Check for updates

OPEN ACCESS

EDITED BY David Sacerdoti, University of Verona, Italy

REVIEWED BY Anna Moles, Newcastle University, United Kingdom Jerzy Beltowski, Medical University of Lublin, Poland

*CORRESPONDENCE Lei Li, ⊠ m13699119545@163.com

[†]These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Gastrointestinal and Hepatic Pharmacology, a section of the journal Frontiers in Pharmacology

RECEIVED 21 March 2022 ACCEPTED 25 November 2022 PUBLISHED 16 December 2022

CITATION

Liu B, Wang S, Xu M, Ma Y, Sun R, Ding H and Li L (2022), The double-edged role of hydrogen sulfide in the pathomechanism of multiple liver diseases. *Front. Pharmacol.* 13:899859. doi: 10.3389/fphar.2022.899859

COPYRIGHT

© 2022 Liu, Wang, Xu, Ma, Sun, Ding and Li. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does

not comply with these terms.

The double-edged role of hydrogen sulfide in the pathomechanism of multiple liver diseases

Bihan Liu^{1†}, Shanshan Wang^{1,2†}, Ming Xu^{3,4}, Yanan Ma¹, Rui Sun¹, Huiguo Ding¹ and Lei Li^{1*}

¹Department of Hepatology and Gastroenterology, Beijing Youan Hospital, Capital Medical University, Beijing, China, ²Beijing Institute of Hepatology, Beijing Youan Hospital, Capital Medical University, Beijing, China, ³Brainnetome Center and National Laboratory of Pattern Recognition, Institute of Automation, Chinese Academy of Sciences, Beijing, China, ⁴School of Artificial Intelligence, University of Chinese Academy of Sciences, Beijing, China

In mammalian systems, hydrogen sulfide (H_2S) —one of the three known gaseous signaling molecules in mammals—has been found to have a variety of physiological functions. Existing studies have demonstrated that endogenous H_2S is produced through enzymatic and non-enzymatic pathways. The liver is the body's largest solid organ and is essential for H_2S synthesis and elimination. Mounting evidence suggests H_2S has essential roles in various aspects of liver physiological processes and pathological conditions, such as hepatic lipid metabolism, liver fibrosis, liver ischemia—reperfusion injury, hepatocellular carcinoma, hepatotoxicity, and acute liver failure. In this review, we discuss the functions and underlying molecular mechanisms of H_2S in multiple liver pathophysiological conditions.

KEYWORDS

hydrogen sulfide, liver fibrosis, liver ischemia-reperfusion injury, hepatocellular carcinoma, acute liver failure, non-alcoholic fatty liver disease

Introduction

Hydrogen sulfide (H₂S) is one of the three recognized gaseous signaling molecules in mammals (Yang et al., 2019b). Existing studies have demonstrated that endogenous H₂S is created in mammalian systems by enzymatic and non-enzymatic mechanisms (Yang et al., 2019b; Loiselle et al., 2020). The enzymatic delivery pathways in mammalian cells and tissues are those related to cystathionine gamma-lyase (CSE) (Jia et al., 2022), cystathionine β -synthase (CBS) (Roy et al., 2012; Zuhra et al., 2020a), and 3-mercaptopyruvate sulfotransferase (3-MST) (Roy et al., 2012; Abdollahi Govar et al., 2020; Dilek et al., 2020) (Figure 1). Moreover, some microorganisms located in the intestine are also capable of producing H₂S (Loiselle et al., 2020; Scammahorn et al., 2021).

Furthermore, hepatic stellate cells can boost H_2S production (Ma et al., 2022). Mounting evidence shows that H_2S plays a critical role in a variety of physiological and pathological processes, including the respiratory, cardiovascular, gastrointestinal, and central nervous systems, as well as in the kidneys (Shackelford et al., 2021) and in the inflammatory response and antioxidant defense systems (Shi et al., 2019).

The liver is the body's largest organ, and it plays an important role in lipid, glucose, and xenobiotic metabolism and in resistance to oxidative stress and defense against invading microorganisms (Norris et al., 2011; Andrade et al., 2019; Chen et al., 2019). It is also important for H₂S synthesis and removal (Dilek et al., 2020). Moreover, CSE, CBS, and 3-MST, which are responsible for H₂S production to varying degrees, have been found in the liver (Bełtowski, 2019). H₂S metabolism in the liver is linked to insulin sensitivity, glucose metabolism, lipoprotein production, and mitochondrial bioenergetics and biogenesis (Sun et al., 2021). Most importantly, H₂S synthesis and signal transmission in the liver are disrupted in a variety of liver disorders, such as non-alcoholic fatty liver disease (NAFLD), liver fibrosis, hepatic ischemia/reperfusion (I/R) injury, and liver cancer (Loiselle et al., 2020).

In recent decades, studies have focused on the importance of H_2S in liver growth and development, and the relationship between H_2S and both liver functions and diseases have partially been explored; thus, the underlying mechanisms of H_2S -mediated hepatoprotection or injury have been gradually revealed. For this article, we retrospectively reviewed recent studies of H_2S in various liver diseases and found that H_2S not only has a positive protective effect against liver disease but also has a non-negligible pathogenic role in certain liver diseases. In particular, we highlight the mechanisms by which H_2S is involved in these metabolic processes and relevant therapeutic targets.

The effects of H₂S in non-alcoholic fatty liver disease

H₂S acts as an essential novel regulator in lipid metabolism and plays a role in many diseases, such as diabetes, obesity, and cardiovascular disorders (Wu et al., 2019; Corvino et al., 2021; Zhao et al., 2022). The liver is an important organ for lipid metabolism in the body and is responsible for the accumulation, storage, and consumption of lipids (Stefan et al., 2019). Researchers have recently given special attention to H₂S and the control of lipid metabolism in the liver-a relationship which has important consequences for the progression of liver disease. Non-alcoholic fatty liver disease (NAFLD) is a broad class of liver diseases that range from steatosis to the more severe form of nonalcoholic steatohepatitis (NASH), a condition that can aggravate liver fibrosis and liver cirrhosis (Abd El-Kader and El-Den Ashmawy, 2015). NASH is now considered a common chronic liver disease that is present in 25% of the global population (Wu et al., 2020).

The involvement of H_2S in liver health has been explored in many studies. A previous study on NAFLD showed that

endogenous H2S levels in hepatocytes of oleic acid-treated mice were lower than the levels in an untreated group. Next, the authors investigated the effect of exogenous H2S on cell growth in the oleic acid group. The results showed that H₂S slowed liver lipid deposition through the activation of farnesol X receptors and increased the proliferation and survival of oleic acid-treated cells (Ruan et al., 2019; Loiselle et al., 2020; Xu et al., 2022). In addition, oleic acid treatment resulted in the arrest of hepatocytes in the G1 phase of the cell cycle-a process which was then reversed by H₂S (Loiselle et al., 2020; Xu et al., 2022). Related experimental results demonstrated that H₂S could promote autophagy and inhibit apoptosis in human hepatocytes through the ROS/PI3K/AKT/mTOR pathway that is mediated by reactive oxygen species (ROS), thereby alleviating the high-fat diet (HFD)-induced NAFLD (Wu et al., 2020). This research shows that H₂S could be applied to treat NAFLD. It is also worth noting that exogenous NaHS can alleviate lipid accumulation and that the 3-MST knockdown has been shown to significantly improve hepatic steatosis in high-fat diet-fed mice (Li et al., 2018). Therefore, further research is needed to fully understand the impact of H₂S on NAFLD.

However, *in vitro* systems cannot accurately simulate the natural *in vivo* physiological environment; therefore, research has increasingly focused on *in vivo* models, which are believed to provide various types of vital evidence for the relationship between H_2S and lipid metabolism that cannot be realized *in vitro*. Various knockout (KO) animal models, including CBS-KO, CSE-KO, and 3-MST-KO mouse models, have revealed critical roles of H_2S -producing enzymes in hepatic lipid metabolism (Loiselle et al., 2020).

A study that used a rat fatty liver ischemia/reperfusion injury (IRI) model revealed that the homozygous CBS mutant mice died within 5 weeks of birth, and further histological examination revealed an enlarged and lipid droplet-filled liver (Zhang et al., 2012). Similar results were shown in another study. Compared to the control group, the CBS-deficient mice had more lipid accumulation in the liver (Li et al., 2018). These results suggested that fatty acids reduced endogenous H₂S levels by inhibiting the CSE-dependent pathway in the liver, which would promote fat accumulation and subsequently lead to NAFLD (Mani et al., 2015). S Mani noted that, in comparison to the wild-type mice, the CSE-KO mice had much higher cholesterol levels in the plasma and liver when fed a HFD. Dyslipidemia, microvascular fat accumulation, changes in liver pigments, and hepatic damage were all observed in the mice. Finally, the CSE-KO mice that were fed a HFD developed a fatty liver (Roehlen et al., 2020).

In conclusion, the effects and mechanisms of H_2S in hepatic lipid metabolism have become increasingly clear. H_2S and its synthase have important protective effects on hepatic lipid metabolism, significantly improving hepatic lipid deposition. Otherwise, the level of H_2S is reduced, which increases lipid deposition and promotes the development of NAFLD. To date, the precise mechanism and clinical significance of H_2S and its synthase have yet to be elucidated.

The effects of H₂S in liver fibrosis

In developed countries, death from fibrotic diseases including chronic kidney disease, liver cirrhosis, idiopathic pulmonary fibrosis, and chronic disease accounts for 45% of human mortality, posing a serious danger to health worldwide (Zou et al., 2009; Song et al., 2015; Ni et al., 2018). Liver fibrosis accounts for a large proportion of these fibrotic diseases. The persistent buildup of extracellular matrix (ECM) over type I collagen is the primary cause of liver fibrosis (Lambrecht et al., 2015). Many molecules are involved in its development, with inflammation and oxidative stress being well-known regulatory targets. In many chronic liver illnesses, such as viral hepatitis, NASH, and NAFLD, liver fibrosis is an unavoidable pathological process (Song et al., 2015). A growing number of studies are demonstrating that the suppression of endogenous H₂S might be related to the progression of fibrosis in humans and that H₂S supplementation has protective and therapeutic effects against fibroproliferative diseases and syndromes of common organs (liver, lung, kidney, and heart), mainly due to its antiinflammatory, antioxidant, and antifibrotic effects (Kabil et al., 2014; Pant et al., 2016; Bai et al., 2019).

H₂S may affect hepatic fibrosis development in two ways (Mohammed et al., 2021): 1) H₂S has been shown to act as an antifibrotic molecule by significantly reducing the levels of TNF- $\alpha,$ IL-1 $\beta,$ IL-6, and soluble intercellular adhesion molecule (ICAM)-1 in the rat serum to suppress the inflammatory response (Tan et al., 2011; Mohammed et al., 2021) and 2) it exerts antioxidant effects by increasing the activity and expression of the catalase, copper-zinc superoxide dismutase and manganese superoxide dismutase, thereby effectively inhibiting the progression of fibrosis (Jung et al., 2013). An in vivo study found that S-allyl-cysteine (SAC), an endogenous provider of H₂S, could relieve CCl4-induced liver fibrosis in rats by inhibiting the STAT3/SMAD3 pathway (Gong et al., 2018). Further experimental studies have shown that SAC treatment reduces the expression of both inflammatory factors and fibrogenic cytokines and increases the expression of antioxidant enzymes (Kodai et al., 2007). H₂S has been shown to induce cell cycle arrest, apoptosis, and vasodilation by activating p53, p21, caspase-3, and MMP-2, by promoting their overexpression and by downregulating the Bcl-xL expression (Ma et al., 2018). H₂S also ameliorates liver fibrosis through its anti-inflammatory and antioxidant properties, thereby alleviating portal hypertension (Zou et al., 2009; Gong et al., 2018; Ma et al., 2018; Damba et al., 2019; Ali et al., 2020).

However, there are several factors associated with H₂S that contribute to the formation of liver fibrosis. A perspective, taking

this aspect into account was proposed by C.G. Zou et al., who found that the precursor of H_2S synthesis, homocysteine, enhances the activation of human hepatic stellate cells by activating the PI3K/Akt signaling pathway (Ali et al., 2020). The T. Damba team further verified this hypothesis. They found that both endogenous and exogenous H_2S can increase the proliferation and activation of hepatic stellate cells by increasing the glycolysis extracellular acidification rate (ECAR) and the oxygen consumption rate (OCR) of mitochondrial oxidative phosphorylation, thus promoting the metabolic activity of hepatic stellate cells (Jiménez-Castro et al., 2019) and further promoting liver fibrosis formation.

These various reports show that additional studies investigating the importance of H_2S in liver fibrosis and H_2S chemical pathways are necessary and that future works should include confirmation with animal experiments and cellular studies.

The effects of H₂S on liver ischemia– reperfusion injury

Ischemia-reperfusion (I/R) is a well-known pathological condition marked by a temporary decrease in blood supply to an organ or region, followed by vascular recovery and downstream tissue damage (Yang et al., 2018b). It is a consequence of hemorrhagic shock and resuscitation, trauma, liver resections, liver transplantation, bowel infarction, and, especially, liver failure. Hepatic I/R damage has become a global health issue (Nastos et al., 2014). Various clinical experiments and basic studies point to diverse molecular mechanisms in this process, including those related to neutrophils and liver Kupffer cells, proinflammatory cytokines, adhesion molecules, mitochondria, oxidative stress, anaerobic metabolism, and intracellular calcium overload (Kang et al., 2009; Zhai et al., 2013). Thus, novel drugs that have antioxidative, anti-inflammatory, and cytoprotective effects may protect the liver from I/R injury (Krylatov et al., 2021). Currently, H₂S is known to be critical in the treatment of liver I/R injury (Wu et al., 2015). Due to the thorough investigation of H₂S and liver I/R injury, the mechanism by which H₂S protects against I/R injury has begun to be elucidated.

There have been significant advances in animal studies focusing on the molecular pathways of H_2S in I/R injury (Lu et al., 2018; Fu et al., 2019; Ibrahim et al., 2021). Recent studies have found that fatty livers are more susceptible to ischemia/ reperfusion (I/R) damage during liver surgery and transplantation (Varela et al., 2011). In a rat fatty liver IRI model, the influence of H_2S on IRI was thoroughly investigated. According to the findings, H_2S mitigated changes in liver pathology and lowered the levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), and malondialdehyde (MDA). Moreover, H_2S decreased oxidative stress levels and the expression of inflammatory factors and slowed the apoptosis of hepatocytes (Younis et al., 2016). In addition, treatment with silymarin protected against hepatic I/R in insulin-resistant rats through anti-inflammatory, antioxidant, and anti-apoptotic effects and the inhibition of H₂S synthesis (Lu et al., 2018; Liu et al., 2020). Therefore, retaining an appropriate level of H₂S in ischemia–reperfusion (I/R) is imperative in protecting the liver from injury. NaHS protects the liver against I/R and, as a donor of H₂S, is protective against hepatic I/R injury, a process associated with the activation of antioxidant enzymes and decreased expression of hepatic tumor necrosis factor-α (TNF-α), MDA, and caspase-3 (Fu et al., 2019).

Cold ischemia–reperfusion injury (IRI) poses a significant threat to the success of solid organ transplantation (SOT) (Muller et al., 2022). A study discussing the molecular mechanisms underlying the role of H_2S donor molecules in liver transplantation showed that H_2S could significantly attenuate IRI during liver transplantation by inhibiting a range of interrelated cells and molecules, including those related to microcirculatory dysfunction and microvascular dysfunction, mitochondrial damage, inflammatory responses, cellular injury, cell death, and other destructive molecular pathways, while promoting the protective pathways (Balaban et al., 2011; Fu et al., 2019). These promising findings will be the basis of the clinical application of H_2S in the future (Dugbartey et al., 2021).

In addition, another piece of evidence from an experimental model of organ transplantation in mice suggests that exogenous administration of H_2S donor molecules during graft preservation significantly improves liver microcirculation, morphology, and function. Moreover, a significant increase in liver antioxidant enzyme levels and activity was also observed. In contrast, lactate dehydrogenase, malondialdehyde (an indicator of lipid peroxidation byproducts and ROS production), and other markers of liver injury were significantly reduced. These new findings suggest that adding H_2S donor molecules during liver transplantation can play an important role in significantly increasing the survival rate of transplanted organs, mitigating liver IRI injury during transplantation, reducing complications, and improving patient prognosis (Balaban et al., 2015).

These findings imply that focusing on H_2S could be a promising new strategy for combating I/R-induced liver damage. The molecular targets of H_2S in liver I/R damage, however, are still unknown. Since different doses of H_2S generated by the donor may have varying therapeutic effects, the optimum dose range should be investigated further for improved efficacy.

The effect of H₂S in hepatocellular carcinoma

Liver cancer is the world's sixth most prevalent malignancy and the fourth leading cause of cancer-related death (Yang et al., 2019c; Li et al., 2021b). Among liver cancers, the most common kind is hepatocellular carcinoma (HCC). The primary risk factors for HCC include viral infection, chronic alcohol consumption, and obesityrelated NASH. HCC pathophysiology is a complex multistep process (Cao et al., 2019). Currently, HCC continues to be a global health threat, with morbidity and mortality rates increasing dramatically. As a consequence, the monitoring of HCC and early detection are regarded as vital methods to improve the effectiveness of treatment (Malik et al., 2020). Studies have revealed that angiogenesis and immune evasion are key core issues in the tumor microenvironment for liver cancer progression and treatment failure (Motz and Coukos, 2011; Fousek et al., 2021; Pinter et al., 2021; Zhou et al., 2021). In a complex tumor microenvironment composed of hepatocytes, liver sinusoidal endothelial cells (LSECs), hepatic stellate cells (HSCs), immune cells, and extracellular matrix, the development of HCC is closely related to the infiltration of immune cells and immune evasion in the tumor microenvironment (TME) (Muñoz et al., 2021). Increasing evidence has shown that H2S plays a key role in the occurrence and development of HCC. Recently, CBS was found to inhibit Treg cell infiltration and induce apoptosis in human HCC cells by suppressing the PRRX2/IL-6/STAT3 signaling pathway. CBS deficiency promoted Treg-mediated immune evasion and tumor growth in mice, suggesting that the CBS/H₂S axis may control immune evasion in the TME (Xu et al., 2021; Zhou et al., 2021). The overexpression and overactivation of the immunosuppressive enzyme indoleamine 2,3-dioxygenase 1 (IDO1) is a key mechanism of immune escape from cancer. In a mouse liver cancer model, exogenous H2S inhibited IDO1 expression by blocking the STAT3 and NF-kB pathways and reduced IDO1 activity through the H₂S/NO crosstalk, effectively stopping tumor progression in mice (Yang et al., 2019a). Furthermore, the H₂S donor effectively suppresses tumor development in mice with hepatocellular carcinoma models (Ngowi et al., 2021). These findings suggest that targeting the CBS/H2S axis might be a novel method for therapeutic immunotherapy in HCC.

In addition to immunotherapy, H_2S can also promote apoptosis of HCC cells directly (Wang et al., 2017). Our previous research revealed that exogenous H_2S could induce HCC cell autophagy and further promote apoptosis by inhibiting the PI3K/AKT signaling pathway (Wang et al., 2017). In the TME, hepatic stellate cells activate the JNK/JunB signaling pathway through the release of H_2S , upregulate the expression of TNFSF14, and promote apoptosis in hepatocellular carcinoma cells (Ma et al., 2022). A similar result was found for endogenous H_2S ; Zhou et al. found that the activation of the CBS/ H_2S axis in HCC cells upregulated the expression of cleaved caspase-3 and promoted HCC cell apoptosis (Sakuma et al., 2019).

In addition, H_2S donors are able to regulate apoptosis by interacting with signaling pathways related to apoptosis in cancer cells (Szadvari et al., 2019). Treatment with GYY4137, an H_2S donor, increased the expression of caspase-9, a marker of apoptosis, in human HCC cells (Zhao et al., 2015). Moreover, treatment with NaHS suppressed the anti-apoptotic markers in B-cell lymphoma 2 (Bcl-2) by regulating the p53 and p38 MAPK pathways (Pan et al., 2014; Zhao et al., 2015), thus promoting cancer cell apoptosis (Zuhra et al., 2020b; Xia et al., 2021). Moreover, H_2S acts as a stimulator of mitochondrial electron transfer, and endogenous H_2S stimulates adenosine triphosphate (ATP) production in cancer cells, which plays an important role in preventing mitochondrial fission and maintaining mitochondrial DNA repair (Fortibui et al., 2021).

There is evidence that metabolic reprogramming of cancer is a determinant in the anticancer-related immune response (Zhang et al., 2020). For example, sulfur-related metabolism is still a novel direction of research in human HCC tumor tissues. Recent studies that focused on gene expression profiling in HCC via Gene Set Enrichment Analysis (GSEA) revealed that sulfur amino acid metabolism in HCC was downregulated. Cell viability experiments showed that H₂S had notable anticancer effects in human HCC cells. H₂S can also provide sulfane sulfur, which mediates reactive sulfur species (RSS)-induced anti-HCC effects in tumor cells. Finally, it was shown that sulfur metabolism in HCC had been reprogrammed and a potential therapeutic strategy for HCC was proposed (Wang et al., 2020b). Acetaldehyde dehydrogenase (ALDH) is the primary enzyme in the liver that regulates acetaldehyde metabolism (Toledo-Guzmán et al., 2019). It has been shown that ALDH can alter a variety of biological features in cancer stem cells and could be used as a cancer stem cell diagnostic marker (Duan et al., 2016; Iciek et al., 2018). Evidence suggests that ALDH plays a significant role in cancer recurrence (Yang et al., 2018a). Based on these findings, it is possible that ALDH is regulated by sulfur substances, which inhibit its enzymatic activity, thus making cancer cells more sensitive to conservative treatment.

In addition, *in vitro* and *in vivo*, a combination of kelp and *Curcuma zedoaria* inhibited the growth and metastasis of liver cancer cells by reducing the generation of endogenous H₂S and regulating the pSTAT3/BCL-2 and VEGF pathways, according to a prior study (Han et al., 2019). Another mechanistic study found that the inhibition of reactive oxygen species (ROS), the activation of the STAT3/Akt/Bcl-2 pathway, and the induced metastatic capacity of HCC cells were the leading causes of enhanced drug resistance in HCC (Wang et al., 2018). Studies have also found that human HCC cells with high CBS expression had low sensitivity to sunitinib and doxorubicin (DOX), while the knockdown of CBS markedly increased the sensitivity of HCC fineness to DOX and sunitinib. Therefore (Wang et al., 2018), it was concluded that CBS overexpression conferred resistance to HCC cells (Stravitz and Lee, 2019).

These findings imply that H_2S has contradictory effects on HCC. Explicitly speaking, exogenous H_2S can cause cancer cell death, while endogenous H_2S can promote cancer. These results indicate that supplementation and restraint of H_2S production are two different ways to treat cancer. As a result, H_2S is expected to play dual roles in the development of HCC (Wu et al., 2017). In the future, it is critical to investigate the mechanism of H_2S in HCC in greater depth (Figure 2).

The effects of H₂S on acute liver failure

Acute liver failure (ALF) is an uncommon and serious complication related to sudden hepatic injury that lasts several days or weeks and is marked by rapid liver destruction, multiorgan failure, and a high death rate (Sen, 2017; Kolodziejczyk et al., 2020). Paracetamol poisoning, hepatic ischemia, viral and autoimmune hepatitis, drug-induced liver injury from medicines, and herbal and nutritional prescription supplements are all potential causes of abrupt liver failure (Kumar and Sandhir, 2018; Yuan et al., 2020). H₂S has been shown to ameliorate liver complications in previous investigations (Dong et al., 2020; Kožich and Stabler, 2020). A recent study explored the possibility of H₂S having a protective impact in ALF. The addition of sodium thiosulfate (STS), an H₂S donor, effectively alleviated D-galactosamine (GalN)- and lipopolysaccharide (LPS)-induced acute liver failure in the wild-type mice by activating Akt- and Nrf2-dependent signaling and by inhibiting GalN/LPS-induced JNK phosphorylation. This suggests that the inhibition of CSE or that the introduction of STS can prevent acute inflammatory liver failure by increasing thiosulfate levels and upregulating antioxidant and anti-apoptotic defenses in the liver (Shirozu et al., 2014). In another study, a thioacetamide (TAA)-induced ALF mouse model was used, and the results showed that NaHS treatment reduced cognitive deficits, enabled the retention of TAA-induced spatial orientation learning, and reduced serum AST and ALT levels and ammonia concentrations in mice (Milewski et al., 2021). These findings suggest the therapeutic potential of H₂S to reduce cognitive deficits and liver dysfunction in ALF mice but the exact biological mechanism remains to be explored (Abo El Gheit et al., 2020; Milewski et al., 2021). Compared to the wild-type mice, the CSE-deficient mice showed a significant attenuation of burn-induced elevations in circulating alkaline aminotransferase and blood urea nitrogen and creatinine levels, suggesting that CSE deficiency has a protective effect against burn-induced impairment of the liver and kidney function. Plasma levels of several burn-induced inflammatory mediators (TNF-a, IL-1β, IL-4, IL-6, IL-10, and IL-12) were significantly lower in the CSE-deficient mice after a burn injury than in the plasma from the wild-type controls. In conclusion, in a mouse burn model, the absence of CSE improved organ function, attenuated the inflammatory response, and effectively limited the progression of multi-organ failure. However, the exact mechanism remains to be explored (Ahmad et al., 2017).

In a study of burn-induced acute liver injury in mice, plasma H_2S levels and H_2S synthesis activity were significantly increased in the liver after a burn injury, and NaHS injections at the time of burn injury also led to a substantial increase in liver myeloperoxidase (MPO) activity and a significant increase in the systemic inflammatory response, inducing multi-organ



Effects of liver-derived H_2S on various liver diseases. H_2S is produced in the liver by the enzymatic reactions of cysteine gamma-catabolase (CSE) (Jia et al., 2022), cystathionine β -synthase (CBS) (Roy et al., 2012; Zuhra et al., 2020a), and 3-mercaptopyruvate (3-MP) *via* L-cysteine (Roy et al., 2012; Abdollahi Govar et al., 2020; Dilek et al., 2020). H_2S can have different effects in various liver diseases.



proliferation and endogenous R_2^{-} play different roles in HCC through uniterent mechanism. (A) Endogenous R_2^{-} s promotes minute escape and proliferation and enhances drug resistance in HCC, which may be related to the promotion of Treg-mediated immune evasion (Xu et al., 2021; Zhou et al., 2021), the stimulation of ATP production (Fortibui et al., 2021), and the activation of the STAT3/Akt/Bcl-2 pathway (Wang et al., 2018). The inhibition of reactive oxygen species (ROS) (Wang et al., 2018). (B) Treatment of hepatoma cells with exogenous H_2 S could induce HCC cell autophagy by activating the JNK/JunB/TNFSF14 signaling pathway (Ma et al., 2022) and the STAT3/Akt/Bcl-2 pathway (Han et al., 2019), upregulating caspase-3/9 (Sakuma et al., 2019), inhibiting the PI3K/AKT/mTOR signaling pathway (Wang et al., 2017), regulating the p53/p58/MAPK pathway (Zhao et al., 2015), blocking the STAT3/NF- κ B/IDO1 pathway, and suppressing anti-apoptotic markers, thus promoting hepatoma cell apoptosis.

damage, including liver injury. This suggests that H_2S can significantly exacerbate burn-induced acute liver injury (Zhang et al., 2010).

These findings imply that H₂S may play a double-edged role in acute liver failure, and more in-depth studies are still needed to validate it further.

The effect of H₂S on acute liver pathology

Liver toxicity refers to the damage to the liver produced by a variety of prescription and over-the-counter medications, including natural medicines, biologics, dietary supplements, nutraceuticals, and some traditional Chinese medicines (Jaeschke, 2015). Drug-induced liver damage is a rare but serious medical issue (Leise et al., 2014; Wang et al., 2020a). In a limited number of patients, the use of multiple drugs has caused serious liver injury and acute liver failure (<1:10,000) (Chalasani et al., 2008). Although its incidence in the population is very low, the high likelihood of acute liver failure in patients with acute liver injury still requires much attention.

A liver toxicity study in mice found that H₂S significantly inhibited oxidative stress, inflammation, and apoptosis induced by polystyrene microplastics (mic-PS). H₂S increased the expression of NAD(P)H:quinone oxidoreductase 1 (NQOl) and heme oxygenase-1 (HO-1) by promoting the nuclear accumulation of the nuclear factor-E2-related factor (Nrf2), thereby reducing the apoptotic and inflammatory responses induced by mic-PS in mouse hepatocytes. This revealed the hepatic toxic effect of mic-PS and the protective effect of H₂S on mic-PS-induced liver injury (Li et al., 2021a). In a study of acetaminophen (APAP)-induced hepatotoxicity in mice, treatment with H₂S significantly reduced serum levels of AST, ALT, IL-33, and TNF-α. It attenuated APAP-induced hepatocyte apoptosis in mice by inhibiting the JNK/MAPK signaling pathway, effectively reducing APAP-induced thus hepatotoxicity (Li et al., 2019; Saleh et al., 2021). In addition, another hepatotoxicity study in rats demonstrated that H₂S protects the liver in methotrexate (MTX)-stimulated rats by acting as anti-inflammatory, antioxidant, and anti-apoptotic agent functions, which are most likely mediated by H₂S through the modulation of the IL-6/STAT3 pathway, initiation of the KATP pathway, and activation of endothelial nitric oxide synthase (eNOS) and transient receptor potential vanilloid 1 (TRPV1) (Fouad et al., 2020). Thus, H₂S has a potential value for treating hepatotoxicity.

Conclusion and perspective

In this review, we summarized and discussed the effects and potential mechanisms of H_2S in the process of liver disease, including NAFLD, liver I/R injury, liver fibrosis, acute liver failure, liver toxicity, and hepatocellular carcinoma. Based on the research results, we found that endogenous and exogenous high and low concentrations of H_2S may exert different effects by regulating different signaling pathways.

The effects of H_2S on NAFLD and liver I/R injury are relatively clear. Under NAFLD conditions, both endogenous and exogenous H_2S were able to reduce lipid deposition to

inhibit the progression of NAFLD, indicating that H_2S plays a protective role in pathological conditions in the liver. Moreover, H_2S alleviated liver I/R injury by reducing the inflammatory reaction. However, H_2S may act as a negative molecule in promoting the progression of liver fibrosis. Both endogenous and exogenous H_2S could activate HSCs to increase the secretion of extracellular matrix, which participates in liver fibrosis. Beyond its roles in NAFLD and liver fibrosis, the roles of H_2S in other liver diseases remain controversial.

Based on the findings of these studies, we determined that H_2S plays a double-edged role in liver diseases. We suppose that there may be a certain balance between the protective and pathogenic effects of H_2S in different liver conditions. Since there is no definite evidence to prove the existence of this balance at present, more studies are needed in the future. Considering the controversial effects of H_2S in different liver diseases, inhibiting the synthesis of endogenous H_2S or providing exogenous H_2S can effectively alleviate the progression of diseases.

Currently, H_2S has been studied as a drug in the cardiovascular field. However, the dual role of H_2S in different diseases should be considered when using H_2S to treat liver diseases. Inhibition of endogenous H_2S synthesis or the administration of exogenous H_2S can play a positive role in the treatment of liver diseases and are, therefore, promising treatment strategies. Therefore, a better understanding of the dual role of H_2S will provide a strong experimental basis for the treatment of different diseases and for drug research.

Author contributions

LL designed the study; BL, SW, and MX wrote the manuscript; HD, RS, and YM contributed to modifying the manuscript. All authors contributed to result interpretation and discussion.

Funding

Financial support for this study was provided by the Beijing Hospital Management Center Gastroenterology Department Synergy Development Center (XXZ0801), Beijing High-level Health Technicians (No. 2013-03-037), Beijing Hospitals Authority Youth Programme (QML20211701), National Natural Science Foundation (81672725), and the Sino-German Mobility Program (M-0200).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

Abd El-Kader, S. M., and El-Den Ashmawy, E. M. (2015). Non-alcoholic fatty liver disease: The diagnosis and management. *World J. Hepatol.* 7, 846–858. doi:10. 4254/wjh.v7.i6.846

Abdollahi Govar, A., Toro, G., Szaniszlo, P., Pavlidou, A., Bibli, S. I., Thanki, K., et al. (2020). 3-Mercaptopyruvate sulfurtransferase supports endothelial cell angiogenesis and bioenergetics. *Br. J. Pharmacol.* 177, 866–883. doi:10.1111/bph. 14574

Abo El Gheit, R. E., Atef, M. M., Badawi, G. A., Elwan, W. M., Alshenawy, H. A., and Emam, M. N. (2020). Role of serine protease inhibitor, ulinastatin, in rat model of hepatic encephalopathy: Aquaporin 4 molecular targeting and therapeutic implication. *J. Physiol. Biochem.* 76, 573–586. doi:10.1007/s13105-020-00762-0

Ahmad, A., Druzhyna, N., and Szabo, C. (2017). Cystathionine-gamma-lyase deficient mice are protected against the development of multiorgan failure and exhibit reduced inflammatory response during burn. *Burns* 43, 1021–1033. doi:10. 1016/j.burns.2017.02.011

Ali, A., Zhang, Y., Fu, M., Pei, Y., Wu, L., Wang, R., et al. (2020). Cystathionine gamma-lyase/H(2)S system suppresses hepatic acetyl-CoA accumulation and nonalcoholic fatty liver disease in mice. *Life Sci.* 252, 117661. doi:10.1016/j.lfs. 2020.117661

Andrade, R. J., Chalasani, N., Björnsson, E. S., Suzuki, A., Kullak-Ublick, G. A., Watkins, P. B., et al. (2019). Drug-induced liver injury. *Nat. Rev. Dis. Prim.* 5, 58. doi:10.1038/s41572-019-0105-0

Bai, Y. W., Ye, M. J., Yang, D. L., Yu, M. P., Zhou, C. F., and Shen, T. (2019). Hydrogen sulfide attenuates paraquat-induced epithelial-mesenchymal transition of human alveolar epithelial cells through regulating transforming growth factor- β 1/Smad2/3 signaling pathway. *J. Appl. Toxicol.* 39, 432–440. doi:10.1002/jat.3734

Balaban, C. L., Rodriguez, J. V., and Guibert, E. E. (2011). Delivery of the bioactive gas hydrogen sulfide during cold preservation of rat liver: Effects on hepatic function in an *ex vivo* model. *Artif. Organs* 35, 508–515. doi:10.1111/j.1525-1594.2011.01256.x

Balaban, C. L., Rodríguez, J. V., Tiribelli, C., and Guibert, E. E. (2015). The effect of a hydrogen sulfide releasing molecule (Na2S) on the cold storage of livers from cardiac dead donor rats. A study in an *ex vivo* model. *Cryobiology* 71, 24–32. doi:10. 1016/j.cryobiol.2015.06.006

Bełtowski, J. (2007). Synthesis, metabolism, and signaling mechanisms of hydrogen sulfide: An overview. *Methods Mol. Biol.* 2007, 1–8. doi:10.1007/978-1-4939-9528-8_1

Cao, X., Ding, L., Xie, Z. Z., Yang, Y., Whiteman, M., Moore, P. K., et al. (2019). A review of hydrogen sulfide synthesis, metabolism, and measurement: Is modulation of hydrogen sulfide a novel therapeutic for cancer? *Antioxid. Redox Signal.* 31, 1–38. doi:10.1089/ars.2017.7058

Chalasani, N., Fontana, R. J., Bonkovsky, H. L., Watkins, P. B., Davern, T., Serrano, J., et al. (2008). Causes, clinical features, and outcomes from a prospective study of drug-induced liver injury in the United States. *Gastroenterology* 135, 1924–1934. doi:10.1053/j.gastro.2008.09.011

Chen, K., Zhong, J., Hu, L., Li, R., Du, Q., Cai, J., et al. (2019). The role of xenobiotic receptors on hepatic glycolipid metabolism. *Curr. Drug Metab.* 20, 29–35. doi:10.2174/1389200219666180918152241

Corvino, A., Frecentese, F., Magli, E., Perissutti, E., Santagada, V., Scognamiglio, A., et al. (2021). Trends in H(2)S-donors chemistry and their effects in cardiovascular diseases. *Antioxidants (Basel)* 10.

Damba, T., Zhang, M., Buist-Homan, M., Van Goor, H., Faber, K. N., and Moshage, H. (2019). Hydrogen sulfide stimulates activation of hepatic stellate cells through increased cellular bio-energetics. *Nitric Oxide* 92, 26–33. doi:10.1016/j. niox.2019.08.004

Dilek, N., Papapetropoulos, A., Toliver-Kinsky, T., and Szabo, C. (2020). Hydrogen sulfide: An endogenous regulator of the immune system. *Pharmacol. Res.* 161, 105119. doi:10.1016/j.phrs.2020.105119

Dong, V., Nanchal, R., and Karvellas, C. J. (2020). Pathophysiology of acute liver failure. *Nutr. Clin. Pract.* 35, 24–29. doi:10.1002/ncp.10459

organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Duan, J. J., Cai, J., Guo, Y. F., Bian, X. W., and Yu, S. C. (2016). ALDH1A3, a metabolic target for cancer diagnosis and therapy. *Int. J. Cancer* 139, 965–975. doi:10.1002/ijc.30091

Dugbartey, G. J., Juriasingani, S., Zhang, M. Y., and Sener, A. (2021). H(2)S donor molecules against cold ischemia-reperfusion injury in preclinical models of solid organ transplantation. *Pharmacol. Res.* 172, 105842. doi:10.1016/j.phrs.2021. 105842

Fortibui, M. M., Yoon, D. W., Lim, J. Y., Lee, S., Choi, M., Heo, J. S., et al. (2021). A cancer cell-specific benzoxadiazole-based fluorescent probe for hydrogen sulfide detection in mitochondria. *Dalton Trans.* 50, 2545–2554. doi:10.1039/d0dt03653f

Fouad, A. A., Hafez, H. M., and Hamouda, A. (2020). Hydrogen sulfide modulates IL-6/STAT3 pathway and inhibits oxidative stress, inflammation, and apoptosis in rat model of methotrexate hepatotoxicity. *Hum. Exp. Toxicol.* 39, 77–85. doi:10. 1177/0960327119877437

Fousek, K., Horn, L. A., and Palena, C. (2021). Interleukin-8: A chemokine at the intersection of cancer plasticity, angiogenesis, and immune suppression. *Pharmacol. Ther.* 219, 107692. doi:10.1016/j.pharmthera.2020.107692

Fu, J., Cheng, X. H., and Zhang, L. (2019). Effect of hydrogen sulphide on inflammatory factors of the mitochondria after limb ischaemia-reperfusion injury in rats. *Int. Wound J.* 16, 595–600. doi:10.1111/iwj.13068

Gong, Z., Ye, H., Huo, Y., Wang, L., Huang, Y., Huang, M., et al. (2018). S-allylcysteine attenuates carbon tetrachloride-induced liver fibrosis in rats by targeting STAT3/SMAD3 pathway. *Am. J. Transl. Res.* 10, 1337–1346.

Han, H., Wang, L., Liu, Y., Shi, X., Zhang, X., Li, M., et al. (2019). Combination of curcuma zedoary and kelp inhibits growth and metastasis of liver cancer *in vivo* and *in vitro* via reducing endogenous H(2)S levels. *Food Funct.* 10, 224–234. doi:10. 1039/c8fo01594e

Ibrahim, S. A., Abdel-Gaber, S. A., Ibrahim, M. A., Amin, E. F., Mohammed, R. K., and Abdelrahman, A. M. (2021). Nitric oxide modulation as a potential molecular mechanism underlying the protective role of NaHS in liver ischemia reperfusion injury. *Curr. Mol. Pharmacol.*, 36, 177.

Iciek, M., Górny, M., Bilska-Wilkosz, A., and Kowalczyk-Pachel, D. (2018). Is aldehyde dehydrogenase inhibited by sulfur compounds? *in vitro* and *in vivo* studies. *Acta Biochim. Pol.* 65, 125–132. doi:10.18388/abp.2017_2324

Jaeschke, H. (2015). Acetaminophen: Dose-Dependent drug hepatotoxicity and acute liver failure in patients. *Dig. Dis.* 33, 464–471. doi:10.1159/000374090

Jia, Y., Wang, Y., Zhang, Y., Wang, J., Pei, Y., Wang, Z., et al. (2022). Profiling cystathionine β / γ -Lyase in complex biosamples using novel activatable fluorogens. *Anal. Chem.* 94, 1203–1210. doi:10.1021/acs.analchem.1c04393

Jiménez-Castro, M. B., Cornide-Petronio, M. E., Gracia-Sancho, J., and Peralta, C. (2019). Inflammasome-mediated inflammation in liver ischemia-reperfusion injury. *Cells* 8, 1131. doi:10.3390/cells8101131

Jung, K. J., Jang, H. S., Kim, J. I., Han, S. J., Park, J. W., and Park, K. M. (2013). Involvement of hydrogen sulfide and homocysteine transsulfuration pathway in the progression of kidney fibrosis after ureteral obstruction. *Biochim. Biophys. Acta* 1832, 1989–1997. doi:10.1016/j.bbadis.2013.06.015

Kabil, O., Motl, N., and Banerjee, R. (2014). H2S and its role in redox signaling. Biochim. Biophys. Acta 1844, 1355-1366. doi:10.1016/j.bbapap.2014.01.002

Kang, K., Zhao, M., Jiang, H., Tan, G., Pan, S., and Sun, X. (2009). Role of hydrogen sulfide in hepatic ischemia-reperfusion-induced injury in rats. *Liver Transpl.* 15, 1306–1314. doi:10.1002/lt.21810

Kodai, S., Takemura, S., Minamiyama, Y., Hai, S., Yamamoto, S., Kubo, S., et al. (2007). S-allyl cysteine prevents CCl(4)-induced acute liver injury in rats. *Free Radic. Res.* 41, 489–497. doi:10.1080/10715760601118361

Kolodziejczyk, A. A., Federici, S., Zmora, N., Mohapatra, G., Dori-Bachash, M., Hornstein, S., et al. (2020). Acute liver failure is regulated by MYC- and microbiome-dependent programs. *Nat. Med.* 26, 1899–1911. doi:10.1038/ s41591-020-1102-2

Kožich, V., and Stabler, S. (2020). Lessons learned from inherited metabolic disorders of sulfur-containing amino acids metabolism. *J. Nutr.* 150, 2506S–2517s. doi:10.1093/jn/nxaa134

Krylatov, A., Maslov, L., Tsibulnikov, S. Y., Voronkov, N., Boshchenko, A., Downey, J., et al. (2021). The role of reactive oxygen species, kinases, hydrogen sulfide, and nitric oxide in the regulation of autophagy and their impact on ischemia and reperfusion injury in the heart. *Curr. Cardiol. Rev.* 17, e230421186874. doi:10. 2174/1573403X16666201014142446

Kumar, M., and Sandhir, R. (2018). Hydrogen sulfide in physiological and pathological mechanisms in brain. CNS Neurol. Disord. Drug Targets 17, 654-670. doi:10.2174/1871527317666180605072018

Lambrecht, J., Mannaerts, I., and Van Grunsven, L. A. (2015). The role of miRNAs in stress-responsive hepatic stellate cells during liver fibrosis. *Front. Physiol.* 6, 209. doi:10.3389/fphys.2015.00209

Leise, M. D., Poterucha, J. J., and Talwalkar, J. A. (2014). Drug-induced liver injury. *Mayo Clin. Proc.* 89, 95–106. doi:10.1016/j.mayocp.2013.09.016

Li, M., Xu, C., Shi, J., Ding, J., Wan, X., Chen, D., et al. (2018). Fatty acids promote fatty liver disease via the dysregulation of 3-mercaptopyruvate sulfurtransferase/hydrogen sulfide pathway. *Gut* 67, 2169–2180. doi:10. 1136/gutjnl-2017-313778

Li, S., Shi, M., Wang, Y., Xiao, Y., Cai, D., and Xiao, F. (2021a). Keap1-Nrf2 pathway up-regulation via hydrogen sulfide mitigates polystyrene microplastics induced-hepatotoxic effects. *J. Hazard. Mat.* 402, 123933. doi:10.1016/j.jhazmat. 2020.123933

Li, X., Lin, J., Lin, Y., Huang, Z., Pan, Y., Cui, P., et al. (2019). Hydrogen sulfide protects against acetaminophen-induced acute liver injury by inhibiting apoptosis via the JNK/MAPK signaling pathway. *J. Cell. Biochem.* 120, 4385–4397. doi:10. 1002/jcb.27724

Li, X., Ramadori, P., Pfister, D., Seehawer, M., Zender, L., and Heikenwalder, M. (2021b). The immunological and metabolic landscape in primary and metastatic liver cancer. *Nat. Rev. Cancer* 21, 541–557. doi:10.1038/s41568-021-00383-9

Liu, X., Yang, R., Bai, W., Xu, X., Bi, F., Hao, Y., et al. (2020). Involvement of amylin B-H2S-connexin 43 signaling pathway in vascular dysfunction and enhanced ischemia-reperfusion-induced myocardial injury in diabetic rats. *Biosci. Rep.* 40, BSR20194154. doi:10.1042/BSR20194154

Loiselle, J. J., Yang, G., and Wu, L. (2020). Hydrogen sulfide and hepatic lipid metabolism - a critical pairing for liver health. *Br. J. Pharmacol.* 177, 757–768. doi:10.1111/bph.14556

Lu, M., Jiang, X., Tong, L., Zhang, F., Ma, L., Dong, X., et al. (2018). MicroRNA-21-Regulated activation of the Akt pathway participates in the protective effects of H(2)S against liver ischemia-reperfusion injury. *Biol. Pharm. Bull.* 41, 229–238. doi:10.1248/bpb.b17-00769

Ma, Y., Wang, S., Wu, Y., Liu, B., Li, L., Wang, W., et al. (2022). Hepatic stellate cell mediates transcription of TNFSF14 in hepatocellular carcinoma cells via H(2)S/ CSE-JNK/JunB signaling pathway. *Cell Death Dis.* 13, 238. doi:10.1038/s41419-022-04678-z

Ma, Y., Yan, Z., Deng, X., Guo, J., Hu, J., Yu, Y., et al. (2018). Anticancer effect of exogenous hydrogen sulfide in cisplatin-resistant A549/DDP cells. *Oncol. Rep.* 39, 2969–2977. doi:10.3892/or.2018.6362

Malik, A., Thanekar, U., Amarachintha, S., Mourya, R., Nalluri, S., Bondoc, A., et al. (2020). Complimenting the complement": Mechanistic insights and opportunities for therapeutics in hepatocellular carcinoma. *Front. Oncol.* 10, 627701. doi:10.3389/fonc.2020.627701

Mani, S., Li, H., Yang, G., Wu, L., and Wang, R. (2015). Deficiency of cystathionine gamma-lyase and hepatic cholesterol accumulation during mouse fatty liver development. *Sci. Bull.* 60, 336–347. doi:10.1007/s11434-014-0722-7

Milewski, K., Czarnecka, A. M., Albrecht, J., and Zielińska, M. (2021). The status of bile acids and farnesoid X receptor in brain and liver of rats with thioacetamide-induced acute liver failure. *Int. J. Mol. Sci.* 22, 7750. doi:10.3390/ jjms21207750

Mohammed, R. A., Shawky, H. M., Rashed, L. A., Elhanbuli, H. M., Abdelhafez, D. N., Said, E. S., et al. (2021). Combined effect of hydrogen sulfide and mesenchymal stem cells on mitigating liver fibrosis induced by bile duct ligation: Role of anti-inflammatory, anti-oxidant, anti-apoptotic, and anti-fibrotic biomarkers. *Iran. J. Basic Med. Sci.* 24, 1753–1762. doi:10.22038/IJBMS. 2021.56477.12604

Motz, G. T., and Coukos, G. (2011). The parallel lives of angiogenesis and immunosuppression: Cancer and other tales. *Nat. Rev. Immunol.* 11, 702–711. doi:10.1038/nri3064

Muller, X., Rossignol, G., Mohkam, K., and Mabrut, J. Y. (2022). Novel strategies in liver graft preservation - the French perspective. *J. Visc. Surg.* 159, 389–398. doi:10.1016/j.jviscsurg.2022.06.006

Muñoz, R., Girotti, A., Hileeto, D., and Arias, F. J. (2021). Metronomic anticancer therapy: A multimodal therapy governed by the tumor microenvironment. *Cancers (Basel)* 13, 5414. doi:10.3390/cancers13215414 Nastos, C., Kalimeris, K., Papoutsidakis, N., Tasoulis, M. K., Lykoudis, P. M., Theodoraki, K., et al. (2014). Global consequences of liver ischemia/reperfusion injury. Oxid. Med. Cell. Longev. 2014, 906965. doi:10.1155/2014/906965

Ngowi, E. E., Afzal, A., Sarfraz, M., Khattak, S., Zaman, S. U., Khan, N. H., et al. (2021). Role of hydrogen sulfide donors in cancer development and progression. *Int. J. Biol. Sci.* 17, 73–88. doi:10.7150/ijbs.47850

Ni, M. M., Wang, Y. R., Wu, W. W., Xia, C. C., Zhang, Y. H., Xu, J., et al. (2018). Novel Insights on Notch signaling pathways in liver fibrosis. *Eur. J. Pharmacol.* 826, 66–74. doi:10.1016/j.ejphar.2018.02.051

Norris, E. J., Culberson, C. R., Narasimhan, S., and Clemens, M. G. (2011). The liver as a central regulator of hydrogen sulfide. *Shock* 36, 242–250. doi:10.1097/SHK. 0b013e3182252ee7

Pan, Y., Ye, S., Yuan, D., Zhang, J., Bai, Y., and Shao, C. (2014). Hydrogen sulfide (H2S)/cystathionine γ -lyase (CSE) pathway contributes to the proliferation of hepatoma cells. *Mutat. Res.* 763, 10–18. doi:10.1016/j.mrfmmm.2014.03.002

Pant, I., Rao, S. G., and Kondaiah, P. (2016). Role of areca nut induced JNK/ ATF2/Jun axis in the activation of TGF- β pathway in precancerous Oral Submucous Fibrosis. *Sci. Rep.* 6, 34314. doi:10.1038/srep34314

Pinter, M., Jain, R. K., and Duda, D. G. (2021). The current landscape of immune checkpoint blockade in hepatocellular carcinoma: A review. *JAMA Oncol.* 7, 113–123. doi:10.1001/jamaoncol.2020.3381

Roehlen, N., Crouchet, E., and Baumert, T. F. (2020). Liver fibrosis: Mechanistic concepts and therapeutic perspectives. *Cells* 9, 875. doi:10.3390/cells9040875

Roy, A., Khan, A. H., Islam, M. T., Prieto, M. C., and Majid, D. S. (2012). Interdependency of cystathione γ -lyase and cystathione β -synthase in hydrogen sulfide-induced blood pressure regulation in rats. *Am. J. Hypertens.* 25, 74–81. doi:10.1038/ajh.2011.149

Ruan, Z., Liang, M., Deng, X., Lai, M., Shang, L., and Su, X. (2019). Exogenous hydrogen sulfide protects fatty liver against ischemia-reperfusion injury by regulating endoplasmic reticulum stress-induced autophagy in macrophage through mediating the class A scavenger receptor pathway in rats. *Cell Biol. Int.* 78, 306–316. doi:10.1002/cbin.11234

Sakuma, S., Minamino, S., Takase, M., Ishiyama, Y., Hosokura, H., Kohda, T., et al. (2019). Hydrogen sulfide donor GYY4137 suppresses proliferation of human colorectal cancer Caco-2 cells by inducing both cell cycle arrest and cell death. *Heliyon* 5, e02244. doi:10.1016/j.heliyon.2019.e02244

Saleh, N. E. H., Saad, A. H., Elbassuoni, E. A., El-Tahawy, N. F., and Abdel-Hakeem, E. A. (2021). Potential benefits of using hydrogen sulfide, vitamin E and necrostatin-1 to counteract acetaminophen-induced hepatotoxicity in rats. *Bratisl. Lek. Listy* 122, 732–738. doi:10.4149/BLL_2021_117

Scammahorn, J. J., Nguyen, I. T. N., Bos, E. M., Van Goor, H., and Joles, J. A. (2021). Fighting oxidative stress with sulfur: Hydrogen sulfide in the renal and cardiovascular systems. *Antioxidants (Basel)* 10 (3), 373. doi:10.3390/antiox10030373

Sen, N. (2017). Functional and molecular insights of hydrogen sulfide signaling and protein sulfhydration. J. Mol. Biol. 429, 543–561. doi:10.1016/j.jmb.2016.12.015

Shackelford, R., Ozluk, E., Islam, M. Z., Hopper, B., Meram, A., Ghali, G., et al. (2021). Hydrogen sulfide and DNA repair. *Redox Biol.* 38, 101675. doi:10.1016/j. redox.2020.101675

Shi, D., Chen, J., Wang, J., Yao, J., Huang, Y., Zhang, G., et al. (2019). Circadian clock genes in the metabolism of non-alcoholic fatty liver disease. *Front. Physiol.* 10, 423. doi:10.3389/fphys.2019.00423

Shirozu, K., Tokuda, K., Marutani, E., Lefer, D., Wang, R., and Ichinose, F. (2014). Cystathionine γ-lyase deficiency protects mice from galactosamine/ lipopolysaccharide-induced acute liver failure. *Antioxid. Redox Signal.* 20, 204–216. doi:10.1089/ars.2013.5354

Song, K., Li, Q., Yin, X. Y., Lu, Y., Liu, C. F., and Hu, L. F. (2015). Hydrogen sulfide: A therapeutic candidate for fibrotic disease? *Oxid. Med. Cell. Longev.* 2015, 458720. doi:10.1155/2015/458720

Stefan, N., Häring, H. U., and Cusi, K. (2019). Non-alcoholic fatty liver disease: Causes, diagnosis, cardiometabolic consequences, and treatment strategies. *Lancet. Diabetes Endocrinol.* 7, 313–324. doi:10.1016/S2213-8587(18)30154-2

Stravitz, R. T., and Lee, W. M. (2019). Acute liver failure. *Lancet* 394, 869–881. doi:10.1016/S0140-6736(19)31894-X

Sun, H. J., Wu, Z. Y., Nie, X. W., Wang, X. Y., and Bian, J. S. (2021). Implications of hydrogen sulfide in liver pathophysiology: Mechanistic insights and therapeutic potential. *J. Adv. Res.* 27, 127–135. doi:10.1016/j.jare.2020.05.010

Szadvari, I., Hudecova, S., Chovancova, B., Matuskova, M., Cholujova, D., Lencesova, L., et al. (2019). Sodium/calcium exchanger is involved in apoptosis induced by H(2)S in tumor cells through decreased levels of intracellular pH. *Nitric Oxide* 87, 1–9. doi:10.1016/j.niox.2019.02.011

Tan, G., Pan, S., Li, J., Dong, X., Kang, K., Zhao, M., et al. (2011). Hydrogen sulfide attenuates carbon tetrachloride-induced hepatotoxicity, liver cirrhosis and portal hypertension in rats. *PLoS One* 6, e25943. doi:10.1371/journal.pone. 0025943

Toledo-Guzmán, M. E., Hernández, M. I., Gómez-Gallegos Á, A., and Ortiz-Sánchez, E. (2019). ALDH as a stem cell marker in solid tumors. *Curr. Stem Cell Res. Ther.* 14, 375–388. doi:10.2174/1574888X13666180810120012

Varela, A. T., Rolo, A. P., and Palmeira, C. M. (2011). Fatty liver and ischemia/ reperfusion: Are there drugs able to mitigate injury? *Curr. Med. Chem.* 18, 4987–5002. doi:10.2174/092986711797535164

Wang, L., Han, H., Liu, Y., Zhang, X., Shi, X., and Wang, T. (2018). Cystathionine β -synthase induces multidrug resistance and metastasis in hepatocellular carcinoma. *Curr. Mol. Med.* 18, 496–506. doi:10.2174/1566524019666181211162754

Wang, M., Liu, C. Y., Wang, T., Yu, H. M., Ouyang, S. H., Wu, Y. P., et al. (2020a). (+)-Clausenamide protects against drug-induced liver injury by inhibiting hepatocyte ferroptosis. *Cell Death Dis.* 11, 781. doi:10.1038/ s41419-020-02961-5

Wang, S. S., Chen, Y. H., Chen, N., Wang, L. J., Chen, D. X., Weng, H. L., et al. (2017). Hydrogen sulfide promotes autophagy of hepatocellular carcinoma cells through the PI3K/Akt/mTOR signaling pathway. *Cell Death Dis.* 8, e2688. doi:10. 1038/cddis.2017.18

Wang, W., Wang, C., Xu, H., and Gao, Y. (2020b). Aldehyde dehydrogenase, liver disease and cancer. Int. J. Biol. Sci. 16, 921–934. doi:10.7150/ijbs.42300

Wu, D. D., Wang, D. Y., Li, H. M., Guo, J. C., Duan, S. F., and Ji, X. Y. (2019). Hydrogen sulfide as a novel regulatory factor in liver health and disease. *Oxid. Med. Cell. Longev.* 2019, 3831713. doi:10.1155/2019/3831713

Wu, D., Li, M., Tian, W., Wang, S., Cui, L., Li, H., et al. (2017). Hydrogen sulfide acts as a double-edged sword in human hepatocellular carcinoma cells through EGFR/ERK/MMP-2 and PTEN/AKT signaling pathways. *Sci. Rep.* 7, 5134. doi:10. 1038/s41598-017-05457-z

Wu, D., Wang, J., Li, H., Xue, M., Ji, A., and Li, Y. (2015). Role of hydrogen sulfide in ischemia-reperfusion injury. *Oxid. Med. Cell. Longev.* 2015, 186908. doi:10.1155/ 2015/186908

Wu, D., Zhong, P., Wang, Y., Zhang, Q., Li, J., Liu, Z., et al. (2020). Hydrogen sulfide attenuates high-fat diet-induced non-alcoholic fatty liver disease by inhibiting apoptosis and promoting autophagy via reactive oxygen species/ phosphatidylinositol 3-kinase/AKT/mammalian target of rapamycin signaling pathway. *Front. Pharmacol.* 11, 585860. doi:10.3389/fphar.2020.585860

Xia, L., Oyang, L., Lin, J., Tan, S., Han, Y., Wu, N., et al. (2021). The cancer metabolic reprogramming and immune response. *Mol. Cancer* 20, 28. doi:10.1186/s12943-021-01316-8

Xu, J., Lin, H., Wu, G., Zhu, M., and Li, M. (2021). IL-6/STAT3 is a promising therapeutic target for hepatocellular carcinoma. *Front. Oncol.* 11, 760971. doi:10. 3389/fonc.2021.760971

Xu, W., Cui, C., Cui, C., Chen, Z., Zhang, H., Cui, Q., et al. (2022). Hepatocellular cystathionine γ lyase/hydrogen sulfide attenuates nonalcoholic fatty liver disease by activating farnesoid X receptor. Hepatology 221, 1794–1810. doi:10.1002/hep.32577

Yang, D., Li, T., Li, Y., Zhang, S., Li, W., Liang, H., et al. (2019a). H(2)S suppresses indoleamine 2, 3-dioxygenase 1 and exhibits immunotherapeutic efficacy in murine hepatocellular carcinoma. *J. Exp. Clin. Cancer Res.* 38, 88. doi:10.1186/s13046-019-1083-5

Yang, J. D., Hainaut, P., Gores, G. J., Amadou, A., Plymoth, A., and Roberts, L. R. (2019c). A global view of hepatocellular carcinoma: Trends, risk, prevention and

management. Nat. Rev. Gastroenterol. Hepatol. 16, 589-604. doi:10.1038/s41575-019-0186-y

Yang, J., Minkler, P., Grove, D., Wang, R., Willard, B., Dweik, R., et al. (2019b). Non-enzymatic hydrogen sulfide production from cysteine in blood is catalyzed by iron and vitamin B(6). *Commun. Biol.* 2, 194. doi:10.1038/s42003-019-0431-5

Yang, T., Gu, J., Liu, T., Ma, H., Ma, X., Tao, J., et al. (2018a). Expression of acetaldehyde dehydrogenase in gefitinib-resistant human lung adenocarcinoma HCC-827/GR cells. *Zhongguo Fei Ai Za Zhi* 21, 431–436. doi:10.3779/j.issn.1009-3419.2018.06.01

Yang, W., Chen, J., Meng, Y., Chen, Z., and Yang, J. (2018b). Novel targets for treating ischemia-reperfusion injury in the liver. *Int. J. Mol. Sci.* 19, 1302. doi:10. 3390/ijms19051302

Younis, N. N., Shaheen, M. A., and Mahmoud, M. F. (2016). Silymarin preconditioning protected insulin resistant rats from liver ischemia-reperfusion injury: Role of endogenous H2S. *J. Surg. Res.* 204, 398–409. doi:10.1016/j.jss.2016. 04.069

Yuan, D. S., Huang, Y. Q., Fu, Y. J., Xie, J., Huang, Y. L., Zhou, S. S., et al. (2020). Hydrogen sulfide alleviates cognitive deficiency and hepatic dysfunction in a mouse model of acute liver failure. *Exp. Ther. Med.* 20, 671–677. doi:10.3892/etm.2020. 8680

Zhai, Y., Petrowsky, H., Hong, J. C., Busuttil, R. W., and Kupiec-Weglinski, J. W. (2013). Ischaemia-reperfusion injury in liver transplantation-from bench to bedside. *Nat. Rev. Gastroenterol. Hepatol.* 10, 79–89. doi:10.1038/nrgastro.2012.225

Zhang, D., Fang, P., Jiang, X., Nelson, J., Moore, J. K., Kruger, W. D., et al. (2012). Severe hyperhomocysteinemia promotes bone marrow-derived and resident inflammatory monocyte differentiation and atherosclerosis in LDLr/CBSdeficient mice. *Circ. Res.* 111, 37–49. doi:10.1161/CIRCRESAHA.112.269472

Zhang, J., Sio, S. W., Moochhala, S., and Bhatia, M. (2010). Role of hydrogen sulfide in severe burn injury-induced inflammation in mice. *Mol. Med.* 16, 417–424. doi:10.2119/molmed.2010.00027

Zhang, X., Chen, M., Ni, X., Wang, Y., Zheng, X., Zhang, H., et al. (2020). Metabolic reprogramming of sulfur in hepatocellular carcinoma and sulfane sulfurtriggered anti-cancer strategy. *Front. Pharmacol.* 11, 571143. doi:10.3389/fphar. 2020.571143

Zhao, L., Wang, Y., Yan, Q., Lv, W., Zhang, Y., and He, S. (2015). Exogenous hydrogen sulfide exhibits anti-cancer effects though p38 MAPK signaling pathway in C6 glioma cells. *Biol. Chem.* 396, 1247–1253. doi:10.1515/hsz-2015-0148

Zhao, M., Cheng, Y., Wang, X., Cui, X., Cheng, X., Fu, Q., et al. (2022). Hydrogen sulfide attenuates high-fat diet-induced obesity: Involvement of mTOR/IKK/NF- κ B signaling pathway. *Mol. Neurobiol.* 59, 6903–6917. doi:10.1007/s12035-022-03004-0

Zhou, Y. F., Song, S. S., Tian, M. X., Tang, Z., Wang, H., Fang, Y., et al. (2021). Cystathionine β -synthase mediated PRRX2/IL-6/STAT3 inactivation suppresses Tregs infiltration and induces apoptosis to inhibit HCC carcinogenesis. *J. Immunother. Cancer* 9, e003031. doi:10.1136/jitc-2021-003031

Zou, C. G., Gao, S. Y., Zhao, Y. S., Li, S. D., Cao, X. Z., Zhang, Y., et al. (2009). Homocysteine enhances cell proliferation in hepatic myofibroblastic stellate cells. J. Mol. Med. 87, 75–84. doi:10.1007/s00109-008-0407-2

Zuhra, K., Augsburger, F., Majtan, T., and Szabo, C. (2020a). Cystathionine-β-Synthase: Molecular regulation and pharmacological inhibition. *Biomolecules* 10, 697. doi:10.3390/biom10050697

Zuhra, K., Panagaki, T., Randi, E. B., Augsburger, F., Blondel, M., Friocourt, G., et al. (2020b). Mechanism of cystathionine- β -synthase inhibition by disulfiram: The role of bis(N, N-diethyldithiocarbamate)-copper(II). *Biochem. Pharmacol.* 182, 114267. doi:10.1016/j.bcp.2020.114267

Glossary

ALDH acetaldehyde dehydrogenase ALF acute liver failure ALT alanine aminotransferase APAP acetaminophen AST aspartate aminotransferase ATP adenosine triphosphate Bcl-2 B-cell lymphoma 2 CBS cystathionine beta-synthase CSE cystathionine gamma-lyase DOX doxorubicin ECAR glycolysis extracellular acidification rate ECM extracellular matrix eNOS endothelial nitric oxide synthase GalN D-galactosamine H₂S hydrogen sulfide HCC hepatocellular carcinoma HFD high-fat diet HO-1 heme oxygenase-1 HSC hepatic stellate cells ICAM intercellular adhesion molecule IDO1 indoleamine 2,3-dioxygenase 1 IRI ischemia/reperfusion injury

KO knockout LPS lipopolysaccharide LSEC liver sinusoidal endothelial cell MDA malondialdehyde mic-PS microplastics MPO myeloperoxidase MTX methotrexate NAFLD non-alcoholic fatty liver disease NASH non-alcoholic steatohepatitis NQO1 NAD(P)H:quinone oxidoreductase 1 Nrf2 nuclear factor-E2-related factor OCR oxygen consumption rate ROS reactive oxygen species **RSS** reactive sulfur species SAC S-allyl-cysteine SOT solid organ transplantation STS sodium thiosulfate TAA thioacetamide TME tumor microenvironment TNF-α tumor necrosis factor-α TRPV1 transient receptor potential vanilloid 1 3-MST 3-mercaptopyruvate sulfotransferase