Check for updates

OPEN ACCESS

EDITED BY Alan Wayne, Children's Hospital of Los Angeles, United States

REVIEWED BY

Ajay Gupta, University at Buffalo, United States Mary Wedekind, National Cancer Institute (NIH), United States Bruce Pawel, Children's Hospital of Los Angeles, United States

*CORRESPONDENCE

Mona M. Mohammed mona.mohammed@57357.org Hanafy A. Hafez hhafez75@cu.edu.eg

RECEIVED 14 August 2024 ACCEPTED 30 September 2024 PUBLISHED 29 October 2024

CITATION

Mohammed MM, Hafez HA, Elnadi EM, Salama AI, Abd Elaziz AS, Ahmed GT, ELwakeel MA, Kamal MK, Kieran MW and Elhaddad AM (2024) Patterns of expression of VEGFR2, PDGFRs and c-Kit in pediatric patients with high grade nonrhabdomyosarcoma soft tissue sarcoma. *Front. Oncol.* 14:1480773. doi: 10.3389/fonc.2024.1480773

COPYRIGHT

© 2024 Mohammed, Hafez, Elnadi, Salama, Abd Elaziz, Ahmed, ELwakeel, Kamal, Kieran and Elhaddad. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Patterns of expression of VEGFR2, PDGFRs and c-Kit in pediatric patients with high grade non-rhabdomyosarcoma soft tissue sarcoma

Mona M. Mohammed^{1*}, Hanafy A. Hafez^{1,2*}, Enas M. Elnadi^{1,3}, Asmaa I. Salama^{4,5}, Abd Elaziz Saad Abd Elaziz⁴, Gehad T. Ahmed^{6,7}, Madeeha A. ELwakeel^{8,9}, Mohamed K. Kamal¹⁰, Mark W. Kieran¹ and Alaa M. Elhaddad¹

¹Department of Pediatric Oncology, Children Cancer Hospital (57357), Cairo, Egypt, ²Department of Pediatric Oncology, National Cancer Institute, Cairo University, Cairo, Egypt, ³Department of Pediatric Oncology, Beni-Swief University, Beni-swief, Egypt, ⁴Department of Pathology, Children Cancer Hospital (57357), Cairo, Egypt, ⁹Department of Pathology, National Cancer Institute, Cairo University, Cairo, Egypt, ⁶Department of Surgical Oncology, Children Cancer Hospital (57357), Cairo, Egypt, ⁷Department of General Surgery, Faculty of Medicine, Helwan University, Cairo, Egypt, ⁸Department of Radio-Diagnosis, Children Cancer Hospital (57357), Cairo, Egypt, Cairo, Egypt, ⁹Department of Radio-Diagnosis, National Cancer Institute, Cairo University, Cairo, Egypt, ¹⁰Department of Clinical Research, Children Cancer Hospital (57357), Cairo, Egypt

Introduction: Activated vascular endothelial growth factor receptors (VEGFRs), platelet-derived growth factor receptors (PDGFRs) and c-Kit have been shown to be involved in the growth, invasion and metastasis of non-rhabdomyosarcoma soft tissue sarcoma tumor (NRSTS) with promising results for targeted therapy. Our aim was to assess the expression of these markers among different histological types and correlate with outcomes.

Material and methods: This retrospective study included pediatric patients aged \leq 18 years diagnosed with high-grade NRSTS who were treated at Children Cancer Hospital Egypt 57357 as per the COG NRSTS protocol (ARST0332). Expression of VEGFR2, PDGFRs (α and β) and c-Kit in tumor tissue was assessed by immunohistochemistry and correlated with clinical outcome.

Results: Of 113 patients, 96 were eligible for the analysis with a median age of 11 years. Overall, 32.3% demonstrated high expression of PDGFR α , 17.7% for PDGFR β , 19.8% for VEGFR2 and 8.3% exhibited positive expression for c-kit on the tumor cells. Most cases of synovial sarcoma (45.8%) and 43.7% of patients with undifferentiated sarcoma exhibited high expression of PDGFR α while 41.6% of MPNST showed high expression to PDGFR β . The 5-year overall survival (OS), event free survival and relapse free survival (RFS) for the whole cohort were 59%, 54% and 60% respectively. In univariate analyses, only PDGFR α had a negative prognostic impact on relapse free survival (RFS) (p=0.03). In multivariate analyses, VEGFR2 was found to have a negative prognostic impact for OS (p = 0.02).

Conclusion: Our findings indicated that tyrosine kinase receptors are upregulated in NRSTS and exhibited a distinct expression pattern within various subgroups. High expression of VEGFR2 and PDGFR α significantly correlated with reduced survival and may guide targeted therapy approaches for this poor prognosis group of patients.

KEYWORDS

VEGFRs, PDGFRs, c-kit, pediatric, sarcoma, immunohistochemistry

1 Introduction

Non-Rhabdomyosarcoma soft tissue sarcomas (NRSTS) are relatively uncommon cancers, accounting for approximately 3.5% of all childhood tumors. They represent a challenge for treatment due to their diverse biology, requiring treatment that has to be tailored to the specific tissue type (1). Histologic grade is one of the most important prognostic factors in the outcome of soft tissue sarcoma (STS) as patients presenting with high-grade lesions are much more likely to develop metastatic disease resulting in poor survival (2). The optimal therapeutic management requires a multidisciplinary approach adapted to each patient and can include surgical resection alone or in combinations of radiotherapy with or without chemotherapy. Neoadjuvant chemotherapy and radiotherapy have been widely used for patients with advanced soft tissue sarcomas (considered deep seated tumors >5cm, those that are unresectable or those with metastatic disease), with radiological response rate of only 35-40% (3, 4). The role of adjuvant chemotherapy in high-grade NRSTS is still controversial due to the small effect on outcome and the variability of response among different histological subtypes (5). Despite intensive multimodality therapy, patients with high-risk metastatic disease have 5-year event-free survival (EFS) of only 21.2% and overall survival (OS) of 35.5%. Given the limited efficacy of chemotherapy in NRSTS and the poor outcomes of a significant fraction of the patient population, and recognizing that further intensification of cytotoxic chemotherapy is not feasible due to severe toxicity, novel therapeutic approaches are needed (6).

Solid tumors are dependent on the vascular system for growth, invasion and metastasis. Tumor angiogenesis has been shown to support solid tumors mainly by supplying oxygen, glucose and cytokines as well as providing the means to the formation of metastases. A number of growth factors, proteases and cytokines have been reported to have pro-angiogenic effects and to induce tumor angiogenesis (7). Human sarcomas express a number of proangiogenic factors that may represent potential therapeutic targets. Certain tyrosine kinases have been found to be expressed in a range of NRSTS subtypes. The vascular endothelial growth factor receptors (VEGFRs, 1-3), platelet-derived growth factor receptors (PDGFRs, α and β), and c-Kit pathways are among the most commonly dysregulated in soft tissue sarcomas (8). The vascular endothelial growth factors (VEGFs) and their receptors play a distinct role in the growth of tumor vasculature through regulation of endothelial cell proliferation and sprouting which drives angiogenesis and lymphangiogenesis. Signaling through VEGFR2 is considered the major angiogenic pathway, prompting endothelial cells (ECs) to proliferate and form tubes (9). Some studies had correlated high expression of VEGFR2 with decreased survival of various soft tissue tumors (10, 11). The PDGFRs are structurally related tyrosine kinase receptors consisting of either α or β chains and function to activate pericytes which support new microvasculature and play a major role in angiogenesis and regulation of tumor stroma (10). Additionally, PDGFRs (α and β) play a key role in activating the PI3K/Akt/mTOR (phosphatidylinositol 3-phosphate kinase/protein kinase B (PKB)/mammalian Target of Rapamycin) pathway in sarcomas which in turn, initiates a cascade of downstream signaling that promotes cell proliferation and survival, making this pathway a critical target for therapeutic development in sarcomas like mTOR inhibitors (12). Activated VEGFRs and PDGFRs work together to guide the microvasculature into tumor lesions, and are reported to be involved in tumor growth, invasion and metastasis (13). The protooncogene c-Kit (CD117) encodes a transmembrane tyrosine kinase receptor and its expression has been detected in a variety of different tumor entities such as gastrointestinal stromal tumors (GIST), malignant melanoma, breast and lung cancer, sarcoma and mastocytosis (14). Activating mutations in c-Kit play a pivotal role in the pathogenesis of the majority (90-95%) of GIST. The standard diagnostic workup of GIST tumors now includes assessment of c-Kit

Abbreviations: VEGFR2, Vascular endothelial growth factor2; PDGFRs, platelet derived growth factor α ; NRSTS, Non-Rhabdomyosarcoma Soft Tissue Sarcoma Histological Subtypes; OS, overall survivor; RFS, relapse free survivor; STS, soft tissue sarcoma; EFS, event-free survival; VEGF, vascular endothelial growth factors; ECs, endothelial cells; GIST, gastrointestinal stromal tumors; CCHE-57357, Children's Cancer Hospital Egypt-57357; COG, Children's oncology group; WHO, World Health Organization guidelines; FNCLCC, French Fédération Nationale des centres de Lutte Contre le Cancer; IRS, Immunoreactive scor; MPNST, malignant peripheral nerve sheath tumor; G1-3, Grades1-3; ELS, Ewing- like sarcomas.

abnormalities and a number of c-kit inhibitors, such as imatinib, have demonstrated significant activity in tumors with these mutations (15).

Several agents targeting VEGFRs, PDGFRs and c-Kit activity are now being introduced for the treatment of sarcomas with promising results like imatinib, sorafenib, sunitinib, anlotinib and pazopanib (16). The multi-targeted tyrosine kinase inhibitor pazopanib, is a potent inhibitor of these pathways, and has shown potential for synergistic interaction with conventional cytotoxic chemotherapy in preclinical studies, suggesting that these drug combinations might overcome chemo-resistance (17, 18). Pazopanib improved outcomes in adults with advanced soft tissue sarcomas and was approved for single-agent use by the US Food and Drug Administration in adults with advanced soft tissue sarcomas based on results from the phase 3 randomized PALETTE study. In this study, patients treated with pazopanib experienced significantly better progression-free survival compared to those given a placebo (median 4.6 months versus 1.6 months, respectively; hazard ratio = 0.31; P < 0.0001). In addition, Cox models revealed a significant benefit for progression-free survival across all histological subtypes receiving pazopanib (19). The COG ARST1321 study added pazopanib to neoadjuvant chemoradiotherapy, which resulted in increased rates of pathological near complete response, suggesting that this is a highly active combination in children as well as adults with advanced soft tissue sarcomas (20). Although pathological response is still not considered a prognostic factor, and long-term data is lacking, it remains crucial to determine whether a greater pathological response rate will translate into improved local control and survival in NRSTS.

Additionally, recent advancements have identified new therapeutic targets in the treatment of STS, with promising new agents currently under clinical investigation. Monoclonal antibodies such as ramucirumab which specifically VEGFR2 (21) and olaratumab, targets PDGFR α (22) are among these agents. Furthermore, nintedanib, a potent triple angiokinase inhibitor targets PDGFR, VEGFR, and FGFR pathways, has demonstrated significant anti-proliferative effects *in vitro* (23). Despite the recent emergence of these attractive molecular targets, there is limited data available regarding the association between these targets and pediatric NRSTS. To address this gap, this study was designed to assess the expression status of VEGFR2, PDGFRs (α and β) and c-Kit in relation to different histological subtypes of high-grade NRSTS and to estimate the impact of overexpression on treatment outcome to traditional therapies.

2 Patients and methodology

2.1 Patient selection

A total of 113 pediatric patients ≤18 years old with newly diagnosed high-grade NRSTS from January 2013 to January 2020 who were treated at Children Cancer Hospital 57357-Egypt (CCHE-57357) as per the COG NRSTS protocol ARST0332 were considered for inclusion. Of these 113, 96 patients met the eligibility criteria for the study. Of the 17 patients who were excluded, 15 had insufficient

tumor material for analysis, 1 patient died before starting treatment and one patient refused treatment. Demographic and clinical data (histological subtype, tumor site, tumor size and stage) were collected from the medical records. The minimum follow up period was 2 years from the end of treatment. Primary tumor tissue was obtained at diagnosis and histologically subtyped according to the World Health Organization guidelines (WHO 2020) and graded according to the French Fédération Nationale des centres de Lutte Contre le Cancer (FNCLCC) system (24, 25). The institutional review board approved this retrospective study.

2.2 Immunohistochemistry

Formalin-fixed and paraffin-embedded primary tumor specimens (FFPE) were obtained from the archive of the Pathology Departments and four um-thick sections were prepared using a microtome (Leica Biosystems) and mounted on positive charged slides. The antibodies used were as follows: vascular endothelial growth factor receptor2 (VEGFR2, 1:50, rabbit monoclonal, D5B1, Cell Signaling), Platelet derived growth factor receptor alpha (PDGFR α , 1:100, rabbit monoclonal, D13C6, Cell Signaling), Platelet derived growth factor receptor beta (PDGFR β , 1:100, rabbit monoclonal, C82A3, Cell Signaling), C-Kit (Ready to use, rabbit monoclonal CD117, YR145, Cell Marque). The cellular localization for all antibodies was cytoplasmic reaction (see Supplementary Figures 1, 2).

2.3 Immunohistochemistry scoring method

For VEGFR2, PDGFR α and PDGFR β biomarkers, the immune reactive scoring (IRS) method as previously published was used to assess the degree of cytoplasmic intensity (negative, weak, intermediate or strong) and percentage of tumor cells expressing the reaction (26); see Supplementary Table 1. Tumor tissues with IRS negative or weak scores were considered to have low expression while those with moderate or strong IRS scores were considered to have high expression. Assessment of c-Kit expression used a minimum cut off of 10% as previously published (27).

2.4 Statistical analysis

Statistical analyses were performed by R software for statistical computing version 4.2.2. Descriptive statistics were calculated and shown for expression of all biomarkers. The Chi-square test and Fishers Exact test were used to examine the association between marker expression and various clinico-pathological parameters. Univariate and multivariate analyses were performed using Cox's proportional hazard model. In multivariate analyses, factors included site, stage, VEGFR2, PDGFRs (α and β) and c-Kit expression. Overall survival (OS) was calculated from the date of diagnosis until the date of death or last contact in the clinic. Event free survival (EFS) was calculated from the date of diagnosis until the first event, including progression, relapse or death. Relapse free

survival (RFS) was calculated from treatment to the time of relapse. The log rank test was used to detect differences between survival curves for stratified variables. Significance was presented by p value if less than 0.05.

3 Results

3.1 Patient's characteristics

The median age of the analysis cohort (n=96) was 11 years (range 0.05-17 years), with a male to female ratio of 0.8:1. More than 50% of the tumors were located in the trunk 61.5% (n=59) while 38.5% of the tumor originated in the extremities (n=37). Of the 96 patients, tumor size was >5 cm in 71.9% (n=69) and \leq 5 cm in 28.1% (n=27). The most common represented histological subtypes were synovial sarcoma (n=24), undifferentiated sarcoma (n=16), malignant peripheral nerve sheath tumor (MPNST) (n=12), epithelioid sarcoma (9) and sarcoma with BCOR genetic alterations (n=7). In accordance with the COG protocol, surgery alone (Arm A) was performed in seven patients (7.3%), surgery followed by radiotherapy (Arm B) in 17.7% (n=17), and surgery followed by adjuvant chemotherapy and radiotherapy (Arm C) in 19.8% (n=19). Fifty-three patients (55.2%) had inoperable tumors, and were treated with neoadjuvant chemotherapy, then surgery followed by adjuvant chemotherapy and radiotherapy (Arm D).

Among the 96 patients, high expression of PDGFR α was demonstrated in 32.3% (n=31) of patients' tissue samples, VEGFR2 in 19.8% (n=19) and PDGFR β in 17.7% (n=17) of cases. Positive expression of c-kit was identified in 8.3% (n=8). Specifically, 45.8% (n=11) of synovial sarcoma cases and 43.7% (n=7) of patients with undifferentiated sarcoma showed high expression of PDGFR α . Furthermore 41.7% (n=5) of patients with MPNST exhibited high expression of PDGFR β . Collectively, expression of one or more of the target markers was documented in 58.4% of synovial sarcoma, 56.3% of undifferentiated sarcoma and 58.3% of MPNST. Additionally, among the seven cases of sarcoma with BCOR genetic alterations, six patients (85.7%) exhibited high expression of various markers (four for c-kit, six for PDGFR α , and two for VEGFR2). Five of the patients with BCOR genetic alterations presented with advanced (III and IV) stage and were treated with neo-adjuvant chemo-radiotherapy with no responses observed. A detailed descriptive analysis of marker expression across different histological subtypes is shown in Table 1.

Among the patients, 53 presented with advanced unresectable disease, (34 with stage III and 19 with stage IV). These patients were treated with neoadjuvant chemotherapy resulting in partial response in 35.8% (n=19) patients, stable disease in 43.5% (n=23), disease progression in 13.2% (n=7) during chemotherapy while four patients (7.5%) succumbed to their disease before achieving local control. When analyzing markers expression in this patient group, high expression of PDGFR α , VEGFR2, PDGFR β and c-kit was exhibited in 37.7% (n=20), 22.6% (n=12), 15% (n=8) and 11.3% (n=6) of tumors, respectively. A correlation with markers expression in patients treated with chemotherapy was not significant, likely due to small number of patients who showed response to chemotherapy.

Correlation between the expression of target markers and the patients' clinic-pathological variables including site (extremities vs. trunk), size (\leq 5cm vs. >5 cm) and stage (stage I+II vs. III vs. IV) of disease revealed no significant differences, summarized in Table 2.

TABLE 1 Descriptive data for tumor expression of c-Kit, VEGFR2, PDGFRα, PDGFRβ between different histological subtypes in the 96 pediatric patients with NRSTS.

Histological subtypes (no)	c-Kit		VEGFR2		PDGFRα		PDGFRβ	
	Negative No. (%)	Positive No. (%)	Low No. (%)	High No. (%)	Low No. (%)	High No. (%)	Low No. (%)	High No. (%)
Synovial Sarcoma (24)	23 (95.8)	1 (4.2)	20 (83.3)	4 (16.7)	13 (54.2)	11 (45.8)	22 (91.7)	2 (8.3)
Undifferentiated Sarcoma (16)	12 (75)	4 (25)	12 (75)	4 (25)	9 (56.3)	7 (43.7)	12 (75)	4 (25)
MPNST (12)	12 (92.3)	-	10 (83.3)	2 (16.7)	11 (91.7)	1 (8.3)	7 (58.3)	5 (41.7)
Epithelioid sarcoma (9)	9 (100)	-	8 (88.9)	1 (11.1)	8 (88.9)	1 (11.1)	6 (66.7)	3 (33.3)
Alveolar soft part sarcoma	6 (100)	-	6 (100)	-	6 (100)	-	6 (100)	-
Sarcoma with BCOR genetic alterations (7)	3 (42.9)	4 (57.1)	5 (72.5)	2 (28.5)	1 (14.3)	6 (85.7)	7 (100)	-
CIC rearranged sarcoma (4)	4 (100)	-	3 (75)	1 (25)	3 (75)	1 (25)	4 (100)	-
Extraskeletal Chondrosarcoma (6)	5 (83.3)	1 (16.7)	5 (83.3)	1 (16.7)	5 (83.3)	1 (16.7)	6 (100)	-
Clear cell sarcoma (5)	5 (100)	-	4 (80)	1 (20)	5 (100)	-	5 (100)	-
Others* (7)	6 (85.7)	1 (14.3)	4 (57.1)	3 (42.9)	4 (57.1)	3 (42.9)	4 (57.1)	3 (42.9)

VEGFR2, vascular endothelial growth factor receptors; PDGFR, platelet-derived growth factor receptors; MPNST, malignant peripheral nerve sheath tumor.

* Others: 1 Leiomyosarcoma, 1 angiosarcoma, 2 Embryonal Sarcoma, 2 Myxoid Liposarcoma, 1 Myxofibrosarcoma.

TABLE 2 Descrip	otive and statisti	cal analysis fo	r the expression	of angiogenic	markers based	on clinic-patho	ological factors	s for the 96 ped	iatric patients w	ith NRSTS.			
	Patient number (%)	c-Kit		P value	VEGFR2		P value	PDGFR0.		P value	PDGFRB		P value
Clinico- pathological variables no (%)	Total n=96 (100%)	Negative No (%)	Positive No (%)		Low No (%)	High No (%)		Low No (%)	High No (%)		Low No (%)	High No (%)	
Site Trunk Extremities	59 (61.5) 37 (38.5)	53 (89.8) 35 (94.5)	6 (10.2) 2 (5.5)	0.48	48 (81.4) 29 (78.4)	11 (18.6) 8 (21.6)	0.79	41 (69.5) 24 (64.9)	18 (30.5) 13 (35.1)	0.66	49 (83) 30 (81)	10 (17) 7 (19)	0.79
Size ≤5 cm >5 cm	27 (28.1) 69 (71.9	24 (88.9) 64 (92.8)	3(11.1) 5 (7.2)	0.68	20 (74) 57 (82.6)	7 (26) 12 (17.4)	0.4	19 (70.4) 46 (66.7)	8 (29.6) 23 (33.3)	0.81	22 (81.5) 57 (82.6)	5 (18.5) 12 (17.4)	1
TNM stage I+II III IV	23 (24) 54 (56.2) 19 (19.8)	21 (91.3) 50 (92.6) 17 (89.5)	$\begin{array}{c} 2 & (8.7) \\ 4 & (17.4) \\ 2 & (10.5) \end{array}$	1	$\begin{array}{c} 17 \ (74) \\ 47 \ (87) \\ 13 \ (68.4) \end{array}$	6 (26) 7 (13) 6 (31.6)	0.38	15 (65.2) 36 (66.7) 14 (73.7)	8 (34.8) 18 (33.3) 5 (26.3)	0.8	18 (78.3) 41 (75.9) 18 (94.7)	$5 (21.7) \\ 11 (24.1) \\ 1 (5.3)$	0.54
VEGFR2, vascular en	dothelial growth fac	tor receptors; PD	GFR, platelet-derived	d growth factor rec	ceptors; TNM, Tun	nor size; Nodal invo	olvement, Metastas	sis.					

3.2 Survival outcome

The median follow-up period was 71.06 months (range 52.86-90.15) with a 5-year OS, EFS and RFS for the whole patient cohort of 59%, 54% and 60% respectively (95% CI) (Figures 1A-C). At the time of data analysis, 54 patients were alive, 39 patients had disease progression and died while only three of the 96 patients were lost to follow-up.

The results suggest that tumors located in the trunk have a negative prognostic impact on OS and EFS compared to those in the extremities. Specifically, OS rates were 48% for trunk tumors versus (vs) 77% for extremity tumors (P<0.01), and EFS rates were 40% for trunk tumors vs 75% for extremity tumors (P<0.01). Regarding RFS, the rates were 49% for trunk tumors and 76% for extremity tumors, with a p value of 0.05, almost reaching statistical significance. For tumor size, larger tumors> 5 cm had poorer outcome versus tumors \leq 5 cm in size for OS, EFS and RFS with rates of 48% vs 89% (P<0.01), 43% vs 80% (P<0.01), and 49% vs 82% (P=0.02) respectively. Additionally, advanced tumor stage was associated with inferior survival outcome for stages I+II vs III vs IV, where OS 91% vs 64% vs 11% (P<0.01), EFS 81% vs 57% vs 11% (P<0.01), RFS 80% vs 59% vs 20% (P<0.01) respectively (Table 3). In univariate analysis, the impact of angiogenic markers on survival outcome was only observed in tumors with high expression of PDGFRa in which a significant reduction in RFS was observed versus patients with low expression (43% vs 70%, CI 95%, P=0.03). None of the other markers had an impact on OS or EFS. The impact of clinico-pathological factors and target markers on survival are summarized in Table 3.

3.3 Multivariate analysis

In multivariate analyses of OS and EFS including site, stage, VEGFR2, PDGFRs (α and β) and c-Kit, tumor stage III-IV (*P*<0.01) and high expression of VEGFR2 (p=0.02), were significant independent prognostic indicators for OS. The other markers including c-Kit (p=0.13), PDGFRa (p=0.9), PDGFRB (p=0.88) and tumors in the trunk site (P=0.05) failed to demonstrate a prognostic impact. Regarding EFS, only tumor site (p=0.015) reached statistical significance while stage III-IV (p=0.05), c-Kit (p=0.16), VEGFR2 (0.29), PDGFR α (p=0.0.25) or PDGFR β expression (p=0.8) failed to reach statistical significance. Results of the multivariate analyses for OS and EFS are presented in Tables 4 and 5 respectively.

4 Discussion

Angiogenesis is important for tumor growth and metastasis. Extensive research on angiogenesis in patents with STS has discovered a series of pro-angiogenic factors that can have direct or indirect influence on tumor angiogenesis (7). Among these, the VEGF receptors, platelet-derived growth factor receptors, and c-Kit are the most commonly dysregulated pathways in STS (28). Effective novel targeted therapies that disrupt the crosstalk between stroma and tumor cells are a promising strategy for cancer treatment (29). Previous retrospective studies in STS have



TABLE 3 Prognostic relevance of clinicopathological variables and angiogenic markers for OS, EFS, RFS in 96 pediatric patients with highgrade NRSTS.

Characters:	Patient number (%)	5-year OS (%)	P value	5-year EFS (%)	P value	5-year RFS (%)	P value			
Site			0.009		0.004		0.05			
Trunk	59 (61.5)	48		40		49				
Extremities	37 (38.5)	77		75		76				
Size			< 0.001		< 0.001		0.02			
≤5 cm	27 (28.1)	89		80		82				
>5 cm	69 (71.9	48		43		49				
TNM stage			< 0.001		<0.001		<0.001			
I+II	23 (24)	91		81		80				
III	54 (56.2)	64		57		59				
IV	19 (19.8)	11		11		20				
Markers expression										
c-Kit			0.4		0.5		0.9			
negative	88 (91.7)	58		53		60				
positive	8 (8.3)	73		56		55				
VEGFR2			0.3		0.7		0.8			
Low	77 (80.2)	62		54		60				
High	19 (19.8)	49		51		62				
PDGFRa			0.9		0.4		0.03			
Low	65 (67.7)	61		56		70				
High	31 (32.3)	56		47		43				
PDGFRβ			0.5		0.7		0.3			
Low	79 (82.3)	61		56		63				
High	17 (17.7)	49		44		47				

OS, overall survivor; EFS, event free survivor; RFS, relapse free survival; VEGFR2, vascular endothelial growth factor receptors; PDGFR, platelet-derived growth factor receptors; TNM, Tumor size, Nodal involvement, Metastasis.

TABLE	4	Muli	ivari	ate	сох	regr	ression	for	OS	including	variables	site,
stage,	c-K	it, V	EGFR	2, 1	PDG	FRα,	PDGF	Rβ.				

Factor	Hazard ratio	Confidence interval (95%)	P value
Site Extremities trunk	2	0.98-4.2	0.05
TNM Stage I-II III-IV	7.2	1.68-31.1	0.008
VEGFR2 Low High	2.6	1.15-5.8	0.02
PDGFRα Low high	1	0.52-2.1	0.9
PDGFRβ Low high	1.1	0.47-2.4	0.8

OS, overall survival; VEGFR2, vascular endothelial growth factor receptors; PDGFR, plateletderived growth factor receptors; TNM, Tumor size, Nodal involvement, Metastasis.

consistently demonstrated a significant correlation between high expression of angiogenic markers and higher-grade sarcomas. However, the majority of these studies primarily included NRSTS of all grades (G1-G3) and focused on adult populations, with limited numbers of pediatric cases owing to the rarity of the disease in this age group (10, 30, 31). Considering that NRSTS are a very heterogeneous group of diseases and the expression of the markers are variable among different histological subtypes, we analyzed the prevalence of VEGFR2, PDGFRs (α and β) and c-Kit in a well-characterized group of pediatric patients with high-grade NRSTS, aiming to identify biomarkers in distinct subgroups

TABLE 5 Multivariate cox regression for EFS including variables site, stage, c-Kit, VEGFR2, PDGFR α , PDGFR β .

Factor	Hazard ratio	Confidence interval (95%)	P value
Site Extremities trunk	2.3	1.18-4.6	0.015
TNM Stage I-II III-IV	2.4	1.00-6.0	0.051
VEGFR2 Low High	1.5	0.7-3.3	0.2
PDGFRα Low high	1.42	0.7-2.6	0.8
PDGFRβ Low high	0.95	0.4-2.0	0.8

EFS, event free survival; VEGFR2, vascular endothelial growth factor receptors; PDGFR, platelet-derived growth factor receptors; TNM, Tumor size, Nodal involvement, Metastasis.

that might benefit from a more individualized therapy approach. In 2015, Kampmann et al. investigated the expression of VEGFRs (1–3) and PDGFRs (α and β) in 275 adult patients with grade 2 and 3 soft tissue sarcoma using immunohistochemistry. Their findings demonstrated that high expression of VEGFR2 (p = 0.032) was an independent poor predictor of long-term survival. By contrast, this effect was not observed for PDGFRs α or β expression (10). Moreover, Kilvaer and coworkers published a study on 181 patients (12 patients were younger than 20 years old) with STS. They demonstrated the influence of VEGFRs on recurrence-free survival, metastasis free survival and disease-specific survival on a variety of subgroups defined by tumor site (31).

In addition to these studies, further investigations have explored these pathways across various histological subtypes, including malignant peripheral nerve sheath tumors (MPNST) and synovial sarcoma. Notably, a study by Perrone et al. demonstrated upstream activation of PDGFR (α and β) and EGFR (endothelial growth factor receptor), alongside downstream RTK signaling activation in MPNST. These findings suggest that combined inhibition of RTK and mTOR pathways could be an effective therapeutic approach for MPNST (32).

Furthermore, a previously published study involving 255 pediatric and adult STS patients examined the differential expression of PDGF ligands and receptors across sarcoma subtypes. The results revealed a significant correlation between PDGF α expression and the risk of metastatic relapse (P = 0.006), further indicating that the expression levels of these ligands and receptors are associated with sarcoma patient outcomes. These findings highlight their potential role as biomarkers for predicting the efficacy of PDGFR α -targeted therapies within this heterogeneous disease group (33).

In line with the findings of the previous studies, we identified VEGFR2 high expression exhibited a negative prognostic impact on OS (P=0.02), while high PDGFR α expression had a negative prognostic impact on RFS (P=0.03) in pediatric patients. Overall, our results emphasize the significance of both VEGFR2 and PDGFR α as prognostic markers of survival, while highlighting the need for additional studies to clarify the role of PDGFR β in this context. Additionally, for c-Kit, immunostaining is a well-established diagnostic tool for GISTs and Imatinib has been shown to be a selective inhibitor of the tyrosine kinase activity of c-Kit (15). There have been occasional reports of its expression in a limited number of soft tissue tumors beyond GISTs (34, 35) and this is supported by the results presented in this study.

In our cohort, we observed IHC staining for c-Kit in a notably restricted number of soft tissue tumors, while no instance of clear cell sarcoma, MPNST, epithelioid sarcoma and alveolar soft part sarcoma exhibited c-Kit positivity. The significance of c-Kit positive staining on patients' outcome is unclear as this did not impact outcome in the limited patient population in this series. This is in concordance with the finding of Hornick and Fletcher who conducted a study of 365 soft tissue sarcoma of different histological subtypes assessing c-Kit expression using IHC staining. Their results indicated that only 22 cases were c-kit positive and none were identified in patients with clear cell sarcoma, MPNST, epithelioid sarcoma or alveolar soft part sarcoma (36). Potti et al. studied the expression of c-Kit in 90 patients with STS with a mean age at diagnosis of 56.9 years. Only 20/90 patients showed overexpression of c-Kit and there was no impact on outcome (34). Along with the studies mentioned above, our data shows the limited utility of c-Kit immunostaining as a prognostic marker in certain subtypes of STS, emphasizing the importance of comprehensive diagnostic approaches tailored to the specific histological characteristics of each tumor subtype. Whether targeting of c-kit can add to the activity of existing therapy in this small subgroup of patients remains to be determined.

When evaluating the expression of the markers among different histological subtypes, distinct patterns emerged. For instance, synovial sarcoma and undifferentiated sarcoma showed high expression for one or more of the markers with predominant high expression of PDGFRa in 58.4% and 56.2% of cases respectively. In cases of MPNST, 58.3% of case showed predominant high expression of PDGFRB. Additionally, more than 55% of patients with epithelioid sarcoma exhibited high marker expression with PDGFRB. Furthermore, we investigated seven cases of sarcoma with BCOR genetic alterations and four patients with CIC-rearranged sarcoma, which are new subsets of rare sarcomas know with poor prognosis, previously known as "Ewing- like sarcomas" (ELS), and now distinguished from Ewing sarcoma in the recent WHO classification, 2020 (24). A multiinstitutional European retrospective analysis conducted by Sparber-Sauer et al. focused on young patients (0-24 years) with CIC-fused (n=31) and BCOR-rearranged (n=29) soft tissue sarcomas. They reported that despite local control achieved by surgery, patients experienced dismal outcomes due to disease progression or relapse. Three-year event-free survival were 44% and 41.2% for CIC and BCOR groups, respectively. Similarly, three- year overall survivals were 46.3% and 67.1% respectively. The study concluded that pediatric patients often present with large tumors and metastatic disease with dismal overall outcome and highlights the need for new treatment options, especially for CIC sarcomas, which often present with advanced and metastatic disease (37).

Interestingly, in our study, among the seven cases of sarcoma with BCOR genetic alterations, six patients (85.7%) exhibited high expression of various markers, five of them presented in advanced stage deemed unfit for upfront resection and four patients succumbed due to disease progression or relapse.

Regarding CIC-rearranged sarcoma, the outcomes observed in our four patients highlight the aggressive nature of this disease, with the three patients presenting with advanced stage succumbing to disease progression or relapse. Through the analysis of markers expression, two patient exhibited high expression of markers, one for VEGFR2 and the other for PDGFR α .

These findings highlight the necessity for further investigation into molecular angiogenic marker expression in these tumors, which are recognized for their poor prognosis. Such studies can help tailor treatment strategies to individual patients based on their specific molecular profiles, ultimately improving treatment outcomes in these challenging tumors. These findings also suggest that comprehensive studies involving larger cohorts of patients, both adult and pediatric) with the rarer BCOR genetic alterations or CIC-rearranged sarcoma are needed. Incorporating a broader range of tyrosine kinase markers are essential to validate and extend these initial findings and establish robust associations between marker expression profiles and clinical outcomes. Understanding the expression patterns of angiogenic markers between different histological subtypes can provide valuable insights into the underlying mechanisms driving tumor progression, especially for those with poor prognosis, and may reveal potential therapeutic targets. This information could be crucial in predicting the response to multi-tyrosine kinase inhibitors such as pazopanib, which are now being incorporated into studies for advanced unresected cases of NRSTS. The integration of tyrosine kinase inhibitors (e.g. pazopanib, sorafenib) alongside chemotherapy regimens holds promise for augmenting local disease control and potentially enhancing treatment outcomes.

Nonetheless, the validation of these preliminary observations necessitates a comprehensive analysis involving a larger pediatric patient cohort. Such efforts will be indispensable for guiding future therapeutic strategies in the management of advanced stage sarcoma.

The major weakness of this study, which is common in pediatric sarcoma studies, is the heterogeneity of the sarcoma population. Even with a relatively large sample cohort with regard to pediatric high-grade NRSTSs, the numbers limit meaningful explorations based on histological subgroups, at least with respect to multivariate analysis.

In conclusion, we demonstrate that tyrosine kinase receptors are upregulated in sarcomas and show a distinct expression pattern in particular subgroups. High expression of VEGFR2 and PDGFR α significantly correlated with reduced patient survival in NRSTS and the identification of these protein signatures suggests that multi-tyrosine kinase inhibitor therapy might be a promising avenue to treat those patients. However, further investigations involving larger cohorts are warranted to explore the functional role of different markers and their impact on treatment outcome in different histological subtypes. This will be critical for informing the development of targeted therapeutic approaches tailored to the specific molecular characteristics of sarcomas, ultimately improving patient care and clinical outcomes.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding authors.

Ethics statement

The study was conducted in accordance with local legislation and institutional requirements and was approved by institutional review board and scientific Advisory Committee at the Children's Cancer Hospital Egypt (CCHE-57357).

Author contributions

MM: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Validation, Visualization, Writing – original draft, Writing – review & editing. HH: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Resources, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. EE: Conceptualization, Writing – review & editing. AS: Data curation, Formal analysis, Investigation, Methodology, Supervision, Validation, Writing – review & editing. AA: Data curation, Investigation, Methodology, Validation, Writing – review & editing. GA: Writing – review & editing. ME: Writing – review & editing. MoK: Formal analysis, Methodology, Software, Writing – review & editing. MaK: Conceptualization, Data curation, Formal analysis, Investigation, Methodology, Project administration, Software, Supervision, Validation, Visualization, Writing – original draft, Writing – review & editing. AE: Supervision, Validation, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare that no financial support was received for the research, authorship, and/or publication of this article.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

References

1. Gronchi A, Ferrari S, Quagliuolo V, Broto JM, Pousa AL, Grignani G, et al. Histotype-tailored neoadjuvant chemotherapy versus standard chemotherapy in patients with high-risk soft-tissue sarcomas (ISG-STS 1001): an international, open-label, randomised, controlled, phase 3, multicentre trial. *Lancet Oncol.* (2017) 18:812–22. doi: 10.1016/S1470-2045(17)30334-0

2. Mancini BR, Roberts KB. Pediatric non-rhabdomyosarcoma soft tissue sarcomas. J Radiat Oncol. (2013) 2:135–48. doi: 10.1007/s13566-012-0081-4

3. Fletcher CDM K, Unni FM. Pathology and genetics of tumours of soft tissue and bone. *Cancer*. (2002) 177:1365–76 p.

4. Pappo AS, Devidas M, Jenkins J, Rao B, Marcus R, Thomas P, et al. Phase II trial of neoadjuvant vincristine, ifosfamide, and doxorubicin with granulocyte colonystimulating factor support in children and adolescents with advanced-stage nonrhabdomyosarcomatous soft tissue sarcomas: A pediatric oncology group study. J Clin Oncol. (2005) 23:4031–8. doi: 10.1200/JCO.2005.03.209

5. Casali PG, Picci P. Adjuvant chemotherapy for soft tissue sarcoma. Curr Opin Oncol. (2005) 17:361-5. doi: 10.1097/01.cco.0000166652.15546.4f

6. Spunt SL, Million L, Chi YY, Anderson J, Merck C, Wales N, et al. Prospective analysis of a risk-based treatment strategy for non-rhabdomyosarcoma soft tissue sarcomas in patients under 30 years of age: A report from children's oncology group study ARST0332 HHS public access. *Lancet Oncol.* (2020) 21:145–61. doi: 10.1016/S1470-2045(19)30672-2

7. Gacche RN, Meshram RJ. Angiogenic factors as potential drug target: Efficacy and limitations of anti-angiogenic therapy. *Biochim Biophys Acta - Rev Cancer*. (2014) 1846:161–79. doi: 10.1016/j.bbcan.2014.05.002

8. Weiss AR, Chen YL, Scharschmidt TJ, Chi YY, Tian J, Black JO, et al. Pathological response in children and adults with large unresected intermediate-grade or high-grade soft tissue sarcoma receiving preoperative chemoradiotherapy with or without pazopanib (ARST1321): a multicentre, randomised, open-label, phase 2 trial. *Lancet Oncol.* (2020) 21:1110–22. doi: 10.1016/S1470-2045(20)30325-9

9. Ahmad A, Nawaz MI. Molecular mechanism of VEGF and its role in pathological angiogenesis. J Cell Biochem. (2022) 123:1938-65. doi: 10.1002/jcb.v123.12

10. Kampmann E, Altendorf-Hofmann A, Gibis S, Lindner LH, Issels R, Kirchner T, et al. VEGFR2 predicts decreased patients survival in soft tissue sarcomas. *Pathol Res Pract.* (2015) 211:726–30. doi: 10.1016/j.prp.2015.04.015

11. Tamura R, Sato M, Morimoto Y, Ohara K, Kosugi K, Oishi Y, et al. Quantitative assessment and clinical relevance of VEGFRs-positive tumor cells in refractory brain tumors. *Exp Mol Pathol.* (2020) 114:104408. doi: 10.1016/j.yexmp.2020.104408

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2024.1480773/ full#supplementary-material.

SUPPLEMENTARY FIGURE 1

high power field showing: (A) Negative expression for Vascular endothelial growth factor2 (VEGFR2). (B) Positive expression for VEGFR2. (C) Negative expression for c-Kit. (D) Positive expression for c-Kit.

SUPPLEMENTARY FIGURE 2

high power field showing: (A) Negative expression for platelet derived growth factor α (PDGFR α). (B) Positive expression for PDGFR α . (C) Negative expression for PDGFR β . (D) Positive expression for PDGFR β .

12. Ho AL, Deraje Vasudeva S, Laé M, Saito T, Barbashina V, Antonescu CR, et al. PDGF receptor alpha is an alternative mediator of rapamycin-induced Akt activation: implications for combination targeted therapy of synovial sarcoma. *Cancer Res.* (2012) 72:4515–25. doi: 10.1158/0008-5472.CAN-12-1319

13. Blume-Jensen P, Hunter T. Oncogenic kinase signalling. Nature. (2001) 411:355-65. doi: 10.1038/35077225

14. Medinger M, Kleinschmidt M, Mross K, Wehmeyer B, Unger C, Schaefer HE, et al. C-kit (CD117) expression in human tumors and its prognostic value: An immunohistochemical analysis. *Pathol Oncol Res.* (2010) 16:295–301. doi: 10.1007/s12253-010-9247-9

15. Rubin BP, Singer S, Tsao C, Duensing A, Lux ML, Ruiz R, et al. KIT activation is a ubiquitous feature of gastrointestinal stromal tumors. *Cancer Res.* (2001) 61:8118–21.

16. Shen G, Zheng F, Ren D, Du F, Dong Q, Wang Z, et al. Anlotinib: A novel multitargeting tyrosine kinase inhibitor in clinical development. *J Hematol Oncol.* (2018) 11:1–12. doi: 10.1186/s13045-018-0664-7

17. Hashimoto K, Man S, Xu P, Cruz-Munoz W, Tang T, Kumar R, et al. Potent preclinical impact of metronomic low-dose oral topotecan combined with the antiangiogenic drug pazopanib for the treatment of ovarian cancer. *Mol Cancer Ther.* (2010) 9:996–1006. doi: 10.1158/1535-7163.MCT-09-0960

18. Li Y, Yang X, Su LJ, Flaig TW. Pazopanib synergizes with docetaxel in the treatment of bladder cancer cells. *Urology*. (2011) 78:233-40. doi: 10.1016/ j.urology.2011.02.041

19. Léon Bérard C, van der Graaf WT A, Blay JY, Chawla SP, Kim DW, Bui-Nguyen B, et al. Pazopanib for metastatic soft-tissue sarcoma (PALETTE): a randomised, double-blind, placebo-controlled phase 3 trial. *Lancet*. (2012) 379:1879–86. doi: 10.1016/S0140-6736(12)60651-5

20. Weiss AR, Chen YL, Scharschmidt TJ, Chi YY, Tian J, Black JO, et al. Pathologic response in children and adults with large unresected intermediate-or high-grade soft tissue sarcoma receiving preoperative chemoradiation with or without pazopanib (ARST1321; PAZNTIS): a multicentre, randomised, open-label, phase 2 trial HHS Public Access. *Lancet Oncol.* (2020) 21:1110–22. doi: 10.1016/S1470-2045(20)30325-9

21. Landgraf Oholendt A, Zadlo JL, Oholendt AL. Ramucirumab: A new therapy for advanced gastric cancer. J Adv Pract Oncol. (2015) 6:71–5. doi: 10.6004/jadpro.2015.6.1.8

22. Lowery CD, Blosser W, Dowless M, Knoche S, Stephens J, Li H, et al. Olaratumab exerts antitumor activity in preclinical models of pediatric bone and soft tissue tumors through inhibition of platelet-Derived growth factor receptor a. *Clin Cancer Res.* (2018) 24:847–57. doi: 10.1158/1078-0432.CCR-17-1258

23. Patwardhan PP, Musi E, Schwartz GK. Preclinical evaluation of nintedanib, a triple angiokinase inhibitor, in soft-tissue sarcoma: potential therapeutic implication for synovial sarcoma. *Mol Cancer Ther.* (2018) 17:2329–40. doi: 10.1158/1535-7163.MCT-18-0319

24. Choi JH, Ro JY. The 2020 WHO classification of tumors of soft tissue: selected changes and new entities. *Adv Anat Pathol.* (2021) 28:44–58. doi: 10.1097/PAP.00000000000284

25. Guillou L, Coindre JM, Bonichon F, Nguyen BB, Terrier P, Collin F, et al. Comparative study of the National Cancer Institute and French Federation of Cancer Centers Sarcoma Group grading systems in a population of 410 adult patients with soft tissue sarcoma. *J Clin Oncol.* (1997) 15:350–62. doi: 10.1200/JCO.1997.15.1.350

26. Specht E, Kaemmerer D, Wirtz RM, Schulz S, Lupp A. Comparison of immunoreactive score, HER2/neu score and H score for the immunohistochemical evaluation of somatostatin receptors in bronchopulmonary neuroendocrine neoplasms. *Histopathology*. (2015) 67:368–77. doi: 10.1111/his.12662

27. Sato O, Wada T, Kawai A, Yamaguchi U, Makimoto A, Kokai Y, et al. Expression of epidermal growth factor receptor, ERBB2 and KIT in adult soft tissue sarcomas A clinicopathologic study of 281 cases. *CANCER*. (2005) 103:9. doi: 10.1002/cncr.20986

28. Le Tourneau C, Faivre S, Raymond E. New developments in multitargeted therapy for patients with solid tumours. *Cancer Treat Rev.* (2008) 34:37-48. doi: 10.1016/j.ctrv.2007.09.003

29. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell.* (2011) 144:646–74. doi: 10.1016/j.cell.2011.02.013

30. Kilvaer TK, Valkov A, Sorbye S, Smeland E, Bremnes RM, Busund LT, et al. Profiling of VEGFs and VEGFRs as prognostic factors in soft tissue sarcoma: VEGFR-3 is an independent predictor of poor prognosis. *PloS One*. (2010) 5(12). doi: 10.1371/ journal.pone.0015368 31. Kilvaer TK, Smeland E, Valkov A, Sorbye SW, Bremnes RM, Busund LT, et al. The VEGF-and PDGF-family of angiogenic markers have prognostic impact in soft tissue sarcomas arising in the extremities and trunk. *BMC Clin Pathol.* (2014) 14:5. doi: 10.1186/1472-6890-14-5

32. Perrone F, Da Riva L, Orsenigo M, Losa M, Jocollè G, Millefanti C, et al. PDGFRA, PDGFRB, EGFR, and downstream signaling activation in Malignant peripheral nerve sheath tumor. *Neuro Oncol.* (2009) 11:725. doi: 10.1215/15228517-2009-003

33. Brahmi M, Lesluyes T, Dufresne A, Toulmonde M, Italiano A, Mir O, et al. Expression and prognostic significance of PDGF ligands and receptors across soft tissue sarcomas. *ESMO Open.* (2021) 6:100037. doi: 10.1016/j.esmoop.2020.100037

34. Potti A, Ganti AK, Foster H, Knox S, Hebert BJ, Tendulkar K, et al. Immunohistochemical detection of HER-2/neu, c-kit (CD117) and vascular endothelial growth factor (VEGF) overexpression in soft tissue sarcomas. *Anticancer Res.* (2004) 24:333–7.

35. Tsuura Y, Hiraki H, Watanabe K, Suzuki T, Igarashi S, Shimamura K, et al. Preferential localization of c-kit product in tissue mast cells, basal cells of skin, epithelial cells of breast, small cell lung carcinoma and seminoma/dysgerminoma in human: immunohistochemical study on formalin-fixed, paraffin-embedded tissues. *Virchows Arch.* (1994) 424:135–41. doi: 10.1007/BF00193492

36. Hornick JL, Fletcher CDM. Immunohistochemical staining for KIT (CD117) in softTissue sarcomas is very limited in distribution. *Am J Clin Pathol.* (2002) 117:188–93. doi: 10.1309/LX9U-F7P0-UWDH-8Y6R

37. Sparber-Sauer M, Corradini N, Affinita MC, Milano GM, Pierron G, Carton M, et al. Clinical characteristics and outcomes for children, adolescents and young adults with "CIC-fused" or "BCOR-rearranged" soft tissue sarcomas: A multi-institutional European retrospective analysis. *Cancer Med.* (2023) 12:14346–59. doi: 10.1002/cam4.6113