



# Interactions, Diffusion, and Membrane Fluctuations in Concentrated Unilamellar Lipid Vesicle Solutions

Elizabeth G. Kelley<sup>1\*</sup>, Emily E. Blick<sup>1†</sup>, Vivek M. Prabhu<sup>2</sup>, Paul D. Butler<sup>1,3,4</sup> and Michihiro Nagao<sup>1,5,6</sup>

<sup>1</sup>Center for Neutron Research, National Institute of Standards and Technology, Gaithersburg, MD, United States, <sup>2</sup>Material Measurement Laboratory, National Institute of Standards and Technology, Gaithersburg, MD, United States, <sup>3</sup>Department of Chemical and Biomolecular Engineering, University of Delaware, Newark, DE, United States, <sup>4</sup>Department of Chemistry, The University of Tennessee, Knoxville, TN, United States, <sup>5</sup>Department of Materials Science and Engineering, University of Maryland, College Park, MD, United States, <sup>6</sup>Department of Physics and Astronomy, University of Delaware, Newark, DE, United States

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### \*Correspondence:

Elizabeth G. Kelley  
egk@nist.gov

### †Present address:

Emily E. Blick,  
Biomedical Engineering Department,  
The University of Wisconsin, Madison,  
WI, United States

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Lipid vesicles are widely used as models for cell membranes, hosts for membrane protein studies, and containers for hydrophilic molecules. The vesicle solutions in these applications are usually prepared at a specific lipid concentration; however, because vesicles are solvent-filled structures, the corresponding volume fraction of vesicles is at least a factor of three times higher than the corresponding lipid volume fraction and critically depends on the vesicle radii. Here we show that these higher than may be expected vesicle volume fractions result in measurable interactions between the vesicles as well as affect the vesicle diffusion. We show that vesicle solutions prepared with lipid mass fractions,  $m_L$ , as low as  $\approx 0.004$ , which correspond to a lipid concentration of  $\approx 4$  mg/mL or 5 mmol/L (mM), not only have a measurable apparent structure factor ( $S'(q)$ ) in small angle neutron scattering (SANS) experiments, but that this repulsive structure factor also affects the measured diffusion coefficient at small scattering vectors ( $q$ ) such as those probed with dynamic light scattering (DLS). The measured diffusion coefficients are further affected by indirect solvent mediated interactions described by a hydrodynamic factor ( $H(q)$ ). Accounting for the concentration-dependence of the vesicle diffusion shows that the lipid concentration dependence measured in neutron spin echo (NSE) spectroscopy is due to differences in the effective vesicle diffusion coefficients and not the membrane fluctuation dynamics. The results have practical implications for static and dynamic scattering experiments as well as provide interesting insights into the interactions between soft lipid vesicles.

**Keywords:** lipid vesicles, small angle neutron scattering (SANS), dynamic light scattering (DLS), neutron spin echo spectroscopy (NSE), structure factor, hydrodynamic factor, membrane dynamics

## 1 INTRODUCTION

Lipid vesicles have many uses. They are used as model systems for biomembranes [1]; [2], environments to reconstitute and study membrane proteins [3]; [4], small volume chemical reactors [5]; [6], as well as containers for hydrophilic guest molecules in personal care products [7], drug delivery vehicles [8], and vaccines [9]. In almost all these applications, the vesicle solutions are formulated at a specific lipid mass concentration to either optimize the signal to noise ratio in

experimental studies of their properties or to ensure sufficient concentrations of the encapsulated cargoes. However, the lipids only form a thin shell layer around a large, solvent-filled, core, and the corresponding vesicle volume fraction is much higher because  $\phi_V = \phi_L + \phi_{internal}$  where  $\phi_L$  is the volume fraction of the lipids that form the shell and  $\phi_{internal}$  is effective volume fraction of the solvent filled cores. As such, the corresponding vesicle volume fraction depends on the vesicle radius and can be anywhere from 2 to 10 times higher than the corresponding lipid volume fraction for standard vesicle sizes created by extruding lipid solutions [10]; [11].

Because most of the volume of lipid vesicles is actually composed of solvent, seemingly low lipid concentrations correspond to significantly higher vesicle volume fractions and closer face-to-face distances between their surfaces than may be expected. These high volume fractions can lead to both direct and indirect solvent mediated hydrodynamic interactions between the particles, as also seen in concentrated colloid [12, 13], micelle [14], and protein solutions [15]; [16]. However, because the lipid membrane shell is soft and undergoes thermal fluctuations, lipid vesicles may have additional repulsive interactions not seen in typical “hard” colloidal systems. In addition to the excluded volume and electrostatic repulsive interactions considered in hard colloidal particle systems, lipid vesicles may also have a hydration repulsion seen at very short distances between membranes, such as seen in multilamellar stacks at low hydration levels and close interlamellar repeat distances [17]; [18], as well as undulation repulsion interactions at relatively long distances that are essential for stabilizing membrane stacks [19]. Importantly, the undulation repulsion force is relatively long range, and direct inter-vesicle interactions may be apparent at lower volume fractions than expected for hard sphere systems. Lipid vesicles are also deformable, which could further affect their interactions at high vesicle volume fractions. As such, it is both fundamentally interesting and practically important to consider the effects of such interactions on the properties of concentrated lipid vesicle solutions.

Here we study the interactions between vesicles as well as the effects of these interactions on the vesicle diffusion and membrane fluctuations in concentrated lipid vesicle solutions prepared with the well-characterized lipid, 1-stearoyl-2-oleoyl-sn-glycero-3-phosphocholine (18:0–18:1 PC, SOPC). We study the direct interactions in concentrated vesicle solutions with small angle neutron scattering (SANS) and show that there are measurable interactions at vesicle volume fractions,  $\phi_V \approx 0.02$ , which corresponds to a seemingly low lipid mass fraction,  $m_L \approx 0.004$  or  $c_L \approx 4$  mg/mL. These direct interactions affect the vesicle diffusion measured at large length scales, such as the length scales accessed with dynamic light scattering (DLS), a technique commonly used to determine the size and size distribution of lipid vesicle preparations. The measured vesicle diffusion constants are further affected by indirect hydrodynamic interactions. While these direct and indirect interactions affect the vesicle diffusion, they do not affect the membrane fluctuation dynamics measured at smaller length scales and faster time scales with neutron spin echo (NSE) spectroscopy over a wide range of  $\phi_V$  that spanned more than two orders of magnitude in lipid

concentration. Together, the results of the combined structural and dynamical scattering methods have important implications for extracting reliable information about the lipid vesicle population from different characterization techniques as well as practical implications for designing efficient scattering experiments.

## 2 MATERIALS AND METHODS

### 2.1 Materials

1-stearoyl-2-oleoyl-sn-glycero-3-phosphocholine (18:0–18:1 PC, SOPC) was purchased from Avanti Polar Lipids. Deuterium Oxide (D<sub>2</sub>O, 99.9% D) was purchased from Cambridge Isotopes. All materials were used without further purification.

### 2.2 Vesicle Preparation

Unilamellar vesicles were prepared by extrusion following protocols in literature [11]. The dry lipid powder was dispersed in D<sub>2</sub>O at a concentration of  $\approx 110$  mg lipid per mL of D<sub>2</sub>O, which corresponded to a lipid weight fraction of  $m_L \approx 0.09$ , and the lipid was hydrated at room temperature to form a suspension of multilamellar vesicles. The multilamellar vesicle solution was subjected to four freeze-thaw cycles and then extruded sequentially through 400 nm (15 ×), 200 nm (15 ×), 100 nm (15 ×), and 50 nm (41 ×) filters at  $\geq 30^\circ\text{C}$ . All samples were prepared by diluting the concentrated unilamellar vesicle stock solution with D<sub>2</sub>O.

### 2.3 Small Angle Neutron Scattering (SANS)

SANS measurements were performed at the NIST Center for Neutron Research (NCNR) on the very small-angle (vSANS) instrument [20]. Measurements were performed with neutron wavelengths ( $\lambda$ ) of 0.6 or 1.1 nm with a wavelength distribution ( $\Delta\lambda/\lambda$ ) of 0.12. The front and middle detector carriages were positioned at 1 and 5 m or 4.5 and 18.5 m, for the 0.6 and 1.1 nm wavelength configurations, respectively, giving access to a combined  $q$ -range of  $0.01 \text{ nm}^{-1} < q < 4 \text{ nm}^{-1}$ , where  $q$  is the magnitude of the scattering vector and is defined as  $q = 4\pi/\lambda \sin(\theta/2)$  with scattering angle,  $\theta$ . Data were collected at  $30.0 \pm 0.2^\circ\text{C}$ . SANS data were reduced using the macros provided by NIST [21] and analysed with the vesicle form factor model in SasView [22].

For homogeneous, monodisperse particles, the measured intensity is given by

$$I(q) = \phi \Delta\rho^2 V P(q) S(q) \quad (1)$$

In which  $\phi$  is the volume fraction of particles,  $\Delta\rho$  is the difference in the neutron scattering length density ( $\rho$ ) between the particle and the surrounding medium,  $V$  is the particle volume,  $P(q)$  is the form factor that describes the shape of the particles and  $S(q)$  is the structure factor that describes the interactions between the particles. At infinite dilution,  $S(q) \rightarrow 1$ , so that  $I(q) = \phi \Delta\rho^2 V P(q)$ .

Importantly,  $S(q)$  as described in Eq. 1 is only strictly true for homogeneous, monodisperse, spherical particles with isotropic interactions. Estimating the apparent structure factor for particle

systems with elongated or anisotropic shapes, anisotropic interactions, or large polydispersities requires using a static decoupling approximation as discussed in several works in literature [23–26]. For the polydisperse vesicles studied here, the measured intensity is more accurately described in terms of an apparent structure factor,  $S'(q)$ , such that

$$I(q) = \phi \Delta \rho^2 VP(q) S'(q) \quad (2)$$

Assuming that the particle form factor is not a function of the concentration (*i.e.* the particle shape is not changing), the only thing that will change with  $\phi$  is  $S'(q)$  and,

$$\frac{I(q, \phi)}{I(q, \phi_0)} = \frac{\phi \Delta \rho^2 VP(q) S'(q)}{\phi_0 \Delta \rho^2 VP(q)} = \frac{\phi}{\phi_0} S'(q) \quad (3)$$

Where  $\phi_0$  corresponds to a particle volume fraction low enough that the interactions between particles are negligible (*i.e.*  $S'(q) \rightarrow 1$ ). Accordingly,  $S'(q)$  can be estimated from the measured scattering intensity as

$$S'(q) \approx \frac{\phi_0}{\phi} \frac{I(q, \phi)}{I(q, \phi_0)} \quad (4)$$

There are a few additional caveats about **Eq. 4**. The first is that the expression assumes that the flat incoherent background has been subtracted from the measured data. While the subtraction is not strictly necessary for comparing the data at low  $q$ , the incoherent background will vary with particle concentration, and the scaled data will not overlap at high  $q$  if the incoherent background is not subtracted from measured intensity. **Eq. 4** also assumes that the particle form factor does not change with concentration, which is not necessarily true for all self-assembled systems. For example, SANS studies of microemulsions showed that the droplets became more monodisperse with increasing concentration, and the structure factor was extracted from the data by varying the scattering length density contrast ( $\Delta\rho$ ) at constant  $\phi$  using the relative form factor method [27]; [28].

## 2.4 Dynamic Light Scattering (DLS)

Dynamic laser light scattering measurements were performed with an LS Instruments AG (Fribourg, Switzerland) 3D cross-correlation laser light scattering system. A 500 mW Cobolt solid state laser provided 532 nm wavelength light ( $\lambda_o$ ) used under Vv polarization conditions. The scattered intensity ( $I$ ) was detected as a function of angle ( $\theta$ ) from 20.0° to 140.0° in steps of 10° *via* a precision goniometer by two optical fiber-coupled avalanche photodiodes under 2D pseudo-cross correlation mode. Measurements were performed with samples in cleaned, dust-free and capped glass cylindrical cells placed in a refractive index-matching vat of decalin that was thermostatically controlled to 30.00°C ± 0.01°C with an external recirculating bath.

For vesicle volume fractions up to  $\phi_V = 0.15$ , the measured normalized intensity-intensity time correlation functions ( $g^{(2)}(q, t) - 1$ ) were well fit by a single exponential with decay rate,  $\Gamma$ , according to

$$g^{(2)}(q, t) - 1 = (A \exp(-\Gamma t))^2, \quad (5)$$

Where  $A$  is the amplitude. The corresponding effective diffusion coefficients,  $D_{eff}$ , were calculated according to  $D_{eff} = \Gamma q^2$ , where for DLS,  $q = 4\pi n/\lambda_o \sin(\theta/2)$  in which  $n$  is the refractive index of the solution and assumed to be equal to that of the D<sub>2</sub>O solvent,  $n = 1.3282$ .

For dilute concentrations with no measurable interactions between vesicles,  $\phi_V \leq 0.016$  where  $S'(q) = 1$ , the translational diffusion coefficients,  $D$ , were calculated from the slopes of plots of  $\Gamma$  vs  $q^2$ . The diffusion coefficient at infinite dilution,  $D_0$ , was then calculated by extrapolating the measured  $D$  values to a concentration of zero using a virial expansion,

$$D = D_0 (1 + k\phi) \quad (6)$$

Where  $k$  is a constant [29].

The corresponding hydrodynamic radius ( $R_H$ ) was calculated from  $D_0$  according to the Stokes-Einstein relation,

$$R_H = \frac{k_B T}{6\pi\eta D_0} \quad (7)$$

Where  $\eta$  is the known solvent viscosity,  $k_B$  is the Boltzmann constant, and  $T$  is temperature.

For the highest concentration vesicle solution,  $\phi_V = 0.28$ , the measured correlation functions did not follow a single exponential decay. These data were fit using a combination of two exponential decay processes,

$$g^{(2)}(q, t) - 1 = (A_{fast} \exp(-\Gamma_{fast} t) + A_{slow} \exp(-\Gamma_{slow} t))^2 \quad (8)$$

Where  $A_{fast}$  and  $A_{slow}$  are the amplitudes of fast and slow modes, and  $\Gamma_{fast}$  and  $\Gamma_{slow}$  are their relaxation rates. The fast decay process was attributed to the effective diffusion coefficient of the vesicles,  $D_{eff} = \Gamma_{fast} q^2$ .

In concentrated solutions at the short time limit, the diffusion of one particle is influenced by the presence of its neighbors by both direct and indirect interactions. These effects are described as,

$$\frac{D_0}{D_{eff}(q)} = \frac{S'(q)}{H(q)} \quad (9)$$

Where  $D_0$  is the diffusion coefficient at infinite dilution,  $S'(q)$  is the apparent static structure factor measured by a technique like SANS and accounts for the direct interactions between particles, and  $H(q)$  is the hydrodynamic factor that accounts for the indirect solvent mediated interactions between particles [12].

There also are two diffusion processes to consider that depend on the length scale probed with the measurement technique relative to the size of the particles. At low  $q$  values (large length scales) relative to the size of the particles, the measured diffusion coefficient describes the relaxation of a concentration gradient, or the collective diffusion coefficient ( $D_c$ ). At high  $q$  values (small length scales) relative to the size of the particles, the measured diffusion coefficient describes the trajectory of a single tracer particle among the other particles and is called the self diffusion coefficient ( $D_s$ ).

Assuming the vesicles arrange on a BCC lattice, the  $q$  value characteristic of the particle spacing can be related to the particle volume fraction according to Bragg's law, where  $q_m = (6\pi^2\phi)^{1/3}/R$ , where  $q_m$  corresponds to the scattering vector at which  $S'(q)$  has a maximum intensity,  $\phi$  is the volume fraction of particles, and  $R$  is the particle radius [30]. The measured effective diffusion coefficient,  $D_{eff}$ , strongly depends on  $q$  in the vicinity of  $q_m$  [31]; [30]. However, at  $q \ll q_m$  such as the  $q$  values typically measured with DLS, the measured diffusion coefficient is the collective diffusion coefficient  $D_c$ . Likewise, at  $q \gg q_m$ , such as the  $q$  values probed with neutron spin echo (NSE) spectroscopy, diffusion is described by the self diffusion coefficient,  $D_s$ .

## 2.5 Neutron Spin Echo Spectroscopy (NSE)

NSE measurements were performed on the NGA NSE Instrument at the NCNR using  $\lambda$  of 0.8 or 1.1 nm with  $\Delta\lambda/\lambda \approx 0.2$  to access a  $q$ -range of  $0.4 \text{ nm}^{-1} < q < 1.1 \text{ nm}^{-1}$  and Fourier times ( $t$ ) from 0.01 to 100 ns [33]. The temperature was controlled at 30.0°C within 0.5°C with a recirculating bath during the measurements. The data were corrected for the instrument resolution by measuring the elastic scattering from a carbon resolution standard as well as the contributions from the  $D_2O$  solvent background to give the normalized intermediate scattering function ( $I(q,t)/I(q,0)$ ) using the DAVE software package [34].

For unilamellar lipid vesicle solutions, the intermediate scattering function measured with NSE can contain contributions from both the vesicle diffusion and the collective membrane motions, depending on the vesicle size and the dynamic range measured during the NSE experiment. Assuming that the dynamic contributions from the vesicle diffusion and membrane fluctuations are independent (*i.e.* the vesicle diffusion does not depend on the rate at which the membrane is fluctuating and *vice versa*), then their contributions to the intermediate scattering function are multiplicative,

$$I_{measured}(q,t) = I_{diffusion}(q,t) \times I_{fluctuations}(q,t) \quad (10)$$

Where  $I_{measured}(q,t)$  is the intermediate scattering function measured for a lipid vesicle solution and  $I_{diffusion}(q,t)$  and  $I_{fluctuations}(q,t)$  are the contributions from the vesicle diffusion and membrane fluctuations, respectively.

Note that polydispersity can also affect the measured intermediate scattering function,  $I_{measured}(q,t)$ , and a dynamic decoupling approximation may be needed to interpret NSE data, particularly when  $S'(q) \neq 1$  in the  $q$  range probed with NSE [26]. However, because NSE measures dynamics at short length scales where  $q \gg q_m$ ,  $S'(q) = 1$  in the NSE  $q$  range studied here, and we do not need to use a decoupling approximation to interpret these NSE data from lipid vesicle solutions.

Also, because NSE measures dynamic processes at short time scales and small length scales where  $q \gg q_m$ , we assume that the vesicle diffusion is described by the self diffusion coefficient,  $D_s$ . At these short length scales and high  $q$  values,  $S'(q) \rightarrow 1$ ; however, the hydrodynamic interactions still affect the self diffusion

**TABLE 1** | Summary of lipid concentrations and corresponding vesicle volume fractions in the studied solutions.

$c_L$ (mg/ml) <sup>a</sup>	$x_L$ (mmol/L) <sup>b</sup>	$m_L$ <sup>c</sup>	$\phi_V$ <sup>d</sup>	$q_m$ (nm <sup>-1</sup> ) <sup>e</sup>
109.8	139.3	0.090	0.28	0.060
48.9	62.0	0.042	0.15	0.049
8.1	10.3	0.0073	0.031	0.029
4.1	5.2	0.0037	0.016	0.023
2.5	3.2	0.0022	0.010	0.020
1.9	2.5	0.0018	0.0076	0.018
1.0	1.3	0.00090	0.0039	0.014
0.5	0.63	0.00045	0.0020	0.012

<sup>a</sup>Mass concentration of lipid in  $D_2O$  with units of mg/ml.

<sup>b</sup>Molar concentration of lipid in  $D_2O$  with units of mmol/L (mM).

<sup>c</sup>Mass fraction of lipid in solution.

<sup>d</sup>Volume fraction of vesicles calculated using  $R_H = 42.4 \text{ nm}$ ,  $d_b = 4.0 \text{ nm}$ , and a lipid volume,  $V_L = 1.309 \text{ nm}^3$  for SOPC at 30°C [32].

<sup>e</sup>Calculated  $q$  value corresponding to the maximum intensity in  $S'(q)$  assuming  $q_m = (6\pi^2\phi_V)^{1/3}/R_H$  [30].

coefficient,  $D_s(q)$ . The hydrodynamic effects in the limit that  $q \rightarrow \infty$  are well characterized in hard sphere systems where it has been shown that

$$\frac{D_s}{D_0} = 1 - 1.832\phi - 0.219\phi^2 + \mathcal{O}(\phi^3) \quad (11)$$

Where **Eq. 11** has been shown to be accurate for  $\phi \leq 0.3$ , and  $D_s/D_0$  is not expected to deviate significantly from **Eq. 11** for the range of vesicle volume fractions studied here. [31]; [35].

Given that the  $q$  values measured with NSE are an order of magnitude larger than the calculated  $q_m$ , we assume that the conditions of  $q \rightarrow \infty$  are met and  $D_s$  can be estimated by **Eq. 11** so that  $I_{diffusion}(q,t)$  simplifies to

$$I_{diffusion}(q,t) = \exp(-D_s q^2 t) \quad (12)$$

Where  $D_s$  was calculated from **Eq. 11** using the experimentally determined  $D_0$  value and the corresponding  $\phi_V$  values listed in **Table 1**.

Most NSE experiments are sensitive to the collective height fluctuations normal to the plane of the membrane, *i.e.* bending fluctuations at a constant membrane thickness [36]; [37]. Specific sample geometries and/or deuteration schemes also can be used to study other dynamic modes with NSE such as in-plane dynamics [38], collective thickness fluctuations [39], and the lipid acyl tail dynamics [40]. Importantly, these fluctuations are equilibrium processes, a direct consequence of lipid membranes being soft, and are therefore controlled by the physical properties of the bilayer and surrounding solvent. For bending fluctuations, the measured  $I_{fluctuations}(q,t)$  should be due to relaxation of the collective height fluctuations with a characteristic relaxation rate that is a balance between the ease of bending the lipid membrane (*i.e.* the membrane stiffness) and the time needed for the associated membrane deformation to relax (*i.e.* the dissipation).

The seminal work by Helfrich describes thermal bending fluctuations by treating the membrane as a thin, structureless sheet [19]. For a tensionless membrane, this treatment predicts



that the membrane fluctuations are hydrodynamically damped with a relaxation rate determined by balance of the membrane bending modulus ( $\kappa$ ) and the viscosity of the surrounding solvent ( $\eta$ ). Zilman and Granek incorporated the dynamic height correlation function predicted by the Helfrich model into the dynamic structure factor and showed that,

$$I_{\text{fluctuations}}(q, t) = \exp(-(\Gamma_{ZG}t)^{2/3}) \quad (13)$$

Where the characteristic relaxation rate,  $\Gamma_{ZG}$ , is given by

$$\Gamma_{ZG} = 0.025\gamma_k \frac{k_B T}{\eta} \sqrt{\frac{k_B T}{\kappa}} \quad (14)$$

And  $\gamma_k \approx 1$  for  $\kappa \gg k_B T$  as expected for most phospholipid membranes [41,42].

The work by Zilman and Granek predicted two important scaling relationships for the membrane dynamics measured at length scales greater than the bilayer thickness and less than the characteristic size scale of the higher order membrane structure ( $\xi$ ), which corresponds to the vesicle radius for unilamellar vesicles. The first is that the intermediate scattering function decays as a stretched exponential with a stretching exponent of  $2/3$  (Eq. 13), and the second is that the corresponding relaxation rate,  $\Gamma_{ZG}$ , scales with  $q^3$  (Eq. 14). Both predicted scaling relationships were confirmed experimentally in early NSE studies of lipid membranes [43]; [44]; however, the extracted values of  $\kappa$  were an order of magnitude larger than those measured with other established characterization techniques.

Watson and Brown suggested that NSE measures the effective bending modulus,  $\tilde{\kappa}$ , originally proposed in work by Seifert and Langer, [45] rather than the bending modulus as traditionally defined by Helfrich. Their derivation showed that incorporating  $\tilde{\kappa}$  into the NSE data analysis framework only modified the expression for the relaxation rate measured with NSE such that,

$$\Gamma_{ZG} = 0.025 \frac{k_B T}{\eta} \sqrt{\frac{k_B T}{\tilde{\kappa}}} q^3 \quad (15)$$

Where the effective bending modulus,  $\tilde{\kappa}$  is defined as  $\tilde{\kappa} = \kappa + 2d^2K_m$  in which  $d$  is the height of the neutral surface and  $K_m$  is the monolayer compressibility modulus [46]. The second additive term in the expression for  $\tilde{\kappa}$  accounts for dissipation within the membrane itself that is important to include at short length scales and fast time scales, where the lipids within the membrane do not fully relax during the time scale of the deformation. As such, it effectively takes more energy to deform the lipid bilayer on the short length scales and fast time scales accessible with NSE. It is now generally accepted that NSE is sensitive to  $\tilde{\kappa}$ , and values of  $\tilde{\kappa}$  range from (4–14) $\kappa$  [46]. However, calculating an absolute value of  $\kappa$  from  $\tilde{\kappa}$  measured in an NSE experiment is highly sensitive to the assumed values of  $d$  and  $K_m$ , and it is not clear how these constants depend on the membrane composition and experimental system [46]; [47]. While more work is needed to extract quantitative values of the membrane mechanical properties from the collective dynamics measured with NSE,

the relative trends in  $\tilde{\kappa}$  measured for vesicles with similar radii appear to be correct [48]; [49].

The NSE data presented here were fit considering contributions from both the vesicle diffusion and the membrane fluctuations following the Zilman-Granek treatment,

$$\frac{I(q, t)}{I(q, 0)} = \exp(-D_s q^2 t) \times \exp(-(\Gamma_{ZG}t)^{2/3}) \quad (16)$$

Where  $D_s$  was calculated using Eq. 11, and  $\Gamma_{ZG}$  was the only fit parameter. Eq. 16 assumes that the amplitude of the undulation motions is 1. While other works also include a  $q$ -dependent relative amplitude of the undulation motion [49], this additional term affects the absolute value of  $\Gamma_{ZG}$ , and consequently the absolute value  $\tilde{\kappa}$ , but not the relative differences in values between samples. As such, here we assume that the amplitude is 1 for simplicity and focus on the relative changes between samples.

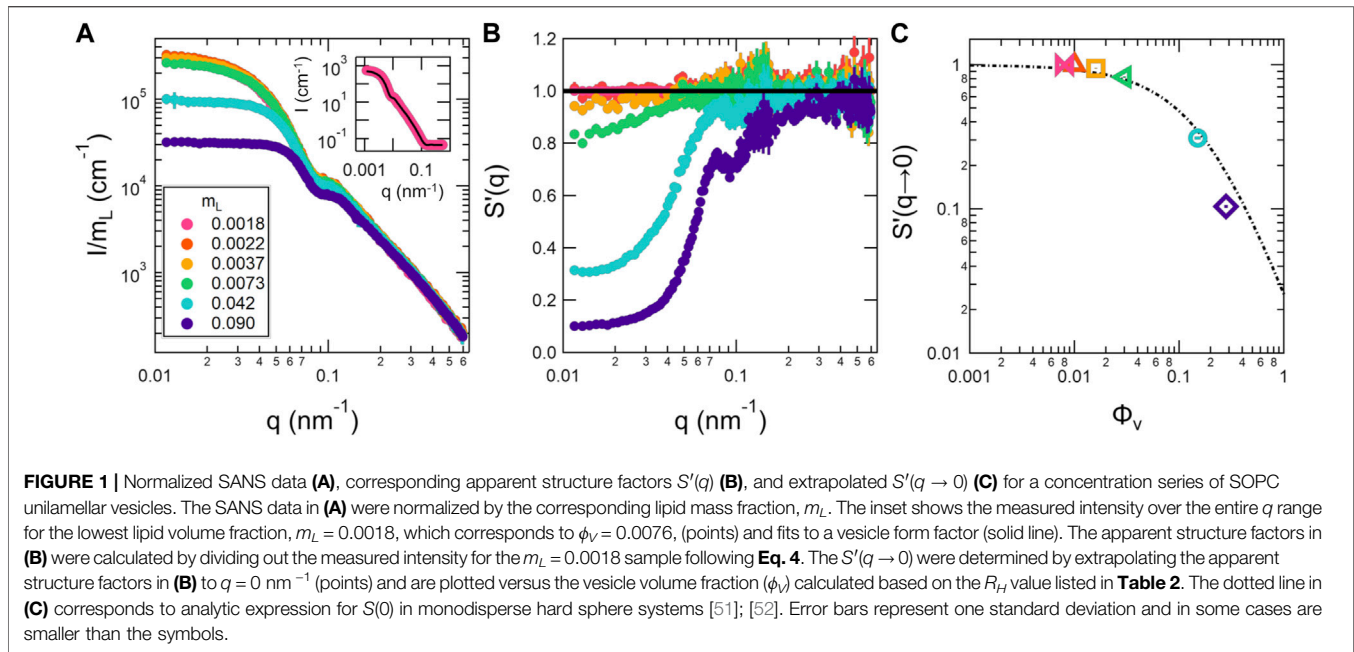
### 3 RESULTS

All presented data were collected on samples made by diluting the same concentrated stock solution of SOPC unilamellar vesicles. The lipid concentration in the most dilute sample,  $m_L = 0.00045$ , corresponded to a concentration of  $\approx 0.6$  mmol/L (mM) and was at least five order of magnitude higher than the critical micelle concentration (CMC) of lipids with at least 16 carbons in the acyl tail which is on the order of  $10^{-6}$  mM or lower [50]. Given that the studied lipid concentrations were well above the CMC and that lipid exchange rates are very slow, we assumed that the lipid vesicle structures were stable upon dilution and any differences in the SANS, DLS, and NSE data were attributed to concentration effects, and not structural differences, in the vesicle solutions.

#### 3.1 Interactions

SANS is most often used to extract detailed information on the structure of the lipid membrane [37]. This information is contained in the data at short length scales at high  $q$  values, while the data at larger length scales and smaller  $q$  values contain information about the vesicle size and polydispersity as well as the direct interactions between vesicles. Shown in **Figure 1** are SANS data measured for a dilution series of unilamellar SOPC vesicles. The inset in **Figure 1A** shows the data for the lowest measured lipid concentration,  $m_L = 0.0018$ , which corresponded to a vesicle volume fraction of  $\phi_V = 0.0076$ , fit with a form factor for unilamellar vesicles. The form factor fits the data well over the entire measured  $q$ -range, supporting that there was not a measurable structure factor in the data at this low lipid concentration. The fit results gave a bilayer thickness of  $d_b \approx 4$  nm and a number-averaged vesicle radius of  $R_n \approx 31$  nm with polydispersity,  $\sigma_R = 0.34$  as summarized in **Table 2**.

The SANS data plotted in **Figure 1A** were normalized by  $m_L$ . The normalized intensities overlapped in all samples at  $q \geq 0.2$  nm<sup>-1</sup> but systematically decreased at  $q \leq 0.1$  nm<sup>-1</sup> with increasing



**TABLE 2** | Summary of lipid vesicle characterization results from SANS and DLS.

T (°C)	$d_b$ (nm)		$R_n$ (nm)	$\sigma_{R_n}$	$R_z$ (nm)	$R_H$ (nm)
	SANS	literature <sup>a</sup>	SANS <sup>b</sup>	SANS <sup>c</sup>	SANS <sup>d</sup>	DLS
30	4.0 ± 0.02	4.0 ± 0.8	30.6	0.34	40.3	42.4

<sup>a</sup>Literature value for the SOPC bilayer thickness [32].

<sup>b</sup>Number average vesicle radius assuming a Schulz distribution of vesicle sizes.

<sup>c</sup>Polydispersity in vesicle radius assuming a Schulz distribution of sizes.

<sup>d</sup>Z-average vesicle radius (intensity-weighted) calculated from  $R_n$  and  $\sigma_{R_n}$ . Uncertainties represent one standard deviation.

concentration for  $m_L \geq 0.0041$ , a hallmark of a repulsive structure factor in scattering data.

Assuming that the vesicle form factor did not change upon dilution, the apparent structure factors,  $S'(q)$ , were calculated by dividing the normalized intensity for the different samples with the normalized intensity measured for the most dilute sample,  $m_L = 0.0018$ , following Eq. 4. The calculated apparent structure factors are plotted in Figure 1B, where the solid line corresponds to  $S'(q) = 1$ , the value expected if the inter-vesicle interactions were negligible. The apparent structure factors deviated from 1 at low  $q$  in the SANS data measured for solutions with lipid mass fractions as low as  $m_L = 0.0041$ , which corresponded to  $\phi_V = 0.016$ , and the values of  $S'(q \rightarrow 0)$  further decreased with increasing concentration.

The value of  $S'(q \rightarrow 0)$  is considered the thermodynamic limit and directly related to the isothermal osmotic compressibility of the solution,

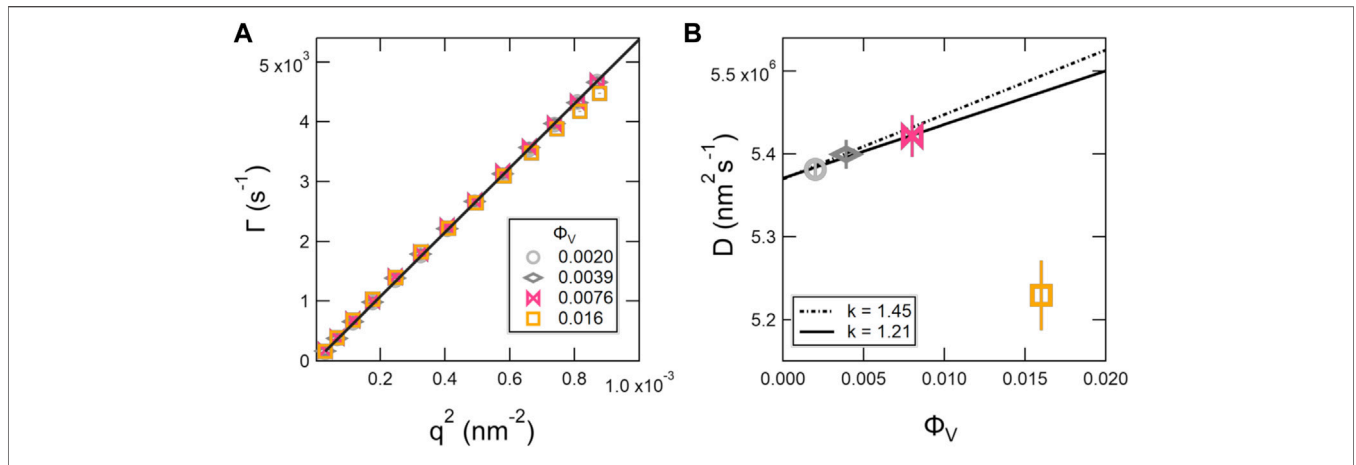
$$\frac{1}{S(q \rightarrow 0)} = \frac{1}{k_B T} \left( \frac{\delta \Pi}{\delta \mathcal{N}} \right)_T \quad (17)$$

Where  $\Pi$  is the osmotic pressure and  $\mathcal{N}$  is the number density of particles in solution. The  $S'(q \rightarrow 0)$  values were estimated by a linear extrapolation of the data for  $q \leq 0.015 \text{ nm}^{-1}$  to  $q = 0 \text{ nm}^{-1}$  and are plotted as the open symbols versus the  $\phi_V$  in Figure 1C. The analytic expression for monodisperse hard spheres,  $S(0)^{-1} = 1 + 8\phi + 30\phi^2 + \dots$  is also shown in Figure 1C as the dotted line for comparison [51]; [52].

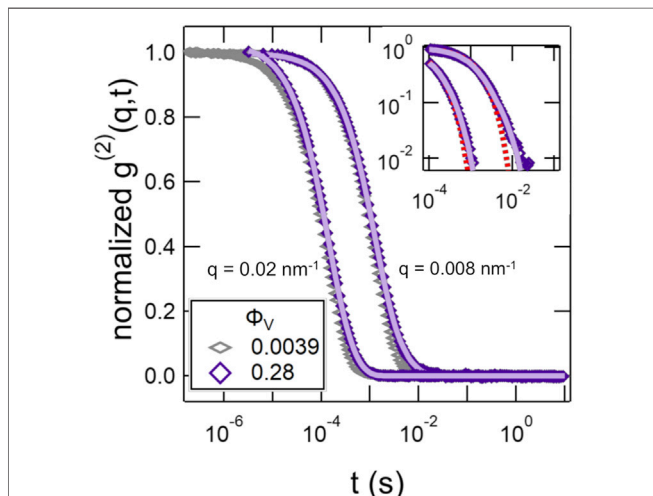
### 3.2 Diffusion

DLS is a bench top characterization technique widely used to determine the size of extruded vesicles; however, DLS does not directly measure the size of the particles in solution, but rather their diffusion. The raw data are intensity-intensity correlation functions that decay due to motions of the particles with a relaxation rate,  $\Gamma$ . Shown in Figure 2A are the relaxation rates,  $\Gamma$ , extracted from single exponential fits to the measured intensity-intensity correlation functions plotted versus  $q^2$  for dilute SOPC vesicle solutions. The data for  $\phi_V \leq 0.0076$  ( $m_L \leq 0.0018$ ) followed the expected  $q^2$  scaling for simple diffusion, while the data for  $\phi_V = 0.016$  ( $m_L = 0.0037$ ) deviated from the expected scaling, particularly at high  $q$ . The corresponding diffusion coefficients,  $D$ , extracted from the best fits to  $\Gamma = Dq^2$  are plotted versus  $\phi_V$  in Figure 2B.

The data for  $\phi_V \leq 0.0076$  in Figure 2B were fit with Eq. 6 to determine the diffusion coefficient at infinite dilutions,  $D_0$ . The best fit to the data is shown as the solid line in Figure 2B and gave values of  $D_0 = 5.37 \times 10^6 \text{ nm}^2 \text{ s}^{-1}$  and  $k = 1.21$ . The corresponding  $R_H$  value calculated from  $D_0$  was 42.4 nm, where  $R_H$  is an intensity-weighted or Z-averaged radius, such that the larger particles are weighted more heavily. Comparing the  $R_H$  with  $R_z$  calculated from the Schulz distribution of vesicle sizes extracted from the SANS data analysis showed that the values were in good agreement (Table 2). Also note that the best fit value



**FIGURE 2** | Decay constants,  $\Gamma$  extracted from single exponential fits to the DLS correlation functions plotted versus  $q^2$  **(A)** and corresponding diffusion coefficients,  $D$  **(B)** for dilute SOPC lipid vesicle solutions. The solid line in **(A)** is a representative linear fit to  $\Gamma = Dq^2$  to determine the  $D$  values plotted in **(B)**. The lines in **(B)** correspond to fits with a virial expansion following Eq. 6 to determine diffusion coefficient at infinite dilution,  $D_0$ , where the solid line is the best fit to the data and the dashed line is the fit with the scaling constant reported for hard spheres,  $k = 1.45$  [29]. The error bars represent one standard deviation and in some cases are smaller than the symbols.



**FIGURE 3** | Normalized intensity-intensity correlation functions,  $g^{(2)}(q, t)$  at two representative  $q$  values for SOPC vesicle solutions with lipid mass fractions of  $\phi_V = 0.0039$  and  $0.28$ . The solid lines are the fits to the data at  $\phi_V = 0.28$  with a double exponential decay using Eq. 8. The inset is the data for  $\phi_V = 0.28$  plotted on a log-log scale and compares the fits with a single (dotted line) and double exponential (solid line) decay.

of  $k = 1.21$  was slightly lower than value reported for hard sphere solutions where  $k = 1.45$  for collective diffusion [29], and the corresponding predictions for hard spheres are shown as the dotted line in Figure 2B.

The intensity-intensity correlation functions deviated from a single exponential decay at the highest vesicle volume fraction,  $\phi_V = 0.28$ , and were better fit by a sum of two exponential decays given by Eq. 8. The normalized correlation functions for  $\phi_V = 0.0039$  and  $0.28$  are compared in Figure 3. A second slower mode

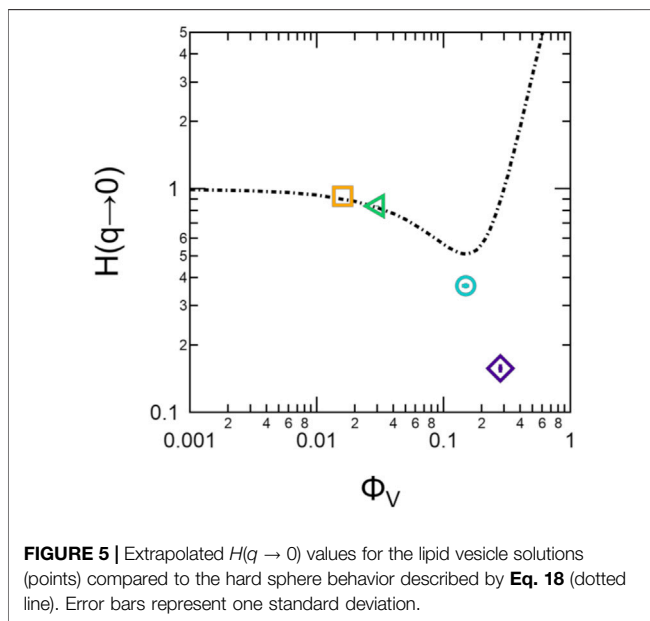
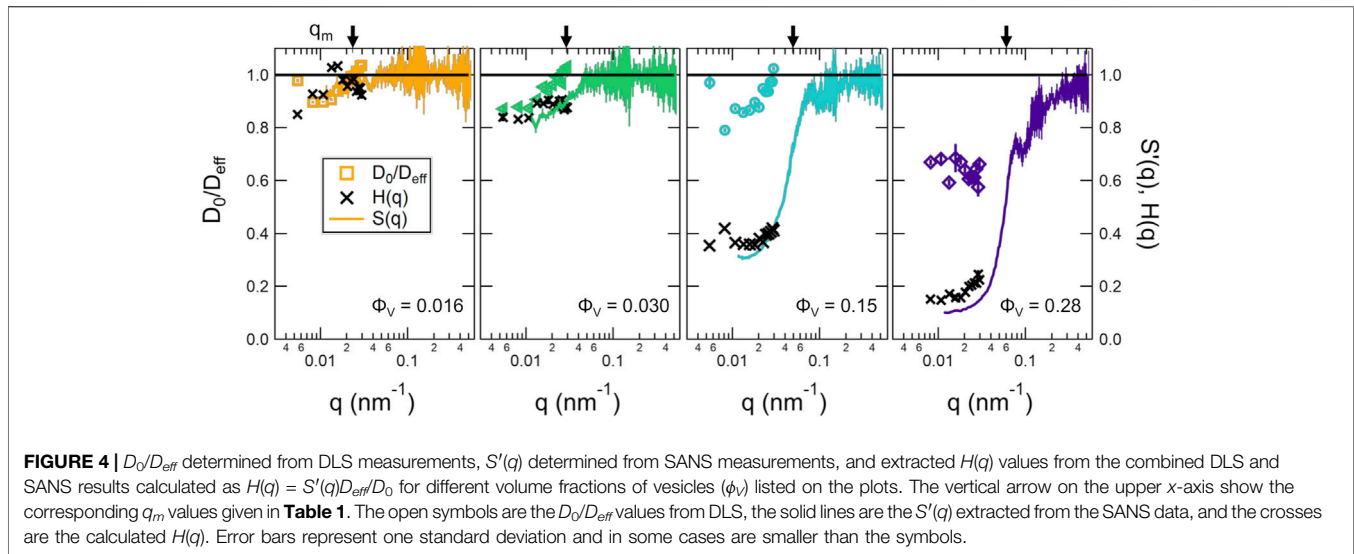
was seen at longer times in the data for  $\phi_V = 0.28$  that was not captured by a single exponential decay (Figure 3 inset).

The short time diffusion of the vesicles also was affected by the inter-vesicle interactions at high  $\phi_V$ . Shown in Figure 4 are plots of  $D_0$  normalized by the effective diffusion coefficients,  $D_{eff}(q)$  from the DLS measurements and the apparent structure factors determined from the SANS measurements in the previous section. The data showed that even at a vesicle volume fraction of  $\phi_V = 0.016$ , which corresponded to  $m_L \approx 0.0037$ , there were measurable effects of the solvent-mediated, indirect vesicle interactions on the DLS data. These indirect solvent mediated interactions are described by the hydrodynamic factor,  $H(q)$ . The corresponding  $H(q)$  were calculated from the combined SANS and DLS data as  $H(q) = S'(q)D_{eff}/D_0$  for the different volume fractions and also are plotted for the different lipid concentrations in Figure 4.

The DLS data measured at the largest length scales corresponded to  $q$  values that were approximately an order of magnitude lower than  $q_m$  for lipid vesicles, which should correspond to the collective diffusion of the vesicles given by  $D_c$ . While there are only a few studies that have measured  $H(q)$  for soft lipid vesicles [30]; [53], these effects on the collective diffusion of hard colloidal particles are well studied. Work by Chichocki et al. calculated  $H(q)$  for hard spheres considering three-particle interactions and showed that in the limit of  $q = 0$ , [54]; [31].

$$H(0) = 1 - 6.546\phi + 21.918\phi^2 + \mathcal{O}(\phi^3). \quad (18)$$

The extrapolated values of  $H(q \rightarrow 0)$  for the lipid vesicle solutions and the calculated values for hard spheres following Eq. 18 are compared in Figure 5. The measured  $H(q)$  for the soft vesicles solutions agreed well with the predictions for hard sphere solutions at low volume fractions, but began to deviate for  $\phi_V \geq 0.15$ .



### 3.3 Membrane Fluctuations

Unilamellar vesicle solutions are often used as model systems for the study of membrane fluctuations on the nanometer lengths scales and nanosecond time scales using neutron spin echo spectroscopy. The contrast between protiated lipids and the deuterated surrounding solvent makes most NSE experiments sensitive to the thermal motions normal to the plane of the bilayer. The relaxation times of these collective height fluctuations, which are also referred to as bending fluctuations, are related to the effective membrane rigidity,  $\tilde{\kappa}$ . Shown in **Figure 6A** are representative NSE data collected for the SOPC solutions with vesicle volume fractions of  $\phi_V = 0.016$  and  $0.15$ , which corresponded to lipid mass fraction of  $m_L = 0.0037$  and  $0.042$ , respectively. The normalized intermediate scattering functions,  $I(q,t)/I(q,0)$ , suggested that the dynamics were slower

in the higher lipid concentration sample, (*i.e.* the curves decay less). If the contributions from the vesicle diffusion were not taken into consideration when analyzing these NSE data, then the slower dynamics corresponded to an approximately 40% increase in  $\tilde{\kappa}$  value for  $\phi_V = 0.15$  compared to  $\phi_V = 0.016$ .

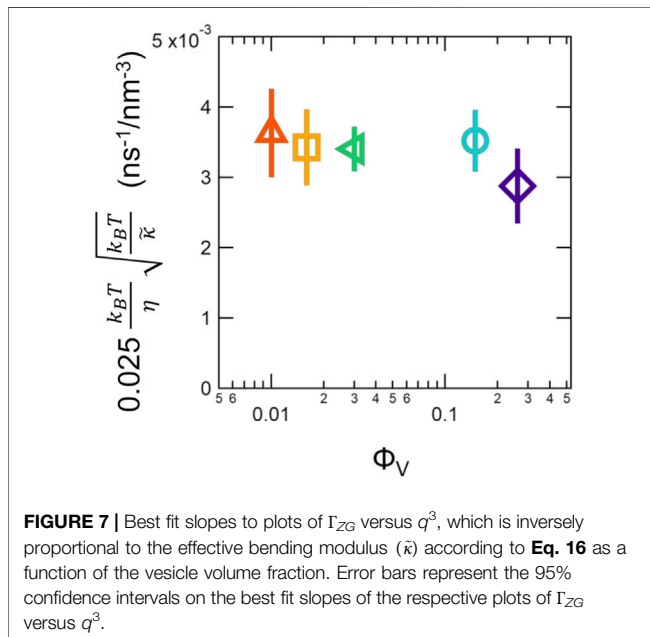
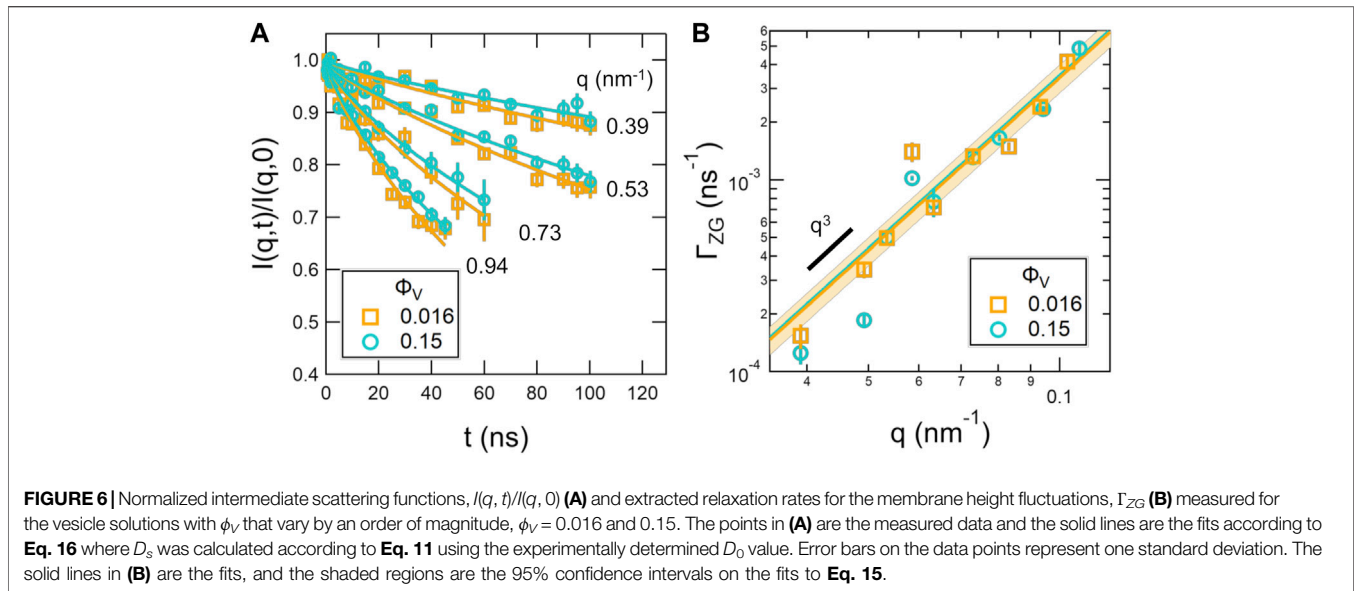
However, the slower dynamics were due to slower self diffusion of the lipid vesicles in the more concentrated sample, not an increase in membrane rigidity. The NSE data were fit assuming that  $D_s$  of the lipid vesicles could be estimated with **Eq. 11** for hard spheres using the experimentally determined  $D_0$  value. The corresponding fits to the NSE data considering both the vesicle diffusion and the membrane fluctuations described by the Zilman-Granek formalism (**Eq. 16**) are shown as the solid lines in **Figure 6A**. The extracted relaxation rates for the membrane fluctuations,  $\Gamma_{ZG}$ , and their fits to a  $q^3$  scaling are shown in **Figure 6B**. The 95% confidence intervals on the fits to the  $\Gamma_{ZG}$  values at the two concentrations overlapped as well as the corresponding values of  $\tilde{\kappa}$ .

The results showed that the membrane fluctuation dynamics were not affected over the wide concentration range studied here. The slopes of the fit to  $\Gamma_{ZG}$  vs  $q^3$  curves, which are proportional to  $\tilde{\kappa}^{-1/2}$ , are plotted versus  $\phi_V$  in **Figure 7**. The differences were not statistically significant within the uncertainty in the experimental data. Though it is important to note that the NSE data were fit using  $D_s$  values calculated according to **Eq. 11** for hard spheres, and the results in **Figure 5** suggest that hydrodynamic interactions between vesicles do not follow the same  $\phi$  dependence as hard spheres at high  $\phi$ . It may be possible that the values calculated with **Eq. 11** overestimate  $D_s$ , particularly for  $\phi_V = 0.28$ , which may also explain the slightly lower value of  $\tilde{\kappa}$  at this high volume fraction.

## 4 DISCUSSION

The combination of scattering methods used here clearly shows that there are both direct and indirect interactions between lipid vesicles in concentrated solutions. Understanding the effects of these interactions has practical implications for designing efficient





scattering experiments as well as extracting reliable information on the vesicle structure and dynamics from the resulting data. Studying these interactions also presents new opportunities for understanding the stability of lipid assemblies in solution as well as using liposomes as models of soft colloids. The practical and potential implications of the presented results are discussed in more detail below.

#### 4.1 Practical Implications for Designing Experiments and Interpreting Scattering Data

The solutions studied here contained unilamellar vesicles that were prepared by extruding a concentrated SOPC solution

through a filter with a nominal pore size of 50 nm. The SANS data measured for these solutions showed evidence of repulsive interactions between the vesicles at vesicle volume fractions as low as  $\phi_V \approx 0.016$ , which corresponded to a lipid mass fraction of  $m_L \approx 0.004$  or  $x_L \approx 5$  mM for the vesicles  $\approx 80$  nm in diameter studied here. Such interactions would appear at even lower lipid mass concentrations in solutions containing larger vesicles, as the volume fraction of the internal solvent core scales with the vesicle radius cubed,  $R^3$ .

Neglecting to properly account for the presence of interactions between vesicles and the associated structure factor, and only fitting small angle scattering data with a form factor, can impact the reliability of the extracted fit results. The peak in a structure factor often occurs at or near the same  $q$  value as the features in the form factor attributed to the vesicle size and size distribution, and as a result, likely can affect the fit values for the vesicle radius and corresponding polydispersity [55]. Not accounting for  $S(q)$  could also affect the fit results for the scattering length density contrast ( $\Delta\rho$  in Eq. 1), which is particularly important in studies aimed at extracting information about the composition of the bilayer, such as structural asymmetry or the location of incorporated peptides or small molecules, where the relevant information is contained in changes in  $\Delta\rho$ . In the dilute limit where  $S'(q) = 1$ , as  $q \rightarrow 0$ ,  $I(q \rightarrow 0) - B = I_0 = \phi(\Delta\rho)^2 V$ , where  $B$  is the incoherent background. As shown in Figure 1,  $S'(q)$  impacts the scattered intensity at low  $q$ , and therefore, fitting low  $q$  data with only a form factor will often underestimate the true  $I_0$  in vesicle solutions with repulsive interactions.

At high enough  $q$ -values relative to the overall vesicle radii,  $qR \gg 1$ , the measured SANS data overlapped, and the  $S'(q)$  correction was not needed to extract information regarding the bilayer thickness. It is important to note that “high”  $q$  will be relative to the radius of the vesicles. Recent work has shown that vesicle prepared by extrusion through a 100 nm filter often are not unilamellar [11], and using smaller filter pore sizes is one way

to ensure that the vesicle solution does not contain a small population of multilamellar vesicles that can affect the interpretation of scattering data at high  $q$ . However, as  $R$  decreases, the absolute value of  $q$  corresponding to  $qR \gg 1$  will increase. Another approach to ensuring that vesicles are unilamellar is to incorporate a small amount of charged lipid [11]. Incorporating charged lipids, particularly in unbuffered solutions, will further increase the repulsive interactions between vesicles and result in an even more pronounced structure factor in the data. To ensure that the  $q$ -range of interest needed to extract information about the bilayer structure is not affected by contributions from  $S'(q)$ , it is good practice to measure one or two dilutions of a sample to ensure that the data overlap.

In addition to the direct interactions measured with SANS, the indirect hydrodynamic interactions also affect the vesicle diffusion coefficient measured with DLS. The data in **Figure 4** show that the diffusion coefficients measured with DLS were affected by the structure factor seen in the SANS data as well as the solvent-mediated hydrodynamic interactions. Because  $D$  is directly used to calculate  $R_H$  (Eq. 7), not accounting for the potential effects of the direct and indirect interactions on the vesicle diffusion will directly impact the vesicle sizes estimated with DLS. It is also important to keep in mind that DLS data are highly sensitive to multiple scattering, especially for visually turbid solutions such as those at high lipid concentrations, which can also affect the reliability of the extracted information about the samples. Here the data were collected on a light scattering instrument specifically designed to correct for the effects of multiple scattering, but in general, DLS data collected on dilute samples ( $\phi_V \leq 0.1\%$  by volume) are more reliable. Like with SANS experiments, collecting DLS data for a dilution series ensures that the measured correlation functions and estimated diffusion coefficients are not changing significantly with concentration and improves the reliability of the results.

At the highest vesicle volume fraction,  $\phi_V \approx 0.3$ , the measured DLS data also did not follow a single exponential decay as shown in **Figure 3**. It is important to keep in mind that DLS measures the diffusion of particles in solution, and that the presence of two decay modes reflects two populations of diffusive processes in this case and not the presence of two vesicle size populations. Previous work by Pusey et al. showed that DLS data measured for even modestly polydisperse particles with a size distribution as small as  $\sigma \approx 0.1$  contained two independent modes with well separated decay times at high volume fractions [56]. The fast mode described the collective diffusion of the particles given by  $D_c$  and discussed above, while the second slower mode was attributed to the local position exchanges of particles with different sizes. At even higher volume fractions, colloidal solutions are also known to undergo caged diffusion, where the slow mode is associated with the time required for the cages formed by neighboring particles to break apart [13]; [57]. Similar behavior was also seen in concentrated lipid vesicle solution studied by Yu et al. in which they observed two distinct slow and fast populations with single-particle fluorescence tracking [58]. While diffusion in concentrated

solutions of hard colloidal particles is well studied, fewer studies have looked at this phenomenon in soft, deformable colloids, and this dynamical heterogeneity may be important for understanding not only diffusion in complex environments but also the rheological properties of concentrated solutions of soft particles.

While the interactions between vesicles affected the diffusion dynamics at large length scales, or equivalently small  $q$  values, the membrane fluctuation dynamics at the nanometer length scale and high  $q$  values measured with NSE were not affected over the entire studied concentration range. While there was a lipid concentration dependence in the measured intermediate scattering functions, as seen in **Figure 6A**, the analysis presented here suggests that these differences in the measured dynamics were due to differences in the vesicle diffusion dynamics, not the membrane fluctuations dynamics.

However, even at the lower vesicle volume fractions, the values of  $\tilde{\kappa}$  calculated from the NSE data using Eq. 15 were approximately an order of magnitude larger than expected. The extracted  $\tilde{\kappa}$  values were  $\approx 1000 k_B T$ . Estimating the effective bending modulus using values  $\kappa = 30 k_B T$  [59],  $K_m = 1/2K_A = 117 \text{ mN m}^{-1}$  [60], and  $d = d_c = 1.5 \text{ nm}$ , where  $d_c$  is the hydrocarbon tail thickness of the bilayer [48]; [32], gives  $\tilde{\kappa} = \kappa + 2d^2 K_m \approx 155 k_B T$ . The order of magnitude larger values determined from fitting the NSE data suggest that the dynamics are relaxing slower than predicted, even when the dissipation within the bilayer is taken into account. One possible reason for the discrepancy is that the NSE data in the present work were treated with the simplest method to account for the vesicle diffusion using Eq. 16. As recently highlighted in work by Hoffmann, it is also possible that the membrane fluctuations have a limited amplitude that depends on  $q$ , and accounting for the finite amplitude would give  $\tilde{\kappa}$  values more in line with what is expected based on values in literature [49]. There could also be other effects that are not accounted for in commonly used NSE data analysis frameworks for lipid vesicles. Work by Monkenbusch et al. showed that taking into account the finite radius of microemulsion droplets affected the calculated intermediate scattering function for surfactant membrane bending fluctuations and the value of the bending modulus estimated from experimental data [61]. Early NSE studies of soft microemulsions, surfactant membranes, and lipid vesicles also suggested that the local dissipation was slower than predicted due an increase in the effective solvent viscosity [62]; [43]; [44]. Nevertheless, the data in **Figure 7** indicate that the membrane fluctuation dynamics are not affected up to vesicle volume fractions of  $\phi_V \approx 0.30$ , where the face-to-face distances between the vesicles are order tens of nanometers.

Another promising implication of these results is that NSE measurements can be performed on higher lipid concentration solutions without affecting the membrane fluctuation dynamics, so long as the vesicle diffusion is properly taken into account. The measured intensity, and therefore the required measurement time, is directly related to the lipid concentration in the sample. The data for the  $\phi_V = 0.016$  and  $0.15$  samples in **Figure 6A** required measurements

times of  $\approx 16$  h and  $\approx 6$  h, respectively, yet the extracted values of  $\Gamma_{ZG}/q^3$  are the same within error. These results show that dilute, non-interacting samples are not necessarily needed for NSE measurements of membrane fluctuations. While the relevant lipid concentration and required NSE counting times will depend on the samples as well as the NSE instrument and instrument configurations used for the experiment, higher concentration samples in general will require shorter measurements times than more dilute samples, which would allow for efficient use of beam time at user facilities.

## 4.2 Potential Opportunities for Probing Interactions in Lipid Systems

Systematically measuring and analyzing the structure factors in concentrated lipid solutions also presents new opportunities to probe the interactions in lipid membrane systems. There are several elegant experimental methods for measuring the interaction potential between two materials as a function of separation distance; however, these methods often require samples that are at least microns in size [63]. Measuring such interactions between things that are nanometers in size is more experimentally challenging. The results presented here for concentrated lipid vesicle solutions highlight the utility of small angle scattering methods for characterizing the interactions between nanometer-sized structures in solution. Future studies aimed at measuring and modeling  $S'(q)$  in concentrated lipid vesicle solutions could provide further insights into the interactions potentials between the liposomes as well as the membrane properties that affect these interactions.

Quite interestingly, the shapes of the calculated apparent structure factors for the lipid vesicles shown in **Figure 1B** did not show the characteristic features seen in structure factors for hard colloidal systems. The structure factors for such systems that are well described by a hard sphere or single Yukawa interaction potential typically show a pronounced maximum where  $S'(q) > 1$  at  $q_m$  that corresponds to the interparticle spacing. The calculated  $S'(q)$  for the lipid vesicles did not show a clear maximum, though we do note that there were peaks at  $q \approx 0.08 \text{ nm}^{-1}$  in the apparent structure factors for  $m_L = 0.042$  and  $0.090$ , which corresponded to a length scale of  $2\pi/q \approx 80 \text{ nm}$ , or approximately the hydrodynamic diameter of the particles. One possible explanation for the atypical shape of the calculated  $S'(q)$  curves is the high polydispersity in the vesicle sizes ( $\sigma_{R_n} = 0.34$ ), and future experiments on more monodisperse vesicle samples would help provide further insights into the shape of the structure factor and the associated interaction potential.

Because the shape of calculated  $S'(q)$  for the lipid vesicles did not match those expected for typical colloidal systems, we were not able to fit the data with a common model and instead chose to compare the extrapolated  $S'(q \rightarrow 0)$  to predictions for hard spheres to gain additional insight into the strength of the repulsive interactions. The present data for  $S'(q \rightarrow 0)$  in

**Figure 1C** suggested that the interactions between lipid vesicles were more repulsive than hard spheres at high volume fractions. Previous measurements of  $S'(q)$  in unilamellar vesicle solutions have also noted deviations from hard sphere behavior [64]. The hard sphere structure factor only considers excluded volume interactions, *i.e.* two spheres can not occupy the same volume. The trends in apparent structure factors suggest that there are additional repulsive interactions in vesicle systems beyond what is predicted by hard sphere interactions. It is interesting to note that  $H(q)$  measured for the lipid vesicles followed the hard sphere predictions at  $\phi_V \leq 0.04$ , but also deviated at higher volume fractions. Previous studies by Haro-Pérez et al. of charged and uncharged liposome solutions have also shown that  $D_{\text{eff}}/D_0$  deviates from hard sphere behavior at vesicle volume fractions on the order of 0.03 [30]; [53]. These deviations in  $H(q)$  may suggest that the additional repulsive interactions also affect hydrodynamic interactions at high  $q$ .

Unlike hard spheres, lipid vesicles are soft and can deform in shape which may affect their interactions at high  $\phi$ . There are also two well-known repulsive interactions found between soft membranes but not colloidal particles: short range hydration repulsion and long range undulation repulsion [63]. The hydration repulsion at sub nanometer length scales prevents the membranes from sticking together [17]; [18], while the undulation repulsion is comparatively long range and arises from entropic confinement of two membranes when they are close together [19]. The strength of the undulation repulsion is therefore related to the membrane rigidity,  $\kappa$ , and the associated membrane fluctuations that are measured with NSE. The balance of these hydration and undulation repulsive forces with Van der Waals attraction is what determines the interlamellar spacing in multilamellar stacks. The undulation repulsion force is also responsible for the unbinding transition seen in multilamellar stacks at high temperatures [65–67] or in the presence of certain salts [68]; [69], and could also lead to increased repulsion between lipid vesicles at very high volume fractions and close face-to-face distances. While more much data at high  $\phi_V$  are needed, measuring the effects of lipid composition, temperature, and ionic strength of the surrounding solvent would provide further insights into the repulsive forces underlying the interactions between vesicles.

When combined with other techniques, measuring  $S'(q)$  in lipid-based assemblies may provide additional insights into the physicochemical properties of the system. For example, we recently showed that the changes in  $S'(q \rightarrow 0)$  measured with SANS and the membrane fluctuation dynamics measured with NSE showed the same temperature-dependent trends for charged lipid membranes, which were both due changes in the effective headgroup charge as the lipids melted [70]. Similarly, extending the  $q$ -range measured during small angle scattering measurements of lipid vesicles with added peptides, drugs, or other small molecules to lower  $q$ , would provide insights into how these additives not only affect the bilayer structure [37], but also the interactions between membranes.

## 5 SUMMARY

Here we show how seemingly low lipid mass concentrations result in measurable interactions between vesicles because the majority of the vesicle volume is composed of the solvent core, and therefore, the corresponding vesicle volume fraction is several times higher than the lipid volume fraction used to prepare the solution. These interactions were seen in solutions of zwitterionic SOPC vesicles  $\approx 80$  nm in diameter at lipid mass concentrations as low as  $\approx 5$  mM, and the onset of interactions would shift to even lower lipid mass concentrations for larger vesicles or those containing charged headgroups. While such interactions would affect the reliability of the vesicle size estimates from SANS data as well as from the vesicle diffusion coefficients measured at low  $q$  values with DLS, the bilayer structure and fluctuation dynamics measured at high  $q$  were unaffected over a wide lipid concentration range. It is also interesting to note that the  $S'(0)$  and  $H(0)$  measured for the concentrated lipid vesicle solutions were in good agreement with the well-studied results for hard sphere solutions up to vesicle volume fractions of  $\phi_V \approx 0.15$ , but deviated at higher volume fractions, suggesting liposomes would be interesting model systems for studies of soft colloidal particles. We hope that the results presented here will help guide the design of more efficient experiments and enable more robust data analysis as well as open new avenues for understanding the interactions between lipid-based assemblies.

## DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found below: NCNR data repository at <http://dx.doi.org/10.18434/T4201B> and the NIST data repository at <https://doi.org/10.18434/mds2-2567>.

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## AUTHOR CONTRIBUTIONS

EK and EB prepared samples. EK, EB, VP, and MN performed experiments. EK, VP, PB, and MN discussed and analyzed data. All authors contributed to the writing and revisions of the manuscript.

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