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\*CORRESPONDENCE Yu Han Stochanyu@zzu.edu.cn

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## The emerging role of osteoclasts in the treatment of bone metastases: rationale and recent clinical evidence

Youjun Liu<sup>1</sup>, Huanshi Chen<sup>1</sup>, Tong Chen<sup>1</sup>, Guowen Qiu<sup>1</sup> and Yu Han<sup>2\*</sup>

<sup>1</sup>Department of Spinal Surgery, Liuzhou Municipal Liutie Central Hospital, Liuzhou, China, <sup>2</sup>Department of Orthopedics, The First Affiliated Hospital of Zhengzhou University, Zhengzhou, China

The occurrence of bone metastasis is a grave medical concern that substantially impacts the quality of life in patients with cancer. The precise mechanisms underlying bone metastasis remain unclear despite extensive research efforts, and efficacious therapeutic interventions are currently lacking. The ability of osteoclasts to degrade the bone matrix makes them a crucial factor in the development of bone metastasis. Osteoclasts are implicated in several aspects of bone metastasis, encompassing the formation of premetastatic microenvironment, suppression of the immune system, and reactivation of quiescent tumor cells. Contemporary clinical interventions targeting osteoclasts have proven effective in mitigating bone-related symptoms in patients with cancer. This review comprehensively analyzes the mechanistic involvement of osteoclasts, and explores clinical evidence regarding interventions targeting osteoclasts.

#### KEYWORDS

osteoclasts, bone metastasis, antiresorptive drugs, therapeutic targets, targeted therapy

## 1 Introduction

An epidemiological investigation revealed that individuals diagnosed with bone metastasis have a prevalence rate of 5.1%, corresponding to an estimated annual incidence of approximately 18.8 cases per 100 000 bone metastasis diagnoses in the United States from 2010 to 2015 (1). Among individuals aged  $\geq$ 25 years, lung cancer has the highest prevalence as the main site for *de novo* bone metastases, with a rate of 8.7 cases per 100 000 diagnoses in 2015, followed by prostate and breast primaries, with rates of 3.19 and 2.38 cases per 100 000 diagnoses, respectively (1). In particular, patients with breast cancer have a bone metastasis risk of 73%, whereas those with prostate cancer have a risk of 68% (2). Furthermore, patients

with lung cancer have a bone metastasis risk of 30%-40%, whereas those with thyroid cancer have a risk of 60% (3). Malignant tumors often spread in bone tissues, with the bone microenvironment commonly considered a pivotal element in the advancement of bone metastasis. The bone microenvironment comprises a diverse array of cellular entities and a complex extracellular matrix (ECM) (4). The intricate interactions between these constituents and tumor cells contribute to the development of bone metastases. The process of bone metastasis development is complex and well-organized, involving several steps. First, a premetastatic microenvironment is established as a breeding ground for seeding disseminated tumor cells (DTCs). Second, DTCs are extravasated from the circulation to settle within the premetastatic microenvironment. Third, DTCs inhabit the bone niche and become dormant to avoid immune surveillance and antitumor therapy. Finally, DTCs are reactivated from the dormant state to develop into clinically detectable metastases (5). The metastasis of cancer cells to the skeletal system can result in the degradation of bone tissue, onset of pain, and an increased susceptibility to fractures (6). The abovementioned process can be considered a pathological mechanism of bone remodeling, involving a delicate balance between osteoclasts and osteoblasts, ultimately leading to bone restructuring (4).

Osteoclasts have been recognized as a potentially effective therapeutic target in various pathological conditions characterized by bone resorption, including osteoporosis and bone metastasis. Several clinical guidelines have recommended drugs targeting osteoclasts as a primary therapeutic approach for bone destructive diseases (7). Systemic antiresorptive drugs that inhibit osteoclasts can provide the desired symptomatic relief. Furthermore, administering bisphosphonates and denosumab can significantly diminish the occurrence of skeletal-related events (SREs) and mitigate the distress caused by pathological bone resorption, particularly in individuals with osteolytic bone metastases (8, 9). Even in prostate cancer cases where osteoblastic lesions are predominant with a concurrent osteolytic component, antiresorptive medications have been found to effectively mitigate the symptoms associated with bone metastases (10). Nevertheless, antiresorptive medications are currently utilized solely as adjuvant therapies in clinical settings.

Osteoclasts contribute significantly to the progression of bone metastasis, as they facilitate the formation of the premetastatic microenvironment, modulate immunity to promote tumor cell immune evasion, and stimulate the proliferation of dormant tumor cells (Figure 1). Various potential targets for regulating the



differentiation and maturation of osteoclasts have been proposed in preclinical studies (Figure 2) (11). The efficacy of corresponding inhibitors or monoclonal antibodies for these targets against bone metastases has been demonstrated in animal models (12). However, most clinical trials have yielded inconclusive results regarding the effectiveness of osteoclast-targeted interventions in the treatment or prevention of bone metastases, with the exception of postmenopausal patients with breast cancer (13-15). The precise mechanism underlying bone metastasis remains elusive, and the precise involvement of osteoclasts in this process remains unclear. Recent single-cell studies have revealed that the heterogeneity of osteoclast phenotypes is significantly greater than previously assumed (16). These knowledge gaps may explain the differences between preclinical research findings and their clinical translations. Thus, this review aimed to summarize recent research findings regarding the precise mechanisms of osteoclasts in bone metastasis. Additionally, the review focused on recent advancements in identifying therapeutic targets for osteoclasts and analyze the current status of clinical trials involving relevant drugs.

# 2 Roles of osteoclasts in bone metastasis

#### 2.1 Fostering the premetastatic niche

The premetastatic microenvironment refers to the structural and physicochemical alterations in distant organs that facilitate the metastatic expansion of a tumor before tumor cell colonization (17). Numerous mechanisms within bone tissue promote the establishment of a premetastatic microenvironment, both in the normal and pathological states (18). As bone metastases are mostly osteolytic, osteoclasts and osteoclastogenesis are considered significant in the development of the premetastatic niche. The activation of osteoclasts by external stimuli triggers a series of reactions involving vascular modifications, immunological modulation, and metabolic alterations (19, 20). Notably, previous research has revealed the presence of osteoclast-mediated osteolytic foci preceding the metastasis of neoplastic cells to osseous tissue (21). Furthermore, besides their involvement in bone tissue, osteoclast precursors and monocytes in the circulation and primary tumor sites can induce peripheral osteoclastogenesis, facilitating the progression of bone metastases through multiple pathways.

The development of bone metastases in breast cancer cases, characterized by osteolytic changes, is frequently facilitated by the manipulation of the premetastatic microenvironment by osteoclasts. According to a previous study, breast cancer cells with a propensity for bone metastasis can secrete S100A4 protein. This protein has been found to play a significant role in promoting osteolysis by directly stimulating osteoclast formation through the surface receptor RAGE. Breast cancer cells can also release R-spondin 2 and receptor activator of nuclear factor kappa-B ligand (RANKL), which are essential proteins for attracting osteoclast precursors and fostering the development of the osteoclastic premetastatic niche (22). Furthermore, tumor cells can control osteoclast precursors, as evidenced by a study demonstrating that HCC cells secreting lectin galactoside-binding soluble 3(LGALS3) can induce the fusion and podosome formation of osteoclasts through the CD98–integrin  $\alpha\nu\beta$ 3



complex, resulting in osteolytic bone remodeling (23). However, the premetastatic bone microenvironment responds differently to LGALS3 released by different tumor cells. LGALS3 can enhance osteoclast differentiation in breast cancer, whereas in prostate cancer, it may only affect osteoblast development without affecting osteoclast differentiation.

In addition to protein secretion, neoplastic cells can discharge exosomes and microvesicles that carry diverse pro-oncogenic molecular cargo, thereby impacting bone homeostasis and forming the premetastatic microenvironment. A previous study revealed that SCP28 breast cancer cells release exosomal miR-21, which can bind to programmed cell death 4 in osteoclasts, thereby stimulating osteoclast differentiation and intensifying bone metastasis (21). Our study findings indicate that patients with breast cancer who have developed bone metastases exhibit higher levels of blood exosome miR-21 than those who have not developed this condition. This finding suggests that miR-21 can potentially serve as an early detection marker for breast cancer bone metastases (21). Osteoblastic bone metastases, including those originating from prostate cancer cells, can regulate osteolysis and hinder osteogenesis through exosomes, thereby promoting the advancement of bone metastasis in tumor cells (24). Several studies have demonstrated that tumor-derived extracellular vesicles containing diverse microRNAs (miRNAs), including miR-152-3p, miR-378a-3p, miR-214-3p, and miR-325-3p, can modulate osteoclast differentiation (25-28). Furthermore, research has shown that exosomal miRNAs are potential indicators of the progression of bone metastases (29). miRNAs hold significant potential in the advancement of novel diagnostic techniques and therapeutic interventions for bone metastases. Previous study has revealed that MRX34, an miR-34 mimic currently undergoing clinical trials for treating hepatocellular carcinoma, protects against the development of osteolytic disease in individuals with metastatic breast cancer (30). However, the use of miRNAs as therapeutic agents for bone metastases presents various challenges, such as suboptimal delivery efficacy and undesired immune responses, which necessitate further investigation (29).

In addition to directly acting on osteoclasts, exosomes can be taken up by myeloid cells in the bone marrow, eliciting various premetastatic responses, including stimulation of NF- $\kappa$ B signaling, accelerated development of osteoclasts, and decreased expression of myeloid thrombospondin-1 (31). The activation of osteoclasts induces a series of osteolytic reactions, resulting in an intensified local inflammatory response. Finally, these inflammatory responses induce changes in vascular permeability and promote the extravasation process of DTCs (32).

Various researchers have effectively cultured osteoclasts from diverse bone marrow and circulating myeloid populations, such as monocytes and dendritic cells (DCs), in humans and animals (33). Emerging evidence indicates that the migration of osteoclast precursors away from the bone considerably influences the development of the premetastatic niche in bone metastasis. In the field of bone metastases, the mechanism through which myeloidderived suppressor cells (MDSCs) serve as progenitors to osteoclasts has attracted significant attention. The proportion of MDSCs in the peripheral blood of patients with breast cancer is approximately 10 times that of healthy controls (34). A strong correlation exists between heightened numbers of circulating MDSCs and the progression of cancer to an advanced stage, as well as the presence of a positive lymph node status (34). MDSCs can express specific integrins, chemokines, and ECM regulatory factors, thereby exerting influence on vascular permeability, modifying the composition of the ECM, recruiting various cell types, suppressing immune responses, and facilitating the survival of DTCs in the bloodstream as well as their migration to distant metastatic sites (35). Upon infiltration of tumor cells into bone metastases sites, MDSCs undergo differentiation into active osteoclasts, facilitating the expansion of osteoclastic lesions (36). The monocyte-macrophage lineage is a significant progenitor of osteoclasts and is found extensively in the tumor and bone microenvironments, as well as in the circulatory system. Similar to the function of MDSCs, the monocyte-macrophage lineage can not only influence the progression of bone metastases through mechanisms such as immune modulation and vascular permeability alteration but also differentiate into osteoclasts under specific circumstances (37). DCs have been recognized as a plausible origin of osteoclast precursors during inflammation. The differentiation of osteoclasts from DCs may serve as an alternate mechanism for osteoclast formation in the bone premetastatic niche. Splenic CD11c+ DCs were effectively induced in mature and activated multinucleated giant cells expressing TRAP and IL-23 when exposed to conditioned media from breast cancer cells (38).

#### 2.2 Immunosuppressive effects

Once DTCs infiltrate bone tissue, the first challenge is whether they can successfully survive immune system surveillance. Several pieces of evidence have demonstrated the involvement of osteoclasts in the immunological regulation of bone metastases. The *in vitro* expansion of natural killer (NK) cells using osteoclasts from patients with cancer resulted in a significant reduction in the quantity and cytotoxicity of NK cells. However, introducing osteoclasts from healthy individuals restored the cytotoxic activity of NK cells (39, 40). The dissimilarities observed in osteoclasts from individuals with tumors and those without tumors suggest that osteoclasts can attenuate immune responses under pathological conditions, thereby preventing the immune system from effectively targeting tumor cells. This section summarizes how osteoclasts contribute to the immunosuppressive impact of bone metastases, including the direct and indirect regulation of cytotoxicity.

The abovementioned clinical finding supports the theory that osteoclasts are immune-competent cells. Multiple *in vitro* investigations have suggested that osteoclasts can directly influence the state of T or NK cells through cell-contactindependent mechanisms. Osteoclasts can directly suppress proliferation and induce apoptosis of T cells by secreting Galectin-9 and CD200, creating an immunosuppressive environment in several patients with myeloma (41). *In vitro* empirical investigations have further confirmed that osteoclasts secrete various cytokines that mitigate the activation, proliferation, cytotoxic activity, and apoptosis of T cells (42). The inactivation of T cells can facilitate osteoclast formation in bone metastases, thereby exacerbating the vicious cyclic progression of bone metastases (43). In addition to directly affecting T cell function, osteoclasts can influence immune activity within bone metastases by regulating the expression of immune checkpoint molecules. A prominent illustration of this phenomenon is the capacity of osteoclasts to facilitate an immunosuppressive environment by secreting a proliferation-induced ligand (APRIL), resulting in enhanced programmed cell death ligand-1(PD-L1) expression in multiple myeloma cells (41). Several studies have established a strong correlation between the expression of immune checkpoints in bone metastases and osteoclasts (44, 45). Furthermore, the regulation of osteoclast activity may be influenced by immune checkpoint expression (41).

Osteoclasts can establish an immunosuppressive microenvironment by interacting with immunomodulatory and immunosuppressive cells, in addition to directly influencing the activity of T and NK cells. Furthermore, osteoclasts contribute to the immunological tolerance of malignancies by inducing regulatory T cell responses (19). Bone marrow-derived inflammatory osteoclasts (Cx3cr1+) secrete immunosuppressive cytokines and polarize CD4+ T cells into immunosuppressive CD4+ Foxp3+ regulatory T cells in an antigendependent manner (46). Another study revealed that PD-L1 is an essential component of the immunosuppressive capacity of Cx3cr1+ osteoclasts and that Cx3cr1 can distinguish between two different subsets of osteoclasts, each with a distinct role in the immune system (47). Moreover, osteoclasts can attract and activate naïve CD8 T cells, resulting in the transcription of CD25 and Foxp3 genes (48).

MDSCs are widely recognized as immunosuppressive cells that can suppress T cell function. As precursors of immature myeloid cells, MDSCs can differentiate into macrophages and osteoclasts, enabling the regulation of bone resorption under various pathological conditions (49). Consequently, they are considered osteoclast precursors. A previous revealed that the accumulation of MDSCs in bone metastases may decrease the effectiveness of bisphosphonates (50). The interaction between MDSCs and osteoclasts holds relevance in the context of bone metastasis (51). However, the existing literature on the modulation of MDSCs by osteoclasts to facilitate immune evasion in bone metastases is limited, necessitating further investigation.

However, certain studies have proposed that osteoclasts can potentially enhance immune responses. As antigen-presenting cells, osteoclasts express both Class I and Class II of the major histocompatibility complex (MHC). Furthermore, they can take up soluble antigens, facilitating the presentation of allogeneic antigens, which, in turn, induce the activation of CD4+ and CD8+ alloreactive T cells in a manner restricted by the MHC (52). However, there is no supporting evidence for the alteration of antigen-presenting activity in osteoclasts throughout bone metastasis. Bone metastases alter the antigen-presenting capacity of DCs, which are osteoclast precursors. In breast cancer animal models, the percentage of plasmacytoid DCs (pDCs) that infiltrated bone metastases was significantly higher, but their antigen-presenting ability was considerably diminished compared to other DCs subtypes (53). Furthermore, the cytotoxic activity of T cells was significantly restored following pDCs depletion.

#### 2.3 Reactivating dormant tumor cells

Metastases may occur at any time, even after the surgical removal of the primary tumor. This delayed metastasis, also known as relapse, is attributed to mechanisms that maintain DTCs in a quiescent or latent state where they cannot proliferate (referred to as "tumor dormancy") (54). The dormancy of malignant cells can be reversed or transitioned between "on" and "off" states by specific signals from the surrounding microenvironment (55). The interaction between cancer cells and their bone microenvironment plays a critical role in regulating cellular dormancy and reactivation. Research has suggested that osteoblasts can induce metastatic tumor cells into a dormant state by secreting growth factors, thereby preventing their elimination via antitumor therapies (56). Understanding the mechanisms underlying tumor dormancy and its reactivation is crucial in preventing metastatic advancement and extending metastasis-free survival. However, the molecular and cellular processes responsible for activating dormant tumor cells remain poorly understood.

A subset of DTCs are located in the endosteal niche, where they enter a quiescent state characterized by G0-G1 cell cycle arrest (55). Some of these DTCs remain in a quiescent state, whereas others continue to multiply and form micrometastases. However, actively proliferating DTCs can be eliminated by the immune system or antitumor therapies, whereas dormant DTCs can persist in the endosteal niche with the potential for reactivation, which can lead to disease relapse (10). Once DTCs reach the bone tissue, they compete with HSCs for the osteoblastic niche and eventually colonize the compartment previously occupied by HSCs (57). Additionally, osteoblasts protect dormant DTCs from environmental stress stimuli through paracrine and juxtacrine secretion (58). Recent studies using single-cell level analysis have further supported the notion that osteoblasts play an essential role in inducing and protecting dormant DTCs, both in vitro and in vivo (59-61). For prostate tumors, the production of Wnt5a in the osteoblastic niche can activate noncanonical ROR2/SIAH2 signaling and induce dormancy in DTCs (62). Recent gene array analyses have identified GDF10 and TGF2 as osteoblast-secreted proteins that induce quiescence in certain prostate tumor cell lines, indicating varying responses to dormancy across prostate cancer cells (60). Furthermore, breast cancer cells interacting with spindleshaped N-Cadherin+ osteoblasts are recognized and induced into a dormant state through a Notch2-dependent process (63).

The transition from a dormant to active state is a critical event in tumor relapse and metastasis. Osteoclasts play a crucial role in bone remodeling and can directly reactivate dormant DTCs by modifying the endosteal niche. For example, myeloma cells can become dormant through direct contact with osteoblasts in the bone marrow niche, but they can be released from this state when the niche is modified in an osteoclast-dependent manner (61). A longitudinal imaging study using intravital two-photon microscopy revealed that dormant myeloma cells could be reactivated by activating osteoclasts with RANKL (61). Another study demonstrated that the combination of VCAM-1 and integrin  $\alpha$ 4 $\beta$ 1 recruits osteoclast progenitors (OPs), inducing a vicious cycle of bone degradation and tumor progression (64). Suppressing osteoclast activity may reduce the risk of dormant tumor cell reactivation and prevent the development of macrometastatic lesions. Dormancy in cancer cells is a severe clinical issue that directly impacts tumor drug resistance and relapse. Thus, targeting tumor dormancy to prevent tumor metastasis and recurrence is an emerging research area.

## 3 Heterogeneity of osteoclasts and bone metastasis

Bone metastases exhibit a lower incidence in distal extremities but are more prevalent in bones with red marrow and trabecular bone, such as the pelvis, vertebrae, and ribs (5). The underlying mechanisms behind site-specific bone metastases are currently unclear. Some hypotheses suggest that differences in trabecular bone structure, bone turnover rates, and vascularization at these sites may contribute to metastasis formation (65). Currently, singlecell transcriptome techniques are being employed to identify variations in bone homeostasis across different bone regions and under various pathological conditions (16). Additionally, heterogeneous osteoclasts have been identified in different bone sites and under various pathological conditions (16).

Osteoclasts can be classified based on their anatomical location (e.g., calvarial osteoclasts, odontoclasts, and vascular-associated osteoclasts) and their association with certain diseases (e.g., arthritis-, obesity-, and fracture-associated osteoclasts) (16). In a murine model, long bone osteoclasts display a more advanced osteoclastic phenotype than calvarial osteoclasts, with a higher expression of membranous and secreted osteoclast proteins (66). Furthermore, osteoclasts from the cranium and appendicular skeletons have distinct characteristics. This heterogeneity in osteoclasts from different sites is reflected in their osteoclastic activity and bone turnover as well as in their response to therapeutic interventions. For example, osteoclasts from various anatomical locations exhibit different responses to bisphosphonate therapy, which is considered one of the mechanisms underlying bisphosphonate-induced osteonecrosis of the jaw (67).

Currently, the scientific literature on the heterogeneity of osteoclasts across different skeletal locations is limited. Several studies have indicated that this osteoclast diversity can be attributed to the regulation of osteoclast development by various cells within the bone, exhibiting site-specific and disease-dependent patterns. One study found that the RANKL/OPG ratio and TNF-a gene expression were significantly higher in cranially isolated osteoblasts than in long bones, yielding a different process of osteoblast-induced osteoclast formation in different bones (68). A single-cell investigation of osteoclastogenesis in bone marrow revealed that, alongside osteoblasts and osteocytes, marrow adipogenic lineage precursors (MALPs) are vital in osteoclast formation (69). The key role of MALP-derived RANKL in bone remodeling at various skeletal sites was confirmed in adipocytespecific RANKL-CKO mice (69). The absence of MALPs within the periosteum indicates that RANKL-CKO mice exhibit reduced bone loss, specifically in the trabecular bone, whereas cortical bone remains unaffected. However, the potential variations in MALPs within bone tissue across distinct anatomical locations remain unexplored. Other studies have suggested that variations in inflammatory signaling pathways, such as IL-1 and TNF- $\alpha$ , within different skeletal sites may contribute to phenotypic differences in osteoclasts (70, 71).

Based on statistical findings from clinical data, bone metastases most frequently occur in the spine, with the pelvis being the subsequent site of occurrence (72). While the correlation between the spread of bone metastases and anatomical characteristics is evident (73), the potential influence of variations in cellular composition within the skeletal system remains uncertain. Furthermore, although osteoclasts vary between anatomical sites, the relationship between osteoclast heterogeneity and the predilection of tumor metastases for specific bone sites has not been well researched. Thus, further research is required to ascertain the precise role of osteoclasts in bone metastasis.

### 4 Therapeutic targets of osteoclasts in preclinical studies

#### 4.1 RANKL

RANKL is a TNF superfamily member generated by osteoblasts, T cells, and stromal cells (74). When RANK binds to RANKL, several genes and pathways involved in osteoclast growth are activated, including NF- $\kappa$ B, mitogen-activated protein kinase, protein kinase C, and Src kinase (75). Extensive research has highlighted the importance of RANKL in bone remodeling and osteoclast development. RANKL-targeted therapy is one of the most effective therapeutic strategies and is currently used to treat osteoporosis, giant bone cell tumors, and bone resorptive disorders.

RANKL is a type II transmembrane protein with a carboxyterminal extracellular domain, which is cleaved by proteases, leading to the release of soluble RANKL into the extracellular environment (74). Recent research has shown that membranebound RANKL is primarily responsible for regulating musculoskeletal and immune functions in a physiological context, whereas soluble RANKL plays a key role in pathological processes such as bone metastasis formation (76). Additionally, soluble RANKL has been demonstrated to be nonessential in physiological processes; however, it facilitates the spread of cancer to the bone by directly inducing tumor cell migration toward the bone. Notably, the absence of soluble RANKL does not impact osteoclasts at the metastatic site or the dissemination of cancer to nonskeletal organs. Based on these findings, targeting soluble RANKL alone could be a highly specific approach for treating bone metastasis (76).

The ability of osteoclasts to reactivate dormant DTCs can be suppressed by treatment with OPG-Fc, which disrupts *in vivo* RANK–RANKL interactions (77). Additionally, RANKL acts as a mediator in the interactions between the skeletal and immune systems. Various inflammatory factors can activate RANKL,

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promoting osteoclastogenesis activity in metastatic lesions (78). Modulating the immune environment of tumors has been reported to decrease osteoclast and osteoclastogenesis activities. Within the local tumor microenvironment, the expression of PD-L1 and CCL2 is increased, with PD-L1 facilitating RANKL-induced osteoclastogenesis through JNK activation and CCL2 release (79). In a murine model of bone metastasis, treatment with the PD-1 monoclonal antibody nivolumab inhibited osteoclast development (79). Further research is warranted to determine the role of RANKL in immunotherapy and the mechanism by which RANKL-targeted medications impact the immunological tumor microenvironment.

#### 4.2 M-CSF

M-CSF, also known as colony-stimulating factor 1 (CSF-1), plays a vital role in osteoclast differentiation in various stages of development. M-CSF promotes the survival and proliferation of OPs by binding to its receptor, CSF1R. Moreover, M-CSF can restore osteoclast deficiency in M-CSF-deficient osteoporotic models (80). It has several isomeric forms and can be expressed as a membrane-bound protein or secreted as proteoglycans and glycoproteins (81). In addition to promoting cell growth, M-CSF protects OPs from undergoing apoptosis (82).

In vitro studies have shown that breast cancer cell lines can stimulate osteoblast differentiation by secreting M-CSF, with M-CSF inhibition potentially reducing the risk of osteolytic metastasis (83). Similarly, a mouse model of lung cancer bone metastasis demonstrated that the knockdown of the CSF-1 gene in A549 cells significantly reduced the number of osteoclasts and inhibited the tendency of tumors to metastasize to bone (84). Osteoclasts and macrophages originate from the same progenitors, which require critical lineage signals to differentiate into distinct cell types. Inhibiting CSF-1/CSF1R signaling can also affect macrophages (85). Therefore, in addition to regulating osteoclast differentiation, considerable preclinical and clinical research efforts have focused on the ability of CSF-1/CSF1R signaling to regulate macrophages. As a relationship exists between macrophages and bone metastasis, the potential antibone metastatic mechanism of CSF-1R may not depend solely on osteoclast inhibition.

While no clinical trials are currently focusing on CSF1R inhibitors, particularly for bone metastasis treatment, several studies are exploring CSF1R inhibitors for various cancer types. These studies might pave the way for further research on patients with bone metastases by elucidating the effectiveness and safety of CSF1R inhibitors in humans.

#### 4.3 Src

Src, a nonreceptor tyrosine kinase, belongs to a family of proteins that bind to the cytoplasmic surface of cellular membranes (86). Unlike M-CSF and RANKL which control the differentiation of osteoclasts, Src is mainly responsible for regulating osteoclast function. In particular, osteoclasts rely on Src to facilitate the rapid assembly and disassembly of podosomes, which serve as attachment structures to aid the transition between migratory and resorbing stages (87). A study revealed that Src expression is significantly correlated with bone metastasis in various tumors (88). Dasatinib, an Src kinase inhibitor, has been shown to inhibit osteoclast recruitment in a xenograft mouse model of breast cancer. In a previous study, bioluminescence scan revealed that dasatinib significantly reduced skeletal metastases in triple-negative breast cancer *in vivo*, whereas  $\mu$ -CT indicated a substantial increase in bone volume in the dasatinib-treated group (89).

The phosphorylation state of Src is influenced by multiple variables. Controlling these variables that regulate Src phosphorylation may limit bone metastasis by suppressing osteoclasts. Macrophage-stimulating protein signals can promote Src phosphorylation by binding to its corresponding receptor, RON tyrosine kinase. RON kinase inhibitors have been employed in preclinical investigations to alter Src phosphorylation in osteoclasts for treating bone metastasis (90). Moreover, factors that may reportedly influence osteoclasts through Src include CXCL12, CCR5, ICAM-1, CX3CL1, and CXCL17 (91–94). These findings demonstrate that Src, similar to RANKL, acts as a link between bone and immune systems and plays an important role in bone metastasis.

#### 4.4 Cathepsin K

Cathepsin K, a cysteine protease highly expressed in osteoclasts, belongs to the cathepsin lysosomal protease family. It acts as an osteolytic enzyme and protease, breaking down bone matrix proteins (95). It is closely associated with osteoclast-mediated bone destruction during tumor metastasis. Researchers have recently developed a system for imaging probes (Osteoadsorptive Fluorogenic Sentinel imaging probe) that can respond to osteoclastsecreted cathepsin K and detect osteolytic reactions at the metastatic site of multiple myeloma in a mouse model (96). Preclinical studies have shown that the inhibition of cathepsin K significantly decreases the ability of prostate cancer cells to induce osteoclastmediated bone resorption, thus decreasing the risk of bone metastasis both in vitro and in vivo (95, 97). Similar results have been reported in preclinical studies on breast, lung, and renal tumors (98). One such study revealed that the inhibition of cathepsin K may directly affect the function of osteoclasts without altering their number (99). Odanacatib, a cathepsin K inhibitor, does not alter the number of osteoclasts but inhibits the mRNA expression of secreted osteolytic factors, such as PTHrP, CXCR-4, and TNF- $\alpha$  (99).

In addition to osteoclasts, cathepsin K expressed by tumor and other cells within the tumor microenvironment can promote bone metastasis. In human colorectal cancer tissues, the overexpression of cathepsin K is consistently associated with increased M2 tumorassociated macrophages (TAMs) in the stroma, which is related to tumor metastasis and poor prognosis. A recent study proposed that cathepsin K secreted by tumor cells binds to TLR4, leading to the M2 polarization of TAMs through an mTOR-dependent mechanism (100).

#### 4.5 mTOR

The mTOR pathway is crucial in regulating skeletal development and homeostasis due to its impact on osteoblasts, osteoclasts, and chondrocytes (101). Although the precise mechanism by which mTOR regulates osteoclasts is being debated, the importance of mTOR signaling in osteoclastogenesis and osteoclast viability has been established. It is widely postulated that several pathways associated with mTOR can influence osteoclastogenesis through diverse mechanisms. Experimental evidence supports the notion that mTOR-mediated restriction Akt signaling regulates osteoclast fusion, whereas the regulation of mTOR-raptor protein translation leads to the cytoplasmic development of individual osteoclasts, independent of fusion (102). Furthermore, mTOR complex 1 (mTORC1) plays a crucial role in modulating bone resorption and maintaining bone homeostasis in pathological scenarios due to its expression in osteoclasts (103). mTORC1 was observed to negatively regulate the expression of NF-KB and NFATc1, which are crucial transcription factors in the differentiation of osteoclasts within osteoclast lineages (104). The mTOR pathway plays a pivotal role as a signaling mechanism that restricts osteoclastic differentiation, suggesting the potential therapeutic application of mTOR inhibitors in managing bone loss-related disorders. Preclinical investigations have reported that the osteoclast population and osteolysis associated with experimental metastases were significantly reduced upon administering Rapamycin (105, 106).

#### 4.6 Siglec-15

The activation of adaptor-receptor complexes of the immunoreceptor tyrosine-based activation motif elicits costimulatory signals that contribute to osteoclastogenesis. Molecules such as cytotoxic T lymphocyte-associated antigen 4 for Fc receptor gamma and Siglec-15 for 12-kD DNAX-activating protein have been identified as prospective candidates for inducing alteration in the suppression of osteoclasts. Siglec-15 is an essential immune suppressor that exhibits varying overexpression across many cancer types, making it a promising candidate for cancer immunotherapy (107).

Siglec-15, a member of the Siglec family, is highly conserved and expressed on osteoclasts, some myeloid cells, and certain cancer cells (108). Research has revealed that Siglec-15 is closely involved in osteoclast differentiation and that conditional knockout of Siglec-15 in mice or treatment with Siglec-15–neutralizing antibodies inhibits the multinucleation process of osteoclasts (109). Siglec-15 is also closely linked to intratumor immunity. Therefore, it has been suggested that Siglec-15 not only regulates osteoclasts in the vicious cycle of bone metastasis but also promotes bone destruction through synergistic effects with TGF- $\beta$  (110). Dou et al. investigate the role of sialylation of TLR2 in osteoclast fusion, demonstrating that Siglec15 binds sialylated TLR2 to activate cell recognition and fusion in osteoclast formation, with implications for bone resorption regulation (111). In subsequent mouse animal

model studies, it was discovered that apoptotic bodies derived from osteoclasts in the bone microenvironment inhibit the activation of CD8 T cells through Siglec15, thereby promoting the metastasis of breast cancer (112). Conversely, the use of anti-Siglec15 therapy reduces the occurrence of secondary metastases and enhances survival rates in cases of breast cancer bone metastasis. However, the detailed molecular processes regulating Siglec-15 expression remain unclear, and there is a lack of relevant preclinical studies confirming the effectiveness of Siglec-15-targeted therapy in inhibiting bone metastasis. In addition, Siglec-15 may have a regulatory effect on osteoblasts; however, further in-depth studies are warranted to confirm these findings.

Siglec-15, a molecule sharing 30% structural similarity with the B7 family (PD-L1/B7-H1), can induce immune-regulatory responses by inhibiting the activation and proliferation of T cells through an IL-10-dependent mechanism (113). Furthermore, no apparent correlation was observed between Siglec-15 and PD-L1 expression, indicating that Siglec-15 might have a significant immunosuppressive function independent of PD-L1 (114). Therefore, Siglec-15 could potentially serve as an additional therapeutic target for managing malignancies with resistance to anti-PD-L1/PD-1 immunotherapy.

# 5 Clinical trials focusing on osteoclast-targeted therapy

The selection of suitable treatment approaches for patients with bone metastases relies on various factors, including the distinct attributes of the primary malignancy, presence or absence of metastases in areas other than the skeletal system, and extent of bone involvement (whether it is confined to a specific area or spread throughout the body) (6). Management of bone metastases requires a comprehensive strategy that incorporates several therapies, such as chemotherapy, radiation therapy, biologically targeted agents, endocrine interventions, and surgical procedures. Osteoclasttargeted therapy strategies are a valuable adjunctive treatment for effectively managing SREs, thus providing a crucial complement to existing therapeutic approaches.

The efficacy of antiosteoclastogenic treatment modalities such as bisphosphonate medications and denosumab in managing bone metastases depends on the inherent attributes of the primary tumor. The present focus of clinical trials is on bone metastases originating from breast and prostate cancer, which have shown promising outcomes. However, in case of bone metastases originating from other tumors, these medications are employed solely to control SREs or as a strategy for symptomatic treatment.

#### 5.1 Bisphosphonates

Bisphosphonates are widely used antiresorptive medications in clinical practice that have demonstrated promising outcomes in treating osteoporosis and Paget's disease of the bone. Their efficacy in preventing SREs has been reported in individuals with metastatic bone cancers, such as breast and prostate cancer. Bisphosphonates predominantly exhibit an affinity toward bones at sites of active bone metabolism and dissolve from the bone matrix during bone resorption. They are then taken up by osteoclasts, limiting their activity and survival and mitigating osteoclast-mediated bone resorption (115). Notably, several bisphosphonate medications have unique mechanisms of action on osteoclasts. Non-nitrogencontaining bisphosphonates such as etidronate induce osteoclast apoptosis by forming a toxic adenosine triphosphate analog. Conversely, nitrogen-containing bisphosphonates such as alendronate, zoledronic acid, and ibandronate target the enzyme farnesyl diphosphate synthase, which is necessary for osteoclast function (116).

Bisphosphonates have been demonstrated to minimize the risk of bone metastases in some patients with breast cancer. However, research on prostate cancer and other solid tumors does not support the use of bisphosphonates in preventing bone metastases (Table 1) (127). Currently, bone-targeted drugs including denosumab, zoledronic acid, and bisphosphonates have not yet been clinically approved to prevent initial bone metastasis in patients with prostate cancer (128). Notwithstanding this fact, bisphosphonates have achieved widespread clinical recognition as a medication for managing and preventing SREs and osteoporosis related to cancer therapy (126, 129, 130). However, adjuvant bisphosphonates should not be considered a replacement for conventional anticancer treatments.

The ASCO-CCO 2021 guidelines provide clear recommendations regarding the use of adjuvant bisphosphonate therapy for postmenopausal patients with primary breast cancer. These recommendations apply to patients eligible for adjuvant systemic therapy, regardless of their hormone receptor status or human epidermal growth factor receptor 2 status (125). The efficacy of bisphosphonates in reducing the incidence of bone metastases in patients with breast cancer was demonstrated through clinical randomized controlled trials as early as the 1990s. In a clinical trial comprising 302 patients with primary breast cancer at high risk for distal metastases, clodronate effectively mitigated the risk of bone and visceral metastases (131). However, a separate trial showed that clodronate was significantly more effective in inhibiting tumor metastasis among patients with breast cancer, specifically those aged  $\geq$ 50 years, compared to the control group receiving a placebo (132). The authors attributed the variations in bisphosphonate effectiveness across various age groups to dissimilarities in estrogen levels and bone turnover mechanisms during the menopausal transition. An increasing number of clinical trials have subsequently aimed to further elucidate the specific subset of patients with breast cancer who may derive therapeutic benefits from bisphosphonate medication. A collaborative metaanalysis conducted by the Early Breast Cancer Trialists' Collaborative Group examined the impact of adjuvant bisphosphonate treatment on breast cancer across multiple trials (133). Their study found that bisphosphonates can reduce the

TABLE 1 Clinical trial of bisphosphonates in the treatment of bone metastases.

Medication	Tumor	Patients	Outcomes	Study	Reference
Clodronate	Breast cancer	Older postmenopausal patients	Preventing metastases	NSABP B-34	(117)
	Breast cancer	Primary breast cancer patients	Preventing metastases, extend OS and DFS		(118)
	Breast cancer	Early-stage breast cancer	Reducing SREs		(119)
	Breast cancer	Node-positive breast cancer	Negative		(120)
	Prostate cancer	Nonmetastatic prostate cancer	Negative	MRC PR04	(121)
	Prostate cancer	Metastatic prostate cancer	Negative	MRC PR05	(117)
Ibandronate	Breast cancer	Patients with postmenopausal ER+ breast cancer	Negative	TEAM-IIB	(122)
	Breast cancer	Patients with high-risk early-stage breast cancer	Negative	GAIN Study	(123)
Zoledronic acid	Breast cancer	Women with established menopause	Preventing metastases	the AZURE trial	(124)
	Breast cancer	Patients with MAF-negative tumor	Preventing metastases and reducing SREs	AZURE trial	(125)
	Breast cancer	After disease recurrence	Reducing SREs	the AZURE trial	(123)
	Prostate cancer	Patients with high-risk nonmetastatic prostate cancer	Negative	ZEUS	(126)
	Prostate cancer	Treatment-naive prostate cancer	Delay the first SREs	ZAPCA	(122)

recurrence of breast cancer in the skeletal system and improve overall survival outcomes for individuals diagnosed with breast cancer. However, these advantages are only noticed in females who commenced the medication after the onset of menopause (133).

A recent clinical trial addressed the question of whether different bisphosphonate agents exhibit varying effectiveness in treating bone metastases in patients with breast cancer. The SWOG/Alliance/Canadian Cancer Trials Group/ECOG-ACRIN/ NRG Oncology Study S0307 evaluated the effectiveness of three bisphosphonates (zoledronic acid, clodronate, and ibandronate) in patients with early-stage breast cancer (15). Neither the overall trial nor subgroup analyses showed any evidence of variations in efficacy among these three bisphosphonates in preventing bone metastatic disease. The authors also concluded that bisphosphonates should be incorporated into the treatment of postmenopausal individuals with early-stage breast cancer (15). However, administering oral ibandronate as an adjuvant treatment did not significantly improve the outcomes of patients with high-risk early-stage breast cancer who had received dose-dense chemotherapy (134). Notably, the outcomes of the TEAM-IIB study do not validate the efficacy of ibandronate in managing metastases among postmenopausal females diagnosed with estrogen receptor-positive (ER+) breast cancer (135).

To further optimize the indications for bisphosphonate treatment for bone metastasis, several clinical trials have investigated the differences in gene expression in subpopulations. The AZURE trial suggested that *MAF* can serve as a biological indicator to identify the potential beneficiary population of bisphosphonate treatment in patients with breast cancer (136). Patients with MAF-negative tumors exhibited prolonged invasive disease-free survival when treated with zoledronic acid (HR = 0.74–0.98; 95% confidence interval [CI] = 0.64–0.98) compared with those who received a placebo. However, patients with MAF positivity did not show this trend. Moreover, in a population of young menopausal patients, zoledronic acid treatment in MAF-negative patients was associated with longer overall survival (HR = 2.27; 95% CI = 1.04–4.93) and invasive disease-free survival (HR =

2.47; 95% CI = 1.23-4.97) compared with the control treatment (136).

#### 5.2 Denosumab

Denosumab, a human monoclonal antibody targeting RANKL, has been approved for treating osteoporosis and giant cell tumors of the bone. It has shown greater effectiveness than bisphosphonates in managing osteoporosis (137). Furthermore, postmenopausal women may safely switch from bisphosphonate treatment to denosumab, as the latter has shown increased effectiveness in enhancing bone mineral density than the continued use of bisphosphonates (138). Moreover, compared with bisphosphonates, denosumab is a more selective inhibitor of osteoclasts, and a phase III clinical trial revealed that denosumab is more effective in preventing malignant bone metastases (Table 2) (142).

Denosumab does not impact overall survival in patients with castration-resistant prostate cancer. However, in a previous study, compared with placebo, denosumab significantly enhanced bone metastasis-free survival (29.5 [95% CI = 25.4-33.3] vs. 25.2 [95% CI = 22.2–29.5] months; HR = 0.85, 95% CI = 0.73–0.98, p = 0.028) and significantly delayed time to first bone metastasis (33.2 [95% CI = 29.5-38.0] vs. 29.5 [22.4-33.1] months; HR = 0.84, 95% CI = 0.71-0.98, p = 0.032) (143). In the ABCSG-18 clinical trial, which exclusively included postmenopausal patients with breast cancer (early-stage, hormone receptor-positive, nonmetastatic adenocarcinoma of the breast), the administration of denosumab as adjuvant therapy improved disease-free survival (14). Nevertheless, the results of this study do not conclusively demonstrate the efficacy of denosumab in preventing the occurrence of bone metastasis in patients with breast cancer, as the distal metastases were not histologically confirmed during the trial.

Recent clinical trial data involving more recruited individuals have also raised doubts about the effectiveness of denosumab in preventing malignant bone metastases. The absence of guideline

Tumor	Patients	Outcomes	Study	Reference
Prostate cancer	Patients with castration-resistant prostate cancer	Increased bone-metastasis-free survival; delayed time to first bone metastasis		(134)
Breast cancer	Patients with stage II or III breast cancer	Negative	D- CARE	(13)
Breast cancer	Postmenopausal patients with hormone receptor-positive breast cancer	Preventing metastases and extending OS and DFS	ABCSG- 18	(14)
Prostate cancer	Patients with castration-resistant prostate cancer, those with no previous bisphosphonate exposure, and those with radiographic evidence of bone metastasis	Reduced the risk of skeletal complications compared with zoledronic acid		(139)
Prostate cancer	Patients with nonmetastatic castration-resistant prostate cancer	Consistently increased bone metastasis-free survival in men, with reduced PSA doubling time		(140)
Breast cancer	Women with breast cancer and skeletal metastases	Administering ZA every 3 months was more cost- effective in reducing the risks of SRE than the monthly administration of denosumab	CALGB 70604	(141)

endorsement for the use of denosumab in preventing bone metastases contrasts with the well-documented effectiveness of bisphosphonates. Following the guidelines released by ASCO-CCO in 2021, the use of adjuvant denosumab to prevent breast cancer recurrence is not recommended due to inconsistent findings in studies involving individuals with early-stage breast cancer (125). A phase III multicenter randomized controlled clinical study involving 4509 patients with breast cancer over a 5-year follow-up period found that adjuvant denosumab did not improve metastasis- and disease-free survival in patients with early-stage breast cancer (13). Additionally, there were no clinical benefits to using denosumab as an adjuvant to chemotherapy in treating nonsmall cell lung cancer (144).

The efficacy of denosumab was found to be superior to that of zoledronic acid regarding SRE prevention in patients with bone metastasis (145). Although the efficacy of denosumab as a therapy for SREs is evident, its increased application has revealed its complications. For example, the termination of denosumab treatment is associated with a rebound effect characterized by an increased likelihood of numerous spontaneous vertebral fractures (146).

#### 5.3 Src inhibitors

The Src family kinases are recognized for their pivotal function in transmitting signals across diverse cellular processes and in bone metastasis formation. Suppressing Src induces a decrease in osteoclastic bone resorption, indicating its potential as a viable treatment strategy for medical disorders characterized by excessive bone resorption, including metastatic bone diseases and osteoporosis (147). Several ongoing clinical trials are assessing the effectiveness of Src kinase inhibitors, including dasatinib, KX2–391, bosutinib, AZD0424, and saracatinib (148).

According to a phase II clinical trial, dasatinib is both safe and effective in safeguarding bone integrity among patients with metastatic castration-resistant prostate cancer who had not received prior chemotherapy (149). The findings of the trial demonstrated that the administration of dasatinib induced a reduction in alkaline phosphatase levels, indicating its effectiveness in mitigating the development of new bone lesions. Nevertheless, the precise impact of dasatinib on bone lesions remains unclear. A subsequent phase III trial reported that dasatinib provided no significant benefit when combined with chemotherapy for prostate cancer (150). In particular, the READY trial demonstrated no significant difference in the survival rates of patients with metastatic castration-resistant prostate cancer who received dasatinib + docetaxel and those who only received docetaxel (21.5 [95% CI = 20.3-22.8] vs. 21.2 [95% CI = 20.0-23.4] months; HR = 0.99, 95.5% CI = 0.87-1.13, p = 0.90) (150). The investigators concluded that the addition of dasatinib to the treatment showed no significant improvement in the levels of bone turnover markers. Furthermore, they revealed that the suppressive impact on bone lesions may be attributed to docetaxel. Owing to the disappointing outcomes of several clinical trials on the effectiveness of dasatinib as a treatment for breast cancer, the feasibility of conducting additional research on its use as an independent therapy remains controversial. Alternatively, studies have suggested that the application of dasatinib may be limited to a certain group of patients exhibiting molecular expression or may be combined with other pharmaceutical medications in the future (151). Moreover, clinical trials have shown that Src enzyme inhibitors, including bosutinib, saracatinib, and KX2–391, exhibit no significant beneficial impact on bone lesions or SREs (152–154).

Although Src inhibitors have shown potential efficacy in alleviating bone resorption in certain contexts, further investigation is warranted to optimize their ability to target bones before realizing their potential advantages in clinical trials.

#### 5.4 Cathepsin K inhibitors

Although preclinical investigations have established a strong correlation between cathepsin K and the occurrence of bone metastases, clinical trials have predominantly prioritized the use of cathepsin K inhibitors for treating osteoporosis and arthritis. To date, no cathepsin K inhibitor has received approval from the Food and Drug Administration. Odanacatib has emerged as the only candidate for inhibiting cathepsin K and has demonstrated considerable therapeutic efficacy in phase III clinical trials in patients with postmenopausal osteoporosis (155). A single shortterm, double-blind clinical trial provisionally demonstrated the effectiveness of odanacatib in managing bone metastases in patients with breast cancer, as evidenced by one of the limited clinical trials conducted on this topic (156). The trial provided evidence regarding the inhibitory effects of odanacatib on the expression levels of osteolysis marker (urinary N-telopeptide of type I collagen corrected for creatinine) in patients with breast cancer, similar to the effects of zoledronic acid. Clinical trials on osteoporosis have recently been discontinued due to the observed increase in cardiovascular events associated with the administration of odanacatib. Consequently, the investigators have decided to refrain from allocating resources toward further research (157). Several cathepsin K inhibitors have been eliminated by researchers due to the severe adverse effects observed during clinical trials.

#### 5.5 Everolimus

Everolimus is an mTOR inhibitor that suppresses the proosteoclast paracrine pathway of tumor cells, thereby inhibiting bone metastasis (158). In the BOLERO-2 trial, patients with bone metastasis who received everolimus showed a significant improvement in bone health, with a considerable reduction in skeletal tumor growth, compared with those in the control group (159). Furthermore, the levels of bone resorptive markers (BSAP, P1NP, and CTX) in patients treated with everolimus decreased significantly, which was attributed to the antitumor action of the drug, inducing a reduction in osteoclast function. The RADAR study suggested that everolimus can provide long-term benefits in patients with bone metastases, as long as no progression occurs during the first 8 weeks of therapy (160).

#### 6 Challenges and future research

The current approach to managing bone metastases involves a combination of several therapeutic modalities. Preclinical research often lack direct applicability to real-world applications, and the use of bisphosphonates and denosumab is limited to their roles as adjuvant therapies. Nevertheless, formulating theoretical frameworks remains the primary method through which we can understand the complexity of bone metastasis. Researchers have recently found that malignant tumors expressing Cyp11a1 produce the steroid pregnenolone, which can promote the growth of bone metastases by enhancing osteoclastogenesis (161).However, researchers encounter various challenges in conducting preclinical and clinical investigations aimed at treating bone metastases.

Regarding clinical studies, the methodology used for the clinical evaluation of bone metastases differs from that used for primary tumors. Evaluating treatment efficacy in metastatic bone disease is also distinctive, focusing on specific criteria that consider both bone repair and deterioration rather than just changes in tumor size. Furthermore, assessing responses in bone metastases poses challenges due to the gradual and inconspicuous nature of the healing process. The predominant clinical investigations for bone metastases rely on imaging modalities, such as X-ray, computer tomography, magnetic resonance imaging, and Positron Emission Tomography. However, the precision of these examinations is often insufficient, limiting their capacity to identify early-stage microlesions and evaluate disease progression. Assessing multifocal bone metastases that affect both the axial and appendicular skeletons can present difficulties and inaccuracies using current imaging modalities. The absence of standardization and harmonization in clinical assessment methods across trials further complicates the precise comparison and interpretation of outcomes. This limitation hinders the ability to ascertain treatment efficacy based solely on individual study results. In the future, the use of nanoprobes holds promise for enhancing the diagnosis and assessment of bone metastases. These nanoprobes can provide extensive insights into several aspects, including anatomical structure, functional metabolism, physiological characteristics, and pathological profiles, with multimodal imaging functions for detecting ultrasmall lesions in bones and multiorgan cancer metastases (162).

The translatability and correlation between therapeutic responses observed in preclinical models and those observed in human clinical trials are often inadequate. This inadequacy can be attributed to the challenges encountered in preclinical studies on bone metastases. Notably, prevalent research models exhibit limitations in accurately replicating human bone metastases. Both animal and *in vitro* models have limitations in fully replicating the components and natural processes present in the bone metastasis microenvironment in humans. Additionally, the precise mechanisms underlying bone metastasis remain elusive, and investigations into these mechanisms are limited by the inability to observe *in vivo* processes accurately. Exploring the dynamic interactions between malignant cells and the bone niche within a living animal through intact bone requires high-resolution deep-tissue imaging, making it an essential area of research. Although high-resolution imaging protocols for deep-tissue observation exist, they are not widely available.

The current understanding of osteoclast heterogeneity is limited and requires further exploration. The osteoclast phenotype fluctuates based on various physiological and pathological conditions, and notable phenotypic differences exist among various bones. The rapid development of single-cell sequencing technology has facilitated these findings and provided a feasible approach for elucidating the mechanisms underlying bone metastasis. The use of single-cell sequencing technologies presents a viable approach for obtaining novel insights into the biological mechanisms underlying the development and advancement of bone metastasis. Moreover, single-cell methodologies have the potential to reveal the molecular mechanisms underlying the metastatic colonization of bones by accounting for significant cellular heterogeneity, plasticity, and evolutionary changes that occur during metastasis and within the complex bone microenvironment. Future in-depth studies on osteoclast heterogeneity can help reveal the mechanisms underlying bone metastasis, improve treatment targeting, and address current clinical challenges in treating bone metastasis. The combination of high-resolution single-cell and spatial technologies in a multidimensional approach has highlighted the significance of the complete cancer ecosystem, which may provide insights into the intricate mechanisms underlying bone metastasis in the future.

### Author contributions

YL: Writing – original draft, Methodology. HC: Writing – review & editing. TC: Writing – review & editing. GQ: Writing – review & editing. YH: Writing – review & editing, Supervision, Investigation, Conceptualization.

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## Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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#### References

1. Ryan C, Stoltzfus KC, Horn S, Chen H, Louie AV, Lehrer EJ, et al. Epidemiology of bone metastases. *Bone*. (2022) 158:115783. doi: 10.1016/j.bone.2020.115783

2. Galasko CS. Monitoring of bone metastases. Schweiz Med Wochenschr. (1981) 111:1873-5.

3. Coleman RE, Roodman, Smith, Body, Suva, Vessella. Clinical features of metastatic bone disease and risk of skeletal morbidity. *Clin Cancer Res.* (2006) 12:6243s-9s. doi: 10.1158/1078-0432.CCR-06-0931

4. Salhotra A, Shah HN, Levi B, Longaker MT. Mechanisms of bone development and repair. *Nat Rev Mol Cell Biol.* (2020) 21:696–711. doi: 10.1038/s41580-020-00279-w

5. Clézardin P, Coleman R, Puppo M, Ottewell P, Bonnelye E, Paycha F, et al. Bone metastasis: mechanisms, therapies, and biomarkers. *Physiol Rev.* (2021) 101:797–855. doi: 10.1152/PHYSREV.00012.2019

6. Coleman RE, Croucher PI, Padhani AR, Clézardin P, Chow E, Fallon M, et al. Bone metastases. Nat Rev Dis Prim. (2020) 6:83. doi: 10.1038/s41572-020-00216-3

7. Qaseem A, Hicks LA, Etxeandia-Ikobaltzeta I, Shamliyan T, Cooney TG, Cross JTJ, et al. Pharmacologic treatment of primary osteoporosis or low bone mass to prevent fractures in adults: A living clinical guideline from the american college of physicians. *Ann Intern Med.* (2023) 176:224–38. doi: 10.7326/M22-1034

8. Jakob T, Tesfamariam YM, Macherey S, Kuhr K, Adams A, Monsef I, et al. Bisphosphonates or RANK-ligand-inhibitors for men with prostate cancer and bone metastases: a network meta-analysis. *Cochrane Database Syst Rev.* (2020) 12: CD013020. doi: 10.1002/14651858.CD013020.pub2

9. Tesfamariam Y, Jakob T, Wöckel A, Adams A, Weigl A, Monsef I, et al. Adjuvant bisphosphonates or RANK-ligand inhibitors for patients with breast cancer and bone metastases: A systematic review and network meta-analysis. *Crit Rev Oncol Hematol.* (2019) 137:1–8. doi: 10.1016/j.critrevonc.2019.02.004

10. Zhang W, Bado I, Wang H, Lo HC, Zhang XHF. Bone metastasis: Find your niche and fit in. Trends Cancer. (2019) 5:95–110. doi: 10.1016/J.TRECAN.2018.12.004

11. Maurizi A, Rucci N. The osteoclast in bone metastasis: Player and target. *Cancers* (*Basel*). (2018) 10(7):218. doi: 10.3390/cancers10070218

12. von Moos R, Costa L, Gonzalez-Suarez E, Terpos E, Niepel D, Body J-J. Management of bone health in solid tumours: From bisphosphonates to a monoclonal antibody. *Cancer Treat Rev.* (2019) 76:57–67. doi: 10.1016/j.ctrv.2019. 05.003

13. Coleman R, Finkelstein DM, Barrios C, Martin M, Iwata H, Hegg R, et al. Adjuvant denosumab in early breast cancer (D-CARE): an international, multicentre, randomised, controlled, phase 3 trial. *Lancet Oncol.* (2020) 21:60–72. doi: 10.1016/S1470-2045(19)30687-4

14. Gnant M, Pfeiler G, Steger GG, Egle D, Greil R, Fitzal F, et al. Adjuvant denosumab in postmenopausal patients with hormone receptor-positive breast cancer (ABCSG-18): disease-free survival results from a randomised, double-blind, placebo-controlled, phase 3 trial. *Lancet Oncol.* (2019) 20:339–51. doi: 10.1016/S1470-2045(18) 30862-3

15. Gralow JR, Barlow WE, Paterson AHG, M'Iao JL, Lew DL, Stopeck AT, et al. Phase III randomized trial of bisphosphonates as adjuvant therapy in breast cancer: S0307. *JNCI J Natl Cancer Inst.* (2020) 112:698. doi: 10.1093/JNCI/DJZ215

16. Tsukasaki M, Takayanagi H. Osteoclast biology in the single-cell era. *Inflamm Regen.* (2022) 42(1):27. doi: 10.1186/S41232-022-00213-X

17. Low V, Blenis J, Gomes AP. Targeting the premetastatic niche: epigenetic therapies in the spotlight. *Signal Transduct Target Ther.* (2020) 5:68. doi: 10.1038/ s41392-020-0165-3

18. Furesi G, Rauner M, Hofbauer LC. Emerging players in prostate cancer-bone niche communication. *Trends Cancer.* (2021) 7:112–21. doi: 10.1016/j.trecan.2020. 09.006

19. Charles JF, Aliprantis AO. Osteoclasts: more than 'bone eaters'. *Trends Mol Med.* (2014) 20:449. doi: 10.1016/J.MOLMED.2014.06.001

20. Han Y, You X, Xing W, Zhang Z, Zou W. Paracrine and endocrine actions of bone-the functions of secretory proteins from osteoblasts, osteocytes, and osteoclasts. *Bone Res.* (2018) 6:16. doi: 10.1038/s41413-018-0019-6

21. Yuan X, Qian N, Ling S, Li Y, Sun W, Li J, et al. Breast cancer exosomes contribute to pre-metastatic niche formation and promote bone metastasis of tumor cells. *Theranostics*. (2021) 11:1429–45. doi: 10.7150/THNO.45351

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22. Yue Z, Niu X, Yuan Z, Qin Q, Jiang W, He L, et al. RSPO2 and RANKL signal through LGR4 to regulate osteoclastic premetastatic niche formation and bone metastasis. *J Clin Invest.* (2022) 132(2):e144579. doi: 10.1172/JCI144579

23. Zhang S, Xu Y, Xie C, Ren L, Wu G, Yang M, et al. RNF219/ $\alpha$ -Catenin/LGALS3 axis promotes hepatocellular carcinoma bone metastasis and associated skeletal complications. Adv Sci (Weinheim Baden-Wurttemberg Ger. (2020) 8(4):2001961. doi: 10.1002/ADVS.202001961

24. Yu L, Sui B, Fan W, Lei L, Zhou L, Yang L, et al. Exosomes derived from osteogenic tumor activate osteoclast differentiation and concurrently inhibit osteogenesis by transferring COL1A1-targeting miRNA-92a-1-5p. *J Extracell vesicles*. (2021) 10:e12056. doi: 10.1002/jev2.12056

25. Liu J, Li D, Dang L, Liang C, Guo B, Lu C, et al. Osteoclastic miR-214 targets TRAF3 to contribute to osteolytic bone metastasis of breast cancer. *Sci Rep.* (2017) 7:40487. doi: 10.1038/SREP40487

26. Chengling L, Yulin Z, Xiaoyu X, Xingchen L, Sen Z, Ziming W, et al. miR-325-3p, a novel regulator of osteoclastogenesis in osteolysis of colorectal cancer through targeting \$100A4. *Mol Med.* (2021) 27(1):23. doi: 10.1186/\$10020-021-00282-7

27. Wang J, Du X, Wang X, Xiao H, Jing N, Xue W, et al. Tumor-derived miR-378a-3p-containing extracellular vesicles promote osteolysis by activating the Dyrk1a/ Nfatc1/Angptl2 axis for bone metastasis. *Cancer Lett.* (2022) 526:76-90. doi: 10.1016/J.CANLET.2021.11.017

28. Ma Q, Liang M, Wu Y, Dou C, Xu J, Dong S, et al. Small extracellular vesicles deliver osteolytic effectors and mediate cancer-induced osteolysis in bone metastatic niche. *J Extracell vesicles*. (2021) 10(4):e12068. doi: 10.1002/JEV2.12068

29. Bravo Vázquez LA, Moreno Becerril MY, Mora Hernández EO, de León Carmona GG, Aguirre Padilla ME, Chakraborty S, et al. The emerging role of MicroRNAs in bone diseases and their therapeutic potential. *Molecules*. (2021) 27 (1):211. doi: 10.3390/molecules27010211

30. Kaboli PJ, Rahmat A, Ismail P, Ling K-H. MicroRNA-based therapy and breast cancer: A comprehensive review of novel therapeutic strategies from diagnosis to treatment. *Pharmacol Res.* (2015) 97:104–21. doi: 10.1016/j.phrs.2015.04.015

31. Henrich SE, McMahon KM, Plebanek MP, Calvert AE, Feliciano TJ, Parrish S, et al. Prostate cancer extracellular vesicles mediate intercellular communication with bone marrow cells and promote metastasis in a cholesterol-dependent manner. *J Extracell vesicles.* (2020) 10(2):e12042. doi: 10.1002/JEV2.12042

32. Strilic B, Offermanns S. Intravascular survival and extravasation of tumor cells. *Cancer Cell*. (2017) 32:282–93. doi: 10.1016/J.CCELL.2017.07.001

33. Veis DJ, O'Brien CA. Osteoclasts, master sculptors of bone. *Annu Rev Pathol.* (2023) 18:257–81. doi: 10.1146/annurev-pathmechdis-031521-040919

34. Safarzadeh E, Hashemzadeh S, Duijf PHG, Mansoori B, Khaze V, Mohammadi A, et al. Circulating myeloid-derived suppressor cells: An independent prognostic factor in patients with breast cancer. *J Cell Physiol.* (2019) 234:3515–25. doi: 10.1002/jcp.26896

35. Wang Y, Ding Y, Guo N, Wang S. MDSCs: Key criminals of tumor premetastatic niche formation. *Front Immunol.* (2019) 10:172. doi: 10.3389/ fimmu.2019.00172

36. He N, Jiang J. Contribution of immune cells to bone metastasis pathogenesis. Front Endocrinol (Lausanne). (2022) 13:1019864. doi: 10.3389/fendo.2022.1019864

37. Yao Y, Cai X, Ren F, Ye Y, Wang F, Zheng C, et al. The macrophage-osteoclast axis in osteoimmunity and osteo-related diseases. *Front Immunol.* (2021) 12:664871. doi: 10.3389/fimmu.2021.664871

38. Monteiro AC, Bonomo A. Dendritic cells development into osteoclast-type APCs by 4T1 breast tumor t cells milieu boost bone consumption. *Bone.* (2021) 143:115755. doi: 10.1016/j.bone.2020.115755

39. Kaur K, Cook J, Park SH, Topchyan P, Kozlowska A, Ohanian N, et al. Novel strategy to expand super-charged NK cells with significant potential to lyse and differentiate cancer stem cells: Differences in NK expansion and function between healthy and cancer patients. *Front Immunol.* (2017) 8:297. doi: 10.3389/ FIMMU.2017.00297

40. Kaur K, Ko MW, Ohanian N, Cook J, Jewett A. Osteoclast-expanded supercharged NK-cells preferentially select and expand CD8+ t cells. *Sci Rep.* (2020) 10 (1):20363. doi: 10.1038/S41598-020-76702-1

41. An G, Acharya C, Feng X, Wen K, Zhong M, Zhang L, et al. Osteoclasts promote immune suppressive microenvironment in multiple myeloma: therapeutic implication. *Blood.* (2016) 128:1590. doi: 10.1182/BLOOD-2016-03-707547

42. Grassi F, Manferdini C, Cattini L, Piacentini A, Gabusi E, Facchini A, et al. T cell suppression by osteoclasts in vitro. *J Cell Physiol.* (2011) 226:982–90. doi: 10.1002/ jcp.22411

43. Arellano DL, Juárez P, Verdugo-Meza A, Almeida-Luna PS, Corral-Avila JA, Drescher F, et al. Bone microenvironment-suppressed t cells increase osteoclast formation and osteolytic bone metastases in mice. J Bone Miner Res Off J Am Soc Bone Miner Res. (2022) 37:1446–63. doi: 10.1002/jbmr.4615

44. Liang H, Zhou L, Hu Z, Ge Y, Zhang T, Chen Q, et al. Siglec15 checkpoint blockade for simultaneous immunochemotherapy and osteolysis inhibition in lung adenocarcinoma spinal metastasis *via* a hollow nanoplatform. *Small.* (2022) 18: e2107787. doi: 10.1002/smll.202107787

45. Liang H, Chen Q, Hu Z, Zhou L, Meng Q, Zhang T, et al. Siglec15 facilitates the progression of non-small cell lung cancer and is correlated with spinal metastasis. *Ann Transl Med.* (2022) 10:281. doi: 10.21037/atm-22-764

46. Ibáñez L, Abou-Ezzi G, Ciucci T, Amiot V, Belaïd N, Obino D, et al. Inflammatory osteoclasts prime TNF $\alpha$ -producing CD4+ t cells and express CX3 CR1. J Bone Miner Res. (2016) 31:1899–908. doi: 10.1002/JBMR.2868

47. Madel MB, Ibáñez L, Ciucci T, Halper J, Rouleau M, Boutin A, et al. Dissecting the phenotypic and functional heterogeneity of mouse inflammatory osteoclasts by the expression of Cx3cr1. *Elife*. (2020) 9:1–22. doi: 10.7554/ELIFE.54493

48. Buchwald ZS, Kiesel JR, DiPaolo R, Pagadala MS, Aurora R. Osteoclast activated FoxP3+ CD8+ t-cells suppress bone resorption in vitro. *PloS One.* (2012) 7(6):e38199. doi: 10.1371/JOURNAL.PONE.0038199

49. Sawant A, Ponnazhagan S. Myeloid-derived suppressor cells as osteoclast progenitors: a novel target for controlling osteolytic bone metastasis. *Cancer Res.* (2013) 73:4606–10. doi: 10.1158/0008-5472.CAN-13-0305

50. Capietto A-H, Lee S, Clever D, Eul E, Ellis H, Ma CX, et al. Effective treatment of established bone metastases can be achieved by combinatorial osteoclast blockade and depletion of granulocytic subsets. *Cancer Immunol Res.* (2021) 9:1400–12. doi: 10.1158/2326-6066.CIR-21-0232

51. Ling Z, Yang C, Tan J, Dou C, Chen Y. Beyond immunosuppressive effects: dual roles of myeloid-derived suppressor cells in bone-related diseases. *Cell Mol Life Sci.* (2021) 78:7161–83. doi: 10.1007/s00018-021-03966-9

52. Li H, Hong S, Qian J, Zheng Y, Yang J, Yi Q. Cross talk between the bone and immune systems: osteoclasts function as antigen-presenting cells and activate CD4+ and CD8+ t cells. *Blood.* (2010) 116:210. doi: 10.1182/BLOOD-2009-11-255026

53. Sawant A, Hensel JA, Chanda D, Harris BA, Siegal GP, Maheshwari A, et al. Depletion of plasmacytoid dendritic cells inhibits tumor growth and prevents bone metastasis of breast cancer cells. *J Immunol.* (2012) 189:4258–65. doi: 10.4049/jimmunol.1101855

54. Lim AR, Ghajar CM. Thorny ground, rocky soil: Tissue-specific mechanisms of tumor dormancy and relapse. *Semin Cancer Biol.* (2022) 78:104–23. doi: 10.1016/J.SEMCANCER.2021.05.007

55. Phan TG, Croucher PI. The dormant cancer cell life cycle. Nat Rev Cancer. (2020) 20:398-411. doi: 10.1038/S41568-020-0263-0

56. Riquelme MA, Cardenas ER, Jiang JX. Osteocytes and bone metastasis. Front Endocrinol (Lausanne). (2020) 11:567844. doi: 10.3389/FENDO.2020.567844

57. Wang H, Yu C, Gao X, Welte T, Muscarella AM, Tian L, et al. The osteogenic niche promotes early-stage bone colonization of disseminated breast cancer cells. *Cancer Cell.* (2015) 27:193. doi: 10.1016/J.CCELL.2014.11.017

58. Hughes R, Chen X, Cowley N, Ottewell PD, Hawkins RJ, Hunter KD, et al. Osteoblast-derived paracrine and juxtacrine signals protect disseminated breast cancer cells from stress. *Cancers (Basel)*. (2021) 13:1–19. doi: 10.3390/CANCERS13061366

59. Khoo WH, Ledergor G, Weiner A, Roden DL, Terry RL, McDonald MM, et al. A niche-dependent myeloid transcriptome signature defines dormant myeloma cells. *Blood.* (2019) 134:30–43. doi: 10.1182/BLOOD.2018880930

60. Yu-Lee LY, Yu G, Lee YC, Lin SC, Pan J, Pan T, et al. Osteoblast-secreted factors mediate dormancy of metastatic prostate cancer in the bone *via* activation of the TGF $\beta$ RIII-p38MAPK-pS249/T252RB pathway. *Cancer Res.* (2018) 78:2911–24. doi: 10.1158/0008-5472.CAN-17-1051

61. Lawson MA, McDonald MM, Kovacic N, Khoo WH, Terry RL, Down J, et al. Osteoclasts control reactivation of dormant myeloma cells by remodelling the endosteal niche. *Nat Commun.* (2015) 6:8983. doi: 10.1038/NCOMMS9983

62. Ren D, Dai Y, Yang Q, Zhang X, Guo W, Ye L, et al. Wnt5a induces and maintains prostate cancer cells dormancy in bone. *J Exp Med.* (2019) 216:428–49. doi: 10.1084/JEM.20180661

63. Capulli M, Hristova D, Valbret Z, Carys K, Arjan R, Maurizi A, et al. Notch2 pathway mediates breast cancer cellular dormancy and mobilisation in bone and contributes to haematopoietic stem cell mimicry. *Br J Cancer.* (2019) 121:157–71. doi: 10.1038/S41416-019-0501-Y

64. Lu X, Mu E, Wei Y, Riethdorf S, Yang Q, Yuan M, et al. VCAM-1 promotes osteolytic expansion of indolent bone micrometastasis of breast cancer by engaging  $\alpha 4\beta 1$ -positive osteoclast progenitors. *Cancer Cell.* (2011) 20:701–14. doi: 10.1016/J.CCR.2011.11.002

65. Fornetti J, Welm AL, Stewart SA. Understanding the bone in cancer metastasis. J Bone Miner Res. (2018) 33:2099–113. doi: 10.1002/JBMR.3618 66. Zenger S, Ek-Rylander B, Andersson G. Long bone osteoclasts display an augmented osteoclast phenotype compared to calvarial osteoclasts. *Biochem Biophys Res Commun.* (2010) 394:743–9. doi: 10.1016/J.BBRC.2010.03.063

67. Vermeer JAF, Jansen IDC, Marthi M, Coxon FP, McKenna CE, Sun S, et al. Jaw bone marrow-derived osteoclast precursors internalize more bisphosphonate than long-bone marrow precursors. *Bone.* (2013) 57:242–51. doi: 10.1016/J.BONE.2013. 08.007

68. Wan Q, Schoenmaker T, Jansen IDC, Bian Z, de Vries TJ, Everts V. Osteoblasts of calvaria induce higher numbers of osteoclasts than osteoblasts from long bone. *Bone*. (2016) 86:10–21. doi: 10.1016/J.BONE.2016.02.010

69. Yu W, Zhong L, Yao L, Wei Y, Gui T, Li Z, et al. Bone marrow adipogenic lineage precursors promote osteoclastogenesis in bone remodeling and pathologic bone loss. *J Clin Invest.* (2021) 131(2):e140214. doi: 10.1172/JCI140214

70. Ascone G, Cao Y, Jansen IDC, Di Ceglie I, van den Bosch MHJ, Blom AB, et al. Increase in the number of bone marrow osteoclast precursors at different skeletal sites, particularly in long bone and jaw marrow in mice lacking IL-1RA. *Int J Mol Sci.* (2020) 21:1–16. doi: 10.3390/IJMS21113774

71. Meirow Y, Jovanovic M, Zur Y, Habib J, Colombo DF, Twaik N, et al. Specific inflammatory osteoclast precursors induced during chronic inflammation give rise to highly active osteoclasts associated with inflammatory bone loss. *Bone Res.* (2022) 10 (1):36. doi: 10.1038/S41413-022-00206-Z

72. Wang C, Shen Y. Study on the distribution features of bone metastases in prostate cancer. *Nucl Med Commun.* (2012) 33:379-83. doi: 10.1097/MNM.0b013e3283504528

73. Zhu M, Liu X, Qu Y, Hu S, Zhang Y, Li W, et al. Bone metastasis pattern of cancer patients with bone metastasis but no visceral metastasis. *J Bone Oncol.* (2019) 15:100219. doi: 10.1016/j.jbo.2019.100219

74. Ono T, Hayashi M, Sasaki F, Nakashima T. RANKL biology: bone metabolism, the immune system, and beyond. *Inflamm Regen*. (2020) 40:2. doi: 10.1186/S41232-019-0111-3

75. Wada T, Nakashima T, Hiroshi N, Penninger JM. RANKL-RANK signaling in osteoclastogenesis and bone disease. *Trends Mol Med.* (2006) 12:17–25. doi: 10.1016/J.MOLMED.2005.11.007

76. Asano T, Okamoto K, Nakai Y, Tsutsumi M, Muro R, Suematsu A, et al. Soluble RANKL is physiologically dispensable but accelerates tumour metastasis to bone. *Nat Metab.* (2019) 1:868–75. doi: 10.1038/S42255-019-0104-1

77. Ottewell PD, Wang N, Brown HK, Fowles CA, Croucher PI, Eaton CL, et al. OPG-fc inhibits ovariectomy-induced growth of disseminated breast cancer cells in bone. *Int J Cancer*. (2015) 137:968–77. doi: 10.1002/IJC.29439

78. Li B, Wang P, Jiao J, Wei H, Xu W, Zhou P. Roles of the RANKL-RANK axis in immunity-implications for pathogenesis and treatment of bone metastasis. *Front Immunol.* (2022) 13:824117. doi: 10.3389/FIMMU.2022.824117

79. Wang K, Gu Y, Liao Y, Bang S, Donnelly CR, Chen O, et al. PD-1 blockade inhibits osteoclast formation and murine bone cancer pain. *J Clin Invest.* (2020) 130:3603–20. doi: 10.1172/JCI133334

80. Yoshida H, Hayashi SI, Kunisada T, Ogawa M, Nishikawa S, Okamura H, et al. The murine mutation osteopetrosis is in the coding region of the macrophage colony stimulating factor gene. *Nature*. (1990) 345:442–4. doi: 10.1038/345442A0

81. Mun SH, Park PSU, Park-Min KH. The m-CSF receptor in osteoclasts and beyond. *Exp Mol Med.* (2020) 52:1239–54. doi: 10.1038/S12276-020-0484-Z

 Bouyer P, Sakai H, Itokawa T, Kawano T, Fulton CM, Boron WF, et al. Colonystimulating factor-1 increases osteoclast intracellular pH and promotes survival via the electroneutral Na/HCO3 cotransporter NBCn1. Endocrinology. (2007) 148:831–40. doi: 10.1210/EN.2006-0547

83. Liverani C, Mercatali L, Spadazzi C, La Manna F, De Vita A, Riva N, et al. CSF-1 blockade impairs breast cancer osteoclastogenic potential in co-culture systems. *Bone.* (2014) 66:214–22. doi: 10.1016/J.BONE.2014.06.017

84. Hung JY, Horn D, Woodruff K, Prihoda T, Lesaux C, Peters J, et al. Colonystimulating factor 1 potentiates lung cancer bone metastasis. *Lab Invest.* (2014) 94:371– 81. doi: 10.1038/LABINVEST.2014.1

85. Fujiwara T, Yakoub MA, Chandler A, Christ AB, Yang G, Ouerfelli O, et al. CSF1/CSF1R signaling inhibitor pexidartinib (PLX3397) reprograms tumor-associated macrophages and stimulates t-cell infiltration in the sarcoma microenvironment. *Mol Cancer Ther.* (2021) 20:1388–99. doi: 10.1158/1535-7163.MCT-20-0591

86. Horne WC, Sanjay A, Bruzzaniti A, Baron R. The role(s) of src kinase and cbl proteins in the regulation of osteoclast differentiation and function. *Immunol Rev.* (2005) 208:106–25. doi: 10.1111/J.0105-2896.2005.00335.X

87. Matsubara T, Addison WN, Kokabu S, Neff L, Horne W, Gori F, et al. Characterization of unique functionalities in c-src domains required for osteoclast podosome belt formation. *J Biol Chem.* (2021) 296:100790. doi: 10.1016/J.JBC.2021.100790

88. Li Y, Bao Y, Zheng H, Qin Y, Hua B. The nonreceptor protein tyrosine kinase src participates in every step of cancer-induced bone pain. *BioMed Pharmacother*. (2021) 141:111822. doi: 10.1016/J.BIOPHA.2021.111822

89. Heilmann T, Rumpf AL, Roscher M, Tietgen M, Will O, Gerle M, et al. Dasatinib prevents skeletal metastasis of osteotropic MDA-MB-231 cells in a xenograft mouse model. *Arch Gynecol Obstet*. (2020) 301:1493–502. doi: 10.1007/S00404-020-05496-4

90. Andrade K, Fornetti J, Zhao L, Miller SC, Randall RL, Anderson N, et al. RON kinase: A target for treatment of cancer-induced bone destruction and osteoporosis. *Sci Transl Med.* (2017) 9(374):eaai9338. doi: 10.1126/SCITRANSLMED.AAI9338

91. Wang K, Jiang L, Hu A, Sun C, Zhou L, Huang Y, et al. Vertebral-specific activation of the CX3CL1/ICAM-1 signaling network mediates non-small-cell lung cancer spinal metastasis by engaging tumor cell-vertebral bone marrow endothelial cell interactions. *Theranostics*. (2021) 11:4770. doi: 10.7150/THNO.54235

92. Conley-LaComb MK, Semaan L, Singareddy R, Li Y, Heath EI, Kim S, et al. Pharmacological targeting of CXCL12/CXCR4 signaling in prostate cancer bone metastasis. *Mol Cancer.* (2016) 15(1):68. doi: 10.1186/S12943-016-0552-0

93. Sicoli D, Jiao X, Ju X, Velasco-Velazquez M, Ertel A, Addya S, et al. CCR5 receptor antagonists block metastasis to bone of v-src-oncogene-transformed metastatic prostate cancer cell lines. *Cancer Res.* (2014) 74:7103. doi: 10.1158/0008-5472.CAN-14-0612

94. Liu W, Xie X, Wu J. Mechanism of lung adenocarcinoma spine metastasis induced by CXCL17. *Cell Oncol (Dordr).* (2020) 43:311–20. doi: 10.1007/S13402-019-00491-7

95. Liang W, Wang F, Chen Q, Dai J, Escara-Wilke J, Keller ET, et al. Targeting cathepsin k diminishes prostate cancer establishment and growth in murine bone. *J Cancer Res Clin Oncol.* (2019) 145:1999. doi: 10.1007/S00432-019-02950-Y

96. Richard ET, Morinaga K, Zheng Y, Sundberg O, Hokugo A, Hui K, et al. Design and synthesis of cathepsin-K-Activated osteoadsorptive fluorogenic sentinel (OFS) probes for detecting early osteoclastic bone resorption in a multiple myeloma mouse model. *Bioconjug Chem.* (2021) 32:916–27. doi: 10.1021/ACS.BIOCONJCHEM. 1C00036

97. Sinha KM, Bagheri-Yarmand R, Lahiri S, Lu Y, Zhang M, Amra S, et al. Oncogenic and osteolytic functions of histone demethylase NO66 in castration-resistant prostate cancer. *Oncogene.* (2019) 38:5038–49. doi: 10.1038/S41388-019-0774-X

98. Qian D, He L, Zhang Q, Li W, Tang D, Wu C, et al. Cathepsin k: A versatile potential biomarker and therapeutic target for various cancers. *Curr Oncol.* (2022) 29:5963. doi: 10.3390/CURRONCOL29080471

99. Vashum Y, Kottaiswamy A, Bupesh G, Singh RP, Kalaiselvan P, Thiagarajan K, et al. Inhibitory effects of cathepsin k inhibitor (ODN-MK-0822) on the paracrine proosteoclast factors of breast cancer cells. *Curr Mol Pharmacol.* (2021) 14:1134–45. doi: 10.2174/1874467214666210211162118

100. Li R, Zhou R, Wang H, Li W, Pan M, Yao X, et al. Gut microbiota-stimulated cathepsin k secretion mediates TLR4-dependent M2 macrophage polarization and promotes tumor metastasis in colorectal cancer. *Cell Death Differ*. (2019) 26:2447–63. doi: 10.1038/S41418-019-0312-Y

101. Chen J, Long F. mTOR signaling in skeletal development and disease. *Bone Res.* (2018) 6:1. doi: 10.1038/S41413-017-0004-5

102. Tiedemann K, Le Nihouannen D, Fong JE, Hussein O, Barralet JE, Komarova SV. Regulation of osteoclast growth and fusion by mTOR/raptor and mTOR/rictor/ Akt. *Front Cell Dev Biol.* (2017) 5:54. doi: 10.3389/fcell.2017.00054

103. Hiraiwa M, Ozaki K, Yamada T, Iezaki T, Park G, Fukasawa K, et al. mTORC1 activation in osteoclasts prevents bone loss in a mouse model of osteoporosis. *Front Pharmacol.* (2019) 10:684. doi: 10.3389/fphar.2019.00684

104. Zhang Y, Xu S, Li K, Tan K, Liang K, Wang J, et al. mTORC1 inhibits NF-κB/ NFATC1 signaling and prevents osteoclast precursor differentiation, *In vitro* and in mice. *J Bone Miner Res Off J Am Soc Bone Miner Res.* (2017) 32:1829–40. doi: 10.1002/ jbmr.3172

105. Abdelaziz DM, Stone LS, Komarova SV. Osteolysis and pain due to experimental bone metastases are improved by treatment with rapamycin. *Breast Cancer Res Treat.* (2014) 143:227–37. doi: 10.1007/s10549-013-2799-0

106. Hussein O, Tiedemann K, Murshed M, Komarova SV. Rapamycin inhibits osteolysis and improves survival in a model of experimental bone metastases. *Cancer Lett.* (2012) 314:176–84. doi: 10.1016/j.canlet.2011.09.026

107. Okada H, Tanaka S. Plasmalemmal interface for calcium signaling in osteoclast differentiation. *Curr Opin Cell Biol.* (2022) 74:55–61. doi: 10.1016/j.ceb.2022.01.001

108. Huang R, Zheng J, Shao Y, Zhu L, Yang T. Siglec-15 as multifunctional molecule involved in osteoclast differentiation, cancer immunity and microbial infection. *Prog Biophys Mol Biol.* (2022). doi: 10.1016/J.PBIOMOLBIO.2022.10.006

109. Zhen G, Dan Y, Wang R, Dou C, Guo Q, Zarr M, et al. An antibody against siglec-15 promotes bone formation and fracture healing by increasing TRAP+ mononuclear cells and PDGF-BB secretion. *Bone Res.* (2021) 9(1):47. doi: 10.1038/S41413-021-00161-1

110. Kang FB, Chen W, Wang L, Zhang YZ. The diverse functions of siglec-15 in bone remodeling and antitumor responses. *Pharmacol Res.* (2020) 155:104728. doi: 10.1016/J.PHRS.2020.104728

111. Dou C, Zhen G, Dan Y, Wan M, Limjunyawong N, Cao X. Sialylation of TLR2 initiates osteoclast fusion. *Bone Res.* (2022) 10:24. doi: 10.1038/s41413-022-00186-0

112. Wu Y, Ai H, Xi Y, Tan J, Qu Y, Xu J, et al. Osteoclast-derived apoptotic bodies inhibit naive CD8(+) t cell activation *via* Siglec15, promoting breast cancer secondary metastasis. *Cell Rep Med.* (2023) 4:101165. doi: 10.1016/j.xcrm.2023.101165

113. Wang J, Sun J, Liu LN, Flies DB, Nie X, Toki M, et al. Siglec-15 as an immune suppressor and potential target for normalization cancer immunotherapy. *Nat Med.* (2019) 25:656–66. doi: 10.1038/s41591-019-0374-x

114. Jiang K-Y, Qi L-L, Liu X-B, Wang Y, Wang L. Prognostic value of siglec-15 expression in patients with solid tumors: A meta-analysis. *Front Oncol.* (2022) 12:1073932. doi: 10.3389/fonc.2022.1073932

115. Coleman RE. Bisphosphonates in breast cancer. Ann Oncol Off J Eur Soc Med Oncol. (2005) 16:687–95. doi: 10.1093/annonc/mdi162

116. Reyes C, Hitz M, Prieto-Alhambra D, Abrahamsen B. Risks and benefits of bisphosphonate therapies. J Cell Biochem. (2016) 117:20–8. doi: 10.1002/JCB.25266

117. Dearnaley DP, Collaborators TMP, Sydes MR, Collaborators TMP, Mason MD, Collaborators TMP, et al. A double-blind, placebo-controlled, randomized trial of oral sodium clodronate for metastatic prostate cancer (MRC PR05 trial). *JNCI J Natl Cancer Inst.* (2003) 95:1300–11. doi: 10.1093/JNCI/DJG038

118. Diel LJ, Jaschke A, Solomayer EF, Gollan C, Bastert G, Sohn C, et al. Adjuvant oral clodronate improves the overall survival of primary breast cancer patients with micrometastases to the bone marrow–a long-term follow-up. *Ann Oncol.* (2008) 19:2007. doi: 10.1093/ANNONC/MDN429

119. Saarto T, Vehmanen L, Blomqvist C, Elomaa I. Ten-year follow-up of 3 years of oral adjuvant clodronate therapy shows significant prevention of osteoporosis in earlystage breast cancer. J Clin Oncol. (2008) 26:4289–95. doi: 10.1200/JCO.2007.15.4997

120. Saarto T, Blomqvist C, Virkkunen P, Elomaa I. Adjuvant clodronate treatment does not reduce the frequency of skeletal metastases in node-positive breast cancer patients: 5-year results of a randomized controlled trial. *J Clin Oncol.* (2001) 19:10–7. doi: 10.1200/JCO.2001.19.1.10

121. Mason MD, Sydes MR, Glaholm J, Langley RE, Huddart RA, Sokal M, et al. Oral sodium clodronate for nonmetastatic prostate cancer-results of a randomized double-blind placebo-controlled trial: Medical research council PR04 (ISRCTN61384873). J Natl Cancer Inst. (2007) 99:765–76. doi: 10.1093/JNCI/DJK178

122. Kamba T, Kamoto T, Maruo S, Kikuchi T, Shimizu Y, Namiki S, et al. A phase III multicenter, randomized, controlled study of combined androgen blockade with versus without zoledronic acid in prostate cancer patients with metastatic bone disease: results of the ZAPCA trial. *Int J Clin Oncol.* (2017) 22:166–73. doi: 10.1007/S10147-016-1037-2

123. Wilson C, Bell R, Hinsley S, Marshall H, Brown J, Cameron D, et al. Adjuvant zoledronic acid reduces fractures in breast cancer patients; an AZURE (BIG 01/04) study. *Eur J Cancer.* (2018) 94:70–8. doi: 10.1016/J.EJCA.2018.02.004

124. Coleman R, Cameron D, Dodwell D, Bell R, Wilson C, Rathbone E, et al. Adjuvant zoledronic acid in patients with early breast cancer: final efficacy analysis of the AZURE (BIG 01/04) randomised open-label phase 3 trial. *Lancet Oncol.* (2014) 15:997–1006. doi: 10.1016/S1470-2045(14)70302-X

125. Eisen A, Somerfield MR, Accordino MK, Blanchette PS, Clemons MJ, Dhesy-Thind S, et al. Use of adjuvant bisphosphonates and other bone-modifying agents in breast cancer: ASCO-OH (CCO) guideline update. *J Clin Oncol.* (2022) 40:787–800. doi: 10.1200/JCO.21.02647

126. Alzahrani M, Stober C, Liu M, Awan A, Ng TL, Pond G, et al. Symptomatic skeletal-related events in patients receiving longer term bone-modifying agents for bone metastases from breast and castration resistant prostate cancers. *Support Care Cancer*. (2022) 30:3977–84. doi: 10.1007/S00520-021-06714-8

127. D'Oronzo S, Coleman R, Brown J, Silvestris F. Metastatic bone disease: Pathogenesis and therapeutic options: Up-date on bone metastasis management. J Bone Oncol. (2019) 15:4. doi: 10.1016/j.jbo.2018.10.004

128. Saylor PJ, Rumble RB, Tagawa S, Eastham JA, Finelli A, Reddy PS, et al. Bone health and bone-targeted therapies for prostate cancer: ASCO endorsement of a cancer care ontario guideline. *J Clin Oncol.* (2020) 38:1736–43. doi: 10.1200/JCO.19.03148

129. Wirth M, Tammela T, Cicalese V, Gomez Veiga F, Delaere K, Miller K, et al. Prevention of bone metastases in patients with high-risk nonmetastatic prostate cancer treated with zoledronic acid: efficacy and safety results of the zometa european study (ZEUS). *Eur Urol.* (2015) 67:482–91. doi: 10.1016/J.EURURO.2014.02.014

130. Saad F, Segal S, Eastham J. Prostate-specific antigen kinetics and outcomes in patients with bone metastases from castration-resistant prostate cancer treated with or without zoledronic acid. *Eur Urol.* (2014) 65:146–53. doi: 10.1016/J.EURURO.2012.05.007

131. Diel IJ, Solomayer E-F, Costa SD, Gollan C, Goerner R, Wallwiener D, et al. Reduction in new metastases in breast cancer with adjuvant clodronate treatment. N Engl J Med. (1998) 339:357–63. doi: 10.1056/NEJM199808063390601

132. Paterson AHG, Anderson SJ, Lembersky BC, Fehrenbacher L, Falkson CI, King KM, et al. Oral clodronate for adjuvant treatment of operable breast cancer (National surgical adjuvant breast and bowel project protocol b-34): a multicentre, placebocontrolled, randomised trial. *Lancet Oncol.* (2012) 13:734–42. doi: 10.1016/S1470-2045 (12)70226-7

133. Coleman R, Gray R, Powles T, Paterson A, Gnant M, Bergh J, et al. Adjuvant bisphosphonate treatment in early breast cancer: Meta-analyses of individual patient data from randomised trials. *Lancet*. (2015) 386:1353–61. doi: 10.1016/S0140-6736(15) 60908-4

134. Von Minckwitz G, Möbus V, Schneeweiss A, Huober J, Thomssen C, Untch M, et al. German adjuvant intergroup node-positive study: a phase III trial to compare oral ibandronate versus observation in patients with high-risk early breast cancer. *J Clin Oncol.* (2013) 31:3531–9. doi: 10.1200/JCO.2012.47.2167

135. Vliek SB, Noordhoek I, Meershoek-Klein Kranenbarg E, Van Rossum AGJ, Dezentje VO, Jager A, et al. Daily oral ibandronate with adjuvant endocrine therapy in postmenopausal women with estrogen receptor-positive breast cancer (BOOG 2006-

04): Randomized phase III TEAM-IIB trial. J Clin Oncol. (2022) 40(25):2934–45. doi: 10.1200/JCO.21.00311

136. Coleman R, Hall A, Albanell J, Hanby A, Bell R, Cameron D, et al. Effect of MAF amplification on treatment outcomes with adjuvant zoledronic acid in early breast cancer: a secondary analysis of the international, open-label, randomised, controlled, phase 3 AZURE (BIG 01/04) trial. *Lancet Oncol.* (2017) 18:1543–52. doi: 10.1016/S1470-2045(17)30603-4

137. Saag KG, Pannacciulli N, Geusens P, Adachi JD, Messina OD, Morales-Torres J, et al. Denosumab versus risedronate in glucocorticoid-induced osteoporosis: Final results of a twenty-Four-Month randomized, double-blind, double-dummy trial. *Arthritis Rheumatol (Hoboken NJ).* (2019) 71:1174–84. doi: 10.1002/art.40874

138. Miller PD, Pannacciulli N, Malouf-Sierra J, Singer A, Czerwiński E, Bone HG, et al. Efficacy and safety of denosumab vs. bisphosphonates in postmenopausal women previously treated with oral bisphosphonates. *Osteoporos Int.* (2020) 31:181–91. doi: 10.1007/S00198-019-05233-X

139. Smith MR, Coleman RE, Klotz L, Pittman K, Milecki P, Ng S, et al. Denosumab for the prevention of skeletal complications in metastatic castration-resistant prostate cancer: comparison of skeletal-related events and symptomatic skeletal events. *Ann Oncol Off J Eur Soc Med Oncol.* (2015) 26:368–74. doi: 10.1093/annonc/mdu519

140. Smith MR, Saad F, Oudard S, Shore N, Fizazi K, Sieber P, et al. Denosumab and bone metastasis-free survival in men with nonmetastatic castration-resistant prostate cancer: exploratory analyses by baseline prostate-specific antigen doubling time. *J Clin Oncol Off J Am Soc Clin Oncol.* (2013) 31:3800–6. doi: 10.1200/JCO.2012.44.6716

141. Shapiro CL, Moriarty JP, Dusetzina S, Himelstein AL, Foster JC, Grubbs SS, et al. Cost-effectiveness analysis of monthly zoledronic acid, zoledronic acid every 3 months, and monthly denosumab in women with breast cancer and skeletal metastases: CALGB 70604 (Alliance). J Clin Oncol Off J Am Soc Clin Oncol. (2017) 35:3949–55. doi: 10.1200/JCO.2017.73.7437

142. Brown JE, Coleman RE. Denosumab in patients with cancer-a surgical strike against the osteoclast. *Nat Rev Clin Oncol.* (2012) 9:110-8. doi: 10.1038/ NRCLINONC.2011.197

143. Smith MR, Saad F, Coleman R, Shore N, Fizazi K, Tombal B, et al. Denosumab and bone-metastasis-free survival in men with castration-resistant prostate cancer: results of a phase 3, randomised, placebo-controlled trial. *Lancet (London England).* (2012) 379:39–46. doi: 10.1016/S0140-6736(11)61226-9

144. Peters S, Danson S, Hasan B, Dafni U, Reinmuth N, Majem M, et al. A randomized open-label phase III trial evaluating the addition of denosumab to standard first-line treatment in advanced NSCLC: The european thoracic oncology platform (ETOP) and european organisation for research and treatment of cancer (EORTC) SPLENDOU. *J Thorac Oncol.* (2020) 15:1647–56. doi: 10.1016/J.JTHO.2020.06.011

145. Lipton A, Fizazi K, Stopeck AT, Henry DH, Smith MR, Shore N, et al. Effect of denosumab versus zoledronic acid in preventing skeletal-related events in patients with bone metastases by baseline characteristics. *Eur J Cancer*. (2016) 53:75–83. doi: 10.1016/J.EJCA.2015.09.011

146. Lamy O, Stoll D, Aubry-Rozier B, Rodriguez EG. Stopping denosumab. Curr Osteoporos Rep. (2019) 17:8–15. doi: 10.1007/S11914-019-00502-4

147. Hannon RA, Clack G, Rimmer M, Swaisland A, Andrew Lockton J, Finkelman RD, et al. Effects of the src kinase inhibitor saracatinib (AZD0530) on bone turnover in healthy men: a randomized, double-blind, placebo-controlled, multiple-ascending-dose phase i trial. *J Bone Miner Res.* (2010) 25:463–71. doi: 10.1359/JBMR.090830

148. Roskoski R. Src protein-tyrosine kinase structure, mechanism, and small molecule inhibitors. *Pharmacol Res.* (2015) 94:9–25. doi: 10.1016/J.PHRS.2015.01.003

149. Yu EY, Wilding G, Posadas E, Gross M, Culine S, Massard C, et al. Phase II study of dasatinib in patients with metastatic castration-resistant prostate cancer. *Clin Cancer Res.* (2009) 15:7421–8. doi: 10.1158/1078-0432.CCR-09-1691

150. Araujo JC, Trudel GC, Saad F, Armstrong AJ, Yu EY, Bellmunt J, et al. Docetaxel and dasatinib or placebo in men with metastatic castration-resistant prostate cancer (READY): a randomised, double-blind phase 3 trial. *Lancet Oncol.* (2013) 14:1307. doi: 10.1016/S1470-2045(13)70479-0

151. Schott AF, Barlow WE, Van Poznak CH, Hayes DF, Moinpour CM, Lew DL, et al. Phase II studies of two different schedules of dasatinib in bone metastasis predominant metastatic breast cancer: SWOG S0622. *Breast Cancer Res Treat.* (2016) 159:87–95. doi: 10.1007/s10549-016-3911-z

152. Campone M, Bondarenko I, Brincat S, Hotko Y, Munster PN, Chmielowska E, et al. Phase II study of single-agent bosutinib, a Src/Abl tyrosine kinase inhibitor, in patients with locally advanced or metastatic breast cancer pretreated with chemotherapy. Ann Oncol Off J Eur Soc Med Oncol. (2012) 23:610–7. doi: 10.1093/ annon/mdr261

153. Antonarakis ES, Heath EI, Posadas EM, Yu EY, Harrison MR, Bruce JY, et al. A phase 2 study of KX2-391, an oral inhibitor of src kinase and tubulin polymerization, in men with bone-metastatic castration-resistant prostate cancer. *Cancer Chemother Pharmacol.* (2013) 71:883–92. doi: 10.1007/s00280-013-2079-z

154. Oswald AJ, Symeonides SN, Wheatley D, Chan S, Brunt AM, McAdam K, et al. Aromatase inhibition plus/minus src inhibitor saracatinib (AZD0530) in advanced breast cancer therapy (ARISTACAT): a randomised phase II study. *Breast Cancer Res Treat.* (2023) 199(1):35–46. doi: 10.1007/S10549-023-06873-8

155. Dai R, Wu Z, Chu HY, Lu J, Lyu A, Liu J, et al. Cathepsin k: The action in and beyond bone. *Front Cell Dev Biol.* (2020) 8:433. doi: 10.3389/fcell.2020.00433

156. Jensen AB, Wynne C, Ramirez G, He W, Song Y, Berd Y, et al. The cathepsin k inhibitor odanacatib suppresses bone resorption in women with breast cancer and established bone metastases: results of a 4-week, double-blind, randomized, controlled trial. *Clin Breast Cancer*. (2010) 10:452–8. doi: 10.3816/CBC.2010.N.059

157. McClung MR, O'Donoghue ML, Papapoulos SE, Bone H, Langdahl BL, Saag KG, et al. Odanacatib for the treatment of postmenopausal osteoporosis: results of the LOFT multicentre, randomised, double-blind, placebo-controlled trial and LOFT extension study. *Lancet Diabetes Endocrinol.* (2019) 7:899–911. doi: 10.1016/S2213-8587(19)30346-8

158. Simone V, Ciavarella S, Brunetti O, Savonarola A, Cives M, Tucci M, et al. Everolimus restrains the paracrine pro-osteoclast activity of breast cancer cells. *BMC Cancer*. (2015) 15:692. doi: 10.1186/S12885-015-1717-8

159. Gnant M, Baselga J, Rugo HS, Noguchi S, Burris HA, Piccart M, et al. Effect of everolimus on bone marker levels and progressive disease in bone in BOLERO-2. *J Natl Cancer Inst.* (2013) 105:654–63. doi: 10.1093/JNCI/DJT026

160. Maass N, Harbeck N, Mundhenke C, Lerchenmüller C, Barinoff J, Lück HJ, et al. Everolimus as treatment for breast cancer patients with bone metastases only: results of the phase II RADAR study. *J Cancer Res Clin Oncol.* (2013) 139:2047–56. doi: 10.1007/S00432-013-1518-X

161. Sandor LF, Huh JB, Benko P, Hiraga T, Poliska S, Dobo-Nagy C, et al. *De novo* steroidogenesis in tumor cells drives bone metastasis and osteoclastogenesis. *Cell Rep.* (2024) 43:113936. doi: 10.1016/j.celrep.2024.113936

162. Yang H, Yu Z, Ji S, Huo Q, Yan J, Gao Y, et al. Targeting bone microenvironments for treatment and early detection of cancer bone metastatic niches. *J Control Release*. (2022) 341:443–56. doi: 10.1016/J.JCONREL.2021.11.005