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## Implications of endoplasmic reticulum stress and autophagy in aging and cardiovascular diseases

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The average lifespan of humans has been increasing, resulting in a rapidly rising percentage of older individuals and high morbidity of aging-associated diseases, especially cardiovascular diseases (CVDs). Diverse intracellular and extracellular factors that interrupt homeostatic functions in the endoplasmic reticulum (ER) induce ER stress. Cells employ a dynamic signaling pathway of unfolded protein response (UPR) to buffer ER stress. Recent studies have demonstrated that ER stress triggers various cellular processes associated with aging and many aging-associated diseases, including CVDs. Autophagy is a conserved process involving lysosomal degradation and recycling of cytoplasmic components, proteins, organelles, and pathogens that invade the cytoplasm. Autophagy is vital for combating the adverse influence of aging on the heart. The present report summarizes recent studies on the mechanism of ER stress and autophagy and their overlap in aging and on CVD pathogenesis in the context of aging. It also discusses possible therapeutic interventions targeting ER stress and autophagy that might delay aging and prevent or treat CVDs.

#### KEYWORDS

ER stress, UPR, autophagy, aging, cardiovascular diseases

## **1** Introduction

The average human lifespan has been increasing, resulting in a rapidly rising percentage of older individuals and high morbidity of aging-associated diseases, especially cardiovascular diseases (CVDs) (Alfaras et al., 2016). Within this aging population, CVDs are responsible for 40% of all deaths and are the primary reason for morbidity and mortality worldwide. The cost associated with CVD treatment continues to increase (Prince et al., 2015; Gaur et al., 2024). Thus, exploring molecular cues underlying the etiology of aging and its relationship with CVDs has become a public health priority (Marian et al., 2018). Current interventions mainly focus on managing and preventing comorbidity while neglecting the nature of the aging process (Alfaras et al., 2016).

The endoplasmic reticulum (ER) is a network of branching tubules and flattened sacs in eukaryotic cells governing protein synthesis, Ca<sup>2+</sup> storage, lipid formation, and signaling (Hetz et al., 2020). Diverse intracellular and extracellular factors interrupting any homeostatic functions in the ER can induce ER stress. To mitigate ER stress and regulate the protein-folding ability of the ER to alter secretory demands, the cells adopt a dynamic signaling pathway of unfolded protein response (UPR) (Hwang and Qi, 2018). However, persistent UPR can cause a maladaptive response (Wiseman et al., 2022). Chronic and sustained ER stress and proteostasis defects are representative features of numerous diseases, including inflammatory diseases, metabolic diseases, cancers, and age-associated diseases like CVDs (Wang and Kaufman, 2012; Uddin et al., 2021).

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Autophagy is a highly conserved process that involves lysosomal degradation and recycling of cytoplasmic components, proteins, organelles, and pathogens that invade the cytoplasm (Levine and Klionsky, 2017). Autophagy's function in maintaining proteostasis, together with the observation that autophagy declines with aging, links it to proteostasis loss, which is a typical characteristic of aging (Kern and Behl, 2019). In addition, this core molecular pathway is critical for orchestrating cellular responses to stress, cellular reprograming, maintaining stemness, fine-tuning senescence, and modulating inflammation (Kaushik et al., 2021). Changes in all of these processes accompany the aging process. Active autophagic activities have been discovered in all cardiovascular tissues (Lavandero et al., 2015). It is also possible that autophagy is vital for combating adverse influences of aging on the heart, while autophagy and autophagic flux generally decrease in aging hearts (Shirakabe et al., 2016).

It is now apparent that ER stress can be a potent trigger for autophagy (Rashid et al., 2015; Jiao et al., 2023), and the interaction of ER stress and autophagy may be implicated in the etiology of aging and CVDs. Therefore, methods of modulating ER stress/UPR, autophagy, and their interaction may serve as therapeutic targets of clinical relevance in aging and aging-related CVDs.

The present report summarizes recent studies on the mechanism of ER stress and autophagy and their overlap in aging and on CVD pathogenesis in the context of aging. Possible therapeutic interventions targeting ER stress and autophagy that could be employed to delay aging and prevent or treat CVDs are also discussed.

The cardiovascular system functions by delivering oxygenated blood to the body's tissues. Maintaining homeostasis is critical for tissue health and organism longevity. With age, the heart undergoes complicated changes that influence cellular function, ultimately transitioning from a compensatory adaptive to a decompensatory maladaptive state. Both the absolute number of cardiomyocytes and their repopulation from cardiac stem cell reserves decline due to an increase in apoptosis and necrosis (Goldspink et al., 2003), leading to a decrease in myocardial contractile capacity, increase in left ventricular wall thickness, and elevation in chamber dimension, prolonging diastole and ultimately damaging the cardiac pump 2018). function (Obas and Vasan, Therefore, aging cardiomyocytes become more susceptible to stress. Aging is also associated with vascular structure changes and declined function, particularly in the great arteries (Safar, 2010; Kovacic et al., 2011). Aging-related impairment of vascular structural changes and functional decline (van der Linden et al., 2024) is a cause of morphological changes that include vascular wall thickening, perivascular fibrosis, collagen deposition, and vessel dilatation (Costantino et al., 2016). Thickening is a critical factor in aged vasculature that promotes arterial stiffness, which, in turn, increases pulsatile and left ventricular systolic loads (Boutouyrie et al., 2021).

## 2 ER stress, UPR, autophagy, and aging

### 2.1 Overview of ER stress and UPR

ER is a double-membrane bound organelle promoting numerous functions, such as protein folding,  $Ca^{2+}$  homeostasis, and lipid/sterol synthesis. Several chaperones, foldases, and cofactors play a central role in carrying out these functions in the ER. Pathological and physiological insults, including oxidative stress, perturbation in  $Ca^{2+}$  homeostasis, release of inflammatory cytokines and toxins, and aging-related insults, can cause a buildup of unfolded and misfolded proteins in the ER. This buildup then leads to downstream disruptions in the integrity and function of the secreted proteome, a condition known as ER stress.

PKR-like ER kinase (PERK) activation through autophosphorylation and homodimerization leads to signaling cascades that include direct phosphorylation downstream of eIF2 $\alpha$ , triggering a total downregulation of translation and inhibition of protein synthesis. The eIF2 $\alpha$  phosphorylation augments ATF4 translation. ATF4 triggers the CCAAT



enhancer-binding protein (C/EBP) homologous protein (CHOP), which translocates to the nucleus. Bcl-2 protein family then induces apoptosis.

Binding immunoglobulin protein (BiP) dissociation triggers IRE1 oligomerization and trans-autophosphorylation, activating endoribonuclease activities and promoting XBP1 mRNA splicing. Spliced XBP1 (sXBP1) is a crucial transcription factor that induces gene expression of ER chaperones and multiple UPR target genes, thereby boosting the ER folding ability.

BiP dissociates from ATF6 $\alpha$ , exposing its Golgi localization signal and resulting in the translocation of ATF6 $\alpha$  from the ER to the Golgi apparatus, where it is sequentially cleaved by site-1 S1P and S2P, liberating the cytoplasmic ATF6 $\alpha$  segment. The activated ATF6 $\alpha$  fragment later binds to cis-acting ER stress response elements in the nucleus, increasing genetic expression of UPR proteins, including BiP, CHOP, GRP94, and ER-associated degradation (ERAD) components, suppressing ER stress, and restoring protein-folding homeostasis.

The adaptive UPR is vital for restoring ER homeostasis after the accumulation of potentially toxic misfolded proteins (Hetz, 2012; Hetz et al., 2020). In mammalian cells, the UPR is regulated by three ER membrane-embedded sensors: PKR-like ER kinase (PERK), inositol-requiring enzyme 1 (IRE1), and activating transcription factor 6 (ATF6) (Figure 1). These proteins can act individually or synergistically to alleviate ER stress on the organism (Spencer and Finnie, 2020). In basal conditions, transmembrane protein activation is restrained by the binding of ER-resident BiP GRP78. When ER stress is present, BiP dissociates and activates PERK, IRE1, and ATF6 (Hetz, 2012). The three mediators directly activate a full transcriptional and translational signaling program, acting on protein secretion, redox homeostasis, lipid biosynthesis, and cell death programs.

PERK is activated via homodimerization and autophosphorylation when BiP is released, resulting in a signaling cascade that includes direct phosphorylation of downstream eukaryotic translation initiation factor 2a (eIF2a) and triggering a full downregulation of translation and protein synthesis inhibition (Hetz et al., 2020). The eIF2a phosphorylation can also increase the translation of specific mRNAs with upstream open reading frames like transcription factor 4 (ATF4) (Vattem and Wek, 2004). ATF4 triggers the induction of one pro-apoptotic transcription factor known as CHOP under sustained stress conditions (Hetz et al., 2020). CHOP translocates to the nucleus and controls the components of the B-cell lymphoma protein 2 (Bcl-2) protein family to induce ER stress-mediated apoptosis. Moreover, phosphorylated eIF2a acts on UPR termination by inducing the growth arrest and DNA damage-inducible protein 34 (GADD34) (Yang et al., 2024).

IRE1 is a type I transmembrane protein with two enzymatic activities: it functions as a serine/threonine kinase and a cytosolic endoribonuclease (Grey et al., 2020). BiP dissociation with IRE1 IRE1 triggers oligomerization and transautophosphorylation, resulting in the activation of endoribonuclease activity (Karagöz et al., 2017) and promoting splicing of the 26-base intron from X-box-binding protein 1 (XBP1) mRNA (Urra et al., 2020). sXBP1 is a crucial transcription factor that induces gene expression of ER chaperones and multiple UPR target genes to boost the ER folding ability. sXBP1 is also involved in cell survival, differentiation, and development (Almanza et al., 2019). Moreover, IRE1 cleaves and downregulates microRNAs and mRNAs via its RNase domain and IRE1-dependent decay (RIDD). When ER stress persists, RIDD activity is increased, and



Autophagy overview. Autophagy is initiated by the ULRI complex and, in turn, activates the class fill PISK complex known as the VPS34 complex, containing VPS34, VPS15, Beclin1, Atg14L, and Ambra1. The VPS34 complex catalyzes the formation of PI3P on the nascent phagophore's membrane. PI3P facilitates phagophore nucleation and recruits WIPI. WIPI then recruits the ubiquitin-like conjugation systems that promote LC3 conjugation to the phagophore membrane. The LC3-PE conjugation site is marked by the Atg12-Atg5-Atg16L complex that is specific for E3-like ligase to generate LC3-PE. The LC3-PE complex acts as a phagophore receptor and facilitates selective uptake by interacting with adapter molecules. The phagophore matures into the autophagosome, eventually fusing with the lysosome and causing the degradation of sequestered substances by diverse hydrolases. The resulting breakdown products are then released into the cytosol for recycling. SNARE-like proteins promote the fusion of autophagosomes and lysosomes.

XBP1 mRNA splicing is decreased, aggravating uncontrolled cell death (Hollien and Weissman, 2006; Wiese et al., 2022).

ATF6 is a type II transmembrane protein with a basic leucine zipper motif. BiP dissociates from ATF6 $\alpha$  upon ER stress, exposing the Golgi localization signal of ATF6 $\alpha$  and causing its translocation from the ER to the Golgi apparatus. This facilitates proteolytic removal of luminal and transmembrane domains by site-1 protease (S1P) and site-2 protease (S2P) (Ye et al., 2000), liberating the cytoplasmic ATF6 $\alpha$  segment, becoming activated to allow interactions with cis-acting ER stress response elements in the nucleus, and enhancing genetic expression of BiP, GRP94, and CHOP to suppress ER stress (Jin et al., 2017).

In the past 5 years, many studies have reported that UPR components have multiple functions beyond maintaining ER proteostasis. UPR binding with novel partners serves as a platform for interorganelle communication and signaling crosstalk to mediate cytoskeleton dynamics, mitochondrial bioenergetics, and membrane contacts (Carreras-Sureda et al., 2019; Shin et al., 2019; Dufey et al., 2020).

### 2.2 Autophagy overview

Autophagy in mammals can be roughly categorized into three groups based on distinct mechanisms of cargo sequestration. Microautophagy is the process of invagination where cytoplasmic components directly enter the lysosomes, while chaperone-mediated autophagy (CMA) includes selective degradation of target proteins that are recognized and translocated to lysosomes by the chaperone complexes. Macroautophagy (hereafter referred to as autophagy) is another mechanism of adaptation to harsh environmental conditions and intrinsic stress (Mizushima, 2018). Autophagy is a well-defined process mediated by numerous proteins encoded by autophagy-related genes (ATGs).

The process of autophagy involves initiation, phagophore formation and elongation, and autophagosome-lysosome fusion (Melia et al., 2020) (Figure 2). Membrane assembly and autophagosome formation as a double-membrane phagophore, which completely or partially sequesters cytoplasmic organelles and then fuses to lysosomes, are the two key steps during autophagic flux. Various biological byproducts are released for energy production and recycling via these processes. Autophagy mediates the breakdown of long-lived proteins, in contrast to the turnover of short-lived proteins, which is mediated by other catabolic mechanisms, such as the ubiquitin-proteasome system (UPS) (Gavilán et al., 2024).

Autophagy can be categorized as non-selective or selective based on the nature of the cargo content. The molecular mechanism of the autophagy process remains basically identical in both types. However, selectivity in selective autophagy is a highly spatiotemporally controlled metabolic pathway guaranteed by specific labeling of every cargo based on stress type and involvement of receptors (or adaptors) (Lamark and Johansen, 2021). Accumulating evidence has shown that abnormalities of selective autophagy are closely related to the occurrence and progression of aging-related diseases, such as cancer, metabolic diseases, neurodegenerative diseases, and CVDs (Ma et al., 2023; Ma et al., 2024).

Autophagy is induced in reaction to diverse stressors, including nutrient or growth factor deprivation, protein and organelle damage, oxidative stress, inflammation, ER stress, and genotoxic stress (Morishita and Mizushima, 2019). Nutrient-sensing pathways, including those involving the mechanistic target of rapamycin complex 1 (mTORC1) and AMP-activated kinase (AMPK), are central to autophagy regulation (Kitada and Koya, 2021). Therefore, identifying the nutrient resources and energy for cells under harsh conditions is important (He, 2022; Feng et al., 2024).

## 2.3 Relationship among UPR, autophagy, and aging

#### 2.3.1 ER stress and UPR during aging

A progressive functional decrease in the ER occurs with age (Bouska et al., 2019). Chaperones, in particular, are progressively oxidized during the aging process, bringing about a functional decline and matching the discovered impairment in adequate cellular reaction to ER stress in cardiac aging (Gude et al., 2018; Li et al., 2019). For example, the function of chaperones like calreticulin, calnexin, and BiP is compromised with age, elevating the number of misfolded or unfolded proteins and protein aggregates. Across the lifetime of Wistar rats, the expression of BiP protein in the brain and peripheral tissues, including lung, kidney, liver, heart, and spleen, was higher in young tissues than in aged tissues (Hussain and Ramaiah, 2007).

Mild and moderate ER stress may induce stress resistance that can extend the lifespan, but prolonged and overwhelming stress may accelerate the aging process (Salminen and Kaarniranta, 2010). On the other hand, aging leads to increased ER stress (Chen et al., 2023). For example, CHOP and caspase-12 can be induced in aged and stressed rats but not in their young and stressed counterparts (Paz Gavilán et al., 2006).

In addition to affecting ER function, aging can influence the ER structure within the cell (Erickson et al., 2006; Moltedo et al., 2019). It has been observed that the highly ordered parallel cisterna of the rough ER, which is a feature of young neurons, becomes dispersed in aged neurons (Hinds and McNelly, 1978). Likewise, an aging and failing heart undergoes architectural changes in the levels of ER and UPR components in cardiomyocytes (Ortega et al., 2014).

#### 2.3.2 Autophagy during aging

Autophagy is crucial for proteostasis maintenance by removing long-lived or damaged proteins and organelles. However, autophagy function goes beyond proteostasis maintenance (Levine and Klionsky, 2017). Autophagic function declines with age, while autophagy dysfunction promotes premature aging. Autophagy inhibition by DBI/ACBP is involved in aging and CVDs (Montégut et al., 2023). Similarly, mice with cardiac-specific deleted ATG5 protein develop dilated cardiomyopathy with serious systolic dysfunction in aging, which accompanies dysfunctional and abnormal mitochondrial accumulation and sarcomeric disarray (Taneike et al., 2010). Mitophagy may also be impaired during cardiac aging. Young Parkin knockout mice exhibited a normal cardiac phenotype, while abnormal mitochondria accumulated in cardiomyocytes with age (Kubli et al., 2013a).

## 2.3.3 Interrelationship among ER stress, UPR, autophagy, and aging

Autophagy correlates with the ER on numerous levels. These two systems are dynamically connected, and changing the function of one affects the other. The ER acts as a potential membrane source and scaffold for autophagosome generation. Molecules responsible for the autophagy cascade from ULK1 to PI3P effectors are localized in the ER and contribute to autophagosome nucleation.

PERK has regulatory functions in autophagy mediated by ATF4 and CHOP, which have been shown to transcriptionally adjust in 12 ATG genes. ATF6 upregulates DAPK1 expression, phosphorylating Beclin-1 and mediating autophagy initiation. IRE1 recruits TRAF2 and ASK1, activating c-Jun N-terminal kinase (JNK), which mediates Bcl-XL/Bcl-2 phosphorylation and causes Beclin-1 dissociation and enhanced basal autophagy. sXBP1 specifically binds to the Beclin-1 promoter and induces transcription, causing autophagy upregulation. ER stress can also activate autophagy by promoting the release of a substantial amount of Ca<sup>2+</sup> from the ER into the cytoplasm.

The ER has been observed in close vicinity to the isolation membrane, serving as a major source of autophagosomes and contributing to autophagosome formation (Senft and Ronai, 2015).

When stress stimuli remain or become extremely strong, UPR and UPS are incapable of restoring the ER to its normal state, and autophagy comes into play, engulfing the damaged ER for degradation. The degraded ER fragments can then be recycled into a fresh ER with restored function. Autophagy becomes the last resort to restore ER homeostasis (Qi and Chen, 2019).

ER stress is a potent trigger for autophagy. In addition to nutrient deprivation and mTOR inhibition, UPR is another factor responsible for autophagy induction (Kapuy, 2024). UPR activation can trigger the change in autophagy that can modulate UPR, exemplifying the crosstalk process. Three UPR arms, including PERK, IRE1 $\alpha$ , and ATF6, each facilitate autophagy differently in ER stress. The signaling pathways operate under diverse stress circumstances and in a contextdependent manner (Figure 3).

The eIF2a phosphorylation in worms is critical for the induction of stress granules, which are required for prolongevity using dietary restriction (Kuo et al., 2020). The eIF2a phosphorylation is indispensable for autophagy activation (Kuo et al., 2020). Autophagy is a major anti-aging mechanism, suggesting an interrelationship between UPR and autophagy in pro-survival signals (López-Otín et al., 2023). The activation of mTOR can inhibit autophagy activation, increasing the severity of ER stress. Therefore, there is reciprocal regulation and interaction between ER stress and autophagy (Shi et al., 2024). During the aging process, the ERAD degradation pathway fails to eliminate aggregated misfolded proteins, and the UPR triggers the autophagy process to facilitate their degradation to maintain proteostasis (Pintado et al., 2017). On the other hand, autophagic functions, such as the synthesis rate of autophagosome and the effectiveness of autophagosome fusion with lysosome, are impaired with aging (Kaushik et al., 2021). UPR hyperactivation in the heart induces Sirtuin 1 (SIRT1), which inhibits the ER stress-triggered apoptosis by regulating the PERK activity and deacetylating eIF2a to decrease the proapoptotic protein CHOP expression (Prola et al., 2017; Pires Da Silva et al., 2020). Moreover, SIRT1 can activate autophagy, playing a positive role in various ER-induced damage processes in the aging heart, decreasing apoptosis, regulating UPR sensors, and decreasing the S-nitrosylation effect of protein disulfide isomerase (PDI) (Hsu





et al., 2017; Prola et al., 2017). Therefore, ER stress correlates with autophagy via SIRT1 in the heart.

# 3 Mechanism of ER stress, UPR, and autophagy in aging and CVDs

ER stress and autophagy dysregulation are closely interrelated with various cellular stress responses, including inflammation,

proteostasis loss, cellular senescence, mitochondrial dysfunction, and oxidative stress (Figure 4).

# 3.1 ER stress, UPR, autophagy, and their overlap in proteostasis

The proteostasis network sustains a normal proteome by integrating protein synthesis, folding, trafficking, and degradation

(Hetz and Saxena, 2017). ERAD, UPR, and the autophagy pathway are the main players in this network. Cellular proteostasis derangement has been proposed to be one of the major hallmarks of aging (López-Otín et al., 2013; López-Otín et al., 2023). Healthy proteostasis in cardiomyocytes ensures proper contractile function (Li et al., 2019). Genetic anomalies or environmental factors often causing wear and tear can lead to loss of protein patency and disrupt protein quality control, triggering proteotoxic stress and ultimately resulting in cardiac aging (Henning and Brundel, 2017).

A disruption in ER stress/UPR and autophagy can result in protein quality control failure (Henning and Brundel, 2017; Li et al., 2019). About a third of the proteome is synthesized in the ER, which is vital for folding and modifying secretory and membrane proteins, thereby sustaining proteostasis. ER stress-induced UPR enhances the ER protein-folding capacity by focusing cellular resources to synthesize the proteins involved in the folding process, such as PDIs and ER-resident chaperones (Kilberg et al., 2009). The ER proteinfolding load is decreased by inhibiting the whole cellular protein synthesis (Kilberg et al., 2009). Therefore, these aspects of UPR eventually facilitate ER stress relief (Ron and Walter, 2007). However, as with numerous other signaling pathways, UPR is susceptible to aging-associated impairment and becomes less efficient over time (Lim et al., 2023). The aging process is accompanied by an imbalance between augmented ER stress and diminished UPR, leading to chronic ER stress (Brown and Naidoo, 2012) and ultimately causing loss of proteostasis. Therefore, mild cardiac ER stress likely helps to clear damaged, unfolded, or old proteins, thus sustaining appropriate heart functions. However, cardiomyocytes may undergo apoptosis when ER stress is prolonged and persistent. Because cardiomyocytes cannot regenerate, apoptosis harms the heart. Therefore, prolonged ER stress must be avoided (Li et al., 2019).

During the extended period of cellular proteotoxic stress, UPR and UPS become overwhelmed and exhausted. Autophagy is activated to clear misfolded proteins to ensure cell homeostasis and survival (Li et al., 2018a). In all cell types, including cardiomyocytes, autophagy has become a proteasome substitution for the clearance of misfolded proteins from the ER via ERAD by coordinating with other components of the proteostasis network (De Leonibus et al., 2019). In the heart, autophagy mediates the effective clearing of abnormal and damaged proteins and organelles from cardiomyocytes and serves as a cellular recycling mechanism by reclaiming lipids, amino acids, and other molecular building blocks with lysosomal acidic hydrolases, thereby maintaining healthy heart function (Sciarretta et al., 2018). Enhancing autophagy in the aged mouse heart can reduce protein damage, attenuate hypertrophy, and restore Ca<sup>2+</sup> homeostasis, thus improving contractile function (Taneike et al., 2010). By eliminating defective/damaged mitochondria, mitophagy facilitates mitochondrial proteostasis in response to mitochondrial stress (Pickles et al., 2018), thus conserving normal cardiac contractility and maintaining heart function. Autophagy impairment can disrupt protein degradation and misfolded or damaged protein processing, consequently contributing to aging, pathogenesis, and development of CVDs. Additionally, impaired autophagy also affects the function of UPS due to decreased degradation of damaged or aged proteasomes, severely increasing the levels of proteasome substrates and leading to heart senescence and cardiac disease onset (Korolchuk et al., 2009).

# 3.2 ER stress, UPR, autophagy, and their overlap in inflammation

Chronic inflammation has been identified as a determinant of aging and plays a central role in most aging-associated chronic disorders (Liberale et al., 2022). A recent study has highlighted the function of inflammation as a critical contributor underlying the mechanism conferring cardiovascular risk (Madhur et al., 2021). For instance, it has been determined that inflammation is a major contributor to atherosclerosis from its initiation to progression and, eventually, to its endpoint (Weber et al., 2023). The term "inflammaging" describes chronic, low-grade, systemic inflammation that occurs in older individuals even without overt infections. It is a risk factor for CVDs and a global indicator of poor health status (Ruparelia et al., 2017). Inflammaging is believed to originate from an imbalance between the generation and disposal of damage-associated molecular patterns (DAMPs) ("enemy within"), including cellular debris, misfolded proteins, and/or misplaced selfmolecules, and it increases with age (Franceschi et al., 2017). DAMP accumulation can also be sensed by the NLRP3 inflammasome, cause NLRP3 oligomerization, and increase caspase 1-dependent secretion of IL-1β and IL-18. Blockade of the NLRP3 inflammasome prolongs the health span, alleviating multiple aging-related degenerative changes associated with inflammaging (Ferrucci et al., 2005; Youm et al., 2013).

Inflammatory mediators can sustain and even exacerbate ER stress within tissues. Chronic ER stress also facilitates chronic inflammation. Therefore, there may be a positive feedback loop between ER stress and inflammation. Both ER stress and maladaptive UPR can boost inflammation and apoptosis in the heart. Proinflammatory cytokines, including CCL2, IL-8, interferony, and TNF, provoke cytotoxicity via IRE1a and XBP1 (Salminen et al., 2020), while cardiac-specific Ccl2 overexpression upregulates the expression of ER stress-associated genes and exacerbates IR injury (Zhang W. et al., 2020). ER stress and UPR take part in inflammation via numerous regulators, including nuclear transcription factor-kB (NF-kB) and the NLRP3 inflammasome, and can intervene in Ca2+ homeostasis. NF-kB activation mediates myocardial and vascular inflammation in aging-related diseases (Seals et al., 2011). Aging-associated NF-KB signaling upregulates the expression of proinflammatory genes, such as ILs, TNF- $\alpha/\beta$ , chemokines, and adhesion molecules (Chung et al., 2019). IRE1a may drive NF-KB activation by cooperating with TRAF2 (Shen et al., 2014). CHOP can also stimulate NF-κB signaling in chronic inflammatory diseases (Willy et al., 2015). ER stress can induce inflammation through TXNIP-NLRP3-IL-1ß signaling and likely participates in CVD pathogenesis (Yang et al., 2020).

Autophagy is a basic biological process that facilitates innate and adaptive immunity. Autophagy perturbation results in repercussions in diseases with inflammatory components, infections, metabolic disorders, neurodegeneration, autoimmunity, cancer, and CVDs (Deretic, 2021). Autophagy controls the burden imposed by exogenous hazards and infection, as well as by endogenous sources of inflammation, such as molecular aggregates and damaged organelles, while simultaneously limiting the inflammatory process (Matsuzawa-Ishimoto et al., 2018). Autophagy facilitates the recycling of cellular content, maintains homeostasis, and prevents recognition of PAMPs by PRRs and consequent inflammation (Zitvogel et al., 2010). Therefore, autophagy directly inhibits the development of the inflammaging process. Specifically, mitochondria are involved in the proper signaling of some innate immunity platforms like mitochondrial antiviral signaling. They can also be the sources of DAMPs under pathological conditions (Youle, 2019). Damaged mitochondria trigger the NLRP3 inflammasome. By effectively sequestering damaged mitochondria leaking DNA and reactive oxygen species (ROS), mitophagy removes the key signal that promotes NLRP3 inflammasome activation (Matsuzawa-Ishimoto et al., 2018).

The autophagic response diminishes with age, and this impairment may result in inflammasome activation and chronic inflammatory responses, accelerate the aging process, and affect diseases with inflammatory components, including CVDs (Deretic, 2021). CMA function becomes impaired during atherosclerosis progression, leading to increases in NLRP3 inflammasome activation and IL-1 $\beta$  secretion, boosting vascular inflammation and atherosclerosis progression (Qiao et al., 2021).

Both ER stress and autophagy are implicated in inflammation and may be interconnected with pathophysiological aspects of the cardiovascular system. For example, recent research has shown that inhibition of transcription factor CCAAT/enhancer-binding protein beta prevents atherosclerosis development and foam cell formation. Its protective role in atherogenesis involves reducing inflammation, ER stress, and apoptosis, promoting autophagy, and inactivating mTOR (Zahid et al., 2020). Similarly, the protective effect of cinnamaldehyde in CVD progression results in the inhibition of inflammation and oxidative stress and modulation of autophagy and ER stress (Lu L. et al., 2022). However, it is unclear whether autophagy is induced by elevated ER stress or is simply a reaction to injury in these pathological conditions. Nonetheless, both the UPR pathway and autophagy are vital for regulating the inflammation process and are implicated in the aging process and pathogenesis of various diseases, including CVDs.

## 3.3 ER stress, autophagy, and their overlap in cellular senescence

Cellular senescence features include irreversible cell-cycle arrest, apoptosis resistance, tumor suppressor pathway activation, altered metabolism and cell shape, and secretion of numerous proinflammatory cytokines, chemokines, and growth factors, collectively named the senescence-associated secretory phenotype (SASP) (Kuilman et al., 2010). Senescence was first identified in human diploid fibroblasts that showed a finite ability for cell division due to telomere shortening (replicative senescence) (Hayflick and Moorhead, 1961). Senescence independent of telomere length was later discovered within numerous aged or damaged tissues and was critically linked with cellular stresses (Kuilman et al., 2010).

Evidence has shown that cellular senescence exists in a variety of cell forms in the cardiovascular system and accumulates in culprit lesions, playing critical roles in CVD onset and outcome (Wang J. et al., 2015; Song et al., 2020). For example, cells in advanced atherosclerotic plaques usually display markers of senescence, including p16<sup>INK4A</sup> and tumor suppressor ARF, and express SASP, which further fuels inflammation and produces metalloproteinases that degrade the extracellular matrix, destabilizing atherosclerotic plaque (Wang M. et al., 2015; Yan et al., 2021).

Cellular senescence is closely related to ER stress and UPR. Various cell types undergoing senescence show structural changes in the ER and elicit UPR activation (Abbadie and Pluquet, 2020). The loss of proteostasis is present in senescent cells, which can be attributed to changes in UPR (Sabath et al., 2020).

UPR activation likely occurs in all forms of senescence under various stresses (Benvenuti et al., 2002; Dörr et al., 2013; Panganiban et al., 2013). In diabetes-induced atherosclerosis, high glucoseinduced senescence, barrier disruption promotion in vascular endothelial cells, and maladaptive UPR constitute a pathway modulating the expression of senescence markers in diabetic conditions via ATF6 a and sXBP1 (Fatima et al., 2022). The DNA damage-inducible transcript 3 (DDIT3) protein level was significantly augmented in aged mouse hearts. Proprotein convertase subtilisin/kexin six is a proteolytic enzyme critical for maintaining cardiac function and vascular homeostasis. Its deficiency promotes cardiomyocyte senescence, which is related to DDIT3-mediated ER stress (Zhan et al., 2022). It can be inferred that ER stress and UPR can be either the reason or the result of senescence, considering that proteotoxic stress resulting from oxidized misfolded proteins or SASP protein overload is induced during senescence. Senescent cells may need optimal UPR activity for long-term survival (Abbadie and Pluquet, 2020). Pharmacological compounds inhibiting UPR subpathways may serve as senolytics for the cardiovascular system, although this remains to be verified.

Autophagy activation first happens in response to the stress stimulus. Therefore, it generally functions as an anti-senescence program. Enhancing autophagy in aged cardiomyocytes improves senescence (Li W. W. et al., 2021). Defective vascular smooth muscle cell (VSMC) autophagy boosts the development of stress-induced premature senescence and atherogenesis (Grootaert et al., 2018). Senescence is promoted in VSMCs. However, autophagy activation can suppress senescence in VSMCs (Li et al., 2014). Blocking autophagic flux and activating mTOR are involved in the mechanism of accelerating vascular senescence by the Yesassociated protein, which has been widely implicated in vascular pathophysiology processes (Pan et al., 2021). Similarly, augmented levels of ROS upon autophagy inhibition somewhat facilitate cellular senescence (Tai et al., 2017). However, another study showed that autophagy inhibition delayed but did not prevent senescence (Young et al., 2009). Autophagy produces a high flux of recycled amino acids and other metabolites, supporting massive synthesis of SASP factors and facilitating senescence (Kang and Elledge, 2016). This contradiction may be explained by the fact that autophagy may modulate some targets acting in an opposing manner to regulate cellular senescence. Autophagy inhibition can lead to diverse consequences depending on its timing, duration, or type.

Autophagy might crosstalk with ER stress and UPR to modulate senescence. Senescent microglia have lower autophagy and higher ER stress levels, possibly facilitating the progression of serious Alzheimer's disease pathology (Angelova and Brown, 2018). Another study showed that UPR promotes autophagy and relieves acid-induced premature senescence in the nuclei of pulposus cells (Zhu et al., 2022). However, this connection is uncertain in CVDs and in the aging process.

# 3.4 ER stress, autophagy, and their overlap in mitochondria

Mitochondria are central organelles in the cell responsible for producing cellular energy. This role is especially crucial in the heart, where mitochondria take up around 30% of the overall cell volume and generate 6 kg of ATP every day for sustaining cardiac mechanical function (Torp et al., 2023). Mitochondria are also the chief source of ROS that modulate physiological processes and control cell death (Panel et al., 2018). The effectiveness of mitochondrial function decreases as the cells and organisms age. The reduction in mitochondrial function may greatly facilitate agedependent changes in the cardiovascular system. For example, both human and animal models exhibited increased mitochondrial damage in atherosclerotic aortas (Ballinger et al., 2002). Mitochondrial dysfunction underlies atrial cardiomyocyte damage and brings about atrial fibrillation (Wiersma et al., 2019). Agedependent mitochondrial DNA damage is a critical factor cardiac arrhythmia pathophysiology (Baris underpinning et al., 2015).

Both the ER and mitochondria are subjected to great metabolic demands (Green et al., 2014; Wang and Kaufman, 2016). Proteostasis and metabolic control are closely connected, and this interrelation is essential for maintaining healthy cardiac function (Murley and Nunnari, 2016). An imbalance in ER and mitochondrial interactions may be a prelude to cardiac aging and disease (Li et al., 2019). For example, although initial ER stress is an adaptive reaction for restoring cell functions, serious ER stress brings about electron transport chain injuries in adult heart mitochondria, contributing to mitochondrial dysfunction in aged hearts (Chen et al., 2020). Attenuating ER stress enhances mitochondrial functions in the aged hearts (Thompson et al., 2020).

Functional interaction between ER and mitochondria occurs through discrete ER-mitochondria contacts known mitochondria-associated membranes (MAMs). Mitochondria in those domains are spatially and functionally organized in close contact with the ER, acting as a signaling hub that regulates cellular physiology (Carreras-Sureda et al., 2017). Commonly researched proteins involved in MAM formation include mitofusin-2 (Mfn2), which is a key component of mitochondrial fusion and fission (Chen et al., 2003). Cardiac-specific Mfn2 ablation decreased ER-mitochondrial tethering by 30%, decreasing mitochondrial Ca2+ uptake and hampering the reaction to physiological stress (Chen et al., 2012). Mfn2-deficient cells exhibit hyperactivated UPR signaling and defective autophagy and apoptosis, indicating the function of MAM integrity in UPR signaling (Muñoz et al., 2013). Another protein in the heart involved in the interaction between the ER and mitochondria is ER Ca2+ sensor stromal interaction molecule 1 (STIM1). It maintains ER Ca2+ levels in normal cardiac homeostasis. Mice with cardiomyocyte-specific STIM1 knockout showed pronounced ER dilatation, smaller mitochondria, and dysfunctional mitochondrial networks. They also developed cardiac fibrosis and dilated cardiomyopathy (Collins et al., 2014), potentially accelerating heart aging.

Mitochondrial quality control can also be achieved through mitophagy. Mitophagy protects cells from loss of mitochondrial function during aging. In the cardiovascular system, mitophagy induction in aged mice improves overall mitochondrial function and prevents arterial wall stiffness (LaRocca et al., 2014). Young mice with Parkin knockout have a normal cardiac phenotype, although abnormal mitochondria accumulate in cardiomyocytes with age (Kubli et al., 2013a). Inhibiting the PINK1/Parkin mitophagy pathway due to insufficient Drp1 induces myocardial apoptosis in the aging heart (Wei et al., 2021). This evidence revealed that the PINK1/Parkin pathway may mediate mitophagy in aging hearts.

Decreased mitophagy is also observed in aged hearts (Hoshino et al., 2013), while ER stress contributes to decreased mitophagy (Wang et al., 2017). Interestingly, MAMs have also been implicated in the control of mitophagy and mitochondrial integrity (Gelmetti et al., 2017). Mitophagy can be initiated in MAMs where PINK1 and Beclin-1 relocalize to promote ER–mitochondria tethering and autophagosome formation (Gelmetti et al., 2017). The mitophagy regulator FUNDC1 is also vital for MAM regulation in CVDs. FUNDC1 can be enriched in MAMs. Therefore, the formation of MAMs regulates cellular Ca<sup>2+</sup> homeostasis and mitochondrial dynamics to prevent heart dysfunction (Li G. et al., 2021).

## 3.5 ER stress, autophagy, and their overlap in oxidative stress

Basal levels of ROS are essential for maintaining numerous cellular functions, including signal transduction pathway, gene expression, defense against invading microorganisms, growth promotion, and death (Finkel, 2011). Excessive ROS levels harm cellular macromolecules, eventually resulting in necrosis and apoptotic cell death. Oxidative stress occurs when the equilibrium of oxidant and antioxidant balance is disrupted and tends toward an oxidative state, where ROS are the main cause of oxidative stress. Oxidative stress is implicated in numerous pathological circumstances, neurodegenerative diseases, CVDs, diabetes, and cancers, most of which are aging-related and accelerate the aging rate (Wu et al., 2024). Excessive oxidative stress leads to endothelial dysfunction, enabling the permeation, trapping, and physicochemical modification of circulating lipoprotein particles in the subendothelial space (Gimbrone and García-Cardeña, 2016). ROS such as O<sup>2-</sup> are able to react with nitric oxide (NO) to deplete NO bioavailability, triggering vasoconstriction in rats and leading to hypertension (Carlisle et al., 2016). Overexpression of mitochondria-targeted catalase protects the aged hearts (Bhuiyan and Fukunaga, 2009).

The interplay of ER and oxidative stress has been demonstrated in numerous pathological and physiological conditions (Zhang and Kaufman, 2008). Disulfide bond formation occurring in the ER is triggered by PDI and ER oxidase 1a (ERO1) (Tu and Weissman, 2004). PDI obtains electrons from protein-folding substrates and oxidizes the thiol group of cysteine residues, leading to disulfide bond formation. ERO1 mediates electron transfer from PDI to

oxygen, resulting in ROS byproducts. To control oxidative stress, UPR pathways are activated by the adaptive mechanism to maintain cellular functions. In the heart, ATF6 serves as a unique bridge for linking ER stress to oxidative stress signaling pathways in cardiovascular stress conditions. In addition to its impact on ER protein folding, ATF6 can have an extensive influence on antioxidant protein expression even outside the ER (Jin et al., 2017). However, severe ER stress promotes ROS production and aggravates oxidative stress (Malhotra and Kaufman, 2007). In the heart, ER stress can diminish complex I activities and augment ROS generation (Chen et al., 2017; Chen Q. et al., 2019). On the other hand, disturbed redox homeostasis in the ER drives ER stress, causing ROS accumulation in the ER and mitochondria (Lei et al., 2017; Mennerich et al., 2019). Therefore, inhibiting ER and oxidative stresses can be a method of delaying heart aging. For example, swimming exercise inhibited oxidative and ER stresses in the hearts of aged mice by increasing cGMP-protein kinase G signaling (Chang et al., 2020).

The interplay between ROS and autophagy is crucial for cellular homeostasis, which is particularly critical in CVDs. Indeed, a physiological ROS level is needed for autophagy induction under conditions of nutrient deprivation and ischemia in cardiomyocytes. Nox4 is quickly activated in the ER in response to glucose deprivation in cardiomyocytes. Nox4-derived ROS boosts autophagy by driving the PERK/ATF4 pathway (Sciarretta et al., 2013). However, excessive oxidative stress can cause impairment in cardiac autophagy. For example, Nox2-derived superoxide activation mediates the impairment of autophagic flux subjected to lipid overload, and superoxide inhibition rescues autophagic flux in cardiomyocytes (Jaishy et al., 2015).

Autophagy is vital for reducing oxidative stress. Endothelial cell treatment with an autophagy inducer reduces oxidative stress (LaRocca et al., 2012). A prior study showed that endogenous CMA activator humanin protected cells from oxidative stress in aging-related CVDs (Cai et al., 2021). Notably, mitochondria are major producers of ROS, and appropriate removal of dysfunctional mitochondria is crucial as mitochondrial damage is linked to numerous cardiovascular pathologies (Lavandero et al., 2015). Through selective sequestration and degradation of dysfunctional mitochondria prior to those mitochondria causing damage or triggering cell death, mitophagy serves as a defense mechanism against ROS generation via aberrant mitochondria (Diao and Gustafsson Å, 2022). Given the importance of mitophagy, further studies are necessary to explore the role of mitophagy in ROS removal in the context of cardiac aging and CVD to identify additional targets for intervention.

The decline in the Nrf2/EpRE signaling system is vital for aging and aging-related cardiovascular oxidative stress (Gounder et al., 2012; Volonte et al., 2013; Li et al., 2023). Autophagy is crucial in ROS scavenging through activation of Nrf2 by p62 (Komatsu et al., 2010). Oxidative stress induces ER stress, amplifying ROS production and causing autophagy upregulation, whereas Nrf2 is tightly linked to ER stress signals (Cominacini et al., 2015; Wang et al., 2020). Nrf2 may be the hub to coordinate UPR and autophagy during oxidative stress, but a direct connection among them needs to be further investigated, particularly in aging-related CVDs.

## 3.6 ER stress, autophagy, and their overlap in nutrient-sensing pathways

Cardiac aging is related to impairment in some nutrient and metabolic pathways, possibly directly or indirectly impacting functional and structural deterioration of the heart (Daneshgar et al., 2021). The mTOR pathway is a critical signaling pathway. It forms two distinct complexes, mTORC1 and mTORC2, where mTORC1 is an important signaling hub coordinating nutrient status and cell growth (Kim and Guan, 2019). The involvement of mTOR during aging was initially demonstrated within a Caenorhabditis elegans model system, showing decreased expression of mTOR homologs or Raptor-extended life span (Jia et al., 2004). Activation of mTOR upregulates the expression of genes involved in growth and metabolism, downregulates genes related to cellular stress adaptation, and suppresses catabolic processes, such as autophagy (Saxton and Sabatini, 2017; Battaglioni et al., 2022). In the heart, chronic mTOR activation may increase cardiac aging. Activating autophagy via mTORC1 inhibition provides potential beneficial effects during aging. Aged mice undergoing 3-month mTOR inhibitor rapamycin treatment showed alleviated cardiac aging pathologies, such as cardiac fibrosis and heart inflammation, and improved cardiovascular function (Flynn et al., 2013). In contrast, chronic Akt1 activation led to activated mTORC1, worsening aging-induced cardiac hypertrophy and myocardial contractile dysfunction via inhibition of autophagy (Hua et al., 2011). Mice with systemic GSK-3a deletion showed cardiac hypertrophy, dysfunction, and sarcomere abnormalities during aging due to deregulated mTORC1 activation and autophagy inhibition (Zhou et al., 2013). However, current research has demonstrated that rapamycin prolongs the lifetime but failed to show that it prevents cardiovascular aging (Neff et al., 2013). This inconsistency in the impact of mTORC1 inhibition on the aging heart is possibly due to diverse cell forms and sources of mTORC1 inhibitors.

ER stress and the mTOR signaling network function in a coordinating manner to regulate various cellular processes, cell development, and survival. ER stress induces mTORC1 activation within the NRK-52E cells, leading to activation of the IRE1-JNK pathway and apoptosis. Inhibition of mTORC1 weakens ER stressinduced apoptosis by selectively suppressing IRE1-JNK signaling and enhancing cellular survival in ER stress (Kato et al., 2012). Constitutive mTORC1 activation by loss of TSC stimulates JNK, contributing to ER stress-induced apoptosis (Ozcan et al., 2008; Bachar et al., 2009). mTORC1 also functions as a feedback loop to regulate ER stress signaling, as observed in intestinal epithelial cells incubated with tunicamycin. This exposure resulted in ER stressinduced caspase-3-dependent apoptotic cell death, which was enhanced by rapamycin (Ji et al., 2018). To date, there is insufficient research devoted to defining ER stress and mTOR signaling networks in cardiac aging and disease. The mTORautophagy-ER stress axis may exist in aging and pathological conditions in the cardiovascular system. In a murine model of cardiac hypertrophy, inhibiting sodium-glucose co-transporter two can reduce left ventricular fibrosis, which is related to reduced cardiac insulin level, augmented AMPK signaling inhibition of cardiac mTOR activation, and a reduction in downstream ER stress, UPR, and apoptosis (Moellmann et al.,

2022). In addition, rapamycin decreased cardiomyocyte apoptosis and boosted cardiomyocyte autophagy. This protective effect was modulated by adjusting crosstalk between the mTOR and ER stress pathways in chronic heart failure (Gao et al., 2020). In blood vessels, inactivation of CCAAT/enhancer-binding protein beta attenuates macrophage foam cell formation in atherogenesis by diminishing inflammation, ER stress, and apoptosis, boosting autophagy, and inactivating mTOR (Zahid et al., 2020). In addition, mTORautophagy dysfunction activated ER stress, thus inducing lipid metabolism, inflammation, and fibrosis (Wang et al., 2022).

## 3.7 ER stress, autophagy, and their overlap in DNA damage

DNA damage accumulation is a strong candidate for the primary cause of aging by inducing cell death, senescence, and tissue dysfunction (Schumacher et al., 2021; Zhao et al., 2023). Therefore, cells have developed an intricate mechanism for confronting and repairing DNA lesions, which is known as DNA damage repair (DDR). It involves one coordinated network that can be triggered by either single-strand breaks (SSBs) or double-strand breaks (DSBs) in the DNA. Reaction to DSBs is mediated by three kinases that are members of the PI3K-related kinase family. They include ATM, ATR, and DNA-PKcs (Blackford and Jackson, 2017), which coordinate the phosphorylation of various proteins, regulating a broad spectrum of cellular processes involved in DNA repair and replication, cell-cycle control, and apoptosis. SSB lesions produced at DSB sites or collapsed replication forks can cause PARP1 and ATR activation (Ciccia and Elledge, 2010). More clinical and preclinical evidence demonstrates the presence of DNA damage and activation of DDR signaling in the growth and/or progression of CVDs, though these processes remain largely underexplored. For example, DDR is activated within postmitotic cells like cardiomyocytes (Botto et al., 2001). Human atherosclerotic plaques show more DSBs and ATM activation than normal tissue (Mahmoudi et al., 2008). An age-associated increase in mitochondrial ROS causes mitochondrial DNA (mtDNA) mutations. Plasma levels of mtDNA fragments and mtDNA damage remain high in CVD patients (Nie et al., 2020), and mtDNA mutations may be highly associated with heart aging (Quan et al., 2020).

Although ER stress, UPR, and DDR are adaptive mechanisms that occur in the ER and nucleus, there is emerging evidence showing that signaling crosstalk exists between UPR stress sensors and DDR. For example, sXBP1 directly controls transcription of various DDR genes and yH2AX levels (Argemí et al., 2017). sXBP1 silencing leads to an increase in yH2AX foci formation and a decrease in the expression of MRN complex proteins and ATM phosphorylation (Lyu et al., 2019). Selective activation of IRE1a under genotoxic stress regulates the repair program and maintains cell survival. Genotoxic agents are involved in RIDD activity without XBP1 mRNA splicing (Dufey et al., 2020). This IRE1a is involved in mRNA degradation and plays a role in DDR, influencing DNA repair, cell-cycle arrest, and cell death. PERK-p-eIF2a-ATF4 signaling activation via SSBs has been shown to support cell survival under nutrient-restricted conditions (Clementi et al., 2020). The molecular connection between UPR and DDR, which maintains both genome integrity and proteostasis in CVDs, is not fully characterized. However, the evidence above suggests a potential coordination between the two pathways that may be relevant to aging and CVDs. Future experimental evidence is needed to explore this idea.

Autophagy is also involved in the reaction to genotoxic stress. Autophagy upregulation has been observed in response to cellular DNA damage (Juretschke and Beli, 2021), while DNA damage accumulation has been shown in autophagy-deficient cells and mice (Schneider et al., 2014). Autophagy modulates the DNA damage response via p62 degradation (Wang et al., 2016). It also directly targets DNA-associated proteins (Chen S. et al., 2015) and DNA resection proteins (Robert et al., 2011). In addition, nucleophagy, which is the process of selective degradation of nuclear components, has been observed in yeast and mammals (Mijaljica et al., 2012; Li and Nakatogawa, 2022). Autophagy upregulation in DNA damage in the heart is mediated via ATM, PARP1, and cGAS-STING signaling. ATM is vital for myocyte apoptosis and cardiac remodeling following myocardial infarction. ATM deficiency results in autophagic impairment in myocardial infarction (Thrasher et al., 2018). PARP1 is involved in autophagy induced by DNA damage (Muñoz-Gámez et al., 2009). Cardiomyocytes in patients suffering from persistent atrial fibrillation demonstrate significant DNA damage that is related to PARP1 activity (Zhang et al., 2019). Inhibiting PARP1 to activate autophagy might serve as an effective strategy to preserve cardiomyocyte function. The cGAS-STING signaling axis senses extranuclear chromatin due to genotoxic stress and DNA released from the mitochondria (Hopfner and Hornung, 2020). The cGAS-STING signaling pathway has been implicated in cellular senescence (Lan et al., 2019) and cardiac hypertrophy (Zhang Y. et al., 2020) through direct and indirect recognition of aberrant self-DNA. Autophagy has a conserved function in the cGAS-STING pathway (Gui et al., 2019). STING-mediated autophagy was discovered to be essential for removing cytosolic DNA following DNA damage. Autophagy/mitophagy inhibits STING signaling and its excessive responses (Oduro et al., 2022). This negative feedback controlling STING signaling ameliorates diabetic cardiomyopathy (Lu Q. B. et al., 2022).

Unc-51-like kinase (ULK) is essential for initiating autophagy in response to DNA damage. ULK1 is a transcriptional p53 target enhancing PARP1 activity, causing sustained autophagy and cell death (Gao et al., 2011; Joshi et al., 2016). ULK also mediates the phosphorylation of its interaction partner SEC16A and regulates the assembly of ER exit sites and ER-to-Golgi trafficking of specific cargo. ULK deficiency results in ER-to-Golgi trafficking defects and activates the UPR pathway (Joo et al., 2016). ULK1 may serve as the nexus that links autophagy, UPR, and DDR. However, direct data demonstrating this mechanism are still needed, particularly regarding its implications in CVDs.

# 4 ER stress, UPR, and autophagy in aging-related CVDs

### 4.1 Atherosclerosis

Advancing age remains the primary risk factor for atherosclerosis (Tyrrell and Goldstein, 2021). Multiple

aging-associated risk factors, including oxidative stress, inflammation, hyperhomocysteinemia, and free cholesterol accumulation in macrophages, are involved in atherosclerosis (Fu et al., 2018; Forman and Zhang, 2021; Tyrrell and Goldstein, 2021). The potential mechanism for these pro-aging risk factors suggests that they couple with activators in ER stress and maladaptive UPR, further facilitating atherosclerosis etiology (Yang et al., 2018; Jiang et al., 2021). ER stress and its response play a role in atherogenesis in coronary artery lesions, as demonstrated by augmented levels of ER stress markers like BiP and CHOP (Myoishi et al., 2007). Mice with CHOP-deficiency crossed with Apoe- or Ldlr-knockout mice experienced decreased necrosis in atherosclerotic plaques and reduced atherosclerotic lesion areas (Thorp et al., 2009). Type 2 diabetes mellitus (T2DM) is hallmarked by accelerated atherosclerosis (La Sala et al., 2019), and nearly half of all individuals with T2DM are older adults. High glucose levels induce more pronounced responses related to maladaptive UPR, senescence, and vascular endothelial cell barrier disruption. Direct evidence for the mechanism of ER stress in aged animal models requires further study.

Autophagy is vital for regulating atherosclerosis. CMA function becomes impaired in atherosclerosis progression, leading to NLRP3 inflammasome activation and IL-1ß secretion and promoting vascular inflammation and atherosclerosis progression (Qiao et al., 2021). Pharmacological activation of CMA provides a novel therapeutic method for atherosclerosis. Aging-related decreases in vascular mitochondrial functions and impaired mitophagy are vital for chronic hyperlipidemia and atherogenesis (Tyrrell and Goldstein, 2021). Autophagy also helps to maintain plaque cells, protecting them against oxidative stress, which can accelerate aging and is a hallmark of advanced atherosclerotic lesions (Yang et al., 2023). Autophagy stimulates the survival of VSMCs, whereas reduced autophagy promotes age-related changes in the vasculature. Defective VSMC autophagy accelerates the development of stress-induced premature senescence and atherogenesis (Grootaert et al., 2018).

Macrophage ATG5 deficiency in fat-fed Ldlr(-/-) mice increases apoptosis and oxidative stress in advanced lesional macrophages, promotes plaque necrosis, and exacerbates lesioned efferocytosis (Liao et al., 2012). Elevated cholesterol levels in mouse macrophages stimulate ER stress-induced apoptosis and promote autophagy using rapamycin-increased cholesterol efflux in oxLDL-loaded macrophages (Zahid et al., 2020). Therefore, autophagy may be a protective stress reaction to ER stress, apoptosis, and inflammation in regulating macrophage foam cell formation.

### 4.2 Cardiac hypertrophy and heart failure

Aging is a major risk factor for heart failure (Florio et al., 2020). Adaptive UPR is able to maintain homeostasis of the heart (Steiger et al., 2018). Aged mouse hearts showed much lower EIF2A phosphorylation levels and lower BiP expression levels while expressing the proapoptotic factor CHOP at high levels. ER stress responses may be the mechanisms involved in the protective effect of folic acid against cardiac aging (Ye et al., 2021). However, ER stress and maladaptive UPR may facilitate cardiac hypertrophy and heart failure via activation of PERK and CHOP signaling. ER and UPR component levels, such as XBP1s, CHOP, and ATF4, are elevated in patients suffering from heart failure (Yao et al., 2017; Schiattarella et al., 2021). Aging leads to increased ER stress, which contributes to mitochondrial dysfunction, while mitochondrial ROS generation also induces ER stress. Targeting ER stress and calpain activation may reverse age-dependent mitochondrial damage in the heart (Thompson et al., 2020).

Autophagy can increase protein degradation, while autophagy inhibition in cardiomyocytes promotes heart failure progression due to proteotoxicity (Delbridge et al., 2017). Mice with Parkin deficiency may exhibit normal cardiac functions while young (Kubli et al., 2013b) and accumulate abnormal mitochondria during aging (Kubli et al., 2013a). Aging-related cardiac remodeling is also related to autophagy dysfunction (Linton et al., 2015; Shirakabe et al., 2016). Autophagy activation via caloric restriction (CR) delays the development of aging-related cardiac disorders (Makino and Maeda, 2021). However, cardiomyocyte-specific autophagy activation through aMHC-Beclin-1 overexpression promotes hypertrophic remodeling and systolic dysfunction (Zhu et al., 2007). Autophagic cell death is also implicated in the pathological process of heart failure (Ding et al., 2020).

UPR signaling and autophagy are interrelated. The PERK/ ATF4 signaling activation and autophagy defects promote cardiomyocyte hypertrophy and cardiac fibroblast activation (Li et al., 2018b). However, ER stress activation through PERK-ATF4 can cause lethal autophagy-mediated cardiac atrophy (Vanhoutte et al., 2021).

### 4.3 Hypertension

The National Health and Nutrition Examination Survey indicated that 70% of people older than 60 have hypertension compared to only 32% of those aged 40–59 years (Mozaffarian et al., 2015).

ER stress is vital for vascular cell phenotype switching, dedifferentiation, calcification, and apoptosis and facilitates endothelial dysfunction and vascular remodeling in hypertension (Shanahan and Furmanik, 2017; Griendling et al., 2021). ER stress inhibition blunts hypertension growth among spontaneously hypertensive rats (Spitler et al., 2013; Naiel et al., 2019) and protects against hypertension-induced vascular dysfunction (Carlisle et al., 2016). Direct data for different effects of ER stress on aged mice or older patients and its potential regulation mechanisms remain limited.

Autophagy is involved in numerous pathological processes related to hypertension and target organ injuries. Aging is associated with downregulation of the Klotho protein, which favors autophagy-suppressed aging (Fernández et al., 2018). Klotho deficiency-triggered arterial stiffening is due to decreased elastin and increased collagen-1 levels and involves compensatory autophagy induction (Chen K. et al., 2015). Autophagy reactivation may remove dysfunctional mitochondria and limit damage to endothelial and cerebral cells in response to high salt level treatment, thereby reducing hypertension-related stroke occurrence (Forte et al., 2020). The relationship between ER stress and autophagy during hypertension remains poorly defined, and future studies are needed to explore their regulation in aging-associated blood vessel changes and hypertension.

## 5 Targeting aging and CVDs by modulating ER stress and autophagy

Due to the pivotal function of ER stress and autophagy during aging and CVD pathogenesis, methods to target proteins and signaling components involved in these cellular processes are attractive therapeutic avenues for disease interventions.

### 5.1 Pharmacological interventions

#### 5.1.1 Metformin

Metformin is a well-established and inexpensive principal antidiabetic drug (Shin et al., 2020). Its use can lead to a 21% reduction in mortality in myocardial infarction and a 16% reduction in mortality in heart failure (Han et al., 2019). Metformin stimulates the AMPK/ PPAR $\delta$  pathway to inhibit ER stress, enhances cardiac injuries, and endothelial dysfunctions and maintain vascular health (Cheang et al., 2014; Ge et al., 2019). In particular, it attenuates ER stress and guards against vascular damage in hypertension (Chen C. et al., 2019). It also inhibits mTOR signaling to promote autophagy and prevent atherosclerosis development (You et al., 2020).

#### 5.1.2 Sirtuin activators

Sirtuins are a family of nicotine adenine dinucleotide (NAD+)dependent histone deacetylases, which are referred to as "the fountain of youth" due to their critical function in longevity and aging anomalies (Kanfi et al., 2012; Verdin, 2015). SIRT1 is a master regulator of cellular stress. It guards cardiomyocytes from ER stressinduced cell death by modulating the PERK/eIF2 $\alpha$  pathway of UPR via deacetylation of eIF2 $\alpha$  on lysine K143 (Prola et al., 2017). Cardioprotective SIRT1 influences are also caused by its capability to affect autophagy by directly deacetylating autophagy machinery, such as ATG5, ATG7, and LC3 (Lee et al., 2008). SIRT1 may play a cardioprotective role in enhancing autophagy induced by ER stress by activating the eEF2K/eEF2 pathway (Pires Da Silva et al., 2020).

#### 5.1.3 Statins

Statins are effective cardioprotective drugs because they can inhibit HMG-CoA reductase. In addition, statins have beneficial pleiotropy anti-thrombotic, anti-oxidative, and anti-inflammatory effects (Oesterle et al., 2017). The beneficial impact of statin therapy on cardiac remodeling due to pressure overload and heart failure in rats is associated with ER stress reduction (Song et al., 2011). Rosuvastatin exerts a cardioprotective effect by enhancing the autophagy pathway by increasing Beclin-1 expression and MAPK activation and inhibiting mTOR signaling (Song et al., 2018).

#### 5.1.4 mTOR inhibition

A previous study discovered that ginkgo biloba leaf extract alleviates atherosclerosis in streptozotocin-induced diabetic

Apo $E^{-/-}$  mice by restraining ER stress via autophagy restoration using the mTOR signaling pathway (Tian et al., 2019). Therefore, this pathway is a possible target for modulating ER stress in CVDs.

### 5.2 CR and lifestyle modification

### 5.2.1 Caloric restriction

Hearts in patients undergoing caloric restriction (CR) exhibit mTOR suppression and activated autophagy. CR-elicited autophagy has a beneficial effect on cardiac aging (Makino and Maeda, 2021). Patients with obesity undergoing CR lose weight and alleviate ER stress, as demonstrated by a decrease in phosphorylated ATF6, CHOP, and JNK levels (López-Domènech et al., 2019). Low-level inhibition of fat consumption in rats can restore AMPK activities in adipose tissues, liver, and peripheral blood mononuclear cells by inhibiting ER stress and UPR modulators CHOP, EIF2A, PERK, and XBP1 (Vega-Martín et al., 2020).

#### 5.2.2 Physical exercise

Swimming has been shown to diminish ER stress and ROS production in mice and attenuate aging-related CVDs and cardiac dysfunctions by activating cGMP-protein kinase G signaling (Chang et al., 2020). Exercise also diminishes atherosclerosis plaques and boosts endothelial function by reducing ER stress, which is closely related to endothelial dysfunctions in diabetes and atherosclerosis (Cheang et al., 2017; Hong et al., 2018). Furthermore, analyses of human skeletal muscle biopsy samples revealed that exercise is related to the adaptive UPR in muscle remodeling and is impaired during aging (Hart et al., 2019).

Exercise augments acute autophagic activities in skeletal muscle as well as several other tissues, including the heart (Escobar et al., 2019). This mechanism involves increased Sirtuin 3 and PINK1/ Parkin levels (Zhao et al., 2018). In addition, the combination of long-term exercise together with CR may be more effective in activating autophagy and thus preventing heart failure than either therapy used alone (Moradi et al., 2019).

### 6 Conclusions and perspectives

Aging is a complex and multifaceted process that results in widespread functional decline affecting every organ and tissue. Determining the mechanisms by which aging enhances CVDs is important to identify new treatments that decrease the burden of aging-related CVDs.

Many biological processes change as a result of aging, contributing to an increased risk of CVDs. ER stress and autophagy are two typical cellular stress response pathways that can function synergistically during aging and aging-associated diseases. As individuals age, many components of the ER stress and autophagy pathways diminish and become less effective, thereby decreasing their reaction potential to various stressors and leading to or exacerbating existing diseases. ER stress response and autophagy pathways and their age-related modifications are also implicated in various other aging-related diseases in a variety of organs, including Alzheimer's disease, Parkinson's disease, type 2 diabetes mellitus, and cancer involving disturbed proteostasis. ER stress and impaired autophagy flux are involved in neuronal degeneration and brain injury. A neuroprotective agent, docosahexanoic acid, has unique functions by reducing ER and oxidative stress and modulating autophagy (Yin et al., 2017). In Alzheimer's disease, ER stress response and impaired autophagy take part in cellular aging and neuroinflammation (Reddy and Oliver, 2019; Uddin et al., 2021). Therefore, coupling UPR, autophagy, and inflammation can facilitate novel therapeutic strategies to mitigate cellular stress and inflammation, which are involved in chronic obstructive pulmonary disease and neurodegenerative disorders (Chipurupalli et al., 2021). Harnessing ER stress response and autophagy is of broader significance for the prevention and treatment of more agerelated diseases than CVDs.

Both ER stress and autophagy are associated with one or more cell death processes (Liu et al., 2023; An et al., 2024). Whether they are connected in aging and CVDs by modulating cell death processes is a topic that warrants further study.

Although our awareness of the roles of these aging processes and their impact on CVD progression has recently improved, detailed molecular mechanisms and cellular pathways remain under investigation, with critical issues still unresolved. Therefore, further exploration of the precise mechanisms governing the interplay of ER stress and autophagy and their overlap with aging risk would help elucidate the pathways involved in cellular homeostasis maintenance in the cardiovascular system. Such research could aid with therapeutic development targeted at maintaining a normal ER stress reaction and autophagy for later life, potentially delaying or preventing aging-related CVDs. This represents an unexplored avenue in CVD prevention.

### References

Abbadie, C., and Pluquet, O. (2020). Unfolded protein response (UPR) controls major senescence hallmarks. *Trends Biochem. Sci.* 45, 371–374. doi:10.1016/j.tibs.2020.02.005

Alfaras, I., Di Germanio, C., Bernier, M., Csiszar, A., Ungvari, Z., Lakatta, E. G., et al. (2016). Pharmacological strategies to retard cardiovascular aging. *Circ. Res.* 118, 1626–1642. doi:10.1161/circresaha.116.307475

Almanza, A., Carlesso, A., Chintha, C., Creedican, S., Doultsinos, D., Leuzzi, B., et al. (2019). Endoplasmic reticulum stress signalling - from basic mechanisms to clinical applications. *Febs J.* 286, 241–278. doi:10.1111/febs.14608

An, Y., Wang, X., Guan, X., Yuan, P., Liu, Y., Wei, L., et al. (2024). Endoplasmic reticulum stress-mediated cell death in cardiovascular disease. *Cell. Stress Chaperones* 29, 158–174. doi:10.1016/j.cstres.2023.12.003

Angelova, D. M., and Brown, D. R. (2018). Altered processing of  $\beta$ -amyloid in SH-SY5Y cells induced by model senescent microglia. ACS Chem. Neurosci. 9, 3137–3152. doi:10.1021/acschemneuro.8b00334

Argemí, J., Kress, T. R., Chang, H. C. Y., Ferrero, R., Bértolo, C., Moreno, H., et al. (2017). X-Box binding protein 1 regulates unfolded protein, acute-phase, and DNA damage responses during regeneration of mouse liver. *Gastroenterology* 152, 1203–1216. doi:10.1053/j.gastro.2016.12.040

Bachar, E., Ariav, Y., Ketzinel-Gilad, M., Cerasi, E., Kaiser, N., and Leibowitz, G. (2009). Glucose amplifies fatty acid-induced endoplasmic reticulum stress in pancreatic beta-cells via activation of mTORC1. *PLoS One* 4, e4954. doi:10.1371/journal.pone.0004954

Ballinger, S. W., Patterson, C., Knight-Lozano, C. A., Burow, D. L., Conklin, C. A., Hu, Z., et al. (2002). Mitochondrial integrity and function in atherogenesis. *Circulation* 106, 544–549. doi:10.1161/01.cir.0000023921.93743.89

Baris, O. R., Ederer, S., Neuhaus, J. F. G., von Kleist-Retzow, J. C., Wunderlich, C. M., Pal, M., et al. (2015). Mosaic deficiency in mitochondrial oxidative metabolism promotes cardiac arrhythmia during aging. *Cell. Metab.* 21, 667–677. doi:10.1016/j. cmet.2015.04.005

Battaglioni, S., Benjamin, D., Wälchli, M., Maier, T., and Hall, M. N. (2022). mTOR substrate phosphorylation in growth control. *Cell*. 185, 1814–1836. doi:10.1016/j.cell. 2022.04.013

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CM: conceptualization, data curation, formal analysis, funding acquisition, investigation, methodology, project administration, resources, software, supervision, validation, visualization, writing–original draft, and writing–review and editing. YL: methodology, supervision, validation, writing–review and editing. ZF: Writing–original draft and writing–review and editing.

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Benvenuti, S., Cramer, R., Quinn, C. C., Bruce, J., Zvelebil, M., Corless, S., et al. (2002). Differential proteome analysis of replicative senescence in rat embryo fibroblasts. *Mol. Cell. Proteomics* 1, 280–292. doi:10.1074/mcp.m100028-mcp200

Bhuiyan, M. S., and Fukunaga, K. (2009). Stimulation of sigma-1 receptor signaling by dehydroepiandrosterone ameliorates pressure overload-induced hypertrophy and dysfunctions in ovariectomized rats. *Expert Opin. Ther. Targets* 13, 1253–1265. doi:10.1517/14728220903264064

Blackford, A. N., and Jackson, S. P. (2017). ATM, ATR, and DNA-PK: the trinity at the heart of the DNA damage response. *Mol. Cell.* 66, 801–817. doi:10.1016/j.molcel. 2017.05.015

Botto, N., Rizza, A., Colombo, M. G., Mazzone, A. M., Manfredi, S., Masetti, S., et al. (2001). Evidence for DNA damage in patients with coronary artery disease. *Mutat. Res.* 493, 23–30. doi:10.1016/s1383-5718(01)00162-0

Bouska, M., Huang, K., Kang, P., and Bai, H. (2019). Organelle aging: lessons from model organisms. J. Genet. Genomics 46, 171–185. doi:10.1016/j.jgg.2019.03.011

Boutouyrie, P., Chowienczyk, P., Humphrey, J. D., and Mitchell, G. F. (2021). Arterial stiffness and cardiovascular risk in hypertension. *Circ. Res.* 128, 864–886. doi:10.1161/circresaha.121.318061

Brown, M. K., and Naidoo, N. (2012). The endoplasmic reticulum stress response in aging and age-related diseases. *Front. Physiol.* 3, 263. doi:10.3389/fphys.2012.00263

Cai, H., Liu, Y., Men, H., and Zheng, Y. (2021). Protective mechanism of humanin against oxidative stress in aging-related cardiovascular diseases. *Front. Endocrinol.* (*Lausanne*) 12, 683151. doi:10.3389/fendo.2021.683151

Carlisle, R. E., Werner, K. E., Yum, V., Tat, V., Memon, M., et al. (2016). Endoplasmic reticulum stress inhibition reduces hypertension through the preservation of resistance blood vessel structure and function. *J. Hypertens.* 34, 1556–1569. doi:10.1097/hjh. 00000000000943

Carreras-Sureda, A., Jaña, F., Urra, H., Durand, S., Mortenson, D. E., Sagredo, A., et al. (2019). Non-canonical function of IRE1a determines mitochondria-associated endoplasmic reticulum composition to control calcium transfer and bioenergetics. *Nat. Cell. Biol.* 21, 755–767. doi:10.1038/s41556-019-0329-y

Carreras-Sureda, A., Pihán, P., and Hetz, C. (2017). The unfolded protein response: at the intersection between endoplasmic reticulum function and mitochondrial bioenergetics. *Front. Oncol.* 7, 55. doi:10.3389/fonc.2017.00055

Chang, P., Zhang, X., Zhang, M., Hu, L., Zhao, H., et al. (2020). Swimming exercise inhibits myocardial ER stress in the hearts of aged mice by enhancing cGMP-PKG signaling. *Mol. Med. Rep.* 21, 549–556. doi:10.3892/mmr.2019.10864

Cheang, W. S., Tian, X. Y., Wong, W. T., Lau, C. W., Lee, S. S. T., Chen, Z. Y., et al. (2014). Metformin protects endothelial function in diet-induced obese mice by inhibition of endoplasmic reticulum stress through 5' adenosine monophosphate-activated protein kinase-peroxisome proliferator-activated receptor  $\delta$  pathway. *Arterioscler. Thromb. Vasc. Biol.* 34, 830–836. doi:10.1161/attybaha.113.301938

Cheang, W. S., Wong, W. T., Zhao, L., Xu, J., Wang, L., Lau, C. W., et al. (2017). PPAR $\delta$  is required for exercise to attenuate endoplasmic reticulum stress and endothelial dysfunction in diabetic mice. *Diabetes* 66, 519–528. doi:10.2337/db15-1657

Chen, C., Kassan, A., Castañeda, D., Gabani, M., Choi, S. K., and Kassan, M. (2019b). Metformin prevents vascular damage in hypertension through the AMPK/ER stress pathway. *Hypertens. Res.* 42, 960–969. doi:10.1038/s41440-019-0212-z

Chen, H., Detmer, S. A., Ewald, A. J., Griffin, E. E., Fraser, S. E., and Chan, D. C. (2003). Mitofusins Mfn1 and Mfn2 coordinately regulate mitochondrial fusion and are essential for embryonic development. *J. Cell. Biol.* 160, 189–200. doi:10.1083/jcb. 200211046

Chen, K., Zhou, X., and Sun, Z. (2015b). Haplodeficiency of Klotho gene causes arterial stiffening via upregulation of scleraxis expression and induction of autophagy. *Hypertension* 66, 1006–1013. doi:10.1161/hypertensionaha.115.06033

Chen, Q., Samidurai, A., Thompson, J., Hu, Y., Das, A., Willard, B., et al. (2020). Endoplasmic reticulum stress-mediated mitochondrial dysfunction in aged hearts. *Biochim. Biophys. Acta Mol. Basis Dis.* 1866, 165899. doi:10.1016/j.bbadis.2020.165899

Chen, Q., Thompson, J., Hu, Y., Das, A., and Lesnefsky, E. J. (2017). Metformin attenuates ER stress-induced mitochondrial dysfunction. *Transl. Res.* 190, 40–50. doi:10.1016/j.trsl.2017.09.003

Chen, Q., Thompson, J., Hu, Y., Das, A., and Lesnefsky, E. J. (2019a). Cardiac specific knockout of p53 decreases ER stress-induced mitochondrial damage. *Front. Cardiovasc Med.* 6, 10. doi:10.3389/fcvm.2019.00010

Chen, Q., Thompson, J., Hu, Y., and Lesnefsky, E. J. (2023). Endoplasmic reticulum stress and alterations of peroxiredoxins in aged hearts. *Mech. Ageing Dev.* 215, 111859. doi:10.1016/j.mad.2023.111859

Chen, S., Wang, C., Sun, L., Wang, D. L., Chen, L., Huang, Z., et al. (2015a). RAD6 promotes homologous recombination repair by activating the autophagymediated degradation of heterochromatin protein HP1. *Mol. Cell. Biol.* 35, 406–416. doi:10.1128/mcb.01044-14

Chen, Y., Csordás, G., Jowdy, C., Schneider, T. G., Csordás, N., Wang, W., et al. (2012). Mitofusin 2-containing mitochondrial-reticular microdomains direct rapid cardiomyocyte bioenergetic responses via interorganelle Ca(2+) crosstalk. *Circ. Res.* 111, 863–875. doi:10.1161/circresaha.112.266585

Chipurupalli, S., Samavedam, U., and Robinson, N. (2021). Crosstalk between ER stress, autophagy and inflammation. *Front. Med. (Lausanne)* 8, 758311. doi:10.3389/fmed.2021.758311

Chung, H. Y., Kim, D. H., Lee, E. K., Chung, K. W., Chung, S., Lee, B., et al. (2019). Redefining chronic inflammation in aging and age-related diseases: proposal of the senoinflammation concept. *Aging Dis.* 10, 367–382. doi:10.14336/ad.2018.0324

Ciccia, A., and Elledge, S. J. (2010). The DNA damage response: making it safe to play with knives. *Mol. Cell.* 40, 179–204. doi:10.1016/j.molcel.2010.09.019

Clementi, E., Inglin, L., Beebe, E., Gsell, C., Garajova, Z., and Markkanen, E. (2020). Persistent DNA damage triggers activation of the integrated stress response to promote cell survival under nutrient restriction. *BMC Biol.* 18, 36. doi:10.1186/s12915-020-00771-x

Collins, H. E., Zou, L., Qu, J., Zhou, L., Litovsky, S. H., et al. (2014). Stromal interaction molecule 1 is essential for normal cardiac homeostasis through modulation of ER and mitochondrial function. *Am. J. Physiol. Heart Circ. Physiol.* 306, H1231–H1239. doi:10.1152/ajpheart.00075.2014

Cominacini, L., Mozzini, C., Garbin, U., Pasini, A., Stranieri, C., Solani, E., et al. (2015). Endoplasmic reticulum stress and Nrf2 signaling in cardiovascular diseases. *Free Radic. Biol. Med.* 88, 233–242. doi:10.1016/j.freeradbiomed.2015.05.027

Costantino, S., Paneni, F., and Cosentino, F. (2016). Ageing, metabolism and cardiovascular disease. J. Physiol. 594, 2061–2073. doi:10.1113/jp270538

Daneshgar, N., Rabinovitch, P. S., and Dai, D. F. (2021). TOR signaling pathway in cardiac aging and heart failure. *Biomolecules* 11, 168. doi:10.3390/biom11020168

Delbridge, L. M. D., Mellor, K. M., Taylor, D. J., and Gottlieb, R. A. (2017). Myocardial stress and autophagy: mechanisms and potential therapies. *Nat. Rev. Cardiol.* 14, 412–425. doi:10.1038/nrcardio.2017.35

De Leonibus, C., Cinque, L., and Settembre, C. (2019). Emerging lysosomal pathways for quality control at the endoplasmic reticulum. *FEBS Lett.* 593, 2319–2329. doi:10.1002/1873-3468.13571

Deretic, V. (2021). Autophagy in inflammation, infection, and immunometabolism. *Immunity* 54, 437–453. doi:10.1016/j.immuni.2021.01.018 Diao, R. Y., and Gustafsson Å, B. (2022). Mitochondrial quality surveillance: mitophagy in cardiovascular health and disease. *Am. J. Physiol. Cell. Physiol.* 322, C218-c230. doi:10.1152/ajpcell.00360.2021

Ding, J., Chen, Y. X., Chen, Y., Mou, Y., Sun, X. T., Dai, D. P., et al. (2020). Overexpression of FNTB and the activation of Ras induce hypertrophy and promote apoptosis and autophagic cell death in cardiomyocytes. *J. Cell. Mol. Med.* 24, 8998–9011. doi:10.1111/jcmm.15533

Dörr, J. R., Yu, Y., Milanovic, M., Beuster, G., Zasada, C., Däbritz, J. H. M., et al. (2013). Synthetic lethal metabolic targeting of cellular senescence in cancer therapy. *Nature* 501, 421–425. doi:10.1038/nature12437

Dufey, E., Bravo-San Pedro, J. M., Eggers, C., González-Quiroz, M., Urra, H., Sagredo, A. I., et al. (2020). Genotoxic stress triggers the activation of IRE1a-dependent RNA decay to modulate the DNA damage response. *Nat. Commun.* 11, 2401. doi:10.1038/ s41467-020-15694-y

Erickson, R. R., Dunning, L. M., and Holtzman, J. L. (2006). The effect of aging on the chaperone concentrations in the hepatic, endoplasmic reticulum of male rats: the possible role of protein misfolding due to the loss of chaperones in the decline in physiological function seen with age. *J. Gerontol. A Biol. Sci. Med. Sci.* 61, 435–443. doi:10.1093/gerona/61.5.435

Escobar, K. A., Cole, N. H., Mermier, C. M., and VanDusseldorp, T. A. (2019). Autophagy and aging: maintaining the proteome through exercise and caloric restriction. *Aging Cell.* 18, e12876. doi:10.1111/acel.12876

Fatima, S., Ambreen, S., Mathew, A., Elwakiel, A., Gupta, A., Singh, K., et al. (2022). ER-stress and senescence coordinately promote endothelial barrier dysfunction in diabetes-induced atherosclerosis. *Nutrients* 14, 2786. doi:10.3390/nu14142786

Feng, Y., Chen, Y., Wu, X., Chen, J., Zhou, Q., Liu, B., et al. (2024). Interplay of energy metabolism and autophagy. *Autophagy* 20, 4–14. doi:10.1080/15548627.2023.2247300

Fernández, Á. F., Sebti, S., Wei, Y., Zou, Z., Shi, M., McMillan, K. L., et al. (2018). Disruption of the beclin 1-BCL2 autophagy regulatory complex promotes longevity in mice. *Nature* 558, 136–140. doi:10.1038/s41586-018-0162-7

Ferrucci, L., Corsi, A., Lauretani, F., Bandinelli, S., Bartali, B., Taub, D. D., et al. (2005). The origins of age-related proinflammatory state. *Blood* 105, 2294–2299. doi:10.1182/blood-2004-07-2599

Finkel, T. (2011). Signal transduction by reactive oxygen species. J. Cell. Biol. 194, 7–15. doi:10.1083/jcb.201102095

Florio, M. C., Magenta, A., Beji, S., Lakatta, E. G., and Capogrossi, M. C. (2020). Aging, MicroRNAs, and heart failure. *Curr. Probl. Cardiol.* 45, 100406. doi:10.1016/j. cpcardiol.2018.12.003

Flynn, J. M., O'Leary, M. N., Zambataro, C. A., Academia, E. C., Presley, M. P., Garrett, B. J., et al. (2013). Late-life rapamycin treatment reverses age-related heart dysfunction. *Aging Cell.* 12, 851–862. doi:10.1111/acel.12109

Forman, H. J., and Zhang, H. (2021). Targeting oxidative stress in disease: promise and limitations of antioxidant therapy. *Nat. Rev. Drug Discov.* 20, 689–709. doi:10.1038/ s41573-021-00233-1

Forte, M., Bianchi, F., Cotugno, M., Marchitti, S., De Falco, E., Raffa, S., et al. (2020). Pharmacological restoration of autophagy reduces hypertension-related stroke occurrence. *Autophagy* 16, 1468–1481. doi:10.1080/15548627.2019.1687215

Franceschi, C., Garagnani, P., Vitale, G., Capri, M., and Salvioli, S. (2017). Inflammaging and 'garb-aging. *Trends Endocrinol. Metab.* 28, 199–212. doi:10.1016/ j.tem.2016.09.005

Fu, Y., Wang, X., and Kong, W. (2018). Hyperhomocysteinaemia and vascular injury: advances in mechanisms and drug targets. *Br. J. Pharmacol.* 175, 1173–1189. doi:10. 1111/bph.13988

Gao, G., Chen, W., Yan, M., Liu, J., Luo, H., Wang, C., et al. (2020). Rapamycin regulates the balance between cardiomyocyte apoptosis and autophagy in chronic heart failure by inhibiting mTOR signaling. *Int. J. Mol. Med.* 45, 195–209. doi:10.3892/ijmm. 2019.4407

Gao, W., Shen, Z., Shang, L., and Wang, X. (2011). Upregulation of human autophagy-initiation kinase ULK1 by tumor suppressor p53 contributes to DNA-damage-induced cell death. *Cell. Death Differ.* 18, 1598–1607. doi:10.1038/cdd.2011.33

Gaur, A., Carr, F., and Warriner, D. (2024). Cardiogeriatrics: the current state of the art. *Heart* 110, 933–939. doi:10.1136/heartjnl-2022-322117

Gavilán, E., Medina-Guzman, R., Bahatyrevich-Kharitonik, B., and Ruano, D. (2024). Protein quality control systems and ER stress as key players in SARS-CoV-2-induced neurodegeneration. *Cells* 13, 123. doi:10.3390/cells13020123

Ge, C. X., Xu, M. X., Qin, Y. T., Gu, T. T., Lou, D. S., Li, Q., et al. (2019). Endoplasmic reticulum stress-induced iRhom2 up-regulation promotes macrophage-regulated cardiac inflammation and lipid deposition in high fat diet (HFD)-challenged mice: intervention of fisetin and metformin. *Free Radic. Biol. Med.* 141, 67–83. doi:10.1016/j. freeradbiomed.2019.05.031

Gelmetti, V., De Rosa, P., Torosantucci, L., Marini, E. S., Romagnoli, A., Di Rienzo, M., et al. (2017). PINK1 and BECN1 relocalize at mitochondria-associated membranes during mitophagy and promote ER-mitochondria tethering and autophagosome formation. *Autophagy* 13, 654–669. doi:10.1080/15548627.2016. 1277309

Gimbrone, M. A., Jr., and García-Cardeña, G. (2016). Endothelial cell dysfunction and the pathobiology of atherosclerosis. *Circ. Res.* 118, 620–636. doi:10.1161/circresaha.115. 306301

Goldspink, D. F., Burniston, J. G., and Tan, L. B. (2003). Cardiomyocyte death and the ageing and failing heart. *Exp. Physiol.* 88, 447–458. doi:10.1113/eph8802549

Gounder, S. S., Kannan, S., Devadoss, D., Miller, C. J., Whitehead, K. J., Whitehead, K. S., et al. (2012). Impaired transcriptional activity of Nrf2 in age-related myocardial oxidative stress is reversible by moderate exercise training. *PLoS One* 7, e45697. doi:10. 1371/journal.pone.0045697

Green, D. R., Galluzzi, L., and Kroemer, G. (2014). Cell biology. Metabolic control of cell death. *Science* 345, 1250256. doi:10.1126/science.1250256

Grey, M. J., Cloots, E., Simpson, M. S., LeDuc, N., Serebrenik, Y. V., De Luca, H., et al. (2020). IRE1 $\beta$  negatively regulates IRE1 $\alpha$  signaling in response to endoplasmic reticulum stress. *J. Cell. Biol.* 219, e201904048. doi:10.1083/jcb.201904048

Griendling, K. K., Camargo, L. L., Rios, F. J., Alves-Lopes, R., Montezano, A. C., and Touyz, R. M. (2021). Oxidative stress and hypertension. *Circ. Res.* 128, 993–1020. doi:10.1161/circresaha.121.318063

Grootaert, M. O. J., Moulis, M., Roth, L., Martinet, W., Vindis, C., Bennett, M. R., et al. (2018). Vascular smooth muscle cell death, autophagy and senescence in atherosclerosis. *Cardiovasc Res.* 114, 622–634. doi:10.1093/cvr/cvy007

Gude, N. A., Broughton, K. M., Firouzi, F., and Sussman, M. A. (2018). Cardiac ageing: extrinsic and intrinsic factors in cellular renewal and senescence. *Nat. Rev. Cardiol.* 15, 523–542. doi:10.1038/s41569-018-0061-5

Gui, X., Yang, H., Li, T., Tan, X., Shi, P., Li, M., et al. (2019). Autophagy induction via STING trafficking is a primordial function of the cGAS pathway. *Nature* 567, 262–266. doi:10.1038/s41586-019-1006-9

Han, Y., Xie, H., Liu, Y., Gao, P., Yang, X., and Shen, Z. (2019). Effect of metformin on all-cause and cardiovascular mortality in patients with coronary artery diseases: a systematic review and an updated meta-analysis. *Cardiovasc Diabetol.* 18, 96. doi:10. 1186/s12933-019-0900-7

Hart, C. R., Ryan, Z. C., Pfaffenbach, K. T., Dasari, S., Parvizi, M., Lalia, A. Z., et al. (2019). Attenuated activation of the unfolded protein response following exercise in skeletal muscle of older adults. *Aging (Albany NY)* 11, 7587–7604. doi:10.18632/aging. 102273

Hayflick, L., and Moorhead, P. S. (1961). The serial cultivation of human diploid cell strains. *Exp. Cell. Res.* 25, 585–621. doi:10.1016/0014-4827(61)90192-6

He, C. (2022). Balancing nutrient and energy demand and supply via autophagy. *Curr. Biol.* 32, R684-r696. doi:10.1016/j.cub.2022.04.071

Henning, R. H., and Brundel, B. (2017). Proteostasis in cardiac health and disease. Nat. Rev. Cardiol. 14, 637-653. doi:10.1038/nrcardio.2017.89

Hetz, C. (2012). The unfolded protein response: controlling cell fate decisions under ER stress and beyond. *Nat. Rev. Mol. Cell. Biol.* 13, 89–102. doi:10.1038/nrm3270

Hetz, C., and Saxena, S. (2017). ER stress and the unfolded protein response in neurodegeneration. *Nat. Rev. Neurol.* 13, 477–491. doi:10.1038/nrneurol.2017.99

Hetz, C., Zhang, K., and Kaufman, R. J. (2020). Mechanisms, regulation and functions of the unfolded protein response. *Nat. Rev. Mol. Cell. Biol.* 21, 421–438. doi:10.1038/ s41580-020-0250-z

Hinds, J. W., and McNelly, N. A. (1978). Dispersion of cisternae of rough endoplasmic reticulum in aging CNS neurons: a strictly linear trend. *Am. J. Anat.* 152, 433–439. doi:10.1002/aja.1001520311

Hollien, J., and Weissman, J. S. (2006). Decay of endoplasmic reticulum-localized mRNAs during the unfolded protein response. *Science* 313, 104–107. doi:10.1126/ science.1129631

Hong, J., Kim, K., Park, E., Lee, J., Markofski, M. M., Marrelli, S. P., et al. (2018). Exercise ameliorates endoplasmic reticulum stress-mediated vascular dysfunction in mesenteric arteries in atherosclerosis. *Sci. Rep.* 8, 7938. doi:10.1038/s41598-018-26188-9

Hopfner, K. P., and Hornung, V. (2020). Molecular mechanisms and cellular functions of cGAS-STING signalling. *Nat. Rev. Mol. Cell. Biol.* 21, 501–521. doi:10. 1038/s41580-020-0244-x

Hoshino, A., Mita, Y., Okawa, Y., Ariyoshi, M., Iwai-Kanai, E., Ueyama, T., et al. (2013). Cytosolic p53 inhibits Parkin-mediated mitophagy and promotes mitochondrial dysfunction in the mouse heart. *Nat. Commun.* 4, 2308. doi:10.1038/ncomms3308

Hsu, Y. J., Hsu, S. C., Hsu, C. P., Chen, Y. H., Chang, Y. L., Sadoshima, J., et al. (2017). Sirtuin 1 protects the aging heart from contractile dysfunction mediated through the inhibition of endoplasmic reticulum stress-mediated apoptosis in cardiac-specific Sirtuin 1 knockout mouse model. *Int. J. Cardiol.* 228, 543–552. doi:10.1016/j.ijcard. 2016.11.247

Hua, Y., Zhang, Y., Ceylan-Isik, A. F., Wold, L. E., Nunn, J. M., and Ren, J. (2011). Chronic Akt activation accentuates aging-induced cardiac hypertrophy and myocardial contractile dysfunction: role of autophagy. *Basic Res. Cardiol.* 106, 1173–1191. doi:10. 1007/s00395-011-0222-8

Hussain, S. G., and Ramaiah, K. V. (2007). Reduced eIF2alpha phosphorylation and increased proapoptotic proteins in aging. *Biochem. Biophys. Res. Commun.* 355, 365–370. doi:10.1016/j.bbrc.2007.01.156

Hwang, J., and Qi, L. (2018). Quality control in the endoplasmic reticulum: crosstalk between ERAD and UPR pathways. *Trends Biochem. Sci.* 43, 593–605. doi:10.1016/j. tibs.2018.06.005

Jaishy, B., Zhang, Q., Chung, H. S., Riehle, C., Soto, J., Jenkins, S., et al. (2015). Lipidinduced NOX2 activation inhibits autophagic flux by impairing lysosomal enzyme activity. *J. Lipid Res.* 56, 546–561. doi:10.1194/jlr.M055152

Ji, Y., Luo, X., Yang, Y., Dai, Z., Wu, G., and Wu, Z. (2018). Endoplasmic reticulum stress-induced apoptosis in intestinal epithelial cells: a feed-back regulation by mechanistic target of rapamycin complex 1 (mTORC1). *J. Anim. Sci. Biotechnol.* 9, 38. doi:10.1186/s40104-018-0253-1

Jia, K., Chen, D., and Riddle, D. L. (2004). The TOR pathway interacts with the insulin signaling pathway to regulate *C. elegans* larval development, metabolism and life span. *Development* 131, 3897–3906. doi:10.1242/dev.01255

Jiang, S., Xu, W., Chen, Z., Cui, C., Fan, X., Cai, J., et al. (2021). Hydrogen sulphide reduces hyperhomocysteinaemia-induced endothelial ER stress by sulfhydrating protein disulphide isomerase to attenuate atherosclerosis. *J. Cell. Mol. Med.* 25, 3437–3448. doi:10.1111/jcmm.16423

Jiao, P., Fan, W., Ma, X., Lin, R., Zhao, Y., Li, Y., et al. (2023). SARS-CoV-2 nonstructural protein 6 triggers endoplasmic reticulum stress-induced autophagy to degrade STING1. *Autophagy* 19, 3113–3131. doi:10.1080/15548627.2023.2238579

Jin, J. K., Blackwood, E. A., Azizi, K., Thuerauf, D. J., Fahem, A. G., Hofmann, C., et al. (2017). ATF6 decreases myocardial ischemia/reperfusion damage and links ER stress and oxidative stress signaling pathways in the heart. *Circ. Res.* 120, 862–875. doi:10. 1161/circresaha.116.310266

Joo, J. H., Wang, B., Frankel, E., Ge, L., Xu, L., Iyengar, R., et al. (2016). The noncanonical role of ULK/ATG1 in ER-to-golgi trafficking is essential for cellular homeostasis. *Mol. Cell.* 62, 491–506. doi:10.1016/j.molcel.2016.04.020

Joshi, A., Iyengar, R., Joo, J. H., Li-Harms, X. J., Wright, C., Marino, R., et al. (2016). Nuclear ULK1 promotes cell death in response to oxidative stress through PARP1. *Cell. Death Differ.* 23, 216–230. doi:10.1038/cdd.2015.88

Juretschke, T., and Beli, P. (2021). Causes and consequences of DNA damage-induced autophagy. *Matrix Biol.* 100-101, 39–53. doi:10.1016/j.matbio.2021.02.004

Kanfi, Y., Naiman, S., Amir, G., Peshti, V., Zinman, G., Nahum, L., et al. (2012). The sirtuin SIRT6 regulates lifespan in male mice. *Nature* 483, 218–221. doi:10.1038/ nature10815

Kang, C., and Elledge, S. J. (2016). How autophagy both activates and inhibits cellular senescence. *Autophagy* 12, 898–899. doi:10.1080/15548627.2015.1121361

Kapuy, O. (2024). Mechanism of decision making between autophagy and apoptosis induction upon endoplasmic reticulum stress. *Int. J. Mol. Sci.* 25, 4368. doi:10.3390/ ijms25084368

Karagöz, G. E., Acosta-Alvear, D., Nguyen, H. T., Lee, C. P., Chu, F., and Walter, P. (2017). An unfolded protein-induced conformational switch activates mammalian IRE1. *Elife* 6, e30700. doi:10.7554/eLife.30700

Kato, H., Nakajima, S., Saito, Y., Takahashi, S., Katoh, R., and Kitamura, M. (2012). mTORC1 serves ER stress-triggered apoptosis via selective activation of the IRE1-JNK pathway. *Cell. Death Differ.* 19, 310–320. doi:10.1038/cdd.2011.98

Kaushik, S., Tasset, I., Arias, E., Pampliega, O., Wong, E., Martinez-Vicente, M., et al. (2021). Autophagy and the hallmarks of aging. *Ageing Res. Rev.* 72, 101468. doi:10.1016/j.arr.2021.101468

Kern, A., and Behl, C. (2019). Special issue on "proteostasis and autophagy. *Cells* 8, 642. doi:10.3390/cells8070642

Kilberg, M. S., Shan, J., and Su, N. (2009). ATF4-dependent transcription mediates signaling of amino acid limitation. *Trends Endocrinol. Metab.* 20, 436–443. doi:10.1016/j.tem.2009.05.008

Kim, J., and Guan, K. L. (2019). mTOR as a central hub of nutrient signalling and cell growth. *Nat. Cell. Biol.* 21, 63–71. doi:10.1038/s41556-018-0205-1

Kitada, M., and Koya, D. (2021). Autophagy in metabolic disease and ageing. Nat. Rev. Endocrinol. 17, 647-661. doi:10.1038/s41574-021-00551-9

Komatsu, M., Kurokawa, H., Waguri, S., Taguchi, K., Kobayashi, A., Ichimura, Y., et al. (2010). The selective autophagy substrate p62 activates the stress responsive transcription factor Nrf2 through inactivation of Keap1. *Nat. Cell. Biol.* 12, 213–223. doi:10.1038/ncb2021

Korolchuk, V. I., Mansilla, A., Menzies, F. M., and Rubinsztein, D. C. (2009). Autophagy inhibition compromises degradation of ubiquitin-proteasome pathway substrates. *Mol. Cell.* 33, 517–527. doi:10.1016/j.molcel.2009.01.021

Kovacic, J. C., Moreno, P., Hachinski, V., Nabel, E. G., and Fuster, V. (2011). Cellular senescence, vascular disease, and aging: Part 1 of a 2-part review. *Circulation* 123, 1650–1660. doi:10.1161/circulationaha.110.007021

Kubli, D. A., Quinsay, M. N., and Gustafsson, A. B. (2013a). Parkin deficiency results in accumulation of abnormal mitochondria in aging myocytes. *Commun. Integr. Biol.* 6, e24511. doi:10.4161/cib.24511

Kubli, D. A., Zhang, X., Lee, Y., Hanna, R. A., Quinsay, M. N., Nguyen, C. K., et al. (2013b). Parkin protein deficiency exacerbates cardiac injury and reduces survival

following myocardial infarction. J. Biol. Chem. 288, 915-926. doi:10.1074/jbc.M112. 411363

Kuilman, T., Michaloglou, C., Mooi, W. J., and Peeper, D. S. (2010). The essence of senescence. *Genes. Dev.* 24, 2463–2479. doi:10.1101/gad.1971610

Kuo, C. T., You, G. T., Jian, Y. J., Chen, T. S., Siao, Y. C., Hsu, A. L., et al. (2020). AMPK-mediated formation of stress granules is required for dietary restriction-induced longevity in *Caenorhabditis elegans. Aging Cell.* 19, e13157. doi:10.1111/acel.13157

Lamark, T., and Johansen, T. (2021). Mechanisms of selective autophagy. Annu. Rev. Cell. Dev. Biol. 37, 143–169. doi:10.1146/annurev-cellbio-120219-035530

Lan, Y. Y., Heather, J. M., Eisenhaure, T., Garris, C. S., Lieb, D., Raychowdhury, R., et al. (2019). Extranuclear DNA accumulates in aged cells and contributes to senescence and inflammation. *Aging Cell.* 18, e12901. doi:10.1111/acel.12901

LaRocca, T. J., Hearon, C. M., Jr., Henson, G. D., and Seals, D. R. (2014). Mitochondrial quality control and age-associated arterial stiffening. *Exp. Gerontol.* 58, 78–82. doi:10.1016/j.exger.2014.07.008

LaRocca, T. J., Henson, G. D., Thorburn, A., Sindler, A. L., Pierce, G. L., and Seals, D. R. (2012). Translational evidence that impaired autophagy contributes to arterial ageing. *J. Physiol.* 590, 3305–3316. doi:10.1113/jphysiol.2012.229690

La Sala, L., Prattichizzo, F., and Ceriello, A. (2019). The link between diabetes and atherosclerosis. *Eur. J. Prev. Cardiol.* 26, 15–24. doi:10.1177/2047487319878373

Lavandero, S., Chiong, M., Rothermel, B. A., and Hill, J. A. (2015). Autophagy in cardiovascular biology. J. Clin. Investig. 125, 55-64. doi:10.1172/jci73943

Lee, I. H., Cao, L., Mostoslavsky, R., Lombard, D. B., Liu, J., Bruns, N. E., et al. (2008). A role for the NAD-dependent deacetylase Sirt1 in the regulation of autophagy. *Proc. Natl. Acad. Sci. U. S. A.* 105, 3374–3379. doi:10.1073/pnas.0712145105

Lei, Y., Li, X., Yuan, F., Liu, L., Zhang, J., Yang, Y., et al. (2017). Toll-like receptor 4 ablation rescues against paraquat-triggered myocardial dysfunction: role of ER stress and apoptosis. *Environ. Toxicol.* 32, 656–668. doi:10.1002/tox.22267

Levine, B., and Klionsky, D. J. (2017). Autophagy wins the 2016 Nobel Prize in Physiology or Medicine: breakthroughs in baker's yeast fuel advances in biomedical research. *Proc. Natl. Acad. Sci. U. S. A.* 114, 201–205. doi:10.1073/pnas.1619876114

Li, D., Si, J., Guo, Y., Liu, B., Chen, X., Qi, K., et al. (2023). Danggui-Buxue decoction alleviated vascular senescence in mice exposed to chronic intermittent hypoxia through activating the Nrf2/HO-1 pathway. *Pharm. Biol.* 61, 1041–1053. doi:10.1080/13880209. 2023.2230753

Li, G., Li, J., Shao, R., Zhao, J., and Chen, M. (2021b). FUNDC1: a promising mitophagy regulator at the mitochondria-associated membrane for cardiovascular diseases. *Front. Cell. Dev. Biol.* 9, 788634. doi:10.3389/fcell.2021.788634

Li, J., Wu, W., Xin, Y., Zhao, M., and Liu, X. (2018b). Inhibition of Nogo-B promotes cardiac hypertrophy via endoplasmic reticulum stress. *Biomed. Pharmacother.* 104, 193–203. doi:10.1016/j.biopha.2018.05.039

Li, J., Zhang, D., Brundel, B., and Wiersma, M. (2019). Imbalance of ER and mitochondria interactions: prelude to cardiac ageing and disease? *Cells* 8, 1617. doi:10.3390/cells8121617

Li, J., Zhang, D., Wiersma, M., and Brundel, B. (2018a). Role of autophagy in proteostasis: friend and foe in cardiac diseases. *Cells* 7, 279. doi:10.3390/cells7120279

Li, W. W., Wang, H. J., Tan, Y. Z., Wang, Y. L., Yu, S. N., and Li, Z. H. (2021a). Reducing lipofuscin accumulation and cardiomyocytic senescence of aging heart by enhancing autophagy. *Exp. Cell. Res.* 403, 112585. doi:10.1016/j.yexcr.2021.112585

Li, X., Xu, M., Pitzer, A. L., Xia, M., Boini, K. M., Li, P. L., et al. (2014). Control of autophagy maturation by acid sphingomyelinase in mouse coronary arterial smooth muscle cells: protective role in atherosclerosis. *J. Mol. Med. Berl.* 92, 473–485. doi:10. 1007/s00109-014-1120-y

Li, Z., and Nakatogawa, H. (2022). Degradation of nuclear components via different autophagy pathways. *Trends Cell. Biol.* 32, 574–584. doi:10.1016/j.tcb.2021.12.008

Liao, X., Sluimer, J. C., Wang, Y., Subramanian, M., Brown, K., Pattison, J. S., et al. (2012). Macrophage autophagy plays a protective role in advanced atherosclerosis. *Cell. Metab.* 15, 545–553. doi:10.1016/j.cmet.2012.01.022

Liberale, L., Badimon, L., Montecucco, F., Lüscher, T. F., Libby, P., and Camici, G. G. (2022). Inflammation, aging, and cardiovascular disease: JACC review topic of the week. *J. Am. Coll. Cardiol.* 79, 837–847. doi:10.1016/j.jacc.2021.12.017

Lim, D., Tapella, L., Dematteis, G., Genazzani, A. A., Corazzari, M., and Verkhratsky, A. (2023). The endoplasmic reticulum stress and unfolded protein response in Alzheimer's disease: a calcium dyshomeostasis perspective. *Ageing Res. Rev.* 87, 101914. doi:10.1016/j.arr.2023.101914

Linton, P. J., Gurney, M., Sengstock, D., Mentzer, R. M., Jr., and Gottlieb, R. A. (2015). This old heart: cardiac aging and autophagy. *J. Mol. Cell. Cardiol.* 83, 44–54. doi:10. 1016/j.yjmcc.2014.12.017

Liu, S., Yao, S., Yang, H., Liu, S., and Wang, Y. (2023). Autophagy: regulator of cell death. *Cell. Death Dis.* 14, 648. doi:10.1038/s41419-023-06154-8

López-Domènech, S., Abad-Jiménez, Z., Iannantuoni, F., de Marañón, A. M., Rovira-Llopis, S., Morillas, C., et al. (2019). Moderate weight loss attenuates chronic endoplasmic reticulum stress and mitochondrial dysfunction in human obesity. *Mol. Metab.* 19, 24–33. doi:10.1016/j.molmet.2018.10.005 López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., and Kroemer, G. (2013). The hallmarks of aging. *Cell*. 153, 1194–1217. doi:10.1016/j.cell.2013.05.039

López-Otín, C., Blasco, M. A., Partridge, L., Serrano, M., and Kroemer, G. (2023). Hallmarks of aging: an expanding universe. *Cell.* 186, 243–278. doi:10.1016/j.cell.2022. 11.001

Lu, L., Xiong, Y., Zhou, J., Wang, G., Mi, B., and Liu, G. (2022a). The therapeutic roles of cinnamaldehyde against cardiovascular diseases. *Oxid. Med. Cell. Longev.* 2022, 9177108. doi:10.1155/2022/9177108

Lu, Q. B., Ding, Y., Liu, Y., Wang, Z. C., Wu, Y. J., Niu, K. M., et al. (2022b). Metrnl ameliorates diabetic cardiomyopathy via inactivation of cGAS/STING signaling dependent on LKB1/AMPK/ULK1-mediated autophagy. *J. Adv. Res.* 51, 161–179. doi:10.1016/j.jare.2022.10.014

Lyu, X., Zhang, M., Li, G., Cai, Y., Li, G., and Qiao, Q. (2019). Interleukin-6 production mediated by the IRE1-XBP1 pathway confers radioresistance in human papillomavirus-negative oropharyngeal carcinoma. *Cancer Sci.* 110, 2471–2484. doi:10. 1111/cas.14094

Ma, W., Lu, Y., Jin, X., Lin, N., Zhang, L., and Song, Y. (2024). Targeting selective autophagy and beyond: from underlying mechanisms to potential therapies. *J. Adv. Res.* doi:10.1016/j.jare.2024.05.009

Ma, Y., Zhou, X., Gui, M., Yao, L., Li, J., Chen, X., et al. (2023). Mitophagy in hypertension-mediated organ damage. *Front. Cardiovasc Med.* 10, 1309863. doi:10. 3389/fcvm.2023.1309863

Madhur, M. S., Elijovich, F., Alexander, M. R., Pitzer, A., Ishimwe, J., Van Beusecum, J. P., et al. (2021). Hypertension: do inflammation and immunity hold the key to solving this epidemic? *Circ. Res.* 128, 908–933. doi:10.1161/circresaha.121.318052

Mahmoudi, M., Gorenne, I., Mercer, J., Figg, N., Littlewood, T., and Bennett, M. (2008). Statins use a novel Nijmegen breakage syndrome-1-dependent pathway to accelerate DNA repair in vascular smooth muscle cells. *Circ. Res.* 103, 717–725. doi:10. 1161/circresaha.108.182899

Makino, N., and Maeda, T. (2021). Calorie restriction delays cardiac senescence and improves cardiac function in obese diabetic rats. *Mol. Cell. Biochem.* 476, 221–229. doi:10.1007/s11010-020-03899-0

Malhotra, J. D., and Kaufman, R. J. (2007). Endoplasmic reticulum stress and oxidative stress: a vicious cycle or a double-edged sword? *Antioxid. Redox Signal* 9, 2277–2293. doi:10.1089/ars.2007.1782

Marian, A. J., Bhatnagar, A., Bolli, R., and Izpisua Belmonte, J. C. (2018). Introduction to cardiovascular aging compendium. *Circ. Res.* 123, 737–739. doi:10.1161/circresaha. 118.313940

Matsuzawa-Ishimoto, Y., Hwang, S., and Cadwell, K. (2018). Autophagy and inflammation. *Annu. Rev. Immunol.* 36, 73–101. doi:10.1146/annurev-immunol-042617-053253

Melia, T. J., Lystad, A. H., and Simonsen, A. (2020). Autophagosome biogenesis: from membrane growth to closure. J. Cell. Biol. 219, e202002085. doi:10.1083/jcb.202002085

Mennerich, D., Kellokumpu, S., and Kietzmann, T. (2019). Hypoxia and reactive oxygen species as modulators of endoplasmic reticulum and Golgi homeostasis. *Antioxid. Redox Signal* 30, 113–137. doi:10.1089/ars.2018.7523

Mijaljica, D., Prescott, M., and Devenish, R. J. (2012). A late form of nucleophagy in Saccharomyces cerevisiae. PLoS One 7, e40013. doi:10.1371/journal.pone.0040013

Mizushima, N. (2018). A brief history of autophagy from cell biology to physiology and disease. *Nat. Cell. Biol.* 20, 521–527. doi:10.1038/s41556-018-0092-5

Moellmann, J., Mann, P. A., Kappel, B. A., Kahles, F., Klinkhammer, B. M., Boor, P., et al. (2022). The sodium-glucose co-transporter-2 inhibitor ertugliflozin modifies the signature of cardiac substrate metabolism and reduces cardiac mTOR signalling, endoplasmic reticulum stress and apoptosis. *Diabetes Obes. Metab.* 24, 2263–2272. doi:10.1111/dom.14814

Moltedo, O., Remondelli, P., and Amodio, G. (2019). The mitochondria-endoplasmic reticulum contacts and their critical role in aging and age-associated diseases. *Front. Cell. Dev. Biol.* 7, 172. doi:10.3389/fcell.2019.00172

Montégut, L., Joseph, A., Chen, H., Abdellatif, M., Ruckenstuhl, C., Martins, I., et al. (2023). DBI/ACBP is a targetable autophagy checkpoint involved in aging and cardiovascular disease. *Autophagy* 19, 2166–2169. doi:10.1080/15548627.2022.2160565

Moradi, F., Imani, A. R., and Faghihi, M. (2019). Effects of regular exercise plus food restriction on left ventricular pathological remodeling in heart failure-induced rats. *Bratisl. Lek. Listy* 120, 243–248. doi:10.4149/bll\_2019\_044

Morishita, H., and Mizushima, N. (2019). Diverse cellular roles of autophagy. Annu. Rev. Cell. Dev. Biol. 35, 453–475. doi:10.1146/annurev-cellbio-100818-125300

Mozaffarian, D., Benjamin, E. J., Go, A. S., Arnett, D. K., Blaha, M. J., Cushman, M., et al. (2015). Heart disease and stroke statistics--2015 update: a report from the American Heart Association. *Circulation* 131, e29–e322. doi:10.1161/cir.000000000000152

Muñoz, J. P., Ivanova, S., Sánchez-Wandelmer, J., Martínez-Cristóbal, P., Noguera, E., Sancho, A., et al. (2013). Mfn2 modulates the UPR and mitochondrial function via repression of PERK. *Embo J.* 32, 2348–2361. doi:10.1038/emboj.2013.168

Muñoz-Gámez, J. A., Rodríguez-Vargas, J. M., Quiles-Pérez, R., Aguilar-Quesada, R., Martín-Oliva, D., de Murcia, G., et al. (2009). PARP-1 is involved in autophagy induced by DNA damage. *Autophagy* 5, 61–74. doi:10.4161/auto.5.1.7272 Murley, A., and Nunnari, J. (2016). The emerging network of mitochondria-organelle contacts. *Mol. Cell.* 61, 648–653. doi:10.1016/j.molcel.2016.01.031

Myoishi, M., Hao, H., Minamino, T., Watanabe, K., Nishihira, K., Hatakeyama, K., et al. (2007). Increased endoplasmic reticulum stress in atherosclerotic plaques associated with acute coronary syndrome. *Circulation* 116, 1226–1233. doi:10.1161/circulationaha.106.682054

Naiel, S., Carlisle, R. E., Lu, C., Tat, V., and Dickhout, J. G. (2019). Endoplasmic reticulum stress inhibition blunts the development of essential hypertension in the spontaneously hypertensive rat. *Am. J. Physiol. Heart Circ. Physiol.* 316, H1214-H1223–h1223. doi:10.1152/ajpheart.00523.2018

Neff, F., Flores-Dominguez, D., Ryan, D. P., Horsch, M., Schröder, S., Adler, T., et al. (2013). Rapamycin extends murine lifespan but has limited effects on aging. *J. Clin. Investig.* 123, 3272–3291. doi:10.1172/jci67674

Nie, S., Lu, J., Wang, L., and Gao, M. (2020). Pro-inflammatory role of cell-free mitochondrial DNA in cardiovascular diseases. *IUBMB Life* 72, 1879–1890. doi:10. 1002/iub.2339

Obas, V., and Vasan, R. S. (2018). The aging heart. Clin. Sci. (Lond) 132, 1367–1382. doi:10.1042/cs20171156

Oduro, P. K., Zheng, X., Wei, J., Yang, Y., Wang, Y., Zhang, H., et al. (2022). The cGAS-STING signaling in cardiovascular and metabolic diseases: future novel target option for pharmacotherapy. *Acta Pharm. Sin. B* 12, 50–75. doi:10.1016/j.apsb.2021. 05.011

Oesterle, A., Laufs, U., and Liao, J. K. (2017). Pleiotropic effects of statins on the cardiovascular system. Circ. Res. 120, 229-243. doi:10.1161/circresaha.116.308537

Ortega, A., Roselló-Lletí, E., Tarazón, E., Molina-Navarro, M. M., Martínez-Dolz, L., González-Juanatey, J. R., et al. (2014). Endoplasmic reticulum stress induces different molecular structural alterations in human dilated and ischemic cardiomyopathy. *PLoS One* 9, e107635. doi:10.1371/journal.pone.0107635

Ozcan, U., Ozcan, L., Yilmaz, E., Düvel, K., Sahin, M., Manning, B. D., et al. (2008). Loss of the tuberous sclerosis complex tumor suppressors triggers the unfolded protein response to regulate insulin signaling and apoptosis. *Mol. Cell.* 29, 541–551. doi:10. 1016/j.molcel.2007.12.023

Pan, X., Wu, B., Fan, X., Xu, G., Ou, C., and Chen, M. (2021). YAP accelerates vascular senescence via blocking autophagic flux and activating mTOR. *J. Cell. Mol. Med.* 25, 170–183. doi:10.1111/jcmm.15902

Panel, M., Ghaleh, B., and Morin, D. (2018). Mitochondria and aging: a role for the mitochondrial transition pore? *Aging Cell*. 17, e12793. doi:10.1111/acel.12793

Panganiban, R. A., Mungunsukh, O., and Day, R. M. (2013). X-irradiation induces ER stress, apoptosis, and senescence in pulmonary artery endothelial cells. *Int. J. Radiat. Biol.* 89, 656–667. doi:10.3109/09553002.2012.711502

Paz Gavilán, M., Vela, J., Castaño, A., Ramos, B., del Río, J. C., Vitorica, J., et al. (2006). Cellular environment facilitates protein accumulation in aged rat hippocampus. *Neurobiol. Aging* 27, 973–982. doi:10.1016/j.neurobiolaging.2005.05.010

Pickles, S., Vigié, P., and Youle, R. J. (2018). Mitophagy and quality control mechanisms in mitochondrial maintenance. *Curr. Biol.* 28, R170-R185-r185. doi:10. 1016/j.cub.2018.01.004

Pintado, C., Macías, S., Domínguez-Martín, H., Castaño, A., and Ruano, D. (2017). Neuroinflammation alters cellular proteostasis by producing endoplasmic reticulum stress, autophagy activation and disrupting ERAD activation. *Sci. Rep.* 7, 8100. doi:10. 1038/s41598-017-08722-3

Pires Da Silva, J., Monceaux, K., Guilbert, A., Gressette, M., Piquereau, J., Novotova, M., et al. (2020). SIRT1 protects the heart from ER stress-induced injury by promoting eEF2K/eEF2-Dependent autophagy. *Cells* 9, 426. doi:10.3390/ cells9020426

Prince, M. J., Wu, F., Guo, Y., Gutierrez Robledo, L. M., O'Donnell, M., Sullivan, R., et al. (2015). The burden of disease in older people and implications for health policy and practice. *Lancet* 385, 549–562. doi:10.1016/s0140-6736(14)61347-7

Prola, A., Pires Da Silva, J., Guilbert, A., Lecru, L., Piquereau, J., Ribeiro, M., et al. (2017). SIRT1 protects the heart from ER stress-induced cell death through eIF2a deacetylation. *Cell. Death Differ.* 24, 343–356. doi:10.1038/cdd.2016.138

Qi, Z., and Chen, L. (2019). Endoplasmic reticulum stress and autophagy. Adv. Exp. Med. Biol. 1206, 167-177. doi:10.1007/978-981-15-0602-4\_8

Qiao, L., Ma, J., Zhang, Z., Sui, W., Zhai, C., Xu, D., et al. (2021). Deficient chaperonemediated autophagy promotes inflammation and atherosclerosis. *Circ. Res.* 129, 1141–1157. doi:10.1161/circresaha.121.318908

Quan, Y., Xin, Y., Tian, G., Zhou, J., and Liu, X. (2020). Mitochondrial ROSmodulated mtDNA: a potential target for cardiac aging. *Oxid. Med. Cell. Longev.* 2020, 9423593. doi:10.1155/2020/9423593

Rashid, H. O., Yadav, R. K., Kim, H. R., and Chae, H. J. (2015). ER stress: autophagy induction, inhibition and selection. *Autophagy* 11, 1956–1977. doi:10.1080/15548627. 2015.1091141

Reddy, P. H., and Oliver, D. M. (2019). Amyloid beta and phosphorylated tauinduced defective autophagy and mitophagy in alzheimer's disease. *Cells* 8, 488. doi:10. 3390/cells8050488 Robert, T., Vanoli, F., Chiolo, I., Shubassi, G., Bernstein, K. A., Rothstein, R., et al. (2011). HDACs link the DNA damage response, processing of double-strand breaks and autophagy. *Nature* 471, 74–79. doi:10.1038/nature09803

Ron, D., and Walter, P. (2007). Signal integration in the endoplasmic reticulum unfolded protein response. *Nat. Rev. Mol. Cell. Biol.* 8, 519–529. doi:10.1038/nrm2199

Ruparelia, N., Chai, J. T., Fisher, E. A., and Choudhury, R. P. (2017). Inflammatory processes in cardiovascular disease: a route to targeted therapies. *Nat. Rev. Cardiol.* 14, 133–144. doi:10.1038/nrcardio.2016.185

Sabath, N., Levy-Adam, F., Younis, A., Rozales, K., Meller, A., Hadar, S., et al. (2020). Cellular proteostasis decline in human senescence. *Proc. Natl. Acad. Sci. U. S. A.* 117, 31902–31913. doi:10.1073/pnas.2018138117

Safar, M. E. (2010). Arterial aging--hemodynamic changes and therapeutic options. *Nat. Rev. Cardiol.* 7, 442–449. doi:10.1038/nrcardio.2010.96

Salminen, A., and Kaarniranta, K. (2010). ER stress and hormetic regulation of the aging process. *Ageing Res. Rev.* 9, 211–217. doi:10.1016/j.arr.2010.04.003

Salminen, A., Kaarniranta, K., and Kauppinen, A. (2020). ER stress activates immunosuppressive network: implications for aging and Alzheimer's disease. J. Mol. Med. Berl. 98, 633–650. doi:10.1007/s00109-020-01904-z

Saxton, R. A., and Sabatini, D. M. (2017). mTOR signaling in growth, metabolism, and disease. Cell. 168, 960–976. doi:10.1016/j.cell.2017.02.004

Schiattarella, G. G., Altamirano, F., Kim, S. Y., Tong, D., Ferdous, A., Piristine, H., et al. (2021). Xbp1s-FoxO1 axis governs lipid accumulation and contractile performance in heart failure with preserved ejection fraction. *Nat. Commun.* 12, 1684. doi:10.1038/s41467-021-21931-9

Schneider, J. L., Suh, Y., and Cuervo, A. M. (2014). Deficient chaperone-mediated autophagy in liver leads to metabolic dysregulation. *Cell. Metab.* 20, 417–432. doi:10. 1016/j.cmet.2014.06.009

Schumacher, B., Pothof, J., Vijg, J., and Hoeijmakers, J. H. J. (2021). The central role of DNA damage in the ageing process. *Nature* 592, 695–703. doi:10.1038/s41586-021-03307-7

Sciarretta, S., Maejima, Y., Zablocki, D., and Sadoshima, J. (2018). The role of autophagy in the heart. *Annu. Rev. Physiol.* 80, 1–26. doi:10.1146/annurev-physiol-021317-121427

Sciarretta, S., Zhai, P., Shao, D., Zablocki, D., Nagarajan, N., Terada, L. S., et al. (2013). Activation of NADPH oxidase 4 in the endoplasmic reticulum promotes cardiomyocyte autophagy and survival during energy stress through the protein kinase RNA-activatedlike endoplasmic reticulum kinase/eukaryotic initiation factor 2α/activating transcription factor 4 pathway. *Circ. Res.* 113, 1253–1264. doi:10.1161/circresaha. 113.301787

Seals, D. R., Jablonski, K. L., and Donato, A. J. (2011). Aging and vascular endothelial function in humans. *Clin. Sci. (Lond)* 120, 357-375. doi:10.1042/cs20100476

Senft, D., and Ronai, Z. A. (2015). UPR, autophagy, and mitochondria crosstalk underlies the ER stress response. *Trends Biochem. Sci.* 40, 141–148. doi:10.1016/j.tibs. 2015.01.002

Shanahan, C. M., and Furmanik, M. (2017). Endoplasmic reticulum stress in arterial smooth muscle cells: a novel regulator of vascular disease. *Curr. Cardiol. Rev.* 13, 94–105. doi:10.2174/1573403x12666161014094738

Shen, C. H., Tung, S. Y., Huang, W. S., Lu, C. C., Lee, K. C., Hsieh, Y. Y., et al. (2014). Exploring the effects of tert-butylhydroperoxide induced liver injury using proteomic approach. *Toxicology* 316, 61–70. doi:10.1016/j.tox.2013.12.007

Shi, Y., Jiang, B., and Zhao, J. (2024). Induction mechanisms of autophagy and endoplasmic reticulum stress in intestinal ischemia-reperfusion injury, inflammatory bowel disease, and colorectal cancer. *Biomed. Pharmacother*. 170, 115984. doi:10.1016/j. biopha.2023.115984

Shin, G. C., Moon, S. U., Kang, H. S., Choi, H. S., Han, H. D., and Kim, K. H. (2019). PRKCSH contributes to tumorigenesis by selective boosting of IRE1 signaling pathway. *Nat. Commun.* 10, 3185. doi:10.1038/s41467-019-11019-w

Shin, J. I., Sang, Y., Chang, A. R., Dunning, S. C., Coresh, J., Inker, L. A., et al. (2020). The FDA metformin label change and racial and sex disparities in metformin prescription among patients with CKD. J. Am. Soc. Nephrol. 31, 1847–1858. doi:10. 1681/asn.2019101119

Shirakabe, A., Ikeda, Y., Sciarretta, S., Zablocki, D. K., and Sadoshima, J. (2016). Aging and autophagy in the heart. *Circ. Res.* 118, 1563–1576. doi:10.1161/circresaha.116. 307474

Song, P., Zhao, Q., and Zou, M. H. (2020). Targeting senescent cells to attenuate cardiovascular disease progression. *Ageing Res. Rev.* 60, 101072. doi:10.1016/j.arr.2020. 101072

Song, X. J., Yang, C. Y., Liu, B., Wei, Q., Korkor, M. T., Liu, J. Y., et al. (2011). Atorvastatin inhibits myocardial cell apoptosis in a rat model with post-myocardial infarction heart failure by downregulating ER stress response. *Int. J. Med. Sci.* 8, 564–572. doi:10.7150/ijms.8.564

Song, Z. C., Chen, L., Zhang, D., Zhang, S. Y., and Lin, X. (2018). Rosuvastatin protects acute myocardial infarction rats through autophagy regulation via AMPK

signaling. Zhonghua Yi Xue Za Zhi 98, 3536–3541. doi:10.3760/cma.j.issn.0376-2491. 2018.43.015

Spencer, B. G., and Finnie, J. W. (2020). The role of endoplasmic reticulum stress in cell survival and death. J. Comp. Pathol. 181, 86–91. doi:10.1016/j.jcpa.2020.10.006

Spitler, K. M., Matsumoto, T., and Webb, R. C. (2013). Suppression of endoplasmic reticulum stress improves endothelium-dependent contractile responses in aorta of the spontaneously hypertensive rat. *Am. J. Physiol. Heart Circ. Physiol.* 305, H344–H353. doi:10.1152/ajpheart.00952.2012

Steiger, D., Yokota, T., Li, J., Ren, S., Minamisawa, S., and Wang, Y. (2018). The serine/threonine-protein kinase/endoribonuclease IRE1a protects the heart against pressure overload-induced heart failure. *J. Biol. Chem.* 293, 9652–9661. doi:10.1074/ jbc.RA118.003448

Tai, H., Wang, Z., Gong, H., Han, X., Zhou, J., Wang, X., et al. (2017). Autophagy impairment with lysosomal and mitochondrial dysfunction is an important characteristic of oxidative stress-induced senescence. *Autophagy* 13, 99–113. doi:10. 1080/15548627.2016.1247143

Taneike, M., Yamaguchi, O., Nakai, A., Hikoso, S., Takeda, T., Mizote, I., et al. (2010). Inhibition of autophagy in the heart induces age-related cardiomyopathy. *Autophagy* 6, 600–606. doi:10.4161/auto.6.5.11947

Thompson, J., Maceyka, M., and Chen, Q. (2020). Targeting ER stress and calpain activation to reverse age-dependent mitochondrial damage in the heart. *Mech. Ageing Dev.* 192, 111380. doi:10.1016/j.mad.2020.111380

Thorp, E., Li, G., Seimon, T. A., Kuriakose, G., Ron, D., and Tabas, I. (2009). Reduced apoptosis and plaque necrosis in advanced atherosclerotic lesions of Apoe-/- and Ldlr-/-mice lacking CHOP. *Cell. Metab.* 9, 474–481. doi:10.1016/j.cmet.2009.03.003

Thrasher, P. R., Scofield, S. L. C., Dalal, S., Crawford, C. C., Singh, M., and Singh, K. (2018). Ataxia telangiectasia mutated kinase deficiency impairs the autophagic response early during myocardial infarction. *Am. J. Physiol. Heart Circ. Physiol.* 315, H48-H57-h57. doi:10.1152/ajpheart.00042.2018

Tian, J., Popal, M. S., Liu, Y., Gao, R., Lyu, S., Chen, K., et al. (2019). Ginkgo biloba leaf extract attenuates atherosclerosis in streptozotocin-induced diabetic ApoE-/- mice by inhibiting endoplasmic reticulum stress via restoration of autophagy through the mTOR signaling pathway. Oxid. Med. Cell. Longev. 2019, 8134678. doi:10.1155/2019/8134678

Torp, M. K., Vaage, J., and Stensløkken, K. O. (2023). Mitochondria-derived damageassociated molecular patterns and inflammation in the ischemic-reperfused heart. *Acta Physiol.* (*Oxf*) 237, e13920. doi:10.1111/apha.13920

Tu, B. P., and Weissman, J. S. (2004). Oxidative protein folding in eukaryotes: mechanisms and consequences. J. Cell. Biol. 164, 341–346. doi:10.1083/jcb.200311055

Tyrrell, D. J., and Goldstein, D. R. (2021). Ageing and atherosclerosis: vascular intrinsic and extrinsic factors and potential role of IL-6. *Nat. Rev. Cardiol.* 18, 58–68. doi:10.1038/s41569-020-0431-7

Uddin, M. S., Yu, W. S., and Lim, L. W. (2021). Exploring ER stress response in cellular aging and neuroinflammation in Alzheimer's disease. *Ageing Res. Rev.* 70, 101417. doi:10.1016/j.arr.2021.101417

Urra, H., Pihán, P., and Hetz, C. (2020). The UPRosome - decoding novel biological outputs of IRE1 $\alpha$  function. J. Cell. Sci. 133, jcs218107. doi:10.1242/jcs. 218107

van der Linden, J., Stefens, S. J. M., Heredia-Genestar, J. M., Ridwan, Y., Brandt, R. M. C., van Vliet, N., et al. (2024). Ercc1 DNA repair deficiency results in vascular aging characterized by VSMC phenotype switching, ECM remodeling, and an increased stress response. *Aging Cell.* 23, e14126. doi:10.1111/acel.14126

Vanhoutte, D., Schips, T. G., Vo, A., Grimes, K. M., Baldwin, T. A., Brody, M. J., et al. (2021). Thbs1 induces lethal cardiac atrophy through PERK-ATF4 regulated autophagy. *Nat. Commun.* 12, 3928. doi:10.1038/s41467-021-24215-4

Vattem, K. M., and Wek, R. C. (2004). Reinitiation involving upstream ORFs regulates ATF4 mRNA translation in mammalian cells. *Proc. Natl. Acad. Sci. U. S. A.* 101, 11269–11274. doi:10.1073/pnas.0400541101

Vega-Martín, E., González-Blázquez, R., Manzano-Lista, F. J., Martín-Ramos, M., García-Prieto, C. F., Viana, M., et al. (2020). Impact of caloric restriction on AMPK and endoplasmic reticulum stress in peripheral tissues and circulating peripheral blood mononuclear cells from Zucker rats. *J. Nutr. Biochem.* 78, 108342. doi:10.1016/j.jnutbio.2020.108342

Verdin, E. (2015). NAD\* in aging, metabolism, and neurodegeneration. *Science* 350, 1208–1213. doi:10.1126/science.aac4854

Volonte, D., Liu, Z., Musille, P. M., Stoppani, E., Wakabayashi, N., Di, Y. P., et al. (2013). Inhibition of nuclear factor-erythroid 2-related factor (Nrf2) by caveolin-1 promotes stress-induced premature senescence. *Mol. Biol. Cell.* 24, 1852–1862. doi:10. 1091/mbc.E12-09-0666

Wang, J., Lu, L., Chen, S., Xie, J., Lu, S., Zhou, Y., et al. (2020). PERK overexpressionmediated Nrf2/HO-1 pathway alleviates hypoxia/reoxygenation-induced injury in neonatal murine cardiomyocytes via improving endoplasmic reticulum stress. *Biomed. Res. Int.* 2020, 6458060. doi:10.1155/2020/6458060

Wang, J., Uryga, A. K., Reinhold, J., Figg, N., Baker, L., Finigan, A., et al. (2015a). Vascular smooth muscle cell senescence promotes atherosclerosis and features of plaque vulnerability. *Circulation* 132, 1909–1919. doi:10.1161/circulationaha.115.016457

Wang, M., and Kaufman, R. J. (2016). Protein misfolding in the endoplasmic reticulum as a conduit to human disease. *Nature* 529, 326-335. doi:10.1038/ nature17041

Wang, M., Kim, S. H., Monticone, R. E., and Lakatta, E. G. (2015b). Matrix metalloproteinases promote arterial remodeling in aging, hypertension, and atherosclerosis. *Hypertension* 65, 698–703. doi:10.1161/hypertensionaha.114. 03618

Wang, S., Chen, X., Nair, S., Sun, D., Wang, X., and Ren, J. (2017). Deletion of protein tyrosine phosphatase 1B obliterates endoplasmic reticulum stress-induced myocardial dysfunction through regulation of autophagy. *Biochim. Biophys. Acta Mol. Basis Dis.* 1863, 3060–3074. doi:10.1016/j.bbadis.2017.09.015

Wang, S., and Kaufman, R. J. (2012). The impact of the unfolded protein response on human disease. J. Cell. Biol. 197, 857–867. doi:10.1083/jcb.201110131

Wang, Y., Zhang, N., Zhang, L., Li, R., Fu, W., Ma, K., et al. (2016). Autophagy regulates chromatin ubiquitination in DNA damage response through elimination of SQSTM1/p62. *Mol. Cell.* 63, 34–48. doi:10.1016/j.molcel.2016.05.027

Wang, Y. L., Zhou, X., Li, D. L., and Ye, J. M. (2022). Role of the mTOR-autophagy-ER stress pathway in high fructose-induced metabolic-associated fatty liver disease. *Acta Pharmacol. Sin.* 43, 10–14. doi:10.1038/s41401-021-00629-0

Weber, C., Habenicht, A. J. R., and von Hundelshausen, P. (2023). Novel mechanisms and therapeutic targets in atherosclerosis: inflammation and beyond. *Eur. Heart J.* 44, 2672–2681. doi:10.1093/eurheartj/ehad304

Wei, X., Wu, Y. E., Wang, W., Zhang, S., Liu, D., and Liu, H. (2021). Decreased dynamin-related protein 1-related mitophagy induces myocardial apoptosis in the aging heart. *Acta Biochim. Biophys. Sin. (Shanghai)* 53, 1354–1366. doi:10.1093/abbs/gmab112

Wiersma, M., van Marion, D. M. S., Wüst, R. C. I., Houtkooper, R. H., Zhang, D., Groot, N. M. S. d., et al. (2019). Mitochondrial dysfunction underlies cardiomyocyte remodeling in experimental and clinical atrial fibrillation. *Cells* 8, 1202. doi:10.3390/ cells8101202

Wiese, W., Siwecka, N., Wawrzynkiewicz, A., Rozpędek-Kamińska, W., Kucharska, E., and Majsterek, I. (2022). IRE1a inhibitors as a promising therapeutic strategy in blood malignancies. *Cancers (Basel)* 14, 2526. doi:10.3390/cancers14102526

Willy, J. A., Young, S. K., Stevens, J. L., Masuoka, H. C., and Wek, R. C. (2015). CHOP links endoplasmic reticulum stress to NF-kB activation in the pathogenesis of nonalcoholic steatohepatitis. *Mol. Biol. Cell.* 26, 2190–2204. doi:10.1091/mbc.E15-01-0036

Wiseman, R. L., Mesgarzadeh, J. S., and Hendershot, L. M. (2022). Reshaping endoplasmic reticulum quality control through the unfolded protein response. *Mol. Cell.* 82, 1477–1491. doi:10.1016/j.molcel.2022.03.025

Wu, Z., Qu, J., Zhang, W., and Liu, G. H. (2024). Stress, epigenetics, and aging: unraveling the intricate crosstalk. *Mol. Cell.* 84, 34–54. doi:10.1016/j.molcel.2023.10.006

Yan, C., Xu, Z., and Huang, W. (2021). Cellular senescence affects cardiac regeneration and repair in ischemic heart disease. *Aging Dis.* 12, 552–569. doi:10. 14336/ad.2020.0811

Yang, J., Ma, X., Niu, D., Sun, Y., Chai, X., Deng, Y., et al. (2023). PCSK9 inhibitors suppress oxidative stress and inflammation in atherosclerotic development by promoting macrophage autophagy. *Am. J. Transl. Res.* 15, 5129–5144.

Yang, M. Y., Wang, Y. B., Han, B., Yang, B., Qiang, Y. W., Zhang, Y., et al. (2018). Activation of aldehyde dehydrogenase 2 slows down the progression of atherosclerosis via attenuation of ER stress and apoptosis in smooth muscle cells. *Acta Pharmacol. Sin.* 39, 48–58. doi:10.1038/aps.2017.81

Yang, Y., Li, J., Han, T. L., Zhou, X., Qi, H., Baker, P. N., et al. (2020). Endoplasmic reticulum stress may activate NLRP3 inflammasomes via TXNIP in preeclampsia. *Cell. Tissue Res.* 379, 589–599. doi:10.1007/s00441-019-03104-9

Yang, Y., Lu, D., Wang, M., Liu, G., Feng, Y., Ren, Y., et al. (2024). Endoplasmic reticulum stress and the unfolded protein response: emerging regulators in progression of traumatic brain injury. *Cell. Death Dis.* 15, 156. doi:10.1038/s41419-024-06515-x

Yao, Y., Lu, Q., Hu, Z., Yu, Y., Chen, Q., and Wang, Q. K. (2017). A non-canonical pathway regulates ER stress signaling and blocks ER stress-induced apoptosis and heart failure. *Nat. Commun.* 8, 133. doi:10.1038/s41467-017-00171-w

Ye, J., Rawson, R. B., Komuro, R., Chen, X., Davé, U. P., Prywes, R., et al. (2000). ER stress induces cleavage of membrane-bound ATF6 by the same proteases that process SREBPs. *Mol. Cell.* 6, 1355–1364. doi:10.1016/s1097-2765(00)00133-7

Ye, S., Zhou, X., Chen, P., and Lin, J. F. (2021). Folic acid attenuates remodeling and dysfunction in the aging heart through the ER stress pathway. *Life Sci.* 264, 118718. doi:10.1016/j.lfs.2020.118718

Yin, Y., Sun, G., Li, E., Kiselyov, K., and Sun, D. (2017). ER stress and impaired autophagy flux in neuronal degeneration and brain injury. *Ageing Res. Rev.* 34, 3–14. doi:10.1016/j.arr.2016.08.008

You, G., Long, X., Song, F., Huang, J., Tian, M., Xiao, Y., et al. (2020). Metformin activates the AMPK-mTOR pathway by modulating lncRNA TUG1 to induce autophagy and inhibit atherosclerosis. *Drug Des. Devel Ther.* 14, 457–468. doi:10. 2147/dddt.S233932

Youle, R. J. (2019). Mitochondria-Striking a balance between host and endosymbiont. *Science* 365, eaaw9855. doi:10.1126/science.aaw9855

Youm, Y. H., Grant, R. W., McCabe, L. R., Albarado, D. C., Nguyen, K. Y., Ravussin, A., et al. (2013). Canonical Nlrp3 inflammasome links systemic low-grade inflammation to functional decline in aging. *Cell. Metab.* 18, 519–532. doi:10.1016/j.cmet.2013.09.010

Young, A. R., Narita, M., Ferreira, M., Kirschner, K., Sadaie, M., Darot, J. F. J., et al. (2009). Autophagy mediates the mitotic senescence transition. *Genes. Dev.* 23, 798–803. doi:10.1101/gad.519709

Zahid, M. D. K., Rogowski, M., Ponce, C., Choudhury, M., Moustaid-Moussa, N., and Rahman, S. M. (2020). CCAAT/enhancer-binding protein beta (C/EBP $\beta$ ) knockdown reduces inflammation, ER stress, and apoptosis, and promotes autophagy in oxLDL-treated RAW264.7 macrophage cells. *Mol. Cell. Biochem.* 463, 211–223. doi:10.1007/s11010-019-03642-4

Zhan, W., Chen, L., Liu, H., Long, C., Liu, J., Ding, S., et al. (2022). Pcsk6 deficiency promotes cardiomyocyte senescence by modulating ddit3-mediated ER stress. *Genes.* (*Basel*) 13, 711. doi:10.3390/genes13040711

Zhang, D., Hu, X., Li, J., Liu, J., Baks-Te Bulte, L., Wiersma, M., et al. (2019). DNA damage-induced PARP1 activation confers cardiomyocyte dysfunction through NAD(+) depletion in experimental atrial fibrillation. *Nat. Commun.* 10, 1307. doi:10.1038/s41467-019-09014-2

Zhang, K., and Kaufman, R. J. (2008). From endoplasmic-reticulum stress to the inflammatory response. *Nature* 454, 455-462. doi:10.1038/nature07203

Zhang, W., Zhu, T., Chen, L., Luo, W., and Chao, J. (2020a). MCP-1 mediates ischemia-reperfusion-induced cardiomyocyte apoptosis via MCPIP1 and CaSR.

Am. J. Physiol. Heart Circ. Physiol. 318, H59-H71-h71. doi:10.1152/ajpheart. 00308.2019

Zhang, Y., Chen, W., and Wang, Y. (2020b). STING is an essential regulator of heart inflammation and fibrosis in mice with pathological cardiac hypertrophy via endoplasmic reticulum (ER) stress. *Biomed. Pharmacother.* 125, 110022. doi:10. 1016/j.biopha.2020.110022

Zhao, D., Sun, Y., Tan, Y., Zhang, Z., Hou, Z., Gao, C., et al. (2018). Short-duration swimming exercise after myocardial infarction attenuates cardiac dysfunction and regulates mitochondrial quality control in aged mice. *Oxid. Med. Cell. Longev.* 2018, 4079041. doi:10.1155/2018/4079041

Zhao, Y., Simon, M., Seluanov, A., and Gorbunova, V. (2023). DNA damage and repair in age-related inflammation. *Nat. Rev. Immunol.* 23, 75–89. doi:10.1038/s41577-022-00751-y

Zhou, J., Freeman, T. A., Ahmad, F., Shang, X., Mangano, E., Gao, E., et al. (2013). GSK-3 $\alpha$  is a central regulator of age-related pathologies in mice. *J. Clin. Investig.* 123, 1821–1832. doi:10.1172/jci64398

Zhu, H., Tannous, P., Johnstone, J. L., Kong, Y., Shelton, J. M., Richardson, J. A., et al. (2007). Cardiac autophagy is a maladaptive response to hemodynamic stress. *J. Clin. Investig.* 117, 1782–1793. doi:10.1172/jci27523

Zhu, L., Xie, Z. Y., Jiang, Z. L., Wang, X. H., Shi, H., Chen, L., et al. (2022). Unfolded protein response alleviates acid-induced premature senescence by promoting autophagy in nucleus pulposus cells. *Cell. Biol. Int.* 46, 568–578. doi:10.1002/cbin.11751

Zitvogel, L., Kepp, O., and Kroemer, G. (2010). Decoding cell death signals in inflammation and immunity. Cell. 140, 798-804. doi:10.1016/j.cell.2010.02.015