Check for updates

OPEN ACCESS

EDITED BY Alison Colquhoun, University of São Paulo, Brazil

REVIEWED BY Shilpa Kuttikrishnan, Hamad Medical Corporation, Qatar Zarema Albakova, University of York, United Kingdom João Pessoa,

University of Aveiro, Portugal

*CORRESPONDENCE

Yan Liu 24947609@qq.com Bo Wang bowang581@gmail.com

[†]These authors have contributed equally to this work and share first authorship

RECEIVED 18 September 2023 ACCEPTED 23 November 2023 PUBLISHED 30 November 2023

CITATION

Xiang Y, Liu X, Sun Q, Liao K, Liu X, Zhao Z, Feng L, Liu Y and Wang B (2023) The development of cancers research based on mitochondrial heat shock protein 90. *Front. Oncol.* 13:1296456. doi: 10.3389/fonc.2023.1296456

COPYRIGHT

© 2023 Xiang, Liu, Sun, Liao, Liu, Zhao, Feng, Liu and Wang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

The development of cancers research based on mitochondrial heat shock protein 90

Yuchu Xiang^{1†}, Xudong Liu^{2†}, Qi Sun^{3†}, Kuo Liao⁴, Xiaohan Liu⁵, Zihui Zhao⁶, Lishuang Feng⁷, Yan Liu⁸* and Bo Wang^{2,9}*

¹West China Hospital of Sichuan University, Sichuan University, Chengdu, China, ²Institute of Medical Microbiology and Hygiene, Faculty of Medicine, University of Freiburg, Freiburg, Germany, ³Department of Pharmacology and Pharmacy, Li Ka Shing Faculty of Medicine, Centre for Safe Medication Practice and Research, The University of Hong Kong, Pok Fu Lam, Hong Kong SAR, China, ⁴School of Biology and Biological Engineering, South China University of Technology, Guangzhou, China, ⁵Multiscale Research Institute of Complex Systems, Fudan University, Shanghai, China, ⁶Shanghai Jiao Tong University School of Medicine, Shanghai, China, ⁷School of Public Health, Li Ka Shing Faculty of Medicine, The University of Hong Kong, Hong Kong, Hong Kong SAR, China, ⁸Department of Organ Transplantation, Guizhou Provincial People's Hospital, Guiyang, Guizhou, China, ⁹Department of Urology, Guizhou Provincial People's Hospital, Guiyang, Guizhou, China

Mitochondrial heat shock protein 90 (mtHsp90), including Tumor necrosis factor receptor-associated protein 1 (TRAP1) and Hsp90 translocated from cytoplasm, modulating cellular metabolism and signaling pathways by altering the conformation, activity, and stability of numerous client proteins, and is highly expressed in tumors. mtHsp90 inhibition results in the destabilization and eventual degradation of its client proteins, leading to interference with various tumor-related pathways and efficient control of cancer cell development. Among these compounds, gamitrinib, a specific mtHsp90 inhibitor, has demonstrated its safety and efficacy in several preclinical investigations and is currently undergoing evaluation in clinical trials. This review aims to provide a comprehensive overview of the present knowledge pertaining to mtHsp90, encompassing its structure and function. Moreover, our main emphasis is on the development of mtHsp90 inhibitors for various cancer therapies, to present a thorough overview of the recent pre-clinical advancements in this field.

KEYWORDS

mtHsp90, TRAP1, molecular chaperone, mitochondria, gamitrinib

Abbreviations: mtHsp90, mitochondrial heat shock protein 90; TRAP1, Tumor necrosis factor receptorassociated protein 1; NTD, N-terminal domain; MD, middle domain; CTD, C-terminal domain; SDHB, B subunit of succinate dehydrogenase; SDH, succinate dehydrogenase; OXPHOS, mitochondrial oxidative phosphorylation; NSCLC, non-small cell lung cancer; gamitrinib–TPP, gamitrinib-triphenylphosphonium; GBM, glioblastoma multiforme; TMZ, temozolomide; CRC, colorectal cancer; ROS, reactive oxygen species.

1 Introduction

Heat shock protein 90 (Hsp90) is a highly conserved family of molecular chaperones that serves a pivotal function in the processes of protein folding, stabilization, and degradation. With several hundred protein clients, Hsp90 is involved in various cellular processes, such as signal transduction, protein trafficking, and DNA repair (1). It functions as an ATP-dependent molecular chaperone, and it's the crucial role of facilitating the appropriate folding, conformational alteration, activation, and regulation of client proteins. During its functional process, it combines and hydrolyzes ATP, while also involving client proteins and cochaperones, which are proteins responsible for regulating the activity and function of Hsp90 (2). Hsp90 exhibits distinct localization patterns inside diverse cellular compartments, each associated with specific functional tasks.

In mammalian cells, the Hsp90 family consists of four highly conserved isoforms: the endoplasmic reticulum-localised Grp94, the cytoplasmic Hsp90 α and Hsp90 β , and TRAP1, which was found in mitochondria (3). These different members exhibit a common molecular chaperone activity pattern, but they selectively bind to diverse client proteins, which is influenced in part by their cellular localization (4). TRAP1 is a Hsp90 homolog and the predominant chaperone in the mitochondria of cancer cells (5). Particularly, Hsp90 compartmentalized in mitochondria is essential regulator of bioenergetics in tumor cells but not normal cells (6). Though many evidences indicated mtHsp90 could be a new potential antitumor target, there is not a review updating its antitumor mechanism and the newest advancements in clinical therapy development.

Due to the increasing number of members of various human heat shock protein (HSP) families, inconsistencies in their nomenclature often result in ambiguity and confusion. In 2009, Kampinga et al. proposed Guidelines for the nomenclature of the human heat shock proteins (7). Among them, the Hsp90 family was renamed as the HSPC family, with the original Hsp90 referred to as HSPC1, Hsp90 α and Hsp90 β as HSPC2 and HSPC3, respectively, and GRP94 renamed as HSPC4 whereas TRAP1/Hsp75 was renamed as HSPC5. We have summarized the agreement of different scholars on the name of Hsp90 through extensive literature review, and hereby we would like to define the concept of Hsp90 mentioned in the article.

Broadly, Hsp90 can refer to all proteins in the Hsp90 family, whereas narrowly, Hsp90 refers to the classical Hsp90 α and Hsp90 β . Hoter et al. (8) in 2018 summarized, based on available studies, that Hsp90 has been customarily used for Hsp90 α and Hsp90 β (9, 10). We have reached the same conclusion through an extensive review of the literature from earlier years. Throughout the discovery history of the Hsp90 family, sequence information for Hsp90 α and Hsp90 β was first revealed in 1984 (11), whereas grp94 was first sequenced in 1986 (12), and TRAP1 in 1995 (13). Since Hsp90 α and Hsp90 β in the cytoplasm were first identified, they have been referred to as Hsp90 by researchers in the subsequent literature, whereas Hsp90 homologous proteins in the endoplasmic reticulum and mitochondria are denoted by grp94 and TRAP1,

respectively. Typically, without specific indication, Hsp90 is referred to as narrowly defined Hsp90, i.e., Hsp90 α and Hsp90 β in the cytoplasm.

In 2007, Kang et al. (14) showed that Hsp90 and its related molecule TRAP1 were abundantly present in the mitochondria of tumor cells, and that Hsp90 was not present in normal mitochondria but was prevalent in the cytoplasm of both normal and tumor cells. However, the exact mechanism of how Hsp90 is imported into mitochondria remains to be fully elucidated. In addition, Boucherat et al, in their study of mitochondrial Hsp90 accumulation and vascular remodeling in pulmonary arterial hypertension, also mentioned that Hsp90 is present in tumor cell mitochondria in addition to being highly expressed in the cytoplasm. Boucherat referred to Hsp90 in mitochondria uniformly as mtHsp90 in his paper [9]. Taken together, we refer to Hsp90 in mitochondria as mtHsp90 for short in our paper, including TRAP1 as well as Hsp90 imported from the cytoplasm.

In this review, our primary focus is on mtHsp90, given its unique subcellular location and potential applications in the development of cancer treatments. In the context of cancer cells, mtHsp90, specifically TRAP1, has the potential to function as either a proto-oncogene or an oncogene (15), though it is commonly observed to be overexpressed in various cancer types, including prostate, breast, lung, and leukemia (16). In addition, we will introduce the basic molecular structure and interaction mechanisms of mtHsp90. Subsequently, we will mainly focus on the application of mtHsp90 inhibitors in cancer treatment.

2 The structure and function mechanisms of mtHsp90

2.1 The structure of mtHsp90

In eukaryotes, Hsp 90 (90 kDa) is a three-domain molecular chaperone protein with a highly conserved biological structure. It contains three domains (Figure 1): an N-terminal domain(NTD) includes an ATP-binding site in a deep pocket on its helical face (17); a middle domain (MD), docking some client proteins and auxiliary protein; and a C-terminal domain(CTD), which contains unique motifs MEEVD or KDEL, depending on its isoform and cellular location and is in charge of binding multiple cochaperones and includes dimerization domain (8, 18). There are also certain inhibitors for each domain. ATP-competitive inhibitors targets NTD, Hsp90-client protein and Hsp90-co-chaperone PPI inhibitors targets MD, while allosteric Hsp90 CTD inhibitors targets CTD (19).

However, in comparison with Hsp90 (or Grp94), the first 59 amino acids in NTD of TRAP1, a mitochondrial targeting sequence at NTD that is cleaved after mitochondrial translocation (20–22); TRAP1 also lacks a highly charged elastic region (CR) between the NTD and MD for regulating chaperone function (23, 24). Among other Hsp90 families, the N-terminal domain contains highly conserved amino acid sequences, such as the MEEVD (Met-Glu-Glu-Val-Asp) sequence, that can interact with other proteins.



interactions with modulators at the end of CTD.

Nevertheless, TRAP1 does not have this sequence, indicating it has no cochaperones (25). Besides, TRAP1 and bacterial chaperone high-temperature protein G (HtpG) can form tetramers as a dimer of dimers, which are related to mitochondrial metabolic regulation (26).

The structure of TRAP1 is not as same as other HSP90 proteins, but the interaction mechanisms with other client proteins are varied. Hsp90 must extensively interplay with cochaperones to coordinate their cycle and recruit clients following the structural changes between open and closed conformations (27, 28). TRAP1 acts as a similar conversed mechanism, but the TRAP1 cochaperones have yet to be reported. Moreover, unlike other Hsp90 proteins, TRAP1 does not need steroid receptors to support signaling protein folding and maturation (28). The amphipathic helix section in Hsp90, which contains E466R, W467R, N470D, M546T, M550A, L553A and F554A, is a client interacting region (29). Furthermore, the clients likely directly interact with open-conformation TRAP1, and the interplay can maintain stably even without ATP contribution (30-32). Therefore, TRAP1 has a certain degree of autonomy. The clients may load into the open-form TRAP1 through a protein-protein way, leading to ATP hydrolysis, changing the conformation, and enhancing the TRAP1 activity (25).

2.2 Mechanism of interaction with client proteins

In comparison to other chaperone protein-requiring Hsp90 (28), TRAP1 is unable to form complexes with known cytoplasmic Hsp90 co-chaperones or to promote maturation of Hsp90 client proteins, suggesting a unique mechanism for TRAP1 (33). The major interactors of TRAP1 are the mitochondrial chaperones mtHSP70 and HSP60 (26). Furthermore, the TRAP1-client interaction itself is unaffected by ATP-mimetic inhibitors that favour the open form and remains stable even in the absence of ATP, indicating that its clients are likely to be loaded by direct interaction with the open conformation TRAP1 (30–32). Mitochondrial contact site and cristae organizing system subunit 60 (MIC60) is a core protein of the mitochondrial cristae, and TRAP1 can directly interact with MIC60 to reduce ubiquitin-dependent degradation in extracellular acidosis. TRAP1 can

directly interact with MIC60 to reduce ubiquitin-dependent degradation in extracellular acidosis, thereby attenuating acidosisinduced mitochondrial and cardiac injury (34). Clients load into the open form of TRAP1 through direct protein-protein interactions, triggering the binding and hydrolysis of ATP, which changes the conformation and enhances the chaperone activity of TRAP1 (25). Therefore, TRAP1 is an autonomous molecular chaperone that functions through interactions with clients and ATP.

Generally, TRAP1 has two different chaperone activities, which are called foldase and holdase. The difference lies in the necessity of ATP hydrolysis in chaperone function (35). Foldase-type activation requires ATP hydrolysis, mainly conformationally remodeling clients and stabilizing, activating, or inactivating them. Eventually may help in the formation of large protein complexes. Holdase-type activity functions independently of ATP hydrolysis and primarily regulates the enzymatic activity of clients and in some cases influences the stability of clients (36). Under normal circumstances, TRAP1 functions in the form of dimers, but recently a tetrametric form of TRAP1 was observed in the response of dysregulated oxidative metabolism (26). It is the most stable stoichiometric TRAP1 complex, the level of which varies with decreasing and increasing OXPHOS (26). The foldase activity of TRAP1 enables it to refold unfolded clients with energy from ATP hydrolysis. Based on broad analyses of the structure of TRAP1 binding its client proteins, it was demonstrated that the unfolded clients' binding surface is usually hydrophobic (37, 38). It was shown that the interaction between TRAP1 and clients is not based on ATP, instead unfolded clients directly bind to a hydrophobic client binding pocket between protomers (30, 31). After clients bind to TRAP1, a conformational transition will be triggered, and TRAP1 turns from an open-form ("apo" state) into a closed state, followed by ATP binding (25). Then an asymmetric homodimeric TRAP1 was formed, in which one protomer was buckled compared to the other. The buckled protomer is better at hydrolyzing ATP, and a subsequent flip in the MD and CTD asymmetry positions the other protomer in the buckled conformation. So, the hydrolysis of two ATP is sequential, and after their hydrolysis, TRAP1 completed the cycle and returned to its symmetric, open state (39). So, in conclusion, it was presumed that the loading of the client triggers the binding of ATP and enhances the ATPase activity of TRAP1 (40). Sequential hydrolysis of two ATPs on TRAP1 helps to remodel the client conformation to release folded or activated clients (25, 41).

To date, the first TRAP1 complex structure with SDHB (B subunit of succinate dehydrogenase) (Figure 2) has been detected (PDB: 7KCM) by cyro-EM (33). In detail, an unfolded SDHB segment interacted with the nonpolar residues located in the TRAP1 MD, such as F531 and F353, and was accommodated into a lumen generated between protomers (25). Actually, F353A and F531A played important roles in the interaction of TRAP1 and other clients, and mutations of these residues would reduce binding and compromise the chaperone functions of TRAP1 (32). Moreover, as a post-translational modification of TRAP1, deletion of TRAP1O-GlcNAcylation impairs TRAP1 binding to ATP and TRAP1 interaction with succinate dehydrogenase (SDHB) (42).

Contrary to the foldase activity, TRAP1 exerts its holdase function in other cases, which means that TRAP1 may not induce client folding, but stabilized clients by binding. In this case, TRAP1 may not need its ATPase activity (26). For example, the major mitochondrial deacetylating enzyme sirtuin-3 (SIRT3) showed increased enzyme activity after binding to TRAP1 in the absence of ATP (30). It is worth noting that SIRT3 binding also increases the ATPase activity of TRAP1, which may subsequently (after the holdase function) operate toward SIRT3 and activate it. Therefore, we can conclude that the holdase and the foldase activities likely operate sequentially or synergetically (25).

2.3 Mechanism of interaction with inhibitors

Tumors are ubiquitously dependent or "addictive" to a heightened protein folding environment (43), which is vital in buffering the proteotoxic stress accompanying in vivo tumor growth (44). Drugs inhibiting or antagonizing mitochondrial Hsp90 and its homolog TRAP1 may have great potential and specificity in treating cancer. Currently, there are several strategies targeting TRAP1. TRAP1 N terminal domain (NTD) inhibition is one of them. NTD is responsible for nucleotide binding and ATPase activity (45). Most inhibitors that have undergone clinical trials as anticancer drugs target the NTD and bind competitively to its ATP binding site to date (46), and representative drugs are Geldanamycin derivatives, guanidine derivatives, and benzamide derivatives. They function by binding the polar residue Asn171 in the middle of the helix of TRAP1 NTD, which locates in the ATP-lid and rotates in to drug binding site (25, 47). Then the foldase activity of TRAP1 is disrupted. Sheperdin is another classical drug targeting TRAP1 by interacting with the ATP-binding pocket in NTD and has cell-penetrating properties (48). However, these competitive inhibitors did not show satisfactory effect in vivo, which may be due to their insufficient concentration in mitochondria (5). Some improvements have been made, like conjugating mitochondrion-targeting moieties with ATPcompetitive inhibitors or increasing mitochondrial permeability (49). SMTIN is one of those drugs. It contains a triphenylphosphonium (TPP) vehicle that targets mitochondria. SMTIN-C10 (a derivative of SMTIN containing C20 linker) was shown to interact strongly with the ATP binding pocket of TRAP1, while its TPP moiety spread out to interact with allosteric sites in the NTD and MD near the ATP binding pocket, enhancing the antitumor activity (31). Electron microscope structural analysis also found that SMTIN-C10 interaction with TRAP1 induced TRAP1 to a tightly packed closed form (31). Another hotspot ATP-mimic inhibitor under research is gamitrinib-TTP (GA mitochondrial matrix inhibitor-TTP), which has shown feasibility and safety in preclinical trials (50), and phase I clinical trial is ongoing(NCT04827810).

As discussed above, the holdase activity of TRAP1 is independent of ATP binding, and client binding was shown to increase ATPase activity (30). Thus, the holdase activity cannot be inhibited by ATP-binding pocket-targeted inhibitors, and these inhibitors become ineffective in blocking the chaperone function



of TRAP1 when it binds to certain client proteins. In these circumstances, the holdase function primarily mediating TRAPclient interaction appears to be essential for the chaperone mechanism. As a result, holdase inhibitors were developed to solve the problem. MitoQ mimics the client and binds to the client-binding site of TRAP1, inhibiting other clients to interact with the chaperone. F353 and F531 residues were shown to participate in the interaction. Although the ATPase activity of TRAP1 is elevated through MitoQ binding, its chaperone function could be fully blocked as other clients were replaced by MitoQ (25).

In addition to the several relatively mature TRAP1 inhibitors mentioned above, many novel compounds have been synthesized and developed with favorable outcomes in recent years. Recently, Merfeld et al. showed a compound 36 that is more than 250 times more selective for TRAP1 than Grp94. In addition, it inhibits OXPHOS, alters cellular metabolism toward glycolysis, destabilizes TRAP1 tetramers, and disrupts mitochondrial membrane potential (51). Kim et al. synthesized a series of pyrazolo[3,4-d] pyrimidine derivatives, in which X-ray diffraction results showed that compounds 47 and 48 interacted with the ATP- binding pocket of TRAP1 protein, and they were shown to exhibit excellent anticancer efficacy in various cancer cells. Moreover, 47 and 48 significantly reduced tumor growth in a mouse PC3 xenograft model (52). Yang et al. developed a series of purine-8one and pyrrolo[2,3d] pyrimidine derivatives based on the TRAP1 structure, of which 5f was 65-fold more selective than Hsp90 α and 13-fold more selective than Grp94, and 6f was 78-fold and 30-fold more selective than Hsp90 α and Grp94, respectively (47). The lead compound synthesized by Sanchez-Martin et al. inhibits TRAP1, but not HSP90 ATPase activity, demonstrating the efficacy of using conformational dynamics to expand the chemical space of molecular chaperone antagonists to TRAP1-specific inhibitors with broad therapeutic opportunities (53).

3 The functions of mtHsp90 in tumor cell

MtHsp90, including Hsp90 and its homologue TRAP1 were abundantly present in the mitochondria of tumor cells (14). TRAP1 performs different functions in different cancer cell types (54). It can act as a proto-oncogene or an oncogene (15). It is highly expressed in hepatocellular carcinoma (HCC) (55), prostate cancer (56), small cell lung cancer (57, 58), nasopharyngeal carcinoma (59), highgrade glioma (60), and breast cancer (61), but it acts as a tumour suppressor in patients with cervical, bladder, and clear cell renal cell carcinomas (62).

TRAP1 is indispensable in the performance of mitochondrial functions and maintenance of structure (Figure 3). It has been suggested that TRAP1 plays an essential role in reprogramming the mitochondrial pathway to critically support tumor growth (63, 64). It interacts with key mitochondrial enzymes (32) and has important roles in shaping mitochondrial function, protecting against apoptosis (14) and oxidative stress (65) to critically support cancer cells. It was also shown to control mitochondrial protein quality (66). Specifically, TRAP1 binds mitochondrial and cytoplasmic ribosomes as well as translation elongation factors and facilitates localized translation in the vicinity of mitochondria (67). Moreover, it has pivotal function in mitochondrial homeostasis and bioenergetics (68). The downregulation or depletion of TRAP1 in cancer cells impairs ATP generation and mitochondrial membrane potential, disturbs mitochondrial function (69), reduce proliferation, and has variable effects on apoptosis (57). Not only dose TRAP1 act on mitochondria, but it also participates in lysosomal-mitochondrial crosstalk to maintain cellular homeostasis, and its activation improves the lysosomal phenotype in the cells of patients with a variety of lysosomal storage disorders, and may be a potential therapeutic target for a wide range of lysosomal diseases (70).

MtHsp90 regulate and activate several cancer-related pathways including metabolic reprogramming (71), tumor cell motility, and

evasion of apoptosis. These effects enhance tumor cells growth in low nutrients environment and metastatic dissemination to other body parts, also arm them with drug resistance. MtHsp90 was suspected to block drug-induced apoptosis in several tumors (57), conferring tumor cells resistance against chemotherapeutic agents (72).

One of the major functions of mtHsp90 is that it played essential role in tumor cells metabolism reprogramming, which is an important driver of tumor adaptation, generation, and progression. One of the hallmarks of tumor cells in energy metabolism is aerobic glycolysis, which is also called Warburg effect. MtHsp90 is important in this pathway. TRAP1 was suggested to inhibit succinate dehydrogenase (SDH) activity to favor aerobic glycolysis (Warburg effect) and inhibit oxidative phosphorylation (OXPHOS) in human colon, cervix, ovary and bone cancers (62, 73). Sciacovelli et al. (74) demonstrated that TRAP1 binds to and inhibits respiratory chain complex II succinate dehydrogenase (SDH). The respiratory downregulation induced by the interaction of TRAP1 with SDH promotes tumorigenesis by initiating the stabilization of the succinate-dependent oncogenic transcription factor HIF1a. Current TRAP1 inhibitors targeting this pathway have achieved promising results in human cells. The study by Sanchez-Martin et al. demonstrated that honokiol DCA (HDCA) (a small molecule antitumor agent) binds to a variant site in TRAP1 and is able to inhibit TRAP1 but not Hsp90 ATPase activity (75). In Human plexiform neurofibroma 95.6 cells, HDCA restored TRAP1-dependent SDH down-regulation, decreased proliferation rate, increased mitochondrial superoxide levels and eliminated tumorigenic growth (75). Despite the facts mentioned



FIGURE 3

The functions of TRAP1 in tumor cells. TRAP1 binds to and inhibits the activity of complex II succinate dehydrogenase (SDH), complex III core component UQCRC2; enhances cellular utilization of glutamine via HIF-2 α ; promotes tumorigenesis by stabilizing HIF-1 α via succinate; facilitates tumor growth and maintenance through stimulation of the TNF, Wnt pathway; promoting tumor metastasis by activating FAK and inhibiting AMPK and ULK1; modulating apoptosis and autophagy by regulating the release of cytochrome c (Cyt C) and the binding of cyclophilin (CypD) to ATP synthase; blocking ROS generation to prevent cellular damage from oxidative stress. Created with BioRender.com.

above, a recent study demonstrated that TRAP1 binds the core component of complex III, UQCRC2, within the mitochondria and regulates complex III activity. This reduces respiration rates under basal conditions, but maintains sustained oxidative phosphorylation during glucose limitation (76). Moreover, TRAP1 interacts with PINK1(*PTEN induced putative kinase 1*) to protect cells from oxidative stress-induced cell death (77).

But it was verified to be insufficient to explain the complexity of metabolic reprogramming in tumor cells (78, 79). There must be other metabolism pathways to support high energy demanding of tumors, such as metastasis and cell proliferation, and that's where mtHsp90 also come into play (68). Under unfavorable nutrient conditions such as nutrient deprivation or amino acid shortage, tumor cells are able to utilized mtHsp90-directed and TRAP1 directed protein folding (14) to maintain ATP production (6), thus attenuating the activation of the nutrient-sensing, tumorsuppressive AMPK pathway (80) and tumor cell autophagy pathway (6). Tumor cells are conferred with a survival and proliferation advantages and their progression and invasion under nutrient deprivation was thus possible then (71). Overexpression of TRAP1 in cancer associated fibroblasts (CAFs) increases basal oxygen consumption rate (OCR) and ATP production. In vivo, overexpression of TRAP1 in CAFs of HSC3 cell xenografts model inhibited tumor growth (81). The mechanisms behind maintained ATP production were the retention of hexokinase II to the mitochondrial outer membrane by regulating cyclophilin D (CypD) folding (6), keeping the stability of oxidative phosphorylation complex II subunit (79) and succinate dehydrogenase. In conclusion, mtHsp90 mediated energetics can maintain a residual level of oxidative phosphorylation to block the activation of tumor-suppressive AMPK signaling and autophagy (6), which are considered vital for tumor progression and drug resistance in vivo. Although the promotion effects of TRAP1 in Warburg effect and oxidative phosphorylation (i.e. inhibit or activate SDH) in cancers seemed to be opposing, it was just a result of different metabolic plasticity under different tumor environments (25). Furthermore, it has been suggested that TRAP1 is essential for malignant transformation of cells but is dispensable in later stages of tumour development (30, 62). Although the function of TRAP1 is controversial, the majority of the literature suggests that TRAP1 is overexpressed in many cancers and regulates metabolic transformation during tumourigenesis, and that TRAP1 attenuation is detrimental to tumor cell survival (16, 61, 82-85). The ultimate function of TRAP1 in pathology is still tumorigenesis. The cellular contexts affecting these contradictory effects are still poorly understood, but upstream regulators and clients of TRAP1 may be one of them, such as the bi-directional interaction between SIRT3 (a NAD⁺-dependent deacetylating enzyme) and TRAP1 (25).

MtHsp90 also promote tumor cell motility and metastasis *in vivo*, especially in case of scarce nutrients. That may explain why these chaperones are ubiquitously overexpressed in advanced tumors. Recent research has found that TRAP1 overexpression (TRAP1 OE) promotes mitochondrial fission, enhanced *in vitro* migration and *in vivo* metastasis of tumor cells, and altered cellular homing properties (86). In a study (71), researchers use

noncytotoxic concentrations of gamitrinib to antagonize the function of Hsp90 in mitochondria. The result showed a nearly complete suppression of invasive length and areas of tumor cells. Next, they use pooled siRNAs to silence TRAP1, the same effect was reproduced, demonstrating mtHsp90 's importance in tumor cell motility. Through further investigation, they found that tumor cell motility was also influenced by cell motility kinases, tumor cell bioenergetics and tumor cell autophagy. These pathways were regulated by mtHsp90 to preserve cytoskeletal dynamics through continuous phosphorylation of multiple cell motility kinases and releasing FAK from inhibition by ULK1-FIP200 autophagyinitiating complex (87). All the above contribute to tumor cell invasion and dissemination, leading to shorted OS in patients. On the contrary, activation of AMPK, ULK1 or autophagy regulators suppressed tumor progression and restricted motility (71). In conclusion, mtHsp90 regulate the crosstalk between bioenergetics stress, AMPK activation and autophagy to preserve tumor cell motility.

MtHsp90 also prevents autophagy, another tumor-suppression mechanism, in cancer cells during nutrient deprivation (71, 88) to preserve tumor cell motility. Inhibition of TRAP1 in tumor cells caused marked increase in lysosome content and autophagy activity (69).

Cancer cells' evasion of apoptosis is mediated partly by mtHsp90, too. TRAP1 improve cancer cells' resistance to various stresses including anticancer therapies by upregulating cell death threshold (89, 90). TRAP1 can protect mitochondria against reactive oxygen species (ROS) and oxidative stress (64) by blocking ROS production (91)(e.g. suppress SDH activity) or scavenging noxious ROS (92)(e.g. activate superoxide dismutase 2 through SIRT3 (30)). There are also other anti-death mechanisms aside from scavenging ROS. For example, it inhibited the release of cvtochrome c (64). It also antagonized cvclophilin D (CvpD) (93) to inhibit the opening of CypD-dependent mitochondrial permeability transition pores (mPTP) (94, 95), and mPTP opening is directly related to cell death. Upon cellular stresses, CypD was shown to irreversibly open mPTP by changing the conformation of complex V, which is one of the components of mPTP (96). So TRAP1 inhibit CypD and preserve the integrity and membrane potential of mitochondria, which is a vital survival mechanism for tumor cells (63). TRAP1 also ameliorates diabetes-induced kidney injury by preventing abnormal opening of mPTP and maintaining mitochondrial structure and function (97).MtHsp90 was reported to interact with CypD to antagonize mitochondrial permeability transition process, too (14). It was also showed to be associated with enhanced DNA repair when patients were treated with cisplatin (98). A study by Zhang et al. in 2021 demonstrated that TRAP1 inhibits cisplatin-induced apoptosis by promoting ROS-dependent mitochondrial dysfunction (99). Combined low activity of DNase1 and Trap1 results in impaired chromatin degradation in vitro, delayed chromatin clearance in vivo, and enhanced immune cell activation (100). Expression of TRAP1 and HSPD1 correlates with DNA replication and mitotic inhibitor sensitivity (101).

However, cancer cells differ from normal cells in many aspects, including altered expression levels and functional states of mitochondrial Hsp90 and TRAP1 (102, 103). Many cancer cells

express higher levels of mitochondrial Hsp90 and TRAP1, upregulate the associated signaling pathways and depend on them for survival, (e.g. TRAP1 promotes neoplastic growth through inhibiting succinate dehydrogenase (104)) making them vulnerable to drugs that inhibit these molecules, normal cells have lower expression levels of mtHsp90, thus less susceptible to drugs targeting mtHsp90. Additionally, some targeted drugs have good tissue distribution and pharmacokinetic properties, which allow them to enter tumor tissues and exert their effects more easily (105). MtHsp90 inhibitors have high affinity and specificity for cancer cells, and the reason may lie in the unique chaperone level, localization (69) and metabolism of cancer cells compared to normal cells. Tumor cells rely far more on Hsp90 than normal cells, leading to their vulnerability to Hsp90 inhibitors. TRAP1 is even more special. It functions only in response to cellular stress instead of maintains housekeeping protein homeostasis, and when this stress-adaptive machinery goes dysregulated, human diseases, such as cancer, may arise (25). TRAP1 is highly expressed in tumors such as glioblastoma, colon, breast, prostate and lung cancers (106), but is low or even undetectable in normal cells (64). As a result, TRAP1 deletion was shown to have no or minimal effect on normal development of mice (68), but can effectively inhibit growth of tumor cells. In another study, TRAP1-depleted tumor cells exhibited decreased cell viability, but the same negative impact of TRAP1 depletion was not observed in normal cells (69). For example, glutamine metabolism is dysregulated in various types of cancers, which are termed "glutamine auxotrophic cancer". They became to rely mainly on glutamine-dependent metabolism for ATP production, and the process was regulated by TRAP1. Dharaskar et al. in a recent research demonstrated that in tumor cells TRAP1 maintains mitochondrial integrity during glucose deprivation and enhances cellular utilization of glutamine for cellular energy requirements via the HIF2\alpha-SLC1A5-GLS axis (107). So TRAP1 inhibition will only influence these addictive cells, posing no threats to normal cells, i.e. glutamineindependent cells (108). Therefore, cancer cells show differential sensitivity to Hsp90 inhibition based on drug metabolism compared with normal cells (109).

In addition to its significant role in the development, metastasis, and maintenance of cancer, aberrant expression of TRAP1 has been associated with a number of diseases, including but not limited to the following: TRAP1 overexpression protects motor neurons from mitochondrial dysfunction and death under conditions of oxidative stress caused by amyotrophic lateral sclerosis (ALS) (110). Ramos Rego et al. describe the role of TRAP1 in CNS cells under physiological and pathological conditions (111). TRAP1 regulates hypoxia-induced cardiomyocyte apoptosis through the COXIImediated mitochondria-dependent apoptotic pathway (112). And in the human renal proximal tubular epithelial cell line HK2, Lin et al. found that TRAP1 is important for the maintenance of mitochondrial function in HK2 cells under pathological conditions and the activation of TRAP1 may be useful in the treatment of renal fibrosis (113).

4 MtHsp90 inhibitors in cancers

Compared to other Hsp90 inhibitors, many new small molecule drugs that target TRAP1 or mitochondria show better performance in controlling cancer cells and reducing cellular toxicity. Extensive safety and effectiveness tests *in vitro* and *in vivo* have been conducted, further demonstrating the potential of mtHsp90 inhibitors in cancer treatment.

4.1 Lung cancer

Lung cancer is one of the most common types of cancer and is responsible for the highest number of cancer-related deaths worldwide. It is a malignant tumor that starts in the cells of the lungs and can quickly spread to other parts of the body. The nonsmall cell lung cancer (NSCLC) is the most common type of lung cancer, accounting for approximately 85% of all cases. The advent of tyrosine kinase inhibitors (TKIs) has profoundly impacted the clinical outcomes of patients with NSCLC that exhibit EGFR-activating mutations, which is one of the most common contributing factors in NSCLC (114, 115). However, despite the initial success of EGFR TKIs, resistance eventually develops in many patients (116). Moreover, patients with NSCLC who harbor other mutation types, such as KRAS mutation, still lack effective treatment options and are more susceptible to developing resistance for current therapies (117). Drug resistance always arises from the evolutionary pressure exerted on NSCLC cancer cells by TKIs, which leads to insensitivity and failure in clinical treatment. Therefore, to induce this drug resistance of TKIs, many different methods have been tried and some of them showed a prospective result for NSCLC control, such as the combined EGFR/Hsp90 inhibition (118).

This special advantage of mtHsp90 inhibitors has been evaluated in many studies and demonstrated its ability to control the development of NSCLC. Tumors collected from Gamitinibtreated animals showed extensive apoptosis in situ and release of cytochrome C in the cytoplasm, while organs collected from nontumor regions showed no significant changes in histology, no change in overall structure, and no evidence of inflammation or hepatic steatosis (119). In another study focused on NSCLC, G-TPP has been shown to increase glutamine synthetase activity and induce cell death in glutamine-dependent NSCLC cells by causing an energy shortage, as evidenced by the phosphorylation of AMPactivated protein kinase (AMPK) (108). Furthermore, due to the specific mitochondrial localisation of TRAP1, TRAP1 selectivity at the cellular level can be achieved by attaching mitochondriatargeted portions to pan Hsp90 inhibitors. Gamitrinibtriphenylphosphonium (Gamitrinib-TTP) and SMTIN-P01 were demonstrated to have cytotoxic effects associated with mitochondrial membrane depolarisation in the lung adenocarcinoma cell line H460 (5, 119). And they also had no effect on Hsp90 client protein levels, suggesting its TRAP1 selectivity. Moreover, Hsp90 inhibitor-mitochondria targeting indocyanine dye conjugate (IR-PU) has high apoptosis-mediated cytotoxicity in the NCI-H460 cancer cell line (120).

Preclinical studies in cell lines and animal models have shown promising results for the use of mtHsp90 inhibitors in NSCLC treatment.

These findings also suggest that combining mtHsp90 inhibitors with other drugs targeting tumor-related pathways may be an option to improve treatment outcomes for NSCLC patients in the future. Recently, gamitrinib-TPP has successfully completed preclinical evaluation and is currently in a Phase I clinical trial in advanced cancer. This study was last updated on February 8, 2023, and it is estimated to be completed initially in December 2028 (ClinicalTrials.gov NCT04827810) (121). Additionally, A recently published article reveals the mechanism of TRAP1-associated cancer treatment with CVM-1118 (foslinanib), a phosphoric ester compound selected from 2phenyl-4-quinolone derivatives. In the NSCLC cell line A549, CVM-1125 reduces cytosolic succinate levels and causes destabilization of HIF-1 α by targeting TRAP1, thereby blocking its downstream signaling and ultimately inducing mitochondrial apoptosis, inhibiting tumor cell growth, and suppressing angiogenic mimicry (122). The drug has completed clinical Phase I (NCT04336124) and is currently recruiting for clinical Phase II (NCT05257590), with an expected primary completion date of September 2025.

4.2 Prostate cancer

Prostate cancer is a prevalent cancer type among men, with over 1.4 million cases diagnosed worldwide annually, which is the second most common solid tumor in men and the fifth cause of cancer mortality over the world (123, 124). It originates in the prostate gland, a small walnut-shaped organ responsible for the production of seminal fluid in males. In its initial stages, prostate cancer frequently manifests no symptoms, but as the cancer progresses and metastasizes, it can cause a range of symptoms, including urinary difficulty, dysuria, hematuria, hematospermia, and pain in the lower back, hips, or thighs (125). Lisanti et al. (82) demonstrated that TRAP1 increases cell proliferation, decreases apoptosis, and promotes cell invasion in prostate cancer without changes in mitochondrial bioenergetics through a common altered Pten +/- context in human prostate cancer.

In the prostate tumor, the mtHsp90 homologous TRAP1 is substantially and differentially expressed in human local and metastatic prostate cancer. In contrast, TRAP-1 is largely undetectable or poorly expressed in normal prostate (56). The treatment of Gamitinib can result in the rapid and complete killing of androgen-dependent or non-androgen-dependent prostate cancer cell types without affecting untransformed prostate epithelial cells. With the loss of organelle membrane potential, release of cytochrome c, and caspase activity, gamitrinib can induce acute mitochondrial dysfunction and then lead to apoptosis in prostate cancer cells (126). In the study of Kang et al., the systemic administration of gamitrinib-G4 was shown inhibition effects on the development of tumor cells in mice with localised and metastatic prostate cancer (127). Furthermore, they found no significant animal weight loss or organ toxicity was observed in response to the continuous gamitrinib-G4 treatment, which confirmed the safety and feasibility of the mtHsp90 inhibitors *in vivo*.

While the results of these preclinical studies are promising, there is still much more research that needs to be done to fully evaluate the potential of mtHsp90 inhibitors for the treatment of prostate cancer in the future. One of the biggest challenges is identifying the most appropriate patient populations and treatment regimens for these inhibitors. Moreover, it is necessary to carefully evaluate the potential adverse events of mtHsp90 inhibitors in humans, as there still is a lack of clinical trial results. However, despite these challenges, the development of mtHsp90 inhibitors represents an exciting new approach to the treatment of prostate cancer in the future. By targeting a critical protein involved in the survival and growth of prostate cancer cells, these inhibitors have the potential to offer a new and effective treatment option for patients with advanced prostate cancer.

4.3 Glioblastoma

Glioblastoma is an aggressive form of brain cancer that arises from the glial cells that support the neurons in the brain. It is the most common primary brain tumor in adults, accounting for approximately half of all malignant brain tumors (128). Despite recent advances in multimodal therapy for glioblastoma, including surgery, radiation therapy, and immunotherapy, the overall prognosis remains poor, with a 5-year survival rate of less than 5% and a median survival of fewer than 2 years, and there is currently no effective drug to prolong median overall survival or control recurrence (129, 130). However, the use of mtHsp90 inhibitors may be one promising area of research in glioblastoma, as it will influence lots of tumor-related pathways.

TRAP1 is a major chaperone of the respiratory complex of the electron transport chain, therefore interfering with TRAP1 disintegrates oxidative phosphorylation (131). Besides, TRAP1 is upregulated in Glioblastoma multiforme (GBM) compared with normal brain cells. The high expression of TRAP1 can affect the cell glycolysis in the tumor microenvironment (132). TRAP1 might take a role in maintaining the stemness of glioblastoma stem cells. TRAP1 protects mitochondrial integrity and prevents apoptosis (133), thus induces the resistance to temozolomide (TMZ), the standard chemotherapy drug for glioma. Therefore, TRAP1 is a promising target for drug design in glioblastoma therapy (134).

Gamitrinib has been reported in glioblastoma that through suppressing TRAP1, gamitrinib can sensitize the GBM cells to temozolomide treatment (134). It inhibits cell proliferation and induces apoptosis and death in 17 primary glioma cell lines, 6 TMZresistant glioma cell lines, 4 neurospheres and 3 PDOs (135).Another research showed that by inducing mtUPR and the subsequent ROS burst, TRAP1 function was inhibited and GBM cells were sensitised to TMZ lysis after gamitrinib treatment (136). Moreover, in cell line–derived xenografts and patient-derived xenografts models implanted with subcutaneous or intracranial tumors, gamitrinib could also significantly delay tumor growth and increased mouse survival. Through integrated analysis of RNAseq and RPPA data, they revealed that gamitrinib exhibited anti-tumor activity mainly by (i) inhibiting mitochondrial biosynthesis, OXPHOS, and cell cycle progression, and (ii) activating energy-sensing AMP-activated protein kinase, DNA damage, and stress response. These preclinical findings provide robust and convincing evidence that supports the potential therapeutic efficacy of gamitrinib in the management and treatment of neuroglial tumors.

Combination therapy has also been studied in glioblastoma. Combining LXR agonist and TRAP1 inhibitor gamitrinib-TPP resulted in increased levels of Bcl-2 family protein expression and higher rates of cell death (136). In addition, the combined use of both gamitrinib and histone deacetylases (HDAC1/2) inhibitors (e.g., Romidepsin or Panobinostat) can further reduce tumor growth in model systems of glioblastoma (137). In murine model systems of patient-derived orthotopic xenografts of human glioblastoma (PDX), the combination of BH3-mimetics and gamitrinib-TPP blunted cellular proliferation in a synergistic manner by massive activation of intrinsic apoptosis (138). These preclinical findings indicate the potential of gamitrinib in treating glioblastoma.

4.4 Breast cancer

Breast cancer is a complex and heterogeneous disease that affects millions of women worldwide. Breast cancer is estimated to be the most common cancer overall and the top 2 cause of death in women (123). It is a type of cancer that begins in the breast tissue and can spread to other parts of the body. Despite the relatively favorable prognosis and advances in the treatment of breast cancer, there is still a need for more effective and targeted therapies for different cancer subtypes. The use of mtHsp90 inhibitors in the treatment of breast cancer may be one of the promising ways.

Liu et al. suggested that TRAP1 expression promotes cell proliferation and tumor growth through the TNF pathway, while its downregulation may lead to reduced proliferation and increased metastatic potential (139). An Aberrant upregulation of TRAP1 has been reported in the tumorigenesis of breast cancer (61). Different from other carsinoma, it has nothing to do with the proliferative capacity of cancer cell (140). Instead, TRAP1 modulates mitochondrial dynamics and function, and links these processes to the tumorigenesis of breast cancer. When TRAP1 is overexpressed, the mitochondria will be less fragmented and more tubular network-shaped and the mitochondrial aerobic respiratory will be upregulated. This suggests that TRAP1 may be a potential target for breast cancer therapy (61).

Several mtHsp90 inhibitors have been developed and shown to have promising activity in preclinical models of breast cancer. For example, gamitrinib has been shown to induce apoptosis in breast cancer cell lines and to inhibit the growth and spread of tumors in mouse models of the disease (119). Gamitrinib-TPP, an inhibitor targeting mtHsp90, increases cell death in MCF7 cells (141). Similarly, SMTIN-P01 has been shown to selectively accumulate in the mitochondria of breast cancer cells and induce cell death (5). However, although mtHsp90 inhibitors demonstrate significant advantages in controlling the development of breast cancer cells and tolerating side effects, there are still some limitations with the tests in other types of cancer. Further research is still needed to ensure the feasibility of these inhibitors before human testing.

4.5 Gastric cancer

In 2020, 769,000 people died from gastric cancer (GC) worldwide (123), ranking as the fourth most common cause of cancer-related deaths. Targeted drugs for gastric cancer have been developed, for instance, Trastuzumab, a monoclonal antibody against HER2. When combined with chemotherapy, it can improve the survival rate of advanced gastric cancer, but the resistance remains to be tackled (142).

Hsp90 plays an important role in gastric carcinogenesis by activating downstream client proteins, including RAF, KIT, EGFR, HER2, etc. (143).TRAP1 is an important member of mtHsp90 that inhibits the survival of reactive oxygen species ROS, thus protecting cells from mitochondrial apoptotic mechanisms and guaranteeing a sustained proliferative state (144, 145).

A study by Ping Han et al. (146). showed that the mRNA and protein expression levels of TRAP1 were significantly higher in cancer tissues than in adjacent normal tissues. In addition, TRAP1 may regulate the malignant biology of cells by increasing the expression of CyclinB1, CyclinD1, CyclinE, MMP-2, and VEGF, leading to the development and progression of gastric cancer, which is an important target for targeted therapy. H. pylori vacuolating cytotoxin A (vacA) is involved in the regulation of apoptosis in human gastric epithelial cells by inducing down-regulation of TRAP1 via the P38MAPK pathway (147). In esophageal cancer (EC) TE-1 cells, shikonin promotes apoptosis and attenuates migration and invasion through inhibition of TRAP1 expression and AKT/mTOR signaling, suggesting that shikonin may be a novel drug for the treatment of EC (148).

Nevertheless, no more targeted drugs are developed for TRAP1 in gastric cancer until now, and the therapeutic field for gastric cancer targeting mtHsp90 remains blank.

4.6 Colorectal cancer

Bowel cancer consists of large bowel cancer and small bowel cancer. Most bowel cancer begins in the colon (149). The colon and rectum make up the large intestine. Cancer that starts here is called colorectal cancer (CRC). Colorectal cancer is the second and the third most commonly occurring cancer in women and men, respectively (150), and one of the most common causes of cancer deaths (151).

According to the investigation, in colorectal cancer samples, Hsp90 expression is highly increased compared with normal epithelial tissues. Hsp90 might take a pathological role in colorectal cancer *in vivo* (103). Particularly, in colorectal cancer, over-expression of TRAP1 might encourage tumor cell invasion (152). TRAP1 is up-regulated in ulcerative colitis associated colorectal cancer (102). TRAP1 has been suggested as a predictive marker for prognosis in colorectal cancer (153), human metastatic colorectal carcinoma (104) and ulcerative colitis-associated colorectal cancer (102).

TRAP1 is possibly involved in the regulation of colorectal cancer through a variety of mechanisms. TRAP1 regulates stemness through Wnt/β-catenin pathway in human colorectal carcinoma (154). Upregulated in 60-70% of human colorectal cancers (CRC), TRAP1 regulates the Wnt/β-Catenin pathway and prevents β-Catenin phosphorylation/degradation by modulating the Wnt ligand receptors LRP5 and LRP6, which facilitates stem cell maintenance (155). Condelli report that TRAP1 can participate in the progression of colorectal cancer through regulating the synthesis and ubiquitination of BRAF (156). In human colorectal cancer cells and tissues, with high TRAP1 background, the protein expression and phosphorylation of p70S6K is reduced (157). When colorectal cancer is under oxygen deprivation, TRAP1 regulates the response of cells to hypoxia and inhibits ribosome biogenesis (158). It is involved in regulating hypoxia-induced HIF-1 α stabilization and glycolytic metabolism. Moreover, glucose transporter protein expression, glucose uptake and lactate production were partially impaired in TRAP1-silenced CRC cells under hypoxic conditions (158). TRAP1 also cooperate with soluble resistance-related calcium-binding protein (sorcin) in human colorectal carcinoma in a survival pathway, which is responsible for inducing multi-drug resistance (72).

For patients with Hsp90-positive rectal cancer, the application of suitable Hsp90 inhibitors would be highly beneficial (103). Since broad spectrum inhibitors of Hsp90 family have demonstrated negative efficacy on TRAP1 inhibitory effect due to its poor mitochondrial permeability, researchers have paid attention to TRAP1 isoform-selective inhibitors. Shepherdin is the first peptidomimetic with the ability to permeate into the mitochondria and target TRAP1 (105). Novel compounds, for example mitochondrial permeating SMTIN-P01 (64) and the most selective TRAP1 inhibitor DN401 (5) are also developed.

Clinically, Hsp90 inhibitors are often combined with chemotherapy to treat with CRC. A previous study revealed that after combined use of AUY-922 with 5-FU, CRC cells exhibited a lower multi-drug resistance (159). Targeting TRAP1 by gamitrinib induces BRAF-driven apoptosis and inhibits colony formation in CRC cells (160). Gamitrinib-TPP, inhibiting TRAP1 signaling pathways in colon cancer, can disrupt redox homeostasis and induce cell death. Under oxidative stress, inhibition of TRAP1 by gamitrinib-TPP induced metabolic reprogramming and heat shock factor 1 (HSF1)-dependent transactivation, with enhanced induction of DNA damage and cell death (161). Combining with BH3-mimetics, gamitrinib-TPP actives intrinsic apoptosis (138). Moreover, combination of LXR agonists and gamitrinib-TPP induces elevation of pro-apoptotic Bcl-2 family and Noxa in HCT116 cells (136). However, colon cancers can induce variable ER stress responses and ROS accumulation to resist gamitrinib-TPP treatment. Tsai report that treatment with both an NRF2 inhibitor and a TRAP1 inhibitor may potentially overcome colon cancer resistance by raising cellular ROS level (162).

Undoubtedly, targeted drugs are becoming more and more important in the field of clinical oncology. However, considering the cytotoxicity and induced multi-drug resistance, there is still a long way to go before drugs targeting mtHsp90 can take a role in the therapy of colorectal cancer.

4.7 Liver cancer

Liver cancer is the third leading cause of cancer-related deaths worldwide. Hepatocellular carcinoma (HCC) is the most common liver cancer, accounting for 90% of liver cancer. It is predicted that 1.3 million people could die from liver cancer in 2040. Traditional treatment methods for liver cancer mainly include: surgery, liver transplantation, ablation therapy, interventional therapy, radiation therapy, and chemotherapy. The new therapies can be mainly divided into the following three categories: targeted drug therapy, immunotherapy, combined treatment strategies, etc. (163).

The study found that the high expression of Heat Shock Protein-90 (Hsp90) was related to the low survival rate of hepatocellular carcinoma (164). Experiments have found that Hsp90 is highly expressed in liver cancer patients (165–168). TRAP1 can regulate mitochondrial integrity, prevent oxidative stress and inhibit cell death (169). Ramos Rego et al. examined the TRAP1 interactome using the tandem affinity purification system and identified 255 unique proteins, which can regulate a variety of cellular processes, including energy metabolism, suggesting that TRAP1 maintains mitochondrial integrity in addition to metabolic rewiring (170).

It is possible that the expression of TRAP1 is associated with autophagy in liver cancer; HepG2 cells exhibited the highest basal level of autophagy and TRAP1 expression with medium invasive ability. Moreover, hepatitis B (HBV) infection of HepG2 cells suppressed autophagy activity and the expression of TRAP1. Treatment with rapamycin also greatly increased autophagy in the 4 liver cancer cell lines and enhanced the expression of TRAP1 in HepG2, Hep3B2.1-7 and Sk-hep1 cells. Therefore, TRAP1 may be related to autophagy in liver cancer, as cell invasiveness, HBV infection and autophagy induction have different effects on TRAP1 expression (171). Another study showed that S-nitrosylation of the mitochondrial chaperone TRAP1 sensitises hepatocellular carcinoma cells to succinate dehydrogenase inhibitors. Chromosomal deletion of GSNOR leads to pathological protein Snitrosylation implicated in human hepatocellular carcinoma (HCC). This study identifies and exploits a metabolic hallmark of aberrant Snitrosylation in HCC, demonstrate that GSNOR deficiency in hepatocytes is characterised by mitochondrial alterations and marked increases in levels and activity of succinate dehydrogenase (SDH). This is dependent on selective S-nitrosylation of Cys501 in the mitochondrial chaperone TRAP1, mediating its degradation. As a consequence, cells and tumours which are GSNOR-deficient are extremely sensitive to SDH inhibition, namely to α -tocopheryl succinate, a molecular targeting SDH, which induced RIP1/ PARP1-mediated necroptosis and inhibited the growth of the tumour (171).

4.8 Leukemia

Leukemia refers to a set of heterogenous hematological malignancies, including acute myeloid leukemia (AML), acute lymphoblastic leukemia (ALL), chronic myeloid leukemia (CML) and chronic lymphocytic leukemia (CLL) (172). It is characterized by the rapid growth and accumulation of abnormal, immature white blood cells (173), which consequently interfere with the production of normal blood cells (174). Treatment may involve chemotherapy, radiotherapy, stem cell transplantation, targeted therapy (175), or a combination of these approaches (176).

Studies have found that Hsp90 is overexpressed in leukemia and its expression has been linked to poor prognosis (177). In particular, TRAP1 is prominently upregulated in pediatric AML patients according to bioinformatics analysis of public databases (178).

The overexpression of TRAP1 can promote neoplastic growth through a variety of mechanisms, including inhibiting succinate dehydrogenase (104) and reducing ROS (179). Therefore, targeting mtHsp90 may be a potential strategy for treating leukemia.

Currently, some drugs targeting at mtHsp90 have been developed. Gamitrinibs (GA mitochondrial matrix inhibitor) is a class of small molecule compound that selectively inhibit the activity of mtHsp90, including gamitrinib-G1–G4 and gamitrinib-TPP (119). Gamitrinibs inhibit AML cells both *in vitro* and *in vivo* preclinical trials. With systemic administration of gamitrinib-G4 to mice, the growth of established human leukemia is inhibited, The lead compound gamitrinib-TPP also shows significant anti-cancer activity against hematologic malignancies (119). Recently, Mathieu et al. designed and synthesized a series of 6BrCaQ-Cn-TPP conjugates as a novel class of TRAP1 inhibitors, among which compound 3a exhibited excellent antiproliferative activity in a variety of cancer cell lines, including human leukemia cells K562 (180).

Overall, there have been several drugs that specifically inhibit mtHsp90. These drugs have shown promising results in leukemia in preclinical studies. However, more research is in need to fully understand the role of these drugs in leukemia and to determine their efficacy and safety.

4.9 Pancreatic cancer

Pancreatic cancer (PC) affects the pancreas, a glandular organ in the abdomen behind the stomach, which plays an essential role in digestion and regulating blood sugar levels. Pancreatic cancer occurs when cells in the pancreas start to grow and divide uncontrollably, forming a tumor. Pancreatic cancer can be challenging to diagnose in its early stages, as symptoms may not appear until it spreads to other body parts. Treatment for pancreatic cancer may include surgery, radiation therapy, chemotherapy, or a combination of these therapies, depending on the stage and location of cancer. The prognosis for pancreatic cancer can be poor, mainly if the cancer has spread to other body parts.

TRAP1 can upgrade the tumor cell death threshold and give them resistance to antineoplastic therapy by scavenging harmful ROS (89, 90, 181, 182) and suppressing mitochondrial permeability transition pore (mPTP) cyclophilin D (CypD; PPIF), which is en essential survival mechanism (14).

Hsp90 inhibitors may enhance pancreatic cancer cells' cytotoxic sensitivity by causing client protein degradation (183). Lang et al. reported that blocking Hsp90 can cause growth-inhibitory by obstructing insulin-like growth factor-I (IGF-I) and interleukin-6 (IL-6), proangiogenic signaling cascades (184). Besides, IPI-504, an Hsp90 inhibitor, showed an anti-proliferative effect on PC growth (185). Moreover, Xianhua Cao introduced a combination therapy of geldanamycin and 3BrPA that enhances efficacy and reduces doselimitation toxicity to treat chemotherapy-resistant PC (186, 187). Tarik Ghadban et al. demonstrated that Hsp90 inhibitors (17-AAG and 17-DMAG) could disrupt gemcitabine and 5- fluorouracil signal cascades in PC, which can promote tumor cell apoptosis (188). These studies showed promising prospects, but few clinical trials assessed them. To date, only one phrase II trial reported that it is not warranted to study targeting Hsp90 inhibiting drugs with gemcitabine in PC treatment (183).

Though there is not enough targeting TRAP1 inhibitors for PC treatment, the ATPase in NTD of other Hsp90 inhibitors have similar structure with TRAP1, one has proved that these Hsp90 inhibitors can competitively inactive TRAP1 ATPase *in vitro* (189), which can be used to develop TRAP1 selective inhibitors in future.

5 Discussion

Despite the increasing maturity of our understanding of mtHsp90, there are still unresolved concerns that require further investigation. Firstly, it is imperative to evaluate the potential relevance of ATPase activity to the function of TRAP1. That's because previous studies have demonstrated that TRAP1, even in a catalytically inactive state, is capable of carrying out its role and reversing mitochondrial dysfunction (26). Secondly, as for how mtHsp90 regulate tumorigenesis and therapy resistance of various cancers at the molecular level, our cognition is still limited. In the future, we may focus more on the molecular functional mechanisms and related signally pathways with mtHsp90. Furthermore, given the lack of clinical success observed in numerous cytosolic Hsp90 inhibitors, it may be prudent to explore the possibility of devising a strategy to concurrently disrupt the mtHsp90 pool in order to evaluate their combined efficacy. MtHsp90 inhibitors have demonstrated the ability to effectively control cancer cell growth both in vitro and in vivo. There are several mtHsp90 inhibitors, such as gamitrinib, SMTIN-P01, SMTIN-C10, and DN401, that have been successfully designed to target mitochondria (Table 1) (5, 31, 119, 191). We have reviewed the research progress of mtHSP90 inhibitors for the treatment of relevant cancers according to the cancer types, which will be helpful for the development of therapeutic drugs for different cancers. Meanwhile, the synthesis and exploration of new Hsp90 drugs are also ongoing, as explained in the reviews by Dernovšek et al. (19) and Xie et al. (85). Moreover, mtHsp90 inhibitors have the potential to selectively accumulate in the mitochondria of cancer cells and limit toxicity to normal tissues. This effectively addresses the primary challenge of other

Cancer type	Cancer cell line <i>in vitro</i>	Animal models	Inhibitors tested	Ref.
Prostate cancer	DU145		gamitrinib	(119)
	22Rv1 and PC3	22Rv1 xenograf	gamitrinib	(90)
	PC3	PC3 Superficial prostate cancer xenograft model, PC3 Orthotopic bone metastatic prostate cancer model	gamitrinib	(126)
	RM1	TRAMP mice	gamitrinib	(127)
	PC3		gamitrinib	(190)
	PC3, LNCaP	TRAMP mice	gamitrinib	(6)
	22Rv1 and PC3		SMTIN-P01	(5)
	PC3		DN401	(191)
	PC3	PC3 xenograft Model	SMTIN-C10	(31)
	DU-145		CVM-1118	(122)
	PC3	PC3 xenograft model	pyrazolo[3,4-d] pyrimidine derivatives	(52)
	PC3		6BrCaQ-Cn- TPP conjugates	(180)
	H460, H1975	H460 lung xenograft mouse model	gamitrinib	(119)
	H1299, H2122, H358, H2073, H460, H2347, H1975, H1395 and H2085		gamitrinib	(108)
	NCI-H460		gamitrinib	(90)
	H1299, A549, H1437, and H1650		gamitrinib	(6)
	NCI-H460		SMTIN-P01	(5)
Lung cancer	NCI-H460		SMTIN-C10	(31)
	A549		CVM-1118	(122)
	NCI-H460		pyrazolo[3,4-d] pyrimidine derivatives	(52)
	NCI-H460		IR-PU	(120)
	MDA-MB-231, MCF-7	MDA-MB-231 breast cancer xenograft mouse model	gamitrinib	(119)
	MDA-MB-231	MDA-MB-231 xenograf mouse	gamitrinib	(90)
	MCF-7		gamitrinib	(6)
	MDA-MB-231		SMTIN-P01	(5)
Breast	MDA-MB-231		DN401	(191)
cancer	BC MCF7		gamitrinib	(160)
	MDA-MB-231, MDA- MB-435, MCF-7		CVM-1118	(122)
	MDA-MB-231		pyrazolo[3,4-d] pyrimidine derivatives	(52)
	MDA-MB-231		6BrCaQ-Cn- TPP conjugates	(180)
Glioblastoma	U87MG		gamitrinib	(119)
Chebhastonia	LN229		gamitrinib	(190)

TABLE 1 Model systems in which mtHsp90 inhibitors were applied successfully, resulting in cell death or growth inhibition.

(Continued)

TABLE 1 Continued

Cancer type	Cancer cell line <i>in vitro</i>	Animal models	Inhibitors tested	Ref.
	LN229		gamitrinib	(6)
	U251MG, A172, M059K, H4, Hs683, M059J, LN18, LN229, U87MG, U118MG, U138MG, DBTRG-05MG, and T98G; D2363PXA, D645PXA, D2159MG, and D2224MG		gamitrinib	(135)
	U87MG, LN229, U251 and T98G	xenograft mouse model	gamitrinib-tpp	(138)
	U87, LN229, T98G, U251, GBM12, GBM14 and GBM43	GBM12 and GBM43 xenograft mouse model	gamitrinib	(137)
	LN229、U87、T98G、GBM12、GBM22		gamitrinib	(136)
	SHG44, U251-MG and U87-MG		gamitrinib-tpp	(134)
	U118MG		CVM-1118	(122)
	HCT116		gamitrinib	(136)
	DLD1, RKO, SW48, HT29 and HCT116		gamitrinib	(162)
Colorectal cancer	RC HCT116, HT29, COLO320, COLO205 and CaCo2		gamitrinib	(160)
	HT-29, HCT-116, COLO205		CVM-1118	(122)
	HCT116		Gamitrinib-TPP	(161)
	HT-29, HCT-116		6BrCaQ-Cn- TPP conjugates	(180)
Leukemia		HL60 leukemia xenograft mouse model	gamitrinib-tpp	(191)
	K562, Raji, THP-1, HL-60	HL60 leukemia xenograft mouse model	gamitrinib-g4	(125)
	K562		6BrCaQ-Cn- TPP conjugates	(180)
Liver cancer	SK-HEP-1		pyrazolo[3,4-d] pyrimidine derivatives	(52)
Gastric cancer	AGS		vacA	(147)

drugs that target the entire Hsp90 in mammals. Another promising area of research is the combination of mtHsp90 inhibitors with other cancer therapies. Recent studies have demonstrated that mtHsp90 inhibitors can enhance the efficacy of other drugs, such as chemotherapy, radiation therapy, and immunotherapy. Combining these therapies could result in more effective treatment regimens with lower toxicity and better patient outcomes. However, despite the preclinical trials continually demonstrating the feasibility and universal validity of mtHsp90 inhibitors, there is still a lack of

TABLE 2 Clinical trials for mtHSP90 inhibitors.

Drug	Date	Phase	Clinical trials.gov
gamitrinib- TTP	Estimated Primary Completion Date: December 2028	PhaseI	NCT04827810
CVM 1119	Actual Study Completion Date: April 8, 2022	PhaseI	NCT04336124
CV IVI-1118	Estimated Primary Completion Date: September 2025	PhaseII	NCT05257590

clinical trials in humans (Table 2). Further research is needed to ensure the feasibility of these inhibitors before human testing.

We anticipate further testing of mtHsp90 inhibitors in a broader range of cancer types beyond those mentioned in this review. While the efficacy of inhibitors targeting the other Hsp90 protein, such as Ganetespib (STA-9090) and luminespib (AUY922), has been demonstrated in certain cancers, including hepatocellular carcinoma (HCC) (192), pancreatic cancer (PC) (193), gastric cancer (GC) (194, 195).

However, there is a lack of research on the use of mtHsp90 inhibitors in hepatocellular carcinoma, pancreatic cancer, gastric cancer (188, 196, 197). Thus, further testing focused on mtHsp90 inhibitors may prove valuable in identifying new strategies for controlling these cancers. In addition, some studies found that the combination of mtHsp90 inhibitors with other available drugs may represent a promising approach to enhance the anticancer activity of different therapies, while minimizing their undesirable side effects. Hence, the combination of mtHsp90 inhibitors with other drugs may unlock their full potential as anticancer agents and expand their use to a wider range of cancer types. In summary, the prospective therapeutic application of mtHsp90 inhibitors in the treatment of cancers is highly promising. Nevertheless, further investigation is needed to comprehensively ascertain their effectiveness and safety in further studies.

6 Conclusion

This review presents a concise summary of the involvement of mtHsp90 in the progression of cancer. The investigation and development of mtHsp90 inhibitors have exhibited their capacity to significantly revolutionize the field of cancer therapy and enhance the overall prognosis of patients. Researchers have demonstrated promising results in utilizing gamitrinib and other mtHsp90 inhibitors as a novel approach to combat cancer, both *in vitro* animal models including prostate cancer and breast cancer, as well as *in vivo* cancer cell lines. Nevertheless, it is important to recognize that there exist numerous obstacles that require resolution. Further investigation in this field has the potential for making breakthroughs in the realm of cancer treatment, leading to the creation of more secure and effective therapeutic approaches in years to come.

Author contributions

YX: Writing – original draft, Writing – review & editing. XuL: Writing – original draft, Writing – review & editing. QS: Writing – original draft, Writing – review & editing. KL: Writing – original draft. XiL: Writing – original draft. ZZ: Writing – original draft. LF: Writing – original draft. YL: Writing – review & editing. BW: Writing – review & editing.

Funding

The author(s) declare financial support was received for the research, authorship, and/or publication of this article. BW is supported by China Scholarship Council (CSC).

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

1. Trepel J, Mollapour M, Giaccone G, Neckers L. Targeting the dynamic hsp90 complex in cancer. *Nat Rev Cancer* (2010) 10(8):537-49. doi: 10.1038/nrc2887

2. Chehab M, Caza T, Skotnicki K, Landas S, Bratslavsky G, Mollapour M, et al. Targeting Hsp90 in urothelial carcinoma. *Oncotarget* (2015) 6(11):8454-73. doi: 10.18632/oncotarget.3502

3. Pearl LH, Prodromou C. Structure and mechanism of the Hsp90 molecular chaperone machinery. *Annu Rev Biochem* (2006) 75:271-94. doi: 10.1146/ annurev.biochem.75.103004.142738

4. Hall JA, Forsberg LK, Blagg BS. Alternative approaches to Hsp90 modulation for the treatment of cancer. *Future Med Chem* (2014) 6(14):1587–605. doi: 10.4155/ fmc.14.89

5. Lee C, Park HK, Jeong H, Lim J, Lee AJ, Cheon KY, et al. Development of a mitochondria-targeted Hsp90 inhibitor based on the crystal structures of human trap1. *J Am Chem Soc* (2015) 137(13):4358–67. doi: 10.1021/ja511893n

6. Chae Young C, Caino MC, Lisanti S, Ghosh Jagadish C, Dohi T, Danial Nika N, et al. Control of tumor bioenergetics and survival stress signaling by mitochondrial Hsp90s. *Cancer Cell* (2012) 22(3):331-44. doi: 10.1016/j.ccr.2012.07.015

7. Kampinga HH, Hageman J, Vos MJ, Kubota H, Tanguay RM, Bruford EA, et al. Guidelines for the nomenclature of the human heat shock proteins. *Cell Stress Chaperones* (2009) 14(1):105–11. doi: 10.1007/s12192-008-0068-7

8. Hoter A, El-Sabban ME, Naim HY. The Hsp90 family: structure, regulation, function, and implications in health and disease. *Int J Mol Sci* (2018) 19(9):2560. doi: 10.3390/ijms19092560

9. Csermely P, Schnaider T, Soti C, Prohászka Z, Nardai G. The 90-Kda molecular chaperone family: structure, function, and clinical applications. A comprehensive review. *Pharmacol Ther* (1998) 79(2):129-68. doi: 10.1016/s0163-7258(98)00013-8

10. Ciocca DR, Arrigo AP, Calderwood SK. Heat shock proteins and heat shock factor 1 in carcinogenesis and tumor development: an update. *Arch Toxicol* (2013) 87 (1):19–48. doi: 10.1007/s00204-012-0918-z

11. Farrelly FW, Finkelstein DB. Complete sequence of the heat shock-inducible Hsp90 gene of saccharomyces cerevisiae. J Biol Chem (1984) 259(9):5745–51. doi: 10.1016/S0021-9258(18)91077-X

 Kulomaa MS, Weigel NL, Kleinsek DA, Beattie WG, Conneely OM, March C, et al. Amino acid sequence of a chicken heat shock protein derived from the complementary DNA nucleotide sequence. *Biochemistry* (1986) 25(20):6244–51. doi: 10.1021/bi00368a061

13. Song HY, Dunbar JD, Zhang YX, Guo D, Donner DB. Identification of a protein with homology to Hsp90 that binds the type 1 tumor necrosis factor receptor. *J Biol Chem* (1995) 270(8):3574–81. doi: 10.1074/jbc.270.8.3574

14. Kang BH, Plescia J, Dohi T, Rosa J, Doxsey SJ, Altieri DC. Regulation of tumor cell mitochondrial homeostasis by an organelle-specific Hsp90 chaperone network. *Cell* (2007) 131(2):257–70. doi: 10.1016/j.cell.2007.08.028

15. Matassa DS, Agliarulo I, Avolio R, Landriscina M, Esposito F. Trap1 regulation of cancer metabolism: dual role as oncogene or tumor suppressor. *Genes (Basel)* (2018) 9(4):195. doi: 10.3390/genes9040195

16. Serapian SA, Sanchez-Martín C, Moroni E, Rasola A, Colombo G. Targeting the mitochondrial chaperone trap1: strategies and therapeutic perspectives. *Trends Pharmacol Sci* (2021) 42(7):566–76. doi: 10.1016/j.tips.2021.04.003

17. Pearl LH. Review: the Hsp90 molecular chaperone-an enigmatic atpase. Biopolymers (2016) 105(8):594-607. doi: 10.1002/bip.22835

18. Rahimi MN, Buckton LK, Zaiter SS, Kho J, Chan V, Guo A, et al. Synthesis and structure-activity relationships of inhibitors that target the C-terminal meevd on heat shock protein 90. ACS Med Chem Lett (2018) 9(2):73–7. doi: 10.1021/acsmedchemlett.7b00310

19. Dernovšek J, Tomašič T. Following the design path of isoform-selective Hsp90 inhibitors: small differences, great opportunities. *Pharmacol Ther* (2023) 245:108396. doi: 10.1016/j.pharmthera.2023.108396

20. Chen B, Piel WH, Gui L, Bruford E, Monteiro A. The Hsp90 family of genes in the human genome: insights into their divergence and evolution. *Genomics* (2005) 86 (6):627–37. doi: 10.1016/j.ygeno.2005.08.012

21. Chen CF, Chen Y, Dai K, Chen PL, Riley DJ, Lee WH. A new member of the Hsp90 family of molecular chaperones interacts with the retinoblastoma protein during mitosis and after heat shock. *Mol Cell Biol* (1996) 16(9):4691–9. doi: 10.1128/mcb.16.9.4691

22. Song HY, Dunbar JD, Zhang YX, Guo D, Donner DB. Identification of a protein with homology to Hsp90 that binds the type 1 tumor necrosis factor receptor. *J Biol Chem* (1995) 270(8):3574–81. doi: 10.1074/jbc.270.8.3574

23. Hainzl O, Lapina MC, Buchner J, Richter K. The charged linker region is an important regulator of Hsp90 function. *J Biol Chem* (2009) 284(34):22559-67. doi: 10.1074/jbc.M109.031658

24. Jahn M, Rehn A, Pelz B, Hellenkamp B, Richter K, Rief M, et al. The charged linker of the molecular chaperone Hsp90 modulates domain contacts and biological function. *Proc Nat Acad Sci* (2014) 111(50):17881–6. doi: 10.1073/pnas.1414073111

25. Kang S, Kang BH. Structure, function, and inhibitors of the mitochondrial chaperone trap1. J Med Chem (2022) 65(24):16155-72. doi: 10.1021/acs.jmedchem.2c01633

26. Joshi A, Dai L, Liu Y, Lee J, Ghahhari NM, Segala G, et al. The mitochondrial hsp90 paralog trap1 forms an oxphos-regulated tetramer and is involved in mitochondrial metabolic homeostasis. *BMC Biol* (2020) 18(1):10. doi: 10.1186/s12915-020-0740-7

27. Krukenberg KA, Street TO, Lavery LA, Agard DA. Conformational dynamics of the molecular chaperone Hsp90. *Q Rev Biophys* (2011) 44(2):229–55. doi: 10.1017/S0033583510000314

28. Schopf FH, Biebl MM, Buchner J. The Hsp90 chaperone machinery. Nat Rev Mol Cell Biol (2017) 18(6):345–60. doi: 10.1038/nrm.2017.20

29. Genest O, Reidy M, Street TO, Hoskins JR, Camberg JL, Agard DA, et al. Uncovering a region of heat shock protein 90 important for client binding in E. Coli and chaperone function in yeast. *Mol Cell* (2013) 49(3):464–73. doi: 10.1016/j.molcel.2012.11.017

30. Park HK, Hong JH, Oh YT, Kim SS, Yin J, Lee AJ, et al. Interplay between trap1 and sirtuin-3 modulates mitochondrial respiration and oxidative stress to maintain stemness of glioma stem cells. *Cancer Res* (2019) 79(7):1369–82. doi: 10.1158/0008-5472.Can-18-2558

31. Hu S, Ferraro M, Thomas AP, Chung JM, Yoon NG, Seol J-H, et al. Dual binding to orthosteric and allosteric sites enhances the anticancer activity of a trap1-targeting drug. *J Medicinal Chem* (2020) 63(6):2930–40. doi: 10.1021/acs.jmedchem.9b01420

32. Yoon NG, Lee H, Kim S-Y, Hu S, Kim D, Yang S, et al. Mitoquinone inactivates mitochondrial chaperone trap1 by blocking the client binding site. *J Am Chem Soc* (2021) 143(47):19684–96. doi: 10.1021/jacs.1c07099

33. Felts SJ, Owen BA, Nguyen P, Trepel J, Donner DB, Toft DO. The hsp90-related protein trap1 is a mitochondrial protein with distinct functional properties. *J Biol Chem* (2000) 275(5):3305–12. doi: 10.1074/jbc.275.5.3305

34. Zhang L, Su N, Luo Y, Chen S, Zhao T. Trap1 inhibits mic60 ubiquitination to mitigate the injury of cardiomyocytes and protect mitochondria in extracellular acidosis. *Cell Death Discovery* (2021) 7(1):389. doi: 10.1038/s41420-021-00786-5

35. Rutledge BS, Choy WY, Duennwald ML. Folding or holding?-Hsp70 and Hsp90 chaperoning of misfolded proteins in neurodegenerative disease. *J Biol Chem* (2022) 298(5):101905. doi: 10.1016/j.jbc.2022.101905

36. Neckers L. Hsp90 inhibitors as novel cancer chemotherapeutic agents. *Trends Mol Med* (2002) 8(4 Suppl):S55–61. doi: 10.1016/s1471-4914(02)02316-x

37. Verba KA, Wang RY, Arakawa A, Liu Y, Shirouzu M, Yokoyama S, et al. Atomic structure of Hsp90-Cdc37-Cdk4 reveals that Hsp90 traps and stabilizes an unfolded kinase. *Science* (2016) 352(6293):1542–7. doi: 10.1126/science.aaf5023

38. Noddings CM, Wang RY, Johnson JL, Agard DA. Structure of Hsp90-P23-Gr reveals the Hsp90 client-remodelling mechanism. *Nature* (2022) 601(7893):465–9. doi: 10.1038/s41586-021-04236-1

39. Elnatan D, Betegon M, Liu Y, Ramelot T, Kennedy MA, Agard DA. Symmetry broken and rebroken during the atp hydrolysis cycle of the mitochondrial Hsp90 trap1. *Elife* (2017) 6:e25235. doi: 10.7554/eLife.25235

40. Motojima-Miyazaki Y, Yoshida M, Motojima F. Ribosomal protein L2 associates with E. Coli Htpg and activates its atpase activity. *Biochem Biophys Res Commun* (2010) 400(2):241–5. doi: 10.1016/j.bbrc.2010.08.047

41. Partridge JR, LA L, Elnatan D, Naber N, Cooke R, Agard DA. A novel N-terminal extension in mitochondrial trap1 serves as a thermal regulator of chaperone activity. *Elife* (2014) 3:e03487. doi: 10.7554/eLife.03487

42. Kim S, Backe SJ, Wengert LA, Johnson AE, Isakov RV, Bratslavsky MS, et al. Oglcnacylation suppresses trap1 activity and promotes mitochondrial respiration. *Cell Stress Chaperones* (2022) 27(5):573–85. doi: 10.1007/s12192-022-01293-x

43. Lin YF, Haynes CM. Metabolism and the upr(Mt). *Mol Cell* (2016) 61(5):677–82. doi: 10.1016/j.molcel.2016.02.004

44. Ruggero D. Translational control in cancer etiology. *Cold Spring Harb Perspect Biol* (2013) 5(2):a012336. doi: 10.1101/cshperspect.a012336

45. Sreedhar AS, Kalmar E, Csermely P, Shen YF. Hsp90 isoforms: functions, expression and clinical importance. *FEBS Lett* (2004) 562(1-3):11-5. doi: 10.1016/s0014-5793(04)00229-7

46. Sanchez J, Carter TR, Cohen MS, Blagg BSJ. Old and new approaches to target the Hsp90 chaperone. *Curr Cancer Drug Targets* (2020) 20(4):253–70. doi: 10.2174/1568009619666191202101330

47. Yang S, Yoon NG, Kim D, Park E, Kim SY, Lee JH, et al. Design and synthesis of trap1 selective inhibitors: H-bonding with asn171 residue in trap1 increases paralog selectivity. *ACS Med Chem Lett* (2021) 12(7):1173-80. doi: 10.1021/acsmedchemlett.1c00213

48. Plescia J, Salz W, Xia F, Pennati M, Zaffaroni N, Daidone MG, et al. Rational design of shepherdin, a novel anticancer agent. *Cancer Cell* (2005) 7(5):457–68. doi: 10.1016/j.ccr.2005.03.035

49. Park HK, Yoon NG, Lee JE, Hu S, Yoon S, Kim SY, et al. Unleashing the full potential of Hsp90 inhibitors as cancer therapeutics through simultaneous inactivation of Hsp90, Grp94, and Trap1. *Exp Mol Med* (2020) 52(1):79–91. doi: 10.1038/s12276-019-0360-x

50. Hayat U, Elliott GT, Olszanski AJ, Altieri DC. Feasibility and safety of targeting mitochondria for cancer therapy – preclinical characterization of gamitrinib, a first-inclass, mitochondrial-targeted small molecule Hsp90 inhibitor. *Cancer Biol Ther* (2022) 23(1):117–26. doi: 10.1080/15384047.2022.2029132

51. Merfeld T, Peng S, Keegan BM, Crowley VM, Brackett CM, Gutierrez A, et al. Elucidation of novel trap1-selective inhibitors that regulate mitochondrial processes. *Eur J Med Chem* (2023) 258:115531. doi: 10.1016/j.ejmech.2023.115531

52. Kim D, Kim SY, Kim D, Yoon NG, Yun J, Hong KB, et al. Development of pyrazolo[3,4-D]Pyrimidine-6-amine-based trap1 inhibitors that demonstrate in vivo anticancer activity in mouse xenograft models. *Bioorg Chem* (2020) 101:103901. doi: 10.1016/j.bioorg.2020.103901

53. Sanchez-Martin C, Moroni E, Ferraro M, Laquatra C, Cannino G, Masgras I, et al. Rational design of allosteric and selective inhibitors of the molecular chaperone trap1. *Cell Rep* (2020) 31(3):107531. doi: 10.1016/j.celrep.2020.107531

54. Rasola A, Neckers L, Picard D. Mitochondrial oxidative phosphorylation trap(1) Ped in tumor cells. *Trends Cell Biol* (2014) 24(8):455–63. doi: 10.1016/j.tcb.2014.03.005

55. Megger DA, Bracht T, Kohl M, Ahrens M, Naboulsi W, Weber F, et al. Proteomic differences between hepatocellular carcinoma and nontumorous liver tissue investigated by a combined gel-based and label-free quantitative proteomics study. *Mol Cell Proteomics* (2013) 12(7):2006–20. doi: 10.1074/mcp.M113.028027

56. Leav I, Plescia J, Goel HL, Li J, Jiang Z, Cohen RJ, et al. Cytoprotective mitochondrial chaperone trap-1 as a novel molecular target in localized and metastatic prostate cancer. *Am J Pathol* (2010) 176(1):393-401. doi: 10.2353/ ajpath.2010.090521

57. Agorreta J, Hu J, Liu D, Delia D, Turley H, Ferguson DJ, et al. Trap1 regulates proliferation, mitochondrial function, and has prognostic significance in nsclc. *Mol Cancer Res* (2014) 12(5):660–9. doi: 10.1158/1541-7786.Mcr-13-0481

58. Lee JH, Kang KW, Kim JE, Hwang SW, Park JH, Kim SH, et al. Differential expression of heat shock protein 90 isoforms in small cell lung cancer. *Int J Clin Exp Pathol* (2015) 8(8):9487–93. doi: 10.26226/morressier.5b4709896f4cb300109519c2

59. Fang W, Li X, Jiang Q, Liu Z, Yang H, Wang S, et al. Transcriptional patterns, biomarkers and pathways characterizing nasopharyngeal carcinoma of southern China. *J Transl Med* (2008) 6:32. doi: 10.1186/1479-5876-6-32

60. Li S, Lv Q, Sun H, Xue Y, Wang P, Liu L, et al. Expression of trap1 predicts poor survival of Malignant glioma patients. *J Mol Neurosci* (2015) 55(1):62–8. doi: 10.1007/s12031-014-0413-5

61. Zhang B, Wang J, Huang Z, Wei P, Liu Y, Hao J, et al. Aberrantly upregulated trap1 is required for tumorigenesis of breast cancer. *Oncotarget* (2015) 6(42):44495–508. doi: 10.18632/oncotarget.6252

62. Yoshida S, Tsutsumi S, Muhlebach G, Sourbier C, Lee MJ, Lee S, et al. Molecular chaperone trap1 regulates a metabolic switch between mitochondrial respiration and aerobic glycolysis. *Proc Natl Acad Sci U S A* (2013) 110(17):E1604–12. doi: 10.1073/pnas.1220659110

63. Masgras I, Laquatra C, Cannino G, Serapian SA, Colombo G, Rasola A. The molecular chaperone trap1 in cancer: from the basics of biology to pharmacological targeting. *Semin Cancer Biol* (2021) 76:45–53. doi: 10.1016/j.semcancer.2021.07.002

64. Altieri DC, Stein GS, Lian JB, Languino LR. Trap-1, the mitochondrial Hsp90. Biochim Biophys Acta (2012) 1823(3):767–73. doi: 10.1016/j.bbamcr.2011.08.007

65. Voloboueva LA, Duan M, Ouyang Y, Emery JF, Stoy C, Giffard RG. Overexpression of mitochondrial Hsp70/Hsp75 protects astrocytes against ischemic injury in vitro. *J Cereb Blood Flow Metab* (2008) 28(5):1009–16. doi: 10.1038/ sj.jcbfm.9600600

66. Costa AC, Loh SH, Martins LM. Drosophila trap1 protects against mitochondrial dysfunction in a pink1/parkin model of Parkinson's disease. *Cell Death Dis* (2013) 4(1):e467. doi: 10.1038/cddis.2012.205

67. Avolio R, Agliarulo I, Criscuolo D, Sarnataro D, Auriemma M, De Lella S, et al. Cytosolic and mitochondrial translation elongation are coordinated through the molecular chaperone trap1 for the synthesis and import of mitochondrial proteins. *Genome Res* (2023) 33(8):1242–57. doi: 10.1101/gr.277755.123

68. Lisanti S, Tavecchio M, Chae YC, Liu Q, Brice AK, Thakur ML, et al. Deletion of the mitochondrial chaperone trap-1 uncovers global reprogramming of metabolic networks. *Cell Rep* (2014) 8(3):671–7. doi: 10.1016/j.celrep.2014.06.061

69. Barbosa IA, Vega-Naredo I, Loureiro R, Branco AF, Garcia R, Scott PM, et al. Trap1 regulates autophagy in lung cancer cells. *Eur J Clin Invest* (2018) 48(4):e12900. doi: 10.1111/eci.12900

70. Chen FW, Davies JP, Calvo R, Chaudhari J, Dolios G, Taylor MK, et al. Activation of mitochondrial trap1 stimulates mitochondria-lysosome crosstalk and correction of lysosomal dysfunction. *iScience* (2022) 25(9):104941. doi: 10.1016/j.isci.2022.104941

71. Caino MC, Chae YC, Vaira V, Ferrero S, Nosotti M, Martin NM, et al. Metabolic stress regulates cytoskeletal dynamics and metastasis of cancer cells. *J Clin Invest* (2013) 123(7):2907–20. doi: 10.1172/jci67841

72. Landriscina M, Laudiero G, Maddalena F, Amoroso MR, Piscazzi A, Cozzolino F, et al. Mitochondrial chaperone trap1 and the calcium binding protein sorcin interact and protect cells against apoptosis induced by antiblastic agents. *Cancer Res* (2010) 70 (16):6577–86. doi: 10.1158/0008-5472.CAN-10-1256

73. Matassa DS, Amoroso MR, Lu H, Avolio R, Arzeni D, Procaccini C, et al. Oxidative metabolism drives inflammation-induced platinum resistance in human ovarian cancer. *Cell Death Differ* (2016) 23(9):1542–54. doi: 10.1038/cdd.2016.39

74. Sciacovelli M, Guzzo G, Morello V, Frezza C, Zheng L, Nannini N, et al. The mitochondrial chaperone trap1 promotes neoplastic growth by inhibiting succinate dehydrogenase. *Cell Metab* (2013) 17(6):988–99. doi: 10.1016/j.cmet.2013.04.019

75. Sanchez-Martin C, Menon D, Moroni E, Ferraro M, Masgras I, Elsey J, et al. Honokiol bis-dichloroacetate is a selective allosteric inhibitor of the mitochondrial chaperone trap1. *Antioxid Redox Signal* (2021) 34(7):505–16. doi: 10.1089/ars.2019.7972

76. Matassa DS, Criscuolo D, Avolio R, Agliarulo I, Sarnataro D, Pacelli C, et al. Regulation of mitochondrial complex iii activity and assembly by trap1 in cancer cells. *Cancer Cell Int* (2022) 22(1):402. doi: 10.1186/s12935-022-02788-4

77. Pridgeon JW, Olzmann JA, Chin L-S, Li L. Pink1 protects against oxidative stress by phosphorylating mitochondrial chaperone trap1. *PLoS Biol* (2007) 5(7):e172. doi: 10.1371/journal.pbio.0050172

78. Ward PS, Thompson CB. Metabolic reprogramming: A cancer hallmark even warburg did not anticipate. *Cancer Cell* (2012) 21(3):297-308. doi: 10.1016/j.ccr.2012.02.014

79. Wallace DC. Mitochondria and cancer. Nat Rev Cancer (2012) 12(10):685–98. doi: 10.1038/nrc3365

80. Mihaylova MM, Shaw RJ. The Ampk signalling pathway coordinates cell growth, autophagy and metabolism. Nat Cell Biol (2011) 13(9):1016-23. doi: 10.1038/ncb2329

81. Xiao L, Hu Q, Peng Y, Zheng K, Zhang T, Yang L, et al. Trap1 suppresses oral squamous cell carcinoma progression by reducing oxidative phosphorylation metabolism of cancer-associated fibroblasts. *BMC Cancer* (2021) 21(1):1329. doi: 10.1186/s12885-021-09049-z

82. Lisanti S, Garlick DS, Bryant KG, Tavecchio M, Mills GB, Lu Y, et al. Transgenic expression of the mitochondrial chaperone tnfr-associated protein 1 (Trap1) accelerates prostate cancer development. *J Biol Chem* (2016) 291(48):25247–54. doi: 10.1074/jbc.M116.745950

83. Ramkumar B, Dharaskar SP, Mounika G, Paithankar K, Sreedhar AS. Mitochondrial chaperone, trap1 as a potential pharmacological target to combat cancer metabolism. *Mitochondrion* (2020) 50:42–50. doi: 10.1016/j.mito.2019.09.011

84. Si T, Yang G, Qiu X, Luo Y, Liu B, Wang B. Expression of tumor necrosis factor receptor-associated protein 1 and its clinical significance in kidney cancer. *Int J Clin Exp Pathol* (2015) 8(10):13090–5.

85. Xie S, Wang X, Gan S, Tang X, Kang X, Zhu S. The mitochondrial chaperone trap1 as a candidate target of oncotherapy. *Front Oncol* (2020) 10:585047. doi: 10.3389/ fonc.2020.585047

86. Purushottam Dharaskar S, Paithankar K, Kanugovi Vijayavittal A, Shabbir Kara H, Amere Subbarao S. Mitochondrial chaperone, trap1 modulates mitochondrial dynamics and promotes tumor metastasis. *Mitochondrion* (2020) 54:92–101. doi: 10.1016/j.mito.2020.08.001

87. Frame MC, Patel H, Serrels B, Lietha D, Eck MJ. The ferm domain: organizing the structure and function of Fak. *Nat Rev Mol Cell Biol* (2010) 11(11):802–14. doi: 10.1038/nrm2996

88. White E. Deconvoluting the context-dependent role for autophagy in cancer. Nat Rev Cancer (2012) 12(6):401–10. doi: 10.1038/nrc3262

89. Masgras I, Ciscato F, Brunati AM, Tibaldi E, Indraccolo S, Curtarello M, et al. Absence of neurofibromin induces an oncogenic metabolic switch via mitochondrial Erk-mediated phosphorylation of the chaperone trap1. *Cell Rep* (2017) 18(3):659–72. doi: 10.1016/j.celrep.2016.12.056

90. Park H-K, Lee J-E, Lim J, Jo D-E, Park S-A, Suh P-G, et al. Combination treatment with doxorubicin and gamitrinib synergistically augments anticancer activity through enhanced activation of Bim. *BMC Cancer* (2014) 14(1):431. doi: 10.1186/1471-2407-14-431

91. Hua G, Zhang Q, Fan Z. Heat shock protein 75 (Trap1) antagonizes reactive oxygen species generation and protects cells from granzyme M-mediated apoptosis. J Biol Chem (2007) 282(28):20553–60. doi: 10.1074/jbc.M703196200

92. Kang BH. Trap1 regulation of mitochondrial life or death decision in cancer cells and mitochondria-targeted trap1 inhibitors. *BMB Rep* (2012) 45(1):1–6. doi: 10.5483/ bmbrep.2012.45.1.1

93. Lebedev I, Nemajerova A, Foda ZH, Kornaj M, Tong M, Moll UM, et al. A novel in vitro cypd-mediated P53 aggregation assay suggests a model for mitochondrial

permeability transition by chaperone systems. J Mol Biol (2016) 428(20):4154-67. doi: 10.1016/j.jmb.2016.08.001

94. Green DR, Kroemer G. The pathophysiology of mitochondrial cell death. *Science* (2004) 305(5684):626–9. doi: 10.1126/science.1099320

95. Xiang F, Huang YS, Shi XH, Zhang Q. Mitochondrial chaperone tumour necrosis factor receptor-associated protein 1 protects cardiomyocytes from hypoxic injury by regulating mitochondrial permeability transition pore opening. *FEBS J* (2010) 277(8):1929–38. doi: 10.1111/j.1742-4658.2010.07615.x

96. Rasola A, Bernardi P. The mitochondrial permeability transition pore and its adaptive responses in tumor cells. *Cell Calcium* (2014) 56(6):437–45. doi: 10.1016/j.ceca.2014.10.003

97. Liu L, Zhang L, Zhao J, Guo X, Luo Y, Hu W, et al. Tumor necrosis factor receptor-associated protein 1 protects against mitochondrial injury by preventing high glucose-induced Mptp opening in diabetes. *Oxid Med Cell Longev* (2020) 2020:6431517. doi: 10.1155/2020/6431517

98. Montesano Gesualdi N, Chirico G, Pirozzi G, Costantino E, Landriscina M, Esposito F. Tumor necrosis factor-associated protein 1 (Trap-1) protects cells from oxidative stress and apoptosis. *Stress* (2007) 10(4):342–50. doi: 10.1080/10253890701314863

99. Zhang X, Dong Y, Gao M, Hao M, Ren H, Guo L, et al. Knockdown of trap1 promotes cisplatin-induced apoptosis by promoting the ros-dependent mitochondrial dysfunction in lung cancer cells. *Mol Cell Biochem* (2021) 476(2):1075–82. doi: 10.1007/s11010-020-03973-7

100. Felux J, Erbacher A, Breckler M, Hervé R, Lemeiter D, Mannherz HG, et al. Deoxyribonuclease 1-mediated clearance of circulating chromatin prevents from immune cell activation and pro-inflammatory cytokine production, a phenomenon amplified by low trap1 activity: consequences for systemic lupus erythematosus. *Front Immunol* (2021) 12:613597. doi: 10.3389/fimmu.2021.613597

101. Badarni M, Gabbay S, Elkabets M, Rotblat B. Gene expression and drug sensitivity analysis of mitochondrial chaperones reveals that hspd1 and trap1 expression correlates with sensitivity to inhibitors of DNA replication and mitosis. *Biol (Basel)* (2023) 12(7):988. doi: 10.3390/biology12070988

102. Chen R, Pan S, Lai K, Lai LA, Crispin DA, Bronner MP, et al. Up-regulation of mitochondrial chaperone trap1 in ulcerative colitis associated colorectal cancer. *World J Gastroenterol* (2014) 20(45):17037–48. doi: 10.3748/wjg.v20.i45.17037

103. Diaz-Diaz A, Roca-Lema D, Casas-Pais A, Romay G, Colombo G, Concha A, et al. Heat shock protein 90 chaperone regulates the E3 ubiquitin-ligase hakai protein stability. *Cancers* (2020) 12(1):215. doi: 10.3390/cancers12010215

104. Condelli V, Crispo F, Pietrafesa M, Lettini G, Matassa DS, Esposito F, et al. Hsp90 molecular chaperones, metabolic rewiring, and epigenetics: impact on tumor progression and perspective for anticancer therapy. *Cells* (2019) 8(6):532. doi: 10.3390/cells8060532

105. Jiang F, Guo A-p, Xu J-c, You Q-D, Xu X-L. Discovery of a potent Grp94 selective inhibitor with anti-inflammatory efficacy in a mouse model of ulcerative colitis. *J Medicinal Chem* (2018) 61(21):9513–33. doi: 10.1021/acs.jmedchem.8b00800

106. Kuchitsu Y, Nagashio R, Igawa S, Kusuhara S, Tsuchiya B, Ichinoe M, et al. Trap1 is a predictive biomarker of platinum-based adjuvant chemotherapy benefits in patients with resected lung adenocarcinoma. *Biomed Res* (2020) 41(1):53–65. doi: 10.2220/biomedres.41.53

107. Dharaskar SP, Amere Subbarao S. The mitochondrial chaperone trap-1 regulates the glutamine metabolism in tumor cells. *Mitochondrion* (2023) 69:159–70. doi: 10.1016/j.mito.2023.02.011

108. Vo VTA, Choi JW, Phan ANH, Hua TNM, Kim MK, Kang BH, et al. Trap1 inhibition increases glutamine synthetase activity in glutamine auxotrophic non-small cell lung cancer cells. *Anticancer Res* (2018) 38(4):2187–93. doi: 10.21873/anticanres.12460

109. Kumar-Sinha C, Kalyana-Sundaram S, Chinnaiyan AM. Landscape of gene fusions in epithelial cancers: seq and ye shall find. *Genome Med* (2015) 7:129. doi: 10.1186/s13073-015-0252-1

110. Clarke BE, Kalmar B, Greensmith L. Enhanced expression of trap1 protects mitochondrial function in motor neurons under conditions of oxidative stress. *Int J Mol Sci* (2022) 23(3):1789. doi: 10.3390/ijms23031789

111. Ramos Rego I, Santos Cruz B, Ambrósio AF, Alves CH. Trap1 in oxidative stress and neurodegeneration. Antioxidants (Basel) (2021) 10(11):1829. doi: 10.3390/antiox10111829

112. Xiang F, Ma SY, Lv YL, Zhang DX, Song HP, Huang YS. Tumor necrosis factor receptor-associated protein 1 regulates hypoxia-induced apoptosis through a mitochondria-dependent pathway mediated by cytochrome C oxidase subunit ii. *Burns Trauma* (2019) 7:16. doi: 10.1186/s41038-019-0154-3

113. Lin MH, Shen KY, Liu BS, Chen IH, Sher YP, Tseng GC, et al. Immunological evaluation of a novel Hla-A2 restricted phosphopeptide of tumor associated antigen, trap1, on cancer therapy. *Vaccine X* (2019) 1:100017. doi: 10.1016/j.jvacx.2019.100017

114. Paez JG, Janne PA, Lee JC, Tracy S, Greulich H, Gabriel S, et al. Egfr mutations in lung cancer: correlation with clinical response to gefitinib therapy. *Science* (2004) 304 (5676):1497–500. doi: 10.1126/science.1099314

115. Lynch TJ, Bell DW, Sordella R, Gurubhagavatula S, Okimoto RA, Brannigan BW, et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* (2004) 350 (21):2129–39. doi: 10.1056/NEJM0a040938

116. Janne PA, Yang JC, Kim DW, Planchard D, Ohe Y, Ramalingam SS, et al. Azd9291 in egfr inhibitor-resistant non-small-cell lung cancer. *N Engl J Med* (2015) 372 (18):1689–99. doi: 10.1056/NEJM0a1411817

117. Suda K, Tomizawa K, Mitsudomi T. Biological and clinical significance of Kras mutations in lung cancer: an oncogenic driver that contrasts with Egfr mutation. *Cancer Metastasis Rev* (2010) 29(1):49–60. doi: 10.1007/s10555-010-9209-4

118. Sawai A, Chandarlapaty S, Greulich H, Gonen M, Ye Q, Arteaga CL, et al. Inhibition of Hsp90 down-regulates mutant epidermal growth factor receptor (Egfr) expression and sensitizes Egfr mutant tumors to paclitaxel. *Cancer Res* (2008) 68 (2):589–96. doi: 10.1158/0008-5472.CAN-07-1570

119. Kang BH, Plescia J, Song HY, Meli M, Colombo G, Beebe K, et al. Combinatorial drug design targeting multiple cancer signaling networks controlled by mitochondrial Hsp90. J Clin Invest (2009) 119(3):454–64. doi: 10.1172/jci37613

120. Thomas AP, Lee AJ, Palanikumar L, Jana B, Kim K, Kim S, et al. Mitochondrial heat shock protein-guided photodynamic therapy. *Chem Commun (Camb)* (2019) 55 (84):12631–4. doi: 10.1039/c9cc06411g

121. Hayat U, Elliott GT, Olszanski AJ, Altieri DC. Feasibility and safety of targeting mitochondria for cancer therapy - preclinical characterization of gamitrinib, a first-inclass, mitochondrial-targeted small molecule hsp90 inhibitor. *Cancer Biol Ther* (2022) 23(1):117–26. doi: 10.1080/15384047.2022.2029132

122. Shen I, Chen YL, Huang CC, Shyu YC, Seftor REB, Seftor EA, et al. Cvm-1118 (Foslinanib), a 2-phenyl-4-quinolone derivative, promotes apoptosis and inhibits vasculogenic mimicry via targeting trap1. *Pathol Oncol Res* (2023) 29:1611038. doi: 10.3389/pore.2023.1611038

123. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: globocan estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/ caac.21660

124. Gandaglia G, Leni R, Bray F, Fleshner N, Freedland SJ, Kibel A, et al. Epidemiology and prevention of prostate cancer. *Eur Urol Oncol* (2021) 4(6):877–92. doi: 10.1016/j.euo.2021.09.006

125. Catalona WJ. Management of cancer of the prostate. N Engl J Med (1994) 331 (15):996–1004. doi: 10.1056/nejm199410133311507

126. Kang BH, Siegelin MD, Plescia J, Raskett CM, Garlick DS, Dohi T, et al. Preclinical characterization of mitochondria-targeted small molecule Hsp90 inhibitors, gamitrinibs, in advanced prostate cancer. *Clin Cancer Res* (2010) 16(19):4779–88. doi: 10.1158/1078-0432.Ccr-10-1818

127. Kang BH, Tavecchio M, Goel HL, Hsieh CC, Garlick DS, Raskett CM, et al. Targeted inhibition of mitochondrial Hsp90 suppresses localised and metastatic prostate cancer growth in a genetic mouse model of disease. *Br J Cancer* (2011) 104 (4):629–34. doi: 10.1038/bjc.2011.9

128. Ostrom QT, Gittleman H, Truitt G, Boscia A, Kruchko C, Barnholtz-Sloan JS. Cbtrus statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2011-2015. *Neuro Oncol* (2018) 20(suppl_4):iv1-iv86. doi: 10.1093/neuonc/noy131

129. Ostrom QT, Cioffi G, Gittleman H, Patil N, Waite K, Kruchko C, et al. Cbtrus statistical report: primary brain and other central nervous system tumors diagnosed in the United States in 2012-2016. *Neuro Oncol* (2019) 21(Suppl 5):v1-v100. doi: 10.1093/ neuonc/noz150

130. Stupp R, Mason WP, van den Bent MJ, Weller M, Fisher B, Taphoorn MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. *N Engl J Med* (2005) 352(10):987–96. doi: 10.1056/NEJMoa043330

131. Zhang Y, Nguyen TTT, Shang E, Mela A, Humala N, Mahajan A, et al. Met inhibition elicits Pgc1 α -dependent metabolic reprogramming in glioblastoma. *Cancer Res* (2020) 80(1):30–43. doi: 10.1158/0008-5472.CAN-19-1389

132. Wu J. Exosomal microrna expression profiling altered by high trap1 expression in glioblastoma. *Neuro-Oncology* (2019) 21:45–. doi: 10.1093/neuonc/noz126.161

133. Wu J, Zhao S. Effects of trap1 on expression of glycolytic enzymes in glioblastoma stem cells. *Neuro-Oncology* (2018) 20:293-. doi: 10.1093/neuonc/ noy139.295

134. Wang N, Zhu P, Huang R, Sun L, Dong D, Gao Y. Suppressing trap1 sensitizes glioblastoma multiforme cells to temozolomide. *Exp Ther Med* (2021) 22(5):1246. doi: 10.3892/etm.2021.10681

135. Wei S, Yin D, Yu S, Lin X, Savani MR, Du K, et al. Antitumor activity of a mitochondrial-targeted Hsp90 inhibitor in gliomas. *Clin Cancer Res* (2022) 28 (10):2180–95. doi: 10.1158/1078-0432.Ccr-21-0833

136. Nguyen TTT, Ishida CT, Shang E, Shu C, Bianchetti E, Karpel-Massler G, et al. Activation of lxr receptors and inhibition of trap1 causes synthetic lethality in solid tumors. *Cancers (Basel)* (2019) 11(6):788. doi: 10.3390/cancers11060788

137. Nguyen TTT, Zhang Y, Shang E, Shu C, Quinzii CM, Westhoff M-A, et al. Inhibition of Hdac1/2 along with trap1 causes synthetic lethality in glioblastoma model systems. *Cells* (2020) 9(7):1661. doi: 10.3390/cells9071661

138. Karpel-Massler G, Ishida CT, Bianchetti E, Shu C, Perez-Lorenzo R, Horst B, et al. Inhibition of mitochondrial matrix chaperones and antiapoptotic Bcl-2 family proteins empower antitumor therapeutic responses. *Cancer Res* (2017) 77(13):3513–26. doi: 10.1158/0008-5472.Can-16-3424

139. Liu D, Hu J, Agorreta J, Cesario A, Zhang Y, Harris AL, et al. Tumor necrosis factor receptor-associated protein 1(Trap1) regulates genes involved in cell cycle and metastases. *Cancer Lett* (2010) 296(2):194–205. doi: 10.1016/j.canlet.2010.04.017

140. Avagliano A, Ruocco MR, Aliotta F, Belviso I, Accurso A, Masone S, et al. Mitochondrial flexibility of breast cancers: A growth advantage and a therapeutic opportunity. *Cells* (2019) 8(5):401. doi: 10.3390/cells8050401

141. Kim H, Yang J, Kim MJ, Choi S, Chung JR, Kim JM, et al. Tumor necrosis factor receptor-associated protein 1 (Trap1) mutation and trap1 inhibitor gamitrinib-triphenylphosphonium (G-tpp) induce a forkhead box O (Foxo)-dependent cell protective signal from mitochondria. *J Biol Chem* (2016) 291(4):1841–53. doi: 10.1074/jbc.M115.656934

142. Pazo Cid RA, Antón A. Advanced Her2-positive gastric cancer: current and future targeted therapies. *Crit Rev Oncology/Hematology* (2013) 85(3):350-62. doi: 10.1016/j.critrevonc.2012.08.008

143. Jhaveri K, Ochiana SO, Dunphy MP, Gerecitano JF, Corben AD, Peter RI, et al. Heat shock protein 90 inhibitors in the treatment of cancer: current status and future directions. *Expert Opin Investigational Drugs* (2014) 23(5):611–28. doi: 10.1517/13543784.2014.902442

144. Zhang P, Lu Y, Yu D, Zhang D, Hu W. Trap1 provides protection against myocardial ischemia-reperfusion injury by ameliorating mitochondrial dysfunction. *Cell Physiol Biochem* (2015) 36(5):2072–82. doi: 10.1159/000430174

145. Kadye R, Kramer AH, Joos-Vandewalle J, Parsons M, Njengele Z, Hoppe H, et al. Guardian of the furnace: mitochondria, trap1, ros and stem cell maintenance. *IUBMB Life* (2014) 66(1):42–5. doi: 10.1002/iub.1234

146. Han P, Wang QL, Zhang X. Expression of trap1 in gastric cancer tissue and its correlation with Malignant biology. *Asian Pac J Trop Med* (2016) 9(1):64–8. doi: 10.1016/j.apjtm.2015.12.014

147. Teng Y, Liu X, Han B, Ma Q, Liu Y, Kong H, et al. Helicobacter pyloridownregulated tumor necrosis factor receptor-associated protein 1 mediates apoptosis of human gastric epithelial cells. *J Cell Physiol* (2019) 234(9):15698–707. doi: 10.1002/ jcp.28223

148. Huang J, Zhao L, Gong C, Wang Y, Qu Y, Ji C, et al. Shikonin promotes apoptosis and attenuates migration and invasion of human esophageal cancer cells by inhibiting tumor necrosis factor receptor-associated protein 1 expression and Akt/mtor signaling pathway. *Evid Based Complement Alternat Med* (2021) 2021:5386050. doi: 10.1155/2021/5386050

149. Deweerdt S. Microbiome microbial mystery. *Nature* (2015) 521(7551):S10–S1. doi: 10.1038/521S10a

150. Sebert M, Sola-Tapias N, Mas E, Barreau F, Ferrand A. Protease-activated receptors in the intestine: focus on inflammation and cancer. *Front Endocrinol* (2019) 10:717. doi: 10.3389/fendo.2019.00717

151. Wolf AMD, Fontham ETH, Church TR, Flowers CR, Guerra CE, LaMonte SJ, et al. Colorectal cancer screening for average-risk adults: 2018 guideline update from the American cancer society. *CA: A Cancer Journal for Clinicians* (2018) 68(4):250–81. doi: 10.3322/caac.21457

152. Pak MG, Koh HJ, Roh MS. Clinicopathologic significance of trap1 expression in colorectal cancer: A large scale study of human colorectal adenocarcinoma tissues. *Diagn Pathol* (2017) 12:6. doi: 10.1186/s13000-017-0598-3

153. Gao J-Y, Song B-R, Peng J-J, Lu Y-M. Correlation between mitochondrial trap-1 expression and lymph node metastasis in colorectal cancer. *World J Gastroenterol* (2012) 18(41):5965–71. doi: 10.3748/wjg.v18.i41.5965

154. Lettini G, Sisinni L, Condelli V, Matassa DS, Simeon V, Maddalena F, et al. Trap1 regulates stemness through Wnt/Beta-catenin pathway in human colorectal carcinoma. *Cell Death Differentiation* (2016) 23(11):1792-803. doi: 10.1038/ cdd.2016.67

155. Lettini G, Condelli V, Pietrafesa M, Crispo F, Zoppoli P, Maddalena F, et al. Trap1 regulates Wnt/B-catenin pathway through Lrp5/6 receptors expression modulation. *Int J Mol Sci* (2020) 21(20):7526. doi: 10.3390/ijms21207526

156. Condelli V, Piscazzi A, Sisinni L, Matassa DS, Maddalena F, Lettini G, et al. Trap1 is involved in braf regulation and downstream attenuation of erk phosphorylation and cell-cycle progression: A novel target for braf-mutated colorectal tumors. *Cancer Res* (2014) 74(22):6693–704. doi: 10.1158/0008-5472.Can-14-1331

157. Amoroso MR, Matassa DS, Agliarulo I, Avolio R, Lu H, Sisinni L, et al. Trap1 downregulation in human ovarian cancer enhances invasion and epithelial-mesenchymal transition. *Cell Death Dis* (2016) 7:e2522. doi: 10.1038/cddis.2016.400

158. Bruno G, Li Bergolis V, Piscazzi A, Crispo F, Condelli V, Zoppoli P, et al. Trap1 regulates the response of colorectal cancer cells to hypoxia and inhibits ribosome biogenesis under conditions of oxygen deprivation. *Int J Oncol* (2022) 60(6):79. doi: 10.3892/ijo.2022.5369

159. Sveen A, Bruun J, Eide PW, Eilertsen IA, Ramirez L, Murumagi A, et al. Colorectal cancer consensus molecular subtypes translated to preclinical models uncover potentially targetable cancer cell dependencies. *Clin Cancer Res* (2018) 24 (4):794–806. doi: 10.1158/1078-0432.Ccr-17-1234

160. Condelli V, Maddalena F, Sisinni L, Lettini G, Matassa DS, Piscazzi A, et al. Targeting trap1 as a downstream effector of braf cytoprotective pathway: A novel strategy for human braf-driven colorectal carcinoma. *Oncotarget* (2015) 6(26):22298–309. doi: 10.18632/oncotarget.4263

161. Bhasin N, Dabral P, Senavirathna L, Pan S, Chen R. Inhibition of trap1 accelerates the DNA damage response, activation of the heat shock response and metabolic reprogramming in colon cancer cells. *Front Biosci (Landmark Ed)* (2023) 28 (9):227. doi: 10.31083/j.fbl2809227

162. Tsai HY, Bronner MP, March JK, Valentine JF, Shroyer NF, Lai LA, et al. Metabolic targeting of Nrf2 potentiates the efficacy of the trap1 inhibitor G-tpp through reduction of ros detoxification in colorectal cancer. *Cancer Lett* (2022) 549:215915. doi: 10.1016/j.canlet.2022.215915

163. Yang C, Zhang H, Zhang L, Zhu AX, Bernards R, Qin W, et al. Evolving therapeutic landscape of advanced hepatocellular carcinoma. *Nat Rev Gastroenterol Hepatol* (2022) 20, 203–22. doi: 10.1038/s41575-022-00704-9

164. Nouri-Vaskeh M, Alizadeh L, Hajiasgharzadeh K, Mokhtarzadeh A, Halimi M, Baradaran B. The role of Hsp90 molecular chaperones in hepatocellular carcinoma. *J Cell Physiol* (2020) 235(12):9110–20. doi: 10.1002/jcp.29776

165. Wei W, Liu M, Ning S, Wei J, Zhong J, Li J, et al. Diagnostic value of plasma Hsp90α Levels for detection of hepatocellular carcinoma. *BMC Cancer* (2020) 20(1):6. doi: 10.1186/s12885-019-6489-0

166. Shi Y, Liu X, Lou J, Han X, Zhang L, Wang Q, et al. Plasma levels of heat shock protein 90 Alpha associated with lung cancer development and treatment responses. *Clin Cancer Res* (2014) 20(23):6016–22. doi: 10.1158/1078-0432.Ccr-14-0174

167. Liu W, Li J, Zhang P, Hou Q, Feng S, Liu L, et al. A novel pan-cancer biomarker plasma heat shock protein 90alpha and its diagnosis determinants in clinic. *Cancer Sci* (2019) 110(9):2941–59. doi: 10.1111/cas.14143

168. Meng J, Liu Y, Han J, Tan Q, Chen S, Qiao K, et al. Hsp90β Promoted endothelial cell-dependent tumor angiogenesis in hepatocellular carcinoma. *Mol Cancer* (2017) 16(1):72. doi: 10.1186/s12943-017-0640-9

169. Im CN. Past, present, and emerging roles of mitochondrial heat shock protein trap1 in the metabolism and regulation of cancer stem cells. *Cell Stress Chaperones* (2016) 21(4):553-62. doi: 10.1007/s12192-016-0687-3

170. Ramos Rego I, Silvério D, Eufrásio MI, Pinhanços SS, Lopes da Costa B, Teixeira J, et al. Trap1 is expressed in human retinal pigment epithelial cells and is required to maintain their energetic status. *Antioxidants (Basel)* (2023) 12(2):381. doi: 10.3390/antiox12020381

171. Sun Y, Zou H, Yang L, Zhou M, Shi X, Yang Y, et al. Effect on the liver cancer cell invasion ability by studying the associations between autophagy and trap1 expression. *Oncol Lett* (2018) 16(1):991–7. doi: 10.3892/ol.2018.8774

172. Gasic V, Karan-Djurasevic T, Pavlovic D, Zukic B, Pavlovic S, Tosic N. Diagnostic and therapeutic implications of long non-coding rnas in leukemia. *Life-Basel* (2022) 12(11):1770. doi: 10.3390/life12111770

173. Bruce M. A systematic and conceptual review of posttraumatic stress in childhood cancer survivors and their parents. *Clin Psychol Rev* (2006) 26(3):233–56. doi: 10.1016/j.cpr.2005.10.002

174. Komiyama T, Ogura A, Hirokawa T, Zhijing M, Kamiguchi H, Asai S, et al. Analysis to estimate genetic variations in the idarubicin-resistant derivative molt-3. *Int J Mol Sci* (2017) 18(1):12. doi: 10.3390/ijms18010012

175. Kantarjian HM, Keating MJ, Freireich EJ. Toward the potential cure of leukemias in the next decade. *Cancer* (2018) 124(22):4301–13. doi: 10.1002/cncr.31669

176. Huang Y, Hong X, Hu J, Lu Q. Targeted regulation of Mir-98 on E2f1 increases chemosensitivity of leukemia cells K562/A02. *Oncotargets Ther* (2017) 10:3233–9. doi: 10.2147/ott.S126819

177. Taha EA, Ono K, Eguchi T. Roles of extracellular hsps as biomarkers in immune surveillance and immune evasion. *Int J Mol Sci* (2019) 20(18):4588. doi: 10.3390/ ijms20184588

178. Andersson A, Ritz C, Lindgren D, Eden P, Lassen C, Heldrup J, et al. Microarray-based classification of a consecutive series of 121 childhood acute leukemias: prediction of leukemic and genetic subtype as well as of minimal residual disease status. *Leukemia* (2007) 21(6):1198–203. doi: 10.1038/sj.leu.2404688

179. Dekker FA, Rudiger SGD. The mitochondrial Hsp90 trap1 and alzheimer's disease. Front Mol Biosci (2021) 8:697913. doi: 10.3389/fmolb.2021.697913

180. Mathieu C, Chamayou Q, Hyen Luong TT, Naud D, Mahuteau-Betzer F, Alami M, et al. Synthesis and antiproliferative activity of 6brcaq-tpp conjugates for targeting the mitochondrial heat shock protein trap1. *Eur J Med Chem* (2022) 229:114052. doi: 10.1016/j.ejmech.2021.114052

181. Park H-K, Lee J-E, Lim J, Kang BH. Mitochondrial Hsp90s suppress calciummediated stress signals propagating from mitochondria to the er in cancer cells. *Mol Cancer* (2014) 13(1):148. doi: 10.1186/1476-4598-13-148

182. Costantino E, Maddalena F, Calise S, Piscazzi A, Tirino V, Fersini A, et al. Trap1, a novel mitochondrial chaperone responsible for multi-drug resistance and protection from apoptotis in human colorectal carcinoma cells. *Cancer Lett* (2009) 279 (1):39–46. doi: 10.1016/j.canlet.2009.01.018

183. Pedersen KS, Kim GP, Foster NR, Wang-Gillam A, Erlichman C, McWilliams RR. Phase ii trial of gemcitabine and tanespimycin (17aag) in metastatic pancreatic cancer: A mayo clinic phase ii consortium study. *Investigational New Drugs* (2015) 33 (4):963–8. doi: 10.1007/s10637-015-0246-2

184. Lang SA, Moser C, Gaumann A, Klein D, Glockzin G, Popp FC, et al. Targeting heat shock protein 90 in pancreatic cancer impairs insulin-like growth factor-I receptor signaling, disrupts an interleukin-6/signal-transducer and activator of transcription 3/ hypoxia-inducible factor-1alpha autocrine loop, and reduces orthotopic tumor growth. *Clin Cancer Res* (2007) 13(21):6459–68. doi: 10.1158/1078-0432.Ccr-07-1104

185. Song D, Chaerkady R, Tan AC, García-García E, Nalli A, Suárez-Gauthier A, et al. Antitumor activity and molecular effects of the novel heat shock protein 90 inhibitor, ipi-504, in pancreatic cancer. *Mol Cancer Ther* (2008) 7(10):3275–84. doi: 10.1158/1535-7163.Mct-08-0508

186. Cao X, Bloomston M, Zhang T, Frankel WL, Jia G, Wang B, et al. Synergistic antipancreatic tumor effect by simultaneously targeting hypoxic cancer cells with hsp90 inhibitor and glycolysis inhibitor. *Clin Cancer Res* (2008) 14(6):1831–9. doi: 10.1158/ 1078-0432.Ccr-07-1607

187. Cao X, Jia G, Zhang T, Yang M, Wang B, Wassenaar PA, et al. Non-invasive mri tumor imaging and synergistic anticancer effect of Hsp90 inhibitor and glycolysis inhibitor in rip1-tag2 transgenic pancreatic tumor model. *Cancer chemotherapy Pharmacol* (2008) 62(6):985–94. doi: 10.1007/s00280-008-0688-8

188. Ghadban T, Dibbern JL, Reeh M, Miro JT, Tsui TY, Wellner U, et al. Hsp90 is a promising target in gemcitabine and 5-fluorouracil resistant pancreatic cancer. *Apoptosis* (2017) 22(3):369–80. doi: 10.1007/s10495-016-1332-4

189. Taldone T, Patel PD, Patel M, Patel HJ, Evans CE, Rodina A, et al. Experimental and structural testing module to analyze paralogue-specificity and affinity in the Hsp90 inhibitors series. *J Medicinal Chem* (2013) 56(17):6803–18. doi: 10.1021/jm400619b

190. Chae YC, Angelin A, Lisanti S, Kossenkov AV, Speicher KD, Wang H, et al. Landscape of the mitochondrial Hsp90 metabolome in tumours. *Nat Commun* (2013) 4 (1):2139. doi: 10.1038/ncomms3139

191. Yang S, Yoon NG, Park MA, Yun J, Im JY, Kang BH, et al. Triphenylphosphonium conjugation to a trap1 inhibitor, 2-amino-6-chloro-7,9dihydro-8h-purin-8-one increases antiproliferative activity. *Bioorg Chem* (2022) 126:105856. doi: 10.1016/j.bioorg.2022.105856

192. Lang SA, Moser C, Fichnter-, Schachtschneider P, Hellerbrand C, Schmitz V, et al. Targeting heat-shock protein 90 improves efficacy of rapamycin in a model of hepatocellular carcinoma in mice. *Hepatology* (2009) 49(2):523–32. doi: 10.1002/hep.22685

193. Xue N, Lai F, Du T, Ji M, Liu D, Yan C, et al. Chaperone-mediated autophagy degradation of Igf-1r β Induced by Nvp-Auy922 in pancreatic cancer. *Cell Mol Life Sci* (2019) 76(17):3433–47. doi: 10.1007/s00018-019-03080-x

194. Chen H, Li L-Q, Pan D. Geldanamycin induces apoptosis in human gastric carcinomas by affecting multiple oncogenic kinases that have synergic effects with Tnf-related apoptosis-inducing ligand. *Oncol Lett* (2015) 10(6):3732–6. doi: 10.1002/jcp.20090

195. Wang D-H, Zhang Y-J, Zhang S-B, Liu H, Liu I, Liu F-L, et al. Geldanamycin mediates the apoptosis of gastric carcinoma cells through inhibition of Epha2 protein expression. *Oncol Rep* (2014) 32(6):2429–36. doi: 10.3892/or.2014.3542

196. Goyal L, Wadlow RC, Blaszkowsky LS, Wolpin BM, Abrams TA, McCleary NJ, et al. A phase I and pharmacokinetic study of ganetespib (Sta-9090) in advanced hepatocellular carcinoma. *Investigational New Drugs* (2015) 33(1):128–37. doi: 10.1007/s10637-014-0164-8.5

197. Augello G, Emma MR, Cusimano A, Azzolina A, Mongiovi S, Puleio R, et al. Targeting Hsp90 with the small molecule inhibitor Auy922 (Luminespib) as a treatment strategy against hepatocellular carcinoma. *Int J Cancer* (2019) 144 (10):2613–24. doi: 10.1002/ijc.31963