Check for updates

OPEN ACCESS

EDITED BY Umberto Malapelle, University of Naples Federico II, Italy

REVIEWED BY

Kanagaraj Palaniyandi, SRM Institute of Science and Technology, India Sapna Deo, University of Miami, United States

*CORRESPONDENCE Lianwen Zheng Mavezheng@sohu.com

SPECIALTY SECTION

This article was submitted to Gynecological Oncology, a section of the journal Frontiers in Oncology

RECEIVED 05 January 2023 ACCEPTED 06 March 2023 PUBLISHED 17 March 2023

CITATION

Wang M, Fu L, Xu Y, Ma S, Zhang X and Zheng L (2023) A comprehensive overview of exosome IncRNAs: Emerging biomarkers and potential therapeutics in gynecological cancers. *Front. Oncol.* 13:1138142. doi: 10.3389/fonc.2023.1138142

COPYRIGHT

© 2023 Wang, Fu, Xu, Ma, Zhang and Zheng. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

A comprehensive overview of exosome lncRNAs: Emerging biomarkers and potential therapeutics in gynecological cancers

Min Wang, Lulu Fu, Ying Xu, Shuai Ma, Xueying Zhang and Lianwen Zheng*

Reproductive Medical Center, Department of Obstetrics and Gynecology, The Second Hospital of Jilin University, Changchun, China

Ovarian, endometrial, and cervical cancer are common gynecologic malignancies, and their incidence is increasing year after year, with a younger patient population at risk. An exosome is a tiny "teacup-like" blister that can be secreted by most cells, is highly concentrated and easily enriched in body fluids, and contains a large number of lncRNAs carrying some biological and genetic information that can be stable for a long time and is not affected by ribonuclease catalytic activity. As a cell communication tool, exosome lncRNA has the advantages of high efficiency and high targeting. Changes in serum exosome IncRNA expression in cancer patients can accurately reflect the malignant biological behavior of cancer cells. Exosome lncRNA has been shown in studies to have broad application prospects in cancer diagnosis, monitoring cancer recurrence or progression, cancer treatment, and prognosis. The purpose of this paper is to provide a reference for clinical research on the pathogenesis, diagnosis, and treatment of gynecologic malignant tumors by reviewing the role of exosome lncRNA in gynecologic cancers and related molecular mechanisms.

KEYWORDS

exosome lncRNA, expression, biomarkers, therapeutics, gynecological cancers

1 Introduction

An exosome is a cell-secreted nanoscale vesicle containing DNA, proteins, lipids, RNA, metabolites, cytokines, transcription factor receptors, and other biologically active substances (1). Its composition is similar to that of parental cells and can be used as a "fingerprint" to identify relevant cells and provide specific signals that can be traced in circulating blood (2). The composition is similar to that of parental cells and can be used to identify relevant cells by providing specific signals that can be traced in circulating blood.

Long noncoding RNAs (lncRNAs) are noncoding RNAs that are abundant in the cytoplasm and nucleus (3). They do not have protein-coding functions, but they can influence cancer development in a variety of ways and can be specifically sorted into the exosome (4). Despite the presence of RNA enzymes in the blood, lncRNAs can persist due to exosome protection (5, 6). Tumor-derived exosomes (TDE) lncRNAs can contribute to cancer progression in a variety of ways by altering the tumor microenvironment, the epithelial-mesenchymal transition (EMT), and angiogenesis, as well as playing a role in cancer growth maintenance and stabilization. Because cancer invasion, metastasis, treatment, and drug resistance are all intertwined, it is of great scientific importance to mine and explores the exosome lncRNAs that affect malignant biological behavior, as this can help to further investigate the mechanism of cancer development and provide new ideas and strategies for cancer treatment (7, 8).

2 Overview of the exosome

2.1 Discovery and distribution of exosome

Exosomes were discovered by Johnstone et al. in the study of extracellular cytoplasmic fusion of reticulocyte multivesicular bodies (9), are 30-150 nm in diameter (10), have a phospholipid bilayer structure, and belong to the extracellular vesicle family. Exosomes released from cells into the extracellular compartment are found in a variety of body fluids, including saliva, breast milk, blood, urine, amniotic fluid, and vaginal/alveolar lavage fluid (11). The endosomal sorting complex required for transport (ESCRT) is made up of the complexes ESCRT-0, I, II, and III, as well as coproteins like apoptosis-linked gene 2-interacting protein X (ALIX) and vacuolar protein sorting 4 (VPS4). Several studies have confirmed the importance of ESCRT in exosome biosynthesis (12). Exosome production, on the other hand, is not entirely dependent on ESCRT mechanisms such as the ceramide mechanism. It was discovered that mouse oligodendrocytes secreted lipoprotein-carrying exosomes normally even after ESCRT inhibition and that cellular exosome secretion was reduced after ceramide synthesis inhibition, implying a regulatory role for ceramide in exosome synthesis. Exosomes are produced by cellular self-selection, and exosomes from different cells can carry different "cargo" (13). Under various physiological and pathological conditions, the same cell can produce multiple exosomes containing additional genetic information (14) (Figure 1).

2.2 Functions of exosome

Exosomes can create a pre-metastatic microenvironment suitable for cancer cell growth, regulate the glucose and lipid metabolism of target cells, counteract the body's immune defense, and promote and cooperate with cancer development by transferring lncRNA to recipient cells and mediating material transport and information exchange.

Exosome has been confirmed as a circulating biomarker for various breast, colorectal, and bladder cancers in numerous studies (15). On January 21, 2016, the first exosome-based cancer diagnostic product was launched in the United States (16). The exosome is a natural lipid vesicle that can be used as a gene therapy carrier and has significant development potential in the field of cancer therapy (17). Exosomes can cross the blood-brain barrier and transport drugs and genes (e.g., proteins, lipids, DNA, and RNA) into tissues, effectively preventing their degradation (18). The drugs could not penetrate the blood-brain barrier in the control group of zebrafish embryos treated with conventional drugs, but in the experimental group, in which the anti-cancer drugs adriamycin and paclitaxel were integrated into the exosome and then introduced into zebrafish embryos, large number of exosomes could penetrate the blood-brain barrier and allow the drugs to reach the cancer cells directly (19). In advanced cancers, clinical trials targeting dendritic cell-derived exosomes (DEX) have been conducted (20). Cancer exosomes are known to play an essential role in the distant compartment effect, a recently discovered mechanism that effectively targets cancers and inhibits distant metastasis (21); As a result, the exosome is expected to be a novel and efficient drug delivery system. Exosomes can be used for gene therapy by transfecting siRNA into the exosome and successfully silencing genes using the exosome as a vector, according to research (22).

3 Overview of IncRNAs

3.1 Biogenesis of IncRNAs

The noncoding region of the human genome contains approximately 88% of single nucleotide polymorphisms. Noncoding RNA is classified into two types based on its length: LncRNA and short-stranded noncoding RNA. LncRNA is a class of single-stranded RNA molecules with sizes less than 200 nt, the majority of which are found in the nucleus and some in the cytoplasm, and are classified as sense lncRNA, antisense lncRNA (AS lncRNA), bidirectional lncRNA, intronic lncRNA, and intergenic lncRNA (23). When compared to most protein-coding genes, lncRNAs have better cell specificity and relatively stable local secondary and tertiary structures, making them easier to detect in body fluids and capable of interacting with DNA, RNA, or proteins. They play an essential role in the physiological and pathological processes of the body (24). LncRNAs participate in a variety of biological pathways, including cell growth, by regulating gene transcription and post-translational expression. By regulating innate and adaptive immunity, lncRNAs can participate in a variety of immune pathways, and dysregulation of their expression levels can disrupt immune homeostasis. It is anticipated that it will be one of the most promising biomarkers for disease diagnosis and prognosis (25) (Figure 2).

3.2 LncRNAs are involved in gene expression

LncRNAs are important regulators at the epigenetic, transcriptional, and post-transcriptional levels (26). Epigenetic



silencing or activation of target genes: lncRNAs can regulate gene expression at the epigenetic level *via* DNA methylation, demethylation, histone modification, and chromosome remodeling (27). MEG3 expression was found to be significantly reduced in glioblastoma due to DNA methyltransferase I-mediated hypermethylation of the MEG3 promoter, which downregulated MEG3 expression in glioblastoma and inhibited p53 protein activation.

Transcriptional level: Long noncoding RNAs (lncRNAs) can interact with transcription factors, enhancers, and promoters to regulate RNA transcription, localization, and stability (28). The lncRNA Gas5 can compete with the glucocorticoid response element (GRE) for binding to the glucocorticoid receptor (GR), preventing GR transcriptional activation and resulting in an autoimmune response. In breast cancer, low Gas5 expression increased cancer cells' survival during starvation. P21-associated noncoding RNA with DNA damage activation (lncRNA PANDA) was found to promote osteosarcoma cell proliferation. Further research revealed that the lncRNA PANDA inhibited apoptosis in normal human fibroblasts by binding to transcription factors that prevented it from binding to apoptosis-related gene promoters. Long noncoding RNA homeobox (HOX) A11 antisense lncRNA (HOXA11-AS) was discovered to bind to transcription factor WD repeat domain 5 (WDR5) in the promoter region, promote -catenin transcription, and activate the Wingless-Type MMTV Integration Site Family (WNT) signaling pathway, accelerating cancer metastasis in vivo.

Post-transcriptionally, lncRNAs can form RNA dimers with target mRNAs *via* complementary base pairing, obstruct transcription factor binding, or directly recruit specific translation

repressor proteins to regulate mRNA shearing, translation, and degradation (29). KLF4 is a transcriptional activator of vascular endothelial growth factor (VEGF). The lncRNA H19 can bind to miR-7, allowing miR-competitive endogenous RNA (ceRNA-7) to release translational repression of KLF4 and activate the KLF4/ VEGF signaling pathway. Stable knockdown of exosome lncRNA H19 can significantly affect KLF4 and VEGF mRNA and protein expression levels, which affect the formation of the pre-metastatic microenvironment, inhibit cancer cell migration and invasion, and regulate the tumor microenvironment and vascular normalization. During the progression of hepatocellular carcinoma, the expression level of long noncoding RNA-activated by transforming growth factor beta (lncRNA-ATB) was increased and directly linked to IL-11, which altered IL-11 tertiary structure, increased the stability of IL-11 mRNA, induced IL-11 autocrine, triggered the signal transducer and activator of transcription 3 (STAT3) pathway, and promoted cancer metastasis and organ colonization. The first IncRNA with trans-activation, HOX transcript antisense RNA (HOTAIR), acts as a pro-oncogene in a variety of cancer cells, including breast cancer and hepatocellular carcinoma (30). HOTAIR, a lncRNA with sponge adsorption for miR-122, can regulate cancer cell epithelial-mesenchymal transition (31).

4 Exosome IncRNAs and tumor

The early and precise diagnosis of malignant cancers has become a hot research topic. Cancer occurrence and progression are dependent on the interaction between cancer cells and the tumor microenvironment. In addition to intercellular contact and the



release of soluble factors, cancer cells can communicate with the tumor microenvironment via exosomes (32). The TDE transports molecules such as content DNA, miRNA, and lncRNA that reflect genetic or signaling changes originating in cancer cells (33). lncRNA enters the recipient cells via the exosome and acts as a signaling mediator to coordinate cellular functions among cancer cells, creating a microenvironment conducive to cancer cell metastasis at a distant site (34-36). In cancer progression, lncRNAs can serve two purposes. MALAT1 (metastasis-associated lung adenocarcinoma transcript1) can promote or inhibit breast cancer metastasis by activating or inactivating neighboring prometastatic transcription factors (37). DEAD-box RNA helicase 3 (DDX3) also plays a dual role in the progression of lung cancer. On the one hand, DDX3 can activate the WNT signaling pathway, facilitating lung cancer metastasis. DDX3, on the other hand, can inhibit lung cancer progression by activating the MDM2/Slug/E-cadherin signaling pathway (38). The lncRNA HOTAIR can affect the co-localization and activity of vesicleassociated membrane protein 3 (VAMP3) and synaptosomalassociated protein 23 (SNAP23) to promote the fusion of MVB with the plasma membrane to promote HCC exosome secretion, confirming that lncRNAs have the function of promoting cancer exosome secretion and providing a new idea for the study of cancer lncRNAs (Figure 3).

4.1 Exosome lncRNAs and tumor microenvironment

The tumor microenvironment is made up of a variety of cells, including cancer cells and stromal cells like endothelial cells, fibroblasts, adipocytes, and mesenchymal stem cells (39). Tumorigenesis, progression, and metastasis are all affected by the characteristics of cancer cells as well as the interaction between cancer cells and stromal cells in the tumor microenvironment (40). The exosome, which is released into the extracellular environment *via* paracrine or autocrine signaling pathways and causes receptor cell-related phenotypic changes (27), is a critical communication mediator for primary tumor microenvironment alterations. Different exosome lncRNA sources play different roles (41). Cancer cells and tumor-associated macrophages (TAMs) may be

important sources of exosomes in the tumor microenvironment (42). TDE, by remodeling the extracellular matrix (ECM) and inducing angiogenesis, creates a microenvironment favorable for cancer cell metastasis at distant sites (43). Cancer parenchymal cells use the exosome to transport biogenetic information to the extracellular space, transforming normal stromal cells and promoting cancer cell proliferation, apoptosis, migration, invasion, and prognosis (44, 45). Carcinoma-associated fibroblasts (CAFs), macrophages, and other cells secrete lncRNA-containing exosomes to promote cancer development and malignancy (46). LINC00092 was found to be significantly elevated in paraneoplastic fibroblasts in OC, along with elevated chemokine (C-X-C motif) ligand 14 (CXCL14), which was associated with metastasis and poor prognosis in OC (47).

4.2 Exosome lncRNA and tumor angiogenesis

The formation of neovascularization is an important environment for cancer genesis and development, and blood vessels provide sufficient oxygen and nutrients for cancer cell metastasis and growth (48). TDE can help with cancer angiogenesis and extracellular matrix remodeling by dynamically regulating different cells in the tumor microenvironment. The cancer vasculature is typically disorganized as a result of adjacent cancer cells compressing new blood vessels, resulting in tortuous and malformed vessels. Endothelial cells are loosely connected, and permeability and leakiness increase, allowing cancer cells to spread quickly into the vasculature and then develop distant metastases (49). As a result, anti-cancer cell angiogenesis will emerge as a novel therapeutic strategy. Cancer cells' exosome lncRNA can act on endothelial cells in the microenvironment to promote cancer angiogenesis. Exosomes secreted by cancer stem cells invade endothelial cells, deliver lncRNA H19 to their target cells, and stimulate HUVEC angiogenesis by synthesizing and releasing VEGF (50). In preparation for cancer growth and metastasis, glioma cells were found to promote angiogenesis by increasing the expression of endothelial cell pro-angiogenic factor VEGFA via exosome lncRNA CCAT and lncRNA HOTAIR (51). The exosome IncRNA Small nucleolar RNA host gene 16 (SNHG16)/miR-4500/ N-acetylgalactosamine-transferase 1 (GALNT1) axis has been linked to tumor angiogenesis.

4.3 Exosome lncRNAs and tumor metastasis

Metastasis is a fundamental challenge in cancer therapy because cancer cells and the tumor microenvironment regulate cancer proliferation and metastasis (52). TDE lncRNAs can promote malignant growth by interacting with the microenvironment and cancer cells, resulting in increased cancer proliferation and metastasis (53). During rapid growth, cancer cells cause internal tissue hypoxia and promote upregulation of hypoxia-inducible factor (HIF-1) expression, stimulating cancer cells to secrete



within the cancer microenvironment. Exosomal lncRNAs from cancer cells can promote immune modulation, angiogenesis, cancer proliferation, metastasis, and drug resistance (35).

exosomes with enhanced angiogenic and metastatic potential and promoting cancer invasion and metastasis (54). According to studies, exosome lncRNA 91H is highly expressed in patients' serums with colorectal cancer and usually decreases after surgery. lncRNA 91H has been shown to promote cancer migration and invasion by regulating the expression of heterogeneous nuclear ribonucleoprotein K (HNRNPK) (55). HOTAIR, an exosome lncRNA, has been linked to bladder cancer progression, and knocking it out in uroepithelial bladder cancer cell lines inhibits EMT (56). MALAT1, an exosome-derived lncRNA that promotes cancer cell migration and prevents cancer cell apoptosis, was found to be positively related to the TNM stage and lymph node metastasis in NSCLC (57). It has been demonstrated that exosome-derived epidermal growth factor receptor (EGFR) protein in lung cancer cells induces the formation of tolerogenic dendritic cells (DCs), which in turn inhibits the anti-cancer effects of CD8+ T cells by inducing the production of regulatory T cells (Treg), and ultimately Treg promotes cancer immune escape (58). Elucidating the molecular mechanisms of cancer metastasis may lead to the development of more effective cancer therapeutic strategies (59).

4.4 Exosome IncRNA and cancer drug resistance

It is critical to investigate the specific mechanisms of innate or acquired drug resistance in cancer cells (60); Cancer cells and stromal cells in the tumor microenvironment can help spread cancer drug resistance by secreting exosomes (61). Exosomes can affect cell sensitivity to drugs *via* the following mechanisms (62). Exosomes directly wrap anti-cancer drugs, reducing their effectiveness. Exosomes transport bioactive molecules that compete for binding targets with anti-cancer drugs. Drugresistant cells transmit drug-resistance information to sensitive cells via exosome-derived bioactive small molecules. Drug sensitivity information is transmitted from sensitive cells to drugresistant cells via exosome-derived bioactive small molecules. Resistance to chemotherapeutic drugs could be improved by interfering with receptor cells with lncRNA, which could be a new therapeutic approach (63). lncRNA regulators of reprogramming (ROR) were found to be highly expressed in hepatocellular carcinoma cells (64). Drug resistance was found to be increased when hepatocellular carcinoma cells were treated with exosomes containing a high concentration of lncRNA ROR (65). Infection of lncRNA ROR in hepatocellular carcinoma cells with RNAi resulted in adriamycin sensitivity, and cancer cells may use exosomes and lncRNA to enhance drug resistance in nearby cells. Celastrol is thought to be therapeutic for a variety of cancers. When compared to free celastrol and celastrol exosome preparations, anticancer activity was significantly increased, with no liver or nephrotoxicity (66). Paclitaxel-resistant breast cancer cells can be induced by delivering the lncRNA SNHG15 to sensitive cells via exosomes. The exosome lncRNA KCNQ1OT1 is a critical molecule mediating radiotherapy resistance in lung cancer A549 cells. The use of CAFs as an entry point for reversing radiotherapy resistance in lung cancer cells provides a critical theoretical foundation. Investigating the effect of exosome lncRNAs on drug resistance will aid in elucidating the molecular mechanism of cancer drug resistance and provide new ideas for overcoming or reversing drug resistance (67).

5 Exosome lncRNA and gynecologic malignancies

5.1 Exosome IncRNA and ovarian cancer

Ovarian cancer (OC) is the most difficult to diagnose and has the worst prognosis of all malignant cancers of the female reproductive system, causing serious health problems in women (68). The pathogenesis of OC is complex, the early clinical symptoms are subtle, and the metastatic potential is high. When most patients are diagnosed, they are already in an advanced stage of the disease (69), so radical surgery cannot be used, the treatment effect is inadequate, and more than 70% of OC patients have a recurrence. The Food and Drug Administration (FDA) has approved only two biomarkers, CA125 and HE4, as diagnostic biomarkers for OC (70). CA125 is widely used in clinical settings, but it has some limitations (71). CA125, for example, is less sensitive in early-stage OC and can be elevated in pregnancy, pelvic inflammatory disease, endometriosis, and other conditions. In the presence of conditions such as acute and chronic renal insufficiency, HE4 can also indicate gynecological diseases and abnormal changes (72). As a result, a new reliable marker is required for the early detection of OC (73).

Exosome lncRNA can be used as a non-invasive diagnostic and screening tool, requiring only a small amount of fresh or frozen blood from OC patients and simultaneously analyzing for DNA,

RNA, and protein. Exosomes can be extracted from the urine and blood of OC patients using a human recombinant S100A8 protein aptamer bound to cell membrane HSP70 (74). CD24 was found in exosomes from malignant ascites and in vitro cancer cells. This marker has been used to predict the prognosis of OC, demonstrating the exosome's utility as a minimally invasive biopsy (75). Experiments with magnetic nanobeads revealed that many HER2-positive exosomes were found in the serum of OC patients (76). Chen et al. (77) discovered that CA125 levels were higher in exosomes than in serum, that serum exosome-derived CA125 improved the sensitivity of OC diagnosis, and that serum HE4 combined with exosome CA125 improved the diagnostic efficiency of OC. Zhang et al. (78) examined the circulating exosomes in the plasma of OC patients and identified seven biomarkers with diagnostic ability, including HER2, EGFR, Folate Receptor (FR), CA-125, Epithelial Cell Adhesion Molecule (EpCAM), CD24, and (CD9+CD63), and demonstrated that these exosome biomarkers not only distinguished OC patients from benign subjects but also differentiated early and advanced OC, indicating the MALAT1 is a long noncoding RNA that is involved in the angiogenesis and metastasis of OC. Sun et al. (79) discovered that the lncRNA MALAT1 plays an important role in the development of OC by mediating the Janus kinase 2 (JAK2)/ STAT3 signaling pathway, promoting OC cell proliferation, and inhibiting cancer cell apoptosis. Jin et al. (80) discovered that the lncRNA MALAT1 could increase OC cell proliferation while inhibiting cancer cell apoptosis via the PI3K-protein kinase B (PKB, AKT) signaling pathway, enhancing OC cell invasion, migration, and EMT function. Some researchers discovered that the expression level of serum exosomes (81) was higher when testing the expression of serum exosome MALAT1. MALAT1 expression was significantly higher in epithelial OC patients than in controls, and it was associated with an advanced International Federation of Gynecology and Obstetrics (FIGO) stage, a high histological grade, and lymph node metastasis. Increased serum exosomeMALAT1 expression was associated with a progressive metastatic epithelial OC phenotype and a poor prognosis, suggesting that it could be used as a prognostic or predictive biomarker for epithelial OC. The HOXA transcript at the distal tip (HOTTIP), a homeobox lncRNA, is critical in the progression of OC. HOTTIP overexpression was found to increase IL-6 expression and secretion in OC cells. IL-6 activated the STAT3 pathway by binding to IL-6 receptors on the surface of neutrophils surrounding cancer cells, increasing the expression of PD-L1 on the surface of neutrophils, inhibiting T cell activity further, accelerating OC immune escape, and ultimately promoting cancer cell growth and metastasis (82). The lncRNA NEAT1 was found to be significantly overexpressed in ovarian cancer cells compared to normal human ovarian epithelial cells. Through sponge adsorption of miR-36, lncRNA NEAT1 may promote ovarian cancer cell proliferation by upregulating fibroblast growth factor (FGF) 9.

The use of exosomes for vaccine preparation is a novel approach in cancer immunotherapy. TDE has low immunogenicity, a low drug attrition rate, and easy tissue diffusion, making it suitable for use as a drug or gene carrier for targeting OC and as a cancer vaccine to inhibit cancer growth. The cytotoxicity of paclitaxelloaded macrophage exosomes against drug-resistant P-gp transfected Manin-Darby canine kidney epithelial cells (MDCKMDR1) cell line was increased more than 50-fold, and the anti-cancer effect of the drug-loaded exosomes was demonstrated (83). Farrukh et al. (84) discovered that exosome delivery of anthocyanin had a strong therapeutic effect on both drug-sensitive and drug-resistant human ovarian cancer cells and that its therapeutic activity was synergistically enhanced when combined with cisplatin. The co-culture of the hypoxic OC cell line exosome (HEX) with cancer cells during cisplatin treatment improved cell survival, according to Kalpana et al. (85). Simultaneously, a known inhibitor, STAT3, inhibited exosome release. Exosome release and cisplatin treatment increased apoptosis, indicating that HEX can promote OC metastasis and increase chemoresistance, and could be a new mechanism for cancer metastasis and chemoresistance, as well as a therapeutic intervention to improve clinical outcomes.

5.2 Exosome IncRNA and endometrial carcinoma

Endometrial carcinoma (EC) is one of the most common malignant cancers of the female reproductive system (86), accounting for 20%-30% of all malignancies of the female genital tract. In recent years, the incidence of EC has been increasing year after year, and the age of onset has gotten younger. EC can be diagnosed clinically based on symptoms such as vaginal bleeding or increased fluid discharge, but a definitive diagnosis requires further examination improvement. Fear of diagnostic scraping and hysteroscopy causes some patients to postpone their investigation, delaying the best time for diagnosis and treatment. Those who do not receive timely treatment at an early stage frequently have poor prognoses and survival rates. When the presence of lesions in the endometrium is determined through diagnostic scraping, the distribution of lesions cannot be accurately grasped, and small local lesions may be missed, increasing the rate of EC misdiagnosis. The clinical treatment of EC is primarily surgical, with the decision to combine radiotherapy based on high-risk factors. There are few adjuvant treatment options for advanced and recurrent cancers. As a result, identifying practical early diagnostic markers and precise therapeutic targets is critical.

Exosome lncRNA regulates EC proliferation and invasion primarily through angiogenesis, EMT, and immune regulation, among other things. Exosome lncRNA promotes the formation of a tumor microenvironment by transforming related cells, which not only speeds up normal cell proliferation but also changes the biological characteristics of nearby and distant non-cancer cells, allowing cancer cells to spread. Through related signaling pathways, some lncRNAs can effectively promote EC cell proliferation and enhance EC cell invasion, migration, and EMT function, thereby promoting cancer growth (87). Some lncRNAs, on the other hand, can effectively inhibit cancer cell apoptosis *via* related signaling pathways, which may be related to the composition of the tumor microenvironment, particularly CAFs (88).

MEG3 is a long noncoding RNA with anti-cancer properties (89). Reduced expression of MEG3, which inhibits cancer cell proliferation, migration, and invasion and promotes apoptosis, has been linked to cancer development and progression (90, 91). MEG3 activity is controlled by both TP53-dependent and TP53independent mechanisms. The TP53 gene is mutated in the majority of human cancers, and it functions as a transcription factor, controlling the expression of many target genes and thus inhibiting cancer development and growth. The differential expression of TP53 in normal and cancerous tissues suggests that MEG3 could be used to assess cancer staging and prognosis (92). Guo et al. examined the expression of MEG3 and Notch signaling molecules in EC tissues and cell lines using real-time quantitative PCR and Western blotting. MEG3 expression was found to be significantly downregulated in EC tissues, whereas Notch protein expression was found to be upregulated in both. MEG3 downregulation inhibits EC proliferation by inhibiting the Notch signaling pathway (93). In EC patients, low expression of exosome IncRNA MEG3 in plasma predicts more high-risk factors, a higher recurrence rate, and a worse prognosis. By comparing the serum expression levels of lncRNA ROR and miR-29 in EC patients and healthy women and using ROC curves to assess the diagnostic value of both in EC, Serum lncRNA ROR and miR-29 levels were found to be significantly higher in EC patients than in healthy women. The expression levels in patients with TNM stages I-II increased dramatically, indicating the combined serum level (94). MALAT1 overexpression is associated with a poor prognosis for EC, implying that MALAT1 could be used as a novel biomarker and diagnostic target for EC (95).

The primary issue with cancer drug therapy is cancer drug resistance, particularly in recurrent cancers where acquired drug resistance renders the therapeutic effect ineffective. Exosome-mediated lncRNA communication in the tumor microenvironment has been shown in studies to be one of the reasons for increased drug resistance. It is possible to inhibit the production or uptake of an exosome-carrying "oncogene" and promote the production or uptake of an exosome-carrying "oncogene" based on the fact that exosomes can transport proteins and nucleic acids related to cancer invasion, metastasis, angiogenesis, and drug resistance. This opens up a new avenue for the future use of exosomes in the treatment of EC. The engineered exosome is more effective in targeting therapy than the original exosome, and it also reduces cytotoxicity and significantly inhibits tumor growth (96). Exosome lncRNAs play an important role in the development of EC, opening up new avenues for the early diagnosis and treatment of EC patients. They may also become an important tool in monitoring the progression and prognosis of EC in the future.

5.3 Exosome IncRNA and cervical cancer

Cervical cancer (CC) is one of the most common cancers in women. According to data, more than 500,000 people are diagnosed with CC each year, with the majority of deaths occurring in developing countries (97, 98). As a result, CC is a global public health issue that should not be underestimated (99). Despite the fact that chemotherapy combined with targeted therapy can improve overall survival (OS), progression-free survival (PFS), and cancer mortality in CC patients, cancer incidence continues to rise. The prognosis of advanced CC, which has a low local control rate and is prone to distant metastasis, is influenced by high-risk factors. The treatment effect is frequently poor, with only a 60% 5-year survival rate (100). As a result, improving early diagnosis of CC, identifying therapeutic targets, and investigating biomarkers that can indicate prognosis have emerged as top priorities in CC basic and clinical research (101).

Because of the impact of exosome LncRNA on the tumor microenvironment and its biological properties, it has the potential to become a cancer biomarker for CC patients, which has important clinical implications in cancer screening, treatment detection, and prognosis evaluation (102). Exosomes from HeLa cells in CC have been shown to promote distant metastasis by inducing endothelial cell endoplasmic reticulum stress and disrupting vascular endothelial cell integrity, thereby disrupting tight endothelial junctions (103). When CC HeLa cell exosomes were injected into mice, they found increased vascular permeability and cancer metastasis. The primary mechanism involved the CC HeLa cell exosome regulating the expression of closed junction proteins. Immunity against CC was improved in a mouse model of CC by increasing the cytotoxic activity of DEX-induced CD8+ T cells against cancer cells, prompting CD8+ T cell proliferation, and increasing IFN secretion (104).

HOXA11 is a recently discovered and researched lncRNA (105). HOXA11-AS has been shown in studies to promote cancer cell proliferation by regulating the expression of miR-124, miR-140-5p, LATS1, PADI2, and other genes (106-108). Exosome lncRNA HOXA11-AS may increase the expression of SRY-related highmobility group box 4 (SOX4) in endothelial cells, increasing the proliferative capacity of endothelial cells involved in cervical cancer. According to the ROC curve, the specificity of lncRNA gradually increased during hepatocarcinogenesis (GIHCG), and the sensitivity was 88.75% in distinguishing between healthy people and CC patients. In the future, lncRNA GIHCG could be used to predict CC (109). HOTAIR and MALAT1 lncRNAs were found to be significantly overexpressed in exosomes isolated from the lavage fluid of CC patients (111.112). The lncRNA MEG3 was found to be significantly reduced and correlated with cancer stage, metastasis, and other factors. Chen et al. demonstrated that MEG3 can inhibit cervical cancer cell proliferation, invasion, and migration by regulating the Rac1 and PI3K/AKT/MMP-2/9 signaling pathways (110). When compared to non-neoplastic cervical tissues, the expression of lncRNA MEG3 was significantly downregulated in the histopathological grading of cervical intraepithelial neoplasia (CIN) in CIN 2 and CIN3. According to one study, MEG3 expression was reduced in cervical tissues, and it was associated with cancer size, lymph node metastasis, high-risk HPV infection, and the FIGO stage. In vitro, ectopic expression of MEG3 may inhibit the proliferation of human CC cells HeLa and C-33A. The researchers discovered that NF-kappaB interacting lncRNA (NKILA) inhibits proliferation and promotes apoptosis in cervical squamous cells by down-regulating miRNA-21 expression.

LncRNA ArfGAP with the RhoGAP domain, ankyrin repeat, and PH domain 1 antisense RNA (ARAP1-AS1) can promote protooncogene c-Myc translation in cervical cancer by separating dimers and promoting tumorigenesis (59). Furthermore, through interactions with recombinant Polypyrimidine Tract Binding Protein 1 (PTBP1), LncRNA surfactant associated 1 (SFTA1P) promoted the degradation of tropomyosin 4 (TPM4) mRNA and the progression of cervical cancer (52). These findings support MEG3's critical role in the molecular etiology of CC and point to MEG3's potential use in the treatment of CC (111) (Table 1).

6 Conclusion

Exosomal lncRNA has a wide range of research applications (114). Exosome lncRNAs regulate a variety of pathophysiological processes, including cancer cell genesis, invasion, metastasis, and vascular neogenesis, as well as mediate cancer drug resistance and

play an important role in cancer development. Exosome lncRNA is thought to be a novel marker for gynecologic cancer diagnosis, efficacy evaluation, and prognosis prediction. Exosome lncRNA research is still in its early stages, and its functions are not fully understood. There are the following flaws: The specificity of exosome lncRNA as a molecular marker for gynecological tumor diagnosis has yet to be determined; The technology of exosome in vitro synthesis as a carrier of targeted therapeutic drugs has yet to be improved; Exosome lncRNAs play a role in a variety of cancers, and multiple exosome lncRNAs have the same cancer action target, but their interaction is still unknown, making it difficult to fully resolve their regulatory network. The above problems can be solved one by one with the rapid development of proteomics, high-throughput sequencing, transcriptomics, and bioinformatics analysis, and researchers will have a better understanding of the mechanisms of exosome-derived lncRNAs in the development of gynecological malignancies and their clinical applications.

TABLE 1 The expression and functions of exosomal lncRNAs in gynecological cancers.

Cancer types	Specimen source	Exosomal IncRNAs	Functions	References
OC	Cells	LINC00092 Metastasis		(47)
OC	Cells	MALAT1	Proliferation	(79)
OC	Cells	HOTTIP	Metastasis	(82)
OC	Cells	MEG3	Drug resistance	(36)
OC	Cells	GIHCG	Proliferation	(34)
OC	Cells	PTAR	Metastasis	(34)
OC	Cells	MORT	proliferation	(73)
OC	Cells	HOTAIR	Metastasis	(86)
OC	Cells	NEAT1	proliferation	(86)
OC	Cells	H19	Proliferation	(86)
OC	Cells	HOXA11 Biomarkers		(105)
EC	Cells	MEG3 Proliferation		(93)
EC	Cells	ROR	Proliferation	(94)
EC	Cells	MALAT1	Biomarkers	(95)
EC	Cells	DLEU1	Metastasis	(87)
EC	Cells	HOTAIR	Metastasis	(86)
EC	Cells	NEAT1	Metastasis	(86)
EC	Cells	H19	Proliferation	(86)
CC	Cells	ARAP1-AS1 Proliferation		(59)
CC	Cells	NKILA Proliferation		(59)
CC	Cells	MORT	proliferation	(73)
CC	Cells	HOXA11	Proliferation	(105)
CC	Cells	GIHCG	Biomarkers	(109)

(Continued)

TABLE 1 Continued

Cancer types	Specimen source	Exosomal IncRNAs	Functions	References
CC	Vaginal Douche	HOTAIR	Metastasis	(112)
CC	Vaginal Douche	MALAT1	Metastasis	(113)
CC	Vaginal Douche	MEG3	Metastasis	(110)
CC	Cells	LINC01305	Metastasis	(102)
CC	Cells	NEAT1	Proliferation	(86)
CC	Serum	H19	Biomarkers	(101)
CC	Cells	SPRY4-IT1	Proliferation	(111)
CC	Cells	GAS5	Proliferation	(111)
CC	Cells	PVT1	Proliferation	(111)
CC	Cells	LINC00675	Metastasis	(111)
CC	Cells	CCST1	Proliferation	(111)
CC	Cells	ZEB1-AS1	Metastasis	(111)

Author contributions

MW, LF, YX, SM, XZ, and LZ performed literature searches and selected the studies and reviews discussed in the manuscript. The first draft of the manuscript was prepared by MW, LF, YX, SM, and XZ and made subsequent amendments. LZ revised the manuscript. All authors contributed to the article and approved the submitted version.

Funding

Funding for this work was provided by the Natural Science Foundation of Jilin Province (No.20210101268JC and No.20210101462JC).

References

1. Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science* (2020) 367(6478):640. doi: 10.1126/science.aau6977

2. Simona F, Laura S, Simona T, Riccardo A. Contribution of proteomics to understanding the role of tumor-derived exosomes in cancer progression: state of the art and new perspectives. *Proteomics* (2013) 14(10-11):1581–94. doi: 10.1002/pmic.201200398

3. Wu P, Mo Y, Peng M, Tang T, Zhong Y, Deng X, et al. Emerging role of tumorrelated functional peptides encoded by lncRNA and circRNA. *Mol Cancer* (2020) 19 (1):22. doi: 10.1186/s12943-020-1147-3

4. Schmitz SU, Grote P, Herrmann BG. Mechanisms of long noncoding RNA function in development and disease. *Cell Mol Life Sci* (2016) 73(13):2491–509. doi: 10.1007/s00018-016-2174-5

5. Zhan Y, Du L, Wang L, Jiang X, Zhang S, Li J, et al. Expression signatures of exosomal long noncoding RNAs in urine serve as novel non-invasive biomarkers for diagnosis and recurrence prediction of bladder cancer. *Mol Cancer* (2018) 17(1):142. doi: 10.1186/s12943-018-0893-y

6. Takahashi K, Ota Y, Kogure T, Suzuki Y, Iwamoto H, Yamakita K, et al. Circulating extracellular vesicle-encapsulated HULC is a potential biomarker for human pancreatic cancer. *Cancer Sci* (2020) 111(1):98–111. doi: 10.1111/cas.14232

7. Morse MA, Garst J, Osada T, Khan S, Hobeika A, Clay TM, et al. A phase I study of dexosome immunotherapy in patients with advanced non-small cell lung cancer. *J Trans Med* (2005) 3(1):9. doi: 10.1186/1479-5876-3-9

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

8. Cheng J, Meng JL, Zhu L, Peng Y. Exosomal noncoding RNAs in glioma: biological functions and potential clinical applications. *Mol Cancer* (2020) 19(1):66. doi: 10.1186/s12943-020-01189-3

9. Pan BT, Johnstone RM. Fate of the transferrin receptor during maturation of sheep reticulocytes *in vitro*: selective externalization of the receptor. *Cell* (1983) 33 (3):967–78. doi: 10.1016/0092-8674(83)90040-5

10. Farooqi AA, Desai NN, Qureshi MZ, Nogueira Librelotto DR, Gasparri ML, Bishayee A, et al. Exosome biogenesis, bioactivities and functions as new delivery systems of natural compounds. *Biotechnol Adv: Int Rev J* (2018) 36(1):328–34. doi: 10.1016/j.biotechadv.2017.12.010

11. Hannafon BN, Gin AL, Xu YF, Bruns M, Calloway CL, Ding WQ. Metastasisassociated protein 1 (MTA1) is transferred by exosomes and contributes to the regulation of hypoxia and estrogen signaling in breast cancer cells. *Cell Communi Signaling CCS* (2019) 17:13. doi: 10.1186/s12964-019-0325-7

12. Henne WM, Stenmark H, Emr SD. Molecular mechanisms of the membrane sculpting ESCRT pathway. *Cold Spring Harbor Perspect Biol* (2013) 5(9):a016766. doi: 10.1101/cshperspect.a016766

13. Pegtel DM, Gould SJ. Exosomes. Annu Rev Biochem (2019) 88(1):487–514. doi: 10.1146/annurev-biochem-013118-111902

14. van Niel G, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nat Rev Mol Cell Biol* (2018) 19(4):213-28. doi: 10.1038/nrm.2017.125

15. Melo S, Sugimoto H, O'Connell J, Kato N, Villanueva A, Vidal A, et al. Cancer exosomes perform cell-independent MicroRNA biogenesis and promote tumorigenesis. *Cancer Cell* (2014) 26(5):707–21. doi: 10.1016/j.ccell.2014.09.005

16. Sheridan C. Exosome cancer diagnostic reaches market. *Nat Biotechnol* (2016) 34(4):358–9. doi: 10.1038/nbt0416-359

17. Sun ZQ, Yang SX, Zhou QB, Wang GX, Song JM, Li Z, et al. Emerging role of exosome-derived long noncoding RNAs in tumor microenvironment. *Mol Cancer* (2018) 17(1):82. doi: 10.1186/s12943-018-0831-z

18. Hu Y, Rao SS, Wang ZX, Cao J, Tan YJ, Luo J, et al. Exosomes from human umbilical cord blood accelerate cutaneous wound healing through miR-21-3p-mediated promotion of angiogenesis and fibroblast function. *Theranostics* (2018) 8 (1):169–84. doi: 10.7150/thno.21234

19. Yang TZ, Martin P, Fogarty B, Brown A, Schurman K, Phipps R, et al. Exosome delivered anti-cancer drugs across the blood-brain barrier for brain cancer therapy in danio rerio. *Pharm Res* (2015) 32(6):2003–14. doi: 10.1007/s11095-014-1593-y

20. Viaud S, Terme M, Flament C, Taieb J, André F, Novault S, et al. Dendritic cellderived exosomes promote natural killer cell activation and proliferation: A role for NKG2D ligands and IL-15Rα. *PloS One* (2009) 4(3):e4942. doi: 10.1371/ journal.pone.0004942

21. Shan ZC, Wang HM, Zhang YJ, Min WP. The role of tumor-derived exosomes in the abscopal effect and immunotherapy. *Life-Basel* (2021) 11(5):381. doi: 10.3390/ life11050381

22. Luan X, Sansanaphongpricha K, Myers I, Chen H, Yuan H, Sun D. Engineering exosomes as refined biologica nanoplatforms for drug delivery. *Acta Pharmacol Sin* (2017) 38):763. doi: 10.1038/aps.2017.12

23. St Laurent G, Wahlestedt C, Kapranov P. The landscape of long noncoding RNA classification. *Trends Genet* (2015) 31(5):239–51. doi: 10.1016/j.tig.2015.03.007

24. Bonasio R, Shiekhattar R. Regulation of transcription by long noncoding RNAs. Annu Rev Genet (2014) 48:433–55. doi: 10.1146/annurev-genet-120213-092323

25. Chen S, Shen X. Long noncoding RNAs: functions and mechanisms in colon cancer. *Mol Cancer* (2020) 19(1):167. doi: 10.1186/s12943-020-01287-2

26. de Almeida VD. Role of IncRNA alterations in cervical oncogenesis. Eurasian J Med Oncol (2022) 6(2):111-20. doi: 10.14744/ejmo.2022.74937

27. Zhao W, Liu Y, Zhang C, Duan C. Multiple roles of exosomal long noncoding RNAs in cancers. *BioMed Res Int* (2019) 2019:1–12. doi: 10.1155/2019/1460572

28. Brandenburger T, Somoza AS, Devaux Y, Lorenzen JM. Noncoding RNAs in acute kidney injury. *Kidney Int* (2018) 94(5):870-81. doi: 10.1016/j.kint.2018.06.033

29. Tripathi V, Ellis JD, Shen Z, Song DY, Prasanth KV. The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell* (2010) 39(6):925-38. doi: 10.1016/j.molcel.2010.08.011

30. Wu YS, Zhang L, Wang Y, Li H, Ren XB, Wei F, et al. Long noncoding RNA HOTAIR involvement in cancer. *Tumor Biol* (2014) 35(10):9531-8. doi: 10.1007/s13277-014-2523-7

31. Cheng D, Deng JG, Zhang B, He XY, Meng Z, Li GL, et al. LncRNA HOTAIR epigenetically suppresses miR-122 expression in hepatocellular carcinoma *via* DNA methylation. *Ebiomedicine* (2018) 36:159–70. doi: 10.1016/j.ebiom.2018.08.055

32. Kirave P, Gondaliya P, Kulkarni B, Rawal R, Kalia K. Exosome mediated miR-155 delivery confers cisplatin chemoresistance in oral cancer cells via epithelialmesenchymal transition. *Oncotarget* (2020) 11(13):1157-71. doi: 10.18632/ oncotarget.27531

33. Zhang L, Yu D. Exosomes in cancer development, metastasis, and immunity. *Biochim Biophys Acta (BBA) - Rev Cancer* (2019) 1871(2):455–68. doi: 10.1016/ j.bbcan.2019.04.004

34. Han SQ, Qi YQ, Luo YM, Chen XP, Liang HF. Exosomal long noncoding RNA: Interaction between cancer cells and non-cancer cells. *Front Oncol* (2021) 10. doi: 10.3389/fonc.2020.617837

35. Zhang WW, Wang QS, Yang Y, Zhou SY, Zhang P, Feng TB. The role of exosomal lncRNAs in cancer biology and clinical management. *Exp Mol Med* (2021) 53 (11):1669–73. doi: 10.1038/s12276-021-00699-4

36. Zhang J, Liu JY, Xu XY, Li L. Curcumin suppresses cisplatin resistance development partly *via* modulating extracellular vesicle-mediated transfer of MEG3 and miR-214 in ovarian cancer. *Cancer Chemother Pharmacol* (2017) 79(3):479–87. doi: 10.1007/s00280-017-3238-4

37. Kim J, Piao HL, Kim BJ, Yao F, Han Z, Wang Y, et al. Long noncoding RNA MALAT1 suppresses breast cancer metastasis. *Nat Genet* (2018) 50:1705–15. doi: 10.1038/s41588-018-0252-3

38. Bol GM, Xie M, Raman V. DDX3, a potential target for cancer treatment. *Mol Cancer* (2015) 14:188. doi: 10.1186/s12943-015-0461-7

39. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. Cell (2011) 144(5):646-74. doi: 10.1016/j.cell.2011.02.013

40. Wu Q, Wu X, Xiang Y, Zhu Q, Wang X. Suppression of endothelial cell migration by tumor associated macrophage-derived exosomes is reversed by epithelial ovarian cancer exosomal lncRNA. *Cancer Cell Int* (2017) 17(1):62. doi: 10.1186/s12935-017-0430-x

41. Chen C-W, Fu M, Du Z-H, Zhao F, Yang W-W, Xu L-H, et al. Long noncoding RNA MRPL23-AS1 promoteoid cystic carcinoma lung metastasis. *Cancer Res* (2020) 80 (11):2273-85. doi: 10.1158/0008-5472.CAN-19-0819

42. Dong R, Zhang B, Tan B, Lin N. Long noncoding RNAs as the regulators and targets of macrophage M2 polarization. *Life Sci* (2021) 266:118895. doi: 10.1016/j.lfs.2020.118895

43. Guo H, Chitiprolu M, Roncevic L, Javalet C, Hemming FJ, Trung MT, et al. Atg5 disassociates the V1V0-ATPase to promote exosome production and tumor metastasis independent of canonical macroautophagy. *Dev Cell* (2017) 43(6):716–30.E7. doi: 10.1016/j.devcel.2017.11.018

44. Hoshino A, Costa-Silva B, Shen T-L, Rodrigues G, Hashimoto A, Mark MT, et al. Tumour exosome integrins determine organotropic metastasis. *Nature* (2015) 527 (7578):329–35. doi: 10.1038/nature15756

45. Rashed MH, Bayraktar E, Helal GK, Abd-Ellah MF, Amero P, Chavez-Reyes A, et al. Exosomes: From garbage bins to promising therapeutic targets. *Int J Mol Sci* (2017) 18(3):538. doi: 10.1158/0008-5472.CAN-16-1615

46. Bradford JR, Cox A, Bernard P, Camp NJ. Consensus analysis of whole transcriptome profiles from two breast cancer patient cohorts reveals long noncoding RNAs associated with intrinsic subtype and the tumour microenvironment. *PloS One* (2016) 11(9):e0163238. doi: 10.1371/journal.pone.0163238

47. Zhao L, Ji G, Le X, Wang C, Xu L, Feng M, et al. Long noncoding RNA LINC00092 acts in cancer-associated fibroblasts to drive glycolysis and progression of ovarian cancer. *Cancer Res* (2017) 77(6):1369–82. doi: 10.1002/jcp.10133

48. Lang HL, Hu GW, Chen Y, Liu Y, Tu W, Lu YM, et al. Glioma cells promote angiogenesis through the release of exosomes containing long noncoding RNA POU3F3. *Eur Rev Med Pharmacol Sci* (2017) 21(5):959–72. doi: 10.1186/s12943-015-0426-x

49. Martin TA, Mansel RE, Jiang WG. Antagonistic effect of NK4 on HGF/SF induced changes in the transendothelial resistance (TER) and paracellular permeability of human vascular endothelial cells. *J Cell Physiol* (2002) 192(3):268–75. doi: 10.3892/ or.2017.5742

50. Conigliaro A, Costa V, Lo Dico A, Saieva L, Buccheri S, Dieli F, et al. CD90+liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lncRNA. *Mol Cancer* (2015) 14:155. doi: 10.1038/s41419-022-05359-7

51. Lang H-L, Hu G-W, Zhang B, Kuang W, Chen Y, Wu L, et al. Glioma cells enhance angiogenesis and inhibit endothelial cell apoptosis through the release of exosomes that contain long noncoding RNA CCAT2. *Oncol Rep* (2017) 38(2):785–98. doi: 10.1186/s12943-021-01411-w

52. Luo AR, Lan XX, Qiu QZ, Zhou Q, Li J, Wu MT, et al. LncRNA SFTA1P promotes cervical cancer progression by interaction with PTBP1 to facilitate TPM4 mRNA degradation. *Cell Death Dis* (2022) 13(11):936. doi: 10.1007/s10555-019-09783-8

53. Jiang CY, Zhang N, Hu XL, Wang HY. Tumor-associated exosomes promote lung cancer metastasis through multiple mechanisms. *Mol Cancer* (2021) 20(1):117. doi: 10.1186/s12935-018-0506-2

54. Logozzi M, Spugnini E, Mizzoni D, Di Raimo R, Fais S. Extracellular acidity and increased exosome release as key phenotypes of malignant tumors. *Cancer Metastasis Rev* (2019) 38(1-2):93–101. doi: 10.1371/journal.pone.0147236

55. Gao T, Liu X, He B, Nie Z, Zhu C, Zhang P, et al. Exosomal lncRNA 91H is associated with poor development in colorectal cancer by modifying HNRNPK expression. *Cancer Cell Int* (2018) 18(1):11. doi: 10.1016/j.bbrc.2017.06.055

56. Berrondo C, Flax J, Kucherov V, Siebert A, Osinski T, Rosenberg A, et al. Expression of the long noncoding RNA HOTAIR correlates with disease progression in bladder cancer and is contained in bladder cancer patient urinary exosomes. *PloS One* (2016) 11(1):e0147236. doi: 10.3109/07357907.2013.789905

57. Zhang R, Xia Y, Wang Z, Zheng J, Chen Y, Li X, et al. Serum long non coding RNA MALAT-1 protected by exosomes is up-regulated and promotes cell proliferation and migration in non-small cell lung cancer. *Biochem Biophys Res Commun* (2017) 490 (2):406–14. doi: 10.1186/s12967-021-02705-9

58. Huang SH, Li Y, Zhang J, Rong J, Ye S. Epidermal growth factor receptorcontaining exosomes induce tumor-specific regulatory T cells. *Cancer Invest* (2013) 31 (5):330–5. doi: 10.1158/0008-5472.CAN-14-3525

59. Zhong QH, Lu MZ, Yuan WQ, Cui YY, Ouyang HQ, Fan Y, et al. Eight-IncRNA signature of cervical cancer were identified by integrating DNA methylation, copy number variation and transcriptome data. *J Trans Med* (2021) 19(1):58. doi: 10.1016/j.canlet.2017.10.040

60. Ran S. The role of TLR4 in chemotherapy-driven metastasis. *Cancer Res* (2015) 75(12):2405–10. doi: 10.1186/s12943-019-0991-5

61. Fan Q, Yang L, Zhang XD, Peng XQ, Wei SB, Su DM, et al. The emerging role of exosome-derived noncoding RNAs in cancer biology. *Cancer Letters* (2018) 414:107–15. doi: 10.1016/j.ijbiomac.2018.12.176

62. Mashouri L, Yousefi H, Aref AR, Ahadi AM, Molaei F, Alahari SK. Exosomes: composition, biogenesis, and mechanisms in cancer metastasis and drug resistance. *Mol Cancer* (2019) 18(1):75. doi: 10.1016/j.gene.2019.144044

63. Gao J, Liu L, Li G, Cai M, Tan C, Han X, et al. LncRNA GAS5 confers the radio sensitivity of cervical cancer cells via regulating miR-106b/IER3 axis. *Int J Biol Macromol* (2018) 126:994–1001. doi: 10.1016/j.fob.2014.04.007

64. He XQ, Yu JJ, Xiong L, Liu YS, Fan L, Li Y, et al. Exosomes derived from liver cancer cells reprogram biological behaviors of LO2 cells by transferring linc-ROR. *Gene* (2019) 719:144044. doi: 10.1016/j.yexmp.2016.05.013

65. Takahashi K, Yan IK, Kogure T, Haga H, Patel T. Extracellular vesicle-mediated transfer of long noncoding RNA ROR modulates chemosensitivity in human hepatocellular cancer. *FEBS Open Bio.* (2014) 4:458–67. doi: 10.1186/s13046-018-0845-9

66. Aqil F, Kausar H, Agrawal AK, Jeyabalan J, Kyakulaga AH, Munagala R, et al. Exosomal formulation enhances therapeutic response of celastrol against lung cancer. *Exp Mol Pathol* (2016) 101(1):12–21. doi: 10.1016/j.yexmp.2016.05.013

67. Kang M, Ren MP, Li Y, Fu YQ, Deng MM, Li CP. Exosome-mediated transfer of IncRNA PART1 induces gefitinib resistance in esophageal squamous cell carcinoma via functioning as a competing endogenous RNA. J Exp Clin Cancer Res (2018) 37(1):171. doi: 10.1186/s13046-018-0845-9

68. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *Ca-a Cancer J Clin* (2021) 71(3):209–49. doi: 10.3322/caac.21660

69. Torre LA, Trabert B, DeSantis CE, Miller KD, Samimi G, Runowicz CD, et al. Ovarian cancer statistics, 2018. *CA A Cancer J Clin* (2018) 68(4):284–96. doi: 10.3322/ caac.21456

70. Montagnana M, Benati M, Danese E. Circulating biomarkers in epithelial ovarian cancer diagnosis: from present to future perspective. *Ann Trans Med* (2017) 5(13):276. doi: 10.21037/atm.2017.05.13

71. Dochez V, Caillon H, Vaucel E, Dimet J, Winer N, Ducarme G. Biomarkers and algorithms for diagnosis of ovarian cancer: CA125, HE4, RMI and ROMA, a review. J Ovarian Res (2019) 12(1):28. doi: 10.1186/s13048-019-0503-7

72. Wang LS, Sun YH, Cai XA, Fu GF. The diagnostic value of human epididymis protein 4 as a novel biomarker in patients with renal dysfunction. *Int Urol Nephrol* (2018) 50(11):2043–8. doi: 10.1007/s11255-018-1930-x

73. Di Fiore R, Suleiman S, Drago-Ferrante R, Felix A, O'Toole SA, O'Leary JJ, et al. LncRNA MORT (ZNF667-AS1) in cancer-is there a possible role in gynecological malignancies? *Int J Mol Sci* (2021) 22(15):7829. doi: 10.3390/ijms22157829

74. Gobbo J, Marcion G, Cordonnier M, Dias AMM, Pernet N, Hammann A, et al. Restoring anti-cancer immune response by targeting tumor-derived exosomes with a HSP70 peptide aptamer. *J Natl Cancer Inst.* 108(3):11. doi: 10.1093/jnci/djv330

75. Nakamura K, Sawada K, Kobayashi M, Miyamoto M, Shimizu A, Yamamoto M, et al. Role of the exosome in ovarian cancer progression and its potential as a therapeutic target. *Cancers* (2019) 11(8):1147. doi: 10.3390/cancers11081147

76. Kabe Y, Suematsu M, Sakamoto S, Hirai M, Koike I, Hishiki T, et al. Development of a highly sensitive device for counting the number of disease-specific exosomes in human sera. *Clin Chem* (2018) 64(10):1463–73. doi: 10.1039/C9SC00961B

77. Chen ZX, Liang QX, Zeng H, Zhao Q, Guo ZD, Zhong RH, et al. Exosomal CA125 as a promising biomarker for ovarian cancer diagnosis. J Cancer (2020) 11 (21):6445–53. doi: 10.2147/OTT.S214689

78. Zhang P, Zhou X, Zeng Y. Multiplexed immunophenotyping of circulating exosomes on nano-engineered ExoProfile chip towards early diagnosis of cancer. *Chem Sci* (2019) 10(21):5495–504. doi: 10.1039/c9sc00961b

79. Sun Q, Li Q, Xie FF. LncRNA-MALAT1 regulates proliferation and apoptosis of ovarian cancer cells by targeting miR-503-5p. *Oncotargets Ther* (2019) 12:6297–307. doi: 10.7150/ijbs.28048

80. Jin Y, Feng SJ, Qiu S, Shao N, Zheng JH. LncRNA MALAT1 promotes proliferation and metastasis in epithelial ovarian cancer via the PI3K-AKT pathway. *Eur Rev Med Pharmacol Sci* (2017) 21(14):3176–84. doi: 10.1186/s13046-019-1394-6

 Qiu JJ, Lin XJ, Tang XY, Zheng TT, Lin YY, Hua KQ. Exosomal metastasisassociated lung adenocarcinoma transcript 1 promotes angiogenesis and predicts poor prognosis in epithelial ovarian cancer. *Int J Biol Sci* (2018) 14(14):1960–73. doi: 10.1016/j.nano.2015.10.012

82. Shang A, Wang W, Gu C, Chen C, Li D. Long non-coding RNA HOTTIP enhances IL-6 expression to potentiate immune escape of ovarian cancer cells by upregulating the expression of PD-L1 in neutrophils. *J Exp Clin Cancer Res* (2019) 38 (1):411. doi: 10.1039/C7FO00882A

83. Kim MS, Haney MJ, Zhao Y, Mahajan V, Deygen I, Klyachko NL, et al. Development of exosome-encapsulated paclitaxel to overcome MDR in cancer cells. *Nanomed-Nanotechnol Biol Med* (2016) 12(3):655–64. doi: 10.1038/s41388-018-0189-0

84. Aqil F, Jeyabalan J, Agrawal AK, Kyakulaga AH, Munagala R, Parker L, et al. Exosomal delivery of berry anthocyanidins for the management of ovarian cancer. *Food Funct* (2017) 8(11):4100–7. doi: 10.3390/cancers13236102

85. Dorayappan KDP, Wanner R, Wallbillich JJ, Saini U, Zingarelli R, Suarez AA, et al. Hypoxia-induced exosomes contribute to a more aggressive and chemoresistant ovarian cancer phenotype: a novel mechanism linking STAT3/Rab proteins. *Oncogene* (2018) 37(28):3806–21. doi: 10.2147/OTT.S262661

86. Naz F, Tariq I, Ali S, Somaida A, Preis E, Bakowsky U. The role of long noncoding RNAs (lncRNAs) in female oriented cancers. *Cancers* (2021) 13(23):6102. doi: 10.1016/j.canlet.2017.03.004

87. Jia JJ, Guo SQ, Zhang D, Tian XH, Xie XM. Exosomal-lncRNA DLEUI accelerates the proliferation, migration, and invasion of endometrial carcinoma cells by regulating microRNA-E2F3. *Oncotargets Ther* (2020) 13:8651–63. doi: 10.1007/s00011-018-1186-z

88. Zhang ZC, Li X, Sun W, Yue SQ, Yang JY, Li JJ, et al. Loss of exosomal miR-320a from cancer-associated fibroblasts contributes to HCC proliferation and metastasis. *Cancer Letters* (2017) 397:33–42. doi: 10.1007/s12253-019-00614-3

89. Wang JL, Xu WQ, He YK, Xia Q, Liu SW. LncRNA MEG3 impacts proliferation, invasion, and migration of ovarian cancer cells through regulating PTEN. *Inflammation Res* (2018) 67(11-12):927–36. doi: 10.1530/JME-12-0008

90. Al-Rugeebah A, Alanazi M, Parine NR. MEG3: an oncogenic long noncoding RNA in different cancers. *Pathol Oncol Res* (2019) 25(3):859–74. doi: 10.18632/ oncotarget.19931

91. Zhou YL, Zhang X, Klibanski A. MEG3 noncoding RNA: a tumor suppressor. J Mol Endocrinol (2012) 48(3):R45–53. doi: 10.1016/j.biopha.2016.02.049

92. He YQ, Luo YH, Liang BY, Ye L, Lu GX, He WM. Potential applications of MEG3 in cancer diagnosis and prognosis. *Oncotarget* (2017) 8(42):73282-95. doi: 10.18632/oncotarget.19931

93. Guo QY, Qian ZD, Yan DD, Li L, Huang LL. LncRNA-MEG3 inhibits cell proliferation of endometrial carcinoma by repressing notch signaling. *Biomed Pharmacother* (2016) 82:589–94. doi: 10.3390/cancers11020234

94. Chen J, Xu YX. Differential expression of serum LncRNA ROR and miR-29 in patients with endometrial carcinoma and their diagnostic value. *Int J Lab Med* (2019) 40(04):484–7. doi: 10.1038/nature22341

95. Dong PX, Xiong Y, Yue JM, Hanley SJB, Kobayashi N, Todo Y, et al. Exploring lncRNA-mediated regulatory networks in endometrial cancer cells and the tumor microenvironment: Advances and challenges. *Cancers* (2019) 11(2):234. doi: 10.3390/ cancers11020234

96. Kamerkar S, LeBleu VS, Sugimoto H, Yang SJ, Ruivo CF, Melo SA, et al. Exosomes facilitate therapeutic targeting of oncogenic KRAS in pancreatic cancer. *Nature* (2017) 546(7659):498. doi: 10.1016/j.semradonc.2016.05.003

97. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. Ca-a Cancer J Clin (2015) 65(2):87-108. doi: 10.3322/caac.21262

98. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer (2015) 136(5):E359–E86. doi: 10.18632/aging.202565

99. Jiangli D, Shannon R, Cordia C. Review of the cervical cancer burden and population-based cervical cancer screening in China. *Asian Pacific J Cancer prevention: APJCP* (2015) 16(17):7401–7. doi: 10.1080/20013078.2020.1722385

100. Verma J, Monk BJ, Wolfson AH. New strategies for multimodality therapy in treating locally advanced cervix cancer. *Semin Radiat Oncol* (2016) 26(4):344–8. doi: 10.1016/j.ijbiomac.2018.02.034

101. He JX, Huang BY, Zhang K, Liu MB, Xu TM. Long noncoding RNA in cervical cancer: From biology to therapeutic opportunity. *Biomed Pharmacother* (2020) 127:110209. doi: 10.1016/j.ygyno.2016.04.216

102. Huang XX, Liu XM, Du B, Liu XL, Xue M, Yan QX, et al. LncRNA LINC01305 promotes cervical cancer progression through KHSRP and exosome-mediated transfer. *Aging-Us* (2021) 13(15):19230–42. doi: 10.1007/s12013-021-01007-7

103. Lin Y, Zhang C, Xiang PP, Shen J, Sun WJ, Yu H. Exosomes derived from HeLa cells break down vascular integrity by triggering endoplasmic reticulum stress in endothelial cells. *J Extracellular Vesicles* (2020) 9(1):1722385. doi: 10.1089/dna.2017.3805

104. Chen SS, Lv MF, Fang S, Ye WX, Gao Y, Xu YS. Poly(I:C) enhanced anticervical cancer immunities induced by dendritic cells-derived exosomes. *Int J Biol Macromol* (2018) 113:1182–7. doi: 10.1016/j.ijbiomac.2018.02.034

105. Kim HJ, Lee JY, Nam EJ, Park SA, Kim S, Kim SW, et al. Upregulation of long noncoding RNA HOXA11 antisense promotes tumor progression and stemness maintenance of cervical cancer cells. *Gynecol Oncol* (2016) 141:75. doi: 10.7150/ jca.28525

106. He YS, Qiu XG. Suppression of lncRNA HOXA11-AS/miR-124 axis inhibits glioma progression. *Cell Biochem Biophys* (2021) 79(4):815–22. doi: 10.2147/ OTT.S167053

107. Cui Y, Yi L, Zhao JZ, Jiang YG. Long noncoding RNA HOXA11-AS functions as miRNA sponge to promote the glioma tumorigenesis through targeting miR-140-5p. DNA Cell Biol (2017) 36(10):822-8. doi: 10.1089/dna.2017.3805

108. Yu J, Hong JF, Kang J, Liao LH, Li CD. Promotion of LncRNA HOXA11-AS on the proliferation of hepatocellular carcinoma by regulating the expression of LATS1. *Eur Rev Med Pharmacol Sci* (2017) 21(15):3402–11.

109. Zhang X, Mao L, Li L, He Z, Wang N, Song Y. Long non-coding RNA GIHCG functions as an oncogene and serves as a serum diagnostic biomarker for cervical cancer. J Cancer (2019) 10(3):672–81. doi: 10.1002/jcla.21990

110. Chen XH, Qu JY. Long noncoding RNA MEG3 suppresses survival, migration, and invasion of cervical cancer. *Oncotargets Ther* (2018) 11:4999–5007. doi: 10.7717/ peerj.4763

111. Tornesello ML, Faraonio R, Buonaguro L, Annunziata C, Starita N, Cerasuolo A, et al. The role of microRNAs, long noncoding RNAs, and circular RNAs in cervical cancer. *Front Oncol* (2020) 10. doi: 10.3389/fonc.2020.00150

112. Chen WT, Huang LS, Zeng DD, Chen ZP, Wu ZX. Effect and mechanism of LncRNA HOTAIR on proliferation, apoptosis and EMT of cervical cancer hela cells. *Int J Lab Med* (2022) 43(06):710–6. doi: 10.3390/ijms18030538

113. Zhang J, Liu SC, Luo XH, Tao GX, Guan M, Yuan H, et al. Exosomal long noncoding RNAs are differentially expressed in the cervicovaginal lavage samples of cervical cancer patients. *J Clin Lab Analysis* (2016) 30(6):1116–21. doi: 10.3322/caac.21262

114. Abak A, Abhari A, Rahimzadeh S. Exosomes in cancer: small vesicular transporters for cancer progression and metastasis, biomarkers in cancer therapeutics. *Peerj* (2018) 6:e4763. doi: 10.1002/ijc.29210

Glossary

lncRNAs	Long non-coding RNAs
TDE	Tumor-derived exosomes
EMT	Epithelial-mesenchymal transition
ESCRT	Endosomal sorting complex required for transport
ALIX	Apoptosis-linked gene 2-interacting protein X
VPS4	Vacuolar protein sorting 4
DEX	Dendritic cell-derived exosomes
AS lncRNA	Antisense lncRNA
MEG3	Maternally expressed gene 3
GRE	Glucocorticoid response element
GR	Glucocorticoid receptor
PANDA	P21-associated noncoding RNA DNA damage-activated
HOX	Homeobox
HOXA11-AS	A11 antisense lncRNA
WDR5	WD repeat domain 5
WNT	Wingless-type MMTV Integration Site Family
KLF4	Kruppel like factor 4
VEGF	Vascular endothelial growth factor
ceRNA	Competitive endogenous RNA
ATB	Activated by transforming growth factor beta
HOTAIR	HOX transcript antisense RNA
MALAT1	Metastasis-associated lung adenocarcinoma transcript 1
DDX3	DEAD-box RNA helicase 3
VAMP3	Vesicle associated membrane protein 3
SNAP23	Synaptosomal-associated protein 23
TAMs	Tumor-associated macrophages
ECM	Extracellular matrix
CAFs	Carcinoma-associated fibroblasts
C-X-C motif	Chemokine
CXCL14	ligand 14
SNHG16	Small nucleolar RNA host gene 16
GALNT1	N-acetylgalactosaminyltransferase 1
HIF-1α	Hypoxia-inducible factor
HNRNPK	Heterogeneous nuclear ribonucleoprotein K
EGFR	Epidermal growth factor receptor
DCs	Dendritic cells
PI3K	Phosphoinositide 3 kinase
ROR	Regulators of Reprogramming

	(Constituted)
(<i>Continuea</i>)

Frontiers	in	Oncol	loav
1 I OIIGICI J		01100	cogy.

Continued

OC	Ovarian Cancer
FDA	Food and Drug Administration
FRα	Folate Receptor-a
EpCAM	Epithelial cell adhesion molecule
JAK2	Janus kinase 2
РКВ	AKT, PI3K-protein kinase B
FIGO	The International Federation of Gynecology and Obstetrics
HOTTIP	HOXA transcript at the distal tip
FGF	Fibroblast growth factor
MDCKMDR1	P-gp transfected Manin-Darby canine kidney epithelial cells
HEX	Hypoxic OC cell line exosome
EC	Endometrial Carcinoma
CC	Cervical Cancer
OS	Overall Survival
PFS	Progression-free Survival
SOX4	SRY-related high-mobility-group box 4
GIHCG	Gradual increase during hepatocarcinogenesis
CIN	Cervical intraepithelial neoplasia
NKILA	NF-kappaB interacting lncRNA
ARAP1-AS1	ArfGAP with RhoGAP domain, ankyrin repeat, and PH domain 1 antisense RNA
SFTA1P	Surfactant associated 1, pseudogene
TPM4	Tropomyosin 4
PTBP1	Polypyrimidine Tract Binding Protein 1.