Check for updates

OPEN ACCESS

EDITED BY Guillaume Leonard, Université de Sherbrooke, Canada

REVIEWED BY Sascha R.A. Alles, University of New Mexico School of Medicine, United States

*CORRESPONDENCE Terence J Coderre is terence.coderre@mcgill.ca

SPECIALTY SECTION

This article was submitted to Pain Mechanisms, a section of the journal Frontiers in Pain Research

RECEIVED 29 November 2022 ACCEPTED 09 January 2023 PUBLISHED 02 February 2023

CITATION

Coderre TJ (2023) Contribution of microvascular dysfunction to chronic pain. Front. Pain Res. 4:111559. doi: 10.3389/fpain.2023.1111559

COPYRIGHT

© 2023 Coderre. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Contribution of microvascular dysfunction to chronic pain

Terence J. Coderre*

Department of Anesthesia and Alan Edwards Centre for Research on Pain, McGill University, Montreal, QC, Canada

There is growing evidence that microvascular dysfunction is a pathology accompanying various injuries and conditions that produce chronic pain and may represent a significant contributing factor. Dysfunction that occurs within each component of the microvasculature, including arterioles, capillaries and venules impacts the health of surrounding tissue and produces pathology that can both initiate pain and influence pain sensitivity. This mini review will discuss evidence for a critical role of microvascular dysfunction or injury in pathologies that contribute to chronic pain conditions such as complex regional pain syndrome (CRPS) and fibromyalgia.

KEYWORDS

endothelial cells, microvascular injury, ischemia-reperfusion, oxidative stress, antioxidants, cytokines

Introduction

While pain physicians and researchers have for centuries concentrated on the role of nerve and inflammatory injury to chronic pain and have more recently studied the potential role of neuron-glia interactions, very little attention has been paid to microvascular function. This is surprising given that a healthy microvasculature is critical for endoneurial health, and microvascular injury plays a significant role in various components of the inflammatory response to tissue damage.

Endoneurial microvessels are essential to the control of ion, solute and water transfer between the bloodstream and the endoneurium. These functions allow for nutrient, macromolecule and leukocyte influx and efflux, as well as contributing to interstitial fluid balance between layers of the endoneurium (1). Alterations in these processes may result in nerve damage that leads to neuropathic pain.

All three element of the microvasculature contribute to inflammation (see Figure 1), with arterioles involved in impaired vasomotor function, capillaries critical for reduced perfusion and poor oxygenation of tissue, and post-capillary venules playing a role in increased vascular permeability (2). These actions trigger various other inflammatory responses including the adhesion of platelets and leukocytes, activation of the coagulation cascade, increased thrombosis, and the enhanced proliferation of blood and lymphatic vessels (3).

In the current minireview, I will assess the role of microvascular dysfunction in two chronic pain conditions, complex regional pain syndrome (CRPS) and fibromyalgia. I concentrate on these two syndromes due to both the elusive nature of their etiologies and/or to the growing evidence of the role played by microvascular dysfunction to their underlying pathologies. A brief section will also discuss the role of the microvasculature in other chronic pain syndromes, such as migraine and neuropathic pain.

Evidence for microvascular dysfunction in CRPS

CRPS, until recently known as reflex sympathetic dystrophy, is a chronic pain condition that usually follows deep-tissue injuries such as fractures, crush injuries or sprains, with incidence

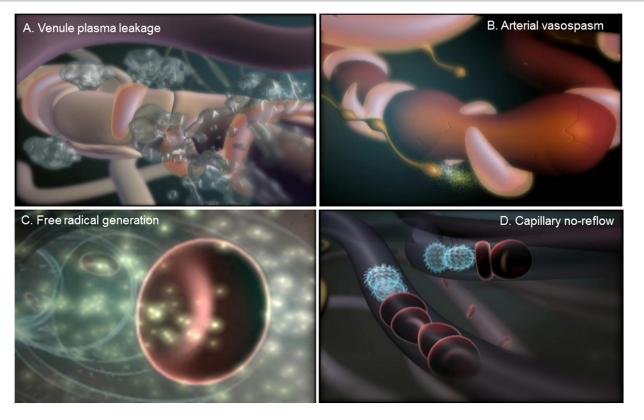


FIGURE 1

Microvascular response to tissue injury. (A) Swelling or edema is caused by a leakage of plasma proteins from microvessels within the post-capillary venules. (B) Extensive edema within damaged tissue can cause vasospasms within arterioles that can produce prolonged tissue ischemia. (C) Reperfusion of microvessels in the ischemic tissue generates the production of oxygen and nitrogen free radicals, which produce injury of the endothelial lining, attraction and adhesion of platelets and leukocytes, activation of the coagulation cascade, and increased thrombosis. (D) The adhesion of platelets and leukocytes leads to a blockade of blood flow within capillaries, or the phenomenon capillary no-reflow, which encourages arteriovenous shunting and the enhanced proliferation of blood and lymphatic vessels.

estimates as high as 26.2 per 100,000 person years (4). Symptoms of CRPS include spontaneous pain ("burning" pain referred to the skin, and "aching" pain referred to deep tissues), and a variety of stimulusevoked abnormal pain sensations, including mechanical allodynia and hyperalgesia, cold allodynia, and sometimes heat hyperalgesia (5). CRPS-I pain is disproportionate to the injury and is not associated with injury of a major nerve, unlike CRPS-II a similar syndrome, but with concomitant major nerve damage. Non-painful symptoms of CRPS include disorders of vasomotor and sudomotor regulation; trophic changes in skin, hair, nails, and bone; and dystonia and other motor abnormalities (5). Vascular abnormalities are prominent symptoms in CRPS patients, with either increased or decreased skin temperature observed, typically corresponding with increased or decreased blood flow (6, 7), and alterations in bone (8, 9). However, it is a common finding that CRPS vascular symptoms change over time. The general picture often reported is a short initial "hot" phase lasting weeks to months, followed by a "mixed hot/cold" phase, and finally a chronic "cold" phase lasting for years (10, 11). During the "hot" phase, the limb displays increased skin temperature, edema, sweating and inflammation, while in the "cold" phase it shows lowered skin temperature, dryness, and cyanosis.

There is a growing body of evidence which suggests that at least some cases of CRPS may also depend on ischemia in deep tissues, and occasionally skin, secondary to limb trauma. Muscle tissue obtained after amputation of the affected limbs of 8 CRPS patients was found to exhibit lipofuscin pigment, atrophic fibers, and collapsed lumens and severely thickened basal membrane layers of the capillaries (12). These findings are consistent with oxidative stress, lipid peroxidation and ischemic conditions resulting from microvascular injury in deep tissue. Although amputation may reflect the most severe cases of CRPS, this conclusion is supported by observations of increased density of perfused vessels; lower capillary filtration capacity, and arteriovenous shunting of peripheral subcutaneous tissue in the intact limbs of patients with CRPS (13, 14). Co-incident with these changes appears to be high arterial flow to the CRPS limb, but low oxygen consumption (indicated by high venous oxygen saturation), as well as high lactate flux-indicative of tissue ischemia despite increased flow in large vessels (15, 16). There is also an impairment of high-energy phosphate metabolism in muscle tissue of CRPS limbs (16, 17), suggestive of mitochondrial oxygen deficiency. A role of oxidative stress in CRPS is suggested by finding that symptoms are relieved in these patients following treatment with antioxidants and free radical scavengers (18, 19). These observations suggest that ischemia in deep tissues may contribute to the induction and maintenance of CRPS. Increased activity in the sympathetic vasoconstrictor innervation of the affected limb would exacerbate any underlying ischemic condition and might be an important contributor to ischemia in some cases. It

has also been shown that skin capillary hemoglobin oxygenation (HbO_2) or skin nutritive flow is reduced in CRPS patients (6, 10, 20) limbs. CRPS patients can have either hot limbs with high blood flow, or cold limbs with reduced blood flow, but whether patients experience pain or not, is independent of the direction of the alteration in skin temperature or thermoregulatory flow (10, 21).

The findings summarized above indicating that there may be microvascular dysfunction and ischemia in deep tissues of CRPS patients, suggest that a disruption of nutritive flow may be a more important factor underlying pain in CRPS patients than changes in thermoregulatory flow. Thus, although thermoregulatory flow may be low or high, reduced nutritive flow to deep tissue in either case would reduce oxygenation leading to tissue ischemia.

Evidence for microvascular dysfunction in fibromyalgia

Fibromyalgia (FM) is a chronic pain condition that affects about 2%-3% of the adult population. The defining feature of FM is chronic widespread pain, but patients often exhibit other symptoms, including fatigue, sleep disturbance, irritable bowel syndrome, mood disorders and headache (22, 23). Although the underlying causes of FM are not well understood, the syndrome has been associated with medical illness, stress, various other pain conditions, as well as disturbances in various neurotransmitter and neuroendocrine systems (23, 24). There is also evidence for both peripheral and central sensitization in FM, as well as dysfunction of descending inhibitory control systems (24, 25). Widespread pain is one of the diagnostic criteria for FM, with the common diagnostic requirement of tenderness in 11 of 18 defined tender points (26). Pain is described as deep, gnawing or burning, and can be accompanied by stiffness, skin tenderness, pain after exercise, paraesthesias, restless legs, and Raynaud's phenomenon, a process in which the fingers or toes become blanched in response to cold or stress (27). Theories about the role of muscle pathology in FM have been controversial (28), although numerous studies have reported histological abnormalities in muscle tissue (29, 30).

The controversy over the role of muscle pathology in FM likely stems from findings that the muscle pathology is subtle, and typically detected by histological rather than functional assessments (28). Studies indicate that muscle pathology includes capillary dysfunction and abnormalities of mitochondria in myocytes (31-33). Mitochondrial abnormalities are significant since mitochondria provide the energy source in the form of ATP through oxidative phosphorylation, and function poorly when not supplied with enough oxygen (34). Recent studies do indicate that there are signs of oxidative stress or increases in oxygen free radicals in FM patients. Thus, in FM patients there is increased serum malondialdehyde and lipid hydroperoxide, indicators of free radicalinduced lipid peroxidation, and decreases in xanthine oxidase, which is normally depleted when oxygen free radicals are produced (35, 36). There are also decreases in the endogenous antioxidants glutathione and superoxide dismutase and total antioxidant status in serum, which would be depleted in response to oxidative stress (35, 37). FM patients also have higher levels of oxygen free radicals in blood mononuclear cells (38), and neutrophils from FM patients produce greater levels of the oxygen free radical hydrogen peroxide (39). This may explain why antioxidants have been useful as analgesics for fibromyalgia in some studies (40).

Additional evidence suggests that oxidative stress in FM patients may occur in response to microvascular dysfunction in affected muscle tissue and possibility in skin. Indeed, it has been shown that FM patients have reduced capillary flow and fewer capillaries in the nail fold (31, 32), perhaps explaining why Raynaud's phenomena is common in FM patients (41). As for muscle, recent microdialysis studies suggests that FM patients have reduced nutritive blood flow and increased lactate in skeletal muscle in response to exercise (42). These results support earlier findings of reduced capillaries, thickened capillary endothelial cells and lowered ATP in biopsies from the vastus lateralis muscle of FM patients (33). These are consistent with findings of reduced muscle blood flow in FM patients as measured using xenon 133 clearance (43), and reduced muscle blood flow in cervical spine trapezius of FM patients following exercise (44). There was also one report indicating that muscle tissue oxygenation is lower in the tender point areas of FM patients (45). Also, while blood flow and skin temperature has been reported to be either lower (46) or higher (47) over tender points in FM patients, there is no correlation between skin temperature and pressure pain threshold (48).

Evidence for microvascular dysfunction in other pain syndromes

The space limitations of mini reviews prevent a detailed description of similar pathology within in other pain syndromes. However, there is considerable evidence for a role of microvascular dysfunction contributing to chronic pain in various syndromes such as angina, frostbite, sickle cell disease, and peripheral vascular disease (49, 50). Data suggest a role for dysfunction in the endoneurial tissues of patients with painful diabetic neuropathy (51, 52), and experimental animals with neuropathic pain (53). There is also evidence that patients with migraine have contributing or associated alterations in microvascular function within the retina (54), dura matter (55), cerebral cortex (56) and coronary tissue (57).

Conclusion

In this mini review, we summarized the potential contribution of microvascular dysfunction to the chronic pain conditions CRPS and fibromyalgia. Evidence of microvascular dysfunction is exhibited by numerous findings of thickened capillary endothelial cells, disruptions in capillary or nutritive blood flow, increases in lactate and oxidative stress, and arteriovenous shunting and abnormalities in mitochondria for both conditions. Numerous studies have concentrated on the effects of mediators released from various immune cells, including mast cells, neutrophils, macrophages and Tcells and their impact on neural function. Far fewer studies have investigated the role of mediators released by or from endothelial cells, including adhesion factors, endothelins, cytokines, chemokines, selectins, additional vasodilatory and vasoconstrictive factors, antithrombotic and procoagulant factors, matrix products, endothelial growth factors, and others, on both immune and neural function (58). It is hoped that future research and targeted therapy will be aimed at microvascular dysfunction and its role in the etiology and maintenance of symptoms in these and potential other chronic pain syndromes. These studies would add to the above-mentioned studies demonstrating the effectiveness of antioxidants or free radical scavengers as analgesics for CRPS (18, 19) and fibromyalgia (40), and why combinations of topical agents aimed at improving both arterial and capillary blood flow (59), or salts and co-crystals comprised of vasodilators and antioxidants are effective analgesics in animal models of CRPS and neuropathic pain (60).

Author contributions

Article was written by TJC. The author was the sole contributor to the article and approved the submitted version.

Acknowledgments

The author wishes to acknowledge the support of the Canadian Institutes of Health Research, the Natural Sciences and Engineering

References

1. Ubogu EE. Biology of the human blood-nerve barrier in health and disease. *Exp Neurol.* (2020) 328:113272. doi: 10.1016/j.expneurol.2020.113272

2. Pober JS, Sessa WC. Inflammation and the blood microvascular system. *Cold Spring Harb Perspect Biol.* (2014) 7:a016345. doi: 10.1101/cshperspect.a016345

3. Granger DN, Senchenkova E. Inflammation and the microcirculation. San rafael: Morgan & Claypool Life Sciences (2010).

4. de Mos M, de Bruijn AG, Huygen FJ, Dieleman JP, Stricker BH, Sturkenboom MC. The incidence of complex regional pain syndrome: a population-based study. *Pain.* (2007) 129:12–20. doi: 10.1016/j.pain.2006.09.008

5. Schwartzman RJ, McLellan TL. Reflex sympathetic dystrophy. A review. Arch Neurol. (1997) 44:555-61. doi: 10.1001/archneur.1987.00520170081028

6. Rosen L, Ostergren J, Fagrell B, Stranden E. Skin microvascular circulation in the sympathetic dystrophies evaluated by videophotometric capillaroscopy and laser Doppler fluxmetry. *Eur J Clin Invest.* (1988) 18:305–8. doi: 10.1111/j.1365-2362.1988.tb01263.x

7. Ackerman WE, Munir MA, Zhang J-M. Assessment of laser Doppler imaging for the diagnosis of complex regional pain syndrome I. J Neuro Pain Symp Pal. (2005) 1:13–20. doi: 10.3109/J426v01n04_03

8. Leitha T, Korpan M, Staudenherz A, Wunderbaldinger P, Fialka V. Five phase bone scintigraphy supports the pathophysiological concept of a subclinical inflammatory process in reflex sympathetic dystrophy. *Q J Nucl Med.* (1996) 40:188–93. PMID: 8909105.

9. Zyluk A. The usefulness of quantitative evaluation of three-phase scintigraphy in the diagnosis of post-traumatic reflex sympathetic dystrophy. J Hand Surg Br. (1999) 24:16–21. doi: 10.1016/S0266-7681(99)90011-6

10. Kurvers HA, Jacobs MJ, Beuk RJ, Van den Wildenberg FA, Kitslaar PJ, Slaaf DW, et al. Reflex sympathetic dystrophy: evolution of microcirculatory disturbances in time. *Pain.* (1995) 60:333–40. doi: 10.1016/0304-3959(94)00133-Y

11. Bruehl S, Harden RN, Galer BS, Saltz S, Backonja M, Stanton-Hicks M. Complex regional pain syndrome: are there distinct subtypes and sequential stages of the syndrome? *Pain.* (2002) 95:119–24. doi: 10.1016/S0304-3959(01)00387-6

12. van der Laan L, ter Laak HJ, Gabreels-Festen A, Gabreels F, Goris RJA. Complex regional pain syndrome type I (RSD): pathology of skeletal muscle and peripheral nerve. *Neurology.* (1998) 51:20–5. doi: 10.1212/WNL.51.1.20

13. Matsumura H, Jimbo Y, Watanabe K. Haemodynamic changes in early phase reflex sympathetic dystrophy. *Scandinavian J Plastic Reconstr Surg and Hand Surg.* (1996) 30:133–8. doi: 10.3109/02844319609056395

14. Schurmann M, Zaspel J, Grandl G, Wipfel A, Christ F. Assessment of the peripheral microcirculation using computer-assisted venous congestion plethysmography in post-traumatic complex regional pain syndrome type I. *J Vasc Res.* (2001) 38:453–61. doi: 10.1159/000051078

Research Council of Canada, and the Louise and Alan Edwards Foundation.

Conflict of interest

The author declares that the conclusion drawn in this review were made in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

15. Goris RJA. Conditions associated with impaired oxygen extraction. In: G Guitierrez, JL Vincent, editors. *Tissue oxygen utilization*. Berlin: Springer Verlag (1991). p. 350-69.

16. Goris RJA. Reflex sympathetic dystrophy: model of a severe regional inflammatory response syndrome. World J Surg. (1998) 22:197–202. doi: 10.1007/s002689900369

17. Heerschap A, den Hollander JA, Reynen H, Goris RJ. Metabolic changes in reflex sympathetic dystrophy: a 31P NMR spectroscopy study. *Muscle and Nerve.* (1993) 16:367–73. doi: 10.1002/mus.880160405

18. Perez RS, Zuurmond WW, Bezemer PD, Zuurmond WW, de Lange JJ. The treatment of complex regional pain syndrome type I with free radical scavengers: a randomized controlled study. *Pain.* (2003) 102:297–307. doi: 10.1016/S0304-3959(02)00414-1

19. Goris RJ, Dongen LM, Winters HA. Are toxic oxygen radicals involved in the pathogenesis of reflex sympathetic dystrophy? *Free Radic Res Commun.* (1987) 3:13–8. doi: 10.3109/10715768709069764

20. Koban M, Leis S, Schultze-Mosgau S, Birklein F. Tissue hypoxia in complex regional pain syndrome. *Pain.* (2003) 104:149–57. doi: 10.1016/S0304-3959(02)00484-0

21. Kurvers HA, Hofstra L, Jacobs MJ, Daemen MA, van den Wildenberg FA, Kitslaar PJ, et al. Reflex sympathetic dystrophy: does sympathetic dysfunction originate from peripheral neuropathy? *Surgery*. (1996) 119:288–96. doi: 10.1016/S0039-6060(96) 80115-2

22. Clauw DJ. Elusive syndromes: treating the biologic basis of fibromyalgia and related syndromes. *Cleve Clin J Med.* (2001) 68:832–4. doi: 10.3949/ccjm.68.10.830

23. Mease P. Fibromyalgia syndrome: review of clinical presentation, pathogenesis, outcome measures, and treatment. J Rheumatol Suppl. (2005) 75:6–21. PMID: 16078356.

24. Bennett RM. Emerging concepts in the neurobiology of chronic pain: evidence of abnormal sensory processing in fibromyalgia. *Mayo Clin Proc.* (1999) 74:385–98. doi: 10. 4065/74.4.385

25. Julien N, Goffaux P, Arsenault P, Marchand S. Widespread pain in fibromyalgia is related to a deficit of endogenous pain inhibition. *Pain.* (2005) 114:295–302. doi: 10. 1016/j.pain.2004.12.032

26. Wolfe F, Smythe HA, Yunus MB, Bennett RM, Bombardier C, Goldenberg DL, et al. The American college of rheumatology (1990 criteria for the classification of fibromyalgia. *Arthritis Rheum*. (1990) 33:160–72. doi: 10.1002/art.1780330203

27. Bennett RM. Fibromyalgia. In: PD Wall, R Melzack, editors. *Textbook of pain*, 4th ed. London: Harcourt Publishers, Churchill Livingstone (1999). p. 579–601.

28. Le Goff P. Is fibromyalgia a muscle disorder? Joint Bone Spine. (2006) 73:239-42. doi: 10.1016/j.jbspin.2005.03.022

29. Kalyan-Raman UP, Kalyan-Raman K, Yunus MB, Masi AT. Muscle pathology in primary fibromyalgia syndrome a light microscopic, histochemical and ultrastructural study. *J Rheumatol.* (1984) 11:808–13. PMID: 6596430

30. Drewes AM, Andreasen A, Schroder HD, Hogsaa B, Jennum P. Pathology of skeletal muscle in fibromyalgia: a histo-immuno-chemical and ultrastructural study. *Br J Rheumatol.* (1993) 32:479–83. doi: 10.1093/rheumatology/32.6.479

31. Grassi W, Core P, Carlino G, Salaffi F, Cervini C. Capillary permeability in fibromyalgia. J Rheumatol. (1994) 21:1328–31. PMID: 7966078.

32. Morf S, Amann-Vesti B, Forster A, Franzeck UK, Koppensteiner R, Uebelhart D, et al. Microcirculation abnormalities in patients with fibromyalgia—measured by capillary microscopy and laser fluxmetry. *Arthritis Res Ther.* (2005) 7:R209–216. doi: 10.1186/ar1459

33. Lindman R, Hagberg M, Bengtsson A, Henriksson KG, Thornell L-E. Capillary structure and mitochondrial volume density in the trapezius muscle of chronic trapezius myalgia, fibromyalgia and healthy subjects. *J Musculoskeletal Pain.* (1995) 3:5–22. doi: 10.1300/J094v03n03_02

34. Campbell NA, Williamson B, Heyden RJ. *Biology: Exploring life.* Boston, Massachusetts: Pearson Prentice Hall (2006).

35. Bagis S, Tamer L, Sahin G, Bilgin R, Guler H, Ercan B, et al. Free radicals and antioxidants in primary fibromyalgia: an oxidative stress disorder? *Rheumatol Int.* (2005) 25:188–90. doi: 10.1007/s00296-003-0427-8

36. Ozgocmen S, Ozyurt H, Sogut S, Akyol O, Ardicoglu O, Yildizhan H. Antioxidant status, lipid peroxidation and nitric oxide in fibromyalgia: etiologic and therapeutic concerns. *Rheumatol Int.* (2006) 26:598–603. doi: 10.1007/s00296-005-0079-y

37. Sendur OF, Turan Y, Tastaban E, Yenisey C, Serter M. Serum antioxidants and nitric oxide levels in fibromyalgia: a controlled study. *Rheumatol Int.* (2009) 29:629–33. doi: 10.1007/s00296-008-0738-x

38. Cordero MD, Moreno-Fernández AM, deMiguel M, Bonal P, Campa F, Jiménez-Jiménez LM, et al. Coenzyme Q10 distribution in blood is altered in patients with fibromyalgia. *Clin Biochem.* (2009) 42:732–5. doi: 10.1016/j.clinbiochem.2008.12.010

39. Kaufmann I, Schelling G, Eisner C, Richter HP, Krauseneck T, Vogeser M, et al. Anandamide and neutrophil function in patients with fibromyalgia. *Psychoneuroendocrinology*. (2008) 33:676–85. doi: 10.1016/j.psyneuen.2008.02.009

40. Fernández-Araque A, Verde Z, Torres-Ortega C, Sainz-Gil M, Velasco-Gonzalez V, González-Bernal JJ, et al. Effects of antioxidants on pain perception in patients with fibromyalgia-A systematic review. *J Clin Med.* (2022) 11:2462. doi: 10.3390/jcm11092462

41. Bennett RM, Clark SR, Campbell SM, Ingram SB, Burckhardt CS, Nelson DL, et al. Symptoms of Raynaud's Syndrome in patients with fibromyalgia. A study utilizing the nielsen test, digital photoplethysmography, and measurements of platelet alpha2adrenergic receptors. *Arthritis Rheum.* (1991) 34:264–9. doi: 10.1002/art.1780340303

42. McIver KL, Evans C, Kraus RM, Ispas L, Sciotti VM, Hickner RC. NO-mediated alterations in skeletal muscle nutritive blood flow and lactate metabolism in fibromyalgia. *Pain.* (2006) 120:161–9. doi: 10.1016/j.pain.2005.10.032

43. Bennett RM, Clark SR, Goldberg L, Nelson D, Bonafede RP, Porter J, et al. Aerobic fitness in patients with fibrositis. A controlled study of respiratory gas exchange and 133xenon clearance from exercising muscle. *Arthritis Rheum.* (1989) 32:454–60. doi: 10.1002/anr.1780320415

44. Larsson SE, Ålund M, Cai H, Öberg PÅ. Chronic pain after soft injury of the cervical spine trapezius muscle blood flow and electromyography at static loads and fatigue. *Pain.* (1994) 57:173–80. doi: 10.1016/0304-3959(94)90221-6

45. Lund N, Bengtsson A, Thorborg P. Muscle tissue oxygen pressure in primary fibromyalgia. Scand J Rheumatol. (1986) 15:165–73. doi: 10.3109/03009748609102084

46. Jeschonneck M, Grohmann G, Hein G, Sprott H. Abnormal microcirculation and temperature in skin above tender points in patients with fibromyalgia. *Rheumatology*. (2000) 39:917–21. doi: 10.1093/rheumatology/39.8.917

47. Fischer AA, Chang CH. Temperature and pressure threshold measurements in trigger points. *Thermology*. (1986) 1:212-5.

48. Radhakrishna M, Burnham R. Infrared skin temperature measurement cannot be used to detect myofascial tender spots. *Arch Phys Med Rehabil.* (2001) 82:902–5. doi: 10. 1053/apmr.2001.23875

49. Queme LF, Ross JL, Jankowski MP. Peripheral mechanisms of ischemic myalgia. *Front Cell Neurosci.* (2017) 11:419. doi: 10.3389/fncel.2017.00419

50. Zook N, Hussmann J, Brown R, Russell R, Kucan J, Roth A, et al. Microcirculatory studies of frostbite injury. *Ann Plast Surg.* (1998) 40:246–53. doi: 10.1097/00000637-199803000-00009

51. Quattrini C, Harris ND, Malik RA, Tesfaye S. Impaired skin microvascular reactivity in painful diabetic neuropathy. *Diabetes Care.* (2007) 30:655–9. doi: 10.2337/dc06-2154

52. Yasuda H, Dyck PJ. Abnormalities of endoneurial microvessels and sural nerve pathology in diabetic neuropathy. *Neurology*. (1987) 37:20-8. doi: 10.1212/WNL.37.1.20

53. Lim TK, Shi XQ, Johnson JM, Rone MB, Antel JP, David S, et al. Peripheral nerve injury induces persistent vascular dysfunction and endoneurial hypoxia, contributing to the genesis of neuropathic pain. *J Neurosci.* (2015) 35:3346–59. doi: 10.1523/JNEUROSCI.4040-14.2015

54. Rose KM, Wong TY, Carson AP, Couper DJ, Klein R, Sharrett AR. Migraine and retinal microvascular abnormalities: the atherosclerosis risk in communities study. *Neurology.* (2007) 68:1694–700. doi: 10.1212/01.wnl.0000261916.42871.05

55. Glinskii OV, Huxley VH, Glinsky VV. Estrogen-Dependent changes in Dura mater microvasculature add new insights to the pathogenesis of headache. *Front Neurol.* (2017) 8:549. doi: 10.3389/fneur.2017.00549

56. Viola S, Viola P, Buongarzone MP, Fiorelli L, Cipulli M, Litterio P. Microvascular vasospasm of cerebral cortex in prolonged aura migraine. *Neurol Sci.* (2018) 39(Suppl 1):95–6. doi: 10.1007/s10072-018-3343-0

57. Elgendy IY, Nadeau SE, Bairey Merz CN, Pepine CJ. American College of cardiology cardiovascular disease in women committee. Migraine headache: an under-appreciated risk factor for cardiovascular disease in women. *J Am Heart Assoc.* (2019) 8:e014546. doi: 10.1161/JAHA.119.014546

58. Galley HF, Webster NR. Physiology of the endothelium. Br J Anaesth. (2004) 93:105-13. doi: 10.1093/bja/aeh163

59. Ragavendran JV, Laferrière A, Xiao WH, Bennett GJ, Padi SS, Zhang J, et al. Topical combinations aimed at treating microvascular dysfunction reduce allodynia in rat models of CRPS-I and neuropathic pain. *J Pain.* (2013) 14:66–78. doi: 10.1016/j. jpain.2012.10.004

60. Fulas OA, Laferrière A, Coderre TJ. Novel co-crystal of pentoxifylline and protocatechuic acid relieves allodynia in rat models of peripheral neuropathic pain and CRPS by alleviating local tissue hypoxia. ACS Chem Neurosci. (2021) 12:3855-63. doi: 10.1021/acschemneuro.1c00312