Check for updates

OPEN ACCESS

EDITED BY Vui King Vincent-Chong, University at Buffalo, United States

REVIEWED BY

Pamela A. Hershberger, University at Buffalo, United States Annie Wai Yeeng Chai, Cancer Research Malaysia, Malaysia Lee Peng Karen-Ng, University of Malaya, Malaysia

*CORRESPONDENCE

Sushmita Ghoshal rtsushmita@gmail.com Arnab Pal pal.arnab@pgimer.edu.in

RECEIVED 01 May 2024 ACCEPTED 16 July 2024 PUBLISHED 02 August 2024

CITATION

Suri S, Boora GS, Kaur R, Chauhan A, Ghoshal S and Pal A (2024) Recent advances in minimally invasive biomarkers of OSCC: from generalized to personalized approach. Front. Oral. Health 5:1426507. doi: 10.3389/froh.2024.1426507

COPYRIGHT

© 2024 Suri, Boora, Kaur, Chauhan, Ghoshal and Pal. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Recent advances in minimally invasive biomarkers of OSCC: from generalized to personalized approach

Smriti Suri¹, Geeta S. Boora¹, Rajandeep Kaur¹, Anshika Chauhan¹, Sushmita Ghoshal^{2*} and Arnab Pal^{1*}

¹Department of Biochemistry, Postgraduate Institute of Medical Education and Research, Chandigarh, India, ²Department of Radiotherapy, Postgraduate Institute of Medical Education and Research, Chandigarh, India

Oral cancer is the 6th most common type of cancer worldwide, and oral squamous cell carcinoma (OSCC) accounts for >90% of oral cancers. It is a major health problem, particularly in low- and middle-income countries (LMICs), due to both its high incidence and significant mortality and morbidity. Despite being a global burden, and even with the significant advancement in the management of OSCC, the overall outcome of the disease is still abysmal. With the advent of time, advanced diagnostic and treatment approaches have come into practice, but the burden of the disease has not improved significantly. Major reasons attributed to the poor outcome are delay in diagnosis, locoregional recurrence and resistance to the currently available treatment regimen. In this review, we have highlighted the existing challenges in the diagnosis and have emphasized the advancements in minimally invasive biomarkers. Additionally, the importance of collaborative multidimensional approaches involving clinicians and researchers has been discussed, as well as the need to redefine and establish better utility and management of existing diagnostic and treatment protocols along with the minimally invasive/non-invasive biomarkers.

KEYWORDS

oral squamous cell carcinoma (OSCC), minimally invasive biomarker, proteomic biomarker, genomic biomarkers, circulating tumour cell (CTC), circulating tumour DNA (ctDNA)

1 Introduction

Oral squamous cell carcinoma (OSCC) is a global health problem and is the sixth most common cancer worldwide, with a global incidence of 389,846 cases in the year 2022. Among those cases, the majority number of cases (258,440) were observed in the Asian subcontinent (1). According to Globocan 2022 estimates, the disease accounts for 188,438 deaths, approximately 75% of them are from Asia (1). With a highly disproportionate burden in low-and middle-income countries (LMICs), it accounts for >25% of all cancer in certain regions globally (2). OSCC is characterized by neoplastic transformation of the epithelium of the oral cavity. The lesions are associated with pain, ulceration, nodularity, and irregular floor and margins, which are often hard upon palpitation (3, 4). Although the disease has a multifactorial etiology, it is majorly caused by smoking, betel nut chewing, alcohol consumption, nutritional deficiencies, poor oral hygiene, traumatic injury of dental origin and viral infections with HPVs and has a significant male preponderance (4). Surprisingly, the combination of alcohol and smoking abuse has led to 11 million deaths annually, leading to a synergistic increase in cases (5).

2 Traditional approaches towards the management of OSCC

The majority of the cases are diagnosed primarily by clinical examination and confirmed with tissue biopsy of the lesion and prognosticated by TNM staging after radiological and histological examination. The current treatment modalities for OSCC are surgery with or without removal of lymph nodes by neck dissection, followed by radiotherapy and/or chemotherapy. In cases of non-metastatic and early-stage disease, surgery remains the primary treatment approach. In contrast, patients with a later disease stage and with high risk of recurrence are treated with radiation therapy (RT) and/or chemoradiotherapy (CRT) in addition to surgery as an adjuvant approach (6). At times, patients who are unfit for surgery are given RT/CRT as the primary treatment modality (7). Systemic therapy involves usage of chemotherapeutic agents like docetaxel and cisplatin where surgery cannot be performed or in higher-stage cancers (8). Surgery, radiotherapy, and chemotherapy are the first line of treatment methods for OSCC, but since last few years, immunotherapy has also come in power. In 2016, the U.S. food and drug administration (FDA) approved two PD-1 immune checkpoint inhibitors nivolumab (Opdivo) and pembrolizumab (Keytruda) to be used in the treatment of relapsed and refractory HNSCC, also few more antibodies against PD-1 have been approved for marketing in the US (9). Compared to other cancers, the 5-year survival rate of OSCC is poor, ranging approximately from 20% to 80% based on the stage at which the disease is diagnosed (10, 11). However, the majority of the cases are diagnosed at a later stage, leading to an overall inferior outcome.

3 Challenges associated with currently available management approaches

Various technological advancements have been made in the field of diagnosis to facilitate timely screening, staging, and monitoring of the disease and initiate therapeutic response, in last two decades. The diagnostic approaches for OSCC can be categorized in two major groups viz tumour imaging techniques and molecular diagnostic techniques (12). X-rays, computed tomography (CT), positron emission tomography (PET), single photon emission computed tomography (SPECT), and magnetic resonance imaging/spectroscopy (MRI/MRS) are among the tumour imaging techniques used for diagnosis of OSCC. In contrast, at the molecular levels, techniques such as next generation sequencing (NGS), assessment of genomic alteration via liquid biopsy using circulating tumour cells (CTCs), circular tumour DNA (ctDNA), non-coding RNAs, exosomes and tissuebased approach via immunohistochemistry (IHC) and spatial transcriptomics are used. Tumour imaging techniques majorly deal with the anatomical aspects including size, shape involvement of nearby tissues etc., the molecular diagnostics report the functional aspect of the tumour in cellular and subcellular levels by analyzing metabolomic, proteomic, genomic and transcriptomic signatures of cancer cells (13–16). However, most of the molecular diagnostic aspects of OSCC are currently confined to laboratory setup (17).

Despite several management approaches, timely diagnosis and treatment of OSCC still remains a challenge. It has been noticed that a significant number of patients are unaware of the condition, and the disease remains undetected till it grows in size and stage and becomes visibly identifiable; this delay might be due to a lack of awareness or seriousness among patient groups towards the initial symptoms (18). Also, at times, there is a sense of denial among patients that leads to delays in seeking medical advice at the incipient stage of the disease, further worsening the condition (19). Furthermore, patients with higher tumour stage (T3-T4), advanced histological grading (moderate or poor differentiation), lymph node positivity (N2-N3) and progressive vascularization show an early relapse of the disease (20, 21). As in a good number of patients, the disease gets diagnosed at later stages, its curative management becomes challenging, and patients often present with incomplete remission or early recurrence.

4 Scope of improvement in diagnostics-emerging role of minimally invasive biomarkers

For a normal healthy cell to become malignant, various alterations in its genome, epigenome, and transcriptome are involved. The pattern of these alterations may vary from patient to patient and will ultimately play an important role in disease progression as well as in the treatment response (22). Many times, the alterations are at different levels simultaneously involving genome, epigenome, transcriptome etc., making the tumour tissue with a heterogeneous population of cancer cells with different characteristics. Moreover, these characteristics are also dynamic in nature and change in temporo-spatial fashions. These heterogeneous characteristics of tumours indicate the imperative need for personalized cancer management. The detection of the pattern in the form of changes in the level of biomarker in conjunction with the current diagnostic protocol is thus important for designing a personalized therapy to improve the disease outcome (23).

4.1 Biomarkers for OSCC

National Cancer Institute (NCI) states that a biomarker is "a biological molecule found in blood, other body fluid, or tissues that is a sign of a normal or abnormal process, or of a condition or disease" (24). Specifically, a cancer biomarker is an entity that quantifies as an indicator of risk and occurrence of cancer and helps predict patient outcomes to a modality (25). Within the body, there can be a varied range of molecules with altered levels and phenotypes indicative of different conditions. These molecules help to differentiate diseased subjects from non-diseased healthy individuals (26). After undergoing various steps in the process of discovery and development, from analytical

validation and evidentiary assessment qualification to utilization, the molecule can serve as a biomarker and qualifies as the primary basis for regulatory approval for marketing (27).

With the advent of time and advancement in the field of tumour diagnostics, several tumour biomarker research experts published "REporting recommendations for Tumour MARKer prognostic studies" (REMARK) criteria, with an aim to provide guidelines while assessing a biomolecule to be considered as a biomarker and reporting of Tumour Biomarker Tests (TBT) associated studies. Along with that, later on, to enhance transparency on biospecimen collection, processing and archiving, Biospecimen Reporting for Improved Study Quality (BRISQ) criteria were developed (28).

A variety of samples can be used to study cancer biomarkers. Tumour tissue, the most invasive among others, is the most reliable and widely used sample type for biopsies so far. Since cancer is a heterogeneously evolving disease associated with metastasis, it is preferred to understand the spatial and temporal evolution by looking at multiple biopsies to get a holistic approach to the disease management in real time (29). However, obtaining resected tissue through invasive procedures at regular intervals for molecular profiling of the disease is associated with various challenges (30). It is not feasible to invasively acquire patient tumour tissue samples repeatedly throughout the course of treatment to monitor response and relapse (29). Considering the various challenges and shortcomings associated with traditional biopsy approaches, current analysis of tumour progression is being complemented by less invasive and more convenient approaches of liquid biopsy (31). Liquid biopsy is a technique of analyzing various biological fluids such as blood, saliva, pleural fluids, urine and cerebrospinal fluids (CSF), to get a real-time picture of tumour status by focusing on different tumour-derived components such as circulating tumour cells (CTCs), circulating tumour DNA (ctDNA), and tumour extracellular vesicles (EVs) (32, 33). Since a great emphasis in current cancer diagnosis is on finding less invasive and costeffective strategy for early diagnosis and improved prognosis, looking for biomarkers using liquid biopsy can provide a more comprehensive management of the disease.

Minimally invasive/non-invasive biomarker approaches are at the forefront of personalized cancer care, which involves customizing diagnostic or prognostic evaluations for the needs of each patient according to the specific features of the cancer and their associated clinical profile (34). Likewise, it has been found that the newer targeted therapy, including inhibitors of *EGFR*, *ALK*, *PI3K/AKT/mTOR*, *RAS-MAPK*, *RET*, *MET*, *BRAF*, and *NTRK/ROS1*, as well as *PD1* and *CTLA4* molecules, for non-small cell lung cancer (NSCLC) have far better treatment responses than the standard therapies (35). Similar strategies such as use of anti-EGFR agents like cetuximab in case of OSCC patients with CTC expressing EGFR may be adopted in the management of OSCC (36).

4.1.1 Types of biomarkers in OSCC

4.1.1.1 Based on the nature of the molecule

Different biological fluids such as blood, saliva, pleural fluid, urine, and cerebrospinal fluid (CSF) can be analyzed to study a wide

array of tumour-derived moieties in the form of biomarkers for OSCC (33). For OSCC, saliva and serum are the most reliable and explored biological fluids for the study of biomarkers. Saliva is the closest contact fluid to the tumour in OSCC and contains an array of analytes such as cytokines, enzymes, and antibodies, which makes it an appropriate indicative body fluid for biomarker analysis and, hence, can be a useful alternative to serum and tissue testing (37). In addition, its accessibility, ease of collection and non-invasive nature make it one of the most extensively used fluids for the investigation of biomarkers in OSCC (38).

Serum, on the other hand, is also widely explored as it contains a pool of biomolecules that have been shed off from the site of the tumour into the circulation, which can be analyzed to get clinically valuable information on both the patient and their underlying malignancy (39). The biomarkers obtained from these biological fluids vary in nature depending upon the type of modifications, such as genomic, epigenomic, transcriptomic, proteomic and cellular.

4.1.1.1.1 Genomic, epigenomic and transcriptomic biomarkers. As is widely known, cancer develops from different types of alterations in DNA, modifying their structure to a great extent. These modifications can be at small scale, such as DNA base insertions, substitution and deletion or large-scale rearrangements in DNA, such as gene duplications/deletions, chromosomal inversions/translocations and loss of heterozygosity (40). Hence, profiling the tumour can help understand the genomic alterations and epigenomic regulations that have influenced the growth and development of cancer.

Alterations in the genome due to a single base pair, such as a point mutation at tumour suppressor gene p53 exon 4 codon 63, have been proven to be significantly associated with the pathophysiology of OSCC and ought to be a good diagnostic biomarker (41). Also, it was reported that the karyotypes of OSCC patients are composed of multiple numerical and structural abnormalities (42). In OSCC patients, heterogeneity in the saliva at the DNA microsatellite level has been correlated with their tumour specimen, and ctDNA isolated from the saliva has also been shown as an early diagnostic biomarker (43). In one of the studies by our group, genomic instability found in tumour DNA isolated from the saliva of OSCC patients was shown to act as a predictive biomarker for response to treatment by radiotherapy (44). Furthermore, DNA methylation also played a major role and can be used as a powerful diagnostic approach to scrutinize OSCC patients from healthy controls. Ferlazzo et al. and Demokan et al., individually in their respective studies, showed a higher frequency of p16, MGMT and K1F1A, and EDNRB promoter methylation in the saliva of their OSCC patients' groups than healthy controls (45, 46).

Methylation of *SHOX2* and *SEPT9* in cfDNA derived from serum samples of HNSCC patients has shown promising diagnostic potential (47). Mydlarz et al., in their study on OSCC patients, concluded that hypermethylation of cfDNA at *EDNRB* gene derived from serum has a role in the detection of OSCC patients with 100% specificity, however with low sensitivity (48). Various studies have also found that levels of cfDNA have a

direct correlation with disease severity. Higher cfDNA in tumour samples from locally advanced HNSCC patients had proportionally gross tumour volume (49). Also, in another group of 121 OSCC patients, a higher plasma level of cfDNA was related to poor prognosis (50). In OSCC, several non-coding RNA family members, such as long non-coding (lnc)RNA and micro RNA (miRNA) and circular RNAs (circRNAs) from various body fluids, such as blood saliva, are found to be associated with disease severity. It was found in the literature that lncRNA such as BLACAT1, AC104041.1, ADAMTS9-AS2, ANRIL, and BANCR, isolated from cancerous cells were dysregulated and are involved in the promotion of growth, metastasis, proliferation, invasion, and migration (51-55). A study by Rajan et al. on miRNA expression profiling in OSCC among three external datasets in comparison to their data, published a list of various upregulated miRNA such as miR-196a, miR-196b, miR-155, miR-21, miR-424, and many downregulated miRNA such as miR-345, miR-101, miR-144, miR-204 (56). In OSCC, circRNA_100290 regulate the expression of CDK6, cause G1/S arrest and inhibit proliferation of the cells (57).

4.1.1.1.2 Proteomic biomarkers. Proteins are the biomolecules translated from genes that regulate major cellular processes. Any change in the genes encoding these proteins, any modification in the splicing process or further post-translational modifications of the translated protein such as phosphorylation, glycosylation, acetylation or proteomic cleavage may regulate the protein functions contributing to diverse human proteome (58). These proteins are differentially secreted in response to changes in the cellular microenvironment such as proteins secreted by tumours are differentially expressed compared to normal tissue and further could provide greater insights into cellular physiology and molecular biology, establishing their role as a biomarker (59). To be used as a potential biomarker, individual proteins, as well as the panel of different proteins, show sensitivity and specificity for their application in diagnostics and/or prognostics by different samples, mainly saliva and serum. For instance, actin, myosin (60), resistin (RETN) (61), angiogenic factors and matrix-metalloproteinases (62), transferrin (63) and many more proteins were found to be the potential salivary biomarkers for early diagnosis of OSCC. Besides the individual proteins, panels of proteins, such as four proteins panel; matrix metalloproteinase 1 (MMP1), kininogen 1 (KNG1), annexin A2 (ANXA2), and heat shock protein family A (Hsp70) Member 5 (HSPA5) (64), another panel of three proteins such as interleukin 1 beta, 6 and 8 (65), and AHSG, KRT6C and AZGP1 (66), a panel of five proteins i.e., M2BP, MRP14, CD59, catalase, and profilin (67) have also been explored in saliva of patients for their diagnostic and prognostic potential in OSCC.

Another relevant biological fluid widely explored for proteomics biomarker identification is the serum of OSCC patients. The serum helps to detect systemic response, i.e., the change in the proteins that correspond to the disease pathogenesis. By using serum, several molecules have been tested clinically to be used in cancer detection termed "cancer markers" such as carbohydrates antigen (CA)19-9 and CA125 (68). Various cancer biomarkers such as *CA19-9*, carcinoembryonic

antigen (*CEA*), squamous cell carcinoma antigen (*SCC Ag*) (69), immunosuppressive acidic protein (*IAP*) and cytokeratin 19 fragment (*Cyfra*) (69, 70) have been explored for the diagnosis of oral cancer. But due to insufficient accuracy, these markers could not be applied to all oral cancer patients for clinical purposes (70).

4.1.1.1.3 Cellular biomarkers. Circulating Tumour Cells (CTCs) and tumour-derived extracellular vesicles (EVs) are among the cellular entities that get activated and released at different time points during carcinogenesis. CTCs represent the entire spectrum of clones present within a tumour. Thus, the transcriptomic and genomic profile of these CTCs provide important information about the heterogeneity of tumour of that patient (38).

Since CTCs are directly disseminated from the primary tumour and preserve its heterogeneity, they have been used as diagnostic biomarkers for micrometastasis and have thus gained a lot of attention from the perspective of personalized biomarkers in a number of malignancies, including OSCC (71). Adding on to this fact, recent work by our group found that CTCs are typically found in OSCC patients even without lymph node metastasis. CTCs presence and their number detected in the patient's blood have been found to be associated with poor prognosis and the recurrence of the disease (72, 73). A higher count of CTCs has been linked to advanced-stage disease, higher risk of metastasis, and decreased overall survival rate (74, 75). They have also been used in real-time monitoring of disease progression, even after resection, to study tumour evolution (76).

EVs contain variety of biological contents such as proteins, lipids, DNAs and RNAs which might play important roles in mediating tumour development and progression in OSCC (77). A study by He et al. showed elevated levels of miR-24-3p in salivary exosomes of OSCC patients (78). Another study on tumour-derived EVs by Ruowei et al. found that EVs regulate inflammatory cytokines such as IL-17A-induced signalling pathways to promote tumour progression in OSCC (79).

4.1.1.2 Classification of biomarkers based on their application in disease analysis

Biomarkers have been utilized in various aspects of disease analysis, such as diagnosis of disease, planning therapy, evaluating the effectiveness of treatment, prognosis, response to treatment and many more. Based on their potential application, biomarkers are further categorized as diagnostic, prognostic, predictive, monitoring, response/ pharmacodynamic, safety, and susceptibility/risk, as discussed in Table 1 below (80).

Different approaches in the management of OSCC and their applications in patient care are discussed in Figure 1.

5 Discussion

OSCC is a heterogeneous group of disorders characterized by neoplastic growth originating from the squamous epithelial lining of the oral mucosa. The prevalence of the disease is ever-rising, and a similar trend is seen in the case of disease-associated mortality. Hence, there is an urgent need to understand the existing challenges and limitations associated with the available

| Types of applications | Purpose | Examples | References |
|-------------------------------|--|---|------------|
| Diagnostic | To detect or confirm the presence of a disease or a medical condition of interest. Marks the foundation of precision medicine | Heat shock protein 70 (HSP70): These are network of molecular chaperons with folding catalyst which assist in protein folding. Promote tumorigenesis by caspase dependent pathway mediated apoptosis suppression. Expression of the gene significantly differ between control, leukoplakia and OSCC cases and within OSCC between different histological grades and are associated with progression to advanced stage tumor and nodal positivity. TP53 mutation early sign of HNSCC: TP53 tumour suppressor gene regulate cell growth and division, at the time of errors initiate apoptosis or senescence. Mutation in TP53 cause invasive progression of lesions, a decrease in survival rate and a poor response to cisplatin and fluorouracil based chemoradiation, hence associated with local recurrence after therapy completion | (82, 83) |
| Prognostic | A biomarker used to associate likelihood of a clinical event, disease progression, recurrence of a disease/medical condition In clinical setup, it is routinely used to set trial entry and exclusion criteria for the stratification of high-risk population | NCBP2 and TFRC: High level of these proteins involved in regulating the proliferation, migration and invasion during the course of OSCC | (84) |
| Predictive | • A change in the biomarkers meant to anticipate the response of patients to a treatment or therapy | Ki-67: Nuclear protein, present in highly proliferating cells but disappear in resting phase of a cell.Its level is predictive of relapse free and overall survival | (85) |
| Monitoring | To assess the status of a disease or medical condition, through the course of the illness.Change in the value helps predict status of the disease | Desmoglein-3: Preferentially maintains structural integrity in the oral epithelium. Keratin 13: Involved in regulating the differentiation of cells and play major role in carcinogenesis | (86, 87) |
| Response/ pharmacodynamics | A biomarker whose level changes in response to exposure to a medicine/therapy or an environment agent. Highly useful in clinics and therapeutic development | Filamin-A(FLNA): Plays a major role in organization of extracellular network that assist in exchange of signals, control DNA double strand break response, cell-ECM interactions, cell signalling. High expression of FLNA in the buccal squamous cell carcinoma (BSCC) patient tissue associated with poor survival -regarded as a novel biomarker for the diagnosis and outcome prediction of OSCC | (88) |
| Susceptibility/risk | A biomarker which can be used to predict the changes of disease development in an apparently healthy individual. Marks the basis of epidemiological studies on risk prediction | Fibrinogen gamma chain: Major function in homeostasis, significantly higher expression in OSCC compared to healthy controls. α-Defensins 1-3: Major constituent of azurophilic granules of neutrophils, not expressed in normal mucosa, their levels rise in physiological states to exhibit innate immune defenses against infectious diseases including epithelial cancers. Acts as an important predictor linking inflammation, angiogenesis and cancer | (89, 90) |

TABLE 1 Types of biomarkers based on their applications (80, 81).

management modalities and what modifications need to be incorporated to improve the outcome. With advancements in technology and a better understanding of the disease, more and more management opportunities focused on the molecular characteristics of cancer cells and the effect of the treatment modality on the patient's disease outcome have become the focus of therapeutics. For proper management of OSCC and enhancing the quality of life of the patient, timely diagnosis and treatment with minimal side effects should be the focus. Liquid biopsybased biomarkers such as cfDNA, non-coding RNAs, proteins and CTCs in the blood, saliva, buccal swabs and other body fluids of the patients are widely being studied for potential use in OSCC. Estimation of the levels of these biomolecules at an early

stage of a disease can help in timely diagnosis and further appropriate management of the disease. At times, the assessment of these biomarkers at regular intervals during the course of the disease management can aid in planning treatment, effectively evaluating the disease, and designing personalized therapy.

On one hand, these biomarkers help in the longitudinal assessment of the disease progression and therapeutics design, but on the other hand, their discovery is also associated with challenges. Most of the biomolecules which have been explored for their biomarker potential are still confined to the research stage and have not been expanded to the clinics due to a lack of sensitivity and specificity as well as technical hurdles such as their identification, standardization, and further validation for clinical



FIGURE 1

Approaches in the diagnosis and prognosis of OSCC: the figure depicting the advancements from traditional and generalised to recent minimally/noninvasive and personalised approach for the early diagnosis and better prognosis of OSCC. TNM, tumour, node, metastasis; CTCs, circulating tumour cells; ctDNA, circulating tumour DNA; EVs, extracellular vesicles.

utility. Moreover, some of these potential markers showing promising results in the early discovery phase failed to reproduce similar successful results during large-scale diagnostic trials.

To attain success in identifying successful biomarkers, including the non-invasive ones, there is a need for the involvement of a multidisciplinary approach comprising experts from clinical oncology, radiology, pathology, and molecular biology specialized in their fields for a collaborative decision-making process, who can harness the available gold standard methods along with these valuable biomarkers and transform the current landscape of OSCC with the best possible solution to the patient.

To summarize, in the era of the ever-growing prevalence of OSCC, prospective use of these biomarkers in the clinical setting will enable early diagnosis and prognosis to enhance the efficacy of cancer management, given that they are highly sensitive, specific and properly validated in large cohorts of patients.

Author contributions

SS: Writing – original draft. GB: Writing – original draft. RK: Writing – original draft. AC: Writing – original draft. SG: Writing – original draft, Writing – review & editing. AP: Conceptualization, Formal Analysis, Funding acquisition, Project administration, Resources, Supervision, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA A Cancer J Clinicians*. (2024) 74(3):229–63.

2. Early Detection, Prevention, and Infections Branch (EPR). (2023). Available online at: https://www.iarc.who.int/branches-epr (cited April 11, 2011).

3. Bagan J, Sarrion G, Jimenez Y. Oral cancer: clinical features. Oral Oncol. (2010) 46 (6):414–7. doi: 10.1016/j.oraloncology.2010.03.009

4. Scully C, Bagan J. Oral squamous cell carcinoma: overview of current understanding of aetiopathogenesis and clinical implications. *Oral Dis.* (2009) 15 (6):388–99. doi: 10.1111/j.1601-0825.2009.01563.x

5. Department of Health. UK Chief Medical Officers' Alcohol Guidelines Review. (2016). Available online at: https://assets.publishing.service.gov.uk/media/ 5a7f51b4e5274a2e87db5206/summary.pdf (Accessed April 11, 2024).

6. Shaw RJ, O'Connell JE, Bajwa M. Basic surgical principles and techniques. In: Warnakulasuriya S, Greenspan JS, editors. *Textbook of Oral Cancer*. Cham: Springer International Publishing (2020). p. 253–82. doi: 10.1007/978-3-030-32316-5_20

7. Hosni A, Chiu K, Huang SH, Xu W, Huang J, Bayley A, et al. Non-operative management for oral cavity carcinoma: definitive radiation therapy as a potential alternative treatment approach. *Radiother Oncol.* (2021) 154:70–5. doi: 10.1016/j. radonc.2020.08.013

8. Schoenfeld JD, Hanna GJ, Jo VY, Rawal B, Chen YH, Catalano PS, et al. Neoadjuvant nivolumab or nivolumab plus ipilimumab in untreated oral cavity squamous cell carcinoma: a phase 2 open-label randomized clinical trial. *JAMA Oncol.* (2020) 6(10):1563. doi: 10.1001/jamaoncol.2020.2955

9. Ru L, Zheng J. Clinical applications and perspectives of immune checkpoint inhibitors in oral squamous cell carcinoma. *Oncologie*. (2024). doi: 10.1515/ oncologie-2024-0086/html

10. Jin L, Lamster I, Greenspan J, Pitts N, Scully C, Warnakulasuriya S. Global burden of oral diseases: emerging concepts, management and interplay with systemic health. *Oral Dis.* (2016) 22(7):609–19. doi: 10.1111/odi.12428

11. Badwelan M, Muaddi H, Ahmed A, Lee KT, Tran SD. Oral squamous cell carcinoma and concomitant primary tumours, what do we know? A review of the literature. *Curr Onco*. (2023) 30(4):3721–34. doi: 10.3390/curroncol30040283

12. Frangioni JV. New technologies for human cancer imaging. JCO. (2008) 26 (24):4012-21. doi: 10.1200/JCO.2007.14.3065

13. Fass L. Imaging and cancer: a review. Mol Oncol. (2008) 2(2):115-52. doi: 10. 1016/j.molonc.2008.04.001

14. Raza F, Zafar H, You X, Khan A, Wu J, Ge L. Cancer nanomedicine: focus on recent developments and self-assembled peptide nanocarriers. *J Mater Chem B.* (2019) 7(48):7639–55. doi: 10.1039/C9TB01842E

15. Walter W, Pfarr N, Meggendorfer M, Jost P, Haferlach T, Weichert W. Nextgeneration diagnostics for precision oncology: preanalytical considerations, technical challenges, and available technologies. *Semin Cancer Biol.* (2022) 84:3–15. doi: 10. 1016/j.semcancer.2020.10.015

16. Tettero JM, Freeman S, Buecklein V, Venditti A, Maurillo L, Kern W, et al. Technical aspects of flow cytometry-based measurable residual disease quantification in acute myeloid leukemia: experience of the European LeukemiaNet MRD working party. *Hemasphere*. (2022) 6(1):e676. doi: 10.1097/HS9.00000000000676

17. Nguyen TTH, Sodnom-Ish B, Choi SW, Jung HI, Cho J, Hwang I, et al. Salivary biomarkers in oral squamous cell carcinoma. *J Korean Assoc Oral Maxillofac Surg.* (2020) 46(5):301–12. doi: 10.5125/jkaoms.2020.46.5.301

18. Yu T, Wood RE, Tenenbaum HC. Delays in diagnosis of head and neck cancers. J Can Dent Assoc. (2008) 74(1):61.

19. Llewellyn CD, Johnson NW, Warnakulasuriya S. Factors associated with delay in presentation among younger patients with oral cancer. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod*. (2004) 97(6):707–13. doi: 10.1016/j.tripleo.2004.01.007

20. Blatt S, Krüger M, Sagheb K, Barth M, Kämmerer PW, Al-Nawas B, et al. Tumor recurrence and follow-up intervals in oral squamous cell carcinoma. *J Clin Med.* (2022) 11(23):7061. doi: 10.3390/jcm11237061

21. Lugano R, Ramachandran M, Dimberg A. Tumor angiogenesis: causes, consequences, challenges and opportunities. *Cell Mol Life Sci.* (2020) 77(9):1745–70. doi: 10.1007/s00018-019-03351-7

22. Ashouri A, Zhang C, Gaiti F. Decoding cancer evolution: integrating genetic and non-genetic insights. *Genes (Basel)*. (2023) 14(10):1856. doi: 10.3390/genes14101856

23. Zhu L, Jiang M, Wang H, Sun H, Zhu J, Zhao W, et al. A narrative review of tumour heterogeneity and challenges to tumour drug therapy. *Ann Transl Med.* (2021) 9(16):1351. doi: 10.21037/atm-21-1948

24. Definition of Biomarker—NCI Dictionary of Cancer Terms—NCI. (2011). Available online at: https://www.cancer.gov/publications/dictionaries/cancer-terms/ def/biomarker (cited April 11, 2024).

25. Sarhadi VK, Armengol G. Molecular biomarkers in cancer. *Biomolecules*. (2022) 12(8):1021. doi: 10.3390/biom12081021

26. Henry NL, Hayes DF. Cancer biomarkers. Mol Oncol. (2012) 6(2):140-6. doi: 10. 1016/j.molonc.2012.01.010

27. Disease I of M (US) C on Q of B and SE in C, Micheel CM, Ball JR. *Evaluation of Biomarkers and Surrogate Endpoints in Chronic Disease*. USA: National Academies Press (2010). Available online at: https://www.ncbi.nlm.nih.gov/books/NBK220297/ (cited 2024 April 11).

28. Hayes DF, Sauerbrei W, McShane LM. REMARK guidelines for tumour biomarker study reporting: a remarkable history. *Br J Cancer*. (2023) 128(3):443–5. doi: 10.1038/s41416-022-02046-4

29. Perakis S, Speicher MR. Emerging concepts in liquid biopsies. *BMC Med.* (2017) 15(1):75. doi: 10.1186/s12916-017-0840-6

30. Siravegna G, Marsoni S, Siena S, Bardelli A. Integrating liquid biopsies into the management of cancer. *Nat Rev Clin Oncol.* (2017) 14(9):531-48. doi: 10.1038/ nrclinonc.2017.14

31. Crowley E, Di Nicolantonio F, Loupakis F, Bardelli A. Liquid biopsy: monitoring cancer-genetics in the blood. *Nat Rev Clin Oncol.* (2013) 10(8):472-84. doi: 10.1038/nrclinonc.2013.110

32. Pantel K, Alix-Panabières C. Circulating tumour cells in cancer patients: challenges and perspectives. *Trends Mol Med.* (2010) 16(9):398–406. doi: 10.1016/j. molmed.2010.07.001

33. Lone SN, Nisar S, Masoodi T, Singh M, Rizwan A, Hashem S, et al. Liquid biopsy: a step closer to transform diagnosis, prognosis and future of cancer treatments. *Mol Cancer*. (2022) 21(1):79. doi: 10.1186/s12943-022-01543-7

34. La Thangue NB, Kerr DJ. Predictive biomarkers: a paradigm shift towards personalized cancer medicine. *Nat Rev Clin Oncol.* (2011) 8(10):587–96. doi: 10. 1038/nrclinonc.2011.121

35. Araghi M, Mannani R, Heidarnejad maleki A, Hamidi A, Rostami S, Safa SH, et al. Recent advances in non-small cell lung cancer targeted therapy; an update review. *Cancer Cell Int.* (2023) 23(1):162. doi: 10.1186/s12935-023-02990-y

36. Kiss F, Pohóczky K, Görbe A, Dembrovszky F, Kiss S, Hegyi P, et al. Addition of epidermal growth factor receptor inhibitors to standard chemotherapy increases survival of advanced head and neck squamous cell carcinoma patients: a systematic review and meta-analysis. *Oral Dis.* (2023) 29(5):1905–19. doi: 10.1111/ odi.14228

37. Yoshizawa JM, Schafer CA, Schafer JJ, Farrell JJ, Paster BJ, Wong DTW. Salivary biomarkers: toward future clinical and diagnostic utilities. *Clin Microbiol Rev.* (2013) 26(4):781–91. doi: 10.1128/CMR.00021-13

38. Armakolas A, Kotsari M, Koskinas J. Liquid biopsies, novel approaches and future directions. *Cancers (Basel).* (2023) 15(5):1579. doi: 10.3390/cancers15051579

39. Huang X, Duijf PHG, Sriram S, Perera G, Vasani S, Kenny L, et al. Circulating tumour DNA alterations: emerging biomarker in head and neck squamous cell carcinoma. *J Biomed Sci.* (2023) 30(1):65. doi: 10.1186/s12929-023-00953-z

40. Stadler ZK, Thom P, Robson ME, Weitzel JN, Kauff ND, Hurley KE, et al. Genome-wide association studies of cancer. *J Clin Oncol.* (2010) 28(27):4255–67. doi: 10.1200/JCO.2009.25.7816

41. Liao PH, Chang YC, Huang MF, Tai KW, Chou MY. Mutation of p53 gene codon 63 in saliva as a molecular marker for oral squamous cell carcinomas. *Oral Oncol.* (2000) 36(3):272-6. doi: 10.1016/S1368-8375(00)00005-1

42. Cowan JM. Cytogenetics in head and neck cancer. Otolaryngol Clin North Am. (1992) 25(5):1073–87. doi: 10.1016/S0030-6665(20)30926-9

43. El-Naggar AK, Mao L, Staerkel G, Coombes MM, Tucker SL, Luna MA, et al. Genetic heterogeneity in saliva from patients with oral squamous carcinomas: implications in molecular diagnosis and screening. *J Mol Diagn*. (2001) 3(4):164–70. doi: 10.1016/S1525-1578(10)60668-X

44. Boora GS, Chauhan A, Kaur R, Kaur H, Chandel J, Bhat SA, et al. Genomic instability detected from the saliva of head and neck squamous cell carcinoma patients: association with clinical implications. *Arch Oral Biol.* (2022) 137:105395. doi: 10.1016/j.archoralbio.2022.105395

45. Ferlazzo N, Currò M, Zinellu A, Caccamo D, Isola G, Ventura V, et al. Influence of MTHFR genetic background on p16 and MGMT methylation in oral squamous cell cancer. *Int J Mol Sci.* (2017) 18(4):724. doi: 10.3390/ijms18040724

46. Demokan S, Chang X, Chuang A, Mydlarz WK, Kaur J, Huang P, et al. KIF1A and EDNRB are differentially methylated in primary HNSCC and salivary rinses. *Int J Cancer.* (2010) 127(10):2351–9. doi: 10.1002/ijc.25248

47. Schröck A, Leisse A, de Vos L, Gevensleben H, Dröge F, Franzen A, et al. Freecirculating methylated DNA in blood for diagnosis, staging, prognosis, and monitoring of head and neck squamous cell carcinoma patients: an observational prospective cohort study. *Clin Chem.* (2017) 63(7):1288–96. doi: 10.1373/clinchem. 2016.270207

48. Mydlarz WK, Hennessey PT, Wang H, Carvalho AL, Califano JA. Serum biomarkers for detection of head and neck squamous cell carcinoma. *Head Neck*. (2015) 38(1):9–14. doi: 10.1002/hed.23842

49. Hilke FJ, Muyas F, Admard J, Kootz B, Nann D, Welz S, et al. Dynamics of cellfree tumour DNA correlate with treatment response of head and neck cancer patients receiving radiochemotherapy. *Radiother Oncol.* (2020) 151:182–9. doi: 10.1016/j. radonc.2020.07.027

50. Lin LH, Chang KW, Kao SY, Cheng HW, Liu CJ. Increased plasma circulating cell-free DNA could be a potential marker for oral cancer. *IJMS*. (2018) 19(11):3303. doi: 10.3390/ijms19113303

51. Gou C, Han P, Li J, Gao L, Ji X, Dong F, et al. Knockdown of lncRNA BLACAT1 enhances radiosensitivity of head and neck squamous cell carcinoma cells by regulating PSEN1. *Br J Radiol.* (2020) 93(1108):20190154. doi: 10.1259/bjr.20190154

52. Li M, Ding X, Zhang Y, Li X, Zhou H, Yang L, et al. Antisense oligonucleotides targeting lncRNA AC104041.1 induces antitumour activity through Wnt2B/ β -catenin pathway in head and neck squamous cell carcinomas. *Cell Death Dis.* (2020) 11(8):672. doi: 10.1038/s41419-020-02820-3

53. Li Y, Wan Q, Wang W, Mai L, Sha L, Mashrah M, et al. LncRNA ADAMTS9-AS2 promotes tongue squamous cell carcinoma proliferation, migration and EMT via the miR-600/EZH2 axis. *Biomed Pharmacother*. (2019) 112:108719. doi: 10.1016/j. biopha.2019.108719

54. Zhang D, Ding L, Li Y, Ren J, Shi G, Wang Y, et al. Midkine derived from cancer- associated fibroblasts promotes cisplatin-resistance via up-regulation of the expression of lncRNA ANRIL in tumour cells. *Sci Rep.* (2017) 7(1):16231. doi: 10. 1038/s41598-017-13431-y

55. Yao C, Kong F, Zhang S, Wang G, She P, Zhang Q. Long non-coding RNA BANCR promotes proliferation and migration in oral squamous cell carcinoma via MAPK signaling pathway. *J Oral Pathol Med.* (2021) 50(3):308–15. doi: 10.1111/jop.12968

56. Rajan C, Roshan VGD, Khan I, Manasa VG, Himal I, Kattoor J, et al. MiRNA expression profiling and emergence of new prognostic signature for oral squamous cell carcinoma. *Sci Rep.* (2021) 11(1):7298. doi: 10.1038/s41598-021-86316-w

57. Chen L, Zhang S, Wu J, Cui J, Zhong L, Zeng L, et al. circRNA_100290 plays a role in oral cancer by functioning as a sponge of the miR-29 family. *Oncogene*. (2017) 36(32):4551-61. doi: 10.1038/onc.2017.89

58. Banks RE, Dunn MJ, Hochstrasser DF, Sanchez JC, Blackstock W, Pappin DJ, et al. Proteomics: new perspectives, new biomedical opportunities. *Lancet.* (2000) 356(9243):1749–56. doi: 10.1016/S0140-6736(00)03214-1

59. Pillai J, Chincholkar T, Dixit R, Pandey M. A systematic review of proteomic biomarkers in oral squamous cell cancer. *World J Surg Onc.* (2021) 19(1):315. doi: 10.1186/s12957-021-02423-y

60. De Jong EP, Xie H, Onsongo G, Stone MD, Chen XB, Kooren JA, et al. Quantitative proteomics reveals myosin and actin as promising saliva biomarkers for distinguishing pre-malignant and malignant oral lesions. *PLoS One.* (2010) 5(6): e11148. doi: 10.1371/journal.pone.0011148

61. Wu C, Chu H, Hsu C, Chang K, Liu H. Saliva proteome profiling reveals potential salivary biomarkers for detection of oral cavity squamous cell carcinoma. *Proteomics.* (2015) 15(19):3394–404. doi: 10.1002/pmic.201500157

62. Cai M, Zheng Z, Bai Z, Ouyang K, Wu Q, Xu S, et al. Overexpression of angiogenic factors and matrix metalloproteinases in the saliva of oral squamous cell carcinoma patients: potential non-invasive diagnostic and therapeutic biomarkers. *BMC Cancer.* (2022) 22(1):530. doi: 10.1186/s12885-022-09630-0

63. Jou YJ, Lin CD, Lai CH, Chen CH, Kao JY, Chen SY, et al. Proteomic identification of salivary transferrin as a biomarker for early detection of oral cancer. *Anal Chim Acta*. (2010) 681(1-2):41-8. doi: 10.1016/j.aca.2010.09.030

64. Yu JS, Chen YT, Chiang WF, Hsiao YC, Chu LJ, See LC, et al. Saliva protein biomarkers to detect oral squamous cell carcinoma in a high-risk population in Taiwan. *Proc Natl Acad Sci U S A.* (2016) 113(41):11549–54. doi: 10.1073/pnas.1612368113

65. Piyarathne NS, Rasnayake RMSGK, Angammana R, Chandrasekera P, Ramachandra S, Weerasekera M, et al. Diagnostic salivary biomarkers in oral cancer and oral potentially malignant disorders and their relationships to risk factors—a systematic review. *Expert Rev Mol Diagn*. (2021) 21(8):789–807. doi: 10. 1080/14737159.2021.1944106

66. Jain A, Kotimoole CN, Ghoshal S, Bakshi J, Chatterjee A, Prasad TSK, et al. Identification of potential salivary biomarker panels for oral squamous cell carcinoma. *Sci Rep.* (2021) 11(1):3365. doi: 10.1038/s41598-021-82635-0

67. Hu S, Arellano M, Boontheung P, Wang J, Zhou H, Jiang J, et al. Salivary proteomics for oral cancer biomarker discovery. *Clin Cancer Res.* (2008) 14 (19):6246–52. doi: 10.1158/1078-0432.CCR-07-5037

68. Shitrit D, Zingerman B, Shitrit ABG, Shlomi D, Kramer MR. Diagnostic value of CYFRA 21-1, CEA, CA 19-9, CA 15-3, and CA 125 assays in pleural effusions: analysis of 116 cases and review of the literature. *Oncologist.* (2005) 10(7):501–7. doi: 10.1634/ theoncologist.10-7-501

69. Yuan C, Yang K, Tang H, Chen D. Diagnostic values of serum tumor markers Cyfra21-1, SCCAg, ferritin, CEA. C:a19–9, and AFP in oral/oropharyngeal squamous cell carcinoma. *Onco Targets Ther.* (2016) 9:3381–6. doi: 10.2147/OTT. S105672

70. Kurokawa H, Tsuru S, Okada M, Nakamura T, Kajiyama M. Evaluation of tumor markers in patients with squamous cell carcinoma in the oral cavity. *Int J Oral Maxillofac Surg.* (1993) 22(1):35–8. doi: 10.1016/S0901-5027(05)80353-4

71. Ju S, Chen C, Zhang J, Xu L, Zhang X, Li Z, et al. Detection of circulating tumour cells: opportunities and challenges. *Biomark Res.* (2022) 10(1):58. doi: 10.1186/s40364-022-00403-2

72. Chauhan A, Pal A, Sachdeva M, Boora GS, Parsana M, Bakshi J, et al. A FACS-based novel isolation technique identifies heterogeneous CTCs in oral squamous cell carcinoma. *Front Oncol.* (2024) 14:1269211. doi: 10.3389/fonc.2024. 1269211

73. Curtin J, Choi SW, Thomson PJ, Lam AKY. Characterization and clinicopathological significance of circulating tumour cells in patients with oral squamous cell carcinoma. *Int J Oral Maxillofac Surg.* (2022) 51(3):289–99. doi: 10. 1016/j.ijom.2021.05.020

74. Capuozzo M, Ferrara F, Santorsola M, Zovi A, Ottaiano A. Circulating tumour cells as predictive and prognostic biomarkers in solid tumours. *Cells.* (2023) 12 (22):2590. doi: 10.3390/cells12222590

75. Nguyen TNA, Huang PS, Chu PY, Hsieh CH, Wu MH. Recent progress in enhanced cancer diagnosis, prognosis, and monitoring using a combined analysis of the number of circulating tumour cells (CTCs) and other clinical parameters. *Cancers (Basel).* (2023) 15(22):5372. doi: 10.3390/cancers15225372

76. Radhakrishnan V, Kaifi JT, Suvilesh KN. Circulating tumour cells: how far have we come with mining these seeds of metastasis? *Cancers (Basel)*. (2024) 16(4):816. doi: 10.3390/cancers16040816

77. Zhang Y, Liu J, Liu S, Yu L, Liu S, Li M, et al. Extracellular vesicles in oral squamous cell carcinoma: current progress and future prospect. *Front Bioeng Biotechnol.* (2023) 11:1149662. doi: 10.3389/fbioe.2023.1149662

78. He L, Ping F, Fan Z, Zhang C, Deng M, Cheng B, et al. Salivary exosomal miR-24-3p serves as a potential detective biomarker for oral squamous cell carcinoma screening. *Biomed Pharmacother*. (2020) 121:109553. doi: 10.1016/j.biopha.2019. 109553

79. Li R, Zhou Y, Zhang M, Xie R, Duan N, Liu H, et al. Oral squamous cell carcinomaderived EVs promote tumour progression by regulating inflammatory cytokines and the IL-17A-induced signaling pathway. *Int Immunopharmacol.* (2023) 118(110094). doi: 10. 1016/j.intimp.2023.110094

80. Califf RM. Biomarker definitions and their applications. *Exp Biol Med (Maywood)*. (2018) 243(3):213–21. doi: 10.1177/1535370217750088

81. Pozzi L. *7 Types of Biomarkers*. Sweden: Atlas Antibodies AB. Available online at: https://www.atlasantibodies.com/knowledge-hub/blog/7-types-of-biomarkers/? language=en_GB 75 (cited April 11, 2024).

82. Elhendawy HA. Clinical implications of heat shock protein 70 in oral carcinogenesis and prediction of progression and recurrence in oral squamous cell carcinoma patients: a retrospective clinicopathological study. *Eur J Med Res.* (2023) 28(1):464. doi: 10.1186/s40001-023-01433-8

83. Rizzo G, Black M, Mymryk J, Barrett J, Nichols A. Defining the genomic landscape of head and neck cancers through next-generation sequencing. *Oral Dis.* (2015) 21(1). doi: 10.1111/odi.12246

84. Arora R, Haynes L, Kumar M, McNeil R, Ashkani J, Nakoneshny SC, et al. NCBP2 And TFRC are novel prognostic biomarkers in oral squamous cell carcinoma. *Cancer Gene Ther.* (2023) 30(5):752–65. doi: 10.1038/s41417-022-00578-8

85. Lopes VKM, Jesus AS de, Souza LL de, Miyahara LAN, Guimarães DM, Pontes HAR, et al. Ki-67 protein predicts survival in oral squamous carcinoma cells: an immunohistochemical study. *Braz Oral Res.* (2017) 31:e66. doi: 10.1590/1807-3107BOR-2017.vol31.0066

86. Patel V, Hood BL, Molinolo AA, Lee NH, Conrads TP, Braisted JC, et al. Proteomic analysis of laser-captured paraffin-embedded tissues: a molecular portrait of head and neck cancer progression. *Clin Cancer Res.* (2008) 14(4):1002–14. doi: 10.1158/1078-0432.CCR-07-1497

87. Enokida T, Fujii S, Takahashi M, Higuchi Y, Nomura S, Wakasugi T, et al. Gene expression profiling to predict recurrence of advanced squamous cell carcinoma of the tongue: discovery and external validation. *Oncotarget.* (2017) 8(37):61786–99. doi: 10. 18632/oncotarget.18692

88. Liu W, Zeng L, Li N, Wang F, Jiang C, Guo F, et al. Quantitative proteomic analysis for novel biomarkers of buccal squamous cell carcinoma arising in background of oral submucous fibrosis. *BMC Cancer.* (2016) 16:584. doi: 10.1186/s12885-016-2650-1

89. Katakura A, Yamamoto N, Sakuma T, Sugahara K, Onda T, Noguchi S, et al. A screening test for oral cancer using saliva samples: proteomic analysis of biomarkers in whole saliva. *J Oral Maxillofac Surg Med Pathol.* (2015) 27(1):1–5. doi: 10.1016/j. ajoms.2013.05.013

90. Roesch-Ely M, Nees M, Karsai S, Ruess A, Bogumil R, Warnken U, et al. Proteomic analysis reveals successive aberrations in protein expression from healthy mucosa to invasive head and neck cancer. *Oncogene.* (2007) 26(1):54–64. doi: 10. 1038/sj.onc.1209770