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*CORRESPONDENCE Piero Pavone ☑ ppavone@unict.it Martino Ruggieri ☑ m.ruggieri@unict.it

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Case report: Structural brain abnormalities in *TUBA1A*tubulinopathies: a narrative review

Piero Pavone^{1,2*}, Pasquale Striano^{3,4}, Giovanni Cacciaguerra¹, Simona Domenica Marino⁵, Enrico Parano², Xena Giada Pappalardo², Raffaele Falsaperla⁵ and Martino Ruggieri^{1*}

¹Section of Pediatrics and Child Neuropsychiatry, Department of Child and Experimental Medicine, University of Catania, Catania, Italy, ²National Council of Research, Institute for Biomedical Research and Innovation (IRIB), Unit of Catania, Catania, Italy, ³Department of Neurosciences, Rehabilitation, Ophthalmology, Genetics, Maternal and Child Health, University of Genoa, Genova, Italy, ⁴Pediatric Neurology and Muscular Diseases Unit, IRCCS Istituto "G. Gaslini", Genova, Italy, ⁵Pediatrics and Pediatric Emergency Department, University Hospital, A.U.O "Policlinico-Vittorio Emanuele", Catania, Italy

Introduction: Tubulin genes have been related to severe neurological complications and the term "tubulinopathy" now refers to a heterogeneous group of disorders involving an extensive family of tubulin genes with *TUBA1A* being the most common. A review was carried out on the complex and severe brain abnormalities associated with this genetic anomaly.

Methods: A literature review of the cases of *TUBA1A*-tubulopathy was performed to investigate the molecular findings linked with cerebral anomalies and to describe the clinical and neuroradiological features related to this genetic disorder. **Results:** Clinical manifestations of *TUBA1A*-tubulinopathy patients are heterogeneous and severe ranging from craniofacial dysmorphism, notable developmental delay, and intellectual delay to early-onset seizures, neuroradiologically associated with complex abnormalities. *TUBA1A*-tubulinopathy may display various and complex cortical and subcortical malformations.

Discussion: A range of clinical manifestations related to different cerebral structures involved may be observed in patients with *TUBA1A*-tubulinopathy. Genotype–phenotype correlations are discussed here. Individuals with cortical and subcortical anomalies should be screened also for pathogenic variants in *TUBA1A*.

KEYWORDS

TUBA1A-tubulinopathy, cerebral anomalies, Dandy–Walker Malformation, DWM phenotype, tubulinopathies

Introduction

The term "tubulinopathy" indicates a heterogeneous group of disorders involving tubulin genes and presenting with complex cerebral malformations and severe neurological manifestations (1–7). Tubulin genes are an extensive family of genes composed of alpha, beta, gamma, delta, and epsilon. The tubulin alpha includes about 15 genes and the tubulin beta about 23 genes, respectively. Tubulin alpha and tubulin beta encode tubulin proteins that form heterodimers that are fundamental components of microtubules. Microtubules play an important role in brain developmental processes

Abbreviations ACC, agenesis of corpus callosum; CBLH, cerebellar hypoplasia; DWM, Dandy–Walker malformation; LIS, lissencephaly; LCH, LIS and cerebellar hypoplasia; PMG, polymicrogyria; SBH, subcortical band heterotopia.

including mitosis, neuronal migration, synaptic connectivity, and axonal transport (3-9). Dysfunction of microtubule-dependent functions from variants in components alpha, beta, and gamma cause severe cerebral malformations and neurological clinical involvement (10). The tubulin genes mainly involved in cerebral malformations are TUBA1A (MIM#602529), TUBA8 (MIM#605742), TUBB2A (MIM#615101), TUBB2B (MIM#612850), TUBB3 (MIM#602661), TUBB5 (MIM#191130), and TUBG1 (MIM#191135) (3-5). The different tubulin isotypes have sequence homology with diverse spatiotemporal expressions prompting a unique, isotype-specific function (10, 11). Among the patients affected by tubulinopathies, the most common mutation is represented by TUBA1A, coding the alpha-1 tubulin that is selectively and mainly expressed in post-mitotic neurons. Keays et al. (1) first reported mutations in alpha-tubulin as a cause of abnormal neuronal migration in mice and of lissencephaly (LIS) in humans. TUBA1A (alpha-tubulin complex) gene is located in chromosome 12q12-q14 and is composed of the N-terminal, intermediate, and C-terminal domains (12, 13). According to Hebebrand et al. (5), the variants in the affected individuals are shown to be distributed in clustering mainly around the Arg 402 residue in exon 4 in the C-terminal domain. Alpha-tubulinopathy manifests with a more severe phenotype than beta-tubulinopathy and accounts for 4%-5% of all the reported cases of lissencephaly (2, 4, 5). The clinical expression of patients affected by TUBA1A-tubulinopathy is various and complex such as craniofacial dysmorphism, severe developmental delay, cerebral palsy, and early-onset epilepsy. Cortical malformations such as lissencephaly, polymicrogyria (PMG), cortical gyral simplification, and gray matter heterotopias are frequently reported as well as anomalies of the corpus callosum, cerebellar vermis, cerebellum, and basal ganglia. Ventricular dilatation, anomalies of brainstem, abnormal hippocampus, and internal capsule are also reported (7, 13-19).

We reviewed the reported cases of *TUBA1A*-tubulinopathy underlying the frequent and severe neurological involvement associated with this genetic abnormality.

Material and methods

CARE checklist and review search strategy

This study was conducted according to the CARE guidelines (www.care-statement.org/checklist). A literature review related to the area of focus was conducted by collecting clinical trials, primary research, and reviews from online bibliographic databases (MEDLINE, Embase, PubMed, Cochrane Central, and Scopus) between January 2007 and September 2022. The key search derived from the medical subject heading terms were the following: Tubulinopathies, or *TUBA1A*-related encephalopathies; or *TUBA1A*-related disorders; or *TUBA1A* and Cerebral Malformation, or *TUBA1A* and cerebellar malformations. After examining 58 articles related to this topic and removing duplicate records, 32 studies were included,

Results

The 32 selected articles on TUBAIA-tubulinopathies were as follows: 12 were related to genetic results, 13 to systemic clinical manifestations, and seven were case reports. Pathogenic variants in *TUBA1A* were associated with cortical and subcortical malformations and with severe neurological complications. The mechanisms of *TUBA1A* mutations in causing these severe cerebral anomalies remain unclear.

Discussion

Pathogenetic variants of TUBAIA, as observed by Fallet-Bianco et al. (15), affect several cerebral cortical and subcortical structures with various and severe clinical manifestations. Clinical features related to subcortical and cerebral TUBAIA-tubulinopathy are reported.

TUBA1A-tubulinopathy and subcortical malformations

In a large-exome sequencing study in 282 individuals from 100 families with Dandy-Walker malformation (DWM) and cerebellar hypoplasia (CBLH), Aldinger et al. (19) obtained a molecular diagnosis in 36 families with a significantly higher number for CBLH (51%) than for DWM (16%). Among the 41 variants, only 27 neurodevelopmental disorder-associated genes were found, thus showing that CBLH and DWM are more often features of monogenic neurodevelopmental disorders. The authors (19) reported a newborn with DWM and TUBA1A mutation. Brain MRI showed small cerebellar hemispheres, an unpaired caudal lobule ("the Dandy-Walker tail"), and a single periventricular nodular heterotopia. Hebebrand et al. (5) identified three patients with heterozygous de novo missense variants in TUBA1A and reviewed the cases reported in the literature. About 166 patients were enrolled with 146 born and 20 fetuses and 107 cases showing available clinical information. The most commonly reported features were developmental delay (98%), anomalies of the corpus callosum (96%), microcephaly (76%), and lissencephaly (agyria-pachygyria) (70%). The authors (5) identified a total of 121 specific variants, including 15 recurrent and three new cases of DWM with a detailed clinical history and presentation of the associated dysmorphic features (cases description is reported in the **Supplementary material**).

TUBA1A-tubulinopathy and related cerebral malformations

Keays et al. (1) first reported two patients with *TUBA3* mutations (the human homolog of *TUBA1A*) both presenting with lissencephaly, pachygyria, and hippocampus anomalies. Among 95 sporadic patients with non-syndromic bilateral PMG,

including 54 patients with perisylvian PMG and 30 with PMG and additional brain abnormalities, Poirier et al. (9) identified three unrelated patients with mutations in TUBA1A representing 3.1% of the PMG group and 10% of PMGs with complex cerebral malformations. The last group included patients with bilateral perisylvian asymmetrical shapes, PMG with dysmorphic basal ganglia, cerebellar vermian dysplasia, and pontine hypoplasia. The authors (9) suggested that in addition to PMG, additional brain abnormalities such as dysmorphic basal ganglia, hypoplastic pons, and cerebellar dysplasia are relevant features for PMG TUBA1A-related diagnosis. The authors (9) concluded that patients with TUBA1A mutations share not only cortical dysgenesis but also cerebellar, hippocampal, corpus callosum, and brainstem abnormalities. Four fetuses with TUBA1A mutations and a prenatal diagnosis of major cerebral malformations with termination of pregnancy were reported by Fallet-Bianco et al. (15). The pathologic studies of the fetuses at 23, 25, 26, and 35 gestational weeks showed a spectrum of abnormalities, which encompassed five brain structures: the neocortex, hippocampus, corpus callosum, cerebellum, and brainstem. Other abnormalities involved the basal ganglia, olfactory bulbs, and germinal zones. Abnormal cortical and hippocampal lamination and heterotopic neurons in the cortex, cerebellum, and brainstem were found during the microscopic examination (15). Six patients with TUBA1A mutations were reported by Bahi-Buisson et al. (4). The authors showed that the TUBA1A-related lissencephaly spectrum ranges from perisylvian pachygyria in the less severe form to posteriorly predominant pachygyria in the most severe, associated with dysgenesis of the anterior limb of the internal capsule and mild to severe cerebellar hypoplasia. Mutation analysis in the TUBA1A gene was performed by Morris-Rosendahl et al. (20) in 46 patients with classical lissencephaly. They identified three novel pathogenic variants and one recurrent mutation in five patients with variable patterns of lissencephaly on the brain MRI. Congenital microcephaly was found in four of the five patients, and all showed dysgenesis of the corpus callosum, cerebellar hypoplasia, and variable cortical malformations, including subtle cortical band heterotopia and absence or hypoplasia of the anterior limb of the internal capsule. The authors maintained that the frequency of mutation in TUBA1A affects approximately 4% of the patients with lissencephaly. Kumar et al. (8) conducted a study of TUBA1A mutations in 125 patients with cortical dysgenesis including 72 patients with classical LIS, 22 with subcortical band heterotopia (SBH), 29 with LIS and cerebellar hypoplasia (LCH), and two with LIS and agenesis of corpus callosum (ACC). The results obtained consisted of missense mutations in five out of 72 patients with classic LIS (7%) and 10 missense mutations in 29 patients with LCH (32%). TUBA1A mutation was also found in one child with ACC and CBLH without LIS. The authors (8) summarized the brain MRI features of 17 patients with TUBA1A mutation in five distinct groups according to the observed variants. Three patients with TUBA1A mutations were reported by Jansen et al. (21) with classic LIS in a boy and polymicrogyria in two sisters whose mother showed somatic mosaicism, thus showing familial recurrence of the mutation. A single case in a

14-month-old girl with TUBA1A mutation and classical lissencephaly was reported by Sohal et al. (14). Marked ventricular dilatation with thin cortex, poorly differentiated basal ganglia, agenesis of corpus callosum, and cerebellar hypoplasia with preserved vermis were described by Okumura et al. (22) in a child with TUBA1A mutation and cerebellar hypoplasia. An extensive neuropathologic analysis showed a lacking lamination in the cerebral cortex, absent corpus callosum without Probst bundle, blurred demarcation among the striatum internal capsule and globus pallidus, irregular running of myelinated fibers, cerebellar hypoplasia with irregular undulation in the dentate nucleus and inferior olivary nucleus, absent olfactory bulbs and tracts, and pyramidal tract hypoplasia. Among 106 patients selected as having complex cortical malformations, Bahi-Buisson et al. (23) identified 45 (42.5%) patients presenting with mutations in TUBA1A, 18 in TUBB2B, (16.9%), 11 in TUBB3 (10.4%), three in TUBB5 (2.8%), and three in TUBG1 (2.8%). Five cortical malformation syndromes were distinguished by the authors (23) in a systemic revision of the data collected by neuroimaging and neuropathologic analysis in patients affected by TUBA1A and other tubulinopathies: microlissence phaly (n =12). lissencephaly (n = 19),central pachygyria and polymicrogyria-like cortical dysplasia (n = 24), generalized polymicrogyria-like cortical dysplasia (n = 6), and simplified gyral pattern with area of focal polymicrogyria (n = 19).

Dysmorphic basal ganglia were reported in 75% of cases and central pachygyria, polymicrogyria-like cortical dysplasia, and simplified gyral malformations syndrome in 100% of the cases. Moreover, the authors (23) highlight a high prevalence of corpus callosum agenesis (32/80; 40%) and mild to severe cerebellar hypoplasia and dysplasia (63/80; 78.7%). Fallet-Bianco et al. (24) selected 19 cases of TUBA1A mutations in a cohort of 60 fetal cases. The authors refer that all the cases of lissencephaly with cerebellar hypoplasia showed distinct TUBA1A mutations, while those with classical lissencephaly harbored recurrent mutations in TUBA1A (three cases) or TUBB2B (one case). The authors (24) concluded that fetal TUBA1A-tubulinopathies most often present with microlissencephaly or classical lissencephaly with corpus callosum agenesis, but polymicrogyria may also occur in contrast to TUBB2B mutations in which either polymicrogyria or microlissencephaly may be found. TUBA1A mutations affect almost exclusively the central nervous system (CNS), but congenital malformations may involve other body organs as reported by Hikita et al. (25) in a 6-year-old girl who presented with lissencephaly, microcephaly, and early-onset epileptic seizures in addition to Hirschsprung disease and inappropriate antidiuretic hormone secretion (SIADH). In a cohort of 156 patients with malformations of cortical development of unknown origin, 79 patients were selected and submitted to a genetic analysis of the TUBA1A, TUBB2B, and TUBB3 (26). Two novel heterozygous mutations were found: a TUBA1A mutation in a 5year-old female with microcephaly, severe intellectual disability, and absence of language and a TUBB2B in a 31-year-old female with microcephaly, spastic tetraparesis, severe intellectual disability, and scoliosis. The patient with TUBA1A mutations in the brain MRI showed pachygyria associated with a diffuse

subcortical band heterotopia that spared the fronto-basal regions only; the heads of caudate nuclei and the putamen were fused; corpus callosum was thin with anterior commissure absent; the hippocampi showed a simplified pattern. Abnormal transition between the medulla and a-flattened pons, thickened mesencephalon, and a thinned pontomesencephalic junction were also present. The cerebellar vermis was mildly hypoplastic. Based on the results of the genetic analysis obtained in the patients, the authors (26) concluded that the frequent observation of hypoplastic and disorganized white matter tracts led to suggest that in addition to defects in neuronal migration, disruption of axon growth and guidance represent a peculiar feature of the disorders linked to tubulinopathy. Shimojima et al. (27) reported a single case of a Japanese baby girl with microcephaly, lissencephaly with cerebellar hypoplasia, and corpus callosum hypoplasia. Genetic analysis disclosed a region anomaly that encodes the N-terminal domain of TUBA1, a region uncommonly involved. The brain MRI showed the presence of colpocephaly, lateral ventricle dilatation, and simplified gyral pattern. There was also hypoplasia of the corpus callosum and the cerebellar vermis. A TUBA1A variant was reported by Bosemani et al. (28) in a 22-month-old girl with developmental delay. Brain MRI showed vermian hypoplasia with abnormally increased rostrocaudal length of the medulla and midbrain

involving the pons, and loss of the normal flat dorsal surface of the brainstem. In a study on patients with hindbrain imaging abnormalities, Oegema et al. (29) identified 10 patients. In seven of 10 patients (78%), targeted sequencing revealed mutations in three different tubulin genes (TUBA1A, TUBB2B, and TUBB3) occurring de novo or inherited from a mosaic parent. Brain MRI revealed cerebellar dysplasia combined with basal ganglia dysplasia in all of the cases (100%) and brainstem dysplasia in almost all (80%), and only irregular patterns of cortical gyri and sulci. With regard to the two patients with TUBA1A mutations, they showed microcephaly, severe developmental delay, and epilepsy. The MRI features in both cases showed diffuse irregular gyration and sulcation of cortex, vermis hypoplasia, asymmetry of pons, medulla dysplasia (in one case), partial agenesis of corpus callosum, basal ganglia hypoplasia, enlarged ventricle, and cranial nerve hypoplasia. A 9-year-old girl affected by epileptic seizures, developmental delay, hypotonia, and microcephaly was reported by Mencarelli et al. (30). The girl showed mild facial dysmorphism including bulbous nasal tip, large mouth, edema of the hands and feet with camptodactyly, bilateral thelarche, hypoplasia of the labia minora, and poor visual and social interaction. The girl had a de novo heterozygous mutation in TUBA1A gene. Brain MRI showed complex cerebral malformations with mild asymmetry and dilatation of lateral

TABLE 1 Brain MRI. Main cerebral malformations in patients with TUBA1A-tubulinopathy.

Authors	Patients with <i>TUBA1A</i> - tubulinopathy	Brain MRI—cerebral malformations
Keays et al. (1)	2	Lissencephaly, pachygyria, hippocampus anomalies
Poirier et al. (9)	3	Bilateral perisylvian asymmetrical shapes, PMG with dysmorphic basal ganglia, cerebellar vermian dysplasia, pontine hypoplasia
Fallet-Bianco et al. (15)	4 fetuses	Pathologic study—five brain structures involved: neocortex, hippocampus, corpus callosum, cerebellum, brainstem
Bahi-Buisson et al. (4)	6/106 complex cortical malformations	Perisylvian pachygyria (less severe form): posteriorly predominant pachygyria (most severe form), internal capsule dysgenesis, mild to severe cerebellar hypoplasia
Morris-Rosendahl et al. (20)	5/46 lissencephaly	Variable pattern of lissencephaly, dysgenesis of corpus callosum, cerebellar hypoplasia, variable cortical malformations
Kumar et al. (8)	125 cortical dysgeneses	 72 classical lissencephaly 22 subcortical band heterotopia 29 lissencephaly + cerebellar hypoplasia 2 lissencephaly + corpus callosum agenesis
Jansen et al. (21)	3/25 cortical development malformations	1 lissencephaly 2 polymicrogyria
Sohal et al. (14)	1	Lissencephaly
Okumura et al. (22)	1	Ventricular dilatation with thin cortex, poorly differentiated basal ganglia, corpus callosum agenesis, cerebellar hypoplasia with preserved vermis
Fallet-Bianco et al. (24)	19/60 fetuses	Pathologic study: lissencephaly and cerebellar hypoplasia distinct features in TUBA1A mutations
Hikita et al. (25)	1	Lissencephaly, microcephaly, Hirschsprung disease
Romaniello et al. (26)	79/156 diagnosis of cortical malformations	Pachygyria, heads of caudate nuclei and the putamen fused, thin corpus callosum, hippocampi simplified pattern, cerebellar vermis mildly hypoplastic
Shimojima et al. (27)	1	Colpocephaly and lateral ventricle dilatation, simplified gyral pattern, hypoplasia corpus callosum and cerebellar vermis
Bosemani et al. (28)	1	Vermian hypoplasia, abnormally increased rostrocaudal medulla and midbrain, loss of flat dorsal surface of the brainstem
Oegema et al. (29)	2/10 hindbrain imaging abnormalities	Vermis hypoplasia, pons asymmetry, partial corpus callosum asymmetry, diffuse irregular gyration and cortex sulcation
Mencarelli et al. (30)	1	Mild asymmetry and dilatation of the lateral ventricles, cortical dysgenesis with dysmorphic frontal lobes- simplified gyral pattern, cerebellar vermis and pons hypotrophy
Romaniello et al. (31)	5/28 harboring tubulin gene variants	Cortical cerebellar dysplasia
Romaniello et al. (32)	48/90 TUBA1A epileptic patients	Extensive brain malformations involving subcortical and midline structures

ventricles and reduced white matter. Cortical dysgenesis with dysmorphic frontal lobes, simplified gyral pattern, and poor development of the Sylvian fissure were also present. The corpus callosum was thin and basal ganglia were hypoplastic. Right caudate nucleus and right lenticular nucleus were dysmorphic. Cerebellar vermis and pons hypoplasia were also reported. A study was conducted by Romaniello et al. (31) on 28 patients harboring 23 heterozygous pathogenic variants in tubulin genes TUBA1A (n = 10), TUBB2B (n = 8), and TUBB3 (n = 5). Neuroimaging patterns of patients showed cerebellar dysplasia and other posterior fossa morphological anomalies. Cerebellar anomalies were found in 24/28 patients (86%): cerebellar dysplasia in 19/28 (68%) including cortical cerebellar dysplasia (18/28), either involving only the cerebellar hemispheres (12/28) or associated with vermis dysplasia (6/28). Cortical cerebellar dysplasia was found only in the right hemisphere in 13/18 including four TUNN2B, four TUBB3, and five TUBA1A mutations. In contrast to the remaining five TUBA1A patients, cortical cerebellar dysplasia was located only in the left hemispheres or in both. The authors refer that the cerebellar involvement in tubulinopathies shows peculiar aspects that may be labeled as "tubulin-related cerebellar dysplasia" (31). Epilepsy is common in patients with TUBA1A. In a cohort of 15 patients and 75 from published studies, Romaniello et al. (32) reported epileptic disorders in 60% of the patients affected by TUBA1A mutations, 74% by TUBB2B, and 25% by TUBB3. In these patients, brain MRIs were consistent with the extensive brain malformations involving subcortical and midline structures (32).

Conclusions

Genotype-phenotype characterization of structural brain abnormalities in patients with *TUBA1A*-tubulinopathies is limited by the large interstudy variability in reporting the clinical features of this disorder. Indeed, previous reports showed the various types of cerebral malformations related to this genetic disorder and their association with severe neurological and systemic manifestations.

Pathogenetic variants in TUBA1A are associated with a broad spectrum of cortical and subcortical brain malformations. Major cortical abnormalities include "classical" lissencephaly, lissencephaly with agenesis/or dysgenesis of the corpus callosum, lissencephaly with cerebellar hypoplasia, polymicrogyria, and mild to moderate dysgyria. Dysmorphic basal ganglia, thalami, and corpus callosum may also be involved as well as cerebellar vermis and cerebellar cortical dysplasia. Brainstem malformations may present with usually asymmetric hypoplasia. Clinical manifestations are variable and usually severe, including motor and cognitive impairment. Developmental delay, intellectual disability, cerebral palsy, and seizures often unresponsive to treatment are widely reported. Undistinctive facial dysmorphism, microcephaly, other associated malformations, and cerebral MRI anomalies may support genetic testing for the suspect of TUBA1A-tubulopathy. Supportive treatments including physical, occupational, speech therapy, and anticonvulsant treatment are necessary although the prognosis remains quite poor, especially in most severe cases (see Table 1).

Data availability statement

The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.

Ethics statement

Written informed consent was obtained from the minor(s)' legal guardian/next of kin for the publication of any potentially identifiable images or data included in this article.

Author contributions

Conceptualization: RF and PP. Methodology: EP. Software: GC. Validation: MR, PP, and XP. Formal analysis: GC. Investigation: RF. Data curation: XP and PP. Writing—original draft preparation: RF, PP, and GC. Writing—review and editing: MR and PP. Visualization: SM. Supervision: EP. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Keays DA, Tian G, Poirier K, Huang GJ, Siebold C, Cleak J, et al. Mutations in alpha-tubulin cause abnormal neuronal migration in mice and lissencephaly in humans. *Cell.* (2007) 128(1):45–57. doi: 10.1016/j.cell.2006.12.017

2. Di Donato N, Timms AE, Aldinger KA, Mirzaa GM, Bennett JT, Collins S, et al. Analysis of 17 genes detects mutations in 81% of 811 patients with lissencephaly. *Genet Med.* (2018) 20(11):1354–64. doi: 10.1038/gim.2018.8

3. Tischfield MA, Cederquist GY, Gupta ML Jr., Engle EC. Phenotypic spectrum of the tubulin- related disorders and functional implications of disease-causing mutations. *Curr Opin Genet Dev.* (2011) 21(3):286–94. doi: 10.1016/j.gde.2011.01.003

4. Bahi-Buisson N, Poirier K, Boddaert N, Saillour Y, Castelnau L, Philip N, et al. Refinement of cortical dysgeneses spectrum associated with TUBA1A mutations. *J Med Genet.* (2008) 45(10):647–53. doi: 10.1136/jmg.2008.058073

5. Hebebrand M, Hüffmeier U, Rollmann R, Keymolen K, Régal L, Romaniello R, et al. The mutational and phenotypic spectrum of TUBA1A-associated tubulinopathy. *Orphanet J Rare Dis.* (2019) 14(1):38. doi: 10.1186/s13023-019-1020-x

6. Kapitein LC, Hoogenraad CC. Building the neuronal microtubule cytoskeleton. Neuron. (2015) 87(3):492-506. doi: 10.1016/j.neuron.2015.05.046

7. Ravelli RB, Gigan B, Curmi PA, Jourdain I, Lachkar S, Sobel A, et al. Insight into tubulin regulation from a complex with colchicine and a stathmin-like domain. *Nature*. (2004) 428(6979):198–202. doi: 10.1038/nature02393

 Kumar RA, Pilz DT, Babatz TD, Cushion TD, Harvey K, Topf M, et al. TUBA1A mutations cause wide spectrum lissencephaly (smooth brain) and suggest that multiple neuronal migration pathways converge on alpha tubulins. *Hum Mol Genet.* (2010) 19(14):2817–27. doi: 10.1093/hmg/ddq182

9. Poirier K, Keays DA, Francis F, Saillour Y, Bahi N, Manouvrier S, et al. Large spectrum of lissencephaly and pachygyria phenotypes resulting from de novo missense mutations in tubulin alpha 1A (TUBA1A). *Hum Mutat.* (2007) 28 (11):1055–64. doi: 10.1002/humu.20572

10. Schröter J, Popp B, Brennenstuhl H, Döring JH, Donze SH, Bijlsma EK, et al. Complementing the phenotypical spectrum of TUBA1A tubulinopathy and its role in early-onset epilepsies. *Eur J Hum Genet.* (2022) 30(3):298–306. doi: 10.1038/s41431-021-01027-0

11. Hausrat TJ, Radwitz J, Lombino FL, Breiden P, Kneussel M. Alpha- and betatubulin isotypes are differentially expressed during brain development. *Develop Neurobiol.* (2021) 81:333–50. doi: 10.1002/dneu.22745

12. Nogales E, Wolf SG, Downing KH. Structure of the alpha beta tubulin dimer by electron crystallography. *Nature*. (1998) 391(6663):199–203. Erratum in: *Nature*. (1998) **393**(6681):191. doi: 10.1038/34465

13. Aiken J, Moore JK, Bates EA. TUBA1A mutations identified in lissencephaly patients dominantly disrupt neuronal migration and impair dynein activity. *Hum Mol Genet.* (2019) 28(8):1227–43. doi: 10.1093/hmg/ddy416

14. Sohal AP, Montgomery T, Mitra D, Ramesh V. TUBA1A mutation-associated lissencephaly: case report and review of the literature. *Pediatr Neurol.* (2012) 46 (2):127–31. doi: 10.1016/j.pediatrneurol.2011.11.017

15. Fallet-Bianco C, Loeuillet I, Poirier K, Loget P, Chapon F, Pasquier L, et al. Neuropathological phenotype of a distinct form of lissencephaly associated with mutations in TUBA1A. *Brain.* (2008) 131(Pt 9):2304–20. doi: 10.1093/brain/awn155

16. Abdollahi MR, Morrison E, Sirey T, Molnar Z, Hayward BE, Carr IM, et al. Mutation of the variant α -tubulin TUBA8 results in polymicrogyria with optic nerve hypoplasia. *Am J Hum Genet.* (2009) 85(5):737–44. doi: 10.1016/j.ajhg.(2009). 10.007

17. Tian G, Jaglin XH, Keays DA, Francis F, Chelly J, Cowan NJ. Disease-associated mutations in TUBA1A result in a spectrum of defects in the tubulin folding and

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fped.2023. 1210272/full#supplementary-material

heterodimer assembly pathway. Hum Mol Genet. (2010) 19(18):3599-613. doi: 10. 1093/hmg/ddq276

18. Hoff KJ, Aiken JE, Gutierrez MA, Franco SJ, Moore JK. TUBA1A tubulinopathy mutants disrupt neuron morphogenesis and override XMAP215/Stu2 regulation of microtubule dynamics. *eLife*. (2022) 11:e76189. doi: 10.7554/eLife.76189

19. Aldinger KA, Timm AE, Thomson Z, Mirzaa GM, Bennett JT, Rosenberg AB, et al. Redefining the etiologic landscape of cerebellar malformations. *Am J Hum Genet.* (2019) 105(3):606–15. doi: 10.1016/j.ajhg.2019.07.019

20. Morris-Rosendahl DJ, Najm J, Lachmeijer AM, Sztriha L, Martins M, Kuechler A, et al. Refining the phenotype of alpha-1a tubulin (TUBA1A) mutation in patients with classical lissencephaly. *Clin Genet.* (2008) 74(5):425–33. doi: 10.1111/j.1399-0004. 2008.01093.x

21. Jansen AC, Oostra A, Desprechins B, De Vlaeminck Y, Verhelst H, Régal L, et al. TUBA1A mutations: from isolated lissencephaly to familial polymicrogyria. *Neurology.* (2011) 76(11):988–92. doi: 10.1212/WNL.0b013e31821043f5

22. Okumura A, Hayashi M, Tsurui H, Yamakawa Y, Abe S, Kudo T, et al. Lissencephaly with marked ventricular dilation, agenesis of corpus callosum, and cerebellar hypoplasia caused by TUBA1A mutation. *Brain Dev.* (2013) 35(3):274–9. doi: 10.1016/j.braindev.2012.05.006

23. Bahi-Buisson N, Poirier K, Fourniol F, Saillour Y, Valence S, Lebrun N, et al. The wide spectrum of tubulinopathies: what are the key features for the diagnosis? *Brain.* (2014) 137(Pt 6):1676–700. doi: 10.1093/brain/awu082

24. Fallet-Bianco C, Laquerrière A, Poirier K, Razavi F, Guimiot F, Dias P, et al. Mutations in tubulin genes are frequent causes of various foetal malformations of cortical development including microlissencephaly. *Acta Neuropathol Commun.* (2014) 2:69. doi: 10.1186/2051-5960-2-69

25. Hikita N, Hattori H, Kato M, Sakuma S, Morotomi Y, Ishida H, et al. A case of *TUBA1A* mutation presenting with lissencephaly and Hirschsprung disease. *Brain Dev.* (2014) 36(2):159–62. doi: 10.1016/j.braindev.2013.02.006

26. Romaniello R, Arrigoni F, Cavallini A, Tenderini E, Baschirotto C, Triulzi F, et al. Brain malformations and mutations in α - and β -tubulin genes: a review of the literature and description of two new cases. *Dev Med Child Neurol.* (2014) 56 (4):354–60. doi: 10.1111/dmcn.12370

27. Shimojima K, Narita A, Maegaki Y, Saito A, Furukawa T, Yamamoto Y. Wholeexome sequencing identifies a de novo *TUBA1A* mutation in a patient with sporadic malformations of cortical development: a case report. *BMC Res Notes*. (2014) 7:465. doi: 10.1186/1756-0500-7-465

28. Bosemani T, Orman G, Boltshauser E, Tekes A, Huisman TA, Poretti A. Congenital abnormalities of the posterior fossa. *Radiographics*. (2015) 35(1):200–20. doi: 10.1148/rg.351140038

29. Oegema R, Cushion TD, Phelps IG, Chung SK, Dempsey JC, Collins S, et al. Recognizable cerebellar dysplasia associated with mutations in multiple tubulin genes. *Hum Mol Genet.* (2015) 24(18):5313–25. doi: 10.1093/hmg/ddv250

30. Mencarelli A, Prontera P, Stangoni G, Mencaroni E, Principi N, Esposito S. Epileptogenic brain malformations and mutations in tubulin genes: a case report and review of the literature. *Int J Mol Sci.* (2017) 18(11):2273. doi: 10.3390/ijms18112273

31. Romaniello R, Arrigoni F, Panzeri E, Poretti A, Micalizzi A, Citterio A, et al. Tubulin-related cerebellar dysplasia: definition of a distinct pattern of cerebellar malformation. *Eur Radiol.* (2017) 27(12):5080–92. Erratum in: *Eur Radiol.* (2017) 27(12):5093. doi: 10.1007/s00330-017-4945-2

32. Romaniello R, Zucca C, Arrigoni F, Bonanni P, Panzeri E, Bassi MT, et al. Epilepsy in tubulinopathy: personal series and literature review. *Cells.* (2019) 8(7):669. doi: 10.3390/cells8070669