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RECEIVED 11 November 2023

ACCEPTED 03 January 2024

PUBLISHED 29 January 2024

CITATION

Zheng H, Wu X, Guo L and Liu J (2024)
MyD88 signaling pathways: role
in breast cancer.
Front. Oncol. 14:1336696.
doi: 10.3389/fonc.2024.1336696

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MyD88 signaling pathways: role in breast cancer

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MyD88 plays a central role in breast cancer, exerting a multitude of effects that carry substantial implications. Elevated MyD88 expression is closely associated with aggressive tumor characteristics, suggesting its potential as a valuable prognostic marker and therapeutic target. MyD88 exerts influence over several critical aspects of breast cancer, including metastasis, recurrence, drug resistance, and the regulation of cancer stem cell properties. Furthermore, MyD88 modulates the release of inflammatory and chemotactic factors, thereby shaping the tumor's immune microenvironment. Its role in immune response modulation underscores its potential in influencing the dynamic interplay between tumors and the immune system. MyD88 primarily exerts intricate effects on tumor progression through pathways such as Phosphoinositide 3-kinases/Protein kinase B (PI3K/Akt), Toll-like Receptor/Nuclear Factor Kappa B (TLR/NF- κ B), and others. Nevertheless, in-depth research is essential to unveil the precise mechanisms underlying the diverse roles of MyD88 in breast cancer. The translation of these findings into clinical applications holds great promise for advancing precision medicine approaches for breast cancer patients, ultimately enhancing prognosis and enabling the development of more effective therapeutic strategies.

KEYWORDS

breast cancer, myeloid differentiation factor 88, epithelial-mesenchymal transition, tumor microenvironment, biomarker

Introduction

Breast cancer (BC) is one of the most prevalent malignant tumors in women (1). The global burden of breast cancer annually amounts to over 2.2 million diagnosed cases, resulting in over 600,000 fatalities (2). Encouragingly, in high-income countries, there has been a decline in breast cancer mortality, largely attributed to advancements in treatment modalities. However, there's a steady rise in breast cancer incidence, and it remains the

most common cause of cancer-related deaths in low-income countries (3). Clinically, specific subtypes of breast cancer are characterized by their histological appearance and the expression of hormone receptors and growth factors, namely estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) (4). Characterized by remarkable heterogeneity, breast cancer is primarily driven by factors such as metastasis, recurrence, and drug resistance, which contribute significantly to patient mortality (3).

Against this backdrop, Myeloid Differentiation Factor 88 (MyD88) assumes a crucial role in breast cancer. The MYD88 gene provides instructions for producing proteins involved in signaling within immune cells. Acting as an adapter, the MyD88 protein plays a crucial role in innate immunity by mediating cell activation through Toll-like receptors (TLRs). TLRs interact with adapter protein MyD88, initiating the activation of two key transcription factors: NF- κ B, a dimeric protein responsible for expressing various inflammatory cytokines, chemokines, adhesion, and co-stimulatory molecules, triggering acute inflammation and stimulating the immune response. IRFs, a group of proteins responsible for expressing type I interferons, to establish the so-called antiviral state of cells (5). While MyD88 is renowned for the role in recognizing and responding to microbial pathogens in innate immunity (6, 7), it also plays a pivotal role in numerous non-immune processes, particularly within the context of tumor development. Currently, MyD88 is regarded as a critical signaling molecule with diverse roles in the development and progression of breast cancer. Engaging in various signaling pathways, MyD88 influences proliferation, migration, and invasion of breast cancer cells. Through cascading responses such as Phosphoinositide 3-kinases/Protein kinase B (PI3K/Akt), Wntless/Integrated (Wnt), Notch, Hedgehog, and Nuclear Factor kappa B (NF- κ B), it orchestrates the epithelial-mesenchymal transition (EMT), conferring migratory and invasive traits upon tumor cells (8–15). By modulating signaling pathways governing the maintenance and functionality of breast cancer stem cells (BCSCs), MyD88 regulates tumor initiation, progression, recurrence, and therapy resistance (16). Activation of MyD88 intricately correlates with self-renewal, cytokine production and secretion, as well as expansion of tumor stem cells, thereby influencing tumor development and progression.

Through modulation of inflammatory responses and immune reactions, MyD88 impacts the immune microenvironment of breast cancer (17). It governs the effects of tumor cell-derived exosomes, stimulating the production of inflammatory factors and influencing the activity of macrophages, T cells, and NK cells. Moreover, the interaction of MyD88 with immune checkpoint therapy plays a pivotal role, significantly impacting breast cancer treatment outcomes. Notably, the expression level of MyD88 level closely associates with disease severity, prognosis, and staging of breast cancer (18). High MyD88 expression correlates intimately with clinical parameters like tumor size, lymph node metastasis status, and histological grade. Patients with high MyD88 expression tend to experience recurrences or metastases, whereas those with low expression exhibit better prognosis.

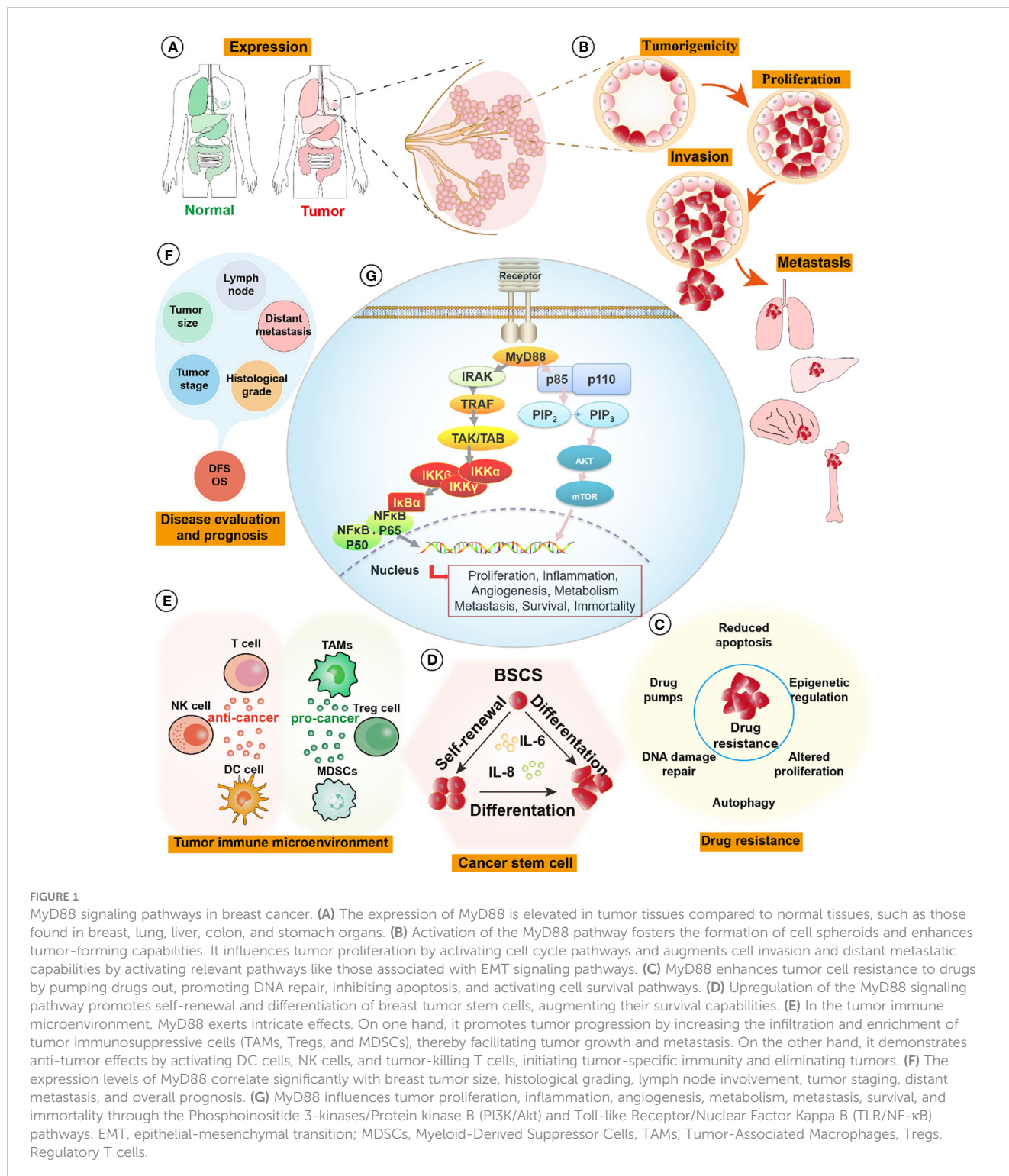
In this review, we provide a comprehensive overview of MyD88's roles in tumorigenesis, metastasis, drug resistance, the

tumor microenvironment, and prognosis in breast cancer (Figure 1). We also explore potential avenues through which this understanding can drive future drug development. By delving deeper into the multifaceted impacts of MyD88 in breast cancer, we anticipate the emergence of novel strategies for the treatment and management of this disease.

The role of MyD88 in clinical disease assessment and prognosis

MyD88 assumes a pivotal role in the clinical evaluation and prognosis of breast cancer. A multitude of studies has revealed a profound association between MyD88 expression levels and the severity and prognostication of breast cancer (Table 1). Elevated MyD88 expression is correlated with larger tumor size, lymph node metastasis, and higher histological grades, suggesting MyD88's potential as a biomarker for appraising breast cancer's invasiveness and progression. The protein expression of MyD88 demonstrates a significant positive correlation with tumor size, stage, axillary lymph node metastasis, and distant metastasis (19). MyD88 protein expression is positively linked with axillary lymph node metastasis and histological grade. Both TLR4 and MyD88 protein expressions are positively correlated with breast cancer cell metastasis (18). Notably, MyD88 protein expression in breast cancer tissue surpasses that in adjacent non-cancerous tissue, and it is positively associated with axillary lymph node metastasis, histological grade, and distant metastasis (18). TLR4 and MyD88 protein expression levels are positively correlated with axillary lymph node metastasis and histological grade, and the co-expression of TLR4 and MyD88 is also positively correlated with breast cancer cell metastasis (18). The expression rate of MyD88 is notably higher in samples from patients with axillary lymph node metastasis (59%) compared to those without metastasis (25.6%). Similarly, the expression rate of MyD88 is higher in patients with stage III disease (65.6%) compared to those with stage I/II disease (44.1%) (18). The expression of MyD88 is intrinsically connected to patient prognosis, with higher MyD88 expression increasing the propensity for recurrence or metastasis. MyD88 expression levels inversely correlate with disease-free survival (DFS) and overall survival (OS), with lower MyD88 expression translating to better patient outcomes (21). A notable divergence in survival rates is evident between high and low MyD88 expression groups, as evidenced by Xiang's study (20). Kaplan-Meier survival curves underscore the significance of MyD88 expression in survival disparities between patients with high MyD88 expression and those with normal expression (20). Furthermore, MyD88's protein expression level exhibits a positive correlation with axillary lymph node metastasis and histological grade, while the co-expression of TLR4 and MyD88 is positively correlated with breast cancer cell metastasis (18).

MyD88, therefore, emerges as a potential diagnostic and therapeutic target in breast cancer. The assessment of MyD88 expression levels offers a means to comprehensively gauge breast cancer invasiveness and predict patient prognosis, thereby



establishing a foundation for informed decisions in personalized treatment.

MyD88 expression in breast cancer

MyD88, a pivotal signaling molecule, has garnered attention for its dysregulated expression in various cancer types, including

colorectal cancer (23), ovarian cancer (24), hepatocellular carcinoma (25), and pancreatic cancer (26), highlighting its role in tumor progression. In breast cancer, the expression of MyD88 varies within breast cancer cells, tumor tissues, and the surrounding microenvironment. While normal breast tissues typically maintain low MyD88 expression levels for physiological functions, breast cancer often exhibits significant upregulation. Studies indicate that adjacent normal tissues and benign breast tumors have lower

TABLE 1 MyD88 expression and clinical evaluation in breast cancer.

Technology	Results	Ref.
IHC	The expression of MyD88 in malignant tumors significantly surpasses that in adjacent normal tissue and benign tumors.	(18–20)
	Significant differences are observed in DFS and OS curves concerning MyD88 expression levels. Patients with lower MyD88 expression exhibit better DFS and OS compared to those with higher expression levels. Multifactorial analysis reveals MyD88 status as an independent risk factor for DFS and OS.	(20, 21)
	MyD88 expression correlates with ER and PR statuses, and patients with higher MyD88 expression experience more frequent recurrence or metastasis.	(21)
	No correlation has been found between MyD88 expression and ER or PR status.	(20)
	MyD88 expression levels relate to breast cancer TNM staging, being more detectable in stage III compared to stage I or II breast cancer patients.	(20)
WB	High expression of MyD88 significantly correlates with tumor size, tumor staging, axillary lymph node metastasis, and distant metastasis.	(19)
	The protein expression level of MyD88 in MDA-MB-231 cells is 1.6 times that of MCF-7 cells and 1.8 times that of MDA-Kb2 cells.	(18)
	High levels of MyD88 protein expression were confirmed in hormone-resistant breast cancer cell lines MDA-MB-231, MDA-MB-231HM, and MDA-MB-468, whereas such confirmation was not observed in MCF-7 and SKBR3 cell lines.	(21)
qRT-PCR	The expression of the MyD88 gene in breast cancer tissues is higher compared to adjacent normal tissues and benign tumors.	(19)
	In MDA-MB-231 cells, the gene expression level of MyD88 is 2.09 times higher than in MCF-7 cells.	(18)
	Zandi et al. assessed the baseline expression levels of MyD88 in different breast cancer cell lines—MCF7, SKBR3, MDA-MB-231, and BT-474—and observed the highest MyD88 mRNA level in MCF-7 cells.	(22)
IF	The average fluorescence intensity of MyD88 in MDA-MB-231 and MCF-7 cells was 0.136 and 0.05, respectively, there was no significant difference between the levels in MCF-7 and MDA-Kb2 cells	(18)

DFS, Disease-Free Survival; IF, Immunofluorescence; IHC: Immunohistochemistry; OS, Overall Survival; qRT-PCR: Quantitative Reverse Transcription Polymerase Chain Reaction; WB, Western Blotting.

MyD88 expression, whereas breast cancer demonstrates heightened and pronounced expression (19).

Differential MyD88 expression may also exist among distinct molecular subtypes of breast cancer. Zandi et al. conducted baseline expression level assessments of MyD88 in various breast cancer cell lines, including MCF7, SKBR3, MDA-MB-231, and BT-474, and found that MyD88 mRNA level was highest in MCF-7 cells (22, 27). However, another study revealed that within the MDA-MB-231 cell

line, both the mRNA and protein expression levels of MyD88 were notably elevated in comparison to MCF-7 and MDA-Kb2 cells (which stably express an androgen- and glucocorticoid-responsive reporter) (18). Furthermore, there is evidence of a connection between MyD88 and tumor cell resistance. Ma et al. confirmed elevated levels of MyD88 protein in hormone-resistant breast cancer cell lines, MDA-MB-231, MDA-MB-231HM, and MDA-MB-468, compared to MCF-7 and SKBR3 cells (18). MyD88 and TLR4 expression and other clinical and pathological parameters. The expression of MyD88 was associated with ER and PR status ($p < 0.001$). TLR4 expression was associated with PR status ($p = 0.015$) (21). In another study, it was demonstrated that there is no correlation between the expression of MyD88 and the status of ER or PR (20). X. Chen et al. found that heightened MyD88 expression is linked to increased malignancy and unfavorable prognosis in breast cancer. Elevated levels of MyD88 expression show a significant correlation with tumor size, staging, axillary lymph node metastasis, and distant metastasis (19).

In summary, MyD88 expression in breast cancer exhibits variability. Understanding the expression patterns of MyD88 contributes to a better comprehension of its roles in breast cancer initiation, progression, and treatment. This knowledge provides essential insights into its potential as a therapeutic target.

Effect of MyD88 on tumorigenicity

The MyD88 protein exerts a profound influence on cancer cell tumorigenesis and the initiation of tumor growth. Serving as a crucial signaling adaptor protein, the TLR/MyD88 pathway plays a pivotal role in tumorigenicity. Activation of the MyD88 pathway promotes tumor formation, while MyD88 knockdown results in reduced clonogenicity in primary ER^{neg} tumors (14). Inhibiting MyD88 leads to decreased clonogenicity, and MyD88 shRNAs result in negative selection in the development of *in vivo* xenograft tumors derived from ER^{neg} breast cancer cells. Depletion of MyD88 in MCF-7 cells markedly inhibits tumor growth in nude mice, with tumor volumes generated from MyD88-deficient MCF-7 cells approximately half the size of those from control cells (20). TLR2, an upstream signaling molecule of MyD88, depletion impedes the tumor-initiating capacity of cells (28). For instance, TLR2-depleted spherical mammospheres cells only gave rise to tumor growth in 5 out of 14 mice, in contrast to control cells, which developed rapidly growing tumors in all mice (28). Additionally, Tlr2^{-/-}MMTV-Wnt1 (Tlr2^{-/-}Wnt1) mice exhibit markedly reduced tumor formation compared with that of their Tlr2^{+/+}MMTV-Wnt1 (Tlr2^{+/+}Wnt1) littermates, with median tumor-free days of 269 and 137, respectively (14). TLR2 neutralizing antibodies also block the growth of two independent ER^{neg} breast cancer xenografts *in vivo*. Short hairpin RNA (shRNA)-mediated knockdown of TLR2 in two breast cell lines (MDA-MB-468 and MDA-MB-231) similarly decreased clonogenic outgrowth (14). Moreover, suppression of constitutive downstream NF- κ B activity in human breast cancer cell lines leads to reduced tumorigenicity (29). Furthermore, inhibiting NF- κ B activity in human breast cancer cell lines can induce cell apoptosis (30) or diminish tumor-forming capability (31). Collectively, these findings underscore the

significance of the TLR2/MyD88/NF- κ B pathway in tumorigenicity. These observations imply that the TLR2/MyD88/NF- κ B pathway is implicated in *de novo* mammary tumor formation.

Effect of MyD88 on breast cancer proliferation

Cell proliferation is a central driver of breast cancer development and progression (32). In breast cancer, MyD88 is recognized as a key regulator with a significant impact on cell proliferation. Our previous studies have demonstrated that lipopolysaccharide (LPS) activates the MyD88 signaling pathway, promoting the *in vitro* proliferation of MCF-7 and MDA-MB-231 cells, as well as enhancing growth in xenograft mouse models *in vivo*. In our preliminary studies, a novel compound (TJ-M2010) was synthesized, drawing inspiration from the MyD88 molecular structure. This compound was engineered to specifically bind to the TIR domain of MyD88, aiming to disrupt the dimerization process of MyD88 (11). The application of TJ-M2010 hinders MyD88/NF- κ B signaling transduction, resulting in the suppression of breast cancer cell proliferation mentioned above (33). Liu et al. employed a small-molecule compound known as 4210, which obstructs MyD88 dimerization (34). Their findings suggested that 4210 inhibition of MyD88 might induce nonspecific cell death in MDA-MB-231 cells (35). Another study demonstrated that treatment using the MyD88 inhibitor (ST2825) resulted in reduced growth of murine mammary carcinomas 4T1, 168, EMT6, and SM1, as well as the human breast cancer cell line MDA-MB-231 (36). In another study, a TLR3 ligand notably increased breast cancer cell proliferation, as indicated by the upregulation of cyclin D1 expression. However, the introduction of a MyD88 inhibitor disrupted the signal transduction of the TLR3-MyD88-NF- κ B (p65)-IL6-Cyclin D1 pathway, leading to reduced breast cancer cell proliferation (37). Holleman et al. also confirmed that reduced MyD88 expression significantly inhibited MCF-7 cell proliferation (38). Similarly, the silencing of the MyD88 gene resulted in an increased G2 phase cell population in MCF-7 cells (20). NF- κ B, a canonical downstream signaling molecule of TLR/MyD88, stimulates the transcription of the cyclin D1 promoter, thereby promoting cell growth (39). Additionally, caffeic acid phenethyl ester inhibited MDA-MB-231 cell proliferation by suppressing the TLR4/MyD88/NF- κ B signaling pathway (40). Furthermore, MyD88 can affect breast cancer proliferation through the PI3K/AKT signaling pathway. The interaction between MyD88 and p85 enhances tumorigenic capabilities through the PI3K/Akt pathway (15). The MyD88 inhibitor TJ-M2010 interferes with the MyD88/PI3K/GSK-3 β axis, consequently restraining breast cancer cell proliferation (33).

In conclusion, MyD88 plays a pivotal role in regulating breast cancer cell proliferation through the activation of multiple key signaling pathways. This highlights MyD88 as a significant potential therapeutic target, and inhibiting its function may contribute to restraining the growth and dissemination of breast cancer cells. However, further research is needed to gain deeper insights into the

regulatory mechanisms of MyD88 in breast cancer cell proliferation, with the goal of developing more precise and effective treatment strategies.

The effect of MyD88 on metastasis

Metastasis represents a pivotal stage in cancer progression, involving the spread of cancer cells from the primary tumor site to distant organs or tissues, where secondary tumors form (41). MyD88 is a molecular orchestrator with multifaceted implications for cellular processes central to cancer metastasis. Multiple studies have underscored the substantial involvement of MyD88 in tumor metastasis. For example, ectopic expression of MyD88 has been shown to enhance the invasion of hepatocellular carcinoma cells, whereas MyD88 inhibition has led to reduced lung metastasis and portal vein tumor thrombosis (15, 25, 42). MyD88 also plays a critical role in breast cancer metastasis. K. Wu et al. found that MyD88 expression was increased in highly invasive BC cell lines compared to low invasive BC cell lines, suggesting that MyD88 could be a potential therapeutic target for the metastasis of BC (18). Research has indicated that the heightened downstream NF- κ B activity resulting from MyD88 elevation leads to increased expression of invasion-associated proteases (43, 44). Moreover, NF- κ B contributes to the induction and maintenance of epithelial-mesenchymal transition (EMT) in Ras-transformed breast epithelial cells, mediating invasive and metastatic tumor phenotypes. Substantial evidence supports the role of NF- κ B in late-stage breast tumor development and metastasis (45). In MDA-MB-231 cells, the downregulation of MyD88 influences NF- κ B nuclear translocation, resulting in a diminished invasion capacity of tumor cells (36). Additionally, by knocking down MyD88, the migration speed of MCF-7 cells is reduced, and their ability to traverse Matrigel is impaired. In another study, tumors with reduced Myd88 levels displayed reduced growth and metastasis (46). MiR-4317 inhibits the proliferation, migration, and invasion of breast cancer cells by targeting MyD88 (47). Advanced glycation end products (AGEs) can promote breast cancer cell migration and invasion through the activation of the RAGE/TLR4/MyD88 signaling cascade (48). Berberine significantly inhibits the TLR9-MyD88-NF- κ B pathway, reversing breast cancer metastasis (49). LPS enhances invasiveness and metastatic potential of breast cancer cells by upregulating the MyD88-BLT2-NF- κ B signaling cascade (50). MCF-7 cell lines, after 48 hours of incubation with 0.5 μ M morphine, exhibit downregulated TLR4/MyD88/NF- κ B pathways, resulting in decreased migration (51, 52). TLR4 silencing reduces the expression of MyD88 and MMP9, significantly inhibiting cell migration and invasion (53). Furthermore, a TLR4 antagonist blocks TLR4 and MyD88 expression, as well as cell invasion and migration (54).

EMT, which refers to the transformation of epithelial cells into mesenchymal-like cells, conferring migratory and invasive properties (55), is a pivotal process in cancer. Various cascading reactions relevant to cancer have been established as critical regulatory signals of EMT (56, 57). The MyD88/NF- κ B signaling

pathway plays a crucial role in Snail-mediated EMT (58). EpRas cells are oncogenic and completely polarized Ha-Ras-transformed EpH4 mammary epithelial cells that undergo EMT, leading to the generation of mesenchyme-like cells (EpRasXT cells). Blocking the MyD88/NF- κ B pathway eliminates the metastatic capability of mammary epithelial cells in murine models. Additionally, the reversal of NF- κ B activity in EpRasXT cells nullifies the EMT process (45). Curcumin inhibits LPS-induced EMT in MDA-MB-231 and MCF-7 cells through the TLR4/MyD88/NF- κ B/Snail signaling pathway (59, 60). The MyD88 inhibitor TJ-M2010-2 reverses TGF- β 1-induced HK-2 cell EMT (61, 62). Similarly, the MyD88 inhibitor TJ-M2010-2 suppresses the proliferation, migration, and invasion of breast cancer cells by intervening in the MyD88/GSK-3 β and MyD88/NF- κ B signaling pathways (33).

In conclusion, MyD88 promotes cancer cell migration, invasion, and the establishment of secondary tumors in distant organs through diverse pathways, significantly impacting breast cancer metastasis. A comprehensive understanding of MyD88's mechanisms in metastasis aids in devising effective strategies to inhibit cancer spread, thereby enhancing patient prognosis.

The effect of MyD88 on drug resistance

Drug resistance poses a formidable challenge in cancer therapy, frequently resulting in reduced treatment effectiveness and disease relapse (63). MyD88 exerts a significant impact on drug resistance in breast cancer cells and is recognized as a key player associated with paclitaxel (PTX) resistance. Xiang et al. demonstrated the reversal of PTX resistance in breast cancer cells by targeting the miRNA-149-5p/MyD88 axis using ursolic acid (UA) (8). The PI3K/Akt pathway, a pivotal signaling route, becomes activated in PTX-resistant breast cancer (64). The observed heightened sensitivity of MCF-7 cells to paclitaxel following MyD88 downregulation implies its impact on inhibiting NF- κ B activation, thereby blocking the PI3K/Akt signaling pathway (20). Elevated MyD88 levels in MDA-MB-231/PTX-resistant cells correlated with enhanced resistance, but MyD88 silencing increased PTX sensitivity (8). Activation of Akt may be linked to reduced paclitaxel-induced apoptosis (20), and MyD88 knockdown significantly suppressed Akt pathway activation in 231/PTX cells (8). The MyD88-regulated PI3K/Akt pathway appeared to mediate paclitaxel resistance in breast cancer cells by modulating Bax/Bcl-2 expression (8). Overexpression of MyD88 protein was associated with acquired paclitaxel resistance in triple-negative breast cancer (TNBC) cells. Dimethoxy-substituted compounds significantly reduced MyD88 overexpression in TLR4⁺ MDA-MB-231 cells and enhanced PTX activity (65). *In vitro* and *in vivo* experiments reversed paclitaxel resistance by inhibiting the target gene MyD88 (66).

Furthermore, the MyD88/NF- κ B signaling pathway plays a central role in drug resistance in breast cancer. Downregulation of MyD88 increased sensitivity to paclitaxel in MCF-7 cells by suppressing NF- κ B activation and blocking the PI3K/Akt pathway (20). The NF- κ B/Notch1 regulatory loop contributes to the

maintenance of breast cancer stem cells and drug resistance (16). Additionally, the NF- κ B pathway promotes the transcription of ABCG2/BCRP transporters (67). This pro-survival signaling pathway becomes more active in response to radiotherapy, chemotherapy, and trastuzumab in breast cancer cells (68–70). Consequently, the MyD88/NF- κ B pathway is involved in mediating drug resistance mechanisms in tumor cells. Despite substantial evidence supporting the regulatory role of MyD88 in drug resistance, some studies have yielded contrasting results, such as LPS pre-treatment sensitizing MyD88-positive cells to docetaxel cytotoxicity. This suggests that pathways induced by LPS stimulation could serve as attractive targets for overcoming therapy-resistant diseases (71).

A comprehensive understanding of MyD88's role in drug resistance is crucial for devising strategies to overcome resistance and enhance the effectiveness of cancer treatment. Targeting MyD88 or its associated signaling pathways might offer a potential approach to increase cancer cell sensitivity to treatment and improve therapeutic outcomes. Further research is required to elucidate the potential mechanisms underlying MyD88-mediated drug resistance in breast cancer and other cancer types, as well as to explore therapeutic intervention methods.

The effect of MyD88 on cancer stem cells

Breast cancer stem cells (BCSCs) constitute a small subset of tumor cells endowed with the unique capacity for self-renewal and differentiation into diverse cell types (72). These cells wield a pivotal role in tumor initiation, progression, recurrence, and resistance to therapy.

MyD88 has been revealed as a regulator of the characteristics and behavior of BCSCs through diverse mechanisms, actively participating in signaling pathways that govern the maintenance and functions of BCSCs. For instance, Conti et al. observed that the stimulation of TLR2 with ligands like Pam3CSK4, LTA, and PGN-SA significantly enhances mammosphere formation. In contrast, the action of the MyD88 homodimerization-inhibitory peptide markedly hinders the formation of spherical mammospheres (28). Silencing the TLR2/MyD88 signaling pathway also reduces mammosphere generation in both mouse (4T1) and human (MDA-MB-231, MCF7) mammary cells (28). The upregulation of the MyD88/NF- κ B signaling pathway acts as a pro-survival route implicated in the self-renewal of BCSCs (16). The NF- κ B protein p65 directly binds to the Gli1 promoter, enhancing its transcription (73). Additionally, the MyD88/NF- κ B signaling pathway governs the expression of various cytokines, notably IL-6 and IL-8, which are intricately linked to tumor progression and the survival of BCSCs (74, 75). Extracellular IL-6 induces malignant characteristics in ductal breast cancer stem/progenitor cells (74), while the overexpression of IL-8 promotes stemness, stromal traits, acquisition of resistance, and the recruitment of immunosuppressive cells that foster tumor growth in the tumor microenvironment (75). Novel therapies targeting IL-8 receptor signaling may serve as a

means to halt tumor progression. Machilin D reduces the secretion of IL-6/IL-8 and inhibits sphere growth (76).

In conclusion, MyD88 plays a significant role in regulating breast cancer stem cell characteristics and functions. A more profound understanding of the mechanisms through which MyD88 influences BCSCs holds the potential for the development of targeted therapies against these aggressive tumor-initiating cells. This offers both theoretical and practical implications for enhancing breast cancer treatment outcomes. Further research is warranted to comprehensively elucidate the mechanisms of MyD88 in BCSCs and its potential value as a therapeutic target.

The influence of MyD88 on the tumor immune microenvironment

The influence of MyD88 on the tumor immune microenvironment is of great significance in the context of breast cancer progression. The tumor microenvironment is a complex ecosystem comprising tumor cells and their immediate surroundings, exercising a profound impact on tumor development (77–79). Through the modulation of inflammatory responses and immune reactions, MyD88 orchestrates changes within the tumor immune microenvironment. Exosomes released by tumor cells can activate the TLR/MyD88/NF- κ B signaling pathway in macrophages, prompting the release of pro-inflammatory cytokines (such as IL-6, TNF- α , GCSF, and CCL2) (80). Within breast cancer stem cells, the HMGB1/TLR2/MyD88 axis governs NF- κ B activation, IL-6 and TGF- β production, and the activation of STAT3 and Smad3. IL-6, stimulated through STAT3, fosters malignant properties, expanding the breast cancer stem cell population (81). The TGF- β signaling pathway contributes to the maintenance, survival, and enhanced migratory capacity of the tumor stem cell population (82, 83). TNF- α promotes the expression of carcinogenic CCL5 variants in breast cancer cells, instigating tumor cell detachment and dissemination (57). Activation of the TLR2/MyD88/NF- κ B pathway elevates the autocrine secretion of VEGF and MMP9 in MDA-MB-231 cells, recognized as pivotal factors for breast cancer cell invasion and adhesion (27). The inhibition of NF- κ B significantly reduces the invasiveness of MDA-MB-231 cells (84, 85). NF- κ B inhibition profoundly diminishes the invasiveness of MDA-MB-231 cells (86, 87).

The MyD88 pathway also mediates immune responses that impact breast cancer progression. For example, Astragalus polysaccharide (APS) may regulate host immunity and exert anti-tumor effects by activating the TLR4-mediated MyD88-dependent signaling pathway (17). A Arazyme activates TLR4-MyD88-TRIF, which reduces primary and metastatic tumor development in the 4T1 murine breast cancer model, thus extending survival (88). Inhibition of proprotein convertases 1/3 steers NR8383 macrophages toward an M1 activation phenotype mainly via TLR4/MyD88 signaling, leading to the secretion of factors that attract immature T helper lymphocytes and promote cytotoxic responses (89). Polysaccharide peptide (PSP) acts by upregulating the TLR4-TIRAP/MAL-MyD88 signaling

pathway in peripheral blood mononuclear cells (PBMCs) from breast cancer patients, inducing the secretion of IL-12, IL-6, and TNF- α , thus serving as an immune adjuvant (90). Activation of the TLR5/MyD88/NME3/NF κ B signaling pathway enhances host immunity, enhances the clearance of tumor xenografts, and potentially augments the effectiveness of immunotherapy, prolonging survival in breast cancer patients (91). Stimulation of TLR8 ligands through the MyD88/IRAK4/TRAF6/p38 signaling pathway reverses the suppressive function of CD4⁺ CD25⁺ Treg cells, eliminating the inhibitory effect of BTIL31 $\gamma\delta$ 1 Treg cells on T cell proliferation and function, as well as dendritic cell maturation and function (92).

Indeed, various immune effects have been observed in other cancers as well. Our previous research demonstrated that TJ-M2010-2 successfully mitigated inflammation induced by AOM/DSS, thereby altering the tumor microenvironment and reducing the incidence of colorectal cancer from 100% to 0% (11). MyD88 plays a critical role in tumor-derived exosome-mediated MDSC expansion and subsequent tumor metastasis (93). Furthermore, Pixatimod (PG545), a novel clinical-stage immune modulator, amplifies T cell infiltration when combined with anti-PD-1 therapy through the MyD88-dependent TLR9 pathway. Simultaneously, it inhibits the infiltration of tumor-associated macrophages (TAMs) and activates dendritic cells (DCs), thereby stimulating natural killer (NK) cells (94). IL-33 has the capacity to promote the differentiation and maturation of DC cells via the MyD88 pathway. This results in upregulated tumor immunity in CD8⁺T cells and NK cells while inhibiting the proliferation of lung cancer cells (95).

The multifaceted influence of MyD88 on the tumor immune microenvironment involves the orchestration of multiple cytokines and intricate signaling pathways. A comprehensive comprehension of the role of MyD88 in shaping the tumor microenvironment greatly enhances our understanding of breast cancer development mechanisms, and it offers a strong theoretical foundation for the development of innovative therapeutic strategies. It is imperative that further research endeavors are undertaken to elucidate the intricate mechanisms through which MyD88 exerts its influence on the tumor immune microenvironment and to explore its potential applications in the development of novel treatments.

Conclusions and prospects

MyD88 assumes multifaceted, pivotal roles in the landscape of breast cancer, influencing diverse aspects such as tumor development, drug resistance, stem cell properties, and the intricacies of the immune microenvironment. A comprehensive understanding of the mechanisms underpinning MyD88's actions in breast cancer not only unravels the molecular complexities governing tumor progression but also unveils fresh perspectives and potential targets for shaping breast cancer treatment strategies. MyD88 emerges as a valuable asset in prognosis assessment, offering guidance for therapeutic choices and the development of innovative treatment modalities. Nonetheless, there exists a need for

further research to unveil the precise workings of MyD88 and translate its applications into clinical practice, with the ultimate aim of achieving more precise and personalized approaches to managing breast cancer.

Author contributions

HZ: Conceptualization, Project administration, Writing – original draft. XW: Funding acquisition, Project administration, Resources, Writing – original draft. LG: Investigation, Methodology, Writing – original draft. JL: Conceptualization, Funding acquisition, Project administration, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. The National Natural Science Foundation of China (Grant No. 81802895), the Natural Science Foundation of Hubei Province,

China (Grant No.2023AFB493), and the Health and Family Planning Commission of Wuhan Municipality (Grant No.WX17Q16) awarded to JL. The Research Projects of Biomedical Center of Hubei Cancer Hospital (2022SWZX09) awarded to XW.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Siegel RL, Miller KD, Wagle NS, Jemal A. Cancer statistics, 2023. *CA Cancer J Clin* (2023) 73(1):17–48. doi: 10.3322/caac.21763
2. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/caac.21660
3. Harbeck N, Gnant M. Breast cancer. *Lancet* (2017) 389(10074):1134–50. doi: 10.1016/S0140-6736(16)31891-8
4. Li Cl, Daling JR, Malone KE. Incidence of invasive breast cancer by hormone receptor status from 1992 to 1998. *J Clin Oncol* (2003) 21(1):28–34. doi: 10.1200/JCO.2003.03.088
5. von Bernuth H, Picard C, Jin Z, Pankla R, Xiao H, Ku CL, et al. Pyogenic bacterial infections in humans with MyD88 deficiency. *Science* (2008) 321(5889):691–6. doi: 10.1126/science.1158298
6. Serbina NV, Kuziel W, Flavell R, Akira S, Rollins B, Pamer EG. Sequential MyD88-independent and -dependent activation of innate immune responses to intracellular bacterial infection. *Immunity* (2003) 19(6):891–901. doi: 10.1016/s1074-7613(03)00330-3
7. Chen H, Jiang Z. The essential adaptors of innate immune signaling. *Protein Cell* (2013) 4(1):27–39. doi: 10.1007/s13238-012-2063-0
8. Xiang F, Fan Y, Ni Z, Liu Q, Zhu Z, Chen Z, et al. Ursolic acid reverses the chemoresistance of breast cancer cells to paclitaxel by targeting miRNA-149-5p/myD88. *Front Oncol* (2019) 9:501. doi: 10.3389/fonc.2019.00501
9. Bohnert KR, Goli P, Roy A, Sharma AK, Xiong G, Gallot YS, et al. The toll-like receptor/myD88/XBP1 signaling axis mediates skeletal muscle wasting during cancer cachexia. *Mol Cell Biol* (2019) 39(15):e00184-19. doi: 10.1128/MCB.00184-19
10. Delitto D, Delitto AE, DiVita BB, Pham K, Han S, Hartlage ER, et al. Human pancreatic cancer cells induce a myD88-dependent stromal response to promote a tumor-tolerant immune microenvironment. *Cancer Res* (2017) 77(3):672–83. doi: 10.1158/0008-5472.CAN-16-1765
11. Xie L, Jiang FC, Zhang LM, He WT, Liu JH, Li MQ, et al. Targeting of myD88 homodimerization by novel synthetic inhibitor TJ-M2010-5 in preventing colitis-associated colorectal cancer. *J Natl Cancer Inst* (2016) 108(4):djv364. doi: 10.1093/jnci/djv364
12. Park GS, Kim JH. LPS up-regulates ICAM-1 expression in breast cancer cells by stimulating a myD88-BLT2-ERK-linked cascade, which promotes adhesion to monocytes. *Mol Cells* (2015) 38(9):821–8. doi: 10.14348/molcells.2015.0174
13. Koch AT, Love-Homan L, Espinosa-Cotton M, Stanam A, Simons AL. MyD88-dependent signaling decreases the antitumor efficacy of epidermal growth factor receptor inhibition in head and neck cancer cells. *Cancer Res* (2015) 75(8):1657–67. doi: 10.1158/0008-5472.CAN-14-2061
14. Scheeren FA, Kuo AH, van Weele LJ, Cai S, Glykofridis I, Sikandar SS, et al. A cell-intrinsic role for TLR2-MYD88 in intestinal and breast epithelia and oncogenesis. *Nat Cell Biol* (2014) 16(12):1238–48. doi: 10.1038/ncb3058
15. Jia RJ, Cao L, Zhang L, Jing W, Chen R, Zhu MH, et al. Enhanced myeloid differentiation factor 88 promotes tumor metastasis via induction of epithelial-mesenchymal transition in human hepatocellular carcinoma. *Cell Death Dis* (2014) 5:e1103. doi: 10.1038/cddis.2014.71
16. Husemann Y, Geigl JB, Schubert F, Musiani P, Meyer M, Burghart E, et al. Systemic spread is an early step in breast cancer. *Cancer Cell* (2008) 13(1):58–68. doi: 10.1016/j.ccr.2007.12.003
17. Zhou L, Liu Z, Wang Z, Yu S, Long T, Zhou X, et al. Astragalus polysaccharides exerts immunomodulatory effects via TLR4-mediated MyD88-dependent signaling pathway *in vitro* and *in vivo*. *Sci Rep* (2017) 7:44822. doi: 10.1038/srep44822
18. Wu K, Zhang H, Fu Y, Zhu Y, Kong L, Chen L, et al. TLR4/MyD88 signaling determines the metastatic potential of breast cancer cells. *Mol Med Rep* (2018) 18(3):3411–20. doi: 10.3892/mmr.2018.9326
19. Chen X, Zhao F, Zhang H, Zhu Y, Wu K, Tan G. Significance of TLR4/MyD88 expression in breast cancer. *Int J Clin Exp Pathol* (2015) 8(6):7034–9.
20. Xiang F, Ni Z, Zhan Y, Kong Q, Xu J, Jiang J, et al. Increased expression of MyD88 and association with paclitaxel resistance in breast cancer. *Tumour Biol* (2016) 37(5):6017–25. doi: 10.1007/s13277-015-4436-5
21. Ma FJ, Liu ZB, Hu X, Ling H, Li S, Wu J, et al. Prognostic value of myeloid differentiation primary response 88 and Toll-like receptor 4 in breast cancer patients. *PLoS One* (2014) 9(10):e111639. doi: 10.1371/journal.pone.0111639
22. Zandi Z, Kashani B, Poursani EM, Bashash D, Kabuli M, Momeny M, et al. TLR4 blockade using TAK-242 suppresses ovarian and breast cancer cells invasion through the inhibition of extracellular matrix degradation and epithelial-mesenchymal transition. *Eur J Pharmacol* (2019) 853:256–63. doi: 10.1016/j.ejphar.2019.03.046
23. Wang EL, Qian ZR, Nakasono M, Tanahashi T, Yoshimoto K, Bando Y, et al. High expression of Toll-like receptor 4/myeloid differentiation factor 88 signals correlates with poor prognosis in colorectal cancer. *Br J Cancer* (2010) 102(5):908–15. doi: 10.1038/sj.bjc.6605558
24. Szajnik M, Szczepanski MJ, Czystowska K, Elishaev E, Mandapathil M, Nowak-Markwitz E, et al. TLR4 signaling induced by lipopolysaccharide or paclitaxel regulates tumor survival and chemoresistance in ovarian cancer. *Oncogene* (2009) 28(49):4353–63. doi: 10.1038/onc.2009.289
25. Liang B, Chen R, Wang T, Cao L, Liu Y, Yin F, et al. Myeloid differentiation factor 88 promotes growth and metastasis of human hepatocellular carcinoma. *Clin Cancer Res* (2013) 19(11):2905–16. doi: 10.1158/1078-0432.CCR-12-1245

26. Ochi A, Nguyen AH, Bedrosian AS, Mushlin HM, Zarbaksh S, Barilla R, et al. MyD88 inhibition amplifies dendritic cell capacity to promote pancreatic carcinogenesis via Th2 cells. *J Exp Med* (2012) 209(9):1671–87. doi: 10.1084/jem.20111706
27. Xie W, Wang Y, Huang Y, Yang H, Wang J, Hu Z. Toll-like receptor 2 mediates invasion via activating NF-kappaB in MDA-MB-231 breast cancer cells. *Biochem Biophys Res Commun* (2009) 379(4):1027–32. doi: 10.1016/j.bbrc.2009.01.009
28. Conti L, Lanzardo S, Arigoni M, Antonazzo R, Radaelli E, Cantarella D, et al. The noninflammatory role of high mobility group box 1/Toll-like receptor 2 axis in the self-renewal of mammary cancer stem cells. *FASEB J* (2013) 27(12):4731–44. doi: 10.1096/fj.13-230201
29. Romieu-Mourez R, Landesman-Bollag E, Seldin DC, Traish AM, Mercurio F, Sonenshein GE. Roles of IKK kinases and protein kinase CK2 in activation of nuclear factor-kappaB in breast cancer. *Cancer Res* (2001) 61(9):3810–8.
30. Sovak MA, Bellas RE, Kim DW, Zanieski GJ, Rogers AE, Traish AM, et al. Aberrant nuclear factor-kappaB/Rel expression and the pathogenesis of breast cancer. *J Clin Invest* (1997) 100(12):2952–60. doi: 10.1172/JCI119848
31. Romieu-Mourez R, Landesman-Bollag E, Seldin DC, Sonenshein GE. Protein kinase CK2 promotes aberrant activation of nuclear factor-kappaB, transformed phenotype, and survival of breast cancer cells. *Cancer Res* (2002) 62(22):6770–8.
32. Booy EP, McRae EK, Koul A, Lin F, McKenna SA. The long non-coding RNA BC200 (BCYRN1) is critical for cancer cell survival and proliferation. *Mol Cancer* (2017) 16(1):109. doi: 10.1186/s12943-017-0679-7
33. Liu JH, Chen C, Li ZY, Zou ZM, Gao DC, Zhang X, et al. The MyD88 inhibitor TJ-M2010-2 suppresses proliferation, migration and invasion of breast cancer cells by regulating MyD88/GSK-3beta and MyD88/NF-kappaB signalling pathways. *Exp Cell Res* (2020) 394(2):112157. doi: 10.1016/j.yexcr.2020.112157
34. Olson MA, Lee MS, Kissner TL, Alam S, Waugh DS, Saikh KU. Discovery of small molecule inhibitors of MyD88-dependent signaling pathways using a computational screen. *Sci Rep* (2015) 5:14246. doi: 10.1038/srep14246
35. Liu X, Yao JJ, Chen Z, Lei W, Duan R, Yao Z. Lipopolysaccharide sensitizes the therapeutic response of breast cancer to IAP antagonist. *Front Immunol* (2022) 13:906357. doi: 10.3389/fimmu.2022.906357
36. Egunsola AT, Zawislak CL, Akuffo AA, Chalmers SA, Ewer JC, Vail CM, et al. Growth, metastasis, and expression of CCL2 and CCL5 by murine mammary carcinomas are dependent upon Myd88. *Cell Immunol* (2012) 272(2):220–9. doi: 10.1016/j.cellimm.2011.10.008
37. Singh A, Devkar R, Basu A. Myeloid differentiation primary response 88-cyclin D1 signaling in breast cancer cells regulates toll-like receptor 3-mediated cell proliferation. *Front Oncol* (2020) 10:1780. doi: 10.3389/fonc.2020.01780
38. Holleman A, Chung I, Olsen RR, Kwak B, Mizokami A, Saijo N, et al. miR-135a contributes to paclitaxel resistance in tumor cells both *in vitro* and *in vivo*. *Oncogene* (2011) 30(43):4386–98. doi: 10.1038/ncr.2011.148
39. Baldwin AS. Control of oncogenesis and cancer therapy resistance by the transcription factor NF-kappaB. *J Clin Invest* (2001) 107(3):241–6. doi: 10.1172/JCI11991
40. Chang H, Wang Y, Yin X, Liu X, Xuan H. Ethanol extract of propolis and its constituent caffeic acid phenethyl ester inhibit breast cancer cells proliferation in inflammatory microenvironment by inhibiting TLR4 signal pathway and inducing apoptosis and autophagy. *BMC Complement Altern Med* (2017) 17(1):471. doi: 10.1186/s12906-017-1984-9
41. Klein CA. Cancer progression and the invisible phase of metastatic colonization. *Nat Rev Cancer* (2020) 20(11):681–94. doi: 10.1038/s41568-020-00300-6
42. Roessler S, Jia HL, Budhu A, Forgues M, Ye QH, Lee JS, et al. A unique metastasis gene signature enables prediction of tumor relapse in early-stage hepatocellular carcinoma patients. *Cancer Res* (2010) 70(24):10202–12. doi: 10.1158/0008-5472.CAN-10-2607
43. Karin M. Nuclear factor-kappaB in cancer development and progression. *Nature* (2006) 441(7092):431–6. doi: 10.1038/nature04870
44. Karin M, Greten FR. NF-kappaB: linking inflammation and immunity to cancer development and progression. *Nat Rev Immunol* (2005) 5(10):749–59. doi: 10.1038/nri1703
45. Huber MA, Azoitei N, Baumann B, Grunert S, Sommer A, Pehamberger H, et al. NF-kappaB is essential for epithelial-mesenchymal transition and metastasis in a model of breast cancer progression. *J Clin Invest* (2004) 114(4):569–81. doi: 10.1172/JCI21358
46. Chalmers SA, Eidelman AS, Ewer JC, Ricca JM, Serrano A, Tucker KC, et al. A role for HMGB1, HSP60 and Myd88 in growth of murine mammary carcinoma *in vitro*. *Cell Immunol* (2013) 282(2):136–45. doi: 10.1016/j.cellimm.2013.04.014
47. Sheng Y, Hu R, Zhang Y, Luo W. MicroRNA-4317 predicts the prognosis of breast cancer and inhibits tumor cell proliferation, migration, and invasion. *Clin Exp Med* (2020) 20(3):417–25. doi: 10.1007/s10238-020-00625-4
48. Zheng X, Zhao Y, Jia Y, Shao D, Zhang F, Sun M, et al. Biomimetic co-assembled nanodrug of doxorubicin and berberine suppresses chemotherapy-exacerbated breast cancer metastasis. *Biomaterials* (2021) 271:120716. doi: 10.1016/j.biomaterials.2021.120716
49. Bourguignon LY, Wong G, Earle CA, Xia W. Interaction of low molecular weight hyaluronan with CD44 and toll-like receptors promotes the actin filament-associated protein 110-actin binding and MyD88-NFkappaB signaling leading to proinflammatory cytokine/chemokine production and breast tumor invasion. *Cytoskeleton (Hoboken)* (2011) 68(12):671–93. doi: 10.1002/cm.20544
50. Park GS, Kim JH. Myeloid differentiation primary response gene 88-leukotriene B4 receptor 2 cascade mediates lipopolysaccharide-potentiated invasiveness of breast cancer cells. *Oncotarget* (2015) 6(8):5749–59. doi: 10.18632/oncotarget.3304
51. Haghjooy-Javanmard S, Ghasemi A, Laher I, Zarrin B, Dana N, Vaseghi G. Influence of morphine on TLR4/ NF-kB signaling pathway of MCF-7 cells. *Bratisl Lek Listy* (2018) 119(4):229–33. doi: 10.4149/BLL_2018_043
52. Andela VB, Schwarz EM, Puzas JE, O'Keefe RJ, Rosier RN. Tumor metastasis and the reciprocal regulation of prometastatic and antimetastatic factors by nuclear factor kappaB. *Cancer Res* (2000) 60(23):6557–62.
53. Pan S, Guan Y, Ma Y, Cui Q, Tang Z, Li J, et al. Advanced glycation end products correlate with breast cancer metastasis by activating RAGE/TLR4 signaling. *BMJ Open Diabetes Res Care* (2022) 10(2):e002697. doi: 10.1136/bmjdr-2021-002697
54. Yang H, Wang B, Wang T, Xu L, He C, Wen H, et al. Toll-like receptor 4 prompts human breast cancer cells invasiveness via lipopolysaccharide stimulation and is overexpressed in patients with lymph node metastasis. *PLoS One* (2014) 9(10):e109980. doi: 10.1371/journal.pone.0109980
55. De Craene B, Berx G. Regulatory networks defining EMT during cancer initiation and progression. *Nat Rev Cancer* (2013) 13(2):97–110. doi: 10.1038/nrc3447
56. Huber MA, Kraut N, Beug H. Molecular requirements for epithelial-mesenchymal transition during tumor progression. *Curr Opin Cell Biol* (2005) 17(5):548–58. doi: 10.1016/j.ceb.2005.08.001
57. Nieto MA, Huang RY, Jackson RA, Thiery JP. EMT: 2016. *Cell* (2016) 166(1):21–45. doi: 10.1016/j.cell.2016.06.028
58. Chua HL, Bhat-Nakshatri P, Clare SE, Morimiya A, Badve S, Nakshatri H. NF-kappaB represses E-cadherin expression and enhances epithelial to mesenchymal transition of mammary epithelial cells: potential involvement of ZEB-1 and ZEB-2. *Oncogene* (2007) 26(5):711–24. doi: 10.1038/sj.onc.1209808
59. Huang T, Chen Z, Fang L. Curcumin inhibits LPS-induced EMT through downregulation of NF-kappaB-Snail signaling in breast cancer cells. *Oncol Rep* (2013) 29(1):117–24. doi: 10.3892/or.2012.2080
60. Chen MC, Chang WW, Kuan YD, Lin ST, Hsu HC, Lee CH. Resveratrol inhibits LPS-induced epithelial-mesenchymal transition in mouse melanoma model. *Innate Immun* (2012) 18(5):685–93. doi: 10.1177/1753425912436589
61. Liu JH, He L, Zou ZM, Ding ZC, Zhang X, Wang H, et al. A novel inhibitor of homodimerization targeting myD88 ameliorates renal interstitial fibrosis by counteracting TGF-beta1-induced EMT *in vivo* and *in vitro*. *Kidney Blood Press Res* (2018) 43(5):1677–87. doi: 10.1159/000494745
62. Zhang LM, Liu JH, Xue CB, Li MQ, Xing S, Zhang X, et al. Pharmacological inhibition of MyD88 homodimerization counteracts renal ischemia reperfusion-induced progressive renal injury *in vivo* and *in vitro*. *Sci Rep* (2016) 6:26954. doi: 10.1038/srep26954
63. Nussinov R, Tsai CJ, Jang H. Anticancer drug resistance: An update and perspective. *Drug Resist Update* (2021) 59:100796. doi: 10.1016/j.drug.2021.100796
64. Zhang W, Zheng X, Meng T, You H, Dong Y, Xing J, et al. SET protein overexpression contributes to paclitaxel resistance in MCF-7/S cells through PI3K/Akt pathway. *J Drug Target* (2017) 25(3):255–63. doi: 10.1080/1061186X.2016.1245307
65. Poh Yen K, Stanslas J, Zhang T, Li H, Wang X, Kok Meng C, et al. Synthesis of small molecules targeting paclitaxel-induced MyD88 expression in triple-negative breast cancer cell lines. *Bioorg Med Chem* (2021) 49:116442. doi: 10.1016/j.bmc.2021.116442
66. Zhang L, Chen T, Yan L, Xu H, Wang Y, Li Y, et al. MiR-155-3p acts as a tumor suppressor and reverses paclitaxel resistance via negative regulation of MYD88 in human breast cancer. *Gene* (2019) 700:85–95. doi: 10.1016/j.gene.2019.02.066
67. Saha S, Mukherjee S, Khan P, Kajal K, Mazumdar M, Manna A, et al. Aspirin suppresses the acquisition of chemoresistance in breast cancer by disrupting an NFkappaB-IL6 signaling axis responsible for the generation of cancer stem cells. *Cancer Res* (2016) 76(7):2000–12. doi: 10.1158/0008-5472.CAN-15-1360
68. Korkaya H, Kim GI, Davis A, Malik F, Henry NL, Ithimakin S, et al. Activation of an IL6 inflammatory loop mediates trastuzumab resistance in HER2+ breast cancer by expanding the cancer stem cell population. *Mol Cell* (2012) 47(4):570–84. doi: 10.1016/j.molcel.2012.06.014
69. Burnett JP, Lim G, Li Y, Shah RB, Lim R, Paholak HJ, et al. Sulforaphane enhances the anticancer activity of taxanes against triple negative breast cancer by killing cancer stem cells. *Cancer Lett* (2017) 394:52–64. doi: 10.1016/j.canlet.2017.02.023
70. Cao N, Li S, Wang Z, Ahmed KM, Degnan ME, Fan M, et al. NF-kappaB-mediated HER2 overexpression in radiation-adaptive resistance. *Radiat Res* (2009) 171(1):9–21. doi: 10.1667/RR1472.1
71. Edwardson DW, Boudreau J, Mapletoff J, Lanner C, Kovala AT, Parissenti AM. Inflammatory cytokine production in tumor cells upon chemotherapy drug exposure or upon selection for drug resistance. *PLoS One* (2017) 12(9):e0183662. doi: 10.1371/journal.pone.0183662
72. Badve S, Nakshatri H. Breast-cancer stem cells-beyond semantics. *Lancet Oncol* (2012) 13(1):e43–8. doi: 10.1016/S1470-2045(11)70191-7
73. Colavito SA, Zou MR, Yan Q, Nguyen DX, Stern DF. Significance of glioma-associated oncogene homolog 1 (GLI1) expression in claudin-low breast cancer and

- crossstalk with the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kappaB) pathway. *Breast Cancer Res* (2014) 16(5):444. doi: 10.1186/s13058-014-0444-4
74. Johnson DE, O'Keefe RA, Grandis JR. Targeting the IL-6/JAK/STAT3 signalling axis in cancer. *Nat Rev Clin Oncol* (2018) 15(4):234–48. doi: 10.1038/nrclinonc.2018.8
75. Waugh DJ, Wilson C. The interleukin-8 pathway in cancer. *Clin Cancer Res* (2008) 14(21):6735–41. doi: 10.1158/1078-0432.CCR-07-4843
76. Zhen X, Choi HS, Kim JH, Kim SL, Liu R, Yun BS, et al. Machilin D, a lignin derived from *Saururus chinensis*, suppresses breast cancer stem cells and inhibits NF-kappaB signaling. *Biomolecules* (2020) 10(2):245. doi: 10.3390/biom10020245
77. Pitt JM, Marabelle A, Eggermont A, Soria JC, Kroemer G, Zitvogel L. Targeting the tumor microenvironment: removing obstruction to anticancer immune responses and immunotherapy. *Ann Oncol* (2016) 27(8):1482–92. doi: 10.1093/annonc/mdw168
78. Zahn LM. Effects of the tumor microenvironment. *Science* (2017) 355(6332):1386–8. doi: 10.1126/science.355.6332.1386-1
79. Joyce JA, Fearon DT. T cell exclusion, immune privilege, and the tumor microenvironment. *Science* (2015) 348(6230):74–80. doi: 10.1126/science.aaa6204
80. Chow A, Zhou W, Liu L, Fong MY, Champer J, Van Haute D, et al. Macrophage immunomodulation by breast cancer-derived exosomes requires Toll-like receptor 2-mediated activation of NF-kB. *Sci Rep* (2014) 4(1):5750. doi: 10.1038/srep05750
81. Korkaya H, Liu S, Wicha MS. Regulation of cancer stem cells by cytokine networks: attacking cancer's inflammatory roots. *Clin Cancer Res* (2011) 17(19):6125–9. doi: 10.1158/1078-0432.CCR-10-2743
82. Liu Z, Bandyopadhyay A, Nichols RW, Wang L, Hinck AP, Wang S, et al. Blockade of autocrine TGF-beta signaling inhibits stem cell phenotype, survival, and metastasis of murine breast cancer cells. *J Stem Cell Res Ther* (2012) 2(1):1–8. doi: 10.4172/2157-7633.1000116
83. Zhao Z, Hao D, Wang L, Li J, Meng Y, Li P, et al. CtBP promotes metastasis of breast cancer through repressing cholesterol and activating TGF-beta signaling. *Oncogene* (2019) 38(12):2076–91. doi: 10.1038/s41388-018-0570-z
84. Omene C, Bandera EV. Anti-VEGF therapy - a role in obesity-related breast cancer. *Nat Rev Endocrinol* (2018) 14(6):329–30. doi: 10.1038/s41574-018-0021-5
85. Sansone P, Storci G, Tavoroli S, Guarnieri T, Giovannini C, Taffurelli M, et al. IL-6 triggers Malignant features in mammospheres from human ductal breast carcinoma and normal mammary gland. *J Clin Invest* (2007) 117(12):3988–4002. doi: 10.1172/JCI32533
86. Sung B, Pandey MK, Nakajima Y, Nishida H, Konishi T, Chaturvedi MM, et al. Identification of a novel blocker of IkappaBalpha kinase activation that enhances apoptosis and inhibits proliferation and invasion by suppressing nuclear factor-kappaB. *Mol Cancer Ther* (2008) 7(1):191–201. doi: 10.1158/1535-7163.MCT-07-0406
87. Park BK, Zhang H, Zeng Q, Dai J, Keller ET, Giordano T, et al. NF-kappaB in breast cancer cells promotes osteolytic bone metastasis by inducing osteoclastogenesis via GM-CSF. *Nat Med* (2007) 13(1):62–9. doi: 10.1038/nm1519
88. Pereira FV, Melo AC, de Melo FM, Mourao-Sa D, Silva P, Berzaghi R, et al. TLR4-mediated immunomodulatory properties of the bacterial metalloprotease arazyme in preclinical tumor models. *Oncoimmunology* (2016) 5(7):e1178420. doi: 10.1080/2162402X.2016.1178420
89. Duhamel M, Rodet F, Delhem N, Vanden Abeele F, Kobeissy F, Nataf S, et al. Molecular consequences of proprotein convertase 1/3 (PC1/3) inhibition in macrophages for application to cancer immunotherapy: A proteomic study. *Mol Cell Proteomics* (2015) 14(11):2857–77. doi: 10.1074/mcp.M115.052480
90. Wang J, Dong B, Tan Y, Yu S, Bao YX. A study on the immunomodulation of polysaccharopeptide through the TLR4-TIRAP/MAL-MyD88 signaling pathway in PBMCs from breast cancer patients. *Immunopharmacol Immunotoxicol* (2013) 35(4):497–504. doi: 10.3109/08923973.2013.805764
91. Zhang W, Xiong Z, Wei T, Li Q, Tan Y, Ling L, et al. Nuclear factor 90 promotes angiogenesis by regulating HIF-1alpha/VEGF-A expression through the PI3K/Akt signaling pathway in human cervical cancer. *Cell Death Dis* (2018) 9(3):276. doi: 10.1038/s41419-018-0334-2
92. Peng G, Wang HY, Peng W, Kiniwa Y, Seo KH, Wang RF. Tumor-infiltrating gamma delta T cells suppress T and dendritic cell function via mechanisms controlled by a unique toll-like receptor signaling pathway. *Immunity* (2007) 27(2):334–48. doi: 10.1016/j.immuni.2007.05.020
93. Liu Y, Xiang X, Zhuang X, Zhang S, Liu C, Cheng Z, et al. Contribution of MyD88 to the tumor exosome-mediated induction of myeloid derived suppressor cells. *Am J Pathol* (2010) 176(5):2490–9. doi: 10.2353/ajpath.2010.090777
94. Hammond E, Haynes NM, Cullinane C, Brennan TV, Bampton D, Handley P, et al. Immunomodulatory activities of pixatimod: emerging nonclinical and clinical data, and its potential utility in combination with PD-1 inhibitors. *J Immunother Cancer* (2018) 6(1):54. doi: 10.1186/s40425-018-0363-5
95. Xu L, Zheng Y, Wang J, Xu Y, Xie Y, Yang ZP. IL33 activates CD8+T and NK cells through MyD88 pathway to suppress the lung cancer cell growth in mice. *Biotechnol Lett* (2020) 42(7):1113–21. doi: 10.1007/s10529-020-02815-2