



## OPEN ACCESS

## EDITED BY

Francesco Sessa,  
University of Catania, Italy

## REVIEWED BY

Charis Liapi,  
National and Kapodistrian University of  
Athens, Greece  
Ravi Sonkar,  
Boston University, United States

## \*CORRESPONDENCE

Nuriye Nuray Ulusu,  
✉ nulusu@ku.edu.tr

## SPECIALTY SECTION

This article was submitted to  
Experimental Pharmacology and  
Drug Discovery, a section of the journal  
Frontiers in Pharmacology

RECEIVED 29 December 2022

ACCEPTED 01 March 2023

PUBLISHED 13 March 2023

## CITATION

Aydemir D and Ulusu NN (2023), The  
impact of the endocrine-disrupting  
chemicals on the glucose-6-phosphate  
dehydrogenase enzyme activity.  
*Front. Pharmacol.* 14:1133741.  
doi: 10.3389/fphar.2023.1133741

## COPYRIGHT

© 2023 Aydemir and Ulusu. This is an  
open-access article distributed under the  
terms of the [Creative Commons  
Attribution License \(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use,  
distribution or reproduction in other  
forums is permitted, provided the original  
author(s) and the copyright owner(s) are  
credited and that the original publication  
in this journal is cited, in accordance with  
accepted academic practice. No use,  
distribution or reproduction is permitted  
which does not comply with these terms.

# The impact of the endocrine-disrupting chemicals on the glucose-6-phosphate dehydrogenase enzyme activity

Duygu Aydemir<sup>1,2</sup> and Nuriye Nuray Ulusu<sup>1,2\*</sup>

<sup>1</sup>Department of Medical Biochemistry, School of Medicine, Koc University, Sariyer, Istanbul, Turkey, <sup>2</sup>Koç University Research Center for Translational Medicine (KUTTAM), Sariyer, Istanbul, Turkey

## KEYWORDS

G6PD, endocrine disruptors, enzyme, environmental pollutant, oxidative stress

## Introduction

Glucose-6-phosphate dehydrogenase (G6PD, glucose 6-phosphate: NADP (+) oxidoreductase, EC1.1.1.49) enzyme is the first and rate-limiting step in the pentose phosphate pathway (PPP). G6PD is an allosteric enzyme found in monomer, homodimer, and homotetramer forms and catalyzes the production of 6-phosphogluconolactone from glucose-6-phosphate (G6P) (Meng et al., 2022). G6PD maintains the reduction of NADP<sup>+</sup> into coenzyme NADPH which is vital for various reductive biosynthetic reactions such as maintaining reduced glutathione (GSH), protection against reactive oxygen species (ROS), and detoxification of xenobiotics (B Tandogan, 2011; Gao et al., 2019). NADPH is used by the glutathione reductase (GR) enzyme to convert oxidized glutathione (GSSG) into reduced glutathione (GSH) which is one of the most powerful antioxidant molecules in the cell. GSH is used by glutathione s-transferase (GST) enzyme to detoxify xenobiotics such as environmental pollutants, drugs, and chemicals used in industrial products (Aydemir et al., 2018, 2019a; 2019b, 2020a).

G6PD activity is tightly regulated *via* NADPH/NADP<sup>+</sup> ratio, extracellular oxidants, and posttranslational modifications such as phosphorylation, acetylation, glycosylation, ubiquitinylation, and glutarylation (Aydemir and Ulusu, 2020a). G6PD is the common player in glycolysis, gluconeogenesis, PPP, and lipid metabolism; also, overexpression of G6PD activity is associated with lipid dysregulation, insulin resistance, increased body weight, and obesity (Park et al., 2005). The increasing incidence of diabetes and obesity is the leading cause of death, and disabilities worldwide are considered a major public health problem (Lin et al., 2020). Excessive exposure to industrial products and environmental pollutants such as processed food and beverages is directly associated with an increased risk of obesity, metabolic disorders, and diabetes (Bhupathiraju and Hu, 2016). A wide variety of synthetic chemicals used in industrial products such as cosmetics, pharmaceuticals, food and beverage packaging, dyes, households, pesticides, and hygiene products interfere with hormone metabolism altering the endocrine system in humans and wildlife; thus, they are called endocrine-disrupting chemicals (EDCs) (Metcalf et al., 2022).

Since EDCs activate peroxisome proliferating activating receptors (PPARs) and exert adverse effects on hormone metabolism, they dysregulate lipid, glucose, and energy metabolisms and are referred to as obesogens (Darbre, 2017). People are exposed to EDCs daily at various concentrations *via* inhalation, digestion, and dermal route. After exposure, these chemicals are metabolized by the liver, kidney, gut, and skin esterases; however, some parts accumulate in the body without metabolizing over time (Aydemir et al., 2020). On the other hand, administration of phthalate and butylparaben altered G6PD

TABLE 1 Impact of the natural and environmental endocrine disrupting chemicals (EDCs) on the G6PD enzyme activity in different species.

Endocrine Disrupting Chemicals (EDCs)	G6PD activity	Species	References
<b>Natural EDCs</b>			
Dehydroepiandrosterone (DHEA)	↓ Breast cancer cells	Human	Song et al. (2022)
17 $\beta$ -estradiol	↑ Breast cancer cells	Human	Shin and Koo (2021)
Genistein	↑ Testis	Mice	Godschalk et al. (2022)
<i>Ferula assafoetida</i>	↓ Uterus	Rat	Keshri et al. (2004)
<i>Melia azedarach</i>	↓ Uterus	Rat	Keshri et al. (2004)
<i>Saraca asoca</i> (Roxb.) de Wilde	↓ Blood	Rat	Swar et al. (2017)
Epicatechin	↓ Pure Enzyme	<i>Leuconostoc mesenteroides</i>	Camara et al. (2016)
Epigallocatechin	↓ Pure Enzyme	<i>Leuconostoc mesenteroides</i>	Camara et al. (2016)
Epicatechin gallate	↓ Pure Enzyme	<i>Leuconostoc mesenteroides</i>	Camara et al. (2016)
Epigallocatechin	↓ Pure Enzyme	<i>Leuconostoc mesenteroides</i>	Camara et al. (2016)
Testosterone	↑ Muscle	Rat	Max and Knudsen (1980)
Estradiol	↑ Muscle	Rat	Max and Knudsen (1980)
<b>Environmental EDCs</b>			
Butylparaben	↓ Liver, ↑ Kidney, ↑ Spleen	Rat	Aydemir et al. (2019)
DEHP	↑ Liver, ↑↓ Kidney	Rat	Aydemir et al. (2018)
Methoxychlor	↑ Uterus	Mice	Ghosh et al. (1999)
Bisphenol A (BPA)	↑ Breast cancer cells	Human	Kim et al. (2003)
4-nonylphenol (NP)	↑ Breast cancer cells	Human	Kim et al. (2003)
4-octylphenol (OP)	↑ Breast cancer cells	Human	Kim et al. (2003)
Zinc	↓ Kidney	Lamb	Tandogan and Ulusu (2006)
Cadmium	↓ Kidney	Lamb	Tandogan and Ulusu (2006)

activity in various tissues and cells, including the liver, kidney, testis, brain, breast, and spleen, according to the literature (Table 1) (Aydemir et al., 2018, 2019b; 2019a, 2020a). Therefore, we suggest that EDCs can adversely affect the G6PD activity associated with increased body weight, obesity, metabolic syndrome, and diabetes.

## Impact of EDCs on the G6PD activity via hormonal regulation

EDCs can be natural or environmental, affecting synthesis, uptake, or hormone-release mechanism in humans and wildlife (Autrup et al., 2020). Natural EDCs can be classified as phytoestrogens and mycoestrogens. Phytoestrogens are the most prominent group mimicking 17  $\beta$ -estradiol (E2) hormones that can bind estrogen receptors. Increased G6PD activity has been reported

in breast cancer cells upon 17  $\beta$ -estradiol administrations (Monet JD, 1987; Shin and Koo, 2021). Genistein is found in soybeans as the primary dietary source of phytoestrogens, and it increases the G6PD in transgenic mice with lower G6PD activity (Atm- $\Delta$ SRI mice) (Zin et al., 2013; Godschalk et al., 2022).

*Ferula assafoetida* (“*Ferula assa-foetida*”) belongs to the Umbelliferae family containing terpenoids and affects the estrogen signaling pathway specifically via estrogen receptor  $\alpha$  and estrogen receptor  $\beta$  same as phytoestrogens (Ikeda et al., 2002). The extracts of *Ferula assafoetida* and Chinaberry (“*Melia azedarach*”) have estrogenic effects, and they cause pregnancy failure when administered to pregnant rats. On the other hand, these extracts exhibit inhibitory effects on the G6PD (Keshri et al., 2004). *Saraca asoca* (Roxb.) de Wilde, Ashok has estrogenic effects, which is very popular in India; however, dose-dependent G6PD activity increased by using this plant’s extracts, according to the literature (Swar et al., 2017). On the other hand, Epicatechin,

epigallocatechin, epicatechin gallate, and epigallocatechin are found in green tea and have anticancer and anti-inflammatory properties. The literature shows they inhibit the G6PD activity (Camara et al., 2016).

G6PD activity is regulated by various hormones (Stanton, 2012), and its activity is considered an indicator of testosterone or estradiol levels. It was proven that injection of the indicated hormones could activate the G6PD activity in a dose-dependent manner (Max and Knudsen, 1980). Xenobiotics, including antibiotics, drugs, and EDCs, can interfere with the secretion, production, metabolism, and transport of estrogen or androgen hormones affecting the G6PD activity. For instance, BPA, dichlorodiphenyltrichloroethane (DDTs), polychlorinated biphenyls (PCBs), and polybrominated diphenyl ethers (PBDEs) have adverse effects on the estrogenic or androgenic pathways associated with reproductive disorders (Amir et al., 2021). Dehydroepiandrosterone (DHEA), androstenedione, is an endogenous steroid hormone precursor. This hormone is produced by the brain, gonads, and adrenal glands and is one of the non-competitive inhibitors of the G6PD in breast cancer cells (Song et al., 2022). Methoxychlor, an insecticide known as an EDC and accepted as a xenoestrogen, can activate the G6PD in mice's uterus (Ghosh et al., 1999).

## Endocrine-disrupting chemicals can cause a metabolic shift in human and wildlife

EDCs, including bisphenol A (BPA), phthalates, perfluoroalkyl substances (PFAS), and aluminum, contribute to obesity *via* increasing fatty acid storage and appetite (Braun, 2017; Tinkov et al., 2019). EDCs-related obesity may be due to the downregulation of mitochondrial pyruvate carriers (MPC) since, in aerobic conditions, MPC transports pyruvate into mitochondria to be metabolized in the Krebs cycle (Ruiz-Iglesias and Mañes, 2021). Downregulation of MPC results in decreased pyruvate levels in the mitochondria, causing a metabolic shift to the triacylglycerol synthesis associated with obesity (Chen et al., 2018; Hodges et al., 2022). Hypoxia-inducible factor 1 $\alpha$  (HIF- $\alpha$ ) is one of the essential regulators of the G6PD. EDCs induce HIF- $\alpha$  expression; for instance, BPA can activate HIF- $\alpha$  and vascular endothelial growth factor (VEGF) by triggering the G-protein estrogen receptor (GPER) (Xu et al., 2017; Yang et al., 2021). Both cellular signaling factors, HIF- $\alpha$  and VEGF, can upregulate the G6PD activity (Leopold et al., 2003).

Bisphenols can regulate the nuclear factor erythroid 2-related factor 2 (Nrf2), which controls antioxidant metabolism upon physiological and pathophysiological outcomes of exposure to oxidative stress. Nrf2 transcriptionally induces G6PD activity to upregulate the PPP pathway involved in the antioxidant response of the cell (Salehabadi et al., 2022; H.-C.; Yang et al., 2021). Another activator of the G6PD Ataxia-telangiectasia mutated kinase (ATM), a DNA damage-inducible protein kinase that can be activated by BPA (Tichý et al., 2010; Ganesan & Keating, 2016; Xu et al., 2017). 2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD) is one of the most toxic EDC with dioxin-like properties inducing TGF- $\beta$ 1-Smad pathway, oxidative stress, and G6PD activity (Jin et al., 2008; H.-

C; Yang et al., 2021). 17 $\beta$ -estradiol (E2), 4-nonylphenol (NP), and BPA induce the G6PD activity in a concentration-dependent manner in the estrogen-sensitive human breast cancer cell (MCF-7 cells) (Kim et al., 2003).

BPA administration reduces the G6PD activity in erythrocytes leading to hemolysis and morphological changes in the erythrocytes. Thus exposure to EDCs can be life-threatening in G6PD enzyme-deficient individuals (Trivedi et al., 2020). On the other hand, the G6PD was used to explain BPA-mediated diseases in colon cancer cells (SW480), mammary glands, and Sertoli cells because the G6PD was accepted as one of the sensitive biomarkers and can be used for the prediction of BPA-mediated diseases (Ryu et al., 2017). Butylparaben-induced and phthalate-induced oxidative stress *via* altered G6PD activity caused tissue damage in rats' liver, kidney, brain, and testis tissue (Aydemir et al., 2020).

Toxic metals such as cadmium, lead, silver nitrate, thallium sulfate, cobalt, nitrate, arsenic oxide, and arsenic, and essential trace metals copper, nickel, and manganese are found to be related to G6PD enzyme inhibition and various organ damage associated with the pathogenesis of multiple disorders (Tandogan & Ulusu, 2006; 2007). For instance, cadmium and zinc inhibit the G6PD enzyme in lamb kidneys (Tandogan and Ulusu, 2006). On the other hand, exposure to various EDCs can change trace elements' homeostasis and affect the activity of antioxidant and detoxification enzymes (Akbay et al., 2004; Aydemir et al., 2018; Aydemir, Öztaşçı, et al., 2019; Aydemir & Ulusu, 2020; Anapali et al., 2022). Metals can interfere with the enzymes' active site, affect the substrate's binding to the active site and cause reversible or irreversible enzyme inhibitions. Since heavy metals can denature enzymes by disturbing the native state of proteins' by destroying secondary and tertiary structures' disulfide bonds, heavy metal ions are accepted as the most effective inhibitors of the G6PD. Workplace exposure to heavy metals such as lead can cause hematotoxicity, hematopoietic malignancies, and mortality in G6PD-deficient individuals (Cocco, 1998; Cocco et al., 2006). G6PD is vital for all living organisms since the detoxification system relies on NADPH production *via* G6PD, 6PGD, and IDH enzymes. On the other hand, individuals with G6PD deficiency are more vulnerable to various diseases, primarily oxidative stress-induced disorders, including endocrine, cardiovascular, and metabolic disorders (Tandogan and Ulusu, 2006; Arese et al., 2012; Aydemir et al., 2018; 2019b; 2019a; 2020; Aydemir and Ulusu, 2020).

## Conclusion

G6PD enzyme is vital for cell proliferation, production of cellular metabolites, cellular energy, oxidative stress status, and antioxidant response in the cell. G6PD is the rate-limiting enzyme of the PPP responsible for NADPH production. NADPH is vital for various reductive biosynthetic reactions, such as maintaining reduced glutathione (GSH), protecting against reactive oxygen species (ROS), and detoxifying xenobiotics, including drugs, environmental pollutants, and environmental pollutants and EDCs. G6PD activity is tightly regulated *via* NADPH/NADP<sup>+</sup> ratio, extracellular oxidants, and

posttranslational modifications such as phosphorylation, acetylation, glycosylation, ubiquitinylation, and glutarylation. Altered G6PD activity is associated with lipid dysregulation, insulin resistance, increased body weight, and obesity since G6PD is the common player in glycolysis, gluconeogenesis, lipid metabolism PPP, and oxidative stress. EDCs are found in most industrial products, including cosmetics, hygiene products, food and beverage packages, toys, and medical devices, and these chemicals interfere with hormone metabolism in humans and wildlife. Since EDCs activate PPARs and adversely affect hormone metabolism, they dysregulate lipid, glucose, and energy metabolisms and are called obesogens. EDCs, including BP, DEHP, BPA, DDT, cadmium, phytoestrogens, PFAS, TCDD, silver nitrate, copper, nickel, manganese, arsenic oxide, and phthalates impair G6PD activity at transcriptional, translational, and posttranslational levels in human and wildlife. Therefore, we suggest that EDCs dysregulate oxidative stress, lipid, and glucose by interfering with G6PD enzyme activity. Individuals with G6PD deficiency can be vulnerable to EDCs-induced adverse health effects, including endocrine, metabolic, and cardiovascular disorders.

## References

- Akbay, E., Nuray Ulusu, N., Töröner, F., Ayvaz, G., Taneri, F., Aktürk, M., et al. (2004). Effects of rosiglitazone treatment on the pentose phosphate pathway and glutathione-dependent enzymes in liver and kidney of rats fed a high-fat diet. *Curr. Ther. Res.* 65, 79–89. doi:10.1016/S0011-393X(04)90007-0
- Amir, S., Shah, S. T. A., Mamoulakis, C., Docea, A. O., Kalantzi, O.-I., Zachariou, A., et al. (2021). Endocrine disruptors acting on estrogen and androgen pathways cause reproductive disorders through multiple mechanisms: A review. *Int. J. Environ. Res. Public Health* 18, 1464. doi:10.3390/ijerph18041464
- Anapali, M., Kaya-Dagistanli, F., Akdemir, A. S., Aydemir, D., Ulusu, N. N., Ulutin, T., et al. (2022). Combined resveratrol and vitamin D treatment ameliorate inflammation-related liver fibrosis, ER stress, and apoptosis in a high-fructose diet/streptozotocin-induced T2DM model. *Histochem Cell Biol.* 158, 279–296. doi:10.1007/s00418-022-02131-y
- Arese, P., Gallo, V., Pantaleo, A., and Turrini, F. (2012). Life and death of glucose-6-phosphate dehydrogenase (G6PD) deficient erythrocytes – role of redox stress and band 3 modifications. *Transfus. Med. Hemotherapy* 39, 328–334. doi:10.1159/000343123
- Autrup, H., Barile, F. A., Berry, S. C., Blaauw, B. J., Boobis, A., Bolt, H., et al. (2020). Human exposure to synthetic endocrine disrupting chemicals (S-EDCs) is generally negligible as compared to natural compounds with higher or comparable endocrine activity: How to evaluate the risk of the S-EDCs? *Arch. Toxicol.* 94, 2549–2557. doi:10.1007/s00204-020-02800-8
- Aydemir, D., Karabulut, G., Gok, M., Barlas, N., and Ulusu, N. N. (2019a). Data the DEHP induced changes on the trace element and mineral levels in the brain and testis tissues of rats. *Data Brief.* 26, 104526. doi:10.1016/j.dib.2019.104526
- Aydemir, D., Karabulut, G., Şimşek, G., Gok, M., Barlas, N., and Ulusu, N. N. (2018). Impact of the di(2-ethylhexyl) phthalate administration on trace element and mineral levels in relation of kidney and liver damage in rats. *Biol. Trace Elem. Res.* 186, 474–488. doi:10.1007/s12011-018-1331-0
- Aydemir, D., Öztaşçı, B., Barlas, N., and Ulusu, N. N. (2019b). Effects of butylparaben on antioxidant enzyme activities and histopathological changes in rat tissues. *Arch. Hig. Rada Toksikol.* 70, 315–324. doi:10.2478/aiht-2019-70-3342
- Aydemir, D., Oztasci, B., Barlas, N., and Ulusu, N. N. (2020). Influence of the butylparaben administration on the oxidative stress metabolism of liver, kidney and spleen. *Turk. J. Biochem.* 45, 689–694. doi:10.1515/tjb-2020-0048
- Aydemir, D., and Ulusu, N. N. (2020). The possible role of the glucose-6-phosphate dehydrogenase enzyme deficiency in the polyneuropathies. *J. Basic Clin. Health Sci.* 4 (3), 212–217. doi:10.30621/jbachs.2020.1151
- B Tandogan, C. S. N. U., Sengencer, C., and Ulusu, N. N. (2011). *In vitro* effects of imatinib on glucose-6-phosphate dehydrogenase and glutathione reductase. *Folia Biol. (Praha)* 57, 57–64.
- Bhupathiraju, S. N., and Hu, F. B. (2016). Epidemiology of obesity and diabetes and their cardiovascular complications. *Circ. Res.* 118, 1723–1735. doi:10.1161/CIRCRESAHA.115.306825
- Braun, J. M. (2017). Early-life exposure to EDCs: Role in childhood obesity and neurodevelopment. *Nat. Rev. Endocrinol.* 13, 161–173. doi:10.1038/nrendo.2016.186
- Camara, M. A., Tian, M., Liu, X., Liu, X., Wang, Y., Yang, J., et al. (2016). Determination of the inhibitory effect of green tea extract on glucose-6-phosphate dehydrogenase based on multilayer capillary enzyme microreactor. *Biomed. Chromatogr.* 30, 1210–1215. doi:10.1002/bmc.3669
- Chen, Y., McCommis, K. S., Ferguson, D., Hall, A. M., Harris, C. A., and Finck, B. N. (2018). Inhibition of the mitochondrial pyruvate carrier by tolylfluaniid. *Endocrinol* 159, 609–621. doi:10.1210/en.2017-00695
- Cocco, P., Fadda, D., Atzeri, S., Avataneo, G., Meloni, M., and Flore, C. (2006). Causes of death among lead smelters in relation to the glucose-6-phosphate dehydrogenase polymorphism. *Occup. Environ. Med.* 64, 414–416. doi:10.1136/oem.2006.028779
- Cocco, P. (1998). Occupational lead exposure and screening of glucose-6-phosphate dehydrogenase polymorphism: Useful prevention or nonvoluntary discrimination? *Int. Arch. Occup. Environ. Health* 71, 148–150. doi:10.1007/s004200050263
- Darbre, P. D. (2017). Endocrine disruptors and obesity. *Curr. Obes. Rep.* 6, 18–27. doi:10.1007/s13679-017-0240-4
- Ganesan, S., and Keating, A. F. (2016). Bisphenol A-induced ovotoxicity involves DNA damage induction to which the ovary mounts a protective response indicated by increased expression of proteins involved in DNA repair and xenobiotic biotransformation. *Toxicol. Sci.* 152, 169–180. doi:10.1093/toxsci/kfw076
- Gao, X., Zhao, L., Liu, S., Li, Y., Xia, S., Chen, D., et al. (2019).  $\gamma$ -6-Phosphogluconolactone, a byproduct of the oxidative pentose phosphate pathway, contributes to AMPK activation through inhibition of PP2A. *Mol. Cell* 76, 857–871. doi:10.1016/j.molcel.2019.09.007
- Ghosh, D., Taylor, J. A., Green, J. A., and Lubahn, D. B. (1999). Methoxychlor stimulates estrogen-responsive messenger ribonucleic acids in mouse uterus through a non-estrogen receptor (Non-ER) $\alpha$  and non-er $\beta$  mechanism. *Endocrinol* 140, 3526–3533. doi:10.1210/endo.140.8.6877
- Godschalk, R. W. L., Janssen, M. C. M., Vanhees, K., Doorn-Khosrovanivanvan, S. B. W., and Schooten, F.-J. van (2022). Maternal exposure to genistein during pregnancy and oxidative DNA damage in testes of male mouse offspring. *Front. Nutr.* 9, 904368. doi:10.3389/fnut.2022.904368
- Hodges, W. T., Jarasvaraparn, C., Ferguson, D., Griffett, K., Gill, L. E., Chen, Y., et al. (2022). Mitochondrial pyruvate carrier inhibitors improve metabolic parameters in diet-induced obese mice. *J. Biol. Chem.* 298, 101554. doi:10.1016/j.jbc.2021.101554
- Ikeda, K., Arai, Y., Otsuka, H., Nomoto, S., Horiguchi, H., Kato, S., et al. (2002). Terpenoids found in the umbelliferae family act as agonists/antagonists for ER(alpha) and ERbeta: Differential transcription activity between ferutinine-liganded ER(alpha) and ERbeta. *Biochem. Biophys. Res. Commun.* 291, 354–360. doi:10.1006/bbrc.2002.6446
- Jin, M. H., Hong, C. H., Lee, H. Y., Kang, H. J., and Han, S. W. (2008). Enhanced TGF-beta1 is involved in 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) induced oxidative stress in C57BL/6 mouse testis. *Toxicol. Lett.* 178, 202–209. doi:10.1016/j.toxlet.2008.03.015
- Keshri, G., Bajpai, M., Lakshmi, V., Setty, B. S., and Gupta, G. (2004). Role of energy metabolism in the pregnancy interceptive action of *Ferula assafoetida* and *Melia*

## Author contributions

NU was responsible for the conceptualization, supervision and writing the original manuscript. DA was responsible for the supporting conceptualization and writing the original manuscript.

## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

## Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

- azedarach extracts in rat. *Contraception* 70, 429–432. doi:10.1016/j.contraception.2004.06.003
- Kim, K. B., Seo, K. W., Kim, Y. J., Park, M., Park, C. W., Kim, P. Y., et al. (2003). Estrogenic effects of phenolic compounds on glucose-6-phosphate dehydrogenase in MCF-7 cells and uterine glutathione peroxidase in rats. *Chemosphere* 50, 1167–1173. doi:10.1016/S0045-6535(02)00628-8
- Leopold, J. A., Walker, J., Scribner, A. W., Voetsch, B., Zhang, Y.-Y., Loscalzo, A. J., et al. (2003). Glucose-6-phosphate dehydrogenase modulates vascular endothelial growth factor-mediated angiogenesis. *J. Biol. Chem.* 278, 32100–32106. doi:10.1074/jbc.M301293200
- Lin, X., Xu, Y., Pan, X., Xu, J., Ding, Y., Sun, X., et al. (2020). Global, regional, and national burden and trend of diabetes in 195 countries and territories: An analysis from 1990 to 2025. *Sci. Rep.* 10, 14790. doi:10.1038/s41598-020-71908-9
- Max, S. R., and Knudsen, J. F. (1980). Effect of sex hormones on glucose-6-phosphate dehydrogenase in rat levator ani muscle. *Mol. Cell Endocrinol.* 17, 111–118. doi:10.1016/0303-7207(80)90123-9
- Meng, Q., Zhang, Y., Hao, S., Sun, H., Liu, B., Zhou, H., et al. (2022). Recent findings in the regulation of G6PD and its role in diseases. *Front. Pharmacol.* 13, 932154. doi:10.3389/fphar.2022.932154
- Metcalfe, C. D., Bayen, S., Desrosiers, M., Muñoz, G., Sauvé, S., and Yargeau, V. (2022). An introduction to the sources, fate, occurrence and effects of endocrine disrupting chemicals released into the environment. *Environ. Res.* 207, 112658. doi:10.1016/j.envres.2021.112658
- Monet Jd, T. M. D. N. B. M. B. C., ThoMas, M., DautigNy, N., BraMi, M., and Bader, C. A. (1987). Effects of 17 beta-estradiol and R5020 on glucose-6-phosphate dehydrogenase activity in MCF-7 human breast cancer cells: A cytochemical assay. *Cancer Res.* 47, 5116–5119.
- Park, J., Rho, H. K., Kim, K. H., Choe, S. S., Lee, Y. S., and Kim, J. B. (2005). Overexpression of glucose-6-phosphate dehydrogenase is associated with lipid dysregulation and insulin resistance in obesity. *Mol. Cell Biol.* 25, 5146–5157. doi:10.1128/MCB.25.12.5146-5157.2005
- Ruiz-Iglesias, A., and Mañes, S. (2021). The importance of mitochondrial pyruvate carrier in cancer cell metabolism and tumorigenesis. *Cancers (Basel)* 13, 1488. doi:10.3390/cancers13071488
- Ryu, D.-Y., Rahman, M., and Pang, M.-G. (2017). Determination of highly sensitive biological cell model systems to screen BPA-related health hazards using pathway studio. *Int. J. Mol. Sci.* 18, 1909. doi:10.3390/ijms18091909
- Salehabadi, A., Farkhondeh, T., Harifi-Mood, M. S., Aschner, M., and Samarghandian, S. (2022). Role of Nrf2 in bisphenol effects: A review study. *Environ. Sci. Pollut. Res.* 29, 55457–55472. doi:10.1007/s11356-022-20996-3
- Shin, E., and Koo, J. S. (2021). Glucose metabolism and glucose transporters in breast cancer. *Front. Cell Dev. Biol.* 9, 728759. doi:10.3389/fcell.2021.728759
- Song, J., Sun, H., Zhang, S., and Shan, C. (2022). The multiple roles of glucose-6-phosphate dehydrogenase in tumorigenesis and cancer chemoresistance. *Life* 12, 271. doi:10.3390/life12020271
- Stanton, R. C. (2012). Glucose-6-phosphate dehydrogenase, NADPH, and cell survival. *IUBMB Life* 64, 362–369. doi:10.1002/iub.1017
- Swar, G., Shailajan, S., and Menon, S. (2017). Activity based evaluation of a traditional Ayurvedic medicinal plant: Saraca asoca (Roxb.) de Wilde flowers as estrogenic agents using ovariectomized rat model. *J. Ethnopharmacol.* 195, 324–333. doi:10.1016/j.jep.2016.11.038
- Tandogan, B., and Ulusu, N. N. (2006). Effects of cadmium and zinc ions on purified lamb kidney cortex glucose-6-phosphate dehydrogenase activity. *J. Enzyme Inhib. Med. Chem.* 21, 225–230. doi:10.1080/14756360500480533
- Tandogan, B., and Ulusu, N. N. (2007). The inhibition kinetics of yeast glutathione reductase by some metal ions. *J. Enzyme Inhib. Med. Chem.* 22, 489–495. doi:10.1080/14756360601162147
- Tichý, A., Vávrová, J., Pejchal, J., and Řezáčová, M. (2010). Ataxia-telangiectasia mutated kinase (ATM) as a central regulator of radiation-induced DNA damage response. *Acta Medica (Hradec Kralove, Czech Repub.)* 53, 13–17. doi:10.14712/18059694.2016.57
- Tinkov, A. A., Skalnaya, M. G., Aaseth, J., Ajsuvakova, O. P., Aschner, M., and Skalny, A. v. (2019). Aluminium levels in hair and urine are associated with overweight and obesity in a non-occupationally exposed population. *J. Trace Elem. Med. Biol.* 56, 139–145. doi:10.1016/j.jtemb.2019.08.005
- Trivedi, M., Vaidya, D., Patel, C., Prajapati, S., and Bhatt, J. (2020). *In silico* and *in vitro* studies to elucidate the role of 1HYN and 1QKI activity in BPA induced toxicity and its amelioration by Gallic acid. *Chemosphere* 241, 125076. doi:10.1016/j.chemosphere.2019.125076
- Xu, F., Wang, X., Wu, N., He, S., Yi, W., Xiang, S., et al. (2017). Bisphenol A induces proliferative effects on both breast cancer cells and vascular endothelial cells through a shared GPER-dependent pathway in hypoxia. *Environ. Pollut.* 231, 1609–1620. doi:10.1016/j.envpol.2017.09.069
- Yang, H.-C., Stern, A., and Chiu, D. T.-Y. (2021). G6PD: A hub for metabolic reprogramming and redox signaling in cancer. *Biomed. J.* 44, 285–292. doi:10.1016/j.bj.2020.08.001
- Zin, S. R. M., Omar, S. Z., Khan, N. L. A., Musameh, N. I., Das, S., and Kassim, N. M. (2013). Effects of the phytoestrogen genistein on the development of the reproductive system of Sprague Dawley rats. *Clinics* 68, 253–262. doi:10.6061/clinics/2013(02)OA21