Check for updates

OPEN ACCESS

EDITED BY Rabia Miray Kisla Ekinci, Ministry of Health, Türkiye

REVIEWED BY Mehmet Yildiz, Istanbul University Cerrahpasa, Türkiye Neslihan Gokcen, Kocaeli University, Türkiye

*CORRESPONDENCE Yanliang Jin Image jinyanliang@scmc.com.cn Yixiao Bao Image baoyixiao@scmc.com.cn

RECEIVED 31 May 2024 ACCEPTED 21 October 2024 PUBLISHED 12 November 2024

CITATION

Huang H, Ding F, Liu C, Bao S, Jin Y, Jin Y and Bao Y (2024) Cytokine correlations in childhood-onset rheumatic diseases with pulmonary involvement. Front. Pediatr. 12:1441890. doi: 10.3389/fped.2024.1441890

COPYRIGHT

© 2024 Huang, Ding, Liu, Bao, Jin, Jin and Bao. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Cytokine correlations in childhood-onset rheumatic diseases with pulmonary involvement

Hua Huang¹, Fei Ding¹, Chenxi Liu¹, Shengfang Bao¹, Yingying Jin¹, Yanliang Jin^{1*} and Yixiao Bao^{2*}

¹Department of Rheumatology and Immunology, Shanghai Children's Medical Center, Shanghai Jiao Tong University School of Medicine, Shanghai, China, ²Shanghai Tonxin Clinic, Shanghai, China

Background: There was less research about cytokines in lung involvement of childhood-onset rheumatic diseases (RD) patients.

Objective: In this investigation, blood levels of 12 cytokines were tested in order to explore their associations with lung involvement in newly diagnosed childhood-onset RD patients.

Methods: High-resolution computed tomography (HRCT) and pulmonary function tests (PFTs) were performed on 53 newly diagnosed RD patients without any treatment in our department between July 2021 and December 2023. Patients were separated into groups based on whether their lungs were involved or not according to abnormalities found on HRCT or PFTs. We examined the serum cytokines with 41 healthy, age and gender-matched controls.

Results: The majority of serum cytokine levels were statistically different between the RD group with lung involvement and the control group. The RD group with lung involvement had considerably higher serum IL-6 and IFN- γ levels than the RD group without lung involvement or control group. The median serum IL-6 levels were 19 pg/ml [interquartile range (IQR) 6.9, 27.6], 4.4 pg/ml (IQR 2.0, 10.5), and 2.0 pg/ml (IQR 2.0, 2.0) in RD group with lung involvement, RD group without lung involvement, and control group, respectively. Similarly, the median serum IFN- γ levels were 6.0 pg/ml (IQR 5.0, 15), 5.0 pg/ml (IQR 5.0, 5.0), and 5.0 pg/ml (IQR 5.0, 5.0) in RD group with lung involvement, RD group without lung involvement, and control group, respectively. The receiver operating characteristic (ROC) curve study showed that IL-6 and IFN- γ had area under the curve (AUC) values of 0.782 and 0.742, respectively, indicating their significant diagnostic potential for lung involvement in RD patients.

Conclusion: Our findings imply that IL-6 and IFN- γ may be associated with the development of lung illnesses and may be involved in the pathophysiology of RD. Thus, in individuals with RD, elevated levels of IFN- γ over 5.42 pg/ml or IL-6 above 6.25 pg/ml may warrant suspicion of lung involvement.

KEYWORDS

rheumatic diseases, connective tissue disease, cytokines, lung, children

1 Introduction

Pediatric rheumatic diseases (RD) encompasses a variety of chronic autoimmune, autoinflammatory, and noninflammatory musculoskeletal conditions (1). No matter what the underlying conditions are, almost all rheumatic diseases could cause pulmonary symptoms, either as a result of the disease itself or as a side effect of pharmacological therapies (2). Pulmonary manifestations maybe present with RD or precede the diseases.

Interstitial lung disease (ILD) is a common manifestation of lung involvement in RD patients, particularly in illnesses such as sclerosis systemic (SSc), rheumatoid arthritis (RA). dermatomyositis (DM), systemic lupus erythematosis (SLE), Sjogren's syndrome (SS), and undifferentiated rheumatoid disorders (3). Up to half childhood-onset SSc patients could have lung involvement, such as ILD and pulmonary arterial hypertension (PAH) (4). Patients with systemic juvenile idiopathic arthritis (SJIA), particularly those with refractory disease histories and exposure to several medications, may develop potentially fatal pulmonary consequences such as pulmonary alveolar proteinosis (PAP), PAH, and ILD (5).

Although pulmonary problems are usually less common in children than in adults, they can lead to functional impairment and increased mortality (6). Clinical presentations might vary from being asymptomatic to having respiratory symptoms and exacerbations, which are more typical clinical presentations (7). The approaches employed for illness identification are relevant to the detection of pulmonary involvement. Determining dependable methods would support early intervention, especially in individuals who are asymptomatic.

The most reliable method for diagnosing RD-ILD is Highresolution computed tomography (HRCT). It is believed that the subclinical radiographic prevalence rate ranges from 33 to 57%, while the total incidence of ILD in connective tissue disease (CTD) is 15% (8). However, in young children, CT examinations pose problems with radiation exposure and anesthesia (9).

Pulmonary function tests (PFTs) is helpful in monitoring the degree and course of respiratory involvement and in identifying subclinical disease (10). But PFT need the cooperation of the patients.

For children, it would be ideal to identify noninvasive, trustworthy biomarkers that may be used to forecast the onset, prognosis, and reaction to therapy of disorders. According to some research, cytokines are crucial in the lung tissue damage, inflammation, and fibrosis that adult CTD-ILD patients experience (8, 11–14). However there was little research about cytokines in the pathogenesis of lung involvement in children with RD (5, 15, 16).

In order to investigate the relationship between alterations in cytokine levels and lung involvement in childhood patients with newly diagnosed RD, we measured the blood levels of 12 cytokines in both the RD children and the healthy controls in this investigation. We try to find some cytokines as potential diagnostics serum biomarkers.

2 Methods

2.1 Study population

This cross-sectional study was carried out between July 2021 and December 2023 at the Department of Rheumatology and Immunology, Shanghai Children's Medical Center, School of medicine, Shanghai Jiao Tong University.

Every participant provided written informed permission. The investigation was approved by the Research Ethics Committee of Shanghai Children's Medical Center, School of Medicine, Shanghai Jiao Tong University (SCMCIRB-W2021040).

Ninty-four participants were enrolled in this study. Fiftythree patients (eighteen patients with juvenile idiopathic arthritis [JIA], eleven with juvenile systemic lupus erythematosus [JSLE], nine with juvenile dermatomyositis [JDM], six with vasculitis (three with IgA vasculitis, one with Takayasu arteritis, one with Behçet's disease, one with ANCAassociated vasculitis), five with Sjogren's syndrome [SS], four with undifferentiated connective tissue disease [UCTD]) were newly diagnosed with RD (mainly including JIA, CTD, vasculitis) and were not on any treatment. All patients fulfilled the internationally accepted criteria of JIA, JSLE, JDM, SS, vasculities and UCTD and underwent evaluation using PFTs and HRCT. Based on the PFTs or HRCT abnormalities, the RD group was divided into RD with lung involvement and RD without lung involvement.

Forty-one healthy controls were recruited from individuals receiving examinations at the developmental and behavioral clinic of Shanghai Children's Medical Center. The controls matched the study group in age and sex distribution. Chronic lung, cardiac, or rheumatic illnesses were not seen in these controls. A medical checkup turned up nothing unusual.

Exclusion criteria encompassed individuals with respiratory symptoms and signs, active systemic diseases like malignancies, infections, metabolic diseases, and a history of chronic respiratory or cardiovascular conditions.

2.2 Laboratory analysis

Upon admission, we examined the serum concentrations of TNF-α, IFN-α, IFN-γ, and IL-1β, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12p70, IL-17. All patients and controls had venous blood samples (3 ml) drawn into standard blood collection tubes, which were then left to solidify for 30 min or more at room temperature. The samples were centrifuged for ten minutes at 1,000 rpm, and then they were kept at -20°C until analysis. Using a commercially available fluorescence flow (MAGPIX[®], assay kit USA), cytometry cvtokine concentrations were determined in accordance with the manufacturer's instructions.

2.3 Disease assessments

All patients underwent PFTs and pulmonary HRCT. PFT measurements were conducted in accordance with standards set by American Thoracic Society/European Respiratory Society (17).

At least three repetitions of each measurement were made, and the highest measurement that was found to be acceptable was compared to the expected normal values.

The definitions of pulmonary dysfunction were as follows: small airway disease (FEF75 or FEF75, and FEF50 < 65% of that predicted), isolated impairment of diffusion capacity (DLco < 80% of that predicted), obstructive dysfunction (FEV1/FVC < 80% of that predicted, and FEV1 < 80% of that predicted), and restrictive dysfunction (VC < 80% of that predicted).

HRCT was evaluated by two trained radiologists. The primary findings on HRCT were considered to be ground-glass opacity, consolidation, septal thickening, honeycombing, crazy paving, nodules, pleural and pericardial effusions, progressive volume loss, and pulmonary embolism (18).

2.4 Statistical analysis

Numerical variables were presented as mean and standard deviation (SD) or median and interquartile range [IQR; (Q1, Q3)]. Categorical variables were presented as the number of cases (percentage). Student's *t*-test for normally distributed values, Mann-Whitney *U*-test for nonnormally distributed data, Chi-square test or Fisher's exact test for categorical variables were uesed to compare patients and controls. To assess the relationships between cytokines and clinical indicators, Spearman correlation analysis was employed. To evaluate cytokines' sensitivity and specificity in predicting lung involvement in patients with RD, receiver operating characteristic (ROC) curve analysis was performed.

IBM SPSS 26.0 software (IBM, Armonk, New York, USA) was used for all statistical analyses. P < 0.05 (two-sided) was used as the statistical significance threshold.

3 Results

3.1 Demographic and clinical characteristics

The demographic and clinical characteristics are listed in Table 1.

Thirty-one female and twenty-two male RD patients made up the participants with a mean age of 10.3 ± 3.4 . Data from 41 healthy control participants (including 23 females and 18 males) with the mean age of 9.8 ± 1.4 were compared with theirs. Age, weight, height, BMI and sex ratio did not significantly differ between the patients and control groups.

Based on subgroup analysis, the RD group included 18 patients with JIA, eleven with SLE, nine with JDM, six with vasculitis (three with IgA vasculitis, one with Takayasu arteritis, one with Behçet's disease, one with ANCA-associated vasculitis), five with SS, and four with UCTD. The median span from the start of the illness to the diagnosis was 60 days [21, 180].

According to the abnormalities of HRCT or PFTs, the RD group was divided into with or without lung involvement. There were 22 patients (mean age 10.7 ± 4.0 years) in the RD group with lung involvement, 16 of whom were female and 6 of whom were male. Thirty days [14, 112] was the median disease duration. There were 31 patients (mean age 10.1 ± 3.0 years) in the RD group without lung involvement, 15 of whom were female and 16 of whom were male. The median disease duration was 60 days [30, 210].

Notably, all RD patients with lung involvement were asymptomatic.

3.2 Pulmonary function tests and HRCT findings in RD patients

HRCT and PFTs were conducted for all patients with RD. Among the 53 patients, nine (17.0%) exhibited abnormal HRCT, and 15 (28.3%) had abnormal PFTs. Overall, 22 patients (41.5%)

Variables	RD group with lung involvement (<i>n</i> = 22)	RD group without lung involvement (<i>n</i> = 31)	P _{ab} -value	RD (<i>n</i> = 53)	Controls (<i>n</i> = 41)	P _{cd} -value
Age (years)	10.7 ± 4.0	10.1 ± 3.0	0.2	10.3 ± 3.4	9.8 ± 1.4	0.0734
Sex (female/ male)	16 (73%)/6 (27%)	15 (48%)/16 (52%)	0.096	31 (58%)/22 (42%)	23 (56%)/18 (44%)	0.8363
Height (cm)	139.1 ± 11.9	136.0 ± 17.2	0.5763	137.2 ± 15.3	131.5 ± 12.8	0.0838
Weight (kg)	36.3 ± 11.6	36.5 ± 12.3	0.8606	36.5 ± 11.9	32.9 ± 7.3	0.1294
BMI	16.2 ± 2.2	17.6 ± 2.6	0.1161	17.1 ± 2.5	17.5 ± 2.5	0.4561
Duration (days)	30 [14, 112]	60 [30, 210]	0.2	60 [21, 180]	NS	
Illness			0.001*		NS	
SLE	10 (45.5%)	1 (3.2%)		11 (20.8%)		
JIA	4 (18.2%)	14 (45.2%)		18 (34.0%)		
JDM	4 (18.2%)	5 (16.1%)		9 (17.0%)		
Others	4 (18.2%)	11 (35.5%)		15 (28.3%)		

TABLE 1 Demographic and clinical characteristics.

BMI, body mass index; SLE, systemic lupus erythematosus; JIA, juvenile idiopathic arthritis; JDM, juvenile dermatomyositis; RD, rheumatic disease. *P<0.05 was considered statistically significant. demonstrated lung involvement. Of the nine patients with abnormal HRCT, 5 (55.6%) were female, while among the fifteen patients with abnormal PFTs, 13 (86.7%) were female. Two patients (3.8%) exhibited HRCT and PFT abnormalities, and both were females.

The three types of radiological abnormalities were pleural thickening (n = 1), linear opacities (n = 1), and ground-glass opacities (n = 7).

PFTs abnormalities included restrictive dysfunction (n = 4), reduction of DLCO (n = 9) and small airway disease (n = 2).

3.3 Concentrations of 12 cytokines in serum

The RD group with lung involvement's serum levels of IL-2, IL-4, IL-5 as well as those of IL-6, IL-8, IL-10, IL-17, IFN- γ were statistically different from the control group's (*P* < 0.05) (Table 2).

Furthermore, the RD group with lung involvement had substantially higher levels of serum IL-6 and IFN- γ than the RD group without lung involvement (*P* < 0.05) (Table 2).

Specifically, 19.0 pg/ml (IQR 6.9, 27.6) in the RD group with lung involvement, 4.4 pg/ml (IQR 2.0, 10.5) in the RD group without lung involvement and 2.0 pg/ml (IQR 2.0, 2.0) in the control group were the median serum IL-6 levels. The RD group with lung involvement had IL-6 levels that were considerably higher than the control group and RD group without lung involvement (P < 0.05).

6.0 pg/ml (IQR 5.0, 15) in the RD group with lung involvement, 5.0 pg/ml (IQR 5.0, 5.0) in the RD group without lung involvement and 5.0 pg/ml (IQR 5.0, 5.0) in the control group were the median serum IFN- γ levels. The RD group with lung involvement had substantially higher IFN- γ levels than both the control group and the RD group without lung involvement (P < 0.05).

These findings imply that the RD group with lung involvement is characterized by elevated blood levels of IL-6 and IFN- γ .

CARLE 2 Differences of 42 entrolines between encourse

3.4 Diagnostic utility of IL-6 and IFN- γ cytokines concentrations for RD-lung involvement patients

We assessed the diagnostic value of IL-6 and IFN- γ for RD-related lung involvement using ROC curves. AUC (area under the curve) for IL-6 was 0.782 (P = 0.005). With a sensitivity of 0.833 and specificity of 0.667, the ideal cut-off value was found to be 6.25 pg/ml. AUC for IFN- γ was 0.742 (P = 0.011), and 5.42 pg/ml was determined to be the ideal cut-off value. This threshold produced results with a 0.636 sensitivity and 0.889 specificity (Table 3).

3.5 Correlations between clinical variables and cytokines in RD group with lung involvement

We conducted a Spearman correlation analysis between IL-6, IFN- γ and clinical variables in RD group with lung involvement (Table 4). There was no discernible association between the parameters of PFT and the levels of IL-6 and IFN- γ (P > 0.05). The IL-6 level and CRP and ESR were shown to be significantly positively correlated. A noteworthy inverse relationship was discovered between IL-6, IFN- γ level and CD4, CD8 (P < 0.05). Intriguingly, there was a strong correlation (P < 0.001) between the level of IFN- γ and IL-6.

4 Discussion

This is the first study about cytokine profile in childhood-onset RD patients with lung involvement in China. Newly diagnosed and treatment naive could guarantee eliminating the influence of drugs.

In the study, We found The RD group with lung involvement had considerably greater levels of IL-6 and IFN- γ than the RD group without lung involvement. The measurement of IL-6 and

Cytokines	RD group with lung involvement (pg/ml) (<i>n</i> = 22)	RD group without lung involvement (pg/ml) (n = 31)	Controls (pg/ ml) (n = 41)	P _{ab} -value	P _{ac} -value	P _{bc} -value
IL-1β	3.0 [2.4, 3.3]	3.0 [2.4, 3.1]	3.1 [3.0, 3.7]	0.813	0.924	0.062
IL-2	4.0 [2.4, 4.3]	4.0 [2.4, 4.0]	4.0 [4.0, 4.0]	0.852	0.036*	0.705
IL-4	2.7 [2.4, 5.5]	3.0 [2.5, 16.9]	16.9 [10.3,33.2]	0.3	0.000*	0.000*
IL-5	3.0 [2.4, 4.0]	3.0 [2.4, 4.9]	6.5 [3.9, 19.5]	0.7	0.000*	0.000*
IL-6	19.0 [6.9, 27.6]	4.4 [2.0, 10.5]	2.0 [2.0, 2.0]	0.010*	0.000*	0.000*
IL-8	23.1 [7.9, 56.4]	22.0 [3.0, 39.5]	3.5 [3.3, 4.8]	0.5	0.000*	0.000*
IL-10	4.6 [2.7, 8.7]	4.5 [2.4, 5.6]	3.6 [3.1, 4.9]	0.5	0.021*	0.832
IL-12p70	2.4 [2.4, 4.6]	4.0 [2.4,8.8]	4.0 [4.0, 4.0]	0.3	0.638	0.169
IL-17	3.7 [2.4, 6.2]	5.0 [2.8, 5.0]	5.0 [5.0, 5.0]	0.782	0.007*	0.937
TNF-α	4.0 [2.4, 4.7]	4.0 [2.4, 4.0]	4.0 [4.0, 4.0]	0.11	0.647	0.384
IFN-α	5.0 [3.1, 16.5]	5.0 [2.4, 5.0]	5.0 [5.0, 11.5]	0.3	0.853	0.037*
IFN-γ	6.0 [5.0, 15]	5.0 [5.0, 5.0]	5.0 [5.0, 5.0]	0.009*	0.000*	0.229

TABLE 2 Differences of 12 cytokines between groups.

IL, interleukin; TNF, tumor necrosis factor; IFN, interferon; RD, rheumatic disease.

*P < 0.05 was considered statistically significant.

TABLE 3 Predictive performance of IL-6 and IFN- γ based on the ROC analysis.

Variables	Cut-off value (pg/ml)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	AUC	P value
IL-6	6.25	83.3	66.7	62.5	85.7	0.782	0.005*
IFN-γ	5.42	63.6	88.9	77.8	80.0	0.742	0.011*

AUC, area under the curve; PPV, positive predictive value; NPV, negative predictive value.

*P < 0.05 was considered statistically significant.

TABLE 4 Spearman correlation between IL-6, IFN- γ levels in RD-lung involvement group.

	Disease duration	CRP	ESR	LDH	CD_4	CD ₈	FVC%	DLco%	IFN-γ
IL-6	-0.21	0.31*	0.54***	0.23	-0.32*	-0.38**	0	-0.31	0.48***
IFN-γ	-0.17	-0.01	0.18	0.15	-0.36*	-0.33*	0.01	-0.09	NS

CRP, C-reactive protein; ESR, erythrocyte sedimentation rate; LDH, lactate dehydrogenase; FVC, forced vital capacity; DLCO, diffusing capacity of the lung for carbon monoxide; IL-6, interleukin-6; IFN-γ, interferon gamma.

*Indicated that statistically significant P < 0.05.

**P < 0.01.

***P < 0.001.

IFN- γ maybe a useful biomarker of lung involvement in RD patients.

Rheumatic disorders often involve activation of autoinflammatory and autoimmune processes, which can lead to pulmonary dysfunction (19). Even though children's symptoms might not be noticeable at first or not at all, they can lead to significant morbidity and mortality (2). While children generally exhibit fewer pulmonary manifestations than adults, PFT abnormalities have been reported in as many as 48% of a pediatric sample (20). Our current investigation revealed a high prevalence (41.5%) of pulmonary involvement in newly diagnosed RD cases, as assessed through functional or structural assessments. This finding aligns with our previous observations (21). Since the patients are asymptomatic, the prevalence of lung involvement varies depending on the detection method. Advanced screening modalities play a crucial role in early detection and intervention.

While the precise pathogenic mechanisms underlying lung involvement remain unclear, some research suggests that cytokines dysregulation—a crucial component of the immune system—plays a major role in the development of lung involvement (22).

Cytokines, active participants in an intricate network of connections, govern immunoregulatory processes and the inflammatory response. In adult-related research, cytokines have been identified as new biomarkers that might help focus screening efforts on people who are most likely to acquire a disease, advance it, and respond well to therapy (3).

Acute-phase pleiotropic inflammatory cytokine (IL-6) is generated by a variety of cells, including fibroblasts, monocytes, and lymphocytes. It contributes to the acute phase response, Tcell activation, and stimulation of megakaryocytic and myeloid progenitors in the hematopoietic system (23). IL-6 levels were reported to be slightly elevated in a rat model of experimental pulmonary fibrosis, which was linked to a proliferative response in fibroblasts from idiopathic pulmonary fibrosis (IPF) (24). Attenuated bleomycin-induced inflammatory cell buildup and subsequent fibrotic lung alterations were observed in IL-6deficient mice (25). Cui et al. identified IL-6 was the primary cause of progressive fibrosis in fibrotic lungs. They demonstrated that fibroblasts expressing *JUN* upregulate IL-6 production and release, which affects the innate and adaptive immune systems (26). Later on in the illness, IL-6 mediates the advancement of fibrosis by activating the IL-6/STAT3 and TGF- β signaling pathways in turn (27).

According to our current investigation, pediatric RD patients with lung involvement had considerably greater amounts of IL-6 in their sera than did those without and healthy controls. This suggests that the elevated level in sera closely corresponds with the emergence of abnormalities in the lung. Higher serum IL-6 levels were helpful in predicting lung involvement, according to ROC curve analysis. According to Lee et al., elevated serum IL-6 levels can independently predict acute exacerbation and serve as a prognostic factor for ILD patients' death (28). Nara et al's findings suggest that serum IL-6 levels may be a helpful prognostic indicator for clinically amyopathic dermatomyositis with rapidly progressive interstitial lung disease, influencing decisions on the intensity of early-phase immunosuppressive treatment (29).

We discovered that IL-6 cytokine was a statistically significant marker for lung involvement in RD patients, which is consistent with the literature.

According to De Lauretis et al., serum IL-6 was predictive of FVC and DL_{CO} declines within the first year in SSc patients, with a threshold of 7.7 pg/ml (30). But we did not find a relationship between IL-6 and PFT findings.

Additionally, compared to RD patients without lung involvement, our study showed a considerably greater level of IFN- γ in lung involvement RD patients.

Type 1 CD4 and CD8 T cells, as well as natural killer (NK) cells, produce the main effector cytokine, IFN- γ (31). Reports on the clinical efficacy of IFN- γ in lung involvement have been contradictory. IFN- γ has been shown by Ishikawa et al. to have a major role in the pathogenesis of dermatomyositis (DM), particularly in the development of pulmonary lesions that are seen in rapidly progressing interstitial lung disease (RP-ILD) (32). By stimulating macrophages and quickening inflammation,

10.3389/fped.2024.1441890

IFN- γ seems to have a pathogenic effect on DM RP-ILD. In Iriguchi S et al's study, they found mice with T-bet overexpression in T lymphocytes, driving IFN- γ production resulting in erythrophagocytosis, as well as alveolar macrophage dysfunction, leading to PAP-like lung disease (33).

Furthermore, IFN- γ can accelerate the development of pulmonary fibrosis via controlling cell apoptosis (34). Prior to Fas activation, exposure to IFN- γ markedly boosted the activity of caspase 3, 7, and 8, as well as the processing of CK-18, a crucial cytoskeletal protein in human lung epithelial cells, by caspase. Additionally, it raised the appearance of apoptotic nuclei (35).

However, the roles of IFN- γ in lung maybe a double-edged sword. According to certain studies, TGF-beta-induced extracellular matrix (ECM) synthesis by myofibroblasts is negatively regulated by IFN- γ , exhibiting anti-fibrotic action (36). IFN- γ prevents fibroblast development into myofibroblasts by opposing TGF- β 1 signaling, in part by upregulating SMAD7 and antagonizing SMAD and the JAK pathway at the nuclear p300/ CBP level (37). The functions and mechanisms of IFN- γ in lung injury associated with RD are yet to be clarified and need more extensive clinical trials.

We discovered a strong inverse relationship between IL-6, IFN- γ , and CD4, CD8 in our investigation. It's interesting to note that IL-6 and IFN- γ were shown to positively correlate in this investigation. According to a research by Huang et al., blood lymphopenia in anti-MDA5 + patients is probably caused by lymphocytes being transferred to the lungs to take part in the local immune response, which lowers the amount of free lymphocytes in the blood (38). We speculate that the combined upregulation of IL-6 and IFN- γ facilitates lymphocyte egress from the circulatory system into the lungs and thus participates in the process of lung pathology. Further studies are necessary to validate these results.

Finally, it is recommended that patients with recently diagnosed RD who have elevated levels of serum IL-6 or IFN- γ pay great attention to any modest pulmonary involvement. Pulmonary specialists can provide guidance on early detection techniques for impaired pulmonary function.

The study has some limitations. First, a somewhat small population from a single center participated in this retrospective study. Second, the study included patients with different subtypes of diseases. Finally, our study did not include patients who were unable to complete the PFT test, introducing a potential selection bias. Thus, a longer-term study with a more extensive RD population might be beneficial for future investigations. To our knowledge, however, this is the first study examining the relationships between cytokines and pulmonary involvement in Chinese children who have recently been diagnosed with rheumatic disorders but have not yet received therapy.

5 Conclusion

A vital component of the immune system, cytokine dysregulation greatly influences the etiology of illness. This study shows that among RD patients with lung involvement, serum levels of IFN- γ and IL-6 were higher. Patients with childhood RD may be at risk for lung illness if they have elevated serum levels of the cytokines IL-6 and IFN- γ .

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving humans were approved by the Research Ethics Committee of Shanghai Children's Medical Center, School of Medicine, Shanghai Jiao Tong University. The studies were conducted in accordance with the local legislation and institutional requirements. The participants provided their written informed consent to participate in this study. Written informed consent was obtained from the minor(s)' legal guardian/next of kin for the publication of any potentially identifiable images or data included in this article.

Author contributions

HH: Writing – review & editing, Writing – original draft. FD: Writing – review & editing. CL: Writing – review & editing. SB: Writing – review & editing. YJ: Writing – review & editing. YaJ: Supervision, Writing – review & editing. YB: Supervision, Funding acquisition, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This study was supported by a grant from National Natural Science Foundation of China [81870024].

Acknowledgments

We would like to thank Shanghai Children's Medical Center for providing clinical data and would like to thank Yabin Hu for helping us doing the statistical analysis.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

References

1. Ogbu EA, Brunner HI. Treatment guidelines in pediatric rheumatic diseases. Rheum Dis Clin North Am. (2022) 48(3):725-46. doi: 10.1016/j.rdc.2022.03.007

2. Buckley MM, Rabinovich CE. Pulmonary manifestations of rheumatic diseases in children. *Pediatr Clin North Am.* (2021) 68(1):147–66. doi: 10.1016/j.pcl.2020.09.005

3. Doyle TJ, Dellaripa PF. Lung manifestations in the rheumatic diseases. Chest. (2017) 152(6):1283-95. doi: 10.1016/j.chest.2017.05.015

4. Roofeh D, Brown KK, Kazerooni EA, Tashkin D, Assassi S, Martinez F, et al. Systemic sclerosis associated interstitial lung disease: a conceptual framework for subclinical, clinical and progressive disease. *Rheumatology*. (2023) 62(5):1877–86. doi: 10.1093/rheumatology/keac557

5. Schulert GS, Yasin S, Carey B, Chalk C, Do T, Schapiro AH, et al. Systemic juvenile idiopathic arthritis-associated lung disease: characterization and risk factors. *Arthritis Rheumatol.* (2019) 71(11):1943–54. doi: 10.1002/art.41073

6. Richardson AE, Warrier K, Vyas H. Respiratory complications of the rheumatological diseases in childhood. *Arch Dis Child*. (2016) 101(8):752–8. doi: 10. 1136/archdischild-2014-306049

7. Clement A, de Blic J, Epaud R, Galeron L, Nathan N, Hadchouel A, et al. Management of children with interstitial lung diseases: the difficult issue of acute exacerbations. *Eur Respir J*. (2016) 48(6):1559-63. doi: 10.1183/13993003.01900-2016

8. Tezcan D, Sivrikaya A, Ergün D, Özer H, Eryavuz Onmaz D, Körez MK, et al. Evaluation of serum interleukin-6 (IL-6), IL-13, and IL-17 levels and computed tomography finding in interstitial lung disease associated with connective tissue disease patients. *Clin Rheumatol.* (2021) 40(11):4713–24. doi: 10.1007/s10067-021-05773-w

9. Ramphul M, Gallagher K, Warrier K, Jagani S, Bhatt JM. Why is a paediatric respiratory specialist integral to the paediatric rheumatology clinic? *Breathe.* (2020) 16(4):200212. doi: 10.1183/20734735.0212-2020

10. Cidon M, Bansal M, Hartl D. Pulmonary manifestations of rheumatologic diseases. *Curr Opin Pediatr.* (2017) 29(3):311–9. doi: 10.1097/MOP.00000000000496

11. Zhang J, Wang D, Wang L, Wang S, Roden AC, Zhao H, et al. Profibrotic effect of IL-17A and elevated IL-17RA in idiopathic pulmonary fibrosis and heumatoid arthritis-associated lung disease support a direct role for IL-17A/IL-17RA in human fibrotic interstitial lung disease. *Am J Physiol Lung Cell Mol Physiol.* (2019) 316(3): L487–97. doi: 10.1152/ajplung.00301.2018

12. Zhang Q, Guo L, Song X, Lv C, Tang P, Li Y, et al. Serum IL-36 cytokines levels in idiopathic pulmonary fibrosis and connective tissue disease-associated interstitial lung diseases. *Clin Chim Acta*. (2022) 530:8–12. doi: 10.1016/j.cca.2022.02.015

13. Kawasumi H, Gono T, Kawaguchi Y, Kaneko H, Katsumata Y, Hanaoka M, et al. IL-6, IL-8, and IL-10 are associated with hyperferritinemia in rapidly progressive interstitial lung disease with polymyositis/dermatomyositis. *Biomed Res Int.* (2014) 2014:815245. doi: 10.1155/2014/815245

14. Jiang S, Wang Z, Ouyang H, Liu Z, Li L, Shi Y. Aberrant expression of cytokine interleukin 9 along with interleukin 4 and interferon γ in connective tissue disease-associated interstitial lung disease: association with severity of pulmonary fibrosis. *Arch Med Sci.* (2016) 12(1):101–6. doi: 10.5114/aoms.2015.47877

15. Kobayashi N, Takezaki S, Kobayashi I, Iwata N, Mori M, Nagai K, et al. Clinical and laboratory features of fatal rapidly progressive interstitial lung disease associated with juvenile dermatomyositis. *Rheumatology*. (2015) 54(5):784–91. doi: 10.1093/ rheumatology/keu385

16. Marstein H, Schwartz T, Aaløkken TM, Lund MB, Flatø B, Sjaastad I, et al. Novel associations between cytokines and pulmonary involvement in juvenile dermatomyositis—a cross-sectional study of long-term disease. *Rheumatology*. (2020) 59(8):1862–70. doi: 10.1093/rheumatology/kez531

17. Beydon N, Davis SD, Lombardi E, Allen JL, Arets HG, Aurora P, et al. An official American thoracic society/European respiratory society statement: pulmonary function testing in preschool children. *Am J Respir Crit Care Med.* (2007) 175 (12):1304–45. doi: 10.1164/rccm.200605-642ST

 García-Peña P, Boixadera H, Barber I, Toran N, Lucaya J, Enríquez G. Thoracic findings of systemic diseases at high-resolution CT in children. *Radiographics*. (2011) 31(2):465–82. doi: 10.1148/rg.312095160

19. Hildebrandt J, Rahn A, Kessler A, Speth F, Fischer DC, Ballmann M. Lung clearance index and diffusion capacity for CO to detect early functional pulmonary impairment in children with rheumatic diseases. *Pediatr Rheumatol Online J.* (2021) 19(1):23. doi: 10.1186/s12969-021-00509-1

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

20. Veiga CS, Coutinho DS, Nakaie CM, Campos LM, Suzuki L, Cunha MT, et al. Subclinical pulmonary abnormalities in childhood-onset systemic lupus erythematosus patients. *Lupus.* (2016) 25(6):645–51. doi: 10.1177/0961203316629554

21. Huang H, Hu Y, Wu Y, Ding F, Xu X, Jin Y, et al. Lung involvement in children with newly diagnosed rheumatic diseases: characteristics and associations. *Pediatr Rheumatol Online J.* (2022) 20(1):71. doi: 10.1186/s12969-022-00731-5

22. Spagnolo P, Distler O, Ryerson CJ, Tzouvelekis A, Lee JS, Bonella F, et al. Mechanisms of progressive fibrosis in connective tissue disease (CTD)-associated interstitial lung diseases (ILDs). *Ann Rheum Dis.* (2021) 80(2):143–50. doi: 10.1136/annrheumdis-2020-217230

23. Bonhomme O, André B, Gester F, de Seny D, Moermans C, Struman I, et al. Biomarkers in systemic sclerosis-associated interstitial lung disease: review of the literature. *Rheumatology*. (2019) 58(9):1534–46. doi: 10.1093/rheumatology/kez230

24. Phan THG, Paliogiannis P, Nasrallah GK, Giordo R, Eid AH, Fois AG, et al. Emerging cellular and molecular determinants of idiopathic pulmonary fibrosis. *Cell Mol Life Sci.* (2021) 78(5):2031–57. doi: 10.1007/s00018-020-03693-7

25. Saito F, Tasaka S, Inoue K, Miyamoto K, Nakano Y, Ogawa Y, et al. Role of interleukin-6 in bleomycin-induced lung inflammatory changes in mice. *Am J Respir Cell Mol Biol.* (2008) 38(5):566–71. doi: 10.1165/rcmb.2007-0299OC

26. Cui L, Chen SY, Lerbs T, Lee JW, Domizi P, Gordon S, et al. Activation of JUN in fibroblasts promotes pro-fibrotic programme and modulates protective immunity. *Nat Commun.* (2020) 11(1):2795. doi: 10.1038/s41467-020-16466-4

27. Shochet G E, Brook E, Bardenstein-Wald B, Shitrit D. TGF- β pathway activation by idiopathic pulmonary fibrosis (IPF) fibroblast derived soluble factors is mediated by IL-6 trans-signaling. *Respir Res.* (2020) 21(1):56. doi: 10.1186/s12931-020-1319-0

28. Lee JH, Jang JH, Park JH, Jang HJ, Park CS, Lee S, et al. The role of interleukin-6 as a prognostic biomarker for predicting acute exacerbation in interstitial lung diseases. *PLoS One.* (2021) 16(7):e0255365. doi: 10.1371/journal.pone.0255365

29. Nara M, Komatsuda A, Omokawa A, Togashi M, Okuyama S, Sawada K, et al. Serum interleukin 6 levels as a useful prognostic predictor of clinically amyopathic dermatomyositis with rapidly progressive interstitial lung disease. *Mod Rheumatol.* (2014) 24(4):633–6. doi: 10.3109/14397595.2013.844390

30. De Lauretis A, Sestini P, Pantelidis P, Hoyles R, Hansell DM, Goh NS, et al. Serum interleukin 6 is predictive of early functional decline and mortality in interstitial lung disease associated with systemic sclerosis. *J Rheumatol.* (2013) 40 (4):435–46. doi: 10.3899/jrheum.120725

31. Chen ES, Greenlee BM, Wills-Karp M, Moller DR. Attenuation of lung inflammation and fibrosis in interferon-gamma-deficient mice after intratracheal bleomycin. *Am J Respir Cell Mol Biol.* (2001) 24(5):545–55. doi: 10.1165/ajrcmb.24. 5.4064

32. Ishikawa Y, Iwata S, Hanami K, Nawata A, Zhang M, Yamagata K, et al. Relevance of interferon-gamma in pathogenesis of life-threatening rapidly progressive interstitial lung disease in patients with dermatomyositis. *Arthritis Res Ther.* (2018) 20(1):240. doi: 10.1186/s13075-018-1737-2

33. Iriguchi S, Kikuchi N, Kaneko S, Noguchi E, Morishima Y, Matsuyama M, et al. T-cell-restricted T-bet overexpression induces aberrant hematopoiesis of myeloid cells and impairs function of macrophages in the lung. *Blood.* (2015) 125(2):370–82. doi: 10.1182/blood-2014-05-575225

34. Gui X, Qiu X, Tian Y, Xie M, Li H, Gao Y, et al. Prognostic value of IFN-γ, sCD163, CCL2 and CXCL10 involved in acute exacerbation of idiopathic pulmonary fibrosis. *Int Immunopharmacol.* (2019) 70:208–15. doi: 10.1016/j.intimp.2019.02.039

35. Coulter KR, Doseff A, Sweeney P, Wang Y, Marsh CB, Wewers MD, et al. Opposing effect by cytokines on Fas-mediated apoptosis in A549 lung epithelial cells. *Am J Respir Cell Mol Biol.* (2002) 26(1):58–66. doi: 10.1165/ajrcmb.26.1.4285

36. Ghosh AK, Yuan W, Mori Y, Chen Sj, Varga J. Antagonistic regulation of type I collagen gene expression by interferon-gamma and transforming growth factor-beta. Integration at the level of p300/CBP transcriptional coactivators. *J Biol Chem.* (2001) 276(14):11041–8. doi: 10.1074/jbc.M004709200

37. Vu TN, Chen X, Foda HD, Smaldone GC, Hasaneen NA. Interferon- γ enhances the antifibrotic effects of pirfenidone by attenuating IPF lung fibroblast activation and differentiation. *Respir Res.* (2019) 20(1):206. doi: 10.1186/s12931-019-1171-2

38. Huang W, Ren F, Luo L, Zhou J, Huang D, Pan Z, et al. The characteristics of lymphocytes in patients positive for anti-MDA5 antibodies in interstitial lung disease. *Rheumatology*. (2020) 59(12):3886–91. doi: 10.1093/rheumatology/keaa266