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# [An insight into pharmacokinetics](https://www.frontiersin.org/articles/10.3389/fphar.2024.1396994/full) [and dose optimization of](https://www.frontiersin.org/articles/10.3389/fphar.2024.1396994/full) [antimicrobials agents in elderly](https://www.frontiersin.org/articles/10.3389/fphar.2024.1396994/full) [patients](https://www.frontiersin.org/articles/10.3389/fphar.2024.1396994/full)

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The global elderly population is on the rise, and infections tend to have a higher mortality rate among older individuals. Aging is associated with the progressive impairment of multi-organ function, which can impact the pharmacokinetics of antimicrobials, potentially leading to the failure of anti-infective therapy. With the increasing life expectancy, a significant growth in the elderly demographic, and the escalating costs of healthcare, gaining a thorough understanding of pharmacokinetic changes in the elderly holds crucial clinical significance. This review compiles findings from published studies, offering a comprehensive overview of the pharmacokinetics of various antimicrobials in both adults and the elderly. Furthermore, it delves into advancements in pharmacokinetic methods specific to the elderly population.

#### KEYWORDS

elderly, pharmacokinetics, antimicrobial agents, drug resistance, anti-infective therapy

## 1 Introduction

The aging of the population, specifically those aged 65 and above, is an undeniable contemporary trend. Thanks to improvements in global living standards and enhanced medical care, the lifespan of the elderly is steadily increasing, resulting in a substantial growth in their numbers. According to the World Social Report 2023 [\(Leaving No One](#page-12-0) [Behind In An Ageing World, 2023\)](#page-12-0), the global population of elderly individuals has more than tripled, rising from approximately 260 million in 1980 to 761 million in 2021. Projections indicate that by 2030, this number is expected to surpass 1 billion and eventually exceed 1.6 billion by 2050. As we approach the end of the 21st century, the world could be home to nearly 2.5 billion elderly people.

Infection stands as a pivotal contributor to escalated disease progression and mortality among the elderly. Typically, the frequency and severity of infections escalate with advancing age ([Bouza et al., 2020\)](#page-11-0). Statistics from 2017 reveal that globally, the proportion of sepsis-related deaths, stemming from infectious diseases, is roughly 10% higher among individuals aged 65–89 than those aged 30–50 [\(Rudd et al., 2020\)](#page-13-0). Moreover, mortality data concerning antimicrobial resistance (AMR) across countries in the WHO Region of the Americas for 2019 illustrate a steep surge in deaths attributed to AMR starting around age 65 [\(Antimicrobial Resistance Collaborators, 2023\)](#page-11-1). Research underscores that within the initial 2 years of the COVID-19 pandemic, over 80% of global COVID-19 fatalities occur among individuals aged 60 and above [\(World Health](#page-14-0) [Statistics, 2023](#page-14-0)). Furthermore, both the prevalence and fatality rates of lung infections are markedly elevated among older adults compared to their younger counterparts, with mortality from community-acquired pneumonia among older hospitalized patients more than doubling from 6% in those aged 65% to 15% in those over 90 ([González-Castillo et al., 2014;](#page-12-1) [Fernández-Sabé](#page-12-2) [et al., 2003\)](#page-12-2). Despite these glaring statistics, a significant treatment gap persists between the elderly and adults, as most drugs are developed with an adult population focus. In clinical practice, antimicrobials deemed effective and safe in adults may exhibit unexpectedly lower efficacy or provoke adverse reactions in elderly patients at standard recommended dosages. Such scenarios heighten the risk of inappropriate drug dosing among the elderly ([Soraci et al., 2023\)](#page-13-1).

As the elderly population continues to grow, the issue of drug safety and effectiveness in this demographic becomes increasingly prominent. The functions of many organs and systems undergo gradual degradation due to aging ([Klotz, 2009\)](#page-12-3). The aging nervous system, for instance, leads to an increased permeability of the bloodbrain barrier, facilitating a greater influx of drugs from the plasma into the central system. Consequently, this amplifies drug distribution within the central system, elevating the incidence of adverse reactions in that area [\(Farrall and Wardlaw, 2009\)](#page-12-4). Moreover, the significant degradation of kidney function in the aging body is noteworthy. This is characterized by reduced glomerular number, glomerular filtration rate, and renal blood flow. These factors impact the metabolism and excretion of drugs primarily processed by the kidneys [\(Thomson, 1995\)](#page-14-1). Consequently, antimicrobial agents eliminated mainly by kidney tend to accumulate in the body, leading to adverse reactions. Additionally, elderly patients are often burdened with a variety of underlying conditions such as hypertension and diabetes. Consequently, the elderly constitute a widely medicated population, with each individual typically taking an average of 2–5 types of drugs daily [\(Kennerfalk et al., 2002\)](#page-12-5). The combination of multiple drugs heightens the likelihood of interactions, presenting additional challenges for doctors when prescribing appropriate dosages of antimicrobials for the elderly ([Gnjidic et al., 2018;](#page-12-6) [Khan and Roberts, 2018](#page-12-7)).

Hence, providing a comprehensive delineation of the pharmacokinetic (PK) parameters of antimicrobials in the elderly population holds immense value in ensuring the delivery of safe and effective anti-infective therapy for elderly patients. This review endeavors to consolidate the understanding of the pharmacokinetics of diverse antimicrobial agents and highlight the advancements in research methodologies pertaining to pharmacokinetics in the elderly population.

#### 2 ADME process in the elderly

The PK characteristics undergo significant alterations with age, particularly in the elderly population. These changes encompass a decline in renal filtration and excretion functions, gastrointestinal absorption function, and hepatic metabolic function ([Koren et al.,](#page-12-8) [2019\)](#page-12-8). As a result, the processes associated with antimicrobial agents, including absorption, distribution, metabolism, and excretion (ADME), experience certain modifications in old age. In general, these changes tend to culminate in an elevated accumulation of drugs within the body.

#### 2.1 Absorption

As age advances, the physiological characteristics of the gastrointestinal tract undergo gradual transformations. These changes predominantly involve a reduction in gastric acid secretion, gastrointestinal smooth muscle motor function, and gastrointestinal blood flow [\(Orr and Chen, 2002\)](#page-13-2). Numerous studies indicate that, whether in a fasting or fed state, the pH value of the stomach increases, and gastric emptying slows in the elderly due to diminished gastric acid secretion and weakened motor function of the gastrointestinal smooth muscle [\(Vinarov](#page-14-2) [et al., 2021;](#page-14-2) [Giarratano et al., 2018](#page-12-9)). For instance, a higher gastric pH and reduced gastrointestinal peristalsis can diminish the solubility of posaconazole oral suspension and shorten its residence time in the stomach, consequently leading to a decrease in its plasma concentration [\(Chen et al., 2020](#page-12-10)). However, for certain oral antimicrobial agents like ciprofloxacin tablets, despite a decreased rate of drug absorption due to increased gastric pH in the elderly, the total amount of drug absorption remains unaffected owing to the prolonged residence time of drugs in the gastrointestinal tract ([Lebel and Bergeron, 1987\)](#page-12-11). Furthermore, the weakened stability of gastrointestinal flora in the elderly accelerates the decline of gastrointestinal function, thereby slowing down drug absorption [\(Tiihonen et al., 2010\)](#page-14-3).

#### 2.2 Distribution

The PK distribution process undergoes substantial changes in the elderly. In comparison to younger individuals, the elderly experience a notable decrease of 10%–15% in body water content and an increase of 20%–38% in adipose tissue ([Beaufrère and Morio,](#page-11-2) [2000\)](#page-11-2). Consequently, this results in a significant reduction in the distribution volume of hydrophilic antibacterial drugs such as aminoglycosides and penicillin, leading to a marked increase in drug peak concentration. It's worth noting that in critically ill patients, the volume of distribution  $(V_d)$  of hydrophilic drugs may increase due to continuous infusion and excessive capillary permeability ([Udy et al., 2013](#page-14-4)). Conversely, the  $V_d$  of lipophilic drugs like tetracycline and amphotericin B increases, leading to a slower elimination ([Shi and Klotz, 2011](#page-13-3)). Furthermore, as age progresses, the plasma protein binding rate of drugs in the body diminishes due to a reduction in plasma protein mass and quantity. Consequently, there is an increased risk of higher concentrations of free drug, a phenomenon more pronounced in individuals over the age of 70 [\(Celestin and Musteata, 2021](#page-11-3)).

#### 2.3 Metabolism

The liver, a pivotal organ in the metabolism of most drugs, including antimicrobials, plays a crucial role in this process. In the elderly, there is a sharp decrease in the number of liver cells and local



#### <span id="page-2-0"></span>TABLE 1 Comparison of pharmacokinetic parameters of some antimicrobial agents in adults and the elderly.

#### Group Study type  $C_{\text{max}}$  $(mg·L^{-1})$  $T_{1/2}$  (h)  $V_{d}$  (L) CL  $(L \cdot h^{-1})$  $CL$  $(mL·min<sup>-1</sup>)$ AUC  $(h \cdot mg \cdot L^{-1})$ Elderly Patients  $(≥65)$ [\(Namkoong](#page-13-11) [et al., 2014\)](#page-13-11) PK STUDY 22.88 6.59 Doripenem (Dosage 500 mg) Adults (22–30) [\(Kim](#page-12-13) [et al., 2018\)](#page-12-13) PK **STUDY** 9.72 1.01 1.01 12.0 Elderly (≥65) [\(Harada](#page-12-14) [et al., 2013\)](#page-12-14) PK STUDY 20.0 1.20 31.6 Monobactams βlactams Aztreonam ([Meyers](#page-13-12) [et al., 1993\)](#page-13-12) (Dosage 2 g) Adults  $(18-30)$ PK **STUDY**  $160 \pm 47.64$  1.80  $\pm$  0.51 17.23  $\pm$ 5.67  $6.60 \pm 1.58$  294.42  $\pm$  64.08 Elderly (≥65) PK **STUDY**  $153.5 \pm 36.31$   $3.10 \pm 0.90$  17.19  $\pm$ 2.14  $4.32 \pm 2.28$   $469.01 \pm$ 144.03 Aminoglycoside antibiotics Amikacin ([Yaoguo](#page-14-5) [et al., 2001\)](#page-14-5) (Dosage 400 mg) Adults  $(27.8 \pm 5.0)$ PK **STUDY**  $43.6 \pm 6.4$  2.1  $\pm$  0.2 16.5  $\pm$  3.4 78.8  $\pm$  10.6 76.5  $\pm$  10.5 Elderly  $(69.8 \pm 5.4)$ PK **STUDY**  $40.7 \pm 6.3$   $3.1 \pm 0.4$   $18.4 \pm 2.7$   $46.5 \pm 10.7$   $99.9 \pm 20.3$ Tobramycin [\(Champoux et al.,](#page-11-5) [1996](#page-11-5)) (Dosage 80 mg) Adults  $(29.4 \pm 11.3)$ PK **STUDY**  $2.5 \pm 0.6$  2.73  $\pm$  1.17 23.6  $\pm$  4.8 6.9  $\pm$  3.12 12.33  $\pm$  3.75 Elderly  $(65.8 \pm 1.9)$ PK STUDY  $2.7 \pm 0.8$   $3.53 \pm 0.97$   $26.2 \pm 8.2$   $5.28 \pm 1.44$  16.33  $\pm 4.8$ Gentamicin Adults [\(Bos et al.,](#page-11-6) [2019](#page-11-6))  $(20–86)$ PPK 20 5.5 Elderly [\(Johnston et al.,](#page-12-15) [2014](#page-12-15))  $(65-96)$ PPK 14.9 2.96 Macrolide antibiotics Telithromycin [Perret et al. \(2002\)](#page-13-13) (Dosage 800 mg) Adults (18–40) PK **STUDY** 1.64 190.3 8.40 Elderly  $(65-85)$ PK STUDY 1.91 10.83 Dirithromycin [Varanese \(1993\)](#page-14-6) (Dosage 500 mg) Adults  $(18–50)$ PK **STUDY**  $0.36 \pm 0.18$  1.83  $\pm 1.09$ Elderly (≥65) PK **STUDY**  $0.85 \pm 0.30$  4.30  $\pm 3.10$ Clarithromycin [Chu et al. \(1992\)](#page-12-16) Adults (18–30) PK STUDY  $2.41 + 0.67$  4.9 28.56 + 6.72  $168 + 35$  18.87 + 5.55 Elderly (65–84) PK **STUDY**  $3.28 \pm 0.80$  7.7 18  $\pm 5.82$  84  $\pm 31$  30.76  $\pm 8.50$ Azithromycin [Coates et al. \(1991\)](#page-12-17) (Dosage 500 mg) Adults (22–39) PK STUDY 0.41 2.5 Elderly  $(67-80)$ PK STUDY  $0.38$  3.0 Tetracycline antibiotics Minocycline Adults Male [\(JB,](#page-12-18) [1983](#page-12-18)) (Dosage 200 mg) PK STUDY 3.5 15.7 115 3.36 ± 0.31 Debilitated Elderly Patients ±6(82 [\(Yamamoto](#page-14-7) [et al., 1999\)](#page-14-7) (Dosage 100 mg) PK **STUDY**  $7.22 \pm 3.53$  25.0  $\pm$  16.4 32.9  $\pm$  13.4 1.14  $\pm$  0.49

#### TABLE 1 (Continued) Comparison of pharmacokinetic parameters of some antimicrobial agents in adults and the elderly.



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Abbreviation: 1. C<sub>max</sub>: maximum concentration; 2. T<sub>1/2</sub>: half-life, V<sub>d</sub>: volume of distribution; 3. CL: clearance; 4. CL: renal clearance; 5. AUC: serum concentration time curve; 6. PBPK: physiologically based pharmacokinetic; 7. PPK: population pharmacokinetics.

blood flow. Studies indicate that individuals at the age of 85 have approximately half the number of liver cells compared to their younger counterparts, coupled with a reduction of about 40 percent in local liver blood flow [\(McLean and Le Couteur, 2004\)](#page-13-23). Consequently, for drugs with high hepatic clearance, the bioavailability may increase as the elderly experience diminished liver clearance of these drugs. However, the relationship between liver clearance and age for drugs with low liver clearance is a matter of controversy, warranting further investigation [\(McLachlan and](#page-13-24) [Pont, 2012\)](#page-13-24). Moreover, numerous drugs commonly prescribed for the elderly may interact with antimicrobial agents through hepatic microsomal enzymes ([Benson, 2017\)](#page-11-8). Take voriconazole, for instance, it serves as an inhibitor of CYP3A4 and CYP2C19 enzymes, influencing the metabolic rate of their substrates. Therefore, the concurrent use of drugs interacting

with CYP3A4 substrates during voriconazole therapy may lead to increased drug concentration, heightening the risk of adverse reactions [\(Theuretzbacher et al., 2006\)](#page-14-16).

## 2.4 Excretion

Most drugs undergo primary elimination through the kidneys, with some being excreted through bile, liver, the respiratory tract, and other pathways. In individuals over 65 years of age, the decline in kidney function exerts a more pronounced impact on pharmacokinetics than any other organ [\(Aymanns et al., 2010\)](#page-11-9). As individuals age, the kidneys undergo a series of changes, including a decrease in the number of glomeruli and glomerular density, along with an increase in the incidence of glomerular sclerosis [\(Esposito et al., 2007](#page-12-28)). Furthermore, the length and volume of renal tubules significantly diminish, and the renal artery gradually hardens while renal blood flow gradually decreases ([Bolignano et al.,](#page-11-11) [2014\)](#page-11-11). Beyond the age of 40, renal blood flow experiences a reduction of 10 percent per decade ([Silva, 2005\)](#page-13-29). Generally, in the elderly, decreased clearance, prolonged elimination half-life, and increased blood drug concentration of antibiotics eliminated by the kidneys are observed. These factors contribute to an extended retention time of the drug and elevate the likelihood of toxic accumulation in the body [\(Mühlberg and Platt, 1999\)](#page-13-30).

#### 3 Pharmacokinetic characteristics of antimicrobial agents in the elderly

In order to guarantee the safe and efficient utilization of antiinfective drugs among the elderly, comparative studies have been conducted for specific types of antimicrobials, encompassing both elderly and adult populations. [Table 1](#page-2-0) encapsulates the crucial PK parameters of anti-infective agents, as published to date, differentiating between older adults and adults. Particular emphasis should be directed towards antimicrobials exhibiting significant alterations in PK parameters within the elderly demographic.

#### 3.1 β-lactam antibiotics (penicillins, cephalosporins, atypical β-lactams, and βlactam inhibitors)

β-lactam antibiotics, recognized as time-dependent agents, exhibit distinct PK characteristics in the elderly, primarily characterized by an augmented area under the serum concentration-time curve (AUC) and prolonged plasma half-life  $(T_{1/2})$ . This phenomenon is evident across various penicillins, including penicillin, piperacillin, ampicillin, and amoxicillin. Notably, the AUC of amoxicillin in elderly subjects surpasses that of their younger counterparts (18,751 ± 3,903 h mg⋅L<sup>-1</sup> vs. 37,542 ± 9,523 h mg·L<sup>-1</sup>), coupled with a lengthened T<sub>1/2</sub> (1.43 ± 0.25 h vs. 1.75 ± 0.22 h) ([Lee et al., 2017;](#page-12-12) [Lee et al., 2020](#page-13-5)). Consistent findings emerge in other penicillin drugs [\(Yaoguo et al., 2001;](#page-14-5) [Meyers et al., 1991](#page-13-4)). Similarly, cephalosporins demonstrate higher AUC and prolonged  $T_{1/2}$  in the elderly [\(Yaoguo et al., 2001](#page-14-5); [Mingji](#page-13-7) [et al., 2007](#page-13-7); [Barbhaiya et al., 1992](#page-11-4)). For instance, a study on cefazolin reveals an elevated AUC in the elderly compared to adult subjects (380.71 ± 61.4 h mg·L<sup>-1</sup> vs. 339.56 ± 79.9 h mg·L<sup>-1</sup>), along with a prolonged  $T_{1/2}$  (2.5 ± 0.4 h vs. 1.9 ± 0.3 h) [\(Yaoguo et al., 2001\)](#page-14-5). Atypical β-lactams, including carbapenems and monocyclic βlactams, exhibit analogous trends [\(Musson et al., 2004](#page-13-8); [Ljungberg](#page-13-9) [and Nilsson-Ehle, 1992](#page-13-9); [Liu et al., 2013](#page-13-10); [Namkoong et al., 2014](#page-13-11); [Kim](#page-12-13) [et al., 2018](#page-12-13); [Harada et al., 2013](#page-12-14)). For example, ertapenem's AUC in the elderly surpasses that of the adult subjects (781.5 ± 95.9 h mg⋅L<sup>-1</sup> vs. 572.1 ± 68.6 h mg·L<sup>-1</sup>), accompanied by a prolonged T<sub>1/2</sub> (5.2 h vs.  $3.8 \pm 0.5$  h) [\(Musson et al., 2004\)](#page-13-8). Similarly, meropenem's AUC in the elderly surpasses that of the adult subjects ( $58.3 \pm 10.0$  h mg·L-1 vs. 39.6 ± 6.8 h mg·L<sup>−1</sup>), accompanied by a prolonged T<sub>1/2</sub> (1.27 h vs. 0.81 h) [\(Ljungberg and Nilsson-Ehle, 1992](#page-13-9)). Another population pharmacokinetics (PPK) study demonstrates that meropenem clearance rate (CL) is significantly lower in the elderly compared to CL reported in younger patients due to the reduced renal function ([Usman et al., 2017\)](#page-14-18). Aztreonam follows suit, with higher AUC in the elderly (469.01 ± 144.03 h mg·L<sup>-1</sup> vs. 294.42 ± 64.08 h mg·L<sup>-1</sup>) and an extended  $T_{1/2}$  (3.10 ± 0.90 h vs. 1.80 ± 0.51 h) ([Meyers et al.,](#page-13-12) [1993\)](#page-13-12). β-lactamase inhibitors, such as sulbactam, clavulanate, and avibactam, don't directly inhibit bacteria but effectively prevent βlactam antibiotics from hydrolysis by β-lactamase. In a PPK study on ceftazidime/avibactam [\(Li et al., 2019a\)](#page-13-6), it has bee observed that avibactam's AUC is higher in the elderly compared to younger subjects (156 h mg·L<sup>-1</sup> vs. 131 h mg·L<sup>-1</sup>). In light of these findings, it is advisable for elderly patients to appropriately reduce dosage and closely monitor serum creatinine levels (Scr). This precaution is crucial as the risk of adverse drug reactions escalates with higher AUC and prolonged  $T_{1/2}$  [\(Berg and Hahn, 2001\)](#page-11-12).

#### 3.2 Aminoglycoside antibiotics

Aminoglycoside antibiotics operate as concentration-dependent agents, implying that a higher peak concentration intensifies their lethality against pathogenic bacteria and accelerates the killing speed. These antibiotics, characterized by low plasma protein binding, exhibit a propensity to accumulate in specific areas such as the renal cortex, inner ear, and perilymph fluid, thereby posing a heightened risk of nephrotoxicity and ototoxicity. Compounding this, the primary elimination route for these drugs is through glomerular filtration. In the elderly and patients with renal failure, the original forms of aminoglycosides can experience an extension of  $T_{1/2}$  by more than 20 to 30 times, elevating the likelihood of drug accumulation and subsequent poisoning in these populations [\(Fujita et al., 1985](#page-12-29)). The PK profile of aminoglycosides in the elderly predominantly manifests as an increased  $T_{1/2}$  and a diminished plasma CL. A PK study on subcutaneous tobramycin injection in both adults and elderly healthy volunteers reveals notable differences ([Champoux et al.,](#page-11-5) [1996\)](#page-11-5). The  $T_{1/2}$  in the aged group is 3.53  $\pm$  0.97 h compared to 2.73  $\pm$ 1.17 h in the adult group at an 80 mg dosage ( $p < 0.05$ ). Additionally, CL in the elderly group significantly decreases to 5.28  $\pm$  1.44 L h<sup>-1</sup> from 6.9  $\pm$  3.12 L h<sup>-1</sup> in the adult group. Analogous outcomes are evident in studies involving gentamicin [\(Bos et al., 2019;](#page-11-6) [Johnston](#page-12-15) [et al., 2014](#page-12-15)) and amikacin [\(Yaoguo et al., 2001](#page-14-5)). Given these substantial PK changes, the elderly face a heightened risk of adverse drug reactions, necessitating strict dosage control based on PK parameters and therapeutic drug monitoring (TDM) during the administration of these medications ([Hodiamont et al., 2022\)](#page-12-30).

#### 3.3 Macrolide antibiotics

With the exception of azithromycin, which falls under concentration-dependent antibiotics, all other macrolides are categorized as time-dependent antibiotics [\(Dorfman et al., 2008\)](#page-12-31). Macrolide antibiotics have undergone three generations of development. The first generation, exemplified by erythromycin and dirithromycin, exhibits low bioavailability and is associated with common gastrointestinal reactions. The second generation, represented by azithromycin and clarithromycin, showcases

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improved bioavailability and a reduced incidence of side effects. The third generation, including telithromycin, demonstrates significantly enhanced bioavailability and antibacterial efficacy, with an absolute oral bioavailability of 57% in both adults and elderly individuals [\(Perret et al., 2002](#page-13-13)). Macrolide antibiotics are rapidly absorbed and widely distributed in extravascular tissues, with concentrations in tissues notably surpassing those in the bloodstream. Generally, in the elderly, age-related reductions in absorption and elimination lead to higher AUC and maximum concentration  $(C_{\text{max}})$  of macrolides compared to adult subjects. This trend is observed in telithromycin [\(Perret et al., 2002\)](#page-13-13), dirithromycin ([Varanese, 1993\)](#page-14-6), and clarithromycin ([Chu et al., 1992](#page-12-16)). However, a study on azithromycin reveals that while AUC is higher in the elderly compared to adult subjects (3.0 h mg·L<sup>-1</sup> vs. 2.5 h mg·L<sup>-1</sup>), Cmax shows only slight changes. Generally, adjusting dosing regimens based solely on age is unnecessary [\(Coates et al., 1991](#page-12-17)).

#### 3.4 Tetracycline antibiotics

Tetracycline antibiotics, including tetracycline, minocycline, doxycycline, and tigecycline, are classified as time-dependent and are predominantly metabolized by the liver. It is crucial for patients with hepatic insufficiency to exercise caution and minimize the use of tetracyclines. Studies investigating the PK characteristics of minocycline reveal significant changes in multiple parameters when comparing adults and elderly groups. In a continuous intravenous infusion of 200 mg minocycline for the adult group and 100 mg for the elderly group, notable differences emerge [\(JB,](#page-12-18) [1983;](#page-12-18) [Yamamoto et al., 1999](#page-14-7)). The C<sub>max</sub> in the elderly group is twice that of the adult group (7.22 ± 3.53 mg  $L^{-1}$  vs. 3.5 mg  $L^{-1}$ ), accompanied by a substantial prolongation of  $T_{1/2}$  in the elderly group (25.0  $\pm$  16.4 h vs. 15.7 h). Additionally, both V<sub>d</sub> and CL are reduced in the elderly group. Similarly, a study on tigecycline has observed a larger C<sub>max</sub> in elderly males compared to adult males (1.017 ± 0.112 mg L<sup>-1</sup> vs. 0.861 ± 0.154 mg L<sup>-1</sup>), with a prolonged T<sub>1</sub>/  $_2$  in the adult group (22.3  $\pm$  15.3 h vs. 19.0  $\pm$  5.0 h) ([Muralidharan](#page-13-14) [et al., 2005](#page-13-14)). However, it's noteworthy that these differences are attributed to variations in body size rather than age. Consequently, adjusting dosing regimens based solely on age appears unnecessary.

#### 3.5 Quinolone antibiotics

Quinolone antibiotics, renowned for their concentrationdependent nature, have evolved into the fourth generation, with the first- and second-generation drugs seeing less use due to their lower blood concentrations. The third-generation quinolones, encompassing ofloxacin, levofloxacin, ciprofloxacin, norfloxacin, among others, and the fourth-generation quinolones like moxifloxacin and gatifloxacin, are widely embraced in clinical applications owing to their expansive antibacterial spectrum, potent antibacterial action, and minimal adverse reactions. Comparatively, the PK characteristics of third- and fourthgeneration quinolones in the elderly differ from those in adult individuals, primarily marked by increased  $C_{\text{max}}$  and  $\text{AUC}$ ([Nicolle, 1999\)](#page-13-31). The elimination routes of quinolones predominantly involve a hepatorenal dual pathway, with renal

elimination taking precedence. In clinical practice, the choice of quinolone is contingent upon the patient's liver and kidney function. For instance, ofloxacin, predominantly eliminated through the kidneys, exhibits larger AUC in the elderly compared to the adults (83.3 ± 16.6 h mg L<sup>-1</sup> vs. 21.5 ± 4.27 h mg L<sup>-1</sup>), along with a prolonged  $T_{1/2}$  (8.5 ± 1.2 h vs. 6.2 ± 0.9 h) [\(Molinaro et al.,](#page-13-15) [1992\)](#page-13-15). Therefore, the dose of ofloxacin for patients over 75 years old should be halved compared to adult patients. Studies on levofloxacin and sitafloxacin, which are mainly eliminated through the kidneys, yield similar results [\(Setiawan et al., 2022](#page-13-16); [Cojutti et al., 2017](#page-12-19); [Yaozh,](#page-14-8) [2023\)](#page-14-8). On the other hand, ciprofloxacin, eliminated through both the liver and kidneys ([Li et al., 2019b\)](#page-13-32), does not undergo significant changes in PK parameters in the elderly [\(Ljungberg and Nilsson-](#page-13-17)[Ehle, 1989](#page-13-17)), rendering dose adjustment generally unnecessary. Novel quinolones antibacterial agents, including garenoxacin ([Van Wart et al., 2004;](#page-14-9) [Ohsaki et al., 2010\)](#page-13-18), nemonoxacin [\(Wu](#page-14-10) [et al., 2015](#page-14-10); [Kang et al., 2019](#page-12-20)), delafloxacin [\(Hoover et al., 2016](#page-12-21)), and lascufloxacin ([Totsuka et al., 2020](#page-14-11)), have not shown more pronounced adverse reactions. Therefore, it is recommended that the elderly use these drugs in the same manner as adult individuals.

#### 3.6 Oxazolidinone agents

Oxazolidinone agents, a class of synthetically derived antibacterial agents that inhibit protein synthesis, are exemplified by oral linezolid, which boasts rapid and complete absorption with exceptionally high bioavailability. A PK study of linezolid indicates that its PK parameters do not significantly differ between the elderly and the adults, suggesting that age-based dose adjustment is unnecessary [\(Sisson et al., 2002](#page-13-19)). Tedizolid, a novel oxazolidinone antibiotic, exhibits a more favorable PK and safety profile compared to linezolid [\(Iqbal et al., 2022\)](#page-12-32). An open-label phase I study on tedizolid reveals similar PK parameters across different age groups, affirming that dosage adjustments based on age are not warranted ([Flanagan et al., 2018](#page-12-22)). Contezolid (MRX-I), another recently approved oxazolidinone agent in 2021 for treating complicated skin and soft tissue infections (cSSTIs) in China [\(Hoy, 2021\)](#page-12-33), demonstrates potent activity against drug-resistant Gram-positive bacteria, with a higher safety profile than linezolid [\(Wu et al., 2020\)](#page-14-19). A PPK study on contezolid suggests that age has no significant impact on the PK parameters, further supporting the conclusion that dosage adjustment based on age is unnecessary [\(Li et al., 2020\)](#page-13-20).

#### 3.7 Polypeptide antibiotics

Polypeptide antibiotics, encompassing polymyxins and glycopeptides, exhibit distinct PK characteristics. Polymyxins, including polymyxin B and polymyxin E (colistin), operate as concentration-dependent antibiotics. PPK studies ([Manchandani](#page-13-21) [et al., 2018](#page-13-21); [Wang et al., 2022a\)](#page-14-12) on polymyxin B in adults and the elderly reveal a non-statistically significant difference in CL between adults and elderly subjects (2.5 ± 1.1 L h<sup>-1</sup> vs. 1.98 ± 0.67 L h<sup>−</sup><sup>1</sup> ). Generally, dose adjustment is not deemed necessary for the elderly; however, TDM is advisable due to potential renal toxicity ([Fiaccadori et al., 2016;](#page-12-34) [Wang et al., 2021](#page-14-20)). Though there's a dearth of studies on colistimethate sodium (CMS) pharmacokinetics in the

elderly, initiating colistin at a low dose and closely monitoring renal function is recommended owing to its nephrotoxicity ([Drugsatfda,](#page-12-35) [2023\)](#page-12-35). On the other hand, glycopeptides, such as vancomycin, norvancomycin, teicoplanin, and travanin, are time-dependent antibiotics. Vancomycin and norvancomycin, primarily excreted by the kidneys, exhibit significantly longer  $T_{1/2}$  in the elderly compared to adults [\(Zhang et al., 2008](#page-14-13); [Guay et al., 1993\)](#page-12-23). Therefore, it is advisable to reduce the dose of vancomycin and norvancomycin and closely monitor renal function in the elderly. Teicoplanin studies ([Thompson et al., 1992](#page-14-14); [Rosina et al., 1988](#page-13-22)) suggest comparable CL and  $T_{1/2}$  in the elderly and adults; however, considerable variation in teicoplanin PK parameters related to renal function among the elderly necessitates dosage and administration interval adjustments based on Scr levels ([Uhart et al., 2013\)](#page-14-21). Travanin, with similar clearance in the elderly and the adults, does not require age-based dosage adjustments [\(Wong et al.,](#page-14-15) [2008;](#page-14-15) [Goldberg et al., 2010\)](#page-12-24).

#### 3.8 Antituberculosis agents

Many medications possess anti-tuberculosis properties, and this section concentrates on the pharmacokinetic profiles of first-line oral anti-tuberculosis agents, including isoniazid, rifampicin, ethambutol, and pyrazinamide. For isoniazid, regardless of whether individuals are fast or slow acetylators, there are no significant differences in pharmacokinetic parameters between adults and elderly subjects [\(Advenier et al., 1980](#page-11-7)). As a result, dosage adjustments for older adults are unnecessary. Early research has shown that when 450 mg of rifampicin is administered orally, there are no significant differences in rifampicin pharmacokinetics between older and middle-aged tuberculosis patients [\(Weng, 1990\)](#page-14-17). Moreover, considering the increasing resistance to rifampicin, future studies should consider higher doses ([Abulfathi et al., 2019\)](#page-11-13). According to a clinical trial involving 100 Tanzanian tuberculosis patients, CL of ethambutol was found to decrease with age, while  $C_{\text{max}}$  and AUC increased with age ([Denti et al., 2015](#page-12-36)). Additionally, a physiologically based pharmacokinetic (PBPK) study on ethambutol indicates that renal function is a significant covariate affecting interindividual variability in the apparent clearance of ethambutol [\(Hung et al.,](#page-12-37) [2023\)](#page-12-37). Therefore, it is recommended that elderly individuals use a lower dosage than adults individuals. Currently, only a PBPK study has demonstrated that CL of pyrazinamide correlated with creatinine clearance ( $p = 0.0349$ ) and showed a trend toward smaller CL values with advancing age  $(p = 0.0778)$  ([Peloquin](#page-13-33) [et al., 1997](#page-13-33)). However, due to the lack of statistical significance in the latter finding, more studies are needed to further determine whether dose adjustments are necessary in elderly patients.

#### 3.9 Antifungal agents

The pharmacokinetics of antifungal agents in the elderly predominantly focus on synthetic antifungal drugs such as triazoles and terbinafine. Elderly patients using antifungals primarily metabolized by the liver are more prone to developing liver diseases ([Chien et al., 1997\)](#page-12-38), necessitating cautious use of these drugs in this population and performance of TDM [\(Ashbee et al.,](#page-11-14) [2014\)](#page-11-14). A study on voriconazole reveals a significant increase in  $C_{\text{max}}$ in the elderly compared to adults (4.31  $\pm$  3.03 mg L<sup>-1</sup> vs. 3.11  $\pm$ 2.13 mg L<sup>−</sup><sup>1</sup> ) ([Cheng et al., 2020](#page-12-25)). Due to reduced liver function in aged patients, lower dosages and longer dosing intervals for voriconazole are recommended ([Li et al., 2023\)](#page-13-34). On the other hand, a study on the pharmacokinetics of posaconazole oral suspension indicates that although  $C_{\text{max}}$ ,  $T_{1/2}$ , and AUC in the elderly are higher than in adult individuals, the differences are nonsignificant, and dosage adjustment based on age is unnecessary ([Sansone-Parsons et al., 2007](#page-13-25)). Similarly, a PPK study on isavuconazole has found that although CL is lower in the elderly than in adult individuals (1.44 L h<sup>-1</sup> vs. 2.33 L h<sup>-1</sup>), the difference is not significant [\(Desai et al., 2019](#page-12-26)). Therefore, adjusting isavuconazole dosage solely based on age is deemed unnecessary. Moreover, despite significant differences in the PK parameters of terbinafine across different ages, the variations are not substantial enough to lead to greater toxicity. Hence, there is no need to alter terbinafine dosage based on age ([Nedelman et al., 1997](#page-13-26)).

#### 3.10 Antiviral agents

Oseltamivir, an anti-influenza drug, exhibits an AUC that is 80% higher in the elderly compared to adult individuals. However, it is well tolerated in very elderly individuals, and dose adjustment is typically not necessary in elderly patients [\(Abe et al., 2006\)](#page-11-10). It's crucial to use antiviral drugs judiciously in older individuals, particularly those living with HIV, due to a higher prevalence of multidrug therapy, potential inappropriate drug use, and drug-drug interactions ([Guaraldi et al., 2018\)](#page-12-39). Bictegravir, a novel integrase inhibitor recommended for most HIV patients by US and European guidelines, does not require dosage adjustment for age, except in older HIV patients with severe comorbidities, according to a recent PBPK study [\(Stader et al., 2021](#page-13-27)). Amidst the COVID-19 pandemic, Paxlovid has emerged as a promising drug for treating the virus ([Amani and Amani, 2023](#page-11-15)). Recent PPK studies indicate that the CL of Paxlovid in adult individuals is approximately twice that in the elderly (9.09 L h<sup>-1</sup> vs. 4.16 L h<sup>-1</sup>). In these studies, creatinine clearance is identified as a significant covariate affecting the CL of nirmatrelvir, suggesting that dosage adjustment for Paxlovid in elderly patients should be considered based on creatinine clearance ([Chan et al., 2023;](#page-12-27) [Qu et al., 2023\)](#page-13-28).

#### 4 Advances in pharmacokinetic research methods of antimicrobials agents in elderly population

Given their compromised immunity and increased susceptibility to underlying diseases, the elderly population faces heightened vulnerability to infectious diseases caused by diverse pathogens. Consequently, this demographic is more prone to encountering adverse drug events during clinical trials, thereby amplifying the intricacies of PK research in the elderly [\(Schlender et al., 2016\)](#page-13-35). Therefore, to efficiently ascertain PK information and deliver precise dosage recommendations for elderly patients, it becomes imperative to explore alternative methods for clinical PK determination.

#### 4.1 PBPK

PBPK model is a mathematical framework that uses NONMEM, PK-Sim and other software to perform PBPK modeling to simulates the absorption, distribution, metabolism, and excretion of drugs within various tissues and organs, based on knowledge from physiology, biochemistry, and anatomy [\(Nestorov, 2003](#page-13-36); [Sager](#page-13-37) [et al., 2015](#page-13-37)). In recent years, thanks to modern modeling and simulation techniques, PBPK models have progressively emerged as a novel alternative for determining pharmacokinetics and pharmacodynamics in the elderly patient population ([Cui](#page-12-40) [et al., 2021](#page-12-40)).

Ciprofloxacin, a widely used fluoroquinolone anti-infective drug, has seen a surge in drug resistance, emphasizing the critical need to select effective doses based on its PK characteristics for rational clinical use. [Schlender et al. \(2018\)](#page-13-38) have meticulously collected and screened clinical PK data for both oral and intravenous ciprofloxacin across all age groups. Subsequently, they have employed PK-Sim and the systems biology platform MoBi to construct PBPK models for both administration methods, validating these models in adults. Finally, they extend the ciprofloxacin PBPK model to encompass elderly and pediatric populations, comparing predicted exposure with clinical observations. Impressively, only 4.82% of intravenous doses and 12.13% of oral doses, on average, deviate from the range of the simulated data. Both the PBPK model for intravenous ciprofloxacin in adults and the PBPK model for oral ciprofloxacin in adults demonstrate reliability. When comparing predicted exposures for the elderly, children, and the entire life cycle with observed exposures, only 7.61% of intravenous dosing and 5.56% of oral dosing are anticipated to deviate beyond twice the average observed exposure. These findings highlight the ciprofloxacin PBPK model's proficiency in accurately predicting exposure variations across different age groups and throughout the entire life cycle. The establishment and validation of this model are pivotal for optimizing dosage schemes and clinical trial designs for ciprofloxacin, particularly in special populations like elderly patients.

#### 4.2 PPK

PPK integrates the PK model with a statistical model to quantitatively analyze the influencing factors of PK parameters in the patient population and the variability of PK when following a standard dose regimen. This analysis aids in proposing a more rational and effective alternative drug administration regimen ([Sheiner and Ludden, 1992](#page-13-39); [Setiawan et al., 2023\)](#page-13-40). Since 1998, the US Food and Drug Administration has permitted the use of population analysis in phase II and III clinical trials of new drugs to assess the PK/PD of special subjects [\(FDA, 1999\)](#page-12-41). Notably, PPK studies of antibiotics in elderly patients have been on the rise.

Polymyxin B, as the last resort of multi-drug-resistant Gramnegative bacteria, pharmacokinetic data are currently scarce in the elderly population. [Wang et al. \(2022b\)](#page-14-22) have established a PPK model using the Phoenix NLME software for critically ill elderly patients undergoing treatment with polymyxin B for multidrugresistant Gram-negative infections. Nonlinear mixed-effect models and stepwise analysis methods are employed to explore the PK parameter covariate. Subsequently, based on the final PPK model, Monte Carlo simulations are conducted on three fixed-dose regimens (50 mg·12 h<sup>-1</sup>, 75 mg 12 h<sup>-1</sup>, and 100 mg 12 h<sup>-1</sup>) and weight-based regimens (1.25 mg kg·12 h $^{-1}$  and 1.5 mg kg·12 h $^{-1})$  to compare the efficacy and renal toxicity of each regimen. Results reveal that albumin is the primary covariate influencing PK parameters of polymyxin B in elderly patients. Simulation outcomes demonstrate that when MIC≤0.5 mg L<sup>−</sup><sup>1</sup> , the probability of reaching the PK/PD target (ACU24 h/MIC >50) for all schemes is 90%. Fixed-dose regimens of 50 mg 12 h<sup>-1</sup> and 75 mg 12 h<sup>−</sup><sup>1</sup> result in a relatively low likelihood of AUCss, 24 h > 100 mg h·L<sup>−</sup><sup>1</sup> , at 53.9% and 66.7%, respectively. Therefore, these fixed-dose regimens of polymyxin B can achieve maximum efficacy while balancing renal toxicity. The PPK model of polymyxin B in elderly patients offers a safer and more effective administration scheme for critically ill elderly patients.

#### 4.3 In vitro to vivo extrapolation (IVIVE)

IVIVE is a method utilized to forecast the rate at which drugs undergo metabolism in the human liver by leveraging in vitro drug metabolism measurements ([Rowland and Pang, 2022\)](#page-13-41). This process involves three key steps: initially measuring intrinsic clearance in vitro, subsequently establishing a correction equation for in vitro liver intrinsic clearance, and ultimately predicting liver clearance within the body [\(Sodhi and Benet, 2021](#page-13-42)). As IVIVE technology advances, its application scope has progressively expanded from animals to humans. [Polasek et al. \(2010\)](#page-13-43) have employed IVIVE to assess the potential interaction between zolpidem and CYP3A-metabolized drugs, predicting its impact on the pharmacokinetics of this CYP3A-metabolized drug in humans. The anticipated results indicate that this interaction elevates the AUC of the drug in the elderly by 1–2 times. However, IVIVE does have limitations, including its tendency to exhibit poor accuracy in predicting in vivo data from in vitro measurements ([Benet and Sodhi, 2022\)](#page-11-16). Combining IVIVE with PBPK models can enhance prediction accuracy to a certain extent and assist in predicting drug clearance in the elderly [\(Polasek](#page-13-44) [et al., 2013](#page-13-44); [Kim and Lee, 2022](#page-12-42)).

#### 4.4 Machine learning (ML)

ML is a powerful method for analyzing and simulating large volumes of data, finding applications in diverse fields such as image recognition and language processing ([Zhou et al., 2018](#page-14-23); [Popel et al.,](#page-13-45) [2020\)](#page-13-45). Its potential in the realm of PK analysis and prediction is considerable and can be harnessed for both adults and the elderly ([Ota and Yamashita, 2022\)](#page-13-46). [Cheng et al. \(2023\)](#page-12-43) have employed 3 ML algorithms (random forest, gradient boosting, and extreme gradient boosting) to predict toxic plasma trough concentrations of Voriconazole (>5 μg mL<sup>−</sup><sup>1</sup> ). Results indicate that the prediction values from models established by these algorithms, including younger adult models, mixed models, and elderly models, are all meaningful. The study concludes that ML models can serve as reliable tools for predicting toxic concentration exposure of Voriconazole ML is broadly categorized into supervised and unsupervised learning. In comparison with PBPK and PPK models, ML possesses the ability to automatically re-establish

mathematical relationships when data are updated, resulting in enhanced predictive performance ([Gill et al., 2022](#page-12-44)). Furthermore, when compared with IVIVE and animal scaling (AS), ML exhibits economic efficiency and the capability to process extensive data and samples [\(Danishuddin et al., 2022\)](#page-12-45). Moreover, innovative approaches involve combining ML with PBPK/PPK or integrating ML with IVIVE/AS. For instance, [Kosugi and Hosea](#page-12-46) [\(2021\)](#page-12-46) have explored a quantitative method for predicting the area under the plasma concentration-time curve after oral administration in rats by combining IVIVE with ML. The findings demonstrate that the amalgamation of ML and IVIVE significantly improves the ability to predict the area under the plasma concentration-time curve after oral administration.

# 5 Conclusion

Given the expanding elderly population and the critical need for precise anti-infection treatment tailored to this demographic, coupled with the limited research on numerous promising antimicrobial agents in the elderly, there is an urgent call to comprehensively delineate the PK characteristics of these agents in elderly patients and formulate rational administration schemes. Furthermore, the current PK study methods for the elderly exhibit certain limitations. Therefore, it is imperative to not only optimize existing PK research methods but also explore innovative approaches to address these limitations and enhance our understanding of drug dynamics in the elderly.

#### Author contributions

GF: Writing–original draft. WS: Writing–original draft. ZT: Data curation, Investigation, Writing–review and editing. BL:

#### References

<span id="page-11-10"></span>Abe, M., Smith, J., Urae, A., Barrett, J., Kinoshita, H., and Rayner, C. R. (2006). Pharmacokinetics of oseltamivir in young and very elderly subjects. Ann. Pharmacother. 40, 1724–1730. doi:[10.1345/aph.1H174](https://doi.org/10.1345/aph.1H174)

<span id="page-11-13"></span>Abulfathi, A. A., Decloedt, E. H., Svensson, E. M., Diacon, A. H., Donald, P., and Reuter, H. (2019). Clinical pharmacokinetics and pharmacodynamics of rifampicin in human tuberculosis. Clin. Pharmacokinet. 58, 1103–1129. doi[:10.1007/s40262-019-](https://doi.org/10.1007/s40262-019-00764-2) [00764-2](https://doi.org/10.1007/s40262-019-00764-2)

<span id="page-11-7"></span>Advenier, C., Saint-Aubin, A., Gobert, C., Houin, G., Albengres, E., and Tillement, J. P. (1980). Pharmacokinetics of isoniazid in the elderly. Br. J. Clin. Pharmacol. 10, 167–169. doi:[10.1111/j.1365-2125.1980.tb01735.x](https://doi.org/10.1111/j.1365-2125.1980.tb01735.x)

<span id="page-11-15"></span>Amani, B., and Amani, B. (2023). Efficacy and safety of nirmatrelvir/ritonavir (Paxlovid) for COVID-19: a rapid review and meta-analysis. J. Med. Virol. 95, e28441. doi:[10.1002/jmv.28441](https://doi.org/10.1002/jmv.28441)

<span id="page-11-1"></span>Antimicrobial Resistance Collaborators (2023). The burden of antimicrobial resistance in the Americas in 2019: a cross-country systematic analysis. Lancet Reg. Health Am. 25, 100561. doi:[10.1016/j.lana.2023.100561](https://doi.org/10.1016/j.lana.2023.100561)

<span id="page-11-14"></span>Ashbee, H. R., Barnes, R. A., Johnson, E. M., Richardson, M. D., Gorton, R., and Hope, W. W. (2014). Therapeutic drug monitoring (TDM) of antifungal agents: guidelines<br>from the British Society for Medical Mycology. J. Antimicrob. Chemother. 69, 1162–1176. doi[:10.1093/jac/dkt508](https://doi.org/10.1093/jac/dkt508)

<span id="page-11-9"></span>Aymanns, C., Keller, F., Maus, S., Hartmann, B., and Czock, D. (2010). Review on pharmacokinetics and pharmacodynamics and the aging kidney. Clin. J. Am. Soc. Nephrol. 5, 314–327. doi:[10.2215/CJN.03960609](https://doi.org/10.2215/CJN.03960609)

<span id="page-11-4"></span>Barbhaiya, R. H., Knupp, C. A., and Pittman, K. A. (1992). Effects of age and gender on pharmacokinetics of cefepime. Antimicrob. Agents Chemother. 36, 1181–1185. doi[:10.1128/aac.36.6.1181](https://doi.org/10.1128/aac.36.6.1181)

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# Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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<span id="page-11-2"></span>Beaufrère, B., and Morio, B. (2000). Fat and protein redistribution with aging: metabolic considerations. Eur. J. Clin. Nutr. 54 (Suppl. 3), S48-S53. doi:[10.1038/sj.](https://doi.org/10.1038/sj.ejcn.1601025) [ejcn.1601025](https://doi.org/10.1038/sj.ejcn.1601025)

<span id="page-11-16"></span>Benet, L. Z., and Sodhi, J. K. (2022). Can in vitro-in vivo extrapolation Be successful? Recognizing the incorrect clearance assumptions. Clin. Pharmacol. Ther. 111, 1022–1035. doi[:10.1002/cpt.2482](https://doi.org/10.1002/cpt.2482)

<span id="page-11-8"></span>Benson, J. M. (2017). Antimicrobial pharmacokinetics and pharmacodynamics in older adults. Infect. Dis. Clin. North Am. 31, 609–617. doi[:10.1016/j.idc.2017.07.011](https://doi.org/10.1016/j.idc.2017.07.011)

<span id="page-11-12"></span>Berg, P., and Hahn, E. G. (2001). Hepatotoxic reactions induced by beta-lactamase inhibitors. Eur. J. Med. Res. 6, 535–542.

<span id="page-11-11"></span>Bolignano, D., Mattace-Raso, F., Sijbrands, E. J., and Zoccali, C. (2014). The aging kidney revisited: a systematic review. Ageing Res. Rev. 14, 65–80. doi:[10.1016/j.arr.2014.](https://doi.org/10.1016/j.arr.2014.02.003) [02.003](https://doi.org/10.1016/j.arr.2014.02.003)

<span id="page-11-6"></span>Bos, J. C., Prins, J. M., Mistício, M. C., Nunguiane, G., Lang, C. N., Beirão, J. C., et al. (2019). Population pharmacokinetics with Monte Carlo simulations of gentamicin in a population of severely ill adult patients from sub-saharan africa. Antimicrob. Agents Chemother. 63, e02328. doi:[10.1128/AAC.02328-18](https://doi.org/10.1128/AAC.02328-18)

<span id="page-11-0"></span>Bouza, E., Brenes, F. J., Díez Domingo, J., Eiros Bouza, J. M., González, J., Gracia, D., et al. (2020). The situation of infection in the elderly in Spain: a multidisciplinary opinion document. Rev. Esp. Quimioter. 33, 327–349. doi[:10.37201/req/057.2020](https://doi.org/10.37201/req/057.2020)

<span id="page-11-3"></span>Celestin, M. N., and Musteata, F. M. (2021). Impact of changes in free concentrations and drug-protein binding on drug dosing regimens in special populations and disease states. J. Pharm. Sci. 110, 3331–3344. doi:[10.1016/j.xphs.2021.05.018](https://doi.org/10.1016/j.xphs.2021.05.018)

<span id="page-11-5"></span>Champoux, N., Du Souich, P., Ravaoarinoro, M., Phaneuf, D., Latour, J., and Cusson, J. R. (1996). Single-dose pharmacokinetics of ampicillin and tobramycin administered by hypodermoclysis in young and older healthy volunteers. Br. J. Clin. Pharmacol. 42, 325–331. doi:[10.1046/j.1365-2125.1996.03967.x](https://doi.org/10.1046/j.1365-2125.1996.03967.x)

<span id="page-12-27"></span>Chan, P. L. S., Singh, R. S. P., Cox, D. S., Shi, H., Damle, B., and Nicholas, T. (2023). Dosing recommendation of nirmatrelvir/ritonavir using an integrated population pharmacokinetic analysis. CPT Pharmacometrics Syst. Pharmacol. 12, 1897–1910. doi[:10.1002/psp4.13039](https://doi.org/10.1002/psp4.13039)

<span id="page-12-10"></span>Chen, L., Krekels, E. H. J., Verweij, P. E., Buil, J. B., Knibbe, C. A. J., and Brüggemann, R. J. M. (2020). Pharmacokinetics and pharmacodynamics of posaconazole. Drugs 80, 671–695. doi:[10.1007/s40265-020-01306-y](https://doi.org/10.1007/s40265-020-01306-y)

<span id="page-12-25"></span>Cheng, L., Xiang, R., Liu, F., Li, Y., Chen, H., Yao, P., et al. (2020). Therapeutic drug monitoring and safety of voriconazole in elderly patients. Int. Immunopharmacol. 78, 106078. doi:[10.1016/j.intimp.2019.106078](https://doi.org/10.1016/j.intimp.2019.106078)

<span id="page-12-43"></span>Cheng, L., Zhao, Y., Liang, Z., You, X., Jia, C., Liu, X., et al. (2023). Prediction of plasma trough concentration of voriconazole in adult patients using machine learning. Eur. J. Pharm. Sci. 188, 106506. doi[:10.1016/j.ejps.2023.106506](https://doi.org/10.1016/j.ejps.2023.106506)

<span id="page-12-38"></span>Chien, R. N., Yang, L. J., Lin, P. Y., and Liaw, Y. F. (1997). Hepatic injury during ketoconazole therapy in patients with onychomycosis: a controlled cohort study. Hepatology 25, 103–107. doi[:10.1002/hep.510250119](https://doi.org/10.1002/hep.510250119)

<span id="page-12-16"></span>Chu, S. Y., Wilson, D. S., Guay, D. R., and Craft, C. (1992). Clarithromycin pharmacokinetics in healthy young and elderly volunteers. J. Clin. Pharmacol. 32, 1045–1049. doi[:10.1002/j.1552-4604.1992.tb03809.x](https://doi.org/10.1002/j.1552-4604.1992.tb03809.x)

<span id="page-12-17"></span>Coates, P., Daniel, R., Houston, A. C., Antrobus, J. H., and Taylor, T. (1991). An open study to compare the pharmacokinetics, safety and tolerability of a multiple-dose regimen of azithromycin in young and elderly volunteers. Eur. J. Clin. Microbiol. Infect. Dis. 10, 850–852. doi[:10.1007/BF01975839](https://doi.org/10.1007/BF01975839)

<span id="page-12-19"></span>Cojutti, P. G., Ramos-Martin, V., Schiavon, I., Rossi, P., Baraldo, M., Hope, W., et al. (2017). Population pharmacokinetics and pharmacodynamics of levofloxacin in acutely hospitalized older patients with various degrees of renal function. Antimicrob. Agents Chemother. 61, e02134. doi[:10.1128/AAC.02134-16](https://doi.org/10.1128/AAC.02134-16)

<span id="page-12-40"></span>Cui, C., Valerie Sia, J. E., Tu, S., Li, X., Dong, Z., Yu, Z., et al. (2021). Development of a physiologically based pharmacokinetic (PBPK) population model for Chinese elderly subjects. Br. J. Clin. Pharmacol. 87, 2711–2722. doi[:10.1111/bcp.14609](https://doi.org/10.1111/bcp.14609)

<span id="page-12-45"></span>Danishuddin, K. V., Faheem, M., and Woo Lee, K. (2022). A decade of machine learning-based predictive models for human pharmacokinetics: advances and challenges. Drug Discov. Today 27, 529–537. doi:[10.1016/j.drudis.2021.09.013](https://doi.org/10.1016/j.drudis.2021.09.013)

<span id="page-12-36"></span>Denti, P., Jeremiah, K., Chigutsa, E., Faurholt-Jepsen, D., PrayGod, G., Range, N., et al. (2015). Pharmacokinetics of isoniazid, pyrazinamide, and ethambutol diagnosed pulmonary TB patients in Tanzania. PLoS One 10, e0141002. doi[:10.](https://doi.org/10.1371/journal.pone.0141002) [1371/journal.pone.0141002](https://doi.org/10.1371/journal.pone.0141002)

<span id="page-12-26"></span>Desai, A. V., Han, D., Kowalski, D. L., Lademacher, C., Pearlman, H., and Yamazaki, T. (2019). No dose adjustment for isavuconazole based on age or sex. Antimicrob. Agents Chemother. 63, e02629. doi[:10.1128/AAC.02629-18](https://doi.org/10.1128/AAC.02629-18)

<span id="page-12-31"></span>Dorfman, M. S., Wagner, R. S., Jamison, T., Bell, B., and Stroman, D. W. (2008). The pharmacodynamic properties of azithromycin in a kinetics-of-kill model and implications for bacterial conjunctivitis treatment. Adv. Ther. 25, 208–217. doi[:10.](https://doi.org/10.1007/s12325-008-0034-x) [1007/s12325-008-0034-x](https://doi.org/10.1007/s12325-008-0034-x)

<span id="page-12-35"></span>Drugsatfda (2023). Instructions for polymyxin E sodium mesylate. Available at: [https://](https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/050108s033lbl.pdf) [www.accessdata.fda.gov/drugsatfda\\_docs/label/2017/050108s033lbl.pdf](https://www.accessdata.fda.gov/drugsatfda_docs/label/2017/050108s033lbl.pdf) (Accessed April 4, 2023).

<span id="page-12-28"></span>Esposito, C., Plati, A., Mazzullo, T., Fasoli, G., De Mauri, A., Grosjean, F., et al. (2007). Renal function and functional reserve in healthy elderly individuals. *J. Nephrol.* 20, 617–625.

<span id="page-12-4"></span>Farrall, A. J., and Wardlaw, J. M. (2009). Blood-brain barrier: ageing and microvascular disease--systematic review and meta-analysis. Neurobiol. Aging 30, 337–352. doi:[10.1016/j.neurobiolaging.2007.07.015](https://doi.org/10.1016/j.neurobiolaging.2007.07.015)

<span id="page-12-41"></span>FDA (1999). FDA guidance for industry on "population pharmacokinetics".

<span id="page-12-2"></span>Fernández-Sabé, N., Carratalà, J., Rosón, B., Dorca, J., Verdaguer, R., Manresa, F., et al. (2003). Community-acquired pneumonia in very elderly patients: causative organisms, clinical characteristics, and outcomes. Med. Baltim. 82, 159–169. doi[:10.](https://doi.org/10.1097/01.md.0000076005.64510.87) [1097/01.md.0000076005.64510.87](https://doi.org/10.1097/01.md.0000076005.64510.87)

<span id="page-12-34"></span>Fiaccadori, E., Antonucci, E., Morabito, S., d'Avolio, A., Maggiore, U., and Regolisti, G. (2016). Colistin use in patients with reduced kidney function. Am. J. Kidney Dis. 68, 296–306. doi:[10.1053/j.ajkd.2016.03.421](https://doi.org/10.1053/j.ajkd.2016.03.421)

<span id="page-12-22"></span>Flanagan, S. D., Minassian, S. L., and Prokocimer, P. (2018). Pharmacokinetics, safety, and tolerability of tedizolid phosphate in elderly subjects. Clin. Pharmacol. Drug Dev. 7, 788–794. doi:[10.1002/cpdd.426](https://doi.org/10.1002/cpdd.426)

<span id="page-12-29"></span>Fujita, K., Sayama, T., Abe, S., Murayama, T., and Tashiro, H. (1985). Age-dependent aminoglycoside nephrotoxicity. J. Urol. 134, 596–597. doi[:10.1016/s0022-5347\(17\)](https://doi.org/10.1016/s0022-5347(17)47308-3) [47308-3](https://doi.org/10.1016/s0022-5347(17)47308-3)

<span id="page-12-9"></span>Giarratano, A., Green, S. E., and Nicolau, D. P. (2018). Review of antimicrobial use and considerations in the elderly population. Clin. Interv. Aging 13, 657–667. doi[:10.](https://doi.org/10.2147/CIA.S133640) [2147/CIA.S133640](https://doi.org/10.2147/CIA.S133640)

<span id="page-12-44"></span>Gill, J., Moullet, M., Martinsson, A., Miljković, F., Williamson, B., Arends, R. H., et al. (2022). Comparing the applications of machine learning, PBPK, and population pharmacokinetic models in pharmacokinetic drug-drug interaction prediction. CPT Pharmacometrics Syst. Pharmacol. 11, 1560–1568. doi:[10.1002/psp4.12870](https://doi.org/10.1002/psp4.12870)

<span id="page-12-6"></span>Gnjidic, D., Husband, A., and Todd, A. (2018). Challenges and innovations of delivering medicines to older adults. Adv. Drug Deliv. Rev. 135, 97–105. doi[:10.](https://doi.org/10.1016/j.addr.2018.08.003) [1016/j.addr.2018.08.003](https://doi.org/10.1016/j.addr.2018.08.003)

<span id="page-12-24"></span>Goldberg, M. R., Wong, S. L., Shaw, J. P., Kitt, M. M., and Barriere, S. L. (2010). Singledose pharmacokinetics and tolerability of telavancin in elderly men and women. Pharmacotherapy 30, 806–811. doi:[10.1592/phco.30.8.806](https://doi.org/10.1592/phco.30.8.806)

<span id="page-12-1"></span>González-Castillo, J., Martín-Sánchez, F. J., Llinares, P., Menéndez, R., Mujal, A., Navas, E., et al. (2014). Guidelines for the management of community-acquired pneumonia in the elderly patient. Rev. Esp. Quimioter. 27, 69–86.

<span id="page-12-39"></span>Guaraldi, G., Pintassilgo, I., Milic, J., and Mussini, C. (2018). Managing antiretroviral therapy in the elderly HIV patient. Expert Rev. Clin. Pharmacol. 11, 1171-1181. doi[:10.](https://doi.org/10.1080/17512433.2018.1549484) [1080/17512433.2018.1549484](https://doi.org/10.1080/17512433.2018.1549484)

<span id="page-12-23"></span>Guay, D. R., Vance-Bryan, K., Gilliland, S., Rodvold, K., and Rotschafer, J. (1993). Comparison of vancomycin pharmacokinetics in hospitalized elderly and young patients using a Bayesian forecaster. J. Clin. Pharmacol. 33, 918–922. doi:[10.1002/j.](https://doi.org/10.1002/j.1552-4604.1993.tb01922.x) [1552-4604.1993.tb01922.x](https://doi.org/10.1002/j.1552-4604.1993.tb01922.x)

<span id="page-12-14"></span>Harada, M., Inui, N., Suda, T., Nakamura, Y., Wajima, T., Matsuo, Y., et al. (2013). Pharmacokinetic analysis of doripenem in elderly patients with nosocomial pneumonia. Int. J. Antimicrob. Agents 42, 149–154. doi:[10.1016/j.ijantimicag.2013.03.012](https://doi.org/10.1016/j.ijantimicag.2013.03.012)

<span id="page-12-30"></span>Hodiamont, C. J., van den Broek, A. K., de Vroom, S. L., Prins, J. M., Mathôt, R. A. A., and van Hest, R. M. (2022). Clinical pharmacokinetics of gentamicin in various patient populations and consequences for optimal dosing for gram-negative infections: an updated review. Clin. Pharmacokinet. 61, 1075–1094. doi:[10.1007/s40262-022-01143-0](https://doi.org/10.1007/s40262-022-01143-0)

<span id="page-12-21"></span>Hoover, R., Hunt, T., Benedict, M., Paulson, S. K., Lawrence, L., Cammarata, S., et al. (2016). Single and multiple ascending-dose studies of oral delafloxacin: effects of food, sex, and age. Clin. Ther. 38, 39–52. doi[:10.1016/j.clinthera.2015.10.016](https://doi.org/10.1016/j.clinthera.2015.10.016)

<span id="page-12-33"></span>Hoy, S. M. (2021). Contezolid: first approval. Drugs 81, 1587–1591. doi:[10.1007/](https://doi.org/10.1007/s40265-021-01576-0) [s40265-021-01576-0](https://doi.org/10.1007/s40265-021-01576-0)

<span id="page-12-37"></span>Hung, T. M., Jayanti, R. P., Lee, H. Y., Kim, H. J., Mok, J., Jang, T. W., et al. (2023). Recommendation of pharmacokinetics/pharmacodynamics target of ethambutol to suppress tuberculosis resistance: a population pharmacokinetics study on a large prospective cohort. Int. J. Antimicrob. Agents 62, 106840. doi:[10.1016/j.ijantimicag.](https://doi.org/10.1016/j.ijantimicag.2023.106840) [2023.106840](https://doi.org/10.1016/j.ijantimicag.2023.106840)

<span id="page-12-32"></span>Iqbal, K., Milioudi, A., and Wicha, S. G. (2022). Pharmacokinetics and pharmacodynamics of tedizolid. Clin. Pharmacokinet. 61, 489–503. doi:[10.1007/](https://doi.org/10.1007/s40262-021-01099-7) [s40262-021-01099-7](https://doi.org/10.1007/s40262-021-01099-7)

<span id="page-12-18"></span>Jb, F. (1983). Pharmacocinetique de Ia minocycline chez I'homme. Gaz. Medicale Fr., 45–52.

<span id="page-12-15"></span>Johnston, C., Hilmer, S. N., McLachlan, A. J., Matthews, S. T., Carroll, P. R., and Kirkpatrick, C. M. (2014). The impact of frailty on pharmacokinetics in older people: using gentamicin population pharmacokinetic modeling to investigate changes in renal drug clearance by glomerular filtration. Eur. J. Clin. Pharmacol. 70, 549–555. doi[:10.](https://doi.org/10.1007/s00228-014-1652-7) [1007/s00228-014-1652-7](https://doi.org/10.1007/s00228-014-1652-7)

<span id="page-12-20"></span>Kang, Y., Li, Y., Xu, F., Zhang, J., Wang, K., Chen, Y., et al. (2019). Population pharmacokinetics study of nemonoxacin among Chinese patients with moderate hepatic impairment. Clin. Ther. 41, 505–517. doi[:10.1016/j.clinthera.2019.01.015](https://doi.org/10.1016/j.clinthera.2019.01.015)

<span id="page-12-5"></span>Kennerfalk, A., Ruigómez, A., Wallander, M. A., Wilhelmsen, L., and Johansson, S. (2002). Geriatric drug therapy and healthcare utilization in the United Kingdom. Ann. Pharmacother. 36, 797–803. doi[:10.1345/aph.1A226](https://doi.org/10.1345/aph.1A226)

<span id="page-12-7"></span>Khan, M. S., and Roberts, M. S. (2018). Challenges and innovations of drug delivery in older age. Adv. Drug Deliv. Rev. 135, 3–38. doi:[10.1016/j.addr.2018.09.003](https://doi.org/10.1016/j.addr.2018.09.003)

<span id="page-12-42"></span>Kim, M. C., and Lee, Y. J. (2022). Analysis of time-dependent pharmacokinetics using in vitro-in vivo extrapolation and physiologically based pharmacokinetic modeling. Pharmaceutics 14, 2562. doi:[10.3390/pharmaceutics14122562](https://doi.org/10.3390/pharmaceutics14122562)

<span id="page-12-13"></span>Kim, S. W., Choe, S., Kim, D. J., Zang, D. Y., and Lee, D. H. (2018). Pharmacokinetics of doripenem in healthy Koreans and Monte Carlo simulations to explore optimal dosage regimens in patients with normal and enhanced renal function. Ther. Drug Monit. 40, 425–434. doi[:10.1097/FTD.0000000000000528](https://doi.org/10.1097/FTD.0000000000000528)

<span id="page-12-3"></span>Klotz, U. (2009). Pharmacokinetics and drug metabolism in the elderly. Drug Metab. Rev. 41, 67–76. doi[:10.1080/03602530902722679](https://doi.org/10.1080/03602530902722679)

<span id="page-12-8"></span>Koren, G., Nordon, G., Radinsky, K., and Shalev, V. (2019). Clinical pharmacology of old age. Expert Rev. Clin. Pharmacol. 12, 749–755. doi[:10.1080/17512433.2019.1632188](https://doi.org/10.1080/17512433.2019.1632188)

<span id="page-12-46"></span>Kosugi, Y., and Hosea, N. (2021). Prediction of oral pharmacokinetics using a combination of in silico descriptors and in vitro ADME properties. Mol. Pharm. 18, 1071–1079. doi[:10.1021/acs.molpharmaceut.0c01009](https://doi.org/10.1021/acs.molpharmaceut.0c01009)

<span id="page-12-0"></span>Leaving No One Behind In An Ageing World (2023). World social Report 2023. world health organization. Available at: [https://www.un.org/development/desa/dspd/wp](https://www.un.org/development/desa/dspd/wp-content/uploads/sites/22/2023/01/2023wsr-fullreport.pdf)[content/uploads/sites/22/2023/01/2023wsr-fullreport.pdf](https://www.un.org/development/desa/dspd/wp-content/uploads/sites/22/2023/01/2023wsr-fullreport.pdf) (Accessed April 4, 2023).

<span id="page-12-11"></span>Lebel, M., and Bergeron, M. G. (1987). Pharmacokinetics in the elderly. Studies on ciprofloxacin. Am. J. Med. 82, 108–114.

<span id="page-12-12"></span>Lee, J., Ji, S. C., Kim, B., Yi, S., Shin, K. H., Cho, J. Y., et al. (2017). Exploration of biomarkers for amoxicillin/clavulanate-induced liver injury: multi-omics approaches. Clin. Transl. Sci. 10, 163–171. doi[:10.1111/cts.12425](https://doi.org/10.1111/cts.12425)

<span id="page-13-5"></span>Lee, S. W., Choi, Y., Jang, K., Yoon, S. H., and Chung, J. Y. (2020). Multiple-dose pharmacokinetics and safety of amoxicillin/clavulanate in healthy elderly subjects. Int. J. Clin. Pharmacol. Ther. 58, 634–641. doi[:10.5414/CP203773](https://doi.org/10.5414/CP203773)

<span id="page-13-34"></span>Li, G., Li, Q., Zhang, C., Yu, Q., Li, Q., Zhou, X., et al. (2023). The impact of gene polymorphism and hepatic insufficiency on voriconazole dose adjustment in invasive fungal infection individuals. Front. Genet. 14, 1242711. doi:[10.3389/fgene.2023.1242711](https://doi.org/10.3389/fgene.2023.1242711)

<span id="page-13-6"></span>Li, J., Lovern, M., Green, M. L., Chiu, J., Zhou, D., Comisar, C., et al. (2019a). Ceftazidime-avibactam population pharmacokinetic modeling and pharmacodynamic target attainment across adult indications and patient subgroups. Clin. Transl. Sci. 12, 151–163. doi:[10.1111/cts.12585](https://doi.org/10.1111/cts.12585)

<span id="page-13-20"></span>Li, L., Wu, H., Chen, Y., Yuan, H., Wu, J., Wu, X., et al. (2020). Population pharmacokinetics study of contezolid (MRX-I), a novel oxazolidinone antibacterial agent, in Chinese patients. Clin. Ther. 42, 818–829. doi[:10.1016/j.clinthera.2020.03.020](https://doi.org/10.1016/j.clinthera.2020.03.020)

<span id="page-13-32"></span>Li, X., Zoller, M., Fuhr, U., Huseyn-Zada, M., Maier, B., Vogeser, M., et al. (2019b). Ciprofloxacin in critically ill subjects: considering hepatic function, age and sex to choose the optimal dose. J. Antimicrob. Chemother. 74, 682–690. doi:[10.1093/jac/](https://doi.org/10.1093/jac/dky485) [dky485](https://doi.org/10.1093/jac/dky485)

<span id="page-13-10"></span>Liu, Y., Li, Z., Yang, C., Zheng, H., Lv, Y., Chen, H., et al. (2013). Tolerability and pharmacokinetics of biapenem following single and multiple intravenous administrations in healthy Chinese subjects: an open-label, randomized, singlecenter study. Drug Res. (Stuttg) 63, 396–403. doi[:10.1055/s-0033-1341498](https://doi.org/10.1055/s-0033-1341498)

<span id="page-13-17"></span>Ljungberg, B., and Nilsson-Ehle, I. (1989). Pharmacokinetics of ciprofloxacin in the elderly: increased oral bioavailability and reduced renal clearance. Eur. J. Clin. Microbiol. Infect. Dis. 8, 515–520. doi:[10.1007/BF01967470](https://doi.org/10.1007/BF01967470)

<span id="page-13-9"></span>Ljungberg, B., and Nilsson-Ehle, I. (1992). Pharmacokinetics of meropenem and its metabolite in young and elderly healthy men. Antimicrob. Agents Chemother. 36, 1437–1440. doi[:10.1128/aac.36.7.1437](https://doi.org/10.1128/aac.36.7.1437)

<span id="page-13-21"></span>Manchandani, P., Thamlikitkul, V., Dubrovskaya, Y., Babic, J. T., Lye, D. C., Lee, L. S., et al. (2018). Population pharmacokinetics of polymyxin B. Clin. Pharmacol. Ther. 104, 534–538. doi:[10.1002/cpt.981](https://doi.org/10.1002/cpt.981)

<span id="page-13-24"></span>McLachlan, A. J., and Pont, L. G. (2012). Drug metabolism in older people--a key consideration in achieving optimal outcomes with medicines. J. Gerontol. A Biol. Sci. Med. Sci. 67, 175–180. doi[:10.1093/gerona/glr118](https://doi.org/10.1093/gerona/glr118)

<span id="page-13-23"></span>McLean, A. J., and Le Couteur, D. G. (2004). Aging biology and geriatric clinical pharmacology. Pharmacol. Rev. 56, 163–184. doi[:10.1124/pr.56.2.4](https://doi.org/10.1124/pr.56.2.4)

<span id="page-13-12"></span>Meyers, B. R., Wilkinson, P., Mendelson, M. H., Bournazos, C., Tejero, C., and Hirschman, S. Z. (1993). Pharmacokinetics of aztreonam in healthy elderly and young adult volunteers. J. Clin. Pharmacol. 33, 470–474. doi:[10.1002/j.1552-4604.1993.](https://doi.org/10.1002/j.1552-4604.1993.tb04690.x) [tb04690.x](https://doi.org/10.1002/j.1552-4604.1993.tb04690.x)

<span id="page-13-4"></span>Meyers, B. R., Wilkinson, P., Mendelson, M. H., Walsh, S., Bournazos, C., and Hirschman, S. Z. (1991). Pharmacokinetics of ampicillin-sulbactam in healthy elderly and young volunteers. Antimicrob. Agents Chemother. 35, 2098–2101. doi[:10.1128/aac.](https://doi.org/10.1128/aac.35.10.2098) [35.10.2098](https://doi.org/10.1128/aac.35.10.2098)

<span id="page-13-7"></span>Mingji, W., Caiyun, Z., Huiming, Q., Yuan, L., Pu, Z., Ming, Z., et al. (2007). Pharmacokinetics of ampicillin-sulbactam in healthy elderly and young volunteers. Antimicrob. Agents Chemother. (105), 28–32.

<span id="page-13-15"></span>Molinaro, M., Villani, P., Regazzi, M. B., Rondanelli, R., and Doveri, G. (1992). Pharmacokinetics of ofloxacin in elderly patients and in healthy young subjects. Eur. J. Clin. Pharmacol. 43, 105–107. doi:[10.1007/BF02280765](https://doi.org/10.1007/BF02280765)

<span id="page-13-30"></span>Mühlberg, W., and Platt, D. (1999). Age-dependent changes of the kidneys: pharmacological implications. Gerontology 45, 243–253. doi[:10.1159/000022097](https://doi.org/10.1159/000022097)

<span id="page-13-14"></span>Muralidharan, G., Fruncillo, R. J., Micalizzi, M., Raible, D. G., and Troy, S. M. (2005). Effects of age and sex on single-dose pharmacokinetics of tigecycline in healthy subjects. Antimicrob. Agents Chemother. 49, 1656–1659. doi:[10.1128/AAC.49.4.1656-1659.2005](https://doi.org/10.1128/AAC.49.4.1656-1659.2005)

<span id="page-13-8"></span>Musson, D. G., Majumdar, A., Holland, S., Birk, K., Xi, L., Mistry, G., et al. (2004). Pharmacokinetics of total and unbound ertapenem in healthy elderly subjects. Antimicrob. Agents Chemother. 48, 521–524. doi:[10.1128/aac.48.2.521-524.2004](https://doi.org/10.1128/aac.48.2.521-524.2004)

<span id="page-13-11"></span>Namkoong, H., Kameyama, Y., Yasuda, H., Nakayama, S., Kaneko, H., Kawashima, C., et al. (2014). The efficacy, safety, and pharmacokinetics of biapenem administered thrice daily for the treatment of pneumonia in the elderly. J. Infect. Chemother. 20, 356–360. doi:[10.1016/j.jiac.2013.12.010](https://doi.org/10.1016/j.jiac.2013.12.010)

<span id="page-13-26"></span>Nedelman, J., Cramer, J. A., Robbins, B., Gibiansky, E., Chang, C. T., Gareffa, S., et al. (1997). The effect of food on the pharmacokinetics of multiple-dose terbinafine in young and elderly healthy subjects. Biopharm. Drug Dispos. 18, 127–138. doi:[10.1002/](https://doi.org/10.1002/(sici)1099-081x(199703)18:2<127::aid-bdd6>3.0.co;2-8) [\(sici\)1099-081x\(199703\)18:2](https://doi.org/10.1002/(sici)1099-081x(199703)18:2<127::aid-bdd6>3.0.co;2-8)<127::aid-bdd6>3.0.co;2-8

<span id="page-13-36"></span>Nestorov, I. (2003). Whole body pharmacokinetic models. Clin. Pharmacokinet. 42, 883–908. doi:[10.2165/00003088-200342100-00002](https://doi.org/10.2165/00003088-200342100-00002)

<span id="page-13-31"></span>Nicolle, L. E. (1999). Quinolones in the aged. Drugs 58 (Suppl. 2), 49–51. doi:[10.2165/](https://doi.org/10.2165/00003495-199958002-00009) [00003495-199958002-00009](https://doi.org/10.2165/00003495-199958002-00009)

<span id="page-13-18"></span>Ohsaki, Y., Morita, K., Takeda, H., Kishino, S., Okumura, S., and Fujiuchi, S. (2010). Pharmacokinetics of garenoxacin in elderly patients with respiratory tract infections. Int. J. Antimicrob. Agents 35, 603–605. doi[:10.1016/j.ijantimicag.2010.01.020](https://doi.org/10.1016/j.ijantimicag.2010.01.020)

<span id="page-13-2"></span>Orr, W. C., and Chen, C. L. (2002). Aging and neural control of the GI tract: IV. Clinical and physiological aspects of gastrointestinal motility and aging. Am. J. Physiol. Gastrointest. Liver Physiol. 283, G1226–G1231. doi[:10.1152/ajpgi.00276.2002](https://doi.org/10.1152/ajpgi.00276.2002)

<span id="page-13-46"></span>Ota, R., and Yamashita, F. (2022). Application of machine learning techniques to the analysis and prediction of drug pharmacokinetics. J. Control Release 352, 961–969. doi[:10.1016/j.jconrel.2022.11.014](https://doi.org/10.1016/j.jconrel.2022.11.014)

<span id="page-13-33"></span>Peloquin, C. A., Jaresko, G. S., Yong, C. L., Keung, A. C., Bulpitt, A. E., and Jelliffe, R. W. (1997). Population pharmacokinetic modeling of isoniazid, rifampin, and pyrazinamide. Antimicrob. Agents Chemother. 41, 2670–2679. doi:[10.1128/AAC.41.](https://doi.org/10.1128/AAC.41.12.2670) [12.2670](https://doi.org/10.1128/AAC.41.12.2670)

<span id="page-13-13"></span>Perret, C., Lenfant, B., Weinling, E., Wessels, D. H., Scholtz, H. E., Montay, G., et al. (2002). Pharmacokinetics and absolute oral bioavailability of an 800-mg oral dose of telithromycin in healthy young and elderly volunteers. Chemotherapy 48, 217–223. doi[:10.1159/000066766](https://doi.org/10.1159/000066766)

<span id="page-13-44"></span>Polasek, T. M., Patel, F., Jensen, B. P., Sorich, M. J., Wiese, M. D., and Doogue, M. P. (2013). Predicted metabolic drug clearance with increasing adult age. Br. J. Clin. Pharmacol. 75, 1019–1028. doi:[10.1111/j.1365-2125.2012.04446.x](https://doi.org/10.1111/j.1365-2125.2012.04446.x)

<span id="page-13-43"></span>Polasek, T. M., Sadagopal, J. S., Elliot, D. J., and Miners, J. O. (2010). In vitro-in vivo extrapolation of zolpidem as a perpetrator of metabolic interactions involving CYP3A. Eur. J. Clin. Pharmacol. 66, 275–283. doi:[10.1007/s00228-009-0760-2](https://doi.org/10.1007/s00228-009-0760-2)

<span id="page-13-45"></span>Popel, M., Tomkova, M., Tomek, J., Kaiser, Ł., Uszkoreit, J., Bojar, O., et al. (2020). Transforming machine translation: a deep learning system reaches news translation quality comparable to human professionals. Nat. Commun. 11, 4381. doi:[10.1038/](https://doi.org/10.1038/s41467-020-18073-9) [s41467-020-18073-9](https://doi.org/10.1038/s41467-020-18073-9)

<span id="page-13-28"></span>Qu, Y., Su, C., Xiang, Z., Wang, Y., Han, J., Pan, J., et al. (2023). Population pharmacokinetic modeling and simulation for nirmatrelvir exposure assessment in Chinese older patients with COVID-19 infection. Eur. J. Pharm. Sci. 189, 106535. doi[:10.](https://doi.org/10.1016/j.ejps.2023.106535) [1016/j.ejps.2023.106535](https://doi.org/10.1016/j.ejps.2023.106535)

<span id="page-13-22"></span>Rosina, R., Villa, G., Danese, A., Cavenaghi, L., Picardi, L., and Salvadeo, A. (1988). Pharmacokinetics of teicoplanin in the elderly. J. Antimicrob. Chemother. 21 (Suppl. A), 39–45. doi:[10.1093/jac/21.suppl\\_a.39](https://doi.org/10.1093/jac/21.suppl_a.39)

<span id="page-13-41"></span>Rowland, M., and Pang, K. S. (2022). Hepatic clearance models and IVIVE predictions. Clin. Pharmacol. Ther. 111, 1205–1207. doi:[10.1002/cpt.2525](https://doi.org/10.1002/cpt.2525)

<span id="page-13-0"></span>Rudd, K. E., Johnson, S. C., Agesa, K. M., Shackelford, K. A., Tsoi, D., Kievlan, D. R., et al. (2020). Global, regional, and national sepsis incidence and mortality, 1990-2017: analysis for the Global Burden of Disease Study. Lancet 395, 200–211. doi:[10.1016/](https://doi.org/10.1016/S0140-6736(19)32989-7) [S0140-6736\(19\)32989-7](https://doi.org/10.1016/S0140-6736(19)32989-7)

<span id="page-13-37"></span>Sager, J. E., Yu, J., Ragueneau-Majlessi, I., and Isoherranen, N. (2015). Physiologically based pharmacokinetic (PBPK) modeling and simulation approaches: a systematic review of published models, applications, and model verification. Drug Metab. Dispos. 43, 1823–1837. doi:[10.1124/dmd.115.065920](https://doi.org/10.1124/dmd.115.065920)

<span id="page-13-25"></span>Sansone-Parsons, A., Krishna, G., Simon, J., Soni, P., Kantesaria, B., Herron, J., et al. (2007). Effects of age, gender, and race/ethnicity on the pharmacokinetics of posaconazole in healthy volunteers. Antimicrob. Agents Chemother. 51, 495–502. doi[:10.1128/AAC.00472-06](https://doi.org/10.1128/AAC.00472-06)

<span id="page-13-35"></span>Schlender, J. F., Meyer, M., Thelen, K., Krauss, M., Willmann, S., Eissing, T., et al. (2016). Development of a whole-body physiologically based pharmacokinetic approach to assess the pharmacokinetics of drugs in elderly individuals. Clin. Pharmacokinet. 55, 1573–1589. doi[:10.1007/s40262-016-0422-3](https://doi.org/10.1007/s40262-016-0422-3)

<span id="page-13-38"></span>Schlender, J. F., Teutonico, D., Coboeken, K., Schnizler, K., Eissing, T., Willmann, S., et al. (2018). A physiologically-based pharmacokinetic model to describe ciprofloxacin pharmacokinetics over the entire span of life. Clin. Pharmacokinet. 57, 1613–1634. doi[:10.1007/s40262-018-0661-6](https://doi.org/10.1007/s40262-018-0661-6)

<span id="page-13-16"></span>Setiawan, E., Abdul-Aziz, M. H., Cotta, M. O., Susaniwati, S., Cahjono, H., Sari, I. Y., et al. (2022). Population pharmacokinetics and dose optimization of intravenous levofloxacin in hospitalized adult patients. Sci. Rep. 12, 8930. doi:[10.1038/s41598-](https://doi.org/10.1038/s41598-022-12627-1) [022-12627-1](https://doi.org/10.1038/s41598-022-12627-1)

<span id="page-13-40"></span>Setiawan, E., Cotta, M. O., Abdul-Aziz, M. H., Widjanarko, D., Sosilya, H., Lukas, D. L., et al. (2023). Population pharmacokinetics and dosing simulations of ampicillin and sulbactam in hospitalised adult patients. Clin. Pharmacokinet. 62, 573–586. doi:[10.1007/](https://doi.org/10.1007/s40262-023-01219-5) [s40262-023-01219-5](https://doi.org/10.1007/s40262-023-01219-5)

<span id="page-13-39"></span>Sheiner, L. B., and Ludden, T. M. (1992). Population pharmacokinetics/dynamics. Annu. Rev. Pharmacol. Toxicol. 32, 185–209. doi[:10.1146/annurev.pa.32.040192.001153](https://doi.org/10.1146/annurev.pa.32.040192.001153)

<span id="page-13-3"></span>Shi, S., and Klotz, U. (2011). Age-related changes in pharmacokinetics. Curr. Drug Metab. 12, 601–610. doi:[10.2174/138920011796504527](https://doi.org/10.2174/138920011796504527)

<span id="page-13-29"></span>Silva, F. G. (2005). The aging kidney: a review--part II. Int. Urol. Nephrol. 37, 419–432. doi[:10.1007/s11255-004-0874-5](https://doi.org/10.1007/s11255-004-0874-5)

<span id="page-13-19"></span>Sisson, T. L., Jungbluth, G. L., and Hopkins, N. K. (2002). Age and sex effects on the pharmacokinetics of linezolid. Eur. J. Clin. Pharmacol. 57, 793–797. doi:[10.1007/](https://doi.org/10.1007/s00228-001-0380-y) [s00228-001-0380-y](https://doi.org/10.1007/s00228-001-0380-y)

<span id="page-13-42"></span>Sodhi, J. K., and Benet, L. Z. (2021). Successful and unsuccessful prediction of human hepatic clearance for lead optimization. J. Med. Chem. 64, 3546–3559. doi:[10.1021/acs.](https://doi.org/10.1021/acs.jmedchem.0c01930) imedchem.0c01930

<span id="page-13-1"></span>Soraci, L., Cherubini, A., Paoletti, L., Filippelli, G., Luciani, F., Laganà, P., et al. (2023). Safety and tolerability of antimicrobial agents in the older patient. Drugs Aging 40, 499–526. doi:[10.1007/s40266-023-01019-3](https://doi.org/10.1007/s40266-023-01019-3)

<span id="page-13-27"></span>Stader, F., Courlet, P., Decosterd, L. A., Battegay, M., and Marzolini, C. (2021). Physiologically-based pharmacokinetic modeling combined with Swiss HIV cohort

study data supports No dose adjustment of bictegravir in elderly individuals living with HIV. Clin. Pharmacol. Ther. 109, 1025–1029. doi:[10.1002/cpt.2178](https://doi.org/10.1002/cpt.2178)

<span id="page-14-16"></span>Theuretzbacher, U., Ihle, F., and Derendorf, H. (2006). Pharmacokinetic/ pharmacodynamic profile of voriconazole. Clin. Pharmacokinet. 45, 649–663. doi[:10.](https://doi.org/10.2165/00003088-200645070-00002) [2165/00003088-200645070-00002](https://doi.org/10.2165/00003088-200645070-00002)

<span id="page-14-14"></span>Thompson, G. A., Smithers, J. A., Kenny, M. T., Dulworth, J. K., Kulmala, H. K., Yuh, L., et al. (1992). Pharmacokinetics of teicoplanin upon multiple dose intravenous administration to normal healthy male volunteers. Biopharm. Drug Dispos. 13, 213–220. doi[:10.1002/bdd.2510130307](https://doi.org/10.1002/bdd.2510130307)

<span id="page-14-1"></span>Thomson, N. M. (1995). Drugs and the kidney in the elderly. Med. J. Aust. 162, 543–547. doi:[10.5694/j.1326-5377.1995.tb138518.x](https://doi.org/10.5694/j.1326-5377.1995.tb138518.x)

<span id="page-14-3"></span>Tiihonen, K., Ouwehand, A. C., and Rautonen, N. (2010). Human intestinal microbiota and healthy ageing. Ageing Res. Rev. 9, 107–116. doi[:10.1016/j.arr.2009.10.004](https://doi.org/10.1016/j.arr.2009.10.004)

<span id="page-14-11"></span>Totsuka, K., Sesoko, S., Fukase, H., Ikushima, I., Odajima, M., and Niwayama, Y. (2020). Pharmacokinetic study of lascufloxacin in non-elderly healthy men and elderly men. J. Infect. Chemother. 26, 231–239. doi[:10.1016/j.jiac.2019.09.010](https://doi.org/10.1016/j.jiac.2019.09.010)

<span id="page-14-4"></span>Udy, A. A., Roberts, J. A., and Lipman, J. (2013). Clinical implications of antibiotic pharmacokinetic principles in the critically ill. Intensive Care Med. 39, 2070–2082. doi[:10.1007/s00134-013-3088-4](https://doi.org/10.1007/s00134-013-3088-4)

<span id="page-14-21"></span>Uhart, M., Leroy, B., Michaud, A., Maire, P., and Bourguignon, L. (2013). Interindividual and intra-individual pharmacokinetic variability during teicoplanin therapy in geriatric patients. Med. Mal. Infect. 43, 295–298. doi:[10.1016/j.medmal.2013.05.008](https://doi.org/10.1016/j.medmal.2013.05.008)

<span id="page-14-18"></span>Usman, M., Frey, O. R., and Hempel, G. (2017). Population pharmacokinetics of meropenem in elderly patients: dosing simulations based on renal function. Eur. J. Clin. Pharmacol. 73, 333–342. doi:[10.1007/s00228-016-2172-4](https://doi.org/10.1007/s00228-016-2172-4)

<span id="page-14-9"></span>Van Wart, S., Phillips, L., Ludwig, E. A., Russo, R., Gajjar, D. A., Bello, A., et al. (2004). Population pharmacokinetics and pharmacodynamics of garenoxacin in patients with community-acquired respiratory tract infections. Antimicrob. Agents Chemother. 48, 4766–4777. doi[:10.1128/AAC.48.12.4766-4777.2004](https://doi.org/10.1128/AAC.48.12.4766-4777.2004)

<span id="page-14-6"></span>Varanese, L. (1993). Pharmacokinetics and safety of dirithromycin in the elderly. J. Antimicrob. Chemother. 31 (Suppl. C), 169–174. doi[:10.1093/jac/31.suppl\\_c.169](https://doi.org/10.1093/jac/31.suppl_c.169)

<span id="page-14-2"></span>Vinarov, Z., Abdallah, M., Agundez, J. A. G., Allegaert, K., Basit, A. W., Braeckmans, M., et al. (2021). Impact of gastrointestinal tract variability on oral drug absorption and pharmacokinetics:<br>an UNGAP review. *Eur. J. Pharm. Sci.* 162, 105812. doi[:10.1016/j.ejps.2021.105812](https://doi.org/10.1016/j.ejps.2021.105812)

<span id="page-14-12"></span>Wang, P., Liu, D., Sun, T., Zhang, X., and Yang, J. (2022a). Pharmacokinetics and pharmacodynamics of polymyxin B and proposed dosing regimens in elderly patients<br>with multi-drug-resistant Gram-negative bacterial infections. I*nt. J. Antimicrob. Agents* 60, 106693. doi:[10.1016/j.ijantimicag.2022.106693](https://doi.org/10.1016/j.ijantimicag.2022.106693)

<span id="page-14-22"></span>Wang, P., Liu, D., Sun, T., Zhang, X., and Yang, J. (2022b). Pharmacokinetics and pharmacodynamics of polymyxin B and proposed dosing regimens in elderly patients with multi-drug-resistant Gram-negative bacterial infections. Int. J. Antimicrob. Agents 60, 106693. doi:[10.1016/j.ijantimicag.2022.106693](https://doi.org/10.1016/j.ijantimicag.2022.106693)

<span id="page-14-20"></span>Wang, P., Zhang, Q., Zhu, Z., Pei, H., Feng, M., Sun, T., et al. (2021). Comparing the population pharmacokinetics of and acute kidney injury due to polymyxin B in Chinese patients with or without renal insufficiency. Antimicrob. Agents Chemother. 65, e01900. doi[:10.1128/AAC.01900-20](https://doi.org/10.1128/AAC.01900-20)

<span id="page-14-17"></span>Weng, F. H. (1990). Comparative study on the pharmacokinetics of rifampicin between the old and middle-aged tuberculosis patients. Zhonghua Jie He He Hu Xi Za Zhi 13, 292–294.

<span id="page-14-15"></span>Wong, S. L., Barriere, S. L., Kitt, M. M., and Goldberg, M. R. (2008). Multiple-dose pharmacokinetics of intravenous telavancin in healthy male and female subjects. J. Antimicrob. Chemother. 62, 780–783. doi[:10.1093/jac/dkn273](https://doi.org/10.1093/jac/dkn273)

<span id="page-14-0"></span>World Health Statistics (2023). World health statistics 2023: monitoring health for the SDGs, sustainable development goals. world health organization. Available at: [https://](https://www.who.int/publications/i/item/9789240074323) [www.who.int/publications/i/item/9789240074323](https://www.who.int/publications/i/item/9789240074323) (Accessed April 4, 2023).

<span id="page-14-19"></span>Wu, J., Cao, G., Wu, H., Chen, Y., Guo, B., Wu, X., et al. (2020). Evaluation of the effect of contezolid (MRX-I) on the corrected QT interval in a randomized, doubleblind, placebo- and positive-controlled crossover study in healthy Chinese volunteers. Antimicrob. Agents Chemother. 64, e02158. doi:[10.1128/AAC.02158-19](https://doi.org/10.1128/AAC.02158-19)

<span id="page-14-10"></span>Wu, X. J., Zhang, J., Guo, B. N., Zhang, Y. Y., Yu, J. C., Cao, G. Y., et al. (2015). Pharmacokinetics and pharmacodynamics of multiple-dose intravenous nemonoxacin in healthy Chinese volunteers. Antimicrob. Agents Chemother. 59, 1446–1454. doi[:10.](https://doi.org/10.1128/AAC.04039-14) [1128/AAC.04039-14](https://doi.org/10.1128/AAC.04039-14)

<span id="page-14-7"></span>Yamamoto, T., Takano, K., Matsuyama, N., Koike, Y., Minshita, S., Sanaka, M., et al. (1999). Pharmacokinetic characteristics of minocycline in debilitated elderly patients. Am. J. Ther. 6, 157–160. doi:[10.1097/00045391-199905000-00006](https://doi.org/10.1097/00045391-199905000-00006)

<span id="page-14-5"></span>Yaoguo, S., Yingyuan, Z., Jing, Z., Ziping, W., Jicheng, Y., and Fu, W. (2001). Pharmacokinetics of antimicrobial agents in the elderly. Chin. J. Infect. Chemother., 3–6.

<span id="page-14-8"></span>Yaozh (2023). Instructions for sitafloxacin tablets. Available at: [https://db.yaozh.com/](https://db.yaozh.com/instruct/60329303.html) [instruct/60329303.html](https://db.yaozh.com/instruct/60329303.html) (Accessed April 4, 2023).

<span id="page-14-13"></span>Zhang, J., Zhang, Y., Shi, Y., Rui, J., Yu, J., Cao, G., et al. (2008). Population pharmacokinetic and pharmacodynamic modeling of norvancomycin. Eur. J. Clin. Microbiol. Infect. Dis. 27, 275–284. doi[:10.1007/s10096-007-0435-9](https://doi.org/10.1007/s10096-007-0435-9)

<span id="page-14-23"></span>Zhou, B., Lapedriza, A., Khosla, A., Oliva, A., and Torralba, A. (2018). Places: a 10 million image database for scene recognition. IEEE Trans. Pattern Anal. Mach. Intell. 40, 1452–1464. doi:[10.1109/TPAMI.2017.2723009](https://doi.org/10.1109/TPAMI.2017.2723009)