Check for updates

OPEN ACCESS

EDITED BY Peng Liu, Institut National de la Santé et de la Recherche Médicale (INSERM). France

REVIEWED BY Lingxiang Jiang, Indiana University, United States Leina Ma, Qingdao University, China

*CORRESPONDENCE Gene Chatman Clark Clarkgc@vcu.edu

RECEIVED 13 October 2023 ACCEPTED 26 December 2023 PUBLISHED 26 February 2024

CITATION

Clark GC, Lai A, Agarwal A, Liu Z and Wang X-Y (2024) Biopterin metabolism and nitric oxide recoupling in cancer. *Front. Oncol.* 13:1321326. doi: 10.3389/fonc.2023.1321326

COPYRIGHT

© 2024 Clark, Lai, Agarwal, Liu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Biopterin metabolism and nitric oxide recoupling in cancer

Gene Chatman Clark^{1,2*}, Alan Lai², Aashri Agarwal³, Zheng Liu^{4,5} and Xiang-Yang Wang^{4,5,6}

¹Department of Biochemistry, Virginia Commonwealth University, Richmond, VA, United States, ²School of Medicine, Virginia Commonwealth University, Richmond, VA, United States, ³Cornell University, Ithaca, NY, United States, ⁴Department of Human Molecular Genetics, Virginia Commonwealth University, Richmond, VA, United States, ⁵Massey Cancer Center, Virginia Commonwealth University, Richmond, VA, United States, ⁶Institute of Molecular Medicine, Virginia Commonwealth University, Richmond, VA, United States

Tetrahydrobiopterin is a cofactor necessary for the activity of several enzymes, the most studied of which is nitric oxide synthase. The role of this cofactorenzyme relationship in vascular biology is well established. Recently, tetrahydrobiopterin metabolism has received increasing attention in the field of cancer immunology and immunotherapy due to its involvement in the cytotoxic T cell response. Past research has demonstrated that when the availability of BH4 is low, as it is in chronic inflammatory conditions and tumors, electron transfer in the active site of nitric oxide synthase becomes uncoupled from the oxidation of arginine. This results in the production of radical species that are capable of a direct attack on tetrahydrobiopterin, further depleting its local availability. This feedforward loop may act like a molecular switch, reinforcing low tetrahydrobiopterin levels leading to altered NO signaling, restrained immune effector activity, and perpetual vascular inflammation within the tumor microenvironment. In this review, we discuss the evidence for this underappreciated mechanism in different aspects of tumor progression and therapeutic responses. Furthermore, we discuss the preclinical evidence supporting a clinical role for tetrahydrobiopterin supplementation to enhance immunotherapy and radiotherapy for solid tumors and the potential safety concerns.

KEYWORDS

tetrahydrobiopterin, radiotherapy, nitric oxide synthase, immunotherapy, immunometabolism, vascular normalization, cancer, S-nitrosylation

Introduction

Nitric oxide synthase (NOS) exhibits two alternative activities depending on the relative availability of its cofactor tetrahydrobiopterin (BH4) (1). (Figure 1) In the presence of sufficient BH4, NOS catalyzes the production of NO and citrulline from the amino acid arginine. A necessary component of this process is the transfer of NADPH-derived

01



NOS Uncoupling and the Generation of Reactive Nitrogen Species (A) Coupled Nitric Oxide Synthase activity leads to signaling dominated by Snitrosylative reactions mediated by nitric oxide. (B) Uncoupled Nitric Oxide Synthase activity leads to signaling dominated by nitrosative reactions mediated by peroxynitrite.

electrons by BH4 to the ferrous dioxygen complex within the NOS active site (2, 3). When the availability of BH4 is low, as it is in chronic inflammatory conditions (4) and tumors (5), electron transfer in the active site becomes uncoupled from the oxidation of arginine, resulting in the production of a superoxide radical (6, 7). In the presence of trace amounts of NO, these two species react to form the short-lived, tyrosine-nitrating radical, peroxynitrite. This further reduces BH4 levels by direct oxidation (1, 8–10), producing a destructive feed-forward loop (11, 12). Due to this evolutionarily conserved (13) phenomenon of "NOS uncoupling," the restriction of this cofactor is reinforced upon depletion of local BH4 until NOS activity ceases.

Due to its capacity for antigen-specific cell killing and immune memory, the T lymphocyte is a major focus in studies on therapeutic anti-tumor immunity. T cells recognize peptides derived from intracellularly degraded proteins and loaded onto Major Histocompatibility Complex (MHC) proteins on the cell surface in a process commonly referred to as antigen presentation (14). Two broad classes of T cells exist and are necessary for antitumor immunity (15, 16). T cells that express the co-receptor CD4 recognize antigens loaded onto MHC-II proteins on the surfaces of APCs. These "helper" T cells organize the adaptive immune response to tumors by secreting pro-inflammatory cytokines and chemokines (16). T cells that carry the co-receptor CD8 on the other hand recognize peptides loaded onto MHC-I molecules on the surfaces of cancer cells and carry out cell killing by secreting cell toxins or presenting cancer cells with FAS Ligand (15). CD8⁺ T cells can also be activated by MHC-I⁺ dendritic cells in a process called cross-presentation, which is crucial for generation of an effective anti-tumor immune response (17). CD4 and CD8 T cells are necessary for both the responses of tumors to immunotherapy and for the immunological memory required for durable responses to anti-cancer therapy (18, 19). Recently, it has been reported that ambient BH4 availability is a rate limiting factor required for the growth and proliferation of T cells after T cell receptor (TCR) engagement (20). Furthermore, pharmacological augmentation of BH4 levels has been demonstrated to enhance T cell function in multiple models including anti-tumor immunity (20). This suggests that mechanisms that alter BH4 metabolism within the tumor microenvironment (TME), such as NOS uncoupling, may play have a significant impact on immune evasion and the resistance of tumors to immunotherapy (14).

Considering the inducible nature of NOS expression within the tissues of the body (21), NOS-mediated BH4 consumption may also, ostensibly, limit the availability of this cofactor for other BH4dependent enzymes such as alkylglycerol monooxygenase (AGMO) and the three aromatic amino acid hydroxylases: phenylalanine hydroxylase, tyrosine hydroxylase, and tryptophan hydroxylase. As reviewed elsewhere (22-24), these enzymes also make contributions to the immunosuppressive TME, making this an attractive area for further study (22-24). NOS uncoupling also has significant tissue level effects beyond the consumption of BH4. NO plays an essential role in the genesis and maturation of nascent vessels as well as the quiescence of mature vasculature (25, 26). As such, its absence in the TME contributes to the irregular structure and dysfunction that is common to tumor vasculature (27-30). This altered vasculature is both a hallmark of tumorigenesis in its own right and leads to therapeutic resistance through multiple mechanisms (31). Thus, exogenous BH4 supplementation may alter the characteristics of TME to improve traditional anti-cancer therapy (32), and also potentially make them more susceptible to immune attack (20, 33-35).

NOS can be effectively "recoupled" by supplementation with BH4 or its precursor sepiapterin. The therapeutic effect of NOS recoupling on endothelial dysfunction in hypertension, diabetes, and myocardial infarction is well recognized (36–40). However, the

therapeutic potential of NOS recoupling in the context of cancer has only just begun to be explored. It has been demonstrated that the BH4/BH2 ratio is low in multiple in vivo tumor models and patient biopsies (5, 27, 32). Reconstitution of coupled NOS signaling with sepiapterin resulted in vasculature normalization, alleviating tumor hypoxia and enhancing the efficacy of chemotherapy (5, 27, 32). Lastly, sepiapterin supplementation has the added benefit of enhancing cytotoxic T cell proliferation and activity in vivo (20). Considering the potential of BH4 supplementation to synergize with existing cancer therapies as well as the excellent and wellestablished safety of both sepiapterin and BH4 in humans (41-45), BH4 supplementation may become an important tool in the anticancer arsenal of the future. In this review, we set out to describe the relevance of local BH4 depletion to existing anti-cancer therapies such as radiotherapy and immune checkpoint blockade with a focus on molecular mechanisms.

NOS uncoupling and BH4 depletion

Biopterins are reductive, enzymatic cofactors required by organisms in all kingdoms of life (13). BH4 is the fully reduced form of biopterin in mammals and its availability is tightly regulated in normal tissues by elegant, cell-type specific control of its synthetic enzymes. Multiple mechanisms of NOS uncoupling and BH4 depletion within tumors have been reported. First, persistent generation of high levels of reactive oxygen and nitrogen species (ROS/RNS) is a hallmark of tumorigenesis (46). BH4 is highly reductive and therefore easily oxidized to its non-productive metabolite 7,8-dihydrobiopterin (BH2) by free radicals such as superoxide or peroxynitrite. Because these two biopterin species bind to the active site of NOS with equal affinities (47), the abundance of BH4 relative to BH2 is a crucial determinant of NOS activity. Additionally, exogenous producers of ROS/RNS, such as radiation, have been demonstrated to initiate NOS uncoupling, leading to reduced BH4 metabolism (48). Second, BH4 levels are regulated by transcriptional control of its synthetic enzymes. In chronic inflammation, BH4 production is limited due to the downregulation of GTP cyclohydrolase-I (49, 50) (GCH-I), the rate limiting enzyme in BH4 synthesis as well as by multiple signaling factors such as the immunosuppressive cytokines IL-10 and TGF- β (13). On the other hand, proinflammatory stimuli such as TNF α or TCR ligation stimulate the expression of GCH-I in their respective cell types (13). This dichotomous regulation of BH4 production by anti-inflammatory and proinflammatory signaling supports a central role of this cofactor in the immune response. Because once NOS uncoupling is initiated by low BH4 levels, its activity directly results in further ROS/RNS-mediated depletion of BH4, the effects of any one of these initial stimuli may be perpetuated until NOS activity ceases or is re-coupled to NO production. This feed-forward loop may represent a molecular switch, flipped off and on by anti-inflammatory and proinflammatory signals, that governs the local availability of BH4 and its impact on the TME and other pathological states (51).

NOS uncoupling is also accompanied by a consequential shift in NO-mediated signaling, which is relevant both to tumor

progression as well as the anti-tumor immune response. The first reported physiological role of NO was discovered when it was found to act as a secondary messenger that binds and activates soluble guanylate cyclase (sGC) in endothelial cells. However, in addition to the activation of sGC and its downstream effectors such as protein kinase G, NO also mediates signaling by oxidative S-nitrosylation of cysteine residues to form S-nitrosothiol groups within target proteins (52). This bona fide post-translational modification, functioning analogously to phosphorylation to activate or inactivate proteins by various physical mechanisms (52). Comparably to phosphorylation, this mechanism modulates proteins of all functional classes and is governed by enzymatic machinery that includes S-nitrosylases and denitrosylases, reflecting its long presence over the course of evolution (53-56). More on the functional role of this post-translational modification on the activation and inactivation of proteins relevant to the immune cascade is provided below.

As NOS becomes uncoupled, an accompanying shift from Snitrosylation to tyrosine nitration occurs and is the direct product of the changing ratio between ambient NO and superoxide produced by the enzyme. Despite its unpaired electron, NO itself is relatively non-reactive, reacting only with transition metals and heme iron. However, NO can also be "activated" via interactions with low levels of superoxide to form more reactive intermediates such as peroxynitrite (ONOO⁻), nitrogen dioxide (NO₂), and dinitrogen trioxide (N₂O₃), all of which are capable of interacting with nucleophiles like cysteine (57). Similar to NO, NO2 is lipophilic, migrating across cell membranes and S-nitrosylating proteins at a relative distance. ONOO⁻, on the other hand, is not lipophilic and reacts rapidly with CO₂ to form a CO₂-ONOO⁻ intermediate that quickly degrades into NO2 and N2O3 in the presence of high levels of NO. For this reason, when NO levels are higher than superoxide levels, as they are when NOS is coupled, cyclic-GMP cyclase, protein kinase G signaling, and nitrosative reactions predominate over the oxidative reactions caused by peroxynitrite (58). In contrast, when superoxide levels are higher than NO levels, as they are when NOS is uncoupled, oxidative reactions mediated by peroxynitrite predominate while NO signaling is inhibited due to its impaired production and rapid local consumption. Due to their charge and short half-life, the oxidative reactions mediated by peroxynitrite, such as tyrosine nitration, are relegated to the immediate area of superoxide production.

This dichotomy of RNS directed signaling has a profound impact on the character and biology of the TME. The baseline NO produced by coupled endothelial NOS plays an important role in maintaining vascular integrity via activation of endothelial sGC as well as the S-nitrosylation and silencing of inflammatory mediators such an NF- κ (59–61). The consequence of these effects is a quiescent and functional vasculature. When NOS is uncoupled, as it often is in the TME, peroxynitrite becomes the predominant RNS species produced. This metabolite exhibits a much different chemistry, preferring to react with immediately available tyrosine groups on nearby proteins, resulting in a relatively long-lived Tyr-nitration. The effect of Tyr-nitration is analogous to Tyr-phosphorylation and it's effects are different for different residues and proteins. Although this mechanism is less

10.3389/fonc.2023.1321326

well studied, peroxynitrite mediated Tyr-nitration and degradation of I $\kappa \alpha$ is an example, and coupled with the loss of NF- κ silencing Snitrosylation, has been suggested to lead to self-perpetuating and potentially unresolved vascular inflammation characteristic of tumors tissues (62, 63). In the setting of the aforementioned T cell dysfunction that comes with NOS mediated BH4 depletion, these effects of NOS uncoupling may greatly contribute to the protumorigenic character of the TME by facilitating immune escape, the development of dysfunctional tumor vasculature, and the propagation of tumor-promoting inflammation.

BH4 metabolism in tumorigenesis

BH4 and immune escape

Although BH4 is necessary for multiple cellular processes, the availability of this essential cofactor is a key determinant of immune cell activity (20, 34, 35). Of the cells of the immune system, the activity of the T lymphocyte is especially susceptible to restriction of BH4, making its control a crucial factor in the adaptive immune response. Since NOS activity is often associated with local BH4 availability, NOS uncoupling within tumor vasculature may represent a novel pathway involved in tumor immune escape.

BH4 availability has been shown to act as a rate-limiting factor for T cell proliferation and activity in diverse models of immunity, including within solid tumors (20, 64). This acceleration in T cell proliferation may occur due to increased iron-redox recycling of T cell mitochondrial cytochrome c which otherwise results in defective oxidative phosphorylation and superoxide production upon TCR stimulation (20, 35). It has been reported that expression of GCH1, the rate-limiting enzyme in BH4 production, is enhanced upon TCR stimulation and its inhibition antagonizes T cell proliferation (13). However, exogenous supplementation of BH4 has been demonstrated to enhance T cell activity in models of autoimmunity, suggesting that endogenous BH4 production by T cells does not support maximum activity and environmental BH4 is rate limiting (20). This supports the idea that the control of BH4 availability by NOS-expressing cells such as endothelial and myeloid cells may be a direct mechanism of impaired T cell activity both within and outside of the TME.

In this way, NOS uncoupling may have an indirect, albeit potent, effect on anti-tumor T cell activity by metabolizing BH4 to non-reducing metabolites. Therefore, supplementation of BH4 may counteract this effect both by supplying BH4 directly to proliferating T cells as well as recoupling NOS, alleviating further NOS-mediated BH4 consumption. Whether BH4 available for use by T cells is represented by free-floating, ambient BH4 or BH4 that is stored in the surrounding stroma has yet to be determined. However, it has been demonstrated that both BH4 production and sequestration occur in various cell types in response to proinflammatory cytokine stimulation (13).

In addition, much evidence exists that the restoration of NO signaling via recoupled NOS may support anti-tumor immunity as well. For instance, human T cells express NOS (65–69) and produce NO upon TCR activation, leading to enhanced activity of the TCR

CD3-zeta chain, the adaptor kinase ZAP-70, as well as N-Ras/ERK pathways mediated by N-nitrolsylation (70–72). Furthermore, endogenous levels of NO, such as those produced by eNOS, promote the differentiation of Th1 T cells via a cGMP dependent pathway (73), while NOS uncoupling and oxidative stress support Th2 differentiation (74, 75). Additionally, FOXP3⁺ regulatory T cells have also been reported to be negatively regulated in a cGMP dependent manner although it is unclear if this is a direct result of NOS activity (76, 77).

Interestingly, two other classic immunosuppressive mechanisms in the TME, Arginase 1 (Arg1) and Indoleamine 2'3-Dioxygenase 1 (IDO1), can also cause BH4 restriction indirectly via NOS uncoupling (11, 78). Arg1 produces arginase, which breaks down ambient arginine that is required for T cell activity and proliferation. The metabolism of arginine by arginase expressing cells such as M2 macrophages and MDSCs, is a well-recognized mechanism causing T functional impairment within the TME (79). Arginine is also the ultimate electron acceptor in the coupled NOS reaction and required for the production of NO. In the absence of arginine, NOS activity will proceed by donating electrons directly to molecular oxygen, generating free radicals (13). This initiates the uncoupled NOS cascade, leading to ambient BH4 depletion and further restriction of T cell activity. IDO1, which causes T cell restraint by restricting ambient tryptophan (79), may also induce NOS-mediated BH4 depletion via a secondary metabolite of tryptophan metabolism, xanthuric acid. This metabolite has an inhibitory effect on sepiapterin reductase (SR), a necessary enzyme in the BH4 salvage pathway that catalyzes the production of BH4 from BH2 (80). The enhanced concentration of BH2 competes with BH4 for binding to NOS, leading to uncoupling of NOS activity and BH4 depletion (48). Considering the evolutionarily conserved phenomenon of NOS uncoupling as well as the sophisticated mechanisms governing its activation, NOS uncoupling may be a deliberate and physiological mechanism of T cell restriction that is co-opted by solid tumors to achieve immune escape (Figure 2).

In addition to depleting available BH4, peroxynitrite produced by uncoupled NOS has been demonstrated to antagonize T cell activity via more direct effects (81-83). This leads to both the direct nitration and inactivation of the TCR as well as nitration and inhibition of the chemokines CCL2 and CCL5, leading to decreased T cell accumulation within tumors (84-86). Taken together, these reports suggest that endogenous NO expression may contribute to immune surveillance in normal tissues and that NOS uncoupling is a necessary step in the development of immune escape. Highlighting the direct antagonistic effect of NOS uncoupling on T cell activity, de Sanctis et al. found that inhibiting ARG1 and NOS2 expression inhibited accumulation of nitro-Tyrosine moieties in T cells and sensitized pancreatic tumor models to attack by TERT specific CAR-T (33). In addition to modulating the negative effects of NOS uncoupling, restoring native NO production and downstream signaling by activated T cells has also been proposed as an attractive therapeutic strategy (87).

Conversely, it should be noted that some studies suggest that very high levels of NO, such as those produced by NO donors or potentially iNOS, may actually negatively regulate Th1 differentiation required for anti-tumor immunity by leading to



the S-nitrosylation of key post-transcriptional regulators of Th1 cytokines and transcription factors (88–90). One report suggests that a FOXP3⁻ population of regulatory T cells may exist whose differentiation may be NO dependent (91). The discrepancy of these effects of NO on T cell phenotype may be explained by the time and dose dependence reported to characterize this pathway. Given the potential of BH4 supplementation to create high levels of NO via forced coupling of iNOS by immune cells within TME, their effects on immune functions need to further investigated.

BH4 and tumor vasculature

One hallmark of tumorigenesis is the acquired ability of tumors to recruit and maintain their own blood supply. Many excellent reviews of our understanding of the mechanisms of tumor vascular recruitment exist (31, 92, 93) and these mechanisms will not be extensively reviewed here. Compared to vasculature elsewhere in the body, tumor vasculature is typically erratic and highly dysfunctional, characterized by blunt ends and inconsistent blood flow, resulting in insufficient tumor perfusion and high interstitial pressure (94–97). These characteristics lead to the poor penetration of chemotherapy (98–100) and can cause intra-tumoral hypoxia and an accumulation of acid (95, 101–104) which in turn antagonizes anti-tumor immunity by stimulating immune checkpoint expression (105, 106) and immunosuppressive myeloid cells. Activity of cytotoxic lymphocytes in the tumor may also be antagonized due to the abnormal nature of tumor vasculature (107–109). As a result, the potential of vasculature normalizing agents to alter the immunosuppressive TME is the subject of considerable ongoing research.

Coupled eNOS activity is a key mediator of vascular function throughout the body. eNOS mediated NO gradients are necessary for the recruitment of pericytes and smooth muscle cells required for vascular integrity (29). NO is also necessary for the migration of endothelial cells, the formation of new capillary structures (26, 110) such as tertiary lymphoid tissues, which are important for the proliferation of anti-tumor lymphocytes, and for the maturation of normal vasculature through the expression of Angiotensin 1 (25, 111–113). Finally, constitutive, physiologic levels of NO provided by eNOS is an important part of maintaining vascular quiescence. Underscoring the importance of NO signaling to vascular function, NOS recoupling with BH4 or sepiapterin has been demonstrated to stimulate neoangiogenesis and normalize the reactivity of dysfunctional endothelium in cardiovascular disease (38, 39, 114, 115).

The positive role of NO in regulating angiogenesis raises the question of whether recoupling NOS and restoring NO signaling

has a beneficial effect on tumor biology. Within the TME, where NOS is uncoupled, NO signaling is absent and vascular inflammation is exacerbated by the effects of peroxynitrite. This leads to endothelial dysfunction, which is typical of tumor vasculature, and may partly explain the irregular structure and leakiness of the TME. Recoupling of endothelial NOS using oral sepiapterin leads to the normalization of tumor vasculature as demonstrated by the enhanced oxygenation of tumors, decreased hypoxia, and increase in pericyte coverage of tumor blood vessels (32). This is likely due to the inhibition of ROS/RNS (29) as well as the vasculature normalizing effect of physiological levels of NO (29, 116-118) enhancing tumor oxygenation and blood flow, thereby contributing to an enhanced efficacy of both radiotherapy and treatment with doxorubicin. Considering their potential synergy with immunotherapy, it is important to note that strategies that aim to restore physiological NO signaling by recoupling NOS appear to lead to vascular normalization as opposed to vascular disruption (27). This is contrary to the NOS inhibitor L-N^G-nitroarginine which disrupts tumor vasculature causing hypoxia and tumor cell death (119-121). For interventions that rely on enhanced immunogenicity of the TME via vasculature modulation, normalization is more likely than vascular disruption to alleviate hypoxic immunosuppression. This is consistent with multiple studies demonstrating that the efficacy of immunotherapy could be enhanced by treatment with combination with anti-angiogenic agents at low doses, but that the use of high dose anti-angiogenic therapy antagonized immunotherapy due to the hypoxia induced by excessive vessel pruning (94, 96, 97).

BH4 and tumor-promoting inflammation

Chronic inflammation is a hallmark of the TME, promoting tumor growth and invasion while paradoxically providing the means for escape from the adaptive immune response (122). While appropriate, physiological inflammation is an essential component of an anti-tumor immune response, the inappropriately prolonged, disordered inflammation associated with the TME actually serves as a barrier to T cell function and immune surveillance (123). It is partially for this reason that initiation of as many as a fifth of human cancers is linked to inflammation caused by chronic infection or exposure to irritants (124) as illustrated by the chemo-preventative effects of daily antiinflammatory drug use on certain types of cancer.

As discussed above, coupled NOS activity plays a major role in maintaining vascular quiescence. This is due to the pleiotropic anti-inflammatory effects of NO-mediated signaling on the cells of the vasculature itself. Possibly the most important pathway regulated by NO is that of the master inflammatory transcription factor NF- κ B, (Figure 3) which is susceptible to silencing by S-nitrosylation. NO-dependent S-nitrosylation of the p65 subunit inhibits the transcriptional activity of NF-kB in multiple cell types (59-61). The p50 subunit is also nitrosylated at a cysteine located in its DNA binding domain, resulting in an inhibition of inflammation (61). IKKB has also been demonstrated to be negatively regulated by S-nitrosylation mediated by NO, resulting in the inhibition of NF- κ B activity (125). In addition to the loss of these silencing mechanisms, uncoupled NOS may directly activate this pathway via the Tyr-nitration and proteosome independent inhibition of IkBa (126). Whether this additional positive regulation of the NF- κ B pathway occurs with NOS uncoupling in vivo remains to be seen.

Other mediators of inflammation are directly inhibited by NO mediated S-nitrosylation as well. CD40 is a member of the TNF receptor family, responsible for activating NF- κ B in response to stimulation with CD40L in what has been called the non-canonical NF- κ B pathway. S-nitrosylation of CD40 on the surface of resting monocytes and macrophages has been observed to inhibit signal transduction in response to CD40 activation, suggesting that denitrosylation may be a prerequisite for activation of this pathway (127). S-nitrosylation of S100A8 has also been suggested to be an important step in the resolution of inflammation by



limiting the endothelial cell-lymphocyte interactions in circulation (128). AP-1 is a transcription factor responsible for cytokine production and proliferation in response to multiple stimuli. It is activated by a MAPK cascade resulting in the activation of the kinase c-Jun N-terminal Kinase (JNK) which phosphorylates and activates c-Jun, one of its components. JNK1 is susceptible to NO mediated S-nitrosylation at Cys-119, further contributing to the anti-inflammatory effect of NO (129, 130). Finally, iNOS itself is inhibited by S-nitrosylation (131, 132), suggesting that NOS uncoupling may be further perpetuated by an increase in iNOS expression under conditions that support superoxide production and BH4 depletion supporting the idea that NOS uncoupling may function as a molecular switch.

NO has also been recognized for its inflammation suppressing effects in quiescent tissues by inhibiting leukocyte adhesion to the vascular wall, preventing the extravasation of immune cells into the underlying tissue (133-135). More recently, it has been suggested that this may be due in part to the suppression of P-selectin expression in the vascular endothelium mediated by constitutive, eNOS dependent sGC activation (136). Previous reports demonstrated that inactivation of sGC is a critical step in the recruitment of myeloid cells to the colon after DSS treatment. It was found by our group that the recruitment of inflammatory macrophages and neutrophils to the colon could be inhibited by treatment with sepiapterin, an effect that was partially inhibited by a cGC inhibitor (4). The consequence of therapeutically augmenting BH4 levels within tumors was the resolution of NF- κ B driven inflammation. This resulted in the inhibition of tumor formation in a spontaneous colon cancer model (4, 5). This finding is relevant to this tumor type for which inflammation plays such a wellrecognized role in progression as well as due to previous findings that suggest BH4 levels are low in human colorectal tumor biopsies when compared to normal tissue (32). All of these are important costimulators of T-cell mediated anti-tumor immunity and extravasation. However, the inappropriate, persistent activity of these inflammatory mediators leads to T cell dysfunction due to



BH4 supplementation has been demonstrated to affect three majo hallmarks of tumorigenesis pertinent to the response to immunotherapy: The normalization of tumor vasculature, T cell proliferation, and the resolution of malignant inflammation. the effects of long-term inflammation characteristic of the TME. The various mechanisms of this effects are expertly reviewed elsewhere (123, 124, 137, 138).

The Effects of BH4 supplementation on Cancer Therapies

Immunotherapy

Immune checkpoint blockade (ICB) has become the standard of care for multiple cancer types and members of this family of drugs have been approved for use as monotherapies for more than a dozen malignancies (101, 139, 140). However, compared to hematological malignancies (102), only a small fraction of patients with solid tumors respond to ICB monotherapy while as many as 87% of patients derive no clinical benefit (102, 141). It is believed that the TME stands as a major impediment to the effector function of CTLs, limiting the efficacy of therapies that rely on their activity. This has led to development of combination strategies designed to alter the TME, making tumors more susceptible to immune attack (123, 142-144). BH4 supplementation represent a novel approach to enhancing immunotherapy both by its direct proliferative effects on T cells outlined above and by its impact on multiple aspects of the TME including its effects on vascular normalization and the resolution of malignant inflammation (Figure 4) (92) As monotherapies, anti-angiogenic agents have also failed to yield clinical benefits, due in part to both the toxic effects that these agents have on normal tissues as well as the rapidity with which tumors develop resistance (102, 145). On the other hand, multiple anti-angiogenesis and immunotherapy combinations have achieved significant clinical results (146-151), demonstrating the potential of this treatment approach. Combination therapy has been shown to sensitize tumors to ICB by enhancing T cell penetration, supporting the formation of high endothelial venules, reducing T cell exhaustion, suppressing immunosuppressive cells types, and increasing the recruitment of activated antigen presenting cells (APCs) to the tumor (144). This illustrates the role that the dysfunctional tumor vasculature has on immune escape. However, combination with existing anti-angiogenic faces significant hurdles. For instance, the efficacy of combination relies on using strategies that normalize tumor vasculature and avoids detrimental vascular pruning as described above (27). That makes combination therapy a delicate balance, which may be difficult to achieve reliably in the clinic. In addition (144), most antiangiogenic strategies are complicated by toxic sequelae due to their effects on normal tissues. This includes hypertension, proteinuria, impaired wound healing, and dose limiting cardiotoxicity (152-156). Finally, most anti-angiogenesis and vasculature normalizing strategies are only effective for a brief window of time (157). This is due to the rapid development of resistance to agents that target angiogenic pathways and represents a major obstacle to their clinical effectiveness.

For these reasons BH4 supplementation maybe be a more robust and reliable agent for combination therapy. Firstly (32, 158) the vascular normalizing function of sepiapterin does not

rely on antagonizing angiogenic pathways but instead causes signaling within tumor vasculature to better resemble that of vasculature elsewhere in the body, as detailed above. Therefore, the risk of inducing vascular pruning as opposed to normalization is minimal and it likewise may not be as susceptible to the same mechanisms of therapeutic resistance as other vasculature normalizing strategies. Additionally, we have been able to demonstrate that enhanced tumor oxygenation caused by oral treatment with sepiapterin lasts for a significant amount of time after treatment has ceased (32), possibly reflecting its different mechanism of action. In addition, it demonstrates none of the toxic effects seen with other agents and has minimal detectable impact on normal tissues where BH4/ BH2 levels are tightly regulated (158). This significantly widens the therapeutic window of BH4 supplementation compared to other agents.

Somewhat paradoxically, the quality of the unresolving, malignant inflammation typical of TME has long been recognized as a hallmark of tumorigenesis that aids in immune escape. As such, many strategies have been attempted to make tumors more susceptible to immunotherapy by limiting this inflammation (123, 159). Despite compelling pre-clinical reports, clinical trials aimed at inhibiting chronic inflammation to enhance therapeutic outcomes of immunotherapy have yet to yield significant results (123). Among other things, this may reflect the necessity of physiological inflammation for the function of immune cells in the TME. Similar to other anti-inflammatory/immunotherapy combination strategies, the resolution of tumor-supporting, malignant inflammation caused by uncoupled NOS may leave tumors vulnerable to more appropriate, T cell mediated responses. However, as with all such strategies timing and dosage need careful consideration, as immunotherapy relies on inflammation driven penetration and activation of both myeloid cells and lymphocytes into tumors. Interventions that limit myeloid cell activation in the TME or immune cell trafficking to the tumor tissue may limit immunotherapy when not given concomitantly with drivers of appropriate inflammation such as ICB or PRR agonists.

Radiotherapy and the optimization of radio-immunotherapy

The majority of cancer patients with solid tumors will receive some form of radiotherapy (RT) during their course of treatment. However, treatment is limited by the toxic effects of off-target radiation on normal tissues. Much research in the field of radiation oncology is focused on limiting these off-target effects or enhancing the efficacy of RT in order to widen its therapeutic window. Numerous studies have demonstrated that NOS can be activated or upregulated by cancer cells in response to exposure to ionizing radiation (160, 161). However, it has also been demonstrated that oxidative stress caused by ionizing radiation may reduce BH4 availability resulting in a shift in BH4/BH2 ratios at both systemic and local levels (48, 162), resulting in uncoupled NOS activity (48). Suggested mechanisms for this phenomenon include the direct oxidation of BH4 by radiation-stimulated ROS production and the upregulation of the negative regulator of GCH1, GTP cyclohydrolase I feedback regulator (GFRP) (163, 164), although other pathways have been implicated (48). It is still unknown which pathway is dominant within the irradiated TME. As a result, multiple investigations are currently studying the role of NOS uncoupling in the response of tumors and normal tissues to RT as well as the therapeutic potential of BH4 supplementation.

The bulk of irradiated cancer cells die due to mitotic crisis, an ill-defined form of necrotic cell death caused by continued cell cycling in the presence of unrepaired DNA double-strand breaks (DSBs). These breaks are caused by either direct attack by highenergy photons or, much more commonly, by reactive oxygen (ROS) released from mitochondria within irradiated cells (165). Because the availability of ambient oxygen is necessary for the majority of DNA damage produced by radiation in the X-ray range, tumor hypoxia reduces the effectiveness of the radiation (157). Multiple studies have demonstrated that the degree of hypoxia directly correlates with reduced radiosensitivity by limiting the oxygen available for the production of ROS (166-169). Due to its ability to normalize tumor vasculature via NOS recoupling, BH4 supplementation has been demonstrated to enhance RT induced cell killing by increasing perfusion and oxygen delivery to tumor cells (32, 170). It is important to note that BH4 itself has been shown to act as a free radical scavenger and inhibitor of apoptosis in vitro, which may directly limit RT cell killing (48). In addition, both positive and negative effects of NO on apoptosis have been reported to function through various mechanisms in vitro as well (171). However, these may not be salient features of BH4 supplementation in vivo as they appear to be overshadowed by the positive effect generated by alleviating tumor hypoxia.

The total dose of RT given to a singular disease site is often limited due to the need to avoid lethal fibrotic side effects in soft tissues such as the heart and lungs. Radiation-induced cardiomyopathy is a dose-dependent, progressive disease characterized by myocardial and pericardial fibrosis, loss of contractile reserve and LV systolic function, and ultimately, premature death (172-175). Radiotherapy-induced lung fibrosis is a fibrotic disease resembling idiopathic pulmonary fibrosis and is one of the most common side effects of thoracic RT. Unfortunately, options for limiting radiation off-target effects or treating exposure are limited (48). The mechanisms behind radiation-induced tissue injury have been reviewed by other authors and will not be deeply examined here (176). However, multiple studies demonstrate that radiation-induced injury to the heart and lungs is mediated in part by chronic inflammatory signaling maintained by persistent endothelial dysfunction (175-181). Radiation-induced oxidative stress has been demonstrated to induce endothelial dysfunction by reducing BH4 levels leading to the uncoupling of NOS in both the heart and lungs (162, 163, 182). Multiple groups have reported that BH4 supplementation has a radioprotective effect on normal tissues through reversing the effects of uncoupling, restoring NO levels, mitigating inflammation, and acting as a free radical scavenger (48, 183). Three clinical trials are currently underway to study the effects of BH4 supplementation on radiation-induced skin injury and enteritis (NCT05299203, NCT05114226,

NCT05138887). The results of these studies may open up a new avenue for the prevention of radiation-induced late effects and their life-threatening fibrotic sequelae, potentially allowing for more aggressive, sub-fractionated therapy without the fear of soft tissue fibrotic effects.

The response rates of solid tumors to immunotherapeutic strategies that have been effective for hematologic malignancies have been disappointing (142). It has been hypothesized that the cause of this discrepancy may lie in the ability of tumor-associated stroma to suppress the effector functions of anti-tumor lymphocytes. One proposed approach to overcoming this immunosuppressive barrier is combination with RT, which is thought to have multiple immunostimulatory effects (142, 165, 184-186). In brief, RT expands the repertoire of tumor-associated antigens (TAAs) available in the TME while directly stimulating type I IFN expression by activating cyclic-GAMP synthase (cGAS) and its downstream signaling partner stimulator of interferon genes (STING). This leads to the necessary cross-presentation of TAAs by newly recruited APCs and subsequent priming of tumor-specific CTLs. It is hoped that the addition of strategies that revitalize T cell function, such as ICB, may have synergistic effects to achieve tumor regression in patients with advanced disease. However, clinical trials exploring the efficacy of a radio-immunotherapy combination have so far failed to yield significant positive results.

Considering the negative effect that RT has on BH4 availability, RT-induced BH4 depletion may be an overlooked restraint to the immune response to irradiated tumors. BH4 supplementation may be exploited to potentiate the combination therapy for at meaningful clinical impact. First, systemic BH4 supplementation may enhance the proliferation of lymphocytes stimulated by APCs in the lymph nodes in response to RT (Figure 5). By enhancing the expansion of tumor-reactive lymphocytes, BH4 may provide for a more robust response to subsequent immune augmentation with ICB. Second, it has been established by multiple groups that durable anti-tumor immune responses to immunotherapy require intra-tumoral proliferation of T cells (187), a process that may be directly inhibited by RT-induced NOS uncoupling and local BH4 depletion. BH4 supplementation is likely to alleviate this immune-inhibitory process, leading to a more robust immune effector response within the irradiated TME.

RT also induces other changes to the TME that are relevant to the discussion of NOS uncoupling and BH4 metabolism. For instance, within a certain dose range, RT has a transient, type I IFN-dependent, normalizing effect on the tumor vasculature. It is thought that this may aid immune cell infiltration into the tumor and synergize with other vasculature normalizing strategies such as BH4 supplementation and immune checkpoint blockade (188). However, with so many vascular modulators at play, this threepart interaction will have to be carefully studied for potential synergy as it may tip the scales from immunostimulatory vascular normalization to immunosuppressive vascular regression and hypoxia. The immunostimulatory effects of RT have also been attributed its ability to recruit lymphocytes and myeloid cells. Many of these cells are likely to carry active iNOS. However, further study is required to understand how forced coupling of this influx of NOS by BH4 saturation may influence the irradiated TME. Finally, the advent of stereotactic body RT has made the immune-mediated effects of RT more clinically relevant by



FIGURE 5

BH4 supplementation and Radiotherapy-Immunotherapy combination. 1) Radiotherapy stimulates an influx of lymphocytes and myeloid cells from the blood stream while simultaneously depleting BH4 in the TME. This depletion represents a potential barrier to ICB response in irradiated tumors. 2) Exogenous supplementation of BH4 leads to normalization of BH4 levels within the tumor, allowing for the intra-tumoral T cell proliferation necessary for the response to ICBs. 3) APCs recruited to the tumor acquire TAAs and migrate to the draining lymph nodes where they present antigen to naïve T cells. 4) Exogenous BH4 supplementation enhances tumor specific T cell expansion both enhancing the number of effector T cells available to attack the tumor as well as to be retained in the form of memory T cells.



allowing the use of more immunogenic doses of radiation (8-15 Gy per fraction) (143). The radioprotective effect of BH4 supplementation on normal tissue may also aid in the use of these more immunogenic doses, leading to better synergy with immunotherapy. How these effects are incorporated into clinical practice will be exciting to see moving forward.

Safety of BH4 supplementation

BH4 supplementation, either with BH4 itself or with its precursor sepiapterin, has a well-established safety record in humans. Both have undergone numerous clinical trials and appear to cause few side effects, even at very high levels (41-45). BH4 is currently indicated in the long-term setting for certain types of phenylketouria (189). This is contrary to other immunomodulators, such as pattern recognition agonists, which cause global immune activation and may have a narrow therapeutic window (190). This superior safety profile is likely a result of BH4s water solubility, its ease of excretion, its highly regulated concentrations in normal tissues (158), and the restricted nature of its positive effects on proliferating T cells (35). T cell proliferation and cytokine production are usually coupled upon activation. Interventions such as pegylated IFN-a, PRR agonism, or ICBs typically enhance both the number and activation state of T cells. However, these therapies enhance T cell activity indiscriminately, putting patients at risk for off target effects and severe reactions caused by T cell cytokine production. These risks pose a major obstacle to the augmentation of cancer immunity in clinical trials. Due to its purely proliferative effect, BH4 supplementation enhances T activity only in those T cells that have already been stimulated by their cognate antigen, without effecting global T cell activation and IFN-y secretion (20). This makes it a much safer immunological option.

However, therapy with ICBs does carry the risk of life threatening immune-related adverse events (irADs) (102). In addition, it has previously been reported in preclinical models that forced overproduction of BH4 exacerbates certain T cell mediated autoimmune disease in mice (20). While it is true that immunological side effects have not been reported with long term BH4 treatment alone, combination therapy with ICB should be considered carefully for the risk of exacerbating the established risk for irADs. However, these events are clinically manageable, and we expect that existing interventions for irADs, namely corticosteroid treatment and ceasing therapy with the offending agent, would be effective in mitigating significant risk for irADs. Finally, due to its anti-inflammatory effect on myeloid cells, which are necessary for anti-tumor immune responses, prolonged BH4 supplementation as a single agent runs the theoretical risk of suppressing anti-tumor immunity, potentially allowing for tumor advancement, and making distant peripheral sites more hospitable to metastases. Although this currently remains a theoretical risk, care should be taken in trials not to expose patients with documented tumors to prolonged BH4 treatment alone.

Conclusions and future directions

In conclusion, the propensity of low intratumoral BH4 levels to affect chronic inflammation, tumor-associated vascular

dysfunction, and proliferation of CTLs contributes significantly to the tumor supportive characteristics of the TME. Here, we have provided pre-clinical evidence that supports exploring the clinical utility of BH4 supplementation to enhance existing anti-cancer therapies such as RT and immunotherapy. Whether BH4 restriction represents a bona fide, physiological mechanism of immune evasion or a pathological consequence of the disordered TME remains to be determined. Multiple strategies for enhancing BH4 production and recycling within cells exist beyond direct supplementation. One of these, supplementation with the BH4 precursor sepiapterin, is especially promising as it can be taken orally and can penetrate cells passively due to its relative hydrophobicity compared to BH4. This also has implications for tumors of the central nervous system as this form of biopterin can more easily pass through the bloodbrain barrier (44). Therefore, BH4 supplementation may one day offer a new therapeutic avenue to oncologists and their patients (Figure 6).

Author contributions

GC: Conceptualization, Writing – original draft, Writing – review & editing. AL: Writing – original draft, Writing – review & editing. AA: Writing – original draft, Writing – review & editing. ZL: Supervision, Writing – original draft, Writing – review & editing. X-YW: Supervision, Writing – original draft, Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. The work was supported in part by Ruth L. Kirschstein Individual Predoctoral NRSA for MD/PhD and other Dual Degree Fellowships (1F30CA239564), National Institute of Health grants (CA175033, AI133595), VCU Massey Cancer Center Research Development Funds, and National Cancer Institute Cancer Center Support Grant to VCU Massey Cancer Center P30CA16059.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Cai S, Khoo J, Channon KM. Augmented BH4 by gene transfer restores nitric oxide synthase function in hyperglycemic human endothelial cells. *Cardiovasc Res* (2005) 65:823–31. doi: 10.1016/j.cardiores.2004.10.040

2. Wever RMF, Van Dam T, Van Rijn HJM, De Groot F, Rabelink TJ. Tetrahydrobiopterin regulates superoxide and nitric oxide generation by recombinant endothelial nitric oxide synthase. *Biochem Biophys Res Commun* (1997) 233:150-3. doi: 10.1006/bbrc.1997.7069

3. Schmidt HHHW, Hofmann H, Schindler U, Shutenko ZS, Cunningham DD, Feelisch M. No NO from NO synthase. *Proc Natl Acad Sci USA* (1996) 93:14492–7. doi: 10.1073/pnas.93.25.14492

4. Cardnell RJG, Rabender CS, Ross GR, Guo C, Howlett EL, Alam A, et al. Sepiapterin ameliorates chemically induced murine colitis and azoxymethane-induced colon cancer. *J Pharmacol Exp Ther* (2013) 347:117–25. doi: 10.1124/jpet.113.203828

5. Rabender CS, Alam A, Sundaresan G, Cardnell RJ, Yakovlev VA, Mukhopadhyay ND, et al. The role of nitric oxide synthase uncoupling in tumor progression. *Mol Cancer Res* (2015) 13:1034–43. doi: 10.1158/1541-7786.MCR-15-0057-T

6. Vásquez-Vivar J, Kalyanaraman B, Martásek P, Hogg N, Masters BSS, Karoui H, et al. Superoxide generation by endothelial nitric oxide synthase: The influence of cofactors. *Proc Natl Acad Sci USA* (1998) 95:9220–5. doi: 10.1073/pnas.95.16.9220

7. Vśquez-Vivar J, Kalyanaraman B, Martásek P. The role of tetrahydrobiopterin in superoxide generation from eNOS: Enzymology and physiological implications. *Free Radic Res* (2003) 37:121–7. doi: 10.1080/1071576021000040655

8. Stuehr D, Pou S, Rosen GM. Oxygen reduction by nitric-oxide synthases. J Biol Chem (2001) 276:14533-6. doi: 10.1074/jbc.R100011200

 Mikkelsen RB, Wardman P. Biological chemistry of reactive oxygen and nitrogen and radiation-induced signal transduction mechanisms. *Oncogene* (2003) 22:5734–54. doi: 10.1038/sj.onc.1206663

10. Alp NJ, Channon KM. Regulation of endothelial nitric oxide synthase by tetrahydrobiopterin in vascular disease. *Arterioscler Thromb Vasc Biol* (2004) 24:413–20. doi: 10.1161/01.ATV.0000110785.96039.f6

11. Alkaitis MS, Crabtree MJ. Recoupling the cardiac nitric oxide synthases: Tetrahydrobiopterin synthesis and recycling. *Curr Heart Fail Rep* (2012) 9:200–10. doi: 10.1007/s11897-012-0097-5

12. Kuzkaya N, Weissmann N, Harrison DG, Dikalov S. Interactions of peroxynitrite, tetrahydrobiopterin, ascorbic acid, and thiols: Implications for uncoupling endothelial nitric-oxide synthase. *J Biol Chem* (2003) 278:22546–54. doi: 10.1074/jbc.M302227200

13. Werner-Felmayer G, Golderer G, Werner E. Tetrahydrobiopterin biosynthesis, utilization and pharmacological effects. *Curr Drug Metab* (2005) 3:159–73. doi: 10.2174/1389200024605073

14. Wculek SK, Cueto FJ, Mujal AM, Melero I, Krummel MF, Sancho D. Dendritic cells in cancer immunology and immunotherapy. *Nat Rev Immunol* (2020) 20:7–24. doi: 10.1038/s41577-019-0210-z

15. Chow A, Perica K, Klebanoff CA, Wolchok JD. Clinical implications of t cell exhaustion for cancer immunotherapy. *Nat Rev Clin Oncol* (2022) 19:775–90. doi: 10.1038/s41571-022-00689-z

16. Zuazo M, Arasanz H, Bocanegra A, Fernandez G, Chocarro L, Vera R, et al. Systemic CD4 immunity as a key contributor to PD-L1/PD-1 blockade immunotherapy efficacy. *Front Immunol* (2020) 11:586907. doi: 10.3389/fimmu.2020.586907

17. Joffre OP, Segura E, Savina A, Amigorena S. Cross-presentation by dendritic cells. *Nat Rev Immunol* (2012) 12:557–69. doi: 10.1038/nri3254

18. Tay RE, Richardson EK, Toh HC. Revisiting the role of CD4+ t cells in cancer immunotherapy-new insights into old paradigms. *Cancer Gene Ther* (2021) 28:5–17. doi: 10.1038/s41417-020-0183-x

19. Raskov H, Orhan A, Christensen JP, Gögenur I. Cytotoxic CD8+ t cells in cancer and cancer immunotherapy. *Br J Cancer* (2021) 124:359–67. doi: 10.1038/s41416-020-01048-4

20. Cronin SJF, Seehus C, Weidinger A, Talbot S, Reissig S, Seifert M, et al. The metabolite BH4 controls t cell proliferation in autoimmunity and cancer. *Nature* (2018). doi: 10.1038/s41586-018-0701-2

21. Bogdan C. Nitric oxide synthase in innate and adaptive immunity: An update. *Trends Immunol* (2015) 36:161–78. doi: 10.1016/j.it.2015.01.003

22. Sailer S, Keller MA, Werner ER, Watschinger K. The emerging physiological role of agmo 10 years after its gene identification. *Life* (2021) 11:88. doi: 10.3390/life11020088

23. Platten M, Nollen EAA, Röhrig UF, Fallarino F, Opitz CA. Tryptophan metabolism as a common therapeutic target in cancer, neurodegeneration and beyond. *Nat Rev Drug Discovery* (2019) 18:379–401. doi: 10.1038/s41573-019-0016-5

24. Gargaro M, Manni G, Scalisi G, Puccetti P, Fallarino F. Tryptophan metabolites at the crossroad of immune-cell interaction *via* the aryl hydrocarbon receptor: Implications for tumor immunotherapy. *Int J Mol Sci* (2021) 22:4644. doi: 10.3390/ ijms22094644

25. Benest AV, Stone OA, Miller WH, Glover CP, Uney JB, Baker AH, et al. Arteriolar genesis and angiogenesis induced by endothelial nitric oxide synthase overexpression results in a mature vasculature. *Arterioscler Thromb Vasc Biol* (2008) 28:1462–8. doi: 10.1161/ATVBAHA.108.169375

26. Sonveaux P, Brouet A, Havaux X, Grégoire V, Dessy C, Balligand JL, et al. Irradiation-induced angiogenesis through the up-regulation of the nitric oxide pathway: Implications for tumor radiotherapy. *Cancer Res* (2003) 63:1012–9. doi: 10.1016/s0167-8140(03)80572-8

27. Rabender C, Bruno N, Wang L, Zweit J, Mikkelsen R, Gobalakrishnan S. Optoacoustic imaging reveals normalized tumor oxygenation following sepiapterin treatment. *J Nucl Med* (2016) 57:1147.

28. Mikkelsen RB, Yakovlev VA, Rabender CS, Alam A. Nitric oxide synthase uncoupling in tumor progression and cancer therapy. *Strategies to Enhance Ther Ratio Radiat as Cancer Treat* (2016), 1034–43. doi: 10.1007/978-3-319-45594-5_6

29. Kashiwagi S, Tsukada K, Xu L, Miyazaki J, Kozin SV, Tyrrell JA, et al. Perivascular nitric oxide gradients normalize tumor vasculature. *Nat Med* (2008) 14:255–7. doi: 10.1038/nm1730

30. Babykutty S, Heishi T, Tsukada K, Huang Y, Kozin S, Conner D, et al. Restoring perivascular nitric oxide gradients normalizes breast cancer vasculature. *Nitric Oxide* (2014) 42:115. doi: 10.1016/j.niox.2014.09.051

31. Schaaf MB, Garg AD, Agostinis P. Defining the role of the tumor vasculature in antitumor immunity and immunotherapy article. *Cell Death Dis* (2018) 9 9(2):115. doi: 10.1038/s41419-017-0061-0

32. Rabender CS, Bruno N, Alam A, Sundaresan G, Zweit J, Mikkelsen RB. Sepiapterin enhances tumor radio- and chemosensitivities by promoting vascular normalization. *J Pharmacol Exp Ther* (2018) 365:536–43. doi: 10.1124/jpet.117.245258

33. De Sanctis F, Lamolinara A, Boschi F, Musiu C, Caligola S, Trovato R, et al. Interrupting the nitrosative stress fuels tumor-specific cytotoxic t lymphocytes in pancreatic cancer. *J Immunother Cancer* (2022) 10:e003549. doi: 10.1136/jitc-2021-003549

34. Cronin SJF, Penninger JM. From t-cell activation signals to signaling control of anti-cancer immunity. *Immunol Rev* (2007) 220:151-68. doi: 10.1111/j.1600-065X.2007.00570.x

35. Cronin SJF, Woolf CJ, Weiss G, Penninger JM. The role of iron regulation in immunometabolism and immune-related disease. *Front Mol Biosci* (2019) 22:116. doi: 10.3389/fmolb.2019.00116

36. Wang S, Xu J, Song P, Wu Y, Zhang J, Choi HC, et al. Acute inhibition of guanosine triphosphate cyclohydrolase 1 uncouples endothelial nitric oxide synthase and elevates blood pressure. *Hypertension* (2008) 52:484–90. doi: 10.1161/ HYPERTENSIONAHA.108.112094

37. Roe ND, He EY, Wu Z, Ren J. Folic acid reverses nitric oxide synthase uncoupling and prevents cardiac dysfunction in insulin resistance: Role of Ca2 +/calmodulin-activated protein kinase II. *Free Radic Biol Med* (2013) 65:234–43. doi: 10.1016/j.freeradbiomed.2013.06.042

38. Shimazu T, Otani H, Yoshioka K, Fujita M, Okazaki T, Iwasaka T. Sepiapterin enhances angiogenesis and functional recovery in mice after myocardial infarction. *Am J Physiol Heart Circ Physiol* (2011) 301:H2061–72. doi: 10.1152/ajpheart.00525.2011

39. Jo H, Otani H, Jo F, Shimazu T, Okazaki T, Yoshioka K, et al. Inhibition of nitric oxide synthase uncoupling by sepiapterin improves left ventricular function in streptozotocin-induced diabetic mice. *Clin Exp Pharmacol Physiol* (2011) 38:485–93. doi: 10.1111/j.1440-1681.2011.05535.x

40. Carnicer R, Duglan D, Ziberna K, Recalde A, Reilly S, Simon JN, et al. BH4 increases nNOS activity and preserves left ventricular function in diabetes. *Circ Res* (2021) 128:585–601. doi: 10.1161/CIRCRESAHA.120.316656

41. Abell T, Garcia LM, Wiener G, Wo JM, Bulat RS, Smith N. Su1719 IMPROVED GASTRIC ACCOMMODATION IN WOMEN WITH MODERATE OR SEVERE DIABETIC GASTROPARESIS RANDOMIZED TO ORAL CNSA-001 (SEPIAPTERIN) VERSUS PLACEBO. *Gastroenterology* (2020) 158:S1–S1603. doi: 10.1016/s0016-5085(20)32247-2

42. Abell TL, Garcia LM, Wiener GJ, Wo JM, Bulat RS, Smith N. Effect of oral CNSA-001 (sepiapterin, PTC923) on gastric accommodation in women with diabetic gastroparesis: A randomized, placebo-controlled, phase 2 trial. *J Diabetes Complications* (2021) 35. doi: 10.1016/j.jdiacomp.2021.107961

43. Bratkovic D, Margvelashvili I, Tchan MC, Nisbet J, Smith N. PTC923 (sepiapterin) lowers elevated blood phenylalanine in subjects with phenylketonuria: a phase 2 randomized, multi-center, three-period crossover, open-label, active controlled, all-comers study. *Metabolism* (2022) 128:155116. doi: 10.1016/j.metabol.2021.155116

44. Smith N, Longo N, Levert K, Hyland K, Blau N. Exploratory study of the effect of one week of orally administered CNSA-001 (sepiapterin) on CNS levels of tetrahydrobiopterin, dihydrobiopterin and monoamine neurotransmitter metabolites in healthy volunteers. *Mol Genet Metab Rep* (2019) 21:100500. doi: 10.1016/j.ymgmr.2019.100500

45. Smith N, Longo N, Levert K, Hyland K, Blau N. Phase i clinical evaluation of CNSA-001 (sepiapterin), a novel pharmacological treatment for phenylketonuria and tetrahydrobiopterin deficiencies, in healthy volunteers. *Mol Genet Metab* (2019) 126:406–12. doi: 10.1016/j.ymgme.2019.02.001

46. Panieri E, Santoro MM. Ros homeostasis and metabolism: A dangerous liason in cancer cells. *Cell Death Dis* (2016) 7:e2253. doi: 10.1038/cddis.2016.105

47. Crabtree MJ, Smith CL, Lam G, Goligorsky MS, Gross SS. Ratio of 5,6,7,8tetrahydrobiopterin to 7,8-dihydrobiopterin in endothelial cells determines glucoseelicited changes in NO vs. superoxide production by eNOS. *Am J Physiol Heart Circ Physiol* (2008) 294:H1530–40. doi: 10.1152/ajpheart.00823.2007

48. Feng Y, Feng Y, Gu L, Liu P, Cao J, Zhang S. The critical role of tetrahydrobiopterin (BH4) metabolism in modulating radiosensitivity: BH4/NOS axis as an angel or a devil. *Front Oncol* (2021) 11:720632. doi: 10.3389/ fonc.2021.720632

49. Zheng JS, Yang XQ, Lookingland KJ, Fink GD, Hesslinger C, Kapatos G, et al. Gene transfer of human guanosine 5'-triphosphate cyclohydrolase i restores vascular tetrahydrobiopterin level and endothelial function in low renin hypertension. *Circulation* (2003) 108:1238–45. doi: 10.1161/01.CIR.0000089082.40285.C3

50. Mitchell BM, Dorrance AM, Webb RC. GTP cyclohydrolase 1 downregulation contributes to glucocorticoid hypertension in rats. *Hypertension* (2003) 41:669–74. doi: 10.1161/01.HYP.0000051889.62249.5D

51. Fanet H, Capuron L, Castanon N, Calon F, Vancassel S. Tetrahydrobioterin (BH4) pathway: From metabolism to neuropsychiatry. *Curr Neuropharmacol* (2020) 19:591–609. doi: 10.2174/1570159x18666200729103529

52. Nathan JA. Nitrosylation rewires metabolism. Nat Chem Biol (2023) 19:253-4. doi: 10.1038/s41589-022-01169-2

53. Zhou HL, Zhang R, Anand P, Stomberski CT, Qian Z, Hausladen A, et al. Metabolic reprogramming by the s-nitroso-CoA reductase system protects against kidney injury. *Nature* (2019) 565:96–100. doi: 10.1038/s41586-018-0749-z

54. Seth D, Hess DT, Hausladen A, Wang L, Juan WY, Stamler JS. A multiplex enzymatic machinery for cellular protein s-nitrosylation. *Mol Cell* (2018). doi: 10.1016/j.molcel.2017.12.025

55. Stomberski CT, Hess DT, Stamler JS. Protein s-nitrosylation: Determinants of specificity and enzymatic regulation of s-Nitrosothiol-Based signaling. *Antioxid Redox Signal* (2019) 30:1331–51. doi: 10.1089/ars.2017.7403

56. Jia J, Arif A, Terenzi F, Willard B, Plow EF, Hazen SL, et al. Target-selective protein s-nitrosylation by sequence motif recognition. *Cell* (2014) 59:623-34. doi: 10.1016/j.cell.2014.09.032

57. Wink DA, Mitchell JB. Chemical biology of nitric oxide: Insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide. *Free Radic Biol Med* (1998) 25:434–56. doi: 10.1016/S0891-5849(98)00092-6

58. Jourd'heuil D, Miranda KM, Kim SM, Espey MG, Vodovotz Y, Laroux S, et al. The oxidative and nitrosative chemistry of the nitric oxide/superoxide reaction in the presence of bicarbonate. *Arch Biochem Biophys* (1999) 365:92–100. doi: 10.1006/abbi.1999.1143

59. Kelleher ZT, Matsumoto A, Stamler JS, Marshall HE. NOS2 regulation of NF-κB by s-nitrosylation of p65. J Biol Chem (2007) 282:30667–72. doi: 10.1074/jbc.M705929200

60. Marshall HE, Hess DT, Stamler JS. S-nitrosylation: Physiological regulation of NF-κB. Proc Natl Acad Sci USA (2004) 101:8841–2. doi: 10.1073/pnas.0403034101

61. Marshall HE, Stamler JS. Inhibition of NF-κB by s-nitrosylation. *Biochemistry* (2001) 40:1688–93. doi: 10.1021/bi002239y

62. Bayden AS, Yakovlev VA, Graves PR, Mikkelsen RB, Kellogg GE. Factors influencing protein tyrosine nitration-structure-based predictive models. *Free Radic Biol Med* (2011) 50:749–62. doi: 10.1016/j.freeradbiomed.2010.12.016

63. Yakovlev VA, Barani IJ, Rabender CS, Black SM, Leach JK, Graves PR, et al. Tyrosine nitration of $I\kappa B\alpha$: A novel mechanism for NF- κ B activation. *Biochemistry* (2007) 46:11671–83.

64. Schmitz K, Trautmann S, Hahnefeld L, Fischer C, Schreiber Y, Wilken-Schmitz A, et al. Sapropterin (BH4) aggravates autoimmune encephalomyelitis in mice. *Neurotherapeutics* (2021) 18:1862–79. doi: 10.1007/s13311-021-01043-4

65. Koh KP, Wang Y, Yi T, Shiao SL, Lorber MI, Sessa WC, et al. T cell-mediated vascular dysfunction of human allografts results from IFN- γ dysregulation of NO synthase. *J Clin Invest* (2004) 114:846–56. doi: 10.1172/JCI21767

66. Choy JC, Wang Y, Tellides G, Pober JS. Induction of inducible NO synthase in bystander human t cells increases allogenic response in the vasculature. *Proc Natl Acad Sci USA* (2007) 104:1313–8. doi: 10.1073/pnas.0607731104

67. Nagy G, Koncz A, Perl A. T cell activation-induced mitochondrial hyperpolarization is mediated by ca 2+ - and redox-dependent production of nitric oxide. *J Immunol* (2003) 171:5188–97. doi: 10.4049/jimmunol.171.10.5188

68. Reiling N, Kröncke R, Ulmer AJ, Gerdes J, Flad HD, Hauschildt S. Nitric oxide synthese: Expression of the endothelial, Ca2+/calmodulin-dependent isoform in human b and t lymphocytes. *Eur J Immunol* (1996). doi: 10.1002/eji.1830260302

69. Sciorati C, Rovere P, Ferrarini M, Heltai S, Manfredi AA, Clementi E. Autocrine nitric oxide modulates CD95-induced apoptosis in $\gamma\delta$ t lymphocytes. *J Biol Chem* (1997) 272:23211–5. doi: 10.1074/jbc.272.37.23211

70. Ibiza S, Víctor VM, Boscá I, Ortega A, Urzainqui A, O'Connor JE, et al. Endothelial nitric oxide synthase regulates t cell receptor signaling at the

immunological synapse. Immunity (2006) 24:753-65. doi: 10.1016/ j.immuni.2006.04.006

71. Ibiza S, Pérez-Rodríguez A, Ortega Á, Martínez-Ruiz A, Barreiro O, García-Domínguez CA, et al. Endothelial nitric oxide synthase regulates n-ras activation on the golgi complex of antigen-stimulated t cells. *Proc Natl Acad Sci USA* (2008) 105:10507– 12. doi: 10.1073/pnas.0711062105

72. Sriskandan S, Evans TJ, Cohen J. Bacterial superantigen-induced human lymphocyte responses are nitric oxide dependent and mediated by IL-12 and IFN-γ. J Immunol (1996) 156(7):2430–5.

73. Niedbala W, Qing WX, Campbell C, Thomson D, Komai-Koma M, Liew FY. Nitric oxide preferentially induces type 1 t cell differentiation by selectively upregulating IL-12 receptor β 2 expression *via* cGMP. *Proc Natl Acad Sci USA* (2002) 99:16186–91. doi: 10.1073/pnas.252464599

74. King MR, Ismail AS, Davis LS, Karp DR. Oxidative stress promotes polarization of human t cell differentiation toward a t helper 2 phenotype. *J Immunol* (2006) 176:2765–72. doi: 10.4049/jimmunol.176.5.2765

75. Chen W, Li L, Brod T, Saeed O, Thabet S, Jansen T, et al. Role of increased guanosine triphosphate cyclohydrolase-1 expression and tetrahydrobiopterin levels upon t cell activation. *J Biol Chem* (2011) 286:13846–51. doi: 10.1074/jbc.M110.191023

76. Lee S-W, Choi H, Eun S-Y, Fukuyama S, Croft M. Nitric oxide modulates TGF- β -directive signals to suppress Foxp3 + regulatory t cell differentiation and potentiate Th1 development. *J Immunol* (2011) 186:6972–80. doi: 10.4049/jimmunol.1100485

77. Brahmachari S, Pahan K. Myelin basic protein priming reduces the expression of Foxp3 in t cells *via* nitric oxide. *J Immunol* (2010) 184:1799–809. doi: 10.4049/jimmunol.0804394

78. Chen CA, Wang TY, Varadharaj S, Reyes LA, Hemann C, Talukder MAH, et al. S-glutathionylation uncouples eNOS and regulates its cellular and vascular function. *Nature* (2010) 468:1115–8. doi: 10.1038/nature09599

79. O'Neill LAJ, Kishton RJ, Rathmell J. A guide to immunometabolism for immunologists. Nat Rev Immunol (2016) 16:553-65. doi: 10.1038/nri.2016.70

80. Haruki H, Hovius R, Pedersen MG, Johnsson K. Tetrahydrobiopterin biosynthesis as a potential target of the kynurenine pathway metabolite xanthurenic acid. J Biol Chem (2016) 291:652–7. doi: 10.1074/jbc.C115.680488

81. Lu T, Ramakrishnan R, Altiok S, Youn JI, Cheng P, Celis E, et al. Tumorinfiltrating myeloid cells induce tumor cell resistance to cytotoxic t cells in mice. *J Clin Invest* (2011) 121:4015–29. doi: 10.1172/JCI45862

82. Lu T, Gabrilovich DI. Molecular pathways: Tumor-infiltrating myeloid cells and reactive oxygen species in regulation of tumor microenvironment. *Clin Cancer Res* (2012) 291:652–7. doi: 10.1158/1078-0432.CCR-11-2939

83. Nagaraj S, Gupta K, Pisarev V, Kinarsky L, Sherman S, Kang L, et al. Altered recognition of antigen is a mechanism of CD8+ t cell tolerance in cancer. *Nat Med* (2007) 13:828–35. doi: 10.1038/nm1609

84. Molon B, Ugel S, Del Pozzo F, Soldani C, Zilio S, Avella D, et al. Chemokine nitration prevents intratumoral infiltration of antigen-specific t cells. *J Exp Med* (2011) 208:1949–62. doi: 10.1084/jem.20101956

85. Sato E, Simpson KL, Grisham MB, Koyama S, Robbins RA. Effects of reactive oxygen and nitrogen metabolites on RANTES- and IL- 5-induced eosinophil chemotactic activity in vitro. *Am J Pathol* (1999) 155:591–8. doi: 10.1016/S0002-9440 (10)65154-1

86. Sato E, Simpson KL, Grisham MB, Koyama S, Robbins RA. Effects of reactive oxygen and nitrogen metabolites on MCP-1-induced monocyte chemotactic activity in vitro. *Am J Physiol Lung Cell Mol Physiol* (1999) 277:L543–9. doi: 10.1152/ajplung.1999.277.3.1543

87. Cunha PP, Bargiela D, Minogue E, Krause LCM, Barbieri L, Brombach C, et al. Infiltration of tumors is regulated by t cell-intrinsic nitric oxide synthesis. *Cancer Immunol Res* (2023) 11:351–63. doi: 10.1158/2326-6066.CIR-22-0387

88. Fiorucci S, Santucci L, Antonelli E, Distrutti E, Del Sero G, Morelli O, et al. NOaspirin protects from t cell-mediated liver injury by inhibiting caspase-dependent processing of Th1-like cytokines. *Gastroenterology* (2000) 18:404–21. doi: 10.1016/ S0016-5085(00)70223-X

89. Fiorucci S, Mencarelli A, Palazzetti B, Del Soldato P, Morelli A, Ignarro LJ. An NO derivative of ursodeoxycholic acid protects against fas-mediated liver injury by inhibiting caspase activity. *Proc Natl Acad Sci USA* (2001) 98:2652–7. doi: 10.1073/ pnas.041603898

90. Huang FP, Niedbala W, Wei XQ, Xu D, Feng GJ, Robinson JH, et al. Nitric oxide regulates Th1 cell development through the inhibition of IL-12 synthesis by macrophages. *Eur J Immunol* (1998) 28:4062–70. doi: 10.1002/(SICI)1521-4141 (199812)28:12<4062::AID-IMMU4062>3.0.CO;2-K

91. Niedbala W, Cai B, Liu H, Pitman N, Chang L, Liew FY. Nitric oxide induces CD4+CD25+ Foxp3- regulatory t cells from CD4+CD25- t cells via p53, IL-2, and OX40. Proc Natl Acad Sci USA (2007) 104:15478–83. doi: 10.1073/pnas.0703725104

92. Liu Z, Wang Y, Huang Y, Kim BYS, Shan H, Wu D, et al. Tumor vasculatures: A new target for cancer immunotherapy. *Trends Pharmacol Sci* (2019) 40:613–23. doi: 10.1016/j.tips.2019.07.001

93. Lee WS, Yang H, Chon HJ, Kim C. Combination of anti-angiogenic therapy and immune checkpoint blockade normalizes vascular-immune crosstalk to potentiate cancer immunity. *Exp Mol Med* (2020) 52:1475–85. doi: 10.1038/s12276-020-00500-y

94. Huang Y, Yuan J, Righi E, Kamoun WS, Ancukiewicz M, Nezivar J, et al. Vascular normalizing doses of antiangiogenic treatment reprogram the immunosuppressive tumor microenvironment and enhance immunotherapy. *Proc Natl Acad Sci USA* (2012). doi: 10.1073/pnas.1215397109

95. Stylianopoulos T, Munn LL, Jain RK. Reengineering the physical microenvironment of tumors to improve drug delivery and efficacy: From mathematical modeling to bench to bedside. *Trends Cancer* (2018) 4:292–319. doi: 10.1016/j.trecan.2018.02.005

96. Mpekris F, Voutouri C, Baish JW, Duda DG, Munn LL, Stylianopoulos T, et al. Combining microenvironment normalization strategies to improve cancer immunotherapy. *Proc Natl Acad Sci USA* (2020) 117:3728–37. doi: 10.1073/pnas.1919764117

97. Jain RK. Normalizing tumor microenvironment to treat cancer: Bench to bedside to biomarkers. J Clin Oncol (2013) 31:2205–18. doi: 10.1200/JCO.2012.46.3653

98. Huang Y, Goel S, Duda DG, Fukumura D, Jain RK. Vascular normalization as an emerging strategy to enhance cancer immunotherapy. *Cancer Res* (2013) 73:2943–8. doi: 10.1158/0008-5472.CAN-12-4354

99. Salmon H, Franciszkiewicz K, Damotte D, Dieu-Nosjean MC, Validire P, Trautmann A, et al. Matrix architecture defines the preferential localization and migration of t cells into the stroma of human lung tumors. *J Clin Invest* (2012) 122:899–910. doi: 10.1172/JCI45817

100. Mariathasan S, Turley SJ, Nickles D, Castiglioni A, Yuen K, Wang Y, et al. TGF β attenuates tumour response to PD-L1 blockade by contributing to exclusion of t cells. *Nature* (2018) 554:544–8. doi: 10.1038/nature25501

101. Munn LL, Jain RK. Vascular regulation of antitumor immunity. Sci (1979) (2019) 365:544–5. doi: 10.1126/science.aaw7875

102. Datta M, Coussens LM, Nishikawa H, Hodi FS, Jain RK. Reprogramming the tumor microenvironment to improve immunotherapy: Emerging strategies and combination therapies. *Am Soc Clin Oncol Educ Book* (2019) 39:165–74. doi: 10.1200/edbk_237987

103. Martin JD, Cabral H, Stylianopoulos T, Jain RK. Improving cancer immunotherapy using nanomedicines: progress, opportunities and challenges. *Nat Rev Clin Oncol* (2020) 17:251–66. doi: 10.1038/s41571-019-0308-z

104. Jain RK, Stylianopoulos T. Delivering nanomedicine to solid tumors. Nat Rev Clin Oncol (2010) 7:653–64. doi: 10.1038/nrclinonc.2010.139

105. Noman MZ, Desantis G, Janji B, Hasmim M, Karray S, Dessen P, et al. PD-L1 is a novel direct target of HIF-1 α , and its blockade under hypoxia enhanced MDSC-mediated t cell activation. *J Exp Med* (2014) 211:781–90. doi: 10.1084/jem.20131916

106. Noman MZ, Hasmim M, Messai Y, Terry S, Kieda C, Janji B, et al. Hypoxia: A key player in antitumor immune response. a review in the theme: Cellular responses to hypoxia. *Am J Physiol Cell Physiol* (2015) 309:C569–79. doi: 10.1152/ajpcell.00207.2015

107. Facciabene A, Peng X, Hagemann IS, Balint K, Barchetti A, Wang LP, et al. Tumour hypoxia promotes tolerance and angiogenesis *via* CCL28 and t reg cells. *Nature* (2011) 475:226–30. doi: 10.1038/nature10169

108. Calcinotto A, Filipazzi P, Grioni M, Iero M, De Milito A, Ricupito A, et al. Modulation of microenvironment acidity reverses anergy in human and murine tumorinfiltrating t lymphocytes. *Cancer Res* (2012) 72:2746–56. doi: 10.1158/0008-5472.CAN-11-1272

109. Palazón A, Aragonés J, Morales-Kastresana A, Ortiz De Landázuri M, Melero I. Molecular pathways: Hypoxia response in immune cells fighting or promoting cancer. *Clin Cancer Res* (2012) 18:1207–13. doi: 10.1158/1078-0432.CCR-11-1591

110. Sonveaux P, Frérart F, Bouzin C, Brouet A, DeWever J, Jordan BF, et al. Irradiation promotes akt-targeting therapeutic gene delivery to the tumor vasculature. *Int J Radiat Oncol Biol Phys* (2007) 67:1155–62. doi: 10.1016/j.ijrobp.2006.11.031

111. Falcón BL, Hashizume H, Koumoutsakos P, Chou J, Bready JV, Coxon A, et al. Contrasting actions of selective inhibitors of angiopoietin-1 and angiopoietin-2 on the normalization of tumor blood vessels. *Am J Pathol* (2009) 175:2159–70. doi: 10.2353/ ajpath.2009.090391

112. Zacharek A, Chen J, Zhang C, Cui X, Roberts C, Jiang H, et al. Nitric oxide regulates Angiopoietin1/Tie2 expression after stroke. *Neurosci Lett* (2006) 404:28–32. doi: 10.1016/j.neulet.2006.05.027

113. Kobayashi H, DeBusk LM, Babichev YO, Dumont DJ, Lin PC. Hepatocyte growth factor mediates angiopoietin-induced smooth muscle cell recruitment. *Blood* (2006) 108:1260–6. doi: 10.1182/blood-2005-09-012807

114. Schulz E, Jansen T, Wenzel P, Daiber A, Münzel T. Nitric oxide, tetrahydrobiopterin, oxidative stress, and endothelial dysfunction in hypertension. *Antioxid Redox Signal* (2008) 10:1115–26. doi: 10.1089/ars.2007.1989

115. Roe ND, Ren J. Nitric oxide synthase uncoupling: A therapeutic target in cardiovascular diseases. *Vascul Pharmacol* (2012) 57:168–72. doi: 10.1016/ j.vph.2012.02.004

116. Fukumura D, Kashiwagi S, Jain RK. The role of nitric oxide in tumour progression. *Nat Rev Cancer* (2006) 6:521–34. doi: 10.1038/nrc1910

117. Kashiwagi S, Izumi Y, Gohongi T, Demou ZN, Xu L, Huang PL, et al. NO mediates mural cell recruitment and vessel morphogenesis in murine melanomas and tissue-engineered blood vessels. *J Clin Invest* (2005) 115:1816–27. doi: 10.1172/JCI24015

118. Schleicher M, Yu J, Murata T, Derakhshan B, Atochin D, Qian L, et al. The Akt1-eNOS axis illustrates the specificity of kinase-substrate relationships in vivo. *Sci Signal* (2009) 2:ra41. doi: 10.1126/scisignal.2000343

119. Cardnell RJG, Mikkelsen RB. Nitric oxide synthase inhibition enhances the antitumor effect of radiation in the treatment of squamous carcinoma xenografts. *PloS One* (2011) 6:e20147. doi: 10.1371/journal.pone.0020147

120. Ng QS, Goh V, Milner J, Stratford MR, Folkes LK, Tozer GM, et al. Effect of nitric-oxide synthesis on tumour blood volume and vascular activity: a phase i study. *Lancet Oncol* (2007) 8:111–8. doi: 10.1016/S1470-2045(07)70001-3

121. Siemann DW. The unique characteristics of tumor vasculature and preclinical evidence for its selective disruption by tumor-vascular disrupting agents. *Cancer Treat Rev* (2011) 37:63–74. doi: 10.1016/j.ctrv.2010.05.001

122. Hanahan D, Weinberg RA. The hallmarks of cancer review douglas. *Cell* (2000) 100:57–70. doi: 10.1016/s0092-8674(00)81683-9

123. Crusz SM, Balkwill FR. Inflammation and cancer: Advances and new agents. Nat Rev Clin Oncol (2015) 12:584–96. doi: 10.1038/nrclinonc.2015.105

124. Kundu JK, Surh YJ. Inflammation: Gearing the journey to cancer. Mutat Res Rev Mutat Res (2008) 659:15–30. doi: 10.1016/j.mrrev.2008.03.002

125. Reynaert NL, Ckless K, Korn SH, Vos N, Guala AS, Wouters EFM, et al. Nitric oxide represses inhibitory κB kinase through s-nitrosylation. *Proc Natl Acad Sci USA* (2004) 101:8945–50. doi: 10.1073/pnas.0400588101

126. Yakovlev VA, Barani IJ, Rabender CS, Black SM, Leach JK, Graves PR, et al. Tyrosine nitration of IκBα: A novel mechanism for NF-κB activation. *Biochemistry* (2007) 46:11671-83. doi: 10.1021/bi701107z

127. Godoy LC, Moretti AI, Jurado MC, Oxer D, Janiszewski M, Ckless K, et al. Loss of CD40 endogenous s-nitrosylation during inflammatory response in endotoxemic mice and patients with sepsis. *Shock* (2010) 33:626–33. doi: 10.1097/SHK.0b013e3181cb88e6

128. Lim SY, Raftery M, Cai H, Hsu K, Yan WX, Hseih H-L, et al. S -nitrosylated S100A8: Novel anti-inflammatory properties. *J Immunol* (2008) 181:5627–36. doi: 10.4049/jimmunol.181.8.5627

129. Park HS, Huh SH, Kim MS, Lee SH, Choi EJ. Nitric oxide negatively regulates cjun n-terminal kinase/stress-activated protein kinase by means of s-nitrosylation. *Proc Natl Acad Sci USA* (2000) 97:14382–7. doi: 10.1073/pnas.97.26.14382

130. Park HS, Mo JS, Choi EJ. Nitric oxide inhibits an interaction between JNK1 and c-jun through nitrosylation. *Biochem Biophys Res Commun* (2006) 351:281–6. doi: 10.1016/j.bbrc.2006.10.034

131. Smith BC, Fernhoff NB, Marletta MA. Mechanism and kinetics of inducible nitric oxide synthase auto- s -nitrosation and inactivation. *Biochemistry* (2012) 51:1028–40. doi: 10.1021/bi201818c

132. Mitchell DA, Erwin PA, Michel T, Marletta MA. S-nitrosation and regulation of inducible nitric oxide synthase. *Biochemistry* (2005) 44:4636–47. doi: 10.1021/bi0474463

133. Lefer DJ, Jones SP, Girod WG, Baines A, Grisham MB, Cockrell AS, et al. Leukocyte-endothelial cell interactions in nitric oxide synthase- deficient mice. *Am J Physiol Heart Circ Physiol* (1999) 276:H1943–50. doi: 10.1152/ajpheart.1999.276.6.h1943

134. Sanz MJ, Hickey MJ, Johnston B, McCafferty DM, Raharjo E, Huang PL, et al. Neuronal nitric oxide synthase (NOS) regulates leukocyte-endothelial cell interactions in endothelial NOS deficient mice. *Br J Pharmacol* (2001) 134:305–12. doi: 10.1038/ sj.bjp.0704234

135. Hickey MJ, Granger DN, Kubes P. Inducible nitric oxide synthase (iNOS) and regulation of leucocyte/endothelial cell interactions: Studies in iNOS-deficient mice. *Acta Physiol Scand* (2001) 173:119–26. doi: 10.1046/j.1365-201X.2001.00892.x

136. Ahluwalia A, Foster P, Scotland RS, McLean PG, Mathur A, Perretti M, et al. Antiinflammatory activity of soluble guanylate cyclase: cGMP-dependent down-regulation of p-selectin expression and leukocyte recruitment. *Proc Natl Acad Sci USA* (2004) 101:1386–91. doi: 10.1073/pnas.0304264101

137. Barber GN. STING: infection, inflammation and cancer. *Nat Rev Immunol* (2015) 15:760–70. doi: 10.1038/nri3921

138. Hou J, Karin M, Sun B. Targeting cancer-promoting inflammation — have anti-inflammatory therapies come of age? *Nat Rev Clin Oncol* (2021) 18:261–79. doi: 10.1038/s41571-020-00459-9

139. Xin Yu J, Hubbard-Lucey VM, Tang J. Immuno-oncology drug development goes global. *Nat Rev Drug Discovery* (2019) 18:899–900. doi: 10.1038/d41573-019-00167-9

140. Li X, Song W, Shao C, Shi Y, Han W. Emerging predictors of the response to the blockade of immune checkpoints in cancer therapy. *Cell Mol Immunol* (2019) 16:28–39. doi: 10.1038/s41423-018-0086-z

141. Haslam A, Prasad V. Estimation of the percentage of US patients with cancer who are eligible for and respond to checkpoint inhibitor immunotherapy drugs. *JAMA Netw Open* (2019) 2:e192535. doi: 10.1001/jamanetworkopen.2019.2535

142. Brooks ED, Chang JY. Time to abandon single-site irradiation for inducing abscopal effects. *Nat Rev Clin Oncol* (2019) 16:123–35. doi: 10.1038/s41571-018-0119-7

143. Bernstein MB, Krishnan S, Hodge JW, Chang JY. Immunotherapy and stereotactic ablative radiotherapy (ISABR): A curative approach? *Nat Rev Clin Oncol* (2016) 13:516–24. doi: 10.1038/nrclinonc.2016.30

144. Fukumura D, Kloepper J, Amoozgar Z, Duda DG, Jain RK. Enhancing cancer immunotherapy using antiangiogenics: Opportunities and challenges. *Nat Rev Clin Oncol* (2018) 15:325–40. doi: 10.1038/nrclinonc.2018.29

145. Jain RK. Antiangiogenesis strategies revisited: From starving tumors to alleviating hypoxia. *Cancer Cell* (2014) 26:605–22. doi: 10.1016/j.ccell.2014.10.006

146. Stein S, Pishvaian MJ, Lee MS, Lee K-H, Hernandez S, Kwan A, et al. Safety and clinical activity of 1L atezolizumab + bevacizumab in a phase ib study in hepatocellular carcinoma (HCC). *J Clin Oncol* (2018) 36. doi: 10.1200/jco.2018.36.15_suppl.4074

147. Motzer RJ, Penkov K, Haanen J, Rini B, Albiges L, Campbell MT, et al. Avelumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. *New Engl J Med* (2019) 280:1103–15. doi: 10.1056/NEJMoa1816047

148. Rini BI, Plimack ER, Stus V, Gafanov R, Hawkins R, Nosov D, et al. Pembrolizumab plus axitinib versus sunitinib for advanced renal-cell carcinoma. *New Engl J Med* (2019) 280:1116–27. doi: 10.1056/NEJMoa1816714

149. Wallin JJ, Bendell JC, Funke R, Sznol M, Korski K, Jones S, et al. Atezolizumab in combination with bevacizumab enhances antigen-specific t-cell migration in metastatic renal cell carcinoma. *Nat Commun* (2016) 7:12624. doi: 10.1038/ ncomms12624

150. Wu X, Giobbie-Hurder A, Liao X, Lawrence D, McDermott D, Zhou J, et al. VEGF neutralization plus CTLA-4 blockade alters soluble and cellular factors associated with enhancing lymphocyte infiltration and humoral recognition in melanoma. *Cancer Immunol Res* (2016) 4:858–68. doi: 10.1158/2326-6066.CIR-16-0084

151. Wu X, Giobbie-Hurder A, Connolly EM, Li J, Liao X, Severgnini M, et al. Anti-CTLA-4 based therapy elicits humoral immunity to galectin-3 in patients with metastatic melanoma. *Oncoimmunology* (2018) 7:e1440930. doi: 10.1080/ 2162402X.2018.1440930

152. Baffert F, Le T, Sennino B, Thurston G, Kuo CJ, Hu-Lowe D, et al. Cellular changes in normal blood capillaries undergoing regression after inhibition of VEGF signaling. *Am J Physiol Heart Circ Physiol* (2006) 290:H547–59. doi: 10.1152/ ajpheart.00616.2005

153. Hurwitz H, Fehrenbacher L, Novotny W, Cartwright T, Hainsworth J, Heim W, et al. Bevacizumab plus irinotecan, fluorouracil, and leucovorin for metastatic colorectal cancer. *New Engl J Med* (2004) 350:2335–42. doi: 10.1056/NEJMoa032691

154. Schrijvers BF, Flyvbjerg A, De Vriese AS. The role of vascular endothelial growth factor (VEGF) in renal pathophysiology. *Kidney Int* (2004) 65:2003–17. doi: 10.1111/j.1523-1755.2004.00621.x

155. Scappaticci FA, Fehrenbacher L, Cartwright T, Hainsworth JD, Heim W, Berlin J, et al. Surgical wound healing complications in metastatic colorectal cancer patients treated with bevacizumab. *J Surg Oncol* (2005) 91:173–80. doi: 10.1002/jso.20301

156. Subbiah IM, Lenihan DJ, Tsimberidou AM. Cardiovascular toxicity profiles of vascular-disrupting agents. *Oncologist* (2011) 16:1120-30. doi: 10.1634/ theoncologist.2010-0432

157. Martin JD, Seano G, Jain RK. Normalizing function of tumor vessels: Progress, opportunities, and challenges. *Annu Rev Physiol* (2019). doi: 10.1146/annurev-physiol-020518-114700

158. Werner ER, Blau N, Thöny B. Tetrahydrobiopterin: Biochemistry and pathophysiology. *Biochem J* (2011) 438:397-414. doi: 10.1042/BJ20110293

159. Coffelt SB, de Visser KE. Immune-mediated mechanisms influencing the efficacy of anticancer therapies. *Trends Immunol* (2015) 36:198–216. doi: 10.1016/j.it.2015.02.006

160. Kim RK, Suh Y, Cui YH, Hwang E, Lim EJ, Yoo KC, et al. Fractionated radiation-induced nitric oxide promotes expansion of glioma stem-like cells. *Cancer Sci* (2013) 104:1172–7. doi: 10.1111/cas.12207

161. Kevin Leach J, Black SM, Schmidt-Ullrich RK, Mikkelsen RB. Activation of constitutive nitric-oxide synthase activity is an early signaling event induced by ionizing radiation. J Biol Chem (2002) 277:15400–6. doi: 10.1074/jbc.M110309200

162. Berbee M, Fu Q, Boerma M, Pathak R, Zhou D, Kumar KS, et al. Reduction of radiation-induced vascular nitrosative stress by the vitamin e analog γ -tocotrienol: Evidence of a role for tetrahydrobiopterin. *Int J Radiat Oncol Biol Phys* (2011) 79:884–91. doi: 10.1016/j.ijrobp.2010.08.032

163. Pathak R, Pawar SA, Fu Q, Gupta PK, Berbée M, Garg S, et al. Characterization of transgenic gfrp knock-in mice: Implications for tetrahydrobiopterin in modulation of normal tissue radiation responses. *Antioxid Redox Signal* (2014) 20:1436–46. doi: 10.1089/ars.2012.5025

164. Cheema AK, Pathak R, Zandkarimi F, Kaur P, Alkhalil L, Singh R, et al. Liver metabolomics reveals increased oxidative stress and fibrogenic potential in gfrp transgenic mice in response to ionizing radiation. *J Proteome Res* (2014) 13:3065–74. doi: 10.1021/pr500278t

165. Rodríguez-Ruiz ME, Vanpouille-Box C, Melero I, Formenti SC, Demaria S. Immunological mechanisms responsible for radiation-induced abscopal effect. *Trends Immunol* (2018) 39:644–55. doi: 10.1016/j.it.2018.06.001

166. Alper T. Adding two components of radiosensitization by oxygen. Int J Radiat Biol (1984) 46:569–85. doi: 10.1080/09553008414551771

167. Watts ME, Hodgkiss RJ, Jones NR, Fowler JF. Radiosensitization of chinese hamster cells by oxygen and misonidazole at low x-ray doses. *Int J Radiat Biol* (1986) 50:1009–21. doi: 10.1080/09553008614551421

168. Stevens G, Joiner M, Joiner B, Johns H, Denekamp J. Radiosensitization of mouse skin by oxygen and depletion of glutathione. *Int J Radiat Oncol Biol Phys* (1995) 33:399–408. doi: 10.1016/0360-3016(95)00070-F

169. Oronsky BT, Knox SJ, Scicinski JJ. Is nitric oxide (NO) the last word in radiosensitization? a review. *Transl Oncol* (2012) 5:66–71. doi: 10.1593/tlo.11307

170. Nagane M, Yasui H, Yamamori T, Zhao S, Kuge Y, Tamaki N, et al. Radiationinduced nitric oxide mitigates tumor hypoxia and radioresistance in a murine SCCVII tumor model. *Biochem Biophys Res Commun* (2013) 437:420–5. doi: 10.1016/j.bbrc.2013.06.093

171. Brüne B. Nitric oxide: NO apoptosis or turning it ON? Cell Death Differ (2003) 10:864–9. doi: 10.1038/sj.cdd.4401261

172. Fajardo LF, Stewart JR, Cohn KE. Morphology of radiation-induced heart disease. Arch Pathol (Chic) (1968) 86:512-9.

173. Kong P, Christia P, Frangogiannis NG. The pathogenesis of cardiac fibrosis. Cell Mol Life Sci (2014) 71:549–74. doi: 10.1007/s00018-013-1349-6

174. Mezzaroma E, Di X, Graves P, Toldo S, Van Tassell BW, Kannan H, et al. A mouse model of radiation-induced cardiomyopathy. *Int J Cardiol* (2012) 156:231–3. doi: 10.1016/j.ijcard.2012.01.038

175. Chello M, Mastroroberto P, Romano R, Zofrea S, Bevacqua I, Marchese AR. Changes in the proportion of types i and III collagen in the left ventricular wall of patients with post-irradiative pericarditis. *Vascular* (1996) 4. doi: 10.1177/096721099600400221

176. Bentzen SM. Preventing or reducing late side effects of radiation therapy: Radiobiology meets molecular pathology. *Nat Rev Cancer* (2006) 6:702–13. doi: 10.1038/ nrc1950

177. Seemann I, Gabriels K, Visser NL, Hoving S, Te Poele JA, Pol JF, et al. Irradiation induced modest changes in murine cardiac function despite progressive structural damage to the myocardium and microvasculature. *Radiotherapy Oncol* (2012) 103:143–50. doi: 10.1016/j.radonc.2011.10.011

178. Schultz-Hector S, Trott KR. Radiation-induced cardiovascular diseases: Is the epidemiologic evidence compatible with the radiobiologic data? *Int J Radiat Oncol Biol Phys* (2007) 67:10–8. doi: 10.1016/j.ijrobp.2006.08.071

179. Anscher MS, Vujaskovic Z. Mechanisms and potential targets for prevention and treatment of normal tissue injury after radiation therapy. *Semin Oncol* (2005) 32: S86–91. doi: 10.1053/j.seminoncol.2005.03.015 180. Zhao W, Robbins M. Inflammation and chronic oxidative stress in radiationinduced late normal tissue injury: Therapeutic implications. *Curr Med Chem* (2008) 16:130–43. doi: 10.2174/092986709787002790

181. Wynn TA. Cellular and molecular mechanisms of fibrosis. J Pathol (2008) 214:199-210. doi: 10.1002/path.2277

182. Berbee M, Fu Q, Sree Kumar K, Hauer-Jensen M. Novel strategies to ameliorate radiation injury: A possible role for tetrahydrobiopterin. *Curr Drug Targets* (2012) 11:1366–74. doi: 10.2174/1389450111009011366

183. Rabender CS, Mezzaroma E, Yakovlev VA, Mauro AG, Bonaventura A, Abbate A, et al. Mitigation of radiation-induced lung and heart injuries in mice by oral sepiapterin after irradiation. *Radiat Res* (2021) 195:463–73. doi: 10.1667/RADE-20-00249.1

184. Herrera FG, Bourhis J, Coukos G. Radiotherapy combination opportunities leveraging immunity for the next oncology practice. *CA Cancer J Clin* (2017) 67:65–85. doi: 10.3322/caac.21358

185. Twyman-Saint Victor C, Rech AJ, Maity A, Rengan R, Pauken KE, Stelekati E, et al. Radiation and dual checkpoint blockade activate non-redundant immune mechanisms in cancer. *Nature* (2015) 520:373–7. doi: 10.1038/nature14292

186. Spiotto M, Fu YX, Weichselbaum RR. The intersection of radiotherapy and immunotherapy: Mechanisms and clinical implications. *Sci Immunol* (2016) 1: EAAG1266. doi: 10.1126/sciimmunol.aag1266

187. Garris CS, Arlauckas SP, Kohler RH, Trefny MP, Garren S, Piot C, et al. Successful anti-PD-1 cancer immunotherapy requires t cell-dendritic cell crosstalk involving the cytokines IFN- γ and IL-12. *Immunity* (2018) 49:1148–1161.e7. doi: 10.1016/j.immuni.2018.09.024

188. Tian L, Goldstein A, Wang H, Lo HC, Kim IS, Welte T, et al. Mutual regulation of tumour vessel normalization and immunostimulatory reprogramming. *Nature* (2017) 544:250–4. doi: 10.1038/nature21724

189. Muntau AC, Röschinger W, Habich M, Demmelmair H, Hoffmann B, Sommerhoff CP, et al. Tetrahydrobiopterin as an alternative treatment for mild phenylketonukia. *New Engl J Med* (2002) 347:2122–32. doi: 10.1056/NEJMoa021654

190. Ong GH, Lian BSX, Kawasaki T, Kawai T. Exploration of pattern recognition receptor agonists as candidate adjuvants. *Front Cell Infect Microbiol* (2021) 11:745016. doi: 10.3389/fcimb.2021.745016