Check for updates

OPEN ACCESS

EDITED BY Elisabetta Caiazzo, University of Naples Federico II, Italy

REVIEWED BY

Mohamed A. Saleh, University of Sharjah, United Arab Emirates Ahmed A. Elmarakby, Augusta University, United States

*CORRESPONDENCE Mahmoud M. El-Mas, mahmoud.elmas@alexu.edu.eg

RECEIVED 08 January 2023 ACCEPTED 17 April 2023 PUBLISHED 25 April 2023

CITATION

Morgaan HA, Sallam MY, El-Gowelli HM, El-Gowilly SM and El-Mas MM (2023), Preeclamptic programming unevenly perturbs inflammatory and renal vasodilatory outcomes of endotoxemia in rat offspring: modulation by losartan and pioglitazone. *Front. Pharmacol.* 14:1140020. doi: 10.3389/fphar.2023.1140020

COPYRIGHT

© 2023 Morgaan, Sallam, El-Gowelli, El-Gowilly and El-Mas. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Preeclamptic programming unevenly perturbs inflammatory and renal vasodilatory outcomes of endotoxemia in rat offspring: modulation by losartan and pioglitazone

Hagar A. Morgaan¹, Marwa Y. Sallam¹, Hanan M. El-Gowelli¹, Sahar M. El-Gowilly¹ and Mahmoud M. El-Mas^{1.2*}

¹Department of Pharmacology and Toxicology, Faculty of Pharmacy, Alexandria University, Alexandria, Egypt, ²Department of Pharmacology and Toxicology, College of Medicine, Health Sciences Center, Kuwait University, Kuwait City, Kuwait

Introduction: Preeclampsia (PE) enhances the vulnerability of adult offspring to serious illnesses. The current study investigated whether preeclamptic fetal programming impacts hemodynamic and renal vasodilatory disturbances in endotoxic adult offspring and whether these interactions are influenced by antenatal therapy with pioglitazone and/or losartan.

Methods: PE was induced by oral administration of L-NAME (50 mg/kg/day) for the last 7 days of pregnancy. Adult offspring was treated with lipopolysaccharides (LPS, 5 mg/kg) followed 4-h later by hemodynamic and renovascular studies.

Results: Tail-cuff measurements showed that LPS decreased systolic blood pressure (SBP) in male, but not female, offspring of PE dams. Moreover, PE or LPS reduced vasodilations elicited by acetylcholine (ACh, 0.01–7.29 nmol) or N-ethylcarboxamidoadenosine (NECA, 1.6–100 nmol) in perfused kidneys of male rats only. The latter effects disappeared in LPS/PE preparations, suggesting a postconditioning action for LPS against renal manifestation of PE. Likewise, elevations caused by LPS in serum creatinine and inflammatory cytokines (TNF α and IL-1 β) as well as in renal protein expression of monocyte chemoattractant protein-1 (MCP-1) and AT1 receptors were attenuated by the dual PE/LPS challenge. Gestational pioglitazone or losartan reversed the attenuated ACh/NECA vasodilations in male rats but failed to modify LPS hypotension or inflammation. The combined gestational pioglitazone/losartan therapy improved ACh/NECA vasodilations and eliminated the rises in serum IL-1 β and renal MCP-1 and AT1 receptors.

Abbreviations: ACh, Acetylcholine; AT1, Angiotensin II type 1 receptor; AUC, Area under the curve, 3, 3'diaminobenzidine; HR, Heart rate; iNOS, Inducible nitric oxide synthase; i.p, intraperitoneal; IL-1 β , Interleukin-1 beta; LPS, Lipopolysaccharides; MCP-1, Monocyte chemoattractant protein-1; NECA, N-ethylcarboxamidoadenosine; L-NAME, N^{\overline}-nitro-L-arginine methyl ester; PPAR- γ , Peroxisome proliferator-activated receptor; PE, Preeclampsia; RAS, Renin-angiotensin system; SBP, Systolic blood pressure; TNF α , Tumor necrosis factor-alpha.

Conclusion: Preeclamptic fetal programming of endotoxic hemodynamic and renal manifestations in adult offspring depends on animal sex and specific biological activity and are reprogrammed by antenatal pioglitazone/losartan therapy.

KEYWORDS

preeclampsia, endotoxemia, adult offspring, blood pressure, renal vasodilation, gestational therapy

1 Introduction

Preeclampsia (PE) is a serious complication of pregnancy that affects 2%-8% of pregnancies and is considered a leading cause of morbidity and mortality. Epidemiological studies indicate that PE accounts for approximately 20% of all maternal deaths in lowincome settings (McCarthy et al., 2018). PE is diagnosed by the presence of new onset hypertension >140/90 after 20 weeks of gestation associated with proteinuria and/or evidence of maternal acute kidney injury, liver impairment, and hematological and neurological signs (Brown et al., 2018). Offspring born to preeclamptic mothers are predisposed to elevated risks of cardiovascular disease, stroke, and mental disorders (Cheng and Sharma, 2016). Evidently, environment disruptions encountered during the fetal developmental phase play a seminal role in determining the offspring's vulnerability to chronic disorders during later stages of childhood and adulthood (Peixoto et al., 2018).

PE management aims at stabilization of mothers and improve fetal maturity till labor. Commonly used therapies include antihypertensives (e.g., hydralazine, labetalol, or nifedipine), anticonvulsants (e.g., magnesium sulfate), and corticosteroids to promote the development of the fetal lung (Scott et al., 2022). Considering the pathogenic role of renin-angiotensin system (RAS) in PE progression (Irani and Xia, 2008; Siddiqui et al., 2010), pharmacologic RAS inhibition was thought to improve PE manifestations. Doering et al. (1998) and Zhou et al. (2008) showed that the PE-associated proteinuria, hypertension, and reduced pup weights in rodents are improved after angiotensin AT1 receptor blockade by losartan. The Peroxisome proliferatoractivated receptor (PPAR-y) negatively modulates RAS signaling and thereby favorably contributes to uteroplacental vascular and metabolic development (McCarthy et al., 2011b; Lane et al., 2019; Nesti et al., 2021). In fact, the blockade of PPAR-y during gestation resulted in PE-like signs such as hypertension and endothelial dysfunction (McCarthy et al., 2011a). Moreover, PE complications are improved after treatment of PE dams with PPAR agonists like rosiglitazone or pioglitazone (McCarthy et al., 2011b; Allam and Masri, 2018; Lane et al., 2019).

Alternatively, sepsis is a life-threatening condition that results from exaggerated systemic immune and inflammatory responses to infections (Su et al., 2017). Systemic administration of LPS of the Gram-negative bacterial membrane is used to model the acute inflammatory response and tissue damage caused by sepsis (El-Mas et al., 2006; El-Lakany et al., 2018; Dickson and Lehmann, 2019). Clinical (Saia et al., 2015; Klein and Flanagan, 2016) and experimental studies (Losonczy et al., 2000) reveal that females are less vulnerable to immunological complications of endotoxemia than males. Likewise, we recently reported that endotoxic features of inflammation, hypotension, and cardiac autonomic neuropathy are demonstrated in male, but not age-matched female, rats (El-Lakany et al., 2018). Moreover, the exposure of rats to the LPS challenge causes sex-unrelated impairment in adenosinergically-mediated renovasclar vasodilations together with amplified mortality and renal inflammation in the male population (Wedn et al., 2020).

Considering the pivotal role of preeclamptic fetal programming in the escalated incidence of chronic illnesses in growing offspring (Cheng and Sharma, 2016; Peixoto et al., 2018), the current investigation determined whether the sexually-related inflammatory, hemodynamic, and renovascular vasodilator responsiveness to endotoxemia could be altered in the offspring of PE rats. Furthermore, given the positive modulation by PPARy of renoprotection prompted by AT1 receptor blockade (Jin and Pan, 2007; Namikoshi et al., 2008; Lai et al., 2011), we asked if antenatal treatment with the combined pioglitazone/losartan therapy would be more influential than individual drugs in reprogramming the developed endotoxic defects.

2 Materials and methods

2.1 Animals

Adult Wistar rats (180–240 g) were used in the current study. Animals were obtained from the Animal facility of the Faculty of Pharmacy, Alexandria University, Egypt, and were maintained under controlled laboratory conditions and allowed free access to standard rat chow and tap water. All experimental protocols were approved by the Institutional Animal Care and Use Committee, Alexandria University, Egypt (Approval No. AU06201957149) and carried out in accordance with the Declaration of Helsinki and the Guide for the Care and Use of Laboratory Animals.

2.2 PE induction

For the induction of PE, N^{ω}-nitro-L-arginine methyl ester (nitric oxide synthase inhibitor, L-NAME) (50 mg/kg/day) was administered via oral gavage for 7 consecutive days starting from day 14 of conception (Pandhi et al., 2001; Abuiessa et al., 2020). The measurements of SBP by the tail-cuff technique (see below) as well as urinary protein level were used to validate PE development.

2.3 The rat isolated perfused kidney

The isolated perfused kidney technique was carried out to assess renal vasodilator capacities to ACh and NECA according to the method described in previous studies (El-Mas et al., 2004; Gohar et al., 2014). The renal perfusion pressure was elevated by continuous infusion with the α_1 -adrenoceptor agonist phenylephrine (20 μ M). Cumulative dose response curves to bolus injections of ACh (0.01–7.29 nmol) and NECA (1.6–100 nmol) were done by direct injection into the perfusate line proximal to the kidney.

2.4 Tail-cuff plethysmography

Non-invasive SBP measurements for conscious pregnant rats and adult offspring using the tail-cuff technique and a computerized data acquisition system with LabChart-7 pro software (Power Lab 4/ 30, model ML866/P, AD Instruments, Bella Vista, Australia) (El-Mas et al., 2015).

2.5 Immunohistochemistry

The method described in our previous studies (Helmy et al., 2015; Abuiessa et al., 2020) was employed to measure the expression of the inflammatory cytokine MCP-1 and Ang II AT1 receptors in glomerular tissues as well as in outer medullary areas of tubular cortex. Sections (4 µm thick) of kidney were deparaffinized in xylene and rehydrated in a series of declining ethanol concentration (100, 95% and 70%). Heat-induced epitope retrieval was carried out by immersing slides in coplin jars containing 10 mM citrate buffer solution and incubated in a microwave at power 100 for 1 min then power 30 for 9 min. Endogenous peroxidases were blocked by 0.3% hydrogen peroxide for 10 min. The rabbit, anti-rat primary antibodies, AT1 (1 µg/µL, Thermo Scientific[®], Berlin, Germany and MCP-1 (1µg/µL, Thermo Scientific[®]) were diluted (1:300), applied to the slides and then sections were incubated at 4°C overnight. The secondary antibody (HRP conjugate) was applied for 30 min. The chromogen 3, 3'diaminobenzidine was prepared and applied as instructed by the manufacturer for protein visualization. Images of glomerular and tubular tissues were used to measure the percentage of chromogen 3, 3'-diaminobenzidine positive stained area in renal tissues.

2.6 Urine analyses

On gestational day 20, pregnant rats were kept in metabolic cages with mesh wire bottom made of stainless steel and allowed access to standard rat chow and water. The 24-h urine samples were collected under light mineral oil and stored at -80°C until processed (Abuiessa et al., 2020). Urinary protein levels were measured by the pyrogallol red method (Yalamati et al., 2016) using Cromatest standard kit (LiNEAR Chemicals, Spain) according to the manufacturer's guidelines.

2.7 Serum analyses

Retro-orbital blood samples were withdrawn from thiopental (50 mg/kg i. p.) anesthetized rats prior to kidney isolation. The collected blood was permitted to coagulate for 15 min at room temperature then centrifuged at 1,200 g for 10 min. The aspirated serum was stored at -80° C for subsequent ELISA determination of IL-1 β (Rat IL-1 β Platinum ELISA Kit, eBioscience TM BMS630, United States) and TNF α (Rat TNF alpha Platinum ELISA Kit, eBioscience TM BMS622, United States) according to the manufacturer's protocol. Creatinine was determined colorimetrically using commercially available kits (BioSystems, Spain).

2.8 Protocols and experimental groups

2.8.1 Hemodynamic and renal effects of endotoxemia in offspring of PE rats

A total of 8 offspring rat groups, 4 males and 4 females (n = 7-8 each, 10 weeks old), were employed in this experiment to investigate the influence of endotoxemia on hemodynamic and renal profiles in offspring of PE and non-PE mothers. The 4 groups of each rat sex were assigned as (i) saline-treated non-PE offspring, (ii) LPS-treated non-PE offspring, (iii) saline-treated PE offspring, and (iv) LPS-treated PE offspring. Endotoxemia was induced in adult offspring by *i. p.* administration of a 5 mg/kg dose of LPS (Lv et al., 2006; Abuiessa et al., 2020). Four hours later, SBP was measured by the tail-cuff technique and rats were anesthetized with thiopental (50 mg/kg i. p). Blood samples were withdrawn from retro orbital plexus for serum analyses, left kidneys were isolated and perfused for the assessment of renal vasodilator to ACh and NECA as detailed above and right kidneys were collected for immunohistochemical determination of MCP-1 and AT1 receptor expression in renal tubular and glomerular tissues.

2.8.2 Modulation by antenatal therapies of PE/LPS manifestations

This experiment investigated the effects of gestational administration of pioglitazone or losartan on hemodynamic and renal defects caused by PE or PE/LPS insult. Studies were performed in the male offspring only since the preceding experiment showed clearer and more significant PE/LPS interactions on renal vasodilations in male compared with female offspring. Losartan (10 mg/kg) and/or pioglitazone (5 mg/kg) were administered to pregnant rats along with L-NAME (50 mg/kg/day) for 7 consecutive days, starting from gestational day 14 till 20. Six groups of male offspring (n = 6-7 each, 10 weeks old) of PE mothers were used: (i) saline-treated male offspring of PE/losartan mothers, (ii) LPS-treated male offspring of PE/losartan mothers (iii) salinetreated male offspring of PE/pioglitazone mothers, (iv) LPS-treated male offspring of PE/pioglitazone mothers,(v) saline-treated male offspring of PE/losartan/pioglitazone mothers and (iv) LPS-treated male offspring of PE/losartan/pioglitazone mothers. LPS (5 mg/kg i. p.) was administered to all adult male offspring and 4 h later, rats were processed for measurement of SBP, blood collection, renovascular and immunohistochemical studies as described in the previous experiment.

2.9 Drugs

Lipopolysaccharide (LPS, from *E coli*, serotype 0111:B4), phenylephrine hydrochloride, 5 -N-ethylcarboxamidoadenosine (NECA, adenosine analogue), acetylcholine (ACh), N_{ω} -Nitro-Larginine methyl ester hydrochloride (L-NAME) (Sigma-Aldrich Co., St. Louis, MO, United States), losartan potassium, pioglitazone hydrochloride (PHARCO Pharmaceutical Co., Alexandria, Egypt), thiopental sodium (Biochemie, Vienna, Austria) and heparin (5000 IU/mL; Nile Pharmaceutical Co., Egypt) were purchased from commercial vendors. ACh and NECA were freshly prepared in distilled water and dimethyl sulfoxide (Loba Chemie Pvt Ltd, India), respectively. LPS, heparin and thiopental were dissolved in saline.

2.10 Statistics

Values are expressed as means \pm S.E.M. The vasodilatory responses to ACh and NECA were expressed as the percentage of the precontraction level induced by 20 µM phenylephrine. The cumulative vasodilatory effects of ACh and NECA were computed by calculating the area under the curve (AUC) for individual experiments using trapezoidal integration and zero line as the baseline. In immunohistochemical studies, the percentages of stained areas were estimated. The one-way ANOVA followed by the Tukey's *post hoc* test was used to assess statistical significance with probability levels <0.05.

3 Results

3.1 L-NAME provokes preeclamptic manifestations in pregnant dams

Daily injection of pregnant dams with L-NAME (50 mg/kg/day) for 7 consecutive days caused a significant rise in SBP compared with saline-treated (non-PE) group on gestational day 20 (138.5 \pm 1.9 mmHg vs. 111.9 \pm 1.3 mmHg). The 24-h urine samples collected on gestational day 20 showed a rise in urine protein levels in PE dams (282 \pm 32 mg/dL vs. 89 \pm 10 mg/dL) compared with control counterparts.

3.2 PE programming of hemodynamics and renal vasodilatory profiles in adult endotoxic offspring

The effects of PE on SBP, HR, and renal vasodilations caused by ACh (0.01–7.29 nmol) and NECA (1.6–100 nmol) in isolated perfused kidneys obtained from adult male offspring in absence and presence of endotoxic insult are shown in Figure 1. Compared to their respective non-PE group, a significant rise in SBP was observed in PE males (Figure 1A). Although the 4-h exposure of male offspring of non-PE dams to LPS (5 mg/kg, i. p) failed to modify SBP, the administration of the same LPS dose to male offspring of PE rats exhibited significant falls in SBP (Figure 1A) that were accompanied by acceleration of the heart (Figure 1D).

In male offspring, the renal vasodilatory responses to cumulative bolus injections of ACh (AUCs) in perfused kidneys were not affected by the PE insult but showed significant reductions in response to the 4-h LPS challenge (Figures 1B, E). To the contrary, the AUCs of NECA vasodilations were attenuated by PE and remained unaffected by LPS (Figures 1C,F). The individual suppressing actions of LPS or PE on ACh and NECA responses, respectively, disappeared in kidney preparations obtained from PE/LPS rats. In fact, the dose-dependent vasodilatory actions and AUCs for both vasodilators in the PE/LPS preparations were not statistically different from those of control non-PE, demonstrating a conditioning effect for the dual PE/LPS challenge against individual insults in male offspring (Figure 1).

Contrary to males, no significant changes in SBP (Figure 2A) or HR (Figure 2D) in response to PE and/or LPS insults were evident in female offspring. Moreover, the renal dose-response curves of ACh (Figure 2B) and AUCs (Figure 2E) were not influenced by PE programming, 4-h LPS exposure, or their combination. On the other hand, a facilitatory effect for LPS on NECA vasodilations was seen in renal preparations of female offspring obtained from non-PE, but not PE or PE/LPS, dams (Figures 2C,F).

3.3 Antenatal pioglitazone and losartan improve PE/LPS outcomes in male offspring

As noted earlier, this experiment was performed in the male offspring in which more significant PE/LPS interactions were observed. Urine analysis showed no statistical differences in urine protein levels between control (non-PE) and PE offspring (197 \pm 13 vs. 234 \pm 19 mg/dL). Moreover, similar levels of proteinuria were seen in PE rats treated gestationally with losartan, pioglitazone, or their combination (196 \pm 25, 162 \pm 16, and 205 \pm 11 mg/dL, respectively). On the other hand, losartan, pioglitazone, or their combination abolished the elevation in SBP caused by PE and restored SBP to values like those found in the non-PE offspring (Figure 3A). However, none of these antenatal therapies affected the drop in SBP demonstrated 4 h post-LPS. Figure 3B illustrates an increase in serum creatinine in LPS-treated male offspring of non-PE rats and this effect was reversed when examined in their LPStreated counterparts from PE dams. Likewise, no rises in serum creatinine occurred in male offspring of PE mothers receiving individual or combined antenatal therapies.

Renovascular studies showed that gestational administration of losartan, pioglitazone, or their combination increased ACh (Figure 4) and NECA (Figure 5) vasodilations in perfused kidneys obtained from PE dams. The amplified vasodilations were exemplified by the upward shifts in the dose-response curves of either vasodilator and significant rises in AUCs of the vasodilatory response (Figures 4, 5). The combined losartan/ pioglitazone therapy caused more exaggerated rises in ACh/ NECA responses in kidney preparations obtained from LPStreated PE dams (Figures 4, 5).

3.4 Antenatal losartan and pioglitazone reverse the inflammatory response

Figures 6-8 depicts the effect of antenatal therapies on circulating and renal cytokines in male offspring. Compared



Effect of PE on tail cuff systolic blood pressure (SBP, panel **A**), heart rate (HR, panel **D**), cumulative vasodilatory dose response curves of ACh and NECA (panels **B**, **C**) and AUCs of the vasodilatory dose response curves (panel **E**, **F**) in phenylephrine preconstricted isolated perfused kidneys of endotoxic male offspring, respectively 4 h post intraperitoneal injection of LPS (5 mg/kg) or equal volume of saline. Data are expressed as the mean \pm SEM of 7-8 measurements. ANOVA followed by the Tukey's *post hoc* was utilized to measure statistical significance. **p* < 0.05 vs. "Control", "*p* < 0.05 vs. "PE" and "*p* < 0.05 vs. "LPS".

with control rats, serum levels of TNF- α (Figure 6A) and IL-1 β (Figure 6B) exhibited significant increases by 3- and 15-fold, respectively, in response to the endotoxic, but not to the PE,

challenge. Moreover, immunohistochemical studies showed that the protein expression of renal tubular AT1 receptors (Figure 7A) and MCP-1 (Figure 7B) was increased by PE



Effect of PE on tail cuff systolic blood pressure (SBP, panel **A**), heart rate (HR, panel **D**), cumulative vasodilatory dose response curves of ACh and NECA (panels **B**, **C**) and AUCs of the vasodilatory dose response curves (panel **E**, **F**) in phenylephrine preconstricted isolated perfused kidneys of endotoxic female offspring, respectively 4 h post intraperitoneal injection of LPS (5 mg/kg) or equal volume of saline. Data are expressed as the mean \pm SEM of 7-8 measurements. ANOVA followed by the Tukey's *post hoc* was utilized to measure statistical significance. *p < 0.05 vs. "Control", "p < 0.05 vs. "LPS".

and LPS, respectively. Comparable changes in the expression of AT1 receptors and MCP-1 were observed in glomerular tissues (Figure 8). Such systemic/renal inflammatory signals incited by

either insult were mostly and significantly attenuated in male offspring exposed to the double PE/LPS challenge (Figures 6-8). In all settings, the residual rises in serum IL-1 β



(Figure 6B) and tubular (Figure 7B) and glomerular (Figure 8B) MCP-1 expression were neutralized in male offspring of dams treated prenatally with the combined losartan/pioglitazone therapy.

4 Discussion

This study reports on the sex specificity of PE fetal programming of hemodynamic and renovascular sequels of endotoxemia in adult rat offspring and the reprogramming potential of prenatal pioglitazone/losartan therapy on these interactions. The data showed that while the female offspring of PE dams was minimally affected, the male progeny exhibited significant rises in SBP, which turned into a robust hypotension when challenged with LPS. Moreover, the renal vasodilatory responses to ACh/NECA were reduced by PE or LPS in male, but not female, offspring and these effects disappeared in PE/LPS preparations. Compared with individual therapies, the simultaneous prenatal exposure to losartan and pioglitazone caused significantly greater enhancement of renal vasodilations and suppression of biomarkers of circulating and renal inflammation in the male population. It is concluded that gestational losartan/pioglitazone boosts the conditioning effect of the dual PE/LPS challenge against individual insults caused by PE or LPS.

PE is a multifactorial gestational disease that is commonly characterized by hypertension, proteinuria and fetal growth retardation (Brown et al., 2018). Various experimental models have been developed to induce PE-like phenotype and imitate the pathological changes observed in humans. In the present study, daily injection of gravid dams with the NOS inhibitor L-NAME for 7 consecutive days elevated SBP and proteinuria compared with control counterparts. This is consistent with previous studies in which gestational NOS inhibition replicated some hallmarks of PE (Yallampalli and Garfield, 1993; Molnár et al., 1994; Chen et al., 2019). Remarkably, NO has been recognized as an important mediator involved in hemodynamic adaptation during gestation (Li et al., 2012).

The current findings favor the presumption that the female offspring are more resistance to hemodynamic and renal complications of endotoxemia and PE because the hypertensive response to PE as well as the reductions caused by LPS and PE in renal vasodilation caused by ACh and NECA, respectively, were all manifest in the male offspring only. These data are consistent with published data that immune, inflammatory, and cardiovascular anomalies caused by PE (Abuiessa et al., 2020; Birukov et al., 2020) or sepsis (Losonczy et al., 2000; Chen et al., 2014; El-Lakany et al., 2018) are sexually dimorphic and display more aggressive profiles in male offspring. The female sex hormones have been implicated, at least in part, in the presumed favorable profile in the female compared with the male population (Grigore et al., 2008; El-Lakany et al., 2018). Nonetheless, contradictory data of no sex differences or even worsened cardiovascular and metabolic outcomes in females than in males in the setting of PE (Habib et al., 2021; Beckers et al., 2022; Kuciene and Dulskiene, 2022; Ushida et al., 2022) and sepsis (Wedn et al., 2020; Fitzgerald et al., 2021) have been also recognized. Such discrepancies can be accounted for by differences in animal species, age, disease model, and specific biological activity under investigation.

In two recent studies from our laboratory (Abuiessa et al., 2020; Abuiessa et al., 2021), we provided the first experimental evidence that the exposure of weaning PE mothers or their male offspring to the endotoxic challenge intensifies cardiac autonomic neuropathy and upregulates the neuroinflammatory signal in the heart as well as in cardiovascular-sensitive nuclei of the brainstem. In the present study, we investigated whether the dual PE/LPS hit could elicit similar exaggerated damaging effects on renovascular reactivity in adult offspring. Contrary to our expectation, the present data in male offspring revealed advantageous sequels for the joint PE/LPS strategy on indices of renal function compared with individual offences. This view is supported by the observations that (i) the attenuated ACh or NECA renal vasodilations caused by LPS and PE, respectively, vanished in kidney preparations obtained from PE/LPS offspring, and (ii) the rises caused by LPS in serum creatinine, a biomarker of kidney injury, was eliminated in the PE-programmed offspring. It is tempting to speculate that the dual PE/LPS



challenge defies renal damage caused by individual insults in male offspring. In this circumstance, while PE programming acts conceivably as a preconditioning stimulus that counterbalances the adverse renal response to consequent endotoxemia, LPS offers a postconditional opportunity that lessens the renal injurious reaction caused by prior PE programming. Remarkably, the phenomenon of pre- or post-insult conditioning has been described in a variety of pathological states. For instance, LPS was found to reduce infarct volume following middle cerebral artery occlusion (Sardari et al., 2020). Postconditioning with repeated mild hypoxia promotes brain rehabilitation and protects neonatal hypoxia-ischemic rats from brain damage (Deng et al., 2018). LPS preconditioning protects against organ damage following ischemic reperfusion in cardiorenal and brain tissues (Ha et al., 2008; Vartanian et al., 2011; He et al., 2018).

The question whether alterations in inflammation and chemotaxis could underlie the renoprotection conferred by the PE/LPS challenge against individual insults in the male offspring was investigated. The inflammatory response to LPS was validated by the substantial increments in circulating TNF α and IL-1 β , two key proinflammatory cytokines that

characterize the early immune response to endotoxemia (Lu et al., 2008; Dickson and Lehmann, 2019). Additionally, the upregulated protein expression of MCP-1, a chemoattractant protein (Singh et al., 2021), in renal tubular and glomerular tissues suggests an enhanced recruitment of monocytes and other immune cells into renal tissues in endotoxic rats. Moreover, MCP-1 provokes systemic inflammation following TLR-4 activation and is readily elevated in animal models of sepsis (Ramnath et al., 2008). Amazingly, these intensified signals of inflammatory and chemoattractant molecules of the endotoxic response were significantly attenuated in rats challenged with the dual PE/LPS insults, suggesting a quashing influence for prior PE programming on cytokine/chemokine responses to endotoxemia. By the same token, this may as well provide the molecular basis for the capacity of the combined PE/LPS maneuver to alleviate the compromised renal vasodilation caused by individual insults. In a similar situation, the preconditioning and neuroprotective actions of LPS against ischemic stroke injury has been attributed to the upregulation of antiinflammatory type I interferonassociated genes and suppression of NFkB activity (Vartanian et al., 2011). Others have correlated the LPS-induced tolerance against renal ischemic reperfusion insult to the activation of



PPAR- γ and attenuation of oxidative stress or nitric oxide generation (Chatterjee et al., 2002; Collino et al., 2005).

Considering the key roles of RAS (Irani and Xia, 2008; Siddiqui et al., 2010) and its negative modulator PPAR-y (McCarthy et al., 2011b; Lane et al., 2019; Nesti et al., 2021) in PE progression, one prime objective of the current study was to investigate if antenatal exposure to a combined regimen of the AT1 receptor antagonist losartan plus the PPAR-y agonist pioglitazone would be more effective than separate therapies in reforming renovascular and inflammatory perturbations observed in the current model system. The rectifying effect of these pharmacologic therapies was verified by the ability of all three regimens to indiscriminately reverse the PE-associated suppression of renovascular dilations in male rats (Figure 5C). More relevantly, the combined losartan/pioglitazone regimen produced super-physiologic levels of renal vasodilations in PE/ LPS rats that were well above increments caused by individual therapies (Figure 4D; Figure 5D). This heightened vasodilatory response was associated with, and possibly motivated by, an intensified antiinflammatory potential as reflected by the elimination of the residual rises in circulating IL-1β and renal MCP-1. Our findings are echoed by published clinical and experimental data that PPAR γ activation enhances the renoprotective effect of AT1 receptor blockers in nephropathic states (Jin and Pan, 2007; Namikoshi et al., 2008; Lai et al., 2011). The negative modulation of RAS signaling (Lai et al., 2011) and oxidative and nitrative stresses (Kong et al., 2012) have been proposed as possible mechanisms for the enhanced renoprotection. Others have demonstrated that a regimen of losartan plus rosiglitazone exhibits additive renoprotection via proportional reductions in renal abundance of transforming growth factor- β and adhesion molecules (Lai et al., 2011).

The observation that gestational therapies failed to modify the hypotensive response to LPS in male offspring of PE dams despite the suppression of the concomitant inflammatory signal deserves a comment. Admittedly, the hypotensive response to endotoxemia is often set off by the developed state of systemic inflammation, which begins with the LPS-mediated upregulation of the TLR-4/NF κ B/TNF α /iNOS cascade and consequent overproduction of NO, diminution of vascular resistance, and widespread systemic vasodilation (El-Mas et al., 2008; Singh et al., 2022). Since the inflammatory response was obliterated by gestational therapies



particularly the combined losartan/pioglitazone regimen, it is likely that factors other than the inflammatory milieu might be responsible for the persistent fall in blood pressure. The upsurges in the renal vasodilatory propensity induced by individual, and more so by the combined, therapies may account, at least partly, for the maintained hypotensive response. The likelihood of this assumption gains credence from the reported intimate relationship between renal and vascular homeostasis (Kuczeriszka and Wąsowicz, 2022). Another possible mechanism may relate to renal AT1 receptors, whose expression was enhanced in PE offspring and restored to control levels after exposure to gestational losartan/pioglitazone therapy (Figure 7A). While the increased abundance of renal AT1 receptors is thought to contribute to the preeclamptic rise in blood pressure in this and other studies (Burke et al., 2016), the restoration of normal expression levels of the peptide receptors would help counterbalance the preeclamptic rises in blood pressure.

Despite the seemingly beneficial effects of targeting AT1 receptors (blockade) and PPAR- γ (activation) in the current study and those reported by others (Doering et al., 1998; Zhou et al.,



on immunohistochemical protein expressions of AT1 receptors (panel A) and MCP-1 (panel B) in renal tubular tissues of male offspring of PE or PE/LPS dams. The One-way ANOVA followed by the Tukey's *post hoc* was utilized to measure statistical significance. Values are expressed as means \pm S.E.M of 5 observations. **p* < 0.05 vs. "Control", "*p* < 0.05 vs. "LPS", "*p* < 0.05 vs. "PE", "*sp* < 0.05 vs. "PE/LPS" values. Representative images for immunostained renal tubular tissues are shown below bar graphs.

2008; McCarthy et al., 2011b; Lane et al., 2019), caution should be taken when extrapolating the current experimental data to the human setting. Arguably, the gestational use of AT1 receptor blockers has often been discouraged because of their teratogenic potential. Fetal RAS blockade is associated with renal failure, fetal growth retardation, pulmonary hypoplasia and limb contractures (Alwan et al., 2005; Bullo et al., 2012). Further, little information is available regarding the safety of thiazolidinediones like pioglitazone during pregnancy (Kung and Henry, 2012). It should be noted, however, that the utilization of drugs such as losartan and pioglitazone as experimental tools could help us unveil important insight into the understanding of the mutual interaction between AT1 receptor and PPAR-γ pathways in PE pathophysiology and reveal possible therapeutic clues for reprogramming renal defects induced by PE in adult offspring.

In conclusion, the study provides novel data that highlight a protective effect for the dual PE/LPS challenge against adverse renal consequences evoked by individual insults in adult offspring. The suppressed renal vasodilatory activity as well as the preconditioning and postconditioning influences of PE and LPS, respectively, are sexually differentiated and appeared in the male offspring only. Gestational therapy of PE dams with the combined losartan/



FIGURE 8

Effect of combined gestational losartan and pioglitazone therapy on immunohistochemical protein expressions of AT1 receptors (panel **A**) and MCP-1 (panel **B**) in renal glomerular tissues of male offspring of PE or PE/LPS dams. The One-way ANOVA followed by the Tukey's *post hoc* was utilized to measure statistical significance. Values are expressed as means \pm S.E.M of 5 observations. *p < 0.05 vs. "Control", "p < 0.05 vs. "LPS", "p < 0.05 vs. "PE/LPS" values. Representative images for immunostained renal glomerular tissues are shown below bar graphs.

pioglitazone regimen are more superior than individual therapies in suppressing the LPS-mediated escalation of the inflammatory state and depression of renovascular reactivity.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material further inquiries can be directed to the corresponding author.

References

Abuiessa, S. A., El-Gowilly, S. M., El-Gowelli, H. M., Helmy, M. M., and El-Mas, M. M. (2021). Short-lived sensitization of cardiovascular outcomes of postpartum endotoxemia in preeclamptic rats: Role of medullary solitary tract neuroinflammation. *Eur. J. Pharmacol.* 910, 174494. doi:10.1016/j.ejphar.2021.174494

Abuiessa, S. A., Wedn, A. M., El-Gowilly, S. M., Helmy, M. M., and El-Mas, M. M. (2020). Pre-eclamptic fetal programming alters neuroinflammatory and cardiovascular consequences of endotoxemia in sex-specific manners. *J. Pharmacol. Exp. Ther.* 373, 325–336. doi:10.1124/jpet.119.264192

Allam, H. I. G., and Masri, A. (2018). The potential therapeutic role of peroxisome ProliferatorActivated receptors agonist in Preeclamptic pregnant rats. *J. Coll. Physicians Surg. Pak* 28, 31–35. doi:10.29271/jcpsp.2018.01.31

Ethics statement

The animal study was reviewed and approved by The Institutional Animal Care and Use Committee, Alexandria University, Egypt (Approval No. AU06201957149).

Author contributions

All authors conceived and designed the research. HM conducted experiments. ME-M and HM analyzed the data and wrote the manuscript. All authors commented on previous versions of the manuscript and all authors read and approved the final manuscript. Funding acquisition by SE-G.

Funding

This study is supported by the Science and Technology Development Fund, Egypt (STDF Grant No. 37026). The funding body had no role in the design of the study, collection, analysis, interpretation of data or writing the manuscript.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fphar.2023.1140020/ full#supplementary-material

Alwan, S., Polifka, J. E., and Friedman, J. M. (2005). Angiotensin II receptor antagonist treatment during pregnancy. *Birth Defects Res. A Clin. Mol. Teratol.* 73, 123–130. doi:10.1002/bdra.20102

Beckers, K. F., Schulz, C. J., Flanagan, J. P., Adams, D. M., Gomes, V. C. L., Liu, C. C., et al. (2022). Sex-specific effects of maternal weight loss on offspring cardiometabolic outcomes in the obese preeclamptic-like mouse model, BPH/5. *Physiol. Rep.* 10, e15444. doi:10.14814/phy2.15444

Birukov, A., Herse, F., Nielsen, J. H., Kyhl, H. B., Golic, M., Kräker, K., et al. (2020). Blood pressure and angiogenic markers in pregnancy: Contributors to pregnancyinduced hypertension and offspring cardiovascular risk. *Hypertension* 76, 901–909. doi:10.1161/hypertensionaha.119.13966 Brown, M. A., Magee, L. A., Kenny, L. C., Karumanchi, S. A., Mccarthy, F. P., Saito, S., et al. (2018). The hypertensive disorders of pregnancy: ISSHP classification, diagnosis and management recommendations for international practice. *Pregnancy Hypertens.* 13, 291–310. doi:10.1016/j.preghy.2018.05.004

Bullo, M., Tschumi, S., Bucher, B. S., Bianchetti, M. G., and Simonetti, G. D. (2012). Pregnancy outcome following exposure to angiotensin-converting enzyme inhibitors or angiotensin receptor antagonists: A systematic review. *Hypertension* 60, 444–450. doi:10.1161/hypertensionaha.112.196352

Burke, S. D., Zsengellér, Z. K., Khankin, E. V., Lo, A. S., Rajakumar, A., Dupont, J. J., et al. (2016). Soluble fms-like tyrosine kinase 1 promotes angiotensin II sensitivity in preeclampsia. *J. Clin. Invest.* 126, 2561–2574. doi:10.1172/jci83918

Chatterjee, P. K., Zacharowski, K., Cuzzocrea, S., Brown, P. A., Stewart, K. N., Mota-Filipe, H., et al. (2002). Lipoteichoic acid from *Staphylococcus aureus* reduces renal ischemia/reperfusion injury. *Kidney Int.* 62, 1249–1263. doi:10.1111/j.1523-1755.2002. kid580.x

Chen, J., Chiazza, F., Collino, M., Patel, N. S., Coldewey, S. M., and Thiemermann, C. (2014). Gender dimorphism of the cardiac dysfunction in murine sepsis: Signalling mechanisms and age-dependency. *PLoS One* 9, e100631. doi:10.1371/journal.pone. 0100631

Chen, Y., Xue, F., Han, C., Yang, H., Han, L., Li, K., et al. (2019). Ferulic acid ameliorated placental inflammation and apoptosis in rat with preeclampsia. *Clin. Exp. Hypertens.* 41, 524–530. doi:10.1080/10641963.2018.1516773

Cheng, S. B., and Sharma, S. (2016). Preeclampsia and health risks later in life: An immunological link. *Semin. Immunopathol.* 38, 699–708. doi:10.1007/s00281-016-0579-8

Collino, M., Patel, N. S., Lawrence, K. M., Collin, M., Latchman, D. S., Yaqoob, M. M., et al. (2005). The selective PPARgamma antagonist GW9662 reverses the protection of LPS in a model of renal ischemia-reperfusion. *Kidney Int*. 68, 529–536. doi:10.1111/j. 1523-1755.2005.00430.x

Deng, Q., Chang, Y., Cheng, X., Luo, X., Zhang, J., and Tang, X. (2018). Postconditioning with repeated mild hypoxia protects neonatal hypoxia-ischemic rats against brain damage and promotes rehabilitation of brain function. *Brain Res. Bull.* 139, 31–37. doi:10.1016/j.brainresbull.2018.02.006

Dickson, K., and Lehmann, C. (2019). Inflammatory response to different toxins in experimental sepsis models. *Int. J. Mol. Sci.* 20, 4341. doi:10.3390/ijms20184341

Doering, T. P., Haller, N. A., Montgomery, M. A., Freeman, E. J., and Hopkins, M. P. (1998). The role of AT1 angiotensin receptor activation in the pathogenesis of preeclampsia. *Am. J. obstetrics Gynecol.* 178, 1307–1312. doi:10.1016/s0002-9378(98) 70337-0

El-Lakany, M. A., Fouda, M. A., El-Gowelli, H. M., El-Gowilly, S. M., and El-Mas, M. M. (2018). Gonadal hormone receptors underlie the resistance of female rats to inflammatory and cardiovascular complications of endotoxemia. *Eur. J. Pharmacol.* 823, 41–48. doi:10.1016/j.ejphar.2018.01.051

El-Mas, M. M., Fan, M., and Abdel-Rahman, A. A. (2008). Endotoxemia-mediated induction of cardiac inducible nitric-oxide synthase expression accounts for the hypotensive effect of ethanol in female rats. *J. Pharmacol. Exp. Ther.* 324, 368–375. doi:10.1124/jpet.107.127498

El-Mas, M. M., Helmy, M. W., Ali, R. M., and El-Gowelli, H. M. (2015). Celecoxib, but not indomethacin, ameliorates the hypertensive and perivascular fibrotic actions of cyclosporine in rats: Role of endothelin signaling. *Toxicol. Appl. Pharmacol.* 284, 1–7. doi:10.1016/j.taap.2015.01.018

El-Mas, M. M., Mohy El-Din, M. M., El-Gowilly, S. M., and Sharabi, F. M. (2004). Regional and endothelial differences in cyclosporine attenuation of adenosine receptormediated vasorelaxations. *J. Cardiovasc Pharmacol.* 43, 562–573. doi:10.1097/ 00005344-200404000-00012

El-Mas, M. M., Zhang, J., and Abdel-Rahman, A. A. (2006). Upregulation of vascular inducible nitric oxide synthase mediates the hypotensive effect of ethanol in conscious female rats. *J. Appl. Physiol. (1985)* 100, 1011–1018. doi:10.1152/japplphysiol.01058. 2005

Fitzgerald, J. C., Ross, M. E., Thomas, N. J., Weiss, S. L., Balamuth, F., Chilutti, M., et al. (2021). Association of early hypotension in pediatric sepsis with development of new or persistent acute kidney injury. *Pediatr. Nephrol.* 36, 451–461. doi:10.1007/ s00467-020-04704-2

Gohar, E. Y., El-Gowilly, S. M., El-Gowelli, H. M., El-Demellawy, M. A., and El-Mas, M. M. (2014). PI3K/Akt-independent NOS/HO activation accounts for the facilitatory effect of nicotine on acetylcholine renal vasodilations: Modulation by ovarian hormones. *PLoS One* 9, e95079. doi:10.1371/journal.pone.0095079

Grigore, D., Ojeda, N. B., and Alexander, B. T. (2008). Sex differences in the fetal programming of hypertension. *Gend. Med.* 5, S121–S132. doi:10.1016/j.genm.2008. 03.012

Ha, T., Hua, F., Liu, X., Ma, J., Mcmullen, J. R., Shioi, T., et al. (2008). Lipopolysaccharide-induced myocardial protection against ischaemia/reperfusion injury is mediated through a PI3K/Akt-dependent mechanism. *Cardiovasc Res.* 78, 546–553. doi:10.1093/cvr/cvn037

Habib, Y. H., Gowayed, M. A., Abdelhady, S. A., El-Deeb, N. M., Darwish, I. E., and El-Mas, M. M. (2021). Modulation by antenatal therapies of cardiovascular and renal programming in male and female offspring of preeclamptic rats. *Naunyn Schmiedeb. Arch. Pharmacol.* 394, 2273–2287. doi:10.1007/s00210-021-02146-7

He, K., Xia, L., and Zhang, J. (2018). LPS ameliorates renal ischemia/reperfusion injury via Hsp27 up-regulation. *Int. Urol. Nephrol.* 50, 571–580. doi:10.1007/s11255-017-1735-3

Helmy, M. M., Helmy, M. W., and El-Mas, M. M. (2015). Additive renoprotection by pioglitazone and fenofibrate against inflammatory, oxidative and apoptotic manifestations of cisplatin nephrotoxicity: Modulation by PPARs. *PLoS One* 10, e0142303. doi:10.1371/journal.pone.0142303

Irani, R. A., and Xia, Y. (2008). The functional role of the renin-angiotensin system in pregnancy and preeclampsia. *Placenta* 29, 763–771. doi:10.1016/j.placenta.2008.06.011

Jin, H. M., and Pan, Y. (2007). Renoprotection provided by losartan in combination with pioglitazone is superior to renoprotection provided by losartan alone in patients with type 2 diabetic nephropathy. *Kidney Blood Press Res.* 30, 203–211. doi:10.1159/000104089

Klein, S. L., and Flanagan, K. L. (2016). Sex differences in immune responses. Nat. Rev. Immunol. 16, 626–638. doi:10.1038/nri.2016.90

Kong, X., Zhang, Y., Wu, H. B., Li, F. X., Zhang, D. Y., and Su, Q. (2012). Combination therapy with losartan and pioglitazone additively reduces renal oxidative and nitrative stress induced by chronic high fat, sucrose, and sodium intake. *Oxid. Med. Cell Longev.* 2012, 856085. doi:10.1155/2012/856085

Kuciene, R., and Dulskiene, V. (2022). Associations of maternal gestational hypertension with high blood pressure and overweight/obesity in their adolescent offspring: A retrospective cohort study. *Sci. Rep.* 12, 3800. doi:10.1038/s41598-022-07903-z

Kuczeriszka, M., and Wąsowicz, K. (2022). Animal models of hypertension: The status of nitric oxide and oxidative stress and the role of the renal medulla. *Nitric Oxide* 125-126, 40–46. doi:10.1016/j.niox.2022.06.003

Kung, J., and Henry, R. R. (2012). Thiazolidinedione safety. Expert Opin. Drug Saf. 11, 565–579. doi:10.1517/14740338.2012.691963

Lai, K. N., Chan, L. Y., Guo, H., Tang, S. C., and Leung, J. C. (2011). Additive effect of PPAR- γ agonist and ARB in treatment of experimental IgA nephropathy. *Pediatr. Nephrol.* 26, 257–266. doi:10.1007/s00467-010-1703-y

Lane, S. L., Blair Dodson, R., Doyle, A. S., Park, H., Rathi, H., Matarrazo, C. J., et al. (2019). Pharmacological activation of peroxisome proliferator-activated receptor γ (PPAR- γ) protects against hypoxia-associated fetal growth restriction. *FASEB J.* 33, 8999–9007. doi:10.1096/fj.201900214R

Li, J., Lamarca, B., and Reckelhoff, J. F. (2012). A model of preeclampsia in rats: The reduced uterine perfusion pressure (RUPP) model. *Am. J. Physiol. Heart Circ. Physiol.* 303, H1–H8. doi:10.1152/ajpheart.00117.2012

Losonczy, G., Kriston, T., Szabó, A., Müller, V., Harvey, J., Hamar, P., et al. (2000). Male gender predisposes to development of endotoxic shock in the rat. *Cardiovasc Res.* 47, 183–191. doi:10.1016/s0008-6363(00)00075-4

Lu, Y. C., Yeh, W. C., and Ohashi, P. S. (2008). LPS/TLR4 signal transduction pathway. *Cytokine* 42, 145–151. doi:10.1016/j.cyto.2008.01.006

Lv, R., Zhou, Z. Q., Wu, H. W., Jin, Y., Zhou, W., and Xu, J. G. (2006). Hydroxyethyl starch exhibits antiinflammatory effects in the intestines of endotoxemic rats. *Anesth. Analg.* 103, 149–155. doi:10.1213/01.ane.0000220906.74517.99

Mccarthy, F. P., Drewlo, S., English, F. A., Kingdom, J., Johns, E. J., Kenny, L. C., et al. (2011a). Evidence implicating peroxisome proliferator-activated receptor-γ in the pathogenesis of preeclampsia. *Hypertension* 58, 882–887. doi:10.1161/HYPERTENSIONAHA.111.179440

Mccarthy, F. P., Drewlo, S., Kingdom, J., Johns, E. J., Walsh, S. K., and Kenny, L. C. (2011b). Peroxisome proliferator-activated receptor-γ as a potential therapeutic target in the treatment of preeclampsia. *Hypertension* 58, 280–286. doi:10.1161/HYPERTENSIONAHA.111.172627

Mccarthy, F. P., Ryan, R. M., and Chappell, L. C. (2018). Prospective biomarkers in preterm preeclampsia: A review. *Pregnancy Hypertens*. 14, 72–78. doi:10.1016/j.preghy. 2018.03.010

Molnár, M., Sütö, T., Tóth, T., and Hertelendy, F. (1994). Prolonged blockade of nitric oxide synthesis in gravid rats produces sustained hypertension, proteinuria, thrombocytopenia, and intrauterine growth retardation. *Am. J. Obstet. Gynecol.* 170, 1458–1466. doi:10.1016/s0002-9378(94)70179-2

Namikoshi, T., Tomita, N., Satoh, M., Haruna, Y., Kobayashi, S., Komai, N., et al. (2008). Pioglitazone enhances the antihypertensive and renoprotective effects of candesartan in Zucker obese rats fed a high-protein diet. *Hypertens. Res.* 31, 745–755. doi:10.1291/hypres.31.745

Nesti, L., Tricò, D., Mengozzi, A., and Natali, A. (2021). Rethinking pioglitazone as a cardioprotective agent: A new perspective on an overlooked drug. *Cardiovasc Diabetol.* 20, 109. doi:10.1186/s12933-021-01294-7

Pandhi, P., Saha, L., and Malhotra, S. (2001). Prolonged blockade of nitric oxide synthesis in pregnant rats as a model of pre-eclampsia. *Indian J. Pharmacol.* 33, 92–95.

Peixoto, A. B., Rolo, L. C., Nardozza, L. M. M., and Araujo Júnior, E. (2018). Epigenetics and preeclampsia: Programming of future outcomes. *Methods Mol. Biol.* 1710, 73–83. doi:10.1007/978-1-4939-7498-6_6

Ramnath, R. D., Ng, S. W., Guglielmotti, A., and Bhatia, M. (2008). Role of MCP-1 in endotoxemia and sepsis. Int. Immunopharmacol. 8, 810-818. doi:10.1016/j.intimp.2008.01.033

Saia, R. S., Garcia, F. M., and Cárnio, E. C. (2015). Estradiol protects female rats against sepsis induced by *Enterococcus faecalis* improving leukocyte bactericidal activity. *Steroids* 102, 17–26. doi:10.1016/j.steroids.2015.06.016

Sardari, M., Dzyubenko, E., Schmermund, B., Yin, D., Qi, Y., Kleinschnitz, C., et al. (2020). Dose-dependent microglial and astrocytic responses associated with postischemic neuroprotection after lipopolysaccharide-induced sepsis-like state in mice. *Front. Cell Neurosci.* 14, 26. doi:10.3389/fncel.2020.00026

Scott, G., Gillon, T. E., Pels, A., Von Dadelszen, P., and Magee, L. A. (2022). Guidelines-similarities and dissimilarities: A systematic review of international clinical practice guidelines for pregnancy hypertension. *Am. J. Obstet. Gynecol.* 226, S1222–s1236. doi:10.1016/j.ajog.2020.08.018

Siddiqui, A. H., Irani, R. A., Blackwell, S. C., Ramin, S. M., Kellems, R. E., and Xia, Y. (2010). Angiotensin receptor agonistic autoantibody is highly prevalent in preeclampsia: Correlation with disease severity. *Hypertension* 55, 386–393. doi:10. 1161/HYPERTENSIONAHA.109.140061

Singh, J., Lee, Y., and Kellum, J. A. (2022). A new perspective on NO pathway in sepsis and ADMA lowering as a potential therapeutic approach. *Crit. Care* 26, 246. doi:10. 1186/s13054-022-04075-0

Singh, S., Anshita, D., and Ravichandiran, V. (2021). MCP-1: Function, regulation, and involvement in disease. Int. Immunopharmacol. 101, 107598. doi:10.1016/j.intimp.2021.107598 Su, B. C., Huang, H. N., Lin, T. W., Hsiao, C. D., and Chen, J. Y. (2017). Epinecidin-1 protects mice from LPS-induced endotoxemia and cecal ligation and puncture-induced polymicrobial sepsis. *Biochim. Biophys. Acta Mol. Basis Dis.* 1863, 3028–3037. doi:10.1016/j.bbadis.2017.08.032

Ushida, T., Cotechini, T., Protopapas, N., Atallah, A., Collyer, C., Toews, A. J., et al. (2022). Aberrant inflammation in rat pregnancy leads to cardiometabolic alterations in the offspring and intrauterine growth restriction in the F2 generation. *J. Dev. Orig. Health Dis.* 13, 706-718. doi:10.1017/s2040174422000265

Vartanian, K. B., Stevens, S. L., Marsh, B. J., Williams-Karnesky, R., Lessov, N. S., and Stenzel-Poore, M. P. (2011). LPS preconditioning redirects TLR signaling following stroke: TRIF-IRF3 plays a seminal role in mediating tolerance to ischemic injury. *J. Neuroinflammation* 8, 140. doi:10.1186/1742-2094-8-140

Wedn, A. M., El-Gowilly, S. M., and El-Mas, M. M. (2020). Time and sex dependency of hemodynamic, renal, and survivability effects of endotoxemia in rats. *Saudi Pharm. J.* 28, 127–135. doi:10.1016/j.jsps.2019.11.014

Yalamati, P., Karra, M. L., and Bhongir, A. V. (2016). Comparison of urinary total proteins by four different methods. *Indian J. Clin. Biochem.* 31, 463–467. doi:10.1007/s12291-016-0551-3

Yallampalli, C., and Garfield, R. E. (1993). Inhibition of nitric oxide synthesis in rats during pregnancy produces signs similar to those of preeclampsia. *Am. J. Obstet. Gynecol.* 169, 1316–1320. doi:10.1016/0002-9378(93)90299-x

Zhou, C. C., Zhang, Y., Irani, R. A., Zhang, H., Mi, T., Popek, E. J., et al. (2008). Angiotensin receptor agonistic autoantibodies induce pre-eclampsia in pregnant mice. *Nat. Med.* 14, 855–862. doi:10.1038/nm.1856