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# Recent advances in nanoparticle applications in respiratory disorders: a review

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Various nanoparticles are used in the discovery of new nanomedicine to overcome the shortages of conventional drugs. Therefore, this article presents a comprehensive and up-to-date review of the effects of nanoparticle-based drugs in the treatment of respiratory disorders, including both basic and clinical studies. Databases, including PubMed, Web of Knowledge, and Scopus, were searched until the end of August 2022 regarding the effect of nanoparticles on respiratory diseases. As a new tool, nanomedicine offered promising applications for the treatment of pulmonary diseases. The basic composition and intrinsic characteristics of nanomaterials showed their effectiveness in treating pulmonary diseases. The efficiency of different nanomedicines has been demonstrated in experimental animal models of asthma, chronic obstructive pulmonary disease (COPD), pulmonary fibrosis (PF), lung cancer, lung infection, and other lung disorders, confirming their function in the improvement of respiratory disorders. Various types of nanomaterials, such as carbon nanotubes, dendrimers, polymeric nanomaterials, liposomes, quantum dots, and metal and metal oxide nanoparticles, have demonstrated therapeutic effects on respiratory disorders, which may lead to new possible remedies for various respiratory illnesses that could increase drug efficacy and decrease side effects.

## KEYWORDS

nanomedicine, nanoparticle, respiratory disorders, drug delivery, treatment

## 1 Introduction

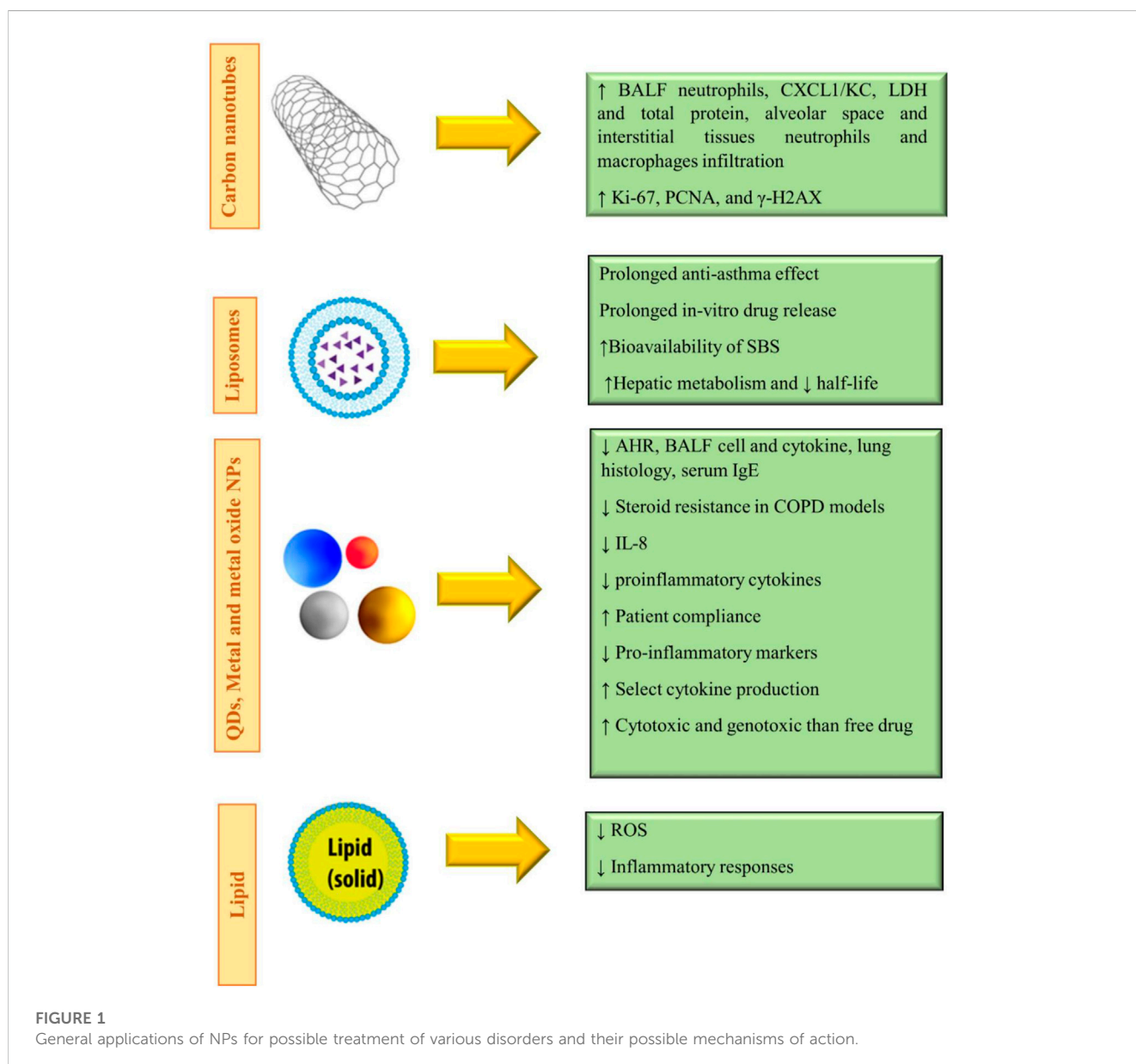
According to the World Health Organization (WHO), respiratory diseases caused the deaths of at least 9 million people worldwide and resulted in numerous disabilities in 2016. These diseases accounted for approximately 15% of all deaths (Horváth et al., 2017). Lower respiratory infections, pneumonia, bronchitis, tuberculosis, pharyngitis, laryngitis, and the common cold, as well as obstructive lung disorders, such as asthma, chronic obstructive pulmonary disease (COPD), and lung cancer, are among the most prevalent respiratory diseases. The main causes of respiratory diseases are air pollution, genetic attributes, and infections caused by viruses and bacteria. The lower respiratory airways could easily be infected by airborne diseases and infections, influencing the lower airways and leading to acute respiratory infections (Thorlund et al., 2020). The recently known acute infection caused by the new coronavirus (COVID-19) and the older ones by severe acute respiratory syndrome coronavirus (SARS-CoV), Middle East respiratory syndrome coronavirus (MERS-CoV), and avian influenza A (H7N9) can cause severe respiratory infection. Chronic respiratory diseases are caused by genetic disorders and air pollution, including cystic

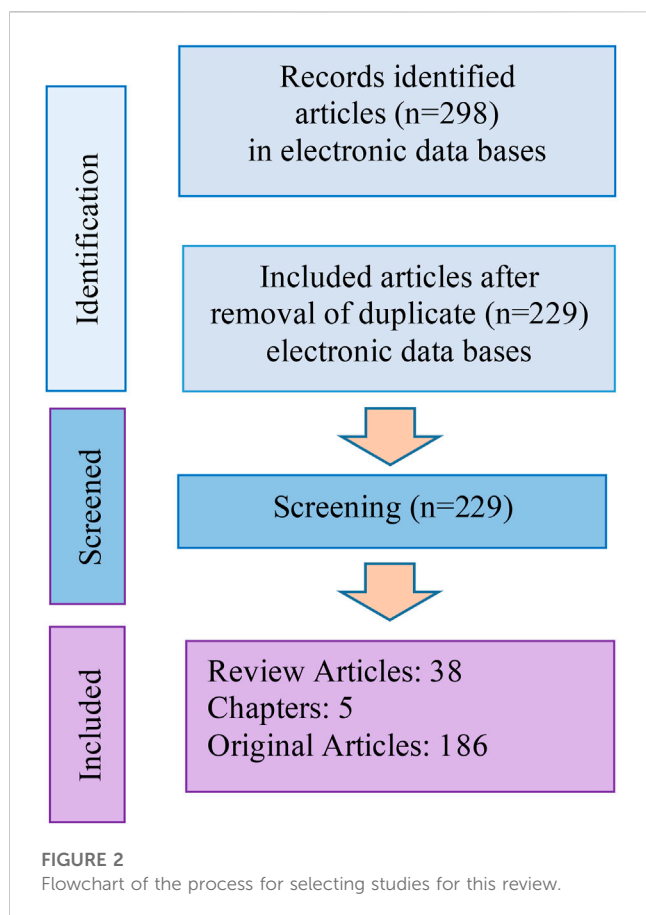
fibrosis, asthma, COPD, hay fever, and lung cancer. The treatment priority goal is to improve patients' quality of life by repairing or restoring the functions of the respiratory system (Thorlund et al., 2020). The main goals in the treatment of respiratory disorders are to provide a suitable treatment for ameliorating disease symptoms, reducing the casualties and mortality rate, increasing the quality of life, and decreasing the side effects of drugs.

The need for innovative treatment methods that can overcome obstacles, including drug resistance, low drug efficiency, side effects, and costs, may justify the use of nano-based drugs. In recent years, new doors have been opened to treat respiratory disorders with the advent of nanomedicine and nanotechnology. Nanoparticles (NPs) have a very high surface-to-volume ratio because of their nano-scale dimensions (Yazdi et al., 2021). This feature allows multiple ligands to be attached to the surface and increases the tendency to develop multiple covalent bonds and simultaneously use in targeted therapy by an appropriate design (Ghorani-Azam et al., 2022). Other

characteristics of nanoparticles are increased chemico-biological stability, the possibility of binding to both hydrophilic and hydrophobic drugs, and the capability to be prescribed via inhalation or injection (Deng et al., 2021; Luo et al., 2021), which depends entirely on the physicochemical characteristics of the nano-carrier and the chemical structure of the reagents. Aerosol drug delivery is one of the popular drug delivery systems that can be used for respiratory and non-respiratory disorders (Bianco et al., 2021; Vartak et al., 2021). The size of nanoparticles would determine the deposition spot in the respiratory tract. For example, 1 nm nanoparticles accumulated over lower airways, but they usually come out from the respiratory system with expired air; 5 nm particles were placed in the tracheal and bronchi; and 20 nm particles were localized in the upper airway regions (Shakerimanesh et al., 2022).

The nano-systems could have higher efficacy than conventional drugs due to functionalized for targeted delivery, increased





biocompatibility, and reduced adverse effects (Mousavi-Kouhi et al., 2021a). Different types of nanoparticles, including organic and inorganic nanoparticles, were available for biological application, which should be explained more thoroughly for designing new nano-systems for specific purposes. In this review article, the NPs that were used in the treatment of respiratory disorders were presented to form new ideas for more convenient future medications. Figure 1 shows the general applications of NPs for the possible treatment of various disorders and their possible mechanisms of action.

## 2 Materials and methods

*In vitro* and *in vivo* studies regarding the influence of NPs on respiratory diseases were searched using online databases, including PubMed, Web of Knowledge, and Scopus, from 2015 to the end of August 2022. For this purpose, keywords including (Nanomedicine), (respiratory disorders), and (respiratory, pulmonary or lung diseases), (name of respiratory diseases individually), (nanoparticles), (inorganic nanocarrier or nanoparticle), (organic nanocarrier or nanoparticle), and (name of nanoparticles individually) were used. Furthermore, the effects of the nanoparticles on lung cancers and lung infections were covered in the current review article. Only articles published in the English language and well-known international journals were included in this review manuscript.

Totally, 298 articles were selected from internet sources, of which 80 articles were duplicates. The study flowchart is shown in Figure 2.

## 3 Nanoparticles for biomedical applications

Nanotechnology has the potential to revolutionize the field of chemotherapy (Vahideh Mohammadzadeh et al., 2022). Nanoscale platforms aim to increase treatment efficacy and localize drug delivery, resulting in reduced dosage, fewer systemic side effects, controlling biodistribution, modulating pharmacokinetics (release kinetics and mechanism), and finally improving patient compliance (Akhter et al., 2018). Nano-architectures could be used to encapsulate different drugs, especially hydrophobic drugs. Nanoparticles can be thought of as tiny packages that contain drugs and can be delivered to specific locations in the body (Mousavi-Kouhi et al., 2021b).

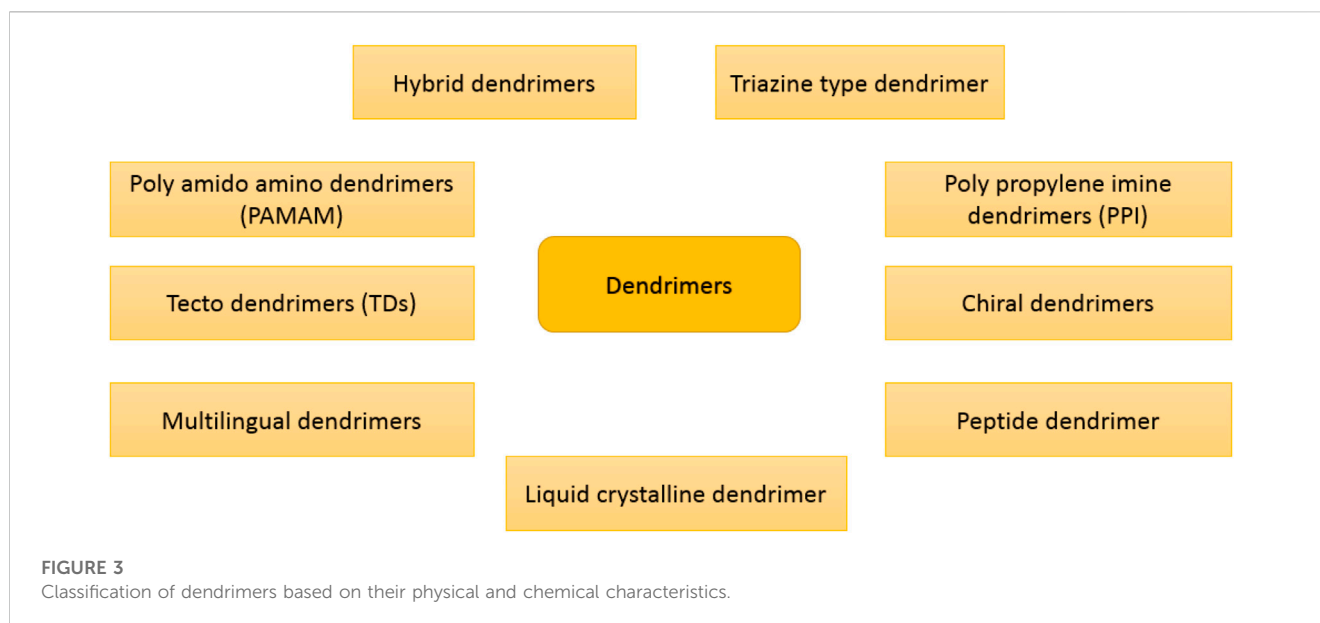
The major organic-based platforms for nanotechnology include carbon nanodiamonds, nanotubes, graphene, graphene oxide, fullerenes, dendrimers, polymeric particles, solid lipid hybrid nanoparticles, liposomes (most popular), pyrosomes, nanoemulsions, and graphene/carbon quantum dots (QDs) (Ashna et al., 2020; Naciute et al., 2020). Inorganic-based drug delivery systems comprise metal and metal oxide nanoparticles (especially gold for diagnosis and magnetic nanoparticles), metallic quantum dots, and porous materials (Javanbakht et al., 2020; Parra-Nieto et al., 2020) (Figure 2).

### 3.1 Nanoparticles used for the treatment of respiratory disorders

Several types of nanoparticle drugs have been used for the treatment of different respiratory disorders, and the most important types are briefly described in the following sections.

#### 3.1.1 Carbon-based nanomaterials

Carbon-based materials play important roles in the advancement of material science, including a) traditional industrial carbon, such as activated carbon, b) new industrial carbon, such as carbon fibers, and c) new carbon nanomaterials, such as graphene and carbon nanotubes. Primary investigation and uses of carbon-based materials are general in the interdisciplinary fields, although macroscopic carbon material lacks a suitable band gap, making it problematic to act as an efficient fluorescent material (Liu et al., 2020). Carbon nanodiamonds and carbon nanotubes are light, strong, highly conductive, and versatile. The drug delivery systems of this type of nanoparticles could be functionalized to prepare robust diagnostic tools (Chauhan et al., 2020). Carbon nanotubes are inhalable but would cause health risks (Oberdörster et al., 2015), such as lung fibrosis and inflammation (Poulsen et al., 2016); hence, they are not suitable for the treatment of respiratory diseases. In recent years, by developing the substantial properties of diverse newly invented carbon-based nanomaterials, these have been modified and widely employed in biology, medicine, and pharmacology (Maiti et al., 2019).



### 3.1.2 Dendrimers

The term “dendrimer” was derived from the Greek word dendron, which means “tree” because dendrimers have a branching structure that resembles a tree (Steinman, 2007). A dendrimer consists of dendrons, which are arms initiated from the central core and can be classified on the basis of physicochemical characteristics (Ambekar et al., 2020). Dendrimers are radially extended hyperbranched architectures with high versatility that can be modified for different medical purposes such as drug delivery, gene delivery, cancer targeting, and diagnosis. By changing the functional groups, the hydrophobic zones could be changed/engineered to hold certain hydrophobic drugs (Wang et al., 2016). In this regard, poly(amidoamine) (PAMAM) is a dendrimer with amide and amine functionalities. Dendrimers can be categorized based on their physicochemical construction, as shown in Figure 3. Monodispersity is one of the exclusive characteristics of dendrimers that can be responsible for the replication of dendrimers at a scalable rank. Dendrimers are extensively used as drug delivery (Zenze et al., 2023), biosensors (Zhou et al., 2023), pharmacology (Pérez-Carrión and Posadas, 2023), antibiotics (Xu et al., 2023), and gene editing (Liu et al., 2023). Today, many types of dendrimers, such as polylysine, polyethylene glycol, and PAMAM, are widely used as nanocarriers for the treatment of diseases (Yousefi et al., 2020). Gold and iron nanoparticles, which are well-known for the treatment of cancer, such as paclitaxel, cisplatin, and doxorubicin are loaded in dendrimer (Khemtong et al., 2009; Zarharan et al., 2023). The solubility of the hydrophobic drugs could be enhanced using embedding in dendrimers (Zhang et al., 2011). Dendrimer-based nanocarriers need to be functionalized with biomolecules to overcome the cationic toxicity of dendrimers, which can be easily carried out due to the large amino groups in dendrimers (Ambekar et al., 2020).

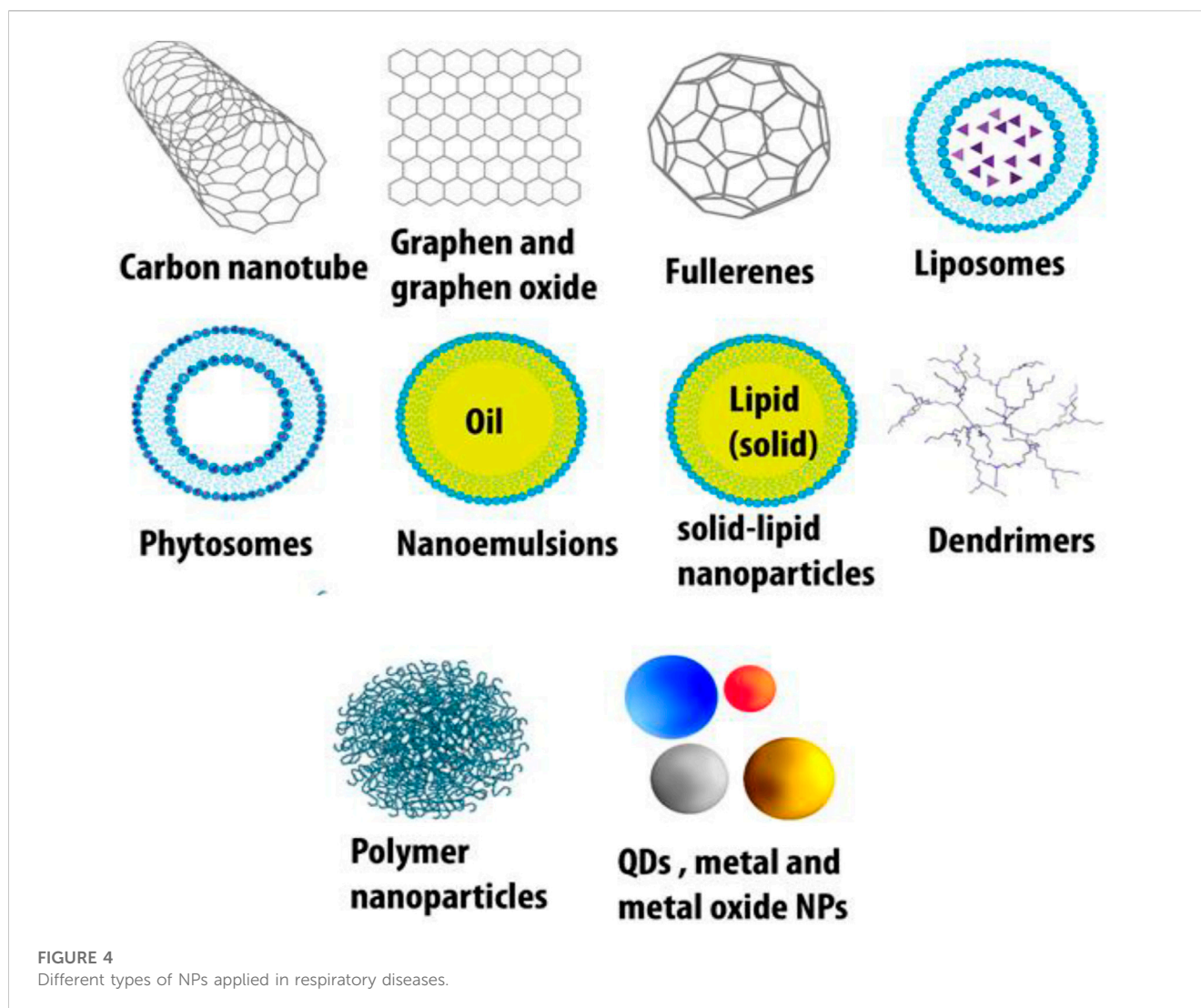
### 3.1.3 Polymeric nanoparticles

Polymeric nanoparticles can be defined as solid colloidal particles, and they are preferably made up of polymers attained from a natural or synthetic basis (Yazdi et al., 2020). Polymeric

nanoparticles can be separated into classes of hydrogels, nanospheres, and nanocapsules, examples of the last being polymeric micelles and polymersomes. Hydrogels are polymeric materials with a significant number of attached hydrophilic groups and have a three-dimensional network that helps them absorb a large amount of water or dissolved drugs chemically or physically. Nanocapsule polymeric NPs are the ones in which the remedial element is encapsulated inside a polymeric capsule shell, whereas nanosphere polymeric NPs are the ones in which drugs or other solid particles are implanted into a polymeric matrix. Polymeric nanoparticles deliver the drug to a specific site and deliver the drug at a specific amount, which is valuable for the treatment of various diseases (Sur et al., 2019).

### 3.1.4 Liposomes

Liposomes (LPs) are vesicles formed from concentric lipid bilayers, so-called lamellae, which are spontaneously formed when certain lipids are hydrated in aqueous media. LPs are very similar to self-assembling colloidal polymeric nanoparticles but constructed from lipid bilayers. Lipids have both hydrophilic and hydrophobic sides that can wrap into spherical or oval-type shapes (Saraf et al., 2020). Nanoparticles similar to liposomes are phytosomes, micelles, nanoemulsions, and solid lipid nanoparticles (Gorain et al., 2020). More specifically, liposomes have an aqueous core that is perfect for encapsulating hydrophilic drugs (New, 1990). In hydrophobic drugs, it could also be placed between the lipid bilayer of the cell membrane in a phagosomal form (Kumar et al., 2020). LPs used in biology and medicine display a range of benefits comprising high drug/lipid proportion loading efficacy, simplicity of synthesis in a size-controlled way, great stability, manageable delivery kinetics, and biocompatibility (Zylberberg and Matosevic, 2016). Moreover, LPs can be extra equipped with functional moieties to increase their performances in terms of increased circulation half-life, targeted delivery, and intracellular permeation capability (Kim, 2016). Consequently, LPs show better pharmacokinetics and biodistribution outlines than



numerous other drug transporters in clinical research, thus improving the pharmacokinetics of encapsulated remedial factors while keeping them and preventing their associated side effects and drug reactions (El-Hammadi and Arias, 2019).

### 3.1.5 Lipid nanocarriers

Lipids make up selectively permeable cell walls; hence, liposomes could be the most permeable and biocompatible nanocarriers in the body. Most of the FDA-approved nanoparticles are based on liposomes, but they encounter major limitations. Liposomes are prone to oxidative degradation and drug leakage when continuously exposed to aqueous solution and body fluids (Lee and Thompson, 2017). Nanoemulsions and solid lipid nanoparticles could be better candidates than liposomes, which cover a broader range of pharmaceuticals and more affordable drug delivery systems (Roy et al., 2022). Lipid-based nanocarriers could dominate the difficulties of low availability for oral treatment with poor water solubility. Lipid NPs have the features of nanosized particles and lipid solubility, leading to increased pharmacokinetic and biocompatibility and decreased harmfulness and enabling scale-up for industrialized fabrication (Dumont et al., 2018). Their

digestion involves the dispersion of fat globules into an emulsion with great surface area, enzyme hydrolysis of the triglyceride lipid at the lipid–water interface, and the dispersion of the bio-product into the absorbed formula (Joyce et al., 2016). Also, the encapsulation of drugs or bio-agents using lipid-based nanocarriers can decrease the food intake effects on absorption and variation among topics because of their controlled release (Hsu et al., 2019).

### 3.1.6 Quantum dots

Quantum dots (QDs) have been known as an innovation in nanoscience for semiconductor inorganic crystals that comprise variable numbers of electrons, which occupy well-defined and distinct quantum statuses (Reshma and Mohanan, 2019). Moreover, QDs are small-sized particles that prepare a wide range of flexible element ratios, which could cause fluorescent properties (Hu et al., 2019). The construction of QDs could be simply controlled while obeying the fundamental of quantum confinement (Bai et al., 2018). Specifically, QDs are small semiconducting particles with typical sizes ranging from 1 to 10 nm. They can make a valuable contribution in an extensive variety of applications and may substitute for many expensive

materials now used for fluorescent tagging and cell imaging because of their facile synthesis, high photo-stability, high resistance to metabolic degradation, and brightness. QDs need little energy to be excited (blue light), and the emitted light from QDs has a narrow emission spectrum at longer wavelengths than the absorbed light. Moreover, the emitted wavelength can be controlled by controlling the size of the QD. QDs could be used for the synthesis of multifunctional nanoparticles that could detect cancer cells and visualize, track, kill, and monitor them. Typical QDs made from carbon, graphene, cadmium or zinc chalcogenides, phosphides, and indium arsenide are placed in both organic and inorganic categories (Ruiyi et al., 2020). Figure 4 shows the role and potential of some QDs in the respiratory system.

### 3.1.7 Metal and metal oxide nanoparticles

Metal and metal oxide nanoparticles are inorganic-based nanocarriers that can be used as a means to transfer small drug molecules and large ones, such as DNA or proteins, throughout the body (Mobaraki et al., 2021). The surface of the nanoparticle could be coated with a protective layer for stability issues in the body fluids and be tagged by different ligands on the outside to attach preferentially to receptors of the cancer cells (Zarei et al., 2021). For example, gold nanoparticles could be PEGylated or functionalized with fluorescent labels, protein, or other targeting agents, which can penetrate the cell membranes while carrying the DNA or the drug (Song et al., 2018). Metal and metal oxide nanoparticles display altered physical and chemical activities and are different from their native bulk materials in some things, which consist of their surface, optical, thermal, and electrical properties. Metal and metal oxide NPs are synthesized using the addition of reducing or oxidizing/precipitating factors during their synthesis, respectively (Hoda Zarharan et al., 2023). Several factors are involved in NP reactivity with biomolecules, which comprise nanoparticles' size, shape, surface activities, stability, and the process of fabrication (Rastogi et al., 2017). Figure 4 demonstrates different types of NPs applied to respiratory diseases.

## 3.2 Methods for NP establishment for lung delivery

A number of conventional methods have been described for the expansion of particulate material for specific delivery to the lung. Nevertheless, there are limitations such as shape, size, and distribution (Patil and Sarasija, 2012). The physicochemical features of any particle are the key agents defining its flow activity, which impacts the particle transport and deposition on the pulmonary structure (Tsuda et al., 2013). However, the influences of inhaled NPs are obviously far for the treatment of non-lung disorders, especially the cardiovascular structure and other highly vascularized organs, such as brain diseases (Mostovenko et al., 2022).

Inhaled powders based on various NP types, such as polymeric nanoparticles, liposomes, solid lipid nanoparticles, and mesoporous silica nanoparticles, were studied for delivery to the respiratory system but mostly for polymeric and lipid nanoparticle types of NPs (Chan et al., 2023). In addition, the clinical study for the liposome inhaler form is also a development for the delivery system (Forest and Pourchez, 2022).

### 3.2.1 Spray drying

In this method, for thermolabile matters such as protein and insulin to be dried, they are atomized into tiny droplets and driven radially into a moving stream of hot drying medium (Pramanik et al., 2021). This method was planned to make submicron particles with greater products in the primary engineering sector, even for massive-scale formats. However, it did not get pleasure with NP preparation due to, for example, little separation capacity. Investigations using inhaled NPs dispersed in watery droplets propose that mucus clearance could be overcome by NPs, probably because of the fast movement of particles to the airway epithelium through surface energetics (Sham et al., 2004). Hence, NPs might be potential vehicles for effectual transporting drugs to the epithelium, although causing undesirable mucociliary clearance.

### 3.2.2 Spray-freeze-drying manner

Spray-freeze-drying (SFD) is a drying manner that first found uses in the arena of pharmacological technologies and has a variety of applications in pharmacological and nutrition technology. This manner is still a distinctive way to increase insufficient water-soluble constituents (Adali et al., 2020). It is an advanced particle establishment manner in which a liquid matter is intended to spray in hot air or nitrogen, followed by lyophilization. The feed rate, which impacts the aerodynamic diameter and size, is the chief cause for decreasing the efficacy of this method (Pramanik et al., 2021). Although some studies have shown the merits and exclusive properties of SFD, the technique has some limitations. SFD has high capital and operational costs, chiefly due to low-pressure and low-temperature necessities. In addition, most developed SFD units are batch-type and unsuitable for commercial usage (Vishali et al., 2019).

### 3.2.3 Solvent precipitation technique

This is a direct-controlled crystallization technique that involves sono-crystallization and microprecipitation to distribute the narrow drug particle size in a crystalline form (Patil and Sarasija, 2012). Sono-crystallization is an exclusive technique for planning an NP system. The energy involved in this procedure will ameliorate the target media's nucleation level more than the others, where this manner is not incorporated (Pramanik et al., 2021).

The use of various types of inhaled NPs was indicated in different sections of this review.

## 4 The effects of nanoparticles on respiratory diseases

### 4.1 Asthma

Asthma is an inflammatory disorder of the lungs characterized by reversible airway obstruction, bronchial hyper-responsiveness, and chronic airway inflammation (Cukic et al., 2012). Long-term exposure to irritants causes an inflammatory response in the lungs, resulting in narrowing the small airways and lung tissue damage (Yang et al., 2019). Anti-inflammatory effects of nanoparticles were shown in various inflammatory disorders previously. Nanoparticles increase the therapeutic effect by facilitating the delivery of the drug to the target tissue, thereby improving the deposition of the drug in

the lungs (Yang et al., 2019). In several *in vitro* and *in vivo* studies, the effects of nanoparticles on asthma were reported.

#### 4.1.1 *In vitro* studies

The poly(lactic acid) nanoparticles were utilized to co-deliver theophylline and budesonide. These two compounds were co-encapsulated in the polymeric nanoparticles through the process of double emulsion solvent diffusion. The poly(lactic acid) nanoparticles can encapsulate both hydrophilic and hydrophobic drugs and be used for co-delivery purposes, although it has a hydrophobic nature. The drug release was investigated using Franz-type diffusion cells using a 1:1 ratio of the simulated lung fluid and methanol in the airway epithelial cell line (16HBE14o-). Inhaled nanoparticles lead to a fine particle fraction of 75% and 48% for theophylline and budesonide, respectively. The results showed that budesonide and theophylline drug-loaded NPs are appropriate drug delivery methods for the treatment of asthma and COPD (Buhecha et al., 2019). In the other work, nanoliposomal salbutamol sulfate (SBS) dry powder inhalation for facilitated asthma therapy was used, and the results showed prolonged *in vitro* drug release of above 90%. In this study, researchers overcome the deprived oral bioavailability of SBS because of its hepatic metabolism and short half-life (Honmane et al., 2019). Elimination of low-solubility issues with curcumin by liposomal curcumin improved patient compliance due to the liposomal form. The suppression of proinflammatory markers, such as IL-6, IL-8, IL-1 $\beta$ , and TNF- $\alpha$ , by curcumin liposomes is the result of this study (Ng et al., 2018).

In another work, quercetin-loaded liquid crystalline NPs (LCN) and surface-modified liquid crystalline NPs (sm-LCN) were formulated, and their anti-inflammatory activity in a human primary bronchial epithelial cell line (BCi-NS1.1) stimulated with lipopolysaccharide (LPS) was investigated. Quercetin LCN and sm-LCN meaningfully reduced the fabrication of IL-1 $\beta$ , IL-6, and IL-8 compared to the LPS-only group. Encapsulation of quercetin into LCN and sm-LCN increased its anti-inflammatory property compared to quercetin in DMSO. Moreover, quercetin LCN and sm-LCN displayed comparable properties to fluticasone in terms of decreasing the fabrication of IL-1 $\beta$  and IL-6. Therefore, quercetin-LCN and sm-LCN can be possible remedial interventions for asthma as they are effective in inhibiting the fabrication of main proinflammatory cytokines related to the development of asthma (Yong et al., 2019).

Treatment with lipid nanoparticles decreased the secretion of proinflammatory cytokines, including IL-4 and IL-13, in asthmatic mice and could significantly diminish allergic asthma by hindering the overexpression of proinflammatory cytokines in the airway (Zhang M. et al., 2022). Treatment of asthmatic mice with poly(lactic-co-glycolic acid) (PLGA)-levamisole NPs reduced airway hyper-responsiveness, eosinophils in the BALF, immunoglobulin levels, and Th2-, Th9-, and Th17-derived cytokines (Fang et al., 2022).

Asthma-induced airway inflammation can be prevented by nanoparticle administration of LNA oligonucleotides. Treatment with oligonucleotide can decrease the gene expression of mmu-miR-145a-5p, which is elevated in the animal model of disease and human asthma (Ahmad, 2022). *In vitro*, the rate of oxygen consumption (OCR) in the mitochondrial respiratory function with exposure to CuONPs (25, 50, and 100  $\mu$ g/ml doses) was

measured. The results indicated that the 25 and 100  $\mu$ g/ml doses of CuONP decreased OCR, but the 50  $\mu$ g/ml dose increased OCR (Bushueva et al., 2023).

In ovalbumin (OVA)-induced asthma model and OVA-severe (lung, heart, liver, kidney, and spleen) models, AuNP-loaded macrophages migrated to the lung tissue. The results showed that macrophages loaded with AuNPs exhibited a sixfold faster migration into the lung tissue in the OVA-severe model compared to the OVA-asthma model. Macrophage phagocytic capacity and the ability to encapsulate drugs and therapeutic particles were also increased (Kang et al., 2022).

#### 4.1.2 *In vivo* studies

Aerosolized liposome formulation for the pulmonary delivery of an anti-asthmatic treatment showed a significantly more prolonged anti-asthma effect of the R-TBH liposome compared to the R-TBH (R-terbutaline hydrochloride) solution (Li Q. et al., 2018). The modulation of an asthmatic response using TiO<sub>2</sub> or AuNPs in a murine model of diisocyanate-induced asthma was studied. The results displayed that a low, intrapulmonary doses of NPs improved pulmonary inflammation and hyperreactivity in a mouse model of asthma (Hussain et al., 2011). Treatment of the house dust mite-induced allergic airway disease model with QDs improved select cytokine production due to QDs (Scoville et al., 2019).

The effect of hyaluronic acid (HA) decorated, ferulic acid (FA)-loaded chitosan (CS) NP (FACHA) aerosolized by vibrating mesh nebulizer as a strategic combination of drug, nanocarrier, and delivery device for effectual asthma control was explored. FACHA displayed a globular structure with proper size ( $164.2 \pm 9.7$  nm) and mass median aerodynamic diameter (MMAD) of  $1.81 \pm 0.15$   $\mu$ m, indicating effective drug deposition. *In vivo* inhalation and toxicology evaluation verified safety, and FACHA prophylactically ameliorated inflammation, airway hypersensitivity, and remodeling in ovalbumin (OVA)-induced mice model of asthma (Dhayanandamoorthy et al., 2020).

The liposomal budesonide treatment in asthmatic mouse models increased efficacy in the reduction of inflammation and showed fewer side effects compared to steroids (Konduri et al., 2005). *In vivo* studies of aerosolized liposomal formulation of SBS also revealed greater drug retention time and anti-inflammatory and anti-asthmatic effects compared with the free drug (Chen et al., 2012).

The administration of liposomes (composition: dioleoylphosphatidylcholine and cholesterol) also reduced IL-6 and stimulated tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) in primary human nasal epithelial cells. *In vivo* studies on allergic airway inflammation induced by OVA have also shown a decline in immune responses and airway responsiveness (AHR) (Komalla et al., 2020).

The effect of bilirubin-based NPs (BRNPs) as nanomedicine for the treatment of allergic lung inflammatory disease was reported. The properties of BRNPs on Th2 (allergen-specific type 2 T-helper) immune responses were studied *in vivo* and *in vitro*. After intravenous injection, BRNPs displayed more serum concentration and a longer circulation time of BR than the intraperitoneal injection of BRNPs or unconjugated bilirubin (UCB). The anti-asthmatic properties of BRNPs were evaluated in a mice model of allergen-induced asthma. Treatment with

BRNPs, in comparison to UCB, inhibited the indices of experimental allergic asthma and intensely improved Th2-related allergic lung inflammation. In agreement with these results, BRNPs also decreased Th2 cell populations and the expression of associated cytokines by antibody-stimulated CD4<sup>+</sup> T Cells *in vitro* (Kim et al., 2017).

Encapsulated and andrographolide (AG) in NP (AGNP) were assessed for anti-asthmatic efficiency by oral/pulmonary delivery. AGNP showed 5.47% drug loading with a sustained drug release *in vitro*. Pharmacokinetic results indicate improved AG bioavailability upon AGNP administration by pulmonary route. Cell numbers; IL-4, IL-5, and IL-13 levels in BALF; and serum IgE content were significantly decreased after administration of AGNP. AGNP-mediated inhibition of NF- $\kappa$ B was also meaningfully higher compared to free-AG. Furthermore, results displayed improved remedial efficiency, and AGNP effectively controlled mild and intense asthma, especially when administered by inhalation. Cell numbers and IL-4, IL-5, and IL-13 levels in the BALF were also significantly reduced by AGNP (Chakraborty et al., 2019).

The capability of an intranasal treatment with NPs targeting IL4Ra to regulate lung inflammation in OVA-sensitized mice was tested. The properties of NPs treatment on the activation of lung inflammatory cells and their capability to proliferate and generate cytokines were identified by fluorescence-activated cell sorting (FACS) examination. Treatment with the anti-IL4Ra NPs meaningfully reduced proinflammatory cytokine expression and release in the BALF and lung tissue. In addition, the numbers of lung tissue lymphocytes, neutrophils, and eosinophils were reduced. Anti-IL4Ra NPs deactivated CD4 and CD8 T cells in lung tissue and inhibited their capacity to generate proinflammatory cytokines meaningfully higher than treatment with free anti-IL4Ra. Moreover, it induced a sustained low level of lung inflammation for 1 week following the last instillation compared to the treatment with free anti-IL4Ra antibodies (Halwani et al., 2016).

The remedial properties of vasoactive intestinal peptide (VIP) when conjugated to  $\alpha$ -alumina NP ( $\alpha$ -AN) to inhibit the enzymatic degradation of VIP in the respiratory system in mice were investigated. The number of eosinophils, serum IgE level, Th2 cytokines, and AHR were reduced by  $\alpha$ -AN-VIP, which was more noticeable than VIP alone. These findings show that  $\alpha$ -AN-VIP could be regarded as an actual nanomedicine for the treatment of asthma (Athari et al., 2016).

The potential of nasal-instilled AuNPs to inhibit allergen-stimulated asthma in a murine model showed that in both mice strains (Swiss-Webster, outbred) and A/J (inbred), AuNPs prevented the allergen-induced gathering of inflammatory cells along with the fabrication of both proinflammatory cytokines and reactive oxygen species (ROS). In A/J mice, known as genetically asthma-prone animals, instilled AuNPs inhibited mucus generation, peribronchiolar fibrosis, and airway hyperreactivity triggered by allergen provocation (Barreto et al., 2015). Treatment with CuONPs in the respiratory structure of BALB/c mice displayed toxic effects, which were related to MAPK phosphorylation. Moreover, CuONP exposure intensified the progress of asthma (Park et al., 2016).

The immune-remedial properties of recombinant *Caryota mitis* profilin (rCmP)-loaded PLGA NPs and the principal mechanisms

involved were investigated. The rCmP-loaded PLGA NPs prevented the production of specific IgE and secretion of the Th2 cytokine interleukin-4; enabled the production of particular IgG2a, the secretion of the Th1 cytokine interferon-gamma (INF- $\gamma$ ), the ratio of Th1/Th2; and improved allergic signs (Xiao et al., 2013). The effect of encapsulation of LPS into PLGA NPs administered by sublingual (SLIT) in asthmatic BALB/c mice was studied. Treatment with LPS-PLGA (200 nm diameter) meaningfully enhanced the cytokine levels. Histological and BALF analysis showed a reduction of eosinophilic and other inflammatory cell infiltration. Therefore, LPS-PLGA NPs can efficiently shift Th1 to Th2/Treg (Khakzad et al., 2019). In another study, the mice were treated with interferon- $\gamma$  (IFN- $\gamma$ ) pDNA-loaded chitosan nanoparticles intranasally. The internalization was increased significantly by macrophages and bronchial epithelial cells. It appears that the AHR was reduced, and the lung morphology was normalized, perhaps via signal transducer and activator of transcription 4 (Kumar et al., 2003).

Grape seed-solid lipid nanoparticle (SLN) administration (30  $\mu$ g/25 g/wt. 1, 2, 3, and 6 days) in C57BL/6 male mice lungs was more effective than grape seed alone in attenuating asthma process. Moreover, SLNs showed a better role in oxidative stress, suppressing airway epithelial cells, indicating the potential treatment of SLNs, drug efficacy, and drug delivery to ameliorate asthma response (Castellani et al., 2018).

In general, *in vitro* studies have shown a reduction of inflammation in lung tissues in asthma and elucidated some molecular mechanisms due to the application of nanoparticles. *In vivo* studies, while confirming the results obtained from the *in vitro* studies, showed higher stability and treatment duration of nanodrugs than ordinary drugs. Table 1 illustrates the effects of various nanoparticles on asthma. Figure 5 demonstrates the molecular mechanisms involved in the effects of nanoparticles on asthma.

## 4.2 Chronic obstructive pulmonary diseases and chronic bronchitis

COPD is a chronic inflammatory lung disease that causes airway obstruction. Its symptoms include difficulty breathing, coughing, and increased mucus secretion. This disease is caused by long-term exposure to irritating or airborne particles, which are often caused by cigarette smoke (Alqahtani, 2022). Airways were narrowed, and their flexibility was reduced in COPD, resulting in air trapping in the lungs (Alqahtani, 2022). Nanomaterial drugs showed therapeutic potential against various respiratory disorders as they are required in small quantities and can be effectively targeted to diseased tissue microenvironment, hence having minimal side effects (Saxena et al., 2022). The effects of nanoparticles on COPD were shown in different *in vitro* and *in vivo* studies.

### 4.2.1 *In vitro* studies

In an earlier study, COPD cells (A549) were treated with a microRNAs (miRNAs)-loaded system using a polyester-based drug delivery platform, poly(glycerol adipate-co- $\omega$ -pentadecalactone), where it was cationic and made better surface interaction with the negatively charged cell membrane. The process of preparing nanoparticles was a single emulsification-solvent evaporation



**TABLE 1 Effects of various nanoparticles on asthma and COPD.**

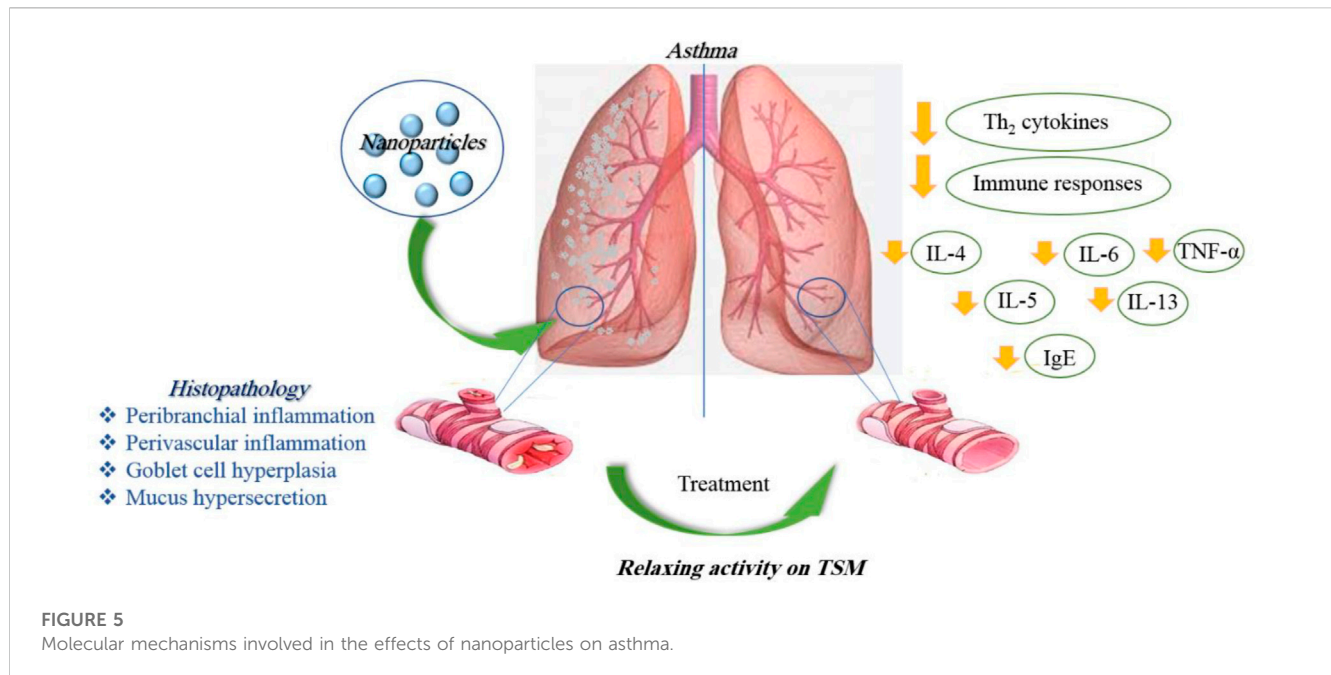
Study type	Study design	Dose and duration	Effect	Ref.
<i>In vitro</i>	Aerosolized R-TBH liposome formulation	1 mg/ml, 2 min	Prolonged anti-asthma effect	Li et al. (2018b)
	SBS dry powder inhalation	200 µg aerosolized, 14 h	Prolonged <i>in vitro</i> drug release ↑ Bioavailability of SBS (↑hepatic metabolism and ↓ half-life)	Honmane et al. (2019)
	TiO <sub>2</sub> or AuNPs in a murine model of asthma	0.4 mg/mL or 0.8 mg/kg, 16 days	↓ AHR, BALF cell and cytokine, lung histology, serum IgE	Hussain et al. (2011)
	Solid lipids for delivering oligomeric proanthocyanidins from grape seed extract	2.5, 5, and 10 µM, 24, 48 h	High cellular uptake and retention time, ↓ ROS ↓ Inflammatory responses	Castellani et al. (2018)
	LPN for treatment of COPD	N/P ratio (N/p ¼ 1, 2, 4, 8, and 16), 1 h	↓ Steroid resistance in COPD models ↓ IL-8	Chikuma et al. (2020)
	Cadmium oxide nanoparticle effects on COPD	1 µg/ml, 24 h	Facilitated post-translational citrullination of proteins in COPD	Hutchinson et al. (2018)
	ZnO-NP toxicity in COPD patient samples	10, 20, and 40 µg/ml, 6 h	↑ Expression of tumor suppressor protein p53, Ras p21	Kumar et al. (2015)
	CaP/PLGA NP-mediated siRNA delivery in	50 µL, containing 5 µg siRNA	↓ Inflammatory effects ↓ CCL-2, IP-10, and IFN-γ	Frede et al. (2017)
	DNA damage protection by nanoforms of quercetin	25 µM, 2 h	↓ Oxidative stress ↑ CAT mRNA expression	Habas et al. (2018)
<i>In vivo</i>	Al <sub>2</sub> O <sub>3</sub> NP treatments for COPD	0, 0.4, 2 mg/ml, 24 h	↓ PTPN6 Phosphorylation of STAT3 lung inflammation	Li et al. (2017)
	ZnO-NP treatments in COPD samples	10, 20, and 40 µg/ml, 6 h	↑ Expression of tumor ↓ Protein p53, Ras p21, JNKs	Kumar et al. (2015)
	DIN for the delivery of siRNA to lung vasculature	2.5 mg/kg Tie2 siRNA after 3 days	↓ Proinflammatory cytokines	Khan et al. (2015)
	LLC for therapeutic intervention in asthma	1 and 5 µg/ml, 24 h	↑ Patient compliance ↓ Proinflammatory markers	Ng et al. (2018)
	QDs in HDM-induced allergic airway disease in mice	100 µg, 140 ng, 10 days	↑ Select cytokine production	Scoville et al. (2019)
	CuONPs-exposed BALB/c mice	25, 50, and 100 lg/kg	↓ AHR, inflammatory cell, cytokines, ROS, MAPK ↑ MAPKs phosphorylation	Park et al. (2016)
	Lungs targeted using IV. Inj of NPs	10 mg/kg, 4 weeks	Significant inhibition of MMPs in the lungs	Parasaram et al. (2016)

R-TBH, R-terbutaline hydrochloride; SBS, nanoliposomal salbutamol sulfate; TiO<sub>2</sub>, titanium dioxide; AHR, airway hyperreactivity; LPN, lipid-polymer nanoparticle; BAL, bronchoalveolar lavage; GNPs, gold nanoparticles; DIN, dendrimer-inspired nanomaterials; LLC, liposomes loaded with curcumin; QDs, quantum dots; HDM, house dust mite; CuONPs, copper oxide nanoparticles; ROS, reactive oxygen species; Dur, duration; MMPs, matrix metalloproteinases; CaP, calcium phosphate.

technique using cationic precursors. The *in vitro* drug delivery method was used to investigate the release profile of miR-146a. The results indicated successful miR-146a delivery to lung cells. It appears that the RNA efficacy remained even after the degradation of nanoparticles (Mohamed et al., 2019).

The potential effect of lipid-polymer nanoparticles (LPNs) for the treatment of COPD was studied. LPNs (N/p = 1, 2, 4, 8, and 16,

72 h) composed of a poly(lactic acid)/Mn-porphyrin dimer/and a cationic lipid (DOTAP) showed an actual tool against COPD. LPNs ameliorated the steroid resistance in COPD models *in vitro*, as proven by the decreased expression levels of IL-8 (Chikuma et al., 2020). Cadmium is a heavy metal that does not have a known physiological function and is often considered a toxicant (Taghavizadeh Yazdi et al., 2021). Cadmium nanoparticles as a



potential cause of citrullination in COPD were shown by the effect of cadmium oxide nanoparticle (CdO NPs, 1  $\mu\text{g}/\text{ml}$ , for 24 h) exposure, which facilitated post-translational citrullination of COPD proteins (Hutchinson et al., 2018). To explore differences in the recovery of exhaled NPs in subjects with COPD and never-smoking controls, the specific and local modulation of the inflammatory response by calcium phosphate (CaP)/PLGA nanoparticle-mediated siRNA (50  $\mu\text{L}$ , containing 5  $\mu\text{g}$  siRNA) delivery showed a promising approach for the treatment of inflammatory disorders of the lung. The NPs protect the siRNA from the environment and can be used to decrease gene expression of CCL-2, IP-10, and IFN- $\gamma$  and, thereby, abrogate inflammatory responses in the lung (Frede et al., 2017). DNA damage protection by nanoforms of quercetin (25  $\mu\text{M}$ , 2 h) in lymphocytes of patients with COPD exposed to the food mutagen 2-amino-3-methylimidazo (Yang et al., 2020) quinolone (IQ) was demonstrated. The ability of flavonoids to reduce oxidative stress and induce human protective enzyme systems, especially an increase in catalase mRNA expression, causes this protection (Habas et al., 2018).

#### 4.2.2 *In vivo* studies

Several drug formulations containing organic polymer-based NPs show more effective results than metallic NPs against COPD *in vivo* (Saxena et al., 2022).  $\text{Al}_2\text{O}_3$  NPs (0, 0.4, 2 mg/ml, 24 h) exposure suppressed PTPN6 and phosphorylation of STAT3. Rescue of PTPN6 expression or application of a STAT3 inhibitor efficiently protected murine lungs from inflammation, as well as, in part, from the induction of COPD-like effects (Li et al., 2017). The majority of the studies concerning ZnONPs toxicity have been performed. The results showed a significant and dose-dependent increase in the expression of tumor suppressor proteins p53, Ras p21, and JNKs (Jun-N-terminal kinase) in blood samples of 31–85-year-old COPD patient samples after 6 h of ZnO treatment at the 10, 20, and 40  $\mu\text{g}/\text{ml}$  concentrations (Kumar et al., 2015). Targeting of nanoparticles to damaged elastin in the lungs and a controlled release of

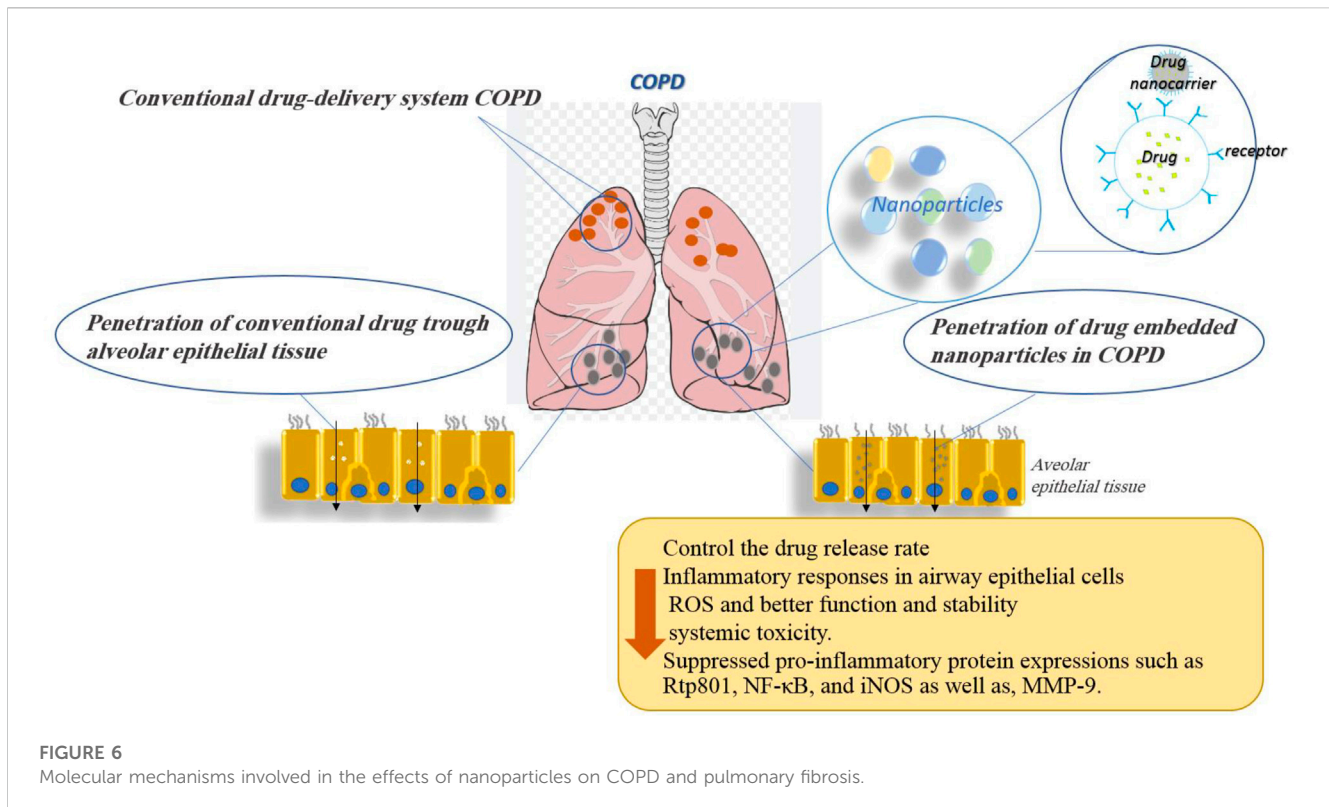
doxycycline played an important role in reducing the MMP activity levels in rats that received single bovine serum albumin (DOX-BSA, 10 mg/kg, for 4 weeks) NP injection. An obvious advantage of such treatment is the requirement for lower dosages. Targeted single-dose delivery of doxycycline NPs significantly inhibited matrix metalloproteinases (MMPs) in the lungs for up to 4 weeks *in vivo* (Parasaram et al., 2016).

It was indicated that the cationic polymeric nanoparticles could be used successfully for drug delivery of miRNAs to the negatively charged cell membranes in COPD and lung diseases. Solid lipids prepared to deliver aerosolized oligomeric proanthocyanidins from grape seed extract (15  $\mu\text{L}$ , 2.5, 5, and 10  $\mu\text{M}$ ) were used to treat COPD in mice. The results exhibited high cellular uptake and retention time, leading to the reduction of ROS and stability at 48 and 72 h compared with the free grape seed extract in both *in vitro* and *in vivo* settings. It also reduced the inflammatory responses in airway epithelial cells (Castellani et al., 2018).

In summary, the use of nanoparticles as drug delivery vehicles in COPD may prove beneficial for NPs as they could provide sustained drug release, overcome airway defenses, and target diseased cells or tissues. Indeed, the nanoparticle-based drug was shown to control the drug release rate and allowed therapeutic doses to be maintained in the lungs for longer periods while decreasing systemic toxicity. The effects of various nanoparticles used for the treatment of COPD disorders are shown in Table 1. Figure 6 demonstrates the molecular mechanisms involved in the effects of nanoparticles on COPD and pulmonary fibrosis (PF).

#### 4.3 Pulmonary fibrosis and noxious agent-induced lung disorders

Idiopathic pulmonary fibrosis (IPF) is a chronic interstitial lung disease characterized by fibrosis, inflammation, and destruction of



lung architecture. Damage to the alveolar epithelium and abnormal wound repair are theorized to be key factors in the development of this disease (Mei et al., 2022). Most often, the cause of pulmonary fibrosis is unknown, which is called idiopathic pulmonary fibrosis (Glass et al., 2022). Various nanoparticles showed therapeutic effects on PF in different *in vitro* and *in vivo* studies.

#### 4.3.1 *In vitro* studies

Nanostructured lipid carriers exhibiting high encapsulation efficiency (over 90%) and exceptional release profiles have the potential to treat cystic fibrosis (Moreno-Sastre et al., 2016). The administration of liposomal amikacin via inhalation for the treatment of cystic fibrosis decreased the bacterial density (Garbuzenko et al., 2014). Polyethylene glycol density and doxorubicin (DOX) were payloads on the interaction of PAMAM dendrimer with an *in vitro* pulmonary epithelium model (Calu-3). The results showed enhanced DOX transport across the pulmonary epithelium and indicated high final particle fractions (>82%) of aerosol formulations (Zhong et al., 2017).

The effect of QD containing a cadmium-selenide (CdSe) core with a zinc sulfide (ZnS) shell functionalized with carboxyl or amine terminal groups in the lung of rats was studied. Lung injuries and inflammation, such as reduced LDH, albumin, and monocyte chemotactic protein-1 (MCP-1) in the BALF, showed dependently improved concentration, which on the seventh day and for 14 hours post-exposure for both forms of QD. Both QDs missed their fluorescent assets and destabilized after 1 week in the lung. No Cd was identified in the liver, spleen, heart, brain, or blood at any time point. Cd appeared in the lung-associated lymph nodes

(LALN) and kidneys beginning at 1–2 weeks after exposure (Roberts et al., 2013).

Administration of Si/SiO<sub>2</sub> QDs on cell redox homeostasis in MRC-5 cells showed a time-dependent reduction in cell viability and biomass due to exposure to QDs. Si/SiO<sub>2</sub> QDs showed a noteworthy influence on the intracellular distribution of glutathione. Si/SiO<sub>2</sub>-QDs also stimulated protein oxidation and actin S-glutathionylation (Stan et al., 2014). In the other research, male Sprague–Dawley rats were exposed to CeO<sub>2</sub> or CeO<sub>2</sub> coated with a nanolayer of amorphous SiO<sub>2</sub> (aSiO<sub>2</sub>/CeO<sub>2</sub>) by a single intratracheal instillation. The results show that the CeO-NPs induce PF, matrix metalloproteinase-9 (MMP-9), and tissue inhibitor of MMP, but SiO<sub>2</sub> coating reduced CeO<sub>2</sub>-induced inflammation, phospholipidosis, and fibrosis. Therefore, SiO<sub>2</sub> coating showed a protective effect on CeO<sub>2</sub>-induced PF (Ma et al., 2015).

The neutrophils in a rat blood sample were stimulated with different concentration gradients of ZnO-NPs (0, 5, 10, 15, and 20 mg/L). The expression levels of the relevant inflammatory factor, such as IL-8, were examined. The results indicated that ZnO-NPs are capable of inducing neutrophil autophagy and regulating ZnO-NPs-induced neutrophil inflammatory response (Zhang et al., 2023).

#### 4.3.2 *In vivo* studies

Treatment of acute lung injury induced by LPS in mice with nanoemulsions containing pequi oil-oleic acid (20 mg/kg), reduced myeloperoxidase (MPO), TNF-α, IL-1β, IL-6, MCP-1, and KC levels and pulmonary leukocyte accumulation if administered orally, which can be a substitute strategy to treat respiratory inflammatory disorders (De Sá Coutinho et al., 2020s).

Complexation of small interfering RNA (siRNA) resulted in greater cellular uptake and enhanced *in vitro* silencing efficiency of TNF- $\alpha$  in LPS-activated mouse macrophage cell line RAW264.7, compared to morpholinium-containing dendriplexes due to higher pKa values leading to improved protection against enzymatic degradation and a higher cellular uptake. In an *in vivo* LPS-induced murine model of acute lung injury, nasal administration of siRNA (2.0 mg/kg in 30  $\mu$ l, 24 h prior to administration of LPS) also showed a strong potential of phosphorus dendriplexes in lung delivery of siRNA for treating inflammatory lung diseases (Bohr et al., 2017). Administration of QD (6  $\mu$ g/kg by oropharyngeal aspiration) induced acute lung inflammation, but lung mechanics were improved by QD exposure in A/J mice (Scoville et al., 2018). The inhaled tacrolimus (Tac, 60  $\mu$ g/mouse) albumin NPs revealed significant anti-fibrotic efficiency in mice with bleomycin-induced PF, which was much better than the effectiveness resulting from intraperitoneal administration of Tac based on histopathological results (Seo et al., 2016). Thymoquinone (TQ)-PLGA-PVA-NPs could attenuate bleomycin-induced PF by inhibiting lung inflammation and suppressing bleomycin-induced oxidative stress. The IL-10 level was significantly reduced in rats treated with bleomycin + TQ-PLGA-PVA-NPs. In addition, the TGF- $\beta$ 1 level was significantly increased in bleomycin-treated rats compared with their levels in TQ-PLGA-PVA-NP-treated rats (Saghir et al., 2019).

Exposure of mice to nano-Co (50  $\mu$ g) induced acute lung inflammation and lung injury indicated by an increased number of neutrophils, CXCL1/KC level, LDH activity and total protein level in the BALF, and neutrophils. Macrophage infiltration in the alveolar space and interstitial tissues, as well as immunostaining of cell proliferation markers, Ki-67 and PCNA, and the DNA damage marker,  $\gamma$ -H2AX, in bronchiolar epithelial cells and hyperplastic type II pneumocytes in the lungs, 7 after exposure was observed. In addition, interstitial fibrosis and inflammatory cell infiltration in the alveolar septa were observed 4 months after exposure. Moreover, nano-Co caused an increased level of 8-OHdG in the genomic DNA of mouse lung tissues. The results indicated the induction of oxidative stress, lung inflammation and injury, and cell proliferation due to nano-Co and showed the potential health effects of nanoparticle exposure (Wan et al., 2017).

Intratracheal instillation of NiO NPs (50, 100, and 200  $\mu$ g/rat) was compared to NiCl<sub>2</sub> (171.1  $\mu$ g) in an ovalbumin-induced allergic airway inflammation model. The results showed that acute neutrophilic inflammation and eosinophils were recruited 3 and 4 days after NiO NP instillation. However, the induction of inflammation was similar to that of NiCl<sub>2</sub> 24 h after instillation (Lee et al., 2015). It was reported that silica nanoparticles (SiNPs) induce PF by enhancing autophagosome accumulation and apoptosis *in vivo* and *in vitro* (Zhao et al., 2019). In addition, CuONPs aggravated pulmonary inflammation in a dose-dependent way. CuONPs induced apoptosis of epithelial cells, which was partially caused by increased ROS. In addition, CuONP exposure promoted collagen accumulation and expression of the progressive fibrosis marker  $\alpha$ -SMA in the lung tissues, indicating that CuONP inhalation could induce PF in C57BL/6 mice (Lai et al., 2018). Progressive massive pulmonary fibrosis, diffuse interstitial fibrosis, and collagen accumulation

involved in the development of pulmonary alveolar proteinosis following nano-indium-tin oxide nanoparticles (ITO) exposure in rats were shown (Liu et al., 2022). Chang et al. (2017) reported extensive lung fibrotic injury in histological investigation and enhanced content of hydroxyproline, collagen types I and III in rat lung tissue exposed to nano-nickel oxide (NiO, 0.015, 0.06, and 0.24 mg/kg; 6 weeks). This study suggested that PF induction by NiO-NPs may be related to TGF- $\beta$ 1 activation. Carbon black nanoparticles (5 mg/m<sup>3</sup> and 30 mg/m<sup>3</sup>; 14, 28, and 90 days) also induced PF in rats, which might be linked with persistent lung inflammation through NLRP3 inflammasome activation (Zhou et al., 2020).

The reviewed articles published on the effects of nanoparticles on PF indicated improvement effects of some nanoparticles on PF. However, other studies showed the induction and aggravation of PF by other nanoparticles. Table 2 shows the effects of various nanoparticles on lung fibrosis. Figure 6 indicates the molecular mechanisms involved in the effects of nanoparticles on COPD and pulmonary fibrosis.

#### 4.4 Lung infections (viral, microbial, parasites, and fungi)

Serious lung infections, such as pneumonia, tuberculosis, and viral lung infections, are serious lung insults and can be life-threatening (Cai et al., 2022). Lung infections involve most of the air sacs and the airways that transmit air to the lungs to a lesser extent (Cai et al., 2022). A variety of treatments or diagnostics have been utilized to manage lung infections, but the emergence of drug-resistant bacteria and the adverse effects experienced once they reach the lung environment have made treatment more difficult (Muhammad et al., 2022). The need for new strategies is felt more than before to control the activities of bacteria. In this field, a promising approach can be the use of nanomaterial science, and the knowledge that is expanding in the field of nanomaterials synthesis should be considered as a global strategy (Muhammad et al., 2022).

##### 4.4.1 *In vitro* studies

Various studies have indicated the effects of NPs on lung infections. To study the anti-tubercular drug, bedaquiline (BDQ) of chitosan nanoparticles (NPs) with an optimized batch showed a particle size of 109.7  $\pm$  9.3 nm and a zeta potential of 36  $\pm$  2.1 mV. *In vitro* deposition studies on non-viable cascade impactors have shown similar toxicity compared to conventional drugs. The most intriguing advantages of NPs were loading capacity, surface properties, stability in enzymes, and excellent pharmacokinetics (Rawal et al., 2018).

*In vitro* microbial studies displaying the dose-dependent manner effects of the complex of phytosome loaded drug delivery of gingerol (PG) on the respiratory infective bacterium, which verified the bactericidal properties (Singh R. P. et al., 2018).

In the United States National Library of Medicine, the amikacin-loaded liposomes were used to treat lung infection to non-tuberculous mycobacteria that have increased the efficacy, tolerance, and safety and shown a promising future for treatment, especially systemic infections (clinicaltrials.gov). Liposomal amikacin was more successful than free amikacin and

TABLE 2 Effects of various nanoparticles on lung fibrosis, lung infection, and noxious agent-induced lung disorders.

Study type	Study design	Dose and dur.	Finding	Ref.
<i>In vitro</i>	PEG and DOX payload interaction with PAMAM	30 $\mu$ l DOX or 50 nM 5.5 h	$\uparrow$ DOX transport High aerosol particle fractions (>82%)	Zhong et al. (2017)
	OSI-420-QDs in NSCLC cell lines	400 $\mu$ L QDs (1 mg/ml) for 72 h	Significantly better efficacy than pure drugs in all tested cell lines	Kulkarni et al. (2019)
	QD-CdSe-ZnS	12.5, 5.0, or 1.25 $\mu$ g/rat on days 0, 1, 3, 5, 7, 14, and 28	Dose-dependent lung injury and inflammation Lose lung fluorescent and destabilize after 1 week	Roberts et al. (2013)
	Si/SiO <sub>2</sub> QDs on cell redox homeostasis MRC-5 cells	25 and 200 $\mu$ g/ml for up to 72 h	$\downarrow$ Cell viability and biomass Impact on the intracellular distribution Protein oxidation and actin S-glutathionylation	Stan et al. (2014)
	nAg and nZnO + CA, TA, and FA in BALF and Gamble solution	0, 3, 10, 30, 100, and 300 mg after 24 h	$\uparrow$ Aggregation of nAg and nZnO Release Zn <sup>2+</sup> promoted by CA and Ag <sup>+</sup>	Zhong et al. (2019)
<i>In vivo</i>	QD-stimulated in C57BL/6J and A/J mice	6 $\mu$ g/kg Cd =	QD-induced acute lung inflammation Impacted lung mechanics in A/J mice only	Scoville et al. (2018)
	Cobalt nanoparticles exposed mice	50 $\mu$ g per at day 1, 3, 7 and 28	$\uparrow$ BALF neutrophils, CXCL1/KC, LDH and total protein, alveolar space and interstitial tissues neutrophils and macrophages infiltration $\uparrow$ Ki-67, PCNA, and $\gamma$ -H2AX	Wan et al. (2017)
	NiO-NPs- exposed Wistar rats	50, 100, and 200 $\mu$ m <sup>2</sup> /rat for 1, 2, 3, and 4 days	Induced neutrophilic inflammation $\uparrow$ Intracellular cotaxin and LDH in alveolar macrophages and normal lung tissue lysis	Lee et al. (2015)

PEG, poly(ethylene glycol); DOX, doxorubicin, BALF; bronchoalveolar lavage fluid; PAMAM, poly(amidoamine) dendrimer; OSI-420, desmethyl erlotinib; NSCLC, non-small cell lung cancer; CdSe, cadmium-selenide; ZnS, zinc sulfide; CA, citric acid; TA, tartaric acid; FA, fulvic acid; ALF, artificial lysosomal fluid; Dur, duration.

can be administered by inhalation under the trade name Arikace™ (Meers et al., 2008). These structures seem to be simply aerosolized and have a noteworthy result on delivery in cell culture. Their capability to better intracellular delivery *in vivo* will be affected by several factors, resulting in lung surfactant on the constancy of the structures and their rate of clearance. The fate of liposomes in the lungs was investigated, and their residence time in the lungs was found to be dependent on the ventilation degree and their structure (Cryan et al., 2006).

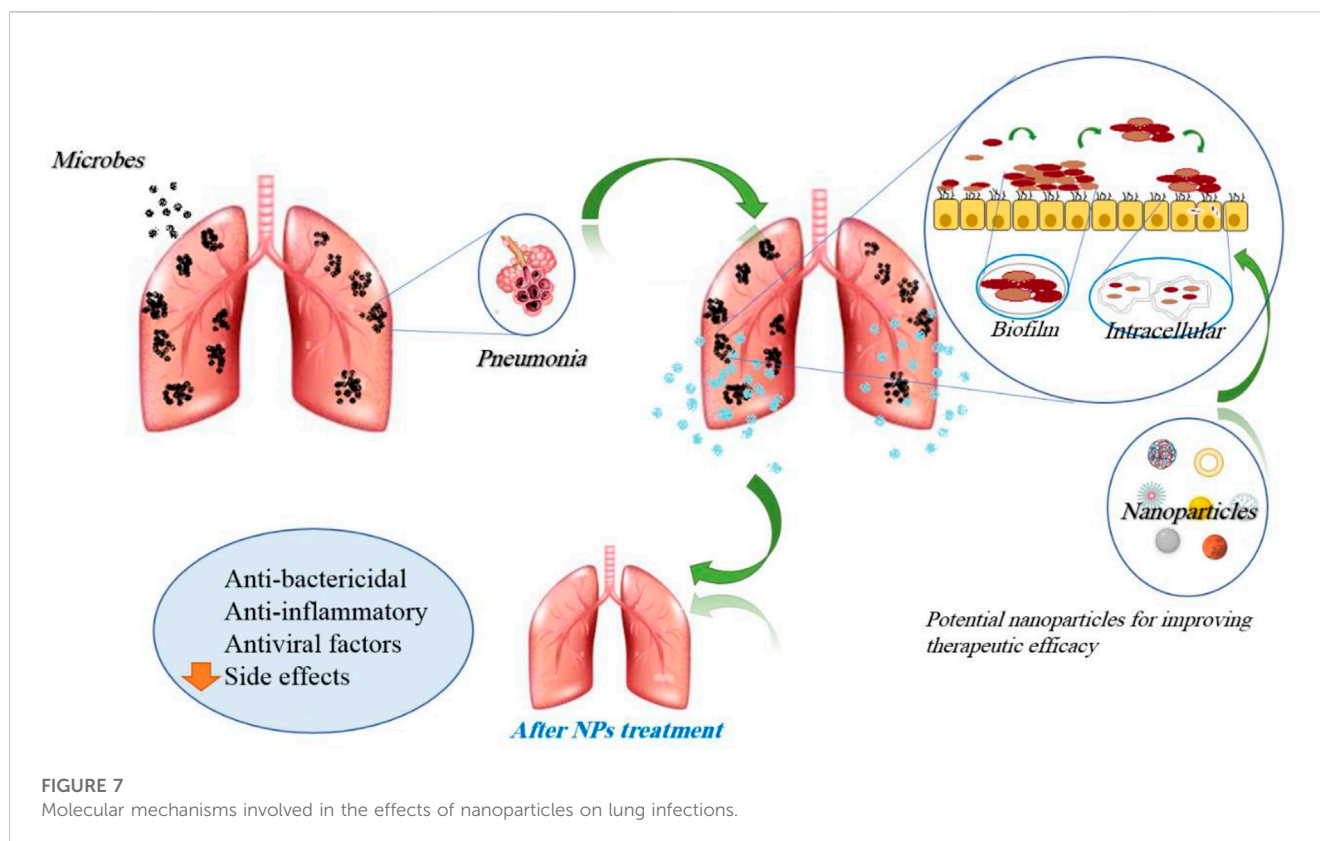
Calcifediol (25(OH) D) in liposomes was encapsulated to enable aerosolization for the capability to inhibit pulmonary infection by *Pseudomonas aeruginosa*, which inhibits *Pseudomonas* infection in human bronchial epithelial cells (Castoldi et al., 2017).

A type of silica nanoparticles loaded with anticancer drug doxorubicin (DOX) and antimicrobial peptide HHC36 (AMP) (MSN@DOX-AMP) significantly treated bacterial infection and eliminated *in situ* lung tumors in a commensal model. Meanwhile, MSN@DOX-AMP encapsulated DOX and AMP were highly efficient via a combined strategy of physical absorption and exhibited hemocompatibility and biocompatibility (Ma et al., 2023). Strong interactions between HA@PLGA-polymyxin B (PMB) NPs with electrical, particle size, and hydrophilicity properties and mucus resulted in more drugs entering deeper into the lung compared to the free drug PMB *in vitro* (Wu et al., 2022).

#### 4.4.2 *In vivo* studies

The pharmacokinetics of the NP formulation of BDQ were compared with the conventional dry powder inhaler and oral drug solutions *in vivo*. The results did not show a better safety profile of NPs compared to conventional DPI and oral solution, but pharmacokinetic studies indicated higher concentrations of BDQ in lungs via developed formulation (Kim et al., 2019). *In vivo* work revealed that the complex of PG ] displayed a noteworthy sustained release profile and improved the oral absorption of gingerol. The hematological characteristics revealed sustained anti-bactericidal and anti-inflammatory properties in bacterial organisms, causing respiratory infections (Chanda et al., 2010). The conjugated form of azithromycin with PAMAM dendrimer with improved membrane penetration was successful against *P. aeruginosa* biofilm associated with chronic lung infection. *In vivo* results also reduced chronic lung infection and lung inflammation (Gao et al., 2020).

Mannosylated solid lipids in the treatment of tuberculosis have increased the rifampicin uptake in macrophages and drug therapeutic efficacy and lowered drug toxicity in J774 murine macrophage cell lines and rats. Similar to the information reviewed in the liposome section, surfactants, such as mannose and synthetic ones, could enhance internalization (Maretti et al., 2019). Evaluation of the D-enantiomeric dendrimers, dG3KL and dTNS18, was studied in relation to tobramycin (Tob) to treat *P. aeruginosa*. The comparable *in vitro* activity of dG3KL and dTNS18



peptide dendrimers show *in vitro* activity comparable to Tob against both *P. aeruginosa* planktonic and biofilm cells at concentrations not toxic *in vivo* (Pompilio et al., 2018).

Mice (C57BL/6J) were treated with ONP-302-NPs (1 mg) via injection (i.v.) on day 3 after infection. The results indicated that daily particle administration from 3 dpi to either 5 or 7 dpi reduced the accumulation of monocytes within the lungs and ameliorated clinical signs during infection. In addition, ONP-302 reduced the levels of MPO, albumin, IL-6, and TNF- $\alpha$  in the lungs of influenza-infected mice (Kelley et al., 2022). In a mouse model of acute *P. aeruginosa* pneumonia, algae-nanoparticles effectively reduced bacterial burden and substantially reduced animal mortality (Zhang F. et al., 2022)

Taken together, the administration of nanoparticle-based medicine is a promising tool for the treatment of lung infections through overcoming low local antibiotics. Additionally, the increased antibacterial activity of nanoparticle-based medicine against bacteria promises decreased doses, thus reducing their side effects. The effects of nanoparticles in lung infections are mentioned in Table 2. Figure 7 demonstrates the molecular mechanisms involved in the effects of nanoparticles on lung infections.

## 4.5 Lung cancers

The most common type of cancer in the world is lung cancer, and nanomaterials play an important role in cancer therapy (Sanaei et al., 2022). Lung cancer, characterized by the uncontrolled growth

of cells in lung tissues, remains a substantial and disturbing origin of universal disease and death. Nano-therapy and regenerative medicine have continued to develop for lung biology and lung disorders (Sanaei et al., 2022). Several studies have shown the effects of NPs on lung cancer.

### 4.5.1 *In vitro* studies

Some studies have shown the effects of NP drugs on increasing delivery to lung cancer cells and diagnosis of lung cancer. Alginate or chitosan-coated single-walled carbon nanotubes (0, 4, 8, 12, 16, and 20  $\mu\text{g/ml}$  for 24 h) were used to deliver curcumin to the A549 lung cancer cells. The loading efficiency and the sustained drug release are two main advantages of the mentioned nanopatforms.

Studies have shown the promising potential of carbon-based nanoparticles, including fullerene, carbon nanotubes, and graphene-based materials, for drug delivery purposes due to high drug loading and outstanding drug release profiles (Li et al., 2014).

Poly(3-hydroxybutyrate-co-3-hydroxyvalerate acid) (PHBV), a biodegradable and biocompatible bacteria-produced biopolymer, was applied to prepare nanoparticles containing sunitinib, a common drug used to treat lung cancer with low selectivity. This NP is a selective epithelium transport and drug internalization, with low side effects due to controlling the release profile. The MTT assay (0.5, 2, 4, 8  $\mu\text{g/ml}$  NPs after 48 h) has shown comparable cytotoxic effects to sunitinib, which demonstrated the promising future of this drug delivery system (Otroj et al., 2020).

Surface-modified ZnS QDs (0, 1, 10, 50, 100  $\mu\text{g/l}$  for 48 h) with adipic dihydrazide-heparin were used to deliver paclitaxel to the A549 cancer cells. *In vivo* and *in vitro* studies have shown a more

controllable drug release profile triggered by acidic conditions, which could enhance drug efficiency and cytotoxicity in cancer treatment (Xie et al., 2020).

In another study, dihydropyrimidinones were formed by conjugation with carbon QDs with various ratios (25, 50, 100, 200, 300, 400, and 500  $\mu\text{M}$ , 24 h). The lower toxicity against healthy cells, fluorescence trackability by bioimaging, and improved tumor-growth inhibition were obvious from the results. QDs could give the nanocomposites multifunctionalities of drug monitoring, trackability, delivery, and medical imaging altogether to the nanocarrier (S et al., 2021). However, sometimes the cytotoxicity of QDs in some cases is an issue that needs special attention (Zhang et al., 2020c). In general, the tuneability, modifiability, stability, and biocompatibility of carbon QDs and inexpensiveness are the main advantages of carbon QDs (Guo et al., 2018). The application of QDs to medical imaging can also facilitate decisions on diagnosing and staging of cancer and consequently choosing the right medication (Jin et al., 2011). Using non-small cell lung cancer biomarkers conjugated with QDs, one could also detect lung cancer metastasis in blood or lymph nodes (Wang et al., 2012).

Magnetic nanoparticles and superparamagnetic iron oxide nanoparticles (SPIONs) are the most usually employed metal oxides in therapy, diagnosis, and separation (Cedrowska et al., 2020). It has been shown that these properties may result from the increased antioxidant effects of these particles. The centerpiece for mag-force cancer therapy is also iron oxide nanoparticles (Gao et al., 2017). The small size of the particle is critical for therapy. Hence, its high density has therapeutic effects when it is injected directly into tumor cells. An external field will cause absorption of the magnetic nanoparticles by cancer cells, after which further oscillation causes drug release, hyperthermia, or cell burst (Alphandéry, 2020).

In lung cancer, metastasis could be detected with sensitive SPIONs by magnetic resonance imaging with no side effects (Wan et al., 2016).

A dendrimer-based nano-drug delivery system was designed to be capable of increasing the penetration of DOX as measured in an *in vitro* 3D lung tumor model and associating those consequences and its efficiency. More specificity for DOX to tumor cells compared to fibroblasts due to conjugation, maintenance activity of the released DOX that reduces tumor similar to free DOX was observed (Almuqbil et al., 2020). The ZnO QD-based pH-responsive drug delivery pattern (0.1, 1, 5, 10, 25, and 50  $\mu\text{g/ml}$ , 48 h) for intracellular controlled release of drugs was designed. Synergistic therapy was observed due to the incorporation of the antitumor results of  $\text{Zn}^{2+}$  and DOX (Cai et al., 2016).

Several other studies indicate the therapeutic effects of NPs on lung cancer. The cytotoxicity of the alginate-coated nanotube against A549 cells was also more efficient than that of the free curcumin (Singh N. et al., 2018). A similar docetaxel-loaded chitosan-coated NPs (40  $\mu\text{g/ml}$  for 1, 2, and 4 h.) led to higher drug bioavailability compared to the free drug, which has demonstrated the growth inhibition of 66% against A549 tumor cells. (Li B. et al., 2018).

Chitosan-coated graphene oxide functionalized with the hyaluronic acid (5, 10, 20, 40, 80, 160, and 320  $\mu\text{g/ml}$  for 24 h) as the targeting agent led to tolerable systemic side effects and

higher internalization and toxicity against A549 cancer cells compared to normal cells (Liu et al., 2018). The nanocomposite of NiO/CuO nitrogen-doped graphene oxide (5, 10, 50, 100, 150, and 200  $\mu\text{g/ml}$ ) has also illustrated the potential to kill A549 cancer cells (Anbu et al., 2021). This study also suggests that A549 cell killing is mediated through ROS generation induced by NiO/CuO NC.

Graphene oxide dots have recently been used as a photosensitizer in photodynamic therapy for lung cancer. It appears that dots were internalized and induced ROS, leading to apoptosis and necrosis (Shih et al., 2021). Fullerenes, another allotrope, are nanocages of carbon that take different hollow shapes from spherical to cylindrical and are very effective radical scavengers. Despite these attractive properties, carbon nanomaterials are toxic and show other problems, such as agglomeration, difficult administration routes, and particle size (Nalepa et al., 2020).

PAMAM was used to transport multiple chemotherapeutics, including cisplatin and small interfering RNAs (siRNA). Folic acid was conjugated for favorable targeting purposes against H1299 lung cancer cells. The target cells expressed folate receptor alpha. The siRNA was deposited electrostatically, and cisplatin was attached via the coordination of the amine group. It appears that the drug damages the DNA and induces apoptosis of cancer cells, which increased synergistically compared to the drugs used individually, and the toxicity also decreased due to targeting using folic acid (Amreddy et al., 2018).

Antisense oligonucleotide (2'-O-methyl-RNA)-loaded chitosan nanoparticles (0.1–2.5  $\text{mg/ml}$  for 6 h) were used to treat A549 cancer cells via telomerase inhibition, which decreased the telomerase activity significantly (50%) and was potentially promising for the treatment of lung cancer (Nafee et al., 2012).

Recently, folate-decorated Ag–In–Zn–S QDs (50  $\mu\text{g ml}^{-1}$ , 24 h) were applied for targeting drug delivery of doxorubicin to A549 cancer cells. The *in vitro* analyses demonstrated that the nanoconjugate was more cytotoxic and genotoxic than the free drug. It appears that the prepared conjugate successfully inhibited the migration of cancer cells (Ruzicka-Ayoush et al., 2021).

The thiolate–Zn interaction was also used to attach a histone deacetylase inhibitor, *N*-hydroxy-*p*-(4-arylbutanamido) benzamides, to the CdSe/ZnS QDs, which formed a water-miscible nanocarrier (20 nM, 48 h) that was employed to treat A549 and H1299 cancer cells. Investigation of the apoptosis cycle showed the breakdown of the cell cycle at the G2-to-M transition while increasing the acetylation of p53 and tubulin, killing cancer cells, or inhibiting tumor growth synergistically and more efficiently than the free active agent (Chen et al., 2021).

Carbon QDs conjugated with  $\text{Bi}_4\text{O}_5\text{Br}_2$  were successfully implemented for tumor photodynamic therapy against A549 cancer cells (0, 0.5, 5, 10, 25, 50, 100, and 200  $\mu\text{g/ml}$ , 24 h). Compared to  $\text{Bi}_4\text{O}_5\text{Br}_2$  nanocrystals, QD nanocomposites with  $\text{Bi}_4\text{O}_5\text{Br}_2$  were water-miscible, improved the uptake of ROS generation, especially  $\text{O}_2^-$  and  $\cdot\text{OH}$  species, and were more effective in killing lung cancer cells. Therefore, it can be used to enhance the efficiency of photocatalytic properties, biocompatibility issues, and internalization of the inorganic active materials (He et al., 2021). Carbon QDs in drug delivery systems can increase the

therapeutic potency of drugs in the treatment of different cancers (Li et al., 2020).

Gold nanoparticles are possible candidates for gene therapy, drug delivery, photothermal therapy, and photodynamic therapy, especially in lung cancer and viral diseases (Mousavi-Kouhi et al., 2021b). In lung cancer, bombesin/GRP receptors were overexpressed; therefore, synthesizing a library of them and their conjugated forms of AuNPs would show high biocompatibility (Chanda et al., 2010). Research has shown that gold nanoparticles bind to drugs by binding to various functional groups, such as carboxyl groups. AuNPs (0.16, 0.31, 0.63, 1.25, 2.50, and 5.00 mg/ml; 24, 48, and 72 h) can enhance ROS production, sensitize mitochondrial membrane potential, and stimulate both primary and late apoptosis in lung cancer cells (Mousavi-Kouhi et al., 2021b).

The conjugated form of Toll-like receptor 4 (TLR4) with AuNPs was used as an adjunct therapy for human adenocarcinoma alveolar basal epithelial cells (A549 cells), which led to the downregulation of the expression of this receptor (Vyas and Goswami, 2019). AuNPs (20, 40, 80, and 100 µg/ml for 24 h) were found to be most effective in preventing ROS generation by TLR4 in A549 cells. RNI produced by unrestrained TLR4 signaling is recognized for the development of lung carcinogenesis, and AuNPs significantly decreased RNI production (Vyas and Goswami, 2019).

It has been reported that the cisplatin-dendrimer nanocomplex has toxic behavior against NCI-H460 lung cancer cells (Nguyen et al., 2015). Assessment of the safety profile and remedial efficiency of the anticancer factor derived from [188Re] rhenium-ligand as radioactive ligand-loaded 5th production poly-L-lysine dendrimer was performed. The results showed a safe and well-tolerated 188-Rhenium-Imdendrim brachytherapy device for lung cancers (Belhadj-Tahar et al., 2019).

Significant improvement of cellular uptake along with gene transfection efficacy of PAM-Ap/pMiR-34a NPs in cultured NSCLC cells due to aptamer conjugation was indicated (Wang H. et al., 2015). Phenethyl isothiocyanate (PEITC) and cisplatin (CDDP) co-encapsulated in a liposomal NP for the treatment of non-small cell lung cancer were previously studied. The results displayed an enhancement of NSCLC cell toxicity due to co-encapsulation of PEITC and CDDP in liposomes. Liposomal-PEITC-CDDP was more toxic against both A549 and H596 human NSCLC cell lines than against WI-38 and BEAS-2B human normal lung cell lines (Sun et al., 2019).

Gefitinib (GEB) encapsulation upon the nanoliposomes (10, 20, 30, 40, and 50 µg/ml, 24 h) for increasing the curative effect of lung cancer was designed. Improved stability, reduced particle size, and higher encapsulation efficacy in nanoliposome compound drug (GL) were observed. In addition, *in vitro*, GL had a promising drug sustained-release result. GL significantly increased the proapoptotic result on A549 cells. The ability of GET to inhibit the proliferation, migration, and invasion of tumor cells has been significantly enhanced after nanoliposome embedding modification (Hu et al., 2020).

Co-administration of curcumin and bromocriptine nanoliposomes (12–20 µM; 12, 24, and 48 h) to human lung cancer cells (QU-DB) *in vitro* showed anticancer effects by affecting apoptosis and cell viability in QU-DB cells but did not show cell

toxicity properties in normal cells (HFLF-PI5) (Sheikhpour et al., 2020). Electroporation preparation of paclitaxel-in-liposome-in-bacteria (LPB) (0.62, 1.25, 2.50, and 5 µmol/L, 24 h) was performed for inhalation treatment of primary lung cancer. LPB prevented the proliferation of A549 lung cancer cells. LPE displayed the maximum anticancer effect on rat primary lung cancer with the downregulation of VEGF and HIF-1α and the improvement of cancer cell apoptosis after intratracheal administration. Bacterial formulations significantly increased the expression of immune markers (TNF-α, IL-4, and IFN-γ) and immune cells (leukocytes and neutrophils) (Zhang et al., 2020b).

The cell toxicity of InP/ZnS QDs (0.62, 1.25, 2.5, 5, 10, 20, 40, 80, and 160 µg/ml, 24, 48 h) with diverse surface clusters (NH<sub>2</sub>, COOH, and OH) against two lung-derived cell lines was reported. All of the InP/ZnS QDs were able to enter the cells, with greater uptake efficacy for InP/ZnS-COOH and InP/ZnS-NH<sub>2</sub> at lower doses. High doses of InP/ZnS QDs initiated the reduction of cell viability, and InP/ZnS-COOH QDs and InP/ZnS-NH<sub>2</sub> QDs seemed more toxic than InP/ZnS-OH QDs. In addition, InP/ZnS QDs developed cell apoptosis and intracellular ROS production after co-culture with cells (Chen et al., 2018).

The green fabrication of CdS QDs using tea leaf extract resulted in the inhibition of bacterial growth and displayed cell toxicity against A549 cancer cells due to CdS QDs. Production of high-contrast fluorescence images of A549 cancer cells due to CdS QDs was another result of this study. CdS QDs arrest the A549 cell growth at the S phase of the cell cycle, preventing more growth of lung cancer cells. CdS-QDs could interact with the P-part in DNA. Then, DNA-replication is inactivated, leading to the inhibition of enzyme functions, which results in loss of cell viability and undergoes cell death through apoptosis (Shivaji et al., 2018).

Black phosphorus quantum dots (BP-QDs, 0, 5, 10, and 20 µg/ml for 24 h) on lung cells (A549 and Beas-2B) induced cell toxicity and reduced oxidative stress and cell cycle arrest in two types of human lung epithelial cells. The prevention of cell uptake and restoration of the cell toxicity of BP-QDs were also shown. Oxidative stress also contributes to cell cycle arrest and cell damage caused by BP-QDs. The results showed that BP-QDs induced LDH (an indicator of cell membrane integrity) leakage from A549 and Beas-2B cells in a concentration-dependent manner (Ruan et al., 2021). Exposing A549 cells to 70 and 420 nm ZnO particles (0, 8, 10, 12, 14, 16, and 18 µg/ml, 24 h) showed dose- and time-dependent cytotoxicity due to exposure of both sizes of NPs. Neither free Zn<sup>2+</sup> nor metal impurities in ZnO NP samples are the cause of cytotoxicity. The cytotoxicity of ZnO is reflected in oxidative stress, lipid peroxidation, cell membrane damage, and oxidative DNA damage (Lin et al., 2009).

Metal and metal oxide nanoparticles (ZnO, CeO, and Ag) (8–128 µg/ml; 1, 6, 24 h) in lung epithelial A549 cells caused metabolic and transcriptional responses, and the great majority of these molecular alterations were usual to both ionic and NP exposures and a characteristic of metal ion exposure. The low toxicity of CeO<sub>2</sub> NPs elicited few molecular changes, displaying few signs of oxidative stress for only one of the four CeO<sub>2</sub> NPs examined (Dekkers et al., 2018). Green AgNP (10–250 µg/ml; 24 h) fabrication using *Cymodocea serrulata* as a potential marine biosource displayed potential cellular toxicity in human lung cancer A549 cells (Palaniappan et al., 2015).



Biosynthesis of gold nanoparticles (AuNPs, 20–100 µg/ml, 24 h) from *Magnolia officinalis* and anticancer activity in A549 lung cancer cells efficiently induced cellular toxicity and apoptosis indicated by apoptotic gene expression. Synthesized AuNPs provoked ROS production in A549 cells, and elevated ROS directed to the fragmentation of the nucleus and depolarization of mitochondria, resulting in oxidative stress inter-ceded apoptosis. AuNPs diminished the expression of Bcl-2 and Bid and elevated the expression of Bax, Beclin-1, and caspase-3 in A549 cells (Zheng et al., 2019).

The ability of nanoconjugates of CdSe/CdS/ZnS-QDs and Dox (330 µl, 24 h) to target alveolar macrophage cells was studied. The release of Dox from the QD-Dox nanoconjugate induced apoptosis. Inflammatory injury parameters, including albumin leakage, proinflammatory cytokines, and neutrophil infiltration, were documented after the administration of QD-Dox (Chakravarthy et al., 2011). In the presence of laser irradiation, GA-AuNPs showed considerable cytotoxicity against A549 cells. Lung tumor treatment with GA-AuNPs followed by laser exposure enhanced the apoptotic pathway. Upregulation of caspase-3, a significant reduction in the levels of the inflammatory mediator TNF-α, and angiogenesis inducer VEGF were also observed. Induction of lipid peroxidation was also reported upon treatment with GA-AuNPs (Gamal-Eldeen et al., 2017). It was indicated that QDs conjugated to desmethyl erlotinib (QD-OSI 420) showed meaningfully better efficiency than pure drugs in non-small cell lung cancer (NSCLC) cell lines, and QD-OSI 420 conjugated significantly increased cell viability compared to pure drugs with an IC<sub>60</sub> of 2.5 µM in erlotinib-resistant A549 cell lines. In addition, in the 3D-SCC model of A549, QD-OSI 420 significantly reduced 3D tumor volume compared to pure drugs *in vitro*. Therefore, these results indicate that conjugating QD-OSI-420 could be considered an alternative to traditional anticancer therapy by improving intracellular drug delivery (Kulkarni et al., 2019).

#### 4.5.2 *In vivo* studies

Paclitaxel-loaded solid lipids (1.0, 10, and 100 µg/ml, 1 h) were used against metastatic mammary tumors in the lungs *in vivo* and *in vitro*. Results revealed drug accumulation in the tumor microenvironment, intracellular targeting, cellular toxicity, increased therapeutic delivery into the cytosol, and prevention of further tumor progression (Videira et al., 2012). In fact, PTX-loaded liposomes did not attain more efficiency but rather induced severe neurotoxicity, proposing the existence of a free drug. Remedial efficacy of a new cationic liposome-nano-formulated all-trans-retinoic acid (0.60 mg/kg/day for 30 days) in the mice model of lung cancer revealed that the formulation of lipo-all-trans-retinoic acid in 1, 2-dioleoyl-3-trimethylammonium-propane (DOTAP) and cholesterol in the ratio of 70:20 was the suitable carrier for ATRA in treating lung cancer (Grace and Viswanathan, 2017). As DOTAP is a cationic lipid, the interaction with a negatively charged cell surface may facilitate an effective fusion and drug delivery in the target site. The liposome-formulated (“antineoplastic” or “cytotoxic”) chemotherapy drug enhances the stability of the ATRA and delivers the drug to the target site effectively. The use of natural polysaccharides is also common to make nanoemulsions to treat different types of cancers. For example, *Brucea javanica* oil with chitosan formed a cationic nanoemulsion (505 mg kg<sup>-1</sup>, 28 days)

that showed high colloidal stability and a higher oral bioavailability (1.6-fold more). The nanoemulsion enhanced the suppression of lung cancer compared to free oil in *in vivo* models (Liu et al., 2016). VEGF, IL-8, and Bcl-2 were downregulated, and P53 was upregulated.

#### 4.5.3 Clinical studies

The efficacy of IFN-γ nanoliposome on peripheral lymphocytes from lung cancer patients was investigated compared to healthy individuals (20 individuals in each group). The lymphocytes from lung cancer patients presented with upper DNA destruction ranked higher than those of healthy individuals. In healthy individuals, IFN-γ liposome did not stimulate any DNA destruction in the lymphocytes. It also decreased DNA destruction in the lymphocytes of lung cancer patients. IFN-γ liposome meaningfully decreased oxidative stress (Alhawmdeh et al., 2021).

In a clinical study, SLIT liposomal cisplatin was administered via inhalation for patients with lung carcinoma. The results indicated the absence of dose-limiting toxicity at the maximum delivered dose, including no hematologic, kidney, ophthalmic, or nervous toxicities (Wittgen et al., 2007).

Overall, various NPs were developed for selective drug delivery to lung cancer and lung metastases based on understanding the biology of the tumor, the microenvironment, and the interaction between malignant cells and nanoparticles. The results showed the NP-based drug as an alternative to traditional anticancer therapy, which is more effective than ordinary drugs by improving intracellular drug delivery. The effects of nanoparticle treatment in lung cancers are documented in Table 3. Figure 8 shows the molecular mechanisms involved in the effects of nanoparticles on lung cancer.

## 5 Discussion

This review article described the impact of different nanoparticles on different respiratory disorders. Nanoparticles showed therapeutic effects on respiratory disorders through various mechanisms such as anti-inflammatory, antioxidant, immunomodulatory, cell viability, proliferation, apoptosis, and necrosis mechanisms. Figure 9 summarizes the effects of nanoparticles on respiratory disorders.

### 5.1 Possible mechanisms of the effects of NPs on lung disorders

#### 5.1.1 Anti-inflammatory, antioxidant, and immunomodulatory effects

The size and structural properties of nanoparticles are among the most important components that can affect lung diseases. Various experiments on the effects of NPs on lung disorders, such as asthma, COPD, and other lung diseases, indicated that nanoparticles suppressed proinflammatory mediators, such as IL-6, IL-8, IL-1β, and TNF-α. In addition, the deactivation of CD4 and CD8 T cells in lung tissue and the reduction of their capability to generate proinflammatory cytokines was reported *in vivo* for lung disorder therapy using NPs (Das et al., 2022).

**TABLE 3 Effects of various nanoparticles on lung cancers.**

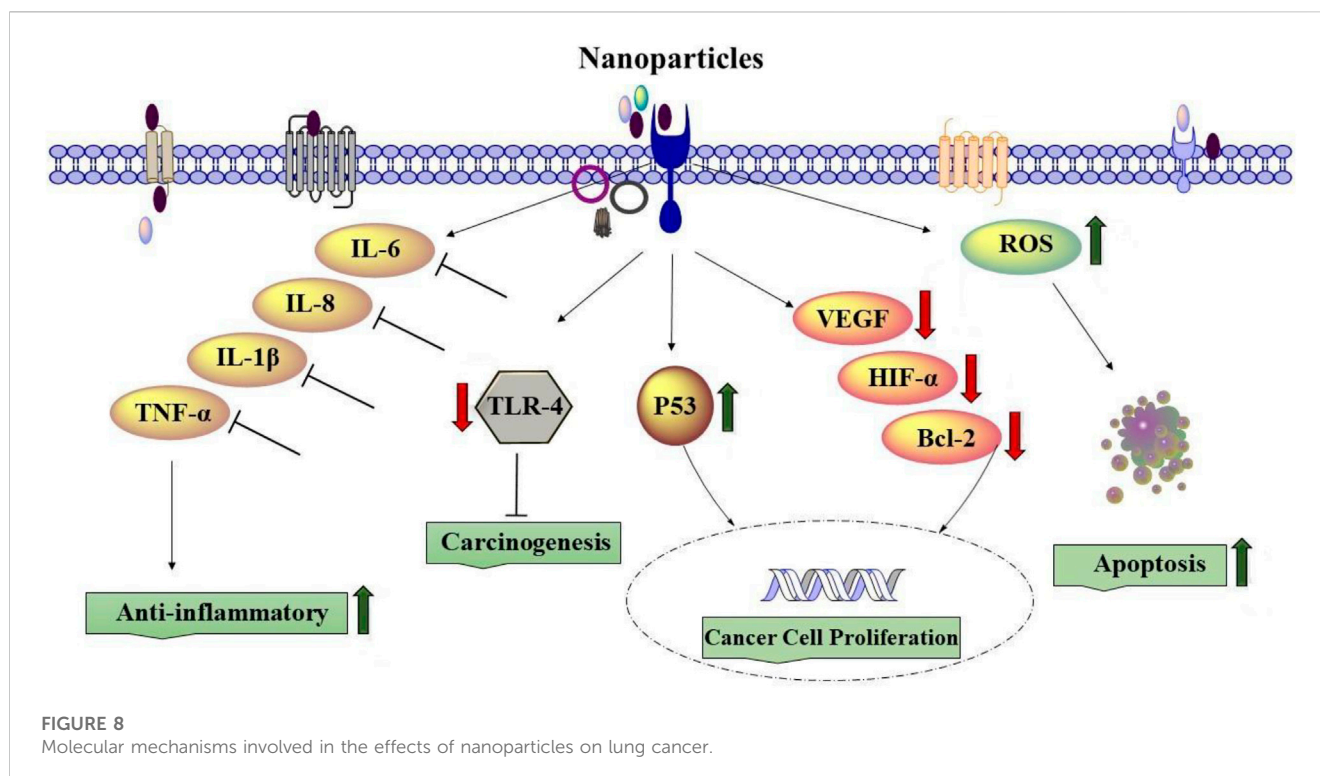
Study	Study design	Dose and time	Finding	Ref.
<i>In vitro</i>	MDM2-exposed rats	0–100 µg/ml for 24 h	Downregulated MDM2 gene, activation of Cas-8 and PARP cleavage, ↑ cell apoptosis, inhibited tumor growth	Huang et al. (2016a)
	CS-coated CNT for delivery and Cur of A549 cells	0, 4, 8, 12, 16, 20 µg/ml for 24 h	Cytotoxicity of alginate-coated nanotube against A549 cells was also more efficient than the free curcumin	Singh et al. (2018a)
	DTX-loaded CS-coated	40 µg/ml for 1, 2, and 4 h	↑ Drug bioavailability	Li et al. (2018a)
			Inhibited A549 tumor cells	
	CS-coated GO against A549 cancer cell	5, 10, 20, 40, 80, 160, and 320 µg/ml, 24 h	↓ Systemic side effects	Liu et al. (2018)
			↑ Internalization and toxicity s	
	NiO/CuO nanocomposite nitrogen-doped GO	5, 10, 50, 100, 150, and 200 µg/ml	↑ ROS generation by NiO/CuO	Anbu et al. (2021)
			↑ Killing of A549 cancer cells	
	ASO (2'-O-methyl-RNA)-loaded CS NPs	0.1–2.5 mg/ml for 6 h	↓ Telomerase activity significantly in A549	Nafee et al. (2012)
	PHBV nanoparticles containing sunitinib	0.5, 2, 4, and 8 µg/ml NPs after 48 h	↓ Side effects	Otroj et al. (2020)
			Controlling the drug release profile	
			Comparable cytotoxic effects of sunitinib	
	Ag–In–Zn–S QDs fin A549 cancer cells	50 µg ml <sup>-1</sup> , 24 h	↑ Cytotoxic and genotoxic than free drug	Ruzycka-Ayoush et al. (2021)
			↓ Migration of cancer cells	
	CdSe/ZnS QD nanocarrier to treat A549 and H1299 cancer cells	20 nM, 48 h	Breakdown of the cell cycle at G2-to-M transition	Chen et al. (2021)
↑ Acetylation of p53 and tubulin				
Killing cancer cells or inhibiting the tumor-growth				
ZnS QDs + ADH in A549 cancer cells	0, 1, 10, 50, and 100 µg/ml for 48 h	↑ Drug efficiency and cytotoxicity in cancer treatment	Xie et al. (2020)	
Encapsulated ZnO-QDs to poly(DL-lactide-co-glycolide)	7.5 µg/ml for 24 h	Selectively cytotoxic against metastatic A549 cells	Kim et al. (2020)	
DHMPs formed a conjugate with CQDs	25, 50, 100, 200, 300, 400, and 500 µM, 24 h	Inhibited tumor growth	Thangamani et al. (2021)	
		Improved rug monitoring, trackability, delivery		
Conjugated Bi <sub>4</sub> O <sub>5</sub> Br <sub>2</sub> + C <sub>2</sub> QDs f against A549 cells	0,0.5, 5, 10, 25, 50, 100, and 200 µg/ml, 24 h	↑ Uptake and reactive oxygen generation in cancer cells leads to killing them	He et al. (2021)	
AuNP treatment in lung cancer cells	0.16, 0.31, 0.63, 1.25, 2.50, and 5.00 mg/ml; 24, 48, 72 h	↑ ROS production	Mousavi-Kouhi et al. (2021b)	
		Improved mitochondrial membrane potential		
		Apoptosis in lung cancer cells		
Toll-like receptor 4 conjugated + AuNPs for A549 lung cancer cells	20, 40, 80, and 100 µg/ml for 24 h	Downregulation of TLR4 expression	Vyas and Goswami (2019)	
		↓ RNI production by unrestrained TLR4 signaling		
GEB encapsulation nano-liposomes for the treatment of lung cancer cells	10, 20, 30, 40, and 50 µg/ml, 24 h	Improved GL stability, encapsulation efficacy	Hu et al. (2020)	
		↓ Particle size, proliferation, migration, and invasion		
		↑ Proapoptotic result on A549 cells		
	100 µg/ml for 48 h		Nguyen et al. (2015)	

(Continued on following page)

**TABLE 3 (Continued) Effects of various nanoparticles on lung cancers.**

Study	Study design	Dose and time	Finding	Ref.
	Cisplatin-stimulated NCI-H460 lung cancer cell		Toxic effect against NCI-H460 lung cancer cell	
	Nano-DDS + ~5.5 DOX	0.1 μM–20 μM	More specificity for DOX to tumor cells Maintenance activity of the released DOX	Almuqbil et al. (2020)
	PAM-Ap/pMiR-34a NP- stimulated NSCLC	-	↑ Cellular uptake, gene transfection efficiency of NPs in cultured NSCLC cells	Wang et al. (2015a)
	LN-encapsulated PEITC and CDDP-stimulated NSCLC	-	↑ Cell toxicity to A549 and H596 human NSCLC cell lines than WI-38 and BEAS-2B HNCL	Sun et al. (2019)
	DOTAP carrier for ATRA- exposed mice	0.60 mg/kg/day for 30 days	Suitable carrier for ATRA in treating lung cancer	Grace and Viswanathan (2017)
	Cur + BR nano-liposomes	BR: 12.5–25 μM Cur: 12–20 μM	Anticancer effects on lung cancer cells No cytotoxicity effects in HFLF-PI5	Sheikhpour et al. (2020)
	Paclitaxel in the liposome model of lung cancer	24 h after 1 ml of LPB	↓ Proliferation of A549 cells, VEGF, and HIF-1α ↑ Effect on the rat primary lung cancer ↑ Expression of TNF-α, IL-4, and IFN-γ	Zhang et al. (2020b)
	IFN-γ liposome peripheral lymphocyte	30 min incubation 100 U/ml	Induce DNA damage in the lymphocytes ↓ DNA damage in lung cancer lymphocytes ↓ Oxidative stress caused by H <sub>2</sub> O <sub>2</sub>	Alhawmdeh et al. (2021)
	ZnO- QD model of lung cancer	0, 0.1, 1, 5, 10, 25, 50 μg/mL, 48 h	Synergistic therapy due to the incorporation of the antitumor effect of Zn <sup>2+</sup> and DOX	Cai et al. (2016)
	InP/ZnS QD-stimulated two lung-derived cell lines	0, 0.62, 1.25, 2.50, 5, 10, 20, 40, 80, and 160 μg/ml, 24 h	↑ Uptake of InP/ZnS-COOH and InP/ZnS-NH <sub>2</sub> ↓ Cell viability ↑ Toxic effect, apoptosis, intracellular ROS	Chen et al. (2018)
	CdS-QD-stimulated A549 cancer cells	0, 10, 20, 30, 40, 50, and 60 μg/ml, 24 h	↓ A549 cell growth at the S cell cycle phase ↑ Cytotoxicity on A549 cancer cells ↑ High-contrast fluorescence A549 cells images	Shivaji et al. (2018)
	BP-QD-stimulated A549 and Beas-2B lung cell	5~20 μg/ml for 24 h	↓ Cytotoxicity, oxidative stress, cell cycle arrest ↓ Cellular uptake restores the cytotoxicity of BP-QDs	Ruan et al. (2021)
	ZnO NP-stimulated A549 lung cells	0,8, 10, 18, and 25 μg/ml in 24 h	Dose- and time-dependent cytotoxicity	Lin et al. (2009)
	ZnO, CeO, and Ag- stimulated A549 cells	8, 15, 10, and 30 μg/ml in 24 h	↑ Metabolic and transcriptional responses Evidence of oxidative stress	Dekkers et al. (2018)
	Green AgNPs synthesis using <i>Cymodocea serrulata</i>	10–250 μg/ml after 24 h	Potential cytotoxicity against human lung cancer A549 cells	Palaniappan et al. (2015)
	AuNP-stimulated A549 lung cancer cells	15 and 20 μg/ml after 24 h	↑ Cytotoxicity and apoptosis by inflecting apoptotic gene expressions in A549 cells	Zheng et al. (2019)
<b>In vivo</b>	CdSe/CdS/ZnS QDs and Dox-stimulated alveolar macrophages	10 nM QD-Dox after 24 h	↑ Cell apoptosis Improved inflammatory cytokines markers	Chakravarthy et al. (2011)

MDM2, mouse double minute-2; CS, chitosan; GO, graphene oxide; CNT, carbon nanotube; Cur, curcumin; DTX, docetaxel; ASO, antisense oligonucleotide; DHMPs, dihydropyrimidinones; CQDs, carbon quantum dots; ADH, adipic dihydrazide-heparin; DOTAP, 1, 2- dioleoyl-3-trimethylammonium-propane; Cas-8, caspase-8; PARP, poly(ADP-ribose) polymerase; miR-34a, microRNA-34a, into PAM-Ap/pmiR-34a NPs, S6 aptamer-conjugated dendrimer nanoparticles; PHBV, poly(3-hydroxybutyrate-co-3-hydroxyvalerate acid); AuNPs, gold nanoparticles; AgNPs, silver nanoparticles; PEITC, phenethyl isothiocyanate; CDDP, cisplatin; GEB, gefitinib; GL, nanoliposome compound drug; LN, liposomal nanoparticle; LPB, liposome-in-bacteria; Cur, curcumin; Dur, duration; BR, bromocriptine; DOXeq, DOX-equivalent; HNCL, human normal lung cell line.



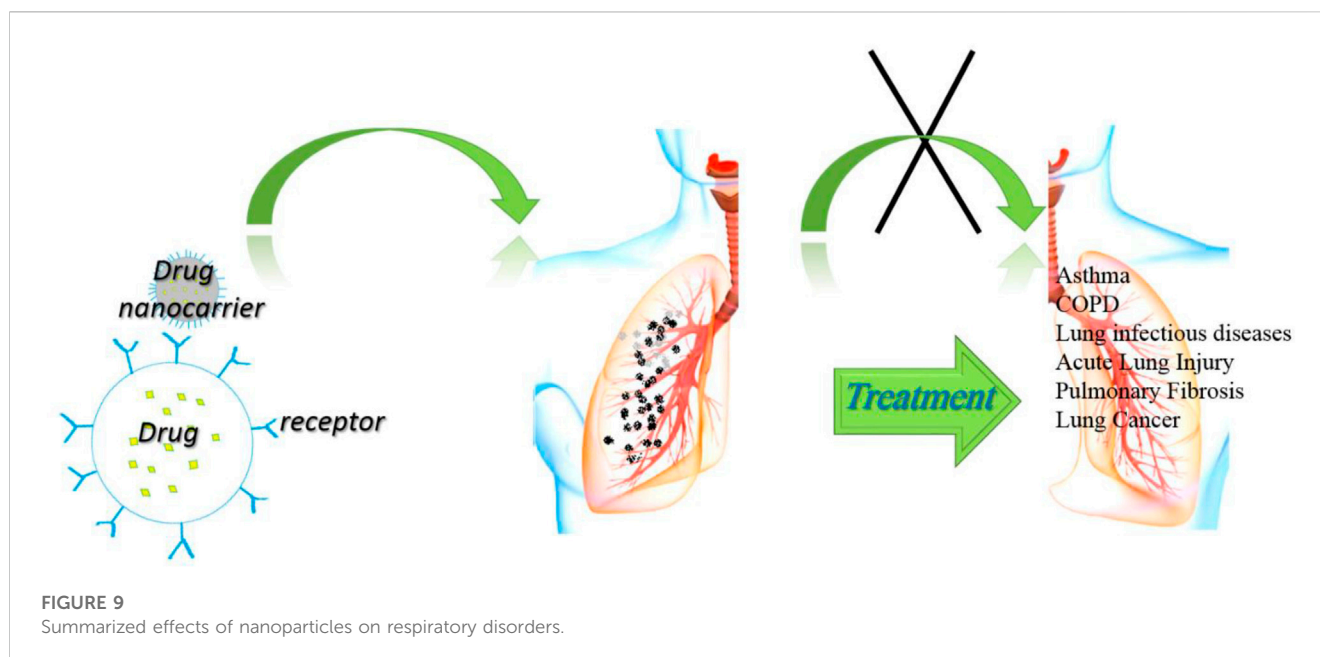
Lung cancer treatment with NPs could be associated with induced ROS, leading to apoptosis and necrosis (Nasrin et al., 2022). Nanoparticles can also downregulate the expression of Toll-like receptor 4, which is known as carcinogenesis in lung cancer cells and affects lung cancer treatment (Wang et al., 2022). In addition, it was indicated that treatment of lung cancer cells with NPs downregulated VEGF, HIF-1 $\alpha$ , IL-8, and Bcl-2 genes but increased the gene expression of p53 (Huang et al., 2016b). Chlorin e6 loaded by iron oxide NPs linked with glucose significantly enhanced the uptake of Ce6 by lung cancer cells and produced more ROS, damaged the DNA of lung cancer cells, and thereby activated STING and upregulated the expression of IFN- $\beta$ , HMGB1, and HSP90, indicating augmented immunogenicity of lung cancer cells (Yu et al., 2022). Silver nanoparticles (AgNP-DT $\alpha$ ) prepared using an aqueous seed extract of *Derris trifoliata* showed antioxidant properties and anti-proliferative activities against A549 lung cancer cell lines (Cyril et al., 2019).

### 5.1.2 Effect on cell viability and proliferation

The reduction effects of Rutin-loaded liquid crystalline nanoparticles (Rutin-LCNs) on NSCLC proliferation and migration were shown (Paudel et al., 2021), which is suggested to result from its inhibitory effect on the MMP-9 expression and prevent the degradation of the extracellular matrix (ECM) component. The combination of cell growth inhibition, cell cycle arrest, suppression of oncoproteins, and impeded cell migration due to HuR-FNP was also reported (Muralidharan et al., 2016). Biosynthesized zinc oxide NPs using the neem tree (100, 150, 200, and 250  $\mu\text{g/ml}$  for 48 h) also significantly reduced cell (A549) viability (Rani et al., 2022). Antiproliferative and cytotoxic effect potentials of chitin and chitosan encapsulated gold nanoparticles (AuNPs) against human lung cancer cell line A549 were demonstrated (Panneer et al., 2022). The green synthesis of AgNPs

using *Streptomyces hirsutus* also decreased the cell viability of A549 via ROS production (Pallavi et al., 2022). The cold atmospheric plasma (CAP) technique combined with iron oxide-based magnetic nanoparticles downregulated epidermal growth factor receptor (EGFR) and inhibited lung cancer cells by depressing pERK and pAKT (Li et al., 2019). Silver nanoparticles (AgNP-DT $\alpha$ ) prepared using an aqueous seed extract of *D. trifoliata* showed antioxidant and anti-proliferative activities against A549 lung cancer cell lines (Cyril et al., 2019). Melatonin (MLT) and palladium nanoparticles (PdNPs) induced cytotoxicity through leakage of lactate dehydrogenase, increased intracellular protease, and reduced membrane integrity (Gurunathan et al., 2020).

Combining PdNPs with MLT elevated the levels of mitochondrial dysfunction by decreasing mitochondrial membrane potential (MMP), ATP content, mitochondrial number, and expression of the main regulators of mitochondrial biogenesis (Gurunathan et al., 2020). The biosynthesized AgNPs using the bark extract of *Toxicodendron vernicifluum* Tv-AgNPs selectively targeted and damaged the cancer cells through ROS generation in human lung carcinoma A549 cells (Saravanakumar et al., 2019). The effects of zinc oxide nanoparticles synthesized from *Euphorbia fischeriana* root (EF-ZnONPs) revealed cytotoxicity activity, inhibited the migration capability of A549 cells, diminished expression of Bcl-2, and enhanced expressions of Bid and Bax (Zhang H. et al., 2020). Cetuximab and doxorubicin-loaded dextran-coated Fe<sub>3</sub>O<sub>4</sub> magnetic NPs suppressed cell proliferation of A549 (Zhang et al., 2019). TiO<sub>2</sub> NPs also inhibited A549 cell proliferation and induced DNA damage and apoptosis via the activation of the intrinsic mitochondrial pathway (Wang Y. et al., 2015). Betulinic acid NPs on lung cancer cells (HKULC2, H1299, and H23) inhibited proliferation and metastatic ability and arrested the cell cycle through the downregulation of ATP-binding cassette and



transporter G1 (ABCG1) oncogene expression (Zhao et al., 2020). Internalized fibroblast growth factor (FGF-2)-loaded NPs increased nuclear ERK1/2 content and resulted in lung cancer cell death (Miao et al., 2020). Amodiaquine (AQ)-loaded NPs (AQ NP) inhibited cell migration, reduced colony growth in A549 cells, inhibited AQ autophagy (increased LC3B-II levels), and improved apoptosis induction (caspase-3 levels) (Parvathaneni et al., 2020). Furthermore, treatment with SeNP-apigenin increased ROS production and oxidative stress in MCF-7 cells (Al-Otaibi et al., 2022). Selenium (Se) NPs also induced ROS-mediated necroptosis in PC-3 cells through TNF activation (Sonkusre and Cameotra, 2017). Generally, AgNP cytotoxicity depends on the dose, time, temperature, surface coatings, size, and cell type (Mousavi-Kouhi et al., 2021a). AgNP exposure reduced antioxidant enzymes, such as glutathione (GSH), elevated intracellular ROS levels and expression of ROS-responsive genes, and lipid peroxidation, leading to DNA damage, necrosis, and apoptosis (Tran and Le, 2013).

### 5.1.3 Effect on apoptosis and necrosis

Tumor cells are characterized by their ability to evade apoptosis, and apoptosis could be used as an indicator for targeted cell treatment in cancer therapy (Akbarzadeh et al., 2022). Cationic poly-l-lysine-assisted magnetic iron oxide nanoparticles (PLL-MNP) in human lung cancer cells resulted in nuclear fragmentation and chromatin condensation (Wang X. et al., 2015). The selenium nanoparticles green-synthesized by apigenin (SeNP-apigenin) treatment in MCF-7 cells induced apoptosis, demonstrating that SeNP-apigenin could directly target Bcl-2, Bax, and caspase-3 and result in the discharge of cytochrome C from mitochondria into the cytosol, accompanied by the initiation of cell death, leading to permanent DNA damage and MCF-7 cell killing. Necrosis as an essential mechanism of cell death was induced by gold NPs in lung cancer cells with a low level of intracellular GSH (Liu et al., 2013). Another mechanism involved in necrosis induced by NPs is the induction of endothelial cell (EC) dysfunction through

the release of VWF (von Willebrand factor). Dysfunction of EC induced by NPs can also result from the formation of ROS, inflammatory cytokines including IL-6 and IL-8, and/or the activation of the system of coagulation (Bauer et al., 2011). It was revealed that AgNPs reduced the apoptosis induced by TNF- $\alpha$  (Fehaid and Taniguchi, 2018). Curcumin nanoparticles (Cur-NPs) also induced apoptosis and caused G2/M arrest in both A549 and Calu-3 cell lines (Lee et al., 2016). Bevacizumab (Avastin, AV) nano-treatments also significantly suppressed A549 cell viability and activated apoptosis by NO level elevation (Abd-Rabou and Ahmed, 2019).

## 5.2 The dosage of NPs affecting respiratory disorders

The doses and duration of different types of NPs used on their effects on respiratory disorders were described in different sections of the review as summarized in Table 4.

## 5.3 Future trends and prospects

The current review article mainly focuses on the effects of NPs on respiratory disorders based on *in vitro* and *in vivo* studies. However, to use different types of NPs in the treatment of various respiratory disorders, further studies, including *in vivo* and mainly clinical trials, should be performed on the following topics:

- Evaluation of the effect of different types of NPs more precisely.
- Accurately determining the duration of the effect of various types of nanoparticles.
- Find out the more effective doses of different types of NPs in particular.

**TABLE 4** Doses and durations used for examining the effects of various nanoparticles on lung disorders.

Nanoparticle	Dose	Duration
AgNPs: silver nanoparticles	8–128 µg/ml	1, 6, 24 h
AuNPs: gold nanoparticles	20, 100 µg/ml–5.00 mg/ml	24, 48, 72 h
Al <sub>2</sub> O <sub>3</sub> NPs	0.4, 2 mg/ml	24 h
Carbon nanotubes	0.4–20 µg/ml	24 h
CdO NPs: cadmium oxide nanoparticles	1 µg/ml	24 h
CdSe: cadmium selenide	330 µl	24 h
CS: chitosan	5 µg/m–2.5 mg/ml	6 and 24 h
LCN: liquid crystalline NPs	10–50 µg/ml	12, 24, and 48 h
LPB: liposome-in-bacteria	0.62–5 µmol/L	24 h
NiO NPs: nickel oxide NPs	5 µg/ml–0.24 mg/kg	6 weeks
ONP-302-NPs	1 mg	
QDs: quantum dots	5–20 µg/ml and 6 µg/kg	24 h
SBS: salbutamol sulfate	200 µg aerosolized	14 h
siRNA: small interfering RNA	5 µg–2.5 mg/kg	24 h and 3 days
SLN: solid lipid nanoparticles	30 µg/25 g/kg	1, 2, 3, and 6 days
TAC: tacrolimus	60 µg/mouse	
ZnO: zinc oxide nanoparticle	10, 20, and 40 µg/ml	

References were provided in the main text.

- Examination of the different routes of administration of NPs, especially the effect of inhaler NPs.
- Assessment of the clinical outcome of different types of NPs in particular.

## 5.4 Conclusion

In this review article, the effects of different types of NPs on various lung disorders, including drug delivery, efficacy, and drug safety, based on the *in vitro* and *in vivo* findings and limited clinical trials were described. In asthma and COPD, different types of NPs reduced inflammatory cells and markers and improved lung oxidative stress and lung pathological changes in *in vitro* and *in vivo* studies. In PF also, although some studies indicated the therapeutic effects of NP drugs, other studies showed the induction of PF by NPs. Several *in vitro* and *in vivo* studies showed the effects of nanoparticle-based medicine as a promising tool for the treatment of lung infections by local antibiotic properties with low doses and reduced side effects. The effects of NPs on lung cancers (both primary and metastatic) were shown by increased drug delivery to selective lung cancer and lung metastases locations, as well as by increasing efficacy on lung cancer cells. The studies also showed that NP-based drugs are more effective than ordinary drugs. Some clinical studies also showed promising effects of NPs on the treatment of lung cancer. Various possible mechanisms of NPs on lung disorders were also summarized. However, more experimental and clinical studies are needed to evaluate the effects of

nanomedicine on respiratory disorders and perform a meta-analysis to systematically assess the results of previous research before the production of nanoparticle-based medicine could be applied for clinical use. In addition, public knowledge regarding the effects of NPs and their benefits on various lung disorders should be encouraged.

## Author contributions

METY, MQ, SB, and MB prepared the first draft of the manuscript and helped in the revision of the final version of the manuscript. MB designed the study and critically edited and revised the manuscript. All authors contributed to the article and approved the submitted version.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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## References

- Abd-Rabou, A. A., and Ahmed, H. H. (2019). Bevacizumab and CCR2 inhibitor nanoparticles induce cytotoxicity-mediated apoptosis in doxorubicin-treated hepatic and non-small lung cancer cells. *Asian Pac. J. Cancer Prev. APJCP* 20, 2225–2238. doi:10.31557/APJCP.2019.20.7.2225
- Adali, M. B., Barresi, A. A., Boccardo, G., and Pisano, R. (2020). Spray freeze-drying as a solution to continuous manufacturing of pharmaceutical products in bulk. *Processes* 8, 709. doi:10.3390/pr8060709
- Ahmad, A. (2022). Pharmacological strategies and recent advancement in nano-drug delivery for targeting asthma. *Life* 12, 596. doi:10.3390/life12040596
- Akbarzadeh, I., Farid, M., Javidfar, M., Zabet, N., Shokoohian, B., Arki, M. K., et al. (2022). The optimized formulation of tamoxifen-loaded niosomes efficiently induced apoptosis and cell cycle arrest in breast cancer cells. *AAPS PharmSciTech* 23, 57–13. doi:10.1208/s12249-022-02212-0
- Akhter, M. H., Rizwanullah, M., Ahmad, J., Ahsan, M. J., Mujtaba, M. A., and Amin, S. (2018). Nanocarriers in advanced drug targeting: Setting novel paradigm in cancer therapeutics. *Artif. cells, nanomedicine, Biotechnol.* 46, 873–884. doi:10.1080/21691401.2017.1366333
- Al-Otaibi, A. M., Al-Gebaly, A. S., Almeer, R., Albasher, G., Al-Qahtani, W. S., and Abdel Moneim, A. E. (2022). Potential of green-synthesized selenium nanoparticles using apigenin in human breast cancer MCF-7 cells. *Environ. Sci. Pollut. Res.* 29, 47539–47548. doi:10.1007/s11356-022-19166-2
- Alhawmdh, M., Isreb, M., Aziz, A., Jacob, B. K., Anderson, D., and Najafzadeh, M. (2021). Interferon- $\gamma$  liposome: A new system to improve drug delivery in the treatment of lung cancer. *ERJ Open Res.* 7, 00555. doi:10.1183/23120541.00555-2020
- Almuqbil, R. M., Heyder, R. S., Bielski, E. R., Durymanov, M., Reineke, J. J., and Da Rocha, S. R. (2020). Dendrimer conjugation enhances tumor penetration and efficacy of doxorubicin in extracellular matrix-expressing 3D lung cancer models. *Mol. Pharm.* 17, 1648–1662. doi:10.1021/acs.molpharmaceut.0c00083
- Alphandéry, E. (2020). Iron oxide nanoparticles for therapeutic applications. *Drug Discov. Today* 25, 141–149. doi:10.1016/j.drudis.2019.09.020
- Alqahtani, J. S. (2022). Prevalence, incidence, morbidity and mortality rates of COPD in Saudi Arabia: Trends in burden of COPD from 1990 to 2019. *PLoS One* 17, e0268772. doi:10.1371/journal.pone.0268772
- Ambekar, R. S., Choudhary, M., and Kandasubramanian, B. (2020). Recent advances in dendrimer-based nanoplatform for cancer treatment: A review. *Eur. Polym. J.* 126, 109546. doi:10.1016/j.eurpolymj.2020.109546
- Amreddy, N., Babu, A., Panneerselvam, J., Srivastava, A., Muralidharan, R., Chen, A., et al. (2018). Chemo-biologic combinatorial drug delivery using folate receptor-targeted dendrimer nanoparticles for lung cancer treatment. *Nanomedicine Nanotechnol. Biol. Med.* 14, 373–384. doi:10.1016/j.nano.2017.11.010
- Anbu, P., Gopinath, S. C., Saravanakumar, K., Vijayakumar, S., Ramanathan, S., and Wang, M.-H. (2021). Bimetallic mixed metal oxide (CuO/NiO) in fusion with nitrogen-doped graphene oxide: An alternate approach for developing potential biocarrier. *J. Environ. Chem. Eng.* 9, 105781. doi:10.1016/j.jece.2021.105781
- Ashna, M., Es-Haghi, A., Karimi Noghondar, M., Al Amara, D., and Yazdi, M. E. T. (2020). Greener synthesis of cerium oxide nanoemulsion using pollen grains of Brassica napus and evaluation of its antitumor and cytotoxicity properties. *Mater. Technol.* 37, 525–532. doi:10.1080/10667857.2020.1863558
- Athari, S. S., Pourpak, Z., Folkerts, G., Garssen, J., Moin, M., Adcock, I. M., et al. (2016). Conjugated Alpha-Alumina nanoparticle with vasoactive intestinal peptide as a Nano-drug in treatment of allergic asthma in mice. *Eur. J. Pharmacol.* 791, 811–820. doi:10.1016/j.ejphar.2016.10.014
- Bai, J., He, Z., Li, L., Dang, S., Li, Y., and Sun, W. (2018). The influence of side-coupled quantum dots on thermoelectric effect of parallel-coupled double quantum dot system. *Phys. B Condens. Matter* 545, 377–382. doi:10.1016/j.physb.2018.06.040
- Barreto, E., Serra, M. F., Dos Santos, R. V., Dos Santos, C. E. A., Hickmann, J., Cotias, A. C., et al. (2015). Local administration of gold nanoparticles prevents pivotal pathological changes in murine models of atopic asthma. *J. Biomed. Nanotechnol.* 11, 1038–1050. doi:10.1166/jbn.2015.2024
- Bauer, A. T., Strozzyk, E. A., Gorzelanny, C., Westerhausen, C., Desch, A., Schneider, M. F., et al. (2011). Cytotoxicity of silica nanoparticles through exocytosis of von Willebrand factor and necrotic cell death in primary human endothelial cells. *Biomaterials* 32, 8385–8393. doi:10.1016/j.biomaterials.2011.07.078
- Belhadj-Tahar, H., Chen, J., Song, P., Zhao, J., Quan, M., Li, C., et al. (2019). Novel CT-guided 188-rhenium brachytherapy device for local primary and secondary lung malignancies. Virginia, United States: American Society of Clinical Oncology.
- Bianco, F., Salomone, F., Milesi, I., Murgia, X., Bonelli, S., Pasini, E., et al. (2021). Aerosol drug delivery to spontaneously-breathing preterm neonates: Lessons learned. *Respir. Res.* 22, 71. doi:10.1186/s12931-020-01585-9
- Bohr, A., Tsapis, N., Andreana, I., Chamarat, A., Foged, C., Delomenie, C., et al. (2017). Anti-inflammatory effect of anti-TNF- $\alpha$  siRNA cationic phosphorus dendrimer nanocomplexes administered intranasally in a murine acute lung injury model. *Biomacromolecules* 18, 2379–2388. doi:10.1021/acs.biomac.7b00572
- Buhecha, M. D., Lansley, A. B., Somavarapu, S., and Pannala, A. S. (2019). Development and characterization of PLA nanoparticles for pulmonary drug delivery: Co-Encapsulation of theophylline and budesonide, a hydrophilic and lipophilic drug. *J. Drug Deliv. Sci. Technol.* 53, 101128. doi:10.1016/j.jddst.2019.101128
- Bushueva, T. V., Panov, V. G., Minigalieva, I. A., Privalova, L. I., Vedernikova, M. S., Gurvich, V. B., et al. (2023). Dose dependence of the separate and combined impact of copper-oxide and selenium-oxide nanoparticles on oxygen consumption by cells *in vitro* with or without the background action of some modulators of the mitochondrial respiratory function. *Dose-Response* 21, 155932582211066. doi:10.1177/15593258221106612
- Cai, X., Luo, Y., Zhang, W., Du, D., and Lin, Y. (2016). pH-Sensitive ZnO quantum dots-doxorubicin nanoparticles for lung cancer targeted drug delivery. *ACS Appl. Mater. interfaces* 8, 22442–22450. doi:10.1021/acsami.6b04933
- Cai, Y., Van Putten, J. P., Gilbert, M. S., Gerrits, W. J., Folkerts, G., and Braber, S. (2022). Galacto-oligosaccharides as an anti-bacterial and anti-invasive agent in lung infections. *Biomaterials* 283, 121461. doi:10.1016/j.biomaterials.2022.121461
- Castellani, S., Trapani, A., Spagnoletta, A., Di Toma, L., Magrone, T., Di Gioia, S., et al. (2018). Nanoparticle delivery of grape seed-derived proanthocyanidins to airway epithelial cells dampens oxidative stress and inflammation. *J. Transl. Med.* 16, 140–215. doi:10.1186/s12967-018-1509-4
- Castoldi, A., Herr, C., Niederstraßer, J., Labouta, H. I., Melero, A., Gordon, S., et al. (2017). Calcifediol-loaded liposomes for local treatment of pulmonary bacterial infections. *Eur. J. Pharm. Biopharm.* 118, 62–67. doi:10.1016/j.ejpb.2016.11.026
- Cedrowska, E., Pruszyński, M., Gaweda, W., Zuk, M., Krysinski, P., Bruchertseifer, F., et al. (2020). Trastuzumab conjugated superparamagnetic iron oxide nanoparticles labeled with (225)Ac as a perspective tool for combined alpha-radioimmunotherapy and magnetic hyperthermia of HER2-positive breast cancer. *Molecules* 25, 1025. doi:10.3390/molecules25051025
- Chakraborty, S., Ehsan, I., Mukherjee, B., Mondal, L., Roy, S., Saha, K. D., et al. (2019). Therapeutic potential of andrographolide-loaded nanoparticles on a murine asthma model. *Nanomedicine Nanotechnol. Biol. Med.* 20, 102006. doi:10.1016/j.nano.2019.04.009
- Chakravarthy, K. V., Davidson, B. A., Helinski, J. D., Ding, H., Law, W.-C., Yong, K.-T., et al. (2011). Doxorubicin-conjugated quantum dots to target alveolar macrophages and inflammation. *Nanomedicine* 7, 88–96. doi:10.1016/j.nano.2010.09.001
- Chan, H. W., Chow, S., Zhang, X., Zhao, Y., Tong, H. H. Y., and Chow, S. F. (2023). Inhalable nanoparticle-based dry powder formulations for respiratory diseases: Challenges and strategies for translational research. *AAPS PharmSciTech* 24, 98. doi:10.1208/s12249-023-02559-y
- Chanda, N., Kattumuri, V., Shukla, R., Zambre, A., Katti, K., Upendran, A., et al. (2010). Bombesin functionalized gold nanoparticles show *in vitro* and *in vivo* cancer receptor specificity. *Proc. Natl. Acad. Sci.* 107, 8760–8765. doi:10.1073/pnas.1002143107
- Chang, X., Zhu, A., Liu, F., Zou, L., Su, L., Liu, S., et al. (2017). Nickel oxide nanoparticles induced pulmonary fibrosis via TGF- $\beta$  1 activation in rats. *Hum. Exp. Toxicol.* 36, 802–812. doi:10.1177/0960327116666650
- Chauhan, S., Jain, N., and Nagaich, U. (2020). Nanodiamonds with powerful ability for drug delivery and biomedical applications: Recent updates on *in vivo* study and patents. *J. Pharm. analysis* 10, 1–12. doi:10.1016/j.jpha.2019.09.003
- Chen, C.-T., Salunke, S., Wei, T.-T., Tang, Y.-A., and Wang, Y.-C. (2021). Fluorescent nanohybrids from ZnS/CdSe quantum dots functionalized with triantennary, N-Hydroxy-p-(4-arylbutanamido)benzamide/Gallamide dendrons that act as inhibitors of histone deacetylase for lung cancer. *ACS Appl. Bio Mater.* 4, 2475–2489. doi:10.1021/acsabm.0c01438
- Chen, T., Li, L., Xu, G., Wang, X., Wang, J., Chen, Y., et al. (2018). Cytotoxicity of InP/ZnS quantum dots with different surface functional groups toward two lung-derived cell lines. *Front. Pharmacol.* 9, 763. doi:10.3389/fphar.2018.00763

- Chen, X., Huang, W., Wong, B. C., Yin, L., Wong, Y. F., Xu, M., et al. (2012). Liposomes prolong the therapeutic effect of anti-asthmatic medication via pulmonary delivery. *Int. J. nanomedicine* 7, 1139–1148. doi:10.2147/IJN.S28011
- Chikuma, K., Arima, K., Asaba, Y., Kubota, R., Asayama, S., Sato, K., et al. (2020). The potential of lipid-polymer nanoparticles as epigenetic and ROS control approaches for COPD. *Free Radic. Res.* 54, 829–840. doi:10.1080/10715762.2019.1696965
- Cryan, S. A., Devocelle, M., Moran, P. J., Hickey, A. J., and Kelly, J. G. (2006). Increased intracellular targeting to airway cells using octaarginine-coated liposomes: *In vitro* assessment of their suitability for inhalation. *Mol. Pharm.* 3, 104–112. doi:10.1021/mp050070i
- Kucik, V., Lovre, V., Dragisic, D., and Ustamujic, A. (2012). Asthma and chronic obstructive pulmonary disease (COPD)—differences and similarities. *Mater. socio-medica* 24, 100–105. doi:10.5455/msm.2012.24.100-105
- Cyril, N., George, J. B., Joseph, L., Raghavamenon, A., and Vp, S. (2019). Assessment of antioxidant, antibacterial and anti-proliferative (lung cancer cell line A549) activities of green synthesized silver nanoparticles from *Derris trifoliata*. *Toxicol. Res.* 8, 297–308. doi:10.1039/c8tx00323h
- Das, S. S., Singh, S. K., Verma, P., Jha, N. K., Gupta, P. K., and Dua, K. (2022). Mitigating inflammation using advanced drug delivery by targeting TNF- $\alpha$  in lung diseases. *Future Med. Chem.* 14, 57–60. doi:10.4155/fmc-2021-0225
- De Sá Coutinho, D., Pires, J., Gomes, H., Raffin Pohlmann, A., Stanisciuaqui Guterres, S., Rodrigues E Silva, P. M., et al. (2020). Pequi (caryocar brasiliense cambess)-loaded nanoemulsion, orally delivered, modulates inflammation in LPS-induced acute lung injury in mice. *Pharmaceutics* 12, 1075. doi:10.3390/pharmaceutics12111075
- Dekkers, S., Williams, T. D., Zhang, J., Zhou, J. A., Vandebriel, R. J., De La Fonteyne, L. J., et al. (2018). Multi-omics approaches confirm metal ions mediate the main toxicological pathways of metal-bearing nanoparticles in lung epithelial A549 cells. *Environ. Sci. Nano* 5, 1506–1517. doi:10.1039/c8en00071a
- Deng, Z., Kalin, G. T., Shi, D., and Kalinichenko, V. V. (2021). Nanoparticle delivery systems with cell-specific targeting for pulmonary diseases. *Am. J. Respir. Cell Mol. Biol.* 64, 292–307. doi:10.1165/rmb.2020-0306TR
- Dhayanandamoorthy, Y., Antoniraj, M. G., Kandregula, C. a. B., and Kandasamy, R. (2020). Aerosolized hyaluronic acid decorated, ferulic acid loaded chitosan nanoparticle: A promising asthma control strategy. *Int. J. Pharm.* 591, 119958. doi:10.1016/j.ijpharm.2020.119958
- Dumont, C., Bourgeois, S., Fessi, H., and Jannin, V. (2018). Lipid-based nanosuspensions for oral delivery of peptides, a critical review. *Int. J. Pharm.* 541, 117–135. doi:10.1016/j.ijpharm.2018.02.038
- El-Hammadi, M. M., and Arias, J. L. (2019). An update on liposomes in drug delivery: A patent review (2014–2018). *Expert Opin. Ther. Pat.* 29, 891–907. doi:10.1080/13543776.2019.1679767
- Fang, L., Nikfarjam, N., Gharagozlou, M., Huang, T., Song, Y., Islambulchilar, Z., et al. (2022). Pulmonary delivery of levamisole nanoparticles as an immunomodulator affecting Th and a potential ADAM10 inhibitor to ameliorate severe allergic asthma. *ACS Biomaterials Sci. Eng.* 8, 4566–4576. doi:10.1021/acsbmaterials.2c00843
- Fehaid, A., and Taniguchi, A. (2018). Silver nanoparticles reduce the apoptosis induced by tumor necrosis factor- $\alpha$ . *Sci. Technol. Adv. Mater.* 19, 526–534. doi:10.1080/14686996.2018.1487761
- Forest, V., and Pourchez, J. (2022). Nano-delivery to the lung-by inhalation or other routes and why nano when micro is largely sufficient? *Adv. Drug Deliv. Rev.* 183, 114173. doi:10.1016/j.addr.2022.114173
- Frede, A., Neuhaus, B., Knuschke, T., Wadwa, M., Kollenda, S., Klopffleisch, R., et al. (2017). Local delivery of siRNA-loaded calcium phosphate nanoparticles abates pulmonary inflammation. *Nanomedicine Nanotechnol. Biol. Med.* 13, 2395–2403. doi:10.1016/j.nano.2017.08.001
- Gamal-Eldeen, A. M., Moustafa, D., El-Daly, S. M., Abo-Zeid, M. A., Saleh, S., Khoobchandani, M., et al. (2017). Gum Arabic-encapsulated gold nanoparticles for a non-invasive photothermal ablation of lung tumor in mice. *Biomed. Pharmacother.* 89, 1045–1054. doi:10.1016/j.biopha.2017.03.006
- Gao, L., Fan, K., and Yan, X. (2017). Iron oxide nanozyme: A multifunctional enzyme mimetic for biomedical applications. *Theranostics* 7, 3207–3227. doi:10.7150/thno.19738
- Gao, Y., Wang, J., Chai, M., Li, X., Deng, Y., Jin, Q., et al. (2020). Size and charge adaptive clustered nanoparticles targeting the biofilm microenvironment for chronic lung infection management. *ACS nano* 14, 5686–5699. doi:10.1021/acsnano.0c00269
- Garbuzenko, O. B., Mainelis, G., Taratula, O., and Minko, T. (2014). Inhalation treatment of lung cancer: The influence of composition, size and shape of nanocarriers on their lung accumulation and retention. *Cancer Biol. Med.* 11, 44–55. doi:10.7497/j.issn.2095-3941.2014.01.004
- Ghorani-Azam, A., Mottaghipisheh, J., Amiri, M. S., Mashreghi, M., Hashemzadeh, A., Haddad-Mashadrizah, A., et al. (2022). Resveratrol-mediated gold-nanoceria synthesis as green nanomedicine for phytotherapy of hepatocellular carcinoma. *Front. Bioscience-Landmark* 27, 227. doi:10.31083/j.fbl2708227
- Glass, D. S., Grossfeld, D., Renna, H. A., Agarwala, P., Spiegler, P., Deleon, J., et al. (2022). Idiopathic pulmonary fibrosis: Current and future treatment. *Clin. Respir. J.* 16, 84–96. doi:10.1111/crj.13466
- Gorain, B., Choudhury, H., Nair, A. B., Dubey, S. K., and Kesharwani, P. (2020). Theranostic application of nanoemulsions in chemotherapy. *Drug Discov. Today* 25, 1174–1188. doi:10.1016/j.drudis.2020.04.013
- Grace, V. B., and Viswanathan, S. (2017). Pharmacokinetics and therapeutic efficiency of a novel cationic liposome nano-formulated all trans retinoic acid in lung cancer mice model. *J. Drug Deliv. Sci. Technol.* 39, 223–236. doi:10.1016/j.jddst.2017.04.005
- Guo, H., You, B., Zhao, S., Wang, Y., Sun, G., Bai, Y., et al. (2018). Full-color tunable photoluminescent carbon dots based on oil/water interfacial synthesis and their applications. *RSC Adv.* 8, 24002–24012. doi:10.1039/c8ra03723j
- Gurunathan, S., Jeyaraj, M., Kang, M.-H., and Kim, J.-H. (2020). Melatonin enhances palladium-nanoparticle-induced cytotoxicity and apoptosis in human lung epithelial adenocarcinoma cells A549 and H1229. *Antioxidants* 9, 357. doi:10.3390/antiox9040357
- Habas, K., Abdulmwli, M., Demir, E., Jacob, B. K., Najafzadeh, M., and Anderson, D. (2018). DNA damage protection by bulk and nano forms of quercetin in lymphocytes of patients with chronic obstructive pulmonary disease exposed to the food mutagen 2-amino-3-methylimidazo [4, 5-f] quinolone (IQ). *Environ. Res.* 166, 10–15. doi:10.1016/j.envres.2018.05.012
- Halwani, R., Shaik, A. S., Ratemi, E., Afzal, S., Kenana, R., Al-Muhsen, S., et al. (2016). A novel anti-IL4Ra nanoparticle efficiently controls lung inflammation during asthma. *Exp. Mol. Med.* 48, e262. doi:10.1038/emmm.2016.89
- He, B., Jin, H.-Y., Wang, Y.-W., Fan, C.-M., Wang, Y.-F., Zhang, X.-C., et al. (2021). Carbon quantum dots/Bi4O5Br2 photocatalyst with enhanced photodynamic therapy: Killing of lung cancer (A549) cells *in vitro*. *Rare Met.* 41, 132–143. doi:10.1007/s12598-021-01762-9
- Hoda Zarharan, M. B., Rokhi, A. S., Bajgiran, R. R., Yousefi, E., Heravian, P., Khazrabig, M. N., et al. (2023). The anti-angiogenesis and antioxidant activity of chitosan-mediated synthesized selenium-gold nanostructure. *Arabian J. Chem.* 16, 104806. doi:10.1016/j.arabj.2023.104806
- Honmane, S., Hajare, A., More, H., Osmani, R. a. M., and Salunkhe, S. (2019). Lung delivery of nanoliposomal salbutamol sulfate dry powder inhalation for facilitated asthma therapy. *J. Liposome Res.* 29, 332–342. doi:10.1080/08982104.2018.1531022
- Horváth, I., Barnes, P. J., Loukides, S., Sterk, P. J., Högman, M., Olin, A.-C., et al. (2017). A European respiratory society technical standard: Exhaled biomarkers in lung disease. *Eur. Respir. J.* 49, 1600965. doi:10.1183/13993003.00965-2016
- Hsu, C.-Y., Wang, P.-W., Alalawi, A., Lin, Z.-C., and Fang, J.-Y. (2019). Use of lipid nanocarriers to improve oral delivery of vitamins. *Nutrients* 11, 68. doi:10.3390/nu11010068
- Hu, Y., Zhang, J., Hu, H., Xu, S., Xu, L., and Chen, E. (2020). Gefitinib encapsulation based on nano-liposomes for enhancing the curative effect of lung cancer. *Cell Cycle* 19, 3581–3594. doi:10.1080/15384101.2020.1852756
- Hu, Z. M., Fei, G. T., and De Zhang, L. (2019). Synthesis and tunable emission of Ga2S3 quantum dots. *Mater. Lett.* 239, 17–20. doi:10.1016/j.matlet.2018.12.046
- Huang, Q., Li, L., Li, L., Chen, H., Dang, Y., Zhang, J., et al. (2016a). MDM2 knockdown mediated by a triazine-modified dendrimer in the treatment of non-small cell lung cancer. *Oncotarget* 7, 44013–44022. doi:10.18632/oncotarget.9768
- Huang, Q., Qian, X., Fan, J., Lv, Z., Zhang, X., Han, J., et al. (2016b). Epigenetic regulation of active Chinese herbal components for cancer prevention and treatment: A follow-up review. *Sci. Rep.* 6, 1–12. doi:10.1016/j.phrs.2016.09.023
- Hussain, S., Vanoirbeek, J. A., Luyts, K., De Vooght, V., Verbeken, E., Thomassen, L. C., et al. (2011). Lung exposure to nanoparticles modulates an asthmatic response in a mouse model. *Eur. Respir. J.* 37, 299–309. doi:10.1183/09031936.00168509
- Hutchinson, D., Müller, J., Mccarthy, J. E., Gun'ko, Y. K., Verma, N. K., Bi, X., et al. (2018). Cadmium nanoparticles citrullinate cyokeratins within lung epithelial cells: Cadmium as a potential cause of citrullination in chronic obstructive pulmonary disease. *Int. J. Chronic Obstr. Pulm. Dis.* 13, 441–449. doi:10.2147/COPD.S152028
- Javanbakht, S., Hemmati, A., Namazi, H., and Heydari, A. (2020). Carboxymethylcellulose-coated 5-fluorouracil@ MOF-5 nano-hybrid as a bio-nanocomposite carrier for the anticancer oral delivery. *Int. J. Biol. Macromol.* 155, 876–882. doi:10.1016/j.ijbiomac.2019.12.007
- Jin, S., Hu, Y., Gu, Z., Liu, L., and Wu, H.-C. (2011). Application of quantum dots in biological imaging. *J. Nanomater.* 2011, 1, 13. doi:10.1155/2011/834139
- Joyce, P., Whitby, C. P., and Prestidge, C. A. (2016). Nanostructuring biomaterials with specific activities towards digestive enzymes for controlled gastrointestinal absorption of lipophilic bioactive molecules. *Adv. Colloid Interface Sci.* 237, 52–75. doi:10.1016/j.cis.2016.10.003
- Kang, S. H., Shin, Y. S., Lee, D.-H., Park, I. S., Kim, S. K., Ryu, D., et al. (2022). Interactions of nanoparticles with macrophages and feasibility of drug delivery for asthma. *Int. J. Mol. Sci.* 23, 1622. doi:10.3390/ijms23031622
- Kelley, W. J., Wragg, K. M., Chen, J., Murthy, T., Xu, Q., Boyne, M. T., et al. (2022). Nanoparticles reduce monocytes within the lungs to improve outcomes after influenza virus infection in aged mice. *JCI insight* 7, e156320. doi:10.1172/jci.insight.156320



- Khakzad, M. R., Hajavi, J., Sadeghdoust, M., and Aligolighasemabadi, F. (2019). "Effects of lipopolysaccharide-loaded PLGA nanoparticles in mice model of asthma by sublingual immunotherapy," in *International journal of polymeric materials and polymeric biomaterials* (United Kingdom: Taylor And Francis).
- Khan, O. F., Zaia, E. W., Jhunjhunwala, S., Xue, W., Cai, W., Yun, D. S., et al. (2015). Dendrimer-inspired nanomaterials for the *in vivo* delivery of siRNA to lung vasculature. *Nano Lett.* 15, 3008–3016. doi:10.1021/nl5048972
- Khemtong, C., Kessinger, C. W., and Gao, J. (2009). Polymeric nanomedicine for cancer MR imaging and drug delivery. *Chem. Commun.* 2009, 3497–3510. doi:10.1039/b21865j
- Kim, D. E., Lee, Y., Kim, M., Lee, S., Jon, S., and Lee, S.-H. (2017). Bilirubin nanoparticles ameliorate allergic lung inflammation in a mouse model of asthma. *Biomaterials* 140, 37–44. doi:10.1016/j.biomaterials.2017.06.014
- Kim, H. B., Meghani, N., Park, M., Lee, S. H., Lee, S. R., Cho, Y.-J., et al. (2020). Electrohydrodynamically atomized pH-responsive PLGA/ZnO quantum dots for local delivery in lung cancer. *Macromol. Res.* 28, 407–414. doi:10.1007/s13233-020-8053-9
- Kim, H., Park, M., Hwang, J., Kim, J. H., Chung, D.-R., Lee, K.-S., et al. (2019). Development of label-free colorimetric assay for MERS-CoV using gold nanoparticles. *ACS sensors* 4, 1306–1312. doi:10.1021/acssensors.9b00175
- Kim, J.-S. (2016). Liposomal drug delivery system. *J. Pharm. investigation* 46, 387–392. doi:10.1007/s40005-016-0260-1
- Komalla, V., Allam, V., Kwok, P. C. L., Sheikholeslami, B., Owen, L., Jaffe, A., et al. (2020). A phospholipid-based formulation for the treatment of airway inflammation in chronic respiratory diseases. *Eur. J. Pharm. Biopharm.* 157, 47–58. doi:10.1016/j.ejpb.2020.09.017
- Konduri, K. S., Nandedkar, S., Rickaby, D. A., Düzgüneş, N., and Gangadharam, P. R. (2005). The use of sterically stabilized liposomes to treat asthma. *Methods Enzym.* 391, 413–427. doi:10.1016/S0076-6879(05)91023-9
- Kulkarni, N. S., Parvathani, V., Shukla, S. K., Barasa, L., Perron, J. C., Yoganathan, S., et al. (2019). Tyrosine kinase inhibitor conjugated quantum dots for non-small cell lung cancer (NSCLC) treatment. *Eur. J. Pharm. Sci.* 133, 145–159. doi:10.1016/j.ejps.2019.03.026
- Kumar, A., Najafzadeh, M., Jacob, B. K., Dhawan, A., and Anderson, D. (2015). Zinc oxide nanoparticles affect the expression of p53, Ras p21 and JNKs: An *ex vivo/in vitro* exposure study in respiratory disease patients. *Mutagenesis* 30, 237–245. doi:10.1093/mutage/geu064
- Kumar, D., Vats, N., Saroha, K., and Rana, A. C. (2020). "Phytosomes as emerging nanotechnology for herbal drug delivery," in *Sustainable agriculture reviews 43* (Heidelberg, Germany: Springer), 217–237.
- Kumar, M., Kong, X., Behera, A. K., Hellermann, G. R., Lockey, R. F., and Mohapatra, S. S. (2003). Chitosan IFN- $\gamma$ -pDNA nanoparticle (CIN) therapy for allergic asthma. *Genet. Vaccines Ther.* 1, 3. doi:10.1186/1479-0556-1-3
- Lai, X., Zhao, H., Zhang, Y., Guo, K., Xu, Y., Chen, S., et al. (2018). Intranasal delivery of copper oxide nanoparticles induces pulmonary toxicity and fibrosis in C57BL/6 mice. *Sci. Rep.* 8, 4499–4512. doi:10.1038/s41598-018-22556-7
- Lee, S., Hwang, S.-H., Jeong, J., Han, Y., Kim, S.-H., Lee, D.-K., et al. (2015). Nickel oxide nanoparticles can recruit eosinophils in the lungs of rats by the direct release of intracellular eotaxin. *Part. fibre Toxicol.* 13, 30–11. doi:10.1186/s12989-016-0142-8
- Lee, W.-H., Loo, C.-Y., Ong, H.-X., Traini, D., Young, P. M., and Rohanzadeh, R. (2016). Synthesis and characterization of inhalable flavonoid nanoparticle for lung cancer cell targeting. *J. Biomed. Nanotechnol.* 12, 371–386. doi:10.1166/jbn.2016.2162
- Lee, Y., and Thompson, D. (2017). Stimuli-responsive liposomes for drug delivery. *Wiley Interdiscip. Rev. Nanomedicine Nanobiotechnology* 9, e1450. doi:10.1002/wnan.1450
- Li, B., Zhang, X.-X., Huang, H.-Y., Chen, L.-Q., Cui, J.-H., Liu, Y., et al. (2018a). Effective deactivation of A549 tumor cells *in vitro* and *in vivo* by RGD-decorated chitosan-functionalized single-walled carbon nanotube loading docetaxel. *Int. J. Pharm.* 543, 8–20. doi:10.1016/j.ijpharm.2018.03.017
- Li, J., Li, M., Tian, L., Qiu, Y., Yu, Q., Wang, X., et al. (2020). Facile strategy by hyaluronic acid functional carbon dot-doxorubicin nanoparticles for CD44 targeted drug delivery and enhanced breast cancer therapy. *Int. J. Pharm.* 578, 119122. doi:10.1016/j.ijpharm.2020.119122
- Li, Q., Zhan, S., Liu, Q., Su, H., Dai, X., Wang, H., et al. (2018b). Preparation of a sustained-release nebulized aerosol of R-terbutaline hydrochloride liposome and evaluation of its anti-asthmatic effects via pulmonary delivery in Guinea pigs. *AAPS PharmSciTech* 19, 232–241. doi:10.1208/s12249-017-0816-z
- Li, W., Yu, H., Ding, D., Chen, Z., Wang, Y., Wang, S., et al. (2019). Cold atmospheric plasma and iron oxide-based magnetic nanoparticles for synergetic lung cancer therapy. *Free Radic. Biol. Med.* 130, 71–81. doi:10.1016/j.freeradbiomed.2018.10.429
- Li, X., Yang, H., Wu, S., Meng, Q., Sun, H., Lu, R., et al. (2017). Suppression of PTPN6 exacerbates aluminum oxide nanoparticle-induced COPD-like lesions in mice through activation of STAT pathway. *Part. fibre Toxicol.* 14, 53–14. doi:10.1186/s12989-017-0234-0
- Li, Y., Liao, K., and Lee, S. (2014). A comparative study of antioxidant-loaded carbon nanoparticles as drug delivery vehicles. *RSC Adv.* 4, 56992–56997. doi:10.1039/c4ra09470k
- Lin, W., Xu, Y., Huang, C.-C., Ma, Y., Shannon, K. B., Chen, D.-R., et al. (2009). Toxicity of nano- and micro-sized ZnO particles in human lung epithelial cells. *J. Nanoparticle Res.* 11, 25–39. doi:10.1007/s11051-008-9419-7
- Liu, J., Li, G., Guo, H., Ni, C., Gao, Y., Cao, X., et al. (2023). Dual-responsive core-shell tecto dendrimers enable efficient gene editing of cancer cells to boost immune checkpoint blockade therapy. *ACS Appl. Mater. Interfaces* 15, 12809–12821. doi:10.1021/acscami.2c22584
- Liu, J., Li, R., and Yang, B. (2020). Carbon dots: A new type of carbon-based nanomaterial with wide applications. *ACS Central Sci.* 6, 2179–2195. doi:10.1021/acscentsci.0c01306
- Liu, M., Gu, X., Zhang, K., Ding, Y., Wei, X., Zhang, X., et al. (2013). Gold nanoparticles trigger apoptosis and necrosis in lung cancer cells with low intracellular glutathione. *J. nanoparticle Res.* 15, 1745–1814. doi:10.1007/s11051-013-1745-8
- Liu, N., Guan, Y., Zhou, C., Wang, Y., Ma, Z., and Yao, S. (2022). Pulmonary and systemic toxicity in a rat model of pulmonary alveolar proteinosis induced by indium-tin oxide nanoparticles. *Int. J. Nanomedicine* 17, 713–731. doi:10.2147/IJN.S338955
- Liu, T.-T., Mu, L.-Q., Dai, W., Wang, C.-B., Liu, X.-Y., and Xiang, D.-X. (2016). Preparation, characterization, and evaluation of antitumor effect of Brucea javanica oil cationic nanoemulsions. *Int. J. nanomedicine* 11, 2515–2529. doi:10.2147/IJN.S101918
- Liu, X., Cheng, X., Wang, F., Feng, L., Wang, Y., Zheng, Y., et al. (2018). Targeted delivery of SNX-2112 by polysaccharide-modified graphene oxide nanocomposites for treatment of lung cancer. *Carbohydr. Polym.* 185, 85–95. doi:10.1016/j.carbpol.2018.01.014
- Luo, M. X., Hua, S., and Shang, Q. Y. (2021). Application of nanotechnology in drug delivery systems for respiratory diseases (Review). *Mol. Med. Rep.* 23, 325. doi:10.3892/mmr.2021.11964
- Ma, J., Mercer, R. R., Barger, M., Schwegler-Berry, D., Cohen, J. M., Demokritou, P., et al. (2015). Effects of amorphous silica coating on cerium oxide nanoparticles induced pulmonary responses. *Toxicol. Appl. Pharmacol.* 288, 63–73. doi:10.1016/j.taap.2015.07.012
- Ma, Z., Wang, H., Shi, Z., Yan, F., Li, Q., Chen, J., et al. (2023). Inhalable GSH-triggered nanoparticles to treat commensal bacterial infection in *in situ* lung tumors. *ACS nano* 17, 5740–5756. doi:10.1021/acsnano.2c12165
- Maiti, D., Tong, X., Mou, X., and Yang, K. (2019). Carbon-based nanomaterials for biomedical applications: A recent study. *Front. Pharmacol.* 9, 1401. doi:10.3389/fphar.2018.01401
- Maretti, E., Costantino, L., Buttini, F., Rustichelli, C., Leo, E., Truzzi, E., et al. (2019). Newly synthesized surfactants for surface mannosylation of respirable SLN assemblies to target macrophages in tuberculosis therapy. *Drug Deliv. Transl. Res.* 9, 298–310. doi:10.1007/s13346-018-00607-w
- Meers, P., Neville, M., Malinin, V., Scotto, A., Sardaryan, G., Kurumunda, R., et al. (2008). Biofilm penetration, triggered release and *in vivo* activity of inhaled liposomal amikacin in chronic *Pseudomonas aeruginosa* lung infections. *J. Antimicrob. Chemother.* 61, 859–868. doi:10.1093/jac/dkn059
- Mei, Q., Liu, Z., Zuo, H., Yang, Z., and Qu, J. (2022). Idiopathic pulmonary fibrosis: An update on pathogenesis. *Front. Pharmacol.* 12, 797292. doi:10.3389/fphar.2021.797292
- Miao, T., Little, A. C., Aronshtam, A., Marquis, T., Fenn, S. L., Hristova, M., et al. (2020). Internalized FGF-2-loaded nanoparticles increase nuclear ERK1/2 content and result in lung cancer cell death. *Nanomaterials* 10, 612. doi:10.3390/nano10040612
- Mobaraki, F., Momeni, M., Yazdi, M. E. T., Meshkat, Z., Toosi, M. S., and Hosseini, S. M. (2021). Plant-derived synthesis and characterization of gold nanoparticles: Investigation of its antioxidant and anticancer activity against human testicular embryonic carcinoma stem cells. *Process Biochem.* 111, 167–177. doi:10.1016/j.procbio.2021.09.010
- Mohamed, A., Kunda, N. K., Ross, K., Hutcheon, G. A., and Saleem, I. Y. (2019). Polymeric nanoparticles for the delivery of miRNA to treat chronic obstructive pulmonary disease (COPD). *Eur. J. Pharm. Biopharm.* 136, 1–8. doi:10.1016/j.ejpb.2019.01.002
- Moreno-Sastre, M., Pastor, M., Esquisabel, A., Sans, E., Viñas, M., Fleischer, A., et al. (2016). Pulmonary delivery of tobramycin-loaded nanostructured lipid carriers for *Pseudomonas aeruginosa* infections associated with cystic fibrosis. *Int. J. Pharm.* 498, 263–273. doi:10.1016/j.ijpharm.2015.12.028
- Mostovenko, E., Canal, C. G., Cho, M., Sharma, K., Erdely, A., Campen, M. J., et al. (2022). Indirect mediators of systemic health outcomes following nanoparticle inhalation exposure. *Pharmacol. Ther.* 235, 108120. doi:10.1016/j.pharmthera.2022.108120
- Mousavi-Kouhi, S. M., Beyk-Khormizi, A., Amiri, M. S., Mashreghi, M., and Yazdi, M. E. T. (2021a). Silver-zinc oxide nanocomposite: From synthesis to antimicrobial and anticancer properties. *Ceram. Int.* 47, 21490–21497. doi:10.1016/j.ceramint.2021.04.160
- Mousavi-Kouhi, S. M., Beyk-Khormizi, A., Mohammadzadeh, V., Ashna, M., Es-Haghi, A., Mashreghi, M., et al. (2021b). Biological synthesis and characterization of

- gold nanoparticles using *Verbascum speciosum* Schrad. and cytotoxicity properties toward HepG2 cancer cell line. *Res. Chem. Intermed.* 48, 167–178. doi:10.1007/s11164-021-04600-w
- Muhammad, W., Zhai, Z., Wang, S., and Gao, C. (2022). Inflammation-modulating nanoparticles for pneumonia therapy. *Wiley Interdiscip. Rev. Nanomedicine Nanobiotechnology* 14, e1763. doi:10.1002/wnan.1763
- Muralidharan, R., Babu, A., Amreddy, N., Basalingappa, K., Mehta, M., Chen, A., et al. (2016). Folate receptor-targeted nanoparticle delivery of HuR-RNAi suppresses lung cancer cell proliferation and migration. *J. nanobiotechnology* 14, 47–17. doi:10.1186/s12951-016-0201-1
- Naciute, M., Niemi, V., Kemp, R. A., and Hook, S. (2020). Lipid-encapsulated oral therapeutic peptide vaccines reduce tumour growth in an orthotopic mouse model of colorectal cancer. *Eur. J. Pharm. Biopharm.* 152, 183–192. doi:10.1016/j.ejpb.2020.04.020
- Nafee, N., Schneider, M., Friebel, K., Dong, M., Schaefer, U. F., Mürdter, T. E., et al. (2012). Treatment of lung cancer via telomerase inhibition: Self-assembled nanoplexes versus polymeric nanoparticles as vectors for 2'-O-Methyl-RNA. *Eur. J. Pharm. Biopharm.* 80, 478–489. doi:10.1016/j.ejpb.2011.11.019
- Nalepa, P., Gawecki, R., Szewczyk, G., Balin, K., Dulski, M., Sajewicz, M., et al. (2020). A [60] fullerene nanoconjugate with gemcitabine: Synthesis, biophysical properties and biological evaluation for treating pancreatic cancer. *Cancer Nanotechnol.* 11, 2–21. doi:10.1186/s12645-020-00058-4
- Nasrin, T., Patra, M., Rahaman, S. M., Das, T. K., and Shaikh, S. (2022). Biosynthesized CdS nanoparticle induces ROS-dependent apoptosis in human lung cancer cells. *Anti-Cancer Agents Med. Chem. Former. Curr. Med. Chemistry-Anti-Cancer Agents* 22, 2156–2165. doi:10.2174/187152062166621115113226
- New, R. R. (1990). *Liposomes*. Oxford, United Kingdom: Oxford University Press.
- Ng, Z. Y., Wong, J.-Y., Panneerselvam, J., Madheswaran, T., Kumar, P., Pillay, V., et al. (2018). Assessing the potential of liposomes loaded with curcumin as a therapeutic intervention in asthma. *Colloids surfaces B Biointerfaces* 172, 51–59. doi:10.1016/j.colsurfb.2018.08.027
- Nguyen, H., Nguyen, N. H., Tran, N. Q., and Nguyen, C. K. (2015). Improved method for preparing cisplatin-dendrimer nanocomplex and its behavior against NCI-H460 lung cancer cell. *J. Nanosci. Nanotechnol.* 15, 4106–4110. doi:10.1166/jnn.2015.9808
- Oberdörster, G., Castranova, V., Asgharian, B., and Sayre, P. (2015). Inhalation exposure to carbon nanotubes (CNT) and carbon nanofibers (CNF): Methodology and dosimetry. *J. Toxicol. Environ. Health, Part B* 18, 121–212. doi:10.1080/10937404.2015.1051611
- Otroj, M., Taymouri, S., Varshosaz, J., and Mirian, M. (2020). Preparation and characterization of dry powder containing sunitinib loaded PHBV nanoparticles for enhanced pulmonary delivery. *J. Drug Deliv. Sci. Technol.* 56, 101570. doi:10.1016/j.jddst.2020.101570
- Palaniappan, P., Sathishkumar, G., and Sankar, R. (2015). Fabrication of nano-silver particles using *Cymodocea serrulata* and its cytotoxicity effect against human lung cancer A549 cells line. *Spectrochimica Acta Part A Mol. Biomol. Spectrosc.* 138, 885–890. doi:10.1016/j.saa.2014.10.072
- Pallavi, S., Rudayni, H. A., Bepari, A., Niazi, S. K., and Nayaka, S. (2022). Green synthesis of Silver nanoparticles using *Streptomyces hirsutus* strain SNPGA-8 and their characterization, antimicrobial activity, and anticancer activity against human lung carcinoma cell line A549. *Saudi J. Biol. Sci.* 29, 228–238. doi:10.1016/j.sjbs.2021.08.084
- Panneer, D. S., Tirunavukkarasu, S., Sadayandi, V., Rajendiran, N., Mohammad, F., Oh, W.-C., et al. (2022). Antiproliferative potentials of chitin and chitosan encapsulated gold nanoparticles derived from unharmed *Artemia* cysts. *Chem. Phys. Lett.* 790, 139345. doi:10.1016/j.cplett.2022.139345
- Parasaram, V., Nosoudi, N., Leclair, R. J., Binks, A., and Vyavahare, N. (2016). Targeted drug delivery to emphysematous lungs: Inhibition of MMPs by doxycycline loaded nanoparticles. *Pulm. Pharmacol. Ther.* 39, 64–73. doi:10.1016/j.pupt.2016.06.004
- Park, J.-W., Lee, I.-C., Shin, N.-R., Jeon, C.-M., Kwon, O.-K., Ko, J.-W., et al. (2016). Copper oxide nanoparticles aggravate airway inflammation and mucus production in asthmatic mice via MAPK signaling. *Nanotoxicology* 10, 445–452. doi:10.3109/17435390.2015.1078851
- Parra-Nieto, J., Del Cid, M. a. G., De Cárcer, I. A., and Baeza, A. (2020). Inorganic porous nanoparticles for drug delivery in antitumoral therapy. *Biotechnol. J.* 16, 2000150. doi:10.1002/biot.202000150
- Parvathaneni, V., Kulkarni, N. S., Chauhan, G., Shukla, S. K., Elbatany, R., Patel, B., et al. (2020). Development of pharmaceutically scalable inhaled anti-cancer nanotherapy—repurposing amodiaquine for non-small cell lung cancer (NSCLC). *Mater. Sci. Eng. C* 115, 111139. doi:10.1016/j.msec.2020.111139
- Patil, J., and Sarasija, S. (2012). Pulmonary drug delivery strategies: A concise, systematic review. *Lung India* 29, 44. doi:10.4103/0970-2113.92361
- Paudel, K. R., Wadhwa, R., Tew, X. N., Lau, N. J. X., Madheswaran, T., Panneerselvam, J., et al. (2021). Rutin loaded liquid crystalline nanoparticles inhibit non-small cell lung cancer proliferation and migration *in vitro*. *Life Sci.* 276, 119436. doi:10.1016/j.lfs.2021.119436
- Pérez-Carrión, M. D., and Posadas, I. (2023). Dendrimers in neurodegenerative diseases. *Processes* 11, 319. doi:10.3390/pr11020319
- Pompilio, A., Geminiani, C., Mantini, P., Siriwardena, T. N., Di Bonaventura, I., Raymond, J. L., et al. (2018). Peptide dendrimers as “lead compounds” for the treatment of chronic lung infections by *Pseudomonas aeruginosa* in cystic fibrosis patients: *In vitro* and *in vivo* studies. *Infect. drug Resist.* 11, 1767–1782. doi:10.2147/IDR.S168868
- Poulsen, S. S., Jackson, P., Kling, K., Knudsen, K. B., Skaug, V., Kyjovska, Z. O., et al. (2016). Multi-walled carbon nanotube physicochemical properties predict pulmonary inflammation and genotoxicity. *Nanotoxicology* 10, 1263–1275. doi:10.1080/17435390.2016.1202351
- Pramanik, S., Mohanto, S., Manne, R., Rajendran, R. R., Deepak, A., Edapully, S. J., et al. (2021). Nanoparticle-based drug delivery system: The magic bullet for the treatment of chronic pulmonary diseases. *Mol. Pharm.* 18, 3671–3718. doi:10.1021/acs.molpharmaceut.1c00491
- Rani, N., Rawat, K., Shrivastava, A., Yadav, S., Gupta, K., Saini, K., et al. (2022). *In vitro* study of green synthesized ZnO nanoparticles on human lung cancer cell lines. *Mater. Today Proc.* 49, 1436–1442. doi:10.1016/j.matpr.2021.07.203
- Rastogi, A., Zivcak, M., Sytar, O., Kalaji, H. M., He, X., Mbarki, S., et al. (2017). Impact of metal and metal oxide nanoparticles on plant: A critical review. *Front. Chem.* 5, 78. doi:10.3389/fchem.2017.00078
- Rawal, T., Patel, S., and Butani, S. (2018). Chitosan nanoparticles as a promising approach for pulmonary delivery of bedaquiline. *Eur. J. Pharm. Sci.* 124, 273–287. doi:10.1016/j.ejps.2018.08.038
- Reshma, V., and Mohanan, P. (2019). Quantum dots: Applications and safety consequences. *J. Luminescence* 205, 287–298. doi:10.1016/j.jlumin.2018.09.015
- Roberts, J. R., Antonini, J. M., Porter, D. W., Chapman, R. S., Scabilloni, J. F., Young, S.-H., et al. (2013). Lung toxicity and biodistribution of Cd/Se-ZnS quantum dots with different surface functional groups after pulmonary exposure in rats. *Part. fibre Toxicol.* 10, 5–17. doi:10.1186/1743-8977-10-5
- Roy, A., Nishchaya, K., and Rai, V. K. (2022). Nanoemulsion-based dosage forms for the transdermal drug delivery applications: A review of recent advances. *Expert Opin. Drug Deliv.* 19, 303–319. doi:10.1080/17425247.2022.2045944
- Ruan, F., Liu, R., Wang, K., Zeng, J., Zuo, Z., He, C., et al. (2021). Cytotoxicity of black phosphorus quantum dots on lung-derived cells and the underlying mechanisms. *J. Hazard. Mater.* 402, 122875. doi:10.1016/j.jhazmat.2020.122875
- Ruiyi, L., Zaijun, L., Xiulan, S., Jan, J., Lin, L., Zhiguo, G., et al. (2020). Graphene quantum dot-rare Earth upconversion nanocages with extremely high efficiency of upconversion luminescence, stability and drug loading towards controlled delivery and cancer theranostics. *Chem. Eng. J.* 382, 122992. doi:10.1016/j.cej.2019.122992
- Ruzjycka-Ayoush, M., Kowalik, P., Kowalczyk, A., Bujak, P., Nowicka, A. M., Wojewodzka, M., et al. (2021). Quantum dots as targeted doxorubicin drug delivery nanosystems in human lung cancer cells. *Cancer Nanotechnol.* 12, 8. doi:10.1186/s12645-021-00077-9
- Saghir, S. A., Al-Gabri, N. A., Khafaga, A. F., El-Shaer, N. H., Alhumaidh, K. A., Elsadek, M. F., et al. (2019). Thymoquinone-PLGA-PVA nanoparticles ameliorate bleomycin-induced pulmonary fibrosis in rats via regulation of inflammatory cytokines and iNOS signaling. *Animals* 9, 951. doi:10.3390/ani9110951
- Sanaei, M.-J., Razi, S., Pourbagheri-Sigaroodi, A., and Bashash, D. (2022). The PI3K/Akt/mTOR pathway in lung cancer; oncogenic alterations, therapeutic opportunities, challenges, and a glance at the application of nanoparticles. *Transl. Oncol.* 18, 101364. doi:10.1016/j.tranon.2022.101364
- Saraf, S., Jain, A., Tiwari, A., Verma, A., Panda, P. K., and Jain, S. K. (2020). Advances in liposomal drug delivery to cancer: An overview. *J. Drug Deliv. Sci. Technol.* 56, 101549. doi:10.1016/j.jddst.2020.101549
- Saravanakumar, K., Chelliah, R., Mubarakali, D., Oh, D.-H., Kathiresan, K., and Wang, M.-H. (2019). Unveiling the potentials of biocompatible silver nanoparticles on human lung carcinoma A549 cells and *Helicobacter pylori*. *Sci. Rep.* 9, 5787–5788. doi:10.1038/s41598-019-42112-1
- Saxena, J., Bisen, M., Misra, A., Srivastava, V. K., Kaushik, S., Siddiqui, A. J., et al. (2022). Targeting COPD with PLGA-based nanoparticles: Current status and prospects. *BioMed Res. Int.* 2022, 5058121. doi:10.1155/2022/5058121
- Scoville, D. K., Nolin, J. D., Ogden, H. L., An, D., Afsharinejad, Z., Johnson, B. W., et al. (2019). Quantum dots and mouse strain influence house dust mite-induced allergic airway disease. *Toxicol. Appl. Pharmacol.* 368, 55–62. doi:10.1016/j.taap.2019.01.018
- Scoville, D. K., White, C. C., Botta, D., An, D., Afsharinejad, Z., Bammler, T. K., et al. (2018). Quantum dot induced acute changes in lung mechanics are mouse strain dependent. *Inhal. Toxicol.* 30, 397–403. doi:10.1080/08958378.2018.1542046
- Seo, J., Lee, C., Hwang, H. S., Kim, B., Lee, E. S., Oh, K. T., et al. (2016). Therapeutic advantage of inhaled tacrolimus-bound albumin nanoparticles in a bleomycin-induced pulmonary fibrosis mouse model. *Pulm. Pharmacol. Ther.* 36, 53–61. doi:10.1016/j.pupt.2016.01.001
- Shakerimanesh, K., Bayat, F., Shahrokh, A., Baradaran, A., Yousefi, E., Mashreghi, M., et al. (2022). Biomimetic synthesis and characterization of homogenous gold nanoparticles and estimation of its cytotoxicity against breast cancer cell line. *Mater. Technol.* 37, 2853–2860. doi:10.1080/10667857.2022.2081287
- Sham, J. O.-H., Zhang, Y., Finlay, W. H., Roa, W. H., and Löbenberg, R. (2004). Formulation and characterization of spray-dried powders containing nanoparticles for

- aerosol delivery to the lung. *Int. J. Pharm.* 269, 457–467. doi:10.1016/j.ijpharm.2003.09.041
- Sheikhpour, M., Sadeghizadeh, M., Yazdian, F., Mansoori, A., Asadi, H., Movafagh, A., et al. (2020). Co-administration of curcumin and bromocriptine nano-liposomes for induction of apoptosis in lung cancer cells. *Iran. Biomed. J.* 24, 24–29. doi:10.29252/ibj.24.1.24
- Shih, C.-Y., Huang, W.-L., Chiang, I.-T., Su, W.-C., and Teng, H. (2021). Biocompatible hole scavenger-assisted graphene oxide dots for photodynamic cancer therapy. *Nanoscale* 13, 8431–8441. doi:10.1039/d1nr01476e
- Shivaji, K., Mani, S., Ponnuragan, P., De Castro, C. S., Lloyd Davies, M., Balasubramanian, M. G., et al. (2018). Green-synthesis-derived CdS quantum dots using tea leaf extract: Antimicrobial, bioimaging, and therapeutic applications in lung cancer cells. *ACS Appl. Nano Mater.* 1, 1683–1693. doi:10.1021/acsnm.8b00147
- Singh, N., Sachdev, A., and Gopinath, P. (2018a). Polysaccharide functionalized single walled carbon nanotubes as nanocarriers for delivery of curcumin in lung cancer cells. *J. Nanosci. Nanotechnol.* 18, 1534–1541. doi:10.1166/jnn.2018.14222
- Singh, R. P., Gangadharappa, H., and Mruthunjaya, K. (2018b). Phytosome complexed with chitosan for gingerol delivery in the treatment of respiratory infection: *In vitro* and *in vivo* evaluation. *Eur. J. Pharm. Sci.* 122, 214–229. doi:10.1016/j.ejps.2018.06.028
- Song, H., Guo, T., Zhao, Z., Wei, Y., Luo, H., Weng, W., et al. (2018). Biocompatible PEGylated Gold nanorods function as cytokinesis inhibitors to suppress angiogenesis. *Biomaterials* 178, 23–35. doi:10.1016/j.biomaterials.2018.06.006
- Sonkusre, P., and Cameotra, S. S. (2017). Biogenic selenium nanoparticles induce ROS-mediated necroptosis in PC-3 cancer cells through TNF activation. *J. Nanobiotechnology* 15, 43–12. doi:10.1186/s12951-017-0276-3
- Stan, M. S., Memet, I., Sima, C., Popescu, T., Teodorescu, V. S., Hermenean, A., et al. (2014). Si/SiO<sub>2</sub> quantum dots cause cytotoxicity in lung cells through redox homeostasis imbalance. *Chemico-biological Interact.* 220, 102–115. doi:10.1016/j.cbi.2014.06.020
- Steinman, R. M. (2007). Lasker basic medical research award. Dendritic cells: Versatile controllers of the immune system. *Nat. Med.* 13, 1155–1159. doi:10.1038/nm1643
- Sun, M., Shi, Y., Dang, U. J., and Di Pasqua, A. J. (2019). Phenethyl isothiocyanate and cisplatin co-encapsulated in a liposomal nanoparticle for treatment of non-small cell lung cancer. *Molecules* 24, 801. doi:10.3390/molecules24040801
- Sur, S., Rathore, A., Dave, V., Reddy, K. R., Chouhan, R. S., and Sadhu, V. (2019). Recent developments in functionalized polymer nanoparticles for efficient drug delivery system. *Nano-Structures Nano-Objects* 20, 100397. doi:10.1016/j.nanos.2019.100397
- Taghavizadeh Yazdi, M. E., Amiri, M. S., Nourbakhsh, F., Rahnama, M., Forouzanfar, F., and Mousavi, S. H. (2021). Bio-indicators in cadmium toxicity: Role of HSP27 and HSP70. *Environ. Sci. Pollut. Res.* 28, 26359–26379. doi:10.1007/s11356-021-13687-y
- Thangamani, S., Mahendran, H. P., Bhattacharjee, R. R., Jeyaraj, S., and Mohanta, K. (2021). Assessment of dihydropyrimidinone-based nanocomposites as multifunctional anti-cancer drug. *Mater. Adv.* 2, 3385–3393.
- Thorlund, K., Dron, L., Park, J., Hsu, G., Forrest, J. I., and Mills, E. J. (2020). A real-time dashboard of clinical trials for COVID-19. *Lancet Digital Health* 2, e286–e287. doi:10.1016/S2589-7500(20)30086-8
- Tran, Q. H., and Le, A.-T. (2013). Silver nanoparticles: Synthesis, properties, toxicology, applications and perspectives. *Adv. Nat. Sci. Nanosci. Nanotechnol.* 4, 033001. doi:10.1088/2043-6262/4/3/033001
- Tsuda, A., Henry, F. S., and Butler, J. P. (2013). Particle transport and deposition: Basic physics of particle kinetics. *Compr. Physiol.* 3, 1437–1471. doi:10.1002/cphy.c100085
- Vahideh Mohammadzadeh, M. B., Sadegh Amiri, M., Yazdi, M. E. T., Hassanisaadi, M., Rahdar, A., Rajender, S., et al. (2022). Applications of plant-based nanoparticles in nanomedicine. *A Sustain. Chem. Pharm.* 25, 100606. doi:10.1016/j.scp.2022.100606
- Vartak, R., Patil, S. M., Saraswat, A., Patki, M., Kunda, N. K., and Patel, K. (2021). Aerosolized nanoliposomal carrier of remdesivir: An effective alternative for COVID-19 treatment *in vitro*. *Nanomedicine* 16, 1187–1202. doi:10.2217/nnm-2020-0475
- Videira, M., Almeida, A. J., and Fabra, A. (2012). Preclinical evaluation of a pulmonary delivered paclitaxel-loaded lipid nanocarrier antitumor effect. *Nanomedicine Nanotechnol. Biol. Med.* 8, 1208–1215. doi:10.1016/j.nano.2011.12.007
- Vishali, D., Monisha, J., Sivakamasundari, S., Moses, J., and Anandharamakrishnan, C. (2019). Spray freeze drying: Emerging applications in drug delivery. *J. Control. Release* 300, 93–101. doi:10.1016/j.jconrel.2019.02.044
- Vyas, S. P., and Goswami, R. (2019). Size-dependent cellular uptake and TLR4 attenuation by gold nanoparticles in lung adenocarcinoma cells. *Nanomedicine* 14, 229–253. doi:10.2217/nnm-2018-0266
- Wan, R., Mo, Y., Zhang, Z., Jiang, M., Tang, S., and Zhang, Q. (2017). Cobalt nanoparticles induce lung injury, DNA damage and mutations in mice. *Part. Fibre Toxicol.* 14, 38–15. doi:10.1186/s12989-017-0219-z
- Wan, X., Song, Y., Song, N., Li, J., Yang, L., Li, Y., et al. (2016). The preliminary study of immune superparamagnetic iron oxide nanoparticles for the detection of lung cancer in magnetic resonance imaging. *Carbohydr. Res.* 419, 33–40. doi:10.1016/j.carres.2015.11.003
- Wang, H., Huang, Q., Chang, H., Xiao, J., and Cheng, Y. (2016). Stimuli-responsive dendrimers in drug delivery. *Biomaterials Sci.* 4, 375–390. doi:10.1039/c5bm00532a
- Wang, H., Zhao, X., Guo, C., Ren, D., Zhao, Y., Xiao, W., et al. (2015a). Aptamer-dendrimer bioconjugates for targeted delivery of miR-34a expressing plasmid and antitumor effects in non-small cell lung cancer cells. *PLoS One* 10, e0139136. doi:10.1371/journal.pone.0139136
- Wang, W., Han, Q., Xu, X., and Wen, C. (2022). miR-146 relieves acute asthma via decreasing epidermal growth factor receptor/toll-like receptor 4 (EGFR/TLR4) and enhancing autophagy. *J. Biomaterials Tissue Eng.* 12, 2030–2037. doi:10.1166/jbt.2022.3144
- Wang, X., Zhang, H., Jing, H., and Cui, L. (2015b). Highly efficient labeling of human lung cancer cells using cationic poly-L-lysine-assisted magnetic iron oxide nanoparticles. *Nano-Micro Lett.* 7, 374–384. doi:10.1007/s40820-015-0053-5
- Wang, Y., Cui, H., Zhou, J., Li, F., Wang, J., Chen, M., et al. (2015c). Cytotoxicity, DNA damage, and apoptosis induced by titanium dioxide nanoparticles in human non-small cell lung cancer A549 cells. *Environ. Sci. Pollut. Res.* 22, 5519–5530. doi:10.1007/s11356-014-3717-7
- Wang, Y., Zhang, Y., Du, Z., Wu, M., and Zhang, G. (2012). Detection of micrometastases in lung cancer with magnetic nanoparticles and quantum dots. *Int. J. Nanomedicine* 7, 2315–2324. doi:10.2147/IJN.S30593
- Wittgen, B. P., Kunst, P. W., Van Der Born, K., Van Wijk, A. W., Perkins, W., Pilkievicz, F. G., et al. (2007). Phase I study of aerosolized SLIT cisplatin in the treatment of patients with carcinoma of the lung. *Clin. Cancer Res.* 13, 2414–2421. doi:10.1158/1078-0432.CCR-06-1480
- Wu, J., Zhai, T., Sun, J., Yu, Q., Feng, Y., Li, R., et al. (2022). Mucus-permeable polymyxin B-hyaluronic acid/poly (lactic-co-glycolic acid) nanoparticle platform for the nebulized treatment of lung infections. *J. Colloid Interface Sci.* 624, 307–319. doi:10.1016/j.jcis.2022.05.121
- Xiao, X., Zeng, X., Zhang, X., Ma, L., Liu, X., Yu, H., et al. (2013). Effects of *Caryota mitis* profilin-loaded PLGA nanoparticles in a murine model of allergic asthma. *Int. J. Nanomedicine* 8, 4553–4562. doi:10.2147/IJN.S51633
- Xie, C., Zhan, Y., Wang, P., Zhang, B., and Zhang, Y. (2020). Novel surface modification of ZnO QDs for paclitaxel-targeted drug delivery for lung cancer treatment. *Dose-Response* 18, 1559325820926739. doi:10.1177/1559325820926739
- Xu, L., Wang, H., Xiao, W., Zhang, W., Stewart, C., Huang, H., et al. (2023). PAMAM dendrimer-based tongue rapidly identifies multiple antibiotics. *Sensors Actuators B Chem.* 382, 133519. doi:10.1016/j.snb.2023.133519
- Yang, B., Choi, H., Kim, S.-H., Yoon, H. J., and Lee, H. (2019). How will nanotechnology lead to better control of asthma? *Ann. Transl. Med.* 7, 515. doi:10.21037/atm.2019.09.129
- Yang, Y., Peng, F., Wang, R., Guan, K., Jiang, T., Xu, G., et al. (2020). The deadly coronaviruses: The 2003 SARS pandemic and the 2020 novel coronavirus epidemic in China. *J. Autoimmun.* 109, 102434. doi:10.1016/j.jaut.2020.102434
- Yazdi, M. E., Amiri, M. S., and Darroudi, M. (2020). “Biopolymers in the synthesis of different nanostructures,” in *Reference module in materials science and materials engineering* (Amsterdam, Netherlands: Elsevier).
- Yazdi, M. E. T., Nourbakhsh, F., Mashreghi, M., and Mousavi, S. H. (2021). Ultrasound-based synthesis of ZnO·Ag<sub>2</sub>O<sub>3</sub> nanocomposite: Characterization and evaluation of its antimicrobial and anticancer properties. *Res. Chem. Intermed.* 47, 1285–1296. doi:10.1007/s11164-020-04355-w
- Yong, D. O. C., Saker, S. R., Wadhwa, R., Chellappan, D. K., Madheswaran, T., Panneerselvam, J., et al. (2019). Preparation, characterization and *in-vitro* efficacy of quercetin loaded liquid crystalline nanoparticles for the treatment of asthma. *J. Drug Deliv. Sci. Technol.* 54, 101297. doi:10.1016/j.jddst.2019.101297
- Yousefi, M., Narmani, A., and Jafari, S. M. (2020). Dendrimers as efficient nanocarriers for the protection and delivery of bioactive phytochemicals. *Adv. colloid interface Sci.* 278, 102125. doi:10.1016/j.cis.2020.102125
- Yu, T.-T., Peng, X.-C., Wang, M.-F., Han, N., Xu, H.-Z., Li, Q.-R., et al. (2022). Harnessing chlorin e6 loaded by functionalized iron oxide nanoparticles linked with glucose for target photodynamic therapy and improving of the immunogenicity of lung cancer. *J. Cancer Res. Clin. Oncol.* 148, 867–879. doi:10.1007/s00432-021-03879-x
- Zarei, M., Karimi, E., Oskoueian, E., Es-Haghi, A., and Yazdi, M. E. T. (2021). Comparative study on the biological effects of sodium citrate-based and apigenin-based synthesized silver nanoparticles. *Nutr. Cancer* 73, 1511–1519. doi:10.1080/01635581.2020.1801780
- Zarharan, H., Bagherian, M., Rokhi, A. S., Bajgirani, R. R., Yousefi, E., Heravian, P., et al. (2023). The anti-angiogenesis and antioxidant activity of chitosan-mediated synthesized selenium-gold nanostructure. *Arabian J. Chem.* 16, 104806. doi:10.1016/j.arabj.2023.104806
- Zenze, M., Daniels, A., and Singh, M. (2023). Dendrimers as modifiers of inorganic nanoparticles for therapeutic delivery in cancer. *Pharmaceutics* 15, 398. doi:10.3390/pharmaceutics15020398

- Zhang, F., Zhuang, J., Li, Z., Gong, H., De Ávila, B. E.-F., Duan, Y., et al. (2022a). Nanoparticle-modified microrobots for *in vivo* antibiotic delivery to treat acute bacterial pneumonia. *Nat. Mater.* 21, 1324–1332. doi:10.1038/s41563-022-01360-9
- Zhang, H., Cai, L., Su, C., Huang, X., and Tong, Y. (2023). Role of autophagy in the regulation of respiratory inflammation caused by zinc oxide nanoparticles.
- Zhang, H., Liang, Z., Zhang, J., Wang, W.-P., Zhang, H., and Lu, Q. (2020a). Zinc oxide nanoparticle synthesized from *Euphorbia fischeriana* root inhibits the cancer cell growth through modulation of apoptotic signaling pathways in lung cancer cells. *Arabian J. Chem.* 13, 6174–6183. doi:10.1016/j.arabjc.2020.05.020
- Zhang, M., Jiang, H., Wu, L., Lu, H., Bera, H., Zhao, X., et al. (2022b). Airway epithelial cell-specific delivery of lipid nanoparticles loading siRNA for asthma treatment. *J. Control. Release* 352, 422–437. doi:10.1016/j.jconrel.2022.10.020
- Zhang, M., Li, M., Du, L., Zeng, J., Yao, T., and Jin, Y. (2020b). Paclitaxel-in-liposome-in-bacteria for inhalation treatment of primary lung cancer. *Int. J. Pharm.* 578, 119177. doi:10.1016/j.ijpharm.2020.119177
- Zhang, M., Wang, Q., Xu, Y., Guo, L., Lai, Z., and Li, Z. (2020c). Graphitic carbon nitride quantum dots as analytical probe for viewing sialic acid on the surface of cells and tissues. *Anal. Chim. Acta* 1095, 204–211. doi:10.1016/j.aca.2019.10.031
- Zhang, Q., Liu, Q., Du, M., Vermorken, A., Cui, Y., Zhang, L., et al. (2019). Cetuximab and Doxorubicin loaded dextran-coated Fe<sub>3</sub>O<sub>4</sub> magnetic nanoparticles as novel targeted nanocarriers for non-small cell lung cancer. *J. Magnetism Magnetic Mater.* 481, 122–128. doi:10.1016/j.jmmm.2019.01.021
- Zhang, X.-Q., Xu, X., Lam, R., Giljohann, D., Ho, D., and Mirkin, C. A. (2011). Strategy for increasing drug solubility and efficacy through covalent attachment to polyvalent DNA–nanoparticle conjugates. *ACS nano* 5, 6962–6970. doi:10.1021/nn201446c
- Zhao, H., Mu, X., Zhang, X., and You, Q. (2020). Lung cancer inhibition by betulinic acid nanoparticles via adenosine 5'-triphosphate (ATP)-binding cassette transporter G1 gene downregulation. *Med. Sci. Monit. Int. Med. J. Exp. Clin. Res.* 26, e922092–e922091. doi:10.12659/MSM.922092
- Zhao, X., Wei, S., Li, Z., Lin, C., Zhu, Z., Sun, D., et al. (2019). Autophagic flux blockage in alveolar epithelial cells is essential in silica nanoparticle-induced pulmonary fibrosis. *Cell death Dis.* 10, 127–216. doi:10.1038/s41419-019-1340-8
- Zheng, Y., Zhang, J., Zhang, R., Luo, Z., Wang, C., and Shi, S. (2019). Gold nano particles synthesized from *Magnolia officinalis* and anticancer activity in A549 lung cancer cells. *Artif. cells, nanomedicine, Biotechnol.* 47, 3101–3109. doi:10.1080/21691401.2019.1645152
- Zhong, L., Hu, X., Cao, Z., Wang, H., Chen, Y., and Lian, H.-Z. (2019). Aggregation and dissolution of engineering nano Ag and ZnO pretreated with natural organic matters in the simulated lung biological fluids. *Chemosphere* 225, 668–677. doi:10.1016/j.chemosphere.2019.03.080
- Zhong, Q., Humia, B. V., Punjabi, A. R., Padilha, F. F., and Da Rocha, S. R. (2017). The interaction of dendrimer-doxorubicin conjugates with a model pulmonary epithelium and their cosolvent-free, pseudo-solution formulations in pressurized metered-dose inhalers. *Eur. J. Pharm. Sci.* 109, 86–95. doi:10.1016/j.ejps.2017.07.030
- Zhou, L., Li, P., Zhang, M., Han, B., Chu, C., Su, X., et al. (2020). Carbon black nanoparticles induce pulmonary fibrosis through NLRP3 inflammasome pathway modulated by miR-96 targeted FOXO3a. *Chemosphere* 241, 125075. doi:10.1016/j.chemosphere.2019.125075
- Zhou, Y., Lü, H., Zhang, D., Xu, K., Hui, N., and Wang, J. (2023). Electrochemical biosensors based on conducting polymer composite and PAMAM dendrimer for the ultrasensitive detection of acetamiprid in vegetables. *Microchem. J.* 185, 108284. doi:10.1016/j.microc.2022.108284
- Zylberberg, C., and Matosevic, S. (2016). Pharmaceutical liposomal drug delivery: A review of new delivery systems and a look at the regulatory landscape. *Drug Deliv.* 23, 3319–3329. doi:10.1080/10717544.2016.1177136

## Glossary

<b>ABCG1</b>	ATP-binding cassette and transporter G1	<b>MMP-9</b>	Matrix metalloproteinase-9
<b>AG</b>	Andrographolide	<b>MPO</b>	Myeloperoxidase
<b>AgNPs</b>	Silver nanoparticles	<b>NF-<math>\kappa</math>B</b>	Nuclear factor kappa B
<b>AHR</b>	Airways responsiveness	<b>NiO NPs</b>	Nickel oxide nanoparticles
<b>AQ</b>	Amodiaquine	<b>NPs</b>	Nanoparticles
<b>AuNPs</b>	Gold nanoparticles	<b>OCR</b>	Rate of oxygen consumption
<b>BALF</b>	Bronchoalveolar lavage fluid	<b>PAMAM</b>	Poly(amidoamine) dendrimer
<b>Bcl-2</b>	B-cell lymphoma 2	<b>PdNPs</b>	Palladium nanoparticles
<b>BDQ</b>	Bedaquiline	<b>PEITC</b>	Phenethyl isothiocyanate
<b>BRNPS</b>	Bilirubin-based NPs	<b>PF</b>	Pulmonary fibrosis
<b>CAP</b>	Cold atmospheric plasma	<b>PG</b>	Gingerol
<b>CDDP</b>	Cisplatin	<b>QDs</b>	Quantum dots
<b>CdO NPs</b>	Cadmium oxide nanoparticles	<b>rCmP</b>	recombinant <i>Caryota mitis</i> profilin
<b>CdSe</b>	Cadmium selenide	<b>SARS-CoV</b>	Severe acute respiratory syndrome coronavirus
<b>COPD</b>	Chronic obstructive pulmonary disease	<b>SBS</b>	Salbutamol sulfate
<b>CS</b>	Chitosan	<b>Si/sio<sub>2</sub></b>	Silicon dioxide
<b>CuONPs</b>	Copper oxide nanoparticles	<b>SiNPs</b>	Silica nanoparticles
<b>Cur-NPs</b>	Curcumin-nanoparticles	<b>siRNA</b>	Small-interfering RNA
<b>CXCL1</b>	Chemokine (C-X-C motif) ligand 1	<b>SLIT</b>	Sublingual
<b>DOX</b>	Doxorubicin	<b>SLN</b>	solid lipid nanoparticles
<b>ECM</b>	Extracellular matrix	<b>SM-LCN</b>	Surface-modified liquid crystalline NPs
<b>EGFR</b>	Epidermal growth factor receptor	<b>SPIONS</b>	Superparamagnetic iron oxide nanoparticles
<b>FA</b>	Ferulic acid	<b>TAC</b>	Tacrolimus
<b>FACS</b>	Fluorescence-activated cell sorting	<b>TLR4</b>	Toll-like receptor 4
<b>GEB</b>	Gefitinib	<b>TNF-<math>\alpha</math></b>	Tumor necrosis factor-alpha
<b>GSH</b>	Glutathione	<b>Tob</b>	Tobramycin
<b>HA</b>	Hyaluronic acid	<b>TQ</b>	Thymoquinone
<b>HIF-1<math>\alpha</math></b>	Hypoxia-inducible factor 1	<b>UCB</b>	Unconjugated bilirubin
<b>IFN-<math>\gamma</math></b>	Interferon- $\gamma$	<b>VEGF</b>	Vascular endothelial growth factor
<b>IL-1<math>\beta</math></b>	Interleukin-1 $\beta$	<b>VIP</b>	Vasoactive intestinal peptide
<b>IL-6</b>	Interleukin-6	<b>VWF</b>	von Willebrand factor
<b>IL-8</b>	Interleukin-8	<b>Zns</b>	Zinc sulfide
<b>INF-<math>\gamma</math></b>	Interferon-gamma	<b><math>\alpha</math>-AN</b>	$\alpha$ -Alumina
<b>KC</b>	Chemokine		
<b>LCN</b>	Liquid crystalline NPs		
<b>LPB</b>	Liposome-in-bacteria		
<b>LPS</b>	Lipopolysaccharide		
<b>MCP-1</b>	Monocyte chemoattractant protein-1		
<b>MERS-CoV</b>	Middle East respiratory syndrome coronavirus		
<b>MMP</b>	Mitochondrial membrane potential		