPPRYDPCAAAAPGAPGP	TOF with patent foramen ovale (PFO) and right aortic arch, a dysplastic pulmonary valve, severe valvar and supravalvar pulmonary stenosis and confluent branch pulmonary arteries	These <i>TBX</i> family variants and <i>CITED2</i> variant, reported in this study, were novel which likely alters the protein function and may increase cardic defects but these possibilities need to be confirmed	Qian et al. (2017)

(Continued on following page)

Name of

References

Role to TOF

#### the gene or genetic alteration TBX2 TOF and AVSD c.2139dupG TBX5 c.409G>T, p.V137L TOF and PDA and PFO CITED2 c.-1AT TOF and PAA ZFPM2/FOG2 c. C452T, p.T151I, c.C1208G, p.A403G, TOF/DORV These variants might have an essential Huang et al. (2014) c.G1552A, p.A518T, c.A2107C, p.M703L, role in pathogenesis of TOF and c.G2482A, p.V828M c.C3239T, p.S1080F DORV, suggesting that ZFPM2/FOG2 genetic variations might be potential biomarkers and therapeutic targets for non-syndromic TOF and DORV. c.3442G>A, p.E1148K; c.3014A>G, p.E1005G TOF and/or PFO These variants may disrupt Oian et al. (2017) interactions with GATA4 causing inhibition of GATA4 protein transcription p.E30G, p.I227V, p.M544I, p.V339I, p.K737E, TOF and Double outlet right ZFPM2 variants can cause De Luca et al. (2010), ventricle (DORV) or ventricular complicated CHD, and may or may Tan et al. (2011) p.A611T and other variants septal defect type of DORV not have a serious effect on protein function FOXC1 p.378GGGdel, p. 488AAAAdel TOF with bicuspid pulmonary These rare gene variants in the second Töpf et al. (2014) valve, valvar and subvalvar stenosis, heart field transcriptional network confluent branch pulmonary impact transcription activity, playing a key role in the growth of right-sided arteries cardiovascular structures and the p.P297S TOF with moderate to severe outflow tract subvalvar pulmonary stenosis, confluent branch pulmonary arteries, right aortic arch, and PFO. FOXC2 TOF with moderate valvar p.O444R pulmonary stenosis, mild subvalvar pulmonary stenosis, confluent branch pulmonary arteries, and pyloric stenosis TOF with severe valvar and FOXH1 p.S113T subvalvar pulmonary stenosis, right aortic arch, and confluent branch pulmonary arteries HAND2 p.A25\_A26insAA TOF with pulmonary atresia and major aortopulmonary collateral arteries, right aortic arch, small atrial septal defect, small central pulmonary arteries, and absent thyroid gland c.414\_415dupGT c.2429C>T, p.P810L TOF with branch PS and PA. TOF JAG1 These variants are predicted to form Bauer et al. (2010) premature termination codons c.3312C>G, p.H1104Q with PS, PE.TOF affecting allele transcripts and resulting in 50% normal JAG1 expression c.1511A>G, p.N504S, c.3038A>T, p.H1013L, TOF and/or patent foramen ovale This finding assumed that JAG1 Oian et al. (2017) c.2906T>C, p.M969T,c.806C>G, p.P269R (PFO), or ASD. variations inhibited the transcriptional activation of HEY proteins via an aberrant signalling pathway and subsequently altered the transcriptional control of GATA family members, which may have an additional impact for the variants in these members This variant might affect gene c.765C>T TOF Safari-Arababadi et al. functions; however, more (2018)investigation needs to be confirmed FLNA c.3415C>T; p.L1139F TOF This variant was found in two Kalayinia et al. (2021)

Cardiac phenotype

#### TABLE 1 (Continued) Non-syndromic TOF genetic variants.

Variant

(Continued on following page)

brothers and further studies need to be performed to confirm the variant frequency in TOF patients in different

populations

Name of the gene	Variant or genetic alteration	Cardiac phenotype	Role to TOF	References
KDR encoding VEGF Receptor 2	p.G345W p.G537R	Absent pulmonary valve syndrome and a double outlet right ventricle- severely hypoplastic pulmonary valve, a right ventricle with two outlets, and a right-sided aortic arch	The findings suggest a role for rare <i>KDR</i> variants in TOF etiology <i>via</i> a loss-of-function mechanism	Škorić-Milosavljević et al. (2021)
NDRG4	p.T256M (NM_020465: c.767C>T, rs144494221)	PA, VSD, patent foramen ovale, patent foramen ovale	NDRG4 p.T256M variant may be implicated in the aetiology of PA/VSD and TOF by affecting the regulation of cardiomyocyte proliferation	Peng et al. (2021)
		TOF, patent foramen ovale		
		TOF, patent foramen ovale, left superior vena cava		
SMARCC2	(NM_003075.5: c.3561del, p.L1188fs)	An enlarged aortic valve ventricular septal defect a narrowing of the right ventricle outflow tract pulmonary stenosis	It was still unclear if the patient's congenital cardiac condition was isolated or syndromic	Sun et al. (2022)

TABLE 1 (Continued) Non-syndromic TOF genetic variants.

Surgical repair of the morphological and physiological alterations in the TOF heart has markedly increased survival rates for infants with TOF. Repair in the first year of life is preferable, and earlier repair improves survival outcomes (Peck et al., 2021). Although patients with repaired TOF have a high chance of surviving into adulthood, the transition into middle life is accompanied by severe morbidity and increased mortality rates (Dennis et al., 2017). Both atrial and ventricular arrhythmias are a common source of morbidity and mortality following TOF repair (Pinsker et al., 2022). Additional causes of mortality for individuals with TOF are heart failure and sudden cardiac death (Norgaard et al., 1996; Oechslin et al., 2000); and (Dennis et al., 2017). Sudden mortality in the repaired TOF population is widely documented, with multiple risk factors, which are assumed to be related to chronic RV volume overload over time.

#### Genetics of TOF

A higher incidence of CHD in first-degree relatives and children of TOF patients (Postoev et al., 2014) suggests a genetic aetiology to this disorder. The genetic analysis of syndromic TOF patients has offered valuable insights into causal genes in specific individuals. Approximately 20% of cases are linked to a recognised condition or chromosomal abnormality (Bailliard and Anderson, 2009). These TOF cases are found in patients with genetic syndromes including 22q11 microdeletions, trisomy 21, Alagille's disorder, Cat Eye syndrome, or CHARGE and VATER/VACTERL syndromes (Goldmuntz et al., 1998; Goldmuntz, 2001; McElhinney et al., 2002; Michielon et al., 2006); and (Marino et al., 2012). About 80% of TOF cases are non-syndromic, and display no indication of familial Mendelian CHD segregation, with no identified aetiology (Goodship et al., 2012; Palomino Doza et al., 2013). Rare, *de novo* genetic variations have been found as substantial factors in TOF (Jin et al., 2017). Notably, variants in the key cardiac transcription factor *NKX2.5* were the first to be identified as a genetic cause of TOF (Goldmuntz et al., 1998; Benson et al., 1999). Subsequent studies have further extended our knowledge of genetic alterations resulting in TOF. Therefeore, this mini review aims to offer an update on the genes and variants reported to be associated with non-syndromic TOF (Table 1). As highlighted in this review, disruption of the balanced interaction of transcription factors which are crucial for cardiogenesis plays a substantial role in the aetiology of non-syndromic TOF.

Whole Exome Sequencing (WES) has been used effectively to find new candidate genes for CHD (Zaidi et al., 2013; Al Turki et al., 2014; Homsy et al., 2015); and (Sifrim et al., 2016). Previous research revealed that ultra-rare non-synonymous variants play a major role in the genetic aetiology of CHD, particularly TOF (Jin et al., 2017; Page et al., 2019); and (Reuter et al., 2019). Although familial cases of TOF are rare, likely due to the severity of the disease, inherited variants in the genes GATA4, NKX2.5, JAG1, FOXC2, TBX5 and TBX1 have been reported (reviewed in (Morgenthau and Frishman, 2018). A recent study of the largest group of non-syndromic TOF patients observed to date, using WES of 829 TOF patients, has found that the NOTCH1 locus, followed by FLT4, is the most common site of genetic variants leading to non-syndromic TOF. Variants in these genes are detected in about 7% of TOF cases (Page et al., 2019). Moreover, rare variants in some specific transcription factors, such as FOXC1 and HAND2, have been identified in nonsyndromic TOF patients (Töpf et al., 2014). Because there are links between normal heart formation and multiple transcription factors, variants in transcription factors required during cardiac development can result in aberrant cardiac morphology or function, thus resulting in CHD.

#### NOTCH family

NOTCH family proteins 1-4 are evolutionarily conserved transmembrane proteins that have a crucial impact on cell fate decisions (Andersson et al., 2011). The sequential expression of Notch components and related genes are required during vascular and cardiac development (Luxán et al., 2016). The *NOTCH* pathway is necessary for the developing heart for the regulation of vital pathways involving the formation of cardiac trabeculae and sequential activation of many *NOTCH* genes promotes myocardial development, formation, and compaction (Luxán et al., 2016).

Since NOTCH signalling is so significant for cardiac and vascular development, variants in genes encoding NOTCH signalling components cause a variety of cardiovascular abnormalities (Eldadah et al., 2001; McElhinney et al., 2002; Tomita-Mitchell et al., 2007; Garg et al., 2005; Stittrich et al., 2014; Kerstjens-Frederikse et al., 2016); and (Luxán et al., 2016). A recent study conducted by (Page et al., 2019) found five new de novo NOTCH1 variants in the TOF cohort; three of these were missense variations, whereas the remaining two were truncating variants (Table 1): p. G200V, p. R448X, p. C1549Y, p. W1638X and p. N1875S. The differences in NOTCH1 signalling observed were not due to the variants' reduced mRNA expression. As a result, two variants, the p. C607Y and p. N1875S, recognised in TOF patients who underwent functional testing, were found to affect canonical NOTCH1 signalling (Page et al., 2019). Furthermore, eight ultra-rare missense NOTCH1 variants were found in the investigation of a 175-member TOF cohort in a study performed by (Manshaei et al., 2020). Their analysis confirmed that those variants were ultra-rare deleterious and involved in the pathogenesis of TOF, suggesting abnormal vascular development and related signalling defects as probable causes of TOF.

## FLT4

Vascular endothelial growth factor receptors (VEGFR1-3) are crucial in the development and long-term maintenance of the cardiovascular and lymphovascular systems. VEGFR3 (Vascular Endothelial Growth Factor 3), encoded by the *FLT4* gene, plays an essential and well-studied role in the development and formation of the circulatory vessels (Olsson et al., 2006; Karaman et al., 2022). Statistical analysis revealed that *FLT4* was among the most common genes predisposing individuals to non-syndromic TOF (Matos-Nieves et al., 2019). Likewise, exome sequence analysis revealed that *FLT4* and *NOTCH1* genes were the most significant genes

that caused non-syndromic TOF in a cohort of 829 TOF patients (Page et al., 2019). Recent analysis of exome sequencing data from 811 TOF probands has found 14 *FLT4* loss of function variants (Reuter et al., 2019). Additionally, (Page et al., 2019); found that several *FLT4* variants in TOF patients are unique and severe, due to causing premature protein truncation and frameshifts located in the FLT4 immunoglobulin (Ig) and VEGFR3 protein kinase domains. Taken together, these findings highlight the role of *FLT4* in TOF and CHD; however, the precise mechanism of how *FLT4* variants provoke the cardiac structural alterations of TOF remains to be elucidated.

#### NKX2.5

The human transcription factor NKX2.5 is the first gene associated with human cardiac development. It sits at the 5q34q35 chromosome five region and is abundantly expressed in the human foetal heart (Shinebourne et al., 2006). An essential role of NKX2.5 during secondary heart field development is its interactions with the broader network of transcriptional factors that regulate cardiac differentiation and fate (Clark et al., 2013). Moreover, deletion of Nkx2.5 has been linked to abnormalities in the structure of the embryonic heart, growth delay, and embryo mortality in mice (Lyons et al., 1995). Several studies have also reported that variants in NKX2.5 cause a wide range of CHDs, including ASD, VSD, TOF, HLH, CoA, TGA, Double Outlet Right Ventricle (DORV), IAA, and cardiac outflow tract (OFT) anomalies; reviewed in (Chung and Rajakumar, 2016). Recently, new NKX2.5 variants were discovered in a Moroccan cohort with non-syndromic TOF; a missense variant R25C and a synonymous variant E21E (EL Bouchikhi et al., 2021). The R25C variant is expected to impair NKX2.5 dimerisation and impede activation of downstream target genes (Kasahara et al., 2000; Dentice et al., 2006). Bioinformatic predictions indicate that the E21E variant might impair NKX2.5 transcript splicing by removing a whole exon from the transcript, resulting in translation of a shortened protein that is most likely inactive. Further in vitro functional tests are needed to validate this finding (EL Bouchikhi et al., 2021).

#### Interaction of transcription factors

In mammals, the master cardiac transcription factors (TFs) such as *GATA4*, *TBX5*, *HAND2*, *KNX2.5*, and *MESP1* control the intricate and complicated process of heart development (Olsson et al., 2006). Pathological variants in essential cardiac transcription factor genes such as *GATA4*, *GATA5* and *GATA6* are associated with CHD (Kodo et al., 2009); Jiang

et al., 2013; and (Granados-Riveron et al., 2011). Moreover (Töpf et al., 2014), reported that rare variants in the transcription factors of the second heart field, such as *HAND2*, *FOXC1*, *FOXC2*, *FOXH1*, and *TBX1*, can be found in non-syndromic TOF patients. These variants affect transcription activity; thereby disrupting the vital role these proteins play in the development of the outflow tract and right-sided cardiovascular components.

Via the general process of combinatorial interactions, a significant subset of cardiac transcription factors can regulate the intricate spatiotemporal control of heart morphogenesis and intermolecular interactions (Durocher and Nemer, 1998; Bruneau, 2008); and (Pradhan et al., 2017). For instance, physical interaction between GATA4, NKX2.5, and TBX5 activates downstream targets in a synergistic manner (Bruneau et al., 2001; Hiroi et al., 2001). Thus, mutations in these transcription factors may damage their interactions, thereby affecting protein function. Interestingly, according to a semi-quantitative study in the TOF cohort, (Qian et al., 2017); found that the interaction between the ZFPM2/FOG2 E1148K mutant protein with GATA4 was significantly damaged and was reduced by 50 per cent compared to the wild-type ZFPM2 protein (Qian et al., 2017). On the other hand, this result is not consistent with the previous findings whereby (Zhang et al., 2014) demonstrated that although the p.M703L and p. T843M ZFPM2 missense variants were assumed to be damaging, they had only a minor effect, if any, on the protein interaction with GATA4. Therefore, the impacts of individual variants may disrupt protein interactions to differing extents, or different variants may involve other unknown mechanisms to provoke CHD.

Moreover, multiple new potential candidate genes for causing TOF, that are linked to the function of FLT4, were identified, such as *CLDN9*, *CCDC168*, *SN2*, *L1TD*, *TACC3*, *CCDC36* and *TTC5* (Manshaei et al., 2020). More recently, it has been noted that twenty-three confirmed TOF genes form an interaction network with KDR/VEGFR2 and NOTCH1, acting as key nodes in the protein interaction network, each directly linking to 11 additional proteins *via* functional biological connections (Reuter et al., 2019). Reanalysis of the accuracy of these protein interactions may help to understand the interaction mechanisms and might inform therapeutic targets of TOF patients.

#### New variants yet to be confirmed

*De novo* variants have a major role in the development of early-onset genetic diseases such as CHD. A screen of a CHD cohort was performed by (Bauer et al., 2010); and found that *JAG1* variants, producing functionally relevant sequence changes, were detected in 3% (2/94) of TOF cases. These individuals did not show signs of any other recognised syndromes. A recent study has concluded that a *JAG1* variant: c.765C>T, is strongly linked to TOF in Iranian patients with a ratio of 60.2% *versus* 5.7% in controls (Safari-Arababadi et al., 2018); this variant may affect gene function. However, additional investigations need to be performed to understand the mechanisms of *JAG1* alterations in TOF.

Furthermore, despite the limited sample size in a study conducted by (Manshaei et al., 2020); burden test findings indicated ultra-rare truncating variants in novel candidate genes, such as ZFAND5 and WNT5A, that might be involved in the genesis of TOF. A novel FLNA variant, p. L1139F, was discovered in two Iranian brothers diagnosed with X-linked recessive TOF. This variant was heterozygous in their mother, whilst their father had the wild type reference allele. Hemizygosity for this variant is therefore predicted to have a role in CHD, particularly TOF (Kalayinia et al., 2021). Nevertheless, further research is required to demonstrate the role of this variant or other FLNA variants in TOF individuals and their frequency in different populations. These new gene associations reveal the need for further investigation of the role of the contribution of rare variants to TOF in a larger patient cohort, which might give new insights into additional genetic causes of non-syndromic TOF.

In addition, it has also been identified that the loss of function of three emerging candidate genes, KDR, IQGAP1, and GDF1 may cause CHD and TOF (Reuter et al., 2019), but further clinical confirmation is needed. Highlighting the genetic variability of TOF; 16 additional patients have been identified to have one pathogenic/likely pathogenic variant (11 loss of function and five missense) in 16 CHD genes (ARHGAP31, ATRX, CACNA1C, CHD7, CSNK2A1, DLL4, EP300, GATAD2B, KAT6A, LZTR1, NF1, NODAL, PIK3CA, RAF1, RASA1, and SMAD2) and one patient has been identified with loss of function variants in two genes (ASXL1 and PSMD12) (Reuter et al., 2019). In addition, recent studies have shown that new variants found in TOF cases in the NDRG4 gene cause multiple heart abnormalities (Peng et al., 2021). Evidence suggests that SMARCC2 is required for cardiac development by promoting cardiac myocyte differentiation and controlling temporal steps in cardiac differentiation (Hota et al., 2019). More recently, in 2022, Sun et al. found a de novo SMARCC2 variant in TOF patients, suggesting there may be a link between CHD and deleterious SMARCC2 variants (Sun et al., 2022). Further research into the pathogenic mechanisms linked with these new variants, and genotype-phenotype associations is required, which may give new therapeutic insights for treating TOF and CHD.

# Discussion

Tetralogy of Fallot is the most common type of cyanotic congenital heart defect and a serious condition that includes

four heart abnormalities: a ventricular septal defect, pulmonary stenosis/right ventricular outflow tract obstruction, the aorta overriding the VSD, and right ventricular hypertrophy (Hota et al., 2019). Despite advanced medical interventions, morbidity and mortality for individuals with TOF remain high due to the severe developmental cardiac abnormalities present in this condition. Additionally, the critical causative genes associated with non-syndromic TOF are poorly understood.

Recent progress in identifying the genetic basis for TOF has shown that deleterious variants in *NOTCH1* and *FLT4* play a significant role in TOF and CHD; however, the precise mechanism of these gene variations in TOF remains unknown. In addition, variants in additional cardiac transcription factors were connected to cardiac developmental defects and non-syndromic TOF (Granados-Riveron et al., 2011); Jiang et al., 2013, and (Töpf et al., 2014), suggesting the impacts of these mutants disrupt protein interactions which disrupts the crucial role these proteins play in the evolution of the cardiovascular components.

Recent research findings showed that several rare variants in many different genes might be associated with TOF in individual patients. Additionally, multiple variants in a single individual may act together to perturb cardiac development and cause TOF. Therefore, there may be a large number of genes that cause TOF in only a small number of individuals. These genes may have links to each other in protein interactions or signalling pathways needed for cardiac development. This finding suggests that more research is needed into the pathophysiological mechanisms associated with these new variants and genotype-phenotype associations. This research may be a promising aspect to provide new therapeutic insights for treating TOF and CHD.

#### References

Al Turki, S., Manickaraj, A. K., Mercer, C. L., Gerety, S. S., Hitz, M.-P., Lindsay, S., et al. 2014). Rare variants in NR2F2 cause congenital heart defects in humans. *Am. J. Hum. Genet.*, 95(1), p.126. doi:10.1016/j.ajhg.2014.06.013

Andersson, E. R., Sandberg, R., and Lendahl, U. (2011) Notch signaling: Simplicity in design, versatility in function, 138. *Development*, (17), pp.3593–3612. doi:10.1242/dev.063610

Bailliard, F., and Anderson, R. H. (2009). Tetralogy of Fallot. Orphanet J. Rare Dis. 4, 2. doi:10.1186/1750-1172-4-2

Bauer, R. C., Laney, A. O., Smith, R., Gerfen, J., Morrissette, J. J. D., Woyciechowski, S., et al. (2010). Jagged1 (Jag1) mutations in patients with tetralogy of Fallot or pulmonic stenosis. *Hum. Mutat.*, 31(5), Pp.594–601. doi:10.1002/Humu.21231

Bedair, R., and Iriart, X. (2019). Educational series in congenital heart disease: Tetralogy of Fallot: Diagnosis to long-term follow-up. *Echo Res. Pract.*, 6(1), pp.R10-R24. doi:10.1530/erp-18-0049

Benson, D. W., Silberbach, G. M., Kavanaugh-McHugh, A., Cottrill, C., Zhang, Y., Riggs, S., et al. (1999). Mutations in the cardiac transcription factor NKX2.5 affect diverse cardiac developmental pathways. *J. Clin. Invest.*, 104(11), pp.1567–1573. doi:10.1172/jci8154

Bruneau, B. G., Nemer, G., Schmitt, J. P., Charron, F., Robitaille, L., Caron, S., et al. (2001). A murine model of holt-oram syndrome defines roles of the T-box

#### Author contributions

NA: Performed Research, Wrote Manuscript KH: Wrote Manuscript, Edited Manuscript, Obtained Funding.

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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transcription factor Tbx5 in cardiogenesis and disease. Cell, 106(6), pp.709-721. doi:10.1016/s0092-8674(01)00493-7

Bruneau, B. G. (2008). The developmental genetics of congenital heart disease. *Nature*, 451(7181), pp.943–948. doi:10.1038/nature06801

Chung, I.-M., and Rajakumar, G. (2016). Genetics of congenital heart defects: The NKX2-5 gene, a key player. *Genes*, 7(2), p.6. doi:10.3390/genes7020006

Clark, C. D., Zhang, B., Lee, B., Evans, S. I., Lassar, A. B., and Lee, K. H. (2013). Evolutionary conservation of Nkx2.5 autoregulation in the second heart field. *Dev. Biol.* 374 (1), 198–209. doi:10.1016/j.ydbio.2012.11.007

De Luca, A., Sarkozy, A., Ferese, R., Consoli, F., Lepri, F., Dentici, M., Vergara, P., De Zorzi, A., Versacci, P., Digilio, M., Marino, B., and Dallapiccola, B. (2010). New mutations in zfpm2/fog2 gene in tetralogy of Fallot and Double outlet right ventricle. *Clin. Genet.*, 80(2), Pp.184–190. doi:10.1111/J.1399-0004.2010.01523.X

Dennis, M., Moore, B., Kotchetkova, I., Pressley, L., Cordina, R., and Celermajer, D. S. (2017). Adults with repaired tetralogy: Low mortality but high morbidity up to middle age. *Open Heart*, 4(1), p.e000564. doi:10.1136/openhrt-2016-000564

Dentice, M., Cordeddu, V., Rosica, A., Ferrara, A. M., Santarpia, L., Salvatore, D., et al. (2006). Missense mutation in the transcription factor NKX2-5: A novel molecular event in the pathogenesis of thyroid dysgenesis. *J. Clin. Endocrinol. Metabolism*, 91(4), pp.1428–1433. doi:10.1210/jc.2005-1350

Dolk, H., Loane, M., and Garne, E. Congenital heart defects in Europe. (2011) congenital heart defects in Europe: Prevalence and perinatal mortality, 2000 to 2005. *Circulation*. 123(8):841–849. doi:10.1161/CIRCULATIONAHA.110.958405

Durocher, D., and Nemer, M. (1998). Combinatorial interactions regulating cardiac transcription. *Dev. Genet.*, 22(3), pp.250–262. doi:10.1002/(sici)1520-

El Bouchikhi, I., Belhassan, K., Moufid, F. Z., Bouguenouch, L., Samri, I., Iraqui Houssaïni, M., et al. (2021). Screening of NKX2.5 gene in Moroccan tetralogy of Fallot (TOF) patients: Worldwide mutation rate comparisons show a significant association between R25C variant and TOF phenotype. *Egypt J. Med. Hum. Genet.*, 22(1). doi:10.1186/s43042-021-00136-1

Eldadah, Z. A., Hamosh, A., Biery, N. J., Montgomery, R. A., Duke, M., Elkins, R., et al. (2001). Familial Tetralogy of Fallot caused by mutation in the jagged1 gene. *Hum. Mol. Genet.* 10 (2), 163–169. [online]. doi:10.1093/hmg/10.2.163

Garg, V., Muth, A. N., Ransom, J. F., Schluterman, M. K., Barnes, R., King, I. N., et al. (2005). Mutations in NOTCH1 cause aortic valve disease. *Nature*. 437 (7056), 270–274. doi:10.1038/nature03940

Goldmuntz, E., Clark, B. J., Mitchell, L. E., Jawad, A. F., Cuneo, B. F., Reed, L., et al. (1998). Frequency of 22q11 deletions in patients with conotruncal defects. *J. Am. Coll. Cardiol.*, 32(2), pp.492–498. doi:10.1016/s0735-1097(98)00259-9

Goldmuntz, E. (2001). The epidemiology and genetics of congenital heart disease. *Clin. Perinatology*, 28(1), pp.1–10. doi:10.1016/s0095-5108(05)70067-1

Goodship, J. A., Hall, D., Topf, A., Mamasoula, C., Griffin, H., Rahman, T. J., et al. 2012). A common variant in the PTPN11 gene contributes to the risk of tetralogy of Fallot. *Circ. Cardiovasc Genet.*, 5(3), pp.287–292. doi:10.1161/circgenetics.111. 962035

Granados-Riveron, J. T., Pope, M., Bu'Lock, F. A., Thornborough, C., Eason, J., Setchfield, K., et al. (2011). Combined mutation screening of NKX2-5, GATA4, and TBX5 in congenital heart disease: Multiple heterozygosity and novel mutations. *Congenit. Heart Dis.*, 7(2), pp.151–159. doi:10.1111/j.1747-0803.2011.00573.x

Hiroi, Y., Kudoh, S., Monzen, K., Ikeda, Y., Yazaki, Y., Nagai, R., et al. (2001). Tbx5 associates with Nkx2-5 and synergistically promotes cardiomyocyte differentiation. *Nat. Genet.*, 28(3), pp.276–280. doi:10.1038/90123

Ho, C. Y., Day, S. M., Ashley, E. A., Michels, M., Pereira, A. C., Jacoby, D., et al. (2019). Response by Ho et al to Letter Regarding Article, "Genotype and Lifetime Burden of Disease in Hypertrophic Cardiomyopathy: Insights From the Sarcomeric Human Cardiomyopathy Registry (SHARe)". *Circulation*, 139(12), pp.1559–1560. doi:10.1161/circulationaha.118.039069

Hoffman, J. I. E., and Kaplan, S. (2002). The incidence of congenital heart disease. J. Am. Coll. Cardiol., 39(12), pp.1890–1900. doi:10.1016/s0735-1097(02)01886-7

Homsy, J., Zaidi, S., Shen, Y., Ware, J. S., Samocha, K. E., Karczewski, K. J., et al. 2015). De novo mutations in congenital heart disease with neurodevelopmental and other congenital anomalies. *Science*, 350(6265), pp.1262–1266. doi:10.1126/science. aac9396

Hota, S. K., Johnson, J. R., Verschueren, E., Thomas, R., Blotnick, A. M., Zhu, Y., et al. (2019), Dynamic BAF chromatin remodeling complex subunit inclusion promotes temporally distinct gene expression programs in cardiogenesis. *Development*. 146, dev174086. doi:10.1242/Dev.174086

Huang, X., Niu, W., Zhang, Z., Zhou, C., Xu, Z., Liu, J., et al. (2014). Identification of novel significant variants of *zfpm2*/fog2 in non-syndromic tetralogy of Fallot and Double outlet right ventricle in A Chinese han population. *Mol. Biol. Rep.*, 41(4), Pp.2671–2677. doi:10.1007/S11033-014-3126-5

Jiang, J. Q., Li, R. G., Wang, J., Liu, X. Y., Xu, Y. J., Fang, W. Y., et al. (2013). Prevalence and spectrum of GATA5 mutations associated with congenital heart disease. *Int. J. Cardiol.* 165 (3), 570–573. doi:10.1016/j.ijcard.2012.09.039

Jin, J. Y., Wu, L. P., Dong, Y., Pi, H. C., Wu, H. F., and Xiang, R. (2022). Identification of a novel GATA binding protein 5 variant (c.830C>T/p.P277L) damaging the nuclear translocation and causing tetralogy of Fallot. *Qjm*,115(4): 256–258. doi:10.1093/Qjmed/Hcab317.Pmid:34904678

Jin, S. C., Homsy, J., Zaidi, S., Lu, Q., Morton, S., DePalma, Sr., et al. (2017). Contribution of rare inherited and de novo variants in 2,871 congenital heart disease probands. *Nat. Genet.*, 49(11):1593–1601. doi:10.1038/Ng.3970

Kalayinia, S., Maleki, M., Mahdavi, M., and Mahdieh, N. (2021) Whole-exome sequencing reveals a novel mutation of FLNA gene in an Iranian family with nonsyndromic tetralogy of Fallot. *Lab. Med.*, 52, 614, 618. doi:10.1093/Labmed/Lmab018

Karaman, S., Paavonsalo, S., Heinolainen, K., Lackman, M. H., Ranta, A., Hemanthakumar, K. A., et al. (2022). Interplay of vascular endothelial growth factor receptors in organ-specific vessel maintenance. *J. Exp. Med.*, 219(3). doi:10. 1084/jem.20210565

Kasahara, H., Lee, B., Schott, J.-J., Benson, D. W., Seidman, J. G., Seidman, C. E., et al. (2000). Loss of function and inhibitory effects of human CSX/

NKX2.5 homeoprotein mutations associated with congenital heart disease. J. Clin. Invest., 106(2), pp.299-308. doi:10.1172/jci9860

Kassab, K., Hariri, H., Gharibeh, L., Fahed, A. C., Zein, M., El-Rassy, I., et al. (2015). Gata5 mutation homozygosity linked to A Double outlet right ventricle phenotype in A Lebanese patient. *Mol. Genet. Genomic Med.*, 4(2), Pp.160–171. doi:10.1002/Mgg3.190

Kerstjens-Frederikse, W. S., van de Laar, I. M. B. H., Vos, Y. J., Verhagen, J. M. A., Berger, R. M. F., Lichtenbelt, K. D., et al. (2016). Cardiovascular malformations caused by NOTCH1 mutations do not keep left: Data on 428 probands with leftsided CHD and their families. *Genet. Med.*, 18(9), pp.914–923. doi:10.1038/gim. 2015.193

Kodo, K., Nishizawa, T., Furutani, M., Arai, S., Yamamura, E., Joo, K., et al. (2009). GATA6 mutations cause human cardiac outflow tract defects by disrupting semaphorin-plexin signaling. *Proc. Natl. Acad. Sci. U.S.A*, 106(33), pp.13933–13938. doi:10.1073/pnas.0904744106

Luxán, G., D'Amato, G., MacGrogan, D., and de la Pompa, J. L. (2016). Endocardial Notch signaling in cardiac development and disease. *Circ. Res.*, 118(1). e1-e18. doi:10.1161/circresaha.115.305350

Lyons, I., Parsons, L. M., Hartley, L., Li, R., Andrews, J. E., Robb, L., et al. (1995). Myogenic and morphogenetic defects in the heart tubes of murine embryos lacking the homeo box gene Nkx2-5. *Genes Dev.* 9 (13), 1654–1666. doi:10.1101/gad.9.13. 1654

Manshaei, R., Merico, D., Reuter, M. S., Engchuan, W., Mojarad, B. A., Chaturvedi, R., et al. 2020). Genes and pathways implicated in tetralogy of Fallot revealed by ultra-rare variant burden analysis in 231 genome sequences. *Front. Genet.*, 11. 957. doi:10.3389/Fgene.2020.00957

Marino, B. S., Lipkin, P. H., Newburger, J. W., Peacock, G., Gerdes, M., Gaynor, J. W., et al. (2012). Neurodevelopmental outcomes in children with congenital heart disease: Evaluation and management. *Circulation*, 126(9), pp.1143–1172. doi:10. 1161/cir.0b013e318265ee8a

Matos-Nieves, A., Yasuhara, J., and Garg, V. (2019). Another Notch in the genetic puzzle of tetralogy of Fallot. *Circ. Res.*, 124(4), pp.462–464. doi:10.1161/circresaha. 118.314520

McElhinney, D. B., Krantz, I. D., Bason, L., Piccoli, D. A., Emerick, K. M., Spinner, N. B., et al. (2002). Analysis of cardiovascular phenotype and genotype-phenotype correlation in individuals with a JAG1 mutation and/or Alagille syndrome. *Circulation*, 106(20), pp.2567–2574. doi:10.1161/01.cir.0000037221.45902.69

Mcleod, G., Shum, K., Gupta, T., Chakravorty, S., Kachur, S., Bienvenu, L., et al. (2018). Echocardiography in congenital heart disease. *Prog. Cardiovasc. Dis.* 61(5-6):468–475. doi:10.1016/j.pcad.2018.11.004

Michielon, G., Marino, B., Formigari, R., Gargiulo, G., Picchio, F., Digilio, M. C., et al. (2006). Genetic syndromes and outcome after surgical correction of tetralogy of Fallot. *Ann. Thorac. Surg.*, 81(3), pp.968–975. doi:10.1016/j.athoracsur.2005. 09.033

Morgenthau, A., and Frishman, W. H. (2018). Genetic origins of tetralogy of Fallot, *Cardiology in review*: 26(2), pp 86–92. doi:10.1097/CRD.000000000000170

Norgaard, M. A., Olsen, P. S., Svendsen, U. G., and Pettersson, G. (1996). Revascularization of the bronchial arteries in lung transplantation: An overview. *Ann. Thorac. Surg.*, 62(4), pp.1215–1221. doi:10.1016/0003-4975(96)00539-5

Oechslin, E. N., Attenhofer Jost, C. H., Rojas, J. R., Kaufmann, P. A., and Jenni, R. (2000). Long-term follow-up of 34 adults with isolated left ventricular noncompaction: A distinct cardiomyopathy with poor prognosis. *J. Am. Coll. Cardiol.*, 36(2), pp.493–500. doi:10.1016/s0735-1097(00)00755-5

Olsson, A.-K., Dimberg, A., Kreuger, J., and Claesson-Welsh, L. (2006). VEGF receptor signalling ? In control of vascular function. *Nat. Rev. Mol. Cell Biol*, 7(5), pp.359–371. doi:10.1038/nrm1911

Page, D. J., Miossec, M. J., Williams, S. G., Monaghan, R. M., Fotiou, E., Cordell, H. J., et al. 2019). Whole exome sequencing reveals the major genetic contributors to nonsyndromic tetralogy of Fallot. *Circ. Res.*, 124(4), Pp.553–563. doi:10.1161/ Circresaha.118.313250

Palomino Doza, J., Topf, A., Bentham, J., Bhattacharya, S., Cosgrove, C., Brook, J. D., et al. (2013). Low-frequency intermediate penetrance variants in the ROCK1 gene predispose to Tetralogy of Fallot. *BMC Genet.*, 14(1). 57. doi:10. 1186/1471-2156-14-57

Park, G., and Thayer, J. F. (2014). From the heart to the mind: Cardiac vagal tone modulates top-down and bottom-up visual perception and attention to emotional stimuli. *Front. Psychol.*, 5. 278. doi:10.3389/fpsyg.2014.00278

Peck, D., Tretter, J., Possner, M., Yutzey, K., Zafar, F., Morales, D., et al. (2021). Timing of repair in tetralogy of Fallot: Effects on outcomes and myocardial health. *Cardiol. Rev.*, 29,(2), Pp.62–67. doi:10.1097/Crd.00000000000293

Peng, J., Wang, Q., Meng, Z., Wang, J., Zhou, Y., Zhou, S., Song, W., Chen, S., Chen, A. F., and Sun, K. (2021). A loss-of-function mutation p.T256M in NDRG4 is

implicated in the pathogenesis of pulmonary atresia with ventricular septal defect (PA/VSD) and tetralogy of Fallot (TOF). *Febs Open Bio*, 11(2), Pp.375–385. doi:10. 1002/2211-5463.13044

Pinsker, B. L., Serfas, J. D., and Krasuski, R. A. (2022). Burden and impact of arrhythmias in repaired tetralogy of Fallot. *Curr. Cardiol. Rep.*, 24(3), Pp.225–234. doi:10.1007/S11886-022-01638-Z

Postoev, V. A., Talykova, L. V., and Vaktskjold, A. (2014). Epidemiology of cardiovascular malformations among newborns in monchegorsk (North-West Russia): A register-based study. *J. Public Health Res.*;3(2):270. doi:10.4081/jphr. 2014.270.PMID:25343136

Pradhan, A., Zeng, X.-X. I., Sidhwani, P., Marques, S. R., George, V., Targoff, K. L., et al. (2017). FGF signaling enforces cardiac chamber identity in the developing ventricle. *Development*, 144(7), pp.1328–1338. doi:10.1242/dev.143719

Qian, Y., Xiao, D., Guo, X., Chen, H., Hao, L., Ma, X., Huang, G., Ma, D., and Wang, H. (2017). Multiple gene variations contributed to congenital heart disease via gata family transcriptional regulation. *J. Transl. Med.*, 15(1), 69. doi:10.1186/S12967-017-1173-0

Reuter, M. S., Jobling, R., Chaturvedi, R. R., Manshaei, R., Costain, G., Heung, T., et al. (2019). Haploinsufficiency of vascular endothelial growth factor related signaling genes is associated with tetralogy of Fallot. *Genet. Med.*, 21(4), Pp.1001–1007. doi:10.1038/S41436-018-0260-9

Safari-Arababadi, A., Behjati-Ardakani, M., Kalantar, S. M., and Jaafarinia, M. (2018). Silencing mutations in JAG1 gene may play crucial roles in the pathogenesis of tetralogy of Fallot. *Cell Mol. Biol. (Noisy-le-grand)*, 64 (4), P.103. doi:10.14715/Cmb/2018.64.4.17

Senst, B., Kumar, A., and Diaz, R. R. Cardiac surgery. In: *Treasure island (FL)*, Tampa, Florida, United States: StatPearls Publishing, 2022, StatPearls [internet].

Shi, L.-M., Tao, J.-W., Qiu, X.-B., Wang, J., Yuan, F., Xu, L., et al. (2014). Gata5 loss-of-function mutations associated with congenital bicuspid aortic valve. *Int. J. Of Mol. Med.*, 33(5), Pp.1219–1226. doi:10.3892/Ijmm.2014.1700

Shinebourne, E. A., Babu-Narayan, S. V., and Carvalho, J. S. (2006) Tetralogy of Fallot: From fetus to adult. *Heart*. 92(9):1353-1359. doi:10.1136/hrt.2005.061143

Sifrim, A., Hitz, M.-P., Hitz, A., Wilsdon, J., Breckpot, S. H. A., Turki, B., et al. 2016). Distinct genetic architectures for syndromic and nonsyndromic congenital heart defects identified by exome sequencing. *Nat. Genet.*, 48(9), pp.1060–1065. doi:10.1038/ng.3627

Škorić-Milosavljević, D., Lahrouchi, N., Bosada, F. M., Dombrowsky, G., Williams, S. G., Lesurf, R., et al. (2021). Rare variants in kdr, encoding vegf receptor 2, are associated with tetralogy of Fallot. *Genet. Med.*, 23(10), Pp.1952–1960. doi:10.1038/S41436-021-01212-Y Stittrich, A.-B., Lehman, A., Bodian, Dale L., Ashworth, J., Zong, Z., Li, H., et al. (2014). Mutations in NOTCH1 cause adams-oliver syndrome. *Am. J. Hum. Genet.*, 95, (3), pp.275–284. doi:10.1016/j.ajhg.2014.07.011

Su, Z., Zou, Z., Hay, S. I., Liu, Y., Li, S., Chen, H., et al. (2022). Global, regional, and national time trends in mortality for congenital heart disease, 1990-2019: An age-period-cohort analysis for the Global Burden of Disease 2019 study. *Eclinicalmedicine*, 43, P.101249. doi:10.1016/J.Eclinm.2021. 101249

Sun, H., Zhang, S., Wang, J., Zhou, X., Zhang, H., Yang, H., et al. (2022). Expanding the phenotype Associated with Smarcc2 variants: A fetus with tetralogy of Fallot. *BMC Med. Genomics*, 15(1). 40. doi:10.1186/S12920-022-01185-0

Suzuki, N., Svensson, K., and Eriksson, U. J. (1996). High glucose concentration inhibits migration of rat cranial neural crest cells *in vitro*. *Diabetologia*, 39(4), pp.401–411. doi:10.1007/bf00400671

Tan, Z-P., Huang, C., Xu, Z-B., Yang, J-F., and Yang, Y-F. (2011). Novelzfpm2/ Fog2variants in patients with Double outlet right ventricle. *Clin. Genet.*, 82(5), Pp.466–471. doi:10.1111/J.1399-0004.2011.01787.X

Tomita-Mitchell, A., Maslen, C. L., Morris, C. D., Garg, V., and Goldmuntz, E. (2007). GATA4 sequence variants in patients with congenital heart disease. J. Med. Genet., 44(12), pp.779–783. doi:10.1136/jmg.2007.052183

Töpf, A., Griffin, H. R., Glen, E., Soemedi, R., Brown, D. L., Hall, D., et al. (2014). Functionally significant, rare transcription factor variants in tetralogy of Fallot. *Plos One*, 9(8), P.E95453. doi:10.1371/Journal.Pone.0095453

Wei, D., Bao, H., Liu, X.-Y., Zhou, N., Wang, Q., Li, R.-G., et al. (2013). Gata5 loss-of-function mutations underlie tetralogy of Fallot. *Int. J. Med. Sci.*, 10(1), Pp.34–42. doi:10.7150/Ijms.5270

Wilson, R., Ross, O., and Griksaitis, M. J. (2019, Tetralogy of Fallot, *BJA Educ.*, 19). (11), Pp.362–369. doi:10.1016/J.Bjae.2019.07.003

Wiputra, H., Chen, C. K., Talbi, E., Lim, G. L., Soomar, S. M., Biswas, A., et al. (2018). Human fetal hearts with tetralogy of Fallot have altered fluid dynamics and forces. Am. J. Of Physiology-Heart Circulatory Physiology, 315(6), Pp.H1649–H1659. doi:10.1152/Ajpheart.00235.2018

Zaidi, S., Choi, M., Wakimoto, H., Ma, L., Jiang, J., Overton, J. D., et al. 2013). De novo mutations in histone-modifying genes in congenital heart disease. *Nature*, 498(7453), pp.220–223. doi:10.1038/nature12141

Zhang, W., Shen, L., Deng, Z., Ding, Y., Mo, X., Xu, Z., et al. (2014). Novel missense variants of zfpm2/fog2 identified in conotruncal heart defect patients do not impair interaction with Gata4. *Plos One*, 9(7), pp.E102379. doi:10.1371/Journal. Pone.0102379