



OPEN ACCESS

EDITED BY

Vuong Trieu,
Oncotelic, Inc., United States

REVIEWED BY

Dawid Sigorski,
University of Warmia and Mazury in Olsztyn,
Poland
Christine Alewine,
National Cancer Institute (NIH), United States
Mayrel Palestino Dominguez,
NIH/NCI Center for Cancer Research/
Laboratory of Molecular Biology,
United States, in collaboration with reviewer
CA

*CORRESPONDENCE

Xin Li
✉ lixin16@sdu.edu.cn

†These authors have contributed
equally to this work and share
first authorship

RECEIVED 12 December 2023

ACCEPTED 15 March 2024

PUBLISHED 28 March 2024

CITATION

Li K, Zhou Y, Zang M, Jin X and Li X (2024)
Therapeutic prospects of nectin-4 in cancer:
applications and value.
Front. Oncol. 14:1354543.
doi: 10.3389/fonc.2024.1354543

COPYRIGHT

© 2024 Li, Zhou, Zang, Jin and Li. This is an
open-access article distributed under the terms
of the [Creative Commons Attribution License
\(CC BY\)](https://creativecommons.org/licenses/by/4.0/). The use, distribution or reproduction
in other forums is permitted, provided the
original author(s) and the copyright owner(s)
are credited and that the original publication
in this journal is cited, in accordance with
accepted academic practice. No use,
distribution or reproduction is permitted
which does not comply with these terms.

Therapeutic prospects of nectin-4 in cancer: applications and value

Kaiyue Li^{1†}, Yujing Zhou^{1†}, Maolin Zang^{2†}, Xin Jin³ and Xin Li^{1*}

¹Department of Nuclear Medicine, Qilu Hospital of Shandong University, Jinan, Shandong, China,

²Department of Urology, Qilu Hospital of Shandong University, Jinan, Shandong, China, ³Imaging Center, Jinan Third People's Hospital, Jinan, Shandong, China

Nectin-4 is a Ca²⁺-independent immunoglobulin-like protein that exhibits significantly elevated expression in malignant tumors while maintaining extremely low levels in healthy adult tissues. In recent years, overexpression of Nectin-4 has been implicated in tumor occurrence and development of various cancers, including breast cancer, urothelial cancer, and lung cancer. In 2019, the Food and Drug Administration approved enfortumab vedotin, the first antibody–drug conjugate targeting Nectin-4, for the treatment of urothelial carcinoma. This has emphasized the value of Nectin-4 in tumor targeted therapy and promoted the implementation of more clinical trials of enfortumab vedotin. In addition, many new drugs targeting Nectin-4 for the treatment of malignant tumors have entered clinical trials, with the aim of exploring potential new indications. However, the exact mechanisms by which Nectin-4 affects tumorigenesis and progression are still unclear, and the emergence of drug resistance and treatment-related adverse reactions poses challenges. This article reviews the diagnostic potential, prognostic significance, and molecular role of Nectin-4 in tumors, with a focus on clinical trials in the field of Nectin-4-related tumor treatment and the development of new drugs targeting Nectin-4.

KEYWORDS

cancer, Nectin-4, therapy, biomarker, ADC

1 Introduction

Tumor-specific biomarkers encompass a wide variety of molecular types, including DNA, metabolites, mRNA, and cell surface molecules (1), which exert distinct influences on tumor development. Cell surface molecules play crucial parts in tumor proliferation and metastasis through adhesion mechanisms (2). Nectin-4 is a type I transmembrane adhesion molecule belonging to the immunoglobulin superfamily (3) and forms part of the nectin protein family together with Nectin-1, Nectin-2, and Nectin-3. Nectin-mediated cell adhesion is characterized by its independence from Ca²⁺ and its ability to engage in both homophilic and heterophilic interactions through its outer domain via *trans*-interactions (4–6). Nectin-1 and Nectin-2 are typically found in immune organs such as bone marrow, thymus, and spleen, whereas Nectin-3

is expressed mainly in the placenta and spermatozoa (4, 5). Under normal physiological conditions, Nectin-4 is expressed in the placenta and embryo, with low levels in healthy tissues and cells of adults (4). However, it has been reported to be highly expressed in various malignant solid tumors and to be associated with cancer progression and prognosis, establishing it as a novel cancer biomarker (7–10). Nectin-4 is involved in tumor cell development, including adhesion, proliferation, migration, and angiogenesis (7–11), and it has prognostic significance in numerous cancers (12–16). In 2019, the Food and Drug Administration (FDA) approved the first antibody-drug conjugate (ADC) targeting Nectin-4, known as enfortumab vedotin (EV), for treating urothelial carcinoma. Its clinical efficacy is a current research hotspot, as are its susceptibility to drug resistance and the potential adverse reactions that it can cause (17–19). In addition, its value in the diagnosis and prognosis of cancer is being widely explored. This review aims to compile and summarize the current state of research on Nectin-4, focusing on its diagnostic utility, prognostic significance, and its role in cancer treatment. Furthermore, the review offers an analysis of potential future research directions and prospective clinical applications for Nectin-4.

2 Overview of Nectin-4

Nectin-4 is a type I transmembrane polypeptide and a member of the immunoglobulin superfamily; it is encoded by the poliovirus receptor related-4 (PVRL4) gene and is 510 amino acids in full length (3, 20). The structure of Nectin-4, similar to other members of the nectin family, comprises an extracellular region, a transmembrane region, and a cytoplasmic tail, with the extracellular domain possessing three immunoglobulin-like loops (1 V-type and 2 C2-types) (21, 22). Notably, the adhesion of nectins does not depend on Ca^{2+} , as it involves a homophilic and heterophilic process mediated by the outer domain through a *trans*-interaction (5). Notably, in addition to the traditional membrane protein form, Nectin-4 has a soluble form. Soluble Nectin-4 (43 kDa) is formed by shedding of the entire Nectin-4 extracellular domain under conditions of cellular hypoxia, a process in which a disintegrin and metalloproteinase (ADAM) has a vital role (23). Soluble Nectin-4 has been shown to interact with integrin-

$\beta 4$ on endothelial cells, promoting angiogenesis through signaling pathways involving Src, PI3K, Akt, and inducible nitric oxide (NO) synthase (24, 25). Initially, Nectin-4 was recognized as a viral entry receptor (22, 26–28). However, subsequent research unveiled its role as a cell adhesion protein closely related to the development of various diseases, in particular, cancers. The anchorage independence between tumor cells is achieved by PVRL4 through driving of intercellular adhesion and stromal independent integrin- $\beta 4$ /Src homology-2-containing protein tyrosine phosphatase 2 (SHP-2)/cellular Src activation, a process facilitated by the extracellular segment of Nectin-4 (7). In general, Nectin-4 shows little expression in healthy adult tissues, but it is highly enriched in embryonic and placental tissues. Nectin-4 is closely related to ectodermal development. Double allelic mutations in the PVRL gene cause ectodermal dysplasia-syndactyly syndrome 1, which is characterized by syndactyly (29–31). Several recent studies have confirmed that Nectin-4 is highly expressed in various solid tumors and is involved in their initiation and progression (32). Challita-Eid et al. used immunohistochemistry to analyze 2394 specimens from patients with various tumors, including bladder, breast, and ovarian cancers, and found that 69% of all samples stained positive for Nectin-4 (33). Although the precise mechanism by which Nectin-4 participates in cancer progression is unclear, some studies have shown that its intracellular region can physically interact with importin- $\alpha 2$ and transfer to the nucleus to enhance DNA repair, whereas the extracellular region can increase angiogenesis through various molecular pathways (34). In addition, several studies have confirmed the involvement of Nectin-4 in migration, adhesion, and proliferation of tumor cells (11, 35, 36). Nectin-4 is also involved in various benign diseases including endometriosis (37, 38). Figure 1 shows the biological functions and structure of Nectin-4.

3 Molecular role of Nectin-4 in carcinogenesis

PI3K/AKT is a crucial signaling pathway involved in tumor development, regulating processes including cell invasion, proliferation, and apoptosis (39–41). Nectin-4 participates in several important processes in tumors through this pathway. For

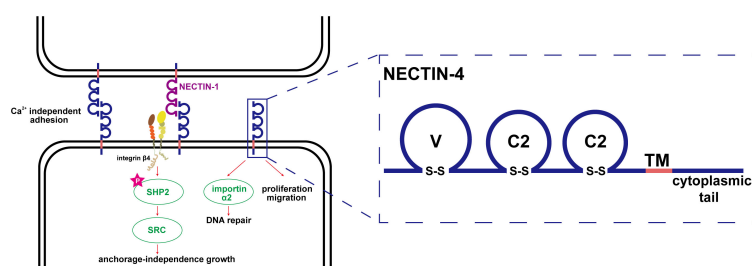


FIGURE 1

Biological function and structure of Nectin-4. The adhesion of nectins is Ca^{2+} independent, which is a homophilic and heterophilic process mediated by the outer domain through *trans*-interaction; the anchorage independence growth between tumor cells is achieved by PVRL4 through driving intercellular adhesion and stromal independent integrin- $\beta 4$ /SHP-2/c-Src activation, a process facilitated by the extracellular segment of Nectin-4; Nectin-4 can physically interact with importin- $\alpha 2$ to transfer to the nucleus to enhance DNA repair; the structure of Nectin-4 was shown in the right part of the figure.

example, soluble Nectin-4, formed by the shedding of the extracellular domain under hypoxic conditions, interacts with integrin- β 4 on endothelial cells to promote angiogenesis; this is achieved through the Src, PI3K, Akt, and endothelial NO synthase (eNOS) pathways (24) and may be related to PI3K-Akt-mediated NO formation. This was confirmed by a further study on oral cancer, in which addition of NO enhanced Nectin-4-mediated expression of eNOS and induced angiogenesis. However, Nectin-4 no longer induced angiogenesis after PI3K-Akt-mediated inhibition of the eNOS pathway (25). In addition, Nectin-4 has been reported to promote progression and metastasis of many tumors through the PI3K/AKT pathway. In breast cancer, Nectin-4 and tyrosine kinase receptor ErbB2 (also known as Her2) are highly expressed and can *cis*-interact with each other to activate the PI3K/AKT signaling pathway for DNA synthesis (42). Wnt/ β -catenin signaling pathway is a self-renewal pathway in breast cancer stem cells. Siddharth et al. found that activation of this pathway was induced by soluble Nectin-4 through the PI3K/AKT axis and promoted the proliferation and metastasis of breast cancer cells (43). However, their study did not explore this process in detail. In August 2023, it was proposed that PI3K α signaling may promote INPP4B-dependent lysosomal degradation, thereby enhancing activation of the Wnt/ β -catenin signaling pathway (44). Moreover, Nectin-4 can downregulate miR-520c-3p, a microRNA, to activate the PI3K/AKT/nuclear factor kappa-B (NF- κ B) pathway, promoting the progression and metastasis of osteosarcoma (45). In gastric and gallbladder cancers, Nectin-4 activates Ras-related C3 botulinum toxin substrate 1 (Rac1) through the PI3K/AKT pathway, promoting tumor proliferation and metastasis (46, 47). In addition to participating in tumor progression, Nectin-4 can mediate drug resistance through the PI3K/AKT pathway. Das et al. demonstrated upregulation of Nectin-4 in colon cancer cells after long-term treatment with standard chemotherapy agent 5-FU (48). Nectin-4 and AFADIN can form a complex that plays a part in intercellular junction formation (49). This complex can activate the PI3K/AKT cascade, leading to enhanced tumor cell survival and suppression of apoptosis (48). Thus, multiple studies have demonstrated that Nectin-4 is implicated in various aspects of tumorigenesis and progression via the PI3K/AKT pathway, suggesting potential novel strategies for future tumor treatment. Figure 2 shows the process by which Nectin-4 regulates cancer development through the PI3K/AKT pathway and promotes angiogenesis after hypoxia.

Nectin-4 is involved in the transduction process of other signaling pathways in addition to the PI3K/AKT pathway. Normally, the extracellular domain of Nectin-4 also *cis*-interacts with the prolactin receptor, which is necessary for mammary follicle development, a process dependent on the Janus kinase 2-signal transducer and activator of transcription 5a (JAK2-STAT5a) signaling pathway (50). The JAK2-STAT5a pathway is vital for regulation of the growth of certain malignant tumors, including breast and prostate cancers (51). Nectin-4 also enhances p95-ErbB2-induced activation of the JAK-STAT3 signaling pathway (42) and cooperates with p95-ErbB2 to regulate expression of the sex determining region Y-box 2 (SOX2) gene, which in turn regulates the proliferation, survival, and differentiation of cancer

cells, thereby enhancing the proliferation of breast cancer T47D cells (52). Moreover, Nectin-4 significantly promotes tumor-induced lymphangiogenesis and lymphatic metastasis by regulating the C-X-C motif chemokine receptor 4 (CXCR4)/C-X-C motif chemokine ligand 12 (CXCL12)-lymphatic vessel endothelial receptor-1 (LYVE-1) axis (53). These findings highlight the multifaceted involvement of Nectin-4 in various molecular pathways that contribute to the development and progression of cancer, underscoring its significance as a potential target for cancer therapy and research.

4 Diagnostic potential of Nectin-4

4.1 Serodiagnosis

Soluble and transmembrane forms of nectin proteins have been described in humans and rodents (4, 54–57). Soluble Nectin-4 was detected in the serum of patients with metastatic breast cancer and in the supernatants of breast cancer cell lines, with similar immunological and biochemical profiles, in a study demonstrating that the overexpression or silencing of tumor necrosis factor- α convert enzyme (TACE; also known as ADAM-17) enhances or reduces Nectin-4 shedding, respectively. Overexpression of TACE in breast cancer indicates that the formation of soluble Nectin-4, both *in vitro* and *in vivo*, can be attributed to TACE (58). Similarly, soluble Nectin-4 has been detected in the serum of patients with ovarian cancer, and it can be used, with CA-125, to identify benign ovarian disease and ovarian cancer (9). There is evidence that serum Nectin-4 levels are associated with the grade and progression of ovarian cancer (59). Notably, Nectin-4 can be detected in the early stages of ovarian cancer, even when CA-125 levels are not significantly elevated in the serum (60). Although soluble Nectin-4 has great potential and value in the diagnosis and monitoring of ovarian cancer, it still requires large-scale prospective studies for further validation and standardization.

Serum Nectin-4 may have diagnostic value for non-small-cell lung cancer (NSCLC). Takano et al. used mouse monoclonal antibodies (mAbs) to establish an enzyme-linked immunosorbent assay system to detect serum Nectin-4 levels. They found that serum levels of Nectin-4 in 164 patients with NSCLC were considerably higher than those of healthy volunteers; the percentage of Nectin-4 positivity in patients with NSCLC was 53.7%, whereas only three of the 131 healthy volunteers displayed false positivity, proving the superiority of Nectin-4 in terms of diagnostic sensitivity and specificity compared with carcinoembryonic antigen (CEA) and cytokeratin 19-fragment (CYFRA21-1) (8). However, it is worth noting that Nectin-2 seems to have an advantage over Nectin-4 in diagnosis of lung cancer (61). Thus, serum Nectin-4 is meaningful for NSCLC, but the evidence is not sufficient to support it as a serological marker.

As well as these potential applications in cancer diagnosis, increased levels of soluble Nectin-4 in serum may contribute to the diagnosis and monitoring of asthma (62) and may also represent a new biomarker for opioid dependence (63, 64). To date, most research has focused on Nectin-4 as a transmembrane protein of tumor cells, with less attention paid to soluble Nectin-4.

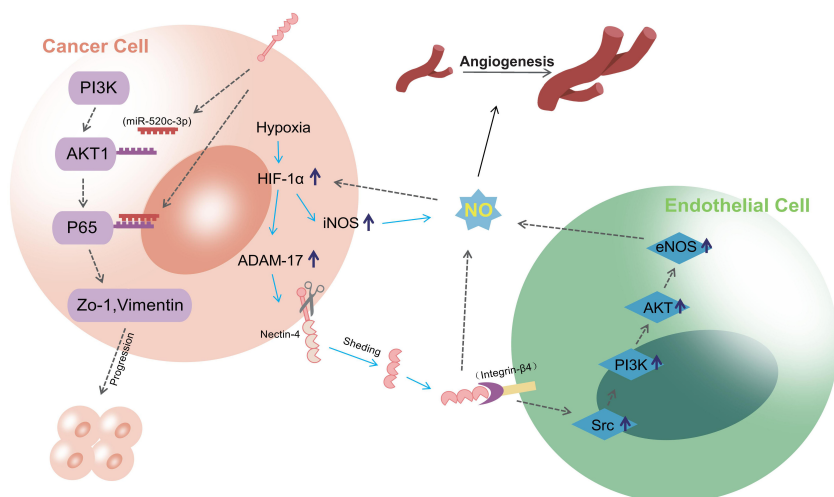


FIGURE 2

the process of Nectin-4 regulating cancer development through PI3K/AKT pathway and promoting angiogenesis after hypoxia. Nectin-4 can downregulate miR-520c-3p to activate the PI3K/AKT/NF- κ B pathway, promoting the progression and metastasis; Soluble nectin-4, which is formed by the shedding of Nectin-4 extracellular domain during hypoxia, can interact with integrin- β 4 on endothelial cells to promote angiogenesis. This progress is realized via the Src, PI3K, Akt, and eNOS pathways, and may be related to PI3K-Akt-mediated NO formation.

More research is required to explore the serological value of soluble Nectin-4 in other cancers.

4.2 Target sites for molecular probes

Dean et al. designed an immuno-positron emission tomography (immunoPET) molecular probe targeting Nectin-4 for preclinical evaluation in tumor-bearing mice, based on the finding that Nectin-4 is highly expressed in several cancers with little expression in normal adult tissue (65). The probe was labeled with [89Zr] for AGS-22M6 (an anti-Nectin-4 mAb) and used for *in vivo* tumor detection in mice and [18F] for assessing the biodistribution of the probe in crab-eating monkeys (65). This was the first experiment in which a molecular probe was developed using Nectin-4 as a target. The new immunoPET probe developed in this study showed good detection of liver and bone lesions and could be used to evaluate expression levels of Nectin-4. In 2022, diagnostic and therapeutic pairs were developed based on an anti-Nectin-4 mAb, namely (99m)Tc-HYNIC-mAb (Nectin-4) and mAb (Nectin-4)-ICG. Of these, (99m)Tc-HYNIC-mAb (Nectin-4) has been used in immunological single-photon emission computed tomography for diagnosis and classification of triple-negative breast cancer (TNBC) and verified to have high targeting properties (66). mAb (Nectin-4)-ICG has been used for photothermal therapy of tumors. These imaging probes all use a mAb as the carrier; this shows good affinity but has a large molecular weight and slow blood clearance rate. To avoid the accumulation of imaging agents in organs, the selection of immune probe vectors may need to focus more on small-molecular-weight proteins such as antibody fragments or nanobodies in the future. A bicyclic peptide-based Nectin-4-targeted PET radiotracer,

68Ga-N188, was recently designed. Preclinical evaluation of this probe was completed in a mouse tumor model, followed by successful clinical translation to patients with advanced urothelial cancer and healthy volunteers (67). This marked the first clinical application of Nectin-4 as an imaging target. Although the value of Nectin-4 as a diagnostic target has been confirmed in clinical practice, the uptake of ⁶⁸Ga-N188 in positive tumor lesions was moderate owing to its low affinity. The structure of N-188 needs to be further modified to improve its affinity with Nectin-4. Non-invasive monitoring of Nectin-4 expression could be beneficial for evaluating tumor progression and guiding treatment. This could benefit cancer patients with high expression of Nectin-4, such as those with locally advanced or metastatic urothelial carcinoma (la/mUC). It is expected that Nectin-4 will continue to be a focus of attention in the field of medical imaging, offering promising opportunities for improved diagnosis and monitoring of various diseases, particularly cancer.

5 Association between Nectin-4 and prognosis

Numerous studies have suggested a possible association between Nectin-4 expression and the prognosis of patients with various cancers, particularly breast cancer. Survivin is expressed in various tumor cells, and in breast cancer cells, it is correlated with the expression of Nectin-4. Both proteins are involved in tumor cell proliferation and differentiation, and in inhibition of tumor cell apoptosis, and they are both independent prognostic markers for breast cancer (68, 69). Therefore, exploring the interaction between Nectin-4 and survivin in breast cancer may provide value for the

treatment of breast cancer in the future. A study on 197 patients with primary unilateral breast cancer (without involved lymph nodes or distant metastasis) found that 34 patients had Nectin-4 expression on the tumor cell membrane. Of the remaining 163 tumors that were negative for membrane Nectin-4 expression, 74.8% showed high cytoplasmic expression (12). This study also found that Nectin-4 positivity on the membrane and in the cytoplasm was associated with various survival indicators, indicating poor outcomes (12). TNBC tumor, which lack estrogen receptor, human epidermal growth factor receptor-2, and progesterone receptor, are more challenging to treat compared with other subtypes of breast cancer. Therefore, identifying new targets is crucial for assessing the prognosis of TNBC patients and guiding their treatment. Protein expression of Nectin-4 appears to be present in around 62% of TNBC cases and shows a strong correlation with mRNA expression levels. Moreover, high PVRL4 mRNA expression has independent negative prognostic value for metastasis-free survival in TNBC patients (13). Zeindler et al. used immunohistochemistry to explore the relationship between Nectin-4 expression and prognosis of TNBC patients. They found that high expression of Nectin-4 protein was correlated with lower tumor stage, better overall survival (OS), and negative lymph nodes and predicted better outcomes in patients with TNBC (70). Owing to the complexity of tumor cells, contradictory results may be obtained when using mRNA and protein expression levels of Nectin-4 to predict prognosis. More studies are needed to clarify the relationship between Nectin-4 and prognosis in TNBC.

Nectin-4 is associated with poor outcomes in various other cancers. For example, in pancreatic cancer, Nectin-4 shows significant correlation with the Ki67 value-added index and vascular endothelial growth factor expression, indicating its association with a prognosis (11). Deng et al. reported that increasing Nectin-4 expression in esophageal cancer cell lines promoted cell viability, migration, invasion, and tumor formation (14). This finding also supported the results of another study in which, among 94 patients with esophageal cancer, patients with increased Nectin-4 expression had shorter OS than those with low expression (71). Nectin-4 is upregulated in colorectal cancer tissues and associated with integrin β -1 expression and vasogenic mimicry formation (36). Vasogenic mimicry serves as an alternative method of arterial supply in tumors and is often associated with a poor prognosis. Strong expression of Nectin-4 is associated with high disease control rate (DCR) (72), and high expression of Nectin-4 in other cancers, including NSCLC, hepatocellular carcinoma, and gastric cancer, has been associated with poor patient outcomes (8, 15, 16). However, not all findings point in the same direction. For example, a study on luminal B breast cancer demonstrated a negative correlation between expression levels of Nectin-4 and survival time, but there was no statistically significant correlation with tumor differentiation, lymph node metastasis, histological subtype, or Ki-67 proliferation index (73). Moreover, Nectin-4 may not be suitable as a prognostic indicator in papillary renal cell carcinoma, in which it appears to have no statistically significant relationship with age, grade, or TNM stage (74). Expression level of a given protein may vary greatly between different tumors and can be affected by many

factors including the diversity of tumor cells and the microenvironment. This could be why Nectin-4 does not show prognostic value in all tumor types. Table 1 shows the associations of Nectin-4 with prognosis in different cancers.

6 Oncological therapeutic value of Nectin-4

6.1 Oncolytic measles virus

Oncolytic viruses, which are attenuated strains that can infect the body without causing serious disease, can utilize the immune system to recognize and kill tumor cells (75). Nectin-4 has long been a focal point for researchers as a specific therapeutic target, and its endocytotic characteristics have served as a foundation for various therapeutic studies (76). Measles virus (MV) has been a particularly active area of research in viral oncolytic therapy. An MV attenuated vaccine is an ideal candidate for oncolytic virus therapy and infects host cells by binding to three receptors: CD46, signaling lymphocytic activation molecule (SLAM), and Nectin-4. Fujiyuki et al. produced a recombinant MV (rMV-SLAMblind) that bound to Nectin-4 but not its original primary receptor SLAM; it also infected and suppressed the growth of subcutaneous xenograft models of various cancers, including breast cancers such as TNBC (77–80). Currently, seven early-stage clinical trials are actively exploring the effects of transgenic and vaccine strains of MV on head and neck cancer, multiple myeloma, and ovarian cancer (81). CD46 and Nectin-4 have been reported to be highly expressed on the surface of ovarian cancer cells, and MV has displayed a remarkable antitumor effect in a mouse model of ovarian cancer. Evanthia et al. used an oncolytic MV encoding the sodium iodide symporter (MV-NIS) gene to treat 16 patients with paclitaxel- and platinum-resistant ovarian cancer (82). The patients' median OS (mOS) reached 26.5 months, and no dose-related adverse reactions were reported during high-dose treatment by injection. Immunological tests suggested that MV-NIS stimulated the body's cellular immunity to mediate the antitumor effect (82). Considering the limited clinical success rate, scholars are exploring methods to strengthen MV-induced viral oncolytic therapy. For example, studies have demonstrated that small molecules such as baicalein and cinnamaldehyde can enhance the oncolytic effects of MV strains on breast cancer (83). Furthermore, ursolic acid (UA) is a phytochemical with great potential in the treatment of breast cancer, and UA nanoparticles developed by nanoemulsification technology have exhibited enhanced drug solubility. A combination of UA and MV therapy was reported to enhance the killing of breast cancer cells by increasing autophagic flux, thereby providing an improved therapeutic effect (84). Although oncolytic viruses have the potential to treat cancer, the complexity and heterogeneity of cancer cells make it impossible to completely kill them by this treatment alone. Developing personalized treatment plans based on patients' immune status and accurately delivering drugs to the tumor site are the most urgent aspects to be addressed to improve therapeutic effects.

TABLE 1 Association between Nectin-4 and tumours prognosis.

Cancer type	Number of patients	Nectin-4+%	prognosis	Year(Ref.)
Gallbladder cancer	68	63.2% +	reduced OS	2016/ (46)
Invasive breast cancer	140	64.3% +	poor prognosis	2011/ (68)
Luminal-A breast cancer	197	1.membrane Nectin-4: 17.3% + 2.cytoplasmic Nectin-4: 62% +	1.poor prognosis, reduced DFS and DRFS 2.prolonged DFS and LRFS	2014/ (12)
TNBC	290		poor prognosis, reduced MFS	2017/ (13)
TNBC	148	58% +	better OS	2019/ (70)
Luminal-B breast cancer (HER2 negative)	147		reduced OS, DFS and DRFS	2017/ (73)
Esophageal cancer	82	74.4% +	poor prognosis, reduced OS	2009/ (14)
Esophageal cancer	82	70% +	reduced OS	2019/ (71)
Urothelial cancer	47	59.6% +	higher DCR	2022/ (72)
Hepatocellular carcinoma	87	Cytoplasmic Nectin-4 67.82% +	reduced DFS and OS	2016/ (8, 15, 16)
Gastric cancer	212	membrane/cytoplasmic Nectin-4 60.4% +	reduced OS	2018/ (8, 15, 16)
Papillary renal cell carcinoma	190(type 1 of pRCC)	48.4% +	prolonged 5 years OS	2022/ (74)
	107(type 2 of pRCC)	36.4% +	independence	

TNBC, Triple-negative breast cancers; OS, overall survival; DFS, disease-free survive; DRFS, distance recurrence-free survive; LRFS, local recurrence-free survive; MFS, metastasis-free survival; DCR, disease control rate; pRCC, Papillary renal cell carcinoma.

6.2 EV-ADC targeting Nectin-4

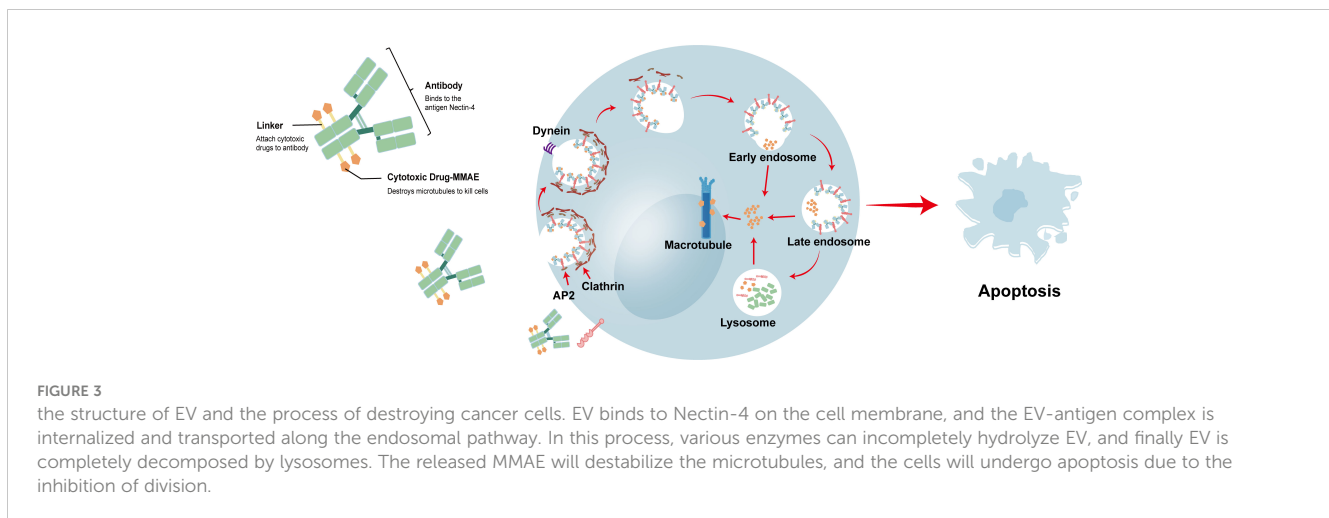
ADCs combine a small-molecule anticancer agent with a mAb to selectively deliver cytotoxic agents to tumor targets with increased efficacy and reduced toxicity. Nectin-4 has emerged as an attractive target for ADC therapy, and in 2019, the FDA approved the first ADC (EV) targeting Nectin-4 for the treatment of urothelial carcinoma (18). EV is a complex formed by the protease-dependent linker binding microtubule-disrupting agent monomethyl auristatin E (MMAE) with anti-Nectin-4 mAb (AGS-22M6) (85). EV binds to Nectin-4 on the cell membrane, and the EV-antigen complex is internalized and transported along the endosomal pathway. During this process, EV is initially hydrolyzed by various enzymes, before finally being completely decomposed by lysosomes. This releases MMAE, which destroys microtubules, causing the cells to undergo apoptosis owing to inhibition of division (18, 76, 86). Figure 3 shows the structure of EV and the process by which it destroys cancer cells. In preclinical research, EV substantially inhibited tumor growth in a mouse xenograft model of human bladder, breast, lung, and pancreatic cancers (33). This outcome has driven the translation of EV into clinical research.

6.3 Clinical trials of EV

EV-101 (NCT02091999) was a trial conducted in patients with mUC, with the primary objectives of evaluating drug safety,

pharmacokinetics, and patient tolerance. A total of 155 patients were recruited for the EV-101 study. In phase I, patients received incremental doses of EV of up to 1.25 mg/kg on days 1, 8, and 15 of each 28-day cycle (87). Encouraging survival data from this phase suggest a recommended phase 2 dose of 1.25 mg/kg on days 1, 8, and 15 of every 28 days. Further, 112 patients received EV at the preliminary recommended phase 2 dose. In this study, the objective response rate (ORR) was 43%, the mOS was 12.3 months, and the median progression-free survival (mPFS) was 5.4 months. The most common treatment-related adverse events (TRAEs) were rash, peripheral neuropathy, and fatigue. Four deaths occurred owing to continued treatment in this study (respiratory failure, urinary tract obstruction, diabetic ketoacidosis, and multiorgan failure) (87). Overall, the EV-101 study provided valuable insights into the potential of EV as a treatment option for mUC, and its safety and efficacy data were promising for further clinical development. In another smaller trial (NCT03070990), 17 Japanese patients with refractory urothelial cancer were randomly assigned to receive either 1 or 1.25 mg/kg of EV for a standard cycle of 28 days. The adverse reactions seen in this study were similar to those reported in the EV-101 phase I trial, with an mPFS of 8.1 months (88).

EV-201 (NCT03219333) study was a global, phase II study designed to evaluate the drug's treatment effect and was key to the FDA's decision to approve EV for treatment of patients with advanced urothelial cancer. This study was a single-arm trial with two separate patient cohorts. Cohort 1 included 125 patients treated with EV at a standard dose; the ORR was 44%, including 12% complete response (89, 90). Cohort 2 included 89 patients (91), who



were treated with EV at standard doses and had an ORR of 52% and a complete response rate of 20%. TRAEs were acute kidney injury, hyperglycemia, decreased appetite, hypotension, diarrhea, neutropenia, and skin reactions. Four patients died of treatment-related toxicity (acute kidney injury, metabolic acidosis, multiorgan failure, and pneumonitis) (91). The EV-201 study played a vital part in demonstrating the efficacy and safety of EV for patients with advanced urothelial carcinoma. The study showed promising clinical outcomes, including significant ORRs and complete responses in both cohorts.

The EV-301 (NCT03474107) trial was a randomized phase III clinical trial designed to evaluate OS in la/mUC patients after treatment with EV. This trial aimed to assess the efficacy of EV compared with standard chemotherapy. It enrolled 608 patients, 301 of whom were treated with EV at standard dose, whereas the other 307 received chemotherapeutic agents (standard doxorubicin, paclitaxel, or vinblastine) on day 1 of a 21-day cycle. The mOS was 12.88 months in the EV group and 8.97 months in the chemotherapy group. The confirmed ORR in the EV group was 40.6%, which was consistent with results of previous studies and substantially higher than the ORR of 17.9% for the chemotherapy group. However, the incidence of adverse reactions in the two groups was similar, indicating that the two treatment modalities had a comparable profile in terms of safety. The study reported a low rate of complete responses (92), although EV was highly effective in reducing tumor size and extending OS. The EV-301 trial provided compelling evidence that EV significantly extends OS in patients with la/mUC.

A combination of EV with pembrolizumab (an immune checkpoint inhibitor) demonstrated enhanced treatment outcomes compared with EV alone (90, 93). EV-103 (NCT03288545) evaluated the efficacy, tolerability, and safety of this combination. The dose escalation phase of the study established the safe dose of EV to be 1.25 mg/kg. Subsequently, 40 patients with la/mUC received EV + pembrolizumab. This trial reported the highest mOS (26.1 months) in a first-line urothelial carcinoma trial (90, 93, 94). EV-103 cohort K enrolled 149 patients who were randomly assigned to receive either EV in combination with pembrolizumab or EV monotherapy (95). After 3 months of

follow-up, the ORR of the EV + pembrolizumab treatment group was 64.5%, and no new safety issues occurred (96). The results of EV-103 confirmed the advantages of immunotherapy combined with ADC therapy compared with ADC monotherapy. EV-302 (NCT04223856) was an open, global, randomized controlled study. In this clinical trial, 886 treatment-naïve patients with la/mUC who met the eligibility criteria for cisplatin or carboplatin were enrolled. They were randomized to receive either EV + pembrolizumab or chemotherapy. The mOS and median progression-free survival of the EV + pembrolizumab group were twice those of the chemotherapy group (97). Thus, the results of EV-302 have changed the standard of care for bladder cancer (98), challenging conventional first-line treatments (including platinum-based chemotherapy) for patients with la/mUC and further validating the efficacy of combined immunotherapy and targeted therapy.

The FDA has progressively approved new indications for EV as clinical trials have been conducted. In 2019, based on the EV-201 cohort 1 study, EV was approved for the treatment of adult patients with la/mUC who had received a PD-1 or PD-L1 inhibitor and platinum-containing chemotherapy. After clinical trials of the EV-201 cohort 2 and EV-301 had been completed, the FDA extended the indications of EV to patients ineligible for cisplatin-containing chemotherapy who had previously received one or more prior lines of therapy. EV-302 showed that EV + pembrolizumab was superior to first-line chemotherapy, which led the FDA to approve EV combined with pembrolizumab for patients with la/mUC. FDA had previously granted accelerated approval to this combination for patients with la/mUC who were ineligible for cisplatin-containing chemotherapy. These consecutive approvals of EV demonstrate its established position in the treatment landscape of la/mUC.

Some new clinical studies are being conducted to broaden the indications of EV. Sacituzumab govitecan (SG), which targets TROP2, and EV are both FDA-approved ADCs. TROP2 and Nectin-4 are highly expressed in mUC. Clinical trial NCT04724018 was the first study to combine two ADCs for the treatment of mUC. This study recruited 24 mUC patients with disease progression after chemotherapy or immunotherapy. Different combinations of SG and EV doses were designed, and the recommended doses (SG: 8 mg/kg, EV: 1.25 mg/kg) for the

phase II study were determined. The ORR in this study was 70%, and 78% of patients had adverse events of grade ≥ 3 (99). Overall, although the SG + EV combination therapy has shown promising clinical significance, the incidence of adverse reactions was high. EV + SG combination therapy may therefore not be suitable for further development. Clinical trials (EV-303; NCT03924895) of EV combined with pembrolizumab as a perioperative neoadjuvant therapy in patients with muscle-invasive bladder cancer are also underway (100) and may provide new options for patients who are not candidates for cisplatin therapy during the perioperative period.

The therapeutic efficacy of EV in patients with urothelial carcinoma has shown variations based on the histological and molecular characteristics of the tumors, particularly expression levels of Nectin-4. In the real-world UNITE dataset, the ORR was 58% in patients with pure urethral histology and 42% in patients with any variant component (101). The expression of Nectin-4 in urothelial carcinoma tumors can vary significantly. Jean et al. detected the expression of Nectin-4 in 169 patients with urothelial carcinoma by immunohistochemistry. Overall, 87% of non-muscle-infiltrating urothelial cancer samples and 58% of muscle-infiltrating tumors were positive. This variation suggests that Nectin-4 expression levels may influence the efficacy of ADC treatment. Nectin-4 expression levels have been associated with different subtypes of urothelial carcinoma. A study using whole-transcriptome RNA sequencing found that most sarcomatoid carcinomas and all but two small-cell carcinomas expressed low levels of Nectin-4 mRNA (102). Nectin-4 expression varies considerably between subtypes, and its expression is correlated with ADC treatment efficacy (103). Considering the differences in tissue expression, testing for Nectin-4 expression levels prior to ADC treatment needs to be considered to optimize treatment efficacy. In the future, EV may be used for the treatment of different tumor subtypes with treatment tailored according to Nectin-4 expression.

Further studies have confirmed that the potential applications of EV are not limited to urothelial carcinoma. Rabet et al. selected one of six mAbs with the best affinity to develop an ADC for Nectin-4, which was used for targeted therapy in TNBC xenograft mouse models. A remarkable therapeutic effect was achieved, which was related to the dose and expression level of Nectin-4 (13). Clinical trials of EV for pancreatic cancer, prostate cancer, and squamous cell carcinoma of the penis are also underway (NCT05915351, NCT04754191, NCT06104618). In addition, EV has shown favorable results in the treatment of skin cancer (104). We anticipate that EV will deliver good efficacy and safety for a broader range of cancers in the future. Table 2 displays the details and results of EV clinical trials.

6.4 EV-related TRAEs

Severe TRAEs are a key factor leading to dose reduction or discontinuation of EV treatment. The predominant adverse events ($\geq 20\%$) of all grades include fatigue, peripheral neuropathy, decreased appetite, rash, alopecia, nausea, dysgeusia, diarrhea, dry eye, pruritus, and dry skin. Warnings for EV encompass

hyperglycemia, peripheral neuropathy, ocular disorders, skin reactions, infusion site extravasations, and embryo-fetal toxicity. The most common AEs leading to discontinuation or dosage reduction are peripheral neuropathy and rash. The types of peripheral neuropathy are diverse and predominantly characterized by sensory abnormalities, which can be attributed to the microtubule toxicity of MMAE. There are many types of rashes, with maculopapular rashes being the most common and severe. Several clinical trials have shown that reducing the dosage and using steroids can have a therapeutic effect on rash. Grade ≥ 3 adverse events include maculopapular rash, febrile neutropenia, anemia, and hyperglycemia. Hyperglycemia is common in patients with a history of hyperglycemia or BMI ≥ 30 kg/m²; therefore, it is important to prevent this complication in overweight patients. Dry eye symptoms were observed in 36% of patients, and 14% of patients experienced blurred vision possibly related to dry eye symptoms. The U.S. Package Insert) recommends the use of artificial tears as a preventive measure for dry eyes or the use of ophthalmic corticosteroids after ophthalmic examinations. During follow-up periods, most patients' TRAEs gradually resolved or improved. There have been a few cases of deaths associated with EV treatment. Each death event was complex and involved multiple confounding factors such as underlying conditions and previous treatments.

6.5 Drug resistance to EV

A majority of patients develop resistance to EV during treatment. Anti-cancer drug resistance encompasses intricate mechanisms that include reduced antigen expression, induction of drug transport proteins, and trafficking defects, as well as alterations in signaling and apoptotic pathways (106). Several studies have demonstrated that membrane Nectin-4 expression is decreased in metastatic tissues, and reduced or absent membrane Nectin-4 expression in mUC tissues may have a crucial role in EV resistance (107, 108). However, certain studies have confirmed the continued high expression of Nectin-4 in new tissue specimens after cancer recurrence (102), which does not support a decrease in antigen levels as key to the emergence of resistance to EV. Cabaud et al. investigated the causes of EV resistance using a mouse model of breast cancer (109); this is the only study of the mechanisms of EV resistance. They reported that ABCB1, which encodes the ATP-binding transporter multidrug resistance 1 (MDR-1)/P-glycoprotein (P-gp), was upregulated in EV-resistant tumors, and sensitivity to EV targeting Nectin-4 was restored in *in vitro* and *in vivo* models after P-gp was inhibited. In addition, the combined use of a third-generation P-gp inhibitor and ADC was well tolerated in mice and restored sensitivity to ADC (109). Considering the limitations of preclinical models, data from clinical patient populations are now urgently needed for further exploration and confirmation.

In summary, resistance to EV treatment is a complex phenomenon that may involve various mechanisms. The role of P-gp and its inhibition in overcoming EV resistance has shown promise in preclinical models (109). However, clinical data are

TABLE 2 Clinical trial of EV for the treatment of patients with UC.

Cinical trials	Number of patients	Characteristic of patients with UC	Purpose	Treatment	Result	Year (Ref.)
EV-101 phaseI (NCT02091999)	155	received chemotherapy or ICI disease progressed	Safety and tolerability	EV, 0.5 -1.25 mg/kg (Schedule A)	ORR: 43%, duration of response: 7.4 months, mOS: 12.3 months	2020/ (87)
EV-101 phaseII (NCT02091999)	112			EV, 1.25 mg/kg (Schedule A)		
Takahashi S et al. (NCT03070990)	17	refractory urothelial cancer patients		EV, 1 or 1.25 mg/kg (Schedule A)	in the whole population: ORR: 35.3%, DCR: 76.5%; in Arm A: ORR 44.4%, DCR 100%; in Arm B: ORR 25%, DCR 50%	2020/ (88)
EV-201 cohort 1 (NCT03219333)	125	received chemotherapy or ICI, and then disease progressed	efficacy and safety	EV, 1.25 mg/kg (Schedule A)	ORR: 44%, complete response: 12%	2019/ (89)
EV-201 cohort 2 (NCT03219333)	89	Contraindication to chemotherapy	efficacy and safety	EV, 1.25 mg/kg (Schedule A)	ORR: 52%, complete response: 20%	2021/ (91)
EV-301 (NCT03474107)	608	received chemotherapy and ICI, and then disease progressed	clinical benefit	EV group: EV, 1.25 mg/kg (Schedule A) Chemotherapy group: doxorubicin, paclitaxel, or vinblastine (Schedule B)	EV group: OS 12.88 months, mPFS 5.55 months, ORR 40.6%; Chemotherapy group: OS 8.97 months, mPFS 3.71 months, ORR 17.9%	2021/ (92)
EV-103 dose escalation/cohort A (NCT03288545)	45	Contraindication to chemotherapy	Safety and tolerability	Dose Escalation Phase (N = 5): EV, 1-1.25 mg/kg (Schedule C) + P, 200mg (Schedule B) Dose Expansion Cohort A (N = 40): EV: EV, 1.25 mg/kg (Schedule C) + P, 200mg (Schedule B)	DCR: 93.3%, ORR: 73.3%. At the first tumor assessment: mPFS 12.3 months, mOS 26.1 months	2023/ (93)
EV-103 cohort K (NCT03288545)	149	Contraindication to chemotherapy	Efficacy and safety	Group1(N=76): EV, 1.25 mg/kg (Schedule C) + P, 200mg (Schedule B) Group2(N=73): EV, 1.25 mg/kg (Schedule C)	Group1: ORR 64.5%, CR 10.5%, PR 53.9%, ORR 49%, mOS 22.3 months Group1: ORR 45.2%, CR 4.1%, PR 41.1%, ORR 33%, mOS 21.7 months	2023 (95, 105)
EV-302 (NCT04223856)	886	previously untreated and met the treatment requirements of cisplatin or carboplatin		Group1: EV, 1.25 mg/kg (Schedule C) + P, 200mg (Schedule B) Group2: gemcitabine with cisplatin or carboplatin	Group1: ORR 67.7%, mOS 31.5 mo, mPFS 12.5 Group2: ORR 44.4%, mOS 16.1 mo, mPFS 6.3	2023 (97)
McGregor BA et al. (NCT04724018)	24	patients with disease progression after chemotherapy or immunotherapy	Safety and tolerability	SG + EV, dose of each group is different (Schedule C)	ORR: 70%	2024 (99)

OS, overall survival; DCR, disease control rate; mOS, median OS; ICI, immune checkpoint inhibitor; ORR, objective response rate; mPFS, median progression-free survival; CR, complete response; PR, partial response; P, pembrolizumab.

Schedule A is days 1, 8 and 15 of a 28-day cycle. Schedule B is day 1 of a 21-day cycle. Schedule C is days 1 and 8 of a 21-day cycle.

essential to fully understand resistance mechanisms and develop effective strategies to combat resistance in patients receiving EV therapy.

6.6 Other ADCs targeting Nectin-4

Other ADCs targeting Nectin-4 include the novel 9MW2821, which consists of an anti-Nectin-4 antibody (MW282), a linker (IDconnect), and MMAE. IDconnect is a novel thioether linker that cross-links the Fab and hinge regions, resulting in a more robust attachment of MMAE (110). The design of IDconnect is the strength of 9MW2821, which shows reduced amounts of off-target drug in circulation and decreased liver toxicity. Compared

with EV, 9MW2821 has a homogeneous drug-antibody ratio (DAR) and has shown better antitumor effects and fewer adverse effects in a preclinical trial (110). Research and development objectives for 9MW2821 are focused on mitigating the adverse reactions caused by EV and improving the therapeutic effect in patients with la/mUC. Clinical trials of 9MW2821 are about to begin (NCT06079112 and NCT06196736). Two clinical studies have expanded from la/mUC patients to patients with other malignant solid tumors (NCT05216965 and NCT05773937). ADRX-0706 and BAT8007 are also ADCs targeting Nectin-4. ADRX-0706 has a DAR of 8 (the DAR of ADCs is usually 2-4), leading to a greatly increased drug concentration in the local area of the tumor. Therefore, the therapeutic effects obtained with high DARs should receive more attention. A clinical trial of ADRX-0706

is being carried out in patients with specific advanced solid tumors, including urothelial carcinoma, head and neck squamous cell carcinoma, breast cancer, cervical cancer, ovarian cancer, NSCLC, and pancreatic cancer (NCT06036121). The cytotoxic drug carried by BAT8007 is a topoisomerase I inhibitor (Exatecan) that interferes with DNA replication and transcription to induce apoptosis. It differs from other ADCs targeting Nectin-4 by replacing MMAE, thereby changing the mechanism of tumor cell killing. At present, a clinical trial of BAT8007 is recruiting patients with advanced solid tumors (NCT05879627). The potential applications of these three new drugs targeting Nectin-4 have already been extended to various malignant solid tumors, underscoring the widespread attention paid to Nectin-4 and recognition of its potential as a therapeutic target. It is expected that research on Nectin-4 in the field of cancer therapy will continue to grow and progress.

6.7 BT8009 and BT7480

Not all ADCs translate perfectly from preclinical models to humans, in part because of poor drug penetration and long drug half-lives (days to weeks). Although antibodies provide high affinity and highly specific carriers for toxin delivery, their large molecular size (150 kDa) hinders their ability to overcome the combined barriers of the endothelial barrier, interstitial pressure, and complex stromal structures in tumors (111). This results in low extravasation rates and slow diffusion outside the cell. Only 0.01–0.10% of injected antibodies may reach solid tumor antigens (111). Bicyclic toxin conjugates (BTCs) targeting Nectin-4 offer an alternative to traditional ADCs by utilizing small hydrophilic peptides instead of mAbs. This allows BTCs to quickly diffuse from the systemic circulation and potentially overcome some of the limitations associated with ADCs. BT8009 is composed of a bicyclic peptide that binds to Nectin-4, a cleavable adaptor system, and a cell-penetrating toxin, MMAE. It has demonstrated superior or comparable antitumor activity compared with EV in several models. BT8009 has the advantage of fast clearance and can penetrate tumors and target tumor cells (112). In summary, BT8009 offers a promising alternative to traditional ADCs for targeting Nectin-4 and other cancer-associated molecules. Clinical trials aimed at further assessing its effectiveness and safety in real-world patient populations are in progress.

BT7480 is composed of two CD137 bicycles and a Nectin-4 bicycle. CD137 is an inducible costimulatory receptor that can participate in the activation of a variety of immune cells and has been reported to be a promising target for cancer immunotherapy (113, 114). Transcriptional analysis has shown that Nectin-4 and CD137 are co-expressed in a variety of cancers. BT7480 targets Nectin-4-expressing tumor cells and then binds to and agonizes CD137 on nearby immune cells to remodel the tumor immune microenvironment, thereby eliminating tumor cells and inhibiting tumor recurrence (115, 116). Compared with CD137 activation alone, immune killing after targeted activation shows a better therapeutic effect and less liver toxicity.

7 Conclusion

This review provides an overview of the role of Nectin-4 in cancers, focusing on clinical trials related to Nectin-4-targeted tumor treatments and the development of novel drugs targeting Nectin-4. Nectin-4 is upregulated in multiple tumor types and promotes tumorigenesis, development, and angiogenesis through pathways such as PI3K/AKT. Nectin-4 has shown clinical significance in serological and molecular imaging diagnosis of some tumor types, such as ovarian cancer, urothelial carcinoma, and breast cancer, but more studies with larger sample sizes are needed to promote its further applications. In 2019, EV was approved by the FDA as the first ADC drug targeting Nectin-4 for the treatment of la/mUC. Subsequently, various clinical trials related to EV have reported encouraging survival data, which has led to the continuing expansion of EV indications in la/mUC. These results have promoted the development of new drugs targeting Nectin-4 and have clearly demonstrated its value as a therapeutic target for tumors. However, TRAEs and drug resistance are aspects that require significant attention. Common TRAEs include rash, peripheral neuropathy, and fatigue, and close monitoring during initial treatment cycles is advisable. Moreover, there has been no definitive study to elucidate the exact underlying causes of drug resistance, which is a key factor impeding further treatment with EV. New therapeutic drugs targeting Nectin-4 have been improved in terms of linkers and DAR compared with EV; however, the relevant clinical trials are still underway, so the treatment efficacy of these alternatives remains unknown.

In conclusion, Nectin-4 is a highly valuable molecular target in cancer, deserving further research and development.

Author contributions

KL: Data curation, Writing – original draft. YZ: Writing – original draft, Funding acquisition. MZ: Writing – review & editing, Visualization, Data curation. XJ: Writing – review & editing. XL: Writing – review & editing, Conceptualization.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. This work was supported by Natural Science Foundation of Shandong Province (ZR2022QH315).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated

organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

- Sawyers CL. The cancer biomarker problem. *Nature*. (2008) 452:548–52. doi: 10.1038/nature06913
- Chothia C, Jones EY. The molecular structure of cell adhesion molecules. *Annu Rev Biochem*. (1997) 66:823–62. doi: 10.1146/annurev.biochem.66.1.823
- Takahashi K, Nakanishi H, Miyahara M, Mandai K, Satoh K, Satoh A, et al. Nectin/PRR: an immunoglobulin-like cell adhesion molecule recruited to cadherin-based adherens junctions through interaction with Afadin, a PDZ domain-containing protein. *J Cell Biol*. (1999) 145:539–49. doi: 10.1083/jcb.145.3.539
- Reymond N, Lecocq E, Adelaide J, Dubreuil P, Lopez M. Nectin4/PRR4, a new afadin-associated member of the nectin family that trans-interacts with nectin1/PRR1 through V domain interaction. *J Biol Chem*. (2001) 276:43205–15. doi: 10.1074/jbc.M103810200
- Fabre S, Reymond N, Cocchi F, Menotti L, Dubreuil P, Campadelli-Fiume G, et al. Prominent role of the Ig-like V domain in trans-interactions of nectins. Nectin3 and nectin 4 bind to the predicted C-C'-D beta-strands of the nectin1 V domain. *J Biol Chem*. (2002) 277:27006–13. doi: 10.1074/jbc.M203228200
- Yasumi M, Shimizu K, Honda T, Takeuchi M, Takai Y. Role of each immunoglobulin-like loop of nectin for its cell-cell adhesion activity. *Biochem Biophys Res Commun*. (2003) 302:61–6. doi: 10.1016/S0006-291X(03)00106-2
- Pavlova NN, Pallasch C, Elia AE, Braun CJ, Westbrook TF, Hemann M, et al. A role for PVRL4-driven cell-cell interactions in tumorigenesis. *eLife*. (2013) 2:e00358. doi: 10.7554/eLife.00358
- Takano A, Ishikawa N, Nishino R, Masuda K, Yasui W, Inai K, et al. Identification of nectin-4 oncoprotein as a diagnostic and therapeutic target for lung cancer. *Cancer Res*. (2009) 69:6694–703. doi: 10.1158/0008-5472.CAN-09-0016
- Derycke MS, Pambuccian SE, Gilks CB, Kaloger SE, Ghidouche A, Lopez M, et al. Nectin 4 overexpression in ovarian cancer tissues and serum: potential role as a serum biomarker. *Am J Clin Pathol*. (2010) 134:835–45. doi: 10.1309/AJCPGXK0FR4MH1HB
- Fabre-Lafay S, Monville F, Garrido-Urbani S, Berruyer-Pouyet C, Ginestier C, Reymond N, et al. Nectin-4 is a new histological and serological tumor associated marker for breast cancer. *BMC cancer*. (2007) 7:73. doi: 10.1186/1471-2407-7-73
- Nishiwada S, Sho M, Yasuda S, Shimada K, Yamato I, Akahori T, et al. Nectin-4 expression contributes to tumor proliferation, angiogenesis and patient prognosis in human pancreatic cancer. *J Exp Clin Cancer Res*. (2015) 34:30. doi: 10.1186/s13046-015-0144-7
- Lattanzio R, Ghasemi R, Brancati F, Sorda RL, Tinari N, Perracchio L, et al. Membranous Nectin-4 expression is a risk factor for distant relapse of T1-T2, N0 luminal-A early breast cancer. *Oncogenesis*. (2014) 3:e118. doi: 10.1038/oncsis.2014.32
- MR M, Cabaud O, Josselin E, Finetti P, Castellano R, Farina A, et al. Nectin-4: a new prognostic biomarker for efficient therapeutic targeting of primary and metastatic triple-negative breast cancer. *Ann Oncol Off J Eur Soc Med Oncol*. (2017) 28:769–76. doi: 10.1093/annonc/mdw678
- Deng H, Shi H, Chen L, Zhou Y, Jiang J. Over-expression of Nectin-4 promotes progression of esophageal cancer and correlates with poor prognosis of the patients. *Cancer Cell Int*. (2019) 19:106. doi: 10.1186/s12935-019-0824-z
- Ma J, Sheng Z, Lv Y, Liu W, Yao Q, Pan T, et al. Expression and clinical significance of Nectin-4 in hepatocellular carcinoma. *OncoTar Ther*. (2016) 9:183–90. doi: 10.2147/OTT
- Zhang Y, Zhang J, Shen Q, Yin W, Huang H, Liu Y, et al. High expression of Nectin-4 is associated with unfavorable prognosis in gastric cancer. *Oncol Lett*. (2018) 15:8789–95. doi: 10.3892/ol
- Wong JL, Rosenberg JE. Targeting nectin-4 by antibody-drug conjugates for the treatment of urothelial carcinoma. *Expert Opin Biol Ther*. (2021) 21:863–73. doi: 10.1080/14712598.2021.1929168
- Heath EI, Rosenberg JE. The biology and rationale of targeting nectin-4 in urothelial carcinoma. *Nat Rev Urol*. (2021) 18:93–103. doi: 10.1038/s41585-020-00394-5
- Bouletfour W, Sargos P, Magne N. Nectin-4: a tumor cell target and status of inhibitor development. *Curr Oncol Rep*. (2023) 25:181–8. doi: 10.1007/s11912-023-01360-1
- Mendelsohn CL, Wimmer E, Racaniello VR. Cellular receptor for poliovirus: molecular cloning, nucleotide sequence, and expression of a new member of the immunoglobulin superfamily. *Cell*. (1989) 56:855–65. doi: 10.1016/0092-8674(89)90690-9
- Rikitake Y, Mandai K, Takai Y. The role of nectins in different types of cell-cell adhesion. *J Cell Sci*. (2012) 125:3713–22. doi: 10.1242/jcs.099572
- Lopez M, Aoubala M, Jordier F, Isnardon D, Gomez S, Dubreuil P. The human poliovirus receptor related 2 protein is a new hematopoietic/endothelial homophilic adhesion molecule. *Blood*. (1998) 92:4602–11. doi: 10.1182/blood.V92.12.4602.424k21_4602_4611
- Buchanan PC, Boylan KLM, Walcheck B, Heinze R, Geller MA, Argenta PA, et al. Ectodomain shedding of the cell adhesion molecule Nectin-4 in ovarian cancer is mediated by ADAM10 and ADAM17. *J Biol Chem*. (2017) 292:6339–51. doi: 10.1074/jbc.M116.746859
- Siddharth S, Nayak A, Das S, Nayak D, Panda J, Wyatt MD, et al. The soluble nectin-4 ecto-domain promotes breast cancer induced angiogenesis via endothelial Integrin-β4. *Int J Biochem Cell Biol*. (2018) 102:151–60. doi: 10.1016/j.jbiocel.2018.07.011
- Chatterjee S, Sinha S, Molla S, Hembram KC, Kundu CN. PARP inhibitor Veliparib (ABT-888) enhances the anti-angiogenic potentiality of Curcumin through deregulation of NECTIN-4 in oral cancer: Role of nitric oxide (NO). *Cell signal*. (2021) 80:109902. doi: 10.1016/j.cellsig.2020.109902
- Laksono BM, de Vries RD, McQuaid S, Duprex WP, de Swart RL. Measles virus host invasion and pathogenesis. *Viruses*. (2016) 8(8):210. doi: 10.3390/v8080210
- Noyce RS, Richardson CD. Nectin 4 is the epithelial cell receptor for measles virus. *Trends Microbiol*. (2012) 20:429–39. doi: 10.1016/j.tim.2012.05.006
- Mühlebach MD, Mateo M, Sinn PL, Prüfer S, Uhlig KM, Leonard VH, et al. Adherens junction protein nectin-4 is the epithelial receptor for measles virus. *Nature*. (2011) 480:530–3. doi: 10.1038/nature10639
- Rotunno R, Diociaiuti A, Dentici ML, Rinelli M, Callea M, Retrosi C, et al. Ectodermal dysplasia-syndactyly syndrome with toe-only minimal syndactyly due to a novel mutation in NECTIN4: A case report and literature review. *Genes*. (2021) 12(5):748. doi: 10.3390/genes12050748
- Dardour L, Cosyng K, Devriendt K. A novel missense variant in the PVRL4 gene underlying ectodermal dysplasia-syndactyly syndrome in a turkish child. *Mol syndromol*. (2017) 9:22–4. doi: 10.1159/000479359
- Raza SI, Nasser Dar R, Shah AA, Ahmad W. A novel homozygous nonsense mutation in the PVRL4 gene and expansion of clinical spectrum of EDSS1. *Ann Hum Genet*. (2015) 79:92–8. doi: 10.1111/ahg.12094
- Sethy C, Goutam K, Nayak D, Pradhan R, Molla S, Chatterjee S, et al. Clinical significance of a pvr1 4 encoded gene Nectin-4 in metastasis and angiogenesis for tumor relapse. *J Cancer Res Clin Oncol*. (2020) 146:245–59. doi: 10.1007/s00432-019-03055-2
- Challita-Eid PM, Satpayev D, Yang P, An Z, Morrison K, Shostak Y, et al. Enfortumab vedotin antibody-drug conjugate targeting nectin-4 is a highly potent therapeutic agent in multiple preclinical cancer models. *Cancer Res*. (2016) 76:3003–13. doi: 10.1158/0008-5472.CAN-15-1313
- Chatterjee S, Kundu CN. Nanoformulated quinacrine regulates NECTIN-4 domain specific functions in cervical cancer stem cells. *Eur J Pharmacol*. (2020) 883:173308. doi: 10.1016/j.ejphar.2020.173308
- Boylan KL, Buchanan PC, Manion RD, Shukla DM, Braumberger K, Bruggemeyer C, et al. The expression of Nectin-4 on the surface of ovarian cancer cells alters their ability to adhere, migrate, aggregate, and proliferate. *Oncotarget*. (2017) 8:9717–38. doi: 10.18632/oncotarget.v8i6
- Zhang J, Liu K, Peng P, Li S, Ye Z, Su Y, et al. Upregulation of nectin-4 is associated with ITGB1 and vasculogenic mimicry and may serve as a predictor of poor prognosis in colorectal cancer. *Oncol Lett*. (2019) 18:1163–70. doi: 10.3892/ol
- Ballester M, Gonin J, Rodenas A, Bernaudin JF, Rouzier R, Coutant C, et al. Eutopic endometrium and peritoneal, ovarian and colorectal endometriotic tissues express a different profile of nectin-1, -3, -4 and nectin-like molecule 2. *Hum Reprod (Oxford England)*. (2012) 27:3179–86. doi: 10.1093/humrep/des304
- Bedir R, Sehitoglu I, Balik G, Kagitci M, Gucer H, Yurdakul C, et al. The role of the adhesion molecule Nectin-4 in the pathogenesis of endometriosis. *Clin Exp Obstet Gynecol*. (2016) 43:463–6. doi: 10.12891/ceog2150.2016
- Bussink J, van der Kogel AJ, Kaanders JH. Activation of the PI3-K/AKT pathway and implications for radioresistance mechanisms in head and neck cancer. *Lancet Oncol*. (2008) 9:288–96. doi: 10.1016/S1470-2045(08)70073-1
- Miricescu D, Totan A, Stanescu S II, Badoiu SC, Stefani C, Greabu M. PI3K/AKT/mTOR signaling pathway in breast cancer: from molecular landscape to clinical aspects. *Int J Mol Sci*. (2020) 22(1):173. doi: 10.3390/ijms22010173
- Fresno Vara JA, Casado E, de Castro J, Cejas P, Belda-Iniesta C, González-Barón M. PI3K/Akt signalling pathway and cancer. *Cancer Treat Rev*. (2004) 30:193–204. doi: 10.1016/j.ctrv.2003.07.007

42. Kedashiro S, Sugiura A, Mizutani K, Takai Y. Nectin-4 cis-interacts with ErbB2 and its trastuzumab-resistant splice variants, enhancing their activation and DNA synthesis. *Sci Rep.* (2019) 9:18997. doi: 10.1038/s41598-019-55460-9
43. Siddharth S, Goutam K, Das S, Nayak A, Nayak D, Sethy C, et al. Nectin-4 is a breast cancer stem cell marker that induces WNT/ β -catenin signaling via PI3K/Akt axis. *Int J Biochem Cell Biol.* (2017) 89:85–94. doi: 10.1016/j.biocel.2017.06.007
44. Rodgers SJ, Mitchell CA, Ooms LM. The mechanisms of class 1A PI3K and Wnt/ β -catenin coupled signaling in breast cancer. *Biochem Soc Trans.* (2023) 51:1459–72. doi: 10.1042/BST20220866
45. Liu Y, Li G, Zhang Y, Li L, Zhang Y, Huang X, et al. Nectin-4 promotes osteosarcoma progression and metastasis through activating PI3K/AKT/NF- κ B signaling by down-regulation of miR-520c-3p. *Cancer Cell Int.* (2022) 22:252. doi: 10.1186/s12935-022-02669-w
46. Zhang Y, Liu S, Wang L, Wu Y, Hao J, Wang Z, et al. A novel PI3K/AKT signaling axis mediates Nectin-4-induced gallbladder cancer cell proliferation, metastasis and tumor growth. *Cancer Lett.* (2016) 375:179–89. doi: 10.1016/j.canlet.2016.02.049
47. Zhang Y, Chen P, Yin W, Ji Y, Shen Q, Ni Q. Nectin-4 promotes gastric cancer progression via the PI3K/AKT signaling pathway. *Hum pathol.* (2018) 72:107–16. doi: 10.1016/j.humpath.2017.10.034
48. Das D, Satapathy SR, Siddharth S, Nayak A, Kundu CN. NECTIN-4 increased the 5-FU resistance in colon cancer cells by inducing the PI3K-AKT cascade. *Cancer Chemother Pharmacol.* (2015) 76:471–9. doi: 10.1007/s00280-015-2794-8
49. Kanzaki N, Ogita H, Komura H, Ozaki M, Sakamoto Y, Majima T, et al. Involvement of the nectin-afadin complex in PDGF-induced cell survival. *J Cell Sci.* (2008) 121:2008–17. doi: 10.1242/jcs.024620
50. Maruoka M, Kedashiro S, Ueda Y, Mizutani K, Takai Y. Nectin-4 co-stimulates the prolactin receptor by interacting with SOCS1 and inhibiting its activity on the JAK2-STAT5a signaling pathway. *J Biol Chem.* (2017) 292:6895–909. doi: 10.1074/jbc.M116.769091
51. Tan SH, Nevalainen MT. Signal transducer and activator of transcription 5A/B in prostate and breast cancers. *Endocrine-related cancer.* (2008) 15:367–90. doi: 10.1677/ERC-08-0013
52. Kedashiro S, Kameyama T, Mizutani K, Takai Y. Nectin-4 and p95-ErbB2 cooperatively regulate Hippo signaling-dependent SOX2 gene expression, enhancing anchorage-independent T47D cell proliferation. *Sci Rep.* (2021) 11:7344. doi: 10.1038/s41598-021-86437-2
53. Sethy C, Goutam K, Das B, Dash SR, Kundu CN. Nectin-4 promotes lymphangiogenesis and lymphatic metastasis in breast cancer by regulating CXCR4-LYVE-1 axis. *Vasc Pharmacol.* (2021) 140:106865. doi: 10.1016/j.vph.2021.106865
54. Lopez M, Cocchi F, Avitabile E, Leclerc A, Adelaide J, Campadelli-Fiume G, et al. Novel, soluble isoform of the herpes simplex virus (HSV) receptor nectin1 (or PRR1-HlgR-HvcC) modulates positively and negatively susceptibility to HSV infection. *J Virol.* (2001) 75:5684–91. doi: 10.1128/JVI.75.12.5684-5691.2001
55. Koike S, Horie H, Ise I, Okitsu A, Yoshida M, Iizuka N, et al. The poliovirus receptor protein is produced both as membrane-bound and secreted forms. *EMBO J.* (1990) 9:3217–24. doi: 10.1002/emboj.1990.9.issue-10
56. Morrison ME, Racaniello VR. Molecular cloning and expression of a murine homolog of the human poliovirus receptor gene. *J Virol.* (1992) 66:2807–13. doi: 10.1128/jvi.66.5.2807-2813.1992
57. Eberlé F, Dubreuil P, Mattei MG, Devillard E, Lopez M. The human PRR2 gene, related to the human poliovirus receptor gene (PVR), is the true homolog of the murine MPH gene. *Gene.* (1995) 159:267–72. doi: 10.1016/0378-1119(95)00180-E
58. Fabre-Lafay S, Garrido-Urbani S, Reymond N, Gonçalves A, Dubreuil P, Lopez M. Nectin-4, a new serological breast cancer marker, is a substrate for tumor necrosis factor- α -converting enzyme (TACE)/ADAM-17. *J Biol Chem.* (2005) 280:19543–50. doi: 10.1074/jbc.M410943200
59. Nabih ES, Abdel Motaleb FI, Salama FA. The diagnostic efficacy of nectin 4 expression in ovarian cancer patients. *Biomarkers Biochem Indic exposure response susceptibility to chemicals.* (2014) 19:498–504. doi: 10.3109/1354750X.2014.940503
60. Rogmans C, Feuerborn J, Trecck L, Tribian N, Flörkemeier I, Arnold N, et al. Nectin-4 as blood-based biomarker enables detection of early ovarian cancer stages. *Cancers.* (2022) 14(23):5867. doi: 10.3390/cancers14235867
61. Erturk K, Karaman S, Dagoglu N, Serilmez M, Duranyildiz D, Tas F. Serum nectin-2 and nectin-4 are diagnostic in lung cancer: which is superior? *Wiener klinische Wochenschrift.* (2019) 131:419–26. doi: 10.1007/s00508-019-01537-4
62. Lee PH, Choi S, An M, Jang AS. Recent patents in allergy and immunology: A quantitative real-time polymerase chain reaction method for diagnosing asthma and asthma exacerbation. *Clin Trans Allergy.* (2022) 12:e12136. doi: 10.1002/ctt2.12136
63. Fang CP, Liu TH, Chung RH, Tsou HH, Kuo HW, Wang SC, et al. Genetic variants in NECTIN4 encoding an adhesion molecule are associated with continued opioid use. *PLoS One.* (2020) 15:e0234549. doi: 10.1371/journal.pone.0234549
64. Liu YL, Kuo HW, Fang CP, Tsung JH, Chen ACH. Adhesion molecules as potential novel biomarkers for opioid dependence. *Curr Pharm design.* (2020) 26:253–9. doi: 10.2174/138161282666200116144147
65. Campbell DO, Noda A, Verlinsky A, Snyder J, Fujita Y, Murakami Y, et al. Preclinical evaluation of an anti-nectin-4 immunoPET reagent in tumor-bearing mice and biodistribution studies in cynomolgus monkeys. *Mol Imaging Biol.* (2016) 18:768–75. doi: 10.1007/s11307-016-0953-x
66. Shao F, Pan Z, Long Y, Zhu Z, Wang K, Ji H, et al. Nectin-4-targeted immunoSPECT/CT imaging and photothermal therapy of triple-negative breast cancer. *J Nanobiotechnol.* (2022) 20:243. doi: 10.1186/s12951-022-01444-3
67. Duan X, Xia L, Zhang Z, Ren Y, Pomper MG, Rowe SP, et al. First-in-human study of the radioligand 68Ga-N188 targeting nectin-4 for PET/CT imaging of advanced urothelial carcinoma. *Clin Cancer Res an Off J Am Assoc Cancer Res.* (2023) 29(17):3395–407. doi: 10.1158/1078-0432.ccr637750.v1
68. Jaiswal PK, Goel A, Mittal RD. Survivin: A molecular biomarker in cancer. *Indian J Med Res.* (2015) 141:389–97. doi: 10.4103/0971-5916.159250
69. Athanassiadou AM, Patsouris E, Tsipis A, Gonidi M, Athanassiadou P. The significance of Survivin and Nectin-4 expression in the prognosis of breast carcinoma. *Folia Histochem Cytobiol.* (2011) 49:26–33. doi: 10.5603/FHC.2011.0005
70. Zeindler J, Soysal SD, Piscuoglio S, Ng CKY, Mechera R, Isaak A, et al. Nectin-4 expression is an independent prognostic biomarker and associated with better survival in triple-negative breast cancer. *Front Med.* (2019) 6:200. doi: 10.3389/fmed.2019.00200
71. Lin X, Hu H, Pan Y, Gao S. The prognostic role of expression of nectin-4 in esophageal cancer. *Med Sci monitor Int Med J Exp Clin Res.* (2019) 25:10089–94. doi: 10.12659/MSM.918288
72. Ueki H, Hinata N, Kitagawa K, Hara T, Terakawa T, Furukawa J, et al. Expressions of PD-L1 and Nectin-4 in urothelial cancer patients treated with pembrolizumab. *Clin Trans Oncol Off Publ Fed Spanish Oncol Soc Natl Cancer Instit Mexico.* (2022) 24:568–77. doi: 10.1007/s12094-021-02717-3
73. Rajc J, Gugić D, Fröhlich I, Marjanović K, Dumenčić B. Prognostic role of Nectin-4 expression in luminal B (HER2 negative) breast cancer. *Pathol Res practice.* (2017) 213:1102–8. doi: 10.1016/j.prp.2017.07.019
74. Zschäbitz S, Mikuteit M, Stöhr C, Herrmann E, Polifka I, Agaimy A, et al. Expression of nectin-4 in papillary renal cell carcinoma. *Discov Oncol.* (2022) 13:90. doi: 10.1007/s12672-022-00558-2
75. Mondal M, Guo J, He P, Zhou D. Recent advances of oncolytic virus in cancer therapy. *Hum Vaccines Immunother.* (2020) 16:2389–402. doi: 10.1080/21645515.2020.1723363
76. Hammood M, Craig AW, Leyton JV. Impact of endocytosis mechanisms for the receptors targeted by the currently approved antibody-drug conjugates (ADCs)-A necessity for future ADC research and development. *Pharm (Basel Switzerland).* (2021) 14(7):674. doi: 10.3390/ph14070674
77. Fujiyuki T, Amagai Y, Shoji K, Kuraishi T, Sugai A, Awano M, et al. Recombinant SLAMF8-measles virus is a promising candidate for nectin-4-positive triple negative breast cancer therapy. *Mol Ther Oncolyt.* (2020) 19:127–35. doi: 10.1016/j.omto.2020.09.007
78. Fujiyuki T, Yoneda M, Amagai Y, Obayashi K, Ikeda F, Shoji K, et al. A measles virus selectively blind to signaling lymphocytic activation molecule shows anti-tumor activity against lung cancer cells. *Oncotarget.* (2015) 6:24895–903. doi: 10.18632/oncotarget.v6i28
79. Awano M, Fujiyuki T, Shoji K, Amagai Y, Murakami Y, Furukawa Y, et al. Measles virus selectively blind to signaling lymphocyte activity molecule has oncolytic efficacy against nectin-4-expressing pancreatic cancer cells. *Cancer Sci.* (2016) 107:1647–52. doi: 10.1111/cas.13064
80. Amagai Y, Fujiyuki T, Yoneda M, Shoji K, Furukawa Y, Sato H, et al. Oncolytic activity of a recombinant measles virus, blind to signaling lymphocyte activation molecule, against colorectal cancer cells. *Sci Rep.* (2016) 6:24572. doi: 10.1038/srep24572
81. Aref S, Bailey K, Fielding A. Measles to the rescue: A review of oncolytic measles virus. *Viruses.* (2016) 8(10):294. doi: 10.3390/v8100294
82. Galanis E, Atherton PJ, Maurer MJ, Knutson KL, Dowdy SC, Cliby WA, et al. Oncolytic measles virus expressing the sodium iodide symporter to treat drug-resistant ovarian cancer. *Cancer Res.* (2015) 75:22–30. doi: 10.1158/0008-5472.CAN-14-2533
83. Kuo YT, Liu CH, Wong SH, Pan YC, Lin LT. Small molecules baicalein and cinnamaldehyde are potentiators of measles virus-induced breast cancer oncolysis. *Phytomed Int J phytother phytopharmacol.* (2021) 89:153611. doi: 10.1016/j.phymed.2021.153611
84. Liu CH, Wong SH, Tai CJ, Tai CJ, Pan YC, Hsu HY, et al. Ursolic acid and its nanoparticles are potentiators of oncolytic measles virotherapy against breast cancer cells. *Cancers.* (2021) 13(1):136. doi: 10.3390/cancers13010136
85. Challita-Eid PM, Satpayev D, Yang P, An Z, Morrison K, Shostak Y, et al. Enfortumab vedotin antibody-drug conjugate targeting nectin-4 is a highly potent therapeutic agent in multiple preclinical cancer models. *Cancer Res.* (2016) 76:3003–13. doi: 10.1158/0008-5472.CAN-15-1313
86. Birrer MJ, Moore KN, Betella I, Bates RC. Antibody-drug conjugate-based therapeutics: state of the science. *J Natl Cancer Instit.* (2019) 111:538–49. doi: 10.1093/jnci/djz035
87. Rosenberg J, Sridhar SS, Zhang J, Smith D, Ruether D, Flaig TW, et al. EV-101: A phase I study of single-agent enfortumab vedotin in patients with nectin-4-positive solid tumors, including metastatic urothelial carcinoma. *J Clin Oncol Off J Am Soc Clin Oncol.* (2020) 38:1041–9. doi: 10.1200/JCO.19.02044
88. Takahashi S, Uemura M, Kimura T, Kawasaki Y, Takamoto A, Yamaguchi A, et al. A phase I study of enfortumab vedotin in Japanese patients with locally advanced

or metastatic urothelial carcinoma. *Invest New Drugs*. (2020) 38:1056–66. doi: 10.1007/s10637-019-00844-x

89. Rosenberg JE, O'Donnell PH, Balar AV, McGregor BA, Heath EI, Yu EY, et al. Pivotal trial of enfortumab vedotin in urothelial carcinoma after platinum and anti-programmed death 1/programmed death ligand 1 therapy. *J Clin Oncol Off J Am Soc Clin Oncol*. (2019) 37:2592–600. doi: 10.1200/JCO.19.01140
90. Hoffman-Censits J, Maldonado L. Targeted treatment of locally advanced and metastatic urothelial cancer: enfortumab vedotin in context. *OncoTar Ther*. (2022) 15:1519–29. doi: 10.2147/OTT.S370900
91. Yu EY, Petrylak DP, O'Donnell PH, Lee JL, van der Heijden MS, Loriot Y, et al. Enfortumab vedotin after PD-1 or PD-L1 inhibitors in cisplatin-ineligible patients with advanced urothelial carcinoma (EV-201): a multicentre, single-arm, phase 2 trial. *Lancet Oncol*. (2021) 22:872–82. doi: 10.1016/S1470-2045(21)00094-2
92. Powles T, Rosenberg JE, Sonpavde GP, Loriot Y, Durán I, Lee JL, et al. Enfortumab vedotin in previously treated advanced urothelial carcinoma. *N Engl J Med*. (2021) 384:1125–35. doi: 10.1056/NEJMoa2035807
93. Hoimes CJ, Flaig TW, Milowsky MI, Friedlander TW, Bilen MA, Gupta S, et al. Enfortumab vedotin plus pembrolizumab in previously untreated advanced urothelial cancer. *J Clin Oncol Off J Am Soc Clin Oncol*. (2023) 41:22–31. doi: 10.1200/JCO.22.01643
94. Gupta S, Rosenberg JE, McKay RR, Flaig TW, Petrylak DP, Hoimes CJ, et al. Study EV-103 dose escalation/cohort A: Long-term outcome of enfortumab vedotin + pembrolizumab in first-line (1L) cisplatin-ineligible locally advanced or metastatic urothelial carcinoma (la/mUC) with nearly 4 years of follow-up. *J Clin Oncol Off J Am Soc Clin Oncol*. (2023) 41(16_suppl):4505. doi: 10.1200/JCO.2023.41.16_suppl.4505
95. O'Donnell PH, Milowsky MI, Petrylak DP, Hoimes CJ, Flaig TW, Mar N, et al. Enfortumab vedotin with or without pembrolizumab in cisplatin-ineligible patients with previously untreated locally advanced or metastatic urothelial cancer. *J Clin Oncol Off J Am Soc Clin Oncol*. (2023) 41:4107–17. doi: 10.1200/JCO.22.02887
96. Friedlander TW, Milowsky MI, O'Donnell PH, Petrylak DP, Hoimes CJ, Flaig TW, et al. Enfortumab vedotin (EV) with or without pembrolizumab (P) in patients (pts) who are cisplatin-ineligible with previously untreated locally advanced or metastatic urothelial cancer (la/mUC): Additional 3-month follow-up on cohort K data. *J Clin Oncol Off J Am Soc Clin Oncol*. (2023) 41(16_suppl):4568. doi: 10.1200/JCO.2023.41.16_suppl.4568
97. Powles TB, Perez Valderrama B, Gupta S, Bedke J, Kikuchi E, Hoffman-Censits J, et al. LBA6 EV-302/KEYNOTE-A39: Open-label, randomized phase III study of enfortumab vedotin in combination with pembrolizumab (EV+P) vs chemotherapy (Chemo) in previously untreated locally advanced metastatic urothelial carcinoma (la/mUC). *Ann Oncol*. (2023) 34:S1340. doi: 10.1016/j.annonc.2023.10.106
98. Fenor de la Maza MD, Villacampa G, Miñana B, Barbas Bernardos G, Andrés G, González-Padilla DA, et al. First-line management of metastatic urothelial cancer: current and future perspectives after the EV-302 and checkMate-901 studies. *Clin Genitourinary Cancer*. (2024) 22(2):330–5. doi: 10.1016/j.clgc.2023.12.001
99. McGregor BA, Sonpavde GP, Kwak L, Regan MM, Gao X, Hvidsten H, et al. The Double Antibody Drug Conjugate (DAD) phase I trial: sacituzumab govitecan plus enfortumab vedotin for metastatic urothelial carcinoma. *Ann Oncol Off J Eur Soc Med Oncol*. (2024) 35:91–7. doi: 10.1016/j.annonc.2023.09.3114
100. Galsky MD, Hoimes CJ, Necchi A, Shore N, Witjes JA, Steinberg G, et al. Perioperative pembrolizumab therapy in muscle-invasive bladder cancer: Phase III KEYNOTE-866 and KEYNOTE-905/EV-303. *Future Oncol (London England)*. (2021) 17:3137–50. doi: 10.2217/fonc-2021-0273
101. Koshkin VS, Henderson N, James M, Natesan D, Freeman D, Nizam A, et al. Efficacy of enfortumab vedotin in advanced urothelial cancer: Analysis from the Urothelial Cancer Network to Investigate Therapeutic Experiences (UNITE) study. *Cancer*. (2022) 128:1194–205. doi: 10.1002/cncr.34057
102. Hoffman-Censits JH, Lombardo KA, Parimi V, Kamanda S, Choi W, Hahn NM, et al. Expression of nectin-4 in bladder urothelial carcinoma, in morphologic variants, and nonurothelial histotypes. *Appl Immunohistochem Mol Morphol AIMM*. (2021) 29:619–25. doi: 10.1097/PAL.0000000000000938
103. Teo MY, Rosenberg JE. NECTIN4 heterogeneity and molecular diversity in bladder cancers: deconstructing the activity of an antibody-drug conjugate. *Clin Cancer Res an Off J Am Assoc Cancer Res*. (2021) 27:4950–2. doi: 10.1158/1078-0432.CCR-21-1807
104. Hashimoto H, Tanaka Y, Murata M, Ito T. Nectin-4: a novel therapeutic target for skin cancers. *Curr Treat opt Oncol*. (2022) 23:578–93. doi: 10.1007/s11864-022-00940-w
105. Milowsky MI, O'Donnell PH, Hoimes CJ, Petrylak DP, Flaig TW, Moon HH, et al. Patient-reported outcomes in patients with advanced urothelial cancer who are ineligible for cisplatin and treated with first-line enfortumab vedotin alone or with pembrolizumab. *J Clin Oncol Off J Am Soc Clin Oncol*. (2024), Jco2301547. doi: 10.1200/JCO.23.01547
106. Loganzo F, Sung M, Gerber HP. Mechanisms of resistance to antibody-drug conjugates. *Mol Cancer Ther*. (2016) 15:2825–34. doi: 10.1158/1535-7163.MCT-16-0408
107. Klümper N, Ralsler DJ, Ellinger J, Roghmann F, Albrecht J, Below E, et al. Membranous NECTIN-4 expression frequently decreases during metastatic spread of urothelial carcinoma and is associated with enfortumab vedotin resistance. *Clin Cancer Res an Off J Am Assoc Cancer Res*. (2023) 29(8):1496–505. doi: 10.1158/1078-0432.CCR-22-1764
108. Aggen DH, Chu CE, Rosenberg JE. Scratching the surface: NECTIN-4 as a surrogate for enfortumab vedotin resistance. *Clin Cancer Res an Off J Am Assoc Cancer Res*. (2023) 29:1377–80. doi: 10.1158/1078-0432.CCR-22-3882
109. Cabaud O, Berger L, Crompot E, Adélaïde J, Finetti P, Garnier S, et al. Overcoming resistance to anti-nectin-4 antibody-drug conjugate. *Mol Cancer Ther*. (2022) 21:1227–35. doi: 10.1158/1535-7163.MCT-22-0013
110. Zhou W, Fang P, Yu D, Ren H, You M, Yin L, et al. Preclinical evaluation of 9MW2821, a site-specific monomethyl auristatin E-based antibody-drug conjugate for treatment of nectin-4-expressing cancers. *Mol Cancer Ther*. (2023) 22:913–25. doi: 10.1158/1535-7163.MCT-22-0743
111. Goldenberg DM. Targeting of cancer with radiolabeled antibodies. *Prospects Imaging Ther Arch Pathol Lab Med*. (1988) 112:580–7.
112. Rigby M, Bennett G, Chen L, Mudd GE, Harrison H, Beswick PJ, et al. BT8009: A nectin-4 targeting bicycle toxin conjugate for treatment of solid tumors. *Mol Cancer Ther*. (2022) 21:1747–56. doi: 10.1158/1535-7163.MCT-21-0875
113. Chester C, Sanmamed MF, Wang J, Melero I. Immunotherapy targeting 4-1BB: mechanistic rationale, clinical results, and future strategies. *Blood*. (2018) 131:49–57. doi: 10.1182/blood-2017-06-741041
114. Sanchez-Paulete AR, Labiano S, Rodriguez-Ruiz ME, Azpilikueta A, Etxeberria I, Bolaños E, et al. Deciphering CD137 (4-1BB) signaling in T-cell costimulation for translation into successful cancer immunotherapy. *Eur J Immunol*. (2016) 46:513–22. doi: 10.1002/eji.201445388
115. Hurov K, Lahdenranta J, Upadhyaya P, Haines E, Cohen H, Repash E, et al. BT7480, a novel fully synthetic Bicycle tumor-targeted immune cell agonistTM (Bicycle TICATM) induces tumor localized CD137 agonism. *J Immunother Cancer*. (2021) 9(11):e002883. doi: 10.1136/jitc-2021-002883
116. Upadhyaya P, Kristensson J, Lahdenranta J, Repash E, Ma J, Kublin J, et al. Discovery and optimization of a synthetic class of nectin-4-targeted CD137 agonists for immuno-oncology. *J Med Chem*. (2022) 65:9858–72. doi: 10.1021/acs.jmedchem.2c00505