



Nanoapproaches to Modifying Epigenetics of Epithelial Mesenchymal Transition for Treatment of Pulmonary Fibrosis

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Idiopathic Pulmonary Fibrosis (IPF) is a chronically progressive interstitial lung that affects over 3 M people worldwide and rising in incidence. With a median survival of 2–3 years, IPF is consequently associated with high morbidity, mortality, and healthcare burden. Although two antifibrotic therapies, pirfenidone and nintedanib, are approved for human use, these agents reduce the rate of decline of pulmonary function but are not curative and do not reverse established fibrosis. In this review, we discuss the prevailing epithelial injury hypothesis, wherein pathogenic airway epithelial cell-state changes known as Epithelial Mesenchymal Transition (EMT) promotes the expansion of myofibroblast populations. Myofibroblasts are principal components of extracellular matrix production that result in airspace loss and mortality. We review the epigenetic transition driving EMT, a process produced by changes in histone acetylation regulating mesenchymal gene expression programs. This mechanistic work has focused on the central role of bromodomain-containing protein 4 in mediating EMT and myofibroblast transition and initial preclinical work has provided evidence of efficacy. As nanomedicine presents a promising approach to enhancing the efficacy of such anti-IPF agents, we then focus on the state of nanomedicine formulations for inhalable delivery in the treatment of pulmonary diseases, including liposomes, polymeric nanoparticles (NPs), inorganic NPs, and exosomes. These nanoscale agents potentially provide unique properties to existing pulmonary therapeutics, including controlled release, reduced systemic toxicity, and combination delivery. NP-based approaches for pulmonary delivery thus offer substantial promise to modify epigenetic regulators of EMT and advance treatments for IPF.

Keywords: fibrosis, epigenetics (MeSH), nanomedicine, epithelial mesenchymal transformation, bromodomain-containing protein 4, lung, aerosol - therapeutic

INTRODUCTION

The Interstitial Lung Diseases (ILDs) are a heterogeneous group of parenchymal diseases with various mixtures of fibrosis and inflammation. Idiopathic Pulmonary Fibrosis (IPF) is the most common and aggressive ILD subtype. IPF is a lethal diagnosis that leads to progressive dyspnea, reduced exercise capacity, and has a median life expectancy of 2–3 years after diagnosis (Raghu et al., 2011). Although there is substantial variability in case definition, comprehensive studies indicate that IPF affects over 3 M people worldwide and the incidence is increasing, particularly in aging populations (Hutchinson et al., 2015).

Mechanistically, it is thought that environmental insults (e.g. cigarette smoking, silicates and burning biofuels) in a genetically predisposed population trigger a chronic epithelial injury-repair response, disrupting alveolar stem cell populations and leading to epithelial growth factor (EGF) stimulation. In response to these factors, epithelial cells undergo cell-state changes to acquire mesenchymal characteristics and promote quiescent fibroblasts to transition into active extracellular matrix (ECM) production. Myofibroblasts perpetuate epithelial injury and further promote ECM deposition resulting in obliteration of alveolar spaces (Torr et al., 2015). Because myofibroblast numbers are inversely related to survival in human IPF (King et al., 2001), and IPF fibroblasts cause lung fibrosis in humanized mice (Geng et al., 2019), therapies reducing myofibroblast activity have emerged as a focus of recent therapeutic approaches.

Several studies have implicated the atypical histone acetyltransferase (HAT) known as bromodomain-containing protein 4 (BRD4) in the EMT and transforming growth factor (TGF)-induced myofibroblast transdifferentiation. In combination with preclinical studies *in vivo* that have observed small molecule BRD4 inhibitors interfere with fibrosis in mouse models of IPF, including bleomycin model (Tang et al., 2013), radiation induced fibrosis (Wang et al., 2018) and TGF- β -induced fibrosis (Tian et al., 2016), BRD4 pathway is a mechanistically relevant and a biologically important mediator of inducible lung fibrosis. However, BRD4 is implicated in a variety of essential cellular functions, including chromatin organization, DNA-damage repair, and post-mitotic gene expression (Dey, 2003; Devaiah et al., 2016b). A major challenge with advancing epigenetic regulators for the treatment of IPF is how to provide controlled delivery to myofibroblastic foci.

In this review, we describe the mechanistic pathways how epithelial inflammation/injury promotes EMT and myofibroblast expansion and the evidence that BRD4 is an epigenetic regulator of EMT and myofibroblast transdifferentiation. Finally, we propose how nanotechnology can advance therapeutics for IPF and ILD. Encapsulation in nanoparticles (NPs) enables selective drug delivery via well-accepted aerosol administration while enhancing overall pharmacokinetics. We propose that nanomedicine can advance the therapeutics of IPF by enhancing therapeutic efficacy and reducing systemic effects, thereby facilitating the development of the next generation of therapeutics for pulmonary fibrosis treatment.

THE ROLE AND REGULATION OF FIBROSIS

The Injury-Inflammation-Fibrosis Paradigm

Fibrosis is defined as pathological state in which tissue and organ function is hindered due to excessive scar formation during the process of wound healing, often associated with a chronic inflammatory state (Wilson and Wynn, 2009). The fibrogenic response usually consists of several phases: 1) activation of the initial response due to organ injury, 2) stimulation of fibroblasts by inflammatory mediators, and 3) deposition of ECM components, resulting in permanent scarring (Mack, 2018).

In the second phase, after the initial injury, inflammatory and profibrogenic cytokines are released such as those from T helper type (Th)-2 cells (IL-4, IL-5, IL-10 and IL-13) or Th-1 cells (IL-2, IFN- γ and TNF). Interleukin (IL)-4 is notable in several studies that the deletion of IL-4 reduces fibrosis in the lung (2). Encompassing a wide range of biological and immunosuppressive activities, IL-4 can activate B cells, induce differentiation of naive CD4⁺ T cells, contribute to the development of M2-type monocytes or macrophages, and inhibit expression of several proinflammatory mediators (e.g., TNF- α ; IL-1, 6, 8,12) (Zhu, 2015). Notably, in bleomycin-induced lung fibrosis, IL-4 was found to suppress inflammation, but was profibrotic. Another important profibrotic cytokine is IL-13 (Karo-Atar et al., 2016). IL-13 has the ability to directly and indirectly activate fibroblasts along with tumor necrosis factor (TNF)- α and IL-4, to produce an upregulation of IL-13Ra2 on macrophages, allowing IL-13 to bind and induce the release of TGF β (Mack, 2018).

Contrary to the effects of Th-2 cytokines, Th-1 cytokines can be both anti- and profibrotic. Classically, the adaptive immune response is triggered when IL-12 interacts with interferon gamma (IFN- γ); it is also important to note that macrophages can also express IFN- γ in a response to microbial stimuli (Zhu, 2015). Once, activated, IFN- γ and IL-12 combine with TGF β to induce collagen (COL) degradation and for ECM remodeling. In contrast, Th-1 polarizing cytokines IL-6, IL-1 β , and TNF- α , are released from monocytes triggering a latent transcription factors such as STAT3 and STAT1, leading to fibroblast migration and recruitment, promoting proliferation and differentiation into myofibroblasts (Mack, 2018).

Myofibroblasts

Myofibroblasts are not normally present in substantial numbers in normal adult tissues but their population rapidly expands in response to injury-repair. In normal conditions, tissue fibroblasts have little secretory activity. However, upon injury, fibroblasts actively produce ECM components [collagen (COL) and fibronectin (FN)] causing a cascade of cytokine release, resulting in chemotaxis of inflammatory cells (Hinz et al., 2007). ECM remodeling triggers local fibroblasts to produce contractile stress fibers made out of cytoplasmic actins, transforming them into “proto-myofibroblasts.” In the presence of high ECM stress, TGF β and FN splice products causing the phosphorylation of focal adhesion kinases (Phan, 2012), the proto-myofibroblast assumes a stable phenotype characterized by α -SMA replacing actin in the stress fibers

(Hinz et al., 2007). Activated myofibroblasts are characterized by a spindle-like morphology with intracytoplasmic stress fibers, a contractile phenotype, expression of α -smooth muscle actin (α -SMA), transgelin (TAGLN/SM22), COL and hyaluronans. Expression of these contractile fibers by myofibroblasts enables wound contraction, and results in wound closure.

When the tissue repair process is completed, myofibroblasts disappear through apoptosis, a process driven by ECM softening (stress release) or pro-apoptotic factors such as IL-1 β , fibroblast growth factor 1, and prostaglandin E2. However, under pathological conditions, such as pulmonary fibrosis, cirrhosis or hypertrophic scar, myofibroblasts persist in fibroblastic foci, producing excess scar and interfering with normal organ function. Myofibroblasts are derived from multiple mesenchymal types. In the lung, myofibroblasts are thought to derive from locally activated tissue fibroblasts, vascular pericytes or recruitment of bone-marrow derived fibrocytes (Hinz et al., 2007; Rock et al., 2011; Phan, 2012). Alternatively, it has been shown that injured epithelial cells can express mesenchymal intermediate filaments (vimentin) and α -SMA through a process of EMT, producing a myofibroblast-like phenotype (Radisky et al., 2007). In cancer, a substantial population of myofibroblasts are thought to arise from EMT. However, because myofibroblast populations are distinct in different organs, the contributions of EMT directly to the lung myofibroblast population is controversial (Rock et al., 2011).

PULMONARY FIBROSIS AND EPITHELIAL MESENCHYMAL TRANSITION

Epithelial Response to Acute and Chronic Lung Injury

Lining the airways and the respiratory compartments, epithelial cells, are exposed to a variety of environmental challenges such as microbial agents, toxic components and particulates leading to inflammation and fibrosis. The airways and distal epithelial cells comprise of a physical and chemical barrier to maintain lung function. Activated airway epithelial cells can modulate their direct barrier function through the release of a variety of defense mechanisms, such as the release of defensins and mucins, as well as by modulating fluid transport (Fehrenbach, 2001). Epithelial cells also have the ability to release multiple mediators and adhesion molecules that can mediate and activate migratory inflammatory and immunological cells. Additionally, epithelial cells interact with other resident cells, such as fibroblasts along with the ECM. The mechanism by which epithelial cells respond to different types of lung injury, acute or chronic, varies drastically from the inflammatory response to fibrosis.

Acute lung injury or acute respiratory distress syndrome (ARDS) can be caused by a variety of factors; pneumonia, trauma, sepsis, aspiration, post-lung transplant, or certain viral infections (Johnson and Matthay, 2010). Alveolar epithelial cells are key for alveolar homeostasis and aid in the proper alveolar fluid clearance through the direction of epithelial sodium channel

(ENaC). When acute injury occurs, permeability of the alveolar-capillary membrane is caused by a disruption in the ENaC either through hypoxia such that the reoxygenation and β -agonists reverse ENaC or transcription factors and inflammation decrease the number of ENaC molecules (Lutter and Spiteri, 2005). When this occurs, fluid enters the alveoli and proinflammatory cytokines (IL-6 and IL-8) are released causing the release of proinflammatory cells such as neutrophils and macrophages. If not treated, this cascade develops into hypercytokinemia releasing IL-1 and TNF- α which will eventually lead to impaired fibrotic repair and lung failure (Johnson and Matthay, 2010).

Differing from acute lung injury, environmental exposures of particulates and chemicals is a much more common cause of chronic epithelial damage. Particulate matter generated from cigarette smoking, diesel emissions, biofuel exposures (including burn pit exposures) contain mixtures of metal, dusts, byproducts of hydrocarbon combustion induce oxidative cellular and DNA damage. These environmental exposures induce alveolar injury and clinical exacerbations in patients with existing chronic airway disease, including IPF, asthma and COPD (Winterbottom et al., 2018). Although the initial mechanism of action is similar in that there is damage to the alveolar wall that causes recruitment of proinflammatory cells, the gradual injury of the epithelium allows time for airway remodeling (Mercer et al., 2006). In asthma and COPD, epithelial cells are exposed to a range of inflammatory mediators over a prolonged period affecting the epithelial cell integrity and metabolism (Athanasio, 2012). Changes in cellular composition of the epithelium can cause goblet cell hyperplasia and pathophysiological of the mucous glands, affecting the function of the airway mucosa (Lutter and Spiteri, 2005).

Epithelial Mesenchymal Transition and the Myofibroblast Expansion

EMT is a multi-step, reversible process observed in several biological contexts: embryonic development (type I EMT), malignant transformation (Type III EMT), and in normal tissues undergoing injury response (Type II EMT). For the purposes of this review, we will focus primarily on type II EMT. Type II EMT begins when oxidative or physical injury to the resting differentiated epithelial cell triggers a phenotypic change characterized by loss of apical-basal polarity and loss of intercellular tight adherens junctions (Kalluri, 2009; Hill et al., 2019). The cells de-differentiate and become motile, enabling the cells to repopulate the injured epithelial surface. EMT can be initiated by the local production of signaling factors; most notable are TGF- β , fibroblast growth factors, EGFs, and the Wnt/ β -catenin signaling pathways (Ijaz et al., 2014). These pleiotropic signaling factors induce the expression of mesenchymal transcription factors such as Snail, Slug, and Twist whose actions repress epithelial genes (E-cadherin) and stimulate mesenchymal genes (N-cadherin, α -SMA, vimentin and fibronectin) (Hill et al., 2019). Consequently, the apical-basal polarity will switch to a front-back polarity in which the tight adherens junctions turn into N-cadherin junctions and vimentin stress fibers, promoting motility.

TABLE 1 | A summary of factors and genes in myofibroblast expansion.

Factors	Representative genes/Pathway	Action
Fibroblast and transforming growth factor signals Changes in ECM stiffness (COL, MMP deposition) Mesenchymal transcription factors Master transcription factors Superenhancers	FGFs, EGFs, and wnt/ β -catenin pathway Integrin activation Snai1, slug, and twist ETS2, HNF4a, JUNB, NF κ B BRD4	Initiates EMT Transition from proto- to myofibroblast Reprogram epigenome: repress epithelial genes and stimulate mesenchymal genes Control partial-EMT state Control expression of gene regulatory networks specifying cell identity

Systems-level studies have led to the conclusion that EMT is a multi-step, dynamic process produced by cooperative effects of master transcription factors in response to EGF and ECM cues. Time series RNA profiles have identified cliques of master transcription factors (such as ETS2, HNF4a and JUNB) that cooperate to control partial-EMT state, as summarized in **Table 1**. These transcription factors control cell type by producing the formation of superenhancers that control mesenchymal gene-regulatory networks. Superenhancers are clusters of functional enrichers enriched in RNA Polymerase, Mediator and acetylated histones that control cell-type specification genes (Whyte et al., 2013). Of relevance here, superenhancers are stabilized by the presence of BRD4 (Shin, 2018). In studies using siRNA-mediated gene knockdowns and small-molecule BRD4 inhibitors, TGF- β -induced partial EMT was reversed. Overall, this data demonstrates that BRD4 maintains epigenetic memory of TGF β stimulation and the EMT phenotype. While EMT is traditionally defined as the conversion of epithelial cells into mesenchymal-like cells, recent studies expanded this cellular reprogramming process to mediate changes in epithelial morphology, motility, and gene expression (Lamouille et al., 2014).

Epithelial Mesenchymal Transition and Myofibroblast Expansion: The “Epithelial Injury” Hypothesis

Under normal conditions, the differentiated epithelial phenotype and the mucosal barrier function is maintained through interactions with the basal lamina and supporting subepithelial fibroblasts, known as the epithelial mesenchymal trophic unit (Evans et al., 1999). Upon exposure to environmental oxidants or insults, distal epithelial injury stimulates activation of the unfolded protein response, and release of fibrogenic growth factors. The consequent EMT transition results in secretion of alternative splice variant of ECM proteins, including FN. Changes in ECM composition (stress) are an important signal for pulmonary mesenchymal cell populations to acquire myofibroblast properties (Tomasek et al., 2002).

Myofibroblasts release inflammatory/profibrotic mediators that perpetuate epithelial injury and further promote ECM deposition (Zhang et al., 1994; Torr et al., 2015). The effects of TGF- β and fibrogenic cytokines further stimulate prosurvival signaling in the myofibroblast population, leading to persistence, resistance to apoptosis and migratory invasiveness (Horowitz et al., 2006). With myofibroblast persistence, deposition of ECM stiffens the lungs, reducing normal elastic properties and pulmonary function. Not only is this cell type primarily responsible for producing ECM and interstitial fibrosis (Hinz et al., 2007; Rock et al., 2011; Hung et al., 2013), these cells form the pathognomonic fibroblastic foci of human IPF (Hinz et al., 2007; Thannickal, 2012). Clinically in IPF, increased myofibroblast numbers are inversely related to survival (King et al., 2001). Therapeutics targeting the myofibroblast foci are therefore an exciting approach for treatment of ILDs.

In IPF, depletion of alveolar epithelial stem cell populations from mitochondrial dysfunction results in tonic oxidative stress,

alveolar cell dysfunction, impaired renewal, senescence and production of profibrotic factors. Compensatory alveolar stress response promotes EMT and myofibroblast transdifferentiation (Kulkarni et al., 2016). Although epithelial cells appear not to be a major source of lung myofibroblasts in IPF, this is not to say that epithelial transition is not a major driver of fibroblastic foci. Injured epithelial cells are a source of TGF β and COL secretion that trigger myofibroblast formation (Table 1). Recent single cell sequencing studies have identified a keratin (KRT)5-/KRT17+ pathologic, ECM-producing epithelial cell population that was highly enriched in IPF lungs (Habermann et al., 2020). Interestingly these epithelial cells exhibited EMT signatures. This finding supports the prevailing epithelial injury hypothesis, illustrated in Figure 1.

New Linkages of Epithelial Innate Signaling on Fibrosis Programs

In addition to environmental oxidant exposure, an emerging body of work has linked respiratory virus infections with epithelial injury and remodeling. Airway remodeling is a collective term that refers to structural changes in the airways resulting in enhanced collagen deposition in the subepithelial basement membrane (lamina reticularis), disruption of the epithelial barrier, epithelial cell-state change (mucous metaplasia and/or mesenchymal transition), and smooth muscle hypertrophy (Bergeron et al., 2010). Collectively, this process narrows the small airways, producing obstruction and reduced lung compliance accounting for enhanced morbidity and mortality (Hough et al., 2020). Respiratory viruses, such as Rhinovirus and Respiratory Syncytial Virus are major triggers of remodeling and chronic lung disease (Jartti and Gern, 2017). These agents primarily replicate in airway epithelial cells producing innate immune response-driven remodeling (Brasier, 2019; Brasier, 2020).

More recently, COVID-19 ARDS has been observed to be presaged by a hyper-inflammatory state characterized by the presence of circulating inflammatory and pro-fibrotic cytokines including IL-6, IL-7, GM-CSF, MCP, and MIP1 α (Ruan et al., 2020). In COVID-19 infections, circulating ECM components (hyaluronans, type III pro-collagen and laminin) are associated with increased with disease severity (Ding et al., 2020). These data suggest that COVID-19 also induces ECM remodeling, a process important in myofibroblast transdifferentiation. In survivors, radiographic evidence of pulmonary scarring (Shi et al., 2020) and functional defects in lung function (diffusion capacity) are commonly seen (Mo et al., 2020; Shi et al., 2020; Wang et al., 2020). In fatal cases, increases in the TGF-ECM networks, including upregulated TGFBR2, COL isoforms, and FN (12) are seen, and pulmonary fibrosis is found upon an autopsy (Zhang T. et al., 2020). These data suggest that fibrosis and impaired pulmonary function is a sequelae of severe COVID-19-ARDS infections. Although the relationship of post-COVID-19 lung scarring/fibrosis with chronic progressive, ILD is not known due to the short-term follow-up, it is significant that post-infectious pulmonary fibrosis is also a known outcome in survivors of SARS, an infection with the closely related virus, SARS-CoV (Hui et al., 2005). We interpret these studies to

indicate that viral epithelial infections may be an important modulator of airway fibrosis.

Mechanisms How Innate Inflammation Produces Epithelial Mesenchymal Transition

Chronic oxidative stress induced by innate signaling triggers adaptive cell-state transitions of the normal epithelium to activate the EMT program. High throughput RNA-seq studies of normal human airway cells have discovered that the core regulatory network of ~3,000 TGF β -induced genes, substantially overlaps with the NF κ B network (Tian et al., 2015). Using selective inducible knockouts in bronchiolar-derived basal cell progenitors, it was found that NF κ B signaling drives the transition to a committed mesenchymal phenotype (Tian et al., 2018c). These findings led to the discovery that tonic (or repetitive) NF κ B signaling in the airway produces EMT *in vivo* (Tian et al., 2017a). Repetitive AEs provoked by exposure to TLR3 agonists activates SNAI expression, EMT, remodeling and expansion of myofibroblast population(s).

Virus-induced NF κ B complexes with the coactivator bromodomain containing (BRD4), a multifunctional protein with intrinsic RNA polymerase kinase (Huang, 2009; Devaiah et al., 2012) and atypical HAT (Huang, 2009; Devaiah et al., 2016a) activity. Through its sequence specific DNA-binding activity, NF κ B redistributes BRD4 to innate inflammatory genes as well as the core EMT regulators SNAI1, ZEB1 and TWIST1 (Brown et al., 2014; Tian et al., 2016), as schematically shown in Figure 2. NF κ B-dependent activation of the core EMT regulators results in expression of COL and FN, key ECM regulators of myofibroblast expansion, airway remodeling and expansion of the subepithelial basement membrane. Supporting this chromatin remodeling mechanism, recent analyses using transposase-accessible-next generation sequencing (ATAC-Seq) in highly differentiated lower airway epithelial cells infected with Respiratory Syncytial Virus demonstrate that the TGF β -FN-JUN pathways controlling EMT are activated by chromatin decondensation at their proximal promoters (Xu et al., 2020).

Although these fundamental studies of the NF κ B-BRD4 pathway were initially developed *in vitro*, evidence that NF κ B mediates mesenchymal transition, myofibroblast expansion, and pulmonary fibrosis has been developed in *in vivo* (Tian et al., 2016). In mice, acute TLR3 activation induces CXCL8/IL-8 expression and neutrophilia, but chronic TLR3 activation produces airway epithelial mesenchymal transition, expansion of the myofibroblast population, and fibrosis (Tian et al., 2016). Activation of the BRD4 kinase and atypical HAT activity in the airway mucosa has been demonstrated in response to TLR3 activation (Tian et al., 2017a) and viral replication (Tian et al., 2017b; Tian et al., 2018d). Here, the unique, BRD4-dependent histone acetylation on Lys 122 is induced by virus or allergen, and this induction is NF κ B-dependent. Viral and allergen-induced NF κ B-BRD4 complex formation was been demonstrated through proximity ligation assays, a technique to detect molecular interactions between the two proteins. Finally, using highly selective BRD4 inhibitors, it was found that inhibition of BRD4 HAT activity interferes with remodeling, airway

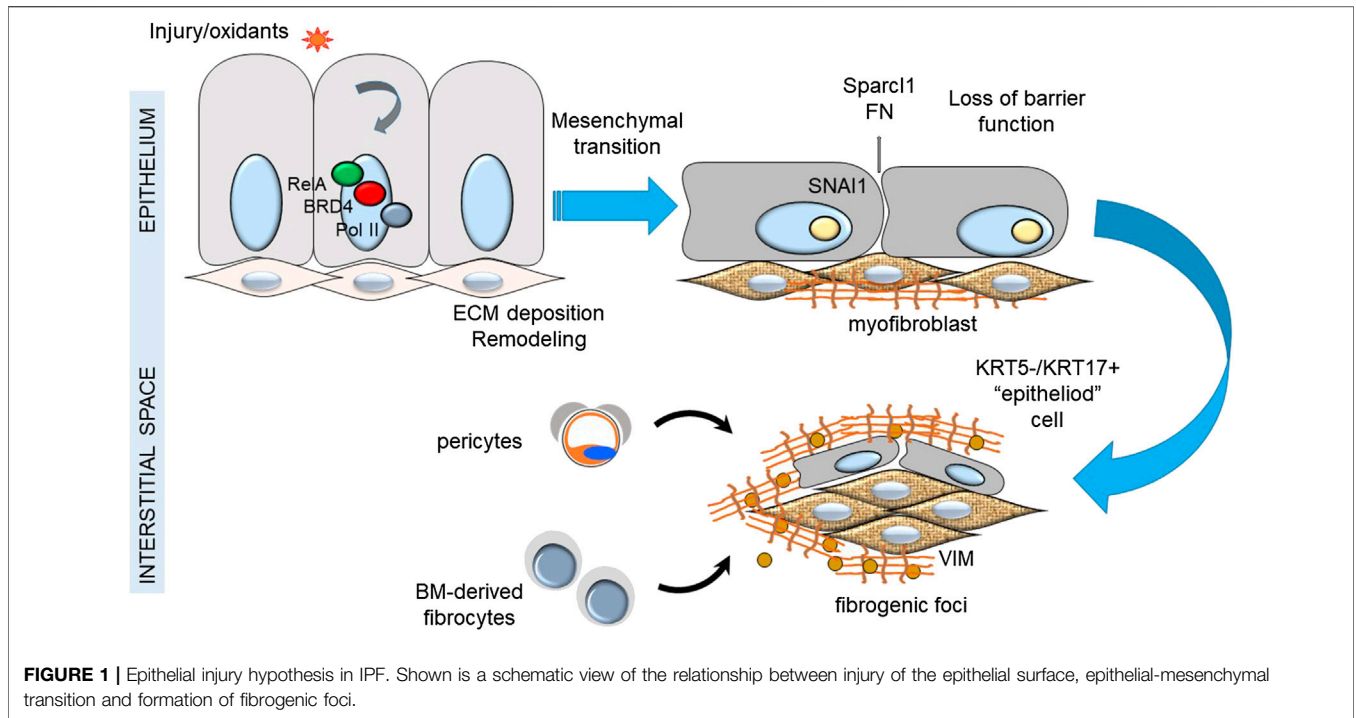


FIGURE 1 | Epithelial injury hypothesis in IPF. Shown is a schematic view of the relationship between injury of the epithelial surface, epithelial-mesenchymal transition and formation of fibrogenic foci.

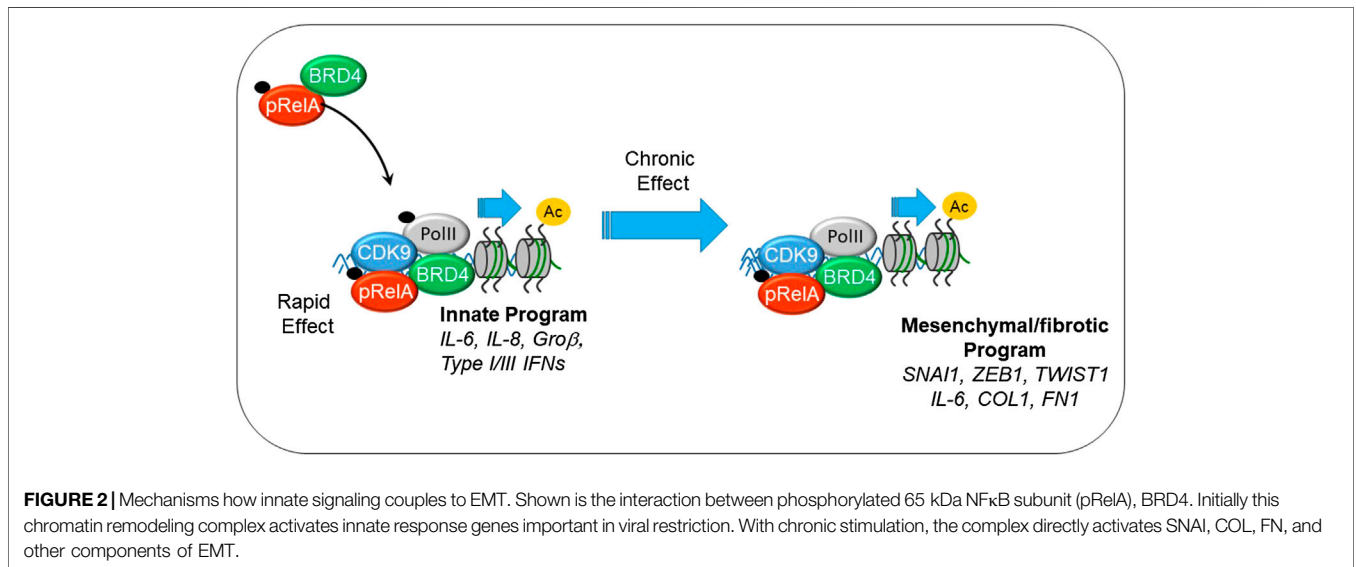


FIGURE 2 | Mechanisms how innate signaling couples to EMT. Shown is the interaction between phosphorylated 65 kDa NFκB subunit (pRelA), BRD4. Initially this chromatin remodeling complex activates innate response genes important in viral restriction. With chronic stimulation, the complex directly activates SNAI1, COL, FN, and other components of EMT.

hyperreactivity and myofibroblast transition *in vivo* (Tian et al., 2018a). These studies validate the central relevant of the NFκB-BRD4 pathway in airway remodeling.

Histone Modifications Underly Epithelial Mesenchymal Transition

In recent years, epigenetics has become the forefront as a major mechanism underlying fibrosis. The term epigenetics refers to altering gene expression without altering the DNA sequence. Extensive research aided by deep sequencing technology, has

identified histone codes to be of interest in EMT. In eukaryotic cells, DNA is condensed in chromatin with the nucleosome, a major part of chromatin, comprised of four different histones to form an octamer (H3, H4, H2A, and H2B) (O'Reilly, 2017). Although histones are globular in nature, they have a "tails" that can possess multiple histone modifications, eight of which are known. Histone tail modifications include methylation, phosphorylation, acetylation, ubiquitination, sumoylation, citrullination, and ADP-ribosylation (O'Reilly, 2017). Additionally, histone modifications are mediated by a family

of enzymes: Histone Deacetylases (HDACs), and HATs. HDACs remove acetyl groups on N terminal lysines, whereas HATs catalyze their addition.

First discovered in hepatic fibrosis, HDACs were shown to enhance the migratory and ECM producing properties of tissue myofibroblasts (de Ruijter et al., 2003). This idea was expanded in pulmonary fibrosis, where HDACs were extensively studied for their role in COL secretion. Inhibition of HDAC induced hyperacetylation in H3 and H4 tails. In idiopathic pulmonary fibroblasts, elevated expression of all class I (HDAC 1, 2, 3, and 8), class IIa (HDAC 4, 5, 7, 9) and class IIb (HDAC 6 and 10) HDACs were discovered to be in both tissue and isolated myofibroblasts (de Ruijter et al., 2003). During hypoxia, another regulator of EMT, cross-talk between chromatin modifiers HDAC3 and WDR5 regulates repression of epithelial genes and activation of mesenchymal genes (Lin and Wu, 2020). The histone mark histone 3 lysine 4 acetylation (H3K4Ac) was shown to bind promoters of several EMT marker genes (e.g., CDH1 and VIM) (Wang J. Q. et al., 2019). Additionally, both H3K4me3 (activator) and H3K27me3 (repressor) were observed in the EMT promoters (Lin and Wu, 2020). This is of much interest as the presence of activation and repressor histone marks allows EMT markers to cycle from expressed to silenced in response to dynamic changes in the extracellular environment. Furthermore, studies using chromatin immunoprecipitation with anti-H3K4Ac antibodies and whole genome sequencing identified new EMT marker genes, glioma-associated oncogene homolog 1 (GLI1) and smoothened (SMO), that are regulated by HDAC3 for in migration and invasion (Wang J. Q. et al., 2019). Although there has been great progress in the use of HDAC-inhibitors demonstrating anti-fibrotic activity in mechanistic studies, the advancement of HDAC inhibitors (HDACi) as therapeutics for IPF has been challenging, discussed below.

The ubiquitous HAT, P300, regulates hundreds of transcription factors such as TGF- β 1 and can cause the hyperacetylation of H4 on the COL promoter. In response to changes in cell substrate stiffness, activated P300 accumulates in the nucleus, enhancing Smad dependent transcription (Dou et al., 2018). Smad2/3 binds directly to gene promoters to induce the transcription of profibrotic molecules, including α -SMA, COL1 and tissue inhibitor of metalloprotein, inducing myofibroblast activation and ECM deposition (O'Reilly, 2017).

A body of work has shown that transdifferentiation, anti-apoptotic activity, ECM production and migration of epithelial-like mesenchymal cells is mediated by BRD4 (Devaiah et al., 2016a). BRD4 is a member of a family of bromodomain and extraterminal (BET) proteins, including BRD2, BRD3 and BRD4 that share two tandem bromodomains (BDs) and an extra-terminal domain, from which the family was named (Taniguchi, 2016). These paralogs are thought to have arisen during a distal evolutionary duplication event. Although the functions of the BET isoforms are tissue specific and pleiotropic, BRD4 activity has been implicated in cell cycle regulation DNA repair, innate inflammation (Brasier et al., 2011; Taniguchi, 2016), and cell-state transition (Chang et al., 2016; Tian et al., 2017a); all processes that play important roles in lung diseases.

Recent work has identified a coenzyme A -dependent atypical histone acetyl transferase activity (HAT) (Devaiah et al., 2016a) and N terminal kinase activity directed for the COOH terminus of RNA Polymerase II (Devaiah et al., 2012). Through these activities, BRD4 plays a central role in the maintenance of cell-type specifying superenhancers, controlling mesenchymal gene regulatory programs by transcriptional elongation discussed above. Small molecule inhibitors and siRNA mediated knockdown experiments have shown that BRD4 plays a central role in myofibroblast transdifferentiation, mediating TGF- β -induced NOX4- α SMA expression, contractility and motility as well (Ijaz et al., 2017). In this way BRD4 mediates both EMT and fibroblast-myofibroblast transdifferentiation.

Deoxyribonucleic Acid Methylation Regulates Epithelial Mesenchymal Transition

Environmental particulate matter emitted by traffic-related air pollution; polyaromatic hydrocarbons, black carbon, ozone (O₃), and nitrogen oxides (NOx), have been shown to be associated with changes in DNA methylation (DNAm). DNAm, one of the most well-known mechanisms of epigenetic gene regulation, is described as the attachment of methyl groups to DNA, commonly at the fifth carbon of cytosines, leading to the formation of 5-methylcytosine (5-mC) (Moen et al., 2015). Methylation on the genome predominantly occurs at C-G dinucleotides (CpGs), resulting in the initiation of transcription, elongation efficiency, and alternative splicing.

Tumor hypoxia, a known inducer of EMT through hypoxia-inducible factor-1 α (HIF-1 α), can directly dysregulate the activity of ten-eleven translocation DNA demethylases. A decrease in DNA demethylation triggers the accumulation of DNA methylation driven by DNA methyltransferases (DNMT) to affect genes encoding cell adhesion functions that are involved in EMT. It has been suggested that DNA methyltransferase 1 associated protein 1 (DNMT1) can directly interact with Snail and suppress E-cadherin in epithelial cells and subsequent activation of epithelial-mesenchymal transition or cell death (Espada et al., 2011). Studies have shown that traffic-related air pollution and smoking can cause increased DNMT1 protein accumulation leading to misexpression of genes to cause cancer.

THERAPEUTIC STRATEGIES FOR PULMONARY FIBROSIS

Glucocorticoids

Glucocorticoids such as cortisone, hydrocortisone, and prednisone, have historically been used to treat lung inflammation. These drugs act by decreasing inflammation both, directly through the glucocorticoid receptor and indirect mechanisms (Barnes, 2009). However, all potent glucocorticosteroids cause side effects such as glaucoma, high blood pressure, psychological effects, osteopenia, and acne

(Newman, 2018). Previously, it was thought that pulmonary fibrosis was a product of immune activity. Therefore, corticosteroids were combined with azathioprine or cyclophosphamide and a mucolytic/antioxidant, N-acetylcysteine. These studies were stopped due to negative effects in varying patient groups and are no longer used (Newman, 2018).

Although inhaled corticosteroids have been effective for several lung diseases such as cystic fibrosis, non-specific interstitial pneumonia (NSIP) and inflammatory diseases such as asthma and COPD, however, it is not effective with IPF (Mapel et al., 1996).

In 2014, oral nintedanib, was a breakthrough therapy approved by the FDA. Nintedanib is a broad spectrum small-molecule inhibitor of receptor tyrosine kinases that reduces fibroblast proliferation and activation. Nintedanib has been subjected to a number of randomized controlled clinical trials. These studies have shown that nintedanib slows the decrease in forced vital capacity (FVC) from IPF and other progressive fibrosing ILDs (Richeldi et al., 2014; Mazzei et al., 2015; Flaherty et al., 2019). Pirfenidone was also licensed in the same year. Pirfenidone is a small molecule with antifibrotic, anti-inflammatory and antioxidant effects that inhibits TGF- β -stimulated collagen production. Several prospective studies have found that pirfenidone also slows the loss of FVC in IPF and improves progression-free survival (Spagnolo et al., 2010). Although these medications represent a substantial advancement for the treatment of IPF, they neither reverse established fibrosis nor are curative.

Histone Deacetylase Inhibitors

Trichostatin A (TSA), a non-selective HDACi, has been shown to attenuate pulmonary fibrosis in several models. Bleomycin-induced pulmonary fibrosis (IPF), is characterized by progressive fibrosis leading to end-stage lung disease and respiratory failure. In the bleomycin model, TSA prevented WIF-1 loss, an important Wnt signaling regulator, resulting in a reduction of collagen overexpression through b-catenin (Yoon et al., 2019). TSA has also been shown to block TGF- β mediated ECM production in corneal fibroblasts. In a separate study, epigenetic defects in COX-2 expression were restored by HDAC inhibition, protecting from pulmonary fibrogenesis (Coward et al., 2009). Although there are currently many ongoing studies with the use of TSA and the reduction of fibrosis *in vivo* with mice and rats, there has yet to be any significant human clinical trials with this drug related to pulmonary fibrosis. A major limitation of HDACi is the lack of isoform specificity and concerns with long-term toxicity (Royce and Karagiannis, 2014), including vascular calcification (Kwon and Kim, 2017).

Bromodomain-Containing Protein 4 Inhibitors

A number of studies have implicated BRD4 as a potential therapeutic target for modifying epigenetic responses in lung disease. It has been shown that inhibition of BRD4 blocks EMT

(Chang et al., 2016) and fibroblast-myofibroblast transdifferentiation *in vitro* (Ijaz et al., 2017). Highly selective BRD4 inhibitors have the ability to block allergen, viral, and growth factor induced remodeling and fibrosis in small animal models *in vivo* (Tian et al., 2016; Tian et al., 2018d; Zhao et al., 2019). Moreover, inhibition of BRD4 interferes with inflammation-induced pericyte-myofibroblast transition *in vivo* and leakage of plasma proteins into the alveolar space (Zhao et al., 2019). Finally, BRD4 activation in myofibroblasts from humans with IPF, and small molecule BRD4 inhibitors can prevent interstitial fibrosis in the bleomycin lung injury model (Tang et al., 2013). Radiation induced lung fibrosis (Wang et al., 2018) and fibrosis mediated by TGF β administration (Tian et al., 2016).

The medicinal chemistry of BRD4 inhibitors has been rapidly advancing (Liu et al., 2018; Tian et al., 2018b; Brasier and Zhou, 2020; Liu et al., 2020). Recently, highly selective inhibitors of BRD4 that interact with distinct regions of the protein have been developed (Liu et al., 2020). These molecules have demonstrated efficacy and exhibit promising medicinal properties (Liu et al., 2020). However, systemic administration of BRD4 inhibitors may be problematic due to its ubiquitous expression and role in chromatin organization, cell stress and innate responses.

We contend that the advancement of epigenetic inhibitors into well-tolerated therapy for chronic lung disease will depend on advancement of strategies that can selectively deliver these agents to the diseased tissue. In the subsequent section, we will review the promise of aerosolized delivery of nanomedicines for addressing this problem.

NANOMEDICINE

NP formulations can provide a wide variety of benefits for the delivery of existing active pharmaceutical ingredients (APIs)—that is, the efficacious molecules identified previously for their anti-fibrotic properties. This includes dramatically improved API solubility, enhanced cellular internalization, controlled or stimuli-responsive release of the API, and enhanced selectivity through targeting, ultimately increasing therapeutic efficacy while reducing toxic side effects (Peer et al., 2007; Alshamsan, 2010; Menon et al., 2013; Patel et al., 2013; Guo, 2014; Menon et al., 2014; Pradhan, 2014; Wang, 2016; Kong, 2017; Song, 2017; Yang, 2018). The peripheral reactive groups of many NPs also provide the opportunity for ligand conjugation and active targeting strategies (Gu et al., 2007; Bugno et al., 2015). Additionally, NPs can provide specific benefits for pulmonary delivery. For example, NPs have been found to facilitate the solubility of APIs in certain pulmonary aerosolization solvents, such as perfluorooctyl bromide and fluorocarbons (Hsu et al., 2003; Lehmler et al., 2008). NPs are also readily internalized by vascular endothelial cells, providing accumulated quantities substantially greater than their micro-sized counterparts (typically $>1 \mu\text{m}$ in diameter) (Foster et al., 2001; Davda and Labhasetwar, 2002). However, despite these potential advantages, EMT-targeting NPs have so far been primarily utilized for anti-cancer strategies, and those targeting EMT in lung fibrosis currently remain limited in the literature. This is the case even

as EMT becomes increasingly implicated as an important pathway for pulmonary fibrosis (Willis and Borok, 2007; Naguchi et al., 2014), with key EMT-markers, such as TGF- β (Burgess et al., 2005; Fernandez and Eickelberg, 2012) and Snail (Wettstein et al., 2013), identified as promising targets for anti-fibrotic therapies.

In the following sections, we review a selection of nanomedicines formulated for pulmonary delivery organized by NP type, with a scope limited to those specifically utilizing aerosolic delivery methods. A general overview of these strategies is presented in **Figure 3**. Nanomedicines targeting EMT-mediated pulmonary fibrosis are especially focused on, with the goal of highlighting current shortcomings in this area of research. To this end, promising directions and potential considerations for future NP formulations are also identified at the end.

Liposomes

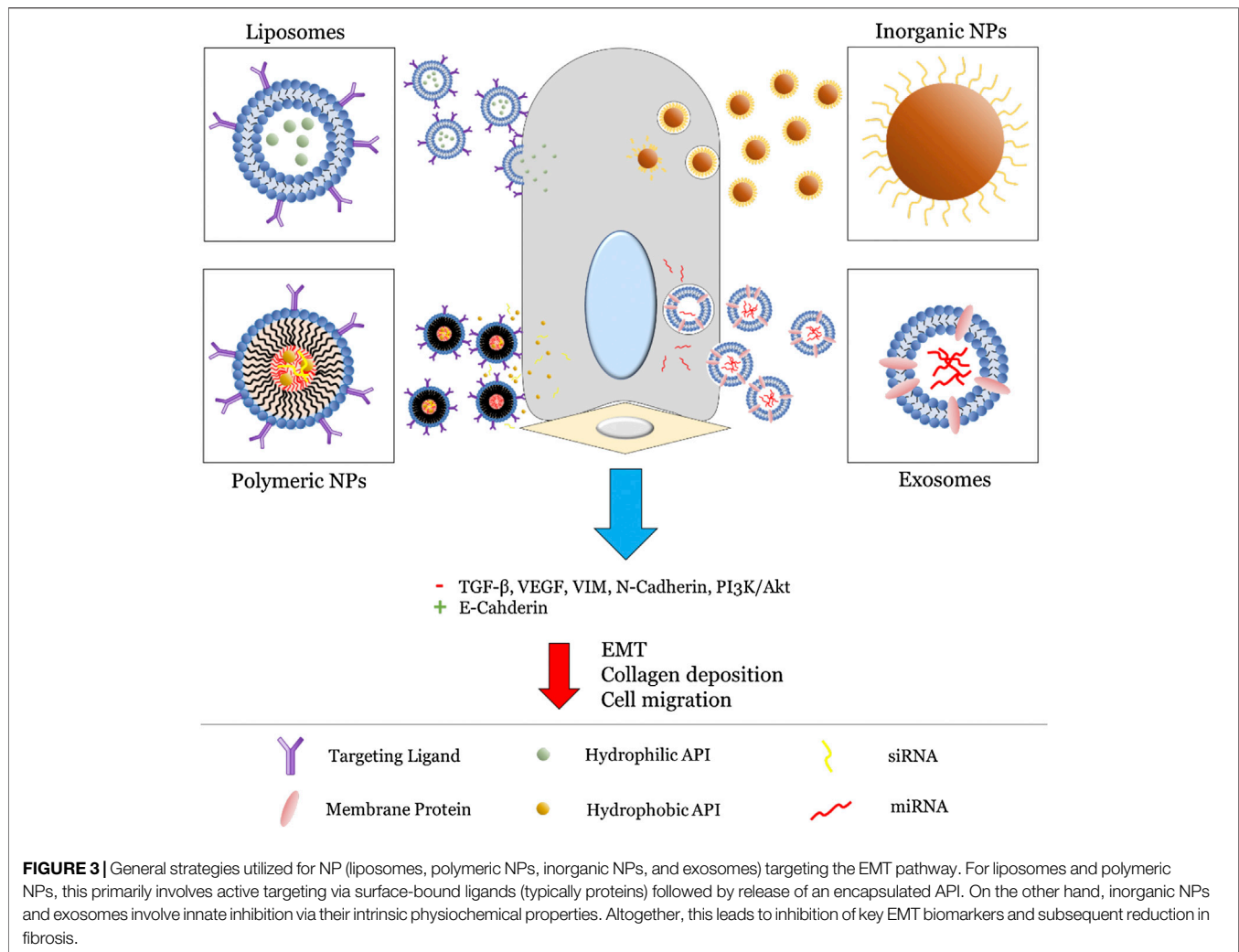
Liposomes are a promising nanocarrier for pulmonary drug delivery as they are primarily composed of natural or synthetic phospholipids either native to or compatible with lung tissues, such as phosphatidylcholine and egg phosphatidylethanolamine (Gao et al., 2013). These nanoformulations are also capable of including additional surfactants and cholesterol. Liposomes are comprised of spontaneously-formed lipid bilayer vesicles with diameters ranging from 30 nm to several micrometers, though those between 50 and 450 nm are favored for therapeutic delivery (Etheridge et al., 2013; Gao et al., 2013). Multiple forms of vesicles exist, including multilamellar vesicles (containing lamellar phase bilayers), small and large unilamellar liposomes (containing a single lipid bilayer), cochleate vesicles, and multivesicular liposomes (one vesicle containing multiple smaller vesicles). Liposomes can also be classified by preparation method, including reverse-phase evaporation and vesicle extrusion (Bozzuto and Molinari, 2015). In general, liposomes consist of a hydrophobic shell encapsulating a hydrophilic solvent. This property enables loading of both lipophilic and hydrophilic APIs with relative ease. Further structural modifications provide additional benefits, such as poly (ethylene glycol) (PEG) or deoxyribonucleic acid (DNA) anchoring to embedded cholesterol for a “stealth effect” or as biosensors (Hosta-Rigau et al., 2013), or the addition of fatty acyl chains to extend transition temperature and increase stability (Bitounis et al., 2012). Circulation time *in vivo* can also be extended by including hydrophilic carbohydrates, e.g., monosialoganglioside (G_{M1}), and polymers, e.g., PEG (Gabizon and Papahadjopoulos, 1988).

Aerosolized liposomes are well represented in the literature for pulmonary delivery, with promising results against multiple cancer models (including lung, renal, breast, and colon) (Knight et al., 1999; Koshkina et al., 2000; Skubitz and Anderson, 2000; Koshkina et al., 2001; Verschraegen et al., 2004), respiratory tract infections (RTIs) (Conley et al., 1997), immunodeficiency (Ten et al., 2002), pulmonary aspergillosis (Allen et al., 1994), cystic fibrosis (Davies et al., 2014), as a method of immunosuppression (Letsou et al., 1999), and as a delivery vehicle of insulin (Huang and Wang, 2006). Aerosolized

liposomes have also been shown to be effectively delivered by multiple common clinical methods, including air-jet, ultrasonic, vibrating-mesh, and continuous-flow nebulizers (Waldrep et al., 1994; Elhissi and Taylor, 2005; Elhissi et al., 2007), and are clinically safe in human patients (Thomas et al., 1991; Vidgren et al., 1995; Gilbert et al., 1997; Waldrep et al., 1997). This demonstrates the high biocompatibility and adaptability of liposomes for pulmonary delivery. In fact, a liposomal formulation is the only currently approved NP formulation of aerosolized drug for pulmonary delivery. This comes in the form of Arikayce, an amikacin liposome nebulized suspension antibiotic for mycobacterium avium complex (MAC) lung disease, developed by Insmid and approved in 2018 (Cipolla et al., 2013; Insmid, 2020). Other notable clinical trials are summarized in **Table 2**.

Despite the available aerosolized liposome formulations for pulmonary delivery, comparatively few utilize an EMT-targeted approach. These are summarized in **Table 3**. The majority of these formulations utilize APIs loaded into the liposome vesicle, including PGE2 (also known as dinoprostone, an oxytocic drug used in pregnancies for smooth muscle contradictions) (Ivanova et al., 2013), paclitaxel (an anti-cancer cytoskeletal drug that targets tubulin) (Zhou et al., 2016; Jin et al., 2019), and simvastatin (a lipid-lowering drug typically used for heart defects) (Jin et al., 2019). One formulation, however, used hyaluronic acid (a ligand of CD44) embedded in the liposome bilayer as both the API and a targeting moiety for overexpressed CD44 (Pandolfi et al., 2019). In all cases, biomarkers of the EMT pathway were subsequently inhibited by the API, including TGF- β , TNF, and VEGF. This led to significant inhibition of EMT, reducing either inflammation and fibrosis of the lungs (Ivanova et al., 2013; Zhou et al., 2016; Pandolfi et al., 2019) or tumor metastasis (Jin et al., 2019). Additionally, the liposomes improved the delivery efficiency through improved cellular entry, tissue retention of API, and selectivity endowed from a surface-decorated ligand.

Even with the broad array of available aerosolized liposomes and initial promising results for EMT-targeted delivery, challenges remain if these NPs are to be further pursued. For one, liposome stability when aerosolized remains a leading concern. When passed through a nebulizer, significant loss of encapsulated API can occur from mechanical-induced fragmentation of vesicles (Taylor et al., 1990). Strategies exist to mitigate this, such as reducing vesicle size by extruding the liposomes through a membrane filter, but it remains a concern for any potential formulation. Additionally, liposomes face several biological challenges for drug delivery. This includes preferential uptake by the reticuloendothelial system (RES) and opsonization (Poste et al., 1976; Senior, 1986), high clearance of large liposomes (Oku and Namba, 1994; Ulrich, 2002), the “accelerated blood clearance” (ABC) phenomenon if PEGylated liposomes are administered repeatedly (Ishida et al., 2006; Dams et al., 2000), and potential triggering of the innate immune response (Szebeni, 2005; Szebeni and Moghimi, 2009). While the unique anatomy and physiology of the lungs could attenuate some of these concerns, they should still be kept in mind when designing a liposome aerosol. For this reason, efforts have increasingly turned



away from simpler formulations like liposomes and into more complex, and robust, NP systems.

Polymeric Nanoparticles

Polymeric NPs, which have increasingly become more explored as pulmonary drug delivery vehicles in the literature, are one such system. While liposomes are primarily composed of phospholipids with the addition of surfactants and cholesterol, polymeric NPs are instead composed of one or more synthetic polymer repeating chains linked together. The most common of these polymers utilized for pulmonary delivery include poly (lactic acid) (PLA), poly (ϵ -caprolactone) (PCL), poly (lactic-co-glycolic acid) (PLGA), chitosan, alginate, and gelatin bases (Menon et al., 2014). While not naturally derived, these polymers are biodegradable, which mitigates some risk of toxicity (Beck-Broichsitter et al., 2012c; Beck-Broichsitter et al., 2014b). Additionally, the nature of polymer chemistry (or polymerization), wherein monomer molecules are consecutively bonded together through step-growth or chain-growth synthesis techniques, affords greater control and diversity for polymeric NPs than

comparable liposomal formulations (Kricheldorf, 1991; Hickey et al., 2015). This ability to finetune molecular weight (MW), chemical composition, and chain structure provides a wide diversity of potential NPs (as detailed later in this review).

The choice of polymer, polymer ratio, or polymer functional groups can also significantly change polymeric NP properties. For example, the rate of *in vivo* biodegradability can vary based on composition (pure vs hybrid NPs) (Singh et al., 2010), type of polymer (Anderson and Shive, 2012), or MW (Mohammad and Reineke, 2013), or even determine the mechanism of biodegradation itself (hydrolysis vs oxidative degradation, for instance) (Ulbricht et al., 2014). Adding another polymeric coating can also change how a polymeric NP interacts physiologically. This includes enhancing tissue penetration (for example, adding a dense PEG coating to infiltrate farther into therapeutically unsusceptible tissues like the brain or through mucus coatings) (Hong et al., 2009; Tang et al., 2009; Nance et al., 2012) and clearance (such as being susceptible to or avoiding the ABC phenomenon) (Alexis et al., 2008; Ishihara et al., 2010; Saadati et al., 2013). Other fine-tunable properties include

TABLE 2 | A summary of current clinical trials for aerosolized liposomes.

NP	APIs	Target disease	Phase	Status	Sponsor	Identifier
9-Nitro-20 (S)-Camptothecin (L9NC) liposomes	N/A	Non-small-cell lung cancer	II	Completed in 2007	University of New Mexico	NCT00250120
9-Nitro-20 (S)-Camptothecin (L9NC) liposomes	N/A	Metastatic endometrial cancer	II	Completed in 2007	University of New Mexico	NCT00249990
AmBisome	N/A	ABPA	II	Completed in 2019	Poitiers university hospital	NCT02273661
AmBisome	Intraconazole	ABPA	III	Recruiting	Poitiers university hospital	NCT03656081
AmBisome	N/A	Lung transplant infections	II	Completed in 2014	University health network, toronto	NCT01254708
AmBisome	N/A	Lung transplant infections	III	Completed in 2007	University of Pittsburgh	NCT00177710
Arikayce	N/A	Cystic fibrosis	I/II	Completed in 2008	Insmed incorporated	NCT00777296
Arikayce	N/A	Cystic fibrosis	I/II	Completed in 2009	Insmed incorporated	NCT00558844
Arikayce	N/A	Cystic fibrosis	II	Completed in 2010	Insmed incorporated	NCT03905642
Arikayce	Mycobacterial multi-drug regimen	Mycobacterium abscessus lung disease	II	Completed in 2019	Kevin winthrop	NCT03038178
Arikayce	Mycobacterial multi-drug regimen	NTM + MAC	III	Completed in 2018	Insmed incorporated	NCT02628600
Bupivacaine liposomes	N/A	Coronary heart disease	III	Completed in 2020	Kathirvel subramaniam	NCT03270514
Cisplatin liposome	N/A	Osteosarcoma metastatic	I/II	Completed in 2008	Insmed incorporated	NCT00102531
Cyclosporin liposome	N/A	Bronchiolitis obliterans syndrome	II/III	Completed in 2014	Pari Pharma GmbH	NCT01334892
Cyclosporin liposome	Tacrolimus + mycophenolate mofetil prednisone + rampamycin	Lung transplant rejection	I/II	Completed in 2015	University of Maryland, college Park	NCT01650545
Pulmaquin	N/A	Non-cystic fibrosis bronchiectasis	III	Completed in 2016	Aradigm corporation	NCT02104245
Pulmaquin	N/A	Non-cystic fibrosis bronchiectasis	III	Completed in 2016	Aradigm corporation	NCT01515007

AmBisome, amphotericin B liposome; ABPA, allergic bronchopulmonary aspergillosis; NTM, nontuberculous mycobacterial lung infection; Pulmaquin, ciprofloxacin liposome.

TABLE 3 | A summary of current erosolized liposome formulations targeting EMT-markers in the literature.

NP	API(s)	Disease	Model(s)	Effect	Ref
Liposome	PGE2	Idiopathic pulmonary fibrosis (IPF)	IPF mice	Inhibition of TGF β -initiated pathways and TNF, restricted inflammation and fibrosis	Ivanova et al. (2013)
Liposome	Paclitaxel	Pulmonary fibrosis (PF)	PF mice	Inhibition of TGF- β 1, reduced collagen levels and alveolar thickness	Zhou et al. (2016)
Liposome	Simvastatin + paclitaxel	Lung cancer	A549T mice	Inhibition of EMT and metastasis	Jin et al. (2019)
DPPE liposome	Hyaluronic acid	CTD-ILD + BOS	A549 + Calu-3 + THP-1 cells	Inhibition of VEGF mRNA	Pandolfi et al. (2019)

DPPE, 1,2-Bis(diphenylphosphino)ethane; CTD-ILD, connective tissue disease-associated interstitial lung disease; BOS, Bohring-Opitz Syndrome.

lipophilicity and drug loading (Johnstone and Lippard, 2013), circulation time and interaction with vascular proteins (Sheng et al., 2009; Sunoqrot et al., 2014; Yang et al., 2014), NP targeting and tissue accumulation (Park et al., 2007), and NP shelf-life.

In addition to polymer choice, polymeric NPs can also be formulated with a diverse range of molecular architectures that further influence their behaviors (Menon et al., 2014), including linear (single chain) polymers, hyperbranched (multiple end group) polymers (including dendrimers), micelles, and crosslinked emulsions (**Box 1; Figure 4**). As such, polymeric NPs typically range from 1 to 1,000 nm in diameter, exist in a variety of morphologies (e.g., spheres, vesicles, rods, tubules, and lamellae), and consist of homopolymers and various copolymers. Of note for drug delivery are amphiphilic copolymers. These polymer chains, consisting of a lipophilic polymer (such as PLA or PCL) conjugated to a hydrophilic polymer (e.g., PEG), spontaneously form a variety of complex morphologies in aqueous solvents due to hydrophobic interactions (Forster and Antonietti, 1999; Letchford and Burt, 2007). Altering the properties of these copolymers, such as constituent chain length, hydrophilic-lipophilic balance (HLB), and charge, can even further alter a NP's properties, including hydrodynamic size (Wittgren et al., 1996), shell thickness (Stubenrauch et al., 2006), and exposed peripheral functional groups (Urbani et al., 2008; Pearson et al., 2016). This in turn allows further finetuning of drug delivery characteristics, such as API solubility, loading capacity, and release profiles (Yan et al., 2011; Glavas et al., 2015). Altogether, this endows polymeric NPs with advantages similar to liposomes (e.g., easy encapsulation and high loading of lipophilic APIs, API protection, enhanced half-life and lowered blood clearance, sustained and controlled release, and the availability of readily modifiable surface end groups for targeting and stealth strategies) while also providing researchers with a more adaptable NP platform for various delivery needs.

The extensive variety of polymeric NPs has given rise to a broad list of aerosolized formulations in the literature. This includes NPs tested against lung tumors (orthotopic and small cell) (Bhattarai et al., 2007; Rosiere et al., 2016; Qiao et al., 2017; Yan et al., 2017; Elbatany et al., 2020; Vaidya et al., 2020), type 2 diabetes (Shrestha et al., 2015), biofilms and lung infections (Cheow et al., 2010; Casciaro et al., 2019), pneumonia (Shah et al., 2013), tuberculosis (Pandey et al., 2003; Costa-Guoveia et al., 2017), asthma (Yoo et al., 2013), and other respiratory diseases (Beck-Broichsitter et al., 2010; Kolte et al., 2017). The majority of these make use of nanospheres, including PLGA, poly(ethylenimine) (PEI), PCL, and porous silicon (PSi) nanospheres, and micelles, including PLGA-co-polyspermine (PSPE) and folate-PEG-dextran copolymers. Much like liposomes, polymeric NPs have also already been extensively tested with common pulmonary delivery methods, like vibrating-mesh (Beck-Broichsitter et al., 2012a; Beck-Broichsitter et al., 2012b; Beck-Broichsitter et al., 2014a), air-jet (Beck-Broichsitter et al., 2012b), and ultrasonic nebulizers (Beck-Broichsitter et al., 2012b). Special attention has been placed on developing dry powder formulations, such as through spray freeze drying, which enhances the stability of sensitive polymer

BOX 1 | Various polymeric nanoparticle formulations

Linear Polymer-based: A single-chain polymeric NP composed of only one polymer type, which is the simplest form of polymeric NP. Can be broadly defined as a solid colloidal particle consisting of a uniform macromolecular component, which typically forms a three-dimensional nanosphere (though can also include nanocapsules, cubes, and plates) encapsulating therapeutic agents (or, in the case of a surface modification such as PEGylation, extend as a directly conjugated chain off the original molecule) (Chan et al., 2010; Bolhassani et al., 2014). As mentioned, these are typically synthesized using either step-growth (mostly via condensation polymerization) or chain-growth (mostly via addition polymerization such as radical, ionic, ring-opening, or living polymerizations) techniques (Kricheldorf, 1991). The polymer is then dispersed in an organic phase and cross-linked to form a spherical structure. A linear polymer can also be attached to an API through precise post-polymerization modifications of semi-telechelic chains, such as chemical ligation of specific antibodies (cysteine, tyrosine, arginine, and histidine), enzyme-mediated ligation, *in situ* polymerization initiated by an API, or through recombinant gene technology (Ekladios et al., 2019). Nanosphere diameters can range from 20–200 nm while single chain modifications can be even smaller, with some of the smallest only 1–10 nm in diameter (Bolhassani et al., 2014; Kroger and Paulussea, 2018).

Hyperbranched Polymers: Polymeric NPs composed of a hyperbranched structure, wherein exponentially increasing numbers of “branch-like” polymer chains extend from a central core to form a surrounding shell. Dendrimers are the most notable example. These complex highly branched monodisperse polymeric NPs form a spherical morphology, including a central core, a hyperbranched body, and peripheral reactive functional groups, the number of which are doubled with an increase of each dendrimer generation (for example, a generation 0 (G0) poly (amidoamine) (PAMAM) dendrimer has a theoretical four surface groups, a G1 would have 8, a G2 would have 16.) (Shi et al., 2006; Hsu et al., 2016; Franiak-Pietryga et al., 2018). Dendrimers can be synthesized through several techniques, including divergent and convergent synthesis, (Holister et al., 2003; Killops et al., 2008; Franc and Kakkar, 2009), as well as click chemistry, such as Diels-Alder (Franc and Kakkar, 2009), azide-alkyne (Franc and Kakkar, 2008), and thiol-ene reactions (Killops et al., 2008). This allows for tight control of the number of branches and results in a homologous NP with predictable properties (Landmark et al., 2008; Yang et al., 2012; Bugno et al., 2016). Unlike other polymeric NPs, the most common dendrimers for delivery are PAMAM and poly (propylene polybenzyl isocyanate) (PPI) dendrimers, with other examples including poly (L-lysine), polyester, and polyether dendrimers (Kesharwani et al., 2014). Diameters can range anywhere from 1–sub-100 nm with increasing generation, but tend to favor smaller sizes of ≤ 20 nm for delivery (Franiak-Pietryga et al., 2018).

Micelles: NP core/shell structures, composed of amphiphilic block copolymer subunits, that self-assemble into uniform morphologies when exposed to aqueous solvents (Croy and Kwon, 2006; Zhang et al., 2014). This leads to the formation of a hydrophobic core surrounded by a hydrophilic corona, with or without peripheral reactive functional groups [that can then be further modified with polymer coatings (e.g. PEGylation) or conjugated to ligands (Choi et al., 2003)]. Differences in copolymer composition, length, and solvent interaction can further alter micelle characteristics, including size and shape, leading to the formation of spheres, vesicles, rods, tubules, or lamellae (Zhang et al., 2014). The copolymer subunits themselves are synthesized similarly to linear polymers. This can involve either site selective chain-growth polymerization utilizing an existing chain as the initiator (de Espinosa and Meier, 2011), limited step-growth polymerization with two polymer species (Delaittre et al., 2009), or click chemistry between two available functional groups [such as an azide and an alkyne in the presence of cationic Cu(I)] (Li et al., 2010; Engler et al., 2013). Spherical micelles can range from 10 to 100 nm in diameter, while polymersomes dimensions can range from 50 nm to 5 μ m (Croy and Kwon, 2006; Zhang et al., 2014; Zhang and Zhang, 2017).

Emulsions: Three-dimensional networks of crosslinked hydrophilic or amphiphilic polymers, including emulsions, nanogels, and hydrogels (Kabanov and Vinogradov, 2009; Chacko et al., 2012; Kousalova and Etrych, 2018). These networks form either through physical interactions, such as hydrophobic effects, or through chemical interactions, such as hydrogen bonding, electrostatic and ionic interactions, or the formation of chemical bonds (Kabanov and Vinogradov, 2009). This generates an extensive “swollen” polymer mesh that displays the properties of both a conventional NP system and a hydrogel. This includes being highly absorbent (Hoshino et al., 2012), possessing upper diameters far larger than other polymeric NPs, an extensive capacity for API loading and release (Chen et al., 2010), viscoelasticity (Mamaghani et al., 2015), and temperature dependent phase transitions (Hoshino et al., 2012). Depending on the base network, emulsions can range widely from 10 to a few thousands of nanometers in size (Kabanov and Vinogradov, 2009).

NPs and allows for finer control of delivered particles (Cheow et al., 2011; D’Addio et al., 2013; Ali and Lamprecht, 2014). However, despite these current advances, many polymeric NP erosolization studies remain in pre-clinical *in vitro* stages. This is most visible for clinical trials, with no currently reported studies utilizing a polymeric NP aerosol. While a promising and robust alternative to liposomes, this demonstrates that polymeric NP aerosols are still in their infancy, with no sign of an approved product in the near future.

Despite this, polymer NPs are comparatively well represented when it comes to EMT-mediated pulmonary delivery, with currently studied formulations including emulsions, nanospheres, and micelles. These are summarized in **Table 4**. Like with liposomes, the majority of these utilize APIs loaded into a lipophilic core, including pirfenidone (a drug used to treat pulmonary fibrosis) (Singh et al., 2019), MDB5 (a novel form of Hedgehog (Hh) inhibitor GDC-0449) (Kumar et al., 2019), salinomycin (an antibiotic with anti-cancer properties) (Sousa et al., 2019), and doxorubicin (an anthracycline chemotherapeutic) (Fang et al., 2014; Seifi-Najmi et al., 2016). Multiple formulations also make use of siRNA targeting genes along the EMT pathway (Fang et al., 2014; Seifi-Najmi et al., 2016; Li et al., 2019; Suresh et al., 2019), including one directly conjugated to PEI to form a

polyplex (Ding et al., 2018; Wang Y. et al., 2019), as well as an inhibitor of CXCR4 (Suresh et al., 2019), a chemokine receptor involved in the promotion of tumor migration and EMT (Lin et al., 2018). These all resulted in the inhibition of key EMT-biomarkers, including PAI-1, TGF- β , VEGF, Hh, Vimentin, and N-Cadherin. This led to decreased collagen deposition (Wang Y. et al., 2019; Kumar et al., 2019; Li et al., 2019), cell migration and metastasis (Fang et al., 2014; Seifi-Najmi et al., 2016; Sousa et al., 2019), and tumor growth (Suresh et al., 2019). Polymeric NPs increased lung penetration and retention, provided stability by a polymer-siRNA conjugate, and facilitated codelivery further enhanced API efficacies.

While a quickly growing subfield in the aerosol delivery of NPs, concerns remain for polymeric NPs that must be addressed before they can properly compete with liposomes. Perhaps the largest of these is induced aggregation. When passed through a nebulizer, polymeric NPs, especially those with high surface hydrophobicity, can exhibit extensive aggregation within aerosol droplets (Dailey et al., 2003b). This effect can be further pronounced depending on the specific nebulizer used—for instance, NPs administered through jet nebulizers appear more susceptible to aggregation than those passed through ultrasonic nebulizers under the same conditions

and gallic acid [an antioxidant capable of inhibiting glioma cell metastasis and radiation-induced reactive oxygen species (ROS)] (Sunil Gowda et al., 2018). Both were added as promising novel inhibitors of EMT markers (specifically PI3K and vimentin, N-cadherin, and Snail-1, respectively) to enhance the existing anti-fibrotic efficacy of their inorganic NPs. In general, common EMT markers targeted by inorganic NPs included N-cadherin, E-cadherin, and TGF- β (Wahab et al., 2016; Li et al., 2018b; Peng et al., 2018). This in turn led to significant inhibition of EMT, reducing cell migration, tumor growth, and treatment-induced radiation toxicity and resistance. Thus, unlike other NP formulations which only promoted the delivery of APIs, the inorganic NPs here provided key inhibition themselves, in addition to other advantages like enhanced stability and cell penetration and accumulation.

Despite the unique and promising advantages provided by inorganic NPs, however, significant disadvantages remain which challenge their application for pulmonary fibrosis treatment. Perhaps the most significant is localized toxicity. As previously mentioned, inorganic NPs are composed of metals which are not naturally found in significant quantities in the lungs. Exposure to these NPs (at industrial and biomedical levels) has been found to cause acute pulmonary inflammation (Cho et al., 2010; Cho et al., 2012a; Cho et al., 2012b) and oxidative stress (Limbach et al., 2007). Specifically, exposure to common inorganic NPs led to elevated neutrophil and eosinophil recruitment and infiltration into lung tissues (Cho et al., 2010; Cho et al., 2012a), the extent of which appears dependent on the relationship between zeta potential and solubility (Cho et al., 2012b). This includes for carbon (C), cerium (Ce), Ti, Si, nickel (Ni), Zn, and Cu NPs, especially in their pure or oxide forms. However, other studies have demonstrated that biomedical quantities of inorganic NPs are no more toxic than naturally occurring dusts and other aerosols (Veranth et al., 2007). In either case, pulmonary toxicity will remain a concern for any potential aerosol formulation. Additionally, inorganic NPs in general can exhibit elevated systemic clearance by the kidneys and the reticuloendothelial system (RES) in the liver, spleen, and lungs, penetration through vascular endothelium and the blood brain barrier (BBB), and potentially toxic tissue accumulation (Li and Chen, 2011). Finally, due to the current lack of relevant examples in the literature, the potential for aerosolized delivery of inorganic NPs is largely unknown currently. However, the existing use of aerosol vapor deposition for inorganic NP films (Palgrave and Parkin, 2006) and other naturally occurring aerosols (Quadros and Marr, 2012) both suggest its feasibility. For these reasons, future treatments for pulmonary fibrosis may lie in formulations that are closer in design, not farther, to naturally occurring lung vesicles.

Exosomes

Exosomes represent perhaps the most novel formulations currently being explored for pulmonary delivery. A type of membrane-bound extracellular vesicle (EV), small vesicles naturally derived from pro- and eukaryotic cells, exosomes are a more homogenous population of EVs—sharing a similar spherical shape and sizes that range from ~50–150 nm—that

are uniquely rich in cholesterol and diacylglycerol (Ellis and Kuehn, 2010; Silverman and Reiner, 2011; Abels and Breakefield, 2016). They are generated when the plasma membrane of a cell “buds” inwardly to form an intermediate endosome-vesicle known as the multivesicular bodies. These multivesicular bodies are then either degraded by lysosomes or fuse with the plasma membrane to be released as exosomes, which in turn intersperses endosomal surface proteins into the exosome bilayer (Fevrier and Raposo, 2004; Colombo et al., 2014). This helps differentiate exosomes as well as provides unique characteristics depending on the origin cell (Thery et al., 2002). However, there is still some debate on the exact definition, and there is considerable overlap in size and morphology between exosomes and the two other categories of EVs: apoptotic bodies and micro vesicles. Additionally, the exact biological functions of exosomes remain somewhat unclear, though they are known to act as miRNA transporters between cells with important regulatory effects for intracellular mRNA activity (Valadi et al., 2007). Other suggested functions include elimination of undegraded endosomal and lysosomal proteins and membranes, regulation of T cell activity, and antigen transfer to dendritic cells (Thery et al., 2002).

As delivery vehicles, exosomes share many similarities with liposomes, including similar size and morphology (as lipid bilayers) and the ability to encapsulate both hydrophilic and lipophilic APIs (Johnsen et al., 2014; van der Meel et al., 2014). However, the complex composition of surface proteins on exosomes provides additional benefits, which most notably includes enhanced organotropism—that is, exosomes are capable of efficient targeting and uptake by specific recipient cells related to the surface proteins on the original plasma membrane (Antimisiaris et al., 2018). Additionally, the unique natural lipid composition of exosomes makes them highly biocompatible, while selection of autologous exosomes can further prevent any adverse patient immune responses (Antimisiaris et al., 2018).

Examples of aerosolized or inhaled exosomes, though limited, do exist in the literature, including as a potential therapy for tuberculosis (Cheng and Schorey, 2013), lung inflammation (Zhang et al., 2018), pulmonary fibrosis (Dinh et al., 2020), and the general delivery of therapeutic miRNA (Zhang et al., 2017). Overwhelmingly, the exosomes used were derived from alveolar macrophages (Cheng and Schorey, 2013; Zhang et al., 2017; Zhang et al., 2018), though one study did utilize exosomes derived from mesenchymal stem cells instead (Dinh et al., 2020). This same study also utilized a compressed air nebulizer for successful aerosol delivery (Dinh et al., 2020), suggesting that clinically relevant nebulizer systems could be viable for a future exosome formulation. However, comprehensive studies characterizing the physical properties of aerosolized exosomes still appear to be missing. Interestingly, despite the overall current lack of studies on the efficacy and patient safety of aerosolized exosome formulations, several clinical studies have recently surfaced due to the coronavirus disease 2019 (COVID-19) pandemic. These are summarized in **Table 6**.

Similarly, the use of exosomes for EMT-targeting appears to still be in its infancy, with no current examples in the literature of

TABLE 4 | A summary of current aerosolized polymeric NP formulations targeting EMT-markers in the literature.

NP	API(s)	Disease	Model(s)	Effects	Ref
Poly (ethylimine) (PEI-O) polyplex	siPAI-1	Pulmonary fibrosis (PF)	PF mice	Inhibition of PAI-1 and TGF- β , decreased collagen deposition in lungs	Ding et al. (2018), Wang Y. et al. (2019)
Perfluorocarbon (PFC) nanoemulsion	CXCR4 antagonist + anti-STAT3 siRNA	Lung metastasis	4T1, Luc mice	Inhibition of VEGF, decreased tumor growth and increased survival	Li et al. (2019)
Poly (ϵ -caprolactone) (PCL) PEG-PCC-g-DC micelles	Pirfenidone MDB5	Pulmonary fibrosis (PF) Liver fibrosis	A549 cells CBDL mice	Inhibition of TGF- β 1 mRNA and EMT Inhibition of hh activity, HSC activation, and decreased collagen deposition	Singh et al. (2019) Kumar et al. (2019)
Pluronic F127 (PM) micelles	Salinomycin (SAL)	Lung cancer	A549 cells	Inhibition of VIM and EMT, reduced cell migration	Sousa et al. (2019)
PEG-PLL-PLLeu Polypeptide micelles	ZEB1 siRNA + doxorubicin	Non-small cell lung cancer	H460 mice	Inhibition of EMT and metastasis	Fang et al. (2014)
CMD-chitosan nanoparticles	siRNA + doxorubicin	Lung cancer	A549 cells	Inhibition of EMT markers, reduced cell migration	Seifi-Najmi et al. (2016)
EGFR-conjugated gelatin nanoparticles	siRNA	Non-small cell lung cancer	H820 + H1975 cells	Inhibition of Vimentin, N-Cadherin, and EMT	Suresh et al. (2019)

erosolized exosomes specific for the pathway. However, examples of non-aerosolized formulations do exist. This includes for intrauterine adhesion (Yao et al., 2019), liver fibrosis (Li et al., 2012), acute lung injury (ALI) (Xiao et al., 2020), and cancer [including colorectal (Zhang X. et al., 2020), liver (Chen et al., 2018), thyroid (Hardin et al., 2018), and lung (Zhao et al., 2018)]. In the majority of these studies, MSC-derived exosomes were used [including from bone marrow (Yao et al., 2019) and umbilical cord (Li et al., 2012; Zhao et al., 2018)], though others involved exosomes derived directly from carcinomas (Chen et al., 2018; Zhang X. et al., 2020) or stem-like cells (Hardin et al., 2018). Like inorganic NPs, most of the exosomes involved no additional API and instead relied on the intrinsic EMT-inhibition of the derived surface proteins. However, one exception did utilize the additional delivery of an exosomal miRNA (Zhang X. et al., 2020). In all instances, EMT inhibition was achieved through the regulation of key biomarkers, including VIM, TGF- β , Smad2/3, FSP-1, E-Cadherin, CK19, NF- κ B, ERK, JNK, and the Akt and hedgehog pathways. This highlights the inherent advantages to using an exosome formulation directly cultured from EMT-associated cells, with the potential for similar results for exosomes derived from fibrotic lung epithelial cells.

Even with the unique advantages provided by exosomes, concerns remain for any future erosol formulations that must be addressed. For one, exosomes have also been linked to the promotion of EMT for a variety of diseases, notably for cancers [including renal (Wang L. et al., 2019), esophageal (Min et al., 2018), pancreatic (Li et al., 2018b), breast (Wang H. et al., 2019), and lung (He et al., 2019)]. Care will need to be taken in selecting a model exosome for fibrosis therapy which does not induce further long-term tissue injury, especially given the unknown physiological functions of many EVs. Additionally, the exosomal surface can quickly become a disadvantage for the design of more robust formulations or combination therapies—due to the complex nature of surface proteins and the current lack of synthetic alternatives to natural biogenesis, the controlled modification of exosomes with other NP strategies, such as PEGylation, is difficult if not impossible (Antimisiaris et al., 2018). This has led to increased interest in engineered exosomes, so called “EV-mimetics,” in recent years as one potential solution to limited natural yields and to facilitate more varied chemistry. Finally, exosomes in general suffer several therapeutic limitations, including rapid *in vivo* blood clearance, methods which result in low drug loading and retention, and difficult and costly synthesis strategies with limited yields (Antimisiaris et al., 2018).

Other Directions and Considerations

Although promising NP formulations for targeting EMT-mediated pulmonary fibrosis already exist, there are additional directions which could be pursued for further benefits. For example, current therapies largely rely on the passive delivery of APIs (whether drugs or genetic material) or the active targeting of the encapsulated moiety itself. Instead, ligands, especially antibodies and synthetic peptides, could be conjugated to the surface of NP formulations to facilitate active targeting. These sequences could be derived from a multitude of EMT-related axis

TABLE 5 | A summary of current erosolized inorganic NP formulations targeting EMT-markers in the literature.

NP	API(s)	Disease	Model(s)	Effect	Ref.
ZnO ₂ and SiO ₂ NPs	N/A	Fibrosis	LX-2 cells	Inhibition of N-Cadherin and EMT, elevation of E-Cadherin	Peng et al. (2018)
PEG-coated Au NPs	Cold plasma	Lung cancer	T98G + A549 cells	Inhibition of PI3K/AKT axis and EMT	Kaushik et al. (2016)
TiO ₂ NPs	N/A	Lung cancer	A549 cells	Inhibition of TGF- β and EMT, reduced cell migration	Li et al. (2018a)
ZnO nanostructures	N/A	Lung cancer	H460 + MRC5 cells	Inhibition of N-Cadherin and EMT	Wahab et al. (2016)
Ag NPs	Gallic acid	Lung cancer	A549 cells	Inhibition of Vimentin, N-Cadherin, and Snail-1	Sunil Gowda et al., (2018)

TABLE 6 | A summary of current clinical trials for erosolized exosomes.

NP	APIs	Target disease	Phase	Status	Sponsor	Identifier
CSTC-derived exosomes	N/A	Coronavirus + pneumonia	I	Active	TC ericyes university	NCT04389385
MPC-derived exosomes	N/A	Drug resistance	I/II	Recruiting	Ruijin hospital	NCT04544215
MSC-derived exosomes	N/A	Coronavirus	I	Completed in 2020	Ruijin hospital	NCT04276987
MSC-derived exosomes	N/A	Healthy tolerance	I	Recruiting	Ruijin hospital	NCT04313647
MSC-derived exosomes	N/A	ARDS	I/II	Not yet recruiting	Ruijin hospital	NCT04602104

CSTC, COVID-19 specific T cell; MPC, mesenchymal stem/progenitor cell; MSC, mesenchymal stem cell; ARDS, acute respiratory distress syndrome.

and phenotypes. For example, a ligand utilizing a passive targeting approach could target an overexpressed protein characteristic of fibrosis, such as fibronectin (Torikata et al., 1985), while a ligand inhibitor utilizing a more active targeting approach could instead target a receptor along the EMT pathway itself, such as TGF- β family receptors, tyrosine kinase receptors, and numerous integrins (Stone et al., 2016). Similar targeting strategies have already demonstrated success in anti-cancer NP formulations, many of which could easily be translated to target the EMT pathway (Shargh et al., 2015; Jeong et al., 2018).

Additionally, more focus could be placed on developing combination therapies. Drug molecules (anti-inflammatory, fibrotic, and oxidation), genetic material (siRNA), and artificial proteins (antibodies and peptides) could all be combined utilizing a robust NP platform. For example, protein-NP conjugates have demonstrated the ability to significantly increase the potential efficacy of existing therapeutic formulations (Jeong et al., 2020). This could lead to the development of more adaptable therapies capable of overcoming the efficacy and toxicity limitations of other, more conventional therapies. This has become even more important in recent years as increased awareness of genetic variation in patients, including for pulmonary fibrosis, has brought into question the past rationale of a “one-size-fits-all” therapeutic (Kaur et al., 2017). Again, this approach has already found promising success in the development of anti-cancer formulations (Nam et al., 2019).

Despite the many promising advantages of nanomedicines, the treatment of EMT-mediated pulmonary fibrosis provides unique challenges for NP therapeutics. For one, NPs have been linked to the facilitation of EMT and lung fibrosis themselves. Though most frequently caused by industrial pollutants such as cerium oxide, silver, and silica NPs (Ma et al., 2017; Wang et al., 2017; Gliga et al., 2018), the elevated infiltration and retention of other NP formulations can still toe the line between beneficial and toxic and must be carefully analyzed (Fattal et al., 2014). Inorganic NPs in particular run the risk of becoming acutely toxic to local tissues, but polymeric NPs and even liposomes and

SLNPs can still lead to lung damage if not safely absorbed and eventually cleared.

The ability of NP formulations to be properly administered using common pulmonary delivery methods, such as through nebulizers, pressurized metered-dose inhalers, and dry powder inhalers (DPIs), can also be a limiting factor (Dailey et al., 2003b). Aggregation, especially, becomes a frequent concern when NPs are aerosolized through fine pores. Liposomes, micelles, and other spontaneously assembled NPs, which rely on maintaining specific concentrations or solvent conditions, can also become unstable when subjected to the erosolization process. Despite this, however, numerous NP formulations have already demonstrated successful aerosolization, including liposomes (Zaru et al., 2009; Arora et al., 2010), polymeric NPs (Doan and Oliver, 2009; Ungaro et al., 2009; Beck-Broichsitter et al., 2010), and even Cu/Zn NPs (Langenback et al., 1999). This indicates that the aerosolization of these novel formulations is certainly a surmountable obstacle worth investigating.

CONCLUSION

In conclusion, the IPF has substantial morbidity and mortality for which no curative treatments are available. In this chapter, we have reviewed the prevailing epithelial injury hypothesis. In this model, epithelial injury-induced EMT drives the expansion of pathogenic myofibroblast populations. Myofibroblasts are principal components of ECM production that result in airspace loss and mortality. We review the substantial mechanistic understanding of EMT, and the recent work demonstrating that reversible histone modifications are an important process regulating gene expression programs controlling mesenchymal transition. This mechanistic work has focused on the central role of BRD4 in mediating EMT and myofibroblast transition and initial preclinical work has provided evidence of efficacy in

multiple models of fibrosis, including bleomycin, radiation, TGF- β induced and viral induced remodeling. Advancing epigenetic regulators as therapeutics will be challenging. Additionally, we review the current state of nanomedicine with a focus on formulations that hold promise for aerosol delivery and EMT-targeting, including liposomes, polymeric NPs, inorganic NPs, and exosomes. These agents can provide a host of benefits, including controlled release for enhanced therapeutic efficacy, reduced systemic toxicity, and intrinsic inhibitory effects against common EMT biomarkers. Many of these NP formulations have already demonstrated success as aerosolized formulations in both the lab and clinic. With further development, NP-based approaches for pulmonary delivery offer the substantial promise to modify epigenetic regulators of EMT and advance treatments for IPF further than current strategies allow. This could open up a new field of study centered on the EMT pathway which combines new physiological understandings and state-of-the-art advances in nanoparticle delivery.

REFERENCES

- Abels, E. R., and Breakefield, X. O. (2016). Introduction to extracellular vesicles: biogenesis, RNA cargo selection, content, release, and uptake. *Cell. Mol. Neurobiol.* 36, 301312. doi:10.1007/s10571-016-0366-z
- Alexis, F., Pridgen, E., Molnar, L. K., and Farokhzad, O. C. (2008). Factors affecting the clearance and biodistribution of polymeric nanoparticles. *Mol. Pharm.* 5, 505–515. doi:10.1021/mp800051m
- Ali, M. E., and Lamprecht, A. (2014). Spray freeze drying for dry powder inhalation of nanoparticles. *Eur. J. Pharm. Biopharm.* 87, 510–517. doi:10.1016/j.ejpb.2014.03.009
- Allen, S. D., Sorensen, K. N., Neal, M. J., Durrant, C., and Proffit, R. T. (1994). Prophylactic efficacy of aerosolized liposomal (AmBisome) and non-liposomal (Fungizone) amphotericin B in murine pulmonary aspergillosis. *J. Antimicrob. Chemother.* 34, 1001–1013. doi:10.1093/jac/34.6.1001
- Alshamsan, A., Haddadi, A., Hamdy, S., Samuel, J., El-Kadi, A. O. S., Uludağ, H., et al. (2010). STAT3 silencing in dendritic cells by siRNA polyplexes encapsulated in PLGA nanoparticles for the modulation of anticancer immune response. *Mol. Pharm.* 7, 1643–1654. doi:10.1021/mp100067u
- Anderson, J. M., and Shive, M. S. (2012). Biodegradation and biocompatibility of PLA and PLGA microspheres. *Adv. Drug Deliv. Rev.* 64, 72–82. doi:10.1016/j.addr.2012.09.004
- Antimisiaris, S. G., Mourtas, S., and Marazioti, A. (2018). Exosomes and exosome-inspired vesicles for targeted drug delivery. *Pharmaceutics* 10 (4), 218. doi:10.3390/pharmaceutics10040218
- Arora, D., K. Goyal, A., R. Paliwal, S., Khurana, B., and P. Vyas, S. (2010). Oral mucosal immunization: recent advancement and future prospects. *Curr. Immunol. Rev.* 6, 234–259. doi:10.2174/157339510791823637
- Arvizo, R. R., Saha, S., Wang, E., Robertson, J. D., Bhattacharya, R., and Mukherjee, P. (2013). Inhibition of tumor growth and metastasis by a self-therapeutic nanoparticle. *Proc. Natl. Acad. Sci. Unit. States Am.* 110, 6700–6705. doi:10.1073/pnas.1214547110
- Athanasio, R. (2012). Airway disease: similarities and differences between asthma, COPD and bronchiectasis. *Clinics* 67 (11), 1335–1343. doi:10.6061/clinics/2012(11)19
- Barnes, P. J. (2009). How corticosteroids control inflammation: quintiles prize lecture 2005. *Br. J. Pharmacol.* 148 (3), 245–254. doi:10.1038/sj.bjp.0706736
- Beck-Broichsitter, M., Gauss, J., Gessler, T., Seeger, W., Kissel, T., and Schmehl, T. (2010). Pulmonary targeting with biodegradable salbutamol-loaded nanoparticles. *J. Aerosol Med. Pulm. Drug Deliv.* 23, 47–57. doi:10.1089/jamp.2009.0759
- Beck-Broichsitter, M., Kleimann, P., Gessler, T., Seeger, W., Kissel, T., and Schmehl, T. (2012a). Nebulization performance of biodegradable sildenafil-loaded nanoparticles using the Aeroneb Pro: formulation aspects and

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- nanoparticle stability to nebulization. *Int. J. Pharm.* 422, 398–408. doi:10.1016/j.ijpharm.2011.10.012
- Beck-Broichsitter, M., Kleimann, P., Schmehl, T., Betz, T., Bakowsky, U., Kissel, T., et al. (2012b). Impact of lyoprotectants for the stabilization of biodegradable nanoparticles on the performance of air-jet, ultrasonic, and vibrating-mesh nebulizers. *Eur. J. Pharm. Biopharm.* 82, 272–280. doi:10.1016/j.ejpb.2012.07.004
- Beck-Broichsitter, M., Schmehl, T., Gessler, T., Seeger, W., and Kissel, T. (2012c). Development of a biodegradable nanoparticle platform for sildenafil: formulation optimization by factorial design analysis combined with application of charge-modified branched polyesters. *J. Contr. Release* 157, 469–477. doi:10.1016/j.jconrel.2011.09.058
- Beck-Broichsitter, M., Knuedeler, M.-C., Oesterheld, N., Seeger, W., and Schmehl, T. (2014a). Boosting the aerodynamic properties of vibrating-mesh nebulized polymeric nanosuspensions. *Int. J. Pharm.* 459, 23–29. doi:10.1016/j.ijpharm.2013.11.040
- Beck-Broichsitter, M., Ruppert, C., Schmehl, T., Günther, A., and Seeger, W. (2014b). Biophysical inhibition of synthetic vs. naturally-derived pulmonary surfactant preparations by polymeric nanoparticles. *Biochim. Biophys. Acta* 1838, 474–481. doi:10.1016/j.bbamem.2013.10.016
- Bergeron, C., Tulic, M. K., and Hamid, Q. (2010). Airway remodelling in asthma: from benchside to clinical practice. *Can. Respir. J. J. Can. Thorac. Soc.* 17 (4), e85–e93. doi:10.1155/2010/318029
- Bhattarai, S. R., Kim, S. Y., Jang, K. Y., Yi, H. K., Lee, Y. H., Bhattarai, N., et al. (2007). Amphiphilic triblock copolymer poly(p-dioxanone-co-L-lactide)-block-poly(ethylene glycol), enhancement of gene expression and inhibition of lung metastasis by aerosol delivery. *Gene Ther.* 14, 476–483. doi:10.1038/sj.gt.3302876
- Bitounis, D., Fanciullino, R., Iliadis, A., and Ciccolini, J. (2012). Optimizing druggability through liposomal formulations: new approaches to an old concept. *ISRN Pharm.* 2012, 738432. doi:10.5402/2012/738432
- Bolhassani, A., Javanad, S., Saleh, T., Hashemi, M., Aghasadeghi, M. R., and Sadat, S. M. (2014). Polymeric nanoparticles. *Hum. Vaccines Immunother.* 10, 321–332. doi:10.4161/hv.26796
- Bozzuto, G., and Molinari, A. (2015). Liposomes as nanomedical devices. *Int. J. Nanomedicine* 10, 975–999. doi:10.2147/ijn.s68861
- Brasier, A. R. (2019). Mechanisms how mucosal innate immunity affects progression of allergic airway disease. *Expert Rev. Respir. Med.* 13 (4), 349–356. doi:10.1080/17476348.2019.1578211
- Brasier, A. R. (2020). RSV reprograms the CDK9BRD4 chromatin remodeling complex to couple innate inflammation to airway remodeling. *Viruses* 12 (4), 472. doi:10.3390/v12040472
- Brasier, A. R., Tian, B., Jamaluddin, M., Kalita, M. K., Garofalo, R. P., and Lu, M. (2011). RelA Ser276 phosphorylation-coupled Lys310 acetylation controls transcriptional elongation of inflammatory cytokines in respiratory syncytial virus infection. *J. Virol.* 85 (22), 11752–11769. doi:10.1128/JVI.05360-11

- Brasier, A. R., and Zhou, J. (2020). Validation of the epigenetic reader bromodomain-containing protein 4 (BRD4) as a therapeutic target for treatment of airway remodeling. *Drug Discov. Today* 25 (1), 126–132. doi:10.1016/j.drudis.2019.11.002
- Brown, J. D., Lin, C. Y., Duan, Q., Griffin, G., Federation, A. J., Paranal, R. M., et al. (2014). NF- κ B directs dynamic super enhancer formation in inflammation and atherogenesis. *Mol. Cell* 56 (2), 219–231. doi:10.1016/j.molcel.2014.08.024
- Bugno, J., Hsu, H.-j., and Hong, S. (2015). Recent advances in targeted drug delivery approaches using dendritic polymers. *Biomater. Sci.* 3, 1025–1034. doi:10.1039/c4bm00351a
- Bugno, J., Hsu, H.-J., Pearson, R. M., Noh, H., and Hong, S. (2016). Size and surface charge of engineered poly(amidoamine) dendrimers modulate tumor accumulation and penetration: a model study using multicellular tumor spheroids. *Mol. Pharm.* 13, 2155–2163. doi:10.1021/acs.molpharmaceut.5b00946
- Burgess, H. A., Daugherty, L. E., Thatcher, T. H., Lakatos, H. F., Ray, D. M., Redonnet, M., et al. (2005). PPAR γ agonists inhibit TGF- β induced pulmonary myofibroblast differentiation and collagen production: implications for therapy of lung fibrosis. *Am. J. Physiol. Lung Cell Mol. Physiol.* 288, L1146–L1153. doi:10.1152/ajplung.00383.2004
- Casciaro, B., d'Angelo, I., Zhang, X., Loffredo, M. R., Conte, G., Cappiello, F., et al. (2019). Poly(lactide-co-glycolide) nanoparticles for prolonged therapeutic efficacy of esculentin-1a-derived antimicrobial peptides against *Pseudomonas aeruginosa* lung infection: *in Vitro* and *in Vivo* studies. *Biomacromolecules* 20, 1876–1888. doi:10.1021/acs.biomac.8b01829
- Chacko, R. T., Ventura, J., Zhuang, J., and Thayumanavan, S. (2012). Polymer nanogels: a versatile nanoscopic drug delivery platform. *Adv. Drug Deliv. Rev.* 64, 836–851. doi:10.1016/j.addr.2012.02.002
- Chan, J. M., Valencia, P. M., Zhang, L., Langer, R., and Farokhzad, O. C. (2010). Polymeric nanoparticles for drug delivery. *Cancer Nanotechnol.* 624, 163–175. doi:10.1007/978-1-60761-609-2_11
- Chang, H., Liu, Y., Xue, M., Liu, H., Du, S., Zhang, L., et al. (2016). Synergistic action of master transcription factors controls epithelial-to-mesenchymal transition. *Nucleic Acids Res.* 44 (6), 2514–2527. doi:10.1093/nar/gkw126
- Chen, L., Guo, P., He, Y., Chen, Z., Chen, L., Luo, Y., et al. (2018). HCC-derived exosomes elicit HCC progression and recurrence by epithelial-mesenchymal transition through MAPK/ERK signalling pathway. *Cell Death Dis.* 9, 513. doi:10.1038/s41419-018-0534-9
- Chen, Y., Zheng, X., Qian, H., Mao, Z., Ding, D., and Jiang, X. (2010). Hollow Core–Porous shell structure poly(acrylic acid) nanogels with a superhigh capacity of drug loading. *ACS Appl. Mater. Interfaces* 2, 3532–3538. doi:10.1021/am100709d
- Cheng, Y., and Schorey, J. S. (2013). Exosomes carrying mycobacterial antigens can protect mice against *Mycobacterium tuberculosis* infection. *Eur. J. Immunol.* 43, 3279–3290. doi:10.1002/eji.201343727
- Cheow, W. S., Chang, M. W., and Hadinoto, K. (2010). Antibacterial efficacy of inhalable antibiotic-encapsulated biodegradable polymeric nanoparticles against *E. coli* biofilm cells. *J. Biomed. Nanotechnol.* 6, 391–403. doi:10.1166/jbn.2010.1116
- Cheow, W. S., Ng, M. L. L., Kho, K., and Hadinoto, K. (2011). Spray-freeze-drying production of thermally sensitive polymeric nanoparticle aggregates for inhaled drug delivery: effect of freeze-drying adjuvants. *Int. J. Pharm.* 404, 289–300. doi:10.1016/j.ijpharm.2010.11.021
- Cho, W.-S., Duffin, R., Poland, C. A., Duschl, A., Oostingh, G. J., MacNee, W., et al. (2012a). Differential pro-inflammatory effects of metal oxide nanoparticles and their soluble ions *in vitro* and *in vivo*; zinc and copper nanoparticles, but not their ions, recruit eosinophils to the lungs. *Nanotoxicology* 6, 22–35. doi:10.3109/17435390.2011.552810
- Cho, W.-S., Duffin, R., Thielbeer, F., Bradley, M., Megson, I. L., MacNee, W., et al. (2012b). Zeta potential and solubility to toxic ions as mechanisms of lung inflammation caused by metal/metal oxide nanoparticles. *Toxicol. Sci.* 126, 469–477. doi:10.1093/toxsci/kfs006
- Cho, W.-S., Duffin, R., Poland, C. A., Howie, S. E. M., MacNee, W., Bradley, M., et al. (2010). Metal oxide nanoparticles induce unique inflammatory footprints in the lung: important implications for nanoparticle testing. *Environ. Health Perspect.* 118, 1699–1706. doi:10.1289/ehp.1002201
- Choi, S. W., Kim, W. S., and Kim, J. H. (2003). Surface modification of functional nanoparticles for controlled drug delivery. *J. Dispersion Sci. Technol.* 24, 475–487. doi:10.1081/dis-120021803
- Cipolla, D., Gonda, I., and Chan, H.-K. (2013). Liposomal formulations for inhalation. *Ther. Deliv.* 4, 1047–1072. doi:10.4155/tde.13.71
- Colombo, M., Raposo, G., and Théry, C. (2014). Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. *Annu. Rev. Cell Dev. Biol.* 30, 255–289. doi:10.1146/annurev-cellbio-101512-122326
- Conley, J., Yang, H., Wilson, T., Blasetti, K., Di Ninno, V., Schnell, G., et al. (1997). Aerosol delivery of liposome-encapsulated ciprofloxacin: aerosol characterization and efficacy against *Francisella tularensis* infection in mice. *Antimicrob. Agents Chemother.* 41, 1288–1292. doi:10.1128/aac.41.6.1288
- Costa-Guoveia, J., Pancani, E., Jouny, S., Machelart, A., Delorme, V., Salzano, G., et al. (2017). Combination therapy for tuberculosis treatment: pulmonary administration of ethionamide and booster co-loaded nanoparticles. *Nature* 7, 5390. doi:10.1038/s41598-017-05453-3
- Coward, W. R., Watts, K., Feghali-Bostwick, C. A., Knox, A., and Pang, L. (2009). Defective histone acetylation is responsible for the diminished expression of cyclooxygenase 2 in idiopathic pulmonary fibrosis. *MCB (Mol. Cell. Biol.)* 29 (15), 4325–4339. doi:10.1128/mcb.01776-08
- Croy, S., and Kwon, G. (2006). Polymeric micelles for drug delivery. *Curr. Pharm. Des.* 12, 4669–4684. doi:10.2174/138161206779026245
- D'Addio, S. M., Chan, J. G., Kwok, P. C., Benson, B. R., Prud'homme, R. K., and Chan, H. K. (2013). Aerosol delivery of nanoparticles in uniform mannitol carriers formulated by ultrasonic spray freeze drying. *Pharm. Res.* 30, 2891–2901. doi:10.1007/s11095-013-1120-6
- Dailey, L. A., Jekel, N., Fink, L., Gessler, T., Schmehl, T., Wittmar, M., et al. (2006). Investigation of the proinflammatory potential of biodegradable nanoparticle drug delivery systems in the lung. *Toxicol. Appl. Pharmacol.* 215, 100–108. doi:10.1016/j.taap.2006.01.016
- Dailey, L. A., Kleeman, E., Wittmar, M., Gessler, T., Schmehl, T., Roberts, C., et al. (2003a). Surfactant-free, biodegradable nanoparticles for aerosol therapy based on the branched polyesters, DEAPA-PVAL-g-PLGA. *Pharmaceutical Res.* 20, 2011–2020. doi:10.1023/b:pham.0000008051.94834.10
- Dailey, L. A., Schmehl, T., Gessler, T., Wittmar, M., Grimminger, F., Seeger, W., et al. (2003b). Nebulization of biodegradable nanoparticles: impact of nebulizer technology and nanoparticle characteristics on aerosol features. *J. Contr. Release* 86, 131–144. doi:10.1016/s0168-3659(02)00370-x
- Dams, E. T., Laverman, P., Oyen, W. J., Storm, G., Scherphof, G. L., van Der Meer, J. W., et al. (2000). Accelerated blood clearance and altered biodistribution of repeated injections of sterically stabilized liposomes. *J. Pharmacol. Exp. Therapeut.* 292, 1071–1079.
- Davda, J., and Labhasetwar, V. (2002). Characterization of nanoparticle uptake by endothelial cells. *Int. J. Pharm.* 233, 51–59. doi:10.1016/s0378-5173(01)00923-1
- Davies, L. A., Nunez-Alonso, G. A., McLachlan, G., Hyde, S. C., and Gill, D. R. (2014). Aerosol delivery of DNA/liposomes to the lung for cystic fibrosis gene therapy. *Hum. Gene Ther. Clin. Dev.* 25, 97–107. doi:10.1089/humc.2014.019
- de Espinosa, L. M., and Meier, M. A. R. (2011). Synthesis of star- and block-copolymers using ADMET: head-to-tail selectivity during step-growth polymerization. *Chem. Commun.* 47, 1908–1910. doi:10.1039/c0cc04161k
- Delaittre, G., Dire, C., Rieger, J., Putaux, J.-L., and Charleux, B. (2009). Formation of polymer vesicles by simultaneous chain growth and self-assembly of amphiphilic block copolymers. *Chem. Commun.* 15, 2887–2889. doi:10.1039/b903040a
- de Ruijter, A. J. M. d., Gennip, A. H. v., Caron, H. N., Kemp, S., and Kuilenburg, A. B. P. v. (2003). Histone deacetylases (HDACs): characterization of the classical HDAC family. *Biochem. J.* 370 (Pt 3), 737–749. doi:10.1042/BJ20021321
- Devaiah, B. N., Case-Borden, C., Geggone, A., Hsu, C. H., Chen, Q., Meerzaman, D., et al. (2016a). BRD4 is a histone acetyltransferase that evicts nucleosomes from chromatin. *Nat. Struct. Mol. Biol.* 23 (6), 540–548. doi:10.1038/nsmb.3228
- Devaiah, B. N., Geggone, A., and Singer, D. S. (2016b). Bromodomain 4: a cellular Swiss army knife. *J. Leukoc. Biol.* 100 (4), 679–686. doi:10.1189/jlb.2RI0616-250R
- Devaiah, B. N., Lewis, B. A., Cherman, N., Hewitt, M. C., Albrecht, B. K., Robey, P. G., et al. (2012). BRD4 is an atypical kinase that phosphorylates serine 2 of the RNA polymerase II carboxy-terminal domain. *Proc. Natl. Acad. Sci. U. S. A.* 109 (18), 6927–6932. doi:10.1073/pnas.1120422109

- Dey, A., Chitsaz, F., Abbasi, A., Misteli, T., and Ozato, K. (2003). The double bromodomain protein Brd4 binds to acetylated chromatin during interphase and mitosis. *Proc. Natl. Acad. Sci. U. S. A.* 100 (15), 8758–8763. doi:10.1073/pnas.1433065100
- Ding, L., Zhu, C., Yu, F., Wu, P., Chen, G., Ullah, A., et al. (2018). Pulmonary delivery of polyplexes for combined PAI-1 gene silencing and CXCR4 inhibition to treat lung fibrosis. *Nanomed. Nanotechnol. Biol. Med.* 14, 1765–1776. doi:10.1016/j.nano.2018.05.005
- Ding, M., Zhang, Q., Li, Q., Wu, T., and Huang, Y.-z. (2020). Correlation analysis of the severity and clinical prognosis of 32 cases of patients with COVID-19. *Respir. Med.* 167, 105981. doi:10.1016/j.rmed.2020.105981
- Dinh, P.-U. C., Paudel, D., Brochu, H., Popowski, K. D., Gracieux, M. C., Coresand, J., et al. (2020). Inhalation of lung spheroid cell secretome and exosomes promotes lung repair in pulmonary fibrosis. *Nat. Commun.* 11, 1064. doi:10.1038/s41467-020-14344-7
- Doan, T. V. P., and Olivier, J. C. (2009). Preparation of rifampicin-loaded PLGA microspheres for lung delivery as aerosol by premix membrane homogenization. *Int. J. Pharm.* 382, 61–66. doi:10.1016/j.ijpharm.2009.08.008
- Dou, C., Dou, C., Liu, Z., Liu, Z., Tu, K., Tu, K., et al. (2018). P300 acetyltransferase mediates stiffness-induced activation of hepatic stellate cells into tumor-promoting myofibroblasts. *Gastroenterology* 154 (8), 2209–2221.e14. doi:10.1053/j.gastro.2018.02.015
- Ekladious, I., Colson, Y. L., and Grinstaff, M. W. (2019). Polymer-drug conjugate therapeutics: advances, insights and prospects. *Nat. Rev. Drug Discov.* 18, 273–294. doi:10.1038/s41573-018-0005-0
- Elbatany, R. S., Parvathaneni, V., Kulkarni, N. S., Shukla, S. K., Chauhan, G., Kunda, N. K., et al. (2020). Afatinib-loaded inhalable PLGA nanoparticles for localized therapy of non-small cell lung cancer (NSCLC)—development and *in-vitro* efficacy. *Drug Deliv. Transl. Res.* 1–17. doi:10.1007/s13346-020-00802-8
- Elhissi, A., Faizi, M., Naji, W., Gill, H., and Taylor, K. (2007). Physical stability and aerosol properties of liposomes delivered using an air-jet nebulizer and a novel micropump device with large mesh apertures. *Int. J. Pharm.* 334, 62–70. doi:10.1016/j.ijpharm.2006.10.022
- Elhissi, A. M. A., and Taylor, K. M. G. (2005). Delivery of liposomes generated from proliposomes using air-jet, ultrasonic, and vibrating-mesh nebulisers. *J. Drug Deliv. Sci. Technol.* 15, 261–265. doi:10.1016/s1773-2247(05)50047-9
- Ellis, T. N., and Kuehn, M. J. (2010). Virulence and immunomodulatory roles of bacterial outer membrane vesicles. *Microbiol. Mol. Biol. Rev.* 74, 81–94. doi:10.1128/membr.00031-09
- Engler, A. C., Lee, H., and Hammond, P. T. (2013). Highly efficient “grafting onto” a polypeptide backbone by “click chemistry”. *Angew. Chem. Int. Ed.* 48 (49), 9334–9338. doi:10.1002/anie.200904070
- Espada, J., Peinado, H., Lopez-Serra, L., Setién, F., Lopez-Serra, P., Portela, A., et al. (2011). Regulation of SNAIL and E-cadherin function by DNMT1 in a DNA methylation-independent context. *Nucleic Acids Res.* 39 (21), 9194–9205. doi:10.1093/nar/gkr658
- Etheridge, M. L., Campbell, S. A., Erdman, A. G., Haynes, C. L., Wolf, S. M., and McCullough, J. (2013). The big picture on nanomedicine: the state of investigational and approved nanomedicine products. *Nanomed. Nanotechnol. Biol. Med.* 9, 1–14. doi:10.1016/j.nano.2012.05.013
- Evans, M. J., Van Winkle, L. S., Fanucchi, M. V., and Plopper, C. G. (1999). The attenuated fibroblast sheath of the respiratory tract epithelial-mesenchymal trophic unit. *Am. J. Respir. Cell Mol. Biol.* 21 (6), 655–657. doi:10.1165/ajrcmb.21.6.3807
- Fang, S., Wu, L., Li, M., Yi, H., Gao, G., Sheng, Z., et al. (2014). ZEB1 knockdown mediated using polypeptide cationic micelles inhibits metastasis and effects sensitization to a chemotherapeutic drug for cancer therapy. *Nanoscale* 6, 10084–10094. doi:10.1039/c4nr01518e
- Fattal, E., Grabowski, N., Mura, S., Vergnaud, J., Tsapis, N., and Hillaireau, H. (2014). Lung toxicity of biodegradable nanoparticles. *J. Biomed. Nanotechnol.* 10, 2852–2864. doi:10.1166/jbn.2014.1939
- Fehrenbach, H. (2001). Alveolar epithelial type II cell: defender of the alveolus revisited. *Respir. Res.* 2 (1), 33–46. doi:10.1186/rr36
- Fernandez, I. E., and Eickelberg, O. (2012). The impact of TGF- β on lung fibrosis. *Proc. Am. Thorac. Soc.* 9, 111–116. doi:10.1513/pats.201203-023aw
- Fevrier, B., and Raposo, G. (2004). Exosomes: endosomal-derived vesicles shipping extracellular messages. *Curr. Opin. Cell Biol.* 16, 415–421. doi:10.1016/j.ceb.2004.06.003
- Flaherty, K. R., Wells, A. U., Cottin, V., Devaraj, A., Walsh, S. L. F., Inoue, Y., et al. (2019). Nintedanib in progressive fibrosing interstitial lung diseases. *N. Engl. J. Med.* 381 (18), 1718–1727. doi:10.1056/NEJMoa1908681
- Foster, K. A., Yazdani, M., and Audus, K. L. (2001). Microparticulate uptake mechanisms of *in-vitro* cell culture models of the respiratory epithelium. *J. Pharm. Pharmacol.* 53, 57–66. doi:10.1211/0022357011775190
- Forster, S., and Antonietti, M. (1999). Amphiphilic block copolymers in structure-controlled nanomaterial hybrids. *Adv. Mater.* 10, 195–217. doi:10.1002/(SICI)1521-4095(199902)10:3<195::AID-ADMA195>3.0.CO;2-V
- Franc, G., and Kakkar, A. (2008). Dendrimer design using CuI-catalyzed alkyne-azide “click-chemistry”. *Chem. Commun.* 42, 5267–5276. doi:10.1039/b809870k
- Franc, G., and Kakkar, A. K. (2009). Diels-alder “click” chemistry in designing dendritic macromolecules. *Chem. Eur J.* 15, 5630–5639. doi:10.1002/chem.200900252
- Franiak-Pietryga, I., Ziemia, B., Messmer, B., and Skowronska-Krawczyk, D. (2018). “Dendrimers as drug nanocarriers: the future of gene therapy and targeted therapies in cancer,” in *Dendrimers—fundamentals and applications*. Editor C. M. Simonescu (London, UK: IntechOpen).
- Gabizon, A., and Papahadjopoulos, D. (1988). Liposome formulations with prolonged circulation time in blood and enhanced uptake by tumors. *Proc. Natl. Acad. Sci. U. S. A.* 85, 6949–6953. doi:10.1073/pnas.85.18.6949
- Gao, W., Hu, C.-M. J., Fang, R. H., and Zhang, L. (2013). Liposome-like nanostructures for drug delivery. *J. Mater. Chem. B* 1 (48), 6569–6585. doi:10.1039/c3tb21238f
- Geng, Y., Liu, X., Liang, J., Habel, D. M., Vrshika, K., Coelho, A. L., et al. (2019). PD-L1 on invasive fibroblasts drives fibrosis in a humanized model of idiopathic pulmonary fibrosis. *JCI Insight* 4 (6), e125326. doi:10.1172/jci.insight.125326
- Gilbert, B. E., Knight, C., Alvarez, F. G., Waldrep, J. C., Rodarte, J. R., Knight, V., et al. (1997). Tolerance of volunteers to cyclosporine A-dilauroylphosphatidylcholine liposome aerosol. *Am. J. Respir. Crit. Care Med.* 156, 1789–1793. doi:10.1164/ajrcm.156.6.9702101
- Glavas, L., Odelius, K., and Albertsson, A. C. (2015). Tuning loading and release by modification of micelle core crystallinity and preparation. *Polym. Adv. Technol.* 26, 880–888. doi:10.1002/pat.3524
- Gliga, A. R., Bucchianico, S. D., Lindvall, J., Fadeel, B., and Karlsson, H. (2018). RNA-sequencing reveals long-term effects of silver nanoparticles on human lung cells. *Nature* 8, 6668. doi:10.1038/s41598-018-25085-5
- Gu, F. X., Karnik, R., Wang, A. Z., Alexis, F., Levy-Nissenbaum, E., Hong, S., et al. (2007). Targeted nanoparticles for cancer therapy. *Nano Today* 2, 14–21. doi:10.1016/s1748-0132(07)70083-x
- Guo, L., Yan, D. D., Yang, D., Li, Y., Wang, X., Zalewski, O., et al. (2014). Combinatorial photothermal and immuno cancer therapy using chitosan-coated hollow copper sulfide nanoparticles. *ACS Nano* 8, 5670–5681. doi:10.1021/nn5002112
- Habermann, A. C., Gutierrez, A. J., Bui, L. T., Yahn, S. L., Winters, N. I., Calvi, C. L., et al. (2020). Single-cell RNA sequencing reveals profibrotic roles of distinct epithelial and mesenchymal lineages in pulmonary fibrosis. *Sci. Adv.* 6 (28), eaba1972. doi:10.1126/sciadv.aba1972
- Hardin, H., Helein, H., Meyer, K., Robertson, S., Zhang, R., Zhong, W., et al. (2018). Thyroid cancer stem-like cell exosomes: regulation of EMT via transfer of lncRNAs. *Lab. Invest.* 98, 1133–1142. doi:10.1038/s41374-018-0065-0
- He, S., Li, Z., Yu, Y., Zeng, Q., Cheng, Y., Ji, W., et al. (2019). Exosomal miR-499a-5p promotes cell proliferation, migration and EMT via mTOR signaling pathway in lung adenocarcinoma. *Exp. Cell Res.* 379, 203–213. doi:10.1016/j.yexcr.2019.03.035
- Hickey, J. W., Santos, J. L., Williford, J.-M., and Mao, H.-Q. (2015). Control of polymeric nanoparticle size to improve therapeutic delivery. *J. Contr. Release* 219, 536–547. doi:10.1016/j.jconrel.2015.10.006
- Hill, C., Jones, M., Davies, D., and Wang, Y. (2019). Epithelial-mesenchymal transition contributes to pulmonary fibrosis via aberrant epithelial/fibroblastic cross-talk. *J. Lung Health Dis.* 3 (2), 31–35. doi:10.29245/2689-999x/2019/2.3149
- Hinz, B., Phan, S. H., Thannickal, V. J., Galli, A., Bochaton-Piallat, M. L., and Gabbiani, G. (2007). The myofibroblast: one function, multiple origins. *Am. J. Pathol.* 170 (176), 1807–1816. doi:10.2353/ajpath.2007.070112

- Holister, P., Roman Vas, C., and Harper, T. (2003). *Dendrimers: technology white papers*. Madrid, Spain: Cientifica.
- Hong, S., Rattan, R., Majoros, I. J., Mullen, D. G., Peters, J. L., Shi, X., et al. (2009). The role of ganglioside GM1 in cellular internalization mechanisms of poly(amidoamine) dendrimers. *Bioconjugate Chem.* 20, 1503–1513. doi:10.1021/bc900029k
- Horowitz, J. C., Cui, Z., Moore, T. A., Meier, T. R., Reddy, R. C., Toews, G. B., et al. (2006). Constitutive activation of prosurvival signaling in alveolar mesenchymal cells isolated from patients with nonresolving acute respiratory distress syndrome. *Am. J. Physiol. Lung Cell Mol. Physiol.* 290 (3), L415–L425. doi:10.1152/ajplung.00276.2005
- Hoshino, Y., Imamura, K., Yue, M., Inoue, G., and Miura, Y. (2012). Reversible absorption of CO₂ triggered by phase transition of amine-containing micro- and nanogel particles. *J. Am. Chem. Soc.* 134, 18177–18180. doi:10.1021/ja3080192
- Hosta-Rigau, L., Zhang, Y., Teo, B. M., Postma, A., Städler, B., Hosta-Rigau, L., et al. (2013). Cholesterol - a biological compound as a building block in bionanotechnology. *Nanoscale* 5, 89–109. doi:10.1039/c2nr32923a
- Hough, K. P., Curtiss, M. L., Blain, T. J., Liu, R.-M., Trevor, J., Deshane, J. S., et al. (2020). Airway remodeling in asthma. *Front. Med.* 7, 191. doi:10.3389/fmed.2020.00191
- Hsu, C.-H., Jay, M., Bummer, P. M., and Lehmler, H. J. (2003). Chemical stability of esters of nicotinic acid intended for pulmonary administration by liquid ventilation. *Pharm. Res.* 20, 918–925. doi:10.1023/a:1023899505837
- Hsu, H.-J., Bugno, J., Lee, S.-r., and Hong, S. (2016). Dendrimer-based nanocarriers: a versatile platform for drug delivery. *WIREs Nanomed. Nanobiotechnol.* 9, e1409. doi:10.1002/wnan.1409
- Huang, B., Yang, X.-D., Zhou, M.-M., Ozato, K., and Chen, L.-F. (2009). Brd4 coactivates transcriptional activation of NF- κ B via specific binding to acetylated RelA. *MCB (Mol. Cell. Biol.)* 29 (5), 1375–1387. doi:10.1128/mcb.01365-08
- Huang, X., Jain, P. K., El-Sayed, I. H., and El-Sayed, M. A. (2008). Plasmonic photothermal therapy (PTT) using gold nanoparticles. *Laser Med. Sci.* 23 (3), 217–228. doi:10.1007/s10103-007-0470-x
- Huang, Y.-Y., and Wang, C.-H. (2006). Pulmonary delivery of insulin by liposomal carriers. *J. Contr. Release* 113, 9–14. doi:10.1016/j.jconrel.2006.03.014
- Hui, D. S., Joynt, G. M., Wong, K. T., Gomersall, C. D., Li, T. S., Antonio, G., et al. (2005). Impact of severe acute respiratory syndrome (SARS) on pulmonary function, functional capacity and quality of life in a cohort of survivors. *Thorax* 60 (5), 401–409. doi:10.1136/thx.2004.030205
- Hung, C., Linn, G., Chow, Y.-H., Kobayashi, A., Mittelsteadt, K., Altemeier, W. A., et al. (2013). Role of lung pericytes and resident fibroblasts in the pathogenesis of pulmonary fibrosis. *Am. J. Respir. Crit. Care Med.* 188 (7), 820–830. doi:10.1164/rccm.201212-2297OC
- Hutchinson, J., Fogarty, A., Hubbard, R., and McKeever, T. (2015). Global incidence and mortality of idiopathic pulmonary fibrosis: a systematic review. *Eur. Respir. J.* 46 (3), 795–806. doi:10.1183/09031936.00185114
- Ijaz, T., Jamaluddin, M., Zhao, Y., Zhang, Y., Jay, J., Finnerty, C. C., et al. (2017). Coordinate activities of BRD4 and CDK9 in the transcriptional elongation complex are required for TGF β -induced Nox4 expression and myofibroblast transdifferentiation *Cell Death Dis.* 8 (2), e2606. doi:10.1038/cddis.2016.434
- Ijaz, T., Pazdrak, K., Kalita, M., Konig, R., Choudhary, S., Tian, B., et al. (2014). Systems biology approaches to understanding Epithelial Mesenchymal Transition (EMT) in mucosal remodeling and signaling in asthma. *World Allergy Organ. J.* 7 (1), 13. doi:10.1186/1939-4551-7-13
- Insmed (2020). Arikayce. Available at: <https://www.arikayce.com/about-arikayce/> (Accessed January 1, 2020).
- Ishida, T., Ichihara, M., Wang, X., Yamamoto, K., Kimura, J., Majima, E., et al. (2006). Injection of PEGylated liposomes in rats elicits PEG-specific IgM, which is responsible for rapid elimination of a second dose of PEGylated liposomes. *J. Contr. Release* 112, 12–25. doi:10.1016/j.jconrel.2006.01.005
- Ishihara, T., Maeda, T., Sakamoto, H., Takasaki, N., Shigyo, M., Ishida, T., et al. (2010). Evasion of the accelerated blood clearance phenomenon by coating of nanoparticles with various hydrophilic polymers. *Biomacromolecules* 11, 2700–2706. doi:10.1021/bm100754e
- Ishihara, T., Takeda, M., Sakamoto, H., Kimoto, A., Kobayashi, C., Takasaki, N., et al. (2009). Accelerated blood clearance phenomenon upon repeated injection of PEG-modified PLA-nanoparticles. *Pharm. Res.* 26, 2270–2279. doi:10.1007/s11095-009-9943-x
- Ivanova, V., Garbuzenko, O. B., Reuhl, K. R., Reimer, D. C., Pozharov, V. P., and Minko, T. (2013). Inhalation treatment of pulmonary fibrosis by liposomal prostaglandin E₂. *Eur. J. Pharm. Biopharm.* 84, 335–344. doi:10.1016/j.ejpb.2012.11.023
- Jartti, T., and Gern, J. E. (2017). Role of viral infections in the development and exacerbation of asthma in children. *J. Allergy Clin. Immunol.* 140 (4), 895–906. doi:10.1016/j.jaci.2017.08.003
- Jeong, W.-j., Bu, J., Han, Y., Drelich, A. J., Nair, A., Král, P., et al. (2020). Nanoparticle conjugation stabilizes and multimerizes β -hairpin peptides to effectively target PD-1/PD-L1 β -sheet-rich interfaces. *J. Am. Chem. Soc.* 142, 1832–1837. doi:10.1021/jacs.9b10160
- Jeong, W., Bu, J., Kubiawicz, L. J., Chen, S. S., Kim, Y., and Hong, S. (2018). Peptide-nanoparticle conjugates: a next generation of diagnostic and therapeutic platforms? *Nano Converg.* 5, 38. doi:10.1186/s40580-018-0170-1
- Jin, H., He, Y., Zhao, P., Hu, Y., Tao, J., Chen, J., et al. (2019). Targeting lipid metabolism to overcome EMT-associated drug resistance via integrin β 3/FAK pathway and tumor-associated macrophage repolarization using legumain-activatable delivery. *Theranostics* 9, 265–278. doi:10.7150/thno.27246
- Johnsen, K. B., Gudbergsson, J. M., Skov, M. N., Pilgaard, L., Moos, T., and Duroux, M. (2014). A comprehensive overview of exosomes as drug delivery vehicles - endogenous nanocarriers for targeted cancer therapy. *Biochim. Biophys. Acta* 1846, 75–87. doi:10.1016/j.bbcan.2014.04.005
- Johnson, E. R., and Matthay, M. A. (2010). Acute lung injury: epidemiology, pathogenesis, and treatment. *J. Aerosol Med. Pulm. Drug Deliv.* 23 (4), 243–252. doi:10.1089/jamp.2009.0775
- Johnstone, T. C., and Lippard, S. J. (2013). The effect of ligand lipophilicity on the nanoparticle encapsulation of Pt(IV) prodrugs. *Inorg. Chem.* 52, 9915–9920. doi:10.1021/ic4010642
- Kabanov, A. V., and Vinogradov, S. V. (2009). Nanogels as pharmaceutical carriers: finite networks of infinite capabilities. *Angew. Chem. Int. Ed.* 48, 5418–5429. doi:10.1002/anie.200900441
- Kalluri, R., and Weinberg, R. A. (2009). The basics of epithelial-mesenchymal transition. *J. Clin. Invest.* 119 (6), 1420–1428. doi:10.1172/JCI39104
- Kanninen, P., Johans, C., Merta, J., and Kontturi, K. (2008). Influence of ligand structure on the stability and oxidation of copper nanoparticles. *J. Colloid Interface Sci.* 318, 88–95. doi:10.1016/j.jcis.2007.09.069
- Karo-Atar, D., Bordowitz, A., Wand, O., Pasmanik-Chor, M., Fernandez, I. E., Itan, M., et al. (2016). A protective role for IL-13 receptor α 1 in bleomycin-induced pulmonary injury and repair. *Mucosal Immunol.* 9 (1), 240–253. doi:10.1038/mi.2015.56
- Kaur, A., Mathai, S. K., and Schwartz, D. (2017). Genetics in idiopathic pulmonary fibrosis pathogenesis, prognosis, and treatment. *Front. Med.* 4, 1–8. doi:10.3389/fmed.2017.00154
- Kaushik, N. K., Kaushik, N., Yoo, K. C., Uddin, N., Kim, J. S., Lee, S. J., et al. (2016). Low doses of PEG-coated gold nanoparticles sensitize solid tumors to cold plasma by blocking the PI3K/AKT-driven signaling axis to suppress cellular transformation by inhibiting growth and EMT. *Biomaterials* 87, 118–130. doi:10.1016/j.biomaterials.2016.02.014
- Kesharwani, P., Jain, K., and Jain, N. K. (2014). Dendrimer as nanocarrier for drug delivery. *Prog. Polym. Sci.* 39, 268–307. doi:10.1016/j.progpolymsci.2013.07.005
- Killops, K. L., Campos, L. M., and Hawker, C. J. (2008). Robust, efficient, and orthogonal synthesis of dendrimers via thiol-ene “click” chemistry. *J. Am. Chem. Soc.* 130, 5062–5064. doi:10.1021/ja8006325
- King, T. E., Jr., Schwarz, M. I., Brown, K., Tooze, J. A., Colby, T. V., Waldron, J. A., Jr., et al. (2001). Idiopathic pulmonary fibrosis. *Am. J. Respir. Crit. Care Med.* 164 (6), 1025–1032. doi:10.1164/ajrccm.164.6.2001056
- Kinnear, C., Moore, T. L., Rodriguez-Lorenzo, L., Rothen-Rutishauser, B., and Petri-Fink, A. (2017). Form follows function: nanoparticle shape and its implications for nanomedicine. *Chem. Rev.* 117, 11476–11521. doi:10.1021/acs.chemrev.7b00194
- Knight, V., Koshkina, N. V., Waldrep, J. C., Giovanella, B. C., and Gilbert, B. E. (1999). Anticancer effect of 9-nitrocaptoprothecin liposome aerosol on human cancer xenografts in nude mice. *Canc. Chemother. Pharmacol.* 44, 177–186. doi:10.1007/s002800050965

- Kolte, A., Patil, S., Lesimple, P., Hanrahan, J. W., and Misra, A. (2017). PEGylated composite nanoparticles of PLGA and polyethylenimine for safe and efficient delivery of pDNA to lungs. *Int. J. Pharm.* 524, 382–396. doi:10.1016/j.ijpharm.2017.03.094
- Kong, M., Tang, J., Qiao, Q., Wu, T., Qi, Y., Tan, S., et al. (2017). Biodegradable hollow mesoporous silica nanoparticles for regulating tumor microenvironment and enhancing antitumor efficiency. *Theranostics* 7, 3276–3292. doi:10.7150/thno.19987
- Koshkina, N. V., Kleinerman, E. S., Waidrep, C., Jia, S. F., Worth, L. L., Gilbert, B. E., et al. (2000). 9-Nitrocamptothecin liposome aerosol treatment of melanoma and osteosarcoma lung metastases in mice. *Clin. Canc. Res.* 6, 2876–2880. doi:10.1111/j.1749-6632.2000.tb07033.x
- Koshkina, N. V., Waldrep, J. C., Roberts, L. E., Golunski, E., Melton, S., and Knight, V. (2001). Paclitaxel liposome aerosol treatment induces inhibition of pulmonary metastases in murine renal carcinoma model. *Clin. Canc. Res.* 7, 3258–3262.
- Kousalova, J., and Etrych, T. (2018). Polymeric nanogels as drug delivery systems. *Physiol. Res.* 67, S305–S317. doi:10.33549/physiolres.933979
- Kricheldorf, H. R. (1991). *Handbook of polymer synthesis*. Hoboken, NJ: John Wiley & Sons.
- Kroger, A. P. P., and Paulusse, J. M. J. (2018). Single-chain polymer nanoparticles in controlled drug delivery and targeted imaging. *J. Contr. Release* 286, 326–347. doi:10.1016/j.jconrel.2018.07.041
- Kulkarni, T., de Andrade, J., Zhou, Y., Luckhardt, T., and Thannickal, V. J. (2016). Alveolar epithelial disintegrity in pulmonary fibrosis. *Am. J. Physiol. Lung Cell Mol. Physiol.* 311 (2), L185–L191. doi:10.1152/ajplung.00115.2016
- Kumar, V., Dong, Y., Kumar, V., Almawash, S., and Mahato, R. I. (2019). The use of micelles to deliver potential hedgehog pathway inhibitor for the treatment of liver fibrosis. *Theranostics* 9, 7537–7555. doi:10.7150/thno.38913
- Kwon, D.-H., Kim, Y.-K., and Kook, H. (2017). New aspects of vascular calcification: histone deacetylases and beyond. *J. Kor. Med. Sci.* 32, 1738–1748. doi:10.3346/jkms.2017.32.11.1738
- Lamouille, S., Xu, J., and Derynck, R. (2014). Molecular mechanisms of epithelial–mesenchymal transition. *Nat. Rev. Mol. Cell Biol.* 15 (13), 178–196. doi:10.1038/nrm3758
- Landmark, K. J., DiMaggio, S., Ward, J., Kelly, C., Vogt, S., Hong, S., et al. (2008). Synthesis, characterization, and *in Vitro* testing of superparamagnetic iron oxide nanoparticles targeted using folic acid-conjugated dendrimers. *ACS Nano*. 2, 773–783. doi:10.1021/nm800034w
- Langenback, E. G., Davis, J. M., Robbins, C., Sahgal, N., Perry, R. J., and Simon, S. R. (1999). Improved pulmonary distribution of recombinant human Cu/Zn superoxide dismutase, using a modified ultrasonic nebulizer. *Pediatr. Pulmonol.* 27, 124–129. doi:10.1002/(sici)1099-0496(199902)27:2<124::aid-ppl9>3.0.co;2-e
- Lehmle, H., Xu, L., Vyas, S., Ojogun, V., Knutson, B., and Ludewig, G. (2008). Synthesis, physicochemical properties and *in vitro* cytotoxicity of nicotinic acid ester prodrugs intended for pulmonary delivery using perfluorooctyl bromide as vehicle. *Int. J. Pharm.* 353, 35–44. doi:10.1016/j.ijpharm.2007.11.011
- Letchford, K., and Burt, H. (2007). A review of the formation and classification of amphiphilic block copolymer nanoparticle structures: micelles, nanospheres, nanocapsules and polymersomes. *Eur. J. Pharm. Biopharm.* 65, 259–269. doi:10.1016/j.ejpb.2006.11.009
- Letsou, G. V., Safi, H. J., Reardon, M. J., Ergenoglu, M., Li, Z., Klonaris, C. N., et al. (1999). Pharmacokinetics of liposomal aerosolized cyclosporine A for pulmonary immunosuppression. *Ann. Thorac. Surg.* 68, 2044–2048. doi:10.1016/s0003-4975(99)01183-2
- Li, T., Yan, Y., Wang, B., Qian, H., Zhang, X., Shen, L., et al. (2013). Exosomes derived from human umbilical cord mesenchymal stem cells alleviate liver fibrosis. *Stem Cell Dev.* 22, 845. doi:10.1089/scd.2012.0395
- Li, X., Song, L., Hu, X., Liu, C., Shi, J., Wang, H., et al. (2018a). Inhibition of epithelial–mesenchymal transition and tissue regeneration by waterborne titanium dioxide nanoparticles. *ACS Appl. Mater. Interfaces* 10, 3449–3458. doi:10.1021/acsami.7b18986
- Li, X., Jiang, P., Li, J., Peng, M., Zhao, X., Zhang, X., et al. (2018b). Tumor-derived exosomal lnc-Sox2ot promotes EMT and stemness by acting as a ceRNA in pancreatic ductal adenocarcinoma. *Oncogene* 37, 3822–3838. doi:10.1038/s41388-018-0237-9
- Li, X., Yuan, W., Gu, S., and Ren, J. (2010). Synthesis and self-assembly of tunable thermosensitive chitosan amphiphilic copolymers by click chemistry. *Mater. Lett.* 64, 2663–2666. doi:10.1016/j.matlet.2010.08.077
- Li, Y.-F., and Chen, C. (2011). Fate and toxicity of metallic and metal-containing nanoparticles for biomedical applications. *Small* 7 (21), 2965–2980. doi:10.1002/smll.201101059
- Li, Z., Chen, G., Ding, L., Wang, Y., Zhu, C., Wang, K., et al. (2019). Increased survival by pulmonary treatment of established lung metastases with dual STAT3/CXCR4 inhibition by siRNA nanoemulsions. *Mol. Ther.* 27, 2100–2110. doi:10.1016/j.ymthe.2019.08.008
- Limbach, L. K., Wick, P., Manser, P., Grass, R. N., Bruinink, A., and Stark, W. J. (2007). Exposure of engineered nanoparticles to human lung epithelial cells: influence of chemical composition and catalytic activity on oxidative stress. *Environ. Sci. Technol.* 41, 4158–4163. doi:10.1021/es062629t
- Lin, Y., Ma, Q., Li, L., and Wang, H. (2018). The CXCL12–CXCR4 axis promotes migration, invasiveness, and EMT in human papillary thyroid carcinoma B-CPAP cells via NF- κ B signaling. *Biochem. Cell. Biol.* 96, 619–626. doi:10.1139/bcb-2017-0074
- Lin, Y.-T., and Wu, K.-J. (2020). Epigenetic regulation of epithelial–mesenchymal transition: focusing on hypoxia and TGF- β signaling. *J. Biomed. Sci.* 27 (1), 39. doi:10.1186/s12929-020-00632-3
- Liu, Z., Chen, H., Wang, P., Li, Y., Wold, E. A., Leonard, P. G., et al. (2020). Discovery of orally bioavailable chromone derivatives as potent and selective BRD4 inhibitors: scaffold hopping, optimization, and pharmacological evaluation. *J. Med. Chem.* 63, 5242. doi:10.1021/acs.jmedchem.0c00035
- Liu, Z., Tian, B., Chen, H., Wang, P., Brasier, A. R., and Zhou, J. (2018). Discovery of potent and selective BRD4 inhibitors capable of blocking TLR3-induced acute airway inflammation. *Eur. J. Med. Chem.* 151, 450–461. doi:10.1016/j.ejmech.2018.04.006
- Lu, L., Lou, B., Zou, S., Kobayashi, H., Liu, J., Xiao, L., et al. (2018). Robust removal of ligands from noble metal nanoparticles by electrochemical strategies. *ACS Catal.* 8, 8484–8492. doi:10.1021/acscatal.8b01627
- Lutter, R., and Spiteri, M. (2005). Current perspectives in epithelial cell injury and repair: consequences for epithelial functions. *Eur. Respir. Rev.* 14, 126–130. doi:10.1183/09059180.05.00009701
- Ma, J., Bishoff, B., Mercer, R. R., Barger, M., Schwegler-Berry, D., and Castranova, V. (2017). Role of epithelial–mesenchymal transition (EMT) and fibroblast function in cerium oxide nanoparticles-induced lung fibrosis. *Toxicol. Appl. Pharmacol.* 323, 16–25. doi:10.1016/j.taap.2017.03.015
- Mack, M. (2018). Inflammation and fibrosis. *Matrix Biol.* 68–69, 106–121. doi:10.1016/j.matbio.2017.11.010
- Mamaghani, P. Y., Kaffashi, B., Salehi, R., and Davaran, S. (2015). Synthesis, characterization, and viscoelastic behavior of thermothickening poly (N-Isopropylacrylamide-Methacrylic acid-Vinylpyrrolidone) nanogels as an injectable biocompatible drug carrier. *Int. J. Polym. Mater. Polym. Biomater.* 64, 55–63. doi:10.1080/00914037.2014.886236
- Mapel, D. W., Samet, J. M., and Coultas, D. B. (1996). Corticosteroids and the treatment of idiopathic pulmonary fibrosis. *Chest* 110 (4), 1058–1067. doi:10.1378/chest.110.4.1058
- Mazzei, M. E., Richeldi, L., and Collard, H. R. (2015). Nintedanib in the treatment of idiopathic pulmonary fibrosis. *Ther. Adv. Respir.* 9 (3), 121–129. doi:10.1177/1753465815579365
- Menon, J. U., Ravikumar, P., Pise, A., Gyawali, D., Hsia, C. C. W., and Nguyen, K. T. (2014). Polymeric nanoparticles for pulmonary protein and DNA delivery. *Acta Biomater.* 10, 2643–2652. doi:10.1016/j.actbio.2014.01.033
- Menon, J. U., Wadajkar, A. S., Xie, Z., and Nguyen, K. T. (2013). “Nanomaterials for management of lung disorders and drug delivery,” in *Nanomaterials in Drug Delivery, Imaging, and Tissue Engineering*. Editor A. Tiwari (Hoboken, NJ: John Wiley & Sons, Inc.), 167–202.
- Mercer, B., Lemaître, V., Powell, C., and D’Armiento, J. (2006). The epithelial cell in lung Health and emphysema pathogenesis. *Curr Respir Med Rev.* 2 (2), 101–142. doi:10.2174/157339806776843085
- Min, H., Sun, X., Yang, X., Zhu, H., Liu, J., Wang, Y., et al. (2018). Exosomes derived from irradiated esophageal carcinoma-infiltrating T cells promote metastasis by inducing the epithelial–mesenchymal transition in esophageal cancer cells. *Pathol. Oncol. Res.* 24, 11–18. doi:10.1007/s12253-016-0185-z

- Mo, X., Jian, W., Su, Z., Chen, M., Peng, H., Peng, P., et al. (2020). Abnormal pulmonary function in COVID-19 patients at time of hospital discharge. *Eur. Respir. J.* 55, 2001217. doi:10.1183/13993003.01217-2020
- Mody, V., Siwale, R., Singh, A., and Mody, H. (2010). Introduction to metallic nanoparticles. *J Pharm Bioanal Sci* 2, 282–289. doi:10.4103/0975-7406.72127
- Moen, E. L., Mariani, C. J., Zullo, H., Jeff-Eke, M., Litwin, E., and Nikitas, J. N. (2015). New themes in the biological functions of 5-methylcytosine and 5-hydroxymethylcytosine. *Immunol. Rev.* 263 (1), 36–49. doi:10.1111/imr.12242
- Mohammad, A. K., and Reineke, J. J. (2013). Quantitative detection of PLGA nanoparticle degradation in tissues following intravenous administration. *Mol. Pharm.* 10, 2183–2189. doi:10.1021/mp300559v
- Naguchi, S., Yamauchi, Y., and Takizawa, H. (2014). Novel therapeutic strategies for fibrotic lung disease: a review with a focus on epithelial-mesenchymal transition. *Recent Pat. Inflamm. Allergy Drug Discov.* 8, 9–18. doi:10.2174/1872213x07666131229131451
- Nam, J., Son, S., Park, K. S., Zou, W., Shea, L. D., and Moon, J. J. (2019). Cancer nanomedicine for combination cancer immunotherapy. *Nat. Rev. Mater.* 4, 398–414. doi:10.1038/s41578-019-0108-1
- Nance, E. A., Woodworth, G. F., Sailor, K. A., Shih, T.-Y., Xu, Q., Swaminatha, G., et al. (2012). A dense poly(ethylene glycol) coating improves penetration of large polymeric nanoparticles within brain tissue. *Sci. Transl. Med.* 4, 149ra119. doi:10.1126/scitranslmed.3003594
- Newman, S. P. (2018). Delivering drugs to the lungs: the history of repurposing in the treatment. *Adv. Drug Deliv. Rev.* 133, 135–118. doi:10.1016/j.addr.2018.04.010
- Niu, Z., and Li, Y. (2014). Removal and utilization of capping agents in nanocatalysis. *Chem. Mater.* 26, 72–83. doi:10.1021/cm4022479
- Oku, N., and Namba, Y. (1994). Long-circulating liposomes. *Crit. Rev. Ther. Drug Carrier Syst.* 11, 231–270. doi:10.4155/tde-2018-0035
- O'Reilly, S. (2017). Epigenetics in fibrosis. *Mol. Aspect. Med.* 54, 89–102. doi:10.1016/j.mam.2016.10.001
- Owens, D. E., and Peppas, N. A. (2006). Opsonization, biodistribution, and pharmacokinetics of polymeric nanoparticles. *Int. Journal Pharm.* 307, 93–102. doi:10.1016/j.ijpharm.2005.10.010
- Palgrave, R. G., and Parkin, I. P. (2006). Aerosol assisted chemical vapor deposition using nanoparticle precursors: a route to nanocomposite thin films. *J. Am. Chem. Soc.* 128, 1587–1597. doi:10.1021/ja055563v
- Pandey, R., Sharma, A., Zahoor, A., Sharma, S., Khuller, G. K., and Prasad, B. (2003). Poly (dl-lactide-co-glycolide) nanoparticle-based inhalable sustained drug delivery system for experimental tuberculosis. *J. Antimicrob. Chemother.* 52, 981–986. doi:10.1093/jac/dkg477
- Pandolfi, L., Frangipane, V., Bocca, C., Marengo, A., Tarro Genta, E., Bozzini, S., et al. (2019). Hyaluronic acid-decorated liposomes as innovative targeted delivery system for lung fibrotic cells. *Molecules* 24, 3291. doi:10.3390/molecules24183291
- Park, K., Kim, J.-H., Nam, Y. S., Lee, S., Nam, H. Y., Kim, K., et al. (2007). Effect of polymer molecular weight on the tumor targeting characteristics of self-assembled glycol chitosan nanoparticles. *J. Contr. Release* 122, 305–314. doi:10.1016/j.jconrel.2007.04.009
- Patel, P., Soni, T., Thakkar, V., and Gandhi, T. (2013). Nanoparticle as an emerging tool in pulmonary drug delivery system. *Micro Nanosys.* 5, 288–302. doi:10.2174/18764029113059990002
- Pearson, R. M., Sen, S., Hsu, H.-j., Pasko, M., Gaske, M., Král, P., et al. (2016). Tuning the selectivity of dendron micelles through variations of the poly(ethylene glycol) corona. *ACS Nano.* 10 (7), 6905–6914. doi:10.1021/acsnano.6b02708
- Peer, D., Karp, J. M., Hong, S., Farokhzad, O. C., Margalit, R., and Langer, R. (2007). Nanocarriers as an emerging platform for cancer therapy. *Nat. Nanotechnol.* 2, 751–760. doi:10.1038/nnano.2007.387
- Peng, F., Tee, J. K., Setyawati, M. I., Ding, X., Yeo, H. L. A., Tan, Y. L., et al. (2018). Inorganic nanomaterials as highly efficient inhibitors of cellular hepatic fibrosis. *ACS Appl. Mater. Interfaces* 10, 31938–31946. doi:10.1021/acsmi.8b10527
- Phan, S. H. (2012). Genesis of the myofibroblast in lung injury and fibrosis. *Proc. Am. Thorac. Soc.* 9 (3), 148–152. doi:10.1513/pats.201201-011aw
- Poste, G., Paphadjopoulos, D., and Vail, W. J. (1976). Chapter 4 lipid vesicles as carriers for introducing biologically active materials into cells. *Methods Cell Biol.* 14, 33–71. doi:10.1016/s0091-679x(08)60468-9
- Pradhan, P., Qin, H., Leleux, J. A., Gwak, D., Sakamaki, I., Kwak, L. W., et al. (2014). The effect of combined IL10 siRNA and CpG ODN as pathogen-mimicking microparticles on Th1/Th2 cytokine balance in dendritic cells and protective immunity against B cell lymphoma. *Biomaterials* 35, 5491–5504. doi:10.1016/j.biomaterials.2014.03.039
- Qiao, J.-B., Jang, Y., Fan, Q.-Q., Chang, S.-H., Xing, L., Cui, P.-F., et al. (2017). Aerosol delivery of biocompatible dihydroergotamine-loaded PLGA-PSPE polymeric micelles for efficient lung cancer therapy. *Polym. Chem.* 8, 1540–1554. doi:10.1039/c7py00024c
- Quadros, M. E., and Marr, L. C. (2012). Environmental and human Health risks of aerosolized silver nanoparticles. *J. Air Waste Manag. Assoc.* 60, 770–781. doi:10.3155/1047-3289.60.7.770
- Radisky, D. C., Kenny, P. A., and Bissell, M. J. (2007). Fibrosis and cancer: do myofibroblasts come also from epithelial cells via EMT? *J. Cell. Biochem.* 101(4), 830–839. doi:10.1002/jcb.21186
- Raghnail, M. N., Bramini, M., Ye, D., Couraud, P.-O., Romero, I. A., Weksler, B., et al. (2014). Paracrine signalling of inflammatory cytokines from an *in vitro* blood brain barrier model upon exposure to polymeric nanoparticles. *Analyst* 139, 923–930. doi:10.1039/c3an01621h
- Raghu, G., Collard, H. R., Egan, J. J., Martinez, F. J., Behr, J., Brown, K. K., et al. (2011). An official ATS/ERS/JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. *Am. J. Respir. Crit. Care Med.* 183 (6), 788–824. doi:10.1164/rccm.2009-040GL
- Richeldi, L., du Bois, R. M., Raghu, G., Azuma, A., Brown, K. K., Costabel, U., et al. (2014). Efficacy and safety of nintedanib in idiopathic pulmonary fibrosis. *N. Engl. J. Med.* 370 (22), 2071–2082. doi:10.1056/NEJMoa1402584
- Rock, J. R., Barkauskas, C. E., Cronce, M. J., Xue, Y., Harris, J. R., Liang, J., et al. (2011). Multiple stromal populations contribute to pulmonary fibrosis without evidence for epithelial to mesenchymal transition. *Proc. Natl. Acad. Sci. U. S. A.* 108 (52), E1475–E1483. doi:10.1073/pnas.1117988108
- Rosiere, R., Van Woensel, M., Mathieu, V., Langer, I., Mathivet, T., Vermeersch, M., et al. (2016). Development and evaluation of well-tolerated and tumor-penetrating polymeric micelle-based dry powders for inhaled anti-cancer chemotherapy. *Int. J. Pharm.* 501, 148–159. doi:10.1016/j.ijpharm.2016.01.073
- Royce, S. G., and Karagiannis, T. C. (2014). Histone deacetylases and their inhibitors. *Curr. Opin. Allergy Clin. Immunol.* 14 (1), 44–48. doi:10.1097/ACI.0000000000000029
- Ruan, Q., Yang, K., Wang, W., Jiang, L., and Song, J. (2020). Clinical predictors of mortality due to COVID-19 based on an analysis of data of 150 patients from Wuhan, China. *Intensive Care Med.* 46 (5), 846–848. doi:10.1007/s00134-020-05991-x
- Saadati, R., Dadashzadeh, S., Abbasian, Z., and Soleimanjahi, H. (2013). Accelerated blood clearance of PEGylated PLGA nanoparticles following repeated injections: effects of polymer dose, PEG coating, and encapsulated anticancer drug. *Pharm. Res.* 30, 985–995. doi:10.1007/s11095-012-0934-y
- Seifi-Najmi, M., Hajivalili, M., Safaralizadeh, R., Sadreddini, S., Esmaili, S., Razavi, R., et al. (2016). SiRNA/DOX loaded chitosan based nanoparticles: development, Characterization and *in vitro* evaluation on A549 lung cancer cell line. *Cell Mol Biol (Noisy-le-grand)* 62, 87.
- Senior, J. H. (1986). Fate and behavior of liposomes *in vivo*: a review of controlling factors. *Crit. Rev. Ther. Drug Carrier Syst.* 3, 123–193.
- Shah, P. N., Lin, L. Y., Smolen, J. A., Tagaev, J. A., Gunsten, S. P., Han, D. S., et al. (2013). Synthesis, characterization, and *in vivo* efficacy of shell cross-linked nanoparticle formulations carrying silver antimicrobials as aerosolized therapeutics. *ACS Nano.* 7, 4977–4987. doi:10.1021/nn400322f
- Shargh, V. H., Hondermarck, H., and Liang, M. (2015). Antibody-targeted biodegradable nanoparticles for cancer therapy. *Futur. Med.* 11, 63–79. doi:10.2217/nmm.15.186
- Sheng, Y., Liu, C., Yuan, Y., Tao, X., Yang, F., Shan, X., et al. (2009). Long-circulating polymeric nanoparticles bearing a combinatorial coating of PEG and water-soluble chitosan. *Biomaterials* 30, 2340–2348. doi:10.1016/j.biomaterials.2008.12.070
- Shi, H., Han, X., Jiang, N., Cao, Y., Alwalid, O., Gu, J., et al. (2020). Radiological findings from 81 patients with COVID-19 pneumonia in Wuhan, China: a descriptive study. *Lancet Infect. Dis.* 20 (4), 425–434. doi:10.1016/S1473-3099(20)30086-4
- Shi, X., Bi, X., Ganser, T. R., Hong, S., Myc, L. A., Desai, A., et al. (2006). HPLC analysis of functionalized poly(amidoamine) dendrimers and the interaction

- between a folate-dendrimer conjugate and folate binding protein. *Analyst* 131, 842–848. doi:10.1039/b602546c
- Shin, H. Y. (2018). Targeting super-enhancers for disease treatment and diagnosis. *Mol Cells* 41 (6), 506–514. doi:10.14348/molcells.2018.2297
- Shrestha, N., Shahbazi, M. A., Araújo, F., Mäkilä, E., Raula, J., Kauppinen, E. I., et al. (2015). Multistage pH-responsive mucoadhesive nanocarriers prepared by aerosol flow reactor technology: a controlled dual protein-drug delivery system. *Biomaterials* 68, 9–20. doi:10.1016/j.biomaterials.2015.07.045
- Silverman, J. M., and Reiner, N. E. (2011). Exosomes and other microvesicles in infection biology: organelles with unanticipated phenotypes. *Cell Microbiol.* 13, 1–9. doi:10.1111/j.1462-5822.2010.01537.x
- Singh, A., Kulshrestha, R., Pandey, A., Kaushik, M., and Dinda, A. K. (2019). "Efficacy of MePEG PCL diblock copolymeric nanoparticles and PCLNanoparticles for drug delivery in lung fibrosis," in American thoracic society 2019 international conference, Dallas, TX, May 17–22, 2019.
- Singh, N. K., Purkayastha, B. D., Roy, J. K., Banik, R. M., Yashpal, M., Singh, G., et al. (2010). Nanoparticle-induced controlled biodegradation and its mechanism in poly(ϵ -caprolactone). *ACS Appl. Mater. Interfaces* 2, 69–81. doi:10.1021/am900584r
- Skubitz, K. M., and Anderson, P. M. (2000). Inhalational interleukin-2 liposomes for pulmonary metastases: a phase I clinical trial. *Anti Canc. Drugs* 11, 555–563. doi:10.1097/00001813-200008000-00006
- Song, Q., Yin, Y., Shang, L., Wu, T., Zhang, D., Kong, M., et al. (2017). Tumor microenvironment responsive nanogel for the combinatorial antitumor effect of chemotherapy and immunotherapy. *Nano Lett.* 17, 6366–6375. doi:10.1021/acs.nanolett.7b03186
- Sousa, C., Gouveia, L., Kreutzer, B., Silva-Lima, B., Maphasa, R., Dube, A., et al. (2019). Polymeric micellar formulation enhances antimicrobial and anticancer properties of salinomycin. *Pharm. Res.* 36, 83. doi:10.1007/s11095-019-2615-6
- Spagnolo, P., Del Giovane, C., Luppi, F., Cerri, S., Balduzzi, S., Walters, E. H., et al. (2010). Non-steroid agents for idiopathic pulmonary fibrosis. *Cochrane Database Syst. Rev.* 9, CD003134. doi:10.1002/14651858.CD003134.pub2
- Stone, R. C., Pastar, I., Ojeh, N., Chen, V., Liu, S., Garzon, K. I., et al. (2016). Epithelial-mesenchymal transition in tissue repair and fibrosis. *Cell Tissue Res.* 365, 495–506. doi:10.1007/s00441-016-2464-0
- Stubenrauch, K., Moitzl, C., Fritz, G., Glatter, O., Trimmel, G., and Stelzer, F. (2006). Precise tuning of micelle, core, and shell size by the composition of amphiphilic block copolymers derived from ROMP investigated by DLS and SAXS. *Macromolecules* 39, 5865–5874. doi:10.1021/ma060451p
- Sunil Gowda, S. N., Rajasowmiya, S., Vadevel, V., Banu Devi, S., Celestin Jerald, A., Marimuthu, S., et al. (2018). Gallic acid-coated silver nanoparticle alters the expression of radiation-induced epithelial-mesenchymal transition in non-small lung cancer cells. *Toxicol. Vitro* 52, 170–177. doi:10.1016/j.tiv.2018.06.015
- Sunoqrot, S., Bugno, J., Lantvit, D., Burdette, J. E., and Hong, S. (2014). Prolonged blood circulation and enhanced tumor accumulation of folate-targeted dendrimer-polymer hybrid nanoparticles. *J. Contr. Release* 191, 115–122. doi:10.1016/j.jconrel.2014.05.006
- Suresh, D., Zambre, A., Mukherjee, S., Ghoshdastidar, S., Jiang, Y., Joshi, T., et al. (2019). Silencing AXL by covalent siRNA-gelatin-antibody nanoconjugate inactivates mTOR/EMT pathway and stimulates p53 for TKI sensitization in NSCLC. *Nanomed. Nanotechnol. Biol. Med.* 20, 102007. doi:10.1016/j.nano.2019.04.010
- Szebeni, J. (2005). Complement activation-related pseudoallergy: a new class of drug-induced acute immune toxicity. *Toxicology* 216, 106–121. doi:10.1016/j.tox.2005.07.023
- Szebeni, J., and Moghimi, S. M. (2009). Liposome triggering of innate immune responses: a perspective on benefits and adverse reactions. *J. Liposome Res.* 19, 85–90. doi:10.1080/08982100902792855
- Tang, B. C., Dawson, M., Lai, S. K., Wang, Y.-Y., Suk, J. S., Yang, M., et al. (2009). Biodegradable polymer nanoparticles that rapidly penetrate the human mucus barrier. *Proc. Natl. Acad. Sci. U. S. A.* 106, 19268–19273. doi:10.1073/pnas.0905998106
- Tang, X., Peng, R., Phillips, J. E., Deguzman, J., Ren, Y., Apparsundaram, S., et al. (2013). Assessment of Brd4 inhibition in idiopathic pulmonary fibrosis lung fibroblasts and *in vivo* models of lung fibrosis. *Am. J. Pathol.* 183 (2), 470–479. doi:10.1016/j.ajpath.2013.04.020
- Taniguchi, Y. (2016). The bromodomain and extra-terminal domain (BET) family: functional anatomy of BET paralogous proteins. *IJMS* 17 (11), 1849. doi:10.3390/ijms17111849
- Taylor, K. M. G., Taylor, G., Kellaway, I. W., and Stevens, J. (1990). The stability of liposomes to nebulisation. *Int. J. Pharm.* 58, 57–61. doi:10.1016/0378-5173(90)90287-e
- Ten, R. M., Anderson, P. M., Zein, N. N., Temesgen, Z., Clawson, M. L., and Weiss, W. (2002). Interleukin-2 liposomes for primary immune deficiency using the aerosol route. *Int. Immunopharm.* 2, 333–344. doi:10.1016/s1567-5769(01)00143-6
- Thannickal, V. J. (2012). Mechanisms of pulmonary fibrosis: role of activated myofibroblasts and NADPH oxidase. *Fibrogenesis Tissue Repair* 5 (1), S23. doi:10.1186/1755-1536-5-s1-s23
- Thery, C., Zitvogel, L., and Amigorena, S. (2002). Exosomes: composition, biogenesis and function. *Nat. Rev. Immunol.* 2, 569–579. doi:10.1038/nri855
- Thomas, D. A., Myers, M. A., Wichert, B., Schreier, H., and Gonzalez-Rothi, R. J. (1991). Acute effects of liposome aerosol inhalation on pulmonary function in healthy human volunteers. *Chest* 99, 1268–1270. doi:10.1378/chest.99.5.1268
- Tian, B., Li, X., Kalita, M., Widen, S. G., Yang, J., Bhavnani, S. K., et al. (2015). Analysis of the TGF β -induced program in primary airway epithelial cells shows essential role of NF- κ B/RelA signaling network in type II epithelial mesenchymal transition. *BMC Genom.* 16 (1), 529. doi:10.1186/s12864-015-1707-x
- Tian, B., Liu, Z., Litvinov, J., Maroto, R., Jamaluddin, M., Rytting, E., et al. (2018a). Efficacy of novel highly specific bromodomain-containing protein 4 inhibitors in innate inflammation-driven airway remodeling. *Am. J. Respir. Cell Mol. Biol.* 60, 68. doi:10.1165/rcmb.2017-0445OC
- Tian, B., Liu, Z., Yang, J., Sun, H., Zhao, Y., Wakamiya, M., et al. (2018b). Selective antagonists of the bronchiolar epithelial NF- κ B-Bromodomain-Containing protein 4 pathway in viral-induced airway inflammation. *Cell Rep.* 23 (4), 1138–1151. doi:10.1016/j.celrep.2018.03.106
- Tian, B., Widen, S. G., Yang, J., Wood, T. G., Kudlicki, A., Zhao, Y., et al. (2018c). NF κ B is a master transcription factor mediating partial-to fully committed epithelial-mesenchymal transition. *J. Biol. Chem.*
- Tian, B., Yang, J., Zhao, Y., Ivanciu, T., Sun, H., Wakamiya, M., et al. (2018d). Central role of the NF- κ B pathway in the Scgb1a1-expressing epithelium in mediating respiratory syncytial virus-induced airway inflammation. *J. Virol.* 92 (11), e00441. doi:10.1128/JVI.00441-18
- Tian, B., Patrikeev, I., Ochoa, L., Vargas, G., Belanger, K. K., Litvinov, J., et al. (2017a). NF- κ B mediates mesenchymal transition, remodeling, and pulmonary fibrosis in response to chronic inflammation by viral RNA patterns. *Am. J. Respir. Cell Mol. Biol.* 56 (4), 506–520. doi:10.1165/rcmb.2016-0259OC
- Tian, B., Yang, J., Zhao, Y., Ivanciu, T., Sun, H., Garofalo, R. P., et al. (2017b). Bromodomain containing 4 (BRD4) couples NF κ B/RelA with airway inflammation and the IRF-RIG-I amplification loop in respiratory syncytial virus infection. *J. Virol.* 91, e00007–e000017. doi:10.1128/JVI.00007-00017
- Tian, B., Zhao, Y., Sun, H., Zhang, Y., Yang, J., and Brasier, A. R. (2016). BRD4 mediates NF- κ B-dependent epithelial-mesenchymal transition and pulmonary fibrosis via transcriptional elongation. *Am. J. Physiol. Lung Cell Mol. Physiol.* 311 (6), L1183–L1201. doi:10.1152/ajplung.00224.2016
- Tomasek, J. J., Gabbiani, G., Hinz, B., Chaponnier, C., and Brown, R. A. (2002). Myofibroblasts and mechano-regulation of connective tissue remodelling. *Nat. Rev. Mol. Cell Biol.* 3 (5), 349–363. doi:10.1038/nrm809
- Torikata, C., Villiger, B., Kuhn, C., and McDonald, J. A. (1985). Ultrastructural distribution of fibronectin in normal and fibrotic human lung. *Lab. Invest.* 52, 399–408.
- Torr, E. E., Ngam, C. R., Bernau, K., Tomasini-Johansson, B., Acton, B., and Sandbo, N. (2015). Myofibroblasts exhibit enhanced fibronectin assembly that is intrinsic to their contractile phenotype. *J. Biol. Chem.* 290 (11), 6951–6961. doi:10.1074/jbc.M114.606186
- Ulbricht, J., Jordan, R., and Luxenhofer, R. (2014). On the biodegradability of polyethylene glycol, polypeptides and poly(2-oxazoline)s. *Biomaterials* 35, 4848–4861. doi:10.1016/j.biomaterials.2014.02.029
- Ulrich, A. S. (2002). Biophysical aspects of using liposomes as delivery vehicles. *Biosci. Rep.* 22, 129–150. doi:10.1023/a:1020178304031
- Ungaro, F., d'Emmanuele di Villa Bianca, R., Giovino, C., Miro, A., Sorrentino, R., Quaglia, F., et al. (2009). Insulin-loaded PLGA/cyclodextrin large porous particles with improved aerosolization properties: *in vivo* deposition and

- hypoglycaemic activity after delivery to rat lungs. *J. Contr. Release* 135, 25–34. doi:10.1016/j.jconrel.2008.12.011
- Urbani, C. N., Bell, C. A., Lonsdale, D., Whittaker, M. R., and Monteiro, M. J. (2008). Self-Assembly of amphiphilic polymeric dendrimers synthesized with selective degradable linkages. *Macromolecules* 41, 76–86. doi:10.1021/ma701993w
- Vaidya, B., Kulkarni, N. S., Shukla, S. K., Parvathaneni, V., Chauhan, G., Damon, J. K., et al. (2020). Development of inhalable quinacrine loaded bovine serum albumin modified cationic nanoparticles: repurposing quinacrine for lung cancer therapeutics. *Int. J. Pharm.* 577, 118995. doi:10.1016/j.ijpharm.2019.118995
- Valadi, H., Ekström, K., Bossios, A., Sjöstrand, M., Lee, J. J., and Lötvall, J. O. (2007). Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 9, 654–659. doi:10.1038/ncb1596
- van der Meel, R., Fens, M. H. A. M., Vader, P., van Solinge, W. W., Eniola-Adefeso, O., and Schiffelers, R. M. (2014). Extracellular vesicles as drug delivery systems: lessons from the liposome field. *J. Contr. Release* 195, 72–85. doi:10.1016/j.jconrel.2014.07.049
- Veranth, J. M., Kaser, E. G., Veranth, M. M., Koch, M., and Yost, G. S. (2007). Cytokine responses of human lung cells (BEAS-2B) treated with micron-sized and nanoparticles of metal oxides compared to soil dusts. Part. *Fibre Toxicol.* 4, 1–18. doi:10.1186/1743-8977-4-2
- Verschraegen, C. F., Gilbert, B. E., Loyer, E., Huaranga, A., Walsh, G., Newman, R. A., et al. (2004). Clinical evaluation of the delivery and safety of aerosolized liposomal 9-nitro-20(S)-Camptothecin in patients with advanced pulmonary malignancies. *Clin. Canc. Res.* 10, 2319–2326. doi:10.1158/1078-0432.ccr-0929-3
- Vidgren, M., Waldrep, J. C., Arppe, J., Black, M., Rodarte, J. A., Cole, W., et al. (1995). A study of 99mtechnetium-labelled beclomethasone dipropionate dilaurylphosphatidylcholine liposome aerosol in normal volunteers. *Int. J. Pharm.* 115, 209–216. doi:10.1016/0378-5173(94)00265-7
- Wahab, R., Kaushik, N., Khan, F., Kaushik, N. K., Choi, E. H., Musarrat, J., et al. (2016). Self-styled ZnO nanostructures promotes the cancer cell damage and suppresses the epithelial phenotype of glioblastoma. *Sci. Rep.* 6, 19950. doi:10.1038/srep19950
- Waldrep, J. C., Gilbert, B. E., Knight, C. M., Black, M. B., Scherer, P. W., Knight, V., et al. (1997). Pulmonary delivery of beclomethasone liposome aerosol in volunteers. *Chest* 111, 316–323. doi:10.1378/chest.111.2.316
- Waldrep, J. C., Keyhani, K., Black, M., and Knight, V. (1994). Operating characteristics of 18 different continuous-flow jet nebulizers with beclomethasone dipropionate liposome aerosol. *Chest* 105, 106–110. doi:10.1378/chest.105.1.106
- Wang, D., Wang, T., Lu, J., Yu, H., Jiao, S., Feng, B., et al. (2016). Acid-activatable versatile micelleplexes for PD-L1 blockade-enhanced cancer photodynamic immunotherapy. *Nano Lett.* 16, 5503–5513. doi:10.1021/acs.nanolett.6b01994
- Wang, D., Yin, Y., Hu, C., Liu, X., Zhang, X., Zhou, S., et al. (2020). Clinical course and outcome of 107 patients infected with the novel coronavirus, SARS-CoV-2, discharged from two hospitals in Wuhan, China. *Crit. Care* 24 (1), 188. doi:10.1186/s13054-020-02895-6
- Wang, H., Wei, H., Wang, J., Li, L., Chen, A., and Li, Z. (2019). MicroRNA-181d-5p-containing exosomes derived from CAFs promote EMT by regulating CDX2/HOXA5 in breast cancer. *Mol. Ther. Nucleic Acids* 19, 654–667. doi:10.1016/j.omtn.2019.11.024
- Wang, J., Zhou, F., Li, Z., Mei, H., Wang, Y., Ma, H., et al. (2018). Pharmacological targeting of BET proteins attenuates radiation-induced lung fibrosis. *Sci. Rep.* 8 (1), 998. doi:10.1038/s41598-018-19343-9
- Wang, J. Q., Yan, F. Q., Wang, L. H., Yin, W. J., Chang, T. Y., Liu, J. P., et al. (2019). Identification of new hypoxia-regulated epithelial-mesenchymal transition marker genes labeled by H3K4 acetylation. *Genes, Chromosomes, and Cancer*, 59 (52), 73–83. doi:10.1002/gcc.22802
- Wang, L., Yang, G., Zhao, D., Wang, J., Bai, Y., Peng, Q., et al. (2019). CD103-positive CSC exosome promotes EMT of clear cell renal cell carcinoma: role of remote MiR-19b-3p. *Mol. Cancer* 18, 86. doi:10.1186/s12943-019-0997-z
- Wang, Y., Ding, L., Li, Z., Chen, G., Sun, M., and Oupicky, D. (2019). Treatment of acute lung injury and early- and late-stage pulmonary fibrosis with combination emulsion siRNA polyplexes. *J. Contr. Release* 314, 12–24. doi:10.1016/j.jconrel.2019.10.030
- Wang, Z., Wang, C., Liu, S., He, W., Wang, L., Gan, J., et al. (2017). Specifically formed corona on silica nanoparticles enhances transforming growth factor β 1 activity in triggering lung fibrosis. *ACS Nano*. 11, 1659–1672. doi:10.1021/acsnano.6b07461
- Wettstein, G., Bellaye, P. S., Kolb, M., Hammann, A., Crestani, B., Soler, P., et al. (2013). Inhibition of HSP27 blocks fibrosis development and EMT features by promoting Snail degradation. *Faseb. J. Am. Soc. Exp. Biol.* 27, 1549–1560. doi:10.1096/fj.12-220053
- Whyte, W. A., Orlando, D. A., Hniz, D., Abraham, B. J., Lin, C. Y., Kagey, M. H., et al. (2013). Master transcription factors and mediator establish super-enhancers at key cell identity genes. *Cell* 153 (2), 307–319. doi:10.1016/j.cell.2013.03.035
- Willis, B. C., and Borok, Z. (2007). TGF- β -induced EMT: mechanisms and implications for fibrotic lung disease. *Am. J. Physiol. Lung Cell Mol. Physiol.* 293, L525–L534. doi:10.1152/ajplung.00163.2007
- Wilson, M. S., and Wynn, T. A. (2009). Pulmonary fibrosis: pathogenesis, etiology and regulation. *Mucosal Immunol.* 2 (2), 103–121. doi:10.1038/mi.2008.85
- Winterbottom, C. J., Shah, R. J., Patterson, K. C., Kreider, M. E., Panettieri, R. A., Jr., Rivera-Lebron, B., et al. (2018). Exposure to ambient particulate matter is associated with accelerated functional decline in idiopathic pulmonary fibrosis. *Chest* 153 (5), 1221–1228. doi:10.1016/j.chest.2017.07.034
- Wittgren, B., Wahlund, K.-G., Dérand, H., and Wesslén, B. (1996). Size characterization of a charged amphiphilic copolymer in solutions of different salts and salt concentrations using flow field-flow fractionation. *Langmuir* 12, 5999–6005. doi:10.1021/la960435t
- Wu, Y., Ali, M. R. K., Dong, B., Han, T., Chen, K., Chen, J., et al. (2018). Gold nanorod photothermal therapy alters cell junctions and actin network in inhibiting cancer cell collective migration. *ACS Nano* 12, 9279–9290. doi:10.1021/acsnano.8b04128
- Xia, Y., Xiong, Y., Lim, B., and Skrabalak, S. E. (2008). Shape-controlled synthesis of metal nanocrystals: simple chemistry meets complex physics? *Angew. Chem. Int. Ed.* 48, 60–103. doi:10.1002/anie.200802248
- Xiao, K., He, W., Guan, W., Hou, F., Yan, P., Xu, J., et al. (2020). Mesenchymal stem cells reverse EMT process through blocking the activation of NF- κ B and Hedgehog pathways in LPS-induced acute lung injury. *Cell Death Dis.* 11, 863. doi:10.1038/s41419-020-03034-3
- Xiong, X., Arvizio, R. R., Saha, S., Robertson, D. J., McMeekin, S., Bhattacharya, R., et al. (2019). Sensitization of ovarian cancer cells to cisplatin by gold nanoparticles. *Cell Stress* 3, 267–279. doi:10.18632/oncotarget.2203
- Xu, X., Qiao, D., Mann, M., Garofalo, R. P., and Brasier, A. R. (2020). Respiratory syncytial virus infection induces chromatin remodeling to activate growth factor and extracellular matrix secretion pathways. *Viruses* 12 (8), 804. doi:10.3390/v12080804
- Yan, J., Ye, Z., Chen, M., Liu, Z., Xiao, Y., Zhang, Y., et al. (2011). Fine tuning micellar core-forming block of poly(ethylene glycol)-block-poly(ϵ -caprolactone) amphiphilic copolymers based on chemical modification for the solubilization and delivery of doxorubicin. *Biomacromolecules* 12, 2562–2572. doi:10.1021/bm200375x
- Yan, Y., Zhou, K., Xiong, H., Miller, J. B., Motea, E. A., Boothman, D. A., et al. (2017). Aerosol delivery of stabilized polyester-siRNA nanoparticles to silence gene expression in orthotopic lung tumors. *Biomaterials* 118, 84–93. doi:10.1016/j.biomaterials.2016.12.001
- Yang, G., Xu, L., Xu, J., Zhang, R., Song, G., Chao, Y., et al. (2018). Smart nanoreactors for pH-responsive tumor homing, mitochondria-targeting, and enhanced photodynamic-immunotherapy of cancer. *Nano Lett.* 18, 2475–2484. doi:10.1021/acs.nanolett.8b00040
- Yang, W., Liu, S., Bai, T., Keefe, A. J., Zhang, L., Ella-Menye, J.-R., et al. (2014). Poly(carboxybetaine) nanomaterials enable long circulation and prevent polymer-specific antibody production. *Nano Today* 9, 10–16. doi:10.1016/j.nantod.2014.02.004
- Yang, Y., Sunoqrot, S., Stowell, C., Ji, J., Lee, C.-W., Kim, J. W., et al. Effect of size, surface charge, and hydrophobicity of poly(amidoamine) dendrimers on their skin penetration. *Biomacromolecules* 13, 2154–2162. doi:10.1021/bm300545b
- Yao, Y., Chen, R., Wang, G., Zhang, Y., and Liu, F. (2019). Exosomes derived from mesenchymal stem cells reverse EMT via TGF- β 1/Smad pathway and promote repair of damaged endometrium. *Stem Cell Res. Ther.* 10, 1–17. doi:10.1186/s13287-019-1332-8

- Yoo, D., Guk, K., Kim, H., Khang, G., Wu, D., and Lee, D. (2013). Antioxidant polymeric nanoparticles as novel therapeutics for airway inflammatory diseases. *Int. J. Pharm.* 450, 87–94. doi:10.1016/j.ijpharm.2013.04.028
- Yoon, S., Kang, G., and Eom, G. (2019). HDAC inhibitors: therapeutic potential in fibrosis-associated human diseases. *IJMS*, 20, 1329. doi:10.3390/ijms20061329
- Zaru, M., Sinico, C., De Logu, A., Caddeo, C., Lai, F., Manca, M. L., et al. (2009). Rifampicin-loaded liposomes for the passive targeting to alveolar macrophages: in vitro and in vivo evaluation. *J. Liposome Res.* 19, 68–76. doi:10.1080/08982100802610835
- Zhang, D., Lee, H., Wang, X., Rai, A., Groot, M., and Jin, Y. (2018). Exosome-mediated small RNA delivery: a novel therapeutic approach for inflammatory lung responses. *Mol. Ther.* 26, 2119–2130. doi:10.1016/j.yjmt.2018.06.007
- Zhang, D., Lee, H., Zhu, Z., Minhas, J. K., and Jin, Y. (2017). Enrichment of selective miRNAs in exosomes and delivery of exosomal miRNAs *in vitro* and *in vivo*. *Am. J. Physiol. Lung Cell Mol. Physiol.* 312, L110–L121. doi:10.1152/ajplung.00423.2016
- Zhang, K., Rekhter, M. D., Gordon, D., and Phan, S. H. (1994). Myofibroblasts and their role in lung collagen gene expression during pulmonary fibrosis. A combined immunohistochemical and *in situ* hybridization study. *Am. J. Pathol.* 145(1), 114–125.
- Zhang, T., Sun, L. X., and Feng, R. E. (2020). Comparison of clinical and pathological features between severe acute respiratory syndrome and coronavirus disease 2019. *Zhonghua Jiehe He Huxi Zazhi* 43 (6), 496–502. doi:10.3760/cma.j.cn112147-20200311-00312
- Zhang, X., Bai, J., Yin, H., Long, L., Zheng, Z., Wang, Q., et al. (2020). Exosomal miR-125b-5p targets human telomerase reverse transcriptase in colorectal cancer cells to suppress epithelial-to-mesenchymal transition. *Mol. Oncol.* 14, 2589–2068. doi:10.1002/1878-0261.12765
- Zhang, X.-y., and Zhang, P.-y. (2017). Polymersomes in nanomedicine—a review. *Cnano* 13, 124–129. doi:10.2174/1573413712666161018144519
- Zhang, Y., Huang, Y., and Li, S. (2014). Polymeric micelles: nanocarriers for cancer-targeted drug delivery. *AAPS PharmSciTech* 15, 862–871. doi:10.1208/s12249-014-0113-z
- Zhao, X., Wu, X., Qian, M., Song, Y., Wu, D., and Zhang, W. (2018). Knockdown of TGF- β 1 expression in human umbilical cord mesenchymal stem cells reverts their exosome-mediated EMT promoting effect on lung cancer cells. *Canc. Lett.* 428, 34–44. doi:10.1016/j.canlet.2018.04.026
- Zhao, Y., Tian, B., Sun, H., Zhang, J., Zhang, Y., Ivannikov, M., et al. (2019). Pharmacoproteomics reveal novel protective activity of bromodomain containing 4 inhibitors on vascular homeostasis in TLR3-mediated airway remodeling. *Journal of Proteomics* 205, 103415. doi:10.1016/j.jpro.2019.103415
- Zhou, Y., Zhu, W. P., Cai, X. J., and Chen, M. (2016). Atomized paclitaxel liposome inhalation treatment of bleomycin-induced pulmonary fibrosis in rats. *Genet. Mol. Res.* 15, 1–11. doi:10.4238/gmr.15027309
- Zhu, J. (2015). T helper 2 (Th2) cell differentiation, type 2 innate lymphoid cell (ILC2) development and regulation of interleukin-4 (IL-4) and IL-13 production. *Cytokine*, 75 (71), 14–24. doi:10.1016/j.cyto.2015.05.010

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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