Check for updates

OPEN ACCESS

EDITED BY Ruiwen Zhang, University of Houston, United States

REVIEWED BY Wei Zhong, University of Kansas Medical Center, United States Demin Cai, Yangzhou University, China Palash Mandal, Charotar University of Science and Technology, India Saritha Gopal Pandit, Central Food Technological Research Institute (CSIR), India

*CORRESPONDENCE Zhi Li, Izhi_swmu@126.com Xin Zhou, Zhouxinmm@swmu.edu.cn Hui Chen, Safairysusan@126.com

¹These authors contributed equally to this work and share first authorship

RECEIVED 13 November 2023 ACCEPTED 18 March 2024 PUBLISHED 08 April 2024

CITATION

He Y-X, Liu M-N, Wang Y-Y, Wu H, Wei M, Xue J-Y, Zou Y, Zhou X, Chen H and Li Z (2024), Hovenia dulcis: a Chinese medicine that plays an essential role in alcohol-associated liver disease. *Front. Pharmacol.* 15:1337633. doi: 10.3389/fphar.2024.1337633

COPYRIGHT

© 2024 He, Liu, Wang, Wu, Wei, Xue, Zou, Zhou, Chen and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Hovenia dulcis: a Chinese medicine that plays an essential role in alcohol-associated liver disease

Yi-Xiang He^{1,2†}, Meng-Nan Liu^{2†}, Yang-Yang Wang^{1,2†}, Hao Wu², Mei Wei^{1,2}, Jin-Yi Xue², Yuan Zou², Xin Zhou^{1,2,3}*, Hui Chen^{1,2,3}* and Zhi Li^{1,2,3}*

¹The Key Laboratory of Integrated Traditional Chinese and Western Medicine for Prevention and Treatment of Digestive System Diseases of Luzhou City, Affiliated Traditional Medicine Hospital of Southwest Medical University, Luzhou, Sichuan, China, ²College of Integrated Chinese and Western Medicine, Southwest Medical University, Luzhou, Sichuan, China, ³Department of Spleen and Stomach Diseases, The Affiliated Traditional Chinese Medicine Hospital of Southwest Medical University, Luzhou, Sichuan, China

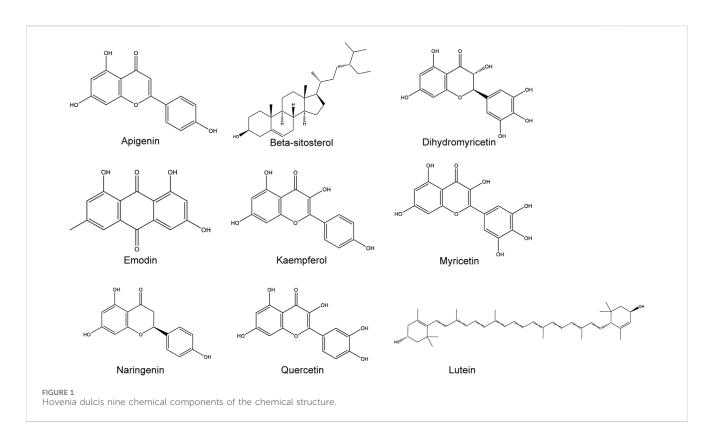
Globally, alcohol-associated liver disease (ALD) has become an increased burden for society. Disulfirams, Benzodiazepines (BZDs), and corticosteroids are commonly used to treat ALD. However, the occurrence of side effects such as hepatotoxicity and dependence, impedes the achievement of desirable and optimal therapeutic efficacy. Therefore, there is an urgent need for more effective and safer treatments. Hovenia dulcis is an herbal medicine promoting alcohol removal clearance, lipid-lowering, anti-inflammatory, and hepatoprotective properties. Hovenia dulcis has a variety of chemical components such as dihydromyricetin, quercetin and beta-sitosterol, which can affect ALD through multiple pathways, including ethanol metabolism, immune response, hepatic fibrosis, oxidative stress, autophagy, lipid metabolism, and intestinal barrier, suggesting its promising role in the treatment of ALD. Thus, this work aims to comprehensively review the chemical composition of Hovenia dulcis and the molecular mechanisms involved in the process of ALD treatment.

KEYWORDS

alcohol-associated liver disease, Hovenia dulcis, mechanisms of action, Chinese herbal, treatment of alcohol-associated liver disease

1 Introduction

Alcohol-associated liver disease (ALD) is strongly associated with chronic excessive alcohol consumption (Liu S. Y. et al., 2021). The World Health Organization (WHO) (Jiaming Chen, 2022) has estimated that approximately 2.3 billion individuals worldwide are currently engaged in alcohol consumption, with a significant proportion found in the United States, Europe, and the Western Pacific. This global trend has significant implications for the progression and outcomes of more than 200 diseases (Liu S. Y. et al., 2021). Moreover, it contributes to increased morbidity, mortality, and financial burden on society, resulting in over 3 million deaths annually (Zhao et al., 2018). The prevalence of ALD in China has significantly risen, making it the second country in terms of high liver disease prevalence after viral hepatitis (Liu et al., 2010). Regional epidemiological studies have indicated that the prevalence of ALD varies from 0.50% to 8.55% in different



regions (Tang et al., 2023). Due to the escalating prevalence and severe implications of ALD, timely intervention and treatment of ALD are crucial for improving its prognosis.

Hovenia dulcis Thunb. (Rhamnaceae) (Hovenia dulcis) (botanical name approved by http://mpns.kew.org) is a kind of traditional Chinese medicine (TCM) widely used in China, Korea, and Japan, and is also used as ingredients for food supplements and nutraceuticals (Sferrazza et al., 2021). In the Materia Medica Compendium (A.D. 1,578), Hovenia dulcis is listed as a TCM with anti-alcoholic properties (Fangfang Xu and Wang, 2011; Liang and Olsen, 2014). In ancient Chinese medicine, its fruits and pedicels were also used as a febrifuge and administered for parasitic infections, antispasmodic, laxative and diuretic. The seeds were used as a diuretic and were also effective in alcoholism (Patricia et al., 2017) Hovenia dulcis is found in open fields and forests in China at an altitude of less than 2,100 m and is classified as a plant used for both food and medicine (Fangfang Xu and Wang, 2011; Linzhi Kang and Li, 2022). Its chemical components include dihydromyricetin (DHM), quercetin, naringenin, etc., which have been shown to have antioxidant and anti-inflammatory properties and are also considered potential therapeutic agents for alcoholassociated liver disease (Qiu et al., 2019; Meng et al., 2020).

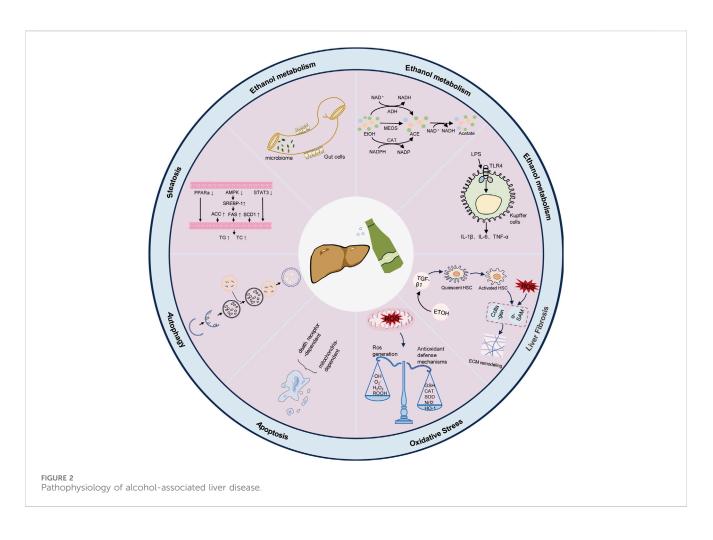
This paper aims to elucidate the fundamental mechanisms underlying ALD and the existing drug treatment strategies, with a specific focus on the impact of Hovenia dulcis's chemical constituents on the pathogenesis of ALD through various pathways and targets. It is our aspiration that this knowledge will contribute to future research endeavors in the field of ALD treatment.

2 Phytochemistry

Research on the chemical composition of Hovenia dulcis began in the late 20th century, and a total of 44 species have been documented on the Herb (http://herb.ac.cn/) and TCMSP platforms, including flavonoids, triterpenoid saponins, and alkaloids (Bao-Jun, 2003; Albano, 2008; Li et al., 2021; Sferrazza et al., 2021; Lv et al., 2023). Flavonoids constitute up to 27.61% as the main bioactive components of Hovenia dulcis, (Fu et al., 2023). Of these, nine chemical constituents such as beta-sitosterol, DHM, quercetin naringenin, lutein, myricetin, kaempferol, emodin, and apigenin, were verified to be related to ALD and their chemical results are shown in Figure 1.

3 Pathophysiology of ALD

ALD is a chronic liver disease that arises from the excessive consumption of alcohol, typically commencing with alcoholic fatty liver disease. Prolonged alcohol intake disrupts the metabolic capacity of hepatocytes, leading to intracellular lipid accumulation and subsequent progression to more severe manifestations of hepatic injury, such as steatohepatitis, liver fibrosis, cirrhosis, and potentially hepatocellular carcinoma (HCC) (Noh et al., 2011; Peng et al., 2013; Liu S. Y. et al., 2021; Liu Y. S. et al., 2021; Wang et al., 2022). As depicted in Figure 2, the pathophysiology of ALD includes ethanol metabolism and mechanisms in ALD development (Dunn and Shah, 2016; Kong et al., 2019).



3.1 Alcohol metabolism

Through the digestive system, alcohol is taken into the bloodstream and affects blood alcohol concentration (BAC), which is mainly processed in hepatocytes (Liu S. Y. et al., 2021). The oxidative conversion of alcohol to acetaldehyde involves three main metabolic processes. The initial step involves the alcohol dehydrogenase (ADH) pathway, facilitated by nicotinamide adenine dinucleotide (NAD) as a cofactor, transforming ethanol into acetaldehyde (Silva et al., 2021). Due to its high reactivity, acetaldehyde can interact with various proteins and form deoxyribonucleic acid (DNA) complexes, thereby influencing protein and genetic material (Shang Zeng and Li, 2022). The second pathway of ethanol metabolism generates reactive oxygen species (ROS) through the microsomal enzyme oxidizing system (MEOS) in the endoplasmic reticulum, leading to various liver reactions, such as oxidative stress and inflammatory damage (Liu S. Y. et al., 2021). The third, more minor pathway is thought to be the catalase (CAT) pathway, which is dependent on nicotinamide adenine dinucleotide phosphate (NADPH) for metabolism (Kong et al., 2019). Subsequently, acetaldehyde is metabolized by the acetaldehyde dehydrogenase enzyme (ALDH) in the mitochondria, leading to the formation of acetate. Acetate is further oxidized to carbon dioxide in various extrahepatic tissues (Chen et al., 2013).

3.2 Mechanisms in ALD development

3.2.1 Alcohol metabolism interferes with the liver's immune response to inflammation

Ethanol triggers unrestrained inflammation and immune turbulence via regulating key cell subsets in the hepatic immune microenvironment. Kupffer cells (KC) (liver-resident macrophages) play a crucial role in the innate immune response against infections and initiating and resolving inflammation (Yao et al., 2017; Chen M. et al., 2023). Chronic alcohol consumption impairs the intestinal mucosal barrier, leading to increase permeability and translocation of intestinal Gram-negative bacteria-derived lipopolysaccharide (LPS) to the liver. As a typical pathogen-associated molecular pattern (PAMP), LPS activates toll-like receptor 4 (TLR4) on macrophages and natural killer cells (NK cells), leading to change macrophage function and increased production of proinflammatory cytokines, which results in T cell activation and inflammation (Ling Yang and Lu, 2022; Fang et al., 2023).

3.2.2 Liver sinusoidal endothelial cell dysfunction (LSEC) and liver fibrosis

LSECs are a highly specialized and distinctive micro-vascular cell type that regulates the hepatic micro-environment (Lafoz et al., 2020). LSEC is the first defense barrier, removing and recycling proteins, lipids, and toxins from the blood through highly permeable

fenestrae when exposed to blood from the intestinal and systemic circulation (Wu et al., 2023). In addition, they are involved in response to specific injuries by regulating neighboring cells, mainly hepatic stellate cells (HSC).

With alcohol consumption, HSCs and hepatic portal fibroblasts transdifferentiate to myofibroblasts. Activated myofibroblasts produce mainly type I and type III collagen, which cross-links and accumulate in the extracellular matrix (ECM), replacing the injured liver parenchyma and forming a fibrous scar (Bellanti et al., 2023). HSCs can be directly activated by alcohol, acetaldehyde, and ROS or in a paracrine manner by various fibrogenic factors (e.g., transforming growth factor- β , platelet-derived growth factor, and inflammatory cytokines) released from damaged hepatocytes, activated Kupffer cells, and infiltrated immune cells (Ha et al., 2022).

3.2.3 Alcohol-induced mitochondrial dysfunction with activated reactive oxygen species (ROS) storm

Mitochondria are double-membrane organelles involved in various physiological functions within the cell (Park et al., 2022). Typically, mitochondrial abnormalities usually occur in the early stages of alcoholic liver injury that continually produce detrimental effects on hepatocytes. When the liver is chronically exposed to ethanol, mitochondria produce large amounts of reactive nitrogen species (RNS) and ROS, which inactivate components of the respiratory chain by destroying mitochondrial DNA (mtDNA), inducing mitochondrial permeability transition, leading to mitochondrial swelling and rupture, and ultimately hepatocyte death (Su et al., 2023). ROS secreted by mitochondria is scavenged by the cellular antioxidant system. Organisms have developed a variety of antioxidant defense systems to scavenge the accumulation of peroxides in the liver to reduce liver injury caused by oxidative stress (Molina et al., 2003).

3.2.4 Alcohol metabolism-mediated cell death and prosurvival pathways

The complex balance between parenchymal and nonparenchymal cells' survival and death pathways is critical for regulating ALD progression. In ALD, programmed cell death (PCD), partly caused by ethanol-induced oxidative stress and innate immune responses, is thought to play a central role in the progression of the injury (Miyata and Nagy, 2020). PCD is a spontaneous and programmed cell death mode associated with multiple physiological and pathological processes (Chen Y. et al., 2023). There are four main modes of PCD: apoptosis, necroptosis, pyroptosis, and ferroptosis (Wu et al., 2023). ALD is influenced by dysregulation or hyperactivation of autophagy, which is linked to cellular death and damage.

3.2.4.1 Apoptosis

Apoptosis is a molecular process that plays an essential role in development and tissue homeostasis (Nössing and Ryan, 2023). Apoptosis is a crucial component of various homeostatic processes, including standard cell renewal and normal development and function of the immune system (Miyata and Nagy, 2020). The mechanisms of apoptosis are complex and are now generally recognized as occurring mainly through the death receptor-dependent extrinsic pathway and the mitochondria-dependent intrinsic pathway (Pistritto et al., 2016). The new guidelines

distinguish between intrinsic caspase-dependent and intrinsic caspase-independent apoptosis (Guicciardi et al., 2013).

3.2.4.2 Ferroptosis

Ethanol feeding results in iron-dependent ferroptotic cell death (FCD), which is characterized by the accumulation of lipid hydroperoxides to lethal levels in the presence of iron (Wu et al., 2023). Ferroptosis is a ROS-dependent cell death with two major biochemical features: iron accumulation and lipid peroxidation (Tang et al., 2021). Excessive oxidative stress and disruption of the redox system cause lipid peroxidation that further leads to mitochondrial dysfunction, which in turn causes FCD. Thus, the central molecular mechanism of iron death is an imbalance between oxidative damage and antioxidant defenses (Chen Y. et al., 2023).

3.2.4.3 Autophagy

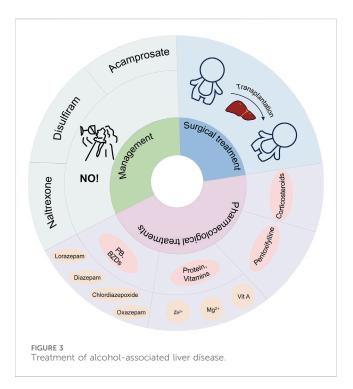
Autophagy is a dynamic process designed to maintain cellular homeostasis by removing lipid droplets, damaged organelles, and faulty proteins from cells to promote cell survival (Dunn and Shah, 2016; Shang Zeng and Li, 2022). There are three main types of autophagy: macro-autophagy, micro-autophagy, and chaperonemediated autophagy (Glick et al., 2010). Macro-autophagy is the main form of cellular autophagy. It has been documented that acute alcohol exposure induces protective autophagy in the liver, whereas chronic alcohol intake impairs autophagy in hepatocytes (Shang Zeng and Li, 2022). Generally, autophagy is considered an early adaptive response to injury that occurs before apoptosis; however, hyperactivation also results in apoptotic cell death through standard regulators, such as beclin1 and Bcl-2 (Wu et al., 2023).

3.2.5 Disorders of lipid metabolism due to ethanol metabolism

In hepatocytes, lipid uptake, esterification, oxidation, and fatty acid secretion occur. These processes are regulated by hormones, nuclear receptors, and transcription factors to maintain hepatic lipid homeostasis (Fu et al., 2021). Alcohol affects hepatic lipid metabolism through synergistic mechanisms, accumulating fatty acids (FA) and triglycerides (Seitz et al., 2023). Ethanol intake decreases signal transducer and activator of transcription 3 (STAT3), AMP-activated protein kinase (AMPK), and peroxisome proliferative activated receptor alpha (PPARa) levels to inhibit fatty acid oxidation. In addition, alcohol intake upregulates sterol regulatory element binding protein-1 expression (SREBP-1) and lipogenic enzymes. All of these dyslipidemia behaviors contribute to and hepatic fat accumulation (Dunn and Shah, 2016; Zhao et al., 2016; Liu S. Y. et al., 2021).

3.2.6 Alcohol metabolism-mediated intestinal barrier and intestinal flora pathways

Tight junctions (TJ), sticky mucus gel layers, and intestinal cells work together to form the intestinal barrier, which prevents bacteria from passing through (Suzuki, 2020). The intestinal barrier also depends on commensal bacteria, sIgA antibodies, and mucins (Guohua Li and Chen, 2022). In comparison to individuals in good health, patients with alcohol-associated liver disease exhibit a decrease in intestinal bacilli and probiotics (e.g., *Lactobacillus*) and increased *E. coli* and *Enterococcus faecalis* (Dunn and Shah, 2016;



Shang Zeng and Li, 2022). Furthermore, studies have demonstrated that augmenting the proportion of intestinal bacilli and firmicutes can serve as a preventive measure against alcoholic liver injury and other chronic liver diseases (Qiu et al., 2019).

4 Management of alcohol-associated liver disease

As shown in Figure 3, the treatment of alcohol-associated liver disease consists of management, medication, and surgery.

4.1 Management of alcohol-associated liver disease

The most important phase of treatment in patients with alcoholassociated liver disease is achieving alcohol abstinence, since patients who drink alcohol after treatment significantly reduce the efficacy of alcohol-related therapies (Singal et al., 2018). Most patients with alcohol-associated liver disease meet the diagnostic criteria for alcohol-use disorder (AUD) which is a chronic relapsing disease (Erickson et al., 2019). Abuse and dependence are clinical characteristics of alcohol use disorder (AUD), characterized by the involuntary pursuit of alcohol stimulation, a lack of control over alcohol consumption, and depressive symptoms during periods of abstinence (Mason, 2017). Medications for AUD include disulfiram, naltrexone, and acamprosate (Jonas et al., 2014; Hartwell and Kranzler, 2019). Patients with AUD often suffer from concomitant liver damage, but disulfiram and naltrexone can exacerbate liver disease. Therefore, medications that can reduce alcohol intake are minimal (Addolorato et al., 2009). Baclofen is a y-Aminobutyric acid type B (GABAB) receptor agonist with a deficient hepatic metabolism (10%-15%), and less reported its side effects on liver (Addolorato et al., 2009; Morley et al., 2013).

4.2 Treatment of alcohol-associated liver disease

4.2.1 Treatment of alcohol withdrawal syndrome (AWS)

Patients with AUD are often accompanied by Alcohol withdrawal syndrome (AWS). AWS is caused by a dramatic decrease in GABA activity and glutamate overactivity a few hours after cessation of alcohol consumption (Airagnes et al., 2019). Benzodiazepines (BZDs) are the generalized medications used to treat alcoholic AWS, including chlordiazepoxide, oxazepam, diazepam, and lorazepam (Holleck et al., 2019). BZDs act therapeutically by stimulating y-aminobutyric acid type A (GABA-A) receptors, mimicking their stimulation by alcohol (Airagnes et al., 2019). Phenobarbital (PB) is a standard alternative or adjunct to BZDs and can reduce AWS complications and ICU admissions. PB exerts its effects through the activation of GABA-A receptors and inhibition of N-methyl-D-aspartate (NMDA) receptors (Mo et al., 2016). However, high doses of BZDs induce liver disease and dependence (Singal et al., 2018). PB also has a low margin of safety (Mo et al., 2016). Therefore, there is a need to explore safer medications for use in AWS. Other detrimental conditions may occur during AWS, such as Wernher's disease, Wernicke-Korsakoff syndrome (WKS) (Airagnes et al., 2019), and alcohol withdrawal delirium (AWD) (Mayo-Smith et al., 2004).

4.2.2 Nutritional therapy

Malnutrition is one of the main adverse consequences of alcohol-associated liver disease, especially in those with severe alcoholic hepatitis. Sufficient protein and vitamins must be administered orally or intravenously as part of nutritional treatment (Griffith and Schenker, 2006). In the case of alcoholism, even if sufficient protein, vitamins, and other nutrients are consumed, the body does not use them effectively because they are not sufficiently absorbed from the intestines into the bloodstream, resulting in malnutrition (Lieber, 2003). Zinc, magnesium, and vitamin A deficiencies are also standard in ALD (McClain et al., 2011).

4.2.3 Corticosteroids

Corticosteroids are used for the treatment of alcoholic hepatitis (AH). One example is prednisolone which is prescribed at a therapeutic dose of 40 mg daily for 4 weeks (Singal et al., 2018). Ramond et al. found that patients treated with prednisolone had a death rate of 12.5%, whereas those given a placebo had a mortality rate of 55% (Ramond et al., 1992). Additionally, prednisolone was found to lower short-term mortality in individuals with severe AH in a clinical investigation (Mathurin et al., 1996). To predict the effect of corticosteroid therapy, the Lille Model was introduced. The Lille Model measures were age, renal insufficiency, albumin, prothrombin time, bilirubin and day 7 bilirubin levels. The model measures from the seventh day of treatment, and patients with scores >0.45 indicate a lack of response to corticosteroids;

directing the need for changing the AH treatment to another suitable option (Louvet et al., 2007).

4.2.4 Pentoxifylline

Pentoxifylline is a non-specific phosphodiesterase inhibitor with anti-inflammatory qualities (Tyagi et al., 2011). It is administered at a dose of 400 mg thrice daily to treat severe AH (Akriviadis et al., 2000). Pentoxifylline attenuates the development of hepatic fibrosis by inhibiting TNF- α , which reduces mortality in AH and may decrease the incidence of hepatorenal syndrome (Tyagi et al., 2011). However, in patients with severe AH, there is no additional survival advantage of using pentoxifylline alone compared to corticosteroids (Sidhu et al., 2012). Pentoxifylline is ineffective in patients who are also unresponsive to corticosteroid therapy (Louvet et al., 2008). According to the European Association for the Study of the Liver (EASL), pentoxifylline is a second-line therapeutic agent for patients with severe AH who have contraindications to corticosteroid therapy (Liver, 2012).

4.3 Surgical treatment of alcohol-associated liver disease

The Lille Model suggests that patients with severe AH who do not respond to medication have a survival rate of only 25% at 6 months and only 10% at 1 year (Mellinger and Stine, 2020). Liver transplantation (LT) is an option for all patients with end-stage alcohol-associated liver disease (Kong et al., 2019). With the application of antiviral therapy to chronic hepatitis C (HCV), the approach for treating liver disease has shifted to treating ALD (Burra et al., 2010). In 1997, the American Society of Transplantation and the American Association for the Study of Liver Diseases made 6 months of abstinence from alcohol in patients with alcoholassociated liver disease, the minimum criteria for inclusion in alcoholic liver transplantation (Dureja and Lucey, 2010). In the context of liver transplantation, the recurrence rate of alcoholassociated liver disease is 10-50 percent, which is mainly due to cardiovascular disease and neoplastic disease (Iruzubieta et al., 2013). Screening and prevention of these factors in alcoholic patients is therefore essential (Mellinger and Stine, 2020). Brian P Lee et al. showed that patients who received liver transplants early in life had a higher survival rate compared to patients who underwent liver transplants after a strict 6-month abstinence from alcohol (Lee et al., 2017). Late-stage liver transplant patients live only 1.5 years, compared to an average of 6.6 years and even up to 10 years for early-stage liver transplants (Mellinger and Stine, 2020).

5 Hovenia dulcis search strategy for the mechanism of action of the treatment of alcohol-associated liver disease

5.1 Hovenia dulcis chemical composition retrieval

In this review, we obtained 44 chemical components of Hovenia dulcis by searching the following databases: TCMSP and Herb, of which the correlation between 9 chemical elements and alcohol-induced liver injury was verified, as shown in Table 1 and Figure 4.

5.2 Literature search

Based on the above Hovenia dulcis search for each chemical constituent, with "a chemical constituent" and "alcohol-associated liver disease" or "ethanol" or "alcohol" as the keywords, in the electronic databases PubMed, Web of Science (WOS), and the China National Knowledge Infrastructure (CNKI) search. The initial search was conducted on 2 April 2023, and the updated search was conducted on 31 August 2023. The search strategy on PubMed is shown in Supplementary Appendix S1 and the results are shown in Figure 5.

5.3 Inclusion exclusion criteria

Inclusion criteria were (a) studies on alcohol-induced liver injury, (b) interventions studied: chemical components of Hovenia dulcis, and (c) studies conducted *in vivo* or *in vitro*. Exclusion criteria were: (a) non-alcohol-induced liver injury, and (b) duplicate studies and titles or abstracts that did not meet the inclusion criteria.

6 Mechanism of action of Hovenia dulcis in the treatment of alcoholassociated liver disease

6.1 Activation of enzymes related to alcohol metabolism

Ethanol causes a decrease in the activity of ADH and ALDH in the liver, leading to toxic acetaldehyde accumulation. Treatment with DHM and naringenin increases the activity of these two enzymes (Jayaraman and Namasivayam, 2011; Hernández-Aquino and Muriel, 2018; Silva et al., 2020; Chen et al., 2021), accelerating alcohol metabolism and sparing hepatocytes from alcohol-induced effects (Chen et al., 2013). Also, the rate of ethanol metabolism by ADH and the rate of metabolism of acetaldehyde to acetic acid by ALDH is critical for the severity of ethanol damage to the liver (Jayaraman and Namasivayam, 2011). NAD, a coenzyme of dehydrogenase, acts as a cofactor molecule to facilitate the ALD to oxidize ethanol to acetaldehyde (Liu S. Y. et al., 2021). Ethanol intake increases the NADH/NAD ratio (Lingshi Wang, 2001), and naringenin reestablishes the ratio (Hernández-Aquino and Muriel, 2018).

6.2 Modulation of inflammatory mediators

6.2.1 LPS/TLR4/NF-κB pathway

Chronic alcohol consumption can damage the intestinal mucosal barrier leading to an increase in its permeability. Lipopolysaccharide (LPS) from intestinal Gram-negative bacteria translocates to the liver, where it binds to Toll-like receptor 4 (TLR4)

No.	Compound	Mechanism							Reported targets	References
		Ethanol metabolism	Inflammatory response	Oxidative stress	Apoptosis	Autophagy	Steatosis	Intestinal barrier		
1	Beta-sitosterol	-	-	+	+	-	-	-	8-OHdG, CYP2E1, MDA, SOD, GPX, CAT, Bax, Caspase-3, Caspase-9	Chen et al. (2020b)
2	Dihydromyricetin/ ampelopsin	+	+	+	+	+	+	+	ADH, ALDH, AMPK, Sirt1, Sirt3, PPARγ, PGC- 1α, TFAM, ACC, SREBP- 1, TNF-α, IL-1β, P62, Keap-1, Nrf2	Shen et al. (2012), Liang and Olsen (2014), Qiu et al. (2017), Zhang et al. (2018), Qiu et al. (2019), Silva et al. (2020), Skotnicová et al. (2020), Chen et al. (2021), Silva et al. (2021), Wang et al. (2022), Janilkarn-Urena et al. (2023)
3	Quercetin	-	+	+	-	+	+	+	Rab7, PLIN2, AMPK, IL- 10, IL-1β, LC3ll, IL-6, TNF-α, ERK, Nrf2, HO-1, Caspase-1, P38, AP-1, NF-κB, GSH, SOD, CAT, P2X7, PI3K, Keap1, Nrf2, Hepc,	Molina et al. (2003), Yao et al. (2007), Vidhya and Indira (2009), Yao et al. (2009), Ambadath et al. (2010), Chen, 2010; Liu et al. (2010), Liu et al. (2012), Tang et al. (2012), Leung and Nieto (2013), Li et al. (2014), Li et al. (2016), Yu et al. (2016), Liu et al. (2018), Zhao et al. (2018), Lee et al. (2019), Zeng et al. (2019), Hou et al. (2020), Pingili et al. (2020), Zhao et al. (2021), Lin et al. (2022), Zhao et al. (2022)
4	Naringenin	+	+	+	+	-	+	+	ADH, ALDH, HDL, TNF- α, IL-6, COX-2, NF-кB, Cyp2e1, ·OH, ROS, RNS, GSH, SOD, CD14	(Seo et al., 2003; Jayaraman and Namasivayam, 2011; Jayaraman et al., 2012; Hernández-Aquino and Muriel, 2018)
5	Lutein	-	+	+	-	-	+	+	TNF-α, IL-1β, NF-κB, COX-2, HO-1, CYP2E1, MDA, SOD, GPX, Nrf-2, Claudin-1, Occludin	(Sindhu et al., 2010; Du et al., 2015; Zhao et al., 2023)
6	Myricetin	-	+	+	-	-	+	+	FAS, SREBP-1c, AMPK, TNF-α, IL-6, TGF-β1, ROS, MDA	(Guo et al., 2019; Ahmad et al., 2022)

TABLE 1 Mechanism of action of Hovenia dulcis chemical constituents in relation to ALD.

on

10.3389/fphar.2024.1337633

No. Compound Me Eth me 7 Kaempferol -									
Kaempferol	Mechanism							Reported targets References	References
	Ethanol metabolism	Inflammatory response	Oxidative stress	Apoptosis	Apoptosis Autophagy Steatosis Intestinal barrier	Steatosis	Intestinal barrier		
			+	I	1	+	1	CYP2E1, ROS, MDA, SP1, ZO-1, Occludin	(Wang et al., 2015; Zhou et al., 2018; Chen et al., 2020a)
8 Emodin -		1	+	1		+		PPAR- γ , CYP2E1, α -SAM Liu et al. (2014)	Liu et al. (2014)
9 Apigenin -		+	+	1	1	+	+	SREBP-1c, FAS, LDL-C, NF-kB, TNF-a, GSH, CYP2E1, MDA, SOD, CAT, GSH, PPARa	Wang et al. (2017)

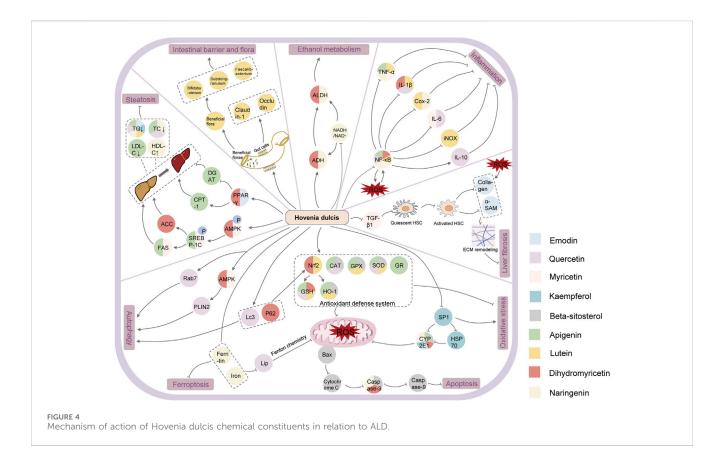
kupffer cells, causing its activation and subsequent release of a series of pro-inflammatory factors, such as tumor necrosis factor-alpha (TNF-alpha) and interleukin-1beta (IL-1β), which result in hepatic inflammation and systemic injury (Y, 2014; Shang Zeng and Li, 2022). Nuclear factor κB (NF- κB) is a key transcription factor in regulating inflammatory responses and a key component of intracellular signaling cascades (Xing et al., 2021). Naringenin, Lutein, Myricetin, and Apigenin reduce ethanol-induced levels of the hepatocyte inflammatory factor NF-κB (Jayaraman et al., 2012; Wang et al., 2017; Guo et al., 2019; Zhao et al., 2023). Activation of NF-kB stimulates the production of various inflammatory mediators, such as IL-1 β , IL-6, TNF- α , COX-2, and inducible nitric oxide synthase (iNOS) (Jayaraman et al., 2012; Du et al., 2015; Zhao et al., 2016). Among them, IL-1β, IL-6, and TNF-α are synthesized and released upon activation of hepatic stellate cells (Li et al., 2013). These are critical inflammatory factors in alcoholic liver injury and induce further necrosis, apoptosis, and exacerbation of inflammatory response in hepatic cells (Yuan et al., 2022). Quercetin reduces the levels of TNF- α , IL-1 β , and IL -6 levels (Chen, 2010); DHM and lutein can reduce TNF- α , and IL-1 β (Shen et al., 2012; Zhao et al., 2023), and myricetin can also reduce IL-6, and TNF- α levels (Guo et al., 2019; Ahmad et al., 2022). Naringenin, and apigenin can only reduce the concentration of TNF-a (Jayaraman et al., 2012; Wang et al., 2017). The essential enzyme that starts inflammation, cyclooxygenase-2 (COX-2), is one of NF-kB's target genes and is expressed by kupffer cells. Normal tissues have low levels of Cox-2 expression; however, an inflammatory stimulation increases Cox-2 expression with subsequent production of prostaglandins (Jayaraman et al., 2012). The inflammatory protective mechanism of naringenin and lutein mainly consists of inhibiting the overexpression of Cox-2 induced by alcohol exposure (Jayaraman et al., 2012; Zhao et al., 2023). Inducible nitric oxide synthase (iNOS) is the enzyme responsible for NO production and is not present in most cells under normal conditions. Its expression is inducible, often associated with inflammation and malignant diseases (Kielbik et al., 2019), and is a downstream gene of NFκB (Du et al., 2015). Lutein supplementation reverses alcoholinduced changes in iNOS, an indicator of inflammation (Zhao et al., 2023).

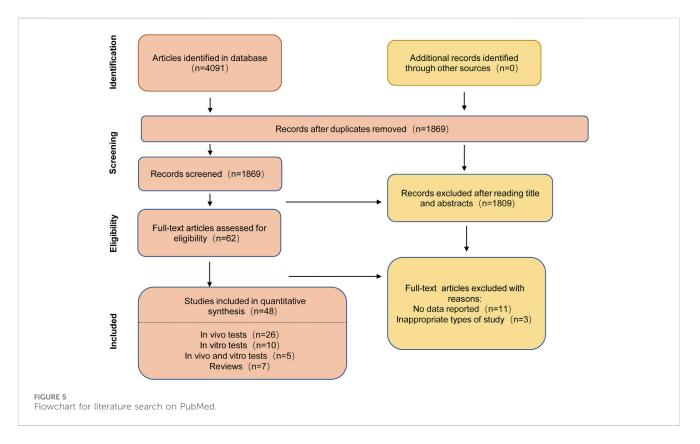
6.2.2 Anti-inflammatory factor pathway

Interleukin 10 (IL-10), a robust anti-inflammatory factor, lessens liver damage (Chen, 2010). Quercetin has been shown to positively affect the liver by up-regulating the expression of Heme Oxygenase-1(HO-1) and IL-10, inhibiting the NOD-like receptor thermal protein domain associated protein 3(NLRP3) inflammatory vesicles (Liu et al., 2018).

6.3 Inhibition of liver fibrosis

Chronic inflammation in the liver underlies the formation of hepatic fibrosis. Patients with severe ASH who persistently consume alcohol can further progress to fibrosis due to repetitive wound-healing processes in response to chronic inflammation and hepatocellular damage (Ha et al., 2022). TGF- β is the pro-fibrotic cytokine in the liver and contains three isoforms: transforming growth factor- β 1 (TGF- β 1), TGF- β 2, and TGF- β 3 (Chang et al.,





2023). A crucial inflammatory mediator, TGF- β 1 mainly stimulates hepatic stellate cells, kupffer, and perisinusoidal cells—all linked to hepatic fibrosis (Song et al., 2019). Myricetin inhibits the higher TGF- β 1 after ethanol exposure, potentially suppressing hepatic fibrosis (Zhao et al., 2018). Alpha-smooth muscle actin (a-SMA) is a marker of activated HSCs, which is responsible for the overproduction of ECM components and the development of fibrosis (Voutilainen et al., 2020). Upon stimulation with TGF- β , HSCs begin to differentiate from a quiescent state into proliferating and fibrotic myofibroblasts that express and secrete α -SMA and type I collagen (CoI-I), CoI-III and CoI-IV, leading to hepatic fibrosis (Chang et al., 2023). Emodin significantly reduces α -SMA and collagen expression in the alcohol-fed mouse group (Liu et al., 2014).

6.4 Maintaining mitochondrial stability and reducing oxidative stress damage induced by ethanol intake

Mitochondria are double-membrane organelles involved in a variety of physiological functions within the cell (Park et al., 2022). Silva et al. showed that DHM improves mitochondrial outcomes in the liver of alcohol-fed mice via the AMPK/Sirt-1/PGC-1 α signaling axis (Silva et al., 2021).

ROS can originate from mitochondria. Small amounts of free radicals usually maintain redox balance in the human body. However, overproduction of ROS due to excessive alcohol consumption may disturb the balance. Subsequently, oxidative stress may become an essential factor in alcoholic liver injury (Su et al., 2023). Oxidative stress is characterized by an increase in lipid peroxidation associated with alterations in the antioxidant system (Molina et al., 2003). Lipid peroxidation plays a crucial role in hepatic oxidative stress, which was determined by assessing malondialdehyde (MDA) enhancement (Chen, 2010). MDA is the end-product of lipid peroxidation, which can respond to the peroxidation of free radicals in vivo and can also indicate the extent to which free radicals indirectly attack cells (Zhao et al., 2010). Betasitosterol, DHM, quercetin, lutein, myricetin, kaempferol, and apigenin, can reduce ethanol-induced increases in MDA levels (Chen, 2010; Wang et al., 2015; Wang et al., 2017; Zhang et al., 2018; Guo et al., 2019; Chen Z. et al., 2020; Zhao et al., 2023).

6.4.1 ROS generation pathway

In addition to ADH, the MEOS metabolizes ethanol to acetaldehyde in ALD. The primary components of MEOS are the cytochrome P450 (CYP450) enzymes found in the smooth endoplasmic reticulum (SER), including CYP2E1 and CYP3A, among others (Lu and Cederbaum, 2018). Prolonged alcohol intake leads to the activation of the MEOS system, which can affect NADPH oxidation and ATP consumption, leading to oxidative stress in hepatocytes (Chen et al., 2013). Within the cytochrome P450 family, CYP2E1 is considered the most relevant to ALD due to its high degree of induction and catalytic activity in response to alcohol (Leung and Nieto, 2013). Heat shock protein 70 (HSP70) facilitates the synthesis of CYP2E1 proteins in the endoplasmic reticulum (ER) and their transfer into mitochondria (Zhou et al., 2018). SP1, a ubiquitously expressed transcription factor, transcribes and activates CYP2E1 and HSP70 (Morgan,

1989; Tang et al., 2019). Upon beta-sitosterol, DHM, naringenin, myricetin, kaempferol, emodin, and apigenin treatment, the levels of endogenous CYP2E1 were reduced in liver tissues, which attenuated hepatocyte injury (Jayaraman and Namasivayam, 2011; Liu et al., 2014; Wang et al., 2017; Zhou et al., 2018; Chen Z. et al., 2020; Silva et al., 2020; Ahmad et al., 2022). Kaempferol reduced ethanolinduced protein levels of CYP2E1 in microsomes and mitochondria and decreased the expression of HSP70 and SP1, reducing oxidative stress and increasing cell viability (Zhou et al., 2018).

The P2X7 receptor (P2X7R), is a critical receptor for oxidative stress and inflammation in ALD, induces the generation of activated ROS and imbalance of the antioxidant defense system. P2X7R is an adenosine triphosphate (ATP)-gated ion channel, that can be activated by high levels of ATP to exacerbate oxidative stress through the PI3K/Keap1/Nrf2 pathway. Quercetin treatment eliminates ethanol-induced overexpression of P2X7R and exerts a cytoprotective effect (Zhao et al., 2021).

6.4.2 Antioxidant defense system

The antioxidant defense system can be used to scavenge the accumulation of peroxides in the liver to reduce hepatic injury caused by oxidative stress. The major endogenous antioxidant enzyme systems include superoxide dismutase (SOD), catalase (CAT), glutathione peroxidase (GPX), and glutathione reductase (GR); non-enzymatic endogenous antioxidants are mainly glutathione (GSH) and vitamin E (Molina et al., 2003). Alcohol intake leads to increased levels of MDA and decreased GSH, SOD, and CAT, leading to liver injury. GSH is the first line of defense against free radicals and is the key to antioxidant activity (Vidhya and Indira, 2009; Tang et al., 2012). Treatment with DHM, quercetin, lutein, and apigenin increases the levels of the antioxidant GSH (Donglin Su and Yao, 2009; Vidhya and Indira, 2009; Sindhu et al., 2010; Wang et al., 2017; Hou et al., 2020). In addition, enzymes such as SOD and GPX are essential in scavenging free radicals to prevent their formation (Jayaraman and Namasivayam, 2011). Alcohol use has been linked to a drop in the liver's MDA and SOD and CAT levels. Treatment with betasitosterol, lutein, and apigenin promoted the elevation of SOD and GPX (Sindhu et al., 2010; Wang et al., 2017; Zhao et al., 2018; Chen Z. et al., 2020; Zhao et al., 2023). However, quercetin only promoted the increase of SOD (Vidhya and Indira, 2009). The activity of CAT responds to the cell's ability to scavenge ROS and its resistance to oxidative damage (Hao and Liu, 2019). The reduction in CAT levels brought on by alcohol can be reversed by beta-sitosterol and apigenin (Wang et al., 2017; Chen Z. et al., 2020). Apigenin treatment can potentially elevate GR levels in alcoholic liver injury (Wang et al., 2017). In the study by Liu et al., the effects of quercetin on GSH, SOD, and CAT levels were guided by pretreatment rather than post-treatment (Liu et al., 2010). Quercetin has also been demonstrated to scavenge oxygen-free radicals more effectively than conventional vitamin C and E forms.

Nrf2 is a protective factor expressed in response to oxidative stress. Excessive ROS disrupt the Keap1-Nrf2 interaction, triggering an effective atomic translocation of Nrf2. The translocated Nrf2 binds to the antioxidant response element (ARE) in blood and enhances the expression of antioxidant and detoxifying enzymes (Lee et al., 2019). Nrf2 has emerged as a potent therapeutic agent for

ALD, aiming to protect downstream targets, namely, HO-1 and GSH. HO-1 is the rate-limiting enzyme for heme catabolism. It can degrade heme to ferrous ions, carbon monoxide (CO), and biliverdin, which play a vital role in combating oxidative stress (Liu et al., 2012). Biliverdin then reductase breaks down biliverdin into bilirubin. Heme has a pro-oxidant nature and increases the susceptibility of cells to oxidative stress, while biliverdin and bilirubin exhibit noteworthy anti-inflammatory, antioxidant, and anti-apoptotic properties (Yao et al., 2009). DHM and lutein can activate the antioxidant system by increasing the expression of Nrf2 and its antioxidant product, HO-1 (Zhang et al., 2018; Silva et al., 2020; Chen et al., 2021; Zhao et al., 2023). Quercetin exerts its protective effect against ALD mainly through ERK/Nrf2-mediated upregulation of HO-1 to reduce ROS production (Liu et al., 2012; Zhao et al., 2021). In the mitogen-activated protein kinase (MAPK) signaling pathway, p38 and extracellular regulated protein kinases (ERK) mediated translocation of nuclear factor-erythroid 2-related factor 2(Nrf2) into the nucleus subsequently induces HO-1 activity, with a more critical mediating role for ERK (Yao et al., 2007). It has also been shown that in addition to Nrf2, HO-1 activation is regulated by other transcription factors, including activator protein-1 (AP-1) and nuclear factor (NF-KB) (Lee et al., 2019).

6.5 Cell death and prosurvival pathways

6.5.1 Inhibition of apoptosis

Chronic alcohol exposure induces multiple forms of cellular stress, including oxidative stress, hypoxia, and endoplasmic reticulum (ER) stress, which activate intrinsic and extrinsic pathways of apoptosis (Miyata and Nagy, 2020). Cell-intrinsic pathways receive tight regulation from the Bcl-2 protein family (Guicciardi et al., 2013). Bax belongs to the Bcl-2 protein family, which plays an essential role in apoptosis (Bruckheimer et al., 1998). Bax appears at the cell membrane in a healthy condition, which gets transferred to the mitochondria in response to apoptotic stressors (DNA damage, misfolded proteins, etc.), increasing the permeability of the cell membrane (Matsuyama et al., 2016). Cytochrome c exists on the inner membrane and then translocates to the cytoplasm (Choi et al., 2022). Bax initiates the intrinsic apoptotic pathway and triggers the release of cytochrome c, which then causes a series of caspase cascade reactions that induce apoptosis (Matsuyama et al., 2016). Beta-sitosterol inhibits alcohol-induced increase in the expression of Bax and inhibits cytochrome c, caspase-3, and caspase-9 elevation (Chen Z. et al., 2020). However, DHM and naringenin only decrease the elevation of caspase-3 after ethanolmediated (Zhao et al., 2018; Silva et al., 2020).

6.5.2 Inhibition of ferroptosis

The core events of iron toxicity include increased iron accumulation, impaired lipid repair systems, and lipid peroxidation, ultimately leading to membrane destruction and cell death (Wu et al., 2021). Numerous studies have shown that alcohol consumption is associated with iron overload in the liver and decreased hepcidin (Hepc) synthesis, which is thought to be a critical iron mediator of homeostasis (Liu S. Y. et al., 2021). Hepc, a small molecule peptide secreted by the liver, plays a role in iron homeostasis by inhibiting iron uptake and mobilization from

tissue stores. Alcohol stimulates iron absorption and hepatic storage by inhibiting the secretion of hepc, which releases free iron from various iron-containing proteins. Free iron in cells exists as a labile iron pool (LIP), i.e., redox-active, weakly chelated iron (Li et al., 2016); that when present in excess catalyzes the Fenton reaction to produce hydroxyl radicals (·OH), exacerbating alcoholic liver injury. Hydroxyl radicals are the most toxic component of ROS (Li et al., 2014; Li et al., 2016). The Fenton reaction occurs within lysosomal compartments, which can accumulate large amounts of redox-active iron and are the primary source of LIP (Li et al., 2016). Quercetin helps to maintain intracellular LIP levels and attenuate intracellular free iron-mediated production of hydroxy radicals (·OH), which can alleviate alcohol-induced liver injury (Li et al., 2014; Li et al., 2016). Ethanol feeding stimulated elevated levels of iron and ferritin in the liver of rats, leading to increased oxidative stress. Whereas naringenin intake significantly reduced the expression of iron and ferritin in the liver (Jayaraman et al., 2012).

6.5.3 Promotion of autophagy

6.5.3.1 Mitophagy

Mitophagy is a selective form of macro-autophagy that mediates mitochondrial degradation in the autolysosomes (Salete-Granado et al., 2023). LC3 is an autophagy-specific protein with three isoforms, LC3A, LC3B, and LC3C, but only LC3-II has been associated with increased phagocytic vesicle membranes. LC3-II is the autophagosomal form of LC3, which is produced by specific protein hydrolysis in the vicinity of the cell and can be a direct response to the degree of active autophagy (Noh et al., 2011). P62 (SQSTM 1) is considered a multifaceted adapter protein. It is thought to be a multidimensional adaptor protein that recognizes ubiquitinated proteins and triggers the autophagic degradation of these proteins via the lysosomal pathway through LC3-II interactions (Qiu et al., 2017; Salete-Granado et al., 2023). Chronic ethanol ingestion leads to impaired autophagy, as evidenced by a reduced LC3. Quercetin restores the levels of LC3. By continuously feeding DHM to C57BL/6 for 6 weeks, Qiu et al., found for the first time that the protective effect of DHM against ethanol-induced liver injury was associated with autophagy induced by the interaction of P62 and the Keap1-Nrf2 system (Qiu et al., 2017). Specifically, DHM can upregulate the expression of P62 and promote ethanol-induced autophagy in mice, activating the positive feedback pathway in the Keap1-Nrf2 pathway, thus exerting its hepatoprotective effect (Qiu et al., 2017; Zhang et al., 2018; Chen et al., 2021).

6.5.3.2 Lipophagy pathway

Lipophagy, which degrades lipid droplets (LDs) mainly through lysosomal acid lipase, is a mode of protection in the body that protects the liver from alcohol-induced steatosis. Specifically, during the lysosomal-centered lipophagy process, LDs in the cytoplasm are engulfed by autophagosomes to be transported to lysosomes, where lipids are broken down by lysosomal acid lipase. Rab7, a critical factor in autophagosome-lysosome fusion, plays an essential role in ALD-related lipophagy. Quercetin intake can restore ethanolinduced impairment of Rab7 cycling to improve lipophagy (Lin et al., 2022). Zeng et al. demonstrated that quercetin reduced perilipin 2 (PLIN2) levels, activated AMPK activity, and increased co-localization of hepatic LC3II and PLIN2 proteins to substantiate the lipophagy-promoting effects of quercetin (Zeng et al., 2019). DHM demonstrated enhanced lipid clearance by way of increased lipophagy activity, shown by the increased interaction and colocalization of p62/SQSTM-1, LC3B, and PLIN-1 proteins (Janilkarn-Urena et al., 2023).

6.6 Reduction of steatosis

6.6.1 Lipid metabolism indicators

Hepatic steatosis represents the liver's earliest and most common response to acute or chronic alcohol exposure (Huo et al., 2018). During steatosis, triglycerides (TG), cholesterol (TC), and phospholipids build up in hepatocytes (Dunn and Shah, 2016). Several studies have shown that quercetin, naringenin, and apigenin reduced alcohol-induced hepatic steatosis. Moreover, these compounds can lower serum TG and TC concentrations, ultimately restoring cell viability (Seo et al., 2003; Zhao et al., 2018; Zhao et al., 2021). Lutein, kaempferol, and emodin only reduced TG levels and had no significant effect on TC (Liu et al., 2014; Wang et al., 2015; Zhao et al., 2023). High-density lipoprotein (HDL-C) is involved in the reverse transport of cholesterol from peripheral tissues and lipids from the vasculature to the liver for clearance (Wierzbicki and Mikhailidis, 2002). Naringin increases the serum HDL-C and HDL-C/total-C ratios, returning these lipid parameters to normal levels and reducing alcohol damage to the liver (Seo et al., 2003). Serum low-density lipoprotein cholesterol (LDL-C) acts oppositely to HDL-C, transporting lipids from the liver into the vasculature and inducing disease. Apigenin reduces the alcohol-induced higher serum values of LDL-C (Wang et al., 2017).

6.6.2 AMPK pathway

AMPK, the classical mammalian upstream serine protein kinase, induces lipolysis and suppresses lipid synthesis. Its activation can lead to a reduction in steatosis and inflammation (Silva et al., 2020; Pang et al., 2021). Ethanol and acetaldehyde inhibit fatty acid oxidation by downregulating the levels of AMPK (Dunn and Shah, 2016; Zhao et al., 2016; Liu S. Y. et al., 2021). DHM and myricetin can decrease AMPK levels (Guo et al., 2019). Increased phosphorylation of AMPK promotes phosphorylation of SREBP-1c (Zhu et al., 2019). One of the critical regulators of adipogenesis, the transcription factor SREBP-1c, plays a role in the transcriptional activation of genes that encode the rate-limiting enzymes in adipogenesis, including acetyl-CoA carboxylase (ACC), fatty acid synthase (FAS), and stearoyl-CoA desaturase 1 (SCD1) (Fang et al., 2019). Myricetin and apigenin significantly inhibited alcoholinduced levels of the liposynthesis gene SREBP-1c (Wang et al., 2017; Guo et al., 2019). FAS catalyzes the condensation of acetyl-CoA and malonyl-CoA to produce long-chain fatty acids in the cytoplasm (Ronnett et al., 2005). Myricetin and apigenin inhibit the elevation of FAS caused by alcohol exposure (Wang et al., 2017; Guo et al., 2019). ACC is the rate-limiting enzyme for the de novo synthesis of fatty acids and catalyzes acetyl-CoA conversion to malonyl-CoA (Pang et al., 2021). In the liver, phosphorylation of AMPK inactivates ACC, which decreases fatty acid synthesis and increases fatty acid oxidation (Liu Y. S. et al., 2021). DHM regulates alcohol-induced hepatic steatosis through the AMPK-ACC pathway, which reduces lipid accumulation (Chen et al., 2021).

6.6.3 PPAR pathway

The peroxisome proliferator-activated receptor (PPAR) belongs to the nuclear hormone receptor superfamily and plays a crucial role in glucose and fatty acid metabolism. There are three known members of this superfamily: PPAR-a, PPAR-b/d, and PPAR-y. Known as the "energy homeostasis receptor," PPAR-y is a crucial regulator in lipid metabolism. PPAR-y regulates several downstream genes involved in fatty acid oxidation, such as carnitine palmitoyltransferase-1 (CPT-1) (Zheng and Cai, 2019). DHM and emodin decreased PPAR-y caused by ethanol exposure (Liang and Olsen, 2014; Liu et al., 2014; Chen et al., 2021). Fatty acid catabolism occurs in mitochondria and is regulated by CPT-1, the rate-limiting step in entering fatty acids into mitochondria (Ronnett et al., 2005). Diacylglycerol acyltransferase (DGAT), a microsomal enzyme widely expressed in mammalian tissues, is also a key enzyme in triglyceride synthesis (Chen and Farese, 2000). Preadministration of apigenin decreased hepatic DGAT protein expression in mice with alcohol-induced liver injury, while hepatic CPT-1 protein expression was inversely increased. These findings show that pretreatment with apigenin reduced hepatic lipid synthesis and increased fatty acid utilization, indicating that apigenin may be able to avoid abnormalities of hepatic lipid metabolism (Wang et al., 2017).

6.7 Protection of the intestinal barrier and intestinal flora

The intestinal barrier system depends on the interactions between several barrier components, including the dense mucus gel layer, immunoglobulin A, commensal bacteria, and intercellular TJs (Suzuki, 2020; Guohua Li and Chen, 2022). TJs play a pivotal role in establishing the physical barrier among these components. The transmembrane proteins claudin and occludin, as well as the intracellular plaque proteins zonula occludens (ZO) and neuregulin, make up the TJs structure (Suzuki, 2020). TJs such as claudin-1 regulate epithelial and endothelial cells by forming TJ chains, paracellular barrier integrity, and paracellular barrier permeability between epithelial cells and endothelial cells (Zwanziger et al., 2015). Occludin, the first transmembrane protein of the epithelial cell tight junction to be identified, plays an essential role in regulating tight junction integrity (Rao, 2009). Alcohol-induced reductions in claudin-1 and occludin can be reversed by supplementation with lutein (Zhao et al., 2023). Short-chain fatty acids (SCFAs) produced by gut flora are the major contributing factors to the development of the TJ. SCFAs are part of the intestinal biological barrier produced in the colon. The concentration of SCFA in the colon's lumen is approximately 60% acetate, 25% propionate, and 15% butyrate. The short-chain fatty acid butyrate improves mucosal inflammation and oxidative status, strengthens the epithelial defense barrier, and modulates visceral sensitivity and intestinal motility (Canani et al., 2011). Kaempferol restores TJ protein ZO-1 and occludin, as well as the levels of butyrate receptor GPR109A and butyrate transporter SLC58A proteins in ethanol-exposed mice (Chen J. et al., 2020).

Chronic alcohol intake leads to bacterial overgrowth and dysbiosis in the small and large intestines of animals and 2015). Beneficial humans (Fukui, intestinal flora like Bifidobacterium and Faecalibacterium contribute to the concentration of butyrate in the digestive system, which reduces intestinal inflammation and preserves intestinal integrity (Ralli et al., 2021). Lutein reduces the amount of damage caused by alcohol while increasing the amount of Bifidobacterium, Subdoligranulum, and Faecalibacterium (Zhao et al., 2023).

7 Conclusion and perspectives

Worldwide, the prevalence of alcohol-associated liver disease is progressively rising. In the early stages, patients can regain their liver function by abstaining from alcohol and practicing exercise. However, as the disease progresses, drug intervention or even liver transplantation is required. Currently, the primary drugs used to treat ALD are disulfiram, BZDs, corticosteroids, and pentoxifylline. However, these drugs have a certain degree of hepatotoxicity and dependence. Therefore, there is still a lack of adequate and safe drugs in this field. Herbal medicine can provide a potential solution for controlling and treating ALD due to its multipathway and multi-target characteristics.

Numerous studies have shown that Hovenia dulcis contains a wide spectrum of pharmacological activities that can be used to treat a variety of liver diseases. Therefore, exploring its pharmacological mechanism of action is beneficial to provide a new direction for the treatment of alcohol-associated liver disease. This review, based on the current research on Hovenia dulcis and its derived natural compounds, retrieved nine chemical components related to ALD, which are betasitosterol, DHM, quercetin, naringenin, lutein, myricetin, kaempferol, emodin, apigenin. The potential protective mechanisms of these nine chemicals against ALD involve ethanol metabolism, immune response, hepatic fibrosis, oxidative stress, autophagy, lipid metabolism, intestinal barrier, and many more. The antioxidant mechanism is involved in every chemical component, lipid metabolism is involved in eight components, and the inflammatory response is involved in six components, so it can be deduced that Hovenia dulcis plays a significant role in the treatment of ALD by detoxifying and lowering lipids, as well as anti-inflammatory and hepatoprotective effects.

Despite this study of the therapeutic effects of Hovenia dulcis' chemical composition on ALD, the current experimental study still has some limitations. (1) Currently, herbs with Hovenia dulcis as the main ingredient are primarily used in the research and development of healthcare products, failing to develop into therapeutic drugs. In the same time, the research on the pharmacological effects of using Hovenia dulcis to treat ALD is mainly *in vivo* and *in vitro* experiments, with a lack of double-blind, multi-center, and randomized controlled clinical trials. Hence, its safety, especially for long-term use, remains to be determined. (2) The material basis of Hovenia dulcis's efficacy in treating ALD is closely related to its unique content of natural compounds. Therefore, it is crucial to clarify the basis of Hovenia dulcis's pharmacological effects and quality standards through

the International Organisation for Standardisation. To overcome these limitations, we should conduct more analyses to find further mechanisms and targets for Hovenia dulcis treatment of alcohol-associated liver disease. (1) Establish a high-quality Hovenia dulcis clinical research protocol to facilitate large-scale, multi-center, controlled trials to assess the efficacy and safety of Hovenia dulcis in treating alcohol-associated liver disease to promote the development and clinical application of the drug. (2) Improve the Hovenia dulcis quality recognition standards to reflect its true quality more accurately. In the same time, exploring the interactions of the various active ingredients in Hovenia dulcis and their compatibility with other natural substances for drugs is also an important direction for future research.

8 Scope statement

We would like to submit the enclosed manuscript entitled "Hovenia dulcis: A Chinese medicine that plays an essential role in alcohol-associated liver disease" for your consideration as a review in "Frontiers in Pharmacology". In this manuscript, we reviewed the mechanism of action of Hovenia dulcis in the treatment of alcoholassociated liver disease and its targets, providing us with new ideas for the treatment of alcohol-associated liver disease. All authors of this research manuscript were directly involved in the planning, implementation, and/or analysis of this study. The authors declare no competing interests. The contents of this manuscript are not copyrighted and have not been previously published. We deeply appreciate your consideration of our manuscript, and we look forward to receiving comments from the reviewers.

Author contributions

Y-XH: Writing-original draft. M-NL: Writing-review and editing. Y-YW: Writing-review and editing. HW: Writing-review and editing. MW: Writing-review and editing. J-YX: Writing-review and editing. YZ: Writing-review and editing. XZ: Writing-review and editing. HC: Writing-review and editing. ZL: Writing-review and editing.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This research was supported by funds from the Sichuan Science and Technology Program (2022YFS0624). Sichuan Provincial Administration of Traditional Chinese Medicine Science and Technology Program (2023zd008 and 2023MS335). Sichuan Provincial Health Department Project (Sichuan Cadre Research 2024-2001).

Acknowledgments

Thanks to Dr. Maryam Mazhar for improving the readability and coherence of the language in the article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Addolorato, G., Russell, M., Albano, E., Haber, P. S., Wands, J. R., and Leggio, L. (2009). Understanding and treating patients with alcoholic cirrhosis: an update. *Alcohol Clin. Exp. Res.* 33 (7), 1136–1144. doi:10.1111/j.1530-0277.2009.00956.x

Ahmad, S. B., Rashid, S. M., Wali, A. F., Ali, S., Rehman, M. U., Maqbool, M. T., et al. (2022). Myricetin (3,3('),4('),5,5('),7-hexahydroxyflavone) prevents ethanol-induced biochemical and inflammatory damage in the liver of Wistar rats. *Hum. Exp. Toxicol.* 41, 9603271211066843. doi:10.1177/09603271211066843

Airagnes, G., Ducoutumany, G., Laffy-Beaufils, B., Le Faou, A. L., and Limosin, F. (2019). Alcohol withdrawal syndrome management: is there anything new? *Rev. Med. Interne* 40 (6), 373–379. doi:10.1016/j.revmed.2019.02.001

Akriviadis, E., Botla, R., Briggs, W., Han, S., Reynolds, T., and Shakil, O. (2000). Pentoxifylline improves short-term survival in severe acute alcoholic hepatitis: a double-blind, placebo-controlled trial. *Gastroenterology* 119 (6), 1637–1648. doi:10.1053/gast.2000.20189

Albano, E. (2008). Oxidative mechanisms in the pathogenesis of alcoholic liver disease. Mol. Asp. Med. 29 (1-2), 9-16. doi:10.1016/j.mam.2007.09.004

Ambadath, V., Venu, R. G., and Madambath, I. (2010). Comparative study of the efficacy of ascorbic acid, quercetin, and thiamine for reversing ethanol-induced toxicity. *J. Med. Food* 13 (6), 1485–1489. doi:10.1089/jmf.2009.1387

Bao-Jun, X. (2003). Chemical compositions of the genus hovenia. *Nat. Product. Sci.* 9 (3), 143–153.

Bellanti, F., Mangieri, D., and Vendemiale, G. (2023). Redox biology and liver fibrosis. Int. J. Mol. Sci. 25 (1), 410. doi:10.3390/ijms25010410

Bruckheimer, E. M., Cho, S. H., Sarkiss, M., Herrmann, J., and McDonnell, T. J. (1998). The Bcl-2 gene family and apoptosis. *Adv. Biochem. Eng. Biotechnol.* 62, 75–105. doi:10.1007/BFb0102306

Burra, P., Senzolo, M., Adam, R., Delvart, V., Karam, V., Germani, G., et al. (2010). Liver transplantation for alcoholic liver disease in Europe: a study from the ELTR (European Liver Transplant Registry). *Am. J. Transpl.* 10 (1), 138–148. doi:10.1111/j. 1600-6143.2009.02869.x

Canani, R. B., Costanzo, M. D., Leone, L., Pedata, M., Meli, R., and Calignano, A. (2011). Potential beneficial effects of butyrate in intestinal and extraintestinal diseases. *World J. Gastroenterol.* 17 (12), 1519–1528. doi:10.3748/wjg.v17.i12.1519

Chang, J., Huang, C., Li, S., Jiang, X., Chang, H., and Li, M. (2023). Research progress regarding the effect and mechanism of dietary polyphenols in liver fibrosis. *Molecules* 29 (1), 127. doi:10.3390/molecules29010127

Chen, H. C., and Farese, R. V., Jr. (2000). DGAT and triglyceride synthesis: a new target for obesity treatment? *Trends Cardiovasc Med.* 10 (5), 188–192. doi:10.1016/s1050-1738(00)00066-9

Chen, J., Wang, X., Xia, T., Bi, Y., Liu, B., Fu, J., et al. (2021). Molecular mechanisms and therapeutic implications of dihydromyricetin in liver disease. *Biomed. Pharmacother.* 142, 111927. doi:10.1016/j.biopha.2021.111927

Chen, J., Xuan, Y. H., Luo, M. X., Ni, X. G., Ling, L. Q., Hu, S. J., et al. (2020a). Kaempferol alleviates acute alcoholic liver injury in mice by regulating intestinal tight junction proteins and butyrate receptors and transporters. *Toxicology* 429, 152338. doi:10.1016/j.tox.2019.152338

Chen, M., Zhong, W., and Xu, W. (2023a). Alcohol and the mechanisms of liver disease. J. Gastroenterol. Hepatol. 38 (8), 1233-1240. doi:10.1111/jgh.16282

Chen, X. (2010). Protective effects of quercetin on liver injury induced by ethanol. *Pharmacogn. Mag.* 6 (22), 135–141. doi:10.4103/0973-1296.62900

Chen, X., Li, R., Liang, T., Zhang, K., Gao, Y., and Xu, L. (2013). Puerarin improves metabolic function leading to hepatoprotective effects in chronic alcohol-induced liver injury in rats. *Phytomedicine* 20 (10), 849–852. doi:10. 1016/j.phymed.2013.04.001

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2024.1337633/ full#supplementary-material

Chen, Y., Fang, Z. M., Yi, X., Wei, X., and Jiang, D. S. (2023b). The interaction between ferroptosis and inflammatory signaling pathways. *Cell Death Dis.* 14 (3), 205. doi:10.1038/s41419-023-05716-0

Chen, Z., Wu, A., Jin, H., and Liu, F. (2020b). β -Sitosterol attenuates liver injury in a rat model of chronic alcohol intake. *Arch. Pharm. Res.* 43 (11), 1197–1206. doi:10.1007/s12272-020-01271-w

Choi, H. S., Lim, E. S., and Baek, K. H. (2022). Deubiquitinating enzyme USP12 regulates the pro-apoptosis protein Bax. *Int. J. Mol. Sci.* 23 (21), 13107. doi:10.3390/ijms232113107

Donglin Su, J. H., and Yao, M. (2009). The acute toxicological evaluation of dihydromyricetin and its control effect for alcoholic hepatic injury. *Hunan Agric. Sci.* 11, 90–93.

Du, S. Y., Zhang, Y. L., Bai, R. X., Ai, Z. L., Xie, B. S., and Yang, H. Y. (2015). Lutein prevents alcohol-induced liver disease in rats by modulating oxidative stress and inflammation. *Int. J. Clin. Exp. Med.* 8 (6), 8785–8793.

Dunn, W., and Shah, V. H. (2016). Pathogenesis of alcoholic liver disease. Clin. Liver Dis. 20 (3), 445–456. doi:10.1016/j.cld.2016.02.004

Dureja, P., and Lucey, M. R. (2010). The place of liver transplantation in the treatment of severe alcoholic hepatitis. J. Hepatol. 52 (5), 759–764. doi:10.1016/j.jhep.2009.12.021

Erickson, E. K., Grantham, E. K., Warden, A. S., and Harris, R. A. (2019). Neuroimmune signaling in alcohol use disorder. *Pharmacol. Biochem. Behav.* 177, 34–60. doi:10.1016/j.pbb.2018.12.007

Fang, K., Wu, F., Chen, G., Dong, H., Li, J., Zhao, Y., et al. (2019). Diosgenin ameliorates palmitic acid-induced lipid accumulation via AMPK/ACC/CPT-1A and SREBP-1c/FAS signaling pathways in LO2 cells. *BMC Complement. Altern. Med.* 19 (1), 255. doi:10.1186/s12906-019-2671-9

Fang, X., Cao, J., Tao, Z., Yang, Z., Dai, Y., and Zhao, L. (2023). Hydroxytyrosol attenuates ethanol-induced liver injury by ameliorating steatosis, oxidative stress and hepatic inflammation by interfering STAT3/iNOS pathway. *Redox Rep.* 28 (1), 2187564. doi:10.1080/13510002.2023.2187564

Fangfang Xu, C. F., and Wang, L. (2011). Chemical constituents from the seeds of Hovenia acerba. J. Jinan Univ. Sci. 32 (3), 304–306.

Fu, K., Wang, C., Ma, C., Zhou, H., and Li, Y. (2021). The potential application of Chinese medicine in liver diseases: a new opportunity. *Front. Pharmacol.* 12, 771459. doi:10.3389/fphar.2021.771459

Fu, X., Tan, Y., Shi, M., Zeng, C., and Qin, S. (2023). Multi-index comprehensive assessment optimized critical flavonoids extraction from semen hoveniae and their *in vitro* digestive behavior evaluation. *Foods* 12 (4), 773. doi:10.3390/foods12040773

Fukui, H. (2015). Gut microbiota and host reaction in liver diseases. *Microorganisms* 3 (4), 759–791. doi:10.3390/microorganisms3040759

Glick, D., Barth, S., and Macleod, K. F. (2010). Autophagy: cellular and molecular mechanisms. J. Pathol. 221 (1), 3–12. doi:10.1002/path.2697

Griffith, C. M., and Schenker, S. (2006). The role of nutritional therapy in alcoholic liver disease. *Alcohol Res. Health* 29 (4), 296–306.

Guicciardi, M. E., Malhi, H., Mott, J. L., and Gores, G. J. (2013). Apoptosis and necrosis in the liver. *Compr. Physiol.* 3 (2), 977–1010. doi:10.1002/cphy.c120020

Guo, C., Xue, G., Pan, B., Zhao, M., Chen, S., Gao, J., et al. (2019). Myricetin ameliorates ethanol-induced lipid accumulation in liver cells by reducing fatty acid biosynthesis. *Mol. Nutr. Food Res.* 63 (14), e1801393. doi:10.1002/mnfr.201801393

Guohua Li, H. L., and Chen, Z. (2022). Research progress on alcoholic liver diseases and gut microbiota. *Res. Prog. Alcohol. liver Dis. gut microbiota* 30 (3), 423–427.

Ha, Y., Jeong, I., and Kim, T. H. (2022). Alcohol-related liver disease: an overview on pathophysiology, diagnosis and therapeutic perspectives. *Biomedicines* 10 (10), 2530. doi:10.3390/biomedicines10102530

Hao, M., and Liu, R. (2019). Molecular mechanism of CAT and SOD activity change under MPA-CdTe quantum dots induced oxidative stress in the mouse primary hepatocytes. *Spectrochim. Acta A Mol. Biomol. Spectrosc.* 220, 117104. doi:10.1016/j. saa.2019.05.009

Hartwell, E. E., and Kranzler, H. R. (2019). Pharmacogenetics of alcohol use disorder treatments: an update. *Expert Opin. Drug Metab. Toxicol.* 15 (7), 553–564. doi:10.1080/17425255.2019.1628218

Hernández-Aquino, E., and Muriel, P. (2018). Beneficial effects of naringenin in liver diseases: molecular mechanisms. *World J. Gastroenterol.* 24 (16), 1679–1707. doi:10. 3748/wjg.v24.i16.1679

Holleck, J. L., Merchant, N., and Gunderson, C. G. (2019). Symptom-triggered therapy for alcohol withdrawal syndrome: a systematic review and meta-analysis of randomized controlled trials. *J. Gen. Intern Med.* 34 (6), 1018–1024. doi:10.1007/s11606-019-04899-7

Hou, W., Qiu, P., Chen, N. J., Yao, P., Liu, S., and Qin, H. (2020). Study on the role and possible mechanism of hemeoxygenase-1/carbon monoxide system in protection of quercetin against ethanol-induced hepatocytes oxidative injury. *Zhonghua Gan Zang Bing Za Zhi* 28 (6), 504–508. doi:10.3760/cma.jcn501113-20200522-00269

Huo, X., Yang, S., Sun, X., Meng, X., and Zhao, Y. (2018). Protective effect of glycyrrhizic acid on alcoholic liver injury in rats by modulating lipid metabolism. *Molecules* 23 (7), 1623. doi:10.3390/molecules23071623

Iruzubieta, P., Crespo, J., and Fábrega, E. (2013). Long-term survival after liver transplantation for alcoholic liver disease. *World J. Gastroenterol.* 19 (48), 9198–9208. doi:10.3748/wjg.v19.i48.9198

Janilkarn-Urena, I., Idrissova, A., Zhang, M., VanDreal, M., Sanghavi, N., Skinner, S. G., et al. (2023). Dihydromyricetin supplementation improves ethanol-induced lipid accumulation and inflammation. *Front. Nutr.* 10, 1201007. doi:10.3389/fnut.2023. 1201007

Jayaraman, J., Jesudoss, V. A., Menon, V. P., and Namasivayam, N. (2012). Antiinflammatory role of naringenin in rats with ethanol induced liver injury. *Toxicol. Mech. Methods* 22 (7), 568–576. doi:10.3109/15376516.2012.707255

Jayaraman, J., and Namasivayam, N. (2011). Naringenin modulates circulatory lipid peroxidation, anti-oxidant status and hepatic alcohol metabolizing enzymes in rats with ethanol induced liver injury. *Fundam. Clin. Pharmacol.* 25 (6), 682–689. doi:10.1111/j. 1472-8206.2010.00899.x

Jiaming Chen, C. J. H. L. (2022). Antialcoholism effects of curcuma botanical beverage in KM mice. *Mod. Food Sci. Technol.* 38 (7), 40–47.

Jonas, D. E., Amick, H. R., Feltner, C., Bobashev, G., Thomas, K., Wines, R., et al. (2014). Pharmacotherapy for adults with alcohol use disorders in outpatient settings: a systematic review and meta-analysis. *Jama* 311 (18), 1889–1900. doi:10.1001/jama.2014. 3628

Kielbik, M., Szulc-Kielbik, I., and Klink, M. (2019). The potential role of iNOS in ovarian cancer progression and chemoresistance. *Int. J. Mol. Sci.* 20 (7), 1751. doi:10. 3390/ijms20071751

Kong, L. Z., Chandimali, N., Han, Y. H., Lee, D. H., Kim, J. S., Kim, S. U., et al. (2019). Pathogenesis, early diagnosis, and therapeutic management of alcoholic liver disease. *Int. J. Mol. Sci.* 20 (11), 2712. doi:10.3390/ijms20112712

Lafoz, E., Ruart, M., Anton, A., Oncins, A., and Hernández-Gea, V. (2020). The endothelium as a driver of liver fibrosis and regeneration. *Cells* 9 (4), 929. doi:10.3390/ cells9040929

Lee, B. P., Chen, P. H., Haugen, C., Hernaez, R., Gurakar, A., Philosophe, B., et al. (2017). Three-year results of a pilot Program in early liver transplantation for severe alcoholic hepatitis. *Ann. Surg.* 265 (1), 20–29. doi:10.1097/sla.00000000001831

Lee, S., Lee, J., Lee, H., and Sung, J. (2019). Relative protective activities of quercetin, quercetin-3-glucoside, and rutin in alcohol-induced liver injury. *J. Food Biochem.* 43 (11), e13002. doi:10.1111/jfbc.13002

Leung, T. M., and Nieto, N. (2013). CYP2E1 and oxidant stress in alcoholic and nonalcoholic fatty liver disease. J. Hepatol. 58 (2), 395–398. doi:10.1016/j.jhep.2012.08.018

Li, M. C., Xie, C. J., Meng, C. W., Zhang, Y. Q., Gao, J. G., Wang, W. H., et al. (2021). Chemical constituents from Hovenia dulcis Thunb. And their chemotaxonomic significance. *Biochem. Syst. Ecol.* 94, 104214. doi:10.1016/j.bse.2020.104214

Li, R., Liang, T., He, Q., Guo, C., Xu, L., Zhang, K., et al. (2013). Puerarin, isolated from Kudzu root (Willd.), attenuates hepatocellular cytotoxicity and regulates the GSK- $3\beta/NF-\kappa B$ pathway for exerting the hepatoprotection against chronic alcohol-induced liver injury in rats. *Int. Immunopharmacol.* 17 (1), 71–78. doi:10.1016/j.intimp.2013. 05.023

Li, Y., Chen, M., Xu, Y., Yu, X., Xiong, T., Du, M., et al. (2016). Iron-mediated lysosomal membrane permeabilization in ethanol-induced hepatic oxidative damage and apoptosis: protective effects of quercetin. *Oxid. Med. Cell Longev.* 2016, 4147610. doi:10.1155/2016/4147610

Li, Y., Deng, Y., Tang, Y., Yu, H., Gao, C., Liu, L., et al. (2014). Quercetin protects rat hepatocytes from oxidative damage induced by ethanol and iron by maintaining

intercellular liable iron pool. Hum. Exp. Toxicol. 33 (5), 534-541. doi:10.1177/0960327113499168

Liang, J., and Olsen, R. W. (2014). Alcohol use disorders and current pharmacological therapies: the role of GABA(A) receptors. *Acta Pharmacol. Sin.* 35 (8), 981–993. doi:10. 1038/aps.2014.50

Lieber, C. S. (2003). Relationships between nutrition, alcohol use, and liver disease. Alcohol Res. Health 27 (3), 220-231.

Lin, H., Guo, X., Liu, J., Liu, P., Mei, G., Li, H., et al. (2022). Improving lipophagy by restoring Rab7 cycle: protective effects of quercetin on ethanol-induced liver steatosis. *Nutrients* 14 (3), 658. doi:10.3390/nu14030658

Lingshi Wang, L. P. (2001). Alcoholic liver disease. Alcohol. liver Dis. 16 (24), 1141-1143.

Ling Yang, S. Z., and Lu, Y. (2022). The exploration of the mechanism of progression in ALD rats based on LPS/TLR4 signaling pathway. *J. Med. Postgraduates* 35 (1).

Linzhi Kang, S. L., and Li, Z. (2022). Research progress on the efficacy of hovenia dulcis thunb. And related food development. J. Shaoguan Univ. 43 (9), 6-10.

Liu, S., Hou, W., Yao, P., Li, N., Zhang, B., Hao, L., et al. (2012). Heme oxygenase-1 mediates the protective role of quercetin against ethanol-induced rat hepatocytes oxidative damage. *Toxicol Vitro* 26 (1), 74–80. doi:10.1016/j.tiv.2011.10.013

Liu, S., Hou, W., Yao, P., Zhang, B., Sun, S., Nüssler, A. K., et al. (2010). Quercetin protects against ethanol-induced oxidative damage in rat primary hepatocytes. *Toxicol Vitro* 24 (2), 516–522. doi:10.1016/j.tiv.2009.03.006

Liu, S., Tian, L., Chai, G., Wen, B., and Wang, B. (2018). Targeting heme oxygenase-1 by quercetin ameliorates alcohol-induced acute liver injury via inhibiting NLRP3 inflammasome activation. *Food Funct.* 9 (8), 4184–4193. doi:10.1039/c8fo00650d

Liu, S. Y., Tsai, I. T., and Hsu, Y. C. (2021a). Alcohol-related liver disease: basic mechanisms and clinical perspectives. *Int. J. Mol. Sci.* 22 (10), 5170. doi:10.3390/ ijms22105170

Liu, Y., Chen, X., Qiu, M., Chen, W., Zeng, Z., and Chen, Y. (2014). Emodin ameliorates ethanol-induced fatty liver injury in mice. *Pharmacology* 94 (1-2), 71-77. doi:10.1159/000363413

Liu, Y. S., Yuan, M. H., Zhang, C. Y., Liu, H. M., Liu, J. R., Wei, A. L., et al. (2021b). Puerariae Lobatae radix flavonoids and puerarin alleviate alcoholic liver injury in zebrafish by regulating alcohol and lipid metabolism. *Biomed. Pharmacother.* 134, 111121. doi:10.1016/j.biopha.2020.111121

Liver, E. A. f.t.S. o. (2012). EASL clinical practical guidelines: management of alcoholic liver disease. *J. Hepatol.* 57 (2), 399-420. doi:10.1016/j.jhep.2012. 04.004

Louvet, A., Diaz, E., Dharancy, S., Coevoet, H., Texier, F., Thévenot, T., et al. (2008). Early switch to pentoxifylline in patients with severe alcoholic hepatitis is inefficient in non-responders to corticosteroids. *J. Hepatol.* 48 (3), 465–470. doi:10.1016/j.jhep.2007. 10.010

Louvet, A., Naveau, S., Abdelnour, M., Ramond, M. J., Diaz, E., Fartoux, L., et al. (2007). The Lille model: a new tool for therapeutic strategy in patients with severe alcoholic hepatitis treated with steroids. *Hepatology* 45 (6), 1348–1354. doi:10.1002/hep. 21607

Lu, Y., and Cederbaum, A. I. (2018). Cytochrome P450s and alcoholic liver disease. Curr. Pharm. Des. 24 (14), 1502–1517. doi:10.2174/1381612824666180410091511

Lv, P., Zhu, F., Lv, M., Bao, J., Lv, L., Xu, X., et al. (2023). Comparative analysis of the liver-protective effects of raw and stir-fried semen of Hovenia dulcis in rats via gas chromatography-mass spectrometry-based serum metabolomic profiling and chemometrics. *Biomed. Chromatogr.* 37 (4), e5578. doi:10.1002/bmc.5578

Mason, B. J. (2017). Emerging pharmacotherapies for alcohol use disorder. *Neuropharmacology* 122, 244-253. doi:10.1016/j.neuropharm.2017.04.032

Mathurin, P., Duchatelle, V., Ramond, M. J., Degott, C., Bedossa, P., Erlinger, S., et al. (1996). Survival and prognostic factors in patients with severe alcoholic hepatitis treated with prednisolone. *Gastroenterology* 110 (6), 1847–1853. doi:10.1053/gast.1996.v110. pm8964410

Matsuyama, S., Palmer, J., Bates, A., Poventud-Fuentes, I., Wong, K., Ngo, J., et al. (2016). Bax-induced apoptosis shortens the life span of DNA repair defect Ku70knockout mice by inducing emphysema. *Exp. Biol. Med. (Maywood)* 241 (12), 1265–1271. doi:10.1177/1535370216654587

Mayo-Smith, M. F., Beecher, L. H., Fischer, T. L., Gorelick, D. A., Guillaume, J. L., Hill, A., et al. (2004). Management of alcohol withdrawal delirium. An evidence-based practice guideline. Arch. Intern Med. 164 (13), 1405–1412. doi:10.1001/archinte.164.13. 1405

McClain, C. J., Barve, S. S., Barve, A., and Marsano, L. (2011). Alcoholic liver disease and malnutrition. *Alcohol Clin. Exp. Res.* 35 (5), 815–820. doi:10.1111/j.1530-0277.2010. 01405.x

Mellinger, J. L., and Stine, J. G. (2020). Early liver transplantation for severe alcoholic hepatitis. Dig. Dis. Sci. 65 (6), 1608-1614. doi:10.1007/s10620-020-06159-9

Meng, X., Tang, G. Y., Zhao, C. N., Liu, Q., Xu, X. Y., and Cao, S. Y. (2020). Hepatoprotective effects of Hovenia dulcis seeds against alcoholic liver injury and related mechanisms investigated via network pharmacology. *World J. Gastroenterol.* 26 (24), 3432–3446. doi:10.3748/wjg.v26.i24.3432

Miyata, T., and Nagy, L. E. (2020). Programmed cell death in alcohol-associated liver disease. *Clin. Mol. Hepatol.* 26 (4), 618–625. doi:10.3350/cmh.2020.0142

Mo, Y., Thomas, M. C., and Karras, G. E., Jr. (2016). Barbiturates for the treatment of alcohol withdrawal syndrome: a systematic review of clinical trials. *J. Crit. Care* 32, 101–107. doi:10.1016/j.jcrc.2015.11.022

Molina, M. F., Sanchez-Reus, I., Iglesias, I., and Benedi, J. (2003). Quercetin, a flavonoid antioxidant, prevents and protects against ethanol-induced oxidative stress in mouse liver. *Biol. Pharm. Bull.* 26 (10), 1398–1402. doi:10.1248/bpb.26.1398

Morgan, W. D. (1989). Transcription factor Sp1 binds to and activates a human hsp70 gene promoter. *Mol. Cell Biol.* 9 (9), 4099–4104. doi:10.1128/mcb.9.9.4099

Morley, K. C., Leung, S., Baillie, A., and Haber, P. S. (2013). The efficacy and biobehavioural basis of baclofen in the treatment of alcoholic liver disease (BacALD): study protocol for a randomised controlled trial. *Contemp. Clin. Trials* 36 (2), 348–355. doi:10.1016/j.cct.2013.08.002

Noh, B. K., Lee, J. K., Jun, H. J., Lee, J. H., Jia, Y., Hoang, M. H., et al. (2011). Restoration of autophagy by puerarin in ethanol-treated hepatocytes via the activation of AMP-activated protein kinase. *Biochem. Biophys. Res. Commun.* 414 (2), 361–366. doi:10.1016/j.bbrc.2011.09.077

Nössing, C., and Ryan, K. M. (2023). 50 years on and still very much alive: 'Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br. J. Cancer* 128 (3), 426–431. doi:10.1038/s41416-022-02020-0

Pang, Y., Xu, X., Xiang, X., Li, Y., Zhao, Z., Li, J., et al. (2021). High fat activates O-GlcNAcylation and affects AMPK/ACC pathway to regulate lipid metabolism. *Nutrients* 13 (6), 1740. doi:10.3390/nu13061740

Park, M. N., Rahman, M. A., Rahman, M. H., Kim, J. W., Choi, M., Kim, J. W., et al. (2022). Potential therapeutic implication of herbal medicine in mitochondria-mediated oxidative stress-related liver diseases. *Antioxidants (Basel)* 11 (10), 2041. doi:10.3390/ antiox11102041

Patricia, M., Dias, M. I., Calhella, R. C., Sánchez-Mata, M. C., Santos-Buelga, C., Barros, L., et al. (2017). Hovenia dulcis Thunb. pseudofruits as functional foods: phytochemicals and bioactive properties in different maturity stages. *Funct. Foods* 29, 37–45. doi:10.1016/j.jff.2016.12.003

Peng, J. H., Cui, T., Huang, F., Chen, L., Zhao, Y., Xu, L., et al. (2013). Puerarin ameliorates experimental alcoholic liver injury by inhibition of endotoxin gut leakage, Kupffer cell activation, and endotoxin receptors expression. *J. Pharmacol. Exp. Ther.* 344 (3), 646–654. doi:10.1124/jpet.112.201137

Pingili, R. B., Challa, S. R., Pawar, A. K., Toleti, V., Kodali, T., and Koppula, S. (2020). A systematic review on hepatoprotective activity of quercetin against various drugs and toxic agents: evidence from preclinical studies. *Phytother. Res.* 34 (1), 5–32. doi:10.1002/ ptr.6503

Pistritto, G., Trisciuoglio, D., Ceci, C., Garufi, A., and D'Orazi, G. (2016). Apoptosis as anticancer mechanism: function and dysfunction of its modulators and targeted therapeutic strategies. *Aging (Albany NY)* 8 (4), 603–619. doi:10.18632/aging.100934

Qiu, P., Dong, Y., Li, B., Kang, X. J., Gu, C., Zhu, T., et al. (2017). Dihydromyricetin modulates p62 and autophagy crosstalk with the Keap-1/Nrf2 pathway to alleviate ethanol-induced hepatic injury. *Toxicol. Lett.* 274, 31–41. doi:10.1016/j.toxlet.2017. 04.009

Qiu, P., Dong, Y., Zhu, T., Luo, Y. Y., Kang, X. J., Pang, M. X., et al. (2019). Semen hoveniae extract ameliorates alcohol-induced chronic liver damage in rats via modulation of the abnormalities of gut-liver axis. *Phytomedicine* 52, 40–50. doi:10. 1016/j.phymed.2018.09.209

Ralli, T., Neupane, Y. R., Saifi, Z., and Kohli, K. (2021). Gut microbiota as an emerging therapeutic avenue for the treatment of nonalcoholic fatty liver disease. *Curr. Pharm. Des.* 27 (46), 4677–4685. doi:10.2174/1389201022666210625141526

Ramond, M. J., Poynard, T., Rueff, B., Mathurin, P., Théodore, C., Chaput, J. C., et al. (1992). A randomized trial of prednisolone in patients with severe alcoholic hepatitis. *N. Engl. J. Med.* 326 (8), 507–512. doi:10.1056/nejm199202203260802

Rao, R. (2009). Occludin phosphorylation in regulation of epithelial tight junctions. *Ann. N. Y. Acad. Sci.* 1165, 62–68. doi:10.1111/j.1749-6632.2009.04054.x

Ronnett, G. V., Kim, E. K., Landree, L. E., and Tu, Y. (2005). Fatty acid metabolism as a target for obesity treatment. *Physiol. Behav.* 85 (1), 25–35. doi:10.1016/j.physbeh.2005.04.014

Salete-Granado, D., Carbonell, C., Puertas-Miranda, D., Vega-Rodríguez, V. J., García-Macia, M., Herrero, A. B., et al. (2023). Autophagy, oxidative stress, and alcoholic liver disease: a systematic review and potential clinical applications. *Antioxidants (Basel)* 12 (7), 1425. doi:10.3390/antiox12071425

Seitz, H. K., Moreira, B., and Neuman, M. G. (2023). Pathogenesis of alcoholic fatty liver a narrative review. *Life (Basel)* 13 (8), 1662. doi:10.3390/life13081662

Seo, H. J., Jeong, K. S., Lee, M. K., Park, Y. B., Jung, U. J., Kim, H. J., et al. (2003). Role of naringin supplement in regulation of lipid and ethanol metabolism in rats. *Life Sci.* 73 (7), 933–946. doi:10.1016/s0024-3205(03)00358-8

Sferrazza, G., Brusotti, G., Zonfrillo, M., Temporini, C., Tengattini, S., Bononi, M., et al. (2021). Hovenia dulcis thumberg: phytochemistry, pharmacology, toxicology and

regulatory framework for its use in the European union. *Molecules* 26 (4), 903. doi:10. 3390/molecules26040903

Shang Zeng, S. L., and Li, Q. (2022). Progress in research of alcoholic liver disease. World Chin. J. Dig. 30 (12), 535–540. doi:10.11569/wcjd.v30.i12.535

Shen, Y., Lindemeyer, A. K., Gonzalez, C., Shao, X. M., Spigelman, I., Olsen, R. W., et al. (2012). Dihydromyricetin as a novel anti-alcohol intoxication medication. *J. Neurosci.* 32 (1), 390–401. doi:10.1523/jneurosci.4639-11.2012

Sidhu, S. S., Goyal, O., Singla, P., Gupta, D., Sood, A., Chhina, R. S., et al. (2012). Corticosteroid plus pentoxifylline is not better than corticosteroid alone for improving survival in severe alcoholic hepatitis (COPE trial). *Dig. Dis. Sci.* 57 (6), 1664–1671. doi:10.1007/s10620-012-2097-4

Silva, J., Spatz, M. H., Folk, C., Chang, A., Cadenas, E., Liang, J., et al. (2021). Dihydromyricetin improves mitochondrial outcomes in the liver of alcohol-fed mice via the AMPK/Sirt-1/PGC-1α signaling axis. *Alcohol* 91, 1–9. doi:10.1016/j.alcohol.2020. 10.002

Silva, J., Yu, X., Moradian, R., Folk, C., Spatz, M. H., Kim, P., et al. (2020). Dihydromyricetin protects the liver via changes in lipid metabolism and enhanced ethanol metabolism. *Alcohol Clin. Exp. Res.* 44 (5), 1046–1060. doi:10.1111/acer.14326

Sindhu, E. R., Firdous, A. P., Preethi, K. C., and Kuttan, R. (2010). Carotenoid lutein protects rats from paracetamol-carbon tetrachloride- and ethanol-induced hepatic damage. *J. Pharm. Pharmacol.* 62 (8), 1054–1060. doi:10.1111/j.2042-7158.2010.01123.x

Singal, A. K., Bataller, R., Ahn, J., Kamath, P. S., and Shah, V. H. (2018). ACG clinical guideline: alcoholic liver disease. *Am. J. Gastroenterol.* 113 (2), 175–194. doi:10.1038/ajg. 2017.469

Skotnicová, A., Boubínová, G., Boštíková, Z., Dušková, Š., Šulc, M., Kutinová-Canová, N., et al. (2020). Does dihydromyricetin impact on alcohol metabolism. *Physiol. Res.* 69 (Suppl. 4), S573–s581. doi:10.33549/physiolres.934606

Song, L., Chen, T. Y., Zhao, X. J., Xu, Q., Jiao, R. Q., Li, J. M., et al. (2019). Pterostilbene prevents hepatocyte epithelial-mesenchymal transition in fructose-induced liver fibrosis through suppressing miR-34a/Sirt1/p53 and TGF- β 1/Smads signalling. *Br. J. Pharmacol.* 176 (11), 1619–1634. doi:10.1111/bph.14573

Su, Z. W., Yan, T. Y., Feng, J., Zhang, M. Y., Han, L., Zhang, H. F., et al. (2023). Protective effects and mechanism of polysaccharides from edible medicinal plants in alcoholic liver injury: a review. *Int. J. Mol. Sci.* 24 (22), 16530. doi:10.3390/ ijms242216530

Suzuki, T. (2020). Regulation of the intestinal barrier by nutrients: the role of tight junctions. *Anim. Sci. J.* 91 (1), e13357. doi:10.1111/asj.13357

Tang, D., Chen, X., Kang, R., and Kroemer, G. (2021). Ferroptosis: molecular mechanisms and health implications. *Cell Res.* 31 (2), 107–125. doi:10.1038/s41422-020-00441-1

Tang, H., Sun, Q., Wang, T., Liao, Y., Wang, G., Zhao, F., et al. (2019). Upregulation of CYP2E1 expression causes oxidative damage induced by 2-chloroethanol in primary cultured rat astrocytes. *Neurotoxicology* 75, 233–244. doi:10.1016/j.neuro.2019.09.016

Tang, Y., Gao, C., Xing, M., Li, Y., Zhu, L., Wang, D., et al. (2012). Quercetin prevents ethanol-induced dyslipidemia and mitochondrial oxidative damage. *Food Chem. Toxicol.* 50 (5), 1194–1200. doi:10.1016/j.fct.2012.02.008

Tang, Z., Ding, Y., Zhang, W., Zhang, R., Zhang, L., Wang, M., et al. (2023). Epidemiological characteristics of alcohol-related liver disease in China: a systematic review and meta-analysis. *BMC Public Health* 23 (1), 1276. doi:10.1186/s12889-023-15645-4

Tyagi, P., Sharma, P., Sharma, B. C., Puri, A. S., Kumar, A., and Sarin, S. K. (2011). Prevention of hepatorenal syndrome in patients with cirrhosis and ascites: a pilot randomized control trial between pentoxifylline and placebo. *Eur. J. Gastroenterol. Hepatol.* 23 (3), 210–217. doi:10.1097/MEG.0b013e3283435d76

Vidhya, A., and Indira, M. (2009). Protective effect of quercetin in the regression of ethanol-induced hepatotoxicity. *Indian J. Pharm. Sci.* 71 (5), 527–532. doi:10.4103/0250-474x.58186

Voutilainen, S. H., Kosola, S. K., Lohi, J., Mutka, A., Jahnukainen, T., Pakarinen, M., et al. (2020). Expression of 6 biomarkers in liver grafts after pediatric liver transplantation: correlations with histology, biochemistry, and outcome. *Ann. Transpl.* 25, e925980. doi:10.12659/aot.925980

Wang, F., Liu, J. C., Zhou, R. J., Zhao, X., Liu, M., Ye, H., et al. (2017). Apigenin protects against alcohol-induced liver injury in mice by regulating hepatic CYP2E1mediated oxidative stress and PPARα-mediated lipogenic gene expression. *Chem. Biol. Interact.* 275, 171–177. doi:10.1016/j.cbi.2017.08.006

Wang, M., Sun, J., Jiang, Z., Xie, W., and Zhang, X. (2015). Hepatoprotective effect of kaempferol against alcoholic liver injury in mice. *Am. J. Chin. Med.* 43 (2), 241–254. doi:10.1142/s0192415x15500160

Wang, Y., Wang, J., Xiang, H., Ding, P., Wu, T., and Ji, G. (2022). Recent update on application of dihydromyricetin in metabolic related diseases. *Biomed. Pharmacother.* 148, 112771. doi:10.1016/j.biopha.2022.112771

Wierzbicki, A. S., and Mikhailidis, D. P. (2002). Beyond LDL-C--the importance of raising HDL-C. *Curr. Med. Res. Opin.* 18 (1), 36–44. doi:10.1185/030079902125000165

Wu, J., Wang, Y., Jiang, R., Xue, R., Yin, X., Wu, M., et al. (2021). Ferroptosis in liver disease: new insights into disease mechanisms. *Cell Death Discov.* 7 (1), 276. doi:10. 1038/s41420-021-00660-4

Wu, X., Fan, X., Miyata, T., Kim, A., Cajigas-Du Ross, C. K., Ray, S., et al. (2023). Recent advances in understanding of pathogenesis of alcohol-associated liver disease. *Annu. Rev. Pathol.* 18, 411–438. doi:10.1146/annurev-pathmechdis-031521-030435

Xing, C., Xu, L., and Yao, Y. (2021). Beneficial role of oleuropein in sepsis-induced myocardial injury. Possible Involvement of GSK- 3β /NF-kB pathway. *Acta Cir. Bras.* 36 (1), e360107. doi:10.1590/acb360107

Y, J. F. C. (2014). Research on alcoholic liver disease. Med. Inf. 27 (7), 670-671.

Yao, K. N., Feng, C. Z., and Zhu, X. H. (2017). Regulatory roles and mechanisms of hovenia dulcis thunb toward polarization of macrophage RAW264.7. *Int. J. Clin. Exp. Med.* 10 (2), 2137–2146.

Yao, P., Hao, L., Nussler, N., Lehmann, A., Song, F., Zhao, J., et al. (2009). The protective role of HO-1 and its generated products (CO, bilirubin, and Fe) in ethanol-induced human hepatocyte damage. *Am. J. Physiol. Gastrointest. Liver Physiol.* 296 (6), G1318–G1323. doi:10.1152/ajpgi.00555.2007

Yao, P., Nussler, A., Liu, L., Hao, L., Song, F., Schirmeier, A., et al. (2007). Quercetin protects human hepatocytes from ethanol-derived oxidative stress by inducing heme oxygenase-1 via the MAPK/Nrf2 pathways. *J. Hepatol.* 47 (2), 253–261. doi:10.1016/j. jhep.2007.02.008

Yu, X., Xu, Y., Zhang, S., Sun, J., Liu, P., Xiao, L., et al. (2016). Quercetin attenuates chronic ethanol-induced hepatic mitochondrial damage through enhanced mitophagy. *Nutrients* 8 (1), 27. doi:10.3390/nu8010027

Yuan, Y., Chen, T., Yongqiang, G., Zhe, W., Meizi, P., Qin, Y., et al. (2022). Generation of mitochondria-rich kidney organoids from expandable intermediate mesoderm progenitors reprogrammed from human urine cells under defined medium. *Food Sci.* 43 (23), 174–179. doi:10.1186/s13578-022-00909-0

Zeng, H., Guo, X., Zhou, F., Xiao, L., Liu, J., Jiang, C., et al. (2019). Quercetin alleviates ethanol-induced liver steatosis associated with improvement of lipophagy. *Food Chem. Toxicol.* 125, 21–28. doi:10.1016/j.fct.2018.12.028

Zhang, J., Chen, Y., Luo, H., Sun, L., Xu, M., Yu, J., et al. (2018). Recent update on the pharmacological effects and mechanisms of dihydromyricetin. *Front. Pharmacol.* 9, 1204. doi:10.3389/fphar.2018.01204

Zhao, L., Wang, Y., Liu, J., Wang, K., Guo, X., Ji, B., et al. (2016). Protective effects of genistein and puerarin against chronic alcohol-induced liver injury in mice via antioxidant, anti-inflammatory, and anti-apoptotic mechanisms. *J. Agric. Food Chem.* 64 (38), 7291–7297. doi:10.1021/acs.jafc.6b02907

Zhao, L., Zhang, N., Yang, D., Yang, M., Guo, X., He, J., et al. (2018). Protective effects of five structurally diverse flavonoid subgroups against chronic alcohol-induced hepatic damage in a mouse model. *Nutrients* 10 (11), 1754. doi:10.3390/nu10111754

Zhao, M., Du, Y. Q., Yuan, L., and Wang, N. N. (2010). Protective effect of puerarin on acute alcoholic liver injury. *Am. J. Chin. Med.* 38 (2), 241–249. doi:10.1142/s0192415x10007816

Zhao, S., Zhang, Y., Ding, H., Hu, S., Wu, X., Ma, A., et al. (2023). Lutein prevents liver injury and intestinal barrier dysfunction in rats subjected to chronic alcohol intake. *Nutrients* 15 (5), 1229. doi:10.3390/nu15051229

Zhao, X., Gong, L., Wang, C., Liu, M., Hu, N., Dai, X., et al. (2021). Quercetin mitigates ethanol-induced hepatic steatosis in zebrafish via P2X7R-mediated PI3K/Keap1/ Nrf2 signaling pathway. *J. Ethnopharmacol.* 268, 113569. doi:10.1016/j.jep.2020.113569

Zhao, X., Wang, C., Dai, S., Liu, Y., Zhang, F., Peng, C., et al. (2022). Quercetin protects ethanol-induced hepatocyte pyroptosis via scavenging mitochondrial ROS and promoting PGC-1a-regulated mitochondrial homeostasis in L02 cells. *Oxid. Med. Cell Longev.* 2022, 4591134. doi:10.1155/2022/4591134

Zheng, F., and Cai, Y. (2019). Concurrent exercise improves insulin resistance and nonalcoholic fatty liver disease by upregulating PPAR- γ and genes involved in the beta-oxidation of fatty acids in ApoE-KO mice fed a high-fat diet. *Lipids Health Dis.* 18 (1), 6. doi:10.1186/s12944-018-0933-z

Zhou, B., Jiang, Z., Li, X., and Zhang, X. (2018). Kaempferol's protective effect on ethanol-induced mouse primary hepatocytes injury involved in the synchronous inhibition of SP1, Hsp70 and CYP2E1. *Am. J. Chin. Med.* 46 (5), 1093–1110. doi:10. 1142/s0192415x1850057x

Zhu, X., Bian, H., Wang, L., Sun, X., Xu, X., Yan, H., et al. (2019). Berberine attenuates nonalcoholic hepatic steatosis through the AMPK-SREBP-1c-SCD1 pathway. *Free Radic. Biol. Med.* 141, 192–204. doi:10.1016/j.freeradbiomed.2019.06.019

Zwanziger, D., Rakov, H., Engels, K., Moeller, L. C., and Führer, D. (2015). Sexdependent claudin-1 expression in the liver of euthyroid and hypothyroid mice. *Eur. Thyroid. J.* 4 (Suppl. 1), 67–73. doi:10.1159/000431316

Glossary

Glossar	У	HDL-C	high-density lipoprotein
		Нерс	ferromodulin
ACC	acetyl-CoA carboxylase	HO-1	heme oxygenase-1
ADH	alcohol dehydrogenase	HSCs	hepatic stellate cells
AH	alcoholic hepatitis	HSP70	heat shock protein 70
ALD	alcohol-associated liver disease	IL-1ß	interleukin-1β
ALDH	acetaldehyde dehydrogenase	r IL-10	interleukin 10
АМРК	AMP-activated protein kinase	iNOS	inducible nitric oxide synthase
AP-1	activator protein-1	LC3	, microtubule-associated protein 1 light chain 3
ARE	antioxidant response element	LDL-C	low-density lipoprotein cholesterol
a-SMA	alpha-smooth muscle actin	LDs	lipid droplets
ATP	adenosine triphosphate	LIP	labile iron pool
AUD	alcohol-use disorder	LPS	lipopolysaccharide
AWD	alcohol withdrawal delirium	LT	liver transplantation
AWS	alcohol withdrawal syndrome	МАРК	mitogen-activated protein kinase
BAC	blood alcohol concentration	MDA	malondialdehyde
BZDs	benzodiazepines	MEOS	microsomal enzyme oxidizing system
CAT	catalase	NAD	nicotinamide adenine dinucleotide
CNKI	the China National Knowledge Infrastructure	NADPH	nicotinamide adenine dinucleotide phosphate
СО	carbon monoxide	NK cells	natural killer cells
CPT-1	carnitine palmitoyltransferase-1	NLRP3	the NOD-like receptor thermal protein domain associated protein 3
CYP2E1	cytochrome P450 2E1	NMDA	N-methyl-D-aspartate
CYP450	cytochrome P450	NF-ĸB	nuclear factor kappa B
DGAT	diacylglycerol acyltransferase	Nrf2	nuclear erythroid 2-related factor 2
DHM	dihydromyricetin	P2X7R	the P2X7 receptor
DNA	deoxyribonucleic acid	РВ	phenobarbital
EASL	the European Association for the Study of the Liver	PLIN2	perilipin 2
ECM	extracellular matrix	PPAR	peroxisome proliferator-activated receptor
ER	endoplasmic reticulum	PPARa	peroxisome proliferative activated receptora
ERK	extracellular regulated protein kinases	ROS	reactive oxygen species
FAS	fatty acid synthase	ROS/RNS	reactive oxygen/reactive nitrogen species
GABA	gamma aminobutyric acid	SCD1	stearoyl-CoA desaturase 1
GABA-A	gamma aminobutyric acid type A	SCFAs	short chain fatty acid
GABAB	gamma aminobutyric acid type B	SOD	superoxide dismutase
GPX	glutathione peroxidase	SER	smooth endoplasmic reticulum
GR	glutathione reductase	SQSTM1/P62	sequestosome 1
GSH	glutathione	SREBP-1	sterol regulatory element binding protein-1 expression
HCC	hepatocellular carcinoma	STAT3	signal transducer and activator of transcription 3
HCV	hepatitis C	TC	cholesterol

TG	triglycerides
TGF-β1	transforming growth factor-β1
TJ	tight junctions
TNF-a	tumor necrosis factor-alpha
WHO	the world health organization
WKS	Wernicke-Korsakoff syndrome
WOS	web of science
Hovenia dulcis	Hovenia dulcis Thunb. (Rhamnaceae)
ZO	zonula occludens
8-OHdG	8-hydroxy-2-deoxyguanosine
ОН	hydroxyl radicals